

Supporting materials

Destruxin A induces and binds HSPs in *Bombyx mori* Bm12 cells

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13 Prokaryotic expression of BmHSPs

14 For cloning of BmHSP genes, the total RNA was isolated by Total RNA Kit II (OMEGA, USA) from
 15 the Bm12 cells in stable phase according to the manufacturer's protocol. The purity and concentration of
 16 total RNA were determined by SimpliNano micro spectrophotometer (GE Healthcare Life Science,
 17 Beijing, China). The cDNA was synthesized using the Reverse Transcriptase (Takara, Japan) following the
 18 manufacturer's protocol. The PCR specific primers were designed based on the *Bombyx mori* HSPs gene
 19 sequences (from NCBI data base) with restriction enzyme sites (Table S1). The PCR reaction system of
 20 25μL contained LA PremixTaq (12.5μL), forward primer (1μL), reverse primer (1μL), template cDNA
 21 (1μL), and distilled water (9.5 μL). The amplification conditions consisted of a pre-denaturation at 94°C for
 22 4 min, followed by 30 cycles of 94°C for 30 s, 56~58°C for 30 s and 72°C for 2.5 min, with a final
 23 extension at 72°C for 10 min. The PCR products were examined with 1% (w/v) gel electrophoresis and
 24 analyzed by sequencing. Additionally, the PCR products of BmHSP genes were gel purified with Universal
 25 DNA Purification Kit (Tiangen, Beijing, China) and used for structuring prokaryotic expression system.

26 **Table S1.** Primers of the silkworm HSP genes for PCR amplification

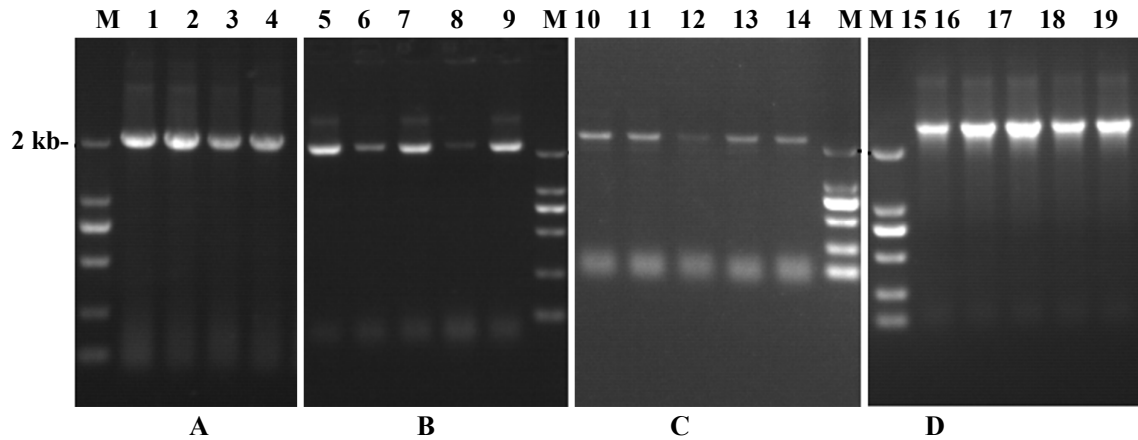
Gene	Gene Accession	Protein Accession	Primer (5'–3')
<i>BmHSP70-3</i> (heat shock protein 70-3)	JF836796.1	Bm_nscaf 2888_391	F: TAGGTACCATGGCAGCGAAGGAAAAGGCA R: CCGCTCGAGTGGTGGTGGTGTGGTGTAAATTC
<i>BmHSP75</i> (heat shock protein 75)	NM_001279432	Bm_nscaf 1898_379	F: TAGGTACCATGTTTGCAGTCCGTCGCCAG R: CCGCTCGAGGTGTTTTCTAAAACCTTGACTAGA
<i>BmHSP83</i> (heat shock protein 83)	NM_001043411.1	Bm_nscaf 2801_21	F: CGATATCATGCCGGAAGAAATGGAG R: CCTCGAGATCAACTTCCTCCATGCG
<i>BmHSCP</i> (heat shock cognate protein 70)	NM_001043427.1	Bm_nscaf 2511_180	F: GTCTAGAATGGCTGCTATGTCCGTCAT R: CGAATTCAATTCAACATCCATTGCTCTTGAG

27 To construct the prokaryotic expression system, the Cloning Vector or PET-30a plasmids (BioVector
 28 NTCC, Beijing, China) and *Escherichia coli* BL21 strain were used. Firstly, target gene fragments were
 29 cloned into the linear purpose vehicle PET-30a plasmids by T4 DNA ligase (TaKaRa, Takara Bio Inc.,
 30 Shiga 525-0058, Japan) linking for static overnight at 16°C. Subsequently, the recombinant plasmids were
 31 transformed into competent host *Escherichia coli* BL21 cells, then, the positive clones were picked up and
 32 validated by sequencing.

34 The results indicated that the cells of *E. coli* BL21 were transformed successfully by recombinant
 35 plasmids. The electrophoretic profiles of PCR products showed the positive clones which contain the target
 36 BmHSPs genes (Figure S1), meanwhile the genes were confirmed by sequencing. Evidently, prokaryotic

37 expression system of BmHSPs were successfully constructed.

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39 **Figure S1** Electrophoresis profiles of recombinant clone by PCR identification. **A**, BmHSP70-3; **B**,
40 BmHSP75; **C**, BmHSP83; **D**, BmHSCP; **M**, Marker; **1-19**, positive clones. The fragment sizes of
41 BmHSP70-3, BmHSP75, BmHSP83 and BmHSCP were respectively about 1.95kb, 2.08kb, 2.15kb
42 and 2.5kb.

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44 **Preparation of BmHSPs**

45 Firstly, the recombinant *E.coli* strains were cultured and induced with IPTG in LB liquid medium
46 (kanamycin resistance) at 37°C and 200 rpm. Then, the broths were centrifuged at 4100×g and 4°C for 10
47 min to respectively collect pellets and supernatants. The following steps were extracted total proteins from
48 the pellets through ultrasonication on ice with cold phosphate buffered saline (PBS) and from the
49 supernatants by centrifugation at 16500×g and 4°C. The total protein samples were dialyzed in the
50 phosphate buffered saline at 4°C and passed through a 0.22 μm micro-pore membrane filter for removing
51 impurities. Then, the samples were used to purify the target BmHSPs by the Nickel affinity
52 chromatography employing BioLogic Duo-Flow system (Bio-Rad, Hercules, CA, USA). The target HSPs
53 were validated by stability test (freeze thawing test), quantification (Bradford) and western blotting¹. The
54 test results showed the four HSPs were all eligible (Figure S2).

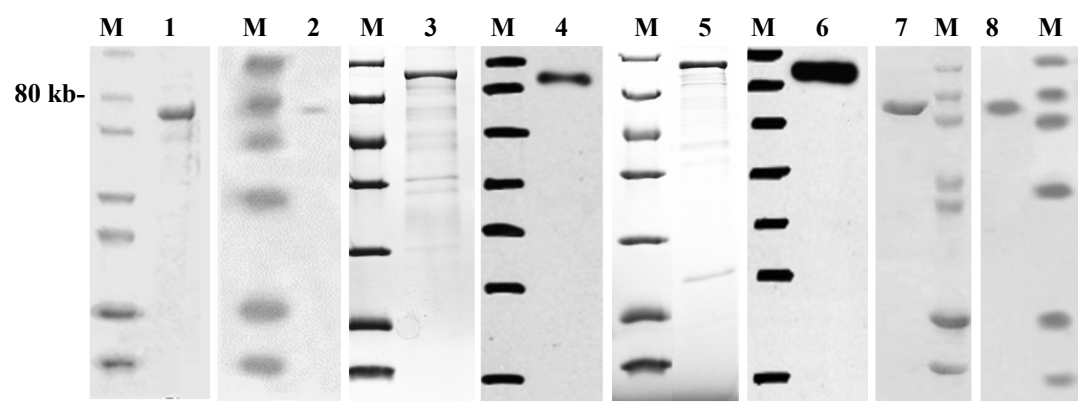
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62 **Figure S2.** Profiles of gel electrophoresis and western blotting (WB) for target proteins. **1/2**, Gel / WB of
63 BmHSP70-3; **3/4**, Gel / WB of BmHSP75; **5/6**, Gel / WB of BmHSP90; **7/8**, Gel / WB of BmHSCP; **M**,
64 protein marker
65

66 **References**

67 1. Guo, Y., Protein electrophoresis experiment technology - Second Edition. China Science Publishing and
68 Media Ltd., Beijing, **2005**.
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