Supporting Information for:

A Probe for the Detection of Hypoxic Cancer Cells

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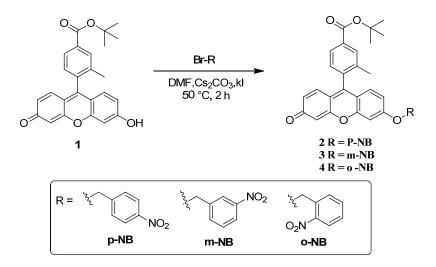
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Synthesis route for intermediates of FBN-1-3



Scheme S1. Synthesis route of compound 2, 3 and 4.

Compound 1 1.0 g (2.5 mmol, 1 eq), Cs_2CO_3 1.7 g (5.0 mmol, 2 eq), and KI 0.4 g (2.5 mmol, 1 eq) in N, N-Dimethylformamide (20 mL) was heated to 50 °C, and then 8 mL of a DMF solution containing nitrobenzyl bromide (5.0 mmol, 2 eq) was added dropwise with stirring. The mixture was stirred for 2 h at 50 °C under an Ar atmosphere. The reaction was monitored with thin-layer chromatography. After the reaction was complete, the solvent was poured into dichloromethane (DCM; 50 mL) and the organic solution was washed with H₂O (80 mL × 5), dried over anhydrous Na₂SO₄, and filtered. The filtrate was added to silica gel and evaporated to dryness. The residue was purified on a silica gel column (DCM/MeOH = $450/1 \rightarrow 150/1$) to generate the target compound.

Compound 2: 1.1 g (2.0 mmol, 81.8%, orange solid). ¹H NMR (400 MHz, CDCl₃) δ 8.29 (d, J = 8.7 Hz, 2H), 8.03 (s, 1H), 8.00 (d, J = 7.9 Hz, 1H), 7.64 (d, J = 8.7 Hz, 2H), 7.25 (d, J = 7.9 Hz, 1H), 7.03 (d, J = 2.4 Hz, 1H), 6.94 (d, J = 8.9 Hz, 1H), 6.88 (d, J = 9.7 Hz, 1H), 6.84 (dd, J = 8.9, 2.4 Hz, 1H), 6.56 (dd, J = 9.7, 1.9 Hz, 1H), 6.45 (d, J = 1.8 Hz, 1H), 5.30 (s, 2H), 2.13 (s, 3H), 1.65 (s, 9H); ¹³C NMR (101 MHz, CDCl₃) δ 185.8, 165.1, 162.6, 158.6, 154.4, 148.0, 147.9, 142.8, 136.6, 136.5, 133.3, 131.5, 130.5, 130.1, 129.3, 129.2, 127.7, 127.2, 124.1, 118.5, 114.5, 113.8, 106.1, 101.6, 81.7, 69.3, 28.2, 19.6; FT-IR: v [cm⁻¹] 3425, 3045, 2978, 2922, 1712, 1643, 1600, 1513, 1442, 1347, 1300, 1260, 1205, 1165, 1100, 1030, 905, 850, 764, 725, 600, 495; MS-ESI: m/z calculated for C₃₂H₂₇NO₇ [M+H]⁺ 538.1866, found 538.1863; melting point: 223.1–224.5 °C.

Compound 3: 1.0 g (1.8 mmol, 74.3%, orange solid). ¹H NMR (400 MHz, CDCl₃) 8.34 (s, 1H), 8.24 (d, J = 7.8 Hz, 1H), 8.03 (s, 1H), 8.00 (d, J = 7.9 Hz, 1H), 7.79 (d, J = 7.7 Hz, 1H), 7.63 (t, J = 7.77.9 Hz, 1H), 7.25 (d, J = 7.9 Hz, 1H), 7.04 (d, J = 2.4 Hz, 1H), 6.95 (d, J = 8.9 Hz, 1H), 6.90 - 6.83 (m, 2H), 6.56 (dd, J = 9.7, 1.9 Hz, 1H), 6.46 (d, J = 1.9 Hz, 1H), 5.28 (s, 2H), 2.14 (s, 3H), 1.65 (s, 2H), 2.14 (s, 3H), 1.65 (s, 2H), 2.14 (s, 2H), 2.19H); ¹³C NMR (CDCl₃) δ: 185.8, 165.1, 162.7, 158.6, 154.4, 148.6, 148.3, 147.9, 137.7, 136.6, 136.6, 133.3, 133.2, 131.5, 130.6, 130.2, 129.9, 129.3 , 129.2, 127.2, 123.5, 122.2, 118.5, 114.5, 113.9, 106.1, 101.6, 81.7, 69.3, 28.2; FT-IR: v [cm⁻¹] 3435, 2975, 2925, 2850, 1710, 1640, 1595, 1530, 1445, 1345, 1290, 1255, 1205, 1160, 1110, 1025, 905, 850, 765, 720, 670, 595, 495; MS-ESI: m/z calculated for $C_{32}H_{27}NO_7 [M+H]^+$ 538.1866, found 538.1871; melting point: 155.4–157.4 °C. Compound 4: 0.8 mg (1.5 mmol, 60.0%, orange solid). The light was avoided during preparation. ¹H NMR (400 MHz, CDCl₃) δ 8.23 (d, *J* = 8.2 Hz, 1H), 8.03 (s, 1H), 8.00 (d, *J* = 8.0 Hz, 1H), 7.84 (d, J = 7.8 Hz, 1H), 7.73 (t, J = 7.5 Hz, 1H), 7.56 (t, J = 7.7 Hz, 1H), 7.24 (s, 1H), 7.06 (d, J = 2.1 Hz), 7.06 (d,Hz, 1H), 6.95 (d, J = 8.9 Hz, 1H), 6.92 – 6.84 (m, 2H), 6.57 (dd, J = 9.7, 1.4 Hz, 1H), 6.46 (d, J = 1.4 Hz, 1H), 5.62 (s, 2H), 2.14 (s, 3H), 1.65 (s, 9H); ¹³C NMR (101 MHz, CDCl₃) δ 185.8, 165.1, 162.7, 158.6, 154.4, 147.9, 147.0, 136.6, 134.2, 133.3, 132.1, 131.5, 130.6, 130.1, 129.3, 129.2, 129.0, 128.5, 127.2, 125.3, 118.5, 114.5, 113.7, 106.2, 101.8, 81.7, 67.6, 29.7, 28.2, 19.6; FT-IR: v [cm⁻¹] 3435, 2970, 2925, 2855, 1715, 1640, 1600, 1525, 1455, 1375, 1335, 1290, 1265, 1205, 1165, 1115, 1030, 910, 855, 730, 595, 500; MS-ESI: m/z calculated for C₃₂H₂₇NO₇ [M+H]⁺ 538.1866, found 538.1866; melting point: 188.1-190.9 °C.

Cell Culture.

HepG-2, A549 and SKOV-3 cells (Bioleaf Biotech Co., Ltd.) were cultured in McCoy's 5A (Gibco), DMEM (Gibco) and RPMI-1640 (Gibco), respectively, at 37 °C under humidified conditions of 95% air and 5% CO₂. All media were supplemented with 100 U penicillin, 10% fetal bovine serum and 0.1 mg of streptomycin (Gibco) per milliliter. The culture media were changed every 2 days to maintain exponential growth of the cells. Cells were passaged using 0.05% Trypsin/EDTA (Sigma) when they reached 80-90% confluence and seeded for experiments. For hypoxia condition experiments, the cells were incubated with FBN-1 (5 μ M) for 30 min at 37 °C, and kept under normoxic (20% O₂) and hypoxic (15%, 8%, and 0.1% O₂) conditions for another 8 h, respectively.

Cytotoxicity Assay.

The cytotoxicity of FBN-1 to HepG-2, A549 and SKOV-3 cells was measured using standard MTT assays. Cells growing in log phase were seeded into 96-well cell-culture plates at 1×10^5 cells/well. The cells were incubated 24 h at 37 °C under humidified conditions of 95% air and 5% CO₂. Then, the three kinds of cells were treated with 1, 2, 4, 8, 16, 32 and 64 µM of FBN-1. Next, one set of plates was kept under dark conditions and MTT solution was added (10 µL, 5 mg/mL in PBS) for another 4 h incubation at 37 °C. The other set of plates were exposed to blue light (LED-light source with 490 nm filter, 0.25 W cm⁻²) for 30 min and subsequently incubated for an additional 30 min. After 1 h, the plates were taken out of the incubator, and the old medium was removed. Next, fresh medium was added, and the cells were allowed to recover for 4 h. Next, MTT solution (10 µL, 5 mg/mL in PBS) was added to the media for another 4 h incubation at 37 °C.

Finally, cell culture media was removed from all plates prior to formazan extraction with 100 μ L DMSO, which was subsequently analysed colorimetrically using a Multi-mode Plate Reader (BioTek, USA) at 570 nm (absorbance value). The following formula was used to calculate the viability of cell growth: Viability (%) = (mean absorbance value of the treatment group-blank/mean absorbance value of the control-blank) × 100.

Fluorescence Quantum Yield Determination.

Fluorescence quantum yields for the FBN-1 and FD were measured by a relative method using Fluorescein ($\Phi_F = 79\%$ in 0.1M NaOH, emission range: 500-600 nm) as a standard. The following equation was used to determine the relative fluorescence quantum yield:

$$\Phi_{\rm F(X)} = \Phi_{\rm F(S)} \times (I_{\rm sample} / I_{\rm standard}) \times (A_{\rm standard} / A_{\rm sample}) \times (n_{\rm sample} / n_{\rm standard})^2$$

Where Φ_F is the fluorescence quantum yield, A is the absorbance at the excitation wavelength (in the range of 0.02-0.05), *I* is the area under the emission curve, *n* is the refractive index of the solvents used in measurements.

HPLC analysis

Agilent SD-1 liquid chromatography (Agilent Technologies, Palo Alto, CA, USA) was performed with a RPC18 HPLC column (4.6×250 mm, 10 μm; Agilent Technologies) and UV detector. The

mobile phase was a gradient of 0-100% of acetonitrile aqueous solution containing 0.1% TFA at a total flow rate of 1 mL/min. The UV absorption wavelength at 254 nm was set for analysis.

Detection limit

The detection limit was calculated based on the fluorescence titration. The fluorescence enhancement of FBN-1 was dose-dependent with respect to NTR. The linear response (y = 1920.1 x + 59.4 with R = 0.992) of fluorescent intensity (y) with respect to the concentration of NTR (x) was established. The lower detection limit (LDL) was calculated following equation. LDL = 3S/m (S is the ratio signal and noise, which is the standard deviation of blank measurements, n = 15; m is the slope of linear equation).

Murine Tumor Model.

All animals were handled in accordance with our laboratory animal handling protocol, and conformed to the Guide for the Care and Use of Laboratory Animals. The HepG-2 cells were washed with PBS (pH 7.4) and harvested with 0.05% trypsin/EDTA (Sigma). After centrifugation, the harvested cells were suspended in PBS (pH 7.4). To create the tumor model, 6-week-old (approximately 20 g) female BALB/c nude mice were implanted subcutaneously on the right flank with 10⁷ HepG-2 cells in 0.2 mL of PBS (pH 7.4). The tumors were allowed to develop for 7 or 35 days before the imaging and ELISA analyses.

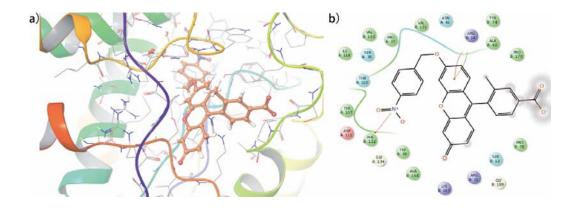


Figure S1. Calculated binding model of FBN-1. a) The substrate binding pocket of NTR (PDB ID: 4DN2) was generated using Schrödinger suites, the C, N, and O atoms of FBN-1 structure are shown in pink, blue, and red, respectively. b) The interactions between the best docking pose of FBN-1 and the active site amino acids of NTR active site in the best docked pose.

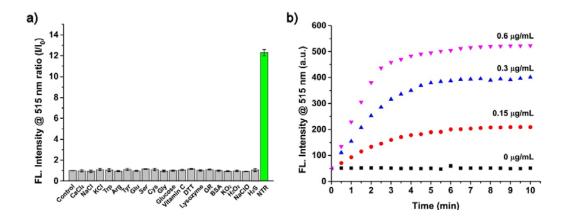


Figure S2. a) Fluorescence responses of FBN-1 (5 μ M) to various species: control (FBN-1 + NADH) and with CaCl₂ (2.5 mM), NaCl (2.5 mM), KCl (50 mM), Trp (1 mM), Arg (1 mM), Tyr (1 mM), Glu (1 mM), Ser (1 mM), Cys (1 mM), Gly (1 mM), glucose (10 mM), vitamin C (1 mM), DTT (1 mM), lysozyme (0.5 mg/mL), GR (10 μ g/mL), BSA (100 mg/mL), KO₂ (1 mM), H₂O₂ (1 mM), NaClO (1 mM), H₂S (1 mM) and NTR (0.6 μ g/mL). b) Time-dependent fluorescence emission intensity of FBN-1 (5 μ M) after reaction with different concentrations of NTR (0, 0.15, 0.3 or 0.6 μ g/mL). $\lambda_{ex} = 490$ nm.

As shown in **Figure S2**, the fluorescence intensity increases linearly for the first ~2 minutes of incubation of 5 μ M FBN-1 with 0.6 μ g/mL NTR, and subsides thereafter. Discrepancies in the final fluorescience intensity are attributed to different concentrations of DMSO used in each run, and associated sensitivities of NTR and NADH to such buffer conditions¹⁻³.

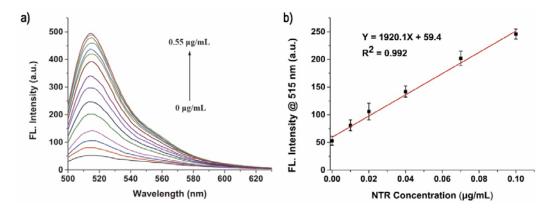


Figure S3. a) Fluorescence spectrum of FBN-1 (5 μ M) response to NTR at varied concentrations (0, 0.01, 0.02, 0.04, 0.07, 0.1, 0.15, 0.2, 0.25, 0.3, 0.35, 0.4, 0.45, 0.5, 0.55 μ g/mL). b) A linear correlation between emission intensity and concentrations of NTR. λ_{ex} = 490 nm.

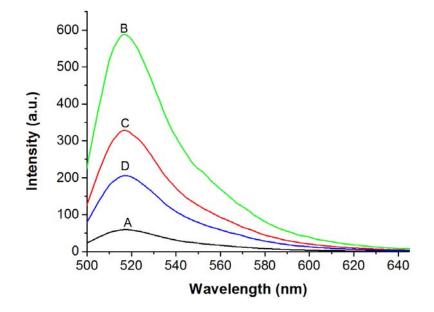


Figure S4. The fluorescence emission spectra of FBN-1 (5 μ M) in the different reaction systems. (A): treated with 500 μ M NADH and without NTR; (B): treated with 1 μ g mL⁻¹ NTR and 500 μ M NADH; (C): B system treated with 0.1 mM dicoumarin; (D): B system treated with 0.2 mM dicoumarin. $\lambda_{ex} = 490$ nm.

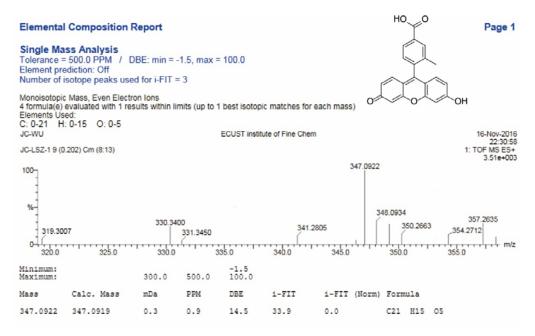


Figure S5. Mass spectrum of the reaction solution of FBN-1 (200 µM) with NTR (20 µg/mL).

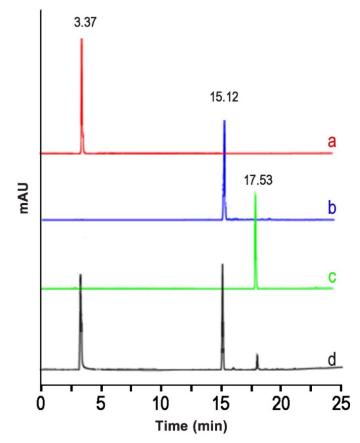


Figure S6. HPLC profiles of (a) 2 mM NADH, (b) 200 μ m fluorescein analogues, (c) 200 μ M FBN-1, (d) 200 μ M FBN-1 mixed with 20 μ g/mL NTR in the presence of 2 mM NADH for 20 min.

| chemical structure | optimized | molecular orbitals | |
|--------------------|---|--|---------------------------|
| | structure | Lumo | Homo |
| | | من خرب بالمن المن المن المن المن المن المن المن | ب بوغو موغو به ه |
| | ۲ ۲ ۲ ۲ ۲ ۲ ۲ ۲ ۲ ۲ ۲ ۲ ۲ ۲ ۲ ۲ ۲ ۲ ۲ | | |

Figure S7. The chemical structure, DFT optimized structure and Molecular orbitals (LUMO and HOMO) of FBN-1 and FD, respectively.

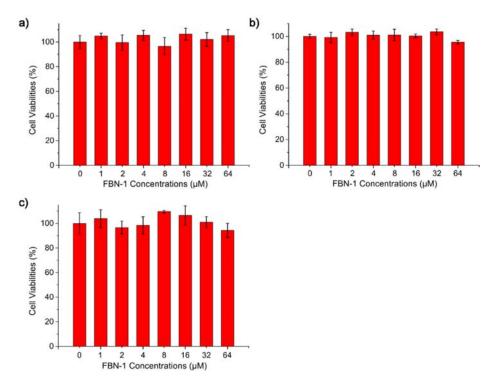


Figure S8. Cell viability (%) estimated by an MTT assay in a) HepG-2 cells, b) A549 cells, and c) SKOV-3 cells. Cells were treated different concentrations of FBN-1 under blue light irradiation (490 nm, 0.25 W cm⁻², 30 min).

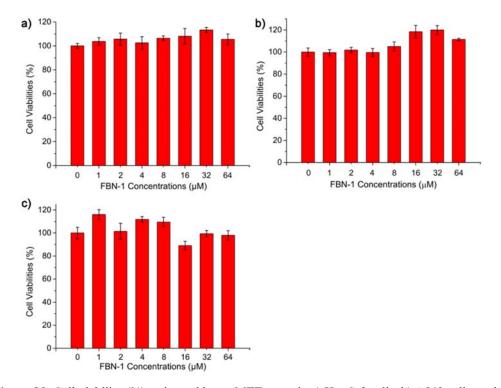


Figure S9. Cell viability (%) estimated by an MTT assay in a) HepG-2 cells, b) A549 cells, and c) SKOV-3 cells. Cells were treated with different concentrations of FBN-1 in the absence of blue light irradiation.

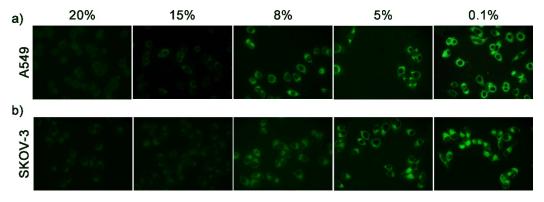


Figure S10. Confocal fluorescence microscopy imaging of a) A549 and b) SKOV-3 cells. A549 and SKOV-3 cells were incubated with FBN-1 (5 μ M) under 20% O₂, 15% O₂, 8% O₂, 5 % O₂ and 0.1 % O₂ conditions for 8 h.

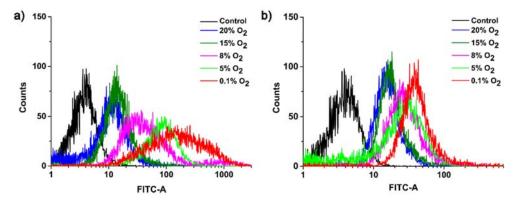


Figure S11. Flow cytometry of a) A549 and b) SKOV-3 cells. A549 and SKOV-3 cells were incubated with FBN-1 (5 μ M) under 20% O₂, 15% O₂, 8% O₂, 5 % O₂ and 0.1 % O₂ conditions for 8 h.

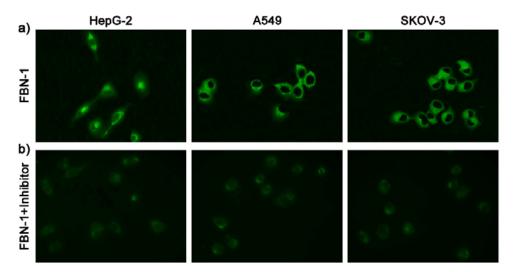


Figure S12. Confocal fluorescence microscopy imaging of HepG-2, A549 and SKOV-3 cells incubated with a) 5 μ M FBN-1 and b) 5 μ M FBN-1 and inhibitor (0.2 mM dicoumarin) under hypoxic condition of 1% O₂ for 8 h.

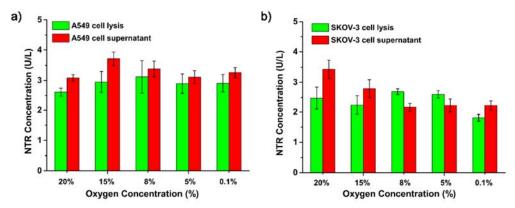


Figure S13. ELISA-based quantification of NTR concentration in a) A549 and b) SKOV-3 cell lysates and supernatant. A549 and SKOV-3 cells were incubated with FBN-1 (5 μ M) under 20% O₂, 15% O₂, 8% O₂, 5 % O₂ and 0.1 % O₂ conditions for 8 h.

Prior findings have claimed that NTR overexpression is likely linked to intensification of hypoxia. However, the reduction process of common NTR probes can be influenced by oxygen and nitroreductase as we show in **Scheme 2**. Our work in direct ELISA quantification of NTR in tumor cell lines, and in excised tumors, both suggest that NTR expression is invariant with degree of hypoxia, an effect which as not probed in prior work⁴⁻⁹.

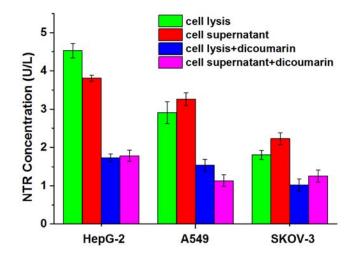


Figure S14. Quantitative detection of NTR levels in cell lysates, cell supernatant, cell lysates with 0.2 mM dicoumarin and cell supernatant with 0.2 mM dicoumarin by ELISA. HepG-2, A549 and SKOV-3 cells were incubated under 0.1 % O_2 conditions for 8 h at 37 °C.

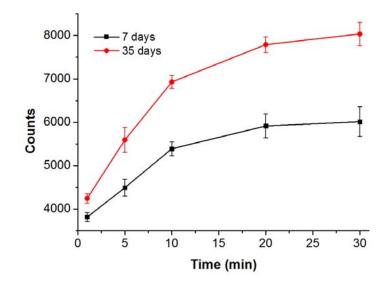
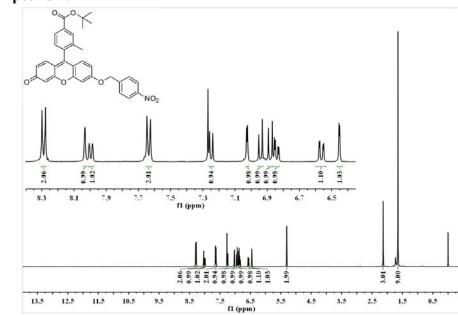


Figure S15. Quantification of the counts of figure 3a.

¹H NMR, ¹³H NMR and Mass Spectra



Compound 2:

Figure S16. ¹H NMR spectrum of compound 2.

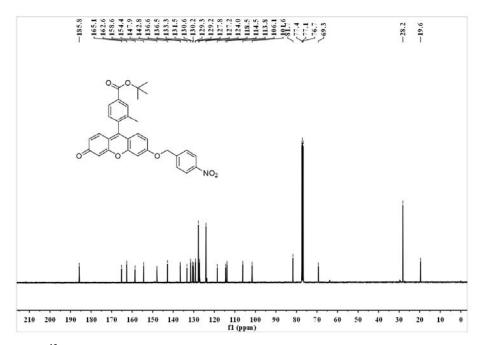


Figure S17. ¹³C NMR spectrum of compound 2.

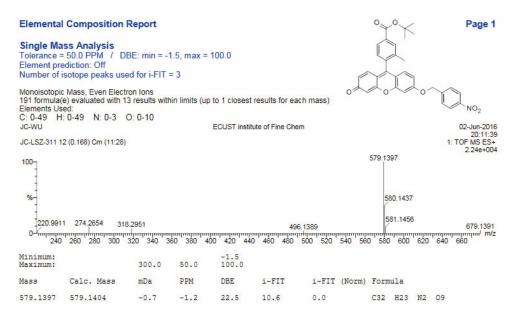


Figure S18. Mass spectrum of compound 2.

Compound 3:

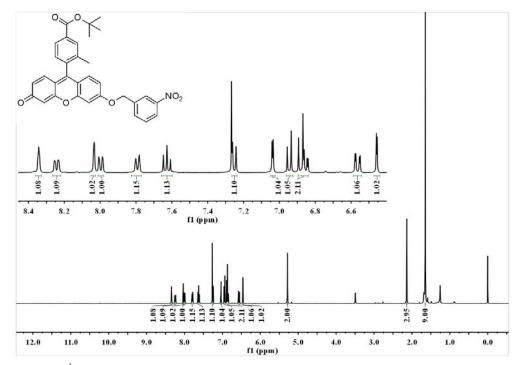


Figure S19. ¹H NMR spectrum of compound 3.

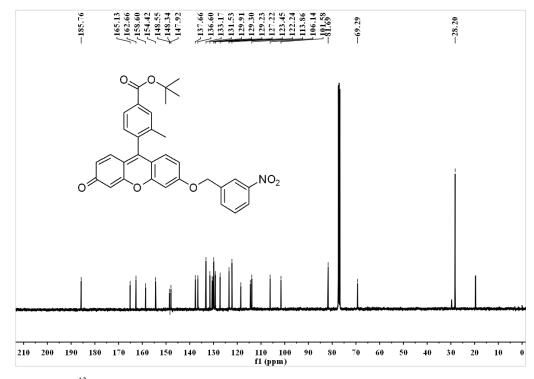
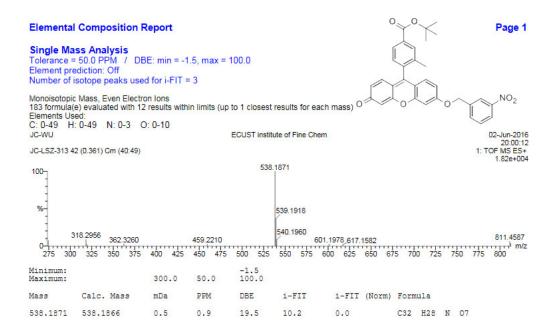
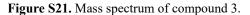


Figure S20. ¹³C NMR spectrum of compound 3.





Compound 4:

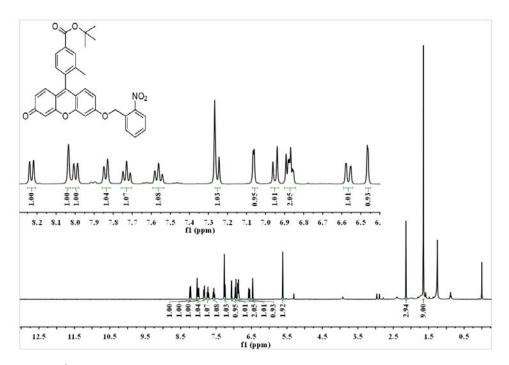


Figure S22. ¹H NMR spectrum of compound 4.

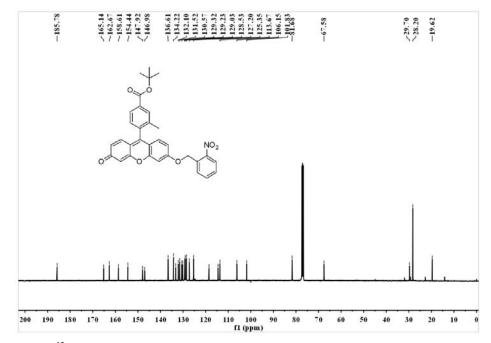


Figure S23. ¹³C NMR spectrum of compound 4.

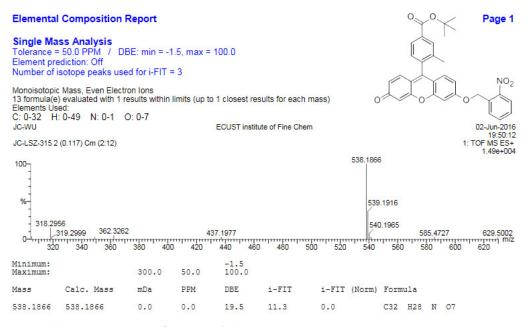


Figure S24. Mass spectrum of compound 4.

FBN-1:

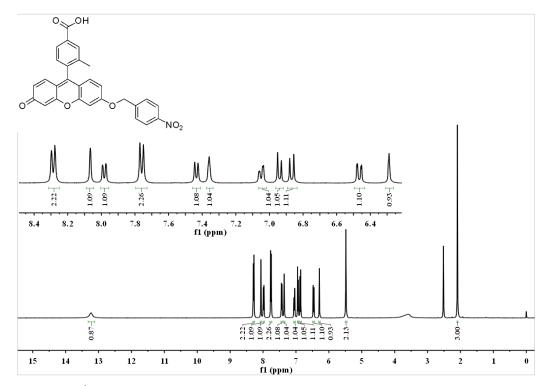


Figure S25. ¹H NMR spectrum of FBN-1.

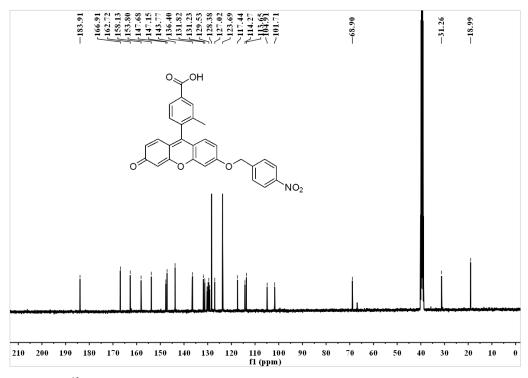


Figure S26. ¹³C NMR spectrum of FBN-1.

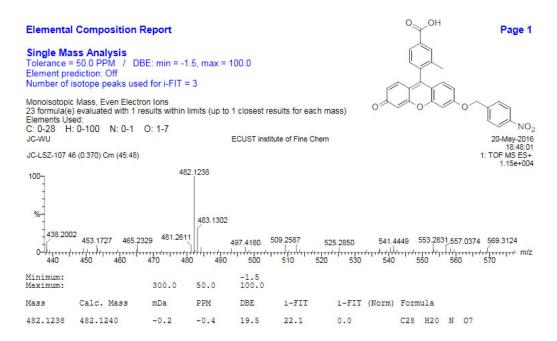


Figure S27. Mass spectrum of the FBN-1.

FBN-2:

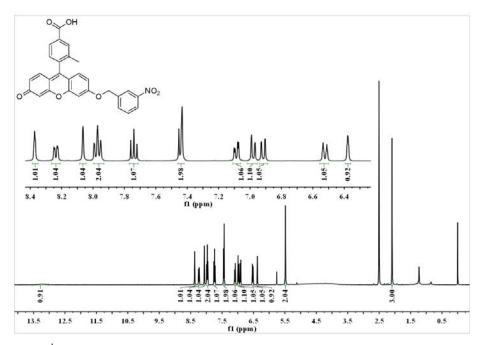


Figure S28. ¹H NMR spectrum of FBN-2.

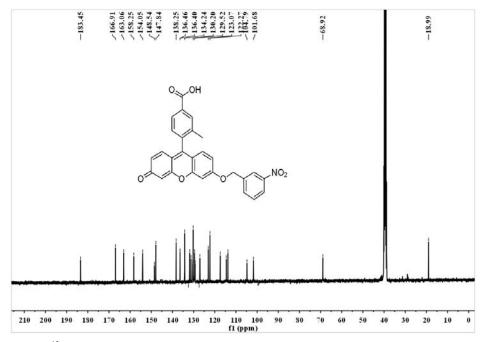


Figure S29. ¹³C NMR spectrum of FBN-2.

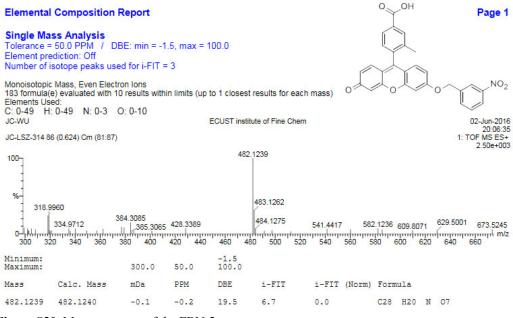


Figure S30. Mass spectrum of the FBN-2.

FBN-3:

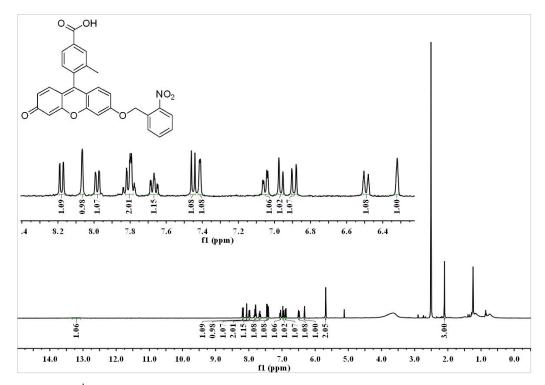


Figure S31. ¹H NMR spectrum of FBN-3.

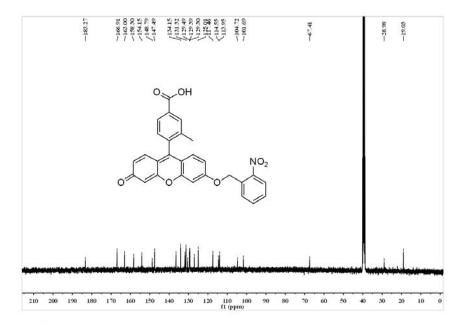


Figure S32. ¹³C NMR spectrum of FBN-3.

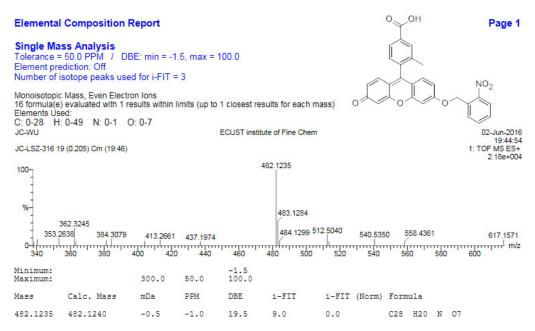


Figure S33. Mass spectrum of the FBN-3.

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