

Residue-Specific Peptide Modification: A Chemist's Guide

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SUPPORTING INFORMATION

Amino acid side-chain modification report card

PROTEIN COMPATIBILITY SCALE				
				
Fully compatible	Compatible	Progress made	Preliminary	Unanswered challenge
<ul style="list-style-type: none"> - protein or antibody substrates <p>Multiple modes of reactivity, each with all of the following:</p> <ul style="list-style-type: none"> - aqueous, mild conditions - fully chemoselective - reproducible 	<ul style="list-style-type: none"> - protein or antibody substrates <p>Any 3 of the following:</p> <ul style="list-style-type: none"> - aqueous, mild conditions - fully chemoselective - multiple modes of reactivity - reproducible 	<ul style="list-style-type: none"> - larger peptide or protein substrates <ul style="list-style-type: none"> - partially chemoselective - non-aqueous or harsh conditions (extreme temp or pH) - one (or few) examples 	<ul style="list-style-type: none"> - small peptides only (di-, tri-, tetra-peptides) <ul style="list-style-type: none"> - proof-of-concept studies - not chemoselective - protecting groups required - not biocompatible (organic reaction media, high temp) 	<ul style="list-style-type: none"> - no peptide examples <ul style="list-style-type: none"> - methods unavailable or demonstrated on single amino acids only - poised for innovation

The amino acid side-chain modification report card provides a graphical overview of available residue-specific modification strategies. Organized by amino acid and subdivided into various reaction classifications (A–L), the report card aims to provide readers with an illustrative summary of available methods and the degree to which they have been evaluated on protein substrates. Historical methods for bioconjugation and recent developments for peptide and protein functionalization were categorized and evaluated on the basis of their suitability for the residue-specific modification of peptides, proteins, and antibodies. The specific parameters used for evaluation are outlined in the “Protein Compatibility Scale” above.

A few notes on the categorization and evaluation process:

- In some cases, a synthetic method falls into more than one reaction classification (A–L). We have generally categorized such methods into all relevant reaction classifications. For example, a Pd-catalyzed arylation might fall under the categories of arylation (**B**), cross-coupling (**H**), and transition-metal functionalization (**L**).

- The compatibility “score” is not meant to be a judgment of the quality of a particular strategy or specific body of work; rather, the score is intended to provide an objective assessment of the suitability of each reaction type, based on available literature, for residue-specific modifications. For example, a “preliminary” functionalization strategy has been demonstrated on small peptides, while a “fully compatible” approach has been successfully applied to the chemoselective modification of proteins and/or antibodies.

A number of comprehensive reviews and literature methods were consulted to assign appropriate compatibility scores. Relevant references are provided in the following lists and amino acid-specific charts. While we have aimed to thoroughly evaluate available methods (particularly those reported in the last decade), the following charts are intended to be representative rather than exhaustive in nature. We refer readers to relevant reviews, where applicable, and have highlighted in purple references that are discussed in more detail in the body of the manuscript.

Select comprehensive reviews:

- (1) Hermanson, G. T. (2013) *Bioconjugate Techniques*, 3rd ed., Academic Press, San Diego, CA, USA.
- (2) Koniev, O., and Wagner, A. (2015) Developments and recent advancements in the field of endogenous amino acid selective bond forming reactions for bioconjugation, *Chem. Soc. Rev.* **44**, 5495–5551.
- (3) McKay, C. S., and Finn, M. G. (2014) Click chemistry in complex mixtures: bioorthogonal bioconjugation, *Chem. Biol.* **21**, 1075–1101.
- (4) Boutureira, O., and Bernardes, G. J. (2015) Advances in chemical protein modification, *Chem. Rev.* **115**, 2174–2195.
- (5) Bondalapati, S., Jbara, M., and Brik, A. (2016) Expanding the chemical toolbox for the synthesis of large and uniquely modified proteins, *Nat. Chem.* **8**, 407–418.
- (6) Spicer, C. D., and Davis, B. G. (2014) Selective chemical protein modification, *Nat. Commun.* **5**, 4740.
- (7) ElSohly, A. M., and Francis, M. B. (2015) Development of oxidative coupling strategies for site-selective protein modification, *Acc. Chem. Res.* **48**, 1971–1978.

- (8) Noisier, A. F., and Brimble, M. A. (2014) C–H functionalization in the synthesis of amino acids and peptides, *Chem. Rev.* **114**, 8775–8806.
- (9) Sletten, E. M., and Bertozzi, C. R. (2009) Bioorthogonal chemistry: Fishing for selectivity in a sea of functionality, *Angew. Chem. Int. Ed.* **48**, 6974–6998.
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- (11) Sengupta, S., and Mehta, G. (2017) Late stage modification of peptides via C–H activation reactions, *Tetrahedron Lett.* **58**, 1357–1372.
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- (13) Schwieter, K. E., and Johnston, J. N. (2016) On-demand complex peptide synthesis: An aspirational (and elusive?) goal for peptide synthesis, *J. Am. Chem. Soc.* **138**, 14160–14169.
- (14) Basle, E., Joubert, N., and Puchault, M. (2010) Protein chemical modification on endogenous amino acids, *Chem. Biol.* **17**, 213–227.
- (15) Lim, R. K., and Lin, Q. (2010) Bioorthogonal chemistry: recent progress and future directions, *Chem. Commun.* **46**, 1589–1600.
- (16) Stephanopoulos, N., and Francis, M. B. (2011) Choosing an effective protein bioconjugation strategy, *Nat. Chem. Biol.* **7**, 876–884.
- (17) Roberts, M. J., Bentley, M. D., and Harris, J. M. (2012) Chemistry for peptide and protein PEGylation, *Adv. Drug Deliv. Rev.* **64**, 116–127.
- (18) Chalker, J. M. (2017) Metal-Mediated Bioconjugation, In *Chemoslective and Bioorthogonal Ligation Reactions*, pp 231–270, Wiley-VCH Verlag GmbH & Co. KGaA, Weinheim, Germany.
- (19) Malins, L. R. (2016) Transition metal-promoted arylation: An emerging strategy for protein bioconjugation, *Aust. J. Chem.* **69**, 1360–1364.
- (20) Yang, M., Li, J., and Chen, P. R. (2014) Transition metal-mediated bioorthogonal protein chemistry in living cells, *Chem. Soc. Rev.* **43**, 6511–6526.
- (21) Antos, J. M., and Francis, M. B. (2006) Transition metal catalyzed methods for site-selective protein modification, *Curr. Opin. Chem. Biol.* **10**, 253–262.
- (22) van Maarseveen, J. H., Reek, J. N., and Back, J. W. (2006) Transition-metal catalysis as a tool for the covalent labeling of proteins, *Angew. Chem. Int. Ed.* **45**, 1841–1843.
- (23) Sambasivarao, K., and Kakali, L. (2005) Post-assembly peptide modifications by chemical methods, *Curr. Med. Chem.* **12**, 849–875.
- (24) Glazer, A. N. (1970) Specific Chemical Modification of Proteins, *Annu. Rev. Biochem.* **39**, 101–130.

		AMINO ACID SIDE-CHAIN MODIFICATION REPORT CARD																				
		Residue	Ala	Cys	Asp/Glu	Phe	Gly	His	Ile	Lys	Leu	Met	Asn/Gln	Pro	Arg	Ser	Thr	Val	Trp	Tyr	Sec	Dha
Method	Residue	Ala	Cys	Asp/Glu	Phe	Gly	His	Ile	Lys	Leu	Met	Asn/Gln	Pro	Arg	Ser	Thr	Val	Trp	Tyr	Sec	Dha	
(A) Alkylation	Ala	●	●	○	○	○	●	●	●	●	●	○	●	○	●	●	●	●	●	●	●	
(B) Arylation	Cys	○	●	●	○	○	●	●	●	●	●	●	●	●	●	●	●	●	●	●	●	
(C) Acylation	Asp/Glu	●	●	●	●	●	●	●	●	●	●	●	●	●	●	●	●	●	●	●	●	
(D) Halogenation	Phe	●	●	●	●	●	●	●	●	●	●	●	●	●	●	●	●	●	●	●	●	
(E) Oxidation	Gly	○	●	●	●	●	●	●	●	●	●	●	●	●	●	●	●	●	●	●	●	
(F) 1,4-Addition	His	●	●	●	●	●	●	●	●	●	●	●	●	●	●	●	●	●	●	●	●	
(G) Condensation	Ile	●	●	●	●	●	●	●	●	●	●	●	●	●	●	●	●	●	●	●	●	
(H) Cross-Coupling	Lys	○	●	○	●	●	●	●	●	●	●	●	●	●	●	●	●	●	●	●	●	
(I) Pericyclic Reaction	Leu	●	●	●	●	●	●	●	●	●	●	●	●	●	●	●	●	●	●	●	●	
(J) Photo Reaction	Met	●	●	●	●	●	●	●	●	●	●	●	●	●	●	●	●	●	●	●	●	
(K) Radical Reaction	Asn/Gln	●	●	●	●	●	●	●	●	●	●	●	●	●	●	●	●	●	●	●	●	
(L) Transition-Metal Functionalization	Pro	●	●	●	●	●	●	●	●	●	●	●	●	●	●	●	●	●	●	●	●	
		Fully compatible - protein or antibody substrates - aqueous, mild conditions - fully chemoselective - multiple modes of reactivity - reproducible (many examples)				●	●	○	○	○	○	●	●	●	●	●	●	●	●	●	●	
		Fully compatible				●	●	●	●	●	●	●	●	●	●	●	●	●	●	●	●	
		Compatible				●	●	●	●	●	●	●	●	●	●	●	●	●	●	●	●	
		Progress made				●	●	●	●	●	●	●	●	●	●	●	●	●	●	●	●	
		Preliminary				●	●	●	●	●	●	●	●	●	●	●	●	●	●	●	●	
		Unanswered challenge - no peptide examples - methods unavailable or single amino acids only - poised for innovation				●	●	●	●	●	●	●	●	●	●	●	●	●	●	●	●	

Amino acid-specific modification charts

ALANINE		
(A) Alkylation		(B) Pd-catalyzed C–H arylation: Gong, W., Zhang, G., Liu, T., Giri, R., and Yu, J.–Q. (2014) <i>J. Am. Chem. Soc.</i> 16940–16946. Tang, J., He, Y., Chen, H., Sheng, W., and Wang, H. (2017) <i>Chem. Sci.</i> 8, 4565–4570.
(B) Arylation		(E) Pd-catalyzed C–H acetoxylation: Gong, W., Zhang, G., Liu, T., Giri, R., and Yu, J.–Q. (2014) <i>J. Am. Chem. Soc.</i> 16940–16946.
(C) Acylation		(H) Pd-catalyzed C–H arylation: Gong, W., Zhang, G., Liu, T., Giri, R., and Yu, J.–Q. (2014) <i>J. Am. Chem. Soc.</i> 16940–16946. Tang, J., He, Y., Chen, H., Sheng, W., and Wang, H. (2017) <i>Chem. Sci.</i> 8, 4565–4570.
(D) Halogenation		(L) Pd-catalyzed C–H arylation: Gong, W., Zhang, G., Liu, T., Giri, R., and Yu, J.–Q. (2014) <i>J. Am. Chem. Soc.</i> 16940–16946. Tang, J., He, Y., Chen, H., Sheng, W., and Wang, H. (2017) <i>Chem. Sci.</i> 8, 4565–4570.
(E) Oxidation		
(F) 1,4-Addition		
(G) Condensation		
(H) Cross-Coupling		
(I) Pericyclic Reaction		
(J) Photo Reaction		
(K) Radicals		
(L) Transition-Metal Functionalization		

CYSTEINE		
Reviews		
A	Alkylation	
B	Arylation	
C	Acylation	
D	Halogenation	
E	Oxidation	
F	1,4-Addition	
G	Condensation	
H	Cross-Coupling	
I	Pericyclic Reaction	
J	Photo Reaction	
K	Radical Reaction	
L	Transition-Metal Functionalization	
(A) Aminoethylation to form Lys mimics:		
Lindley, H. (1956) <i>Nature</i> , 178 , 647–648. Rafferty, M. A. and Cole, R. D. (1966) <i>J. Biol. Chem.</i> , 241 , 3457–3461. Smith, H. B. and Hartman, F. C. (1987) <i>J. Biol. Chem.</i> , 263 , 4921–4925. Simon, M. D., Chu, F., Racki, L. R., de la Cruz, C. C., Burlingame, A. L., Panning, B., Narlikar, G. J., and Shokat, K. M. (2007) <i>Cell</i> , 128 , 1003–1012. Huang, R., Holbert, M. A., Tarrant, M. K., Curtef, S., Colquhoun, D. R., Dancy, B. M., Dancy, B. C., Hwang, Y., Tang, Y., Meeth, K., Marmorstein, R., Cole, R. N., Khochbin, S., and Cole, P. A. (2010) <i>J. Am. Chem. Soc.</i> , 132 , 9986–9987.		
Cross metathesis:		
Lin, Y. A., Chalker, J. M., Floyd, N., Bernardes, G. J. L., and Davis, B. G. (2008) <i>J. Am. Chem. Soc.</i> , 130 , 9642–9643. Chalker, J. M., Lin, Y. A., Boutureira, O., and Davis, B. G. (2009) <i>Chem. Commun.</i> , 3714–3716.		
Arylpropionitriles:		
Koniev, O., Leriche, G., Nothisen, M., Remy, J.-S., Strub, J.-M., Schaeffer-Reiss, C., Van Dorsselaer, A., Baati, R., Wagner, A. (2014) <i>Bioconjugate Chem.</i> , 25 , 202–206. Koloddy, S., Koniev, O., Baatarkhuu, Z., Bonnefoy, J.-Y., Debaene, F., Cianfrani, S., Van Dorsselaer, A., Wagner, A. (2015) <i>Bioconjugate Chem.</i> , 26 , 197–200.		
Allenamides: Abbas, A., Xing, B., and Loh, T.-P. (2014) <i>Angew. Chem. Int. Ed.</i> , 53 , 7491–7494.		
α-halocarbonyls:		
Goddard, D. R. and Michaelis, L. (1935) <i>J. Biol. Chem.</i> , 112 , 361–371. Mackworth, J. F. (1948) <i>Biochem. J.</i> , 42 , 82–90. Gerwin, B. I. (1966) <i>J. Biol. Chem.</i> , 242 , 451–456. Clark, P. I. and Lowe, G. (1977) <i>J. Chem. Soc. Chem. Commun.</i> , 923–924. Clark, P. I. and Lowe, G. (1977) <i>Eur. J. Biochem.</i> , 84 , 293–299. Davis, N. J. and Flitsch, S. L. (1991) <i>Tet. Lett.</i> , 32 , 6793–6796. Kim, J.-R., Yoon, H. W., Kwon, K.-S., Lee, S.-R., and Rhee, S. G. (2000) <i>Anal. Biochem.</i> , 283 , 214–221. Schelte, P., Boeckler, C., Frisch, B., and Schuber, F. (2000) <i>Bioconjugate Chem.</i> , 11 , 118–123. Macmillan, D., Bill, R. M., Sage, K. A., Fern, D., and Flitsch, S. L. (2001) <i>Chem. Biol.</i> , 8 , 133–145. Macmillan, D., Bill, R. M., Sage, K. A., Fern, D., Flitsch, S. L. (2001) <i>Chem. Biol.</i> , 8 , 133–145. Swanson, R. S., Daines, A. M., Tey, L.-H., Flitsch, S. L., and Allemann, R. K. (2005) <i>ChemBioChem</i> , 6 , 1338–1340. Grant, G. A. (2017) <i>Curr. Protoc. Protein.</i> , 87 :15.1.1–15.1.23.		
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Disulfide bridges:		
Balan, S., Choi, J.-w., Godwin, A., Teo, I., Laborde, C. M., Heidelberger, S., Zloh, M., Shaunak, S., Broccolini, S. (2007) <i>Bioconjugate Chem.</i> , 18 , 61–76. Lewis, A., Tang, Y., Broccolini, S., Choi, J.-w., Godwin, A. (2008) <i>Bioconjugate Chem.</i> , 19 , 2144–2155. Smith, M. E. B., Schumacher, F. F., Ryan, C. P., Tedaldi, L. M., Papaioannou, D., Waksman, G., Caddick, S., Baker, J. R. (2010) <i>J. Am. Chem. Soc.</i> , 132 , 1960–1965. Smith, M. E. B., Schnumacher, F. F., Ryan, C. P., Tedaldi, L. M., Papaioannou, D., Waksman, G., Caddick, Baker, J. R. (2010) <i>J. Am. Chem. Soc.</i> , 132 , 1960–1965. Jones, M. W., Strickland, R. A., Schumacher, F. F., Caddick, S., Baker, J. R., Gibson, M. I., Haddleton, D. M. (2012) <i>J. Am. Chem. Soc.</i> , 134 , 1847–1852. Collins, J., Tanaka, J., Wilson, P., Kempe, K., Davis, T. P., McIntosh, M. P., Whittaker, M. R., Haddleton, D. M. (2015) <i>Bioconjugate Chem.</i> , 26 , 633–638. Wang, T., Wu, Y., Kuan, S. L., Dumelle, O., Lamia, M., Ng, D. Y. W., Arzt, M., Thomas, J., Mueller, J. O., Barner-Kowollik, C., Weil, T. (2015) <i>Chem. Eur. J.</i> , 21 , 228–238.		
Pd-mediated cross-coupling:		
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CYSTEINE		
(A)	Alkylation	
(B)	Arylation	
(C)	Acylation	
(D)	Halogenation	
(E)	Oxidation	
(F)	1,4-Addition	
(G)	Condensation	
(H)	Cross-Coupling	
(I)	Pericyclic Reaction	
(J)	Photo Reaction	
(K)	Radical Reaction	
(L)	Transition-Metal Functionalization	
(B) Stapling:		
Tucker, M. J., Courter, J. R., Chen, J., Atasoylu, O., Smith, A. B., III, Hochstrasser, R. M. (2010) <i>Angew. Chem. Int. Ed.</i> 49 , 3612–3616. Spokoyny, A. M., Zou, Y., Ling, J. J., Yu, H., Lin, Y.-S., Pentelute, B. L. (2013) <i>J. Am. Chem. Soc.</i> 135 , 5946–5949. Brown, S. P. and Smith, A. B., III. (2015) <i>J. Am. Chem. Soc.</i> 137 , 4034–4037. Zhang, C., Welborn, M., Zhu, T., Yang, T., Yang, N. J., Santos, M. S., Van Voorhis, T., and Pentelute, B. L. (2016) <i>Nat. Chem.</i> 8 , 120–128. Julia-Kocienski-like reagents: Toda, N., Asano, S., and Barbas, C. F., III. (2013) <i>Angew. Chem. Int. Ed.</i> 52 , 12592–12596.		
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Strain-release functionalization: Gianatassio, Y., Lopchuk, J. M., Wang, J., Pan, C.-M., Malins, L. R., Prieto, L., Brandt, T. A., Collins, M. R., Gallego, G. M., Sach, N. W., Spangler, J. E., Zhu, H., Zhu, J., and Baran, P. S. (2016) <i>Science</i> 351 , 241–246.		
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Arylsulfones: Patterson, J. T., Asano, S., Li, X., Rader, C., and Barbas, C. F., III. (2014) <i>Bioconjugate Chem.</i> 25 , 1402–1407.		
(D) Trifluoromethylation: Capone, S., Kieltsch, I., Flögel, O., Lelais, G., Togni, A., and Seebach, D. (2008) <i>Helv. Chim. Acta</i> 91 , 2035–2056. Bottecchia, C., Wei, X.-J., Kuijpers, K. P. L., Hessel, V., Noel, T. (2016) <i>J. Org. Chem.</i> 81 , 7301–7307.		
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G	Condensation	
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(C) Acylation		(D) Ortho-iodination: Espuña, G., Arsequell, G., Valencia, G., Barluenga, J., Alvarez-Gutiérrez, J. M., Ballesteros, A., and González, J. M. (2004) <i>Angew. Chem. Int. Ed.</i> , 43 , 325–329. Photochemical β-fluorination: Bume, D. D., Pitts, C. R., Jokhai, R. T., and Lectka, T. (2016) <i>Tetrahedron</i> 72 , 6031–6036.
(D) Halogenation		(E) Pd-catalyzed oxidative amidation: Yang, M., Jiang, X., and Shi, Z.-J. (2015) <i>Org. Chem. Front.</i> 2 , 51–54.
(E) Oxidation		(H) Iodination–Suzuki–Miyaura reaction: Espuña, G., Arsequell, G., Valencia, G., Barluenga, J., Alvarez-Gutiérrez, J. M., Ballesteros, A., and González, J. M. (2004) <i>Angew. Chem. Int. Ed.</i> , 43 , 325–329. Pd-catalyzed β-arylation: Mondal, B., Roy, B., and Kazmaier, U. (2016) <i>J. Org. Chem.</i> 81 , 11646–11655. Pd-catalyzed oxidative amidation: Yang, M., Jiang, X., and Shi, Z.-J. (2015) <i>Org. Chem. Front.</i> 2 , 51–54.
(F) 1,4-Addition		(J) Photochemical β-fluorination: Bume, D. D., Pitts, C. R., Jokhai, R. T., and Lectka, T. (2016) <i>Tetrahedron</i> 72 , 6031–6036.
(G) Condensation		(K) Photochemical β-fluorination: Bume, D. D., Pitts, C. R., Jokhai, R. T., and Lectka, T. (2016) <i>Tetrahedron</i> 72 , 6031–6036.
(H) Cross-Coupling		(L) Dirhodium metalloclopeptide functionalization: Popp, B. V. and Ball, Z. T. (2010) <i>J. Am. Chem. Soc.</i> 132 , 6660–6662. Popp, B. V., and Ball, Z. T. (2011) <i>Chem. Sci.</i> 2 , 690–695. Iodination–Suzuki–Miyaura reaction: Espuña, G., Arsequell, G., Valencia, G., Barluenga, J., Alvarez-Gutiérrez, J. M., Ballesteros, A., and González, J. M. (2004) <i>Angew. Chem. Int. Ed.</i> , 43 , 325–329. Pd-catalyzed β-arylation: Mondal, B., Roy, B., and Kazmaier, U. (2016) <i>J. Org. Chem.</i> 81 , 11646–11655.
(I) Pericyclic Reaction		Pd-catalyzed oxidative amidation: Yang, M., Jiang, X., and Shi, Z.-J. (2015) <i>Org. Chem. Front.</i> 2 , 51–54.
(J) Photo Reaction		
(K) Radical Reaction		
(L) Transition-Metal Functionalization		

GLYCINE		
(A)	Alkylation	
(B)	Arylation	
(C)	Acylation	
(D)	Halogenation	
(E)	Oxidation	
(F)	1,4-Addition	
(G)	Condensation	
(H)	Cross-Coupling	
(I)	Pericyclic Reaction	
(J)	Photo Reaction	
(K)	Radical Reaction	
(L)	Transition-Metal Functionalization	

(A) **Photoalkylation:**

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(B) **Cross-dehydrogenative coupling:**

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(D) **Radical bromination:** Easton, C. J., Tan, E. W., and Hay, M. P. (1989) *J. Chem. Soc., Chem. Commun.*, 385–386.

(E) **Cross-dehydrogenative coupling:** Zhao, L., Basle, O., and Li, C. J. (2009) *Proc. Natl. Acad. Sci. U. S. A.* **106**, 4106–4111.

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 Schwarzberg, M., Sperling, J., and Elad, D. (1973) *J. Am. Chem. Soc.* **95**, 6418–6426.

Zhu, S., and Rueping, M. (2012) *Chem. Commun.* **48**, 11960–11962.

Photoarylation: Zhu, S. and Rueping, M. (2012) *Chem. Commun.* **48**, 11960–11962.

(H) **Cross-dehydrogenative coupling:**

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 Zhao, L., Basle, O., and Li, C. J. (2009) *Proc. Natl. Acad. Sci. U. S. A.* **106**, 4106–4111.

(K) **Samarium diiodide functionalization:** Ricci, M., Blakskjaer, P., and Skrydstrup, T. (2000) *J. Am. Chem. Soc.* **122**, 12413–12421.

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HISTIDINE		
(A)	Alkylation	
(B)	Arylation	
(C)	Acylation	
(D)	Halogenation	
(E)	Oxidation	
(F)	1,4-Addition	
(G)	Condensation	
(H)	Cross-Coupling	
(I)	Pericyclic Reaction	
(J)	Photo Reaction	
(K)	Radical Reaction	
(L)	Transition-Metal Functionalization	

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(I) Radical Reaction:

- Trifluoromethylation: Labroo, V. M., Labroo, R. B., and Cohen, L. A. (1990) *Tet. Lett.* **31**, 5705–5708.

(J) Transition-Metal Functionalization:

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(A) Alkylation		
(B) Arylation		(B) Pd-catalyzed arylation: Rodriguez, N., Romero-Revilla, J. A., Fernandez-Ibanez, M. A., and Carretero, J. C. (2013) <i>Chem. Sci.</i> 4, 175–179.
(C) Acylation		(E) Fe-catalyzed oxidation: Gormisky, P. E. and White, M. C. (2013) <i>J. Am. Chem. Soc.</i> 135, 14052–14055.
(D) Halogenation		(H) Pd-catalyzed arylation: Rodriguez, N., Romero-Revilla, J. A., Fernandez-Ibanez, M. A., and Carretero, J. C. (2013) <i>Chem. Sci.</i> 4, 175–179.
(E) Oxidation		(L) Fe-catalyzed oxidation: Gormisky, P. E. and White, M. C. (2013) <i>J. Am. Chem. Soc.</i> 135, 14052–14055. Pd-catalyzed arylation: Rodriguez, N., Romero-Revilla, J. A., Fernandez-Ibanez, M. A., and Carretero, J. C. (2013) <i>Chem. Sci.</i> 4, 175–179.
(F) 1,4-Addition		
(G) Condensation		
(H) Cross-Coupling		
(I) Pericyclic Reaction		
(J) Photo Reaction		
(K) Radical Reaction		
(L) Transition-Metal Functionalization		

LYSINE		
(A) Alkylation		(A) Reductive amination: Means, G. E. and Feeney, R. E. (1968) <i>Biochemistry</i> 7, 2192–2201. Gray, G. R. (1974) <i>Arch. Biochem. Biophys.</i> 163, 426–428. Schwartz, B. A. and Gray, G. R. (1977) <i>Arch. Biochem. Biophys.</i> 181, 542–549. Lee, R. T. and Lee, Y. C. (1980) <i>Biochemistry</i> 19, 156–163. Gildersleeve, J. C., Oyelaran, O., Simpson, J. T., Allred, B. (2008) <i>Bioconjugate Chem.</i> 19, 1485–1490. McFarland, J. M. and Francis, M. B. (2005) <i>J. Am. Chem. Soc.</i> 127, 13490–13491.
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(C) Acylation		α-halo carbonyls: Gurd, F. R. N. (1971) <i>Methods Enzymol.</i> 25, 424–438.
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(G) Condensation		Pd-catalyzed arylation: Lee, H. G., Lautrette, G., Pentelute, B. L., and Buchwald, S. L. (2017) <i>Angew. Chem. Int. Ed.</i> 56, 3177–3181.
(H) Cross-Coupling		(C) Potassium acyltrifluoroborates: Noda, H., Eros, G., and Bode, J. W. (2014) <i>J. Am. Chem. Soc.</i> 136, 5611–5614. Isocyanates: Creech, H. J. and Jones, R. N. (1941) <i>J. Am. Chem. Soc.</i> 63, 1670–1673.
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(J) Photo Reaction		(E) Iminoboronates: Cal, P. M. S. D., Vicente, J. B., Pires, E., Coelho, A. V., Veiros, L. F., Cordeiro, C., and Gois, P. M. P. (2012) <i>J. Am. Chem. Soc.</i> 134, 10299–10305. Oxidative coupling with anilines: Hooker, J. M., Esser-Kahn, A. P., and Francis, M. B. (2006) <i>J. Am. Chem. Soc.</i> 128, 15558–15559.
(K) Radical Reaction		(F) Valero, E., Tambalo, S., Marzola, P., Ortega-Muñoz, M., López-Jaramillo, F. J., Santoyo-González, F., de Dios López, J., Delgado, J. J., Calvino, J. J., Cuesta, R., Domínguez-Vera, J. M., Gálvez, N. (2011) <i>J. Am. Chem. Soc.</i> 133, 4889–4895. Maleic anhydride: Butler, P. J. G., Harris, J. I., Hartley, B. S., and Leberman, R. (1969) <i>Biochem. J.</i> 112, 679–689.
(L) Transition-Metal Functionalization		(G) Citraconic anhydride: Dixon, H. B. F. and Perham, R. N. (1968) <i>Biochem. J.</i> 109, 312–314. Vinyl sulfones: Megia-Fernandez, A., Hernandez-Mateo, F., and Santoyo-Gonzalez, F. (2013) <i>Org. Biomol. Chem.</i> 11, 2586–2596. Reductive amination: Means, G. E. and Feeney, R. E. (1968) <i>Biochemistry</i> 7, 2192–2201. Gray, G. R. (1974) <i>Arch. Biochem. Biophys.</i> 163, 426–428. Schwartz, B. A. and Gray, G. R. (1977) <i>Arch. Biochem. Biophys.</i> 181, 542–549.

LYSINE		
(A) Alkylation		Reductive amination (continued): Lee, R. T. and Lee, Y. C. (1980) <i>Biochemistry</i> 19, 156–163. McFarland, J. M. and Francis, M. B. (2005) <i>J. Am. Chem. Soc.</i> 127, 13490–13491. Gildersleeve, J. C., Oyelaran, O., Simpson, J. T., Allred, B. (2008) <i>Bioconjugate Chem.</i> 19, 1485–1490. With appropriate N-terminus protection, Lys condensation reactions to form the corresponding amide can generally be achieved under standard peptide coupling conditions.
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(C) Acylation		Kinetically controlled amidation: Chen, X., Muthoosamy, K., Pfisterer, A., Neumann, B., and Weil, T. (2012) <i>Bioconjugate Chem.</i> 23, 500–508.
(D) Halogenation		Imidoesters: Hunter, M. J. and Ludwig, M. L. (1962) <i>J. Am. Chem. Soc.</i> 84, 3491–3502. Traut, R. R., Bollen, A., Sun, T.-T., Hershey, J. W. B., Sundberg, J., Pierce, L. R. (1973) <i>Biochemistry</i> 12, 3266–3273. Lee, Y.-C., Stowell, C. P., Krantz, M. J. (1976) <i>Biochemistry</i> 15, 3956–3963. Denny, J. B. and Roberts, R. M. (1982) <i>J. Biol. Chem.</i> 257, 2460–2468. Mehlhorn, R., Swanson, M., Packer, L., Smith, P. (1980) <i>Arch. Biochem. Biophys.</i> 204, 471–476. Wood, F. T., Wu, M. M., Gerhart, J. C. (1975) <i>Anal. Biochem.</i> 69, 339–349.
(E) Oxidation		H Pd-catalyzed arylation: Lee, H. G., Lautrette, G., Pentelute, B. L., and Buchwald, S. L. (2017) <i>Angew. Chem. Int. Ed.</i> 56, 3177–3181.
(F) 1,4-Addition		I 6π-electrocyclization: Tanaka, K., Masuyama, T., Hasegawa, K., Tahara, T., Mizuma, H., Wada, Y., Watanabe, Y., Fukase, K. (2007) <i>Angew. Chem. Int. Ed.</i> 47, 102–105. Tanaka, K., Fujii, Y., and Fukase, K. (2008) <i>ChemBioChem</i> 9, 2392–2397. Tanaka, K., Fukase, K., and Katsumura, S. (2011) <i>Synlett</i> 2115–2139.
(G) Condensation		L Pd-catalyzed arylation: Lee, H. G., Lautrette, G., Pentelute, B. L., and Buchwald, S. L. (2017) <i>Angew. Chem. Int. Ed.</i> 56, 3177–3181.
(H) Cross-Coupling		Technetium-mediated isothiocyanation Linder, K. E., Wen, M. D., Nowotnik, D. P., Malley, M. F., Gougoutas, J. A., Nunn, A. D., and Eckelman, W. C. (1991) <i>Bioconjugate Chem.</i> 2, 160–170.
(I) Pericyclic Reaction		Misc. Sulfonates: Gais, H.-J. and Ruppert, S. (1995) <i>Tet. Lett.</i> 36, 3837–3838.
(J) Photo Reaction		Sulfonyl halides: Lefevre, C., Kang, H. C., Haugland, R. P., Malekzadeh, N., Arttamangkul, S., and Haugland, R. P. (1996) <i>Bioconjugate Chem.</i> 7, 482–489. Gray, W. R. (1967) <i>Methods Enzymol.</i> 11, 139–151. Titus, J. A., Haugland, R., Sharroow, S. O., and Segal, D. M. (1982) <i>J. Immunol. Methods</i> 50, 193–204. Herzig, D. J., Rees, A. W., and Day, R. A. (1964) <i>Biopolymers</i> 2, 349–360. Gros, C. and Labouresse, B. (1969) <i>Eur. J. Biochem.</i> 7, 463–470. Lin, J.-K. and Chang, J.-Y. (1975) <i>Anal. Chem.</i> 47, 1634–1638. Nilsson, K. and Mosbach, K. (1981) <i>Biochem. Biophys. Res. Commun.</i> 102, 449–457.
(K) Radical Reaction		Glutaldehyde functionalization: Ikeda, Y., Katamachi, J., Kawasaki, H., Nagasaki, Y. (2013) <i>Bioconjugate Chem.</i> 24, 1824–1827.
(L) Transition-Metal Functionalization		Isothiocyanates: Amante, L., Ancona, A., Forni, L. (1972) <i>J. Immunol. Methods</i> 1, 289–301. Huang, K.-H. and Cantor, C. R. (1972) <i>J. Mol. Biol.</i> 67, 265–275. Edidin, M., Zagynsky, Y., Lardner, T. J. (1976) <i>Science</i> 191, 466–468.

LYSINE		
(A)	Alkylation	
(B)	Arylation	
(C)	Acylation	
(D)	Halogenation	
(E)	Oxidation	
(F)	1,4-Addition	
(G)	Condensation	
(H)	Cross-Coupling	
(I)	Pericyclic Reaction	
(J)	Photo Reaction	
(K)	Radical Reaction	
(L)	Transition-Metal Functionalization	

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		Review: Deming, T. J. (2017) <i>Bioconjugate Chem.</i> , 28, 691–700.
(A)	Alkylation	
(B)	Arylation	
(C)	Acylation	
(D)	Halogenation	
(E)	Oxidation	
(F)	1,4-Addition	
(G)	Condensation	
(H)	Cross-Coupling	
(I)	Pericyclic Reaction	
(J)	Photo Reaction	
(K)	Radical Reaction	
(L)	Transition-Metal Functionalization	
(A)	Formation of sulfonium salt:	
	Toennies, G. (1940) <i>J. Biol. Chem.</i> 132, 455–456.	
	Toennies, G. and Kolb, J. J. (1945) <i>J. Am. Chem. Soc.</i> 67, 849–851.	
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	Gharakhanian, E. G. and Deming, T. J. (2015) <i>Biomacromolecules</i> 16, 1802–1806.	
	Gharakhanian, E. G. and Deming, T. J. (2016) <i>Chem. Commun.</i> 52, 5336–5339.	
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(E)	Via ReACT reagents:	
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ASPARAGINE/GLUTAMINE		
(A)	Alkylation	
(B)	Arylation	
(C)	Acylation	
(D)	Halogenation	
(E)	Oxidation	
(F)	1,4-Addition	
(G)	Condensation	
(H)	Cross-Coupling	
(I)	Pericyclic Reaction	
(J)	Photo Reaction	
(K)	Radical Reaction	
(L)	Transition-Metal Functionalization	

(A) Rh-catalyzed diazo reactions: Popp, B. V., and Ball, Z. T. (2011) *Chem. Sci.* 2, 690–695.

(C) Acylation with KAT reagents: Galvez, A. O., Schaack, C. P., Noda, H., and Bode, J. W. (2017) *J. Am. Chem. Soc.* 139, 1826–1829.

(G) Condensation with formaldehyde: Metz, B., et al. (2004) *J. Biol. Chem.* 279, 6235–6243.

(L) Rh-catalyzed diazo reactions: Popp, B. V., and Ball, Z. T. (2011) *Chem. Sci.* 2, 690–695.

PROLINE		
(A)	Alkylation	
(B)	Arylation	
(C)	Acylation	
(D)	Halogenation	
(E)	Oxidation	
(F)	1,4-Addition	
(G)	Condensation	
(H)	Cross-Coupling	
(I)	Pericyclic Reaction	
(J)	Photo Reaction	
(K)	Radical Reaction	
(L)	Transition-Metal Functionalization	

- (B) Cu-catalyzed oxidative indolation: Wu, X., Zhang, D., Zhou, S., Gao, F., and Liu, H. (2015) *Chem. Commun.* 51, 12571–12573.
 Fe-catalyzed oxidative arylation: Osberger, T. J., Rogness, D. C., Kohrt, J. T., Stepan, A. F., and White, M. C. (2016) *Nature* 537, 214–219.
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 Fe-catalyzed oxidative arylation: Osberger, T. J., Rogness, D. C., Kohrt, J. T., Stepan, A. F., and White, M. C. (2016) *Nature* 537, 214–219.
- (H) Cu-catalyzed oxidative indolation: Wu, X., Zhang, D., Zhou, S., Gao, F., and Liu, H. (2015) *Chem. Commun.* 51, 12571–12573.
 Fe-catalyzed oxidative arylation: Osberger, T. J., Rogness, D. C., Kohrt, J. T., Stepan, A. F., and White, M. C. (2016) *Nature* 537, 214–219.
 Pd-catalyzed arylation: Mondal, B., Roy, B., and Kazmaier, U. (2016) *J. Org. Chem.* 81, 11646–11655.
- (L) Cu-catalyzed oxidative indolation: Wu, X., Zhang, D., Zhou, S., Gao, F., and Liu, H. (2015) *Chem. Commun.* 51, 12571–12573.
 Fe-catalyzed oxidative arylation: Osberger, T. J., Rogness, D. C., Kohrt, J. T., Stepan, A. F., and White, M. C. (2016) *Nature* 537, 214–219.
 Pd-catalyzed arylation: Mondal, B., Roy, B., and Kazmaier, U. (2016) *J. Org. Chem.* 81, 11646–11655.

ARGININE		
(A) Alkylation		
(B) Arylation		
(C) Acylation		
(D) Halogenation		
(E) Oxidation		
(F) 1,4-Addition		
(G) Condensation		
(H) Cross-Coupling		
(I) Pericyclic Reaction		
(J) Photo Reaction		
(K) Radical Reaction		
(L) Transition-Metal Functionalization		

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- (A) Rh-catalyzed diazo reactions: Popp, B. V., and Ball, Z. T. (2011) *Chem. Sci.* 2, 690–695.
- (C) Guanidine acylation:
Grundler, V., and Gademann, K. (2014) *ACS Med. Chem. Lett.* 5, 1290–1295.
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- (G) Condensation with dicarbonyl reagents (e.g., cyclohexanedione, malonaldehyde, glyoxal reagents):
King, T. P. (1966) *Biochemistry* 5, 3454–3459.
Toi, K., Bynum, E., Norris, E., and Itano, H. A. (1967) *J. Biol. Chem.* 242, 1036–1043.
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- (L) Rh-catalyzed diazo reactions: Popp, B. V., and Ball, Z. T. (2011) *Chem. Sci.* 2, 690–695.

SERINE		
(A) Alkylation		Reviews: (2011) Modification of Hydroxyl and Carboxyl Functional Groups in Proteins, In <i>Chemical Modification of Biological Polymers</i> , pp 115–166, CRC Press Trader, D. J., and Carlson, E. E. (2012) <i>Mol. BioSyst.</i> 8, 2484–2493. (A) Rh-catalyzed diazo reactions: Popp, B. V., and Ball, Z. T. (2011) <i>Chem. Sci.</i> 2, 690–695.
(B) Arylation		(C) Acylation (e.g. depsipeptide synthesis): [Review] Hamada, Y., and Shioiri, T. (2005) <i>Chem. Rev.</i> 105, 4441–4482. [Review] Tsakos, M., Schaffert, E. S., Clement, L. L., Villadsen, N. L., and Poulsen, T. B. (2015) <i>Nat. Prod. Rep.</i> 32, 605–632. Ferreira, P. M. T., Monteiro, L. S., Pereira, G., Ribeiro, L., Sacramento, J., and Silva, L. (2007) <i>Eur. J. Org. Chem.</i> 5934–5949. Acylation with NHS esters: Miller, B. T., Collins, T. J., Nagle, G. T., and Kurosky, A. (1992) <i>J. Biol. Chem.</i> 267, 5060–5069. Miller, B. T., and Kurosky, A. (1993) <i>Biochem. Biophys. Res. Commun.</i> 196, 461–467. Miller, B. T., Rogers, M. E., Smith, J. S., and Kurosky, A. (1994) <i>Anal. Biochem.</i> 219, 240–248. Miller, B. T. (1996) <i>Biochem. Biophys. Res. Commun.</i> 218, 377–382. Miller, B. T., Collins, T. J., Rogers, M. E., and Kurosky, A. (1997) <i>Peptides</i> 18, 1585–1595. Kalkhof, S., and Sinz, A. (2008) <i>Anal. Bioanal. Chem.</i> 392, 305–312. Mäder, S., Gschwind, S., and Zenobi, R. (2010) <i>Anal. Biochem.</i> 398, 123–125.
(C) Acylation		(E) Fitzner-Moffatt oxidation: D'Angeli, F., Giormani, V., Filira, F., and Di Bello, C. (1967) <i>Biochem. Biophys. Res. Commun.</i> 28, 809–814. Marzotto, A., and Giormani, V. (1970) <i>Experientia</i> 26, 833–834.
(D) Halogenation		(F) Pb(OAc) ₄ oxidation: Apitz, G., and Steglich, W. (1991) <i>Tetrahedron Lett.</i> 32, 3163–3166. Apitz, G., Jäger, M., Jaroch, S., Kratzel, M., Schäffeler, L., and Steglich, W. (1993) <i>Tetrahedron</i> 49, 8223–8232.
(E) Oxidation		(G) Sulfenylation: Gold, A. M. (1965) <i>Biochemistry</i> 4, 897–901. [Review] Narayanan, A., and Jones, L. H. (2015) <i>Chem. Sci.</i> 6, 2650–2659. Sulfonylation-displacement (Ser to Cys conversion): Zioudrou, C., Wilchek, M., and Patchornik, A. (1965) <i>Biochemistry</i> 4, 1811–1822. Polgar, L., and Bender, M. L. (1966) <i>J. Am. Chem. Soc.</i> 88, 3153–3154. Neet, K. E., and Koshland, D. E., Jr. (1966) <i>Proc. Natl. Acad. Sci. U. S. A.</i> 53, 1606–1611. Martin, J., Slade, A., Aitken, A., Arche, R., and Virden, R. (1991) <i>Biochem. J.</i> 280, 659–662.
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(G) Condensation		Phosphorylation: Zioudrou, C., Wilchek, M., and Patchornik, A. (1965) <i>Biochemistry</i> 4, 1811–1822. DiPersio, L. P., Fontaine, R. N., and Hui, D. Y. (1990) <i>J. Biol. Chem.</i> 265, 16801–16806. Sachdeva, A., and Silverman, S. K. (2010) <i>Chem. Commun.</i> 46, 2215–2217. Wong, O. Y., Pradeepkumar, P. I., and Silverman, S. K. (2011) <i>Biochemistry</i> 50, 4741–4749.
(H) Cross-Coupling		Oxazoline formation: [Review] Gant, T. G., and Meyers, A. I. (1994) <i>Tetrahedron</i> 50, 2297–2360. Phillips, A. J., Uto, Y., Wipf, P., Reno, M. J., and Williams, D. R. (2000) <i>Org. Lett.</i> 2, 1165–1168. [Review] Hamada, Y., and Shioiri, T. (2005) <i>Chem. Rev.</i> 105, 4441–4482.
(I) Pericyclic Reaction		(L) Rh-catalyzed diazo reactions: Popp, B. V., and Ball, Z. T. (2011) <i>Chem. Sci.</i> 2, 690–695.
(J) Photo Reaction		
(K) Radical Reaction		
(L) Transition-Metal Functionalization		

THREONINE		
(A)	Alkylation	
(B)	Arylation	
(C)	Acylation	
(D)	Halogenation	
(E)	Oxidation	
(F)	1,4-Addition	
(G)	Condensation	
(H)	Cross-Coupling	
(I)	Pericyclic Reaction	
(J)	Photo Reaction	
(K)	Radical Reaction	
(L)	Transition-Metal Functionalization	

Reviews: (2011) Modification of Hydroxyl and Carboxyl Functional Groups in Proteins, In *Chemical Modification of Biological Polymers*, pp 115–166, CRC Press
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(C) Acylation (e.g. depsipeptide synthesis):

[Review] Hamada, Y., and Shioiri, T. (2005) *Chem. Rev.* 105, 4441–4482.

[Review] Tsakos, M., Schaffert, E. S., Clement, L. L., Villadsen, N. L., and Poulsen, T. B. (2015) *Nat. Prod. Rep.* 32, 605–632.
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Acylation with NHS esters:

Miller, B. T., Rogers, M. E., Smith, J. S., and Kurosky, A. (1994) *Anal. Biochem.* 219, 240–248.

Miller, B. T. (1996) *Biochem. Biophys. Res. Commun.* 218, 377–382.

Kalkhof, S., and Sinz, A. (2008) *Anal. Bioanal. Chem.* 392, 305–312.

Mädler, S., Gschwind, S., and Zenobi, R. (2010) *Anal. Biochem.* 398, 123–125.

(E) Pfitzner-Moffatt oxidation:

D'Angeli, F., Scoffone, E., Filira, F., and Giormani, V. (1966) *Tetrahedron Lett.* 7, 2745–2748.

D'Angeli, F., Giormani, V., Filira, F., and Di Bello, C. (1967) *Biochem. Biophys. Res. Commun.* 28, 809–814.

Marzotto, A., and Giormani, V. (1970) *Experientia* 26, 833–834.

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Apitz, G., and Steglich, W. (1991) *Tetrahedron Lett.* 32, 3163–3166.

Apitz, G., Jäger, M., Jaroch, S., Kratzel, M., Schäffeler, L., and Steglich, W. (1993) *Tetrahedron* 49, 8223–8232.

(G) Oxazoline formation:

[Review] Gant, T. G., and Meyers, A. I. (1994) *Tetrahedron* 50, 2297–2360.

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Reactions with sulfonyl fluoride reagents: [Review] Narayanan, A., and Jones, L. H. (2015) *Chem. Sci.* 6, 2650–2659.

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(A) Alkylation		(B) Pd-catalyzed C–H arylation: Rodriguez, N., Romero-Revilla, J. A., Fernandez-Ibanez, M. A., and Carretero, J. C. (2013) <i>Chem. Sci.</i> 4, 175–179.
(B) Arylation		(E) Fe-catalyzed C–H oxidation: Osberger, T. J., Rogness, D. C., Kohrt, J. T., Stepan, A. F., and White, M. C. (2016) <i>Nature</i> 537, 214–219. TFDO Oxidation: Rella, M. R., and Williard, P. G. (2007) <i>J. Org. Chem.</i> 72, 525–531. Annese, C., Fanizza, I., Calvano, C. D., D'Accolti, L., Fusco, C., Curci, R., and Williard, P. G. (2011) <i>Org. Lett.</i> 13, 5096–5099.
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(D) Halogenation		
(E) Oxidation		
(F) 1,4-Addition		
(G) Condensation		
(H) Cross-Coupling		
(I) Pericyclic Reaction		
(J) Photo Reaction		
(K) Radical Reaction		
(L) Transition-Metal Functionalization		

TRYPTOPHAN		
(A)	Alkylation	
(B)	Arylation	
(C)	Acylation	
(D)	Halogenation	
(E)	Oxidation	
(F)	1,4-Addition	
(G)	Condensation	
(H)	Cross-Coupling	
(I)	Pericyclic Reaction	
(J)	Photo Reaction	
(K)	Radical Reaction	
(L)	Transition-Metal Functionalization	

(A) Alkylation with Koshland's reagent (2-hydroxy-5-nitrobenzyl bromide):

Koshland, D. E., Karkhanis, Y. D., and Latham, H. G. (1964) *J. Am. Chem. Soc.* **86**, 1448–1450.
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Antos, J. M., and Francis, M. B. (2004) *J. Am. Chem. Soc.* **126**, 10256–10257.
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(B) Pd-catalyzed arylation:

Chapman, C. J., Matsuno, A., Frost, C. G., and Willis, M. C. (2007) *Chem. Commun.* 3903–3905.
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Ru-catalyzed arylation: Schischko, A., Ren, H., Kaplaneris, N., and Ackermann, L. (2017) *Angew. Chem. Int. Ed.* **56**, 1576–1580.

(C) N-acyl tryptophan: Popov, V., Panda, S. S., and Katritzky, A. R. (2013) *Org. Biomol. Chem.* **11**, 1594–1597.

(D) Oxidation through halogenated intermediates:

Viswanatha, T., Lawson, W. B., and Witkop, B. (1960) *Biochim. Biophys. Acta* **40**, 216–224.
 Spande, T. F., Green, N. M., and Witkop, B. (1966) *Biochemistry* **5**, 1926–1933.
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 Bell, J. E., Castellino, F. J., Trayer, I. P., and Hill, R. L. (1975) *J. Biol. Chem.* **250**, 7579–7585.
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 Savige, W. E., and Fontana, A. (1977) *Methods Enzymol.* **47**, 442–453.

(E) CAN oxidative aniline coupling: Seim, K. L., Obermeyer, A. C., and Francis, M. B. (2011) *J. Am. Chem. Soc.* **133**, 16970–16976.

Oxindole synthesis (DMSO, HCl):

Savige, W. E., and Fontana, A. (1976) *J. Chem. Soc., Chem. Commun.* 599–600.
 Savige, W. E., and Fontana, A. (1977) *Methods Enzymol.* **47**, 442–453.

TRYPTOPHAN		
(A)	Alkylation	
(B)	Arylation	
(C)	Acylation	
(D)	Halogenation	
(E)	Oxidation	
(F)	1,4-Addition	
(G)	Condensation	
(H)	Cross-Coupling	
(I)	Pericyclic Reaction	
(J)	Photo Reaction	
(K)	Radical Reaction	
(L)	Transition-Metal Functionalization	

Oxindole synthesis (NBS):

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(C)	Acylation	
(D)	Halogenation	
(E)	Oxidation	
(F)	1,4-Addition	
(G)	Condensation	
(H)	Cross-Coupling	
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(H)	Cross-Coupling	
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(C) Acylation		Oxidative cross-linking: [Review] Kodadek, T., Duroux-Richard, I., and Bonnafous, J.-C. (2005) <i>Trends Pharmacol. Sci.</i> 26, 210–217.
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(I) Pericyclic Reaction		
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(L) Transition-Metal Functionalization		

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(A)	Alkylation	
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(G)	Condensation	
(H)	Cross-Coupling	
(I)	Pericyclic Reaction	
(J)	Photo Reaction	
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(G) Condensation		
(H) Cross-Coupling		
(I) Pericyclic Reaction		
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