

# Supporting Information

## to

### Sulfurization of Dissolved Organic Matter Increases Hg-S-DOM Bioavailability to a Hg-Methylating Bacterium

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**Table SI-1.** Recovery and UV-VIS spectral characteristics of sulfurized DOM by solid phase extraction (SPE). Slope ratio ( $S_R$ ) is the ratio of the slope of the natural log transformed spectra in the wavelength range 275-295 nm divided by the slope in the range 350-400 nm.  $S_R$  is strongly correlated with the size and aromaticity of DOM as described in Helms et al. (2008).<sup>1</sup> The reported error on  $S_R$  was determined based on the relative standard errors of the linear fits to the natural log transformed spectra in each wavelength range.

Sample	Measured S/C ratio	SPE Recovery (%)	$S_R$ (slope ratio)
SRHA (unsulfurized)	3.42	33.3	0.67±0.02
SRHA	4.12	34.6	0.67±0.02
SRHA	4.73	34.8	0.68±0.02
SRHA	6.12	32.8	0.66±0.02
SRHA	5.83	31.6	0.70±0.02
IHSS SRHA (no SPE)	not measured	not applicable	0.65±0.02
SRFA (unsulfurized)	1.88	67.3	0.69±0.02
SRFA	3.80	54.2	0.72±0.02
SRFA	4.22	53.5	0.69±0.02
SRFA	4.08	55.8	0.63±0.02
SRFA	5.69	52.5	0.70±0.02
IHSS SRFA (no SPE)	not measured	not applicable	0.82±0.05
NLFA (unsulfurized)	3.12	46.2	0.78±0.06
NLFA	3.98	46.6	0.76±0.06
NLFA	8.83	29.4	0.74±0.09
NLFA	12.8	44.9	0.84±0.05
IHSS NLFA (no SPE)	not measured	not applicable	0.65±0.04
PLFA (unsulfurized)	11.0	78.6	0.99±0.08
PLFA	10.4	75.6	0.79±0.06
PLFA	14.5	75.0	0.76±0.05
PLFA	12.5	79.9	0.85±0.06
PLFA	12.2	81.3	0.96±0.09
IHSS PLFA (no SPE)	not measured	not applicable	0.91±0.06

**Table SI-2.** Quality control data for total Hg (THg) and methylmercury (MeHg) analyses. Instrument detection limit determined as three times standard deviation of blank.

<i>Parameter</i>	<i>Result</i>
Me <sup>201</sup> Hg instrument detection limit	0.11±0.18 pg (0.02 ng/L for 5 mL sample)
Distillation blanks for Me <sup>201</sup> Hg	0.02±0.02 ng/L
Relative percent difference for duplicate MeHg analyses	7.4±6.2% ( <i>n</i> = 5 pairs)
MeHg recovery for NIST 1566b (oyster tissue)	139±8% ( <i>n</i> = 6 determinations)
<sup>201</sup> THg instrument detection limit	0.37±0.38 ng/L
Digestion blanks for <sup>201</sup> Hg	0.02±0.04 ng/L
Relative percent difference for duplicate THg analyses	5.4±2.7% ( <i>n</i> = 4 pairs)
THg recovery for NIST 2709a (San Joaquin soil)	91.6±22.0% ( <i>n</i> = 8 determinations)

Instrument detection limit calculated as three times the standard deviation of reagent blanks.

### Description of Equilibrium Speciation Modeling

Equilibrium speciation modeling was performed in MINEQL+ v. 4.6 (Environmental Research Software). Equilibrium constants were critically selected using the most up to date information on  $\text{Hg(II)}_i$  complexation in natural waters. The solubility product for metacinnabar ( $\text{HgS(s)}$ ) was recently reevaluated by Drott *et al.*<sup>2</sup> and reported as  $\log K = 36.8$ , 1.2 log units lower than that reported in the NIST Critical Database.<sup>3</sup> Following Skyllberg,<sup>4</sup> we have assumed that  $\text{Hg(II)}_i$  forms linear two-coordinate complexes with DOM thiols with a  $\log K = 42.0$ . In this approach, we ignore the contribution of weaker O- and N- donor ligands in the DOM pool. This approach is justified for two reasons: 1) DOM/Hg ratios are sufficiently high in these experiments, such that binding will be dominated by stronger S-donor ligands<sup>5</sup>; 2) All solutions contain  $\mu\text{M}$  concentrations of sulfide further diminishing the contributions of weak  $\text{Hg(II)}_i$ -binding ligands.  $[\text{RSH}]_T$  was estimated based upon  $[\text{DOC}]$ , the measured S/C ratio, and the assumption that strong  $\text{Hg(II)}$ -binding thiols could be estimated based on the concentration of exocyclic sulfur in each DOM sample. Manceau and Nagy<sup>6</sup> determined S speciation using X-ray absorption near edge spectroscopy (XANES) for 3 out of the 4 isolates used in this study (and S speciation for the humic acid fraction of the Nordic Lake sample). The percentage of total S as reduced exocyclic S ranged from 23.6 to 46.9% (mean =  $32.2 \pm 10.5\%$ ). We further assume that DOM S speciation is independent of total S content – recent data from Hoffmann *et al.*<sup>7</sup> and Poulin *et al.*<sup>8</sup> suggests, however, that the fraction of total S in reduced forms increases with increasing sulfurization. In that case, our application of a single conversion factor for total S to reduced S may underestimate the true contribution of DOM thiols to  $\text{Hg(II)}_i$  binding. Other input parameters for modeling can be found in Table SI-3 below; for sulfide concentration, the mean of initial and final ( $t=3$  h) concentrations were input into the speciation model. In modeling Hg-cysteine complexation, some reports suggest the possibility of a tris  $\text{Hg(cys)}$  complex (likely  $\text{Hg(Hcys)}_3^-$ ).<sup>9</sup> Unfortunately, no thermodynamic data are available for this purported complex. Kőszegi-Szalai and Paál<sup>10</sup> reported equilibrium constants for Hg-penicillamine complexes, including a  $\text{Hg(Hpen)}_3^-$  complex with a  $\log K$  of 75.3 at  $I = 0$  M. Given their similar structures (differing only in the two  $\text{CH}_3$ —substituents at the 3-position for penicillamine), we can evaluate the potential contributions of a  $\text{Hg(Hcys)}_3^-$  complex to  $\text{Hg(II)}_i$  speciation using the  $\log K$  for the  $\text{Hg(Hpen)}_3^-$  complex. Using this approach, we find that  $\text{Hg(Hcys)}_3^-$  is not likely to be a significant species under our experimental conditions ( $[\text{}^{201}\text{Tg}]$ ,  $[\text{H}_2\text{S}]_T$ ,  $[\text{cys}]_T$ , and pH), and we do not include this species in our modeling. A summary of important thermodynamic data for speciation modeling can be found in Table SI-3 below.

**Table SI-3.** Thermodynamic data for equilibrium speciation modeling. Equilibrium constants for Hg-Cl and Hg-OH complexes were taken directly from the MINEQL+ database.

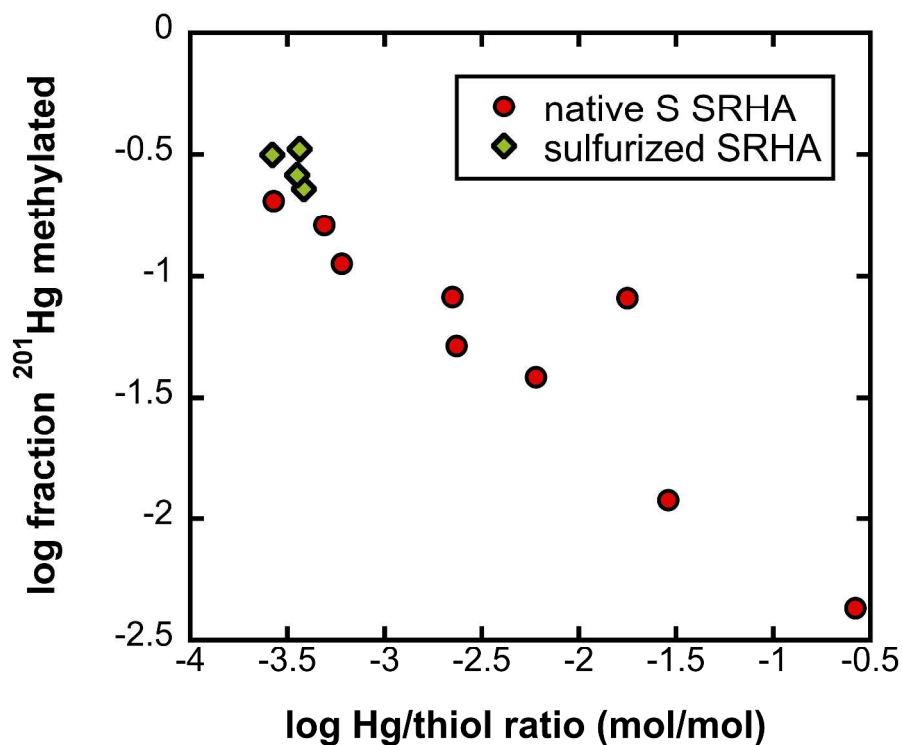
<i>Reaction</i>	<i>log K</i>	<i>Reference</i>
<i>Hg-sulfide Aqueous Speciation</i>		
$\text{Hg}^{2+} + 2\text{HS}^- = \text{Hg}(\text{SH})_2^0$	39.1	Drott <i>et al.</i> <sup>2</sup>
$\text{Hg}^{2+} + 2\text{HS}^- = \text{HgS}_2\text{H}^- + \text{H}^+$	32.5	Drott <i>et al.</i> <sup>2</sup>
$\text{Hg}^{2+} + 2\text{HS}^- = \text{HgS}_2^{2-} + 2\text{H}^+$	23.2	Drott <i>et al.</i> <sup>2</sup>
<i>Metacinnabar Precipitation</i>		
$\text{Hg}^{2+} + \text{HS}^- = \text{HgS}(\text{s}) + \text{H}^+$	36.8	Drott <i>et al.</i> <sup>2</sup>
<i>Hg-DOM Complexation</i>		
$\text{Hg}^{2+} + 2\text{RS}^- = \text{Hg}(\text{SR})_2$	42.0	Skylberg <sup>4</sup>
$\text{RS}^- + \text{H}^+ = \text{RSH}$	10.0	Skylberg <sup>4</sup>
<i>Hg-CYS Complexation</i>		
$\text{Hg}^{2+} + 2\text{H}^+ + 2\text{CYS}^{2-} = \text{Hg}(\text{HCYS})_2^0$	64.1	Starý and Kratzer. <sup>11</sup>
$\text{Hg}^{2+} + 2\text{CYS}^{2-} = \text{Hg}(\text{CYS})_2^{2-}$	43.9	Starý and Kratzer. <sup>11</sup>

**Table SI-4.** Summary of experimental variables in Hg methylation assays with *Desulfovibrio desulfuricans* ND132 in the presences of sulfurized DOM samples. DOM isolates were sulfurized as described in the main text, resulting in the S/C ratios reported in the table below. Reported values are means and standard deviations ( $n = 3$ , excepting NLFA experiments, where  $n = 2$ ). n.d. = not determined due to lost samples. Cell density is average cell density measured at beginning and end of 3h incubation which typically increased less than 5% over the duration of the experiment.

DOM Isolate or Control	[DOC] (mg/L)	Measured S/C ratio (mmol S/mol C)	Cell density ( $\times 10^8$ cells/mL)	pH	Initial sulfide ( $\mu$ M)	Final sulfide ( $\mu$ M)	Total $^{201}\text{Hg}$ in medium (nM)	Total filterable $^{201}\text{Hg}$ (nM)	Total Me $^{201}\text{Hg}$ in medium (pM)
SRHA	9.19	3.42	5.68 $\pm$ 0.21	7.27 $\pm$ 0.01	2.25 $\pm$ 0.05	3.62 $\pm$ 0.12	0.30 $\pm$ 0.16	0.21 $\pm$ 0.01	39.6 $\pm$ 4.9
SRHA	9.52	4.12	5.57 $\pm$ 0.22	7.31 $\pm$ 0.01	2.67 $\pm$ 0.26	3.92 $\pm$ 0.02	0.30 $\pm$ 0.05	0.29 $\pm$ 0.04	66.4 $\pm$ 1.3
SRHA	9.58	4.73	5.32 $\pm$ 0.31	7.25 $\pm$ 0.01	3.36 $\pm$ 0.08	4.15 $\pm$ 0.11	0.32 $\pm$ 0.06	0.26 $\pm$ 0.04	80.7 $\pm$ 1.5
SRHA	9.59	6.12	5.67 $\pm$ 0.19	7.20 $\pm$ 0.01	4.18 $\pm$ 0.24	4.40 $\pm$ 0.06	0.31 $\pm$ 0.01	0.23 $\pm$ 0.004	95.8 $\pm$ 1.2
SRHA	9.24	5.83	5.43 $\pm$ 0.09	7.33 $\pm$ 0.02	3.72 $\pm$ 0.17	4.53 $\pm$ 0.10	0.39 $\pm$ 0.02	0.24 $\pm$ 0.01	128.4 $\pm$ 1.9
SRFA	10	1.88	4.55 $\pm$ 0.82	7.28 $\pm$ 0.04	2.33 $\pm$ 0.10	3.01 $\pm$ 0.29	0.057 $\pm$ 0.003	0.028 $\pm$ 0.004	24.0 $\pm$ 0.6
SRFA	10	3.80	5.75 $\pm$ 1.51	7.28 $\pm$ 0.04	2.79 $\pm$ 0.29	3.22 $\pm$ 0.20	0.070 $\pm$ 0.008	0.034 $\pm$ 0.007	40.5 $\pm$ 1.6
SRFA	10	4.22	5.15 $\pm$ 0.38	7.29 $\pm$ 0.02	2.88 $\pm$ 0.10	3.36 $\pm$ 0.06	0.083 $\pm$ 0.007	0.054 $\pm$ 0.014	52.5 $\pm$ 2.9
SRFA	10	4.08	5.03 $\pm$ 0.52	7.22 $\pm$ 0.02	2.87 $\pm$ 0.03	3.44 $\pm$ 0.14	0.13 $\pm$ 0.01	0.088 $\pm$ 0.007	86.6 $\pm$ 0.2
SRFA	10	5.69	4.74 $\pm$ 0.26	7.26 $\pm$ 0.02	3.58 $\pm$ 0.25	3.76 $\pm$ 0.20	0.26 $\pm$ 0.001	0.12 $\pm$ 0.02	160 $\pm$ 8
NLFA	10	3.12	1.44 $\pm$ 0.21	7.52 $\pm$ 0.12	0.26 $\pm$ 0.18	0.95 $\pm$ 0.83	0.15 $\pm$ 0.07	0.067 $\pm$ 0.006	36.9 $\pm$ 11.4
NLFA	10	3.98	1.50 $\pm$ 0.15	7.74 $\pm$ 0.05	0.10 $\pm$ 0.04	0.33 $\pm$ 0.06	0.099 $\pm$ 0.003	0.086 $\pm$ 0.006	53.4 $\pm$ 13.6
NLFA	10	8.83	1.36 $\pm$ 0.18	7.61 $\pm$ 0.10	0.05 $\pm$ 0.01	0.42 $\pm$ 0.31	0.10 $\pm$ 0.01	0.090 $\pm$ 0.01	49.5 $\pm$ 13.4
NLFA	10	12.8	1.66 $\pm$ 0.07	7.66 $\pm$ 0.04	0.30 $\pm$ 0.06	0.42 $\pm$ 0.11	0.15 $\pm$ 0.01	0.17 $\pm$ 0.01	138 $\pm$ 9
PLFA	8.3	11.0	4.79 $\pm$ 0.23	7.35 $\pm$ 0.02	3.99 $\pm$ 0.27	6.44 $\pm$ 0.78	0.13 $\pm$ 0.004	0.018 $\pm$ 0.002	39.2 $\pm$ 4.0
PLFA	8.3	10.4	5.11 $\pm$ 0.45	7.35 $\pm$ 0.05	5.13 $\pm$ 0.52	6.87 $\pm$ 0.47	0.13 $\pm$ 0.003	0.036 $\pm$ 0.006	64.2 $\pm$ 6.3
PLFA	8.3	14.5	4.76 $\pm$ 0.33	7.34 $\pm$ 0.05	5.13 $\pm$ 0.52	7.02 $\pm$ 0.28	0.12 $\pm$ 0.01	0.038 $\pm$ 0.002	53.0 $\pm$ 7.0
PLFA	8.3	12.5	5.28 $\pm$ 0.37	7.28 $\pm$ 0.06	4.78 $\pm$ 0.19	6.72 $\pm$ 0.54	0.11 $\pm$ 0.01	0.030 $\pm$ 0.008	62.1 $\pm$ 10.7
PLFA	8.3	12.2	5.80 $\pm$ 0.89	7.32 $\pm$ 0.02	5.13 $\pm$ 0.52	6.56 $\pm$ 0.26	0.11 $\pm$ 0.01	0.049 $\pm$ 0.008	57.8 $\pm$ 0.8
500 $\mu$ M L-cysteine control (SRHA)	N/A	N/A	5.74 $\pm$ 0.13	7.28 $\pm$ 0.00	5.78 $\pm$ 0.11	19.3 $\pm$ 0.1	0.46 $\pm$ 0.02	0.07 $\pm$ 0.01	362 $\pm$ 6
500 $\mu$ M L-cysteine control (SRFA)	N/A	N/A	4.04 $\pm$ 0.67	7.14 $\pm$ 0.00	5.55 $\pm$ 0.72	31.1 $\pm$ 4.9	0.37 $\pm$ 0.02	0.28 $\pm$ 0.02	308 $\pm$ 11
500 $\mu$ M L-cysteine control (NLFA)	N/A	N/A	1.43 $\pm$ 0.17	7.30 $\pm$ 0.06	0.28 $\pm$ 0.10	4.08 $\pm$ 3.1	0.13 $\pm$ 0.02	n.d.	43.4 $\pm$ 13.4
500 $\mu$ M L-cysteine control (PLFA)	N/A	N/A	5.15 $\pm$ 0.45	7.23 $\pm$ 0.04	5.90 $\pm$ 0.88	17.0 $\pm$ 0.7	0.31 $\pm$ 0.07	0.39 $\pm$ 0.01	347 $\pm$ 20
No DOM control (SRHA)	N/A	N/A	5.31 $\pm$ 0.34	7.40 $\pm$ 0.01	3.64 $\pm$ 0.13	2.75 $\pm$ 0.1	0.28 $\pm$ 0.02	0.25 $\pm$ 0.01	2.2 $\pm$ 0.2
No DOM control (SRFA)	N/A	N/A	3.81 $\pm$ 0.10	7.20 $\pm$ 0.12	2.74 $\pm$ 0.25	3.22 $\pm$ 0.53	0.064 $\pm$ 0.007	0.007 $\pm$ 0.005	6.5 $\pm$ 1.2

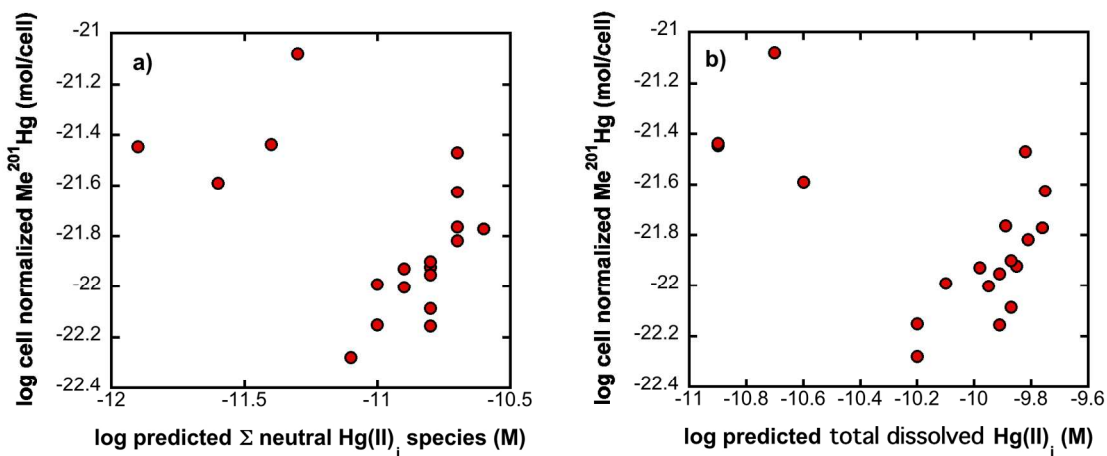
**Table SI-5.** Predicted equilibrium speciation of inorganic Hg(II) based on measured total  $^{201}\text{Hg}$  in medium, pH, sulfide, DOC, and S/C ratio of DOM.  $\text{Hg}(\text{SR})_2$  is a two-coordinate complex of  $\text{Hg}(\text{II})_i$  with organic thiols;  $\text{Hg}(\text{SH})_2$  is the equivalent complex with inorganic sulfide.

DOM Isolate or Control	$[\text{RSH}]_T$ ( $\mu\text{M}$ )	[Meta-cinnabar] (M)	$[\text{Hg}(\text{SR})_2]$ (M)	$[\text{Hg}(\text{SH})_2]$ (M)	$[\text{HgS}_2\text{H}^-] + [\text{HgS}_2^{2-}]$ (M)	Total dissolved Hg (M)
SRHA	0.62	$1.77 \times 10^{-10}$	$7.28 \times 10^{-15}$	$1.65 \times 10^{-11}$	$1.06 \times 10^{-10}$	$1.23 \times 10^{-10}$
SRHA	0.77	$1.60 \times 10^{-10}$	$1.08 \times 10^{-14}$	$1.74 \times 10^{-11}$	$1.23 \times 10^{-10}$	$1.40 \times 10^{-10}$
SRHA	0.89	$1.65 \times 10^{-10}$	$1.15 \times 10^{-14}$	$2.18 \times 10^{-11}$	$1.33 \times 10^{-10}$	$1.55 \times 10^{-10}$
SRHA	1.15	$1.36 \times 10^{-10}$	$1.55 \times 10^{-14}$	$2.69 \times 10^{-11}$	$1.47 \times 10^{-10}$	$1.74 \times 10^{-10}$
SRHA	1.06	$2.14 \times 10^{-10}$	$1.69 \times 10^{-14}$	$2.10 \times 10^{-11}$	$1.55 \times 10^{-10}$	$1.76 \times 10^{-10}$
SRFA	0.40	undersaturated	$1.70 \times 10^{-15}$	$7.50 \times 10^{-12}$	$4.94 \times 10^{-11}$	$5.69 \times 10^{-11}$
SRFA	0.80	undersaturated	$6.44 \times 10^{-15}$	$9.24 \times 10^{-12}$	$6.09 \times 10^{-11}$	$7.01 \times 10^{-11}$
SRFA	0.88	undersaturated	$9.12 \times 10^{-15}$	$1.07 \times 10^{-11}$	$7.18 \times 10^{-11}$	$8.25 \times 10^{-11}$
SRFA	0.85	undersaturated	$1.19 \times 10^{-14}$	$1.90 \times 10^{-11}$	$1.09 \times 10^{-10}$	$1.28 \times 10^{-10}$
SRFA	1.19	$1.02 \times 10^{-10}$	$2.13 \times 10^{-14}$	$2.10 \times 10^{-11}$	$1.32 \times 10^{-10}$	$1.53 \times 10^{-10}$
NLFA	0.84	$1.21 \times 10^{-10}$	$1.00 \times 10^{-13}$	$2.18 \times 10^{-12}$	$2.54 \times 10^{-11}$	$2.77 \times 10^{-11}$
NLFA	1.06	$8.73 \times 10^{-11}$	$6.80 \times 10^{-13}$	$5.20 \times 10^{-13}$	$1.03 \times 10^{-11}$	$1.15 \times 10^{-11}$
NLFA	2.36	$9.30 \times 10^{-11}$	$3.00 \times 10^{-12}$	$5.81 \times 10^{-13}$	$8.42 \times 10^{-12}$	$1.20 \times 10^{-11}$
NLFA	3.42	$1.32 \times 10^{-10}$	$3.77 \times 10^{-12}$	$9.70 \times 10^{-13}$	$1.59 \times 10^{-11}$	$2.06 \times 10^{-11}$
PLFA	3.55	undersaturated	$9.18 \times 10^{-14}$	$1.53 \times 10^{-11}$	$1.19 \times 10^{-10}$	$1.34 \times 10^{-10}$
PLFA	3.39	undersaturated	$6.34 \times 10^{-14}$	$1.53 \times 10^{-11}$	$1.19 \times 10^{-10}$	$1.35 \times 10^{-10}$
PLFA	4.70	undersaturated	$1.08 \times 10^{-13}$	$1.44 \times 10^{-11}$	$1.09 \times 10^{-10}$	$1.24 \times 10^{-10}$
PLFA	5.75	undersaturated	$7.14 \times 10^{-14}$	$1.38 \times 10^{-11}$	$9.11 \times 10^{-11}$	$1.05 \times 10^{-10}$
PLFA	5.85	undersaturated	$7.21 \times 10^{-14}$	$1.35 \times 10^{-11}$	$9.74 \times 10^{-11}$	$1.11 \times 10^{-10}$
500 $\mu\text{M}$ L-cysteine control (SRHA)	500	undersaturated	$4.60 \times 10^{-10}$	$2.43 \times 10^{-19}$	$1.68 \times 10^{-14}$	$4.60 \times 10^{-10}$
500 $\mu\text{M}$ L-cysteine control (SRFA)	500	undersaturated	$1.27 \times 10^{-10}$	$7.12 \times 10^{-17}$	$9.94 \times 10^{-15}$	$1.27 \times 10^{-10}$
500 $\mu\text{M}$ L-cysteine control (NLFA)	500	undersaturated	$3.09 \times 10^{-10}$	$6.09 \times 10^{-17}$	$3.56 \times 10^{-16}$	$3.09 \times 10^{-10}$
500 $\mu\text{M}$ L-cysteine control (PLFA)	500	undersaturated	$3.71 \times 10^{-10}$	$2.59 \times 10^{-15}$	$1.23 \times 10^{-14}$	$3.71 \times 10^{-10}$
No DOM control (SRHA)	0	$1.33 \times 10^{-10}$	0	$1.44 \times 10^{-11}$	$1.26 \times 10^{-10}$	$1.40 \times 10^{-10}$
No DOM control (SRFA)	0	undersaturated	0	$9.91 \times 10^{-12}$	$5.41 \times 10^{-11}$	$6.40 \times 10^{-11}$



**Figure SI-1.** Relationship between log Hg/thiol ratio and log fraction <sup>201</sup>Hg methylated in solutions containing Suwannee River humic acid (SRHA), sulfide, and <sup>201</sup>HgCl<sub>2</sub>. Data for native SRHA include data from this study and from Graham et al.<sup>12</sup> Data for sulfurized SRHA from this study only. Thiol concentrations were estimated based on measured S/C ratio for SRHA samples and the assumption that 70% of total DOM S was thiols.





**Figure SI-2.** Correlations between the sum of neutral Hg(II)<sub>i</sub> species (**panel a**) or total dissolved Hg(II)<sub>i</sub> (**panel b**) and cell-normalized MeHg production. MeHg production cell-normalized due to significant differences in cell density between experiments. Data log-transformed due to non-normal distributions.

## References for Supporting Information

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