

Supporting Information

Nickel-Catalyzed Hydrogenolysis and Conjugate Addition Of 2-Hydroxymethylpyridines via Organozinc Intermediates

Luke E. Hanna, Michael R. Harris, Kenji Domon, Elizabeth R. Jarvo*

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I. General Procedures

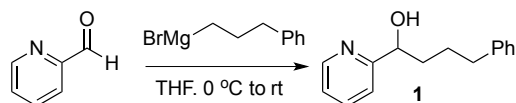
All reactions were carried out under an atmosphere of N₂, or Ar when noted. All glassware was oven- or flame-dried prior to use. Tetrahydrofuran (THF), diethyl ether (Et₂O), dichloromethane (CH₂Cl₂), and toluene (PhMe) were degassed with Ar and then passed through two 4 x 36 inch columns of anhydrous neutral A-2 alumina (8 x 14 mesh; LaRoche Chemicals; activated under a flow of argon at 350 °C for 12 h) to remove H₂O. All other solvents utilized were purchased “anhydrous” commercially, or purified as described. ¹H NMR spectra were recorded on Bruker DRX-400 (400 MHz ¹H, 100 MHz ¹³C, 376.5 MHz ¹⁹F), GN-500 (500 MHz ¹H, 125.7 MHz ¹³C), or CRYO-500 (500 MHz ¹H, 125.7 MHz ¹³C) spectrometers. Proton chemical shifts are reported in ppm (δ) relative to internal tetramethylsilane (TMS, δ 0.00). Data are reported as follows: chemical shift (multiplicity [singlet (s), broad singlet (br s), doublet (d), doublet of doublets (dd), triplet (t), doublet of triplets (dt), doublet of doublet of triplets (ddt), triplet of triplets (tt), quartet (q), quintet (quin), apparent doublet (ad), apparent triplet (at), multiplet (m)], coupling constants [Hz], integration). Carbon chemical shifts are reported in ppm (δ) relative to TMS with the respective solvent resonance as the internal standard (CDCl₃, δ 77.16 ppm). Unless otherwise indicated, NMR data were collected at 25 °C. Infrared (IR) spectra were obtained on a Thermo Scientific Nicolet iS5 spectrometer with an iD5 ATR tip (neat) and are reported in terms of frequency of absorption (cm⁻¹). Analytical thin-layer chromatography (TLC) was performed using Silica Gel 60 F₂₅₄ precoated plates (0.25 mm thickness). Visualization was accomplished by irradiation with a UV lamp and/or staining with KMnO₄, ceric ammonium molybdate (CAM), or *p*-anisaldehyde (PAA) solutions. Flash chromatography was performed using Silica Gel 60 (170-400 mesh) from Fisher Scientific. Melting points (m.p.) were obtained using a Mel-Temp melting point apparatus and are uncorrected. High resolution mass spectrometry was performed by the University of California, Irvine Mass Spectrometry Center.

[1,2-Bis(diphenylphosphino)ethane]dichloronickel(II) was purchased from Strem, stored in a glovebox under an atmosphere of N₂, and used as received. Diethylzinc (ZnEt₂) and diethyl chlorophosphate were purchased from Sigma and used as received. 1-isoquinolinecarboxaldehyde was prepared from selenium (IV) oxide oxidation of 1-methylisoquinoline by a procedure reported by Long.¹ All other reagents were purchased commercially and used as received.

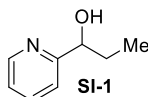
¹ Cao, B.; Wang, Y.; Ding, K.; Neamati, N.; Long, Y-Q. *Org. Biomol. Chem.* **2012**, *10*, 1239.

II. Synthesis and Characterization of Benzylic Alcohols

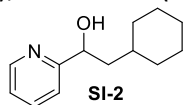
General Procedure A. Grignard addition to aldehydes.



1. In a flame-dried round-bottom flask, a solution of 2-pyridinecarboxaldehyde (1.43 mL, 15.0 mmol, 1.00 equiv) in THF (30 mL) was cooled to 0 °C and 3-(phenylpropyl)magnesium bromide (1.6 M in THF, 10 mL, 16 mmol, 1.1 equiv) was added. The reaction mixture was stirred at room temperature for 1 h, then cooled to 0 °C and saturated ammonium chloride (25 mL) was added. The reaction was extracted with EtOAc (3 x 25 mL). The combined organic layers were washed with brine (1 x 40 mL), dried over Na₂SO₄, and concentrated in vacuo. The product was purified by column flash chromatography (25–40% EtOAc/hexanes) to afford the title compound as a yellow solid (1.70 g, 7.50 mmol, 50%). **TLC** *R*_f = 0.16 (20% EtOAc/hexanes, UV active); **m.p.** = 58–60 °C; **¹H NMR** (500 MHz, CDCl₃) δ 8.53 (d, *J* = 4.8 Hz, 1H), 7.66 (td, *J* = 7.8, 1.8 Hz, 1H), 7.29–7.23 (m, 2H), 7.22–7.13 (m, 5H), 4.75 (s, 1H), 4.19 (s, 1H), 2.71–2.59 (m, 2H), 1.91–1.82 (m, 1H), 1.80–1.67 (m, 3H); **¹³C NMR** (125 MHz, CDCl₃) δ 161.9, 148.2, 142.4, 136.7, 128.5, 128.3, 125.8, 122.3, 120.4, 72.5, 38.2, 35.8, 26.9; **IR** (neat) 3192, 2925, 2861, 1594, 1492, 1452 cm⁻¹; **HRMS** (TOF MS ES+) *m/z* calcd for C₁₅H₁₇NONa (M + Na)⁺ 227.1310, found 227.1315.



SI-1. Prepared according to a procedure reported by Braga², the following amounts of reagents were used: 2-pyridinecarboxaldehyde (0.38 mL, 4.0 mmol, 1.0 equiv), ethylmagnesium bromide (2.4 M in Et₂O, 2.0 mL, 4.8 mmol, 1.2 equiv), and THF (15 mL). Analytical data is consistent with literature values.³ **¹H NMR** (400 MHz, CDCl₃) δ 8.55 (dt, *J* = 4.9, 1.3 Hz, 1H), 7.68 (td, *J* = 7.6, 1.8 Hz, 1H), 7.25 (dd, *J* = 7.8 Hz, 0.5 Hz, 1H), 7.20 (ddd, *J* = 7.6, 4.9, 0.5 Hz, 1H), 4.73–4.66 (m, 1H), 4.17 (d, *J* = 5.5 Hz, 1H), 1.95–1.84 (m, 1H), 1.78–1.66 (m, 1H), 0.95 (t, *J* = 7.3 Hz, 3H).

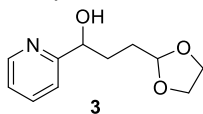


SI-2. Using General Procedure A, outlined above, the following amounts of reagents were used: 2-pyridinecarboxaldehyde (0.48 mL, 5.0 mmol, 1.0 equiv), (cyclohexylmethyl)magnesium bromide (0.56 M in THF, 9.8 mL, 11 mmol, 1.1 equiv), and THF (10 mL). The product was purified by column flash chromatography (20% EtOAc/hexanes) to afford the title compound as a yellow solid (0.453 g, 2.20 mmol, 44%). **TLC** *R*_f = 0.3 (20% EtOAc/hexanes, UV active); **¹H NMR** (500 MHz, CDCl₃) δ 8.51 (d, *J* = 4.7 Hz, 1H), 7.66 (t, *J* = 7.7 Hz, 1H), 7.26 (d, *J* = 7.7 Hz, 1H), 7.19 (t, *J* = 6.0 Hz, 1H), 4.82 (t, *J* = 6.3, 1H), 4.28 (br s, 1H), 1.94 (d, *J* = 12.7 Hz, 1H), 1.75–1.54 (m, 7H) 1.32–1.11 (m, 3H), 1.02–0.90 (m, 2H); **¹³C NMR** (125 MHz, CDCl₃) δ 163.4, 148.3, 136.7, 122.2,

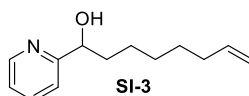
² Braga, A. L.; Paixao, M. W.; Ludtke, D. S.; Silveira, C. C.; Rodrigues, O. E. D. *Org. Lett.* **2003**, 5, 2365.

³ Moody, C. J.; Morfitt, C. N. *Synthesis* **1998**, 7, 1039.

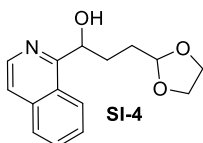
120.4, 70.7, 46.9, 34.4, 34.2, 32.7, 26.7, 26.5, 26.2; **IR** (neat) 3217, 2919, 1594 cm^{-1} ; **HRMS** (TOF MS ES+) m/z calcd for $\text{C}_{13}\text{H}_{19}\text{NONa}$ ($\text{M} + \text{Na}$)⁺ 228.1364, found 228.1364.



3. Using General Procedure A, outlined above, the following amounts of reagents were used: 2-pyridinecarboxaldehyde (0.95 mL, 10 mmol, 1.0 equiv), hept-6-en-1-ylmagnesium bromide (1.2 M in THF, 9.2 mL, 11 mmol, 1.1 equiv), and THF (10 mL). The product was purified by column flash chromatography (25–40% EtOAc/hexanes) to afford the title compound as a yellow oil (1.31 g, 6.38 mmol, 64%). Analytical data is consistent with literature values.⁴ **TLC** R_f = 0.1 (50% EtOAc/hexanes, UV active); **¹H NMR** (500 MHz, CDCl_3) δ 8.55–8.50 (m, 1H), 7.67 (td, J = 7.8, 1.8 Hz, 1H), 7.29 (d, J = 7.8 Hz, 1H), 7.21–7.15 (m, 1H), 4.93–4.88 (m, 1H), 4.83–4.75 (m, 1H), 4.33 (d, J = 5.4 Hz, 1H), 3.99–3.80 (m, 4H), 2.07–1.95 (m, 1H), 1.87–1.74 (m, 3H); **¹³C NMR** (125 MHz, CDCl_3) δ 161.9, 148.3, 136.7, 122.3, 120.4, 104.4, 72.4, 64.9, 32.5, 29.5.



SI-3. Using General Procedure A, outlined above, the following amounts of reagents were used: 2-pyridinecarboxaldehyde (0.95 mL, 10 mmol, 1.0 equiv), hept-6-en-1-ylmagnesium bromide (1.2 M in THF, 9.2 mL, 11 mmol, 1.1 equiv), and THF (10 mL). The product was purified by column flash chromatography (25–40% EtOAc/hexanes) to afford the title compound as a red oil (1.31 g, 6.38 mmol, 64%). **TLC** R_f = 0.4 (40% EtOAc/hexanes, UV active); **¹H NMR** (500 MHz, CDCl_3) δ 8.53 (d, J = 4.7 Hz 1H), 7.67 (t, J = 7.6 Hz, 1H), 7.26 (d, J = 7.6 Hz, 1H), 7.19 (t, J = 5.5 Hz, 1H), 5.79 (ddt, J = 17.2, 10.2, 6.9 Hz, 1H), 4.98 (d, J = 17.2 Hz, 1H), 4.92 (d, J = 10.2 Hz, 1H), 4.73 (dd, J = 7.7, 4.4 Hz, 1H), 4.25 (br s, 1H), 2.03 (dd, J = 10.2, 6.9 Hz, 2H), 1.87–1.77 (m, 1H) 1.74–1.65 (m, 1H), 1.50–1.24 (m, 6H); **¹³C NMR** (125 MHz, CDCl_3) δ 162.4, 148.3, 139.2, 136.7, 122.3, 120.4, 114.3, 72.9, 38.7, 33.8, 29.2, 28.9, 25.2; **IR** (neat) 3260, 2926, 1594 cm^{-1} ; **HRMS** (TOF MS ES+) m/z calcd for $\text{C}_{13}\text{H}_{19}\text{ONNa}$ ($\text{M} + \text{Na}$)⁺ 228.1364, found 228.1364.

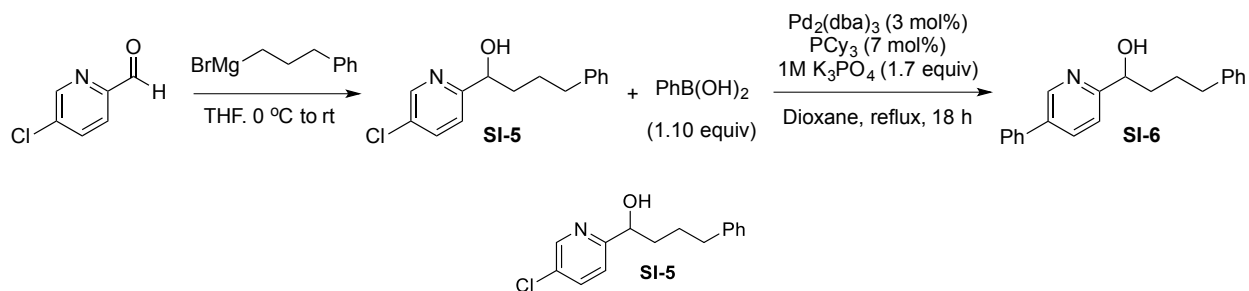


SI-4. Using General Procedure A, outlined above, the following amounts of reagents were used: 1-Isoquinolinecarboxaldehyde (0.47 g, 3.0 mmol, 1.0 equiv), (2-(1,3-dioxolan-2-yl)ethyl)magnesium bromide (0.95 M in THF, 3.5 mL, 3.3 mmol, 1.1 equiv), and THF (10 mL). The product was purified by column flash chromatography (40% EtOAc/hexanes) to afford the title compound as a tan solid (249 mg, 0.960 mmol, 32%). **TLC** R_f = 0.2 (40% EtOAc/hexanes, UV active); **m.p.** = 53–55 °C; **¹H NMR** (500 MHz, CDCl_3) δ 8.43 (d, J = 5.8 Hz, 1H), 8.07 (d, J = 8.4 Hz, 1H), 7.85 (d, J = 8.4 Hz, 1H), 7.69 (t, J = 7.7 Hz, 1H), 7.61 (t, J = 7.7 Hz, 1H), 7.58 (d, J = 5.8 Hz, 1H), 5.52 (d, J = 7.1 Hz, 1H), 5.18 (br s, 1H), 4.93 (t, J = 4.8 Hz, 1H), 4.00–3.89 (m, 2H), 3.87–3.78

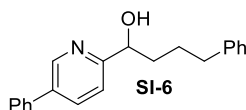
⁴ Gebert, A.; Barth, M.; Linden, A.; Widmer, U.; Heimgartner, H. *Helv. Chim. Acta* **2012**, 95, 737.

(m, 2H) 2.12–2.02 (m, 1H), 2.06–1.97 (m, 1H), 1.90–1.81 (m, 1H), 1.80–1.72 (m, 1H); ^{13}C NMR (125 MHz, CDCl_3) δ 161.1, 140.4, 136.5, 130.3, 126.6, 126.5, 124.9, 124.4, 120.6, 104.5, 69.2, 65.0, 64.9, 33.3, 29.6; IR (neat) 3390, 2957, 2882 cm^{-1} ; HRMS (TOF MS ES+) m/z calcd for $\text{C}_{15}\text{H}_{17}\text{O}_3\text{NNa}$ ($M + \text{Na}$) $^+$ 282.1106, found 282.1104.

i. Synthesis of Alcohol Precursor of 11 (Scheme 3)



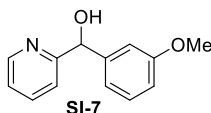
SI-5. Using General Procedure A, outlined above, the following amounts of reagents were used: 5-chloro-2-formylpyridine (0.85 g, 6.0 mmol, 1.0 equiv), (3-phenylpropyl)magnesium bromide (1.6 M in THF, 4.1 mL, 6.6 mmol, 1.1 equiv), and THF (20 mL). The product was purified by column flash chromatography (20% EtOAc/hexanes) to afford the title compound as a red oil (0.88 g, 3.4 mmol, 56%). TLC R_f = 0.3 (20% EtOAc/hexanes, UV active); ^1H NMR (500 MHz, CDCl_3) δ 8.47 (d, J = 1.7 Hz, 1H), 7.62 (dd, J = 8.4, 2.3 Hz, 1H), 7.26 (t, J = 7.5 Hz, 2H), 7.21–7.12 (m, 4H), 4.76–4.71 (m, 1H), 3.80 (br s, 1H), 2.72–2.58 (m, 2H), 1.87–1.78 (m, 1H), 1.77–1.68 (m, 3H); ^{13}C NMR (125 MHz, CDCl_3) δ 160.6, 147.3, 142.2, 136.6, 130.6, 128.5, 128.4, 125.9, 121.2, 72.7, 38.0, 35.8, 27.0; IR (neat) 3359, 2936, 2859 cm^{-1} ; HRMS (TOF MS ES+) m/z calcd for $\text{C}_{15}\text{H}_{16}\text{ClN}$ ($M + \text{Na}$) $^+$ 284.0818, found 284.0810.



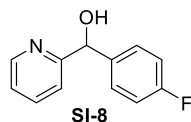
SI-6. The product was prepared according to a modified procedure by Fu.⁵ Tris(dibenzylideneacetone)dipalladium (69 mg, 0.076 mmol, 3 mol %) and tricyclohexylphosphine (50 mg, 0.18 mmol, 7 mol %) were charged into a flame round bottom flask inside a glovebox. The flask was fitted with a septum, removed from the glovebox, and phenylboronic acid (0.337 g, 2.77 mmol, 1.10 equiv), **4.22** (0.66 g, 2.5 mmol, 1.0 equiv), aqueous potassium phosphate (1.3 M in H_2O , 3.3 mL, 4.3 mmol, 1.7 equiv) and dioxane (13 mL) were added. The reaction flask was fitted with a reflux condenser and heated to 95 $^\circ\text{C}$ for 18 h. After cooling, the solvent was removed under reduced pressure. The resultant residue was purified by flash column chromatography (30% EtOAc/hexane) to afford the title compound as a pale yellow solid (0.488 g, 1.60 mmol, 64%). TLC R_f = 0.4 (40% EtOAc/hexanes, UV active); $m.p.$ = 99–101 $^\circ\text{C}$; ^1H NMR (500 MHz, CDCl_3) δ 8.75 (d, J = 2.0 Hz, 1H), 7.85 (dd, J = 8.0, 2.0 Hz, 1H), 7.57 (d, J = 7.2 Hz, 2H), 7.48 (t, J = 7.2 Hz, 2H), 7.40 (t, J = 7.2 Hz, 1H), 7.31–7.23 (m, 3H), 7.21–7.14 (m, 3H), 7.85 (m, 1H), 4.15 (d, J = 5.1 Hz, 1H), 2.73–2.61 (m, 2H), 1.96–1.86 (m, 1H), 1.84–1.73 (m, 3H); ^{13}C NMR (125 MHz, CDCl_3) δ 161.0, 146.7, 142.4, 137.7, 135.5, 135.3, 129.2,

⁵ Kudo, N.; Perseghini, M.; Fu, G. C. *Angew. Chem. Int. Ed.* **2006**, *45*, 1282.

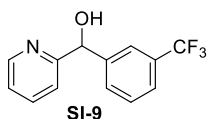
128.6, 128.4, 128.2, 127.2, 125.8, 120.3, 72.6, 38.2, 35.9, 27.1; **IR** (neat) 3286, 2946, 2914 cm^{-1} ; **HRMS** (TOF MS ES+) m/z calcd for $\text{C}_{21}\text{H}_{21}\text{NONa}$ ($M + \text{Na}$)⁺ 326.1521, found 326.1516.



SI-7. Using General Procedure A, outlined above, the following amounts of reagents were used: 2-pyridinecarboxaldehyde (0.95 mL, 10 mmol, 1.0 equiv), (3-methoxyphenyl)magnesium bromide (0.92 M in THF, 13 mL, 12 mmol, 1.2 equiv), and THF (20 mL). The product was purified by column flash chromatography (20–40% EtOAc/hexanes) to afford the title compound as a yellow oil (0.62 g, 3.1 mmol, 62%). Analytical data is consistent with literature values.⁶ **TLC** R_f = 0.2 (20% EtOAc/hexanes, UV active); **¹H NMR** (500 MHz, CDCl_3) δ 8.56 (d, J = 4.4 Hz, 1H), 7.62 (td, J = 7.5, 1.3 Hz, 1H), 7.29–7.14 (m, 3H), 6.96 (d, J = 6.7 Hz, 1H), 6.93 (s, 1H), 6.81 (d, J = 7.2 Hz, 1H), 5.27 (d, J = 3.6 Hz, 1H), 5.29 (d, J = 4.3 Hz, 1H); **¹³C NMR** (125 MHz, CDCl_3) δ 160.6, 159.8, 147.8, 144.8, 136.9, 129.6, 122.5, 121.4, 119.5, 113.5, 112.5, 74.9, 55.3.



SI-8. Using General Procedure A, outlined above, the following amounts of reagents were used: 2-pyridinecarboxaldehyde (0.36 mL, 3.8 mmol, 1.1 equiv), (4-fluorophenyl)magnesium bromide (0.27 M in THF, 15 mL, 4.0 mmol, 1.2 equiv), and THF (20 mL). The product was purified by column flash chromatography (20–40% EtOAc/hexanes) to afford the title compound as a yellow oil (0.60 g, 3.0 mmol, 78%). Analytical data is consistent with literature values.⁶ **TLC** R_f = 0.2 (20% EtOAc/hexanes, UV active); **¹H NMR** (500 MHz, CDCl_3) δ 8.56 (d, J = 4.7 Hz, 1H), 7.62 (dt, J = 7.7, 1.8 Hz, 1H), 7.36–7.30 (m, 2H), 7.23–7.17 (m, 1H), 7.11 (d, J = 7.9 Hz, 1H), 7.05–6.97 (m, 2H), 5.73 (s, 1H), 5.31 (s, 1H); **¹³C NMR** (125 MHz, CDCl_3) δ 163.4 (d, J_{CF} = 245.0 Hz), 160.6, 147.9, 139.1 (d, J_{CF} = 2.77 Hz), 137.0, 128.9 (d, J_{CF} = 8.3 Hz), 122.6 (d, J_{CF} = 161.8 Hz), 115.6 (d, J_{CF} = 21.5 Hz), 74.31.



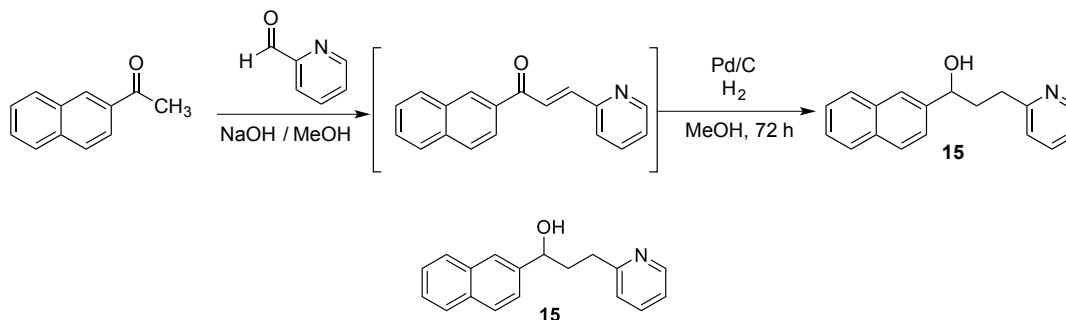
SI-9. Using General Procedure A, outlined above, the following amounts of reagents were used: 2-pyridinecarboxaldehyde (0.95 mL, 10 mmol, 1.0 equiv), (3-(trifluoromethyl)phenyl)magnesium bromide (0.50 M in THF, 20 mL, 10 mmol, 1.2 equiv), and THF (20 mL). The product was purified by column flash chromatography (20–40% EtOAc/hexanes) to afford the title compound as a yellow oil (0.60 g, 3.0 mmol, 78%). Analytical data is consistent with literature values.⁷ **TLC** R_f = 0.2 (20% EtOAc/hexanes, UV active); **¹H NMR** (500 MHz, CDCl_3) δ 8.57 (d, J = 5.0 Hz, 1H), 7.69–7.62 (m, 2H), 7.57 (d, J = 7.8 Hz, 1H), 7.53 (d, J = 7.6 Hz, 1H), 7.45 (t, J = 8.0 Hz, 1H), 7.25–7.20 (m, 1H), 7.14 (d, J = 8.0 Hz, 1H), 5.80 (s, 1H), 5.44

⁶ Kamitani, M.; Ito, M.; Itazaki, M.; Nakazawa, H. *Chem. Commun.* **2014**, 50, 7941.

⁷ Agai, B.; Prosenyak, A.; Tarkanyi, G.; Vida, L.; Faigl, F. *Eur. J. Org. Chem.* **2004**, 3623.

(s, 1H); ^{13}C NMR (125 MHz, CDCl_3) δ 159.9, 148.0, 144.2, 137.1, 130.8 (q J = 32.4 Hz), 130.4, 129.0, 124.7 (q, J = 4.16 Hz), 124.0 (q, J = 270 Hz), 123.8 (q, J = 3.70 Hz), 122.8, 121.2, 74.4.

ii. Synthesis of Substrate 15 for Scheme 4



15. The product was prepared according to modified procedures by Attar⁸ and Hirota.⁹ To a 100 mL round bottomed flask equipped with a stir bar was added 2-acetonaphthone (6.81 g, 40.0 mmol, 1.00 equiv), MeOH (10 mL), and 10% NaOH (20 mL). The mixture was stirred for 15 min at room temperature before addition of 2-pyridinecarboxaldehyde (6.85 mL, 72.0 mmol, 1.80 equiv). The reaction was stirred until judged complete by TLC analysis (2 h). Ice water was added to the reaction and the solid was filtered, washed with Et₂O, and dried in vacuo. The unpurified solid enone was then redissolved in MeOH (40 mL) and transferred to a round bottom flask containing a stir bar and 10% Pd/C (0.68 g, 10% by weight relative to 2-acetonaphthone). The flask was flushed with nitrogen and then equipped with a balloon of hydrogen gas. The reaction was stirred under an atmosphere of hydrogen for 72 h. The reaction mixture was flushed with N₂, then filtered through celite, washing with MeOH. The filtrate was poured into sat. aqueous NaHCO₃ (60 mL) and extracted with EtOAc (3 x 50 mL). The combined organics were washed with brine, dried with Na₂SO₄, filtered and concentrated in vacuo. The residue was purified by flash column chromatography (10–30% EtOAc/hexanes) to afford alcohol **15** as a white solid (1.63 g, 6.20 mmol, 16%). **TLC** R_f = 0.2 (20% EtOAc/hexanes, UV active); **m.p.** = 76–77 °C; ^1H NMR (500 MHz, CDCl_3) δ 8.46 (d, J = 4.8 Hz, 1H), 7.84 (s, 1H), 7.55 (td, J = 7.5, 1.3 Hz, 1H), 7.48 (d, J = 8.2 Hz, 1H), 7.46–7.39 (m, 2H), 7.13–7.05 (m, 2H), 5.98 (br s, 1H), 4.96 (dd, J = 7.8, 4.5 Hz, 1H), 2.97 (t, J = 6.6 Hz, 2H), 2.31–2.17 (m, 2H); ^{13}C NMR (125 MHz, CDCl_3) δ 161.4, 148.6, 142.9, 136.9, 133.4, 132.8, 128.0, 127.7, 126.0, 125.5, 124.2, 123.3, 121.3, 73.7, 38.0, 34.5; **IR** (neat) 3250, 3054, 2919 cm^{-1} ; **HRMS** (TOF MS ES+) m/z calcd for $\text{C}_{18}\text{H}_{17}\text{ONNa}$ ($M + \text{Na}$)⁺ 286.1208, found 286.1199.

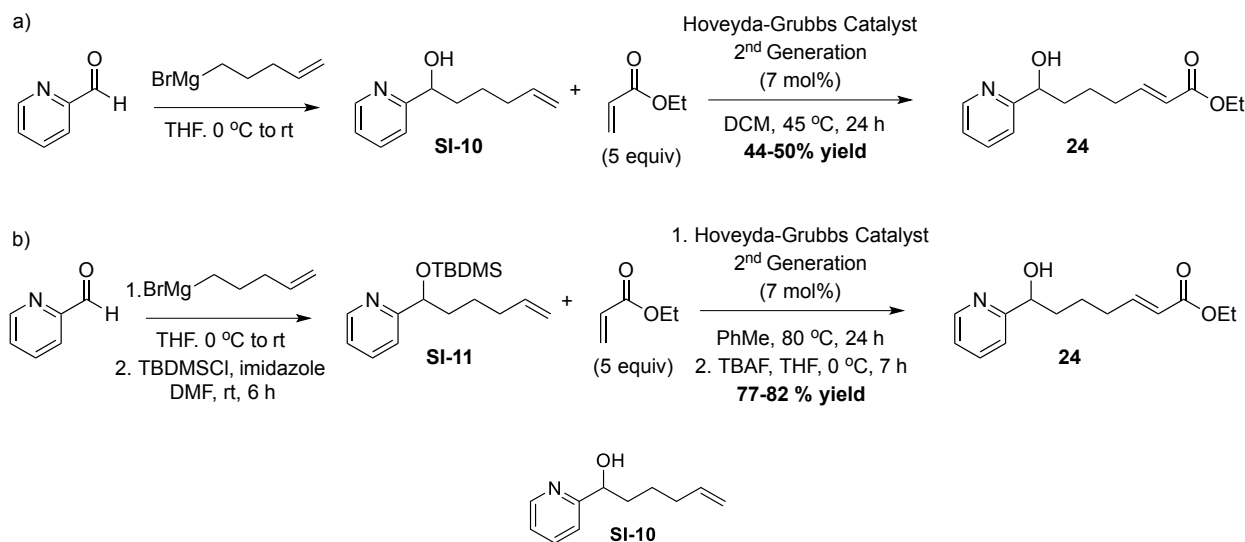
⁸ Attar, S.; O'Brien, Z.; Alhaddad, H.; Golden, M. L.; Calderon-Urrea, A. *Bioorg. Med. Chem.* **2011**, *19*, 2055.

⁹ Hattori, K.; Sajiki, H.; Hirota, K. *Tetrahedron* **2001**, *57*, 4817.

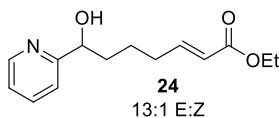
iii. Synthesis of Substrates **24** and **26** for Scheme 5

Compounds **24** and **26** were prepared by cross-metathesis of the corresponding alcohols with ethyl acrylate. In general, yields in the cross-metathesis step were higher and more reproducible when the alcohol was protected as the TBDMS ether (cf Scheme SI-1a and b).

Scheme SI-1 Synthesis of **24**.



SI-10. Using General Procedure A, outlined above, the following amounts of reagents were used: 2-pyridinecarboxaldehyde (0.95 mL, 10 mmol, 1.0 equiv), pent-4-en-1-ylmagnesium bromide (1.1 M in THF, 10 mL, 11 mmol, 1.1 equiv), and THF (15 mL). The product was purified by column flash chromatography (20–40% EtOAc/hexanes) to afford the title compound as a yellow oil (0.784 g, 4.40 mmol, 44%). **TLC** R_f = 0.4 (40% EtOAc/hexanes, UV active); **$^1\text{H NMR}$** (500 MHz, CDCl_3) δ 8.53 (d, J = 4.5 Hz, 1H), 7.68 (td, J = 7.6, 1.5 Hz, 1H), 7.25 (d, J = 8.1 Hz, 1H), 7.19 (dd, J = 6.9, 5.1 Hz, 1H), 5.79 (ddt, J = 17.1, 10.3, 6.7 Hz, 1H), 4.99 (dd, J = 17.1, 1.6 Hz, 1H), 4.94 (d, J = 10.3 Hz, 1H), 4.80–4.78 (m, 1H), 4.25 (s, 1H), 2.16–2.02 (m, 2H), 1.89–1.78 (m, 1H), 1.74–1.62 (m, 1H), 1.58–1.47 (m, 2H); **$^{13}\text{C NMR}$** (125 MHz, CDCl_3) δ 162.3, 148.3, 138.8, 136.8, 122.4, 120.4, 114.7, 72.7, 38.1, 33.8, 24.6; **IR** (neat) 3260, 2936, 1594 cm^{-1} ; **HRMS** (TOF MS ES+) m/z calcd for $\text{C}_{11}\text{H}_{15}\text{ONa}$ ($M + \text{Na}$)⁺ 200.1051, found 200.1056.

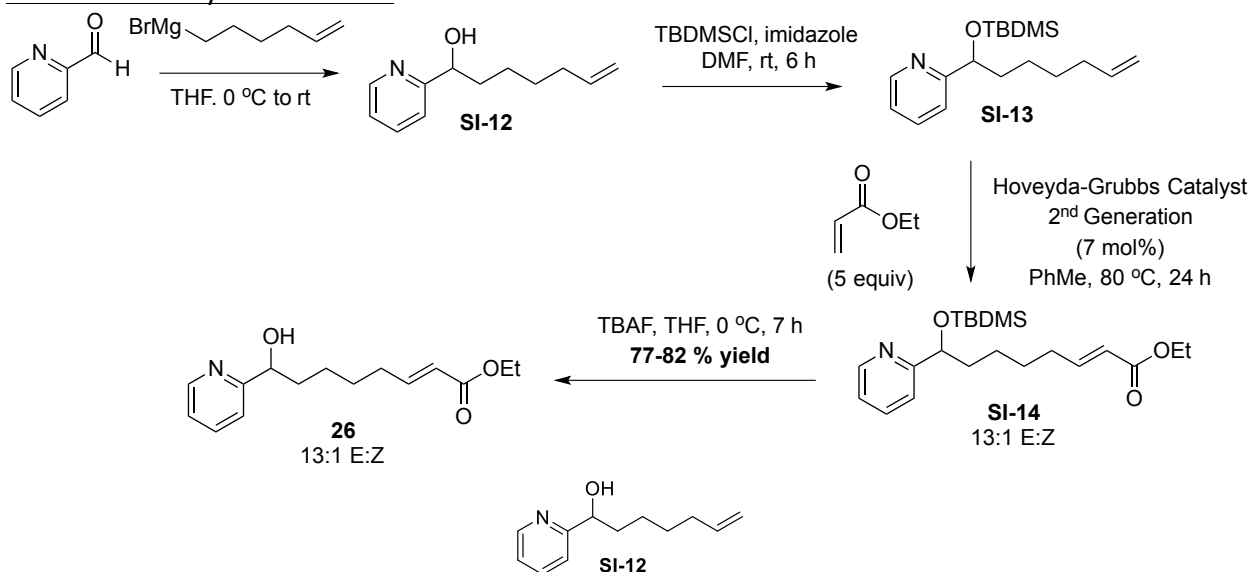


24. The title compound was prepared according to a modified procedure reported by Grubbs.¹⁰ In a glovebox, a flame-dried bomb flask was charged with a stir bar, **SI-10** (0.71 g, 4.0 mmol, 1.0 equiv), and Hoveyda-Grubbs Catalyst 2nd Generation (178 mg, 0.280 mmol, 0.0700 equiv). The flask was removed from the glovebox, and anhydrous CH_2Cl_2 (50 mL) and ethyl acrylate (2.2 mL,

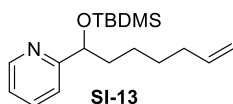
¹⁰ Chatterjee, A. K.; Choi, T.-L.; Sanders, D. P.; Grubbs, R. H. *J. Am. Chem. Soc.* **2003**, 125, 11360.

20 mmol, 5.0 equiv) were added. The flask was sealed and heated to reflux over 24 h. The flask was then cooled to ambient temperature, and the solvent was removed in vacuo. The residue was purified by flash column chromatography to afford the title compound as a pale yellow oil (125 mg, 0.501 mmol, 13%, 13:1 E:Z). **TLC** R_f = 0.1 (40% EtOAc/hexanes, UV active); **^1H NMR** (500 MHz, CDCl_3) δ 8.53 (d, J = 4.4 Hz, 1H), 7.68 (td, J = 7.6, 1.5 Hz, 1H), 7.26 (d, J = 7.7 Hz, 1H), 7.20 (dd, J = 6.9, 5. Hz, 1H), 6.93 (dt, J = 15.5, 7.1 Hz, 1H), 5.80 (dt, J = 15.5, 1.5 Hz, 1H), 4.75 (dd, J = 7.2, 4.3 Hz, 1H), 4.33 (br s, 1H), 4.16 (q, J = 7.2 Hz, 2H), 2.28–2.18 (m, 2H), 1.90–1.81 (m, 1H), 1.75–1.66 (m, 1H), 1.64–1.55 (m, 2H), 1.28 (t, J = 7.2 Hz, 3H); **^{13}C NMR** (125 MHz, CDCl_3) δ 166.8, 162.0, 148.9, 148.3, 136.8, 122.4, 121.7, 120.4, 72.5, 60.2, 38.0, 32.1, 23.7, 14.3; **IR** (neat) 2938, 1730 cm^{-1} ; **HRMS** (TOF MS ES+) m/z calcd for $\text{C}_{14}\text{H}_{19}\text{NO}_3\text{Na}$ ($M + \text{Na}$) $^+$ 272.1263, found 272.1271.

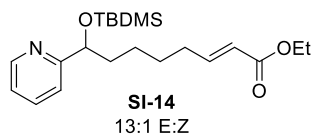
Scheme SI-2. Synthesis of 26.



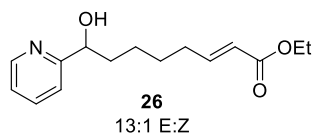
SI-12. Using General Procedure A, outlined above, the following amounts of reagents were used: 2-pyridinecarboxaldehyde (4.3 mL, 45 mmol, 1.0 equiv), hex-5-en-1-ylmagnesium bromide (1.1 M in THF, 50 mL, 50 mmol, 1.1 equiv), and THF (60 mL). The product was purified by column flash chromatography (20–40% EtOAc/hexanes) to afford the title compound as a yellow oil (4.4 g, 23.6 mmol, 53%).



SI-13. Following the synthetic route outlined in Scheme SI-2, **SI-12** (2.0 g, 10 mmol, 1.0 equiv) was dissolved in DMF (20 mL) with TBDMSCl (12 mmol, 1.8 g, 1.2 equiv) and stirred at room temperature for 6 h. The crude reaction mixture was diluted with EtOAc (60 mL), washed with water (3x100 mL), then brine (3x100 mL) to remove DMF, dried over MgSO_4 , and the solvent was removed to afford the product as a pale yellow oil (2.8 g, 9.3 mmol, 93%). The compound was taken forward without further purification.



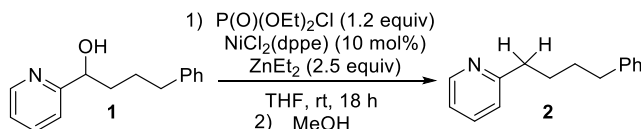
SI-14. The title compound was prepared according to a modified procedure reported by Grubbs.⁹ In a glovebox, a flame-dried bomb flask was charged with a stir bar, **SI-13** (1.5 g, 5.0 mmol, 1.0 equiv), and Hoveyda-Grubbs Catalyst 2nd Generation (350 mg, 0.560 mmol, 0.7 mol %). The flask was removed from the glovebox, and anhydrous PhMe (50 mL) and ethyl acrylate (2.7 mL, 25 mmol, 5.0 equiv) were added. The flask was sealed and heated to reflux over 24 h. The flask was then cooled to ambient temperature, and the solvent was removed in vacuo. The residue was purified by flash column chromatography (10-20% EtOAc/Hex) to afford the title compound as a pale yellow oil (1.45 g, 3.85 mmol, 77%, 13:1 *E:Z*).



26. SI-14 (1.0 g, 2.6 mmol, 1.0 equiv) was dissolved in THF (20 mL) and cooled to 0 °C. TBAF (1.0 M in THF, 3.6 mL, 3.6 mmol, 1.4 equiv) was then added slowly and the reaction was stirred for 5 to 7 h until judged complete by TLC. The reaction mixture was diluted with EtOAc (45 mL) and washed with saturated NH₄Cl (30 mL), water (30 mL), brine (30 mL), dried over MgSO₄, and the solvent was removed. The residue was purified by flash column chromatography (10-20% EtOAc, 3% triethyl amine in hexanes) to afford the title compound as a pale yellow oil. **TLC** R_f = 0.1 (1% TEA, 20% EtOAc/hexanes, UV active) **¹H NMR** (500 MHz, CDCl₃) δ 8.54 (m, 1H), 7.7 (td, *J* = 7.7 Hz, 1.7 Hz, 1H), 7.22 (m, 2H), 6.9 (dt, *J* = 15.6, 7.0 Hz, 1H), 5.8 (dt, *J* = 15.6, 1.6 Hz, 1H), 4.7 (m, 1H), 4.2 (m, 1H), 2.2 (m, 1H), 1.8 (m, 1H), 1.7 (m, 1H), 1.5 (m, 1H), 1.3 (t, *J* = 7.0 Hz, 3H). **¹³C NMR** (125 MHz, CDCl₃) δ 166.7, 162.8, 149.2, 148.2, 136.8, 122.3, 121.4, 120.3, 72.9, 60.1, 38.2, 32.1, 28.0, 24.9, 14.3; **HRMS** (TOF MS ES+) *m/z* calcd for C₁₅H₂₁NO₃Na (M + Na)⁺ 286.1419, found 286.1414.

III. Nickel-Catalyzed Hydrogenolysis of Benzylic Carbinols and Characterization Data for Products

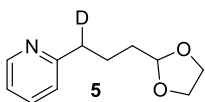
General Procedure B. Hydrogenolysis of benzylic carbinols



2. In a glovebox, a flame-dried 7 mL vial equipped with a stir bar was charged with $\text{NiCl}_2(\text{dppe})$ (11 mg, 0.020 mmol, 10 mol %), alcohol **1** (45 mg, 0.20 mmol, 1.0 equiv) and diethyl chlorophosphate (35 μL , 0.24 mmol, 1.2 equiv) and suspended in THF (1.6 mL). The reaction vial was capped with a screw-cap fitted with a septum and removed from the glove box. The reaction was placed under an atmosphere of N_2 and stirred at room temperature for five minutes. Diethyl zinc (1 M in PhMe, 0.50 mL, 0.50 mmol, 2.5 equiv) was added via syringe and an immediate color change from slightly orange to transparent yellow occurred. The reaction was then sealed with parafilm and allowed to stir for 18 h at 24 °C after which the reaction typically became dark brown or black. The reaction was quenched with MeOH and filtered through a plug of silica gel (washing with 100% EtOAc). The solvent was removed under reduced vacuum and the product was purified by flash column chromatography (12% EtOAc, 1% triethyl amine in hexanes) to afford the title compound as a pale yellow oil (29 mg, 0.13 mmol, 69%). **TLC** R_f = 0.3 (20% EtOAc/hexanes, UV active); **^1H NMR** (500 MHz, CDCl_3) δ 8.52 (ddd, J = 4.9, 1.9, 1.0 Hz, 1H), 7.56 (td, J = 2.0, 7.7 Hz, 1H), 7.29–7.23 (m, 2H), 7.19–7.14 (m, 3H), 7.13–7.06 (m, 2H), 2.81 (t, J = 7.3 Hz, 2H), 2.65 (t, J = 7.7 Hz, 2H), 1.83–1.74 (m, 2H) 1.74–1.64 (m, 2H); **^{13}C NMR** (125 MHz, CDCl_3) δ 162.2, 149.3, 142.6, 136.3, 128.5, 128.3, 125.7, 122.7, 120.9, 38.3, 35.8, 31.3, 29.6; **IR** (neat) 3025, 2926, 2855, 2359, 2341, 1598 cm^{-1} ; **HRMS** (TOF MS Cl^+) m/z calcd for $\text{C}_{16}\text{H}_{18}\text{ONH}_4$ ($\text{M} + \text{NH}_4$) $^+$ 211.1361, found 211.1351.

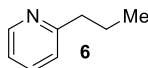
One-gram scale formation of 2. In a glovebox, a flame-dried round-bottomed flask equipped with a stir bar was charged with $\text{NiCl}_2(\text{dppe})$ (244.4 mg, 0.4444 mmol, 10 mol %), alcohol **1** (1.00 g, 4.44 mmol, 1.0 equiv) and diethyl chlorophosphate (777 μL , 5.33 mmol, 1.2 equiv) and suspended in THF (35.5 mL). The flask was capped with a septum and removed from the glove box. The reaction was placed under an atmosphere of N_2 and stirred at room temperature. Diethyl zinc (1 M in PhMe, 11.1 mL, 11.1 mmol, 2.5 equiv) was added via syringe and an immediate color change from slightly orange to transparent yellow occurred. The reaction was allowed to stir for 18 h at 24 °C after which the reaction typically became dark brown or black. The reaction was quenched with MeOH and filtered through a plug of silica gel (washing with 100% EtOAc). The solvent was removed under reduced vacuum and the product was purified by flash column chromatography (12% EtOAc, 1% triethyl amine in hexanes) to afford the title compound as a pale yellow oil (0.672 g, 3.01 mmol, 72%).

i. Deuterium incorporation (Scheme 2)

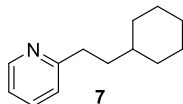


5. Using General Procedure B, outlined above, the following amounts of reagents were used: $\text{NiCl}_2(\text{dppe})$ (11 mg, 0.020 mmol, 10 mol %), alcohol **3** (42 mg, 0.20 mmol, 1.0 equiv), diethyl chlorophosphate (35 μL , 0.24 mmol, 1.2 equiv), THF (1.6 mL), and diethyl zinc (1.0 M in PhMe, 0.50 mL, 0.50 mmol, 2.5 equiv). The reaction was quenched with CH_3OD from a sealed ampoule (1 mL) and the product was purified by flash column chromatography (12 % EtOAc, 1% triethyl amine, in hexane) to afford the title compound as a pale yellow oil (30 mg, 0.15 mmol, 78%). **TLC** R_f = 0.1 (20% EtOAc/hexanes, UV active); $^1\text{H NMR}$ (500 MHz, CDCl_3) δ 8.52 (d, J = 4.6 Hz, 1H), 7.58 (td, J = 7.7, 1.9 Hz, 1H), 7.15 (d, J = 7.7 Hz, 1H), 7.11–7.08 (m, 1H), 4.89 (t, J = 4.7 Hz, 1H), 4.01–3.79 (m, 4H), 2.87–2.79 (m, 1H), 1.91–1.83 (m, 2H), 1.76–1.68 (m, 2H); $^2\text{H NMR}$ (500 MHz, CHCl_3) 3.10 (s, 1D); $^{13}\text{C NMR}$ (125 MHz, CDCl_3) δ 161.8, 149.3, 136.3, 122.8, 121.1, 104.5, 64.9, 38.1, 33.4, 24.2; **IR** (neat) 2922, 2874, 1591, 1567, 1473, 1410 cm^{-1} ; **HRMS** (TOF MS Cl^+) m/z calcd for $\text{C}_{16}\text{H}_{18}\text{ONH}_4$ ($M + \text{NH}_4$) $^+$ 195.1244, found 195.1251.

ii. Synthesis of Products in Scheme 3 (6–14)



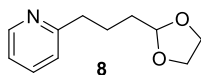
6. Using General Procedure B, outlined above, the following amounts of reagents were used: $\text{NiCl}_2(\text{dppe})$ (11 mg, 0.020 mmol, 10 mol %), alcohol **SI-1** (28 mg, 0.20 mmol, 1.0 equiv), diethyl chlorophosphate (35 μL , 0.24 mmol, 1.2 equiv), THF (1.6 mL), and diethyl zinc (1.0 M in PhMe, 0.50 mL, 0.50 mmol, 2.5 equiv). The product was purified by flash column chromatography (15% EtOAc/hexanes) to afford the title compound as a pale yellow oil (16 mg, 0.13 mmol, 67%). Analytical data is consistent with literature values.¹¹ **TLC** R_f = 0.3 (20% EtOAc/hexanes, UV active); $^1\text{H NMR}$ (500 MHz, CDCl_3) δ 8.53 (d, J = 4.4 Hz, 1H), 7.58 (t, J = 7.7 Hz, 1H), 7.14 (d, J = 7.7 Hz, 1H), 7.09 (t, J = 6.5 Hz, 1H), 2.77 (t, J = 7.8 Hz, 2H), 1.76 (sextet, J = 7.7 Hz, 2H), 0.97 (t, J = 7.7 Hz, 3H); $^{13}\text{C NMR}$ (125 MHz, CDCl_3) δ 162.4, 149.3, 136.3, 122.9, 121.0, 40.5, 23.2, 14.0.



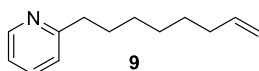
7. Using General Procedure B, outlined above, the following amounts of reagents were used: $\text{NiCl}_2(\text{dppe})$ (11 mg, 0.020 mmol, 10 mol %), alcohol **SI-2** (41 mg, 0.20 mmol, 1.0 equiv), diethyl chlorophosphate (35 μL , 0.24 mmol, 1.2 equiv), THF (1.6 mL), and diethyl zinc (1.0 M in PhMe, 0.50 mL, 0.50 mmol, 2.5 equiv). The product was purified by flash column chromatography (15% EtOAc/hexanes) to afford the title compound as a pale yellow oil (27 mg, 0.14 mmol, 70%). **TLC** R_f = 0.2 (10% EtOAc/hexanes, UV active); $^1\text{H NMR}$ (500 MHz, CDCl_3) δ 8.52 (d, J = 4.5 Hz, 1H), 7.57 (td, J = 7.7, 1.7 Hz, 1H), 7.14 (d, J = 7.7 Hz, 1H), 7.08 (dd, J = 7.7, 5.2 Hz, 1H), 2.83–2.76 (m, 2H), 1.78 (ad, J = 13.3 Hz, 2H), 1.74–1.67 (m, 2H), 1.67–1.58 (m, 3H), 1.34–1.10 (m, 4H),

¹¹ Groenhagen, U.; Maczka, M.; Dickschat, J. S.; Schulz, S. *Beilstein J. Org. Chem.* **2014**, *10*, 1421.

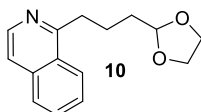
1.00–0.90 (m, 2H); ¹³C NMR (125 MHz, CDCl₃) δ 163.0, 149.3, 136.4, 122.8, 120.9, 37.8, 37.7, 36.0, 33.4, 26.8, 26.5; IR (neat) 2920, 1589 cm⁻¹; HRMS (TOF MS ES+) *m/z* calcd for C₁₃H₁₉NH (M + H)⁺ 190.1596, found 190.1594.



8. Using General Procedure B, outlined above, the following amounts of reagents were used: NiCl₂(dppe) (11 mg, 0.020 mmol, 10 mol %), alcohol **3** (42 mg, 0.20 mmol, 1.0 equiv), diethyl chlorophosphate (35 μL, 0.24 mmol, 1.2 equiv), THF (1.6 mL), and diethyl zinc (1.0 M in PhMe, 0.50 mL, 0.50 mmol, 2.5 equiv). The product was purified by flash column chromatography (12% EtOAc/hexanes) to afford the title compound as a pale yellow oil (31 mg, 0.15 mmol, 74 %). Analytical data are consistent with literature values.¹² TLC R_f = 0.1 (20% EtOAc/hexanes, UV active); ¹H NMR (500 MHz, CDCl₃) δ 8.52 (d, *J* = 4.9 Hz, 1H), 7.57 (td, *J* = 7.6, 1.6 Hz, 1H), 7.14 (d, *J* = 7.8 Hz, 1H), 7.10 (dd, *J* = 7.6, 5.3 Hz, 1H), 4.89 (t, *J* = 6 Hz, 1H), 4.00–3.79 (m, 4H), 2.84 (t, *J* = 7.7 Hz, 2H), 1.92–1.82 (m, 2H), 1.76–1.68 (m, 2H); ¹³C NMR (125 MHz, CDCl₃) δ 161.8, 149.3, 136.3, 122.8, 121.1, 104.5, 64.9, 38.1, 33.4, 24.2.



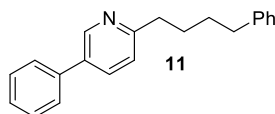
9. Using General Procedure B, outlined above, the following amounts of reagents were used: NiCl₂(dppe) (11 mg, 0.020 mmol, 10 mol %), alcohol **SI-3** (41 mg, 0.20 mmol, 1.0 equiv), diethyl chlorophosphate (35 μL, 0.24 mmol, 1.2 equiv), THF (1.6 mL), and diethyl zinc (1.0 M in PhMe, 0.50 mL, 0.50 mmol, 2.5 equiv). The product was purified by flash column chromatography (15% EtOAc/hexanes) to afford the title compound as a pale yellow oil (29 mg, 0.15 mmol, 77%). TLC R_f = 0.2 (20% EtOAc/hexanes, UV active); ¹H NMR (500 MHz, CDCl₃) δ 8.52 (d, *J* = 4.5 Hz, 1H), 7.58 (td, *J* = 7.6, 1.5 Hz, 1H), 7.14 (d, *J* = 7.6 Hz, 1H), 7.09 (dd, *J* = 7.3, 5.5 Hz, 1H), 5.80 (ddt, *J* = 17.1, 10.4, 3.3 Hz, 1H), 4.98 (dd, *J* = 17.1, 1.1 Hz, 1H), 4.92 (d, *J* = 10.4 Hz, 1H), 2.78 (t, *J* = 7.8 Hz, 2H), 2.03 (dd, *J* = 13.5, 6.5 Hz, 2H), 1.72 (quint, *J* = 7.6 Hz, 2H), 1.44–1.30 (m, 6H); ¹³C NMR (125 MHz, CDCl₃) δ 162.2, 149.3, 139.3, 136.3, 122.8, 121.0, 114.3, 38.6, 33.9, 30.0, 29.6, 29.1, 28.9; IR (neat) 3075, 2925, 1589 cm⁻¹; HRMS (TOF MS ES+) *m/z* calcd for C₁₃H₁₉NNa (M + Na)⁺ 212.1415, found 212.1406.



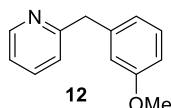
10. Using General Procedure B, outlined above, the following amounts of reagents were used: NiCl₂(dppe) (11 mg, 0.020 mmol, 10 mol %), alcohol **SI-4** (52 mg, 0.20 mmol, 1.0 equiv), diethyl chlorophosphate (35 μL, 0.24 mmol, 1.2 equiv), THF (1.6 mL), and diethyl zinc (1.0 M in PhMe, 0.50 mL, 0.50 mmol, 2.5 equiv). The product was purified by flash column chromatography (30–50% EtOAc/hexanes) to afford the title compound as a pale yellow oil (24 mg, 0.099 mmol, 49%). TLC R_f = 0.2 (40% EtOAc/hexanes, UV active); ¹H NMR (500 MHz, CDCl₃) δ 8.43 (d, *J* = 5.6

¹² Kitbunnadaj, R.; Zuiderveld, O.P.; Christophe, B.; Hulscher, S.; Menge, W.M.P.B.; Gelens, E.; Snip, E.; Bakker, R.A.; Celanire, S.; Gillard, M.; Talaga, P.; Timmerman, H.; Leurs, R. *J. Med. Chem.* **2004**, *47*, 2414.

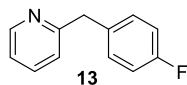
Hz, 1H), 8.17 (d, J = 8.4 Hz, 1H), 7.81 (d, J = 8.4 Hz, 1H), 7.66 (t, J = 7.5 Hz, 1H), 7.59 (t, J = 7.5 Hz, 1H), 7.50 (d, J = 5.6 Hz, 1H), 4.93 (t, J = 4.7 Hz, 1H), 4.00–3.92 (m, 2H), 3.89–3.81 (m, 2H), 3.36 (t, J = 8.0 Hz, 2H), 2.07–1.98 (m, 2H) 1.88–1.81 (m, 2H); ^{13}C NMR (125 MHz, CDCl_3) δ 161.9, 142.1, 136.4, 129.9, 127.5, 127.14, 127.05, 125.4, 119.4, 104.5, 65.0, 35.2, 33.8, 24.1; IR (neat) 3050, 2877, 1562 cm^{-1} ; HRMS (TOF MS ES+) m/z calcd for $\text{C}_{15}\text{H}_{17}\text{O}_2\text{NNa}$ ($\text{M} + \text{Na}$) $^+$ 266.1157, found 266.1160.



11. Using General Procedure B, outlined above, the following amounts of reagents were used: $\text{NiCl}_2(\text{dppe})$ (11 mg, 0.020 mmol, 10 mol %), alcohol **SI-6** (61 mg, 0.20 mmol, 1.0 equiv), diethyl chlorophosphate (35 μL , 0.24 mmol, 1.2 equiv), THF (1.6 mL), and diethyl zinc (1.0 M in PhMe, 0.50 mL, 0.50 mmol, 2.5 equiv). The product was purified by flash column chromatography (30–50% EtOAc/hexanes) to afford the title compound as a white solid (48 mg, 0.17 mmol, 83%). TLC R_f = 0.2 (10% EtOAc/hexanes, UV active); m.p. = 60 $^\circ\text{C}$; ^1H NMR (500 MHz, CDCl_3) δ 8.75 (s, 1H), 7.76 (dd, J = 8.1, 2.0 Hz, 1H), 7.56 (d, J = 8.0 Hz, 2H), 7.45 (t, J = 7.5 Hz, 2H), 7.37 (t, J = 7.2 Hz, 1H), 7.26 (t, J = 7.5 Hz, 2H), 7.18 (d, J = 8.1 Hz, 4H), 2.86 (t, J = 8.2 Hz, 2H), 2.67 (t, J = 7.6 Hz, 2H), 1.82 (quint, J = 7.6 Hz, 2H), 1.72 (quint, J = 7.5 Hz, 2H); ^{13}C NMR (125 MHz, CDCl_3) δ 161.1, 147.7, 142.6, 138.0, 134.8, 134.0, 129.1, 128.5, 128.4, 127.9, 127.1, 125.8, 122.7, 38.0, 35.9, 31.3, 29.6; IR (neat) 3056, 2854 cm^{-1} ; HRMS (TOF MS ES+) m/z calcd for $\text{C}_{21}\text{H}_{21}\text{NH}$ ($\text{M} + \text{H}$) $^+$ 288.1752, found 288.1758.



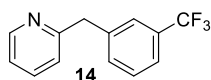
12. Using General Procedure B, outlined above, the following amounts of reagents were used: $\text{NiCl}_2(\text{dppe})$ (11 mg, 0.020 mmol, 10 mol %), alcohol **SI-7** (43 mg, 0.20 mmol, 1.0 equiv), diethyl chlorophosphate (35 μL , 0.24 mmol, 1.2 equiv), THF (1.6 mL), and diethyl zinc (1.0 M in PhMe, 0.50 mL, 0.50 mmol, 2.5 equiv). The product was purified by flash column chromatography (20 % EtOAc, 10% DCM, in hexanes) to afford the title compound as a pale yellow oil (21.5 mg, 0.120 mmol, 54%). Analytical data are consistent with literature values.¹³ TLC R_f = 0.3 (20% EtOAc/hexanes, UV active); ^1H NMR (500 MHz, CDCl_3) δ 8.54 (d, J = 4.3 Hz, 1H), 7.57 (td, J = 7.6, 1.7, 1H), 7.21 (t, J = 7.9 Hz, 1H), 7.12 (s, 1H), 7.10 (t, J = 3.2 Hz, 1H), 6.85 (d, J = 7.4 Hz, 1H), 6.81 (s, 1H), 6.76 (dd, J = 8.4, 2.4 Hz, 1H), 4.13 (s, 2H), 3.77 (s, 3H); ^{13}C NMR (125 MHz, CDCl_3) δ 160.9, 159.8, 149.4, 141.1, 136.6, 129.6, 123.2, 121.5, 121.3, 114.9, 111.9, 55.2, 44.8.



13. Using General Procedure B, outlined above, the following amounts of reagents were used: $\text{NiCl}_2(\text{dppe})$ (11 mg, 0.020 mmol, 10 mol %), alcohol **SI-8** (43 mg, 0.20 mmol, 1.0 equiv), diethyl chlorophosphate (35 μL , 0.24 mmol, 1.2 equiv), THF (1.6 mL), and diethyl zinc (1.0 M in PhMe, 0.50 mL, 0.50 mmol, 2.5 equiv). The crude product was pre-absorbed to a minimal amount of

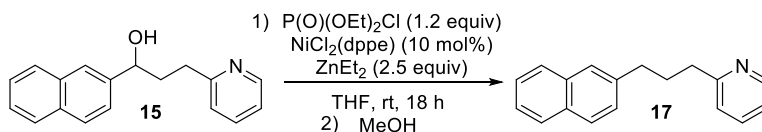
¹³ Chen, X.; Zhou, L.; Li, Y.; Xie, T.; Zhou, S. *J. Org. Chem.* **2014**, 79, 230.

silica and purified by flash column chromatography (12 % EtOAc, 1% triethyl amine, in hexane) to afford the title compound as a pale yellow oil (20.3 mg, 0.120 mmol, 60%). Analytical data are consistent with literature values.¹⁴ **TLC** R_f = 0.3 (20% EtOAc/hexanes, UV active); **¹H NMR** (500 MHz, CDCl₃) δ 8.55 (ddd, J = 4.9, 1.9, 1.0 Hz, 1H), 7.58 (td, J = 7.6, 1.9 Hz, 1H), 7.24–7.19 (m, 2H), 7.14–7.07 (m, 2H), 7.01–6.95 (m, 2H), 4.12 (s, 2H); **¹³C NMR** (125 MHz, CDCl₃) δ 162.5 (d, J_{CF} = 244.14 Hz), 160.6, 149.4, 136.6, 135.1 (d, J_{CF} = 3.24 Hz), 130.5 (d, J_{CF} = 7.87 Hz), 123.0, 121.3, 115.3 (d, J_{CF} = 21.2 Hz), 43.8.



14. Using General Procedure B, outlined above, the following amounts of reagents were used: NiCl₂(dppe) (11 mg, 0.020 mmol, 10 mol %), alcohol **SI-9** (51 mg, 0.20 mmol, 1.0 equiv), diethyl chlorophosphate (35 μ L, 0.24 mmol, 1.2 equiv), THF (1.6 mL), and diethyl zinc (1.0 M in PhMe, 0.50 mL, 0.50 mmol, 2.5 equiv). The product was purified by flash column chromatography (30–50% EtOAc/hexanes) to afford the title compound as a pale yellow oil (29.4 mg, 0.124 mmol, 62%). Analytical data are consistent with literature values.¹⁰ **TLC** R_f = 0.2 (10% EtOAc/hexanes, UV active); **¹H NMR** (500 MHz, CDCl₃) δ 8.56 (d, J = 4.9 Hz, 1H), 7.61 (dt, J = 2.0, 7.6 Hz, 1H), 7.53 (s, 1H), 7.50–7.38 (m, 3H), 7.17–7.09 (m, 2H), 4.21 (s, 2H); **¹³C NMR** (125 MHz, CDCl₃) δ 159.9, 149.6, 140.4, 136.8, 132.5, 130.8 (q, J_{CF} = 31.2 Hz), 129.0, 125.7, 124.2 (q, J_{CF} = 270 Hz) 123.4, 123.3, 121.6, 44.3.

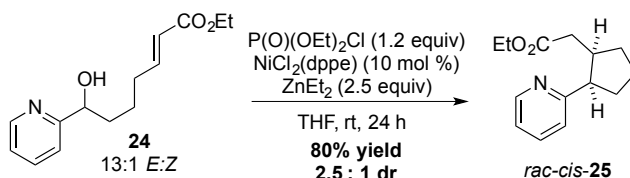
iii. Hydrogenolysis of **15** (Scheme 4)



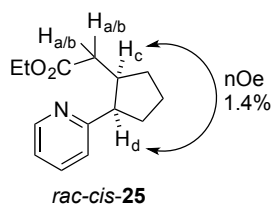
17. Using General Procedure B, outlined above, the following amounts of reagents were used: NiCl₂(dppe) (11 mg, 0.020 mmol, 10 mol %), alcohol **15** (55 mg, 0.20 mmol, 1.0 equiv), diethyl chlorophosphate (35 μ L, 0.24 mmol, 1.2 equiv), THF (1.6 mL), and diethyl zinc (1.0 M in PhMe, 0.50 mL, 0.50 mmol, 2.5 equiv). The product was purified by flash column chromatography (20% EtOAc/hexanes) to afford the title compound as a pale yellow oil (40 mg, 0.16 mmol, 81%). **TLC** R_f = 0.2 (20% EtOAc/hexanes, UV active); **¹H NMR** (500 MHz, CDCl₃) δ 8.54 (d, J = 4.4 Hz, 1H), 7.79 (d, J = 8.0 Hz, 1H), 7.76 (d, J = 7.7 Hz, 2H), 7.62 (s, 1H), 7.56 (t, J = 7.7 Hz, 1H), 7.47–7.38 (m, 2H), 7.34 (d, J = 8.3 Hz, 1H), 7.12 (d, J = 7.7 Hz, 1H), 7.09 (at, J = 6.4 Hz, 1H), 2.90–2.81 (m, 4H), 2.16 (quint, J = 7.0 Hz, 2H); **¹³C NMR** (125 MHz, CDCl₃) δ 162.0, 149.4, 139.8, 136.3, 133.7, 132.1, 128.0, 127.7, 127.53, 127.48, 126.6, 126.0, 125.2, 122.9, 121.1, 38.0, 35.8, 31.4; **IR** (neat) 3051, 2856, 1590 cm⁻¹; **HRMS** (TOF MS ES+) m/z calcd for C₁₈H₁₇NNa (M + Na)⁺ 270.1259, found 270.1260.

¹⁴ De Houwer, J.; Tehrani, K. A.; Maes, B. U. W. *Angew. Chem. Int. Ed.* **2012**, *51*, 2745.

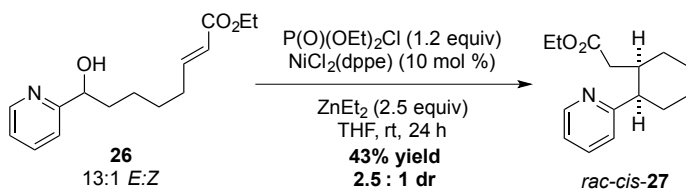
IV. Nickel-Catalyzed Intramolecular 1,4-addition (Scheme 5)



25. Using General Procedure B, outlined above, the following amounts of reagents were used: NiCl₂(dppe) (11 mg, 0.020 mmol, 10 mol %), alcohol **24** (50 mg, 0.20 mmol, 1.0 equiv), diethyl chlorophosphate (35 μ L, 0.24 mmol, 1.2 equiv), THF (1.6 mL), and diethyl zinc (1.0 M in PhMe, 0.50 mL, 0.50 mmol, 2.5 equiv). The product was purified by flash column chromatography (20% EtOAc/hexanes) to afford the title compound as a 2.5:1 mixture of diastereomers (37 mg, 0.16 mmol, 80%). H_c was integrated to determine the ratio of diastereomers. Flash column chromatography (5–10% EtOAc/hexanes) was performed a second time to isolate analytically pure *cis* diastereomer (19 mg, 0.081 mmol, 40%) and a 1:1 mixture of diastereomers (16 mg, 0.070 mmol, 34%). The relative configuration of the *cis* diastereomer was assigned based on the nOe correlation shown.

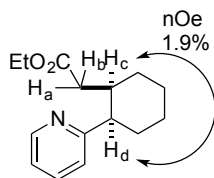


TLC R_f = 0.1 (10% EtOAc/hexanes, UV active); **¹H NMR** (500 MHz, CDCl₃) δ 8.55 (d, *J* = 4.7 Hz, 1H), 7.56 (td, *J* = 7.7, 2.0 Hz, 1H), 7.12–7.07 (m, 2H), 4.00 (q, *J* = 7.2 Hz, 2H), 3.43 (q, *J* = 8.0 Hz, 1H), 2.79–2.71 (m, 1H), 2.14–2.04 (m, 2H), 2.02–1.90 (m, 4H), 1.75–1.67 (m, 1H), 1.63–1.54 (m, 1H), 1.17 (t, *J* = 7.2 Hz, 3H); **¹³C NMR** (125 MHz, CDCl₃) δ 173.6, 163.2, 149.2, 136.0, 123.5, 121.2, 60.2, 49.8, 40.8, 36.2, 31.9, 30.2, 24.2, 14.3; **IR** (neat) 2935, 1716 cm⁻¹; **HRMS** (TOF MS ES+) *m/z* calcd for C₁₄H₁₉NO₂Na (M + Na)⁺ 256.1313, found 256.1316.



27. Using General Procedure B, outlined above, the following amounts of reagents were used: NiCl₂(dppe) (11 mg, 0.020 mmol, 10 mol %), alcohol **26** (53 mg, 0.20 mmol, 1.0 equiv), diethyl chlorophosphate (35 μ L, 0.24 mmol, 1.2 equiv), THF (1.6 mL), and diethyl zinc (1.0 M in PhMe, 0.50 mL, 0.50 mmol, 2.5 equiv). The product was purified by flash column chromatography (10–20% EtOAc/hexanes) to afford the title compound as a 2.5:1 mixture of diastereomers (21.3 mg, 0.086 mmol, 43%). H_c was integrated to determine the ratio of diastereomers. Flash column chromatography (5–10% EtOAc/hexanes) was performed a second time to isolate the

analytically pure *cis* diastereomer (11 mg, 0.045 mmol, 23%) and a 1:1 mixture of diastereomers (10 mg, 0.04 mmol, 20%). The relative configuration of the *cis* diastereomer was assigned based on the nOe correlation shown below.



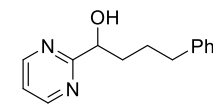
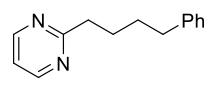
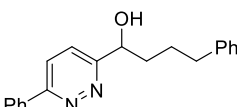
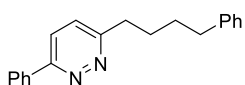
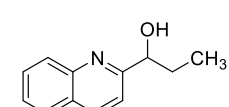
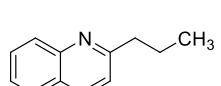
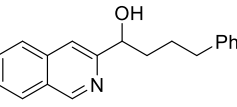
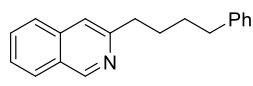
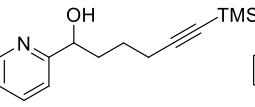
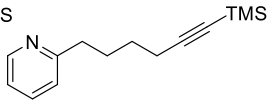
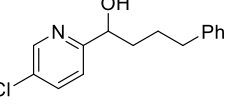
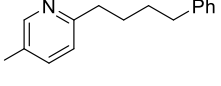
rac-cis-27

TLC R_f = 0.5 (20% EtOAc/hexanes, UV active); **^1H NMR** (500 MHz, CDCl_3) δ 8.54 (d, J = 4.3 Hz, 1H), 7.59 (t, J = 7.6, Hz, 1H), 7.12–7.05 (m, 2H), 3.97 (q, J = 7.3 Hz, 2H), 3.05 (m, 1H), 2.7 (m, 1H), 2.34 (dd, J = 9.8, 5.38, 1H), 2.00 (dd, J = 4.3, 4.7 1H), 1.95–1.75 (m, 4H), 1.69 (m, 1H), 1.5 (m, 2H), 1.4 (m, 1H) 1.16 (t, J = 7.14 Hz, 3H); **^{13}C NMR** (125 MHz, CDCl_3) δ 173.8, 164.3, 149.3, 136.4, 122.4, 121.4, 60.4, 48.1, 36.3, 32.8, 30.6, 26.2, 25.4, 21.0, 14.5; **HRMS** (TOF MS ES+) m/z calcd for $\text{C}_{15}\text{H}_{21}\text{NO}_2\text{Na}$ ($M + \text{Na}$) $^+$ 270.1470, found 270.1471.

IV. Additional Information Regarding Substrate Scope

The identity of the functional groups and heteroarenes of the benzylic alcohols were varied to explore the functional group tolerance of the reaction (Table SI-1). Heterocycles containing more than a single nitrogen, such as pyrimidine and pyridazine, formed the desired product in low yields (entries 1 and 2). Likewise, 2-quinoline and 3-isoquinoline moieties were not well tolerated under the reaction conditions (entries 3 and 4). A 2-pyridyl carbinol containing a tethered TMS protected alkyne produced the desired deoxygenated product in modest yield (entry 5). Finally, halogen substitution within the pyridine ring proved detrimental to the desired transformation (entry 6).

Table SI-1. Evaluation of additional substrate classes

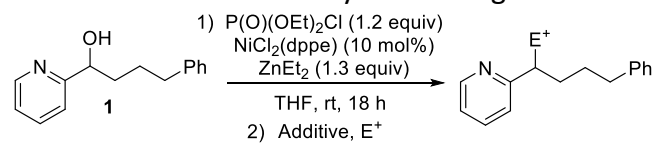
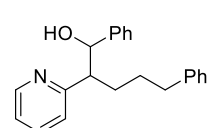
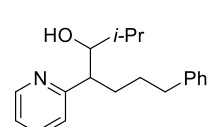
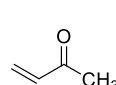
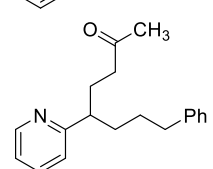
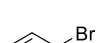
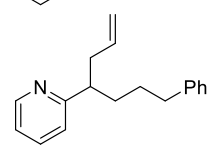
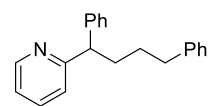
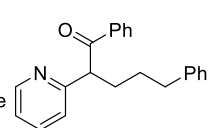
$ \begin{array}{c} \text{HetAr}-\text{CH}(\text{OH})-\text{R} \\ \xrightarrow[\text{2) MeOH}]{\begin{array}{l} \text{P(O)(OEt)}_2\text{Cl (1.2 equiv)} \\ \text{NiCl}_2(\text{dppe}) \text{ (10 mol\%)} \\ \text{ZnEt}_2 \text{ (1.3 equiv)} \\ \text{THF, rt, 18 h} \end{array}} \\ \text{HetAr}-\text{CH}_2-\text{R} \\ \textbf{A} \end{array} $			
Entry	Substrate	Deoxygenation Product	yield A (%) ^a
1			8
1			16
3			11
4			20
5 ^b			45
6			15

^aDetermined by ¹H NMR using an internal standard (PhSiMe₃).

^bReaction run at 70 °C

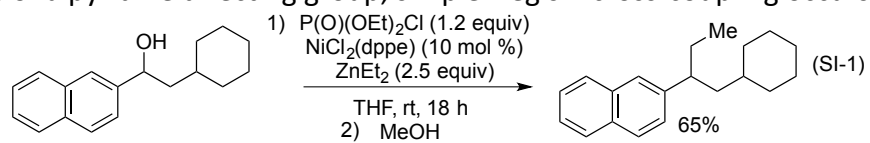
The addition of benzylic zinc reagents formed under our reaction conditions to a variety of electrophiles was examined (Table SI-2). Representative examples are shown. Titanium- and copper-assisted additions of zinc reagent into aldehydes proceeds in modest yield (entries 1 and 2). An intermolecular 1,4-addition to methyl vinyl ketone afford the desired product in 30% yield (entry 3). Addition of zinc reagent to allyl bromide resulted in low yield of the alkylated product (entry 4). Cross-coupling of the benzylic zinc reagent with iodobenzene failed to produce desired product (entry 5). Finally, nickel-catalyzed coupling of the zinc reagent with benzoyl chloride results in low yields of the desired ketone (entry 6).

Table SI-2. Addition of heterobenzylic zinc reagent to electrophiles

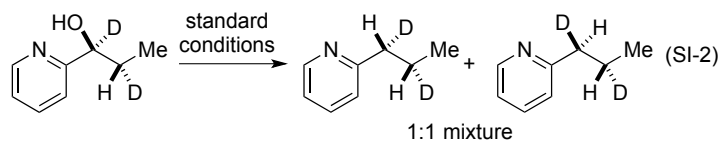
				
Entry	Additive	E ⁺	Product	yield A (%) ^a
1	Ti(O <i>i</i> -Pr) ₄	PhCHO		21
2	CuCN•2LiCl	<i>i</i> -PrCHO		32
3	none			30
4	CuCN•2LiCl			12
5	none	PhI		< 5
6	none	benzoyl chloride		11

^aDetermined by ¹H NMR using an internal standard (PhSiMe₃).

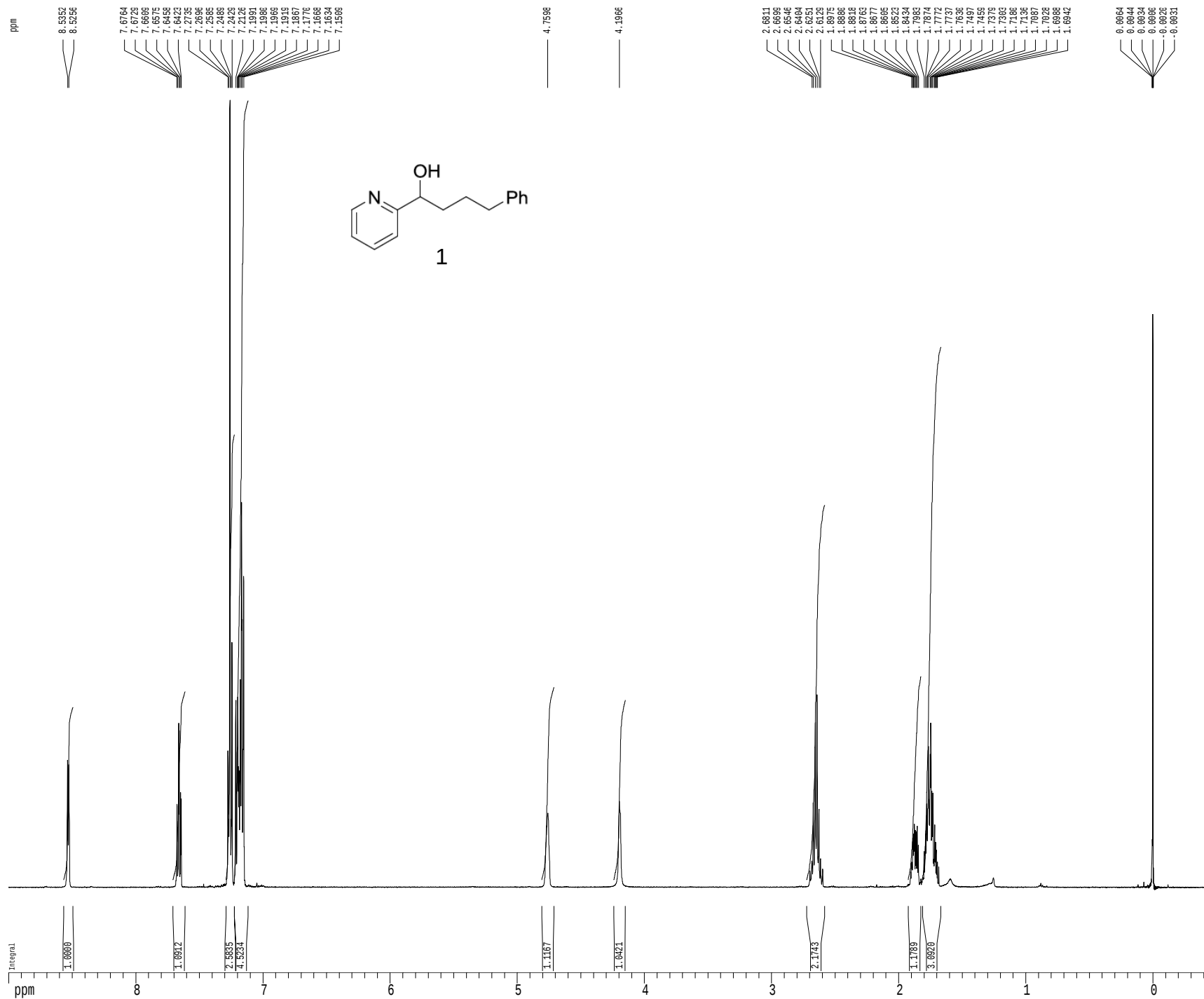
In the absence of a pyridine directing group, simple Negishi cross-coupling occurs (eq SI-1).



The hydrogenolysis is stereoablative, as demonstrated by reduction of a deuterated test substrate.



¹H spectrum



Current Data Parameters
 USER lhanna
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 EXPNO 1
 PROCNO 1

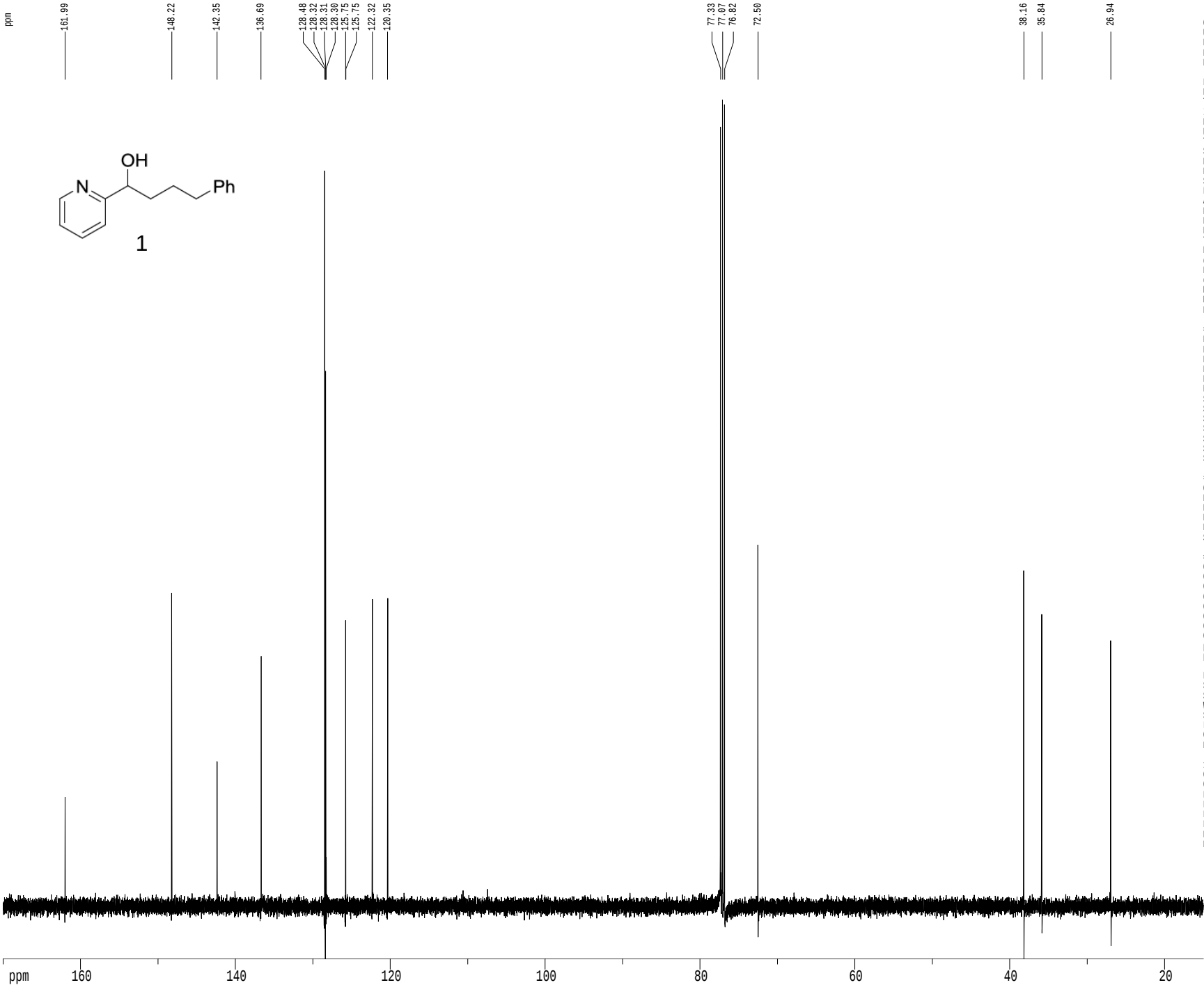
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 Time 18.59
 INSTRUM cryo500
 PROBHD 5 mm CPTCI 1H-
 PULPROG zg30
 TD 32648
 SOLVENT CDCl3
 NS 8
 DS 2
 SWH 8012.820 Hz
 FIDRES 0.250026 Hz
 AQ 1.9998451 sec
 RG 5.7
 DW 62.400 usec
 DE 6.00 usec
 TE 298.0 K
 D1 0.10000000 sec
 MCREST 0.00000000 sec
 MCWRR 0.01500000 sec

===== CHANNEL f1 =====
 NUC1 1H
 P1 7.50 usec
 PL1 1.60 dB
 SF01 500.2235015 MHz

F2 - Processing parameters
 SI 65536
 SF 500.2200321 MHz
 WDW no
 SSB 0
 LB 0.00 Hz
 GB 0
 PC 4.00

1D NMR plot parameters
 CX 22.80 cm
 CY 15.00 cm
 F1P 9.000 ppm
 F1 4501.98 Hz
 F2P -0.500 ppm
 F2 -250.11 Hz
 PPMCM 0.41667 ppm/cm
 HZCM 208.42502 Hz/cm

Z-restored spin-echo 13C spectrum with 1H decoupling



Current Data Parameters

USER lhanna
NAME LEH-5-MRH-102-C13
EXPNO 1
PROCNO 1

F2 - Acquisition Parameters

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Time 19.07
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PROBHD 5 mm CPTCI 1H-
PULPROG SpinEchopg30gp.prd
TD 65536
SOLVENT CDCl3
NS 513
DS 16
SWH 30303.031 Hz
FIDRES 0.462388 Hz
AQ 1.0813940 sec
RG 10321.3
DW 16.500 usec
DE 6.00 usec
TE 298.0 K
D1 0.25000000 sec
d11 0.03000000 sec
D16 0.00020000 sec
d17 0.00019600 sec
MCREST 0.00000000 sec
MCMRK 0.01500000 sec
P2 33.10 usec

===== CHANNEL f1 =====

NUC1 13C
P1 16.55 usec
P11 500.00 usec
P12 2000.00 usec
PL0 120.00 dB
PL1 -1.00 dB
SF01 125.7942548 MHz
SP1 2.70 dB
SP2 2.70 dB
SPNAM1 Crp60,0.5,20.1
SPNAM2 Crp60comp.4
SPOFF1 0.00 Hz
SPOFF2 0.00 Hz

===== CHANNEL f2 =====

CPDPRG2 waltz16
NUC2 1H
PCPD2 100.00 usec
PL2 1.60 dB
PL12 24.50 dB
SF02 500.2225011 MHz

===== GRADIENT CHANNEL =====

GP1AM1 SINE.100
GP1AM2 SINE.100
GPX1 0.00 %
GPX2 0.00 %
GPY1 0.00 %
GPY2 0.00 %
GPZ1 30.00 %
GPZ2 50.00 %
p15 500.00 usec
p16 1000.00 usec

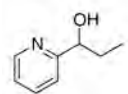
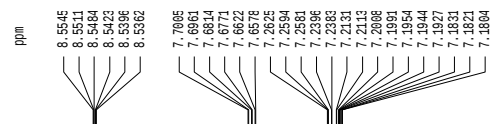
F2 - Processing parameters

SI 65536
SF 125.7804190 MHz
WDW NO
SSB 0
LB 0.00 Hz
GB 0
PC 2.00

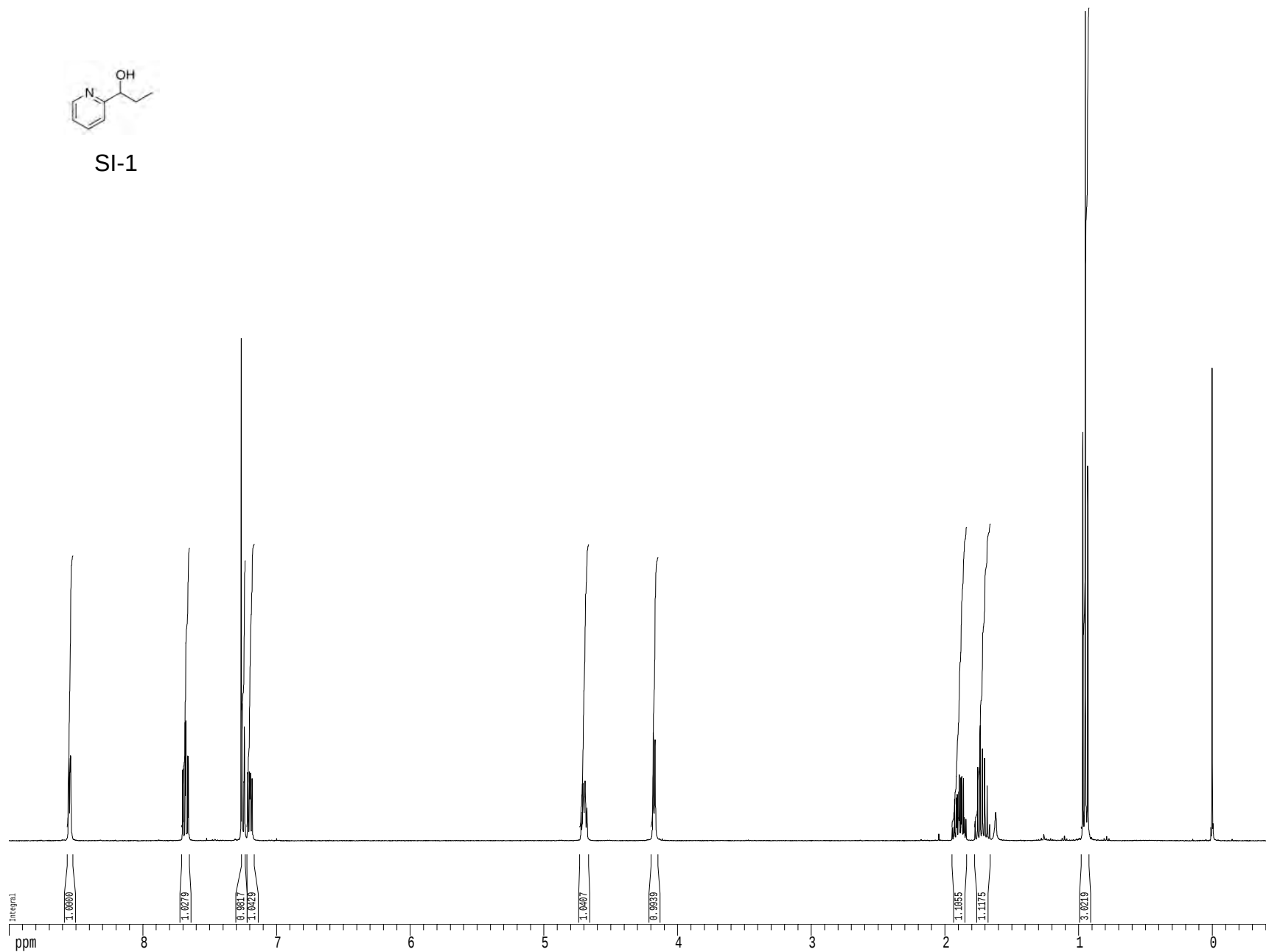
1D NMR plot parameters

CX 22.00 cm
CY 15.65 cm
F1P 170.000 ppm
F1 21382.67 Hz
F2P 15.000 ppm
F2 1886.71 Hz
PPMCM 6.79825 ppm/cm
HZCM 855.08618 Hz/cm

¹H spectrum



SI-1



Current Data Parameters
 USER mharri
 NAME MRH-VII-181-A
 EXPNO 1
 PROCNO 1

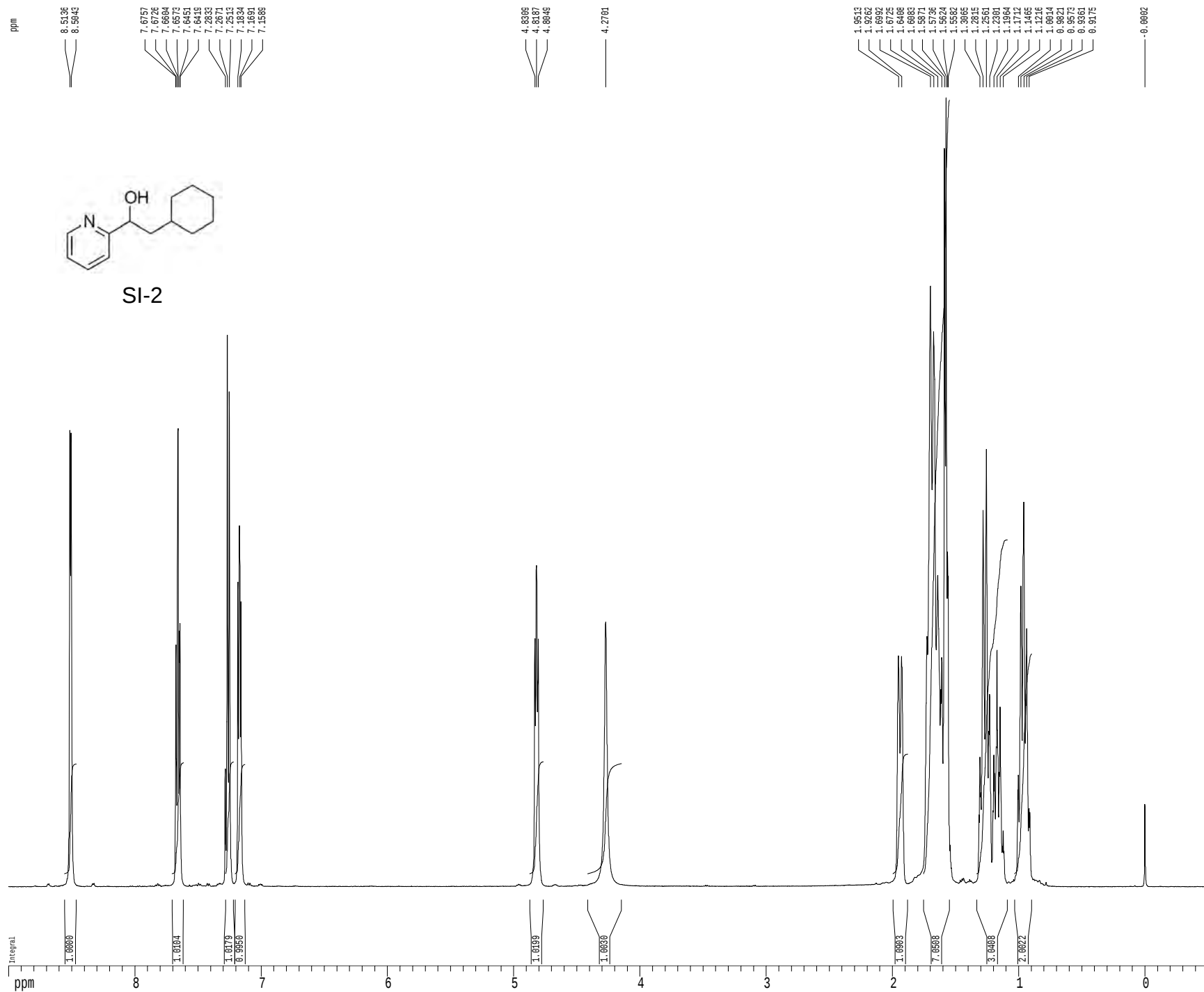
F2 - Acquisition Parameters
 Date_ 20141028
 Time 15.24
 INSTRUM drx400
 PROBHD 5 mm QNP H/F/P
 PULPROG zg30
 TD 65536
 SOLVENT CDC13T
 NS 8
 DS 2
 SWH 6410.256 Hz
 FIDRES 0.097813 Hz
 AQ 5.1118579 sec
 RG 456.1
 DW 78.000 usec
 DE 4.50 usec
 TE 298.0 K
 D1 0.10000000 sec
 MCREST 0.00000000 sec
 MCWRK 0.01500000 sec

===== CHANNEL f1 =====
 NUC1 1H
 P1 12.00 usec
 PL1 0.00 dB
 SF01 400.1328009 MHz

F2 - Processing parameters
 SI 65536
 SF 400.1300199 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 2.00

1D NMR plot parameters
 CX 22.00 cm
 CY 15.00 cm
 F1P 9.000 ppm
 F1 3691.17 Hz
 F2P -0.500 ppm
 F2 -200.06 Hz
 PPMCM 0.41667 ppm/cm
 HZCM 166.72084 Hz/cm

¹H spectrum



Current Data Parameters
 USER mharri
 NAME MRH-VII-299-1HNMR
 EXPNO 1
 PROCNO 1

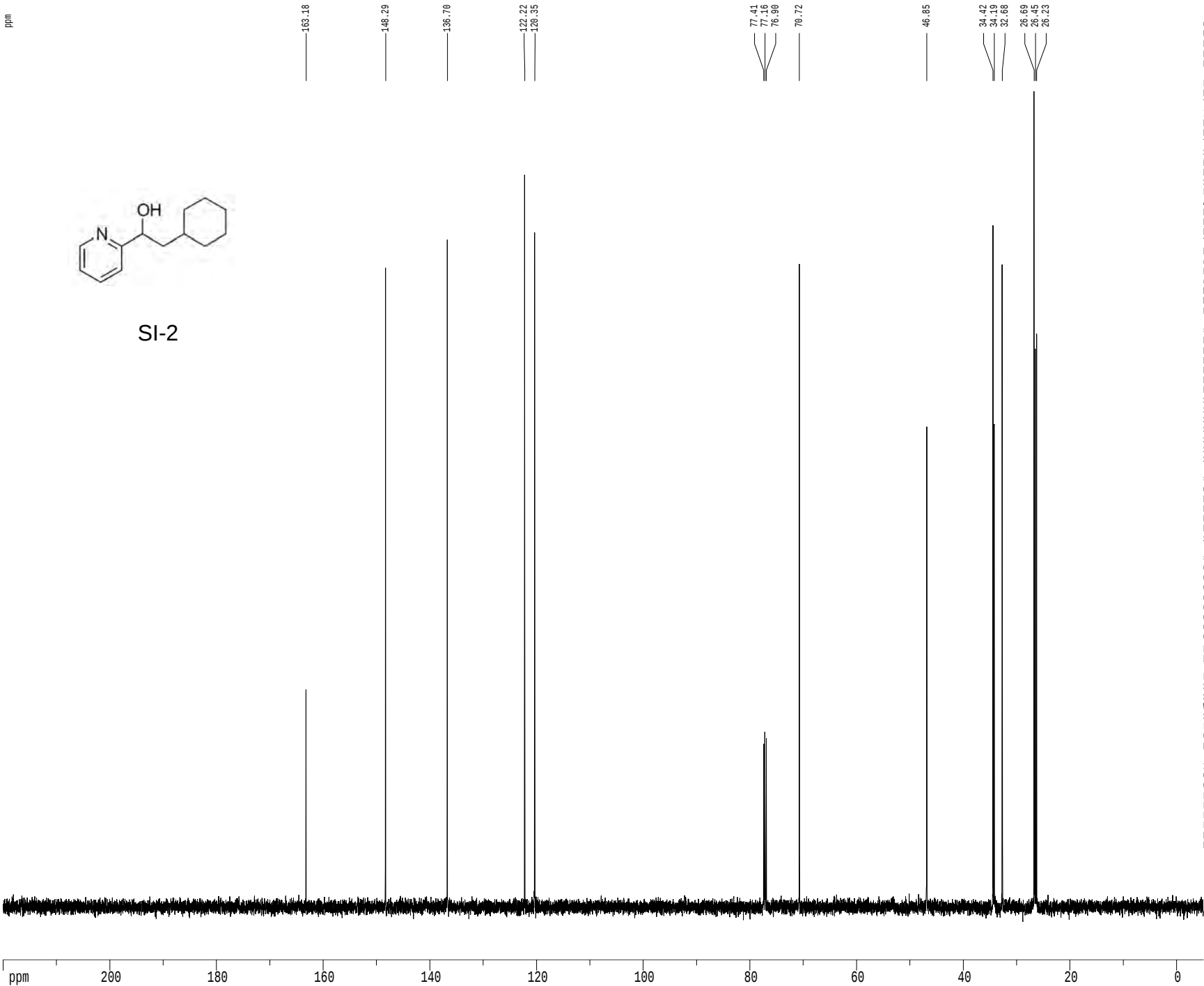
F2 - Acquisition Parameters
 Date_ 20150304
 Time 17.20
 INSTRUM cryo500
 PROBHD 5 mm CPTCI 1H-
 PULPROG zg30
 TO 81728
 SOLVENT CDCl3
 NS 8
 DS 2
 SWH 8012.820 Hz
 FIDRES 0.090843 Hz
 AQ 5.0998774 sec
 RG 5
 DW 62.400 usec
 DE 6.00 usec
 TE 298.0 K
 D1 0.10000000 sec
 MCREST 0.00000000 sec
 MCNRK 0.01500000 sec

===== CHANNEL f1 =====
 NUC1 1H
 P1 7.50 usec
 PL1 1.00 dB
 SF01 500.2235015 MHz

F2 - Processing parameters
 SI 65536
 SF 500.2200197 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 4.00

1D NMR plot parameters
 CX 22.00 cm
 CY 15.00 cm
 F1P 9.000 ppm
 F1 4501.98 Hz
 F2P -0.500 ppm
 F2 -250.11 Hz
 PPMCM 0.41667 ppm/cm
 HZCM 208.42502 Hz/cm

Z-restored spin-echo 13C spectrum with 1H decoupling



Current Data Parameters
USER mharri
NAME MRH-VII-299-13CNMR
EXPNO 1
PROCNO 1

F2 - Acquisition Parameters
Date_ 20150304
Time 17.26
INSTRUM cryo500
PROBHD 5 mm CPTCI 1H-
PULPROG SpinEchopg30gp.prd
TD 65536
SOLVENT CDCl3
NS 41
DS 16
SWH 30303.031 Hz
FIDRES 0.462388 Hz
AQ 1.0813940 sec
RG 5792.6
DW 16.500 usec
DE 6.00 usec
TE 298.0 K
D1 0.25000000 sec
d11 0.03000000 sec
D16 0.00020000 sec
d17 0.00019600 sec
MCREST 0.00000000 sec
MCWRK 0.01500000 sec
P2 33.10 usec

===== CHANNEL f1 =====
NUC1 13C
P1 16.55 usec
P11 500.00 usec
P12 2000.00 usec
PL0 120.00 dB
PL1 -1.00 dB
SF01 125.7942548 MHz
SP1 2.70 dB
SP2 2.70 dB
SPNAM1 Crp60,0.5,20.1
SPNAM2 Crp60comp.4
SPOFF1 0.00 Hz
SPOFF2 0.00 Hz

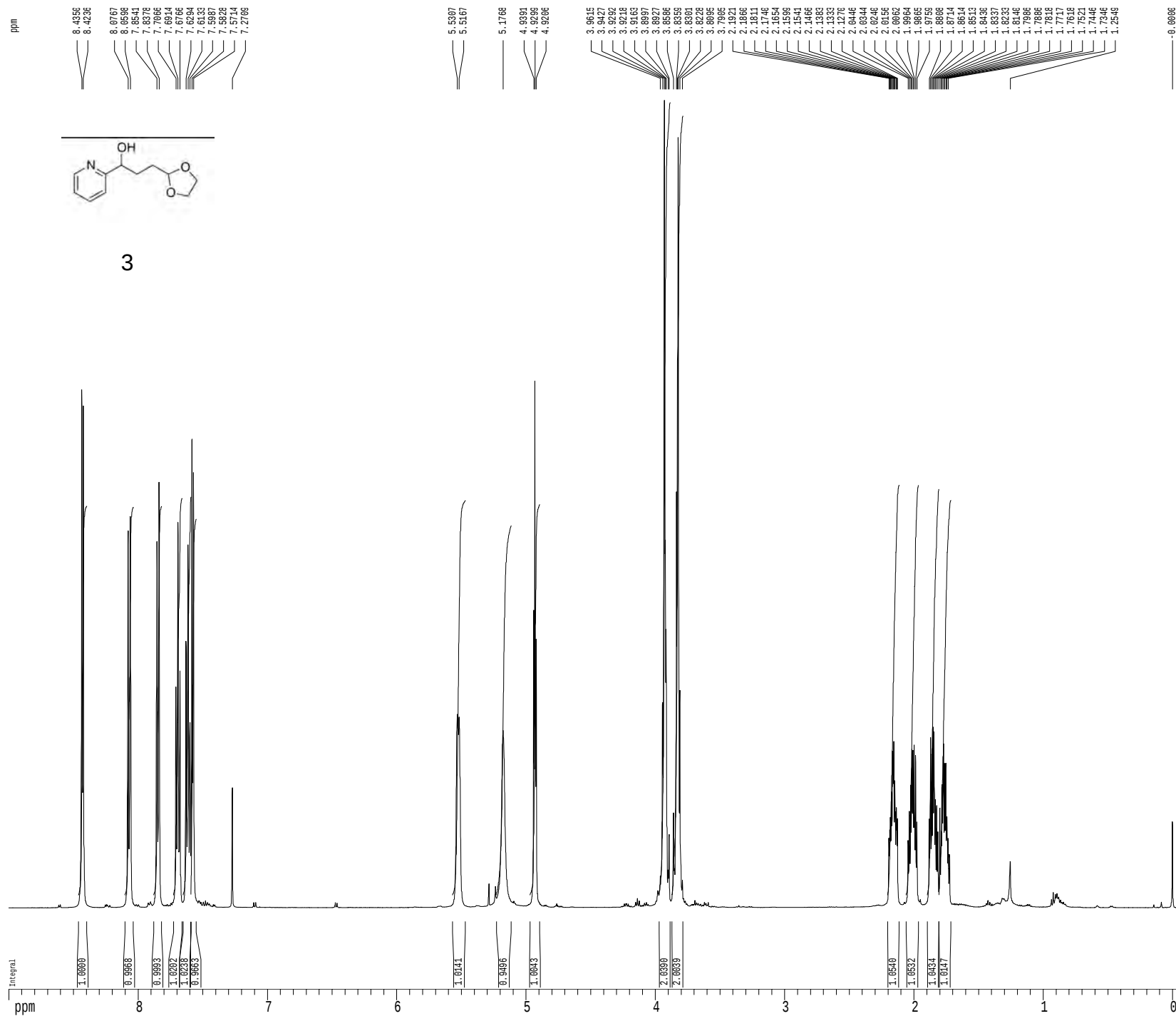
===== CHANNEL f2 =====
CPDPRG2 waltz16
NUC2 1H
PCPD2 100.00 usec
PL2 -1.00 dB
PL12 24.50 dB
SF02 500.2225011 MHz

===== GRADIENT CHANNEL =====
GPNAM1 SINE.100
GPNAM2 SINE.100
GPX1 0.00 %
GPX2 0.00 %
GPY1 0.00 %
GPY2 0.00 %
GPZ1 30.00 %
GPZ2 50.00 %
p15 500.00 usec
p16 1000.00 usec

F2 - Processing parameters
SI 65536
SF 125.7804150 MHz
EN
WDW 0
SSB 0
LB 1.00 Hz
GB 0
PC 2.00

1D NMR plot parameters
CX 22.00 cm
CY 15.65 cm
F1P 220.000 ppm
F1 27671.69 Hz
F2P -5.000 ppm
F2 -628.90 Hz
PPMCM 9.86842 ppm/cm
HZCM 1241.25415 Hz/cm

¹H spectrum



Current Data Parameters
 USER mharri
 NAME MRH-VII-208-1HNMR
 EXPNO 1
 PROCNO 1

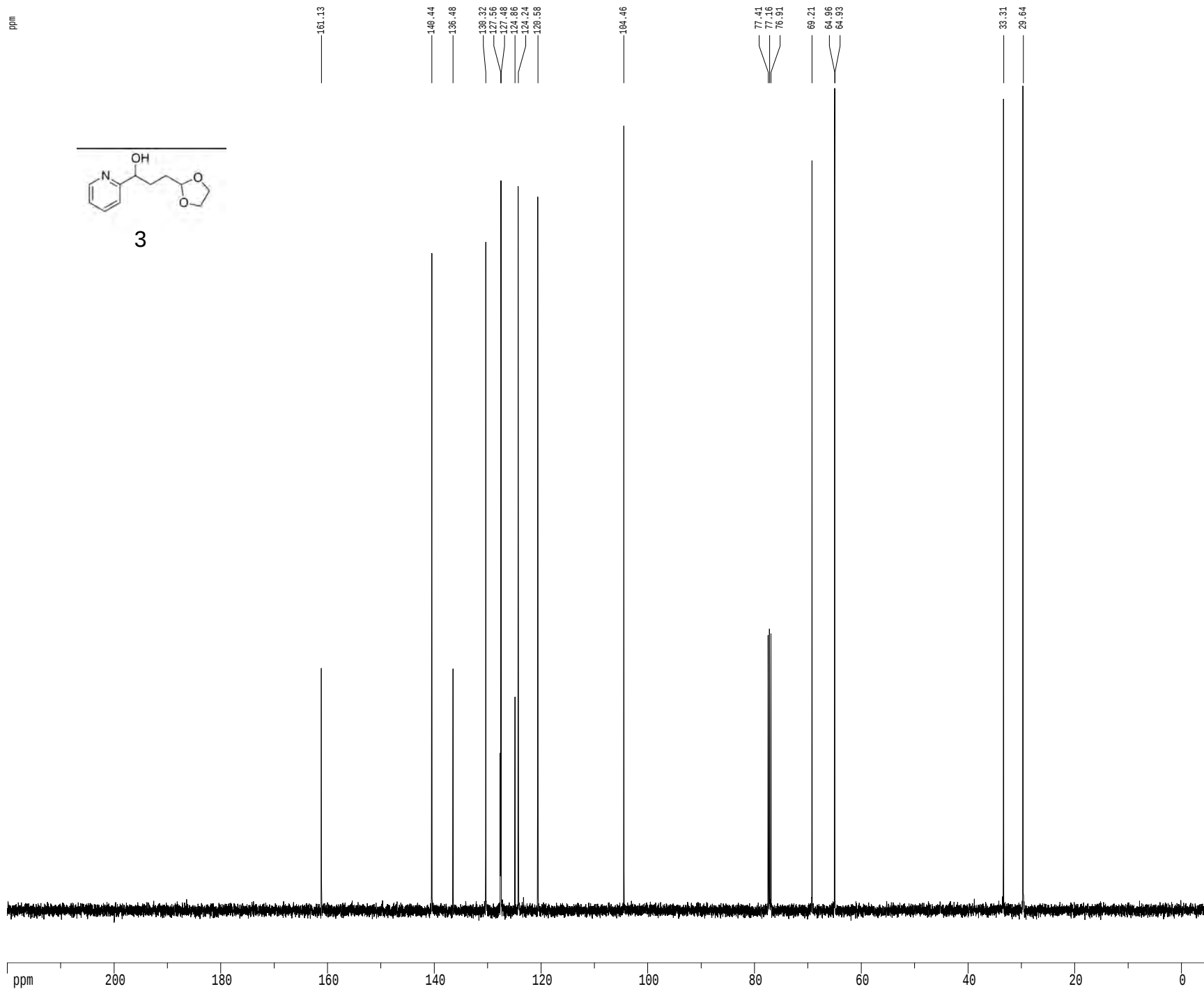
F2 - Acquisition Parameters
 Date_ 20150213
 Time 19.20
 INSTRUM cryo500
 PROBHD 5 mm CPTCI 1H-
 PULPROG zg30
 TO 81728
 SOLVENT CDCl3
 NS 8
 DS 2
 SWH 8012.820 Hz
 FIDRES 0.098043 Hz
 AQ 5.0998774 sec
 RG 6.3
 DW 62.400 usec
 DE 6.00 usec
 TE 298.0 K
 D1 0.10000000 sec
 MCREST 0.00000000 sec
 MCNRK 0.01500000 sec

===== CHANNEL f1 =====
 NUC1 1H
 P1 7.50 usec
 PL1 1.00 dB
 SFO1 500.2235015 MHz

F2 - Processing parameters
 SI 65536
 SF 500.2200249 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 4.00

1D NMR plot parameters
 CX 22.00 cm
 CY 15.00 cm
 F1P 9.000 ppm
 F1 4501.98 Hz
 F2P -0.500 ppm
 F2 -250.11 Hz
 PPMCM 0.41667 ppm/cm
 HZCM 208.42502 Hz/cm

Z-restored spin-echo 13C spectrum with 1H decoupling



Current Data Parameters
 USER mharri
 NAME MRH-VII-208-13CNMR
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20150213
 Time_ 19.23
 INSTRUM cryo500
 PROBHD 5 mm CPTCI 1H-
 PULPROG SpinEchopg30gp.prd
 TD 65536
 SOLVENT CDCl3
 NS 51
 DS 16
 SWH 30303.031 Hz
 FIDRES 0.462388 Hz
 AQ 1.0813940 sec
 RG 6502
 DW 16.500 usec
 DE 6.00 usec
 TE 298.0 K
 D1 0.25000000 sec
 d11 0.03000000 sec
 D16 0.00020000 sec
 d17 0.00019600 sec
 MCREST 0.00000000 sec
 MCWRR 0.01500000 sec
 P2 33.10 usec

===== CHANNEL f1 =====
 NUC1 13C
 P1 16.55 usec
 P11 500.00 usec
 P12 2000.00 usec
 PL0 120.00 dB
 PL1 -1.00 dB
 SF01 125.7942548 MHz
 SP1 2.70 dB
 SP2 2.70 dB
 SPNAM1 Crp60,0.5,20.1
 SPNAM2 Crp60comp.4
 SPOFF1 0.00 Hz
 SPOFF2 0.00 Hz

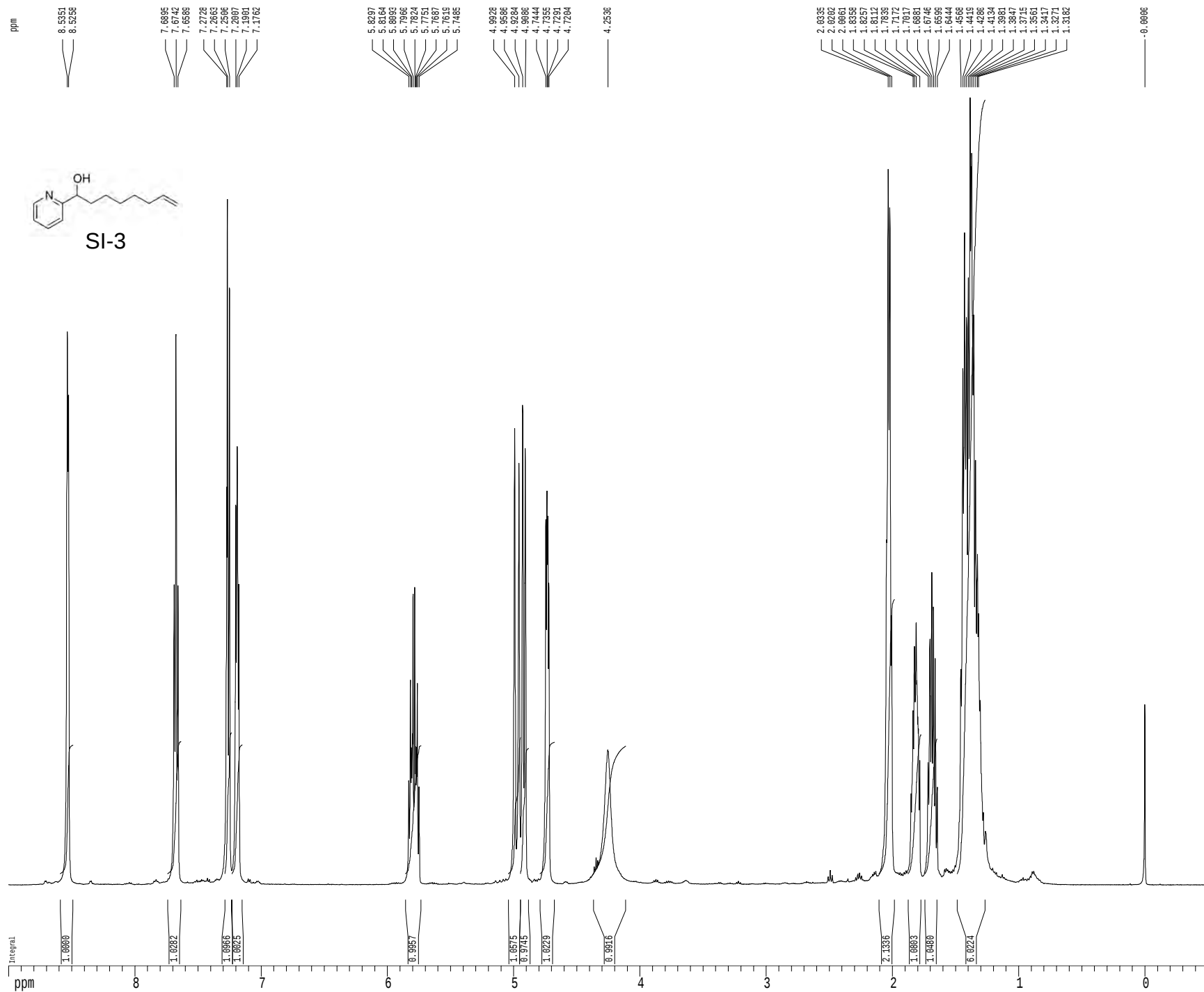
===== CHANNEL f2 =====
 CPDPRG2 waltz16
 NUC2 1H
 PCPD2 100.00 usec
 PL2 1.00 dB
 PL12 24.50 dB
 SF02 500.2225011 MHz

===== GRADIENT CHANNEL =====
 GPNAM1 SINE.100
 GPNAM2 SINE.100
 GPX1 0.00 %
 GPX2 0.00 %
 GPY1 0.00 %
 GPY2 0.00 %
 GPZ1 30.00 %
 GPZ2 50.00 %
 p15 500.00 usec
 p16 1000.00 usec

F2 - Processing parameters
 SI 65536
 SF 125.7804164 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 2.00

1D NMR plot parameters
 CX 22.00 cm
 CY 15.65 cm
 F1P 220.000 ppm
 F1 27671.69 Hz
 F2P -5.000 ppm
 F2 -628.90 Hz
 PPMCM 9.86842 ppm/cm
 HZCM 1241.25415 Hz/cm

¹H spectrum



Current Data Parameters
 USER mharri
 NAME MRH-VII-223-1HNMR
 EXPNO 1
 PROCNO 1

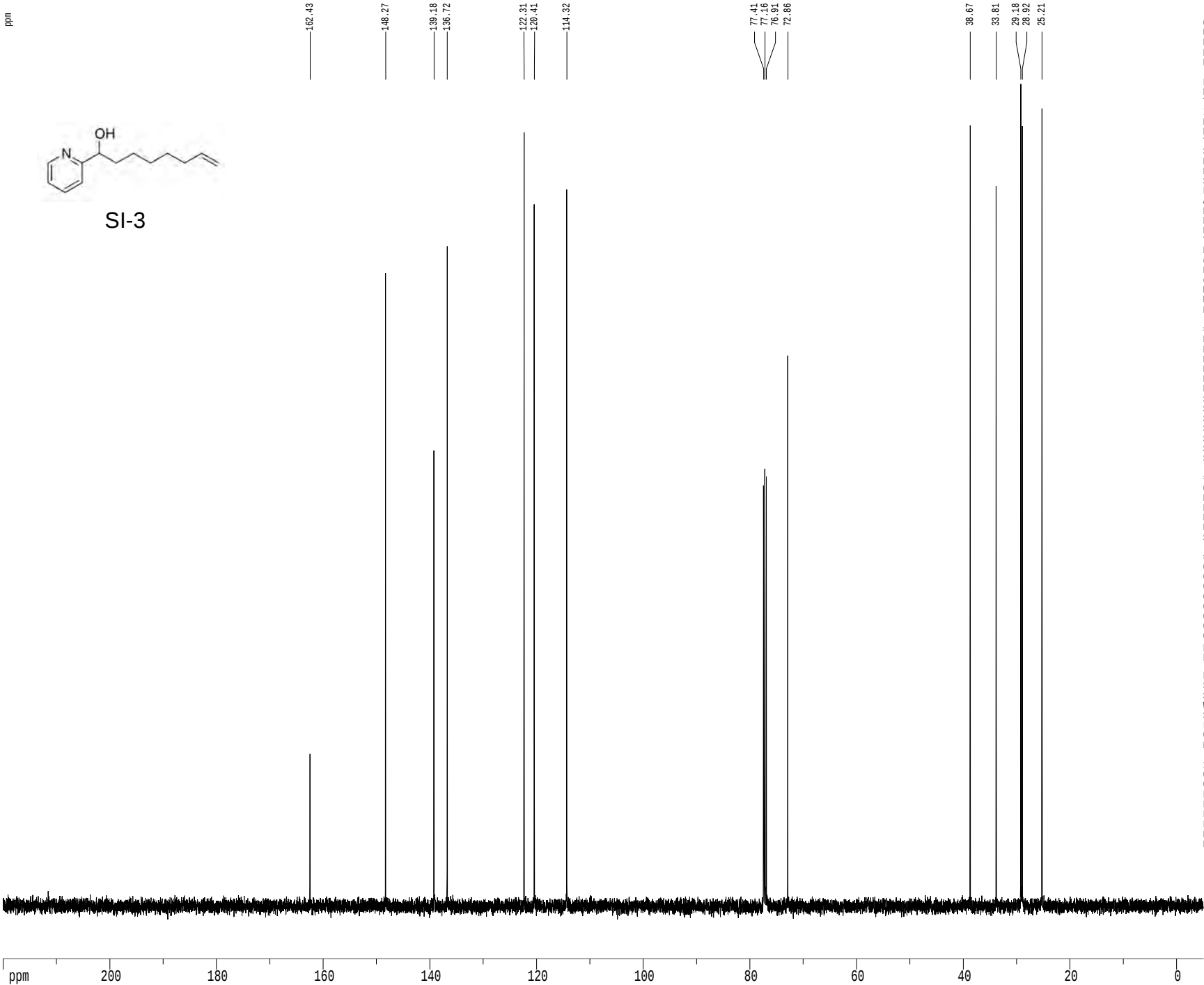
F2 - Acquisition Parameters
 Date_ 20150213
 Time 19.26
 INSTRUM cryo500
 PROBHD 5 mm CPTCI 1H-
 PULPROG zg30
 TO 81728
 SOLVENT CDCl3
 NS 8
 DS 2
 SWH 8012.820 Hz
 FIDRES 0.098043 Hz
 AQ 5.0998774 sec
 RG 6.3
 DW 62.400 usec
 DE 6.00 usec
 TE 298.0 K
 D1 0.10000000 sec
 MCREST 0.00000000 sec
 MCNWK 0.01500000 sec

===== CHANNEL f1 =====
 NUC1 1H
 P1 7.50 usec
 PL1 1.00 dB
 SF01 500.2235015 MHz

F2 - Processing parameters
 SI 65536
 SF 500.2200238 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 4.00

1D NMR plot parameters
 CX 22.00 cm
 CY 15.00 cm
 F1P 9.000 ppm
 F1 4501.98 Hz
 F2P -0.500 ppm
 F2 -250.11 Hz
 PPMCM 0.41667 ppm/cm
 HZCM 208.42502 Hz/cm

Z-restored spin-echo 13C spectrum with 1H decoupling



Current Data Parameters
USER mharri
NAME MRH-VII-223-13CNMR
EXPNO 1
PROCNO 1

F2 - Acquisition Parameters
Date_ 20150213
Time 19.28
INSTRUM cryo500
PROBHD 5 mm CPTCI 1H-
PULPROG SpinEchopg30gp.prd
TD 65536
SOLVENT CDCl3
NS 66
DS 16
SWH 30303.031 Hz
FIDRES 0.462388 Hz
AQ 1.0813940 sec
RG 3649.1
DW 16.500 usec
DE 6.00 usec
TE 298.0 K
D1 0.25000000 sec
d11 0.03000000 sec
D16 0.00020000 sec
d17 0.00019600 sec
MCREST 0.00000000 sec
MCWRR 0.01500000 sec
P2 33.10 usec

===== CHANNEL f1 =====
NUC1 13C
P1 16.55 usec
P11 500.00 usec
P12 2000.00 usec
PL0 120.00 dB
PL1 -1.00 dB
SF01 125.7942548 MHz
SP1 2.70 dB
SP2 2.70 dB
SPNAM1 Crp60,0.5,20.1
SPNAM2 Crp60comp.4
SPOFF1 0.00 Hz
SPOFF2 0.00 Hz

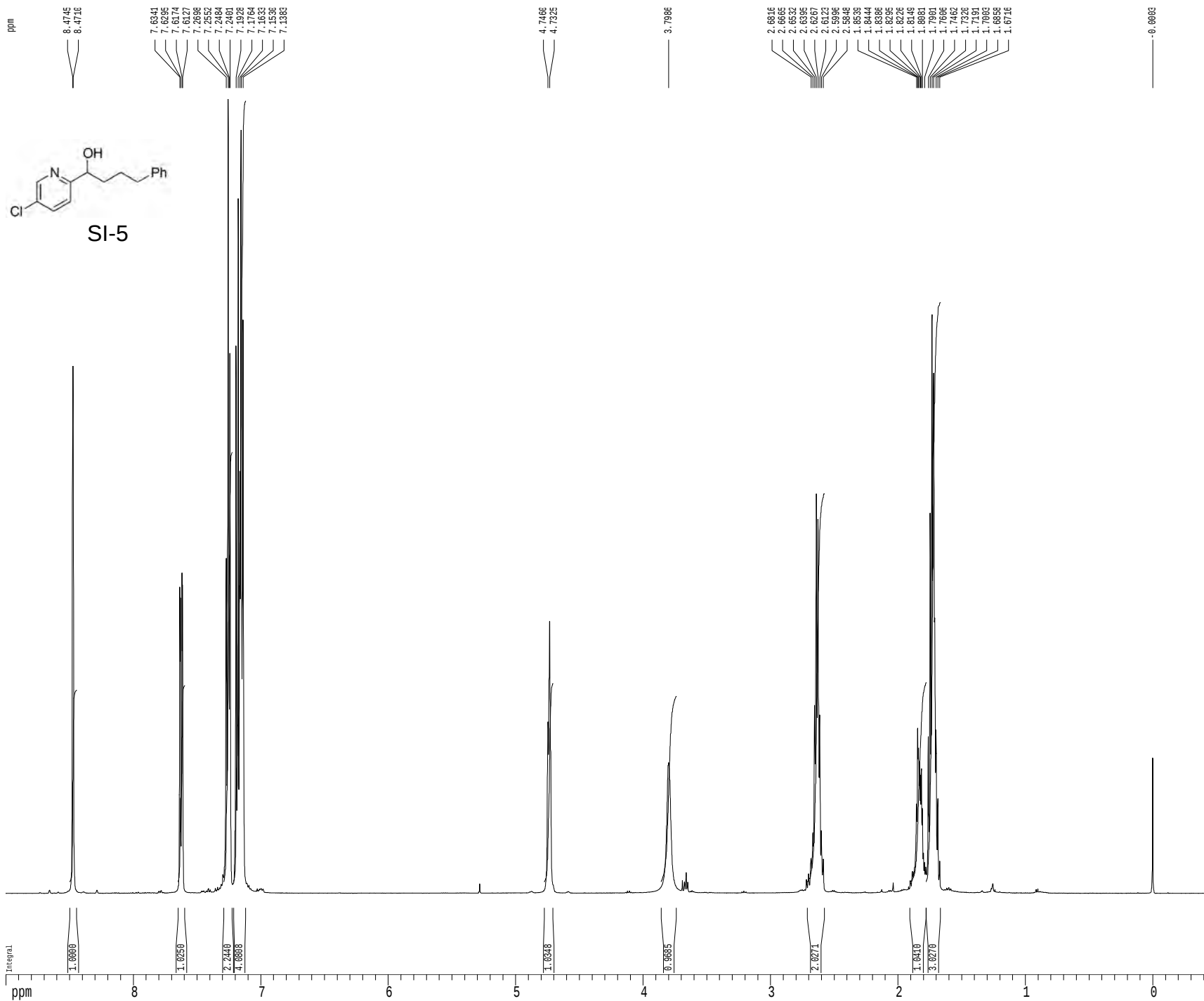
===== CHANNEL f2 =====
CPDPRG2 waltz16
NUC2 1H
PCPD2 100.00 usec
PL2 -1.00 dB
PL12 24.50 dB
SF02 500.2225011 MHz

===== GRADIENT CHANNEL =====
GPNAM1 SINE.100
GPNAM2 SINE.100
GPX1 0.00 %
GPX2 0.00 %
GPY1 0.00 %
GPY2 0.00 %
GPZ1 30.00 %
GPZ2 50.00 %
p15 500.00 usec
p16 1000.00 usec

F2 - Processing parameters
SI 65536
SF 125.7804113 MHz
EN
WDW 0
SSB 0
LB 1.00 Hz
GB 0
PC 2.00

1D NMR plot parameters
CX 22.00 cm
CY 15.65 cm
F1P 220.000 ppm
F1 27671.69 Hz
F2P -5.000 ppm
F2 -628.90 Hz
PPMCM 9.86842 ppm/cm
HZCM 1241.25415 Hz/cm

¹H spectrum



Current Data Parameters
 USER mharri
 NAME MRH-VII-269-1HNMR
 EXPNO 1
 PROCNO 1

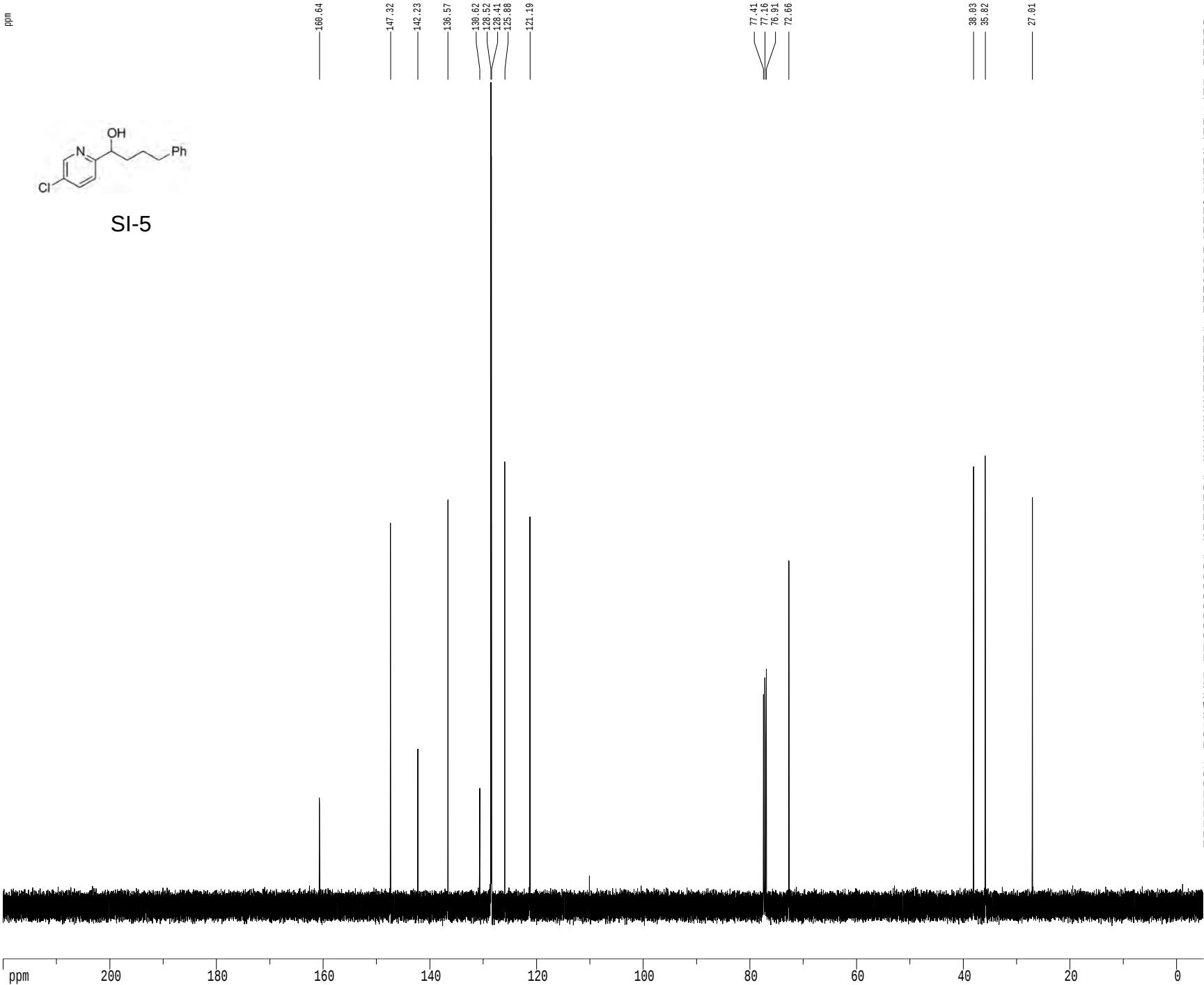
F2 - Acquisition Parameters
 Date_ 20150213
 Time 19.15
 INSTRUM cryo500
 PROBHD 5 mm CPTCI 1H-
 PULPROG zg30
 TO 81728
 SOLVENT CDCl3T
 NS 8
 DS 2
 SWH 8012.820 Hz
 FIDRES 0.098043 Hz
 AQ 5.0998774 sec
 RG 6.3
 DW 62.400 usec
 DE 6.00 usec
 TE 298.0 K
 D1 0.10000000 sec
 MCREST 0.00000000 sec
 MCWRR 0.01500000 sec

===== CHANNEL f1 =====
 NUC1 1H
 P1 7.50 usec
 PL1 1.00 dB
 SFO1 500.2235015 MHz

F2 - Processing parameters
 SI 65536
 SF 500.2200365 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 4.00

1D NMR plot parameters
 CX 22.00 cm
 CY 15.00 cm
 F1P 9.000 ppm
 F1 4501.98 Hz
 F2P -0.500 ppm
 F2 -250.11 Hz
 PPMCM 0.41667 ppm/cm
 HZCM 208.42502 Hz/cm

Z-restored spin-echo 13C spectrum with 1H decoupling



Current Data Parameters

USER mharri
NAME MRH-VII-269-13CNMR
EXPNO 1
PROCNO 1

F2 - Acquisition Parameters

Date_ 20150213
Time_ 19.17
INSTRUM cryo500
PROBHD 5 mm CPTCI 1H-
PULPROG SpinEchopg30gp.prd
TD 65536
SOLVENT CDCl3
NS 63
DS 16
SWH 30303.031 Hz
FIDRES 0.462388 Hz
AQ 1.0813940 sec
RG 2048
DW 16.500 usec
DE 6.00 usec
TE 298.0 K
D1 0.25000000 sec
d11 0.03000000 sec
D16 0.00020000 sec
d17 0.00019600 sec
MCREST 0.00000000 sec
MCWRR 0.01500000 sec
P2 33.10 usec

===== CHANNEL f1 =====

NUC1 13C
P1 16.55 usec
P11 500.00 usec
P12 2000.00 usec
PL0 120.00 dB
PL1 -1.00 dB
SF01 125.7942548 MHz
SP1 2.70 dB
SP2 2.70 dB
SPNAM1 Crp60,0.5,20.1
SPNAM2 Crp60comp.4
SPOFF1 0.00 Hz
SPOFF2 0.00 Hz

===== CHANNEL f2 =====

CPDPRG2 waltz16
NUC2 1H
PCPD2 100.00 usec
PL2 1.00 dB
PL12 24.50 dB
SF02 500.2225011 MHz

===== GRADIENT CHANNEL =====

GPVAM1 SINE.100
GPVAM2 SINE.100
GPX1 0.00 %
GPX2 0.00 %
GPY1 0.00 %
GPY2 0.00 %
GPZ1 30.00 %
GPZ2 50.00 %
p15 500.00 usec
p16 1000.00 usec

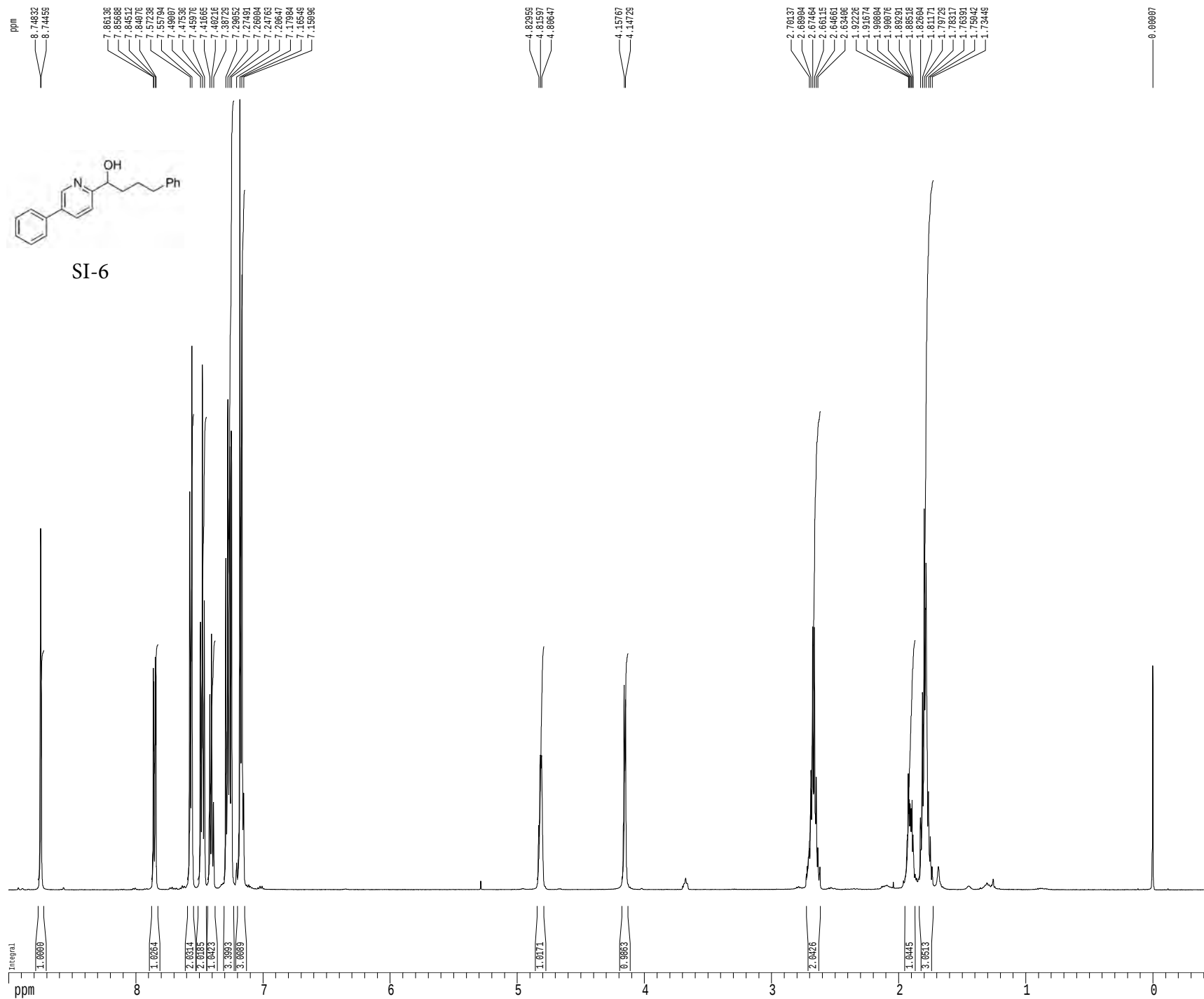
F2 - Processing parameters

SI 65536
SF 125.7804130 MHz
EN
WDW 0
SSB 0
LB 1.00 Hz
GB 0
PC 2.00

1D NMR plot parameters

CX 22.00 cm
CY 15.65 cm
F1P 220.000 ppm
F1 27671.69 Hz
F2P -5.000 ppm
F2 -628.90 Hz
PPMCM 9.86842 ppm/cm
HZCM 1241.25415 Hz/cm

¹H spectrum



```

Current Data Parameters
USER      mharri
NAME      MRH-VII-258-1HNMR
EXPNO     1
PROCNO    1

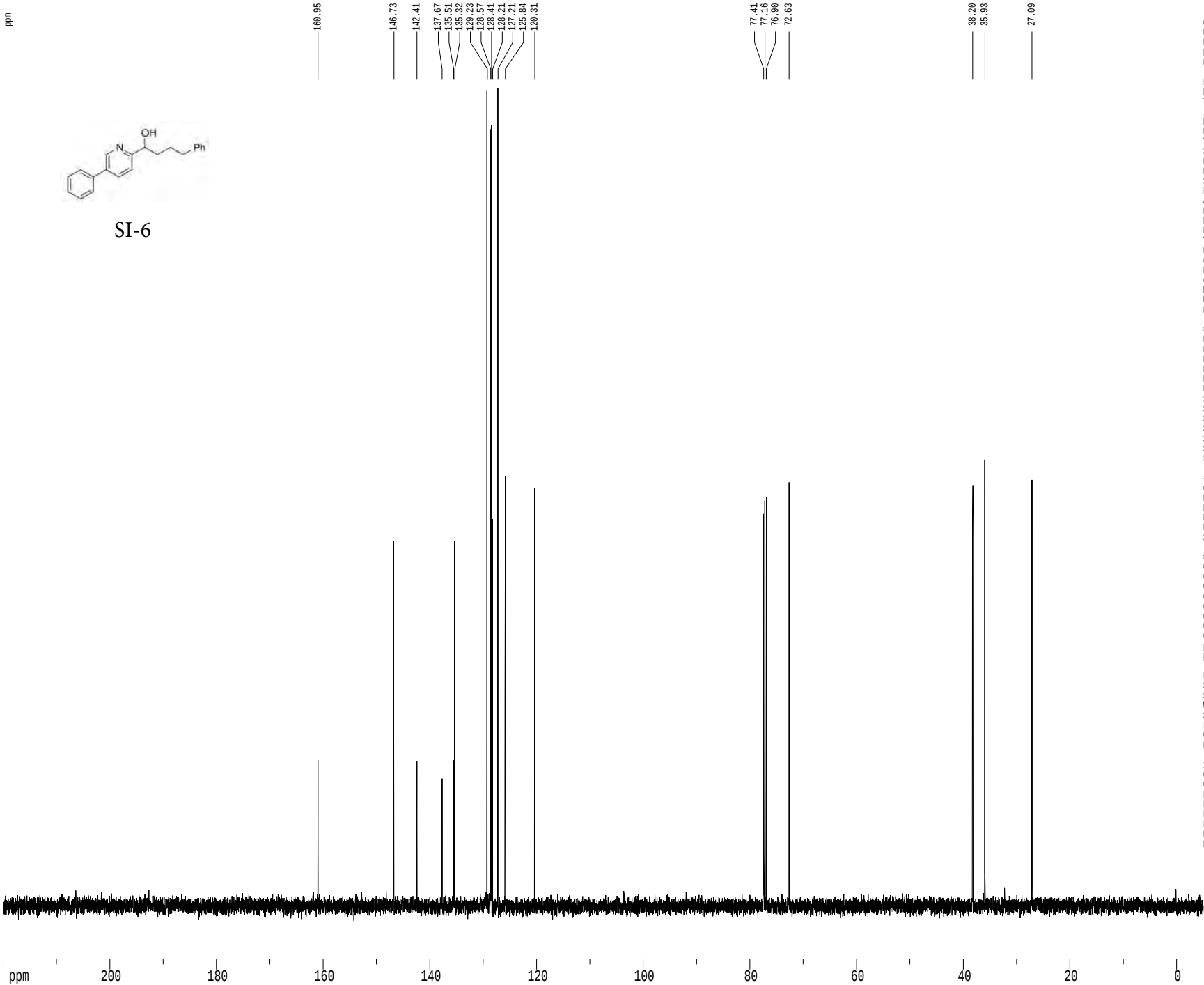
F2 - Acquisition Parameters
Date_     20150213
Time      19.02
INSTRUM    cryo500
PROBHD     5 mm CPTCI 1H-
PULPROG    zg30
TD         81728
SOLVENT    CDCl3
NS         8
DS         2
SWH         8012.820 Hz
FIDRES     0.098043 Hz
AQ         5.0998774 sec
RG         6.3
DW         62.400 usec
DE         6.00 usec
TE         298.0 K
D1         0.10000000 sec
MCREST     0.00000000 sec
MCNRK      0.01500000 sec

===== CHANNEL f1 =====
NUC1       1H
P1         7.50 usec
PL1        1.00 dB
SF01       500.2235015 MHz

F2 - Processing parameters
SI         65536
SF         500.2200366 MHz
WDW        EM
SSB        0
LB         0.30 Hz
GB         0
PC         4.00

1D NMR plot parameters
CX         22.00 cm
CY         15.00 cm
F1P        9.000 ppm
F1         4501.98 Hz
F2P        -0.500 ppm
F2         -250.11 Hz
PPMCM      0.41667 ppm/cm
HZCM       208.42502 Hz/cm
    
```

Z-restored spin-echo 13C spectrum with 1H decoupling



Current Data Parameters

USER mharri
NAME MRH-VII-258-13CNMR
EXPNO 1
PROCNO 1

F2 - Acquisition Parameters

Date_ 20150213
Time 19.05
INSTRUM cryo500
PROBHD 5 mm CPTCI 1H-
PULPROG SpinEchopg30gp.prd
TD 65536
SOLVENT CDCl3
NS 92
DS 16
SWH 30303.031 Hz
FIDRES 0.462388 Hz
AQ 1.0813940 sec
RG 6502
DW 16.500 usec
DE 6.00 usec
TE 298.0 K
D1 0.25000000 sec
d11 0.03000000 sec
D16 0.00020000 sec
d17 0.00019600 sec
MCREST 0.00000000 sec
MCWRR 0.01500000 sec
P2 33.10 usec

===== CHANNEL f1 =====

NUC1 13C
P1 16.55 usec
P11 500.00 usec
P12 2000.00 usec
PL0 120.00 dB
PL1 -1.00 dB
SF01 125.7942548 MHz
SP1 2.70 dB
SP2 2.70 dB
SPNAM1 Crp60,0.5,20.1
SPNAM2 Crp60comp.4
SPOFF1 0.00 Hz
SPOFF2 0.00 Hz

===== CHANNEL f2 =====

CPDPRG2 waltz16
NUC2 1H
PCPD2 100.00 usec
PL2 1.00 dB
PL12 24.50 dB
SF02 500.2225011 MHz

===== GRADIENT CHANNEL =====

GP1AM1 SINE.100
GP1AM2 SINE.100
GPX1 0.00 %
GPX2 0.00 %
GPY1 0.00 %
GPY2 0.00 %
GPZ1 30.00 %
GPZ2 50.00 %
p15 500.00 usec
p16 1000.00 usec

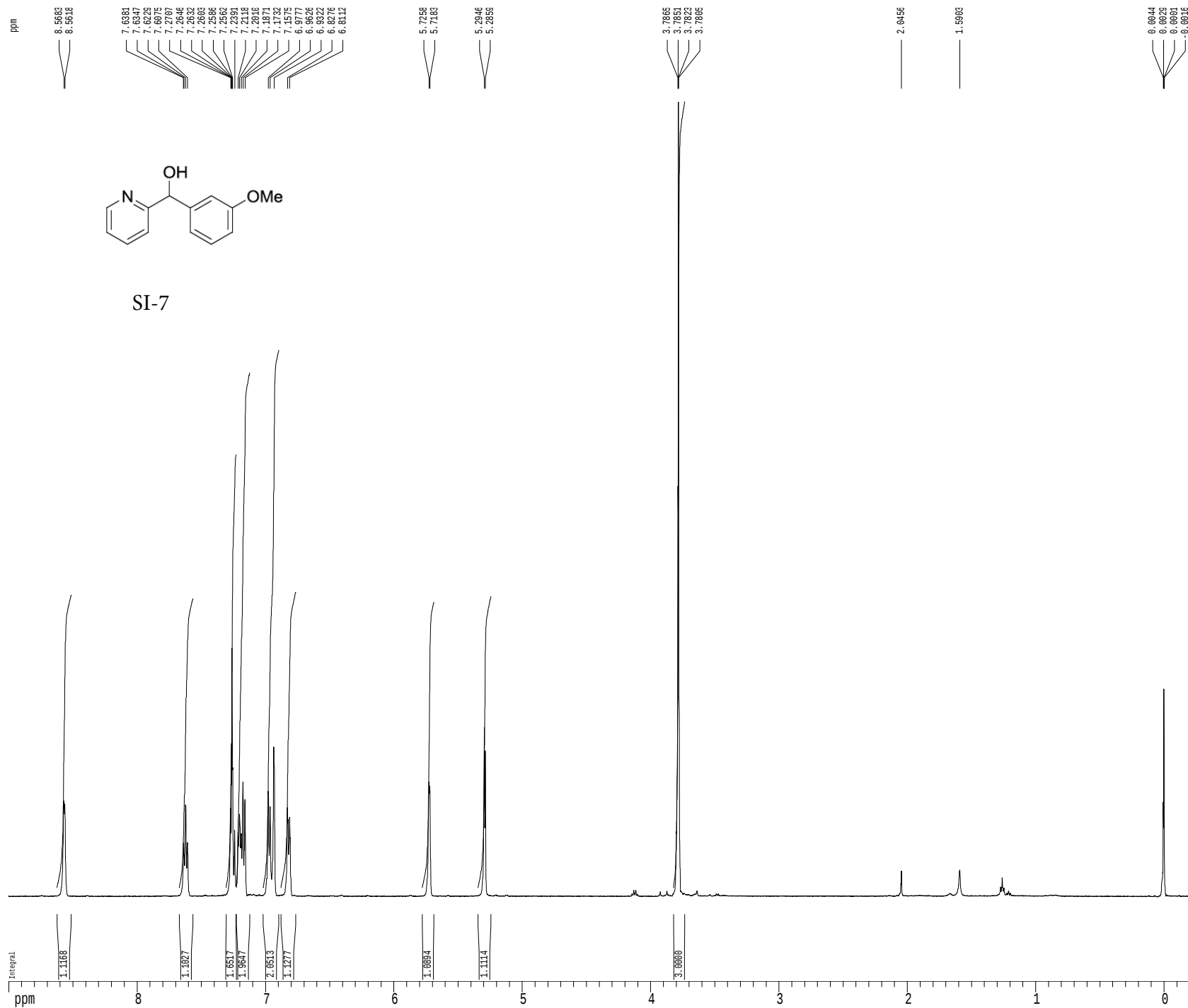
F2 - Processing parameters

SI 65536
SF 125.7804117 MHz
EN
WDW 0
SSB 0
LB 1.00 Hz
GB 0
PC 2.00

1D NMR plot parameters

CX 22.00 cm
CY 15.65 cm
F1P 220.000 ppm
F1 27671.69 Hz
F2P -5.000 ppm
F2 -628.90 Hz
PPMCM 9.86842 ppm/cm
HZCM 1241.25415 Hz/cm

¹H spectrum



```

Current Data Parameters
USER          lhanna
NAME          LEH-5-037-H1
EXPNO         1
PROCNO        1

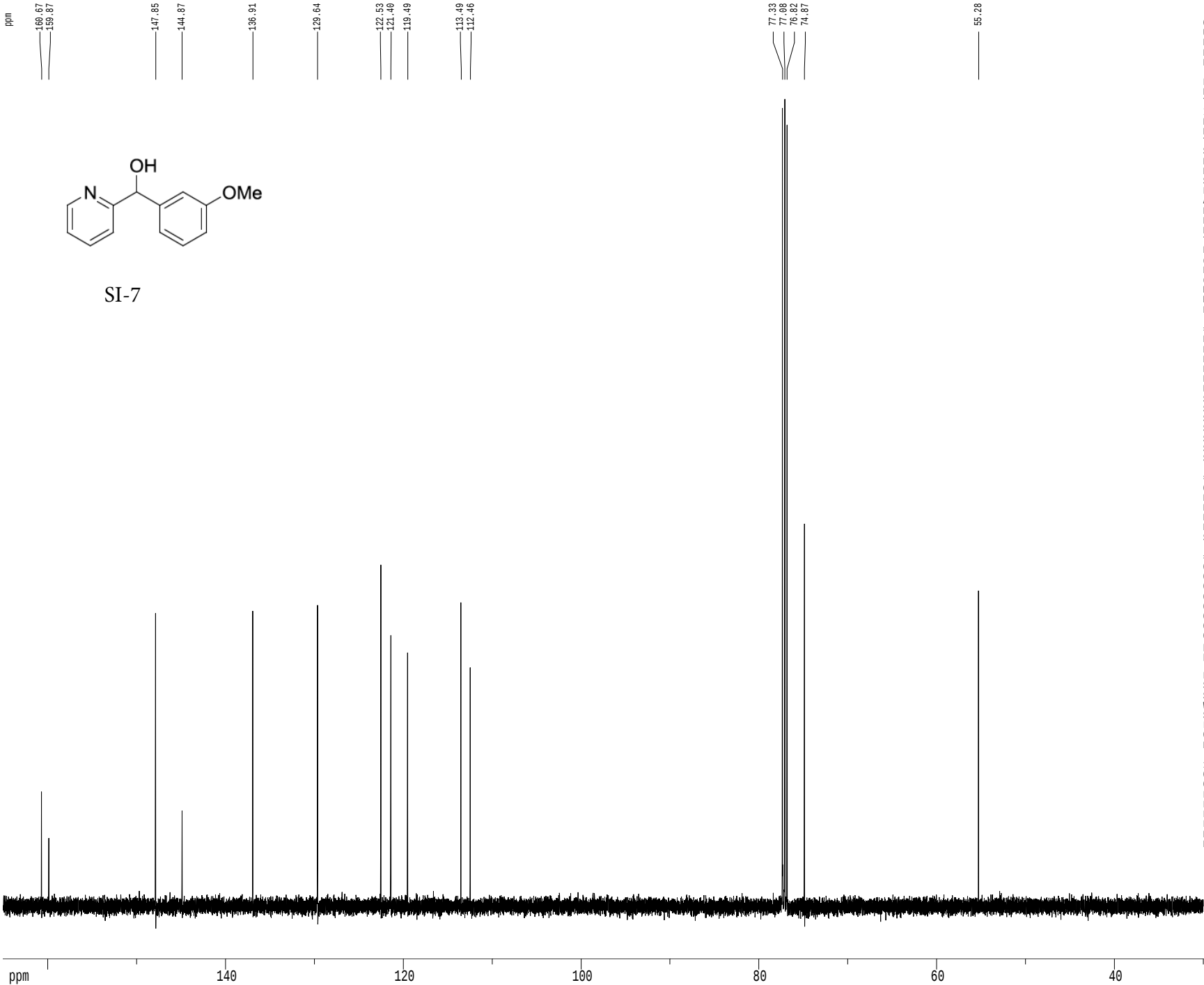
F2 - Acquisition Parameters
Date_         20150114
Time          18.24
INSTRUM       cryo500
PROBHD        5 mm CPTCI 1H-
PULPROG       zg30
TD            32648
SOLVENT       CDCl3
NS            8
DS            2
SWH           8012.820 Hz
FIDRES        0.250026 Hz
AQ            1.9998451 sec
RG            5.7
DW            62.400 usec
DE            6.00 usec
TE            298.0 K
D1            0.10000000 sec
MCREST        0.00000000 sec
MCWRK         0.01500000 sec

===== CHANNEL f1 =====
NUC1           1H
P1             7.50 usec
PL1            1.60 dB
SF01          500.2235015 MHz

F2 - Processing parameters
SI            65536
SF            500.2200314 MHz
WDW           no
SSB           0
LB            0.00 Hz
GB            0
PC            4.00

1D NMR plot parameters
CX            22.00 cm
CY            15.00 cm
F1P           9.000 ppm
F1            4501.98 Hz
F2P          -0.500 ppm
F2            -250.11 Hz
PPMCM         0.41667 ppm/cm
HZCM          208.42502 Hz/cm
    
```

Z-restored spin-echo 13C spectrum with 1H decoupling



Current Data Parameters

USER lhanna
NAME LEH-5-037-C13
EXPNO 1
PROCNO 1

F2 - Acquisition Parameters

Date_ 20150114
Time 18.28
INSTRUM cryo500
PROBHD 5 mm CPTCI 1H-
PULPROG SpinEchopg30gp.prd
TD 65536
SOLVENT CDCl3
NS 513
DS 16
SWH 30303.031 Hz
FIDRES 0.462388 Hz
AQ 1.0813940 sec
RG 7298.2
DW 16.500 usec
DE 6.00 usec
TE 298.0 K
D1 0.25000000 sec
d11 0.03000000 sec
D16 0.00020000 sec
d17 0.00019600 sec
MCREST 0.00000000 sec
MCWRK 0.01500000 sec
P2 33.10 usec

===== CHANNEL f1 =====

NUC1 13C
P1 16.55 usec
P11 500.00 usec
P12 2000.00 usec
PL0 120.00 dB
PL1 -1.00 dB
SF01 125.7942548 MHz
SP1 2.70 dB
SP2 2.70 dB
SPNAM1 Crp60,0.5,20.1
SPNAM2 Crp60comp.4
SPOFF1 0.00 Hz
SPOFF2 0.00 Hz

===== CHANNEL f2 =====

CPDPRG2 waltz16
NUC2 1H
PCPD2 100.00 usec
PL2 1.00 dB
PL12 24.50 dB
SF02 500.2225011 MHz

===== GRADIENT CHANNEL =====

GP1AM1 SINE.100
GP1AM2 SINE.100
GPX1 0.00 %
GPX2 0.00 %
GPY1 0.00 %
GPY2 0.00 %
GPZ1 30.00 %
GPZ2 50.00 %
p15 500.00 usec
p16 1000.00 usec

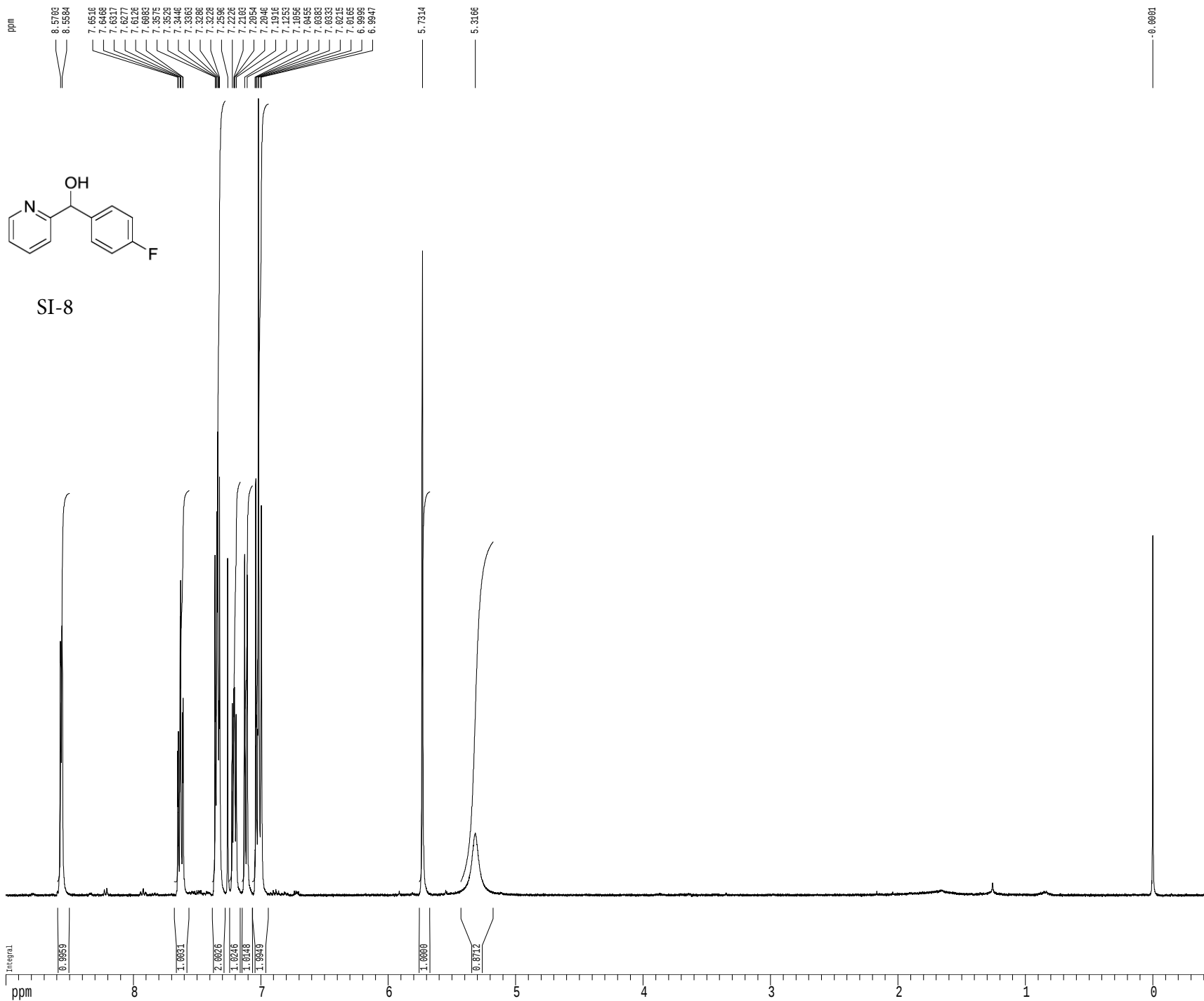
F2 - Processing parameters

SI 65536
SF 125.7804190 MHz
WDW NO
SSB 0
LB 0.00 Hz
GB 0
PC 2.00

1D NMR plot parameters

CX 22.00 cm
CY 15.65 cm
F1P 165.000 ppm
F1 20753.77 Hz
F2P 30.000 ppm
F2 3773.41 Hz
PPMCM 5.92105 ppm/cm
HZCM 744.75250 Hz/cm

¹H spectrum



Current Data Parameters
 USER Inama
 NAME LEH-5-149-2
 EXPNO 1
 PROCNO 1

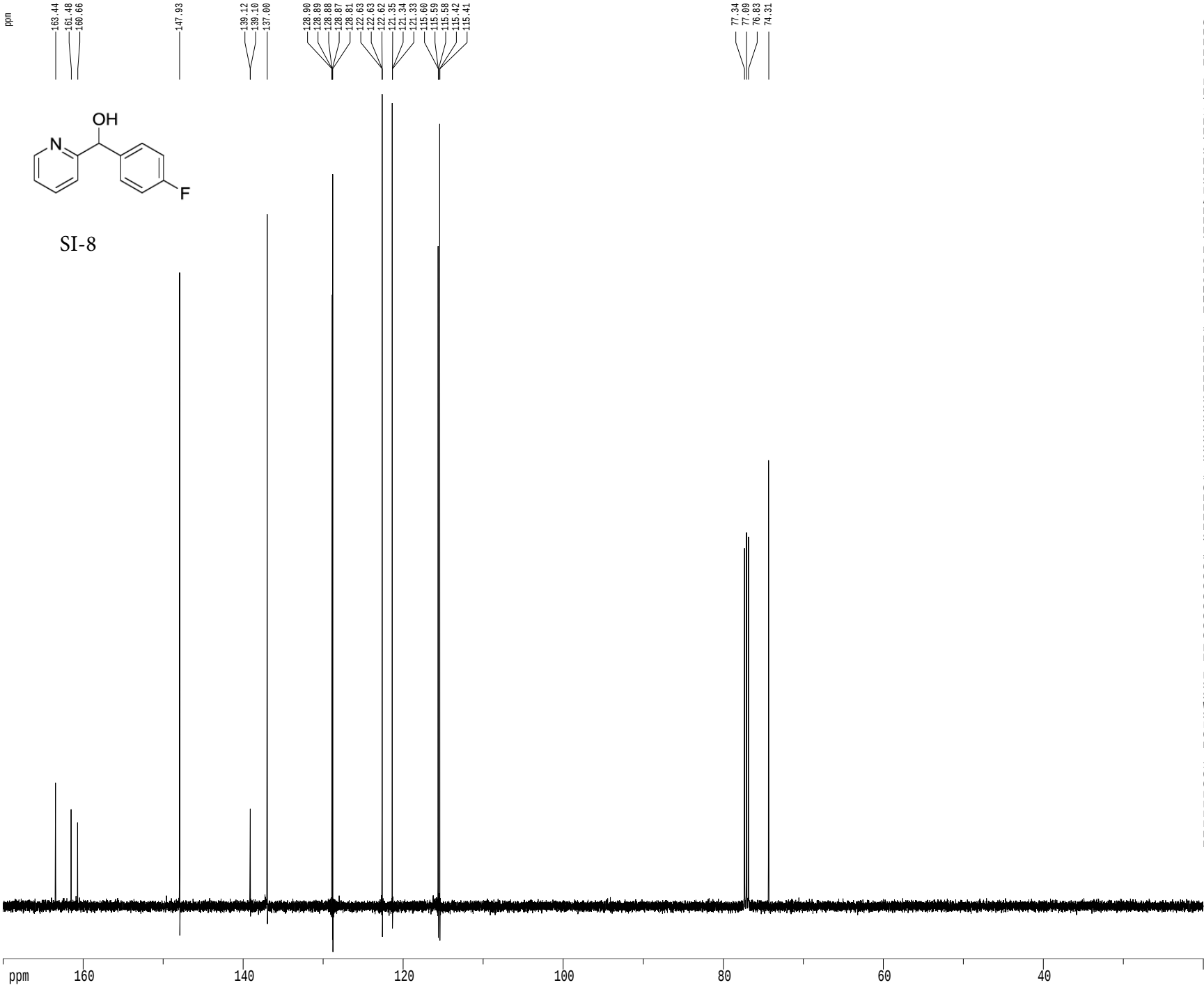
F2 - Acquisition Parameters
 Date_ 20150320
 Time 13.41
 INSTRUM drx400
 PROBHD 5 mm QNP H/F/P
 PULPROG zg30
 TD 65536
 SOLVENT CDCl3
 NS 8
 DS 2
 SWH 6410.256 Hz
 FIDRES 0.097813 Hz
 AQ 5.1118579 sec
 RG 287.4
 DW 78.000 usec
 DE 4.50 usec
 TE 298.0 K
 D1 0.10000000 sec
 MCREST 0.00000000 sec
 MCWRK 0.01500000 sec

===== CHANNEL f1 =====
 NUC1 1H
 P1 12.00 usec
 PL1 0.00 dB
 SF01 400.1328009 MHz

F2 - Processing parameters
 SI 65536
 SF 400.1300211 MHz
 WDW no
 SSB 0
 LB 0.00 Hz
 GB 0
 PC 2.00

1D NMR plot parameters
 CX 22.00 cm
 CY 15.00 cm
 F1P 9.000 ppm
 F1 3691.17 Hz
 F2P -0.500 ppm
 F2 -200.06 Hz
 PPMCM 0.41667 ppm/cm
 HZCM 166.72086 Hz/cm

Z-restored spin-echo 13C spectrum with 1H decoupling



Current Data Parameters

USER lhanna
NAME LEH-5-149-c13
EXPNO 1
PROCNO 1

F2 - Acquisition Parameters

Date_ 20150320
Time 14.42
INSTRUM cryo500
PROBHD 5 mm CPTCI 1H-
PULPROG SpinEchopg30gp.prd
TD 65536
SOLVENT CDCl3
NS 594
DS 16
SWH 30303.031 Hz
FIDRES 0.462388 Hz
AQ 1.0813940 sec
RG 7298.2
DW 16.500 usec
DE 6.00 usec
TE 298.0 K
D1 0.25000000 sec
d11 0.03000000 sec
D16 0.00020000 sec
d17 0.00019600 sec
MCREST 0.00000000 sec
MCWRK 0.01500000 sec
P2 33.10 usec

===== CHANNEL f1 =====

NUC1 13C
P1 16.55 usec
P11 500.00 usec
P12 2000.00 usec
PL0 120.00 dB
PL1 -1.00 dB
SF01 125.7942548 MHz
SP1 2.70 dB
SP2 2.70 dB
SPNAM1 Crp60,0.5,20.1
SPNAM2 Crp60comp,4
SPOFF1 0.00 Hz
SPOFF2 0.00 Hz

===== CHANNEL f2 =====

CPDPRG2 waltz16
NUC2 1H
PCPD2 100.00 usec
PL2 1.00 dB
PL12 24.50 dB
SF02 500.2225011 MHz

===== GRADIENT CHANNEL =====

GPVAM1 SINE.100
GPVAM2 SINE.100
GPX1 0.00 %
GPX2 0.00 %
GPY1 0.00 %
GPY2 0.00 %
GPZ1 30.00 %
GPZ2 50.00 %
p15 500.00 usec
p16 1000.00 usec

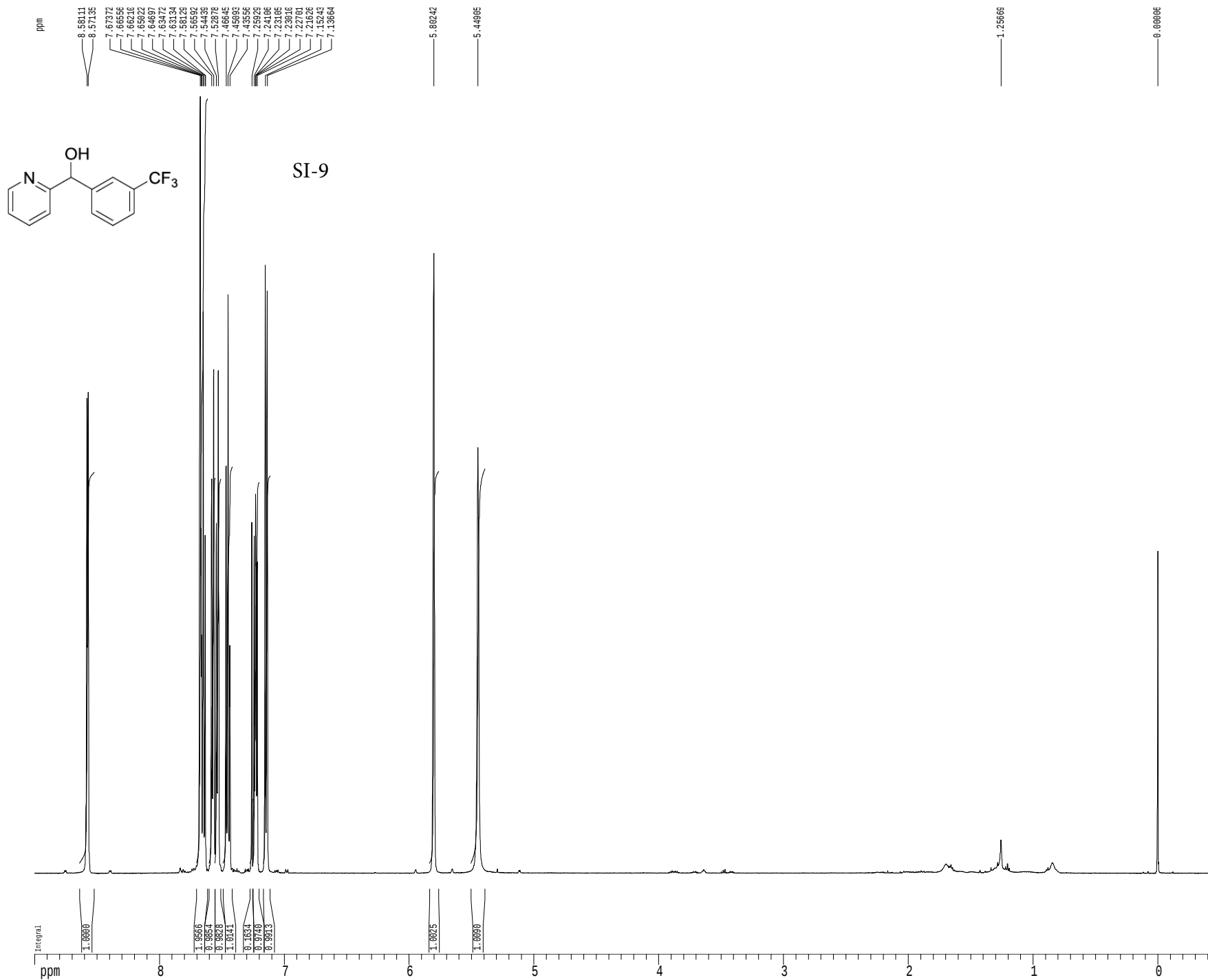
F2 - Processing parameters

SI 65536
SF 125.7804190 MHz
WDW no
SSB 0
LB 0.00 Hz
GB 0
PC 2.00

1D NMR plot parameters

CX 22.00 cm
CY 15.65 cm
F1P 170.000 ppm
F1 21382.67 Hz
F2P 20.000 ppm
F2 2515.61 Hz
PPMCM 6.57895 ppm/cm
HZCM 827.50281 Hz/cm

¹H spectrum



Current Data Parameters
 USER lhanna
 NAME LEH-5-067-H1
 EXPNO 1
 PROCNO 1

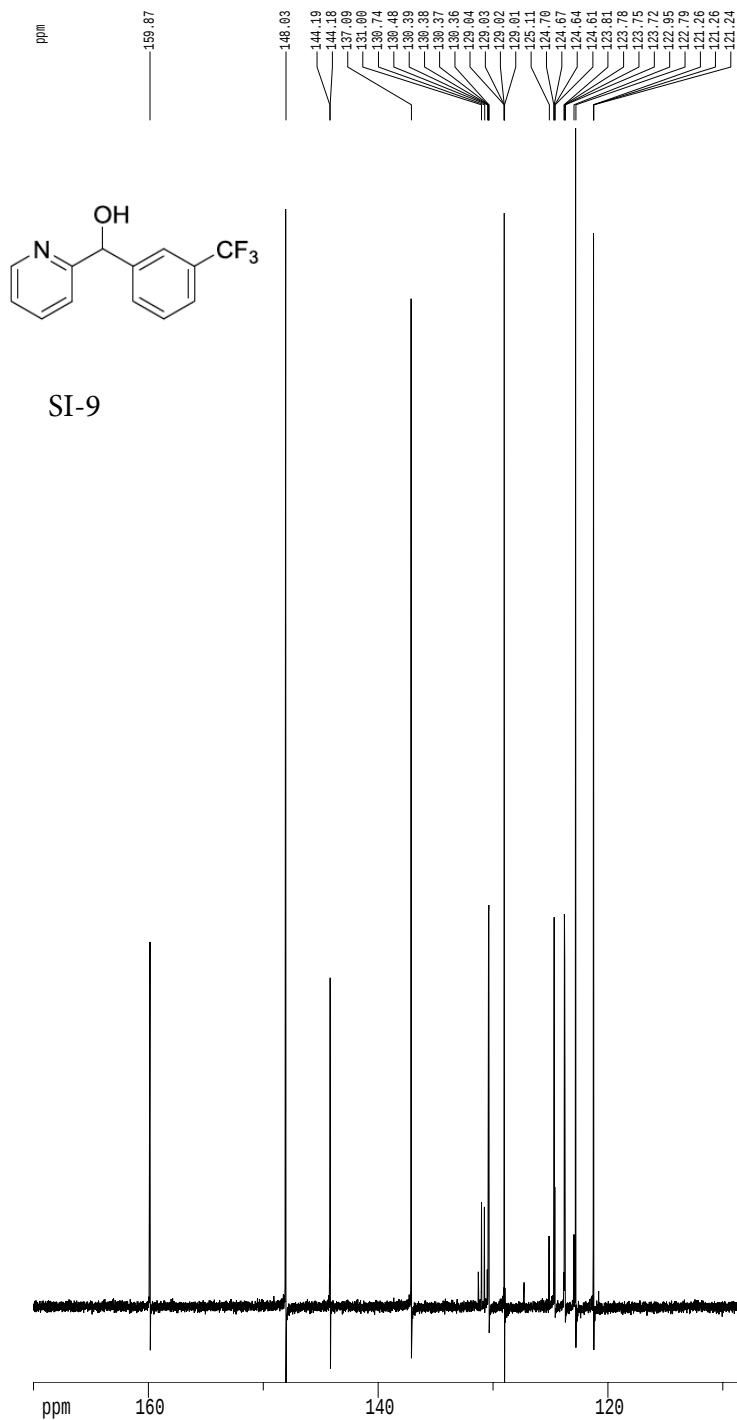
F2 - Acquisition Parameters
 Date_ 20150122
 Time 18.35
 INSTRUM cryo500
 PROBHD 5 mm CPTCI 1H-
 PULPROG zg30
 TD 32648
 SOLVENT CDCl3
 NS 8
 DS 2
 SWH 8012.820 Hz
 FIDRES 0.250026 Hz
 AQ 1.9998451 sec
 RG 10.1
 DW 62.400 usec
 DE 6.00 usec
 TE 298.0 K
 D1 0.10000000 sec
 MCREST 0.00000000 sec
 MCWRR 0.01500000 sec

===== CHANNEL f1 =====
 NUC1 1H
 P1 7.50 usec
 PL1 1.60 dB
 SF01 500.2235015 MHz

F2 - Processing parameters
 SI 65536
 SF 500.2200315 MHz
 WDW no
 SSB 0
 LB 0.00 Hz
 GB 0
 PC 4.00

1D NMR plot parameters
 CX 22.80 cm
 CY 15.00 cm
 F1P 9.000 ppm
 F1 4501.98 Hz
 F2P -0.500 ppm
 F2 -250.11 Hz
 PPMCM 0.41667 ppm/cm
 HZCM 208.42502 Hz/cm

Z-restored spin-echo 13C spectrum with 1H decoupling



Current Data Parameters

USER	lhanna
NAME	LEH-5-067-C13
EXPNO	1
PROCNO	1

F2 - Acquisition Parameters

Date_	20150122
Time	18.41
INSTRUM	cryo500
PROBHD	5 mm CPTCI 1H-
PULPROG	SpinEchopg30gp.prd
TD	65536
SOLVENT	CDCl3
NS	771
DS	16
SWH	30303.031 Hz
FIDRES	0.462388 Hz
AQ	1.0813940 sec
RG	11585.2
DW	16.500 usec
DE	6.00 usec
TE	298.0 K
D1	0.25000000 sec
d11	0.03000000 sec
D16	0.00020000 sec
d17	0.00019600 sec
MCREST	0.00000000 sec
MCWRK	0.01500000 sec
P2	33.10 usec

===== CHANNEL f1 =====

NUC1	13C
P1	16.55 usec
P11	500.00 usec
P12	2000.00 usec
PL0	120.00 dB
PL1	-1.00 dB
SFO1	125.7942548 MHz
SP1	2.70 dB
SP2	2.70 dB
SPNAM1	Crp60,0.5,20.1
SPNAM2	Crp60comp.4
SPOFF1	0.00 Hz
SPOFF2	0.00 Hz

===== CHANNEL f2 =====

CPDPRG2	waltz16
NUC2	1H
PCPD2	100.00 usec
PL2	-1.00 dB
PL12	24.50 dB
SFO2	500.2225011 MHz

===== GRADIENT CHANNEL =====

GPVAM1	SINE.100
GPVAM2	SINE.100
GPX1	0.00 %
GPX2	0.00 %
GPY1	0.00 %
GPY2	0.00 %
GPZ1	30.00 %
GPZ2	50.00 %
p15	500.00 usec
p16	1000.00 usec

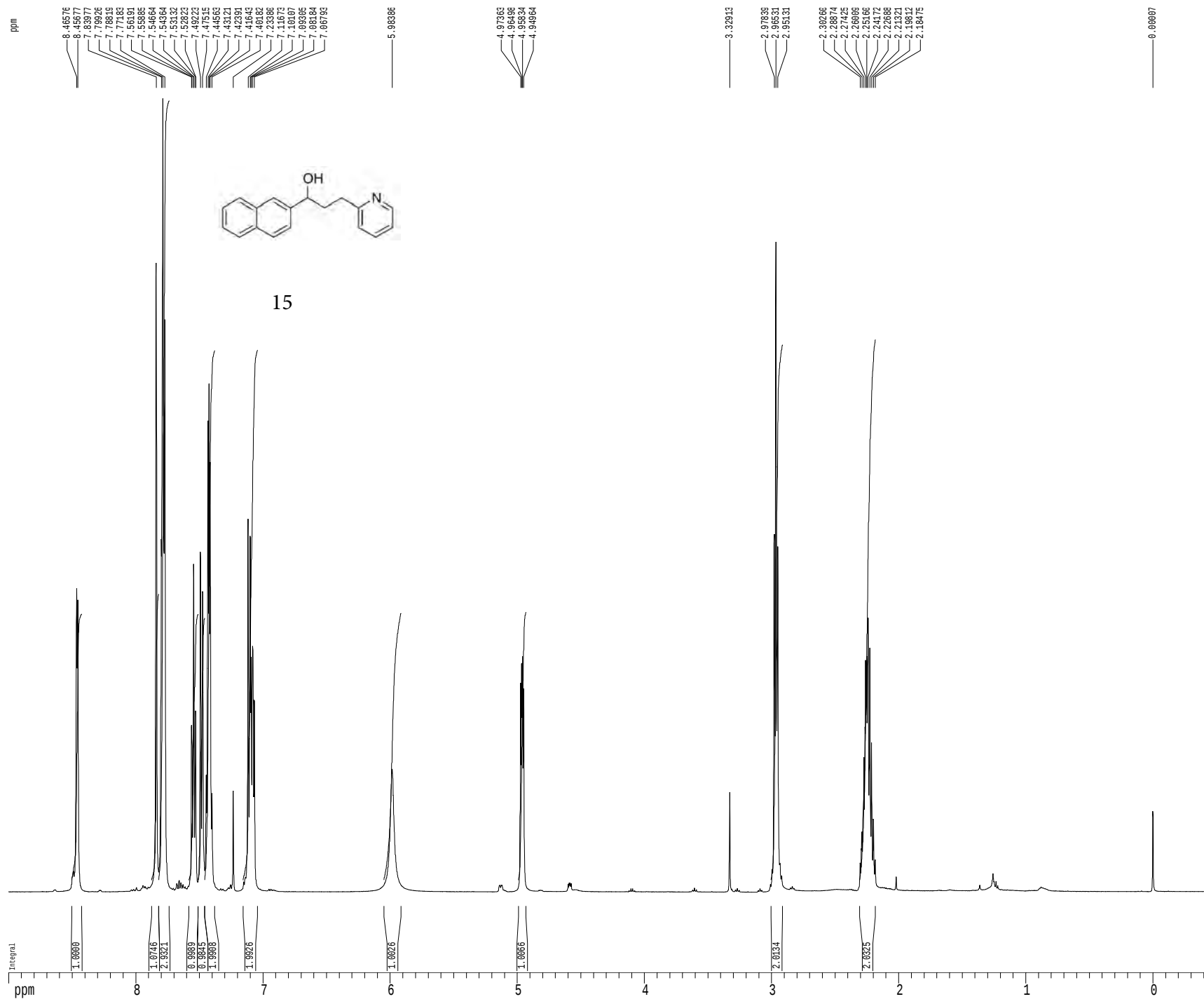
F2 - Processing parameters

SI	65536
SF	125.7804298 MHz
WDW	no
SSB	0
LB	0.00 Hz
GB	0
PC	2.00

1D NMR plot parameters

CX	22.00 cm
CY	15.65 cm
F1P	170.000 ppm
F1	21382.67 Hz
F2P	20.000 ppm
F2	2515.61 Hz
PPMCM	6.57895 ppm/cm
HZCM	827.50281 Hz/cm

¹H spectrum



Current Data Parameters
 USER mharri
 NAME MRH-VII-236-1HNMR
 EXPNO 2
 PROCNO 1

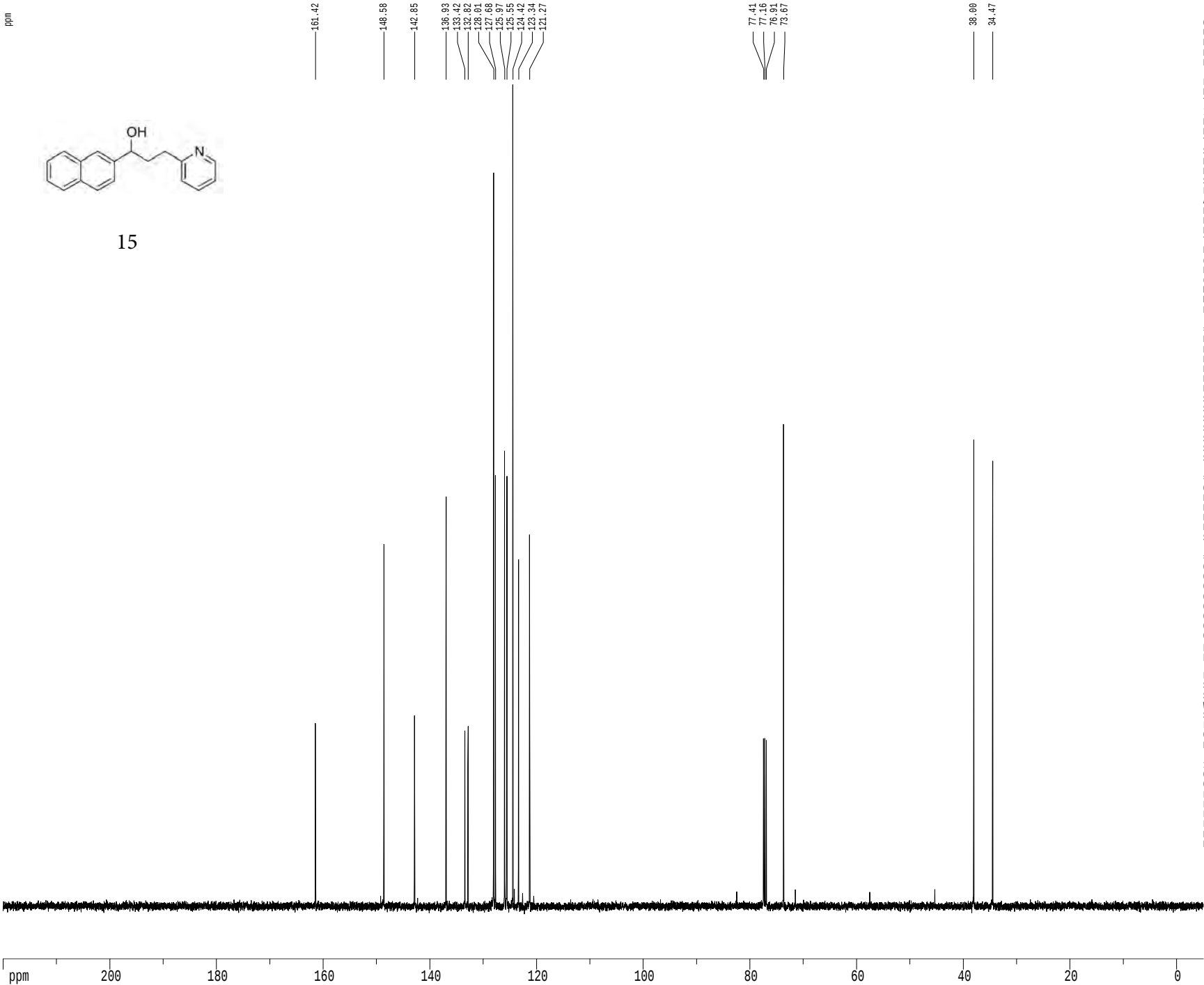
F2 - Acquisition Parameters
 Date_ 20150223
 Time 17.13
 INSTRUM cryo500
 PROBHD 5 mm CPTCI 1H-
 PULPROG zg30
 TO 81728
 SOLVENT CDCl3
 NS 8
 DS 2
 SWH 8012.820 Hz
 FIDRES 0.098043 Hz
 AQ 5.0998774 sec
 RG 5
 DW 62.400 usec
 DE 6.00 usec
 TE 298.0 K
 D1 0.10000000 sec
 MCREST 0.00000000 sec
 MCNRK 0.01500000 sec

===== CHANNEL f1 =====
 NUC1 1H
 P1 7.50 usec
 PL1 1.00 dB
 SFO1 500.2235015 MHz

F2 - Processing parameters
 SI 65536
 SF 500.2200448 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 4.00

1D NMR plot parameters
 CX 22.00 cm
 CY 15.00 cm
 F1P 9.000 ppm
 F1 4501.98 Hz
 F2P -0.500 ppm
 F2 -250.11 Hz
 PPMCM 0.41667 ppm/cm
 HZCM 208.42502 Hz/cm

Z-restored spin-echo 13C spectrum with 1H decoupling



Current Data Parameters

USER mharri
NAME MRH-VII-236-13CNMR
EXPNO 2
PROCNO 1

F2 - Acquisition Parameters

Date_ 20150223
Time 17.15
INSTRUM cryo500
PROBHD 5 mm CPTCI 1H-
PULPROG SpinEchopg30gp.prd
TD 65536
SOLVENT CDCl3
NS 48
DS 16
SWH 30303.031 Hz
FIDRES 0.462388 Hz
AQ 1.0813940 sec
RG 7298.2
DW 16.500 usec
DE 6.00 usec
TE 298.0 K
D1 0.25000000 sec
d11 0.03000000 sec
D16 0.00020000 sec
d17 0.00019600 sec
MCREST 0.00000000 sec
MCMRK 0.01500000 sec
P2 33.10 usec

===== CHANNEL f1 =====

NUC1 13C
P1 16.55 usec
P11 500.00 usec
P12 2000.00 usec
PL0 120.00 dB
PL1 -1.00 dB
SF01 125.7942548 MHz
SP1 2.70 dB
SP2 2.70 dB
SPNAM1 Crp60,0.5,20.1
SPNAM2 Crp60comp.4
SPOFF1 0.00 Hz
SPOFF2 0.00 Hz

===== CHANNEL f2 =====

CPDPRG2 waltz16
NUC2 1H
PCPD2 100.00 usec
PL2 1.00 dB
PL12 24.50 dB
SF02 500.2225011 MHz

===== GRADIENT CHANNEL =====

GPVAM1 SINE.100
GPVAM2 SINE.100
GPX1 0.00 %
GPX2 0.00 %
GPY1 0.00 %
GPY2 0.00 %
GPZ1 30.00 %
GPZ2 50.00 %
p15 500.00 usec
p16 1000.00 usec

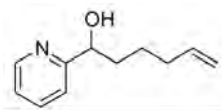
F2 - Processing parameters

SI 65536
SF 125.7804214 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 2.00

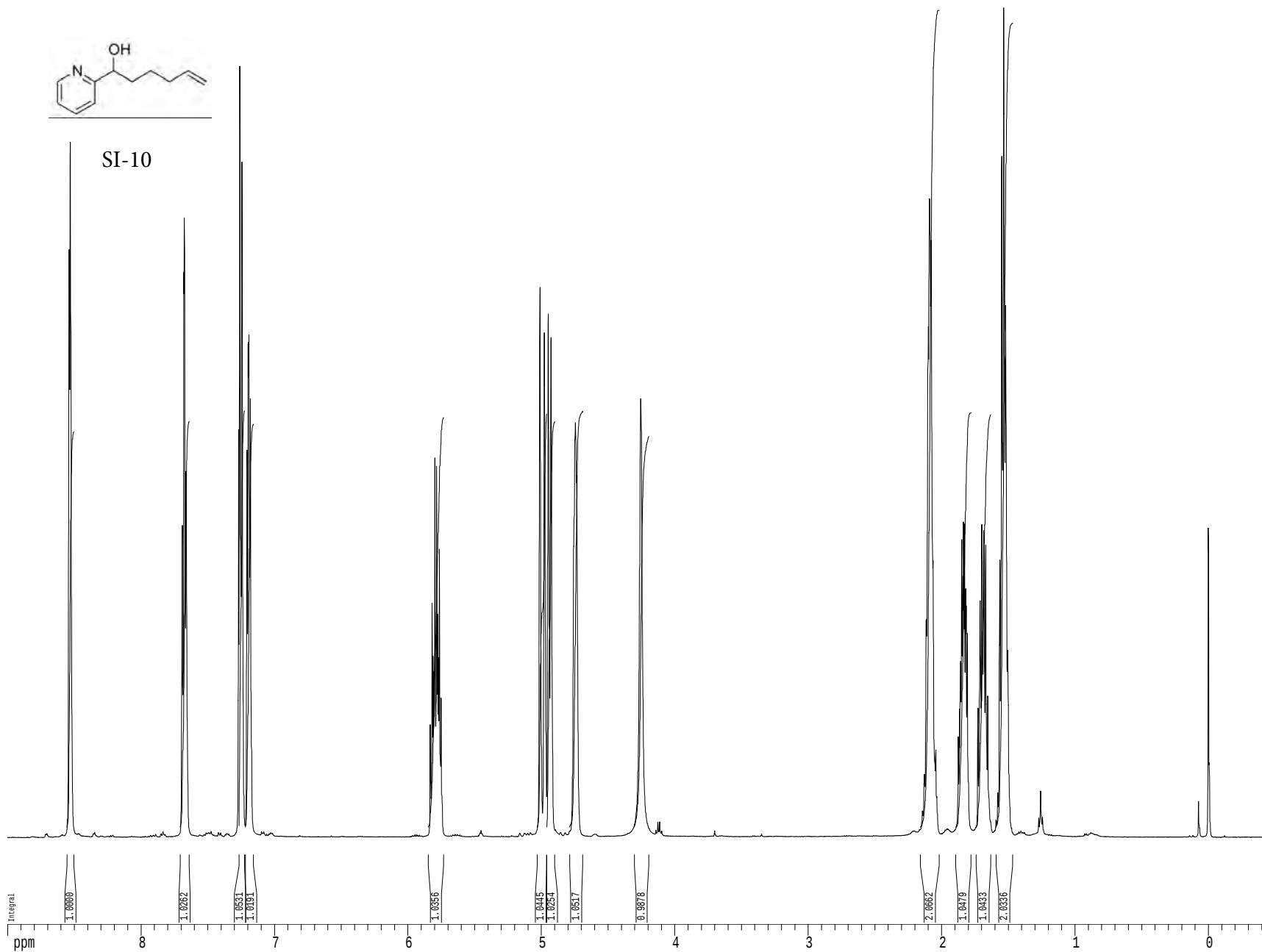
1D NMR plot parameters

CX 22.00 cm
CY 15.65 cm
F1P 220.000 ppm
F1 27671.69 Hz
F2P -5.000 ppm
F2 -628.90 Hz
PPMCM 9.86842 ppm/cm
HZCM 1241.25415 Hz/cm

Figure 1 displays two ^{13}C NMR spectra. The top spectrum is for poly(2,2,2-trifluoroethyl acrylate) (PTEA), showing peaks at 8.53856, 8.52959, 7.69335, 7.69016, 7.67794, 7.61484, 7.62277, 7.65946, 7.26903, 7.26603, 7.24495, 7.28566, 7.19576, 7.19165, and 7.14184 ppm. The bottom spectrum is for poly(2,2,2-trifluoroethyl methacrylate) (PTMA), showing peaks at 5.83174, 5.81843, 5.81133, 5.80596, 5.77931, 5.78439, 5.77711, 5.77688, 5.76585, 5.75936, 5.01264, 5.00941, 4.97846, 4.97516, 4.94784, 4.97236, 4.97286, and 4.25405 ppm. The x-axis is labeled 'ppm'.



SI-10



```
Current Data Parameters
USER          mharri
NAME          MRH-VII-212-1HNMR
EXPNO         1
PROCNO        1
```

```

F2 - Acquisition Parameters
Date_          20150213
Time           19.31
INSTRUM        CRYOS000
PROBHD         5 mm  CPTCI 1H-
PULPROG        zg30
TD             81728
SOLVENT        CDCl3
NS              8
DS              2
SWH            8012.820 Hz
FIDRES         0.098043 Hz
AQ             5.0998774 sec
RG              9
DW             62.400 usec
DE             6.00 usec
TE             298.0 K
D1             0.10000000 sec
MCREST         0.00000000 sec
MCWRRK         0.01500000 sec

```

```
===== CHANNEL f1 =====
NUC1                1H
P1                  7.50 usec
PL1                 1.60 dB
SF01               500.2235015 MHz
```

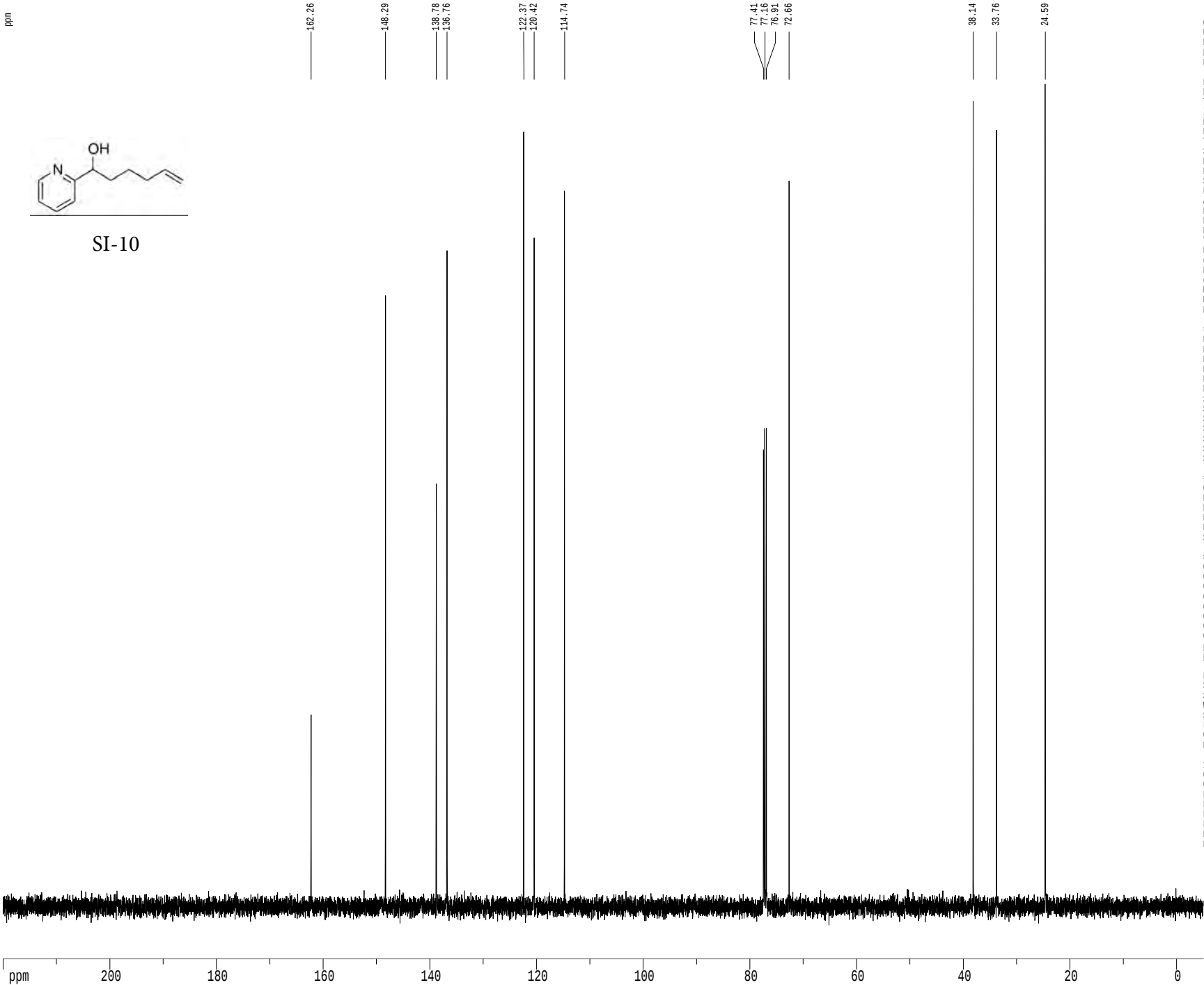
```
F2 - Processing parameters
SI                65536
SF                500.2200264 MHz
WDW               EM
SSB               0
LB                0.30 Hz
GB                0
PC                4.00
```

```

1D NMR plot parameters
CX                22.80 cm
CY                15.00 cm
F1P              9.000 ppm
F1               4501.98 Hz
F2P             -0.500 ppm
F2             -250.11 Hz
PPMCM           0.41667 ppm/cm
HZCM           208.42502 Hz/cm

```

Z-restored spin-echo 13C spectrum with 1H decoupling



Current Data Parameters

USER mharri
NAME MRH-VII-212-13CNMR
EXPNO 1
PROCNO 1

F2 - Acquisition Parameters

Date_ 20150213
Time 19.33
INSTRUM cryo500
PROBHD 5 mm CPTCI 1H-
PULPROG SpinEchopg30gp.prd
TD 65536
SOLVENT CDCl3
NS 83
DS 16
SWH 38383.831 Hz
FIDRES 0.462388 Hz
AQ 1.0813940 sec
RG 7298.2
DW 16.500 usec
DE 6.00 usec
TE 298.0 K
D1 0.25000000 sec
d11 0.03000000 sec
D16 0.00020000 sec
d17 0.00019600 sec
MCREST 0.00000000 sec
MCWRR 0.01500000 sec
P2 33.10 usec

===== CHANNEL f1 =====

NUC1 13C
P1 16.55 usec
P11 500.00 usec
P12 2000.00 usec
PL0 120.00 dB
PL1 -1.00 dB
SF01 125.7942548 MHz
SP1 2.70 dB
SP2 2.70 dB
SPNAM1 Crp60,0.5,20.1
SPNAM2 Crp60comp.4
SPOFF1 0.00 Hz
SPOFF2 0.00 Hz

===== CHANNEL f2 =====

CPDPRG2 waltz16
NUC2 1H
PCPD2 100.00 usec
PL2 1.00 dB
PL12 24.50 dB
SF02 500.2225011 MHz

===== GRADIENT CHANNEL =====

GPVAM1 SINE.100
GPVAM2 SINE.100
GPX1 0.00 %
GPX2 0.00 %
GPY1 0.00 %
GPY2 0.00 %
GPZ1 30.00 %
GPZ2 50.00 %
p15 500.00 usec
p16 1000.00 usec

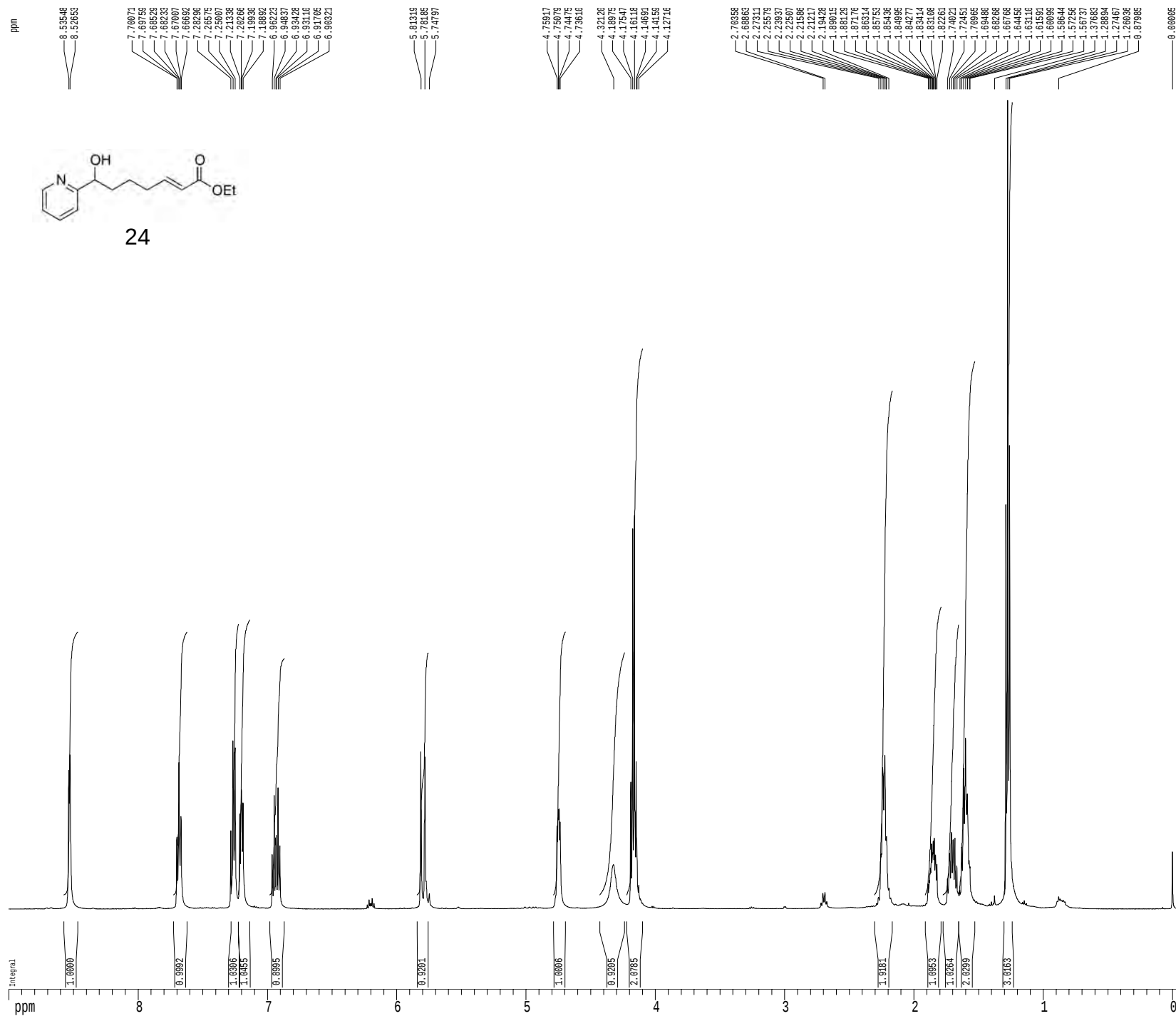
F2 - Processing parameters

SI 65536
SF 125.7804113 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 2.00

1D NMR plot parameters

CX 22.00 cm
CY 15.65 cm
F1P 220.000 ppm
F1 27671.69 Hz
F2P -5.000 ppm
F2 -628.90 Hz
PPMCM 9.86842 ppm/cm
HZCM 1241.25415 Hz/cm

¹H spectrum



Current Data Parameters
 USER mharri
 NAME MRH-VII-260-1HNMR
 EXPNO 1
 PROCNO 1

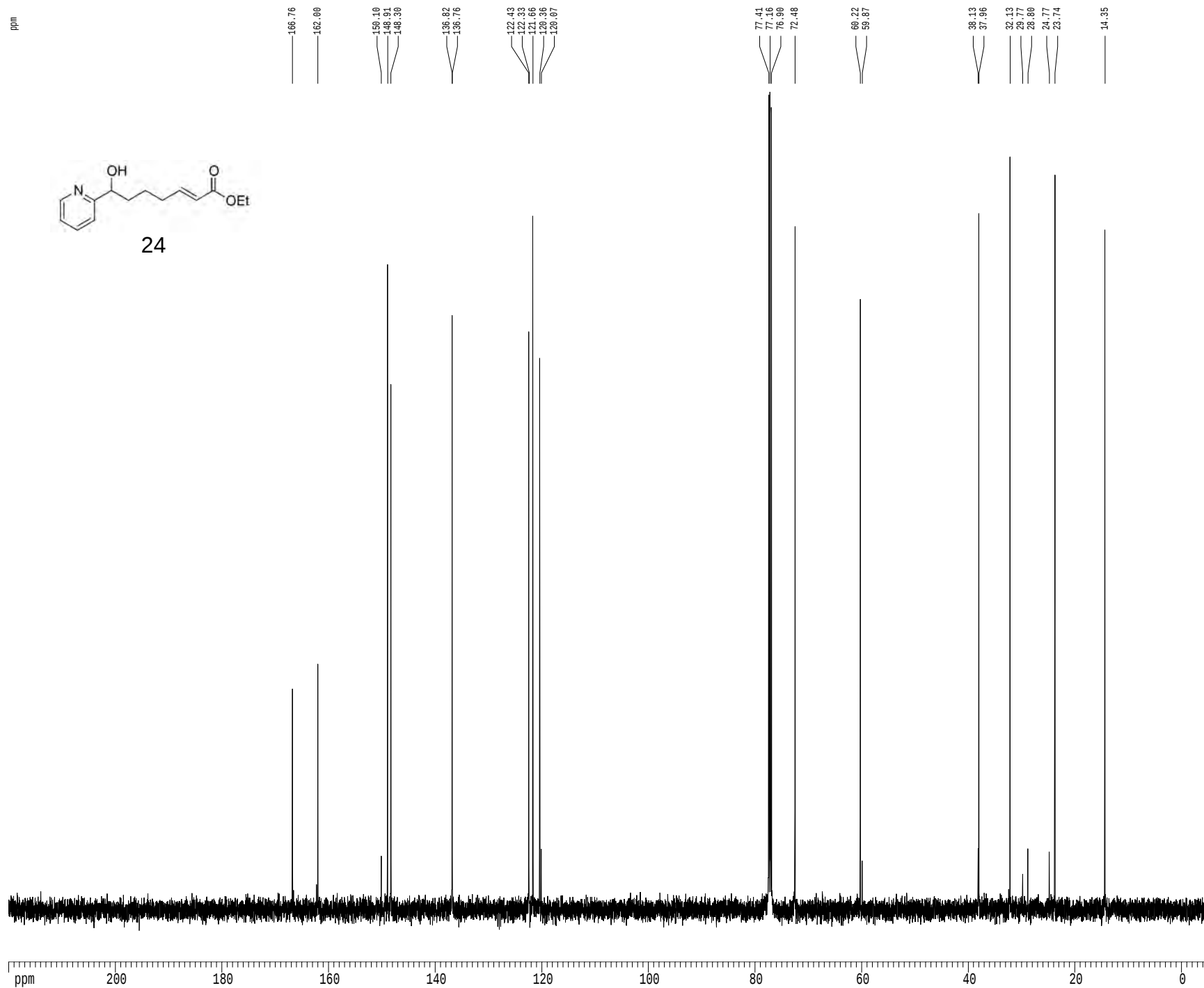
F2 - Acquisition Parameters
 Date_ 20150204
 Time 19.01
 INSTRUM gn500
 PROBHD 5 mm broadband
 PULPROG zg30
 TD 81728
 SOLVENT CDCl3T
 NS 8
 DS 2
 SWH 8012.820 Hz
 FIDRES 0.098043 Hz
 AQ 5.0998774 sec
 RG 64
 DW 62.400 usec
 DE 6.00 usec
 TE 298.0 K
 D1 0.10000000 sec
 MCREST 0.00000000 sec
 MCWRK 0.01500000 sec

===== CHANNEL f1 =====
 NUC1 1H
 P1 12.00 usec
 PL1 -5.00 dB
 SF01 499.1834943 MHz

F2 - Processing parameters
 SI 65536
 SF 499.1800153 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00

1D NMR plot parameters
 CX 22.80 cm
 CY 15.00 cm
 F1P 9.980 ppm
 F1 4492.62 Hz
 F2P -0.580 ppm
 F2 -249.59 Hz
 PPMCM 0.41667 ppm/cm
 HZCM 207.99168 Hz/cm

¹³C spectrum with ¹H decoupling



Current Data Parameters
 USER mharri
 NAME MRH-VII-266-13CNMR
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20150204
 Time 19.05
 INSTRUM gn500
 PROBHD 5 mm broadband
 PULPROG zgpg30
 TD 65536
 SOLVENT CDCl3
 NS 201
 DS 4
 SWH 30303.031 Hz
 FIDRES 0.462388 Hz
 AQ 1.0013940 sec
 RG 6502
 DW 16.500 usec
 DE 4.50 usec
 TE 298.0 K
 D1 0.25000000 sec
 d11 0.03000000 sec
 MCREST 0.00000000 sec
 MCWRR 0.01500000 sec

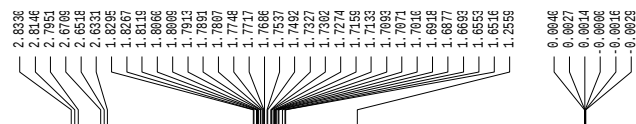
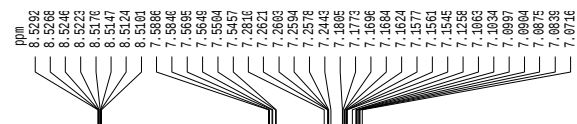
===== CHANNEL f1 =====
 NUC1 13C
 P1 9.00 usec
 PL1 -0.60 dB
 SF01 125.5327181 MHz

===== CHANNEL f2 =====
 CPDPRG2 waltz16
 NUC2 1H
 PCPD2 80.00 usec
 PL2 -3.00 dB
 PL12 12.80 dB
 SF02 499.1824959 MHz

F2 - Processing parameters
 SI 65536
 SF 125.5189030 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 2.00

1D NMR plot parameters
 CX 22.80 cm
 CY 15.65 cm
 F2P 220.000 ppm
 F1 27614.16 Hz
 F2P -5.000 ppm
 F2 -627.59 Hz
 PPMCM 9.86842 ppm/cm
 HZCM 1238.67346 Hz/cm

¹H spectrum



Current Data Parameters
 USER lhamma
 NAME LEH-5-054-c1-f007
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20150114
 Time 20.33
 INSTRUM drx400
 PROBHD 5 mm QNP H/F/P
 PULPROG zg30
 TD 25640
 SOLVENT CDCl3
 NS 8
 DS 2
 SWH 6410.256 Hz
 FIDRES 0.250010 Hz
 AQ 1.9999700 sec
 RG 406.4
 DW 78.000 usec
 DE 4.50 usec
 TE 290.0 K
 D1 0.10000000 sec
 MCREST 0.00000000 sec
 MCWRK 0.01500000 sec

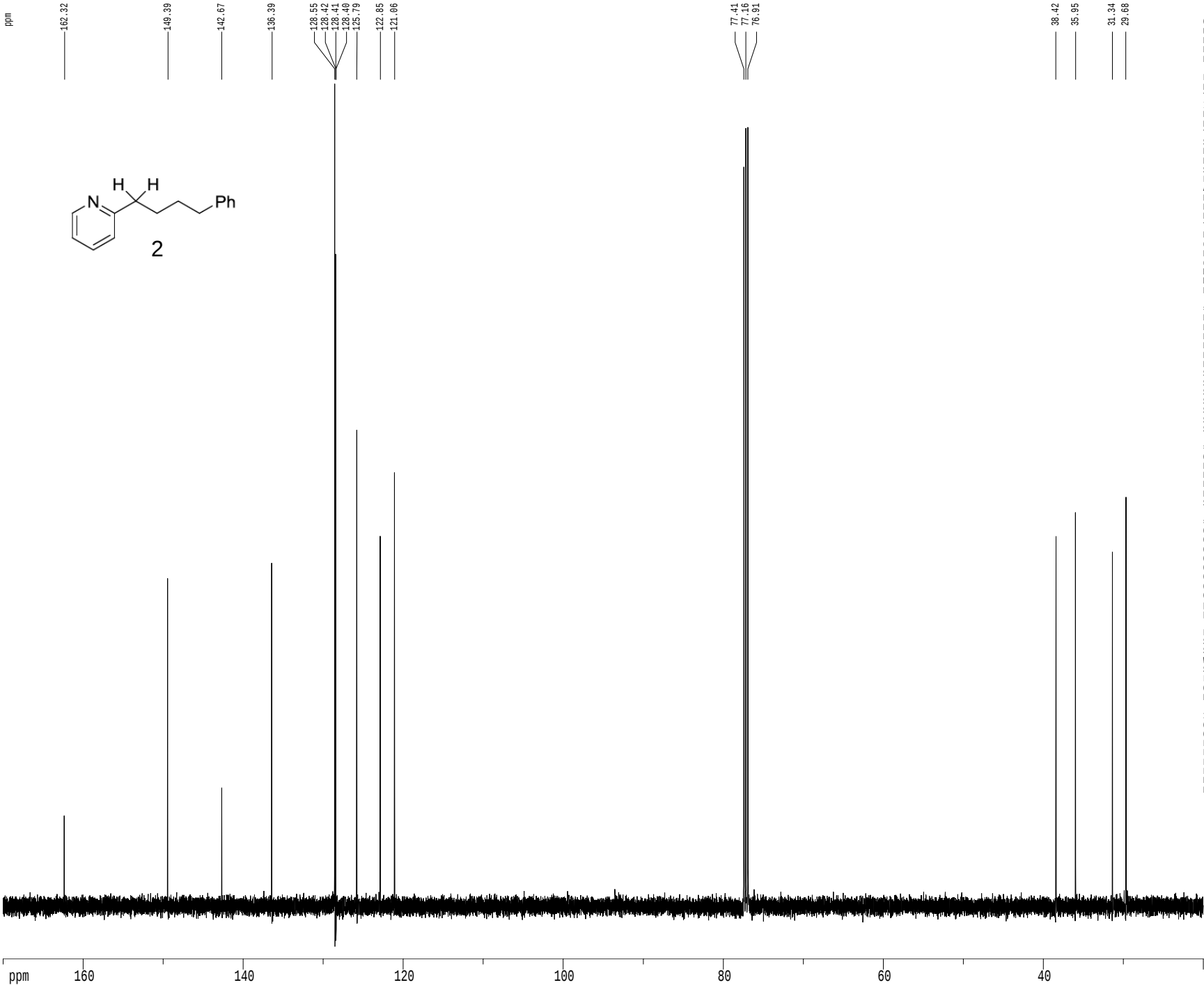
===== CHANNEL f1 =====
 NUC1 1H
 P1 12.00 usec
 PL1 0.00 dB
 SFO1 400.1328009 MHz

F2 - Processing parameters
 SI 65536
 SF 400.1308216 MHz
 WDW no
 SSB 0
 LB 0.00 Hz
 GB 0
 PC 2.00

1D NMR plot parameters
 CX 22.80 cm
 CY 15.00 cm
 F1P 9.000 ppm
 F1 3601.17 Hz
 F2P -0.580 ppm
 F2 -200.86 Hz
 PPMCM 0.41667 ppm/cm
 HZCM 166.72086 Hz/cm



Z-restored spin-echo 13C spectrum with 1H decoupling



Current Data Parameters

USER ljhanna
NAME LEH-5-054-c1-f008-C13
EXPNO 1
PROCNO 1

F2 - Acquisition Parameters

Date_ 20150113
Time 18.34
INSTRUM cryo500
PROBHD 5 mm CPTCI 1H-
PULPROG SpineEcho300p.prd
TD 65536
SOLVENT CDCl3
NS 1024
DS 16
SWH 30303.031 Hz
FIDRES 0.462388 Hz
AQ 1.0813940 sec
RG 13004
DW 16.500 usec
DE 6.00 usec
TE 298.0 K
D1 0.25000000 sec
d11 0.03000000 sec
D16 0.00020000 sec
D17 0.00019000 sec
MCOREST 0.00000000 sec
MCORRK 0.01500000 sec
P2 33.10 usec

===== CHANNEL f1 =====

NUC1 13C
P1 16.55 usec
P11 500.00 usec
P12 2000.00 usec
PL0 120.00 dB
PL1 -1.00 dB
SFO1 125.7942540 MHz
SP1 2.70 dB
SP2 2.70 dB
SPNAM1 Crp60,0.5,20.1
SPNAM2 Crp60comp.4
SPOFF1 0.00 Hz
SPOFF2 0.00 Hz

===== CHANNEL f2 =====

CPDPRG2 waltz16
NUC2 1H
PCPD2 100.00 usec
PL2 1.00 dB
PL12 24.50 dB
SFO2 500.2225011 MHz

===== GRADIENT CHANNEL =====

SPNAM1 SINE.100
SPNAM2 SINE.100
GPX1 0.00 %
GPX2 0.00 %
GPY1 0.00 %
GPY2 0.00 %
GPZ1 30.00 %
GPZ2 50.00 %
p15 500.00 usec
p16 1000.00 usec

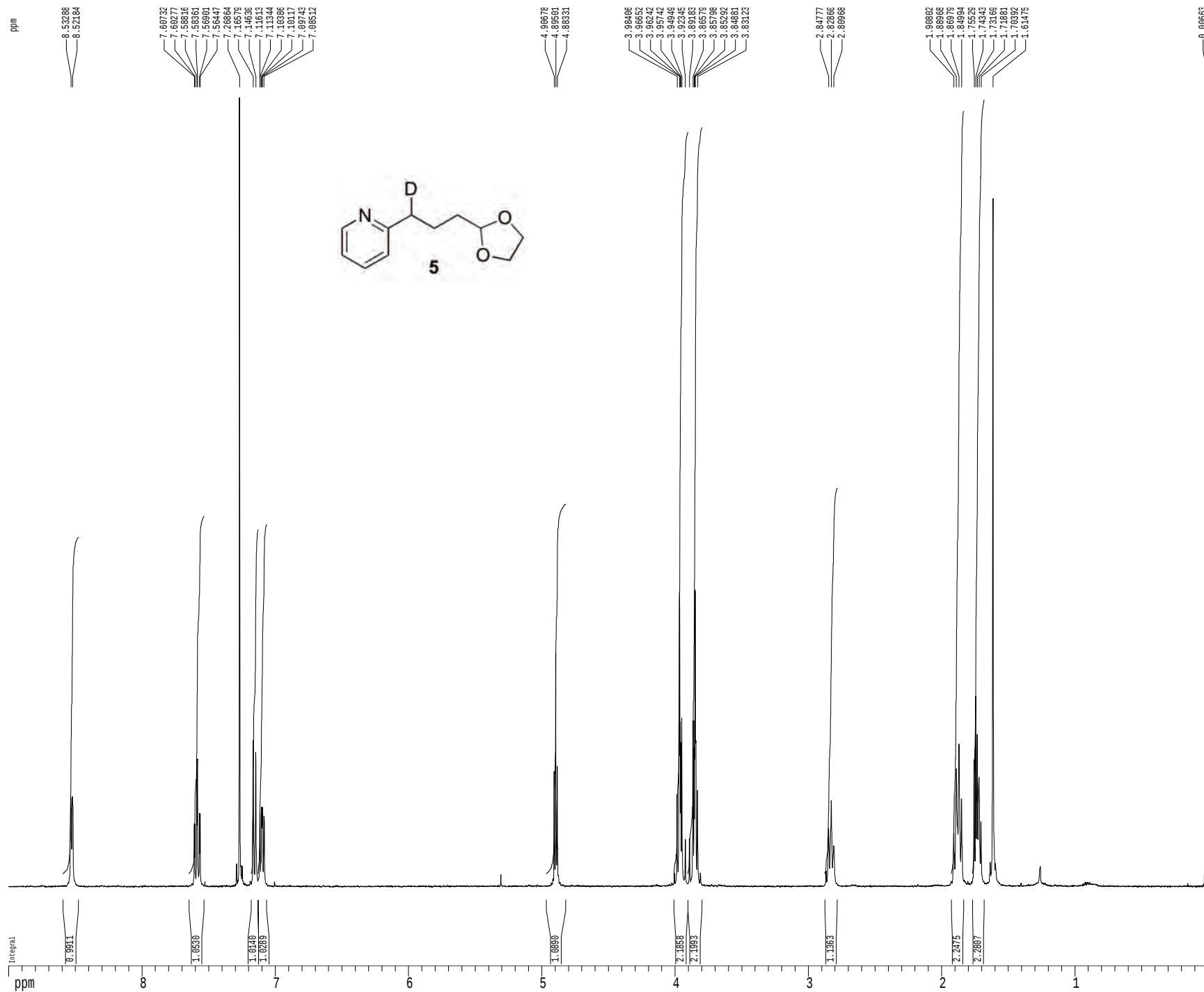
F2 - Processing parameters

SI 65536
SF 125.7804085 MHz
WDW no
SSB 0
LB 0.00 Hz
GB 0
PC 2.00

1D NMR plot parameters

CX 22.00 cm
CY 15.65 cm
F1P 170.000 ppm
F1 21302.67 Hz
F2P 20.000 ppm
F2 2515.61 Hz
PPMCM 6.57895 ppm/cm
HZCM 827.50275 Hz/cm

¹H spectrum



Current Data Parameters
 USER lhanna
 NAME LEH-5-044-c3-f021
 EXPNO 1
 PROCNO 1

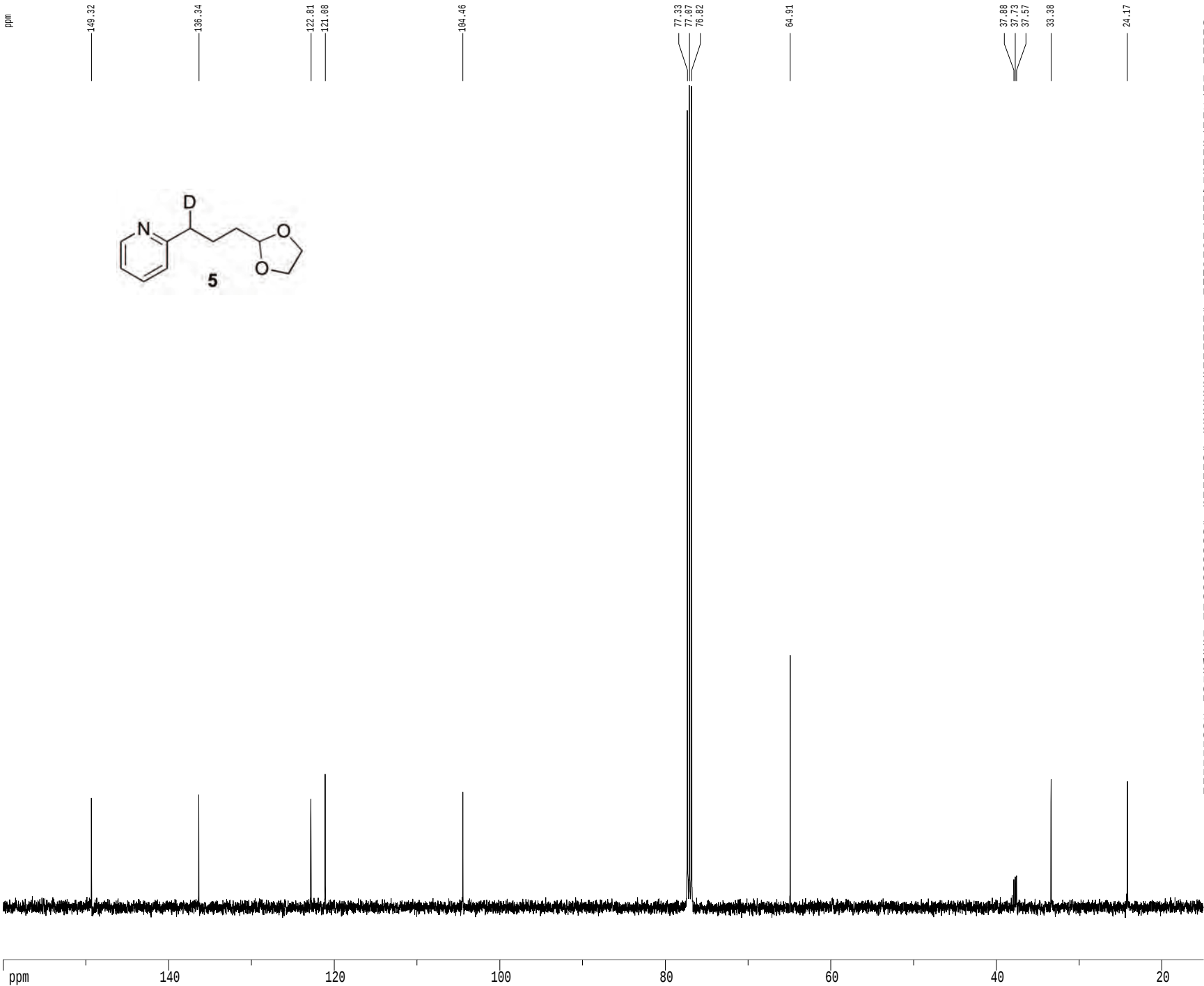
F2 - Acquisition Parameters
 Date_ 20150113
 Time 13.46
 INSTRUM drx400
 PROBHD 5 mm QNP H/F/P
 PULPROG zg30
 TD 25640
 SOLVENT CDCl3
 NS 16
 DS 2
 SMH 6410.256 Hz
 FIDRES 0.250010 Hz
 AQ 1.9999700 sec
 RG 1024
 DW 78.000 usec
 DE 4.50 usec
 TE 290.0 K
 D1 0.10000000 sec
 MCREST 0.00000000 sec
 MCWRK 0.01500000 sec

===== CHANNEL f1 =====
 NUC1 1H
 P1 12.00 usec
 PL1 0.00 dB
 SF01 400.1328009 MHz

F2 - Processing parameters
 SI 65536
 SF 400.1300175 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 2.00

1D NMR plot parameters
 CX 22.00 cm
 CY 15.00 cm
 F1P 9.000 ppm
 F1 3601.17 Hz
 F2P 0.000 ppm
 F2 0.00 Hz
 PPMCM 0.39474 ppm/cm
 HZCM 157.94606 Hz/cm

Z-restored spin-echo 13C spectrum with 1H decoupling



Current Data Parameters
USER lhanna
NAME LEH-5-044-c3-f020-C13
EXPNO 1
PROCNO 1

F2 - Acquisition Parameters
Date_ 20150113
Time 12.24
INSTRUM cryo500
PROBHD 5 mm CPTCI 1H-
PULPROG SpineEcho300p.prd
TD 65536
SOLVENT CDCl3
NS 1024
DS 16
SWH 30303.031 Hz
FIDRES 0.462388 Hz
AQ 1.0813940 sec
RG 13004
DW 16.500 usec
DE 6.00 usec
TE 298.0 K
D1 0.25000000 sec
d11 0.03000000 sec
D16 0.00020000 sec
D17 0.00019000 sec
MCREST 0.00000000 sec
MCWRK 0.01500000 sec
P2 33.10 usec

===== CHANNEL f1 =====
NUC1 13C
P1 16.55 usec
P11 500.00 usec
P12 2000.00 usec
PL0 120.00 dB
PL1 -1.00 dB
SP01 125.7942540 MHz
SP1 2.70 dB
SP2 2.70 dB
SPNAM1 Crp60,0.5,20.1
SPNAM2 Crp60comp.4
SPOFF1 0.00 Hz
SPOFF2 0.00 Hz

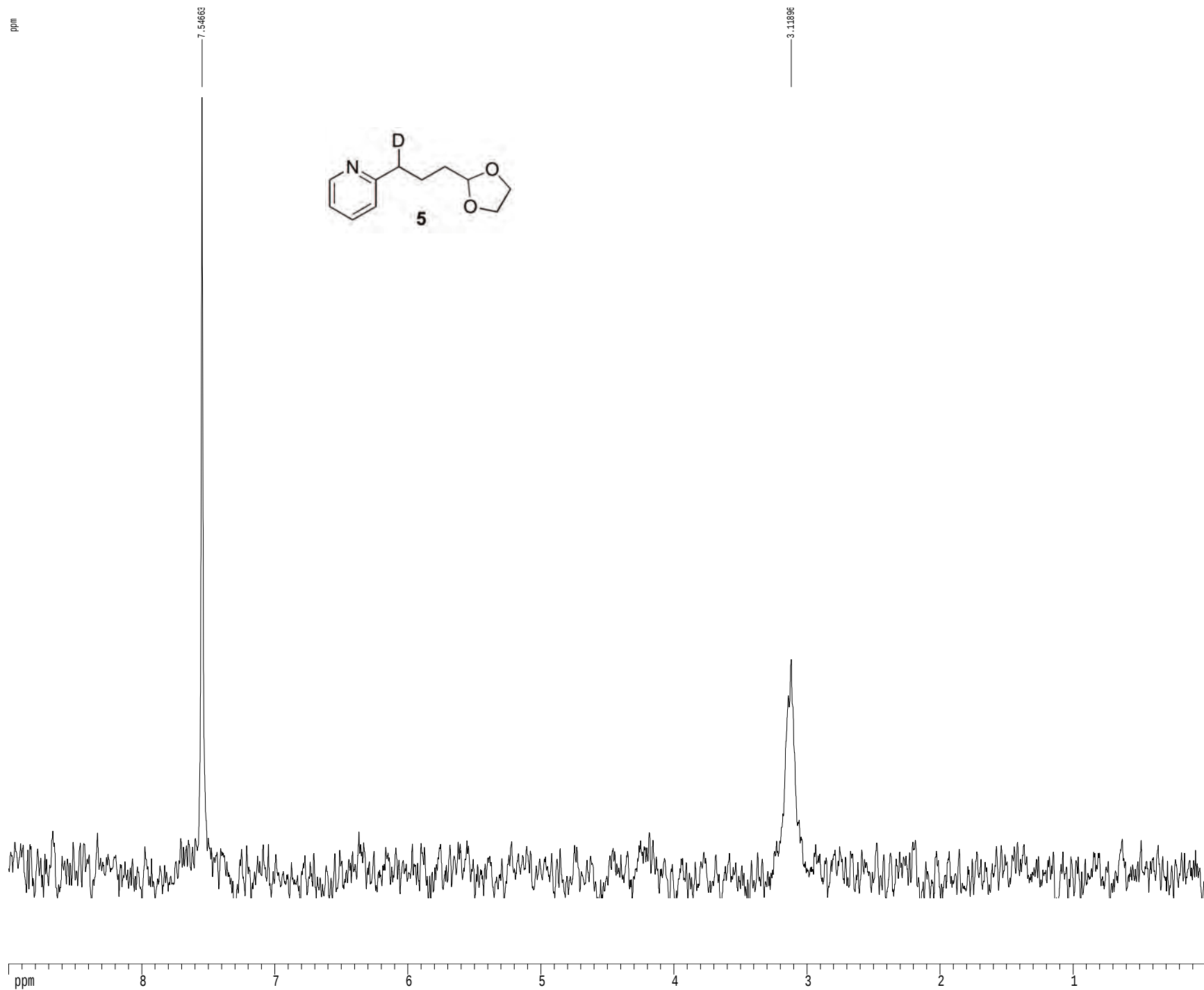
===== CHANNEL f2 =====
CPDPRG2 waltz16
NUC2 1H
PCPD2 100.00 usec
PL2 1.00 dB
PL12 24.50 dB
SF02 500.2225011 MHz

===== GRADIENT CHANNEL =====
GPNAM1 SINE.100
GPNAM2 SINE.100
GPX1 0.00 %
GPX2 0.00 %
GPY1 0.00 %
GPY2 0.00 %
GPZ1 30.00 %
GPZ2 50.00 %
p15 500.00 usec
p16 1000.00 usec

F2 - Processing parameters
SI 65536
SF 125.7804190 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 2.00

1D NMR plot parameters
CX 22.00 cm
CY 15.65 cm
F1P 160.000 ppm
F1 20124.87 Hz
F2P 15.000 ppm
F2 1886.71 Hz
PPMCH 6.35965 ppm/cm
HZCM 799.91937 Hz/cm

2H spectrum (measure via lock channel without changing any cables)



Current Data Parameters
USER lhanna
NAME LEH-5-044-D1
EXPNO 1
PROCNO 1

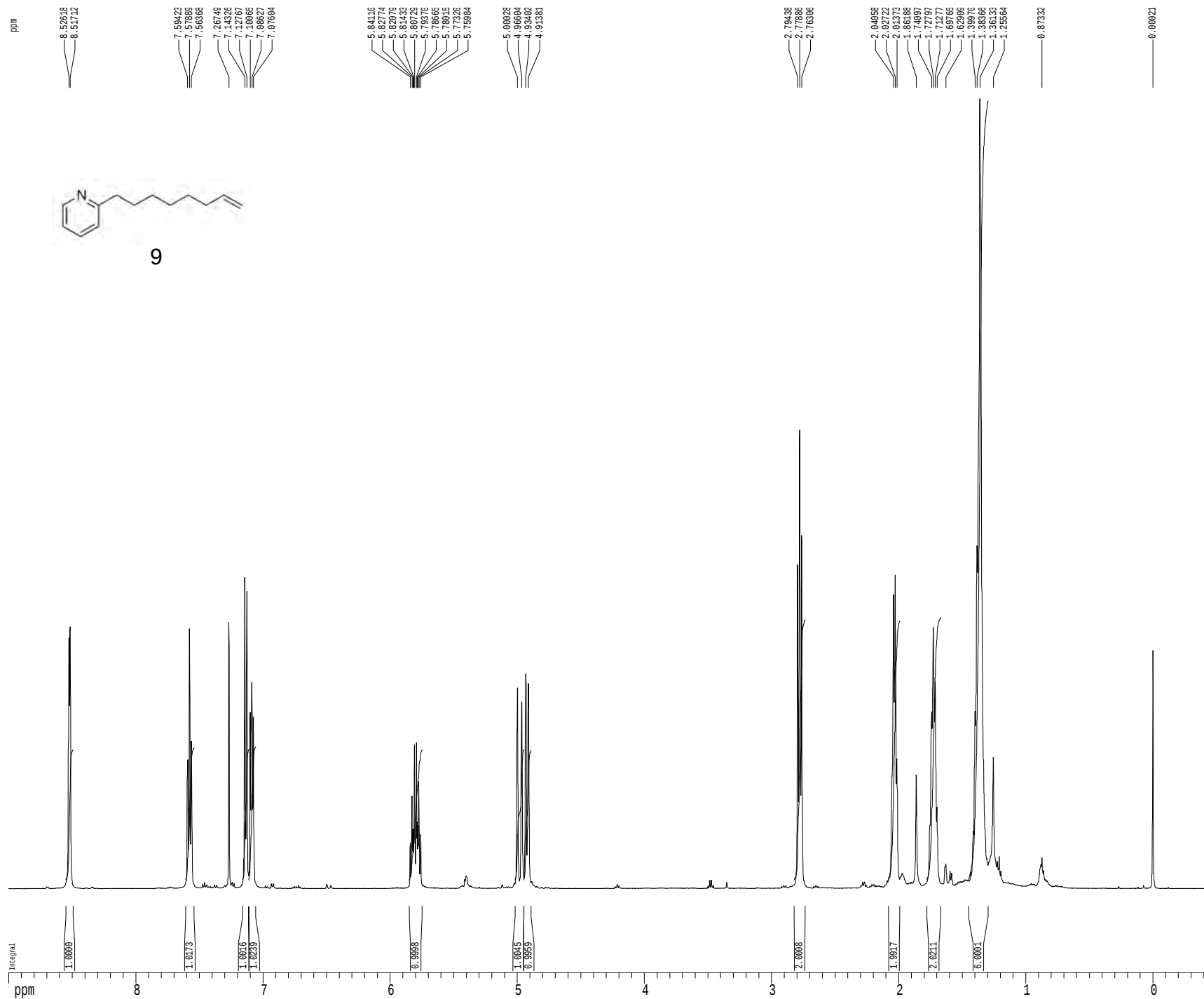
F2 - Acquisition Parameters
Date_ 20150115
Time 17.23
INSTRUM drx400
PROBHD 5 mm QNP H/F/P
PULPROG zg30h2lock
TD 10022
SOLVENT CDC13
NS 243
DS 2
SWH 982.704 Hz
FIDRES 0.098055 Hz
AQ 5.0992436 sec
RG 64
DW 508.800 usec
DE 20.30 usec
TE 298.0 K
D1 0.10000000 sec
d11 0.03000000 sec

===== CHANNEL f1 =====
NUC1 2H
P1 70.00 usec
PL1 -6.00 dB
SF01 61.4227670 MHz

F2 - Processing parameters
SI 65536
SF 61.4223370 MHz
WDW EM
SSB 0
LB 0.50 Hz
GB 0
PC 1.00

1D NMR plot parameters
CX 22.00 cm
CY 15.00 cm
F1P 9.000 ppm
F1 552.00 Hz
F2P 0.000 ppm
F2 0.00 Hz
PPMCM 0.39474 ppm/cm
HZCM 24.24566 Hz/cm

¹H spectrum



Current Data Parameters

USER	mharri
NAME	MRH-VII-232-1HNMR
EXPNO	1
PROCNO	1

F2 - Acquisition Parameters

Date_	20150116
Time	19.00
INSTRUM	Cryo500
PROBHD	5 mm CPTCI 1H-
PULPROG	zg30
TD	81728
SOLVENT	CDCl3
NS	8
DS	2
SWH	8012.820 Hz
FIDRES	0.098043 Hz
AQ	5.0998774 sec
RG	4.5
DW	62.400 usec
DE	6.00 usec
TE	298.0 K
D1	0.10000000 sec
MCREST	0.00000000 sec
MCNWK	0.01500000 sec

===== CHANNEL f1 =====

NUC1	1H
P1	7.50 usec
PL1	1.00 dB
SFO1	500.2235015 MHz

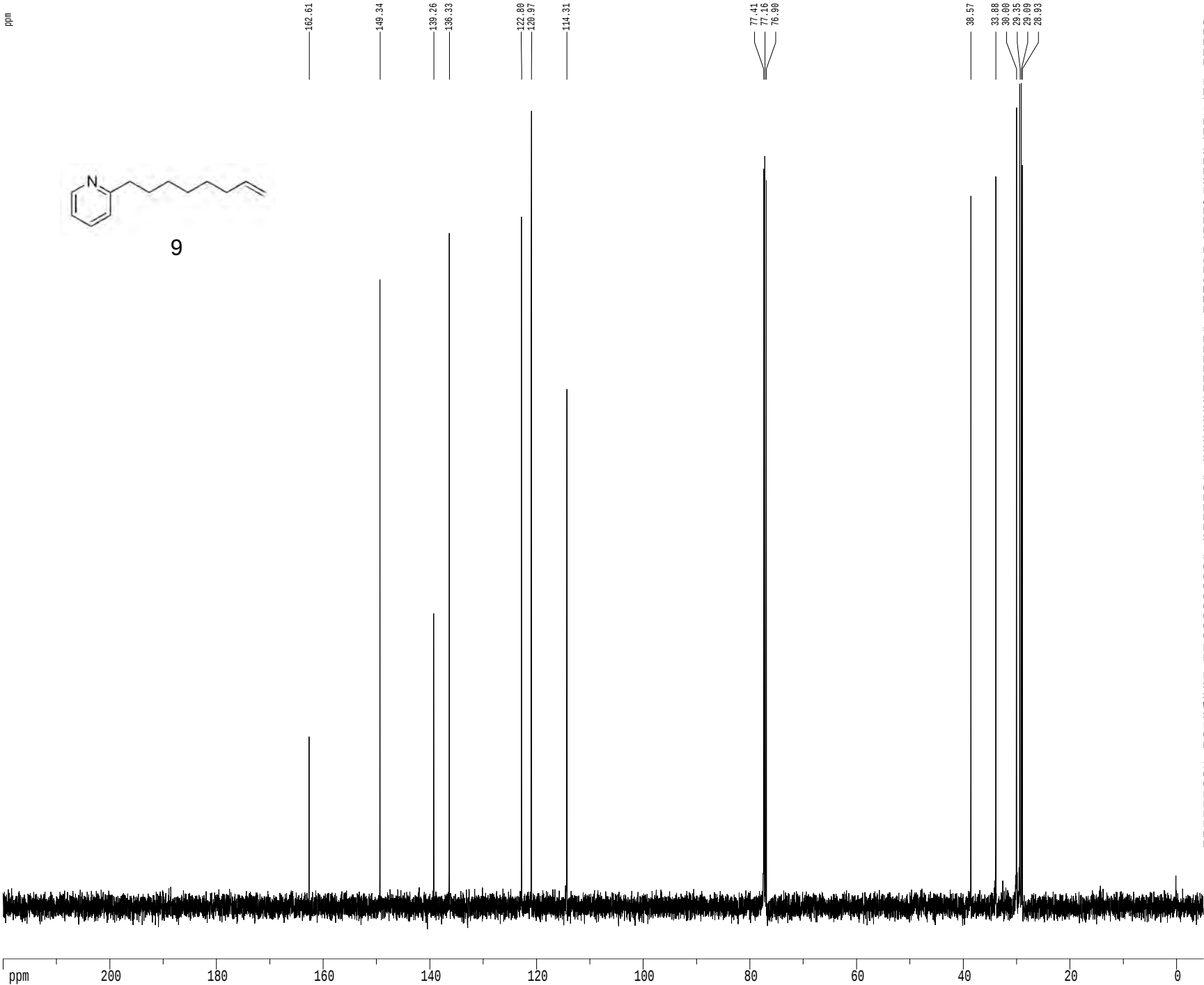
F2 - Processing parameters

SI	65536
SF	500.2200280 MHz
WDW	EM
SSB	0
LB	0.30 Hz
GB	0
PC	4.00

1D NMR plot parameters

CX	22.00 cm
CY	15.00 cm
F1P	9.000 ppm
F1	4501.98 Hz
F2P	-0.500 ppm
F2	-250.11 Hz
PPMCM	0.41667 ppm/cm
HZCM	208.42502 Hz/cm

Z-restored spin-echo 13C spectrum with 1H decoupling



Current Data Parameters

USER mharri
NAME MRH-VII-232-13CNMR
EXPNO 1
PROCNO 1

F2 - Acquisition Parameters

Date_ 20150116
Time 19.05
INSTRUM cryo500
PROBHD 5 mm CPTCI 1H-
PULPROG SpinEchopg30gp.prd
TD 65536
SOLVENT CDCl3
NS 152
DS 16
SWH 30303.031 Hz
FIDRES 0.462388 Hz
AQ 1.0813940 sec
RG 7298.2
DW 16.500 usec
DE 6.00 usec
TE 298.0 K
D1 0.25000000 sec
d11 0.03000000 sec
D16 0.00020000 sec
d17 0.00019600 sec
MCREST 0.00000000 sec
MCWRK 0.01500000 sec
P2 33.10 usec

===== CHANNEL f1 =====

NUC1 13C
P1 16.55 usec
P11 500.00 usec
P12 2000.00 usec
PL0 120.00 dB
PL1 -1.00 dB
SF01 125.7942548 MHz
SP1 2.70 dB
SP2 2.70 dB
SPNAM1 Crp60,0.5,20.1
SPNAM2 Crp60comp.4
SPOFF1 0.00 Hz
SPOFF2 0.00 Hz

===== CHANNEL f2 =====

CPDPRG2 waltz16
NUC2 1H
PCPD2 100.00 usec
PL2 1.00 dB
PL12 24.50 dB
SF02 500.2225011 MHz

===== GRADIENT CHANNEL =====

GPVAM1 SINE.100
GPVAM2 SINE.100
GPX1 0.00 %
GPX2 0.00 %
GPY1 0.00 %
GPY2 0.00 %
GPZ1 30.00 %
GPZ2 50.00 %
p15 500.00 usec
p16 1000.00 usec

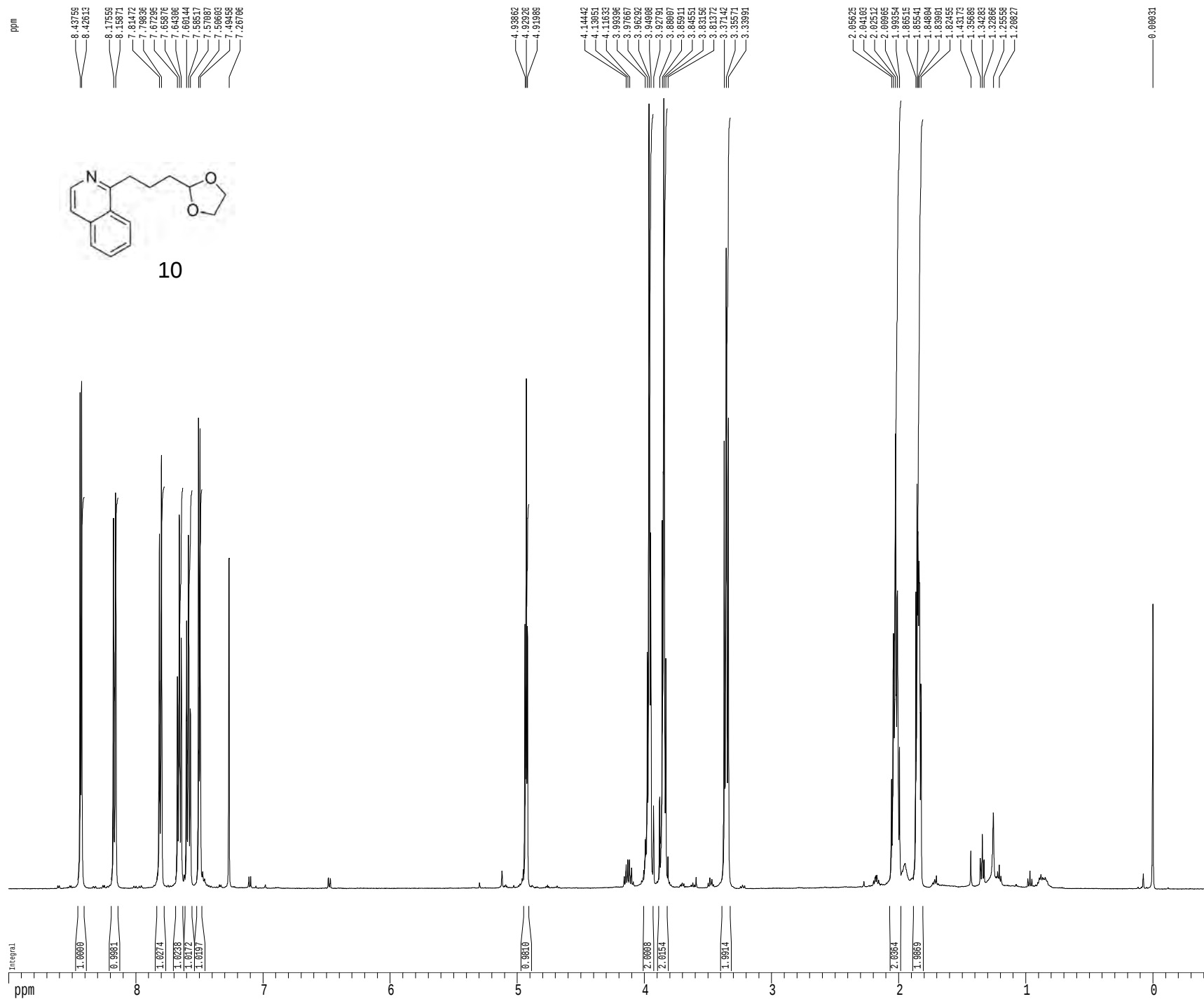
F2 - Processing parameters

SI 65536
SF 125.7804094 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 2.00

1D NMR plot parameters

CX 22.00 cm
CY 15.65 cm
F1P 220.000 ppm
F1 27671.69 Hz
F2P -5.000 ppm
F2 -628.90 Hz
PPMCM 9.86842 ppm/cm
HZCM 1241.25415 Hz/cm

¹H spectrum



Current Data Parameters
 USER mharri
 NAME MRH-VII-241-1HNMR
 EXPNO 1
 PROCNO 1

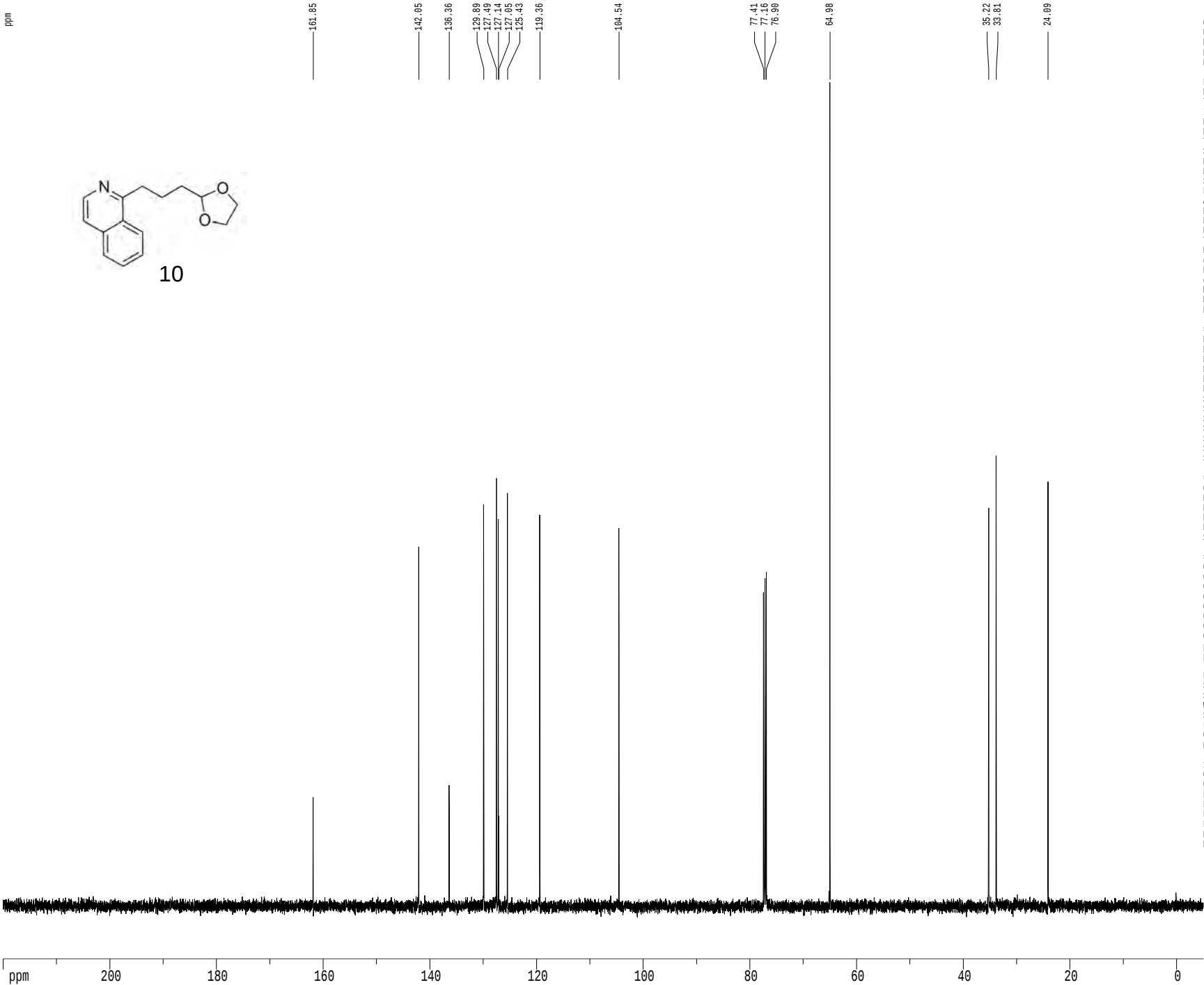
F2 - Acquisition Parameters
 Date_ 20150116
 Time 19.09
 INSTRUM cryo500
 PROBHD 5 mm CPTCI 1H-
 PULPROG zg30
 TO 81728
 SOLVENT CDCl3
 NS 8
 DS 2
 SWH 8012.820 Hz
 FIDRES 0.098043 Hz
 AQ 5.0998774 sec
 RG 5.7
 DW 62.400 usec
 DE 6.00 usec
 TE 298.0 K
 D1 0.10000000 sec
 MCREST 0.00000000 sec
 MCNRK 0.01500000 sec

===== CHANNEL f1 =====
 NUC1 1H
 P1 7.50 usec
 PL1 1.00 dB
 SFO1 500.2235015 MHz

F2 - Processing parameters
 SI 65536
 SF 500.2200280 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 4.00

1D NMR plot parameters
 CX 22.00 cm
 CY 15.00 cm
 F1P 9.000 ppm
 F1 4501.98 Hz
 F2P -0.500 ppm
 F2 -250.11 Hz
 PPMCM 0.41667 ppm/cm
 HZCM 208.42502 Hz/cm

Z-restored spin-echo 13C spectrum with 1H decoupling



Current Data Parameters

USER mharri
NAME MRH-VII-241-13CNMR
EXPNO 1
PROCNO 1

F2 - Acquisition Parameters

Date_ 20150116
Time 19.12
INSTRUM cryo500
PROBHD 5 mm CPTCI 1H-
PULPROG SpinEchopg30gp.prd
TD 65536
SOLVENT CDCl3
NS 127
DS 16
SWH 30303.031 Hz
FIDRES 0.462388 Hz
AQ 1.0813940 sec
RG 7298.2
DW 16.500 usec
DE 6.00 usec
TE 298.0 K
D1 0.25000000 sec
d11 0.03000000 sec
D16 0.00020000 sec
d17 0.00019600 sec
MCREST 0.00000000 sec
MCWRK 0.01500000 sec
P2 33.10 usec

===== CHANNEL f1 =====

NUC1 13C
P1 16.55 usec
P11 500.00 usec
P12 2000.00 usec
PL0 120.00 dB
PL1 -1.00 dB
SF01 125.7942548 MHz
SP1 2.70 dB
SP2 2.70 dB
SPNAM1 Crp60,0.5,20.1
SPNAM2 Crp60comp.4
SPOFF1 0.00 Hz
SPOFF2 0.00 Hz

===== CHANNEL f2 =====

CPDPRG2 waltz16
NUC2 1H
PCPD2 100.00 usec
PL2 1.00 dB
PL12 24.50 dB
SF02 500.2225011 MHz

===== GRADIENT CHANNEL =====

GPVAM1 SINE.100
GPVAM2 SINE.100
GPX1 0.00 %
GPX2 0.00 %
GPY1 0.00 %
GPY2 0.00 %
GPZ1 30.00 %
GPZ2 50.00 %
p15 500.00 usec
p16 1000.00 usec

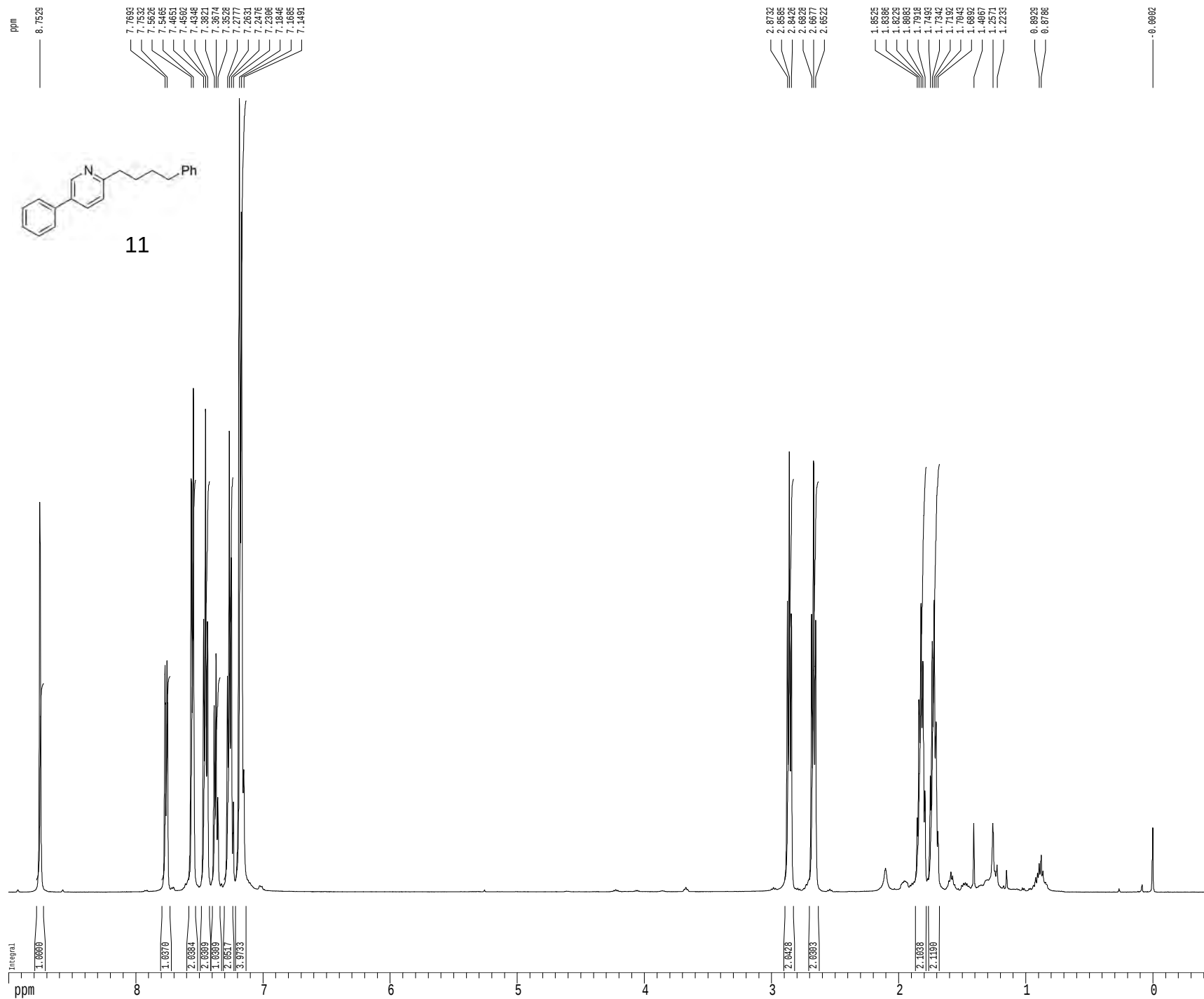
F2 - Processing parameters

SI 65536
SF 125.7804100 MHz
EN
WDW 0
SSB 0
LB 1.00 Hz
GB 0
PC 2.00

1D NMR plot parameters

CX 22.00 cm
CY 15.65 cm
F1P 220.000 ppm
F1 27671.69 Hz
F2P -5.000 ppm
F2 -628.90 Hz
PPMCM 9.86842 ppm/cm
HZCM 1241.25415 Hz/cm

¹H spectrum



Current Data Parameters
 USER mharri
 NAME MRH-VII-261-1HNMR
 EXPNO 1
 PROCNO 1

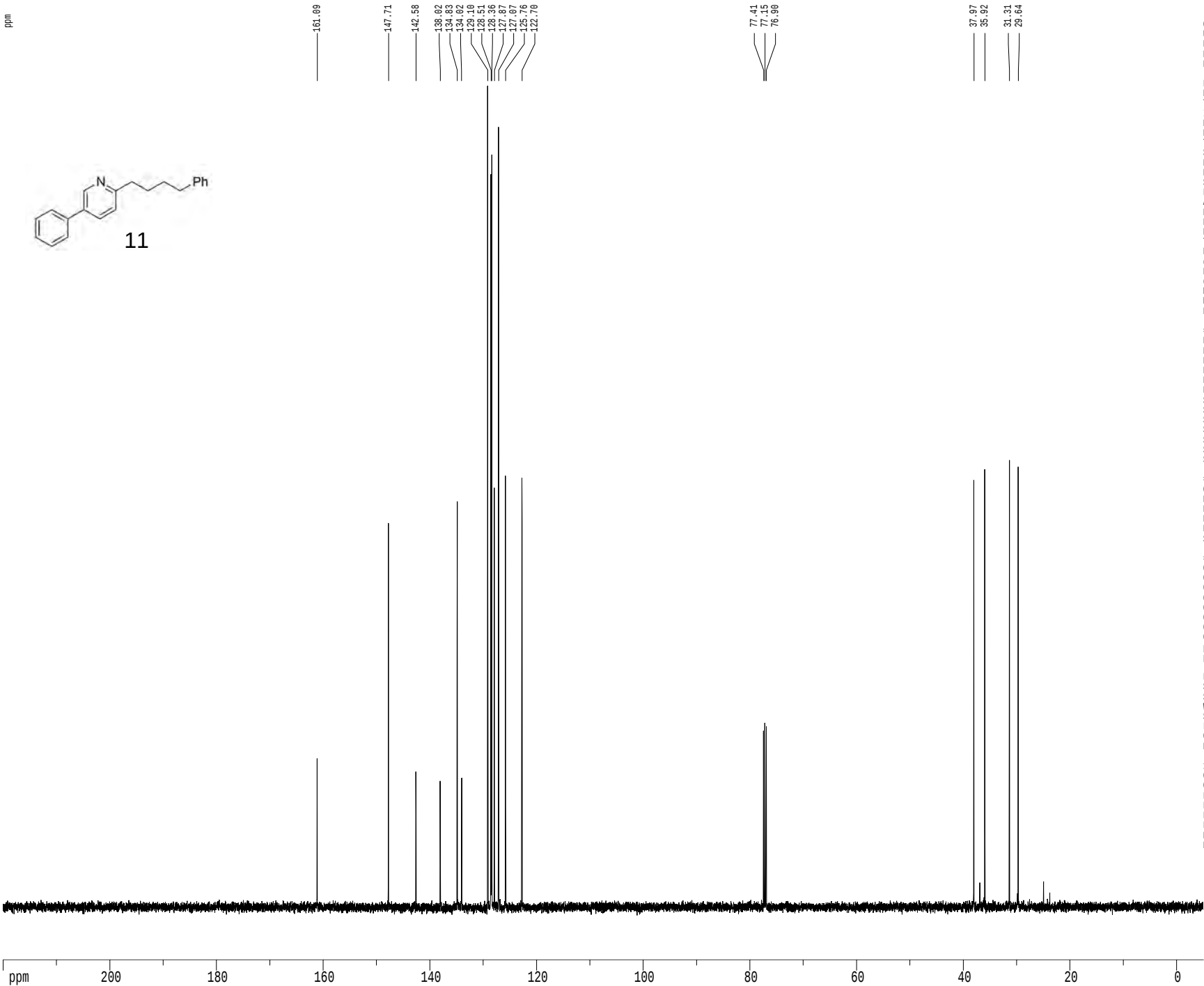
F2 - Acquisition Parameters
 Date_ 20150131
 Time 20.40
 INSTRUM cryo500
 PROBHD 5 mm CPTCI 1H-
 PULPROG zg30
 TD 81728
 SOLVENT CDCl3T
 NS 8
 DS 2
 SWH 8012.820 Hz
 FIDRES 0.098043 Hz
 AQ 5.0998774 sec
 RG 4
 DW 62.400 usec
 DE 6.00 usec
 TE 298.0 K
 D1 0.10000000 sec
 MCREST 0.00000000 sec
 MCNRK 0.01500000 sec

===== CHANNEL f1 =====
 NUC1 1H
 P1 7.50 usec
 PL1 1.00 dB
 SF01 500.2235015 MHz

F2 - Processing parameters
 SI 65536
 SF 500.2200454 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 4.00

1D NMR plot parameters
 CX 22.00 cm
 CY 15.00 cm
 F1P 9.000 ppm
 F1 4501.98 Hz
 F2P -0.500 ppm
 F2 -250.11 Hz
 PPMCM 0.41667 ppm/cm
 HZCM 208.42502 Hz/cm

Z-restored spin-echo 13C spectrum with 1H decoupling



Current Data Parameters

USER mharri
NAME MRH-VII-261-13CNMR
EXPNO 1
PROCNO 1

F2 - Acquisition Parameters

Date_ 20150131
Time 20.43
INSTRUM cryo500
PROBHD 5 mm CPTCI 1H-
PULPROG SpinEchopg30gp.prd
TD 65536
SOLVENT CDCl3
NS 45
DS 16
SWH 30303.031 Hz
FIDRES 0.462388 Hz
AQ 1.0813940 sec
RG 5792.6
DW 16.500 usec
DE 6.00 usec
TE 298.0 K
D1 0.25000000 sec
d11 0.03000000 sec
D16 0.00020000 sec
d17 0.00019600 sec
MCREST 0.00000000 sec
MCWRR 0.01500000 sec
P2 33.10 usec

===== CHANNEL f1 =====

NUC1 13C
P1 16.55 usec
P11 500.00 usec
P12 2000.00 usec
PL0 120.00 dB
PL1 -1.00 dB
SF01 125.7942548 MHz
SP1 2.70 dB
SP2 2.70 dB
SPNAM1 Crp60,0.5,20.1
SPNAM2 Crp60comp.4
SPOFF1 0.00 Hz
SPOFF2 0.00 Hz

===== CHANNEL f2 =====

CPDPRG2 waltz16
NUC2 1H
PCPD2 100.00 usec
PL2 1.00 dB
PL12 24.50 dB
SF02 500.2225011 MHz

===== GRADIENT CHANNEL =====

GPVAM1 SINE.100
GPVAM2 SINE.100
GPX1 0.00 %
GPX2 0.00 %
GPY1 0.00 %
GPY2 0.00 %
GPZ1 30.00 %
GPZ2 50.00 %
p15 500.00 usec
p16 1000.00 usec

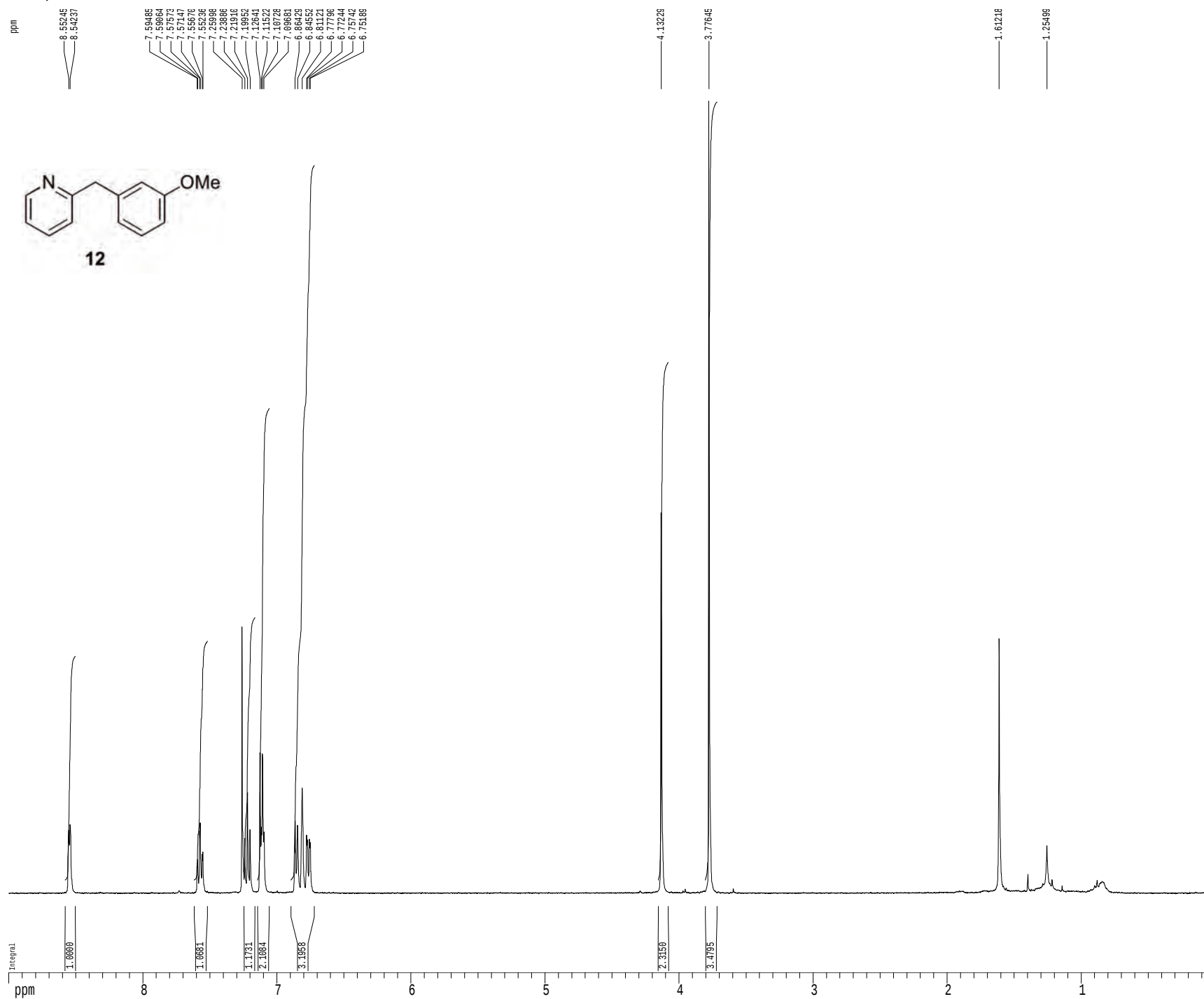
F2 - Processing parameters

SI 65536
SF 125.7804187 MHz
EN
WDW 0
SSB 0
LB 1.00 Hz
GB 0
PC 2.00

1D NMR plot parameters

CX 22.00 cm
CY 15.65 cm
F1P 220.000 ppm
F1 27671.69 Hz
F2P -5.000 ppm
F2 -628.90 Hz
PPMCM 9.86842 ppm/cm
HZCM 1241.25415 Hz/cm

¹H spectrum



Current Data Parameters
 USER jhanna
 NAME LEH-5-043-c1-f12
 EXPNO 1
 PROCNO 1

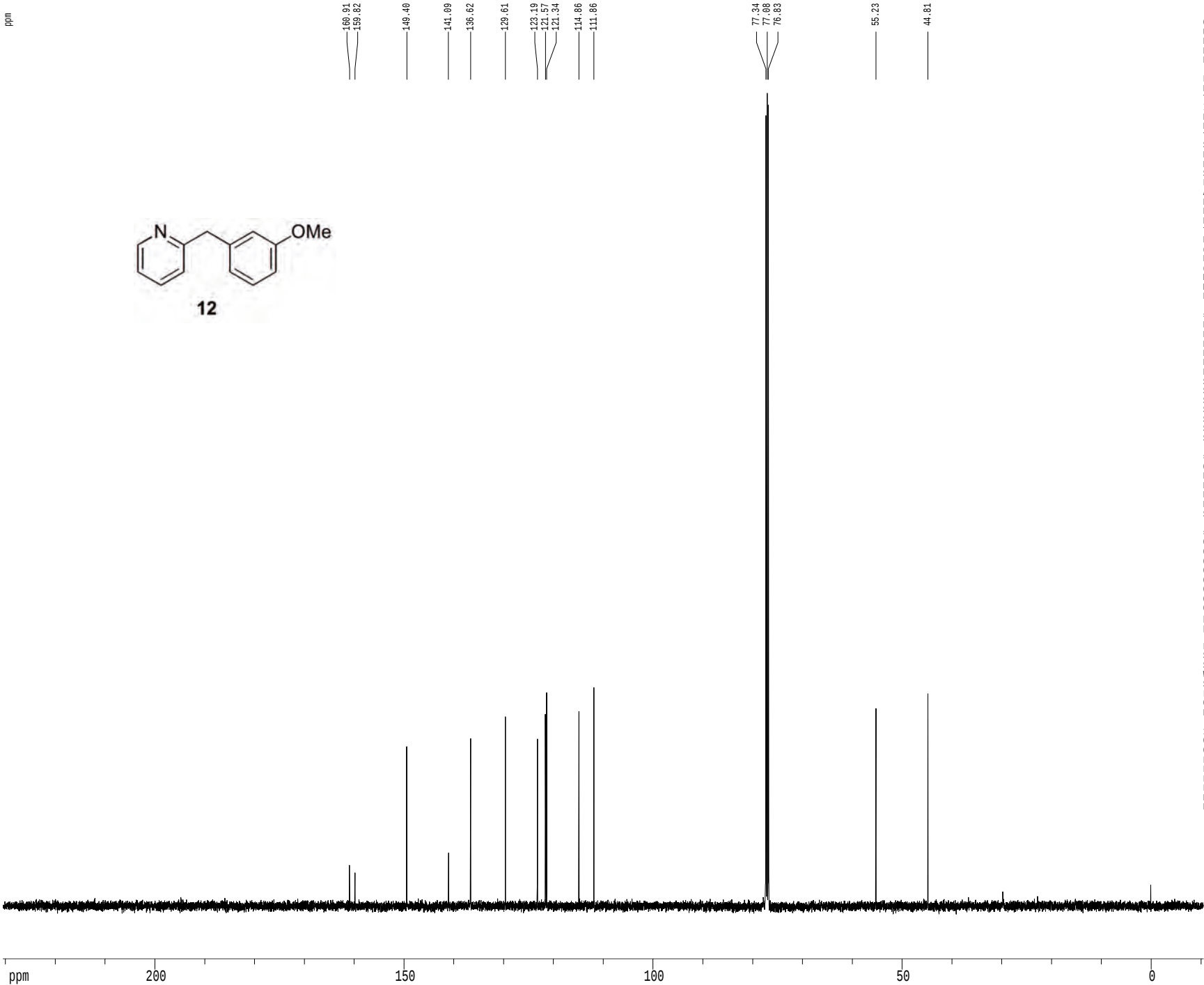
F2 - Acquisition Parameters
 Date_ 20141212
 Time 14.02
 INSTRUM drx400
 PROBHD 5 mm QNP H/F/P
 PULPROG zg30
 TD 25640
 SOLVENT CDCl3
 NS 8
 DS 2
 SWH 6410.256 Hz
 FIDRES 0.250010 Hz
 AQ 1.9999700 sec
 RG 912.3
 DW 78.000 usec
 DE 4.50 usec
 TE 298.0 K
 D1 0.10000000 sec
 MCREST 0.00000000 sec
 MCWRK 0.01500000 sec

===== CHANNEL f1 =====
 NUC1 1H
 P1 12.00 usec
 PL1 0.00 dB
 SF01 400.1328009 MHz

F2 - Processing parameters
 SI 65536
 SF 400.1300209 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 2.00

1D NMR plot parameters
 CX 22.00 cm
 CY 15.00 cm
 F1P 9.000 ppm
 F1 3691.17 Hz
 F2P 0.000 ppm
 F2 0.00 Hz
 PPMCM 0.39474 ppm/cm
 HZCM 157.94608 Hz/cm

Z-restored spin-echo 13C spectrum with 1H decoupling



Current Data Parameters

USER lhanna
NAME LEH-5-043-cl-f12-C13
EXPNO 1
PROCNO 1

F2 - Acquisition Parameters

Date_ 20141212
Time 16.30
INSTRUM cryo500
PROBHD 5 mm CPTCI 1H-
PULPROG SpineEcho30ppr
TO 65536
SOLVENT CDCl3
NS 1024
DS 16
SWH 30303.031 Hz
FIDRES 0.462388 Hz
AQ 1.0813940 sec
RG 2896.3
DW 16.500 usec
DE 6.00 usec
TE 298.0 K
D1 0.25000000 sec
d11 0.03000000 sec
D16 0.00020000 sec
d17 0.00019000 sec
MCREST 0.00000000 sec
MCWRK 0.01500000 sec
P2 33.10 usec

===== CHANNEL f1 =====

NUC1 13C
P1 16.55 usec
P11 500.00 usec
P12 2000.00 usec
PL0 120.00 dB
PL1 0.00 dB
SF01 125.7942548 MHz
SP1 2.70 dB
SP2 2.70 dB
SPNAM1 Crp60,0.5,20.1
SPNAM2 Crp60comp,4
SPOFF1 0.00 Hz
SPOFF2 0.00 Hz

===== CHANNEL f2 =====

CPDPRG2 waltz16
NUC2 1H
PCPD2 100.00 usec
PL2 1.00 dB
PL12 24.50 dB
SF02 500.2225011 MHz

===== GRADIENT CHANNEL =====

GP1AM1 SINE.100
GP1AM2 SINE.100
GPX1 0.00 %
GPX2 0.00 %
GPY1 0.00 %
GPY2 0.00 %
GPZ1 30.00 %
GPZ2 50.00 %
p15 500.00 usec
p16 1000.00 usec

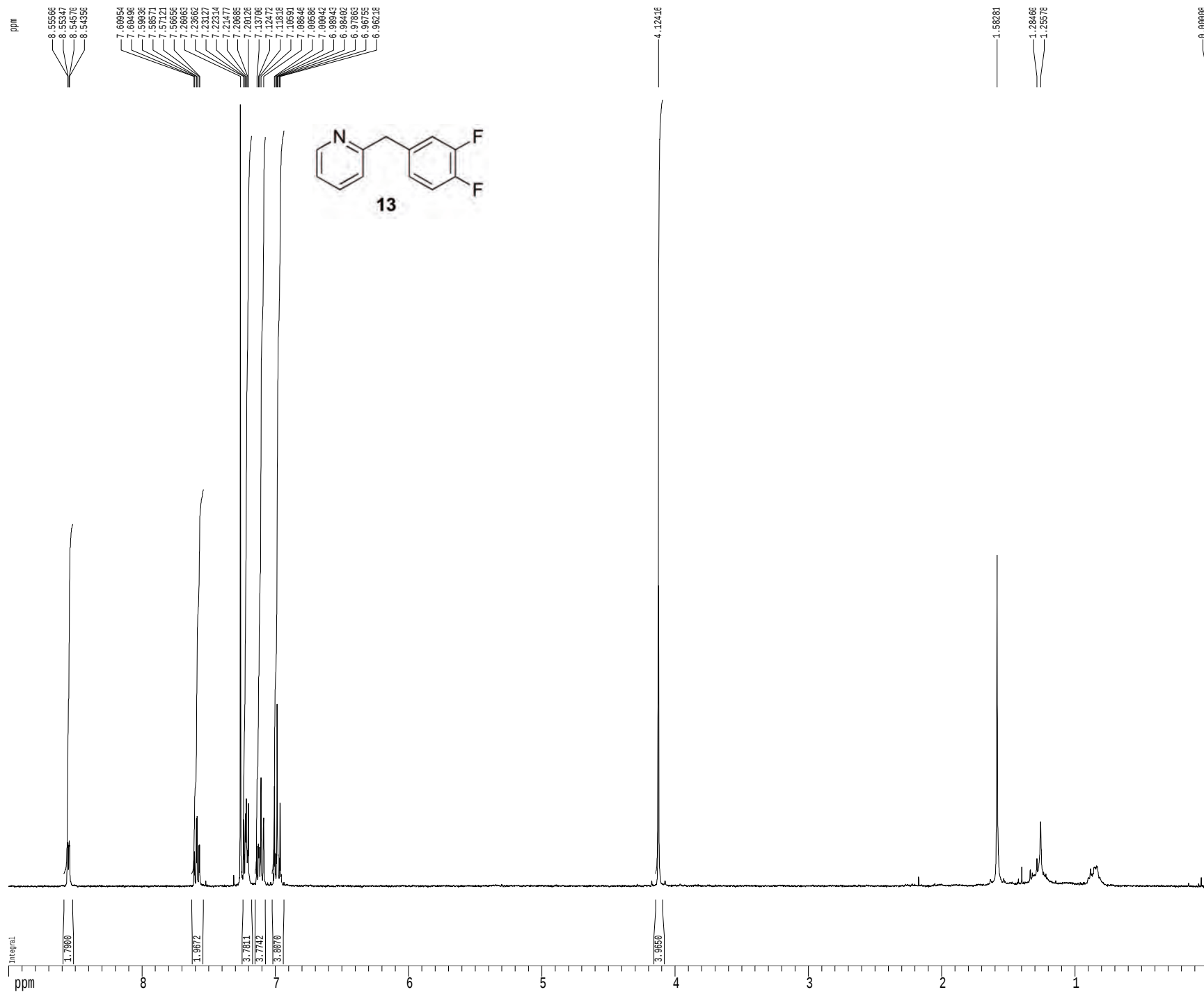
F2 - Processing parameters

SI 65536
SF 125.7804190 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 2.00

1D NMR plot parameters

CX 22.80 cm
CY 15.65 cm
F1P 230.460 ppm
F1 28987.37 Hz
F2P -10.460 ppm
F2 -1315.66 Hz
PPMCM 10.5667 ppm/cm
HZCM 1329.68032 Hz/cm

¹H spectrum



Current Data Parameters
 USER jhanna
 NAME LEH-5-150-c1-f24
 EXPNO 1
 PROCNO 1

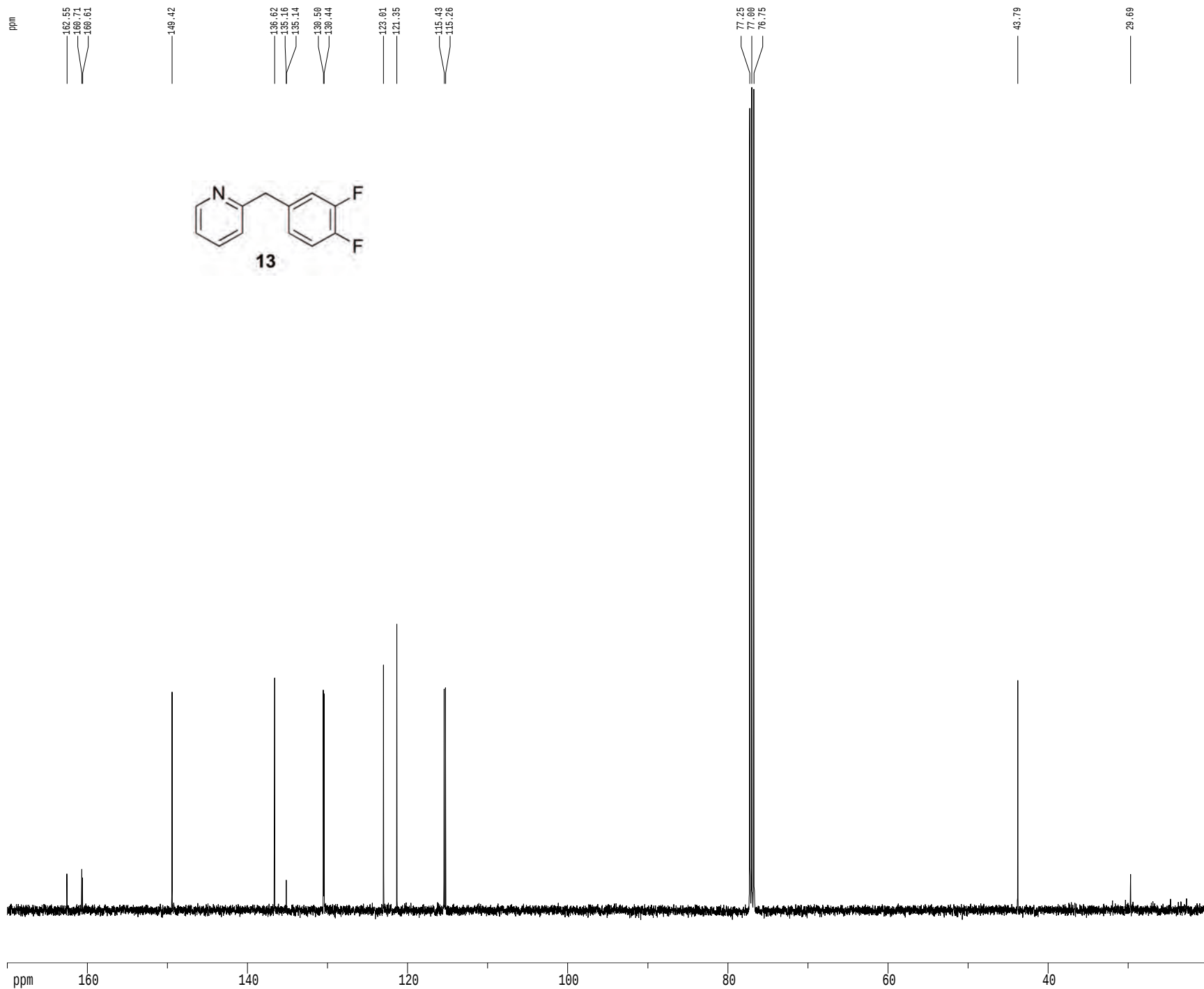
F2 - Acquisition Parameters
 Date_ 20150318
 Time 12.36
 INSTRUM drx400
 PROBHD 5 mm QNP H/F/P
 PULPROG zg30
 TD 25640
 SOLVENT CDCl3
 NS 8
 DS 2
 SWH 6410.256 Hz
 FIDRES 0.250010 Hz
 AQ 1.9999700 sec
 RG 1024
 DW 78.000 usec
 DE 4.50 usec
 TE 298.0 K
 D1 0.10000000 sec
 MCREST 0.00000000 sec
 MCWRR 0.01500000 sec

===== CHANNEL f1 =====
 NUC1 1H
 P1 12.00 usec
 PL1 0.00 dB
 SF01 400.1328009 MHz

F2 - Processing parameters
 SI 65536
 SF 400.1300206 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 2.00

1D NMR plot parameters
 CX 22.00 cm
 CY 15.00 cm
 F1P 9.000 ppm
 F1 3601.17 Hz
 F2P 0.000 ppm
 F2 0.00 Hz
 PPMCM 0.39474 ppm/cm
 HZCM 157.94608 Hz/cm

Z-restored spin-echo 13C spectrum with 1H decoupling



```

Current Data Parameters
USER          lhanna
NAME          LEH-5-150-C13-r1
EXPNO         1
PROCNO        1

F2 - Acquisition Parameters
Date_         20150320
Time          14.11
INSTRUM       cryo500
PROBHD        5 mm CPTCI 1H-
PULPROG       SpinEchopg30gp.prd
TD            65536
SOLVENT       CDCl3
NS            1024
DS            16
SWH           30303.031 Hz
FIDRES        0.462388 Hz
AQ            1.0813940 sec
RG            7298.2
DW            16.500 usec
DE            6.00 usec
TE            298.0 K
D1            0.25000000 sec
d11           0.03000000 sec
D16           0.00020000 sec
d17           0.00019600 sec
MCREST        0.00000000 sec
MCWRK         0.01500000 sec
P2            33.10 usec

===== CHANNEL f1 =====
NUC1           13C
P1            16.55 usec
P11           500.00 usec
P12           2000.00 usec
PL0           120.00 dB
PL1           -1.00 dB
SF01          125.7942548 MHz
SP1           2.70 dB
SP2           2.70 dB
SFO1          125.7942548 MHz
SFO2          125.7942548 MHz
SFO3          125.7942548 MHz
SFO4          125.7942548 MHz
SFO5          125.7942548 MHz
SFO6          125.7942548 MHz
SFO7          125.7942548 MHz
SFO8          125.7942548 MHz
SFO9          125.7942548 MHz
SFO10         125.7942548 MHz
SFO11         125.7942548 MHz
SFO12         125.7942548 MHz
SFO13         125.7942548 MHz
SFO14         125.7942548 MHz
SFO15         125.7942548 MHz
SFO16         125.7942548 MHz
SFO17         125.7942548 MHz
SFO18         125.7942548 MHz
SFO19         125.7942548 MHz
SFO20         125.7942548 MHz
SFO21         125.7942548 MHz
SFO22         125.7942548 MHz
SFO23         125.7942548 MHz
SFO24         125.7942548 MHz
SFO25         125.7942548 MHz
SFO26         125.7942548 MHz
SFO27         125.7942548 MHz
SFO28         125.7942548 MHz
SFO29         125.7942548 MHz
SFO30         125.7942548 MHz
SFO31         125.7942548 MHz
SFO32         125.7942548 MHz
SFO33         125.7942548 MHz
SFO34         125.7942548 MHz
SFO35         125.7942548 MHz
SFO36         125.7942548 MHz
SFO37         125.7942548 MHz
SFO38         125.7942548 MHz
SFO39         125.7942548 MHz
SFO40         125.7942548 MHz
SFO41         125.7942548 MHz
SFO42         125.7942548 MHz
SFO43         125.7942548 MHz
SFO44         125.7942548 MHz
SFO45         125.7942548 MHz
SFO46         125.7942548 MHz
SFO47         125.7942548 MHz
SFO48         125.7942548 MHz
SFO49         125.7942548 MHz
SFO50         125.7942548 MHz
SFO51         125.7942548 MHz
SFO52         125.7942548 MHz
SFO53         125.7942548 MHz
SFO54         125.7942548 MHz
SFO55         125.7942548 MHz
SFO56         125.7942548 MHz
SFO57         125.7942548 MHz
SFO58         125.7942548 MHz
SFO59         125.7942548 MHz
SFO60         125.7942548 MHz
SFO61         125.7942548 MHz
SFO62         125.7942548 MHz
SFO63         125.7942548 MHz
SFO64         125.7942548 MHz
SFO65         125.7942548 MHz
SFO66         125.7942548 MHz
SFO67         125.7942548 MHz
SFO68         125.7942548 MHz
SFO69         125.7942548 MHz
SFO70         125.7942548 MHz
SFO71         125.7942548 MHz
SFO72         125.7942548 MHz
SFO73         125.7942548 MHz
SFO74         125.7942548 MHz
SFO75         125.7942548 MHz
SFO76         125.7942548 MHz
SFO77         125.7942548 MHz
SFO78         125.7942548 MHz
SFO79         125.7942548 MHz
SFO80         125.7942548 MHz
SFO81         125.7942548 MHz
SFO82         125.7942548 MHz
SFO83         125.7942548 MHz
SFO84         125.7942548 MHz
SFO85         125.7942548 MHz
SFO86         125.7942548 MHz
SFO87         125.7942548 MHz
SFO88         125.7942548 MHz
SFO89         125.7942548 MHz
SFO90         125.7942548 MHz
SFO91         125.7942548 MHz
SFO92         125.7942548 MHz
SFO93         125.7942548 MHz
SFO94         125.7942548 MHz
SFO95         125.7942548 MHz
SFO96         125.7942548 MHz
SFO97         125.7942548 MHz
SFO98         125.7942548 MHz
SFO99         125.7942548 MHz
SFO100        125.7942548 MHz

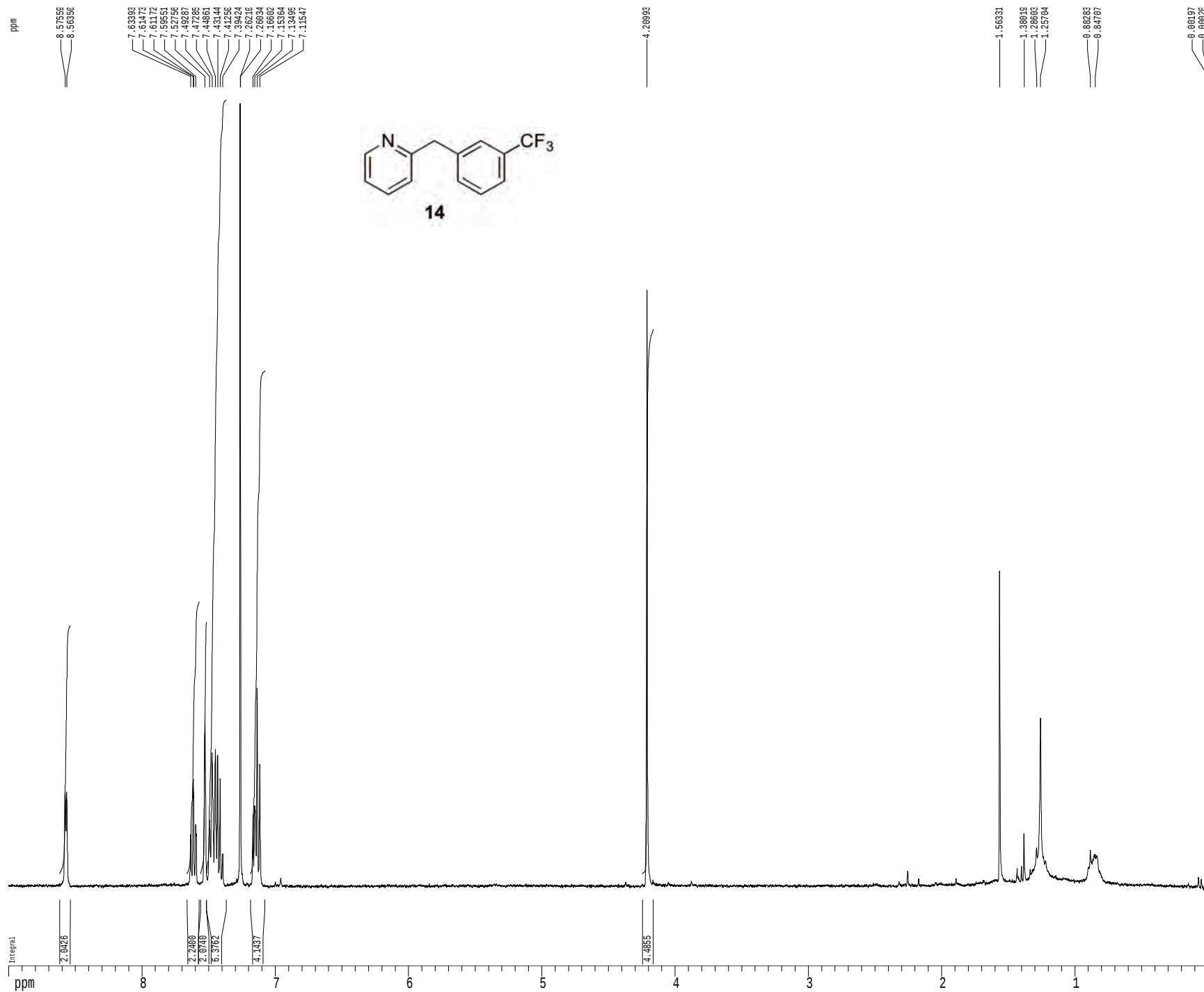
===== CHANNEL f2 =====
CPDPRG2       waltz16
NUC2           1H
PCPD2         100.00 usec
PL2           1.00 dB
PL12          24.50 dB
SF02          500.2225011 MHz

===== GRADIENT CHANNEL =====
GPNAM1        SINE.100
GPNAM2        SINE.100
GPX1          0.00 %
GPX2          0.00 %
GPY1          0.00 %
GPY2          0.00 %
GPZ1          30.00 %
GPZ2          50.00 %
p15           500.00 usec
p16           1000.00 usec

F2 - Processing parameters
SI            65536
SF            125.7804282 MHz
WDW           EM
SSB           0
LB            1.00 Hz
GB            0
PC            2.00

1D NMR plot parameters
CX            22.00 cm
CY            15.65 cm
F1P           170.000 ppm
F1            21382.67 Hz
F2P           20.000 ppm
F2            2515.61 Hz
PPMCM         6.57895 ppm/cm
HZCM          827.50281 Hz/cm
    
```

¹H spectrum



```

Current Data Parameters
USER          jhanna
NAME          LEH-5-068-c2-f16
EXPNO         1
PROCNO        1

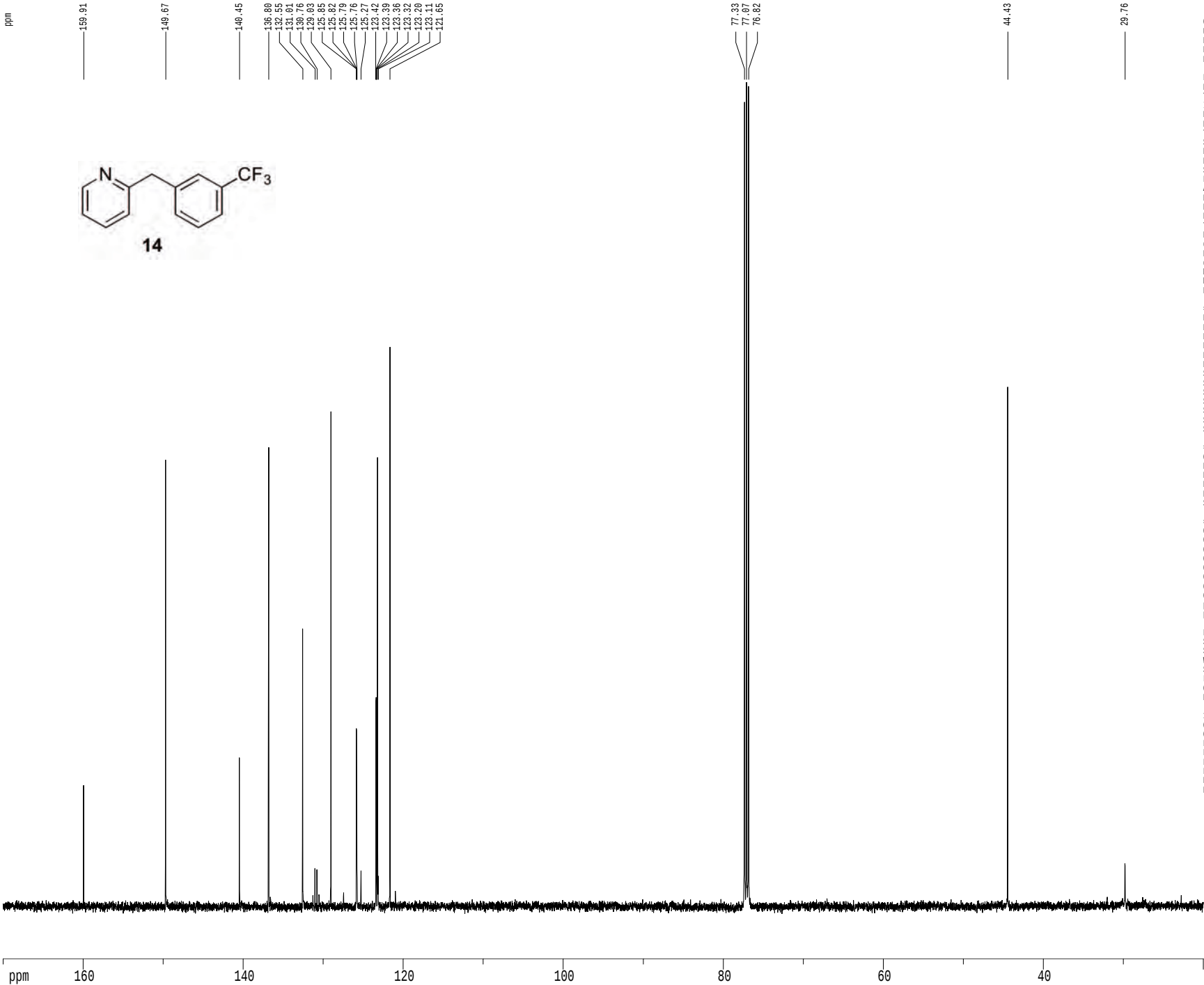
F2 - Acquisition Parameters
Date_         20150124
Time          17.48
INSTRUM       drx400
PROBHD        5 mm QNP H/F/P
PULPROG       zg30
TD            25640
SOLVENT       CDCl3
NS            8
DS            2
SWH           6410.256 Hz
FIDRES        0.250010 Hz
AQ            1.9999700 sec
RG            1149.4
DW            78.000 usec
DE            4.50 usec
TE            298.0 K
D1            0.10000000 sec
MCREST        0.00000000 sec
MCWRK         0.01500000 sec

===== CHANNEL f1 =====
NUC1          1H
P1            12.00 usec
PL1           0.00 dB
SF01          400.1328009 MHz

F2 - Processing parameters
SI            65536
SF            400.1300199 MHz
WDW           EM
SSB           0
LB            0.30 Hz
GB            0
PC            2.00

1D NMR plot parameters
CX            22.00 cm
CY            15.00 cm
F1P           9.000 ppm
F1            3681.17 Hz
F2P           0.000 ppm
F2            0.00 Hz
PPMCM         0.39474 ppm/cm
HZCM          157.94606 Hz/cm
    
```

Z-restored spin-echo 13C spectrum with 1H decoupling



Current Data Parameters

USER lhanna
NAME LEH-5-068-C2-C13-real
EXPNO 1
PROCNO 1

F2 - Acquisition Parameters

Date_ 20150129
Time 15.01
INSTRUM cryo500
PROBHD 5 mm CPTCI 1H-
PULPROG Spinecho300g.prd
TD 65536
SOLVENT CDCl3
NS 1281
DS 16
SWH 30303.031 Hz
FIDRES 0.462388 Hz
AQ 1.0813940 sec
RG 5792.6
DW 16.500 usec
DE 6.00 usec
TE 298.0 K
D1 0.50000000 sec
d11 0.03000000 sec
D16 0.00020000 sec
D17 0.00019000 sec
MCOREST 0.00000000 sec
MCORRK 0.01500000 sec
P2 33.10 usec

===== CHANNEL f1 =====

NUC1 13C
P1 16.55 usec
P11 500.00 usec
P12 2000.00 usec
PL0 120.00 dB
PL1 -1.00 dB
SFO1 125.7942540 MHz
SP1 2.70 dB
SP2 2.70 dB
SPNAM1 Crp60,0.5,20.1
SPNAM2 Crp60comp.4
SPOFF1 0.00 Hz
SPOFF2 0.00 Hz

===== CHANNEL f2 =====

CPDPRG2 waltz16
NUC2 1H
PCPD2 100.00 usec
PL2 1.00 dB
PL12 24.50 dB
SFO2 500.2225011 MHz

===== GRADIENT CHANNEL =====

GP1AM1 SINE.100
GP1AM2 SINE.100
GPX1 0.00 %
GPX2 0.00 %
GPY1 0.00 %
GPY2 0.00 %
GPZ1 30.00 %
GPZ2 50.00 %
p15 500.00 usec
p16 1000.00 usec

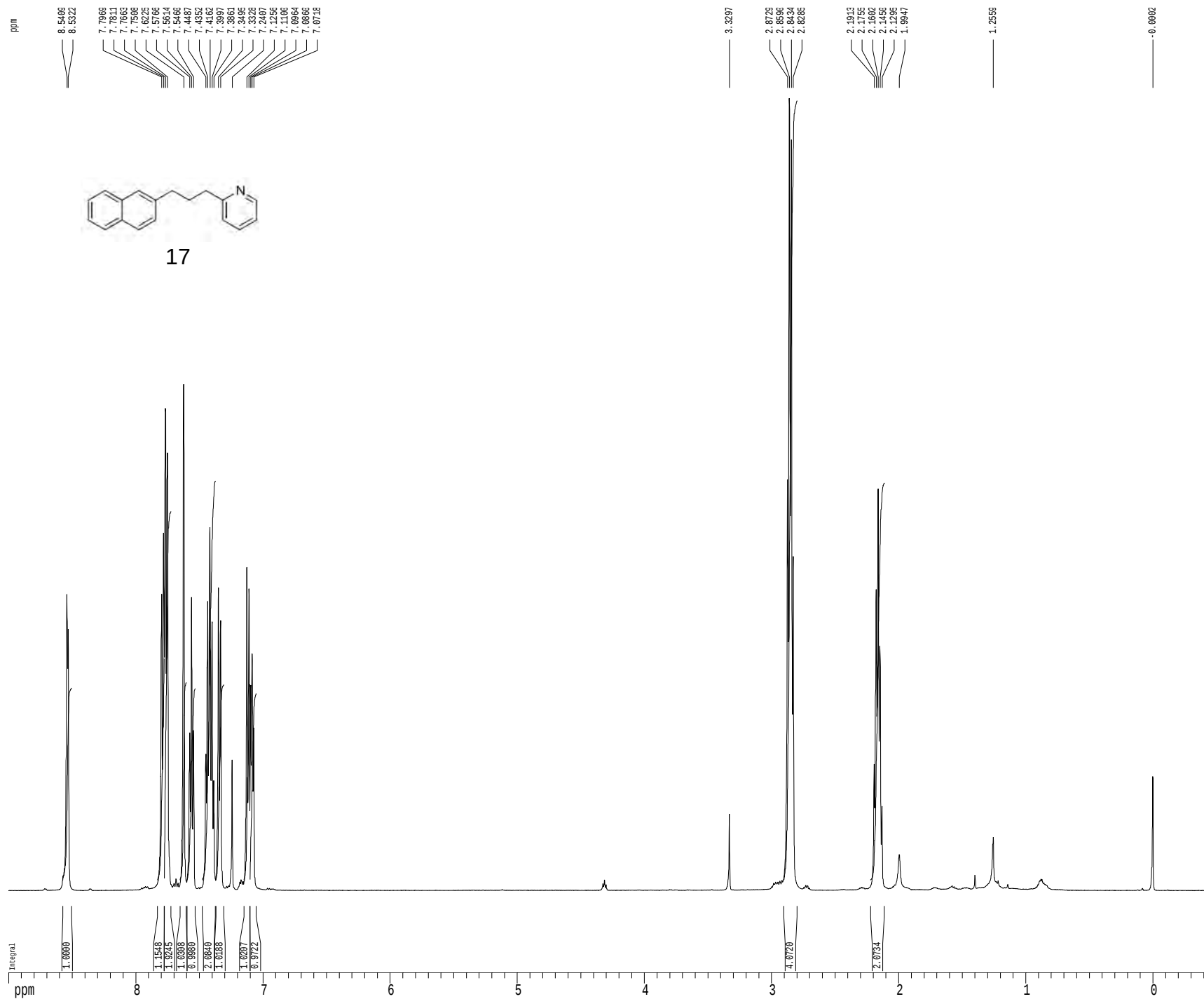
F2 - Processing parameters

SI 65536
SF 125.7804190 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 2.00

1D NMR plot parameters

CX 22.00 cm
CY 15.65 cm
F1P 170.000 ppm
F1 21302.67 Hz
F2P 20.000 ppm
F2 2515.61 Hz
PPMCM 6.57895 ppm/cm
HZCM 827.50281 Hz/cm

¹H spectrum



```

Current Data Parameters
NAME      MRH-VII-247-1HNMR
EXPNO     2
PROCNO    1

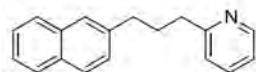
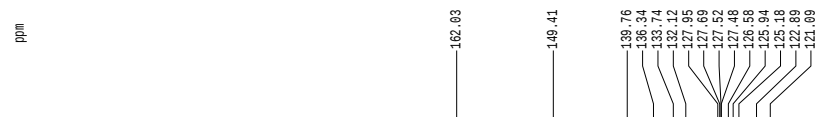
F2 - Acquisition Parameters
Date_     20150126
Time      11.03
INSTRUM    cryo500
PROBHD     5 mm CPTCI 1H-
PULPROG    zg30
TD         81728
SOLVENT    CDCl3
NS          8
DS          2
SWH         8012.820 Hz
FIDRES      0.098043 Hz
AQ         5.0998774 sec
RG          4.5
DW         62.400 usec
DE          6.00 usec
TE         298.0 K
D1         0.10000000 sec
MCREST     0.00000000 sec
MCNRK      0.01500000 sec

===== CHANNEL f1 =====
NUC1        1H
P1          7.50 usec
PL1         1.00 dB
SFO1       500.2235015 MHz

F2 - Processing parameters
SI          65536
SF         500.2200398 MHz
WDW         EM
SSB         0
LB          0.30 Hz
GB          0
PC          4.00

1D NMR plot parameters
CX          22.00 cm
CY          15.00 cm
F1P         9.000 ppm
F1          4501.98 Hz
F2P        -0.500 ppm
F2         -250.11 Hz
PPMCM       0.41667 ppm/cm
HZCM        208.42502 Hz/cm
    
```

¹³C spectrum with ¹H decoupling



17

Current Data Parameters
 USER mharri
 NAME MRH-VII-247-13CNMR
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20150124
 Time 17.09
 INSTRUM gn500
 PROBHD 5 mm broadband
 PULPROG zgpg30
 TD 65536
 SOLVENT CDCl3T
 NS 145
 DS 4
 SWH 30303.031 Hz
 FIDRES 0.462388 Hz
 AQ 1.0813849 sec
 RG 23170.5
 DW 16.500 usec
 DE 4.50 usec
 TE 298.0 K
 D1 0.25000000 sec
 d11 0.03000000 sec
 MCREST 0.00000000 sec
 MCWRR 0.01500000 sec

===== CHANNEL f1 =====
 NUC1 13C
 P1 9.00 usec
 PL1 -0.60 dB
 SF01 125.5327181 MHz

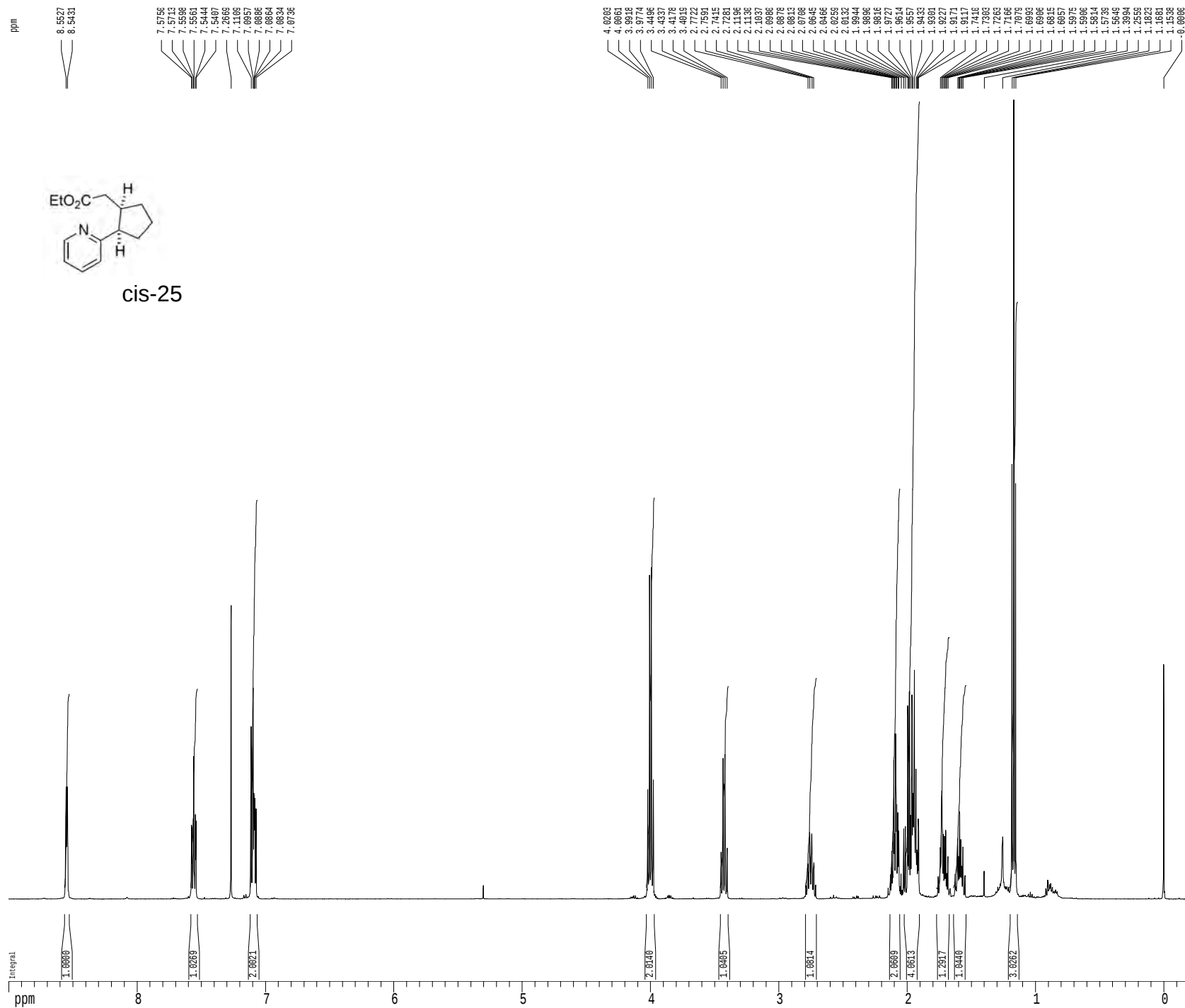
===== CHANNEL f2 =====
 CPDPRG2 waltz16
 NUC2 1H
 PCPD2 80.00 usec
 PL2 -3.00 dB
 PL12 12.80 dB
 SF02 499.1824959 MHz

F2 - Processing parameters
 SI 65536
 SF 125.5189062 MHz
 WDW EN
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 2.00

1D NMR plot parameters
 CX 22.80 cm
 CY 15.65 cm
 F2P 220.000 ppm
 F1 27614.16 Hz
 F2P -5.000 ppm
 F2 -627.59 Hz
 PPMCM 9.86842 ppm/cm
 HZCM 1238.67346 Hz/cm



¹H spectrum



Current Data Parameters
 USER mharri
 NAME MRH-VII-267-1HNMR
 EXPNO 1
 PROCNO 1

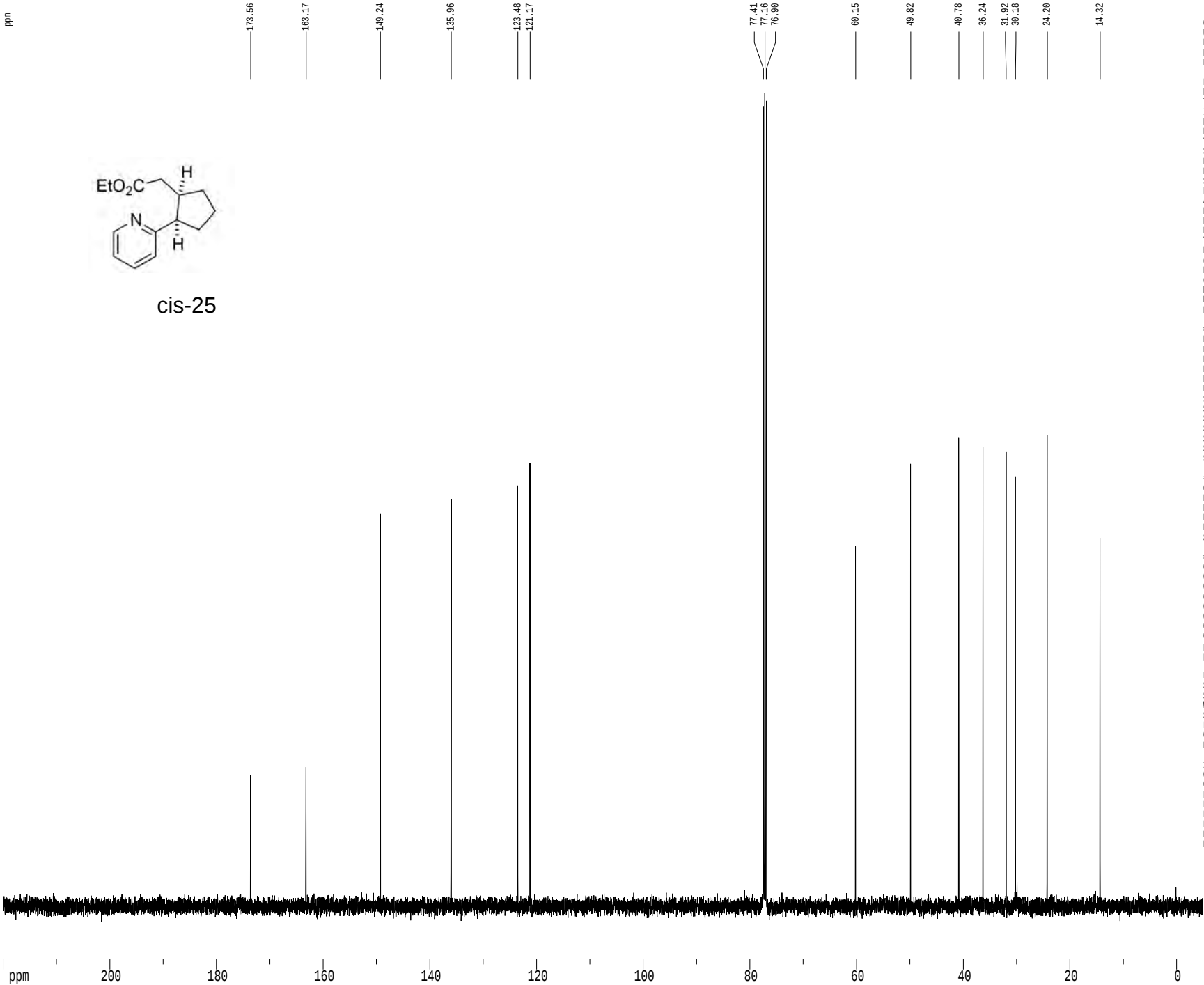
F2 - Acquisition Parameters
 Date_ 20150206
 Time 19.46
 INSTRUM cryo500
 PROBHD 5 mm CPTCI 1H-
 PULPROG zg30
 TO 81728
 SOLVENT CDCl3
 NS 8
 DS 2
 SWH 8012.820 Hz
 FIDRES 0.098043 Hz
 AQ 5.0998774 sec
 RG 7.1
 DW 62.400 usec
 DE 6.00 usec
 TE 298.0 K
 D1 0.10000000 sec
 MCREST 0.00000000 sec
 MCNRK 0.01500000 sec

===== CHANNEL f1 =====
 NUC1 1H
 P1 7.50 usec
 PL1 1.00 dB
 SF01 500.2235015 MHz

F2 - Processing parameters
 SI 6536
 SF 500.2200276 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 4.00

1D NMR plot parameters
 CX 22.00 cm
 CY 15.00 cm
 F1P 9.000 ppm
 F1 4501.98 Hz
 F2P -0.500 ppm
 F2 -250.11 Hz
 PPMCM 0.41667 ppm/cm
 HZCM 208.42502 Hz/cm

Z-restored spin-echo 13C spectrum with 1H decoupling



Current Data Parameters
USER mharri
NAME MRH-VII-267-13CNMR
EXPNO 1
PROCNO 1

F2 - Acquisition Parameters
Date_ 20150206
Time 19.49
INSTRUM cryo500
PROBHD 5 mm CPTCI 1H-
PULPROG SpinEchopg30gp.prd
TD 65536
SOLVENT CDCl3
NS 144
DS 16
SWH 30303.031 Hz
FIDRES 0.462388 Hz
AQ 1.0813940 sec
RG 3649.1
DW 16.500 usec
DE 6.00 usec
TE 298.0 K
D1 0.25000000 sec
d11 0.03000000 sec
D16 0.00020000 sec
d17 0.00019600 sec
MCREST 0.00000000 sec
MCWRR 0.01500000 sec
P2 33.10 usec

===== CHANNEL f1 =====
NUC1 13C
P1 16.55 usec
P11 500.00 usec
P12 2000.00 usec
PL0 120.00 dB
PL1 -1.00 dB
SF01 125.7942548 MHz
SP1 2.70 dB
SP2 2.70 dB
SPNAM1 Crp60,0.5,20.1
SPNAM2 Crp60comp.4
SPOFF1 0.00 Hz
SPOFF2 0.00 Hz

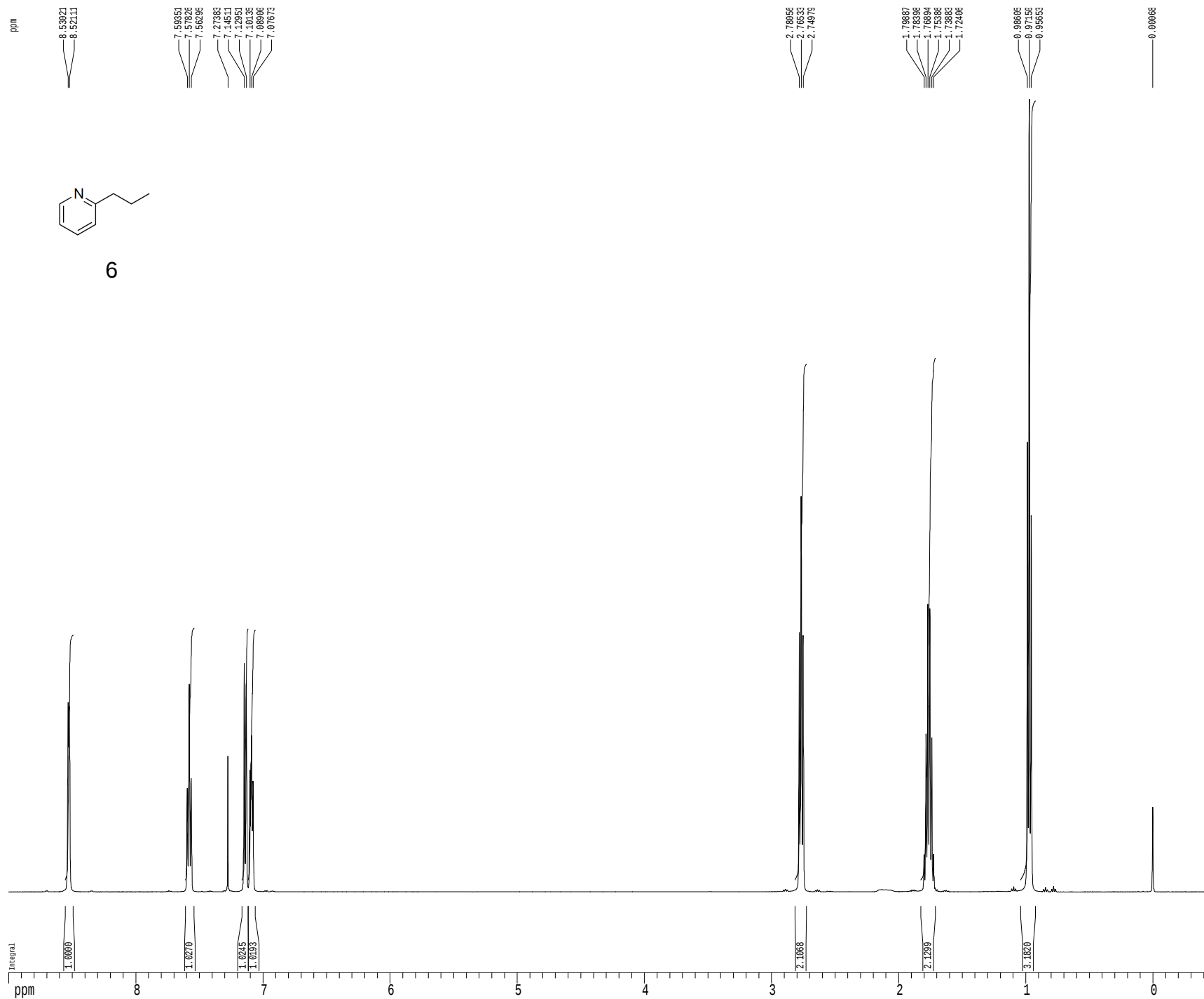
===== CHANNEL f2 =====
CPDPRG2 waltz16
NUC2 1H
PCPD2 100.00 usec
PL2 1.00 dB
PL12 24.50 dB
SF02 500.2225011 MHz

===== GRADIENT CHANNEL =====
GPNAM1 SINE.100
GPNAM2 SINE.100
GPX1 0.00 %
GPX2 0.00 %
GPY1 0.00 %
GPY2 0.00 %
GPZ1 30.00 %
GPZ2 50.00 %
p15 500.00 usec
p16 1000.00 usec

F2 - Processing parameters
SI 65536
SF 125.7804088 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 2.00

1D NMR plot parameters
CX 22.00 cm
CY 15.65 cm
F1P 220.000 ppm
F1 27671.69 Hz
F2P -5.000 ppm
F2 -628.90 Hz
PPMCM 9.86842 ppm/cm
HZCM 1241.25415 Hz/cm

¹H spectrum



```

Current Data Parameters
USER          mharri
NAME          MRH-VII-283-1HNMR
EXPNO         3
PROCNO        1

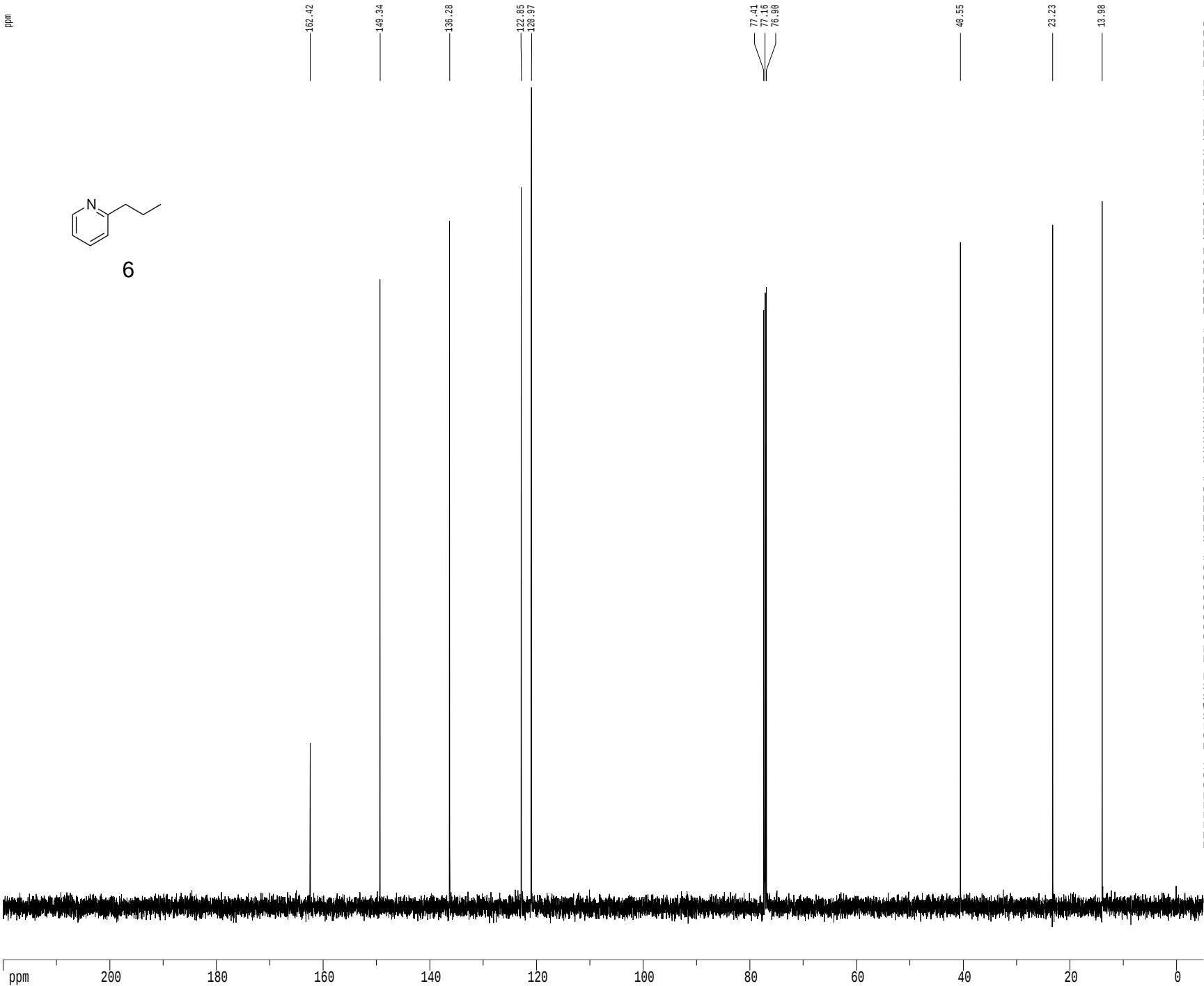
F2 - Acquisition Parameters
Date_         20150304
Time          17.30
INSTRUM       cryo500
PROBHD        5 mm CPTCI 1H-
PULPROG       zg30
TD            81728
SOLVENT       CDCl3
NS            8
DS            2
SWH           8012.820 Hz
FIDRES        0.098043 Hz
AQ            5.0998774 sec
RG            8
DM            62.400 usec
DE            6.00 usec
TE            298.0 K
D1            0.10000000 sec
MCREST        0.00000000 sec
MCWRK         0.01500000 sec

===== CHANNEL f1 =====
NUC1          1H
P1            7.50 usec
PL1           1.00 dB
SFO1          500.2235015 MHz

F2 - Processing parameters
SI            65536
SF            500.2200244 MHz
WDW           EM
SSB           0
LB            0.30 Hz
GB            0
PC            4.00

1D NMR plot parameters
CX            22.00 cm
CY            15.00 cm
F1P           9.000 ppm
F1            4501.98 Hz
F2P          -0.500 ppm
F2           -250.11 Hz
PPMCM         0.41667 ppm/cm
HZCM          208.42502 Hz/cm
    
```

Z-restored spin-echo 13C spectrum with 1H decoupling



Current Data Parameters

USER mharri
NAME MRH-VII-283-13CNMR
EXPNO 3
PROCNO 1

F2 - Acquisition Parameters

Date_ 20150304
Time 17.33
INSTRUM cryo500
PROBHD 5 mm CPTCI 1H-
PULPROG SpinEchopg30gp.prd
TD 65536
SOLVENT CDCl3
NS 54
DS 16
SWH 30303.031 Hz
FIDRES 0.462388 Hz
AQ 1.0813940 sec
RG 3649.1
DW 16.500 usec
DE 6.00 usec
TE 298.0 K
D1 0.25000000 sec
d11 0.03000000 sec
d16 0.00020000 sec
d17 0.00019600 sec
MCREST 0.00000000 sec
MCWRK 0.01500000 sec
P2 33.10 usec

===== CHANNEL f1 =====

NUC1 13C
P1 16.55 usec
P11 500.00 usec
P12 2000.00 usec
PL0 120.00 dB
PL1 -1.00 dB
SF01 125.7942548 MHz
SP1 2.70 dB
SP2 2.70 dB
SPNAM1 Crp60,0.5,20.1
SPNAM2 Crp60comp.4
SPOFF1 0.00 Hz
SPOFF2 0.00 Hz

===== CHANNEL f2 =====

CPDPRG2 waltz16
NUC2 1H
PCPD2 100.00 usec
PL2 1.00 dB
PL12 24.50 dB
SF02 500.2225011 MHz

===== GRADIENT CHANNEL =====

GPVAM1 SINE.100
GPVAM2 SINE.100
GPX1 0.00 %
GPX2 0.00 %
GPY1 0.00 %
GPY2 0.00 %
GPZ1 30.00 %
GPZ2 50.00 %
p15 500.00 usec
p16 1000.00 usec

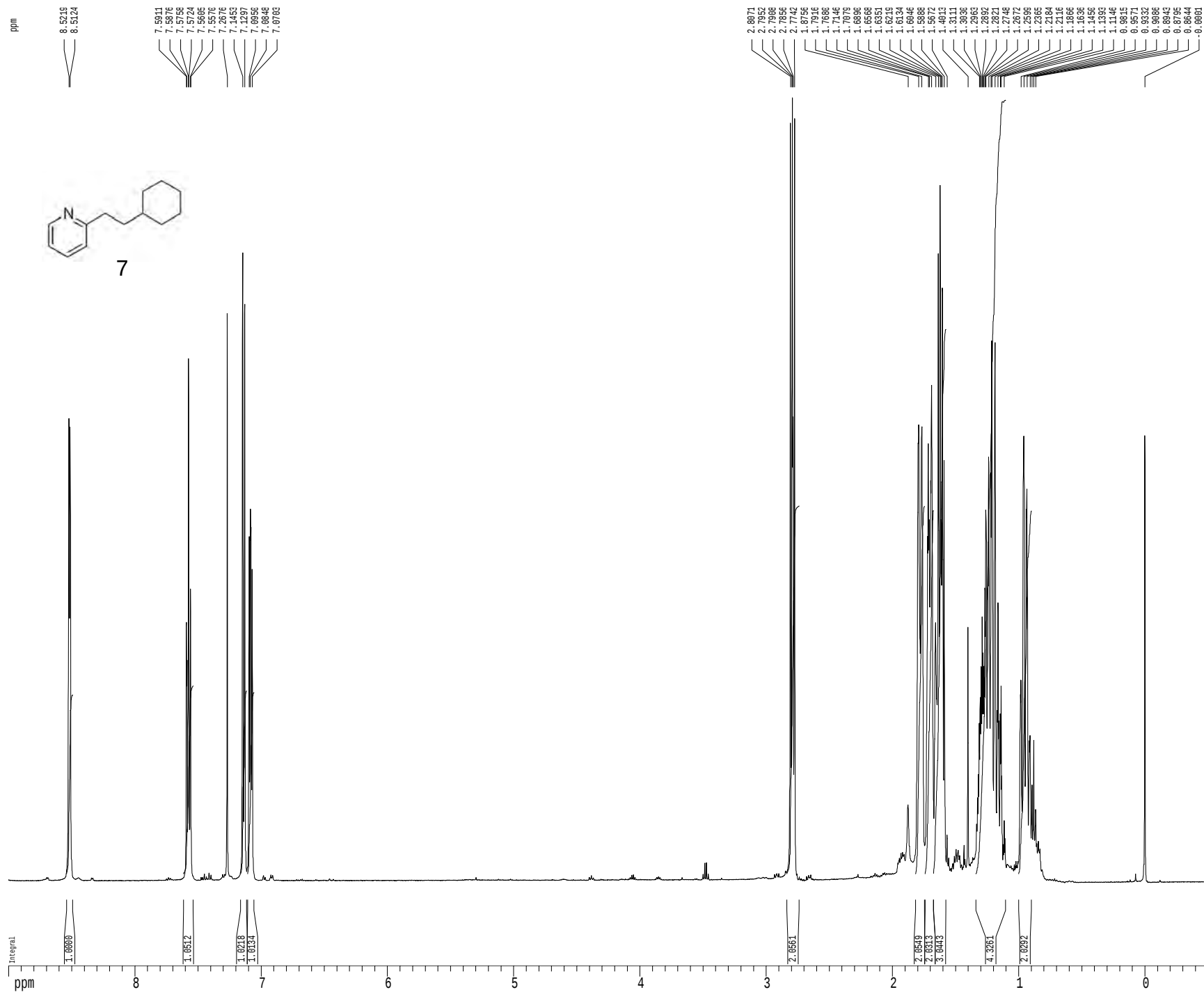
F2 - Processing parameters

SI 65536
SF 125.7804099 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 2.00

1D NMR plot parameters

CX 22.00 cm
CY 15.65 cm
F1P 220.000 ppm
F1 27671.69 Hz
F2P -5.000 ppm
F2 -628.90 Hz
PPMCM 9.86842 ppm/cm
HZCM 1241.25415 Hz/cm

¹H spectrum



Current Data Parameters
 USER mharri
 NAME MRH-VII-300-1HNMR
 EXPNO 1
 PROCNO 1

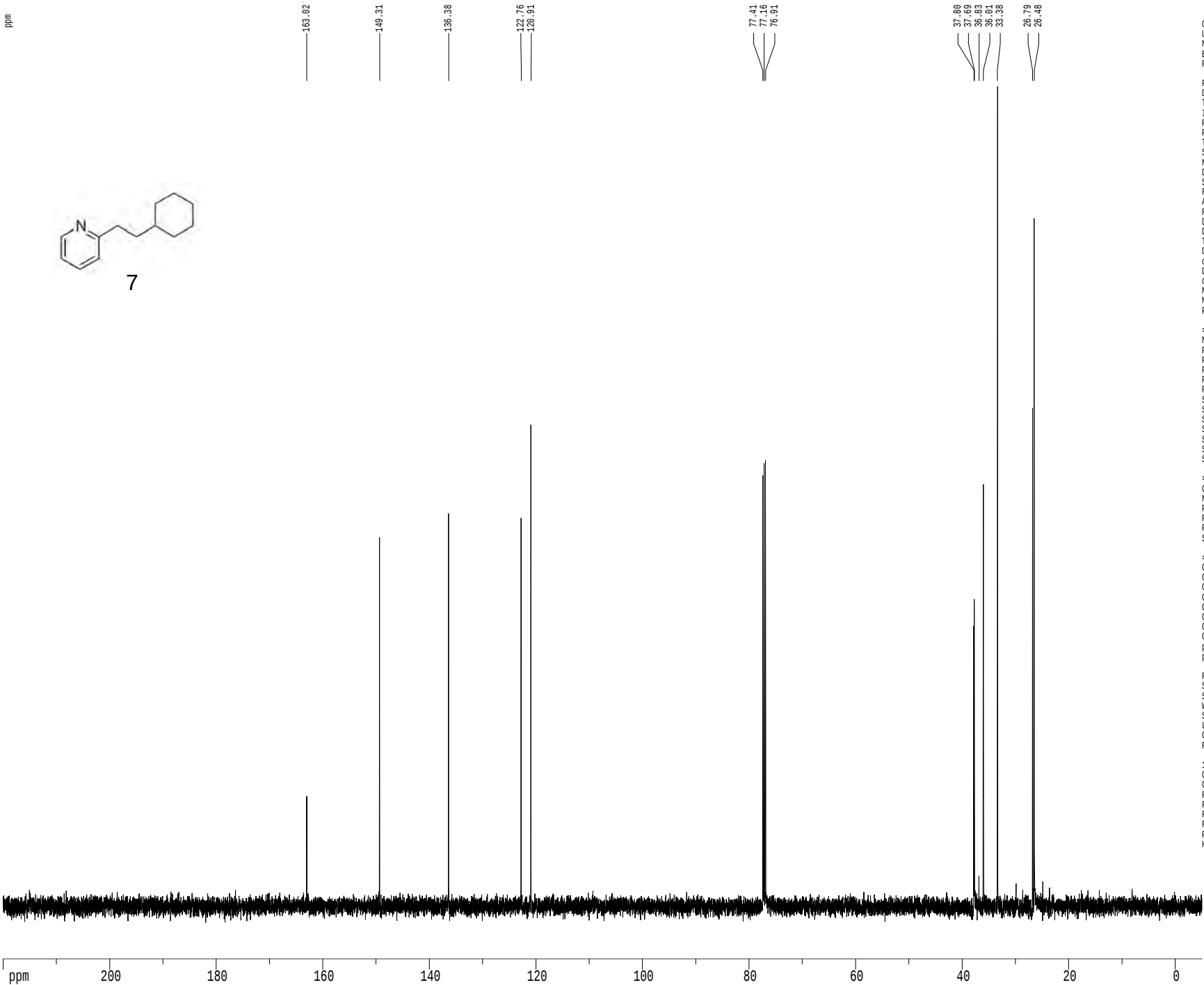
F2 - Acquisition Parameters
 Date_ 20150304
 Time 17.13
 INSTRUM cryo500
 PROBHD 5 mm CPTCI 1H-
 PULPROG zg30
 TO 81728
 SOLVENT CDCl3
 NS 8
 DS 2
 SMH 8012.820 Hz
 FIDRES 0.098043 Hz
 AQ 5.0998774 sec
 RG 5.7
 DW 62.400 usec
 DE 6.00 usec
 TE 298.0 K
 D1 0.10000000 sec
 MCREST 0.00000000 sec
 MCNWK 0.01500000 sec

===== CHANNEL f1 =====
 NUC1 1H
 P1 7.50 usec
 PL1 1.00 dB
 SFO1 500.2235015 MHz

F2 - Processing parameters
 SI 65536
 SF 500.2200275 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 4.00

1D NMR plot parameters
 CX 22.00 cm
 CY 15.00 cm
 F1P 9.000 ppm
 F1 4501.98 Hz
 F2P -0.500 ppm
 F2 -250.11 Hz
 PPMCM 0.41667 ppm/cm
 HZCM 208.42502 Hz/cm

Z-restored spin-echo 13C spectrum with 1H decoupling



Current Data Parameters

USER mharri
NAME MRH-VII-300-13CNMR
EXPNO 1
PROCNO 1

F2 - Acquisition Parameters

Date_ 20150304
Time 17.16
INSTRUM cryo500
PROBHD 5 mm CPTCI 1H-
PULPROG SpinEchopg30gp.prd
TD 65536
SOLVENT CDCl3
NS 85
DS 16
SWH 30303.031 Hz
FIDRES 0.462388 Hz
AQ 1.0813940 sec
RG 7298.2
DW 16.500 usec
DE 6.00 usec
TE 298.0 K
D1 0.25000000 sec
d11 0.03000000 sec
D16 0.00020000 sec
d17 0.00019600 sec
MCREST 0.00000000 sec
MCWRR 0.01500000 sec
P2 33.10 usec

===== CHANNEL f1 =====

NUC1 13C
P1 16.55 usec
P11 500.00 usec
P12 2000.00 usec
PL0 120.00 dB
PL1 -1.00 dB
SF01 125.7942548 MHz
SP1 2.70 dB
SP2 2.70 dB
SPNAM1 Crp60,0.5,20.1
SPNAM2 Crp60comp.4
SPOFF1 0.00 Hz
SPOFF2 0.00 Hz

===== CHANNEL f2 =====

CPDPRG2 waltz16
NUC2 1H
PCPD2 100.00 usec
PL2 -1.00 dB
PL12 24.50 dB
SF02 500.2225011 MHz

===== GRADIENT CHANNEL =====

GP1AM1 SINE.100
GP1AM2 SINE.100
GPX1 0.00 %
GPX2 0.00 %
GPY1 0.00 %
GPY2 0.00 %
GPZ1 30.00 %
GPZ2 50.00 %
p15 500.00 usec
p16 1000.00 usec

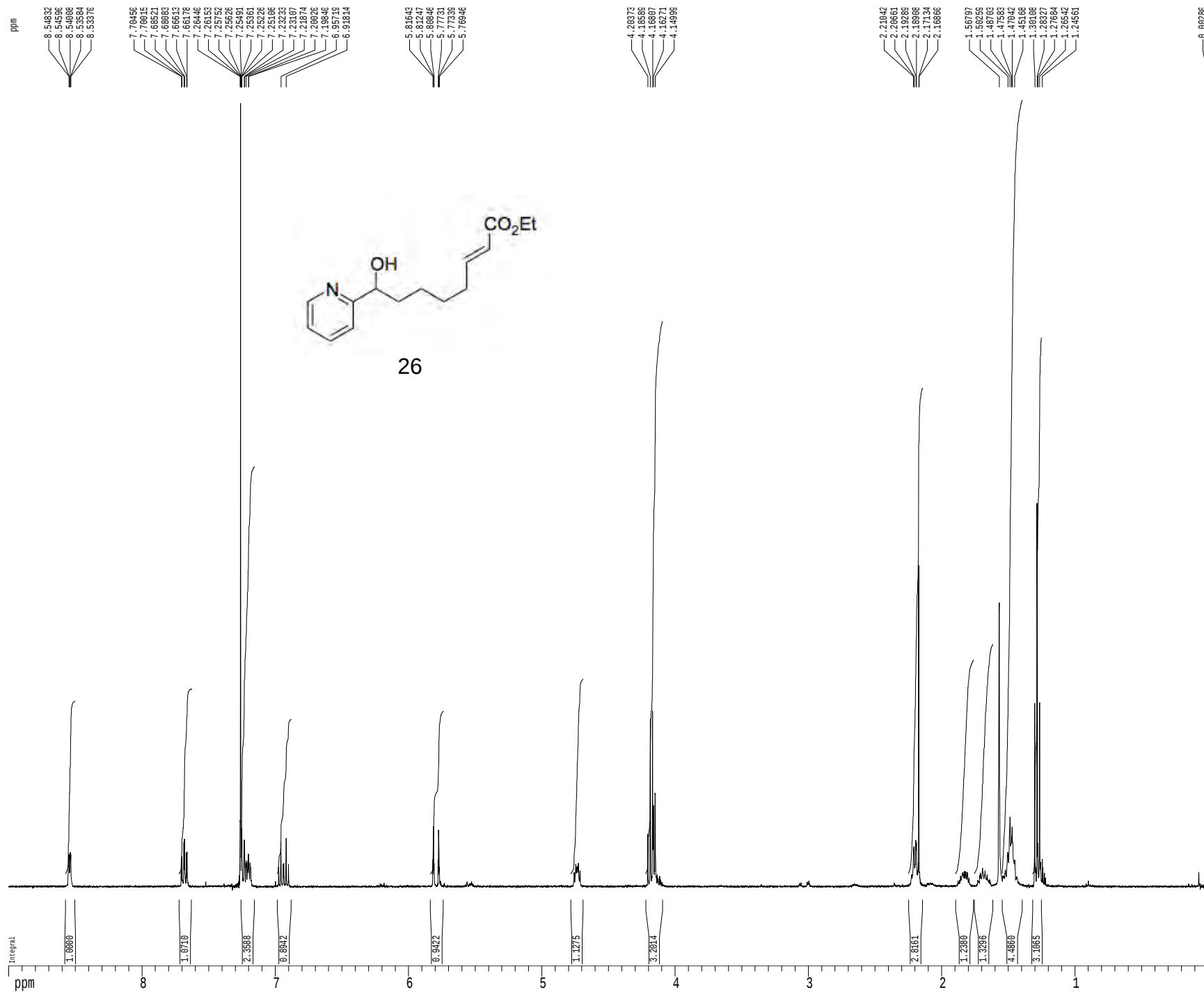
F2 - Processing parameters

SI 65536
SF 125.7804090 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 2.00

1D NMR plot parameters

CX 22.00 cm
CY 15.65 cm
F1P 220.000 ppm
F1 27671.69 Hz
F2P -5.000 ppm
F2 -628.90 Hz
PPMCM 9.86842 ppm/cm
HZCM 1241.25415 Hz/cm

¹H spectrum



Current Data Parameters
 USER lnama
 NAME KD-1-083cl
 EXPNO 1
 PROCNO 1

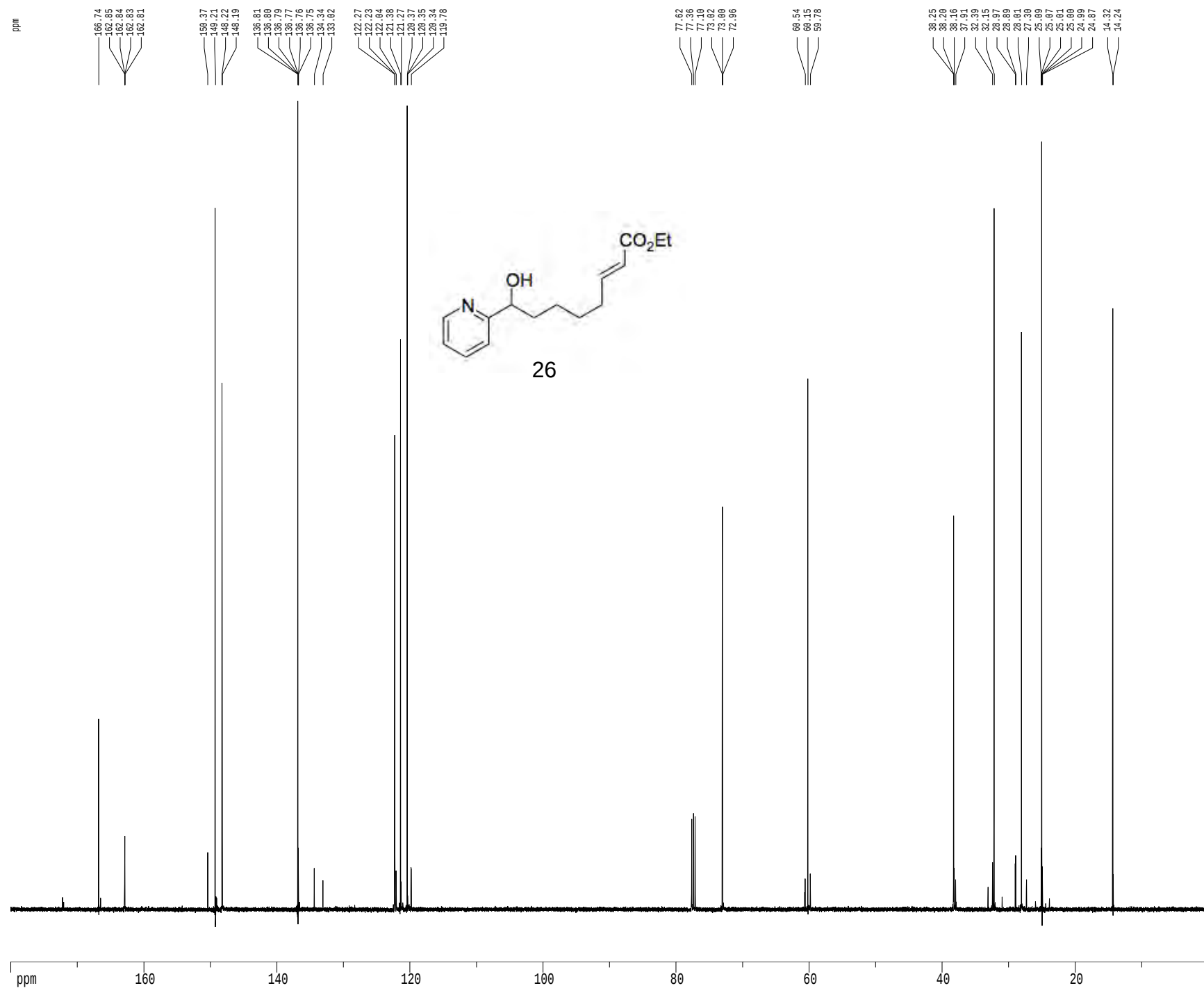
F2 - Acquisition Parameters
 Date_ 20161019
 Time 15.33
 INSTRUM drx400
 PROBHD 5 mm QNP H/F/P
 PULPROG zg30
 TD 25640
 SOLVENT CDCl3
 NS 8
 DS 2
 SWH 6410.256 Hz
 FIDRES 0.250010 Hz
 AQ 1.9999700 sec
 RG 812.7
 DW 78.000 usec
 DE 4.50 usec
 TE 297.9 K
 D1 0.10000000 sec
 MCREST 0.00000000 sec
 MCWRK 0.01500000 sec

===== CHANNEL f1 =====
 NUC1 1H
 P1 12.00 usec
 PL1 0.00 dB
 SF01 400.1328009 MHz

F2 - Processing parameters
 SI 65536
 SF 400.1300210 MHz
 MDW no
 SSB 0
 LB 0.00 Hz
 GB 0
 PC 2.00

1D NMR plot parameters
 CX 22.00 cm
 CY 15.00 cm
 F1P 9.000 ppm
 F1 3601.17 Hz
 F2P 0.000 ppm
 F2 0.00 Hz
 PPMCM 0.39474 ppm/cm
 HZCM 157.94608 Hz/cm

Z-restored spin-echo 13C spectrum with 1H decoupling



Current Data Parameters

USER	lhanna
NAME	KD-1-083-6MEM-C13
EXPNO	1
PROCNO	1

F2 - Acquisition Parameters

Date_	20161220
Time	13.34
INSTRUM	cryo500
PROBHD	5 mm CPTCI 1H-
PULPROG	SpinEchopg30gp.prd
TD	65536
SOLVENT	CDCl3
NS	115
DS	10
SWH	30303.031 Hz
FIDRES	0.462388 Hz
AQ	1.0813940 sec
RG	7298.2
DW	16.500 usec
DE	6.00 usec
TE	298.0 K
D1	0.25000000 sec
d11	0.03000000 sec
D16	0.00020000 sec
d17	0.00019600 sec
MCREST	0.00000000 sec
MCWRK	0.01500000 sec
P2	33.10 usec

===== CHANNEL f1 =====

NUC1	13C
P1	16.55 usec
P11	500.00 usec
P12	2000.00 usec
PL0	120.00 dB
PL1	-1.00 dB
SFO1	125.7942548 MHz
SP1	2.70 dB
SP2	2.70 dB
SPNAM1	Crp60,0.5,20.1
SPNAM2	Crp60comp.4
SPOFF1	0.00 Hz
SPOFF2	0.00 Hz

===== CHANNEL f2 =====

CPDPRG2	waltz16
NUC2	1H
PCPD2	100.00 usec
PL2	-1.00 dB
PL12	24.50 dB
SFO2	500.2225011 MHz

===== GRADIENT CHANNEL =====

GPVAM1	SINE.100
GPVAM2	SINE.100
GPX1	0.00 %
GPX2	0.00 %
GPY1	0.00 %
GPY2	0.00 %
GPZ1	30.00 %
GPZ2	50.00 %
p15	500.00 usec
p16	1000.00 usec

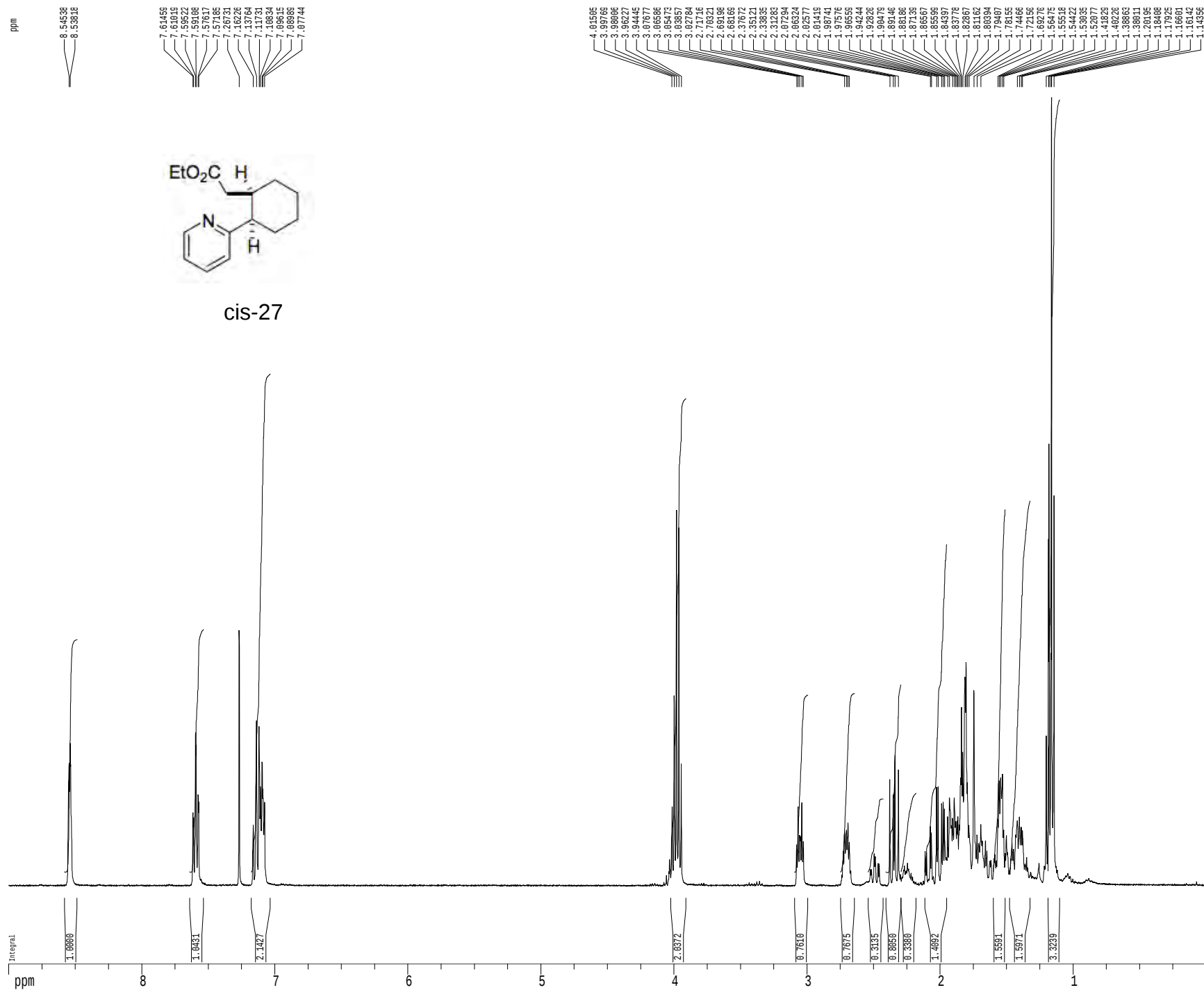
F2 - Processing parameters

SI	65536
SF	125.7804069 MHz
WDW	no
SSB	0
LB	0.00 Hz
GB	0
PC	2.00

1D NMR plot parameters

CX	22.00 cm
CY	15.65 cm
F1P	180.000 ppm
F1	22640.47 Hz
F2P	0.000 ppm
F2	0.00 Hz
PPMCM	7.89474 ppm/cm
HZCM	993.00323 Hz/cm

¹H spectrum



Current Data Parameters
 USER Inanna
 NAME LEH-7-086-c1-f1
 EXPNO 1
 PROCNO 1

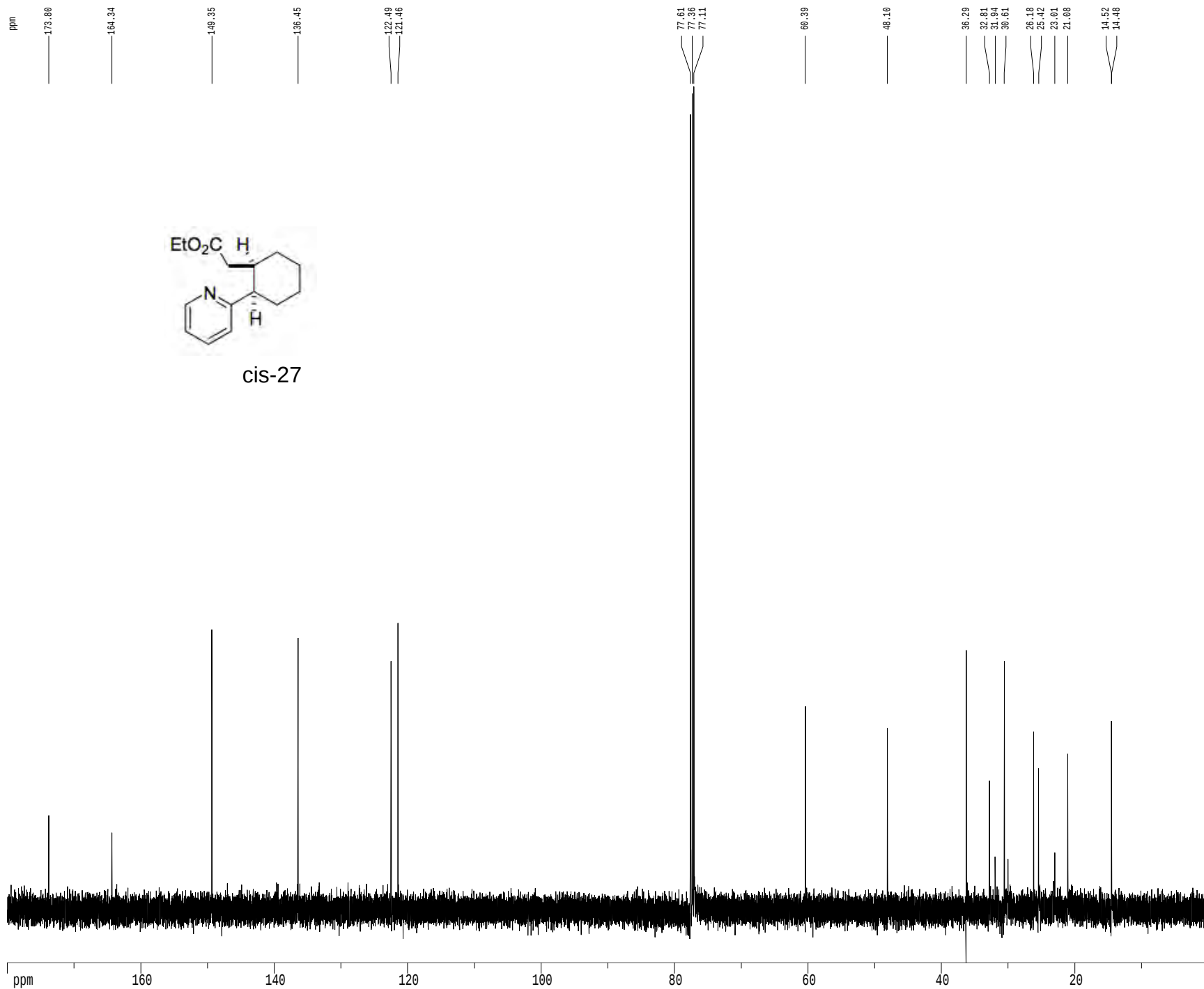
F2 - Acquisition Parameters
 Date_ 20161213
 Time 14.33
 INSTRUM drx400
 PROBHD 5 mm QNP H/F/P
 PULPROG zg30
 TD 25640
 SOLVENT CDCl3
 NS 8
 DS 2
 SWH 6410.256 Hz
 FIDRES 0.250010 Hz
 AQ 1.9999700 sec
 RG 228.1
 DW 78.000 usec
 DE 4.50 usec
 TE 298.0 K
 D1 0.10000000 sec
 MCREST 0.00000000 sec
 MCWRK 0.01500000 sec

===== CHANNEL f1 =====
 NUC1 1H
 P1 12.00 usec
 PL1 0.00 dB
 SF01 400.1328009 MHz

F2 - Processing parameters
 SI 65536
 SF 400.1300189 MHz
 WDW no
 SSB 0
 LB 0.00 Hz
 GB 0
 PC 2.00

1D NMR plot parameters
 CX 22.00 cm
 CY 15.00 cm
 F1P 9.000 ppm
 F1 3601.17 Hz
 F2P 0.000 ppm
 F2 0.00 Hz
 PPMCM 0.39474 ppm/cm
 HZCM 157.94606 Hz/cm

Z-restored spin-echo 13C spectrum with 1H decoupling



Current Data Parameters
 USER lhanna
 NAME LEH-7-099-c1-MJR Dr-C13
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20161222
 Time 11:10
 INSTRUM cryo500
 PROBH0 5 mm CPTCI 1H-
 PULPROG SpinEcho9389p.prd
 TD 65536
 SOLVENT CDCl3
 NS 800
 DS 16
 SWH 30303.031 Hz
 FIDRES 0.462300 Hz
 AQ 1.0813940 sec
 RG 5792.6
 DW 16.500 usec
 DE 6.00 usec
 TE 298.0 K
 D1 0.25000000 sec
 d11 0.03000000 sec
 D16 0.00020000 sec
 d17 0.00019000 sec
 MCREST 0.00000000 sec
 MCWIK 0.01500000 sec
 P2 33.10 usec

===== CHANNEL f1 =====
 NUC1 13C
 P1 16.55 usec
 P11 500.00 usec
 P12 2000.00 usec
 PL0 120.00 dB
 PL1 -1.00 dB
 SF01 125.7942548 MHz
 SP1 2.70 dB
 SP2 2.70 dB
 SPANM1 Crp60,0.5,20.1
 SPANM2 Crp60comp,4
 SPOFF1 0.00 Hz
 SPOFF2 0.00 Hz

===== CHANNEL f2 =====
 CPDPRG2 waltz16
 NUC2 1H
 PCPD2 100.00 usec
 PL2 1.00 dB
 PL12 24.50 dB
 SF02 500.2226011 MHz

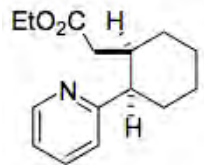
===== GRADIENT CHANNEL =====
 GPMAM1 SINE.100
 GPMAM2 SINE.100
 GPX1 0.00 %
 GPX2 0.00 %
 GPY1 0.00 %
 GPY2 0.00 %
 GPZ1 30.00 %
 GPZ2 50.00 %
 p15 500.00 usec
 p16 1000.00 usec

F2 - Processing parameters
 SI 65536
 SF 125.7803820 MHz
 WDW no
 SSB 0
 LB 0.00 Hz
 GB 0
 PC 2.00

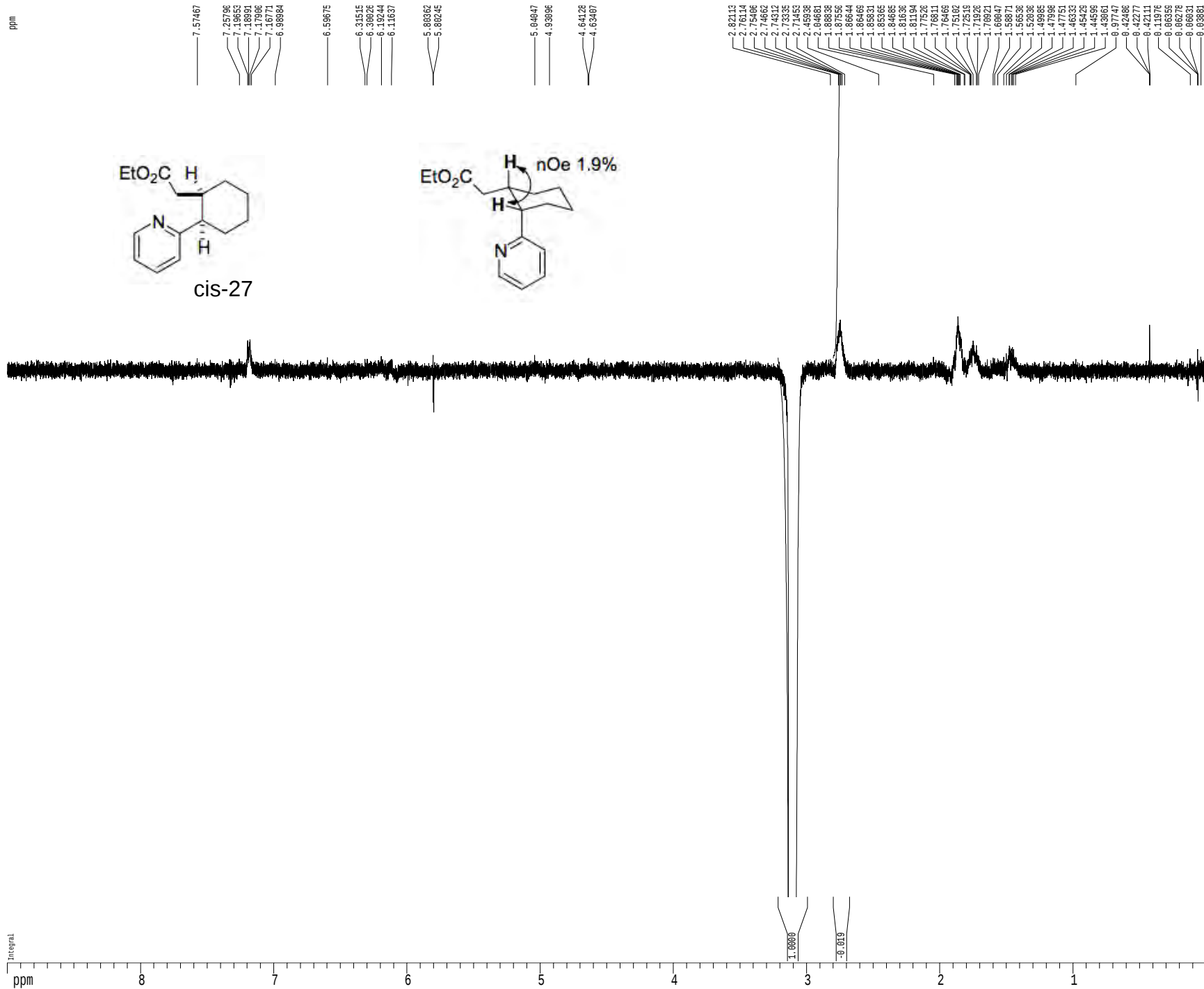
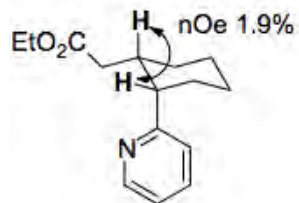
1D NMR plot parameters
 CX 22.00 cm
 CY 15.65 cm
 F1P 190.000 ppm
 F1 22640.47 Hz
 F2P 0.000 ppm
 F2 0.00 Hz
 PPMCM 7.89474 ppm/cm
 HZCM 993.00305 Hz/cm

gnoe

ppm



cis-27



Current Data Parameters
 USER jhanna
 NAME LEH-7-88-c2-f2-NOE
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20161217
 Time 13.48
 INSTRUM cryo500
 PROBHD 5 mm CPTCI 1H-
 PULPROG gnoe1cc.wu
 TD 65536
 SOLVENT CDCl3
 NS 256
 DS 8
 SWH 8012.820 Hz
 FIDRES 0.122266 Hz
 AQ 4.0894966 sec
 RG 64
 DW 62.400 usec
 DE 6.00 usec
 TE 298.0 K
 D1 1.00000000 sec
 D8 0.50000000 sec
 D16 0.00020000 sec
 d21 0.33376500 sec
 d22 0.16399699 sec
 p2 15.00 usec

===== CHANNEL f1 =====
 NUC1 1H
 P1 7.50 usec
 P3 22.50 usec
 P4 30.00 usec
 P5 20.00 usec
 P12 40000.00 usec
 PL1 1.60 dB
 SF01 500.2215568 MHz
 SP1 61.60 dB
 SPNAM1 gauss1.512
 SPOFF1 0.00 Hz

===== GRADIENT CHANNEL =====
 GPNAM1 sine.100
 GPNAM2 sine.100
 GPNAM3 sine.100
 GPNAM4 sine.100
 GPX1 0.00 %
 GPX2 0.00 %
 GPX3 0.00 %
 GPX4 0.00 %
 GPY1 0.00 %
 GPY2 0.00 %
 GPY3 0.00 %
 GPY4 0.00 %
 GRZ1 7.00 %
 GRZ2 3.00 %
 GRZ3 2.30 %
 GRZ4 2.30 %
 P16 1000.00 usec

F2 - Processing parameters
 SI 65536
 SF 500.2200000 MHz
 WDW no
 SSB 0
 LB 0.00 Hz
 GB 0
 PC 1.00

1D NMR plot parameters
 CX 22.80 cm
 CY 50.00 cm
 F1P 9.000 ppm
 F1 4501.98 Hz
 F2P 0.000 ppm
 F2 0.00 Hz
 PPMCM 0.39474 ppm/cm
 HZCM 197.45528 Hz/cm