

**–Supporting Information –**

**Ruthenium(II)-Catalyzed *ortho*-C–H Chalcogenation of Benzoic Acids via  
Weak *O*-Coordination: Synthesis of Chalcogenoxanthenes**

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## Table of Contents

	Page
<b>General information</b>	<b>S03</b>
<b>Screening of the reaction conditions</b>	<b>S03</b>
<b>General procedure : Ru(II)-catalyzed direct C–H selenylation of aromatic carboxylic acids</b>	<b>S05</b>
<b>General procedure : Ru(II)-catalyzed direct C–H sulfenylation of aromatic carboxylic acids</b>	<b>S06</b>
<b>General procedure : Sequential synthesis of chalcogenoxanthenes</b>	<b>S07</b>
<b>Control experiments</b>	<b>S07</b>
<b>Crystallographic experimental section</b>	<b>S11</b>
<b>Spectroscopic data</b>	<b>S14</b>
<b>NMR spectra</b>	<b>S22</b>

## General information

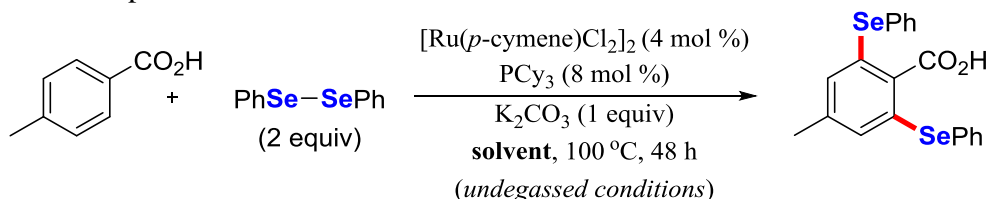
All non-aqueous reactions were carried out under an air atmosphere in flame-dried glassware and were stirred using a magnetic stir plate. All reactions were carried out using anhydrous solvent unless otherwise noted. DMSO and DMF were purchased from Acros Organic Company. Dry toluene, xylene, tetrahydrofuran and 1,4-dioxane were prepared by distilling over sodium ketyl.  $[\text{Ru}(p\text{-cymene})\text{Cl}_2]_2$  was purchased from Alfa-Aesar Company.

All reactions were monitored by thin layer chromatography (TLC) on WhatmanPartisil® K6F TLC plates (silica gel 60 Å, 0.25 mm thickness) and visualized using a UV lamp (366 or 254 nm) or by use of one of the following visualization reagents: PMA: 10 g phosphomolybdic acid/ 100 mL ethanol,  $\text{KMnO}_4$ : 0.75 g potassium permanganate, 5 g  $\text{K}_2\text{CO}_3$  / 100mL water. Products were isolated by column chromatography (Merck silica gel 100-200µm). Yields refer to chromatographically and spectroscopically homogenous materials unless noted otherwise.  $^{13}\text{C}$  and  $^1\text{H}$  NMR spectra were recorded on a Bruker 400 or Bruker 500 MHz spectrometers. Chemical shift values ( $\delta$ ) are reported in ppm and calibrated to the residual solvent peak  $\text{CDCl}_3$   $\delta = 7.260$  ppm for  $^1\text{H}$ ,  $\delta = 77.160$  ppm for  $^{13}\text{C}$ ,  $\text{DMSO}-d_6$   $\delta = 2.500$  ppm for  $^1\text{H}$ ,  $\delta = 39.500$  ppm for  $^{13}\text{C}$  or calibrated to tetramethylsilane ( $\delta = 0.00$ ). All NMR spectra were recorded at ambient temperature (290 K) unless otherwise noted.  $^1\text{H}$  NMR spectra are reported as follows: chemical shift (multiplicity, coupling constant, integration). The following abbreviations are used to indicate multiplicities: s, singlet; d, doublet; t, triplet; q, quartet; quint, quintet; sext, sextet; sept, septet; m, multiplet; dd, doublet of doublet; dt, doublet of triplet; dq, doublet of quartet; td, triplet of doublet; tt, triplet of triplet; dq, doublet of quartet; br, broad; app, apparent.

Mass spectra were recorded by electron spray ionization (ESI) method on a Q-TOF Micro with lock spray source. The crystal data were collected and integrated using a BrukerAxs kappa apex2 CCD diffractometer, with graphite monochromated Mo-K $\alpha$  radiation.

## Screening of the reaction conditions:

**Table S1: Solvent optimization**

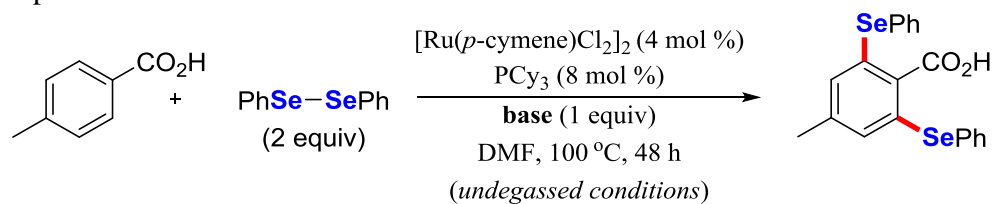


entry	solvent	yield (%)
1	DMF	40
2	DMSO	-
3	Xylene	trace
4	Toluene	trace
5	1,4-Dioxane	14 (27) <sup>a</sup>
6	$\text{CH}_3\text{OH}$	-

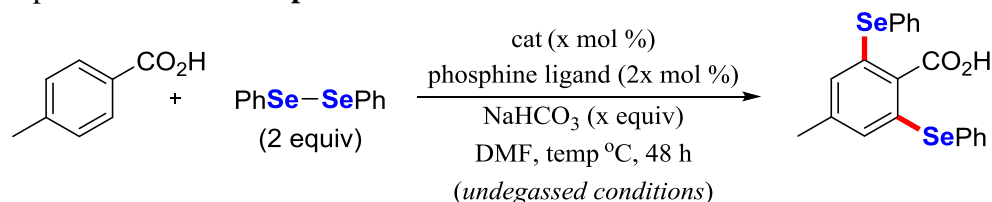
7	THF	-
8	DCE	-
9	DME	-
10	CH <sub>3</sub> CN	-
11	DMAc	-
12	Trifluorotoluene	12 (20) <sup>a</sup>
13	1,3-Bis(trifluoromethyl)benzene	8

<sup>a</sup>NaHCO<sub>3</sub> was used instead of K<sub>2</sub>CO<sub>3</sub> and yields are given in parenthesis.

**Table S2:** Optimization of **bases**



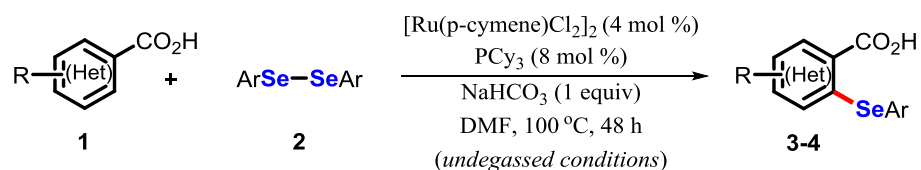
Entry	base	yield (%)
1	Na <sub>2</sub> CO <sub>3</sub>	90
2	Guanidine carbonate	-
3	Li <sub>2</sub> CO <sub>3</sub>	trace
4	Mn <sub>2</sub> CO <sub>3</sub>	trace
5	Cupric carbonate	-
6	Ag <sub>2</sub> CO <sub>3</sub>	trace
7	KHCO <sub>3</sub>	56
<b>8</b>	<b>NaHCO<sub>3</sub></b>	<b>96</b>
9	NH <sub>4</sub> HCO <sub>3</sub>	-
10	K <sub>3</sub> PO <sub>4</sub>	80
11	Na <sub>2</sub> HPO <sub>4</sub>	trace
12	K <sub>2</sub> HPO <sub>4</sub>	51
13	LiO <sup>t</sup> Bu	45

**Table S3: Optimization of other parameters**

entry	[Ru] (mol %)	NaHCO <sub>3</sub> (equiv)	ligands (mol %)	temp °C	yield (%)
1	[Ru( <i>p</i> -cymene)Cl <sub>2</sub> ] <sub>2</sub> (4 mol %)	1.0	P( <i>o</i> -tolyl) <sub>3</sub> (8 mol %)	100	81
2	"	"	PPh <sub>3</sub> (8 mol %)	"	72
3	"	"	-	"	83
4	[Ru( <i>p</i> -cymene)Cl <sub>2</sub> ] <sub>2</sub> (4 mol %)	"	PCy <sub>3</sub> (8 mol %)	"	48 <sup>a</sup>
5	"	0.5	"	"	65
6	"	-	"	100	-
7	"	1.0	"	120	75
8	"	"	"	80	55
9	-	"	-	100	-
10	[Ru( <i>p</i> -cymene)Cl <sub>2</sub> ] <sub>2</sub> (2 mol %)	"	PCy <sub>3</sub> (4 mol %)	"	63
11	[Ru( <i>p</i> -cymene)Cl <sub>2</sub> ] <sub>2</sub> (4 mol %)	"	PCy <sub>3</sub> (8 mol %)	"	71 <sup>b</sup>
12	( <i>p</i> -cymene)RuCl <sub>2</sub> PPh <sub>3</sub> (4 mol %)	"	-	"	45
13	[Ru( <i>p</i> -cymene)Cl <sub>2</sub> ] <sub>2</sub> (4 mol %)	"	Cy <sub>3</sub> PO (8 mol %)	"	94

<sup>a</sup>under N<sub>2</sub> atmosphere, <sup>b</sup>PhSeSePh (1.5 equiv)

### General procedure: Ru(II)-catalyzed direct C–H selenylation of aromatic carboxylic acids

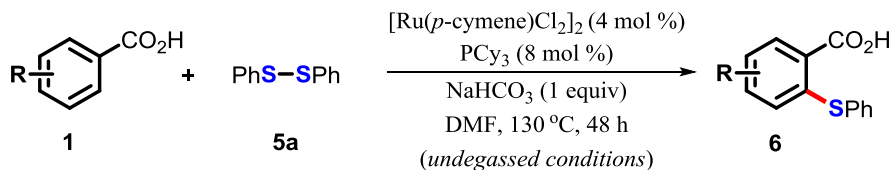


The benzoic acids **1** (0.2 mmol), diselenide derivatives **2** (2 equiv), and [Ru(*p*-cymene)Cl<sub>2</sub>]<sub>2</sub> (4 mol%), PCy<sub>3</sub> (8 mol%), and sodium bicarbonate (1 equiv) were taken in a dried screw cap reaction tube with a magnetic stir bar under open air. Then, dry DMF (1.5mL) was added with a syringe, the tube was capped,

and the resulting mixture was heated at 100 °C for 48 h. After completion of the reaction, it was allowed to cool to room temperature, quenched with AcOH and diluted with brine solution and ethyl acetate. The mixture was extracted with ethyl acetate (10 mL, three times). The organic layer was dried over anhydrous sodium sulfate and concentrated under reduced pressure. The resulting residue was purified by silica gel column chromatography (hexane : ethyl acetate : acetic acid = 90 : 10 :1) to provide pure selenylation products **3-4**.

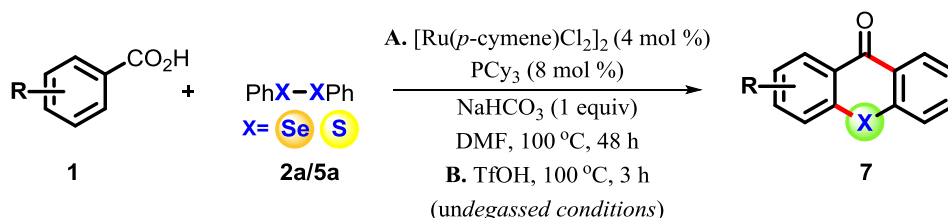
Ru(II)-catalyzed selenylation reaction of **1a** was also performed in 1.1 mmol scale. The benzoic acids **1a** (1.1 mmol, 150 mg, 1 equiv), diphenyl diselenide **2a** (2 equiv), and [Ru(*p*-cymene)Cl<sub>2</sub>]<sub>2</sub> (4 mol%), PCy<sub>3</sub> (8 mol%), and sodium bicarbonate (1 equiv) were taken in a dried screw cap reaction tube with a magnetic stir bar under open air. Then, dry DMF (1.5mL) was added with a syringe, the tube was capped, and the resulting mixture was heated at 100 °C for 48 h. After completion of the reaction, it was allowed to cool to room temperature, quenched with AcOH and diluted with brine solution and ethyl acetate. The mixture was extracted with ethyl acetate (10 mL, three times). The organic layer was dried over anhydrous sodium sulfate and concentrated under reduced pressure. The resulting residue was purified by silica gel column chromatography (hexane : ethyl acetate : acetic acid = 90 : 10 :1) to provide pure selenylation product **3a** as white solid (452 mg, 92% yield).

### General procedure: Ru(II)-catalyzed direct C–H sulfenylation of aromatic carboxylic acids



The benzoic acids **1** (0.2 mmol), diphenyl disulfide **5a** (2 equiv), and [Ru(*p*-cymene)Cl<sub>2</sub>]<sub>2</sub> (4 mol%), PCy<sub>3</sub> (8 mol%), and sodium bicarbonate (1 equiv) were taken in a dried screw cap reaction tube with a magnetic stir bar under open air. Then, DMF (1.5mL) was added with a syringe, the tube was capped, and the resulting mixture was heated at 130 °C for 48 h. After completion of the reaction, it was cooled to room temperature, quenched with AcOH and diluted with brine solution and ethyl acetate. The mixture was extracted with ethyl acetate (10 mL, three times). The organic layer was dried over anhydrous sodium sulfate and concentrated under reduced pressure. The resulting residue was purified by silica gel column chromatography (hexane : ethyl acetate : acetic acid = 90 : 10 :1) to provide pure sulfenylation products **6**.

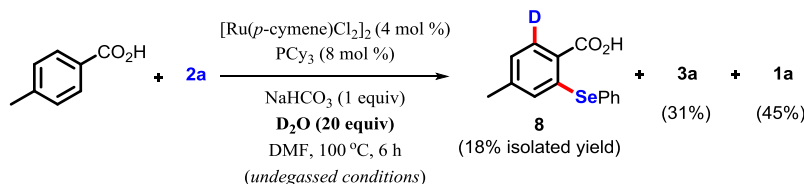
## General procedure: Sequential synthesis of chalcogenoxanthones

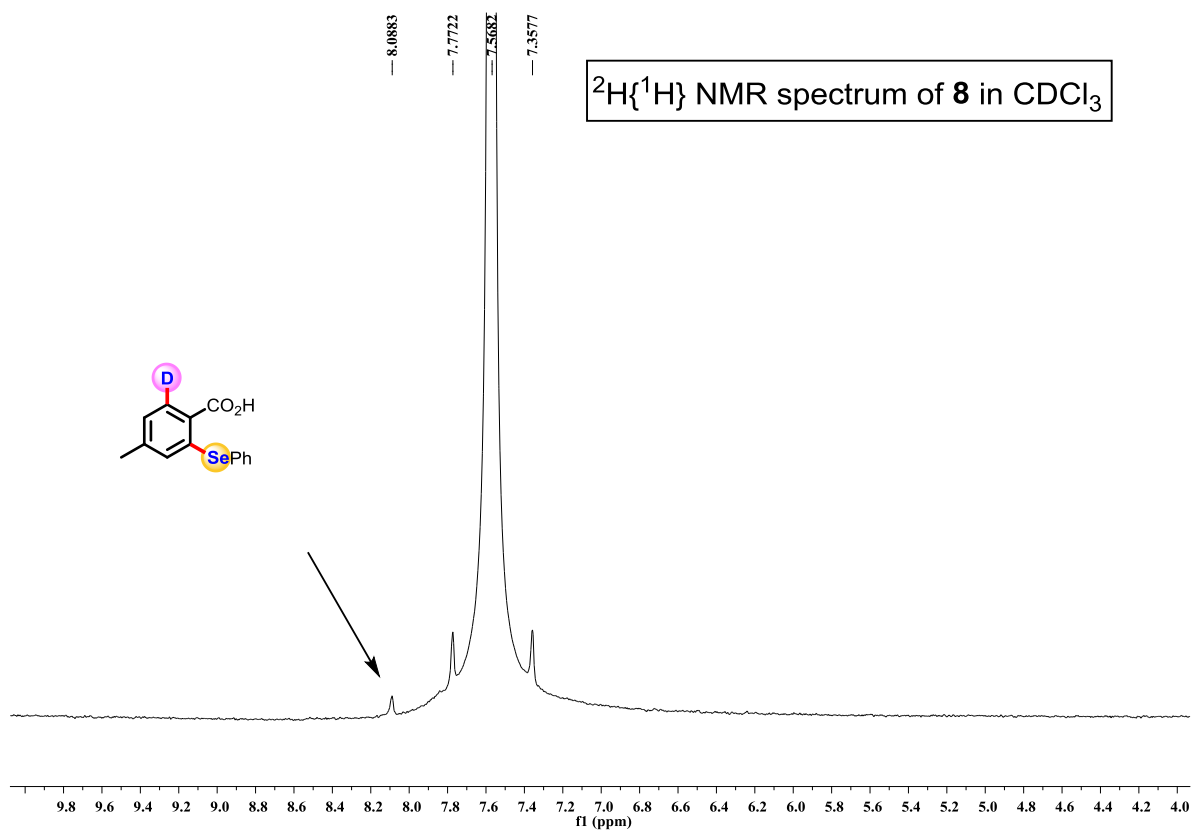
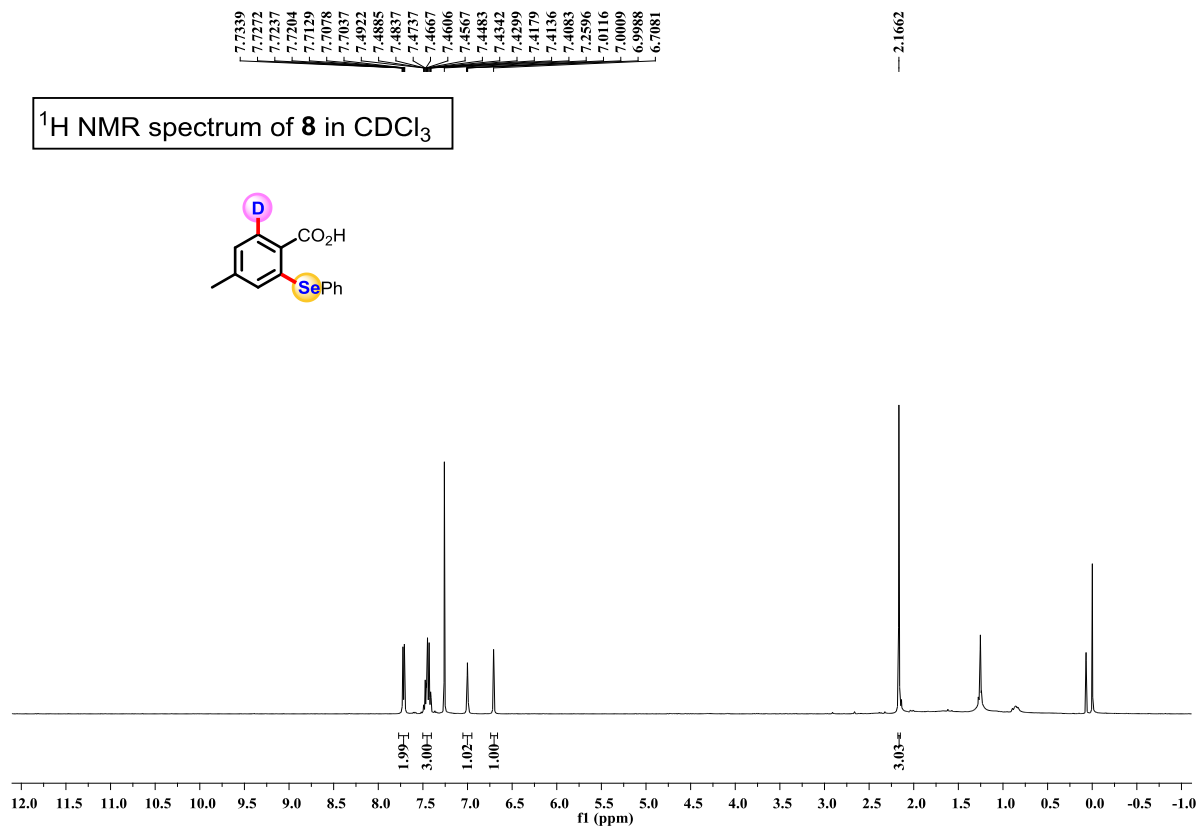


The benzoic acids **1** (0.2 mmol), diphenyl diselenide (**2a**) or disulfide (**5a**) (2 equiv), and  $[\text{Ru}(p\text{-cymene})\text{Cl}_2]_2$  (4 mol%),  $\text{PCy}_3$  (8 mol%), and sodium bicarbonate (1 equiv), were taken in a dried screw cap reaction tube with a magnetic stir bar under open air. Then, DMF (1.5 mL) was added with a syringe, the tube was capped, and the resulting mixture was heated at 100 °C for 48 h (at 130 °C for **5a**). After completion of the reaction, it was cooled to room temperature, quenched with AcOH and diluted with brine solution and ethyl acetate. The mixture was extracted with ethyl acetate (10 mL, three times). The organic layer was dried over anhydrous sodium sulfate and concentrated under reduced pressure. The resulting residue was treated with triflic acid at 100 °C for 3 h. After that, it was allowed to cool to room temperature and quenched with water. The mixture was extracted with dichloromethane (DCM) and the organic layer was dried over anhydrous sodium sulfate and concentrated under reduced pressure. The resulting residue was purified by silica gel column chromatography (hexane : ethyl acetate = 95 : 5) to provide pure chalcogenoxanthones **7**.

### Control experiments:

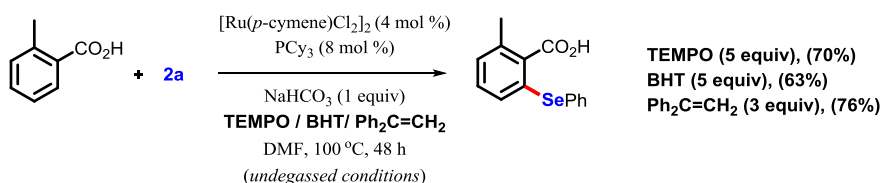
#### 1) Deuterium exchange experiments







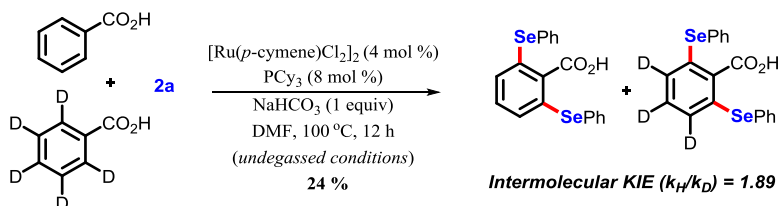
## 2) Radical scavenging experiment:



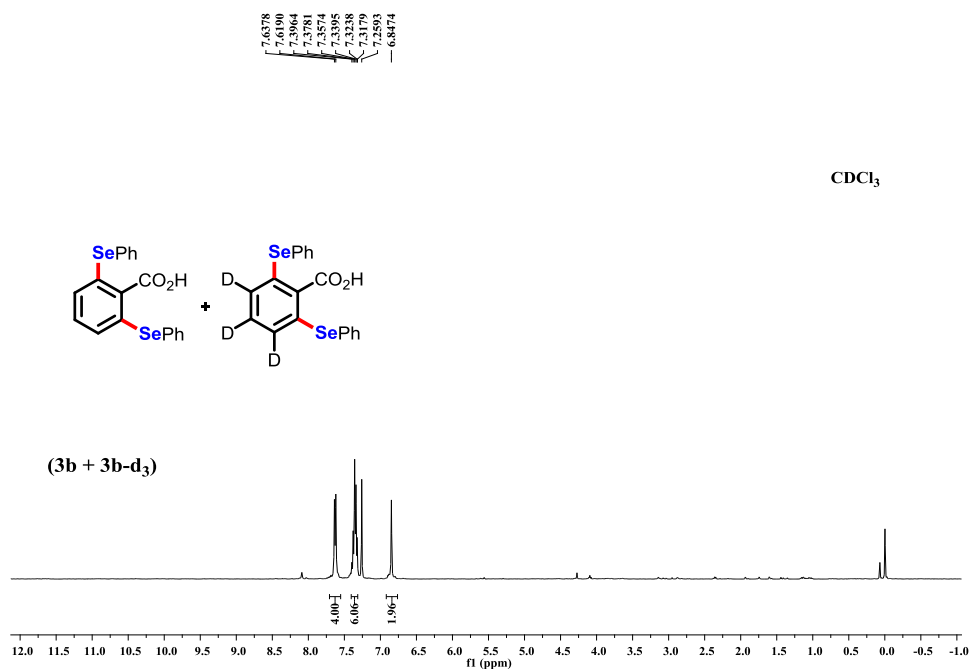
## 3) Kinetic Isotope Effect:

### a) Intermolecular Kinetic Isotopic Experiment:

To demonstrate the intermolecular kinetic isotope effect (KIE), a 1:1 mixture of benzoic acid (**1b**) and the d<sub>5</sub>-benzoic acid (**1b-d<sub>5</sub>**) was subjected to the standard reaction condition for 12 h and the products were isolated by column chromatography. From the <sup>1</sup>H NMR, analysis of the H/D ratio of the benzene ring revealed an intermolecular KIE of  $k_H/k_D = 1.89$ . This result indicates that *ortho* C–H bond breaking may be involved in the rate determining step.

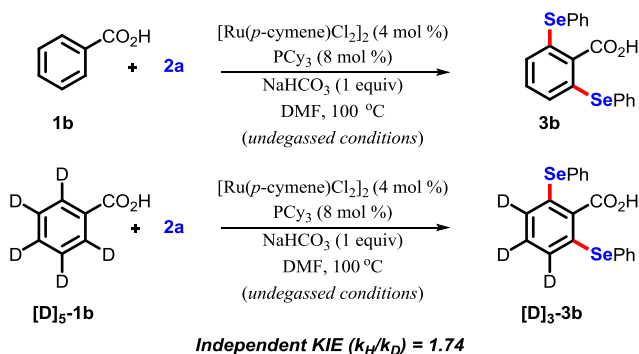


### <sup>1</sup>H NMR spectrum of products of the intermolecular KIE experiment:

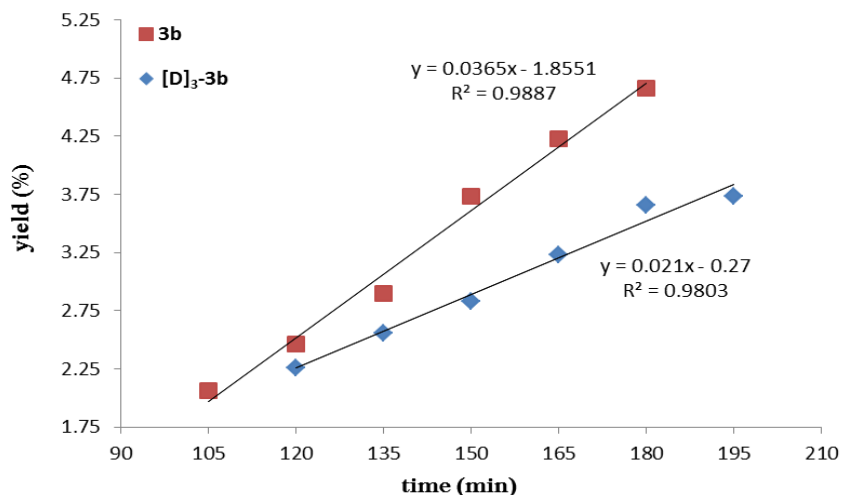


## b) Kinetic Isotope Effect by independent experiments:

Two independent reactions with **1b** and **[D]<sub>5</sub>-1b** under the standard conditions were performed: Following the general procedure of selenylation, **1b** (0.3 mmol) or **[D]<sub>5</sub>-1b** (0.3 mmol), diphenyldiselenide **2a** (2 equiv), [Ru(*p*-cymene)Cl<sub>2</sub>]<sub>2</sub> (4 mol %), PCy<sub>3</sub> (8 mol %), and sodium bicarbonate (1 equiv), were stirred at 100 °C in 3 ml DMF solvent under open atmosphere. After the reaction times indicated below, aliquots of 0.3 mL were taken out of the reaction mixture. The aliquots were acidified, worked up with brine/ethyl acetate mixture. The organic layer was dried over anhydrous sodium sulfate and concentrated under reduced pressure. Yields of products were determined by <sup>1</sup>H NMR spectroscopy using dibromomethane as an internal standard.

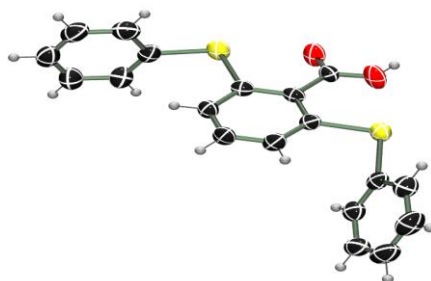


t (min)	105	120	135	150	165	180	195
<b>3b</b> (%)	2.06	2.46	2.90	3.73	4.23	4.66	-
<b>[D]<sub>5</sub>-3b</b> (%)	-	2.26	2.56	2.83	3.23	3.66	3.73



## Crystallographic experimental section:

Crystal structure of compound **3b**: CCDC1534856 (Ellipsoid probability 50%)



(The crystal structure contained two molecules of **3b**)

Table 1. Crystal data and structure refinement for **3b** (CCDC 1534856).

Identification code	993
Empirical formula	C <sub>38</sub> H <sub>28</sub> O <sub>4</sub> Se <sub>4</sub>
Formula weight	864.44
Temperature	296(2) K
Wavelength	0.71073 Å
Crystal system, space group	Triclinic, P-1
Unit cell dimensions	$a = 10.1260(3) \text{ Å}$ $\alpha = 77.9530(10)^\circ$ $b = 12.9083(3) \text{ Å}$ $\beta = 80.0760(10)^\circ$ $c = 13.2893(4) \text{ Å}$ $\gamma = 82.4580(10)^\circ$
Volume	1665.16(8) Å <sup>3</sup>
Z, Calculated density	2, 1.724 Mg/m <sup>3</sup>
Absorption coefficient	4.447 mm <sup>-1</sup>
F(000)	848
Crystal size	0.250 x 0.220 x 0.100 mm
Theta range for data collection	1.585 to 24.998°
Limiting indices	-12 ≤ h ≤ 11, -15 ≤ k ≤ 11, -15 ≤ l ≤ 15

Reflections collected / unique	21799 / 5855 [R(int) = 0.0260]
Completeness to theta = 24.998	99.8 %
Absorption correction	None
Refinement method	Full-matrix least-squares on F <sup>2</sup>
Data / restraints / parameters	5855 / 0 / 424
Goodness-of-fit on F <sup>2</sup>	1.029
Final R indices [I>2sigma(I)]	R1 = 0.0270, wR2 = 0.0550
R indices (all data)	R1 = 0.0425, wR2 = 0.0592
Extinction coefficient	0.00130(14)
Largest diff. peak and hole	0.391 and -0.466 e.Å <sup>-3</sup>

**Crystal structure of compound 3k:** CCDC1534855 (Ellipsoid probability 25%)

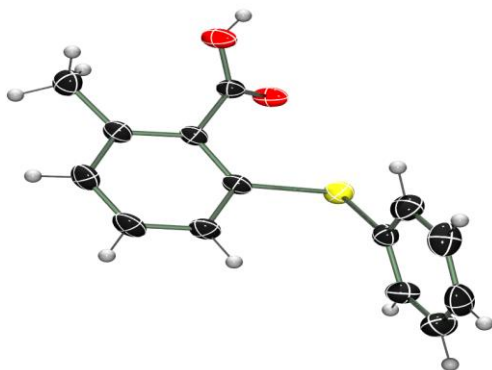
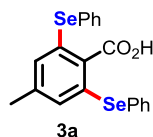


Table 2. Crystal data and structure refinement for **3k** (CCDC 1534855).

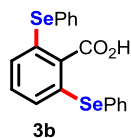
Identification code	914
Empirical formula	C <sub>14</sub> H <sub>12</sub> O <sub>2</sub> Se
Formula weight	291.20
Temperature	296(2) K
Wavelength	0.71073 Å

Crystal system, space group	Monoclinic, P2(1)/c
Unit cell dimensions	$a = 7.4743(6) \text{ \AA}$ $\alpha = 90^\circ$ $b = 17.3909(14) \text{ \AA}$ $\beta = 107.933(3)^\circ$ $c = 10.0665(6) \text{ \AA}$ $\gamma = 90^\circ$
Volume	1244.92(16) $\text{\AA}^3$
Z, Calculated density	4, 1.554 $\text{Mg/m}^3$
Absorption coefficient	3.002 $\text{mm}^{-1}$
F(000)	584
Crystal size	0.220 x 0.120 x 0.100 mm
Theta range for data collection	2.342 to 24.750 deg.
Limiting indices	$-8 \leq h \leq 8$ , $-20 \leq k \leq 20$ , $-11 \leq l \leq 11$
Reflections collected / unique	14211 / 2126 [R(int) = 0.0470]
Completeness to theta = 24.750	99.6 %
Absorption correction	None
Refinement method	Full-matrix least-squares on $F^2$
Data / restraints / parameters	2126 / 0 / 159
Goodness-of-fit on $F^2$	1.101
Final R indices [ $I > 2\sigma(I)$ ]	$R_1 = 0.0339$ , $wR_2 = 0.0798$
R indices (all data)	$R_1 = 0.0608$ , $wR_2 = 0.0891$
Extinction coefficient	n/a
Largest diff. peak and hole	0.253 and -0.293 $\text{e.\AA}^{-3}$

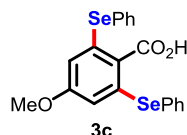
## Spectroscopic data:



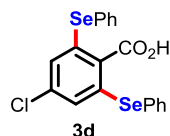
**3a:** White solid, **Yield:** 96% (86 mg);  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ )  $\delta$  9.99 (s, 1H), 7.79–7.52 (m, 4H), 7.50–7.31 (m, 6H), 6.70 (s, 2H), 1.96 (s, 3H);  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ )  $\delta$  172.68, 142.37, 138.79, 136.42, 130.25, 129.76, 129.69, 128.90, 126.82, 21.50; **HRMS** (TOF MS  $\text{ES}^+$ )  $\text{C}_{20}\text{H}_{16}\text{O}_2\text{Se}_2\text{Na}^+$   $m/z$  (%) = 470.9361 ( $[\text{M}+\text{Na}]^+$ , 100%).



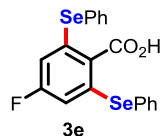
**3b:** White solid, **Yield:** 82% (71 mg);  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ )  $\delta$  7.68–7.61 (m, 4H), 7.46–7.31 (m, 6H), 6.90 (s, 3H);  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ )  $\delta$  172.41, 138.73, 136.49, 131.68, 130.11, 129.83, 128.98, 128.93; **HRMS** (TOF MS  $\text{ES}^+$ )  $\text{C}_{19}\text{H}_{14}\text{O}_2\text{Se}_2\text{Na}^+$   $m/z$  (%) = 456.9248 ( $[\text{M}+\text{Na}]^+$ , 100%).



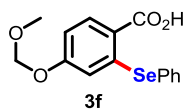
**3c:** White solid, **Yield:** 86% (79 mg);  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ )  $\delta$  7.72–7.65 (m, 4H), 7.49–7.34 (m, 6H), 6.23 (s, 2H), 3.26 (s, 3H);  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ )  $\delta$  165.57, 161.39, 142.73, 137.11, 130.07, 129.76, 129.14, 119.99, 113.68, 54.84; **HRMS** (TOF MS  $\text{ES}^+$ )  $\text{C}_{20}\text{H}_{16}\text{O}_3\text{Se}_2\text{H}^+$   $m/z$  (%) = 464.9539 ( $[\text{M}+\text{H}]^+$ , 100%).



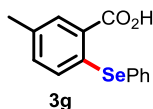
**3d:** White solid, **Yield:** 61% (57 mg);  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ )  $\delta$  7.81–7.56 (m, 4H), 7.50–7.31 (m, 6H), 6.74 (s, 2H);  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ )  $\delta$  171.82, 142.08, 138.91, 136.92, 130.14, 129.63, 129.25, 127.62, 126.02; **HRMS** (TOF MS  $\text{ES}^+$ )  $\text{C}_{19}\text{H}_{13}\text{O}_2\text{ClSe}_2\text{Na}^+$   $m/z$  (%) = 490.8818 ( $[\text{M}+\text{Na}]^+$ , 100%).



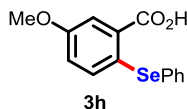
**3e:** White solid, **Yield:** 55% (50 mg);  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ )  $\delta$  7.70–7.64 (m, 4H), 7.55–7.34 (m, 6H), 6.42 (d,  $J$  = 9.2 Hz, 2H);  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ )  $\delta$  171.61, 164.21 (d,  $J$  = 257.9 Hz), 144.80 (d,  $J$  = 8.1 Hz), 137.33, 130.18, 129.76, 129.29, 114.52 (d,  $J$  = 25.2 Hz); **HRMS** (TOF MS  $\text{ES}^+$ )  $\text{C}_{19}\text{H}_{13}\text{O}_2\text{FSe}_2\text{Na}^+$   $m/z$  (%) = 474.9154 ( $[\text{M}+\text{Na}]^+$ , 100%).



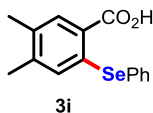
**3f:** White solid, **Yield:** 70% (47 mg); **<sup>1</sup>H NMR** (400 MHz, CDCl<sub>3</sub>) δ 8.13 (d, *J* = 8.8 Hz, 1H), 7.75 – 7.70 (m, 2H), 7.57 – 7.33 (m, 3H), 6.85 (dd, *J* = 8.7, 2.4 Hz, 1H), 6.53 (d, *J* = 2.4 Hz, 1H), 4.97 (s, 2H), 3.33 (s, 3H); **<sup>13</sup>C NMR** (100 MHz, CDCl<sub>3</sub>) δ 171.52, 161.32, 144.07, 137.82, 134.38, 129.96, 129.50, 128.85, 119.87, 116.60, 112.60, 94.16, 56.27; **HRMS** (TOF MS ES<sup>+</sup>) C<sub>15</sub>H<sub>14</sub>O<sub>4</sub>SeNa<sup>+</sup> *m/z* (%) = 360.9936 ([M+Na]<sup>+</sup>, 100%).



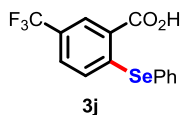
**3g:** White solid, **Yield:** 93% (54 mg); **<sup>1</sup>H NMR** (400 MHz, CDCl<sub>3</sub>) δ 8.03 (d, *J* = 1.4 Hz, 1H), 7.76 – 7.66 (m, 2H), 7.50 – 7.37 (m, 3H), 7.08 (dd, *J* = 8.3, 1.6 Hz, 1H), 6.83 (d, *J* = 8.2 Hz, 1H), 2.32 (s, 3H); **<sup>13</sup>C NMR** (100 MHz, CDCl<sub>3</sub>) δ 172.46, 137.80, 137.65, 134.88, 134.68, 132.94, 129.87, 129.25, 129.05, 126.18, 20.63; **HRMS** (TOF MS ES<sup>+</sup>) C<sub>14</sub>H<sub>12</sub>O<sub>2</sub>SeNa<sup>+</sup> *m/z* (%) = 314.9929 ([M+Na]<sup>+</sup>, 100%).



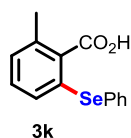
**3h:** White solid, **Yield:** 75% (46 mg); **<sup>1</sup>H NMR** (400 MHz, CDCl<sub>3</sub>) δ 7.70 (dd, *J* = 9.0, 2.5 Hz, 3H), 7.49 – 7.35 (m, 3H), 6.93 – 6.77 (m, 2H), 3.81 (s, 3H); **<sup>13</sup>C NMR** (100 MHz, CDCl<sub>3</sub>) δ 171.72, 157.47, 137.50, 131.39, 130.52, 129.86, 129.22, 129.19, 127.28, 121.19, 116.19, 55.69; **HRMS** (TOF MS ES<sup>+</sup>) C<sub>14</sub>H<sub>12</sub>O<sub>3</sub>SeNa<sup>+</sup> *m/z* (%) = 330.9820 ([M+Na]<sup>+</sup>, 100%).



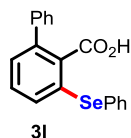
**3i:** White solid, **Yield:** 82% (50 mg); **<sup>1</sup>H NMR** (400 MHz, CDCl<sub>3</sub>) δ 7.97 (s, 1H), 7.77 – 7.67 (m, 2H), 7.52 – 7.37 (m, 3H), 6.67 (s, 1H), 2.23 (s, 3H), 2.08 (s, 3H); **<sup>13</sup>C NMR** (100 MHz, CDCl<sub>3</sub>) δ 172.38, 143.49, 138.05, 137.62, 133.79, 133.38, 130.24, 129.81, 129.19, 129.15, 123.93, 20.30, 19.13; **HRMS** (TOF MS ES<sup>+</sup>) C<sub>15</sub>H<sub>14</sub>O<sub>2</sub>SeNa<sup>+</sup> *m/z* (%) = 329.0065 ([M+Na]<sup>+</sup>, 100%).



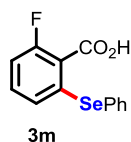
**3j:** White solid, **Yield:** 74% (51 mg); **<sup>1</sup>H NMR** (500 MHz, CDCl<sub>3</sub>) δ 8.43 (d, *J* = 1.4 Hz, 1H), 7.77 – 7.67 (m, 2H), 7.58 – 7.40 (m, 4H), 7.05 (d, *J* = 8.5 Hz, 1H); **<sup>13</sup>C NMR** (125 MHz, CDCl<sub>3</sub>) δ 170.81, 147.40, 137.69, 130.31, 129.98, 129.85, 129.51 (q, *J* = 3.4 Hz), 129.33 (q, *J* = 3.7 Hz), 128.04, 127.61 (q, *J* = 33.5 Hz), 126.36, 123.81 (q, *J* = 271.8 Hz); **HRMS** (TOF MS ES<sup>+</sup>) C<sub>14</sub>H<sub>9</sub>O<sub>2</sub>F<sub>3</sub>SeK<sup>+</sup> *m/z* (%) = 384.9346 ([M+K]<sup>+</sup>, 100%).



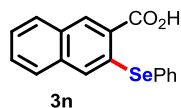
**3k:** White solid, **Yield:** 90% (52 mg);  $^1\text{H NMR}$  (400 MHz,  $\text{CDCl}_3$ )  $\delta$  9.84 (s, 1H), 7.73 – 7.50 (m, 2H), 7.44 – 7.23 (m, 3H), 7.21 – 6.97 (m, 3H), 2.55 (s, 3H);  $^{13}\text{C NMR}$  (100 MHz,  $\text{CDCl}_3$ )  $\delta$  174.27, 138.45, 135.30, 133.95, 132.90, 131.04, 130.75, 130.21, 129.62, 129.32, 128.37, 21.35; **HRMS** (TOF MS  $\text{ES}^+$ )  $\text{C}_{14}\text{H}_{12}\text{O}_2\text{SeNa}^+$   $m/z$  (%) = 314.9900 ( $[\text{M}+\text{Na}]^+$ , 100%).



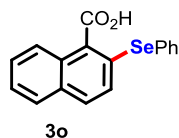
**3l:** White solid, **Yield:** 83% (59 mg);  $^1\text{H NMR}$  (400 MHz,  $\text{CDCl}_3$ )  $\delta$  7.65 – 7.54 (m, 2H), 7.47 – 7.29 (m, 8H), 7.26 – 7.18 (m, 3H);  $^{13}\text{C NMR}$  (100 MHz,  $\text{CDCl}_3$ )  $\delta$  173.62, 141.87, 140.38, 135.31, 133.25, 132.80, 131.41, 130.65, 130.27, 129.68, 128.77, 128.57, 128.48, 128.43, 127.89; **HRMS** (TOF MS  $\text{ES}^+$ )  $\text{C}_{19}\text{H}_{14}\text{O}_2\text{SeK}^+$   $m/z$  (%) = 392.9771 ( $[\text{M}+\text{K}]^+$ , 100%).



**3m:** White solid, **Yield:** 75% (44 mg);  $^1\text{H NMR}$  (400 MHz,  $\text{CDCl}_3$ )  $\delta$  7.73 – 7.62 (m, 2H), 7.49 – 7.32 (m, 3H), 7.18 (td,  $J$  = 8.2, 5.4 Hz, 1H), 6.94 (m, 1H), 6.83 – 6.75 (m, 1H);  $^{13}\text{C NMR}$  (100 MHz,  $\text{CDCl}_3$ )  $\delta$  169.82, 162.77 (d,  $J$  = 260.9 Hz), 141.61, 137.03, 133.37 (d,  $J$  = 9.6 Hz), 129.99, 129.39, 129.23, 126.01 (d,  $J$  = 3.3 Hz), 117.69, 113.58 (d,  $J$  = 23.1 Hz); **HRMS** (TOF MS  $\text{ES}^+$ )  $\text{C}_{13}\text{H}_9\text{O}_2\text{FSeNa}^+$   $m/z$  (%) = 318.9657 ( $[\text{M}+\text{Na}]^+$ , 100%).

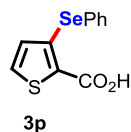


**3n:** White solid, **Yield:** 66% (43 mg);  $^1\text{H NMR}$  (400 MHz,  $\text{DMSO-d}_6$ )  $\delta$  8.67 (s, 1H), 8.06 – 8.00 (m, 1H), 7.75 – 7.69 (m, 2H), 7.58 – 7.41 (m, 6H), 7.21 (s, 1H);  $^{13}\text{C NMR}$  (125 MHz,  $\text{DMSO-d}_6$ )  $\delta$  168.18, 137.06, 134.72, 133.83, 132.30, 130.21, 130.11, 129.44, 129.15, 129.08, 129.03, 127.04, 126.70, 126.40, 126.24; **HRMS** (TOF MS  $\text{ES}^+$ )  $\text{C}_{17}\text{H}_{12}\text{O}_2\text{SeNa}^+$   $m/z$  (%) = 350.9906 ( $[\text{M}+\text{Na}]^+$ , 100%).

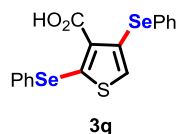


**3o:** White solid, **Yield:** 78% (51 mg);  $^1\text{H NMR}$  (400 MHz,  $\text{CDCl}_3$ )  $\delta$  8.33 (d,  $J$  = 8.5 Hz, 1H), 7.77 (d,  $J$  = 8.1 Hz, 1H), 7.69 – 7.59 (m, 3H), 7.56 (t,  $J$  = 7.6 Hz, 1H), 7.48 (t,  $J$  = 7.4 Hz, 1H), 7.39 – 7.20 (m, 4H);  $^{13}\text{C NMR}$  (100 MHz,  $\text{CDCl}_3$ )  $\delta$  172.42, 135.35, 133.06, 132.08, 131.10, 130.65, 129.64, 129.02, 128.44, 128.32, 127.83, 126.30, 125.42; **HRMS** (TOF MS  $\text{ES}^+$ )  $\text{C}_{17}\text{H}_{12}\text{O}_2\text{SeNa}^+$   $m/z$  (%) = 350.9912 ( $[\text{M}+\text{Na}]^+$ , 100%).

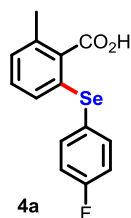




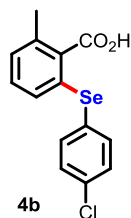
**3p:** White solid, **Yield:** 88% (50 mg); **<sup>1</sup>H NMR** (400 MHz, DMSO-*d*<sub>6</sub>) δ 7.95 – 7.60 (m, 3H), 7.60 – 7.19 (m, 3H), 6.24 (d, *J* = 5.2 Hz, 1H); **<sup>13</sup>C NMR** (100 MHz, DMSO-*d*<sub>6</sub>) δ 163.34, 138.57, 136.43, 132.37, 129.88, 129.40, 128.58, 127.93, 123.27; **HRMS** (TOF MS ES<sup>+</sup>) C<sub>11</sub>H<sub>8</sub>O<sub>2</sub>SSeNa<sup>+</sup> *m/z* (%) = 306.9330 ([M+Na]<sup>+</sup>, 100%).



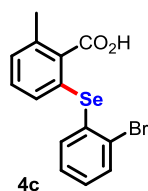
**3q:** White solid, **Yield:** 69% (61 mg); **<sup>1</sup>H NMR** (400 MHz, CDCl<sub>3</sub>) δ 7.77 (d, *J* = 7.1 Hz, 2H), 7.69 (d, *J* = 6.2 Hz, 2H), 7.53 – 7.31 (m, 6H), 6.14 (s, 1H); **<sup>13</sup>C NMR** (100 MHz, CDCl<sub>3</sub>) δ 168.21, 150.70, 137.35, 136.73, 131.52, 130.28, 130.00, 129.84, 129.14, 128.57, 127.92, 125.55, 120.08; **HRMS** (TOF MS ES<sup>+</sup>) C<sub>17</sub>H<sub>12</sub>O<sub>2</sub>SSe<sub>2</sub>Na<sup>+</sup> *m/z* (%) = 462.8777 ([M+Na]<sup>+</sup>, 100%).



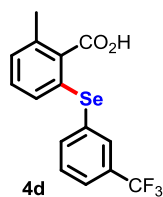
**4a:** White solid, **Yield:** 72% (45 mg); **<sup>1</sup>H NMR** (500 MHz, CDCl<sub>3</sub>) δ 7.65 – 7.55 (m, 2H), 7.18 – 7.08 (m, 2H), 7.07 – 6.97 (m, 3H), 2.54 (s, 3H); **<sup>13</sup>C NMR** (125 MHz, CDCl<sub>3</sub>) δ 173.42, 163.22 (d, *J* = 7.7 Hz), 138.84, 137.86, 137.80, 134.67, 132.19, 131.20, 129.49 (d, *J* = 29.2 Hz), 125.25 (d, *J* = 3.5 Hz), 116.93 (d, *J* = 21.5 Hz), 21.51; **HRMS** (TOF MS ES<sup>+</sup>) C<sub>14</sub>H<sub>11</sub>O<sub>2</sub>FSeNa<sup>+</sup> *m/z* (%) = 332.9819 ([M+Na]<sup>+</sup>, 100%).



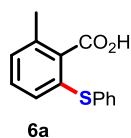
**4b:** White solid, **Yield:** 70% (46 mg); **<sup>1</sup>H NMR** (400 MHz, DMSO-*d*<sub>6</sub>) δ 7.45 (dd, *J* = 6.6, 1.6 Hz, 2H), 7.34 (d, *J* = 6.7 Hz, 2H), 7.14 (s, 2H), 7.05 (s, 1H), 2.36 (s, 3H); **<sup>13</sup>C NMR** (100 MHz, DMSO-*d*<sub>6</sub>) δ 169.49, 136.83, 135.96, 135.44, 133.17, 130.25, 129.98, 129.48, 129.40, 129.30, 20.03; **HRMS** (TOF MS ES<sup>+</sup>) C<sub>14</sub>H<sub>11</sub>O<sub>2</sub>ClSeNa<sup>+</sup> *m/z* (%) = 348.9501 ([M+Na]<sup>+</sup>, 100%).



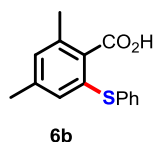
**4c:** White solid, **Yield:** 64% (47 mg); **<sup>1</sup>H NMR** (400 MHz, CDCl<sub>3</sub>) δ 7.56 (dd, *J* = 7.7, 1.1 Hz, 1H), 7.31 – 7.26 (m, 1H), 7.25 – 7.19 (m, 2H), 7.20 – 7.06 (m, 3H), 2.51 (s, 3H); **<sup>13</sup>C NMR** (100 MHz, CDCl<sub>3</sub>) δ 172.63, 137.97, 136.09, 135.32, 133.87, 133.04, 132.69, 131.09, 130.64, 129.67, 128.69, 128.19, 126.22, 20.97; **HRMS** (TOF MS ES<sup>+</sup>) C<sub>14</sub>H<sub>11</sub>O<sub>2</sub>BrSeNa<sup>+</sup> *m/z* (%) = 392.8985 ([M+Na]<sup>+</sup>, 100%).



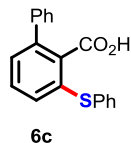
**4d:** White solid, **Yield:** 68% (49 mg); **<sup>1</sup>H NMR** (400 MHz, CDCl<sub>3</sub>) δ 7.79 (s, 1H), 7.67 (d, *J* = 7.0 Hz, 1H), 7.54 (d, *J* = 7.7 Hz, 1H), 7.38 (t, *J* = 7.4 Hz, 1H), 7.20 – 7.05 (m, 3H), 2.51 (s, 3H); **<sup>13</sup>C NMR** (100 MHz, CDCl<sub>3</sub>) δ 173.54, 138.43(m), 137.79, 132.49, 131.92, 131.59, 131.12, 130.96 (q, *J* = 3.61 Hz), 130.88, 130.02, 129.84, 124.8 (q, *J* = 3.56 Hz), 123.77 (q, *J* = 271.27 Hz), 21.19; **HRMS** (TOF MS ES<sup>+</sup>) C<sub>15</sub>H<sub>11</sub>O<sub>2</sub>F<sub>3</sub>SeNa<sup>+</sup> *m/z* (%) = 382.9787([M+Na]<sup>+</sup>, 100%).



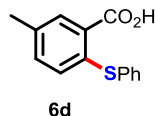
**6a:** White solid, **Yield:** 72% (35 mg); **<sup>1</sup>H NMR** (400 MHz, CDCl<sub>3</sub>) δ 7.39 – 7.35 (m, 2H), 7.32 – 7.24 (m, 3H), 7.18 (d, *J* = 7.7 Hz, 1H), 7.13 – 7.08 (m, 2H), 2.44 (s, 3H); **<sup>13</sup>C NMR** (100 MHz, CDCl<sub>3</sub>) δ 172.49, 136.34, 135.82, 133.81, 131.72, 130.16, 130.12, 129.36, 129.31, 127.42, 20.16; **HRMS** (TOF MS ES<sup>+</sup>) C<sub>14</sub>H<sub>12</sub>O<sub>2</sub>SNa<sup>+</sup> *m/z* (%) = 267.0479 ([M+Na]<sup>+</sup>, 100%).



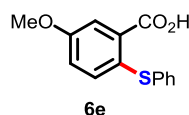
**6b:** White solid, **Yield:** 75% (39 mg); **<sup>1</sup>H NMR** (400 MHz, CDCl<sub>3</sub>) δ 7.36 (dt, *J* = 3.3, 2.0 Hz, 2H), 7.32 – 7.20 (m, 3H), 6.94 (s, 2H), 2.42 (s, 3H), 2.22 (s, 3H); **<sup>13</sup>C NMR** (100 MHz, CDCl<sub>3</sub>) δ 173.19, 140.50, 136.56, 136.01, 133.88, 132.72, 131.60, 130.70, 130.46, 129.27, 127.31, 21.27, 20.26; **HRMS** (TOF MS ES<sup>+</sup>) C<sub>15</sub>H<sub>14</sub>O<sub>2</sub>SK<sup>+</sup> *m/z* (%) = 297.0320 ([M+K]<sup>+</sup>, 100%).



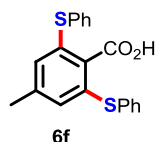
**6c:** White solid, **Yield:** 64% (39 mg);  $^1\text{H NMR}$  (400 MHz,  $\text{CDCl}_3$ )  $\delta$  7.45 – 7.39 (m, 4H), 7.38 – 7.27 (m, 6H), 7.25 – 7.18 (m, 3H);  $^{13}\text{C NMR}$  (100 MHz,  $\text{CDCl}_3$ )  $\delta$  173.41, 141.02, 139.86, 135.05, 134.91, 132.42, 130.88, 130.32, 129.46, 128.82, 128.62, 128.49, 128.19, 127.99, 127.87; **HRMS** (TOF MS  $\text{ES}^+$ )  $\text{C}_{19}\text{H}_{14}\text{O}_2\text{SNa}^+$   $m/z$  (%) = 329.0613 ( $[\text{M}+\text{Na}]^+$ , 100%).



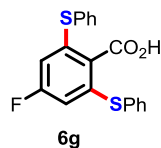
**6d:** White solid, **Yield:** 68% (33 mg);  $^1\text{H NMR}$  (400 MHz,  $\text{CDCl}_3$ )  $\delta$  7.94 (d,  $J$  = 1.5 Hz, 1H), 7.58 – 7.53 (m, 2H), 7.47 – 7.37 (m, 3H), 7.11 (dd,  $J$  = 8.3, 1.8 Hz, 1H), 6.75 (d,  $J$  = 8.3 Hz, 1H), 2.32 (s, 3H);  $^{13}\text{C NMR}$  (100 MHz,  $\text{CDCl}_3$ )  $\delta$  171.16, 140.58, 135.49, 134.60, 134.30, 132.88, 132.60, 129.86, 129.14, 128.00, 125.72, 20.68; **HRMS** (TOF MS  $\text{ES}^+$ )  $\text{C}_{14}\text{H}_{12}\text{O}_2\text{SNa}^+$   $m/z$  (%) = 267.0465 ( $[\text{M}+\text{Na}]^+$ , 100%).



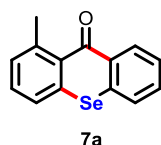
**6e:** White solid, **Yield:** 66% (34 mg);  $^1\text{H NMR}$  (400 MHz,  $\text{CDCl}_3$ )  $\delta$  7.63 (d,  $J$  = 1.9 Hz, 1H), 7.55 – 7.46 (m, 2H), 7.43 – 7.33 (m, 3H), 6.96 – 6.77 (m, 2H), 3.81 (s, 3H);  $^{13}\text{C NMR}$  (125 MHz,  $\text{CDCl}_3$ )  $\delta$  171.08, 157.30, 134.53, 133.71, 130.50, 129.75, 129.16, 128.84, 128.72, 120.62, 115.96, 55.72; **HRMS** (TOF MS  $\text{ES}^+$ )  $\text{C}_{14}\text{H}_{12}\text{O}_3\text{SNa}^+$   $m/z$  (%) = 283.0430 ( $[\text{M}+\text{Na}]^+$ , 100%).



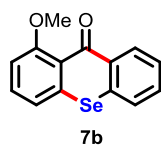
**6f:** White solid, **Yield:** 60% (42 mg);  $^1\text{H NMR}$  (400 MHz,  $\text{CDCl}_3$ )  $\delta$  7.42 – 7.36 (m, 4H), 7.33 – 7.23 (m, 6H), 6.94 (s, 2H), 2.12 (s, 3H);  $^{13}\text{C NMR}$  (125 MHz,  $\text{CDCl}_3$ )  $\delta$  171.53, 141.00, 135.82, 135.15, 132.14, 131.79, 129.42, 129.32, 127.74, 21.32; **HRMS** (TOF MS  $\text{ES}^+$ )  $\text{C}_{20}\text{H}_{16}\text{O}_2\text{S}_2\text{Na}^+$   $m/z$  (%) = 375.0500 ( $[\text{M}+\text{Na}]^+$ , 100%).



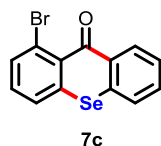
**6g:** White solid, **Yield:** 54% (39 mg);  $^1\text{H NMR}$  (400 MHz,  $\text{CDCl}_3$ )  $\delta$  7.49 (m, 4H), 7.39 – 7.33 (m, 6H), 6.49 (d,  $J$  = 9.0 Hz, 2H);  $^{13}\text{C NMR}$  (125 MHz,  $\text{CDCl}_3$ )  $\delta$  163.30 (d,  $J$  = 254.7 Hz), 141.90 (d,  $J$  = 7.5 Hz), 134.36, 132.69, 129.91, 129.63, 129.19, 114.35 (d,  $J$  = 24.8 Hz); **HRMS** (TOF MS  $\text{ES}^+$ )  $\text{C}_{19}\text{H}_{13}\text{O}_2\text{FS}_2\text{Na}^+$   $m/z$  (%) = 379.0231 ( $[\text{M}+\text{Na}]^+$ , 100%).



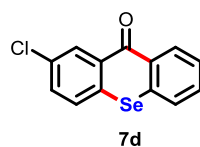
**7a:** Yellow solid, **Yield:** 86% (47 mg); **<sup>1</sup>H NMR** (400 MHz, CDCl<sub>3</sub>) δ 8.43 – 8.32 (m, 1H), 7.57 (dd, *J* = 7.7, 1.0 Hz, 1H), 7.51 – 7.45 (m, *J* = 7.4, 1.7 Hz, 2H), 7.44 – 7.39 (m, 1H), 7.33 (t, *J* = 7.6 Hz, 1H), 7.23 (dd, *J* = 7.4, 0.5 Hz, 1H), 2.80 (s, 3H); **<sup>13</sup>C NMR** (100 MHz, CDCl<sub>3</sub>) δ 185.83, 144.21, 135.53, 134.32, 133.25, 131.66, 131.04, 130.99, 130.92, 130.49, 127.71, 126.77, 126.58, 24.44; **HRMS** (TOF MS ES<sup>+</sup>) C<sub>14</sub>H<sub>10</sub>OSeNa<sup>+</sup> *m/z* (%) = 296.9764 ([M+Na]<sup>+</sup>, 100%).



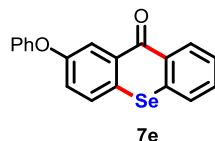
**7b:** Yellow solid, **Yield:** 60% (35 mg); **<sup>1</sup>H NMR** (400 MHz, CDCl<sub>3</sub>) δ 8.41 – 8.30 (m, 1H), 7.59 – 7.51 (m, 1H), 7.49 – 7.40 (m, 2H), 7.40 – 7.36 (m, 1H), 7.19 (dd, *J* = 7.9, 0.9 Hz, 1H), 6.94 (d, *J* = 8.3 Hz, 1H), 3.98 (s, 3H); **<sup>13</sup>C NMR** (100 MHz, CDCl<sub>3</sub>) δ 184.06, 162.26, 136.72, 134.90, 132.61, 132.52, 131.52, 130.88, 127.49, 126.90, 121.64, 120.55, 110.11, 56.45; **HRMS** (TOF MS ES<sup>+</sup>) C<sub>14</sub>H<sub>10</sub>O<sub>2</sub>SeH<sup>+</sup> *m/z* (%) = 290.9926 ([M+H]<sup>+</sup>, 100%).



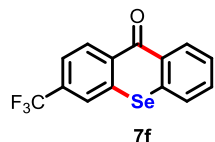
**7c:** Yellow solid, **Yield:** 65% (44 mg); **<sup>1</sup>H NMR** (400 MHz, CDCl<sub>3</sub>) δ 8.32 (dd, *J* = 7.8, 1.7 Hz, 1H), 7.71 (dd, *J* = 7.8, 1.2 Hz, 1H), 7.60 – 7.53 (m, 2H), 7.49 (td, *J* = 7.5, 1.6 Hz, 1H), 7.46 – 7.40 (m, 1H), 7.23 (dd, *J* = 14.9, 7.1 Hz, 1H); **<sup>13</sup>C NMR** (100 MHz, CDCl<sub>3</sub>) δ 183.86, 136.45, 134.37, 133.68, 132.23, 132.00, 131.46, 130.95, 129.78, 127.96, 127.63, 127.21, 124.45; **HRMS** (TOF MS ES<sup>+</sup>) C<sub>13</sub>H<sub>7</sub>OBrSeNa<sup>+</sup> *m/z* (%) = 360.8763 ([M+Na]<sup>+</sup>, 100%).



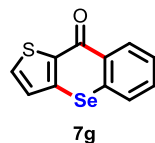
**7d:** Yellow solid, **Yield:** 67% (39 mg); **<sup>1</sup>H NMR** (400 MHz, CDCl<sub>3</sub>) δ 8.71 – 8.55 (m, 2H), 7.67 – 7.58 (m, 1H), 7.58 – 7.51 (m, 2H), 7.50 – 7.41 (m, 2H); **<sup>13</sup>C NMR** (100 MHz, CDCl<sub>3</sub>) δ 181.04, 134.89, 133.24, 133.17, 132.61, 132.47, 132.06, 131.67, 131.03, 130.52, 129.74, 128.42, 127.16; **HRMS** (TOF MS ES<sup>+</sup>) C<sub>13</sub>H<sub>7</sub>OClSeNa<sup>+</sup> *m/z* (%) = 316.9222 ([M+Na]<sup>+</sup>, 100%).



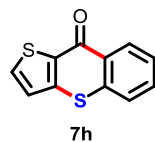
**7e:** Yellow solid, **Yield:** 62% (44 mg); **<sup>1</sup>H NMR** (400 MHz, CDCl<sub>3</sub>) δ 8.62 (d, *J* = 8.1, 1.6 Hz, 1H), 8.26 (d, *J* = 2.8 Hz, 1H), 7.70 – 7.59 (m, 2H), 7.57 – 7.51 (m, 1H), 7.49 – 7.43 (m, 1H), 7.41 – 7.34 (m, 2H), 7.31 (dd, *J* = 8.6, 2.9 Hz, 1H), 7.19 – 7.13 (m, 1H), 7.09 – 7.03 (m, 2H); **<sup>13</sup>C NMR** (100 MHz, CDCl<sub>3</sub>) δ 181.63, 156.83, 156.64, 135.23, 132.33, 132.26, 131.64, 130.50, 130.16, 129.89, 128.65, 128.46, 126.94, 124.29, 124.15, 119.89, 119.39; **HRMS** (TOF MS ES<sup>+</sup>) C<sub>19</sub>H<sub>12</sub>O<sub>2</sub>SeH<sup>+</sup> *m/z* (%) = 353.0100 ([M+H]<sup>+</sup>, 100%).



**7f:** Yellow solid, **Yield:** 58% (38 mg); **<sup>1</sup>H NMR** (400 MHz, CDCl<sub>3</sub>) δ 8.91 (d, *J* = 1.8 Hz, 1H), 8.65 (dd, *J* = 8.1, 1.7 Hz, 1H), 7.80 – 7.70 (m, 2H), 7.64 (dd, *J* = 7.9, 1.6 Hz, 1H), 7.61 – 7.55 (m, 1H), 7.54 – 7.46 (m, 1H); **<sup>13</sup>C NMR** (100 MHz, CDCl<sub>3</sub>) δ 181.19, 139.51, 134.56, 132.92, 131.74, 131.11, 130.64, 129.40 (q, *J* = 33.8 Hz), 129.30, 128.65 (q, *J* = 3.9 Hz), 128.42, 128.08 (q, *J* = 3.3 Hz), 127.52, 123.82 (q, *J* = 272.0 Hz); **HRMS** (TOF MS ES<sup>+</sup>) C<sub>14</sub>H<sub>7</sub>OF<sub>3</sub>SeH<sup>+</sup> *m/z* (%) = 328.9662 ([M+H]<sup>+</sup>, 100%).



**7g:** Yellowish solid, **Yield:** 81% (43 mg); **<sup>1</sup>H NMR** (500 MHz, CDCl<sub>3</sub>) δ 8.80 – 8.69 (m, 1H), 7.90 (d, *J* = 5.2 Hz, 1H), 7.73 (dd, *J* = 7.1, 2.0 Hz, 1H), 7.64 – 7.52 (m, 2H), 7.33 (d, *J* = 5.2 Hz, 1H); **<sup>13</sup>C NMR** (100 MHz, CDCl<sub>3</sub>) δ 176.58, 135.51, 134.91, 134.47, 134.31, 131.78, 130.72, 130.59, 128.91, 127.38, 127.34; **HRMS** (TOF MS ES<sup>+</sup>) C<sub>11</sub>H<sub>6</sub>OSSeNa<sup>+</sup> *m/z* (%) = 288.9219 ([M+Na]<sup>+</sup>, 100%).



**7h:** White solid, **Yield:** 70% (31 mg); **<sup>1</sup>H NMR** (400 MHz, CDCl<sub>3</sub>) δ 8.68 (dd, *J* = 8.4, 1.5 Hz, 1H), 7.89 (d, *J* = 5.3 Hz, 1H), 7.72 – 7.62 (m, 2H), 7.61 – 7.54 (m, 1H), 7.29 (d, *J* = 5.3 Hz, 1H); **<sup>13</sup>C NMR** (125 MHz, CDCl<sub>3</sub>) δ 175.25, 138.73, 137.29, 134.77, 133.62, 131.80, 129.14, 127.08, 126.77, 125.53; **HRMS** (TOF MS ES<sup>+</sup>) C<sub>11</sub>H<sub>6</sub>OS<sub>2</sub>Na<sup>+</sup> *m/z* (%) = 240.9774 ([M+Na]<sup>+</sup>, 100%).

# NMR Spectra

