

Halogen-Bond-Promoted Photoactivation of Perfluoroalkyl Iodides:

A Photochemical Protocol for Perfluoroalkylation Reactions

Yaxin Wang,^a Junhua Wang,^a Guo-Xing Li,^a Gang He,^a and Gong Chen^{*a,b}

^a State Key Laboratory and Institute of Elemento-Organic Chemistry, Collaborative Innovation Center of Chemical Science and Engineering (Tianjin), Nankai University, Tianjin 300071, China.

^b Department of Chemistry, The Pennsylvania State University, University Park, Pennsylvania 16802, United States.

Supporting Information

Content

| | |
|---|-----|
| 1. Reagents..... | S2 |
| 2. Instruments..... | S2 |
| 3. Synthesis of substrates..... | S3 |
| 4. Evaluation of synthesis phenanthridine 3 from 2-isocyanobiphenyl and C ₄ F ₉ -I via a radical cascade process..... | S15 |
| 5. Evaluation of amine promoters for synthesis of 3 under CFL irradiation..... | S16 |
| 6. General procedures and substrate scope of phenanthridine synthesis..... | S18 |
| 7. General procedures and substrate scope for addition of perfluorobutyl iodide to alkenes and alkynes..... | S25 |
| 8. General procedure and substrate scope for C-H perfluorobutylation of electron-rich arene and heteroarene..... | S30 |
| 9. Titration experiment of C ₁₀ F ₂₁ I with TEEDA..... | S36 |
| 10. Determination of binding stoichiometry of halogen bond complex..... | S37 |
| 11. Determination of the association constant (K_a) | S41 |
| 12. Evaluation of different light source..... | S47 |
| 13. References..... | S49 |
| 14. Spectra..... | S50 |

1. Reagents

All commercial materials were used as received unless otherwise noted. Perfluoroalkyl iodides (CF_3I , $\text{C}_3\text{F}_7\text{I}$, $\text{C}_4\text{F}_9\text{I}$, $\text{C}_6\text{F}_{13}\text{I}$, $\text{C}_8\text{F}_{17}\text{I}$, $\text{C}_{10}\text{F}_{21}\text{I}$) and $\text{BrCF}_2\text{COOCH}_3$ were purchased from J&K Chemical, Energy Chemical or TCI. *N,N,N',N'*-tetraethylenediamine (TEEDA) was purchased from Energy Chemical. Dry solvents (THF (99.5%), 2-MeTHF (99.0%), THP (99.0%), dioxane (99.5%), Et_2O (99.0%), CH_2Cl_2 (99.0%), $\text{ClCH}_2\text{CH}_2\text{Cl}$ (98.9%), CHCl_3 (99.0%), MeCN (99.9%)) and deuterated solvents (CDCl_3 , CD_3OD) were purchased from J&K Chemical. TLC were performed on silica gel Huanghai HSGF254 plates and visualization of the developed chromatogram was performed by fluorescence quenching ($\lambda_{\text{max}} = 254 \text{ nm}$). Flash chromatography was performed using Silica gel (200-300 mesh) purchased from Qingdao Haiyang Chemical Co., China.

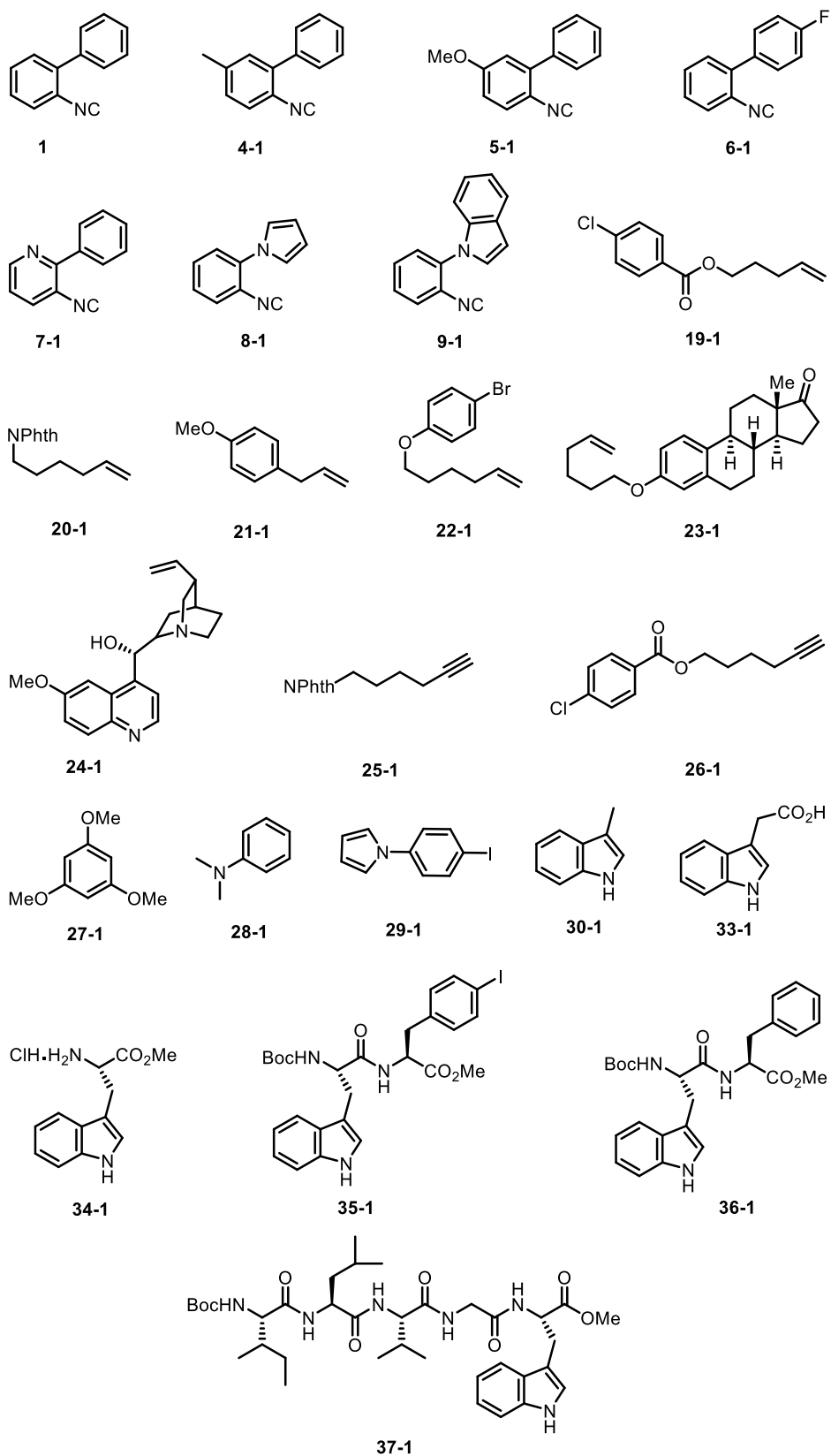
2. Instruments

NMR spectra were recorded on Bruker AVANCE AV 400 instruments and all NMR experiments were reported in units, parts per million (ppm), using residual solvent peaks (CDCl_3 ($\delta = 7.26 \text{ ppm}$) or CD_3OD ($\delta = 3.31 \text{ ppm}$) for ^1H NMR, chloroform ($\delta = 77.16 \text{ ppm}$) CD_3OD ($\delta = 49.00 \text{ ppm}$) for ^{13}C NMR) as internal reference. Multiplicities are recorded as: s = singlet, d = doublet, t = triplet, dd = doublet of doublets, m = multiplet. HRMS were obtained on Varian 7.0T FTMS instrument. UV-vis data were recorded on a HITACHI U-3900 spectrophotometer.

All reactions were carried out in a 4 mL glass vial (Thermo SCIENTIFIC National B7999-2, made from superior quality 33 expansion borosilicate clear glass), sealed with PTEF cap on bench top.

Lights: PHILIPS TORNADO 25W CFL, Cnlight 220V/25W UV (254nm), Cnlight 220V/25W UV (365nm), Cnlight LED lights 25 W (red LED, yellow LED, green LED, blue LED, purple LED, white LED) were used in the perfluoroalkylation reactions

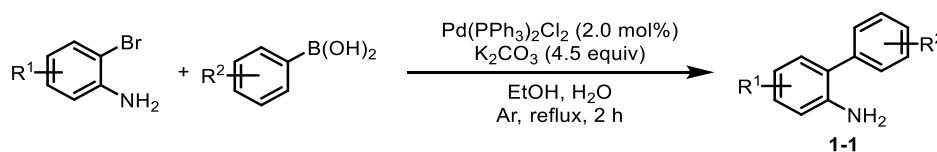
3. Synthesis of substrates



Scheme S1. List of all substrates used in this study

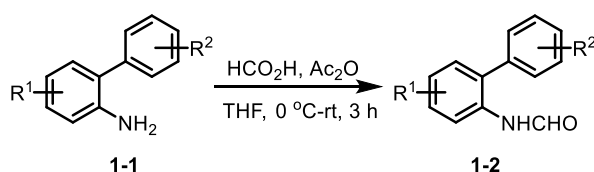
Compounds **21-1**, **24-1**, **27-1**, **28-1**, **29-1**, **30-1**, **33-1**, and **34-1** were commercial available and used as received. Compounds **1¹**, **4-1¹**, **5-1¹**, **6-1¹**, **7-1²**, **8-1³**, **9-1³**, **19-1⁴**, **20-1⁶**, **25-1⁷**, **26-1⁵** and **36-1⁸** were known compounds and were synthesized following the reported procedures. Compounds **22-1**, **23-1**, **35-1**, **37-1** were new compounds.

3.1 General procedure for synthesis of 2-Isocyanobiaryl substrates¹



Scheme S2

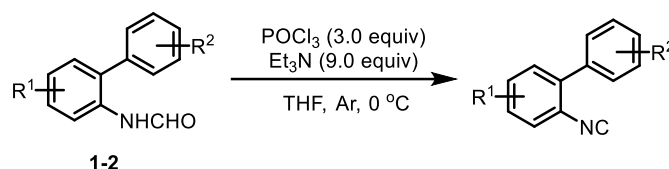
2-Bromoaniline (10.0 mmol, 1.0 equiv), aryl boronic acid (12.0 mmol, 1.2 equiv), K₂CO₃ (6.2 g, 45.0 mmol, 4.5 equiv) and Pd (PPh₃)₂Cl₂ (140.4 mg, 0.2 mmol, 0.02 equiv) were added to a mixture of EtOH (20 mL) and water (20 mL) at room temperature. The mixture was heated to reflux for 2 h under Ar. After been cooled to room temperature, the mixture was extracted with EtOAc. The combined organic layer was washed with brine, dried over anhydrous Na₂SO₄ and concentrated *in vacuo*. The residue was purified by chromatography on silica gel (eluted with hexane/acetone (v/v 40:1)) to afford compound **1-1**.



Scheme S3

Acetic formic anhydride (18.0 mmol, 6.0 equiv), which was newly prepared from the reaction of acetic anhydride (1.7 mL, 18.0 mmol) with formic acid (0.8 mL, 20.0 mmol) at 55 °C for 2 h, was added dropwise to a mixture of **1-1** (3.0 mmol, 1.0 equiv) in THF

(6.0 mL) at 0 °C. After the addition was completed, the mixture was warmed to room temperature and stirred for 3 h. Then, the reaction was quenched with Sat. NaHCO₃ and extracted with EtOAc. The combined organic layer was washed with brine, dried over anhydrous Na₂SO₄ and concentrated *in vacuo* to give compound **1-2**. These compound were used for the subsequent dehydration reaction without further purification.

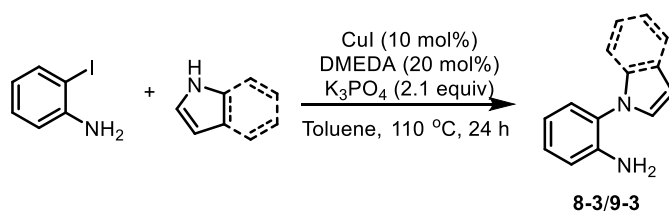


Scheme S4

POCl₃ (0.8 mL, 9.0 mmol, 3.0 equiv) was added via syring pump to a mixture of Et₃N (3.8 mL, 27.0 mmol, 9.0 equiv) and **1-2** (3.0 mmol, 1.0 equiv) in THF (6 mL) at 0 °C within 2 hours. After the addition was completed, the resulting mixture was stirred at 0 °C for another 2 hours. Then, the mixture was quenched with Sat. NaHCO₃ and extracted with CHCl₃. The combined organic layer was washed with brine, dried over anhydrous Na₂SO₄ and concentrated *in vacuo*. The residue was purified by chromatography on silica gel (eluted with hexane/acetone (v/v 100:1)) to afford desired product.

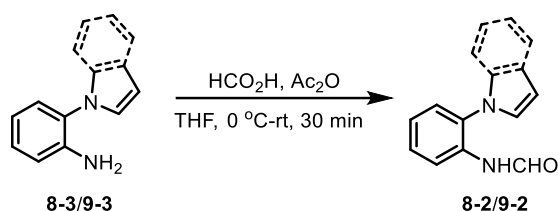
Compound **1** (0.46 g, 85% yield, 3 steps)¹, **4-1** (0.50 g, 86% yield, 3 steps)¹, **5-1** (0.47 g, 75% yield, 3 steps)¹, **6-1** (0.50 g, 85% yield, 3 steps)¹, and **7-1** (0.48 g, 88% yield, 3 steps)² were synthesized following the above procedure and spectra data are consistent with those reported in literature.

3.2 Synthesis of compound **8-1** and **9-1**³



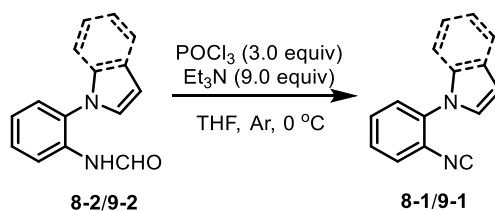
Scheme S5

Indole (702 mg, 6.0 mmol, 1.2 equiv) or pyrrole (403 mg, 6.0 mmol, 1.2 equiv) was added to a mixture of CuI (95 mg, 0.5 mmol, 0.1 equiv), K₃PO₄ (2.3 g, 10.5 mmol, 2.1 equiv), 2-iodoaniline (1.1 g, 5.0 mmol, 1.0 equiv) and *N,N*-dimethylethane-1,2-diamine (DMEDA) (88.1 mg, 1.0 mmol, 0.2 equiv) in toluene (5 mL) at room temperature. The reaction tube was purged with Argon and sealed with PTFE cap. After been heated at 110 °C for 24 h, the mixture was cooled to room temperature, diluted with ethyl acetate (50 mL) and filtered through a plug of celite. The filtrate was concentrated *in vacuo* and the resulting residue was purified by silica gel column chromatography to give the 2-heteroaryl aniline, which was subsequently subjected to the next step.



Scheme S6

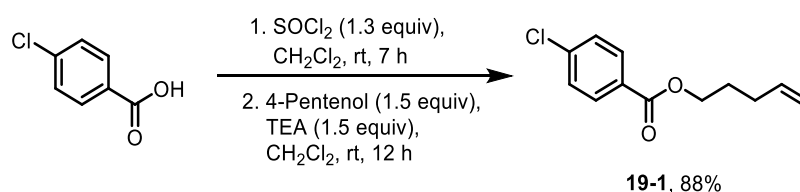
Acetic formic anhydride (18.0 mmol, 6.0 equiv), which was newly prepared from the reaction of acetic anhydride (1.7 mL, 18.0 mmol) with formic acid (0.8 mL, 20.0 mmol) at 55 °C for 2 h, was added dropwise to a mixture of **8-2** (475 mg, 3.0 mmol, 1.0 equiv) or **9-2** (625 mg, 3.0 mmol, 1.0 equiv) in THF (7 mL) at 0 °C. After the addition was completed, the mixture was warmed to room temperature and stirred for 30 mins. Then, the volatiles were removed *in vacuo* to afford (2-heteroaryl)-formanilide quantitatively. This product was directly used for the subsequent dehydration reaction without further purification.



Scheme S7

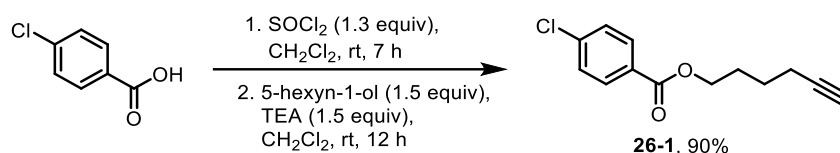
POCl₃ (0.8 mL, 9.0 mmol, 3.0 equiv) was added via syringe pump to a mixture of Et₃N (3.8 mL, 27.0 mmol, 9.0 equiv) and (2-heteroaryl)-formanilide (3.0 mmol, 1.0 equiv) in THF (6 mL) at 0 °C within 2 hour. After the addition was completed, the resulting mixture was stirred at 0 °C for another 2 hour. Then, the mixture was quenched with Sat. NaHCO₃ and extracted with ethyl acetate (10 mL x 3). The combined organic layer was washed with brine, dried over anhydrous Na₂SO₄ and concentrated *in vacuo*. The residue was purified by chromatography on silica gel (eluted with hexane/acetone (v/v 100:1)) to give compound **8-1** (0.40 g, 78% yield, 3 steps)³ and **9-1** (0.52 g, 80% yield, 3 steps)³. The spectra data are consistent with those reported in literature.

3.3 Synthesis of compound **19-1** and **26-1**



Scheme S8

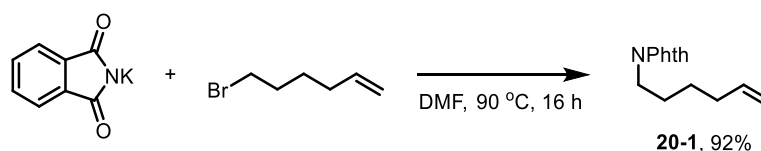
SOCl₂ (2.3 mL, 32.0 mmol, 1.3 equiv) was added dropwise to a solution of 4-chlorobenzoic acid (3.9 g, 25.0 mmol, 1.0 equiv) in dry CH₂Cl₂ (50 mL) at room temperature. The mixture was stirred for 7 h under N₂. DCM and excess of SOCl₂ were then removed under reduced pressure. The crude acyl chloride was dissolved in CH₂Cl₂ (50 mL), and 4-pentenol (3.2 g, 37.5 mmol, 1.5 equiv), TEA (4.9 mL, 37.5 mmol, 1.5 equiv) were added to this solution at 0 °C. The mixture was stirred at room temperature for 12 hour and then quenched with H₂O. The organic layer was separated, washed with brine, dried over anhydrous Na₂SO₄ and concentrated *in vacuo*. The resulting residue was purified by silica gel column chromatography (eluted with hexane) to afford compound **19-1** in 88% yield (4.93 g). The spectra data are consistent with those reported in literature.⁴



Scheme S9

To a solution of 4-chlorobenzoic acid (3.9 g, 25.0 mmol, 1.0 equiv) in dry CH_2Cl_2 (50 mL) was added SOCl_2 (2.3 mL, 32.0 mmol, 1.3 equiv). The mixture was stirred at room temperature for 7 h under N_2 . DCM and excess of SOCl_2 were then removed under reduced pressure. The crude acyl chloride was dissolved in CH_2Cl_2 (50 mL), and 5-hexyn-1-ol (3.7 g, 37.5 mmol, 1.5 equiv), TEA (4.9 mL, 37.5 mmol, 1.5 equiv) were added to this solution at 0 °C. The mixture was stirred at room temperature for 12 hour and then quenched with H_2O . The organic layer was washed with brine, dried over anhydrous Na_2SO_4 and concentrated *in vacuo*. The resulting residue was purified by silica gel column chromatography (eluted with hexane) to afford compound **26-1** in 90% yield (5.31 g). The spectra data are consistent with those reported in literature.⁵

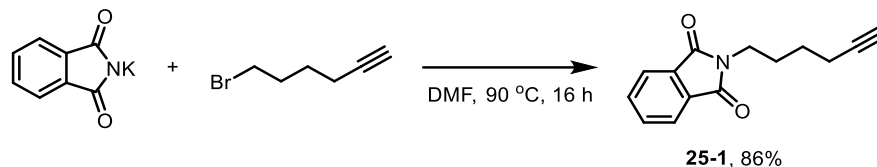
3.4 Synthesis of compound **20-1** and **25-1**



Scheme S10

Potassium phthalimide (3.9 g, 21.0 mmol, 1.1 equiv) was added to a solution of 6-bromohexene (3.2 g, 19.6 mmol, 1.0 equiv) in anhydrous DMF (23 mL) at room temperature. The mixture was heated to 90 °C for 16 h. After been cooled to room temperature, the reaction mixture was poured into water (75 mL) and extracted with CH_2Cl_2 (3 x 50 mL). The combined organic phase was washed with 100 mL of aq. KOH (0.2 M) and water. The organic phase was dried over anhydrous Na_2SO_4 , filtered and concentrated *in vacuo*. The resulting residue was purified by silica gel column

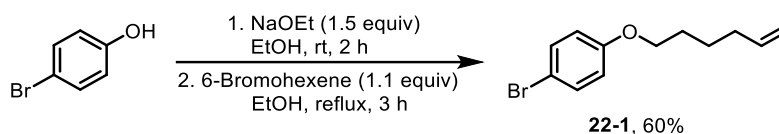
chromatography (eluted with hexane/acetone (v/v 5:1)) to afford the compound **20-1** in 92% yield (4.13 g). The spectra data are consistent with those reported in literature.⁶



Scheme S11

Potassium phthalimide (3.9 g, 21.0 mmol, 1.1 equiv) was added to a solution of 6-bromo-1-hexyne (3.1 g, 19.6 mmol, 1.0 equiv) in anhydrous DMF (23 mL) at room temperature. The mixture was heated to 90 °C for 16 h. After being cooled to room temperature, the reaction mixture was poured into water (75 mL) and extracted with CH₂Cl₂ (3 x 50 mL). The combined organic phase was washed with 100 mL of aq. KOH (0.2 M) and water. The organic phase was dried over anhydrous Na₂SO₄, filtered and concentrated *in vacuo*. The resulting residue was purified by silica gel column chromatography (eluted with hexane/acetone (v/v 5:1)) to afford compound **25-1** in 86% yield (3.83 g). The spectra data are consistent with those reported in literature.⁷

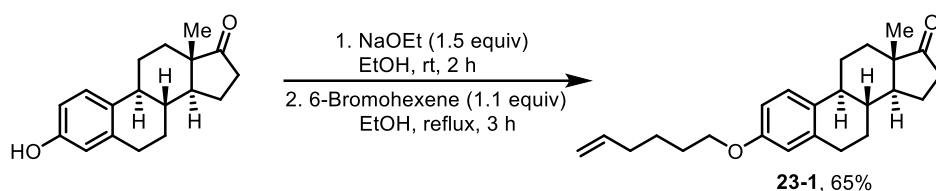
3.5 Synthesis of compound **22-1** and **23-1**



Scheme S12

A mixture of *p*-BrC₆H₄OH (0.5 g, 3.0 mmol, 1.0 equiv) and NaOEt (0.3 g, 4.5 mmol, 1.5 equiv) in EtOH (4 mL) was stirred at room temperature for 2 hour. Then 6-bromohexene (0.5 g, 3.3 mmol, 1.1 equiv) was added slowly. The resulting mixture was heated at reflux for 3 h. After being cooled to room temperature, the solvent was

removed under reduced pressure. The residue was dissolved in ethyl acetate and washed with water and brine. The organic phase was dried over anhydrous Na_2SO_4 and concentrated *in vacuo*. The residue was purified by silica gel column chromatography (eluted with hexane) to afford compound **22-1** as a colorless oil in 60% yield (0.46 g, R_f = 0.7, hexane). **^1H NMR** (400 MHz, CDCl_3) δ 7.36 (d, J = 9.0 Hz, 2H), 6.77 (d, J = 9.0 Hz, 2H), 5.83 (m, 1H), 5.04 (dd, J = 17.1, 1.7 Hz, 1H), 4.99 (d, J = 8.0 Hz, 1H), 3.92 (t, J = 6.5 Hz, 2H), 2.13 (q, J = 7.2 Hz, 2H), 1.85-1.74 (m, 2H), 1.61-1.52 (m, 2H); **^{13}C NMR** (101 MHz, CDCl_3) δ 158.28, 138.52, 132.28, 116.35, 114.93, 112.68, 68.07, 33.51, 28.70, 25.36; **HRMS** Calcd for $\text{C}_{12}\text{H}_{15}\text{BrO}$ [M]: 254.0306, Found: 254.0302.

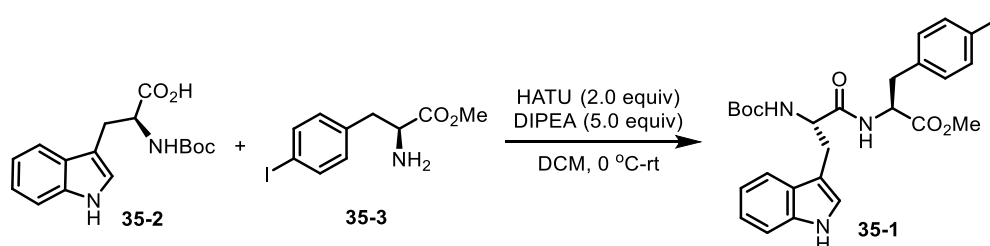


Scheme S13

A mixture of compound **23-2** (0.8 g, 3.0 mmol, 1.0 equiv) and NaOEt (0.3 g, 4.5 mmol, 1.5 equiv) in EtOH (4 mL) was stirred at room temperature for 2 hour. Then 6-bromohexene (0.5 g, 3.3 mmol, 1.1 equiv) was added to this solution. The resulting mixture was heated at reflux for 3 h. After been cooled to room temperature, the solvent was removed under reduced pressure. The residue was dissolved in ethyl acetate and washed with water and brine. The organic phase was dried over anhydrous Na_2SO_4 and concentrated *in vacuo*. The residue was purified by silica gel column chromatography (eluted with hexane) to afford compound **23-1** as a white solid in 65% yield (0.69 g, R_f = 0.3, hexane). **^1H NMR** (400 MHz, CDCl_3) δ 7.20 (d, J = 8.6 Hz, 1H), 6.72 (dd, J = 8.6, 2.6 Hz, 1H), 6.65 (d, J = 2.5 Hz, 1H), 5.89-5.79 (m, 1H), 5.04 (dd, J = 17.1, 1.7 Hz, 1H), 4.98 (dd, J = 10.2, 0.7 Hz, 1H), 3.94 (t, J = 6.4 Hz, 2H), 3.00-2.80 (m, 2H), 2.51 (dd, J = 18.8, 8.6 Hz, 1H), 2.44-2.36 (m, 1H), 2.25 (dd, J = 13.6, 7.1 Hz, 1H), 2.18-1.93 (m, 6H), 1.86-1.73 (m, 2H), 1.69-1.53 (m, 5H), 1.52-1.38 (m, 3H), 0.92 (s, 3H); **^{13}C NMR** (101 MHz, CDCl_3) δ 220.95, 157.17, 138.63, 137.73, 131.91, 126.34,

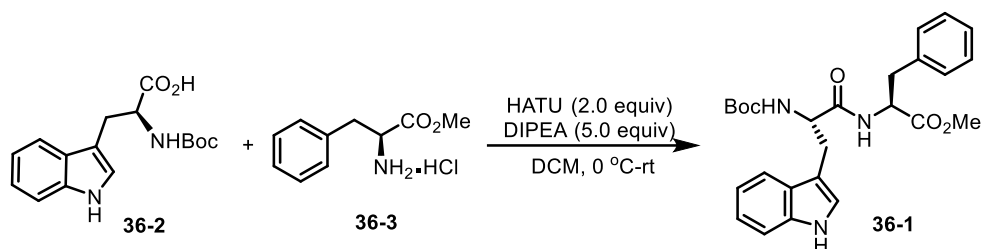
114.77, 114.58, 112.15, 67.70, 50.46, 48.06, 44.04, 38.45, 35.93, 33.51, 31.65, 29.72, 28.84, 26.64, 25.99, 25.41, 21.65, 13.92; **HRMS** Calcd for C₂₄H₃₂O₂ [M]: 352.2402; Found: 352.2401.

3.6 Synthesis of peptides **35-1** and **36-1**



Scheme S14

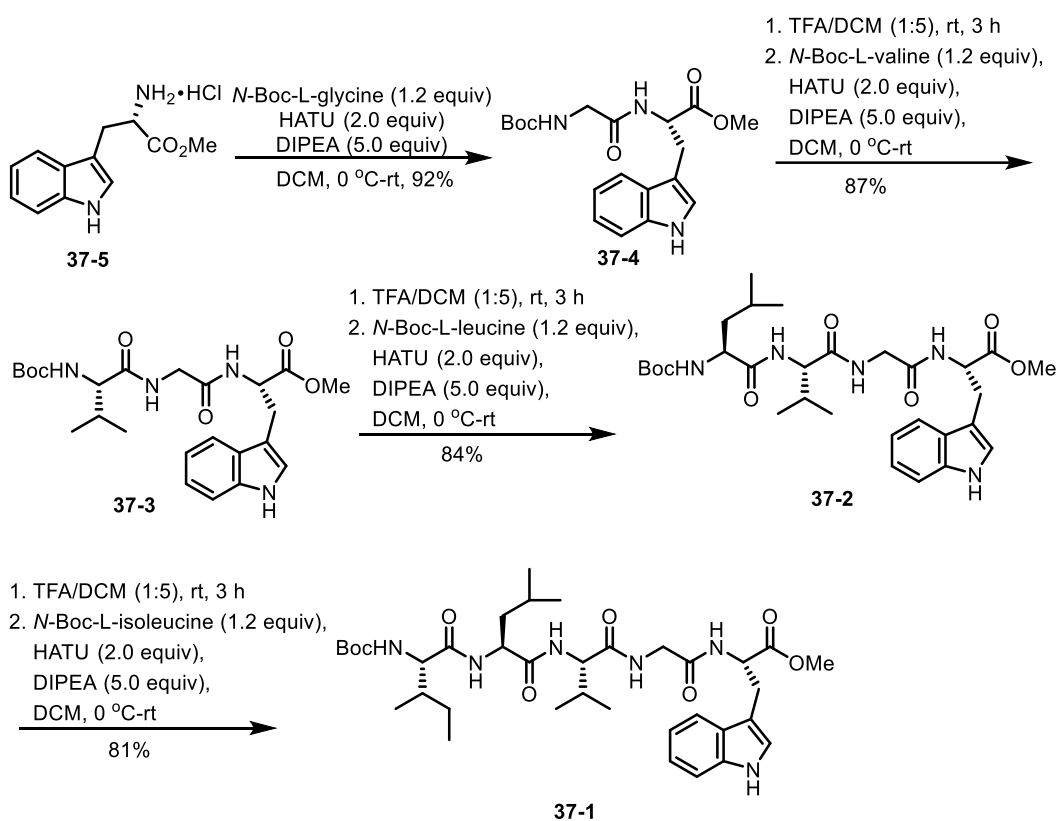
Compound **35-1** was prepared following the general amide coupling procedure: HATU (3.8 g, 10.0 mmol, 2.0 equiv) were added to a solution of compound **35-2** (1.5 g, 5.0 mmol, 1.0 equiv) and compound **35-3** (1.8 g, 6.0 mmol, 1.2 equiv) in dry DCM (25 mL) at 0 °C. After 10 min, DIPEA (4.1 mL, 25.0 mmol, 5.0 equiv) was added to the reaction mixture. The resulting solution was warmed to room temperature and stirred for 1 hour. Then the reaction mixture was washed with water and brine successively. The organic phase was dried over anhydrous Na₂SO₄ and concentrated *in vacuo*. The residue was purified by silica gel flash chromatography (eluted with hexane/acetone (v/v 3:2)) to give the desired product **35-1** in 89% yield (2.63 g, *R_f* = 0.2, 20% acetone in hexane). ¹H NMR (400 MHz, CDCl₃) δ 8.26 (s, 1H), 7.66 (d, *J* = 7.8 Hz, 1H), 7.42 (d, *J* = 8.1 Hz, 2H), 7.37 (d, *J* = 8.1 Hz, 1H), 7.20 (t, *J* = 7.4 Hz, 1H), 7.13 (t, *J* = 7.4 Hz, 1H), 7.00 (s, 1H), 6.51 (d, *J* = 7.8 Hz, 2H), 6.25 (s, 1H), 5.13 (s, 1H), 4.70 (s, 1H), 4.43 (s, 1H), 3.60 (s, 3H), 3.33 (d, *J* = 10.8 Hz, 1H), 3.11 (dd, *J* = 14.5, 7.2 Hz, 1H), 2.86 (d, *J* = 5.2 Hz, 2H), 1.43 (s, 9H); ¹³C NMR (101 MHz, CDCl₃) δ 171.53, 171.06, 155.46, 137.45, 136.34, 135.32, 131.19, 127.38, 123.45, 122.24, 119.70, 118.83, 111.38, 110.14, 92.57, 80.24, 55.18, 53.00, 52.31, 38.67, 37.28, 30.97, 28.32, 28.20; **HRMS** Calcd for C₂₆H₃₀IN₃O₅Na [M+Na⁺]: 614.1122, Found: 614.1126.



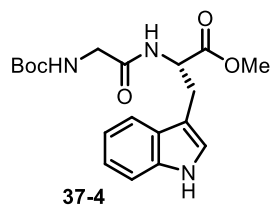
Scheme S15

Compound **36-1** was prepared following the same amide coupling procedure as compound **35-1** in 87% yield (2.02 g). The spectra data are consistent with those reported in literature.⁸

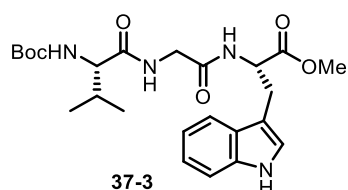
3.7 Synthesis of compound **37-1**



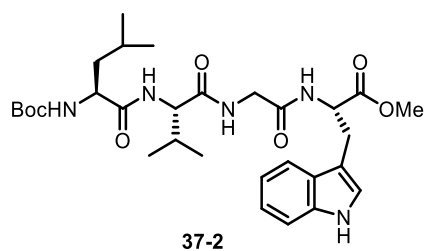
Scheme S16



Compound **37-4** was synthesized by coupling **37-5** with *N*-Boc-L-glycine following the amide coupling procedure in 92% yield (1.72 g). The spectra data are consistent with those reported in literature.⁹

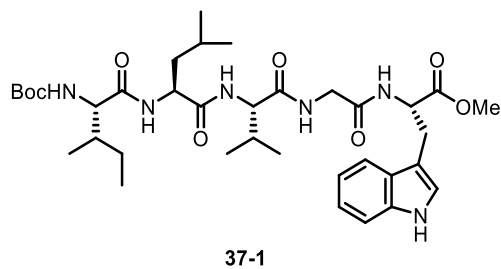


Trifluoroacetic acid (5.0 mL) was added to a solution of compound **37-4** (1.9 g, 5.0 mmol, 1.0 equiv) in dry dichloromethane (25 mL). The mixture was stirred for 3 hour at room temperature. Then the solvent was removed under reduced pressure and the residue was treated with *N*-Boc-L-valine (1.3 g, 6.0 mmol, 1.2 equiv), HATU (3.8 g, 10.0 mmol, 2.0 equiv) and DIPEA (4.1 mL, 25.0 mmol, 5.0 equiv) following the general amide coupling procedure to give **37-3** ($R_f = 0.4$, 60% acetone in hexane) in 87% yield (2.06 g). **¹H NMR** (400 MHz, CDCl₃) δ 9.09 (s, 1H), 7.43 (d, $J = 7.6$ Hz, 1H), 7.26 (d, $J = 7.9$ Hz, 1H), 7.16-6.98 (m, 4H), 6.94 (s, 1H), 5.43 (d, $J = 8.6$ Hz, 1H), 4.79 (d, $J = 5.7$ Hz, 1H), 4.07-3.94 (m, 1H), 3.77 (dd, $J = 16.6, 5.0$ Hz, 1H), 3.63 (s, 3H), 3.38-3.13 (m, 2H), 1.98 (d, $J = 6.2$ Hz, 1H), 1.43 (s, 9H), 1.28 (d, $J = 6.1$ Hz, 1H), 0.90 (d, $J = 6.6$ Hz, 3H), 0.82 (d, $J = 6.6$ Hz, 3H); **¹³C NMR** (101 MHz, CDCl₃) δ 172.57, 172.29, 168.92, 156.30, 136.28, 127.31, 123.84, 121.99, 119.37, 118.35, 111.63, 108.99, 80.16, 59.73, 52.48, 52.32, 42.88, 38.66, 31.23, 28.42, 27.33, 26.97, 19.33, 17.50; **HRMS** Calcd for C₂₄H₃₄N₄O₆Na [M+Na⁺]: 497.2371; Found: 497.2373.



Trifluoroacetic acid (5 mL) was added to a solution of compound **37-3** (2.06 g, 4.3 mmol,

1.0 equiv) in dry dichloromethane (25 mL). The mixture was stirred for 3 hour at room temperature. Then the solvent was removed under reduced pressure and the residue was treated with *N*-Boc-L-leucine (1.2 g, 5.2 mmol, 1.2 equiv), HATU (3.3 g, 8.6 mmol, 2.0 equiv) and DIPEA (3.5 mL, 21.5 mmol, 5.0 equiv) following the general amide coupling procedure to give **37-2** ($R_f = 0.4$, 5% CH₃OH in DCM) in 84% yield (2.12 g). **¹H NMR** (400 MHz, MeOD) δ 7.48 (d, $J = 7.8$ Hz, 1H), 7.31 (d, $J = 8.1$ Hz, 1H), 7.06 (t, $J = 6.0$ Hz, 2H), 6.99 (t, $J = 7.0$ Hz, 1H), 4.71 (t, $J = 6.5$ Hz, 1H), 4.16 (d, $J = 6.8$ Hz, 1H), 4.09 (q, $J = 9.8$, 1H), 3.85 (q, $J = 16.8$ Hz, 2H), 3.61 (s, 3H), 3.30-3.28 (m, 1H), 3.28-3.16 (m, 2H), 2.04 (m, 1H), 1.65-1.56 (m, 1H), 1.53-1.43 (m, 2H), 1.40 (s, 10H), 1.26 (s, 1H), 1.00-0.76 (m, 14H); **¹³C NMR** (101 MHz, MeOD) δ 174.43, 172.55, 172.36, 169.71, 156.68, 136.57, 127.29, 123.23, 121.08, 118.50, 117.74, 111.01, 109.07, 79.32, 58.76, 54.40, 53.41, 53.14, 51.38, 41.88, 40.28, 30.60, 27.36, 27.09, 24.48, 22.15, 20.48, 18.38, 17.31, 17.18, 15.86; **HRMS** Calcd for C₂₄H₃₅N₄O₆ [M+H⁺]: 587.3319; Found: 587.3320.



Trifluoroacetic acid (5.0 mL) was added to a solution of compound **37-2** (2.12 g, 3.6 mmol, 1.0 equiv) in dry dichloromethane (25 mL). The mixture was stirred for 3 h at room temperature. Then the solvent was removed under reduced pressure and the residue was treated with *N*-Boc-L-isoleucine (1.0 g, 4.3 mmol, 1.2 equiv), HATU (2.7 g, 7.2 mmol, 2.0 equiv) and DIPEA (3.0 mL, 18.0 mmol, 5.0 equiv) following the general amide coupling procedure to give **37-1** ($R_f = 0.4$, 5% CH₃OH in DCM) in 81% yield (2.04 g). **¹H NMR** (400 MHz, MeOD) δ 7.47 (d, $J = 7.8$ Hz, 1H), 7.30 (d, $J = 8.1$ Hz, 1H), 7.06 (t, 2H), 6.98 (t, $J = 7.3$ Hz, 1H), 4.73 (t, $J = 6.4$ Hz, 1H), 4.59 (m, 1H), 4.21 (d, $J = 7.2$ Hz, 1H), 3.94 (d, $J = 7.6$ Hz, 1H), 3.88 (d, $J = 9.0$ Hz, 2H), 3.58 (s, 3H), 3.29 (s, 1H), 3.26 (d, $J = 5.8$ Hz, 1H), 3.22 (d, $J = 7.0$ Hz, 1H), 2.02 (q, $J = 4.0$ Hz, 1H), 1.79-1.68 (m, 1H), 1.63 (m, 1H), 1.59-1.46 (m, 3H), 1.40 (s, 10H), 1.26 (s, 1H), 1.17-

1.04 (m, 1H), 0.88 (m, 21H); ^{13}C NMR (101 MHz, MeOD) δ 174.46, 174.37, 173.87, 173.69, 170.94, 157.78, 137.88, 128.65, 124.55, 122.38, 119.81, 119.06, 112.33, 110.38, 80.39, 60.44, 60.32, 55.73, 54.81, 52.76, 52.68, 43.25, 41.80, 38.04, 31.86, 28.73, 28.52, 25.87, 25.66, 23.52, 22.17, 19.74, 18.96, 17.20, 16.09, 11.39; HRMS Calcd for $\text{C}_{24}\text{H}_{35}\text{N}_4\text{O}_6$ $[\text{M}+\text{H}^+]$: 701.4232; Found: 701.4235.

4. Evaluation of synthesis phenanthridine 3 from 2-isocyanobiphenyl and $\text{C}_4\text{F}_9\text{I}$ via a radical cascade process

All screening reactions were carried out at a 0.2 mmol scale in a 4 mL glass vial (Thermo Scientific, National B7999-2) sealed with PTEF cap and stirred on bench top. 2-Isocyanobiphenyl **1**, $\text{C}_4\text{F}_9\text{I}$ and other specified reagents were dispersed in 2 mL of solvent and the resulting mixture was purged with Ar (if necessary) and vigorously stirred at specified temperature under light irradiation (laid 10 CM away from the vial, if necessary) for 20 hour. The solvent of the reaction mixture was removed under reduced pressure. The resulting residue was dissolved in 1 mL of CDCl_3 along with $\text{Cl}_2\text{CHCHCl}_2$ (20 μL) as an external standard for ^1H -NMR analysis. The composition of reaction mixture was based on the Ar-H at 8.73 ppm (d, $J = 8.4$ Hz, 1H) for compound **3**.

Table S1. Perfluoroalkylation reaction of 2-isocyanobiphenyl **1** with perfluorobutyl iodide **2**

| Entry | Reagents (equiv)/atmosphere ^a | Solvent | t (°C) | Time (h) | Yield (%) ^b |
|-------|---|---------|--------|----------|------------------------|
| 1 | NiCl_2 (0.05), Cs_2CO_3 (1.5), Ar ^c | dioxane | 100 | 20 | 62 |
| 2 | Cs_2CO_3 (1.5), Ar ^c | dioxane | 100 | 20 | 60 |

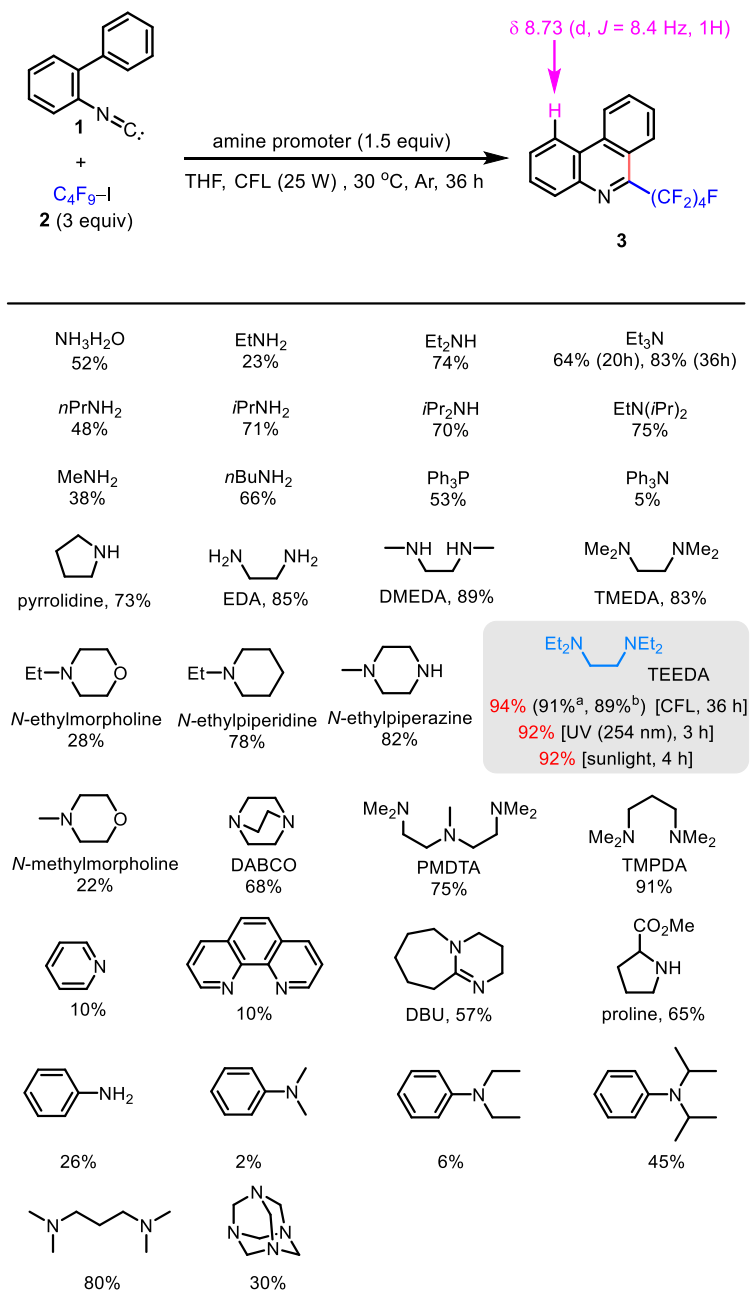
| | | | | | |
|----|--|--------------------|-----|----|----|
| 3 | no additives, Ar ^c | dioxane | 100 | 20 | <2 |
| 4 | Cs ₂ CO ₃ (1.5), Ar ^c | dioxane | 30 | 20 | <2 |
| 5 | no additives, CFL, Ar | dioxane | 30 | 20 | 5 |
| 6 | no additives, CFL, Ar | THF | 30 | 20 | 10 |
| 7 | no additives, CFL, Ar | THP | 30 | 20 | 7 |
| 8 | no additives, CFL, Ar | DCM | 30 | 20 | 4 |
| 9 | no additives, CFL, Ar | CHCl ₃ | 30 | 20 | 5 |
| 10 | no additives, CFL, Ar | CH ₃ CN | 30 | 20 | <2 |
| 11 | no additives, CFL, Ar | DMF | 30 | 20 | <2 |
| 12 | no additives, in dark, Ar | THF | 30 | 20 | <2 |
| 13 | Cs ₂ CO ₃ (1.5), CFL, Ar | THF | 30 | 20 | 29 |
| 14 | K ₂ CO ₃ (1.5), CFL, Ar | THF | 30 | 20 | 10 |
| 15 | Na ₂ CO ₃ (1.5), CFL, Ar | THF | 30 | 20 | 7 |
| 16 | NaOMe (1.5), CFL, Ar | THF | 30 | 20 | 4 |
| 17 | KOt-Bu (1.5), CFL, Ar | THF | 30 | 20 | 20 |
| 18 | Et ₃ N (1.5), CFL, Ar | THF | 30 | 20 | 64 |
| 19 | Et ₃ N (1.5), UV-Hg (254nm), Ar | THF | 30 | 20 | 73 |
| 20 | Et ₃ N (1.5), sunlight (direct irradiation), Ar | THF | 30 | 4 | 76 |
| 21 | UV-Hg (254nm), no Et ₃ N, Ar | THF | 30 | 20 | 15 |
| 22 | Et ₃ N (1.5), CFL, Ar | THP | 30 | 20 | 57 |
| 23 | Et ₃ N (1.5), CFL, Ar | 2-MeTHF | 30 | 20 | 52 |
| 24 | Et ₃ N (1.5), CFL, Ar | Et ₂ O | 30 | 20 | 28 |
| 25 | Et ₃ N (1.5), CFL, Ar | MTBE | 30 | 20 | 40 |
| 26 | Et ₃ N (1.5), CFL, Ar | dioxane | 30 | 20 | 38 |
| 27 | Et ₃ N (1.5), CFL, Ar | CHCl ₃ | 30 | 20 | 30 |
| 28 | Et ₃ N (1.5), CFL, Ar | CH ₃ CN | 30 | 20 | 46 |
| 29 | Et ₃ N (1.5), CFL, Ar | H ₂ O | 30 | 20 | 20 |
| 30 | Et ₃ N (0.2), CFL, Ar | THF | 30 | 20 | 16 |
| 31 | Et ₃ N (0.2), K ₂ CO ₃ (1.5), CFL, Ar | THF | 30 | 20 | 32 |
| 32 | Et ₃ N (1.5), in dark, Ar | THF | 30 | 20 | <2 |
| 33 | Et ₃ N (1.5), CFL, O ₂ (1 atm) | THF | 30 | 20 | 4 |
| 34 | Et ₃ N (1.5), CFL, air | THF | 30 | 20 | 32 |
| 35 | I ₂ (0.1), CFL, Ar | THF | 30 | 20 | <2 |
| 36 | I ₂ (0.1), CFL, Ar | DCM | 30 | 20 | <2 |

a) CFL: household compact fluorescent lamp, 25W; UV (254 nm): low-pressure Hg-vapor lamp, 25W; b) Yield are based on ¹H-NMR analysis of crude reaction mixture on a 0.2 mmol scale; c) Conducted in the presence of ambient laboratory light.

5. Evaluation of amine promoters for synthesis of **3** under CFL irradiation

2-Isocyanobiphenyl **1** (36 mg, 0.2 mol, 1.0 equiv), C₄F₉I (208 mg, 0.6 mol, 3.0 equiv) and the specific amine (0.3 mol, 1.5 equiv) were dispersed in 2 mL of solvent. The

reaction vial was then purged with Ar for 1 min, sealed with PTEF cap. The reaction mixture was vigorously stirred at 30 °C under CFL (25W, laid 10 CM away from the vial) irradiation for 36 hour. Then THF was removed under reduced pressure. The resulting residue was dissolved in 1 mL of CDCl₃ along with Cl₂CHCHCl₂ (20 μL) as an external standard for ¹H-NMR analysis. The composition of reaction mixture was based on the Ar-H at 8.73 ppm (d, *J* = 8.4 Hz, 1H) for compound **3**.



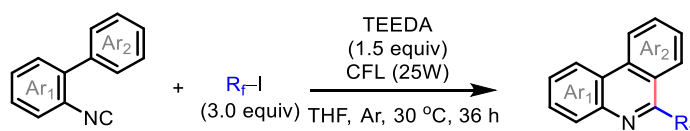
CFL: 25 W; UV (254 nm): low-pressure Hg-vapor lamp, 25 W; sunlight: direct irradiation for 4 h. Yields are based on ¹H-NMR analysis of crude reaction mixture on a 0.2 mmol scale performed

in 4 mL borosilicate glass vial under Ar. a) Isolated yield on 0.2 mmol scale. b) Isolated yield on 1 mmol scale.

Scheme S17

Procedure of phenanthridine synthesis in sunlight: 2-Isocyanobiphenyl **1** (36 mg, 0.2 mol, 1.0 equiv), C₄F₉I (208 mg, 0.6 mol, 3.0 equiv) and *N,N,N',N'*-Tetraethylenediamine (TEEDA) (52 mg, 0.3 mmol, 1.5 equiv) were dispersed in 2 mL of THF. The reaction vial was then purged with Ar for 1 min and sealed with PTEF cap. The reaction mixture was vigorously stirred at 30 °C in sunlight for 4 hour. Then THF was removed under reduced pressure. The resulting residue was dissolved in 1 mL of CDCl₃ along with Cl₂CHCHCl₂ (20 µL) as an external standard for ¹H-NMR analysis.

6. General procedures and substrate scope of phenanthridine synthesis



Scheme S18

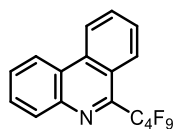
General procedure A for the synthesis of compound 4, 5, 6, 7, 8, 9, 10, 11, 12, 13 and 15:

2-Isocyanobiaryl compound (0.2 mmol, 1.0 equiv), perfluoroalkyl iodide (0.6 mmol, 3.0 equiv) and *N,N,N',N'*-Tetraethylenediamine (TEEDA) (0.3 mmol, 1.5 equiv) were dispersed in 2 mL of THF. The reaction vial was then purged with Ar for 1 min and sealed with PTEF cap. The reaction mixture was vigorously stirred at 30 °C under irradiation of CFL lamp (25W, laid 10 CM away from the vial) for 36 hour. Then the mixture was concentrated *in vacuo*, and the residue was purified by silica gel flash chromatography to give the desired product.

General procedure B for the synthesis of compound 16, 17 and 18:

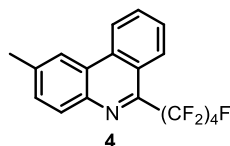
2-Isocyanobiaryl compound (0.2 mmol, 1.0 equiv), alkyl bromide (1.0 mmol, 5.0 equiv) and *N,N,N',N'*-Tetraethylenediamine (TEEDA) (0.6 mmol, 3.0 equiv) were dispersed in 2 mL of THF. The reaction vial was then purged with Ar for 1 min and sealed with

PTEF cap. The reaction mixture was vigorously stirred at 30 °C under irradiation of CFL lamp (25W, laid 10 CM away from the vial) for 36 h. Then the mixture was concentrated *in vacuo*, and the residue was purified by silica gel flash chromatography to give the desired product.



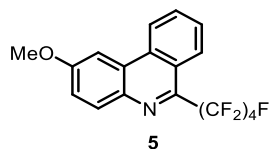
3 $R_f = 0.5$, 1% acetone in Hexane

Compound **3** was isolated in 91% yield (72 mg) following the general procedure A. **¹H NMR** (400 MHz, CDCl₃) δ 8.73 (d, $J = 8.4$ Hz, 1H), 8.67-8.58 (m, 1H), 8.47 (d, $J = 8.4$ Hz, 1H), 8.33-8.23 (m, 1H), 7.93 (t, $J = 7.7$ Hz, 1H), 7.86-7.72 (m, 3H); **¹³C NMR** (101 MHz, CDCl₃) δ 146.79 (t, $J = 24.1$ Hz), 141.87, 134.13, 131.34, 131.32, 129.54, 129.44, 128.13, 126.36-126.22(m), 124.95, 123.08, 122.75, 122.16, 120-100 (m); **¹⁹F NMR** (376 MHz, CDCl₃) δ -80.97 (t, $J = 10.6$ Hz, 3F), -103.16 – -106.93 (m, 2F), -118.09 – -121.23 (m, 2F), -122.18 – -125.63 (m, 2F); **HRMS** Calcd for C₁₇H₉F₉N [M+H⁺]: 398.0586; Found: 398.0588.



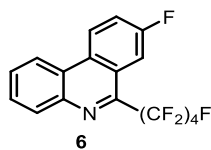
4 $R_f = 0.5$, 1% acetone in Hexane

Compound **4** was isolated in 85% yield (70 mg) following the general procedure A. **¹H NMR** (400 MHz, CDCl₃) δ 8.70 (d, $J = 8.4$ Hz, 1H), 8.44 (d, $J = 8.4$ Hz, 1H), 8.39 (s, 1H), 8.16 (d, $J = 8.4$ Hz, 1H), 7.89 (t, $J = 7.3$ Hz, 1H), 7.74 (t, $J = 7.8$ Hz, 1H), 7.63 (dd, $J = 8.4, 1.4$ Hz, 1H), 2.66 (s, 3H); **¹³C NMR** (101 MHz, CDCl₃) δ 145.72 (t, $J = 24.6$ Hz), 140.24, 139.88, 133.77, 131.19, 131.00, 127.93, 126.23-126.09 (m), 124.80, 123.16, 122.67, 121.69, 120-100 (m), 22.32; **¹⁹F NMR** (376 MHz, CDCl₃) δ -80.98 (t, $J = 10.6$ Hz, 3F), -102.98 – -107.39 (m, 2F), -118.41 – -121.54 (m, 2F), -122.31 – -125.62 (m, 2F); **HRMS** Calcd for C₁₈H₁₁F₉N [M+H⁺]: 412.0742; Found: 412.0745.



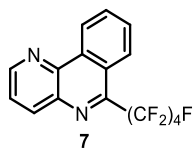
5 $R_f = 0.6$, 1% acetone in Hexane

Compound **5** was isolated in 71% yield (61 mg) following the general procedure A. **¹H NMR** (400 MHz, CDCl₃) δ 8.62 (d, *J* = 8.4 Hz, 1H), 8.44 (d, *J* = 8.4 Hz, 1H), 8.18 (d, *J* = 9.0 Hz, 1H), 7.95-7.84 (m, 2H), 7.75 (t, *J* = 7.8 Hz, 1H), 7.42 (dd, *J* = 9.0, 2.4 Hz, 1H), 4.04 (s, 3H); **¹³C NMR** (101 MHz, CDCl₃) δ 160.41, 143.91(t, *J* = 24.9 Hz), 137.20, 133.35, 132.78, 130.68, 128.11, 126.35, 126.16-126.03 (m), 123.23, 122.64, 120-100 (m), 119.61, 102.60, 55.76; **¹⁹F NMR** (376 MHz, CDCl₃) δ -80.98 (t, *J* = 10.6 Hz, 3F), -113.62 – -106.12 (m, 2F), -118.72 – -121.21 (m, 2F), -122.31 – -125.62 (m, 2F); **HRMS** Calcd for C₁₈H₁₁F₉NO [M+H⁺]: 428.0691; Found: 428.0692.



R_f = 0.5, 1% acetone in Hexane

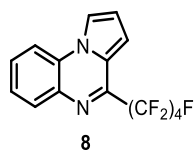
Compound **6** was isolated in 94% yield (78 mg) following the general procedure A. **¹H NMR** (400 MHz, CDCl₃) δ 8.72 (dd, *J* = 9.1, 5.4 Hz, 1H), 8.61-8.48 (m, 1H), 8.32-8.21 (m, 1H), 8.09 (d, *J* = 10.1 Hz, 1H), 7.81 (dd, *J* = 6.0, 3.4 Hz, 2H), 7.68 (t, *J* = 8.4 Hz, 1H); **¹³C NMR** (101 MHz, CDCl₃) δ 161.56 (d, *J* = 249.5 Hz), 145.91 (t, *J* = 24.6 Hz), 141.52, 131.40, 130.81, 129.97, 129.33, 125.25 (d, *J* = 8.8 Hz), 124.49, 124.06 (d, *J* = 8.9 Hz), 121.85, 120.90, 120.66, 120-100 (m), 111.12 (dt, *J* = 23.8, 7.1 Hz); **¹⁹F NMR** (376 MHz, CDCl₃) δ -80.97 (t, *J* = 10.7 Hz, 3F), -104.39 – -106.93 (m, 2F), -109.92 (s, 1F), -118.54 – -121.53 (m, 2F), -122.52 – -125.31 (m, 2F); **HRMS** Calcd for C₁₇H₈F₁₀N [M+H⁺]: 416.0492, Found: 416.0495.



R_f = 0.4, 1% acetone in Hexane

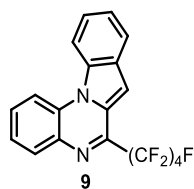
Compound **7** was isolated in 74% yield (59 mg) following the general procedure A. **¹H NMR** (400 MHz, CDCl₃) δ 9.37 (d, *J* = 7.7 Hz, 1H), 9.12 (dd, *J* = 4.3, 1.6 Hz, 1H), 8.55 (dd, *J* = 8.3, 1.7 Hz, 1H), 8.47 (d, *J* = 8.5 Hz, 1H), 8.07-7.97 (m, 1H), 7.88 (ddd, *J* = 8.4, 7.1, 1.3 Hz, 1H), 7.76 (dd, *J* = 8.3, 4.3 Hz, 1H); **¹³C NMR** (101 MHz, CDCl₃) δ 151.90, δ 147.63 (t, *J* = 25.0 Hz), 141.46, 138.30, 136.75, 135.06, 131.84, 129.80, 125.73-125.55 (m), 125.20, 124.60, 124.42, 120-100 (m); **¹⁹F NMR** (376 MHz, CDCl₃) δ -80.95 (t, *J* = 10.4 Hz, 3F), -103.44 – -107.07 (m, 2F), -117.91 – -121.22 (m, 2F), -

122.63 – -125.32 (m, 2F); **HRMS** Calcd for C₁₆H₈F₉N₂ [M+H⁺]: 399.0538, Found: 399.0536.



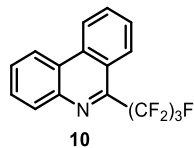
8 R_f = 0.3, 1% acetone in Hexane

Compound **8** was isolated in 59% yield (46 mg) following the general procedure **A**. **¹H NMR** (400 MHz, CDCl₃) δ 8.09-8.05 (m, 2H), 7.90 (d, J = 8.2 Hz, 1H), 7.64 (t, J = 7.5 Hz, 1H), 7.51 (t, J = 7.6 Hz, 1H), 7.17-7.10 (m, 1H), 7.03-6.93 (m, 1H); **¹³C NMR** (101 MHz, CDCl₃) 142.74 (t, J = 26.2 Hz), 134.25, 131.35, 130.16, 127.77, 125.91, 122.86, 120-100 (m), 115.50, 115.09, 113.90, 108.88; **¹⁹F NMR** (376 MHz, CDCl₃) δ -79.24 – 82.55 (m, 3F), -111.80 – -114.94 (m, 2F), -120.59 – -122.95 (m, 2F), -124.20 – -126.72 (m, 2F); **HRMS** Calcd for C₁₅H₈F₉N₂ [M+H⁺]: 387.0538, Found: 387.0542.



9 R_f = 0.4, 1% acetone in Hexane

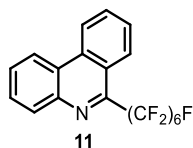
Compound **9** was isolated in 56% yield (49 mg) following the general procedure **A**. **¹H NMR** (400 MHz, CDCl₃) δ 8.53 (d, J = 8.4 Hz, 1H), 8.47 (d, J = 8.7 Hz, 1H), 8.11 (dd, J = 8.0, 1.4 Hz, 1H), 8.01 (d, J = 8.0 Hz, 1H), 7.73 (t, J = 7.9 Hz, 1H), 7.61 (t, J = 7.3 Hz, 1H), 7.52-7.47 (m, 2H), 7.44 (s, 1H); **¹³C NMR** (101 MHz, CDCl₃) δ 144.40 (t, J = 25.5 Hz), 134.17, 132.70, 131.86, 131.17, 130.95, 129.21, 125.72, 125.47, 124.69, 123.50, 123.43, 120-100 (m), 114.90, 114.59, 102.33; **¹⁹F NMR** (376 MHz, CDCl₃) δ -78.78 – -82.87 (m, 3F), -111.17 – -114.63 (m, 2F), -119.64 – -122.95 (m, 2F), -123.59 – -127.19 (m, 2F); **HRMS** Calcd for C₁₉H₁₀F₉N₂ [M+H⁺]: 437.0695, Found: 437.0698.



10 R_f = 0.5, 1% acetone in Hexane

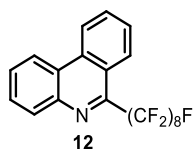
Compound **10** was isolated in 71% yield (49 mg) following the general procedure **A**. **¹H NMR** (400 MHz, CDCl₃) δ 8.72 (d, J = 8.3 Hz, 1H), 8.67-8.57 (m, 1H), 8.47 (d, J = 8.3 Hz, 1H), 8.34-8.22 (m, 1H), 7.92 (t, J = 7.6 Hz, 1H), 7.86-7.70 (m, 3H); **¹³C NMR** (101 MHz, CDCl₃) δ 146.67 (t, J = 24.3 Hz), 141.94, 134.15, 131.38, 131.32, 129.53,

129.44, 128.12, 126.42-126.43 (m), 124.96, 123.13, 122.75, 122.16, 120-100 (m); **¹⁹F NMR** (376 MHz, CDCl₃) δ -79.05 (t, *J* = 10.2 Hz, 3F), -104.57 – -107.21 (m, 2F), -123.66 (t, *J* = 7.2 Hz, 2F); **HRMS** Calcd for C₁₆H₉F₇N [M+H⁺]: 348.0618, Found: 348.0620.



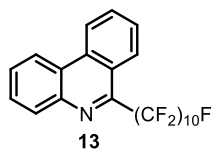
R_f = 0.5, 1% acetone in Hexane

Compound **11** was isolated in 93% yield (92 mg) following the general procedure A. **¹H NMR** (400 MHz, CDCl₃) δ 8.73 (d, *J* = 8.4 Hz, 1H), 8.66-8.58 (m, 1H), 8.47 (d, *J* = 8.4 Hz, 1H), 8.34-8.25 (m, 1H), 7.92 (t, *J* = 7.3 Hz, 1H), 7.86-7.70 (m, 3H); **¹³C NMR** (101 MHz, CDCl₃) δ 146.77 (t, *J* = 24.9 Hz), 141.89, 134.09, 131.32, 131.25, 129.49, 129.40, 128.09, 126.28 (t, *J* = 6.9 Hz), 124.91, 123.13, 122.69, 122.10, 120-100 (m); **¹⁹F NMR** (376 MHz, CDCl₃) δ -80.82 (t, *J* = 11.6 Hz, 3F), -103.30 – -106.62 (m, 2F), -116.99 – -119.18 (m, 2F), -119.18 – -121.05 (m, 2F), -121.36 – -124.22 (m, 2F), -124.82 – -127.82 (m, 2F); **HRMS** Calcd for C₁₉H₉F₁₃N [M+H⁺]: 498.0522, Found: 498.0526.



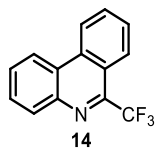
R_f = 0.6, 1% acetone in Hexane

Compound **12** was isolated in 93% yield (111 mg) following the general procedure A. **¹H NMR** (400 MHz, CDCl₃) δ 8.73 (d, *J* = 8.3 Hz, 1H), 8.66-8.58 (m, 1H), 8.47 (d, *J* = 8.4 Hz, 1H), 8.33-8.24 (m, 1H), 7.92 (t, *J* = 7.7 Hz, 1H), 7.86-7.70 (m, 3H); **¹³C NMR** (101 MHz, CDCl₃) δ 146.80 (t, *J* = 24.9 Hz), 141.90, 134.11, 131.33, 131.25, 129.49, 129.40, 128.09, 126.29 (t, *J* = 6.8 Hz), 124.92, 123.15, 122.69, 122.11, 120-100 (m); **¹⁹F NMR** (376 MHz, CDCl₃) δ -80.63 – -81.09 (m, 3F), -104.82 – -105.07 (m, 2F), -118.50 – -119.29 (m, 2F), -119.75 (s, 2F), -121.13 – -122.26 (m, 4F), -122.76 (s, 2F), -125.73 – -126.59 (m, 2F); **HRMS** Calcd for C₂₁H₉F₁₇N [M+H⁺]: 598.0458, Found: 598.0462.



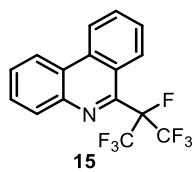
$R_f = 0.7$, 1% acetone in Hexane

Compound **13** was isolated in 94% yield (131 mg) following the general procedure A. **^1H NMR** (400 MHz, CDCl_3) δ 8.72 (d, $J = 8.3$ Hz, 1H), 8.66-8.54 (m, 1H), 8.47 (d, $J = 8.3$ Hz, 1H), 8.35-8.21 (m, 1H), 7.91 (t, $J = 7.6$ Hz, 1H), 7.86-7.65 (m, 3H); **^{13}C NMR** (101 MHz, CDCl_3) δ 146.81 (t, $J = 24.7$ Hz), 141.90, 134.10, 131.33, 131.23, 129.47, 129.38, 128.07, 126.29 (t, $J = 6.8$ Hz), 124.91, 123.15, 122.68, 122.09, 120-100 (m); **^{19}F NMR** (376 MHz, CDCl_3) δ -80.82 (t, $J = 10.6$ Hz, 3F), -104.63 – -105.33 (m, 2F), -119.02 (s, 2F), -119.76 (s, 2F), -120.30 – -122.55 (m, 8F), -122.77 (s, 2F), -125.89 – -126.61 (m, 2F); **HRMS** Calcd for $\text{C}_{23}\text{H}_9\text{F}_{21}\text{N}$ [$\text{M}+\text{H}^+$]: 698.0394, Found: 698.0395.



$R_f = 0.7$, 1% acetone in Hexane

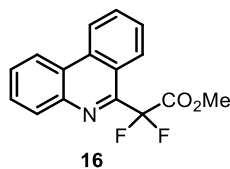
2-Isocyanobiphenyl **1** (36 mg, 0.2 mmol, 1.0 equiv) and N,N,N',N' -Tetraethylenediamine (TEEDA) (52 mg, 0.3 mmol, 1.5 equiv) were dispersed in 2 mL of THF. The reaction vial was then purged with CF_3I for 5 min and sealed with PTEF cap. The reaction mixture was stirred at 30 °C under irradiation of CFL lamp (25W, laid 10 CM away from the vial) for 36 h. Then the mixture was concentrated *in vacuo*, and the residue was purified by silica gel flash chromatography to give compound **14** in 66% yield (33 mg). **^1H NMR** (400 MHz, CDCl_3) δ 8.65 (d, $J = 8.4$ Hz, 1H), 8.60-8.52 (m, 1H), 8.42-8.33 (m, 1H), 8.31-8.24 (m, 1H), 7.89 (t, $J = 7.7$ Hz, 1H), 7.84-7.68 (m, 3H); **^{13}C NMR** (101 MHz, CDCl_3) δ 146.61 (q, $J = 33.0$ Hz), 141.84, 134.05, 131.48, 131.21, 129.44, 129.32, 128.16, 126.02 (q, $J = 3.1$ Hz), 125.21, 123.44, 122.63, 122.17; **^{19}F NMR** (376 MHz, CDCl_3) δ -63.44 (s, 3F); **HRMS** Calcd for $\text{C}_{14}\text{H}_9\text{F}_3\text{N}$ [$\text{M}+\text{H}^+$]: 248.0682, Found: 248.0685.



$R_f = 0.6$, 1% acetone in Hexane

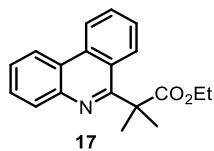
Compound **15** was isolated in 80% yield (56 mg) following the general procedure A.

¹H NMR (400 MHz, CDCl₃) δ 8.73 (d, *J* = 8.3 Hz, 1H), 8.64-8.60 (m, 2H), 8.29-8.14 (m, 1H), 7.90 (t, *J* = 7.6 Hz, 1H), 7.84-7.66 (m, 3H); **¹³C NMR** (101 MHz, CDCl₃) δ 145.90 (d, *J* = 23.9 Hz), 141.71 (d, *J* = 2.6 Hz), 134.25, 131.22, 131.15, 129.31 (d, *J* = 5.5 Hz), 128.05 (d, *J* = 3.3 Hz), 126.60, 126.39, 124.39, 124.25 (d, *J* = 4.0 Hz), 122.78, 122.07; **¹⁹F NMR** (376 MHz, CDCl₃) δ -72.48 (d, *J* = 6.9 Hz, 6F), -174.77 – -175.55 (m, 1F); **HRMS** Calcd for C₁₆H₉F₇N [M+H⁺]: 348.0618, Found: 348.0621.



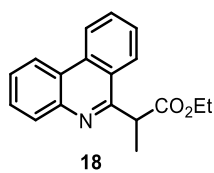
R_f = 0.6, 1% acetone in Hexane

Compound **16** was isolated in 66% yield (38 mg) following the general procedure **B**. **¹H NMR** (400 MHz, CDCl₃) δ 8.71 (d, *J* = 8.4 Hz, 1H), 8.64-8.53 (m, 2H), 8.14 (dt, *J* = 7.1, 3.7 Hz, 1H), 7.93 (t, *J* = 7.4 Hz, 1H), 7.82-7.71 (m, 3H), 4.08 (s, 3H); **¹³C NMR** (101 MHz, CDCl₃) δ 150.19, 141.88, 134.07, 131.43, 131.20, 129.18, 129.11, 128.08, 126.41 (t, *J* = 4.8 Hz), 125.04, 122.69, 122.43, 122.19, 53.622; **¹⁹F NMR** (376 MHz, CDCl₃) δ -98.55 (s, 2F); **HRMS** Calcd for C₁₆H₁₂F₂NO₂ [M+H⁺]: 288.0831, Found: 288.0835.



R_f = 0.6, Hexane

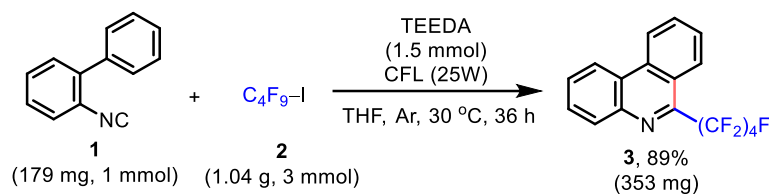
Compound **17** was isolated in 71% yield (42 mg) following the general procedure **B**. **¹H NMR** (400 MHz, CDCl₃) δ 8.67 (d, *J* = 8.3 Hz, 1H), 8.55 (d, *J* = 8.1 Hz, 1H), 8.17 (dd, *J* = 8.1, 0.9 Hz, 1H), 8.05 (d, *J* = 8.4 Hz, 1H), 7.78 (t, *J* = 7.7 Hz, 1H), 7.73 (t, *J* = 7.6 Hz, 1H), 7.67-7.55 (m, 2H), 4.11 (q, *J* = 7.1 Hz, 2H), 1.88 (s, 6H), 1.00 (t, *J* = 7.1 Hz, 3H); **¹³C NMR** (101 MHz, CDCl₃) δ 178.21, 161.34, 143.17, 133.61, 130.47, 129.83, 128.59, 126.96, 126.16, 124.41, 123.91, 122.95, 121.88, 100.07, 61.12, 50.05, 26.70, 14.00; **HRMS** Calcd for C₁₉H₂₀NO₂ [M+H⁺]: 294.1489, Found: 294.1490.



R_f = 0.7, Hexane

Compound **18** was isolated in 94% yield (53 mg) following the general procedure **B**. **¹H NMR** (400 MHz, CDCl₃) δ 8.65 (d, *J* = 8.3 Hz, 1H), 8.54 (d, *J* = 8.1 Hz, 1H), 8.22 (d, *J* = 8.3 Hz, 1H), 8.16 (d, *J* = 8.1 Hz, 1H), 7.82 (t, *J* = 7.6 Hz, 1H), 7.76-7.59 (m, 3H), 4.75 (q, *J* = 7.1 Hz, 1H), 4.31-4.05 (m, 2H), 1.79 (d, *J* = 7.1 Hz, 3H), 1.16 (t, *J* = 7.1 Hz, 3H); **¹³C NMR** (101 MHz, CDCl₃) δ 173.83, 159.64, 143.60, 133.33, 130.44, 130.27, 128.68, 127.48, 126.92, 125.68, 124.72, 123.80, 122.76, 121.94, 61.06, 45.67, 16.55, 14.23; **HRMS** Calcd for C₁₈H₁₈NO₂ [M+H⁺]: 280.1332, Found: 280.1335.

1 mmol scale reaction of 2-Isocyanobiphenyl 1 with perfluorobutyl iodide 2.

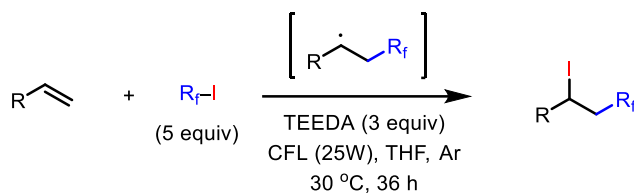


Scheme S19

In a 40 mL borosilicate glass vial, 2-isocyanobiphenyl **1** (179 mg, 1.0 mmol, 1.0 equiv), perfluorobutyl iodide **2** (1.04 g, 3.0 mmol, 3.0 equiv) and *N,N,N',N'*-tetraethylenediamine (TEEDA) (258 mg, 1.5 mmol, 1.5 equiv) were dispersed in 10 mL of THF. The reaction vial was then purged with Ar for 1 min and sealed with PTEF cap. The reaction mixture was vigorously stirred at 30 °C under irradiation of CFL lamp (25W, laid 10 CM away from the vial) and monitored by TLC analysis. After 36 hour, all the 2-Isocyanobiphenyl **1** was consumed. Then the mixture was concentrated *in vacuo*, and the residue was purified by silica gel flash chromatography to give compound **3** in 89% yield (353 mg).

7. General procedures and substrate scope for addition of perfluorobutyl iodide to alkenes and alkynes

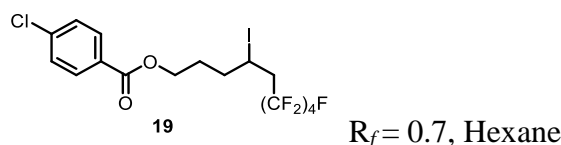
7.1 General procedure and substrate scope for addition of perfluorobutyl iodide to alkenes



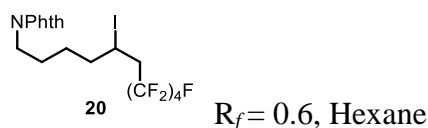
Scheme S20

General procedure C for the synthesis of compound 19, 20, 21, 22, 23, and 24:

Alkene (0.2 mmol, 1.0 equiv), perfluoroalkyl iodide (1.0 mmol, 5.0 equiv) and *N,N,N',N'*-tetraethylenediamine (TEEDA) (0.6 mmol, 3.0 equiv) were dispersed in 2 mL of THF. The reaction vial was then purged with Ar for 1 min and sealed with PTEF cap. The reaction mixture was vigorously stirred at 30 °C under irradiation of CFL lamp (25W, laid 10 CM away from the vial) for 36 h. Then the mixture was concentrated *in vacuo*, and the residue was purified by silica gel flash chromatography to give the desired product.

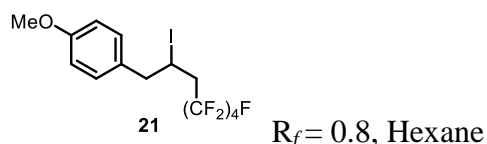


Compound **19** was isolated in 90% yield (103 mg) following the general procedure **C** (89% yield (102 mg) was obtained under the irradiation of UV (254nm) lamp for 20 hours; 74% yield (84 mg) was obtained after irradiated in sunlight for 4 hours). **¹H NMR** (400 MHz, CDCl₃) δ 7.96 (d, $J = 8.6$ Hz, 2H), 7.42 (d, $J = 8.6$ Hz, 2H), 4.47-4.29 (m, 3H), 3.15-2.61 (m, 2H), 2.12-1.86 (m, 4H); **¹³C NMR** (101 MHz, CDCl₃) δ 165.78, 139.67, 131.08, 128.91, 128.65, 63.93, 41.71 (t, $J = 21.1$ Hz), 36.91 (d, $J = 2.9$ Hz), 29.05, 19.52; **¹⁹F NMR** (376 MHz, CDCl₃) δ -80.11 – -81.80 (m, 3F), -109.86 – -112.76 (m, 1F), -113.86 – -116.17 (m, 1F), -124.42 (s, 2F), -125.83 (q, $J = 15.8$ Hz, 2F); **HRMS** Calcd for C₁₆H₁₃ClF₉IO₂Na [M+Na⁺]: 592.9397, Found: 592.9400.

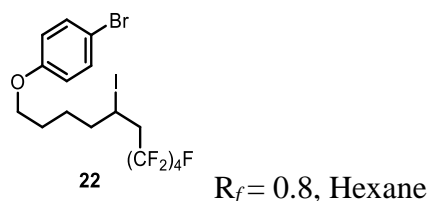


Compound **20** was isolated in 86% yield (99 mg) following the general procedure **C**. **¹H NMR** (400 MHz, CDCl₃) δ 7.89-7.80 (m, 2H), 7.76-7.65 (m, 2H), 4.39-4.18 (m,

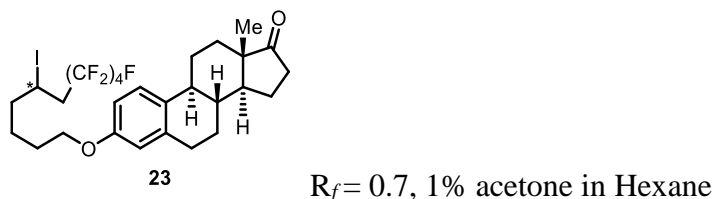
1H), 3.70 (t, $J = 7.1$ Hz, 2H), 3.04-2.59 (m, 2H), 1.90-1.63 (m, 5H), 1.54-1.36 (m, 1H); **^{13}C NMR** (101 MHz, CDCl_3) δ 168.53, 134.10, 132.23, 123.38, 41.72 (t, $J = 20.9$ Hz), 39.81, 37.66, 27.60, 27.09, 20.06; **^{19}F NMR** (376 MHz, CDCl_3) δ -79.40 – -82.59 (m, 3F), -113.24 – -116.44 (m, 2F), -123.29 – -124.90 (m, 2F), -125.30 – -127.56 (m, 2F); **HRMS** Calcd for $\text{C}_{18}\text{H}_{15}\text{F}_9\text{INO}_2\text{Na}$ [$\text{M}+\text{Na}^+$]: 597.9896, Found: 597.9902.



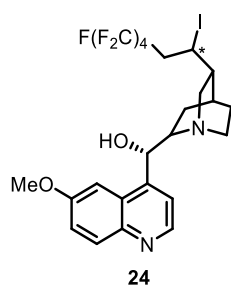
Compound **21** was isolated in 90% yield (89 mg) following the general procedure **C**. **^1H NMR** (400 MHz, CDCl_3) δ 7.12 (d, $J = 8.6$ Hz, 2H), 6.88 (d, $J = 8.7$ Hz, 2H), 4.46-4.38 (m, 1H), 3.81 (s, 3H), 3.25-3.11 (m, 2H), 3.02-2.72 (m, 2H); **^{13}C NMR** (101 MHz, CDCl_3) δ 158.95, 130.79, 130.18, 114.12, 100.13, 55.37, 46.37, 20.29; **^{19}F NMR** (376 MHz, CDCl_3) δ -81.01 (s, 3F), -110.65 – -113.14 (m, 1F), -113.19 – -115.24 (m, 1F), -124.55 (s, 2F), -125.93 (s, 2F); **HRMS** Calcd for $\text{C}_{14}\text{H}_{12}\text{F}_9\text{IONa}$ [$\text{M}+\text{Na}^+$]: 516.9681, Found: 516.9680.



Compound **22** was isolated in 91% yield (109 mg) following the general procedure **C**. **^1H NMR** (400 MHz, CDCl_3) δ 7.37 (d, $J = 9.0$ Hz, 2H), 6.77 (d, $J = 9.0$ Hz, 2H), 4.38-4.32 (m, 1H), 3.95 (t, $J = 6.1$ Hz, 2H), 3.07-2.62 (m, 2H), 1.98-1.69 (m, 5H), 1.68-1.56 (m, 1H); **^{13}C NMR** (101 MHz, CDCl_3) δ 158.15, 132.38, 116.40, 112.95, 67.81, 41.69 (t, $J = 20.9$ Hz), 40.06, 28.23, 26.52, 20.34; **^{19}F NMR** (376 MHz, CDCl_3) δ -81.19 (s, 3F), -110.63 – -113.25 (m, 1F), -114.02 – -116.26 (m, 1F), -124.73 (s, 2F), -126.10 (q, $J = 14.8$ Hz, 2F); **HRMS** Calcd for $\text{C}_{16}\text{H}_{15}\text{BrF}_9\text{IONa}$ [$\text{M}+\text{Na}^+$]: 622.9099, Found: 622.9096.



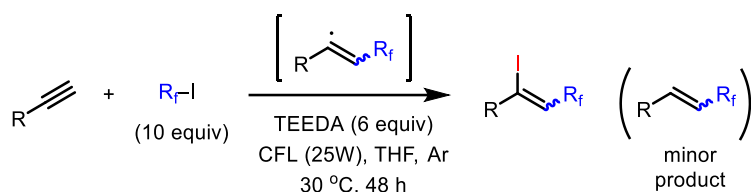
Compound **23** was isolated in 80% yield (112 mg) following the general procedure **C**. **¹H NMR** (400 MHz, CDCl₃) δ 7.20 (d, *J* = 8.6 Hz, 1H), 6.72 (dd, *J* = 8.5, 2.4 Hz, 1H), 6.65 (d, *J* = 2.2 Hz, 1H), 4.44-4.22 (m, 1H), 3.96 (t, *J* = 6.0 Hz, 2H), 3.03-2.69 (m, 4H), 2.56- 2.36 (m, 2H), 2.25 (t, *J* = 10.3 Hz, 1H), 2.21-1.69 (m, 10H), 1.68-1.36 (m, 6H), 0.91 (s, 3H); **¹³C NMR** (101 MHz, CDCl₃) δ 220.99, 157.07, 137.88, 132.20, 126.45, 114.70, 112.21, 67.50, 50.54, 48.13, 44.11, 41.69 (t, *J* = 20.8 Hz), 40.13, 38.50, 35.98, 31.71, 29.77, 28.36, 26.68, 26.61, 26.05, 21.70, 20.35, 13.96; **¹⁹F NMR** (376 MHz, CDCl₃) δ -80.82 (s, 3F), -109.99 – -112.70 (m, 1F), -113.59 – -116.17 (m, 1F), -124.36 (s, 2F), -125.71 (m, 2F); **HRMS** Calcd for C₂₈H₃₃F₉IO₂ [M+H⁺]: 699.1376, Found: 699.1379.



R_f = 0.7, 5% CH₃OH in CH₂Cl₂

Compound **24** was isolated in 70% yield (94 mg) following the general procedure **C**. **¹H NMR** (400 MHz, CDCl₃) δ 8.62 (d, *J* = 3.8 Hz, 1H), 8.00 (d, *J* = 9.1 Hz, 1H), 7.54 (d, *J* = 3.8 Hz, 1H), 7.44-7.32 (m, 1H), 5.65 (s, 1H), 4.11-4.02 (m, 1H), 4.01-3.90 (m, 3H), 3.59 (s, 1H), 3.30-3.09 (m, 2H), 3.08-2.78 (m, 2H), 2.70 (d, *J* = 11.5 Hz, 1H), 2.59 (d, *J* = 10.1 Hz, 1H), 2.14 (s, 2H), 1.88 (s, 2H), 1.53 (s, 1H), 1.44-1.24 (m, 2H); **¹³C NMR** (101 MHz, CDCl₃) δ 158.07, 147.54, 147.40, 144.15, 131.55, 126.54, 121.75, 118.46, 101.38, 71.33, 61.94, 60.12, 56.00, 44.24, 43.11, 39.05 (t, *J* = 20.8 Hz), 27.51, 26.58, 24.16, 20.03; **¹⁹F NMR** (376 MHz, CDCl₃) δ -80.96 (s, 3F), -110.95 – -116.63 (m, 2F), -123.23 – -125.12 (m, 2F), -125.28 – -126.74 (m, 2F); **HRMS** Calcd for C₂₄H₂₅F₉IN₂O₂ [M+H⁺]: 671.0812, Found: 671.0815.

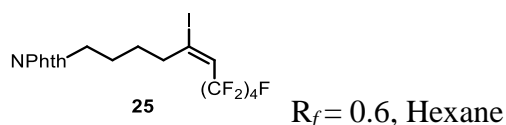
7.2 General procedure and substrate scope for addition of perfluorobutyl iodide to alkynes



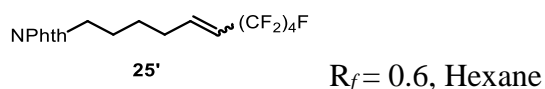
Scheme S21

General procedure D for the synthesis of compound 25 and 26:

Alkynes (0.2 mmol, 1.0 equiv), perfluoroalkyl iodide (2.0 mmol, 10.0 equiv) and *N,N,N',N'*-tetraethylenediamine (TEEDA) (1.2 mmol, 6.0 equiv) were dispersed in 2 mL of THF. The reaction vial was purged with Ar for 1 min and then sealed with PTEF cap. The reaction mixture was vigorously stirred at 30 °C under irradiation of CFL lamp (25W, laid 10 CM away from the vial) for 36 h. After that, the mixture was concentrated *in vacuo*, and the residue was purified by silica gel flash chromatography to give the desired product.

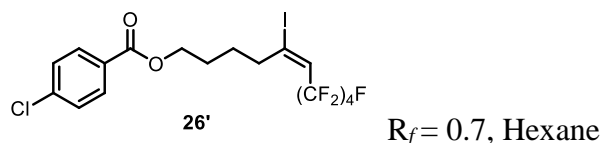


Compound **25**⁵ was isolated in 72% yield (83 mg) following the general procedure **D**. **¹H NMR** (400 MHz, CDCl₃) δ 7.85-7.82 (m, 2H), 7.75-7.66 (m, 2H), 6.33 (t, *J* = 14.4 Hz, 1H), 3.71 (t, *J* = 6.8 Hz, 2H), 2.67 (t, *J* = 6.6 Hz, 2H), 1.82-1.55 (m, 4H); **¹⁹F NMR** (376 MHz, CDCl₃) δ -81.58 (s, 3F), -106.10 (s, 2F), -124.75 (s, 2F), -126.39 (s, 2F); **HRMS** Calcd for C₁₈H₁₃F₉INO₂Na [*M*+Na⁺]: 595.9739, Found: 595.9846.

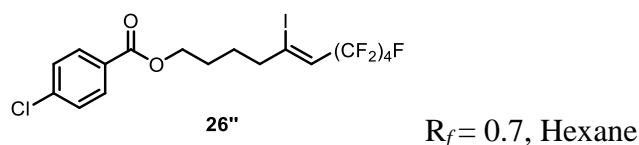


Compound **25'** was isolated in 19% yield (17 mg) following the general procedure **D**. **¹H NMR** (400 MHz, CDCl₃) δ 7.90-7.80 (m, 2H), 7.76-7.67 (m, 2H), 6.41-6.34 (m, 0.7 H), 6.12-6.03 (m, 0.3 H), 5.75-5.37 (m, 1H), 3.80-3.57 (m, 2H), 2.43-2.12 (m, 2H), 1.78-1.61 (m, 2H), 1.54-1.46 (m, 2H); **¹³C NMR** (101 MHz, CDCl₃) δ 168.54, 144.84, 142.60 (t, *J* = 8.8 Hz), 134.09, 132.26, 123.38, 117.45 (t, *J* = 22.9 Hz), 116.69 (t, *J* = 23.4 Hz), 37.79, 37.61, 31.60, 28.21, 28.08, 26.42, 25.38; **¹⁹F NMR** (376 MHz, CDCl₃) δ -80.10 – -81.73 (m, 3F), -106.45 – -107.38 (m, 0.6), -110.35 – -112.81 (m, 1.4), -

123.25 – -125.04 (m, 2F), -124.99 – -126.54 (m, 2F); **HRMS** Calcd for C₁₈H₁₄F₉NO₂Na [M+Na⁺]: 470.0773, Found: 470.0778.

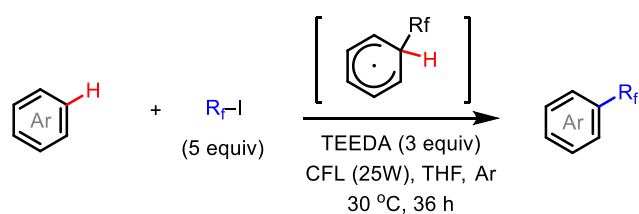


Compound **26'**⁵ was isolated in 61% yield (71 mg) following the general procedure **D**. **¹H NMR** (400 MHz, CDCl₃) δ 7.98 (d, $J = 8.2$ Hz, 2H), 7.41 (d, $J = 8.2$ Hz, 2H), 6.37 (t, $J = 14.4$ Hz, 1H), 4.34 (t, $J = 5.6$ Hz, 2H), 2.72 (t, $J = 7.6$ Hz, 2H), 1.87-1.70 (m, 4H); **¹⁹F NMR** (376 MHz, CDCl₃) δ -77.39 – -83.32 (m, 3F), -102.67 – -107.14 (m, 2F), -120.37 – -124.27 (m, 2F), -124.03 – -127.69 (m, 2F).



Compound **26''**⁵ was isolated in 11% yield (13 mg) following the general procedure **D**. **¹H NMR** (400 MHz, CDCl₃) δ 7.98 (d, $J = 8.4$ Hz, 2H), 7.42 (d, $J = 8.4$ Hz, 2H), 6.29 (t, $J = 13.0$ Hz, 2H), 4.34 (s, 3H), 2.74 (s, 2H), 1.77 (m, 4H); **¹⁹F NMR** (376 MHz, CDCl₃) δ -80.46 (s, 3F), -108.33 (s, 2F), -123.34 (s, 2F), -125.29 (s, 2F).

8. General procedure and substrate scope for C-H perfluorobutylation of electron-rich arene and heteroarene

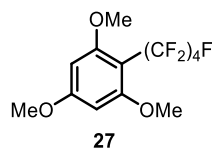


Scheme S22

General procedure E for the synthesis of compound 27-37:

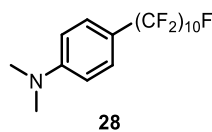
Electron-rich arene or heteroarene (0.2 mmol, 1.0 equiv), perfluoroalkyl iodide (1.0 mmol, 5.0 equiv) and *N,N,N',N'*-tetraethylenediamine (TEEDA) (0.6 mmol, 3.0 equiv) were dispersed in 2 mL of THF. The reaction vial was purged with Ar for 1 min then sealed with PTEF cap. The reaction mixture was vigorously stirred at 30 °C under

irradiation of CFL lamp (25W, laid 10 CM away from the vial) for 36 h. After that, the mixture was concentrated *in vacuo*, and the residue was purified by silica gel flash chromatography to give the desired product.



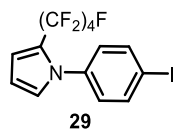
$R_f = 0.4$, Hexane

Compound **27** was isolated in 56% yield (43 mg; and 32% of starting material **27-1** was recovered) following the general procedure **E**. **^1H NMR** (400 MHz, CDCl_3) δ 6.14 (s, 2H), 3.84 (s, 3H), 3.80 (s, 6H); **^{13}C NMR** (101 MHz, CDCl_3) δ 163.98, 161.88 (t, $J = 2.2$ Hz), 98.60 (t, $J = 21.8$ Hz), 91.79, 56.44, 55.47; **^{19}F NMR** (376 MHz, CDCl_3) δ -80.67 (t, $J = 11.6$ Hz, 3F), -101.07 – -104.45 (m, 2F), -122.76 – -123.27 (q, $J = 9.6$ Hz, 2F), -125.36 – -127.17 (m, 2F); **HRMS** Calcd for $\text{C}_{13}\text{H}_{12}\text{F}_9\text{O}_3$ $[\text{M}+\text{H}^+]$: 387.0637, Found: 387.0640.



$R_f = 0.8$, Hexane

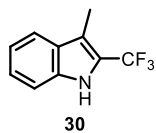
Compound **28** was isolated in 52% yield (67 mg; and 34% of starting material **28-1** was recovered) following the general procedure **E**. **^1H NMR** (400 MHz, CDCl_3) δ 7.40 (d, $J = 8.8$ Hz, 2H), 6.72 (d, $J = 8.9$ Hz, 2H), 3.02 (s, 6H); **^{13}C NMR** (101 MHz, CDCl_3) δ 152.53, 128.01 (t, $J = 6.3$ Hz), 115.43, 111.28, 40.18; **^{19}F NMR** (376 MHz, CDCl_3) δ -80.06 – -81.37 (m, 3F), -108.23 – -110.06 (m, 2F), -118.80 – -124.76 (m, 14F), -125.57 – -127.21 (m, 2F); **HRMS** Calcd for $\text{C}_{18}\text{H}_{11}\text{F}_{21}\text{N}$ $[\text{M}+\text{H}^+]$: 640.0551, Found: 640.0557.



$R_f = 0.7$, Hexane

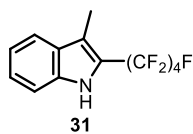
Compound **29** was isolated in 55% yield (54 mg; and 40% of starting material **29-1** was recovered) following the general procedure **E**. **^1H NMR** (400 MHz, CDCl_3) δ 7.76 (d, $J = 8.0$ Hz, 2H), 7.09 (d, $J = 8.0$ Hz, 2H), 6.84 (s, 1H), 6.74 (s, 1H), 6.33 (s, 1H); **^{13}C NMR** (101 MHz, CDCl_3) δ 139.48, 138.04, 129.28, 129.04, 120.13 (t, $J = 29.9$ Hz),

115.67, 109.27, 94.26; **¹⁹F NMR** (376 MHz, CDCl₃) δ -80.73 (t, *J* = 11.2 Hz, 3F), -99.87 – -101.89 (m, 2F), -121.16 (q, *J* = 9.5 Hz, 2F), -124.11 – -127.25 (m, 2F); **HRMS** Calcd for C₁₄H₇F₉IN [M+H⁺]: 486.9479, Found:486.9483.



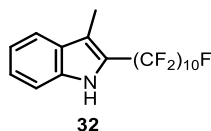
R_f = 0.4, 2% acetone in Hexane

3-Methyl indole **30-1** (0.2 mmol, 1.0 equiv) and *N,N,N',N'*-tetraethylenediamine (TEEDA) (0.6 mmol, 3.0 equiv) were dispersed in 2 mL of THF. The reaction vial was purged with CF₃I for 5 min and then sealed with PTEF cap. The reaction mixture was stirred at 30 °C under the irradiation of CFL lamp (25W, laid 10 CM away from the vial) for 36 h. After that, the mixture was concentrated *in vacuo*, and the residue was purified by silica gel flash chromatography to give compound **30**¹⁰ in 66% yield (26 mg). **¹H NMR** (400 MHz, CDCl₃) δ 8.19 (s, 1H), 7.64 (d, *J* = 8.0 Hz, 1H), 7.39 (d, *J* = 8.2 Hz, 1H), 7.32 (t, *J* = 7.6 Hz, 1H), 7.19 (t, *J* = 7.5 Hz, 1H), 2.44 (d, *J* = 1.7 Hz, 3H); **¹⁹F NMR** (376 MHz, CDCl₃) δ -58.10 (s, 3F).



R_f = 0.4, 2% acetone in Hexane

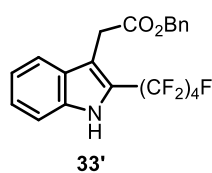
Compound **31** was isolated in 80% yield (56 mg) following the general procedure **E**. **¹H NMR** (400 MHz, CDCl₃) δ 8.18 (s, 1H), 7.68 (d, *J* = 8.0 Hz, 1H), 7.40 (d, *J* = 8.2 Hz, 1H), 7.35 (t, *J* = 7.5 Hz, 1H), 7.22 (t, *J* = 7.2 Hz, 1H), 2.46 (s, 3H); **¹³C NMR** (101 MHz, CDCl₃) δ 136.07, 128.46, 125.09, 120.52, 120.22, 119.54 116.86 (t, *J* = 3.2 Hz), 111.65, 8.71; **¹⁹F NMR** (376 MHz, CDCl₃) δ -77.96 – -83.88 (m, 3F), -106.64 – -111.68 (m, 2F), -120.94 – -124.85 (m, 2F), -124.85 – -127.69 (m, 2F); **HRMS** Calcd for C₁₃H₇F₉N [M-H⁺]: 348.0440, Found: 348.0438.



R_f = 0.5, 2% acetone in Hexane

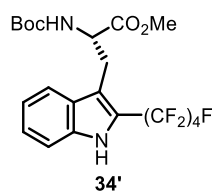
Compound **32** was isolated in 91% yield (118 mg) following the general procedure **E** (90% yield (117 mg) was obtained under the irradiation of UV lamp (254 nm, 25W) for

36 hours; 68% yield (88 mg) was obtained after irradiated in sunlight for 4 hours). **¹H NMR** (400 MHz, CDCl₃) δ 8.19 (s, 1H), 7.66 (d, *J* = 8.0 Hz, 1H), 7.40 (d, *J* = 8.2 Hz, 1H), 7.34 (t, *J* = 7.5 Hz, 1H), 7.21 (t, *J* = 7.2 Hz, 1H), 2.44 (s, 3H); **¹³C NMR** (101 MHz, CDCl₃) δ 136.08, 128.47, 125.08, 120.50, 120.22, 119.67 (t, *J* = 28.4 Hz), 116.84 (t, *J* = 3.3 Hz), 111.65, 8.65; **¹⁹F NMR** (376 MHz, CDCl₃) δ -80.42 – -81.05 (m, 3F), -108.59 (s, 2F), -119.29 – -121.97 (m, 10F), -122.11 (m, 2F), -122.70 (m, 2F), -126.08 (m, 2F); **HRMS** Calcd for C₁₉H₇F₂₁N [M-H⁺]: 648.0248, Found: 648.0238.



R_f = 0.4, 2% acetone in Hexane

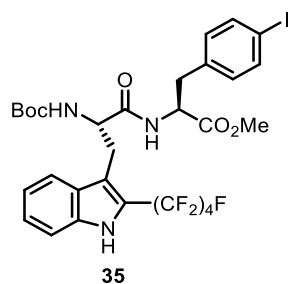
Compound **33-1** (0.2 mmol, 1.0 equiv), perfluoroalkyl iodide **2** (1.0 mmol, 5.0 equiv) and *N,N,N',N'*-tetraethylenediamine (TEEDA) (0.6 mmol, 3.0 equiv) were dispersed in 2 mL of THF. The reaction vial was purged with Ar for 1 min and then sealed with PTEF cap. The reaction mixture was stirred at 30 °C under the irradiation of CFL lamp (25W, laid 10 CM away from the vial) for 36 h. After that, the mixture was concentrated *in vacuo*, and the residue was dissolved in acetone (15 mL). BnBr (0.3 mmol, 1.5 equiv) was added, the resulting mixture was stirred at 30 °C for another 12 hour. Then the mixture was concentrated *in vacuo*, and the residue purified by silica gel flash chromatography to give compound **33'** in 67% yield (65 mg). **¹H NMR** (400 MHz, CDCl₃) δ 8.48 (s, 1H), 7.65 (d, *J* = 8.1 Hz, 1H), 7.43-7.27 (m, 7H), 7.20 (t, *J* = 7.3 Hz, 1H), 5.16 (s, 2H), 3.98 (s, 2H); **¹³C NMR** (101 MHz, CDCl₃) δ 170.60, 136.07, 135.82, 128.61, 128.33, 128.24, 127.73, 125.37, 121.17, 120.47, 113.09, 111.88, 66.96, 30.16; **¹⁹F NMR** (376 MHz, CDCl₃) δ -80.10 – -80.89 (m, 3F), -106.79 – -108.95 (m, 2F), -120.92 – -123.91 (m, 2F), -124.09 – -127.75 (m, 2F); **HRMS** Calcd for C₂₁H₁₈F₉N₂O₂ [M+NH₄⁺]: 501.1219, Found: 501.1216.



R_f = 0.4, 4% acetone in Hexane

Compound **34-1** (0.2 mmol, 1.0 equiv), perfluoroalkyl iodide **2** (1.0 mmol, 5.0 equiv)

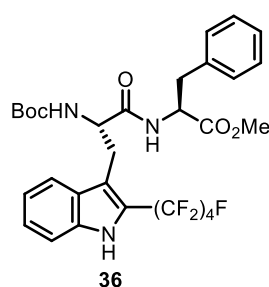
and *N,N,N',N'*-tetraethylenediamine (TEEDA) (0.6 mmol, 3.0 equiv) were dispersed in 2 mL of THF. The reaction vial was purged with Ar for 1 min and then sealed with PTEF cap. The reaction mixture was stirred at 30 °C under the irradiation of CFL lamp (25W, laid 10 CM away from the vial) for 36 hour. After that, the mixture was concentrated *in vacuo*, and the residue was dissolved in THF (2 mL). Boc₂O (2.0 mmol, 10.0 equiv), DIPEA (0.4 mmol, 2.0 equiv) were added, and the resulting mixture was stirred at 30 °C for another 12 hour. The reaction was quenched with 0.3 M HCl 20 mL and extracted with dichloromethane. Then organic phase was separated, washed with brine, dried over anhydrous Na₂SO₄, and concentrated *in vacuo*. The residue was purified by silica gel flash chromatography to give compound **34'** in 65% yield (70 mg). **¹H NMR** (400 MHz, CDCl₃) δ 8.68 (s, 1H), 7.75 (d, *J* = 8.0 Hz, 1H), 7.38 (d, *J* = 8.1 Hz, 1H), 7.32 (t, *J* = 7.5 Hz, 1H), 7.19 (t, *J* = 7.4 Hz, 1H), 5.15 (d, *J* = 8.3 Hz, 1H), 4.67 (q, *J* = 7.1 Hz, 1H), 3.64 (s, 3H), 3.40-3.24 (m, 2H), 1.36 (s, 9H); **¹³C NMR** (101 MHz, CDCl₃) δ 172.60, 155.12, 136.27, 129.69, 127.66, 125.31, 121.03, 120.47, 115.47, 111.91, 80.03, 54.26, 52.37, 29.35, 28.29, 28.03; **¹⁹F NMR** (376 MHz, CDCl₃) δ -80.67 (t, *J* = 11.0 Hz, 3F), -105.25 – -109.24 (m, 2F), -121.92 – -123.14 (m, 2F), -125.03 – -126.30 (m, 2F); **HRMS** Calcd for C₂₁H₂₁F₉N₂O₄Na [M+Na⁺]: 559.1250, Found: 559.1252



R_f = 0.3, 20% acetone in Hexane

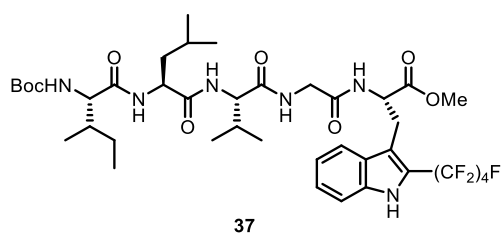
Compound **35** was isolated in 68% yield (110 mg; and 23% of starting material **35-1** was recovered) following the general procedure E. **¹H NMR** (400 MHz, CDCl₃) δ 8.51 (s, 1H), 7.81 (d, *J* = 7.6 Hz, 1H), 7.52 (d, *J* = 8.0 Hz, 2H), 7.40 (d, *J* = 8.2 Hz, 1H), 7.33 (t, *J* = 7.4 Hz, 1H), 7.21 (t, *J* = 7.4 Hz, 1H), 6.71 (d, *J* = 8.1 Hz, 2H), 6.08 (d, *J* = 6.7 Hz, 1H), 5.16 (s, 1H), 4.67 (q, *J* = 12.2, 1H), 4.50-4.30 (m, 1H), 3.58 (s, 3H), 3.28 (d, *J* = 5.2 Hz, 2H), 3.06-2.83 (m, 2H), 1.37 (s, 9H); **¹³C NMR** (101 MHz, CDCl₃) δ 170.98,

170.83, 155.28, 137.66, 136.31, 135.39, 131.36, 127.36, 125.54, 121.30, 120.93, 120.65, 120.36, 115.73, 111.88, 92.75, 80.26, 55.35, 53.17, 52.40, 37.70, 28.29; **¹⁹F NMR** (376 MHz, CDCl₃) δ -79.81 – -81.92 (m, 3F), -105.42 – -109.99 (m, 2F), -122.70 (d, J = 10.3 Hz, 2F), -124.60 – -127.32 (m, 2F); **HRMS** Calcd for C₃₀H₂₉F₉IN₃O₅Na [M+Na⁺]: 832.0900, Found: 832.0905.



R_f = 0.3, 20% acetone in Hexane

Compound **36** was isolated in 65% yield (89 mg; and 20% of starting material **36-1** was recovered) following the general procedure **E**. **¹H NMR** (400 MHz, CDCl₃) δ 9.40 (s, 1H), 7.79 (d, J = 6.7 Hz, 1H), 7.40 (d, J = 8.2 Hz, 1H), 7.29 (t, J = 7.5 Hz, 1H), 7.23-7.14 (m, 4H), 6.98 (s, 2H), 6.20 (d, J = 5.8 Hz, 1H), 5.16 (d, J = 6.1 Hz, 1H), 4.73 (q, J = 13.3, 5.9 Hz, 1H), 4.45 (d, J = 6.3 Hz, 1H), 3.58 (s, 3H), 3.37-3.15 (m, 2H), 3.07-2.94 (m, 2H), 2.81 (s, 9H); **¹³C NMR** (101 MHz, CDCl₃) δ 171.24, 170.89, 165.95, 155.22, 136.58, 135.74, 129.33, 128.54, 127.37, 127.15, 125.16, 120.94, 120.45, 115.38, 112.05, 80.09, 55.45, 53.37, 52.20, 38.66, 38.18, 28.16; **¹⁹F NMR** (376 MHz, CDCl₃) δ -80.90 (s, 3F), -104.70 – -110.11 (m, 2F), -122.68 (s, 2F), -125.82 (s, 2F); **HRMS** Calcd for C₃₀H₃₀F₉N₃O₅Na [M+Na⁺]: 706.1934, Found: 706.1938.



R_f = 0.3, 5% CH₃OH in DCM

Compound **37-1** (0.1 mmol, 1.0 equiv), C₄F₉I (0.5 mmol, 5.0 equiv) and *N,N,N',N'*-tetraethylenediamine (TEEDA) (0.3 mmol, 3.0 equiv) were dispersed in 2 mL of THF. The reaction vial was purged with Ar for 1 min and the sealed with PTEF cap. The reaction mixture was stirred at 30 °C under the irradiation of CFL lamp (25W, laid 10 CM away from the vial). After 12 hour, C₄F₉I (0.5 mmol, 5.0 equiv) and *N,N,N',N'*-

tetraethylenediamine (TEEDA) (0.3 mmol, 3.0 equiv) were added and the resulting mixture was stirred at 30 °C under the irradiation of CFL lamp (25W, laid 10 CM away from the vial) for another 24 hour under Ar atmosphere. After that, the mixture was concentrated *in vacuo*, and the residue was purified by silica gel flash chromatography to give the compound **37** in 50% yield (46 mg; and 35% of the starting material compound **37-1** was recovered). **¹H NMR** (400 MHz, MeOD) δ 7.66 (d, J = 8.1 Hz, 1H), 7.41 (d, J = 8.2 Hz, 1H), 7.25 (t, J = 7.5 Hz, 1H), 7.12 (t, J = 7.5 Hz, 1H), 4.68 (t, J = 7.4 Hz, 1H), 4.50 (s, 1H), 4.10 (d, J = 7.1 Hz, 1H), 3.92-3.73 (m, 3H), 3.44 (s, 3H), 3.42-3.31 (m, 2H), 2.09-1.97 (m, 1H), 1.78-1.48 (m, 5H), 1.40 (s, 9H), 1.18-1.08 (m, 1H), 0.97-0.78 (m, 18H); **¹³C NMR** (101 MHz, MeOD) δ 174.56, 173.90, 173.33, 170.89, 157.91, 138.19, 128.65, 125.69, 121.34, 120.93, 115.65, 113.14, 80.51, 60.54, 55.25, 52.87, 52.57, 43.20, 41.67, 38.07, 31.79, 28.70, 28.09, 25.89, 25.67, 23.46, 22.02, 19.71, 18.94, 16.06, 11.36; **¹⁹F NMR** (376 MHz, CDCl₃) δ -78.51 (s, 3F), -101.53 – -108.27 (m, 2F), -119.77 (s, 2F), -123.06 (s, 2F); **HRMS** Calcd for C₄₀H₅₅F₉N₆O₈Na [M+Na⁺]: 941.3830, Found: 941.3832.

9. Titration experiment of C₁₀F₂₁I with TEEDA

¹⁹F NMR spectra of eight samples of mixtures of C₁₀F₂₁I and TEEDA in CDCl₃ were recorded at 298 K. The total volume of the mixture was 0.5 mL, the amount of C₁₀F₂₁I was kept constant at 0.01 mmol (6.5 mg) while that of TEEDA was varied from 0 to 0.1 mmol. The molar ratios of C₁₀F₂₁I:TEEDA were 1:0, 1:1, 1:2, 1:3, 1:4, 1:5, 1:7, 1:10. Fluorobenzene ($\delta_{\text{F-Ph}}$ = -113.066) was used as internal standard.

The ¹⁹F NMR signal of -CF₂I shifted upfield when the amount of TEEDA increased, indicating the formation of halogen bond between C₁₀F₂₁I and TEEDA (Figure S1).

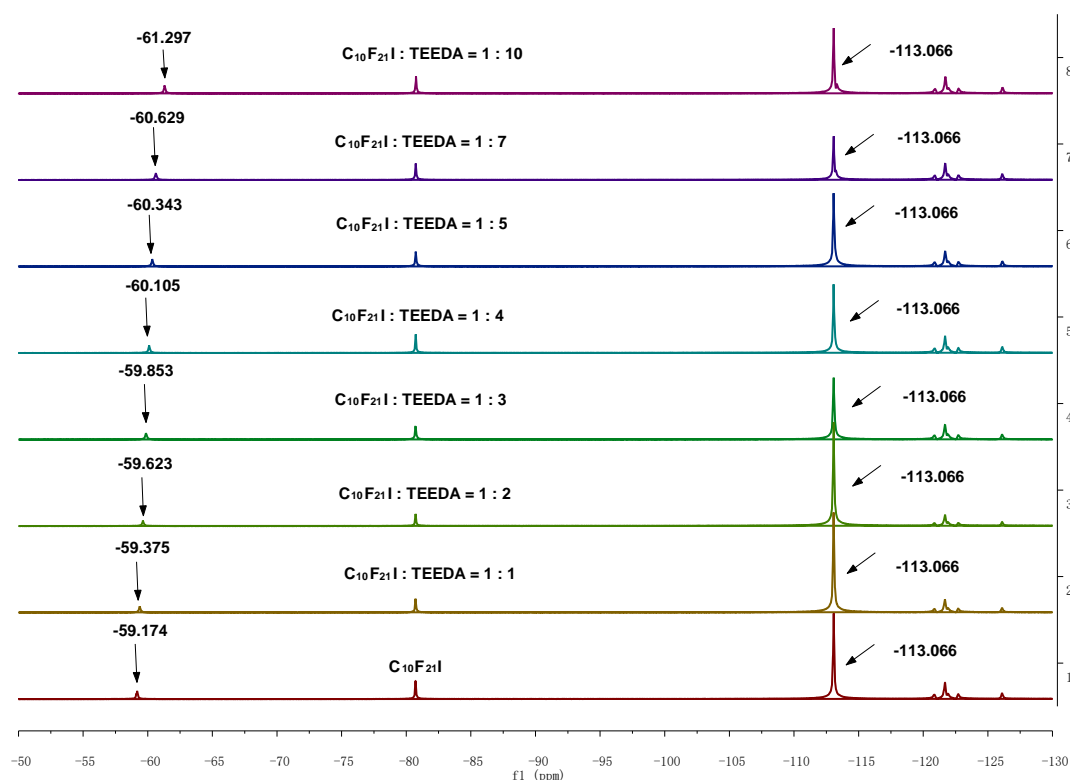


Figure S1. ^{19}F NMR shift of $\text{C}_{10}\text{F}_{21}\text{I}$ with TEEDA

10. Determination of binding stoichiometry of halogen bond complex¹¹

The binding stoichiometry between $\text{C}_{10}\text{F}_{21}\text{I}$ and halogen bond acceptors (XB acceptor: TEEDA, TEA, THF, dioxane) were evaluated using Job's plot analysis¹⁰: ^{19}F NMR spectra of eleven samples of mixtures of $\text{C}_{10}\text{F}_{21}\text{I}$ and XB acceptor in CDCl_3 were recorded at 298 K. Fluorobenzene ($\delta_{\text{F-Ph}} = -113.066$) was used as internal standard. The total volume of the mixture was 0.5 mL, and the total amount of $\text{C}_{10}\text{F}_{21}\text{I}$ and halogen bond acceptor was kept constant at 0.25 mmol (0.5 M), while the amount of $\text{C}_{10}\text{F}_{21}\text{I}$ was varied from 0 to 0.25 mmol (0-0.5 M). The molar ratios of $\text{C}_{10}\text{F}_{21}\text{I}/(\text{C}_{10}\text{F}_{21}\text{I} + \text{XB acceptor})$ were 0.0, 0.1, 0.2, 0.3, 0.4, 0.5, 0.6, 0.7, 0.8, 0.9, 1.0. ^{19}F NMR for each sample was recorded and the chemical shifts differences ($\Delta\delta$) for $-\text{CF}_2\text{I}$ were used to draw the plot. The stoichiometry was determined by plotting ratios of $[\text{C}_{10}\text{F}_{21}\text{I}] \times \Delta\delta$ against ratios of $[\text{C}_{10}\text{F}_{21}\text{I}]/[\text{C}_{10}\text{F}_{21}\text{I} + \text{XB acceptor}]$ to afford a maximum at

ratio $[C_{10}F_{21}I]/[C_{10}F_{21}I + XB \text{ acceptor}] = 0.5$, which meant a 1:1 complex ratio between $C_{10}F_{21}I$ and XB acceptor.

10.1 Determination of binding stoichiometry of $C_{10}F_{21}I$ with TEEDA

| | $[C_{10}F_{21}I]$ (M) | $\Delta\delta$ (ppm) | $[C_{10}F_{21}I]/[C_{10}F_{21}I + TEEDA]$ | $[C_{10}F_{21}I] \times \Delta\delta$ (M.ppm) |
|----|-----------------------|----------------------|---|---|
| 1 | 0 | 0 | 0 | 0 |
| 2 | 0.05 | 4.117 | 0.1 | 0.2059 |
| 3 | 0.10 | 3.663 | 0.2 | 0.3663 |
| 4 | 0.15 | 3.215 | 0.3 | 0.4823 |
| 5 | 0.20 | 2.831 | 0.4 | 0.5662 |
| 6 | 0.25 | 2.144 | 0.5 | 0.5360 |
| 7 | 0.30 | 1.965 | 0.6 | 0.5895 |
| 8 | 0.35 | 1.384 | 0.7 | 0.4844 |
| 9 | 0.40 | 0.987 | 0.8 | 0.3948 |
| 10 | 0.45 | 0.458 | 0.9 | 0.2061 |
| 11 | 0.50 | 0 | 1.0 | 0 |

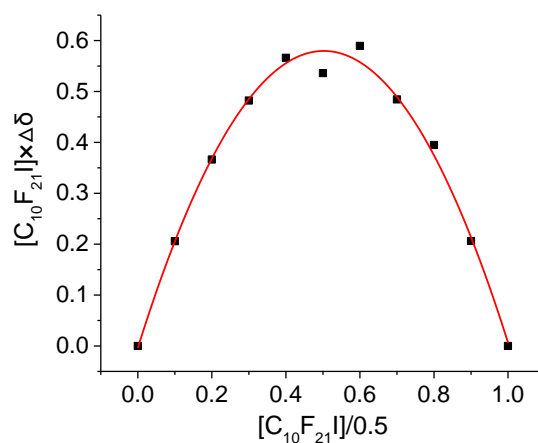


Figure S2

$$y = -0.0039 + 2.3239x - 2.3135x^2, x_{\max} = 2.3239/(-2 \times (-2.3135)) = 0.50$$

10.2 Determination of binding stoichiometry of $C_{10}F_{21}I$ with TEA

| | $[C_{10}F_{21}I]$ (M) | $\Delta\delta$ (ppm) | $[C_{10}F_{21}I]/[C_{10}F_{21}I + TEA]$ | $[C_{10}F_{21}I] \times \Delta\delta$ (M.ppm) |
|---|-----------------------|----------------------|---|---|
| 1 | 0 | 0 | 0 | 0 |
| 2 | 0.05 | 4.006 | 0.1 | 0.2003 |
| 3 | 0.10 | 3.493 | 0.2 | 0.3493 |
| 4 | 0.15 | 2.851 | 0.3 | 0.4277 |
| 5 | 0.20 | 2.505 | 0.4 | 0.5010 |
| 6 | 0.25 | 2.261 | 0.5 | 0.5653 |
| 7 | 0.30 | 1.758 | 0.6 | 0.5274 |

| | | | | |
|----|------|-------|-----|--------|
| 8 | 0.35 | 1.393 | 0.7 | 0.4876 |
| 9 | 0.40 | 0.975 | 0.8 | 0.3900 |
| 10 | 0.45 | 0.335 | 0.9 | 0.1507 |
| 11 | 0.50 | 0 | 1.0 | 0 |

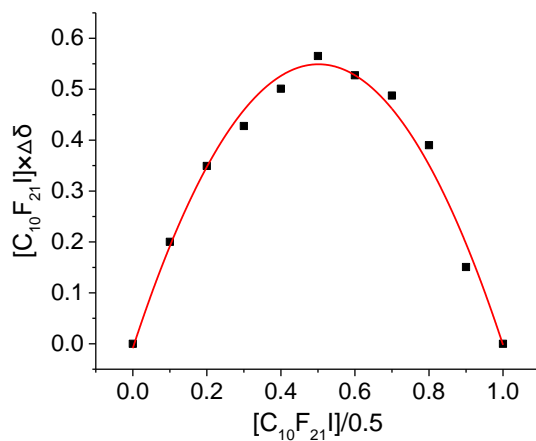


Figure S3

$$y = -0.00872 + 2.22459x - 2.21822x^2, x_{\max} = 2.22459/(-2 \times (-2.21822)) = 0.50$$

10.3 Determination of binding stoichiometry of C₁₀F₂₁I with THF

| | [C ₁₀ F ₂₁ I] (M) | Δδ (ppm) | [C ₁₀ F ₂₁ I]/ [C ₁₀ F ₂₁ I +THF] | [C ₁₀ F ₂₁ I]×Δδ (M.ppm) |
|----|---|----------|---|--|
| 1 | 0 | 0 | 0 | 0 |
| 2 | 0.05 | 0.430 | 0.1 | 0.0215 |
| 3 | 0.10 | 0.325 | 0.2 | 0.0325 |
| 4 | 0.15 | 0.282 | 0.3 | 0.0423 |
| 5 | 0.20 | 0.254 | 0.4 | 0.0508 |
| 6 | 0.25 | 0.196 | 0.5 | 0.0490 |
| 7 | 0.30 | 0.165 | 0.6 | 0.0495 |
| 8 | 0.35 | 0.132 | 0.7 | 0.0462 |
| 9 | 0.40 | 0.097 | 0.8 | 0.0388 |
| 10 | 0.45 | 0.020 | 0.9 | 0.0090 |
| 11 | 0.50 | 0 | 1.0 | 0 |

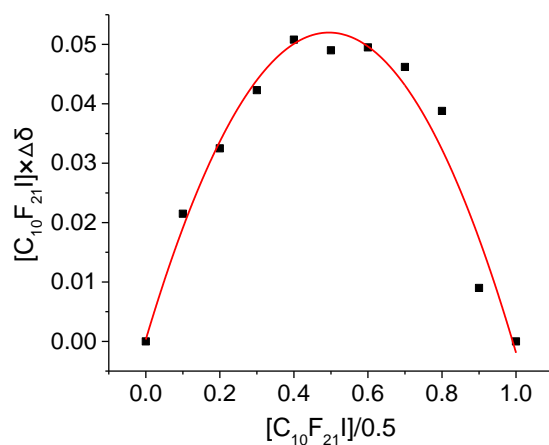


Figure S4

$$y = 0.00031 + 0.20895x - 0.21119x^2, x_{\max} = 0.20895/(-2 \times (-0.21119)) = 0.50$$

10.4 Determination of binding stoichiometry of C₁₀F₂₁I with dioxane

| | [C ₁₀ F ₂₁ I] (M) | Δδ (ppm) | [C ₁₀ F ₂₁ I]/ [C ₁₀ F ₂₁ I +Dioxane] | [C ₁₀ F ₂₁ I]×Δδ (M.ppm) |
|----|---|----------|---|--|
| 1 | 0 | 0 | 0 | 0 |
| 2 | 0.05 | 0.392 | 0.1 | 0.0196 |
| 3 | 0.10 | 0.327 | 0.2 | 0.0327 |
| 4 | 0.15 | 0.302 | 0.3 | 0.0453 |
| 5 | 0.20 | 0.255 | 0.4 | 0.0510 |
| 6 | 0.25 | 0.198 | 0.5 | 0.0495 |
| 7 | 0.30 | 0.173 | 0.6 | 0.0519 |
| 8 | 0.35 | 0.153 | 0.7 | 0.0535 |
| 9 | 0.40 | 0.090 | 0.8 | 0.0360 |
| 10 | 0.45 | 0.047 | 0.9 | 0.0211 |
| 11 | 0.50 | 0 | 1.0 | 0 |

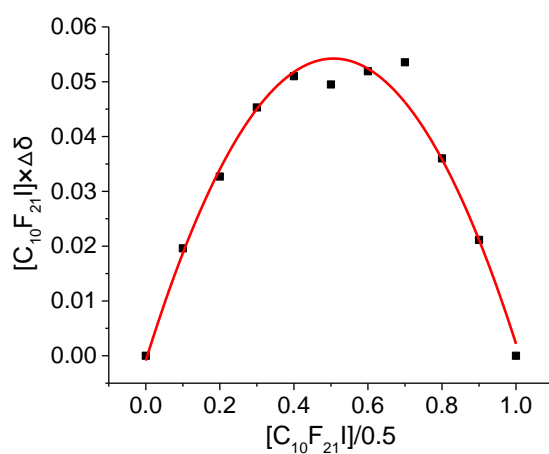


Figure S5

$$y = 0.00087 + 0.21731x - 0.21427x^2, x_{\max} = 0.21731/(-2 \times (-0.21427)) = 0.50$$

11. Determination of the association constant (K_a)

The association constant (K_a) was calculated using Hanna and Ashbaugh's¹²⁻¹⁵ Graphical method: For ideal behaviour in an interaction between an halogen bond donor molecule **C₁₀F₂₁I** and an lewis base molecule **LB** to form a halogen bond complex **XB**, we may define an association constant **K**, where

$$K = [\text{XB}]/[\text{LB}][\text{C}_{10}\text{F}_{21}\text{I}] \quad (1)$$

For 1:1 molecular complexes, when the concentration of lewis base molecule is in large excess we have

$$\delta_{\text{obs}} - \delta_0 = \frac{[\text{XB}]}{[\text{XB}] + [\text{C}_{10}\text{F}_{21}\text{I}]} (\delta_{\text{XB}} - \delta_0) \quad (2)$$

where δ_{obs} is the observed fluorine chemical shift of the -CF₂I group in the complexing medium, δ_0 is the fluorine chemical shift of the -CF₂I group in the uncomplexed state and δ_{XB} is the fluorine chemical shift of the -CF₂I group in the pure complex. If $\Delta = \delta_{\text{obs}} - \delta_0$ and $\Delta_0 = \delta_{\text{XB}} - \delta_0$, then substituting Eq. (1) into Eq. (2), we obtain Eq. (3) as follows:

$$\Delta = \frac{[\text{LB}]K}{1 + [\text{LB}]K} \Delta_0 \quad (3)$$

This assumes that δ_{obs} is the result of molecular complexing between halogen bond donor molecule **C₁₀F₂₁I** and lewis base molecule **LB**, and that there are no significant solvent effects on chemical shifts of the various species.

To obtain K from a series of measurements of Δ for various values of $[\text{LB}]$, the Eq. (3) may be rewritten as:

$$\frac{1}{\Delta} = \frac{1}{K\Delta_0} \frac{1}{[\text{LB}]} + \frac{1}{\Delta_0} \quad (4)$$

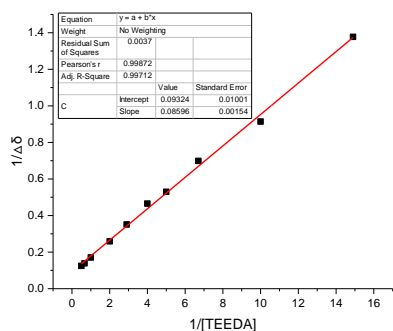
Plots of $1/\Delta$ against $1/[\text{LB}]$ should be a straight line, K may be obtained from the gradient using the value of $1/\Delta_0$ from the intercept of the line with the ordinate:

$$K = \left(\frac{1}{\Delta_0} \right) / \left(\frac{1}{K\Delta_0} \right) = \text{intercept} / \text{gradient} \quad (5)$$

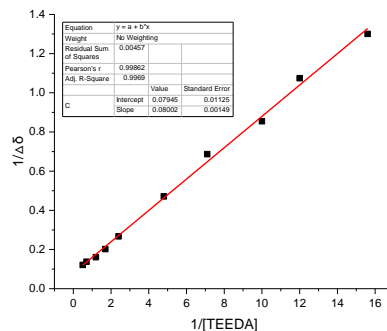
11.1 Determination of the association constant of C₁₀F₂₁I with TEEDA (K_{TEEDA})

^{19}F NMR spectra of ten samples of mixtures of $\text{C}_{10}\text{F}_{21}\text{I}$ and TEEDA in CDCl_3 were recorded at 298 K (Fluorobenzene ($\delta_{\text{F-Ph}} = -113.066$) was used as internal standard). The total volume of the mixture was 0.6 mL, the amount of $\text{C}_{10}\text{F}_{21}\text{I}$ was kept constant at 0.03 mmol (19.4 mg), while that of TEEDA was varied from 0.03 to 1.2 mmol. ^{19}F NMR for each sample was recorded and the chemical shifts differences ($\Delta\delta$) for $-\text{CF}_2\text{I}$ were used to draw the plot.

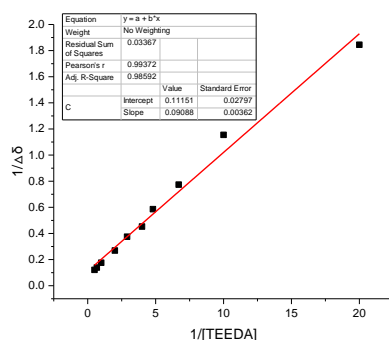
| | trial 1 | | | | trial 2 | | | | trial 3 | | | |
|----|---|---------------------------------------|-------------------------|------------------------------------|---|---------------------------------------|-------------------------|------------------------------------|---|---------------------------------------|-------------------------|------------------------------------|
| | TEEDA (mol/L) | $\frac{1}{[\text{TEEDA}]}$ (L/mol) | $\Delta\delta$ (ppm) | $1/\Delta\delta$ (ppm $^{-1}$) | TEEDA (mol/L) | $\frac{1}{[\text{TEEDA}]}$ (L/mol) | $\Delta\delta$ (ppm) | $1/\Delta\delta$ (ppm $^{-1}$) | TEEDA (mol/L) | $\frac{1}{[\text{TEEDA}]}$ (L/mol) | $\Delta\delta$ (ppm) | $1/\Delta\delta$ (ppm $^{-1}$) |
| 1 | 2.00 | 0.5 | 8.006 | 0.1249 | 2.00 | 0.5 | 8.237 | 0.1214 | 2.00 | 0.5 | 8.264 | 0.1210 |
| 2 | 1.49 | 0.67 | 7.199 | 0.1389 | 1.43 | 0.7 | 7.278 | 0.1374 | 1.49 | 0.67 | 7.194 | 0.1390 |
| 3 | 1.00 | 1.0 | 5.865 | 0.1705 | 0.83 | 1.2 | 6.211 | 0.1610 | 1.00 | 1.0 | 5.682 | 0.1760 |
| 4 | 0.50 | 2.0 | 3.860 | 0.2591 | 0.59 | 1.7 | 4.950 | 0.2020 | 0.50 | 2.0 | 3.717 | 0.2690 |
| 5 | 0.34 | 2.9 | 2.849 | 0.3510 | 0.42 | 2.4 | 3.745 | 0.2670 | 0.34 | 2.9 | 2.667 | 0.3750 |
| 6 | 0.25 | 4.0 | 2.148 | 0.4655 | 0.21 | 4.8 | 2.123 | 0.4710 | 0.25 | 4 | 2.208 | 0.4529 |
| 7 | 0.20 | 5.0 | 1.888 | 0.5297 | 0.14 | 7.1 | 1.456 | 0.6868 | 0.21 | 4.8 | 1.706 | 0.5862 |
| 8 | 0.15 | 6.7 | 1.431 | 0.6988 | 0.10 | 10.0 | 1.170 | 0.8547 | 0.15 | 6.7 | 1.292 | 0.7740 |
| 9 | 0.10 | 10.0 | 1.094 | 0.9141 | 0.083 | 12.0 | 0.931 | 1.0741 | 0.10 | 10 | 0.866 | 1.1547 |
| 10 | 0.067 | 14.9 | 0.726 | 1.3774 | 0.064 | 15.6 | 0.769 | 1.3004 | 0.05 | 20 | 0.542 | 1.8450 |
| | $K_{\text{TEEDA}} = 0.09324/0.08596 = 1.0847$ | | | | $K_{\text{TEEDA}} = 0.07945/0.08002 = 0.9929$ | | | | $K_{\text{TEEDA}} = 0.11151/0.09088 = 1.2270$ | | | |



Trial 1



Trial 2



Trial 3

Figure S6

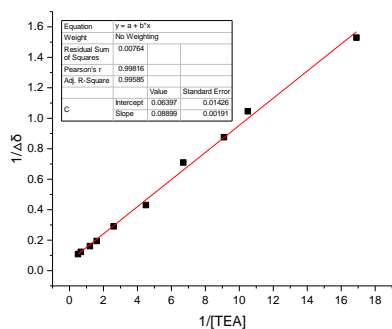
The association constant of C₁₀F₂₁I and TEEDA (K_{TEEDA}) was calculated (average of three experiments) to be 1.1 ($K_{TEEDA} = 1.1 \text{ M}^{-1}$).

11.2 Determination of the association constant of C₁₀F₂₁I and TEA (K_{TEA})

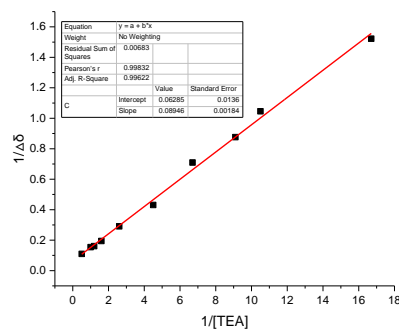
¹⁹F NMR spectra of ten samples of mixtures of C₁₀F₂₁I and TEA in CDCl₃ were recorded at 298 K (Fluorobenzene ($\delta_{\text{F-Ph}} = -113.066$) was used as internal standard). The total volume of the mixture was 0.6 mL, the amount of C₁₀F₂₁I was kept constant at 0.03 mmol (19.4 mg), while that of TEA was varied from 0.036 to 1.2 mmol. ¹⁹F NMR for each sample was recorded and the chemical shifts differences ($\Delta\delta$) for -CF₂I were used to draw the plot.

| | trial 1 | | | | trial 2 | | | | trial 3 | | | |
|---|----------------|-------------------------------------|-------------------------|--|----------------|-------------------------------------|-------------------------|--|----------------|-------------------------------------|-------------------------|--|
| | TEA (mol/L) | $\frac{1}{[\text{TEA}]}$ (L/mol) | $\Delta\delta$ (ppm) | $1/\Delta\delta$ (ppm ⁻¹) | TEA (mol/L) | $\frac{1}{[\text{TEA}]}$ (L/mol) | $\Delta\delta$ (ppm) | $1/\Delta\delta$ (ppm ⁻¹) | TEA (mol/L) | $\frac{1}{[\text{TEA}]}$ (L/mol) | $\Delta\delta$ (ppm) | $1/\Delta\delta$ (ppm ⁻¹) |
| 1 | 2.00 | 0.5 | 9.225 | 0.1084 | 2.00 | 0.5 | 9.115 | 0.1097 | 2.00 | 0.5 | 9.311 | 0.1074 |
| 2 | 1.49 | 0.67 | 8.104 | 0.1234 | 1.00 | 1.0 | 6.493 | 0.1540 | 1.49 | 0.67 | 8.032 | 0.1245 |
| 3 | 0.83 | 1.2 | 6.215 | 0.1609 | 0.83 | 1.2 | 6.215 | 0.1609 | 1.0 | 1.0 | 6.510 | 0.1536 |
| 4 | 0.63 | 1.6 | 5.141 | 0.1945 | 0.63 | 1.6 | 5.141 | 0.1945 | 0.53 | 1.9 | 4.290 | 0.2331 |
| 5 | 0.38 | 2.6 | 3.439 | 0.2908 | 0.38 | 2.6 | 3.438 | 0.2909 | 0.34 | 2.9 | 2.946 | 0.3394 |
| 6 | 0.22 | 4.5 | 2.323 | 0.4305 | 0.22 | 4.5 | 2.321 | 0.4308 | 0.26 | 3.8 | 2.381 | 0.4200 |

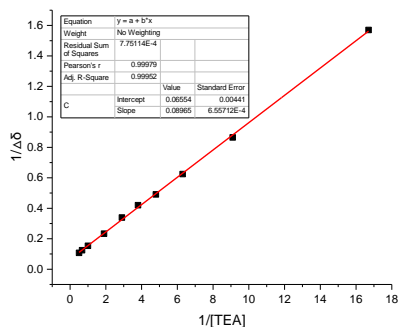
| | | | | | | | | | | | | |
|--------------------------------------|-------|------|-------|--------|--------------------------------------|------|-------|--------|--------------------------------------|------|-------|--------|
| 7 | 0.15 | 6.7 | 1.409 | 0.7097 | 0.15 | 6.7 | 1.409 | 0.7097 | 0.21 | 4.8 | 2.036 | 0.4912 |
| 8 | 0.11 | 9.1 | 1.142 | 0.8757 | 0.11 | 9.1 | 1.141 | 0.8764 | 0.16 | 6.3 | 1.601 | 0.6246 |
| 9 | 0.095 | 10.5 | 0.956 | 1.0460 | 0.095 | 10.5 | 0.956 | 1.0460 | 0.11 | 9.1 | 1.156 | 0.8651 |
| 10 | 0.059 | 16.9 | 0.654 | 1.5291 | 0.06 | 16.7 | 0.657 | 1.5221 | 0.06 | 16.7 | 0.637 | 1.5700 |
| $K_{TEA} = 0.06397/0.08899 = 0.7188$ | | | | | $K_{TEA} = 0.06285/0.08946 = 0.7025$ | | | | $K_{TEA} = 0.06554/0.08965 = 0.7311$ | | | |



Trial 1



Trial 2



Trial 3

Figure S7

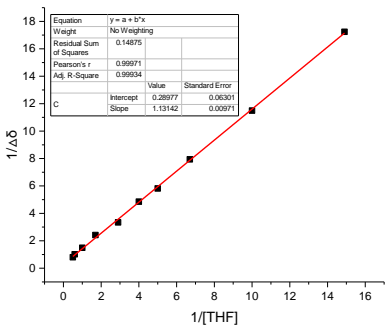
The association constant of $C_{10}F_{21}I$ and TEA (K_{TEA}) was calculated (average of three experiments) to be 0.72 ($K_{TEA} = 0.72 M^{-1}$).

11.3 Determination of the association constant of $C_{10}F_{21}I$ and THF (K_{THF})

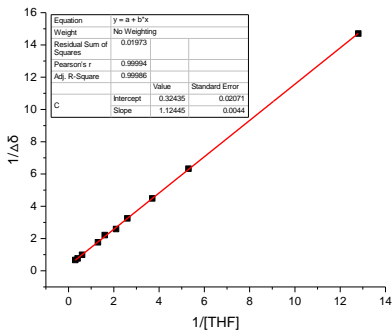
^{19}F NMR spectra of ten samples of mixtures of $C_{10}F_{21}I$ and THF in $CDCl_3$ were recorded at 298 K (Fluorobenzene ($\delta_{F-Ph} = -113.066$) was used as internal standard). The total volume of the mixture was 0.6 mL, the amount of $C_{10}F_{21}I$ was kept constant

at 0.03 mmol (19.4 mg), while that of THF was varied from 0.04 to 2.0 mmol. ¹⁹F NMR for each sample was recorded and the chemical shifts differences (Δδ) for -CF₂I were used to draw the plot.

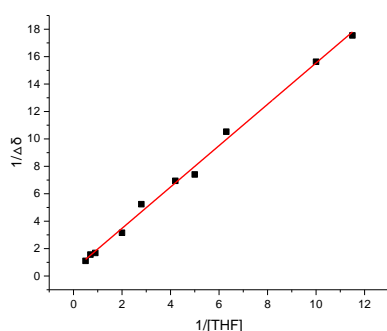
| | trial 1 | | | | trial 2 | | | | trial 3 | | | |
|----|---|------------------------------|-------------|------------------------------|---|------------------------------|-------------|------------------------------|---|------------------------------|-------------|------------------------------|
| | THF (mol/L) | $\frac{1}{[THF]}$ (L/mol) | Δδ (ppm) | 1/Δδ (ppm ⁻¹) | THF (mol/L) | $\frac{1}{[THF]}$ (L/mol) | Δδ (ppm) | 1/Δδ (ppm ⁻¹) | THF (mol/L) | $\frac{1}{[THF]}$ (L/mol) | Δδ (ppm) | 1/Δδ (ppm ⁻¹) |
| 1 | 2.00 | 0.5 | 1.247 | 0.8019 | 3.33 | 0.3 | 1.488 | 0.6720 | 2.00 | 0.5 | 0.906 | 1.1037 |
| 2 | 1.67 | 0.6 | 0.985 | 1.0152 | 2.50 | 0.4 | 1.305 | 0.7662 | 1.43 | 0.7 | 0.640 | 1.5625 |
| 3 | 1.00 | 1.0 | 0.671 | 1.4903 | 1.67 | 0.6 | 1.010 | 0.9900 | 1.11 | 0.9 | 0.594 | 1.6835 |
| 4 | 0.59 | 1.7 | 0.413 | 2.4213 | 0.77 | 1.3 | 0.566 | 1.7668 | 0.50 | 2.0 | 0.317 | 3.1546 |
| 5 | 0.34 | 2.9 | 0.299 | 3.3444 | 0.63 | 1.6 | 0.452 | 2.2124 | 0.36 | 2.8 | 0.191 | 5.2356 |
| 6 | 0.25 | 4.0 | 0.206 | 4.8544 | 0.48 | 2.1 | 0.386 | 2.5907 | 0.24 | 4.2 | 0.144 | 6.9444 |
| 7 | 0.20 | 5.0 | 0.172 | 5.8140 | 0.38 | 2.6 | 0.308 | 3.2468 | 0.20 | 5.0 | 0.135 | 7.4074 |
| 8 | 0.15 | 6.7 | 0.126 | 7.9365 | 0.27 | 3.7 | 0.223 | 4.4843 | 0.16 | 6.3 | 0.095 | 10.5263 |
| 9 | 0.10 | 10.0 | 0.087 | 11.4943 | 0.19 | 5.3 | 0.158 | 6.3291 | 0.10 | 10.0 | 0.064 | 15.6250 |
| 10 | 0.067 | 14.9 | 0.058 | 17.2414 | 0.078 | 12.8 | 0.068 | 14.7059 | 0.087 | 11.5 | 0.057 | 17.5439 |
| | K _{THF} = 0.28977/1.13142 = 0.2561 | | | | K _{THF} = 0.32435/1.12445 = 0.2885 | | | | K _{THF} = 0.46041/1.50758 = 0.3054 | | | |



Trial 1



Trial 2



Trial 3

Figure S8

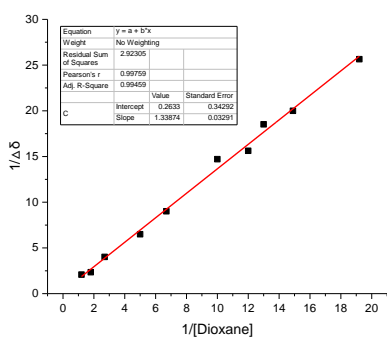
The association constant of $C_{10}F_{21}I$ and TEEDA (K_{THF}) was calculated (average of three experiments) to be 0.28 ($K_{THF} = 0.28 M^{-1}$).

11.4 Determination of the association constant of $C_{10}F_{21}I$ and dioxane ($K_{Dioxane}$)

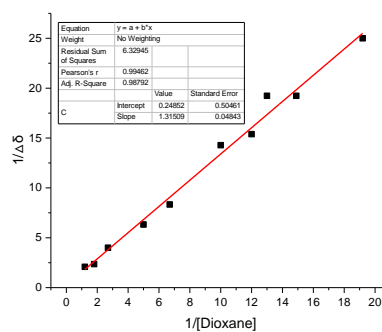
^{19}F NMR spectra of ten samples of mixtures of $C_{10}F_{21}I$ and dioxane in $CDCl_3$ were recorded at 298 K (Fluorobenzene ($\delta_{F-Ph} = -113.066$) was used as internal standard). The total volume of the mixture was 0.6 mL, the amount of $C_{10}F_{21}I$ was kept constant at 0.03 mmol (19.4 mg), while that of dioxane was varied from 0.03 to 0.05 mmol. ^{19}F NMR for each sample was recorded and the chemical shifts differences ($\Delta\delta$) for $-CF_2I$ were used to draw the plot.

| | trial 1 | | | | trial 2 | | | | trial 3 | | | |
|---|--------------------|----------------------------------|-------------------------|------------------------------------|--------------------|----------------------------------|-------------------------|------------------------------------|--------------------|----------------------------------|-------------------------|------------------------------------|
| | Dioxane (mol/L) | $\frac{1}{[Dioxane]}$ (L/mol) | $\Delta\delta$ (ppm) | $1/\Delta\delta$ (ppm $^{-1}$) | Dioxane (mol/L) | $\frac{1}{[Dioxane]}$ (L/mol) | $\Delta\delta$ (ppm) | $1/\Delta\delta$ (ppm $^{-1}$) | Dioxane (mol/L) | $\frac{1}{[Dioxane]}$ (L/mol) | $\Delta\delta$ (ppm) | $1/\Delta\delta$ (ppm $^{-1}$) |
| 1 | 0.83 | 1.2 | 0.480 | 2.0833 | 0.83 | 1.2 | 0.479 | 2.0877 | 0.83 | 1.2 | 0.464 | 2.1552 |
| 2 | 0.56 | 1.8 | 0.427 | 2.3419 | 0.56 | 1.8 | 0.424 | 2.3585 | 0.56 | 1.8 | 0.420 | 2.3810 |
| 3 | 0.37 | 2.7 | 0.249 | 4.0161 | 0.37 | 2.7 | 0.250 | 4.000 | 0.37 | 2.7 | 0.250 | 4.000 |
| 4 | 0.20 | 5.0 | 0.154 | 6.4935 | 0.20 | 5.0 | 0.158 | 6.3291 | 0.20 | 5.0 | 0.151 | 6.6225 |
| 5 | 0.15 | 6.7 | 0.111 | 9.0090 | 0.15 | 6.7 | 0.120 | 8.3333 | 0.15 | 6.7 | 0.110 | 9.0909 |

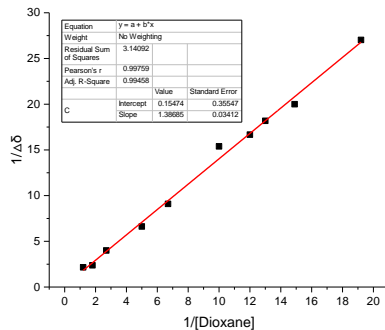
| | | | | | | | | | | | | |
|----|--|------|-------|---------|---|------|-------|---------|---|------|-------|---------|
| 6 | 0.10 | 10.0 | 0.068 | 14.7059 | 0.10 | 10.0 | 0.070 | 14.2857 | 0.10 | 10.0 | 0.065 | 15.3846 |
| 7 | 0.083 | 12.0 | 0.064 | 15.6250 | 0.083 | 12.0 | 0.065 | 15.3846 | 0.083 | 12.0 | 0.060 | 16.6667 |
| 8 | 0.077 | 13.0 | 0.054 | 18.5185 | 0.077 | 13.0 | 0.052 | 19.2308 | 0.077 | 13.0 | 0.055 | 18.1818 |
| 9 | 0.067 | 14.9 | 0.050 | 20.0000 | 0.067 | 14.9 | 0.052 | 19.2308 | 0.067 | 14.9 | 0.050 | 20.0000 |
| 10 | 0.052 | 19.2 | 0.039 | 25.6410 | 0.052 | 19.2 | 0.040 | 25.0000 | 0.052 | 19.2 | 0.037 | 27.0270 |
| | K _{Dioxane} = 0.2633/1.33874 = 0.1967 | | | | K _{Dioxane} = 0.24852/1.31509 = 0.1890 | | | | K _{Dioxane} = 0.15474/1.38685 = 0.1116 | | | |



Trial 1



Trial 2



Trial 3

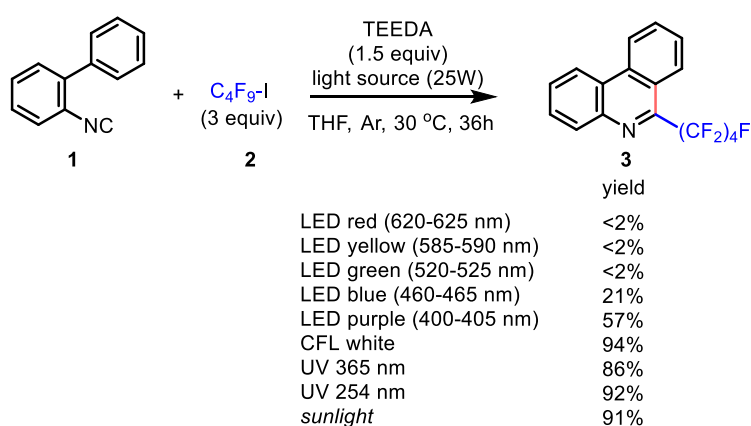
Figure S9

The association constant of $\text{C}_{10}\text{F}_{21}\text{I}$ and TEEDA (K_{Dioxane}) was calculated (average of three experiments) to be 0.17 ($K_{\text{Dioxane}} = 0.17 \text{ M}^{-1}$).

12. Evaluation of different light source

2-Isocyanobiphenyl **1** (35.8 mg, 0.2 mol, 1.0 equiv), $\text{C}_4\text{F}_9\text{I}$ (207.5 mg, 0.6 mol, 3.0 equiv) and TEEDA (0.3 mol, 1.5 equiv) were dispersed in 2 mL of THF. The reaction

vial was then purged with Ar for 1 min, sealed with PTEF cap. The reaction mixture was vigorously stirred at 30 °C under the irradiation of different light source (25W, laid 10 CM away from the vial) for 36 h. Then THF was removed under reduced pressure. The resulting residue was dissolved in 1 mL of CDCl₃ along with Cl₂CHCHCl₂ (20 μL) as an external standard for ¹H-NMR analysis. The composition of reaction mixture was based on the Ar-H at 8.73 ppm (d, *J* = 8.4 Hz, 1H) for compound **3**.



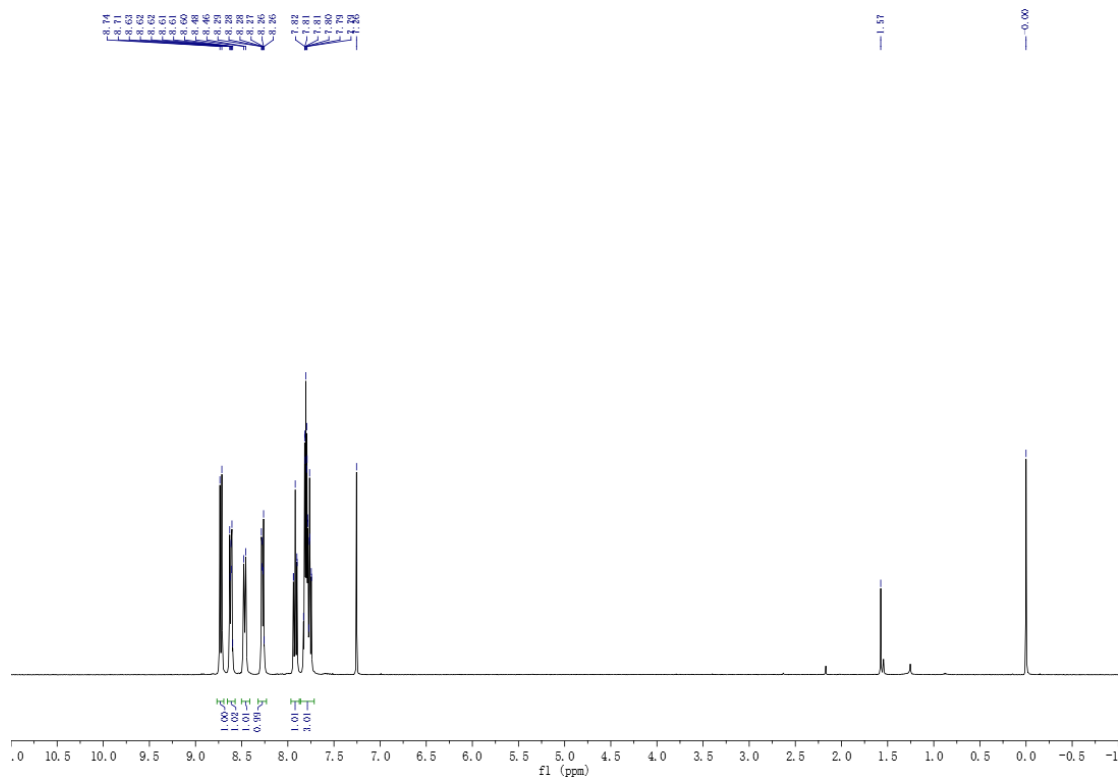
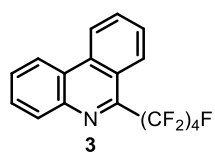
Scheme S23

As shown in Scheme S23, the source of light clearly has an impact to the reaction of **1** and **2** for synthesis of **3**. Irradiation by LED lamp of red, yellow and green colors (25W) gave little product **3**; the blue LED lamp with relatively shorter emission wavelength (25W) gave 21% yield, the purple LED lamp with relatively shorter emission wavelength (25W) gave 57% yield. The irradiation of UV lamp (254 or 360nm), CFL of the same power intensity or sunlight gave excellent yields. It is known that CFL emit small amount of UV light. These results suggest that the low-intensity irradiation in the UV region is responsible for the XB-promoted photochemical activation of Rf-I.

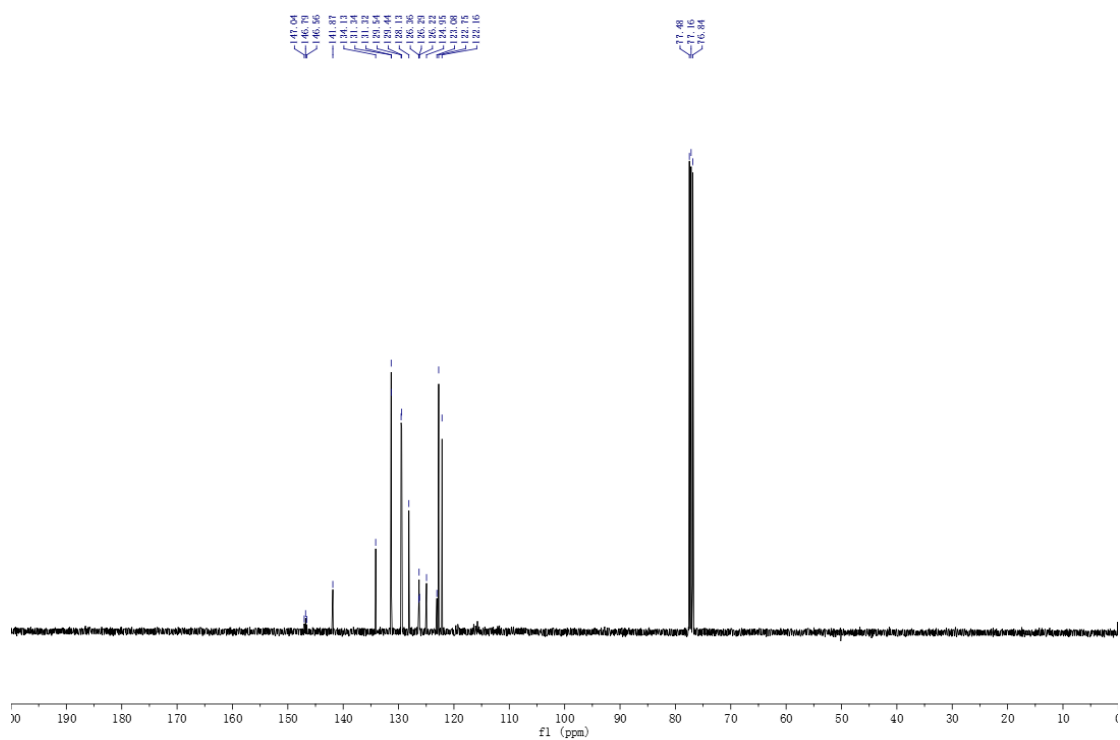
13. References

1. Rong, J.; Deng, L.; Tan, P.; Ni, C. F.; Gu, Y. C.; Hu, J. B. *Angew. Chem., Int. Ed.* **2016**, *55*, 2743.
2. Cao, J. J.; Wang, X.; Wang, S. Y.; Ji, S. J. *Chem. Commun.* **2014**, *50*, 12892.
3. He, Z.; Bae, M.; Wu, J.; Jamison, T. F. *Angew. Chem., Int. Ed.* **2014**, *53*, 14451.
4. Chighine, A.; Crosignani, S.; Marie-Claire Arnal, M. C.; Bradley, M.; Bruno Linclau, B. *J. Org. Chem.* **2009**, *74*, 4753.
5. Xu, T.; Cheung, C. W.; Hu, X. L. *Angew. Chem., Int. Ed.* **2013**, *53*, 4910.
6. Collins, B. S. L.; Suero, M. G.; Gaunt, M. J. *Angew. Chem., Int. Ed.* **2013**, *52*, 5799.
7. Rozkiewicz, D. I.; Jańczewski, D.; Verboom, W.; Ravoo, B. J.; Reinhoudt, D. N. *Angew. Chem., Int. Ed.* **2006**, *45*, 5292.
8. Coste, A.; Toumi, M.; Wright, K.; Razafimahaleo, V.; Couty, F.; Marrot, J.; Evano, G. *Org. Lett.* **2008**, *10*, 3841.
9. Naskar, J.; Drew, M. G. B.; Deb, I.; Das, S.; Banerjee, A. *Org. Lett.* **2008**, *10*, 2625.
10. Li, L.; Mu, X. Y.; Liu, W. B.; Wang, Y. C.; Mi, Z.; Li, C. -J. *J. Am. Chem. Soc.* **2016**, *138*, 5809.
11. Job, P. *Justus Liebigs Ann. Chem.* **1928**, *9*, 113.
12. Berkeley; Hanna. *J. Physic. Chem.* **1963**, *67*, 846.
13. Hanna; Ashbaugh. *J. Physic. Chem.* **1964**, *68*, 811.
14. Foster, R.; Fyfe, C. A. *Trans. Faraday Soc.* **1965**, *61*, 1626.
15. Foster, R.; Fyfe, C. A. *J. Chem. Soc., Chem. Commun.* **1965**, 642.

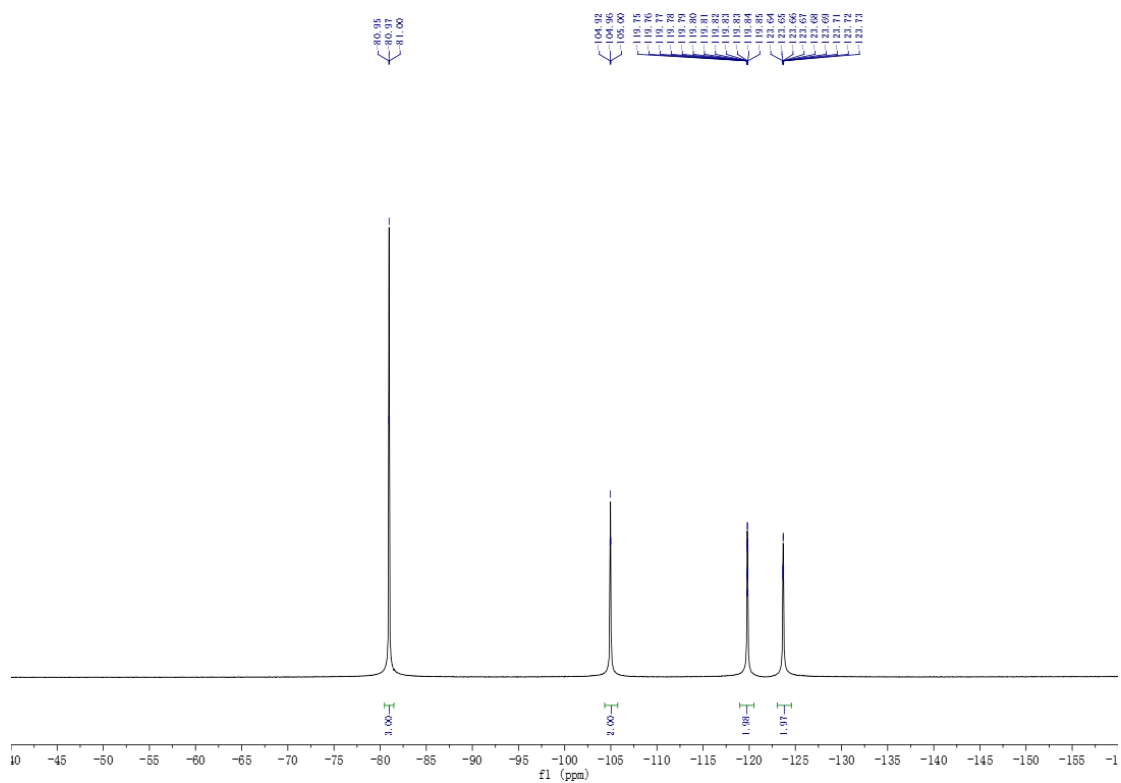
14. Spectra



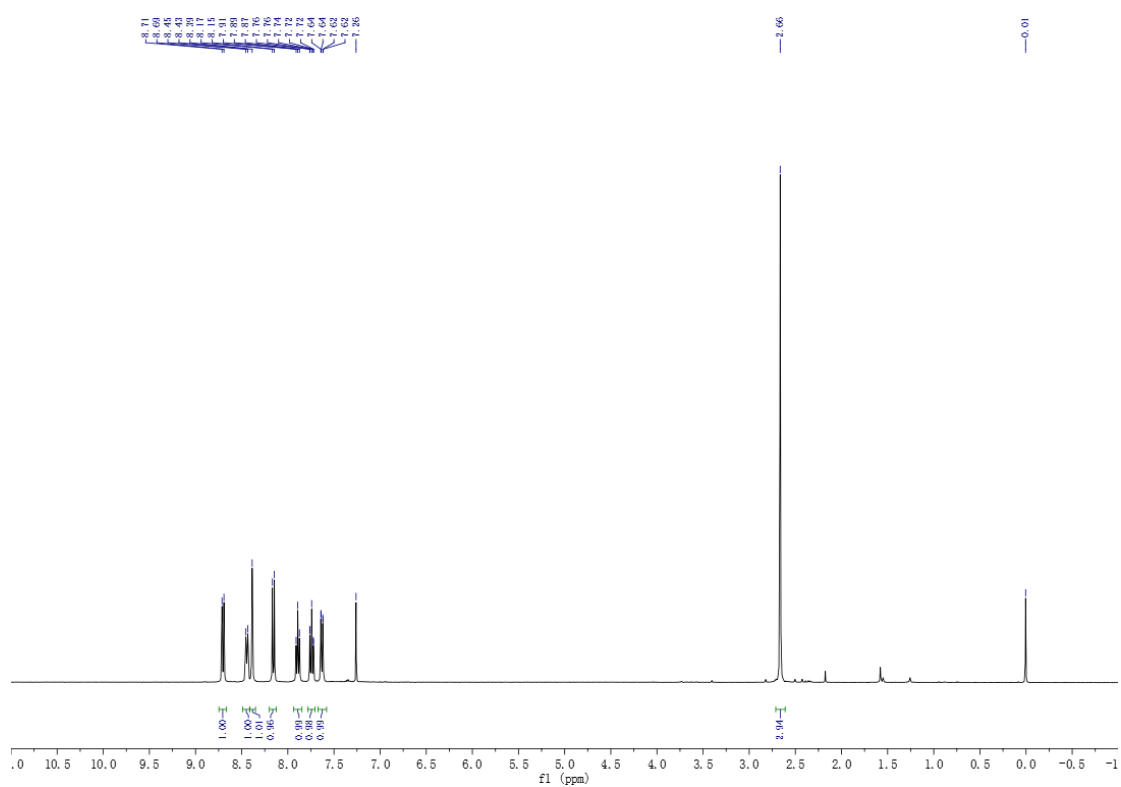
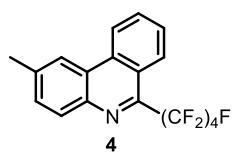
¹H NMR of compound **3**



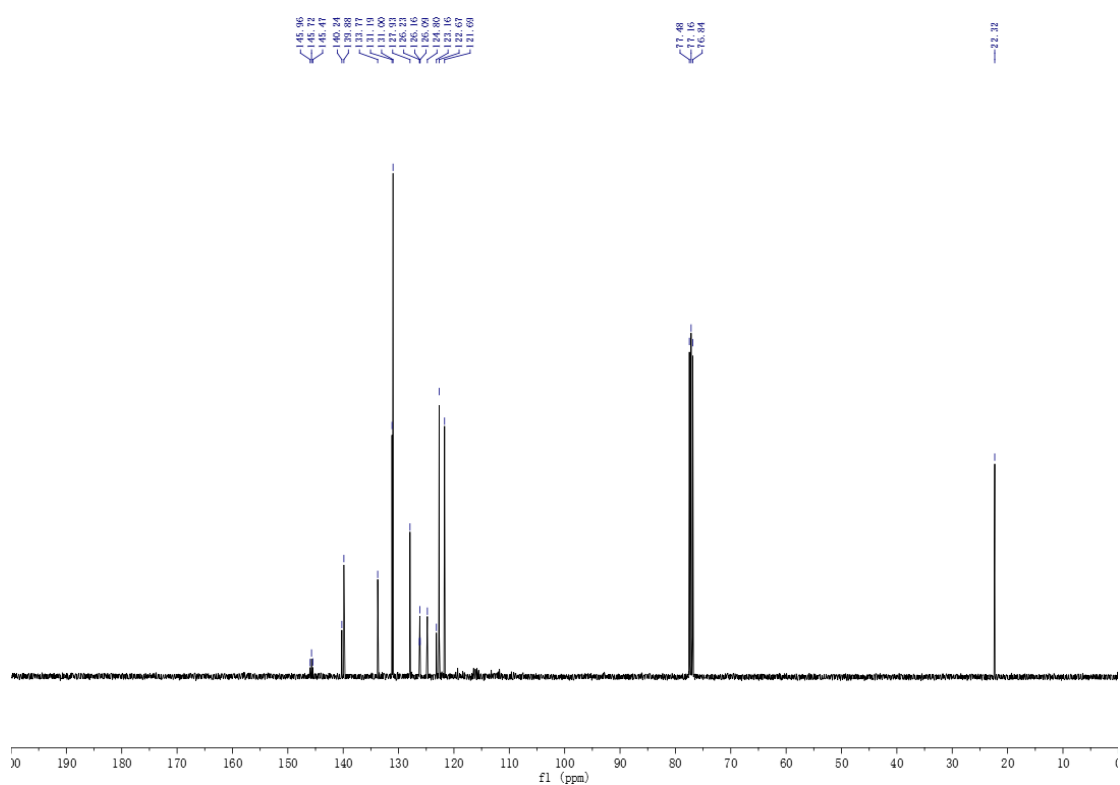
¹³C NMR of compound **3**



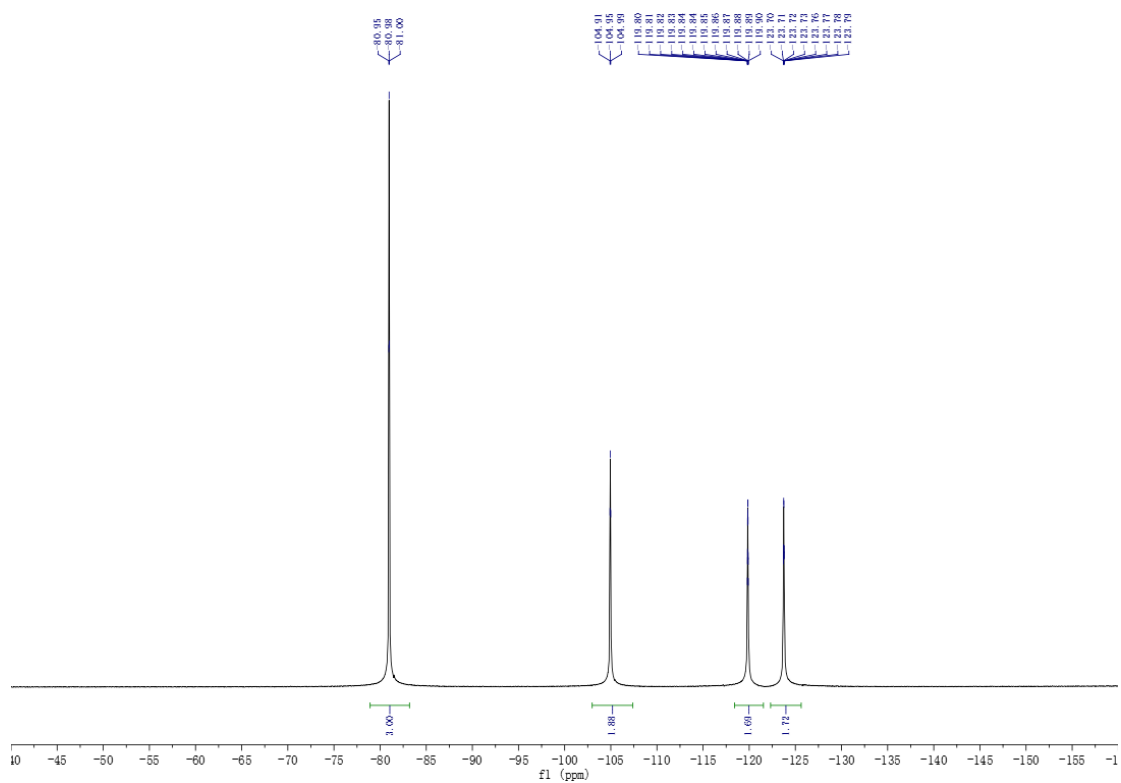
¹⁹F NMR of compound **3**



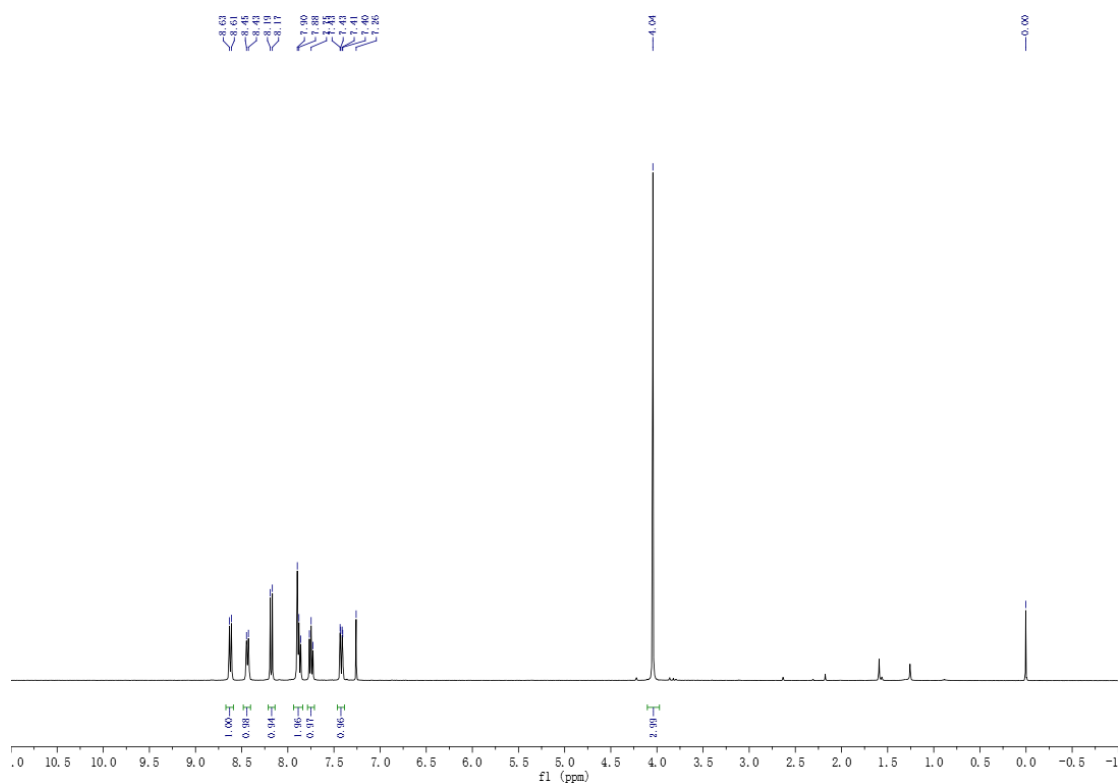
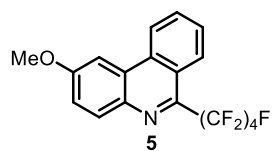
¹H NMR of compound **4**



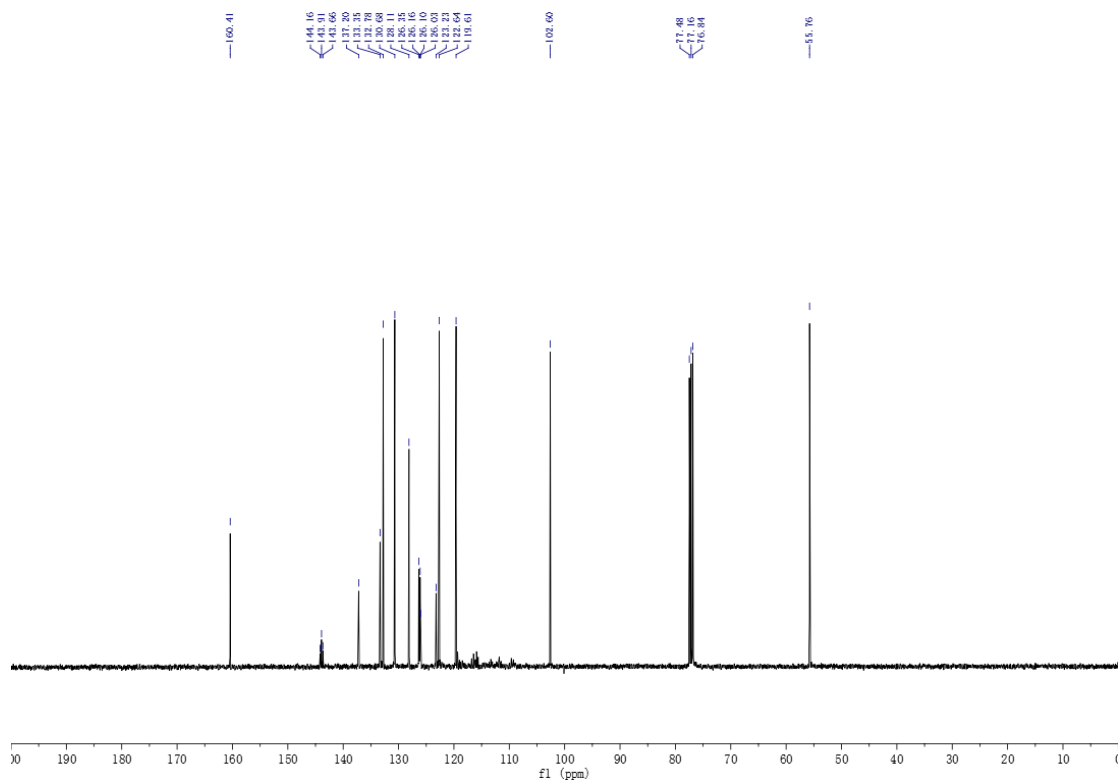
¹³C NMR of compound **4**



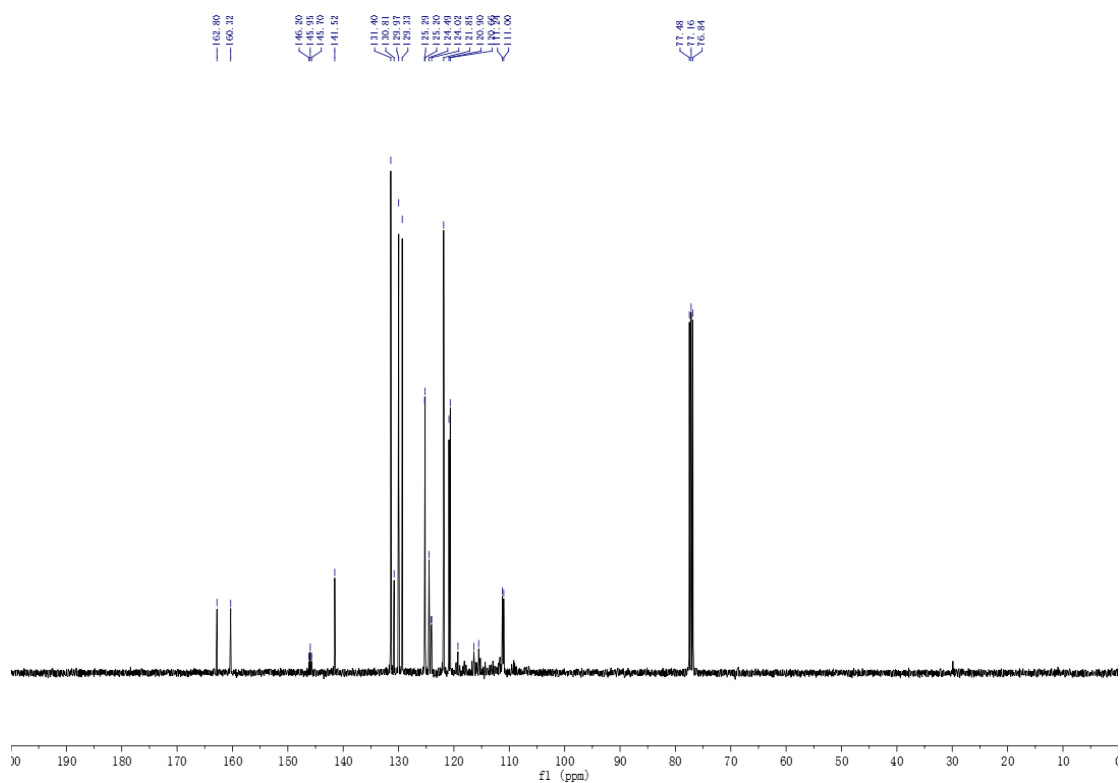
¹⁹F NMR of compound **4**



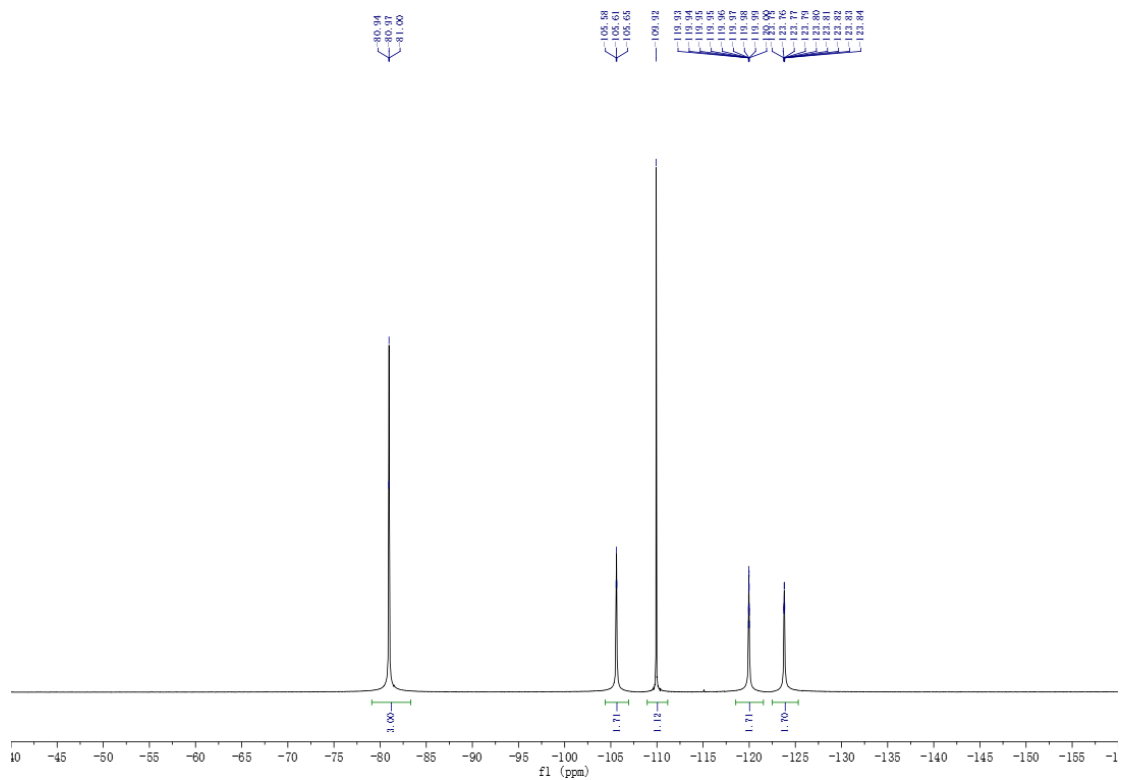
¹H NMR of compound **5**



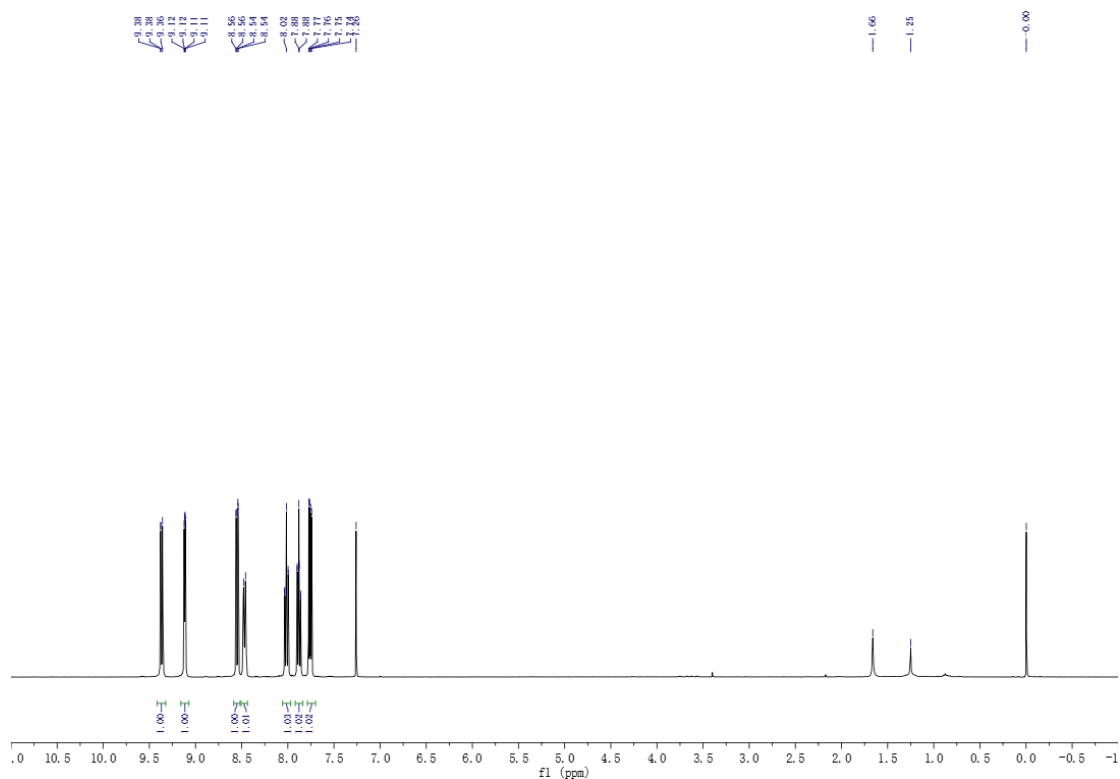
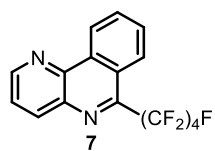
¹³C NMR of compound **5**



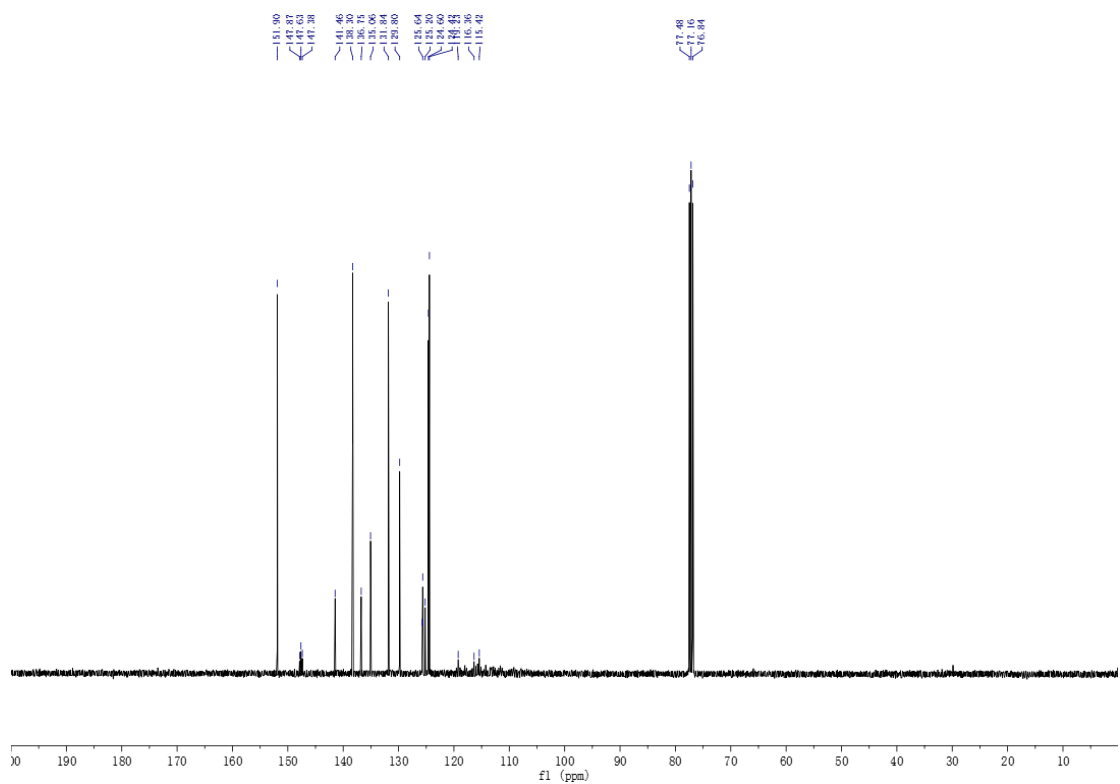
¹³C NMR of compound 6



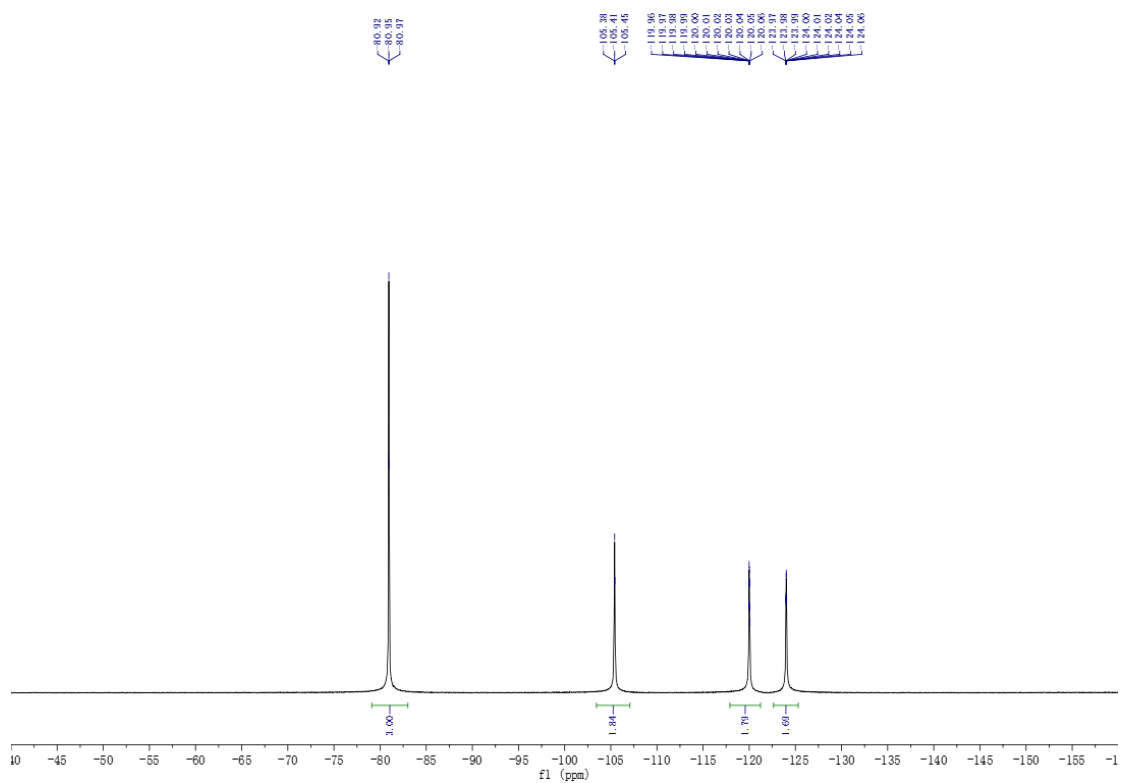
¹⁹F NMR of compound 6



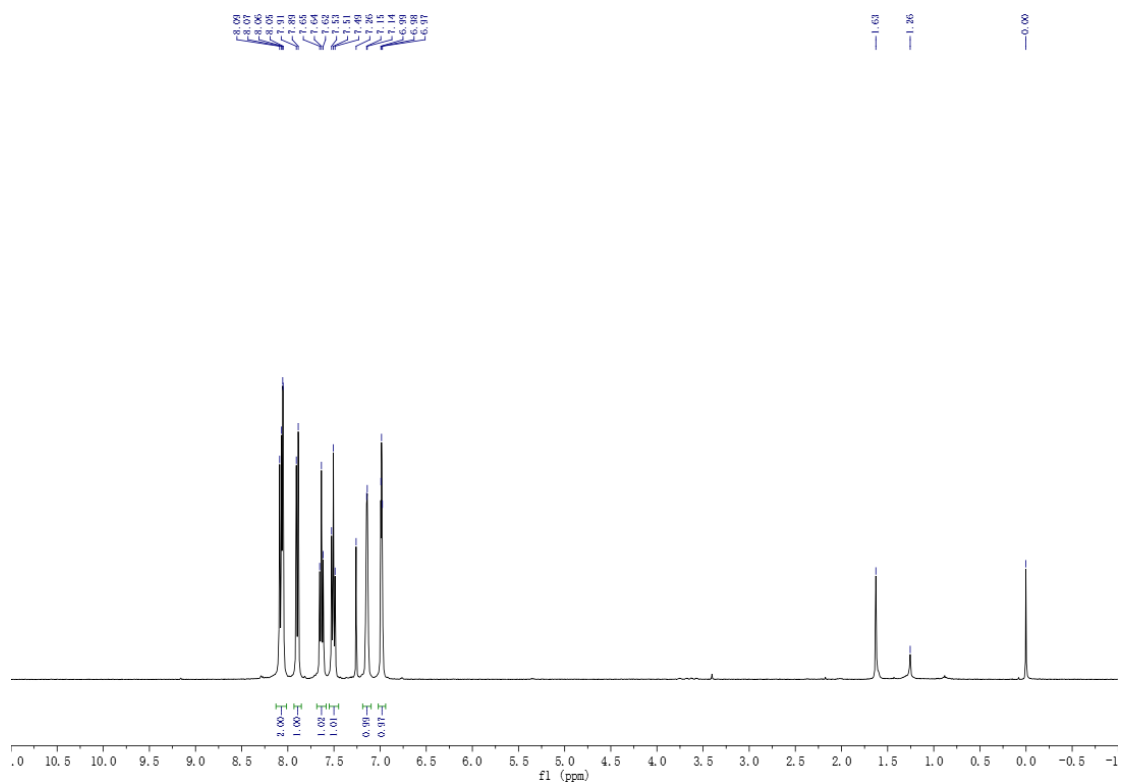
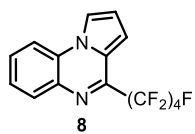
¹H NMR of compound **7**



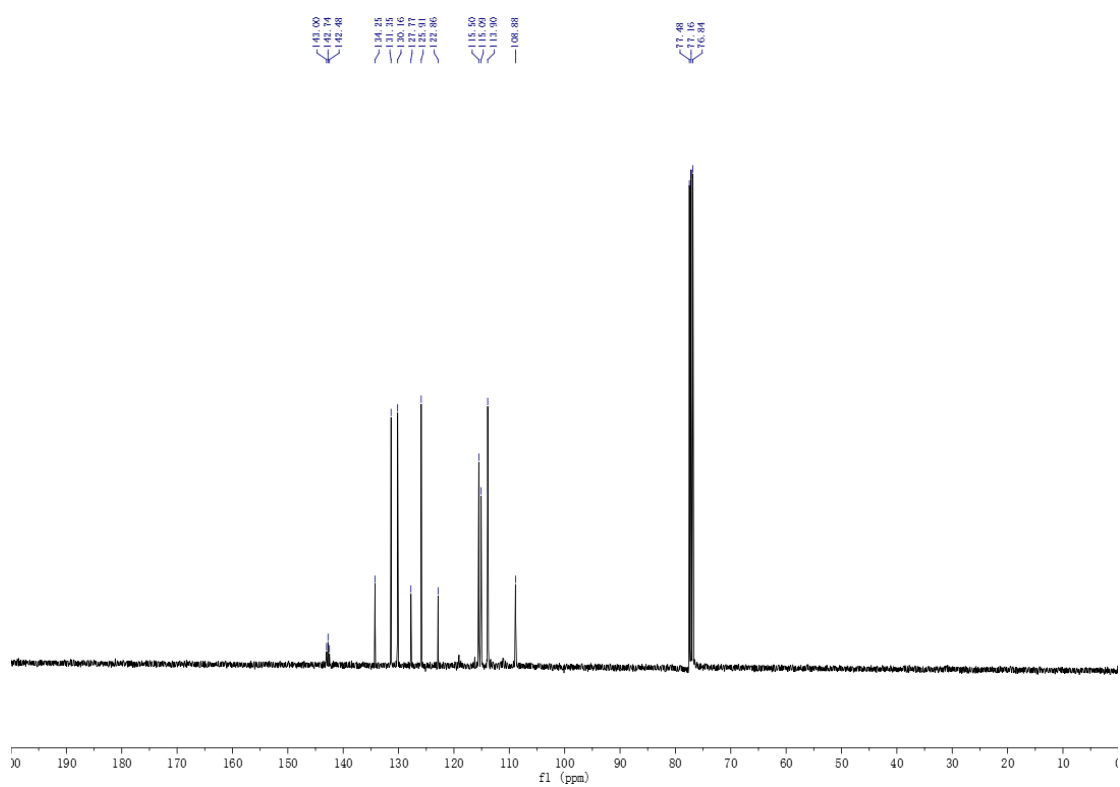
¹³C NMR of compound **7**



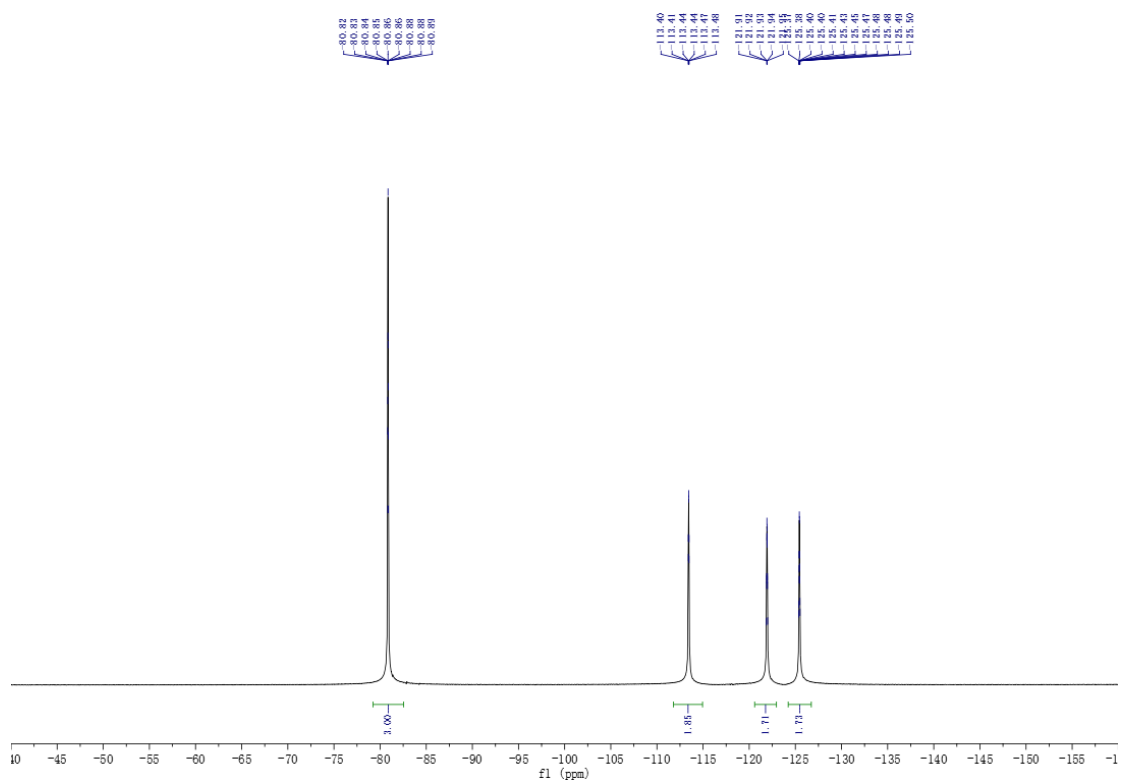
^{19}F NMR of compound **7**



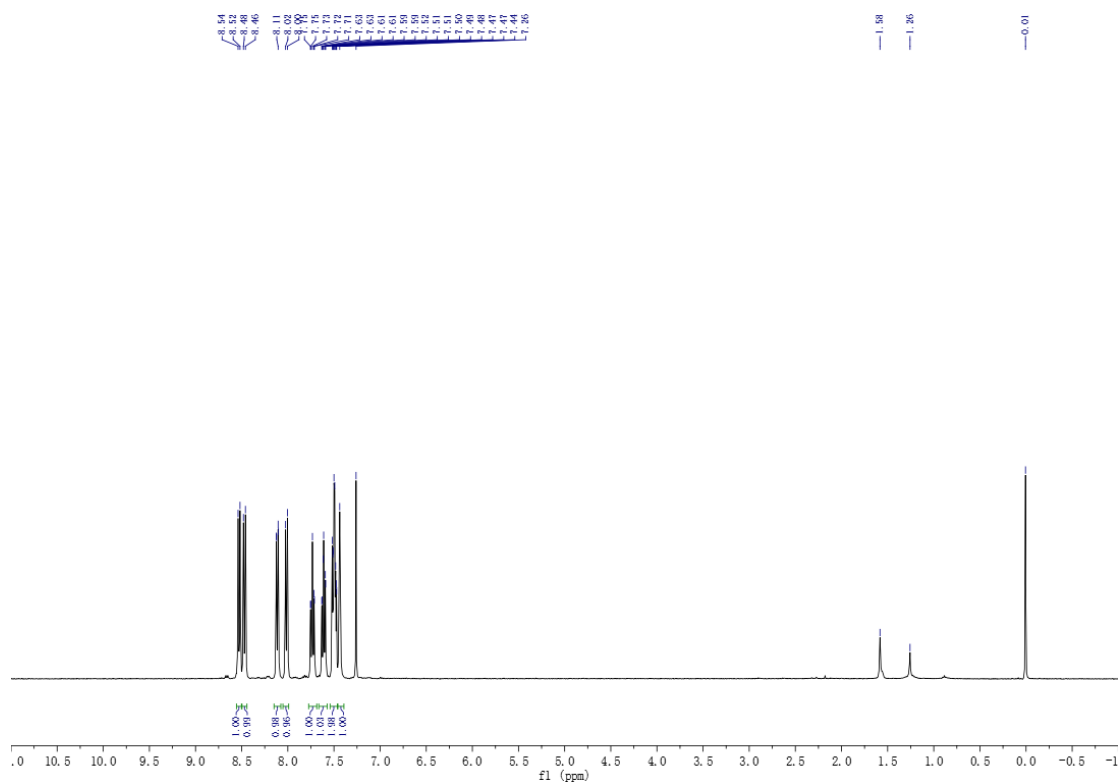
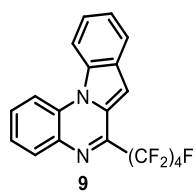
^1H NMR of compound **8**



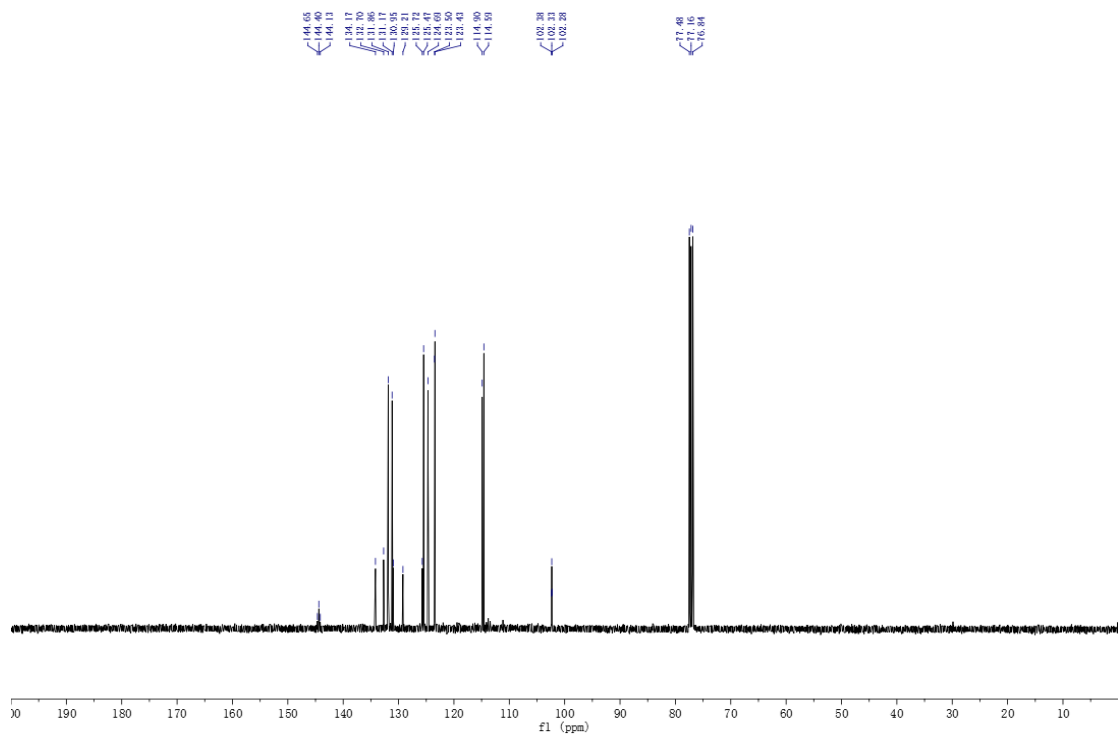
^{13}C NMR of compound **8**



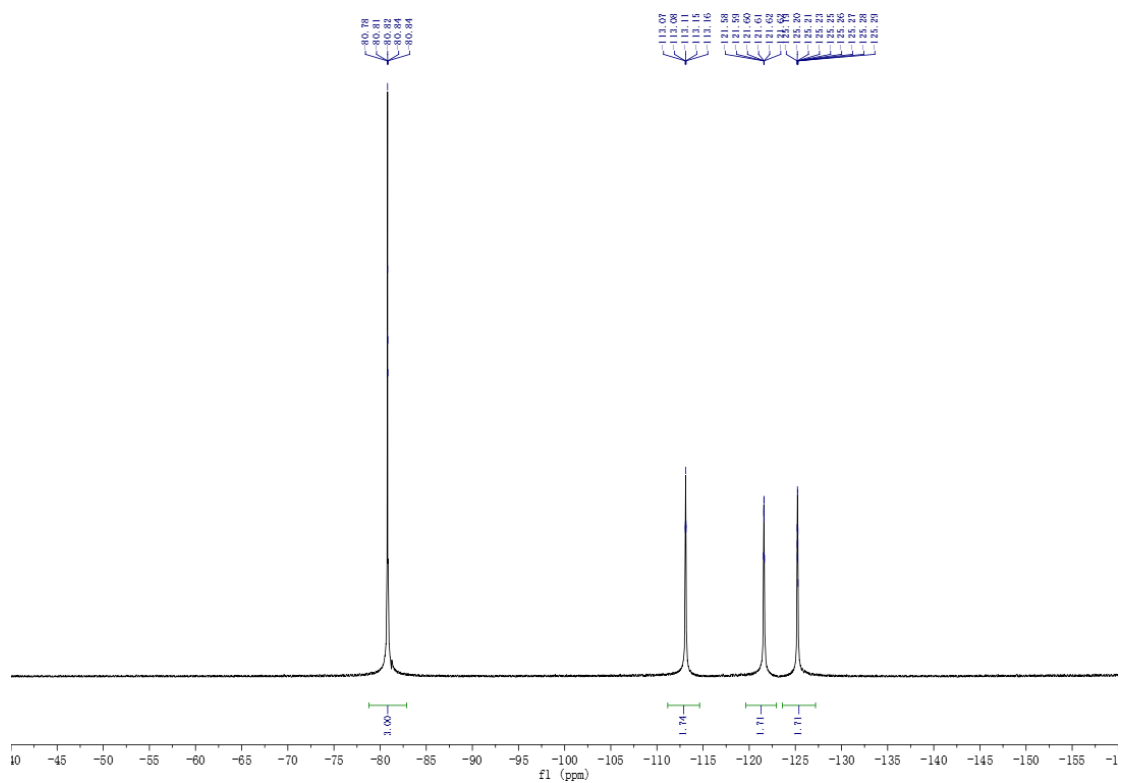
^{19}F NMR of compound **8**



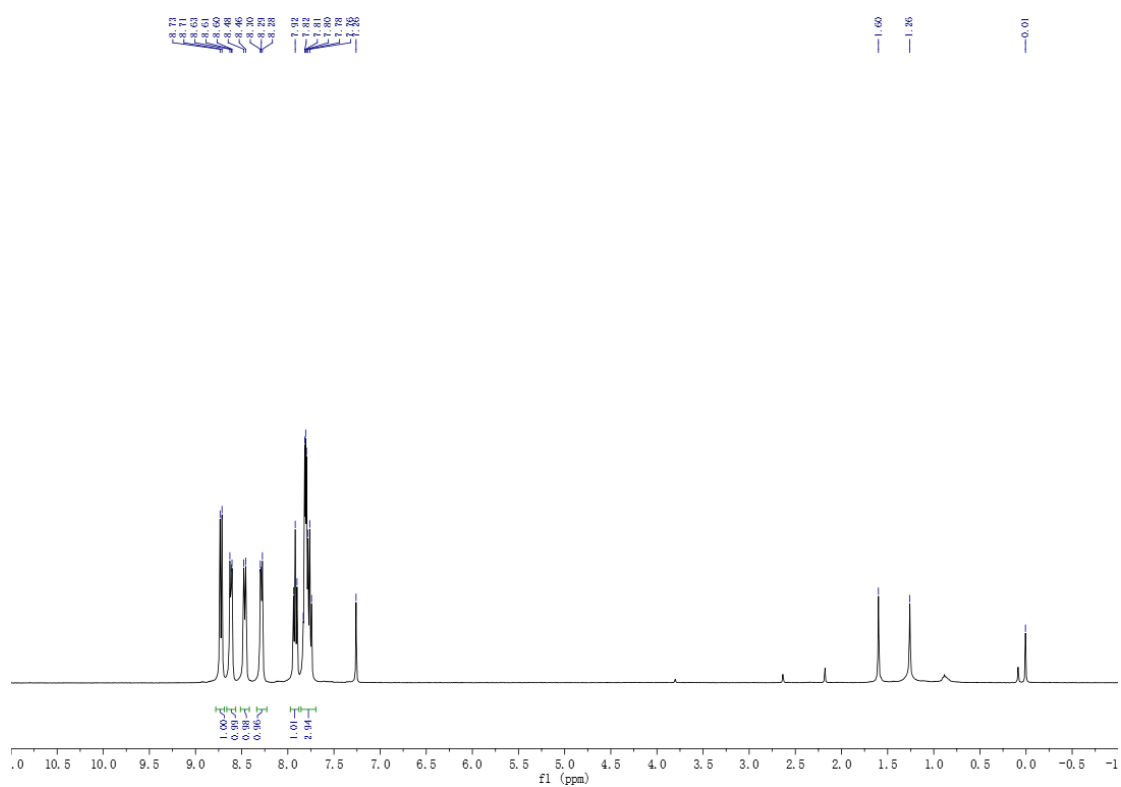
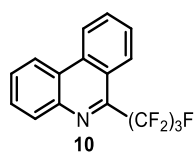
¹H NMR of compound **9**



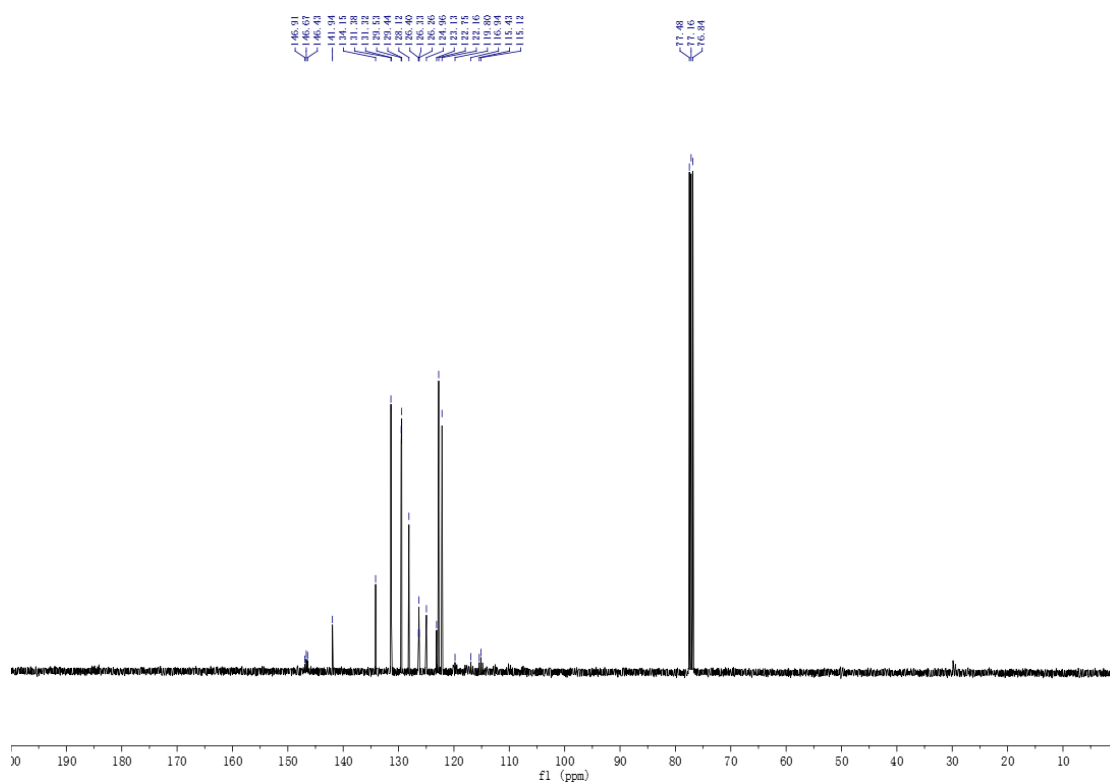
¹³C NMR of compound **9**



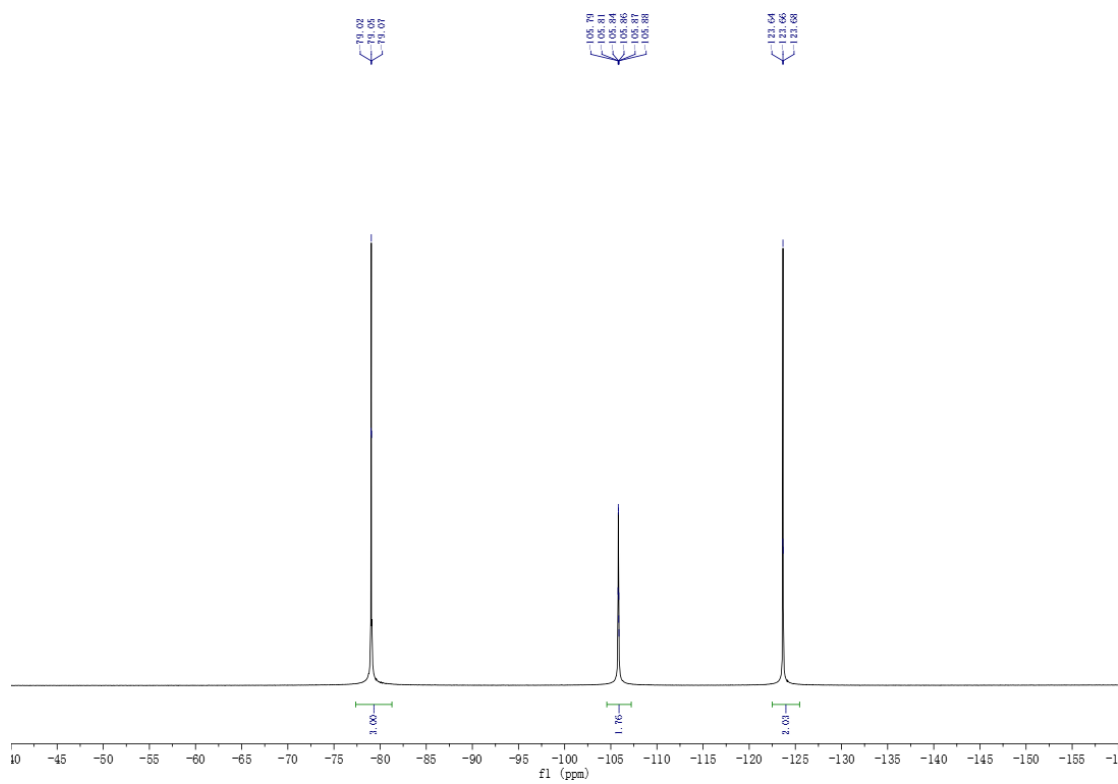
^{19}F NMR of compound **9**



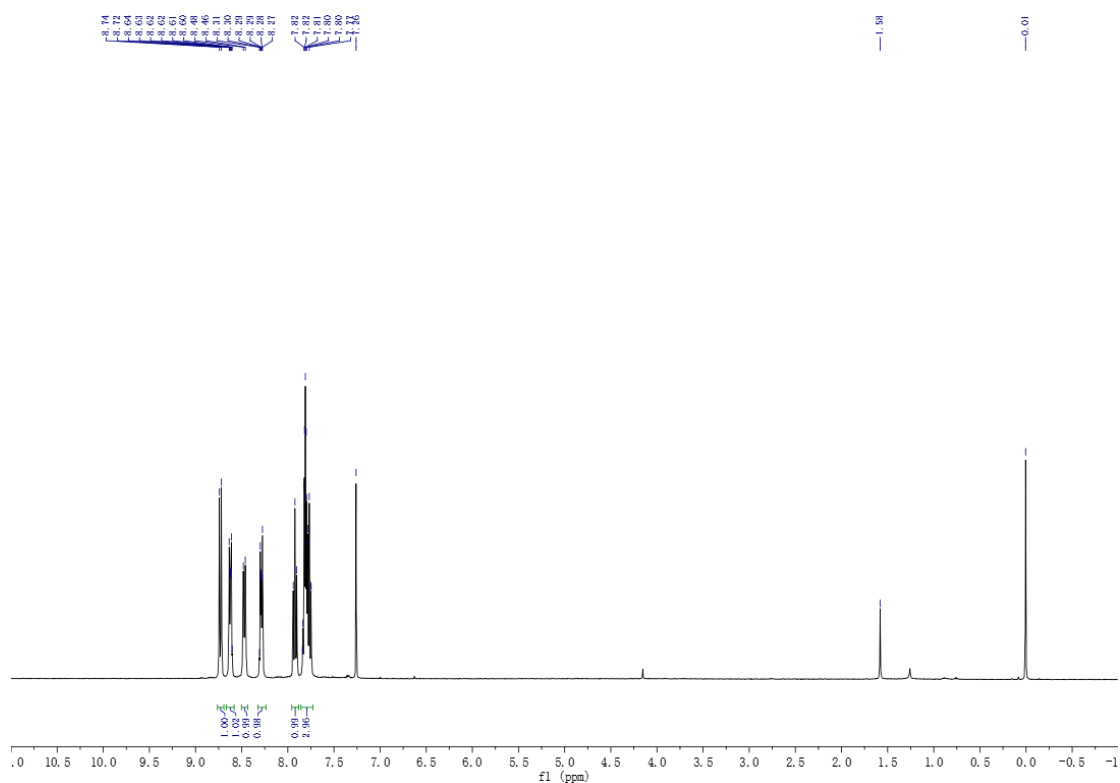
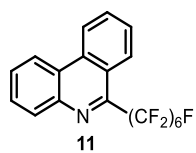
^1H NMR of compound **10**



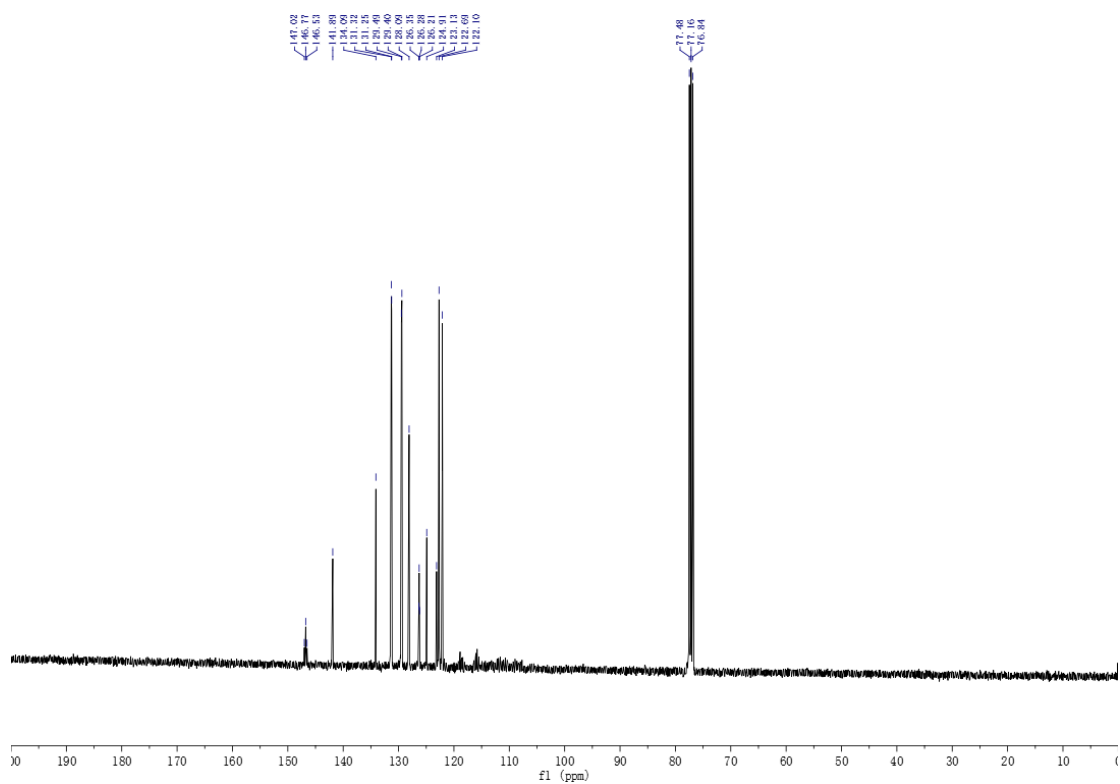
¹³C NMR of compound **10**



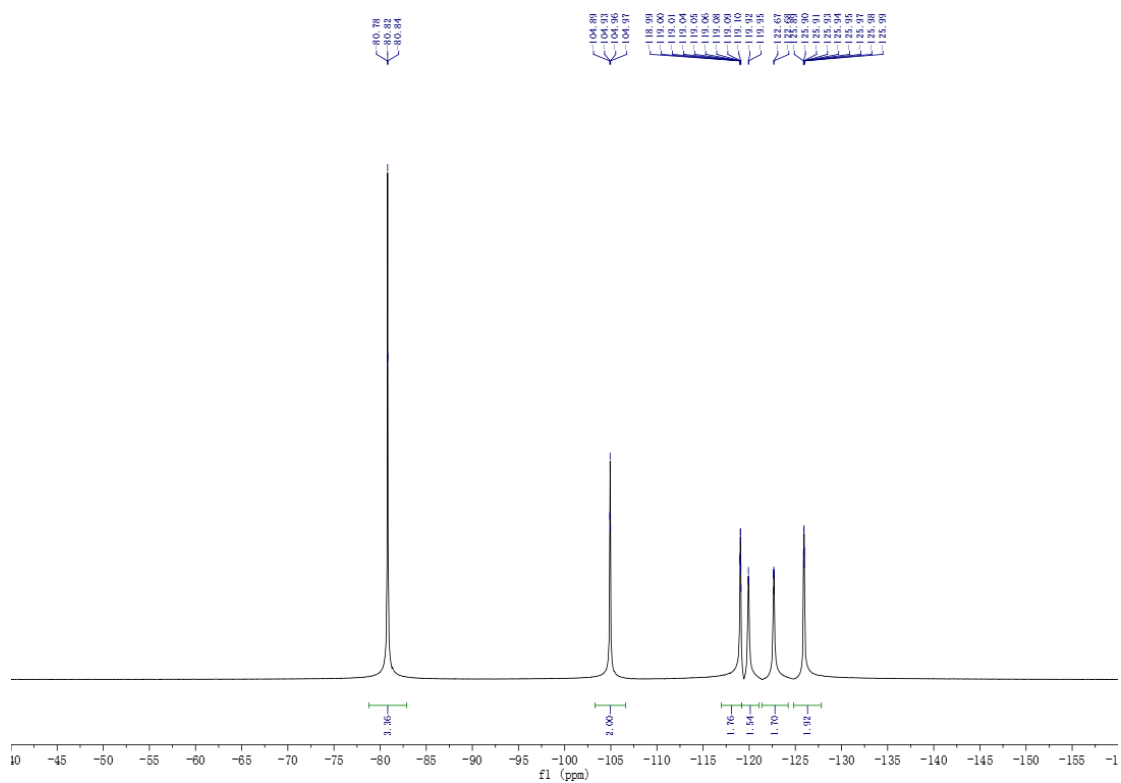
¹⁹F NMR of compound **10**



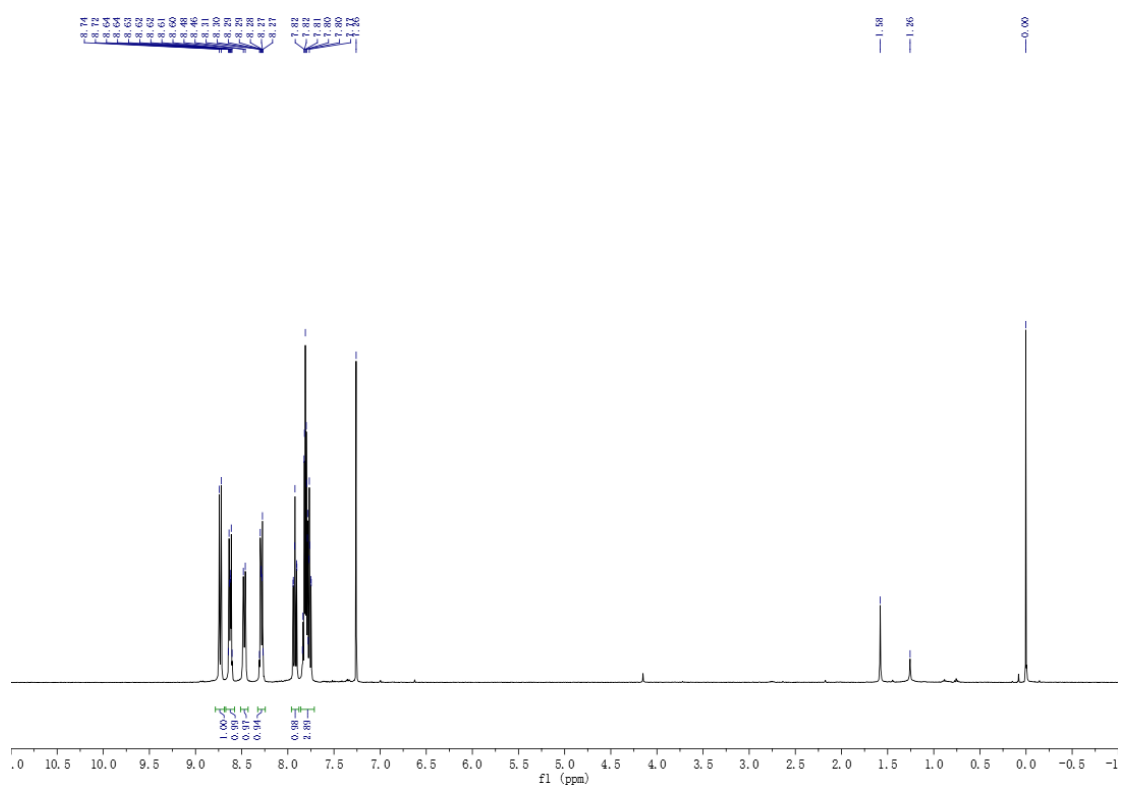
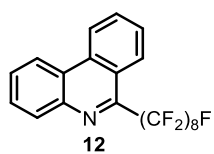
¹H NMR of compound **11**



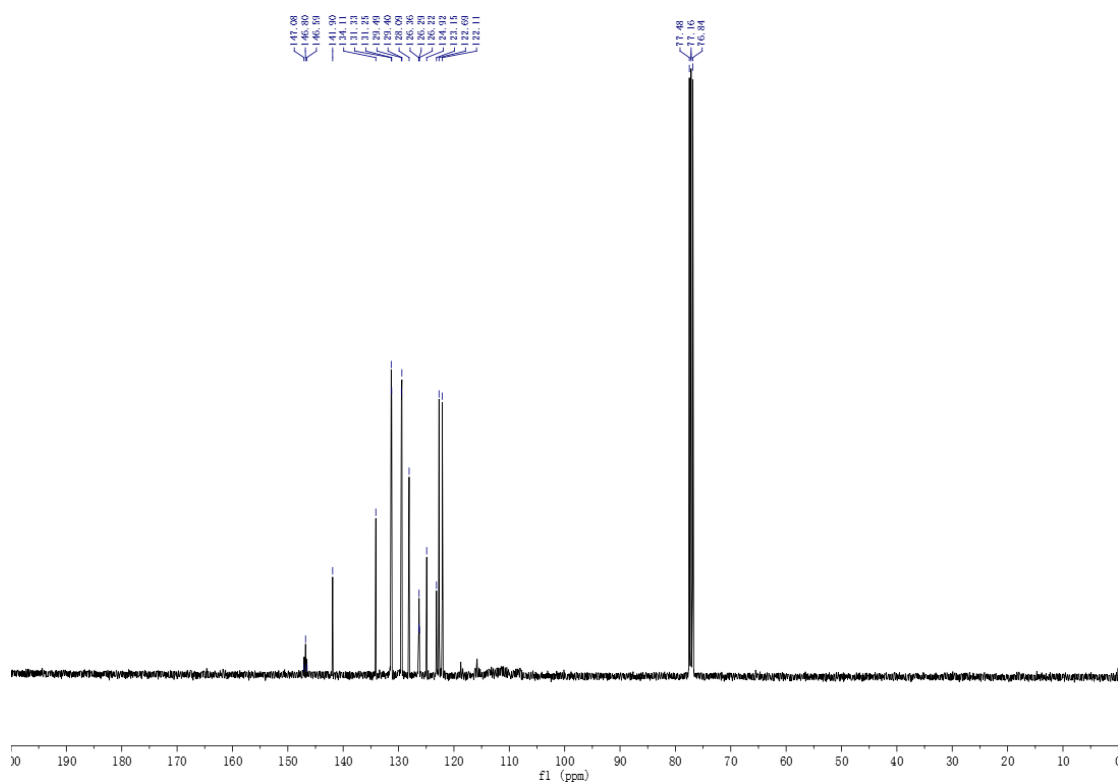
¹³C NMR of compound **11**



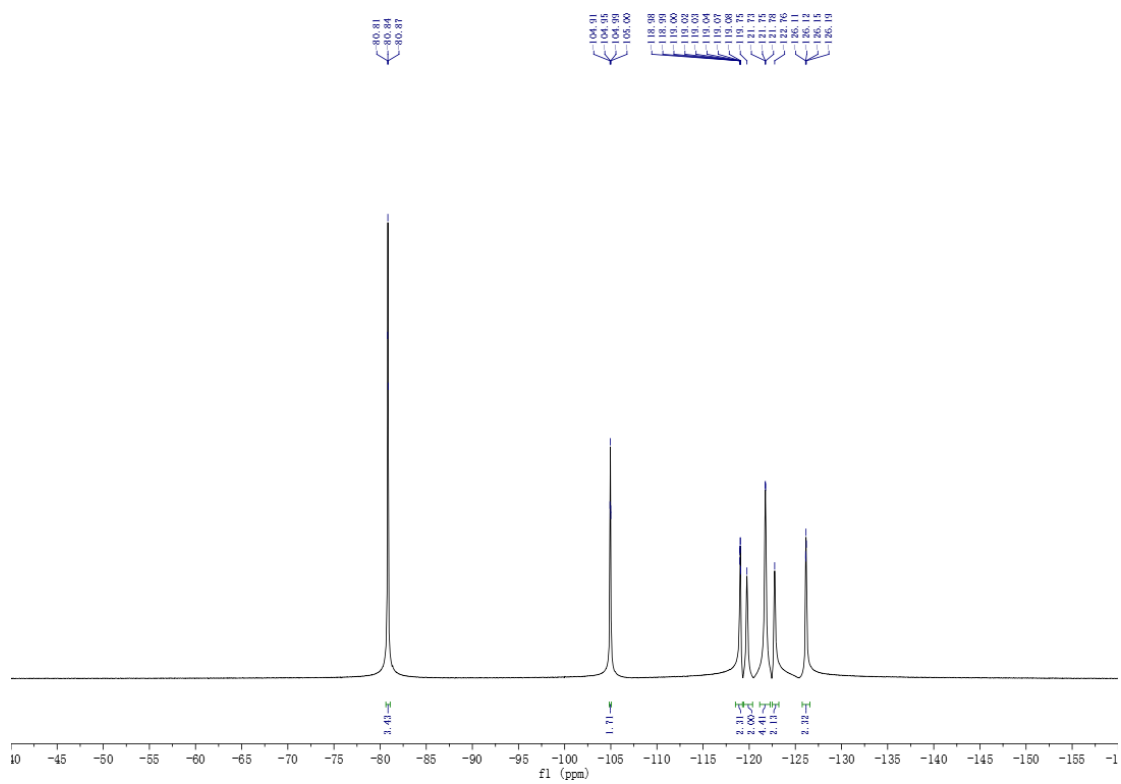
^{19}F NMR of compound **11**



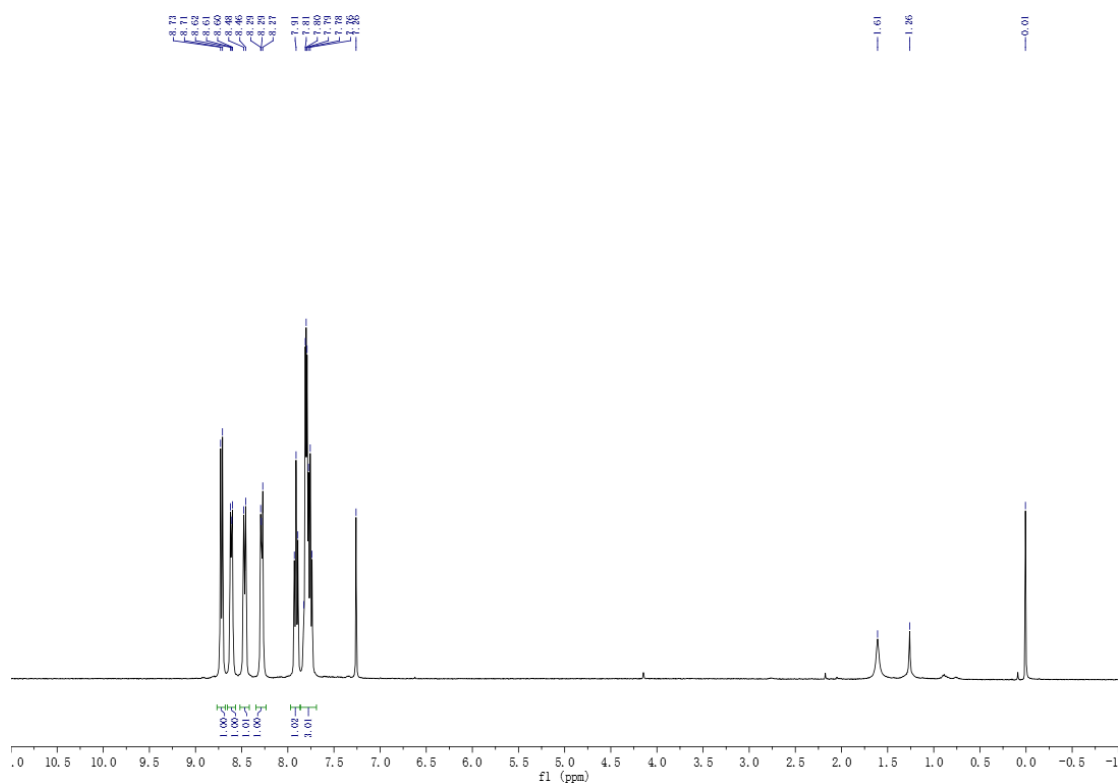
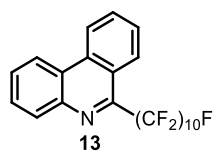
^1H NMR of compound **12**



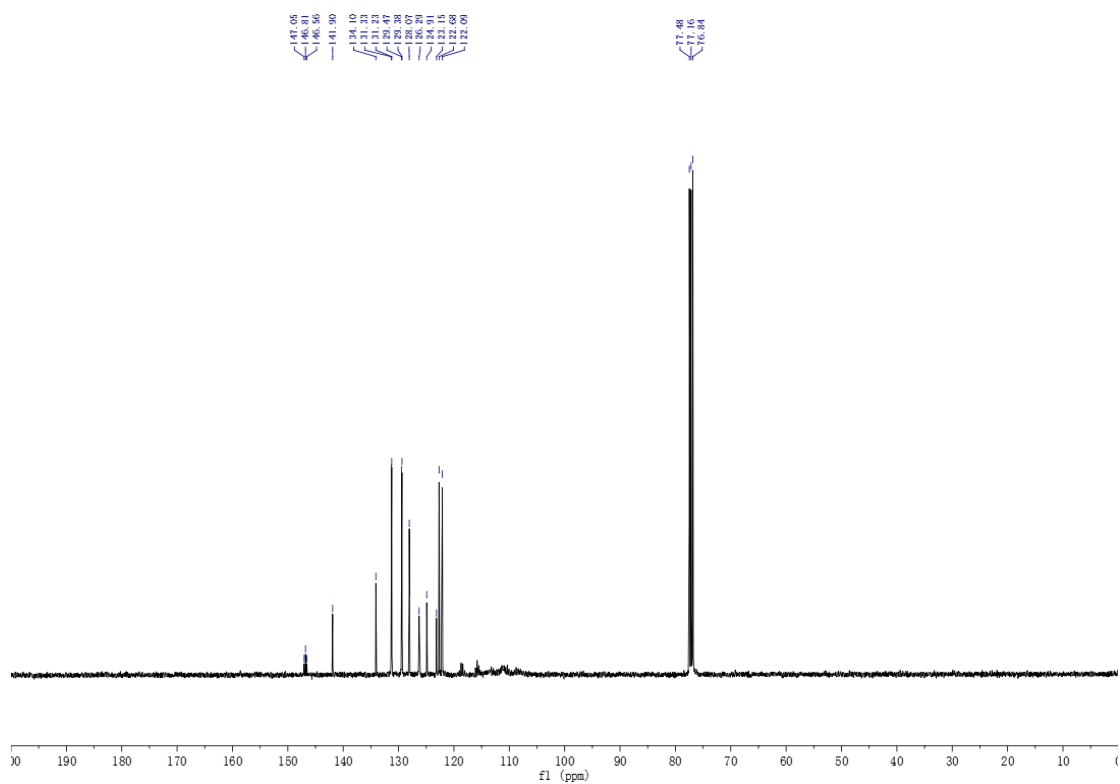
^{13}C NMR of compound **12**



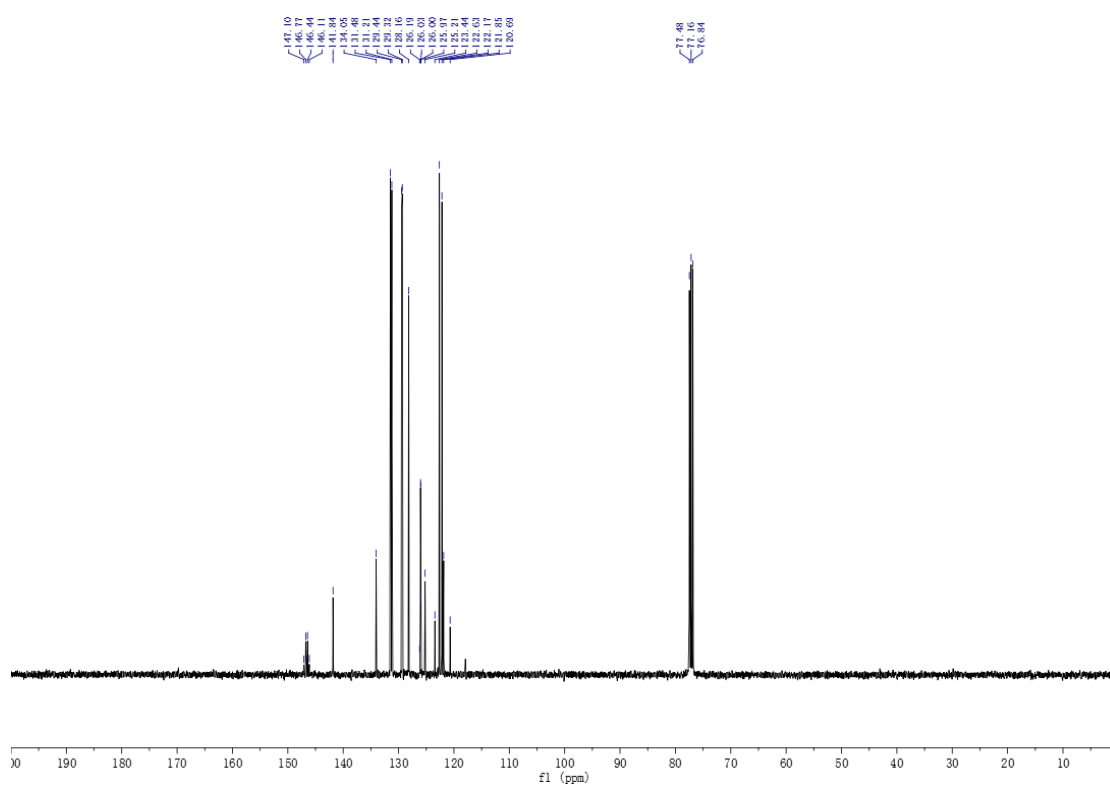
^{19}F NMR of compound **12**



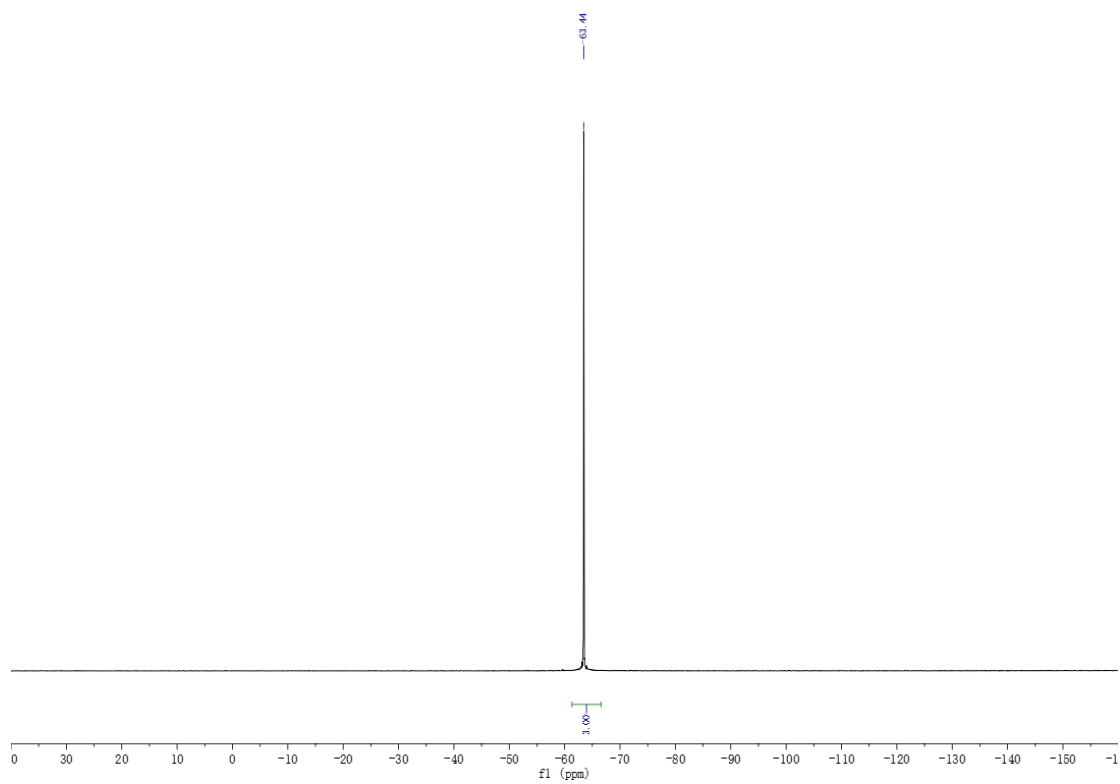
¹H NMR of compound **13**



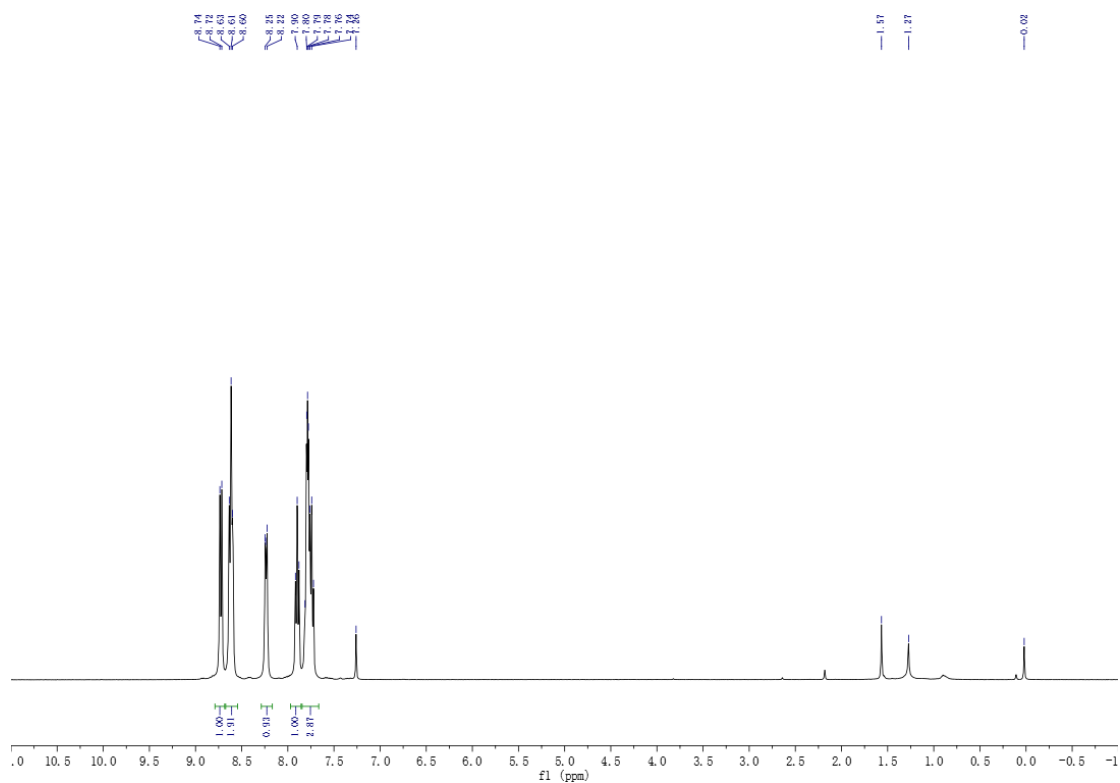
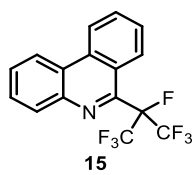
¹³C NMR of compound **13**



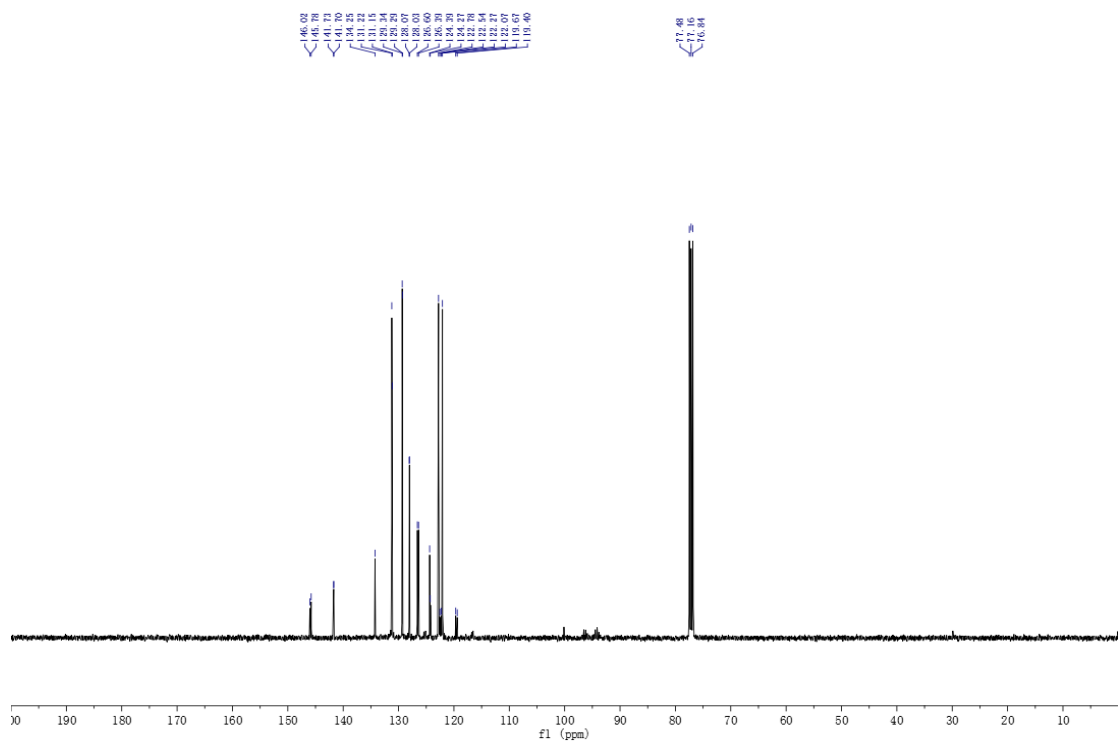
^{13}C NMR of compound **14**



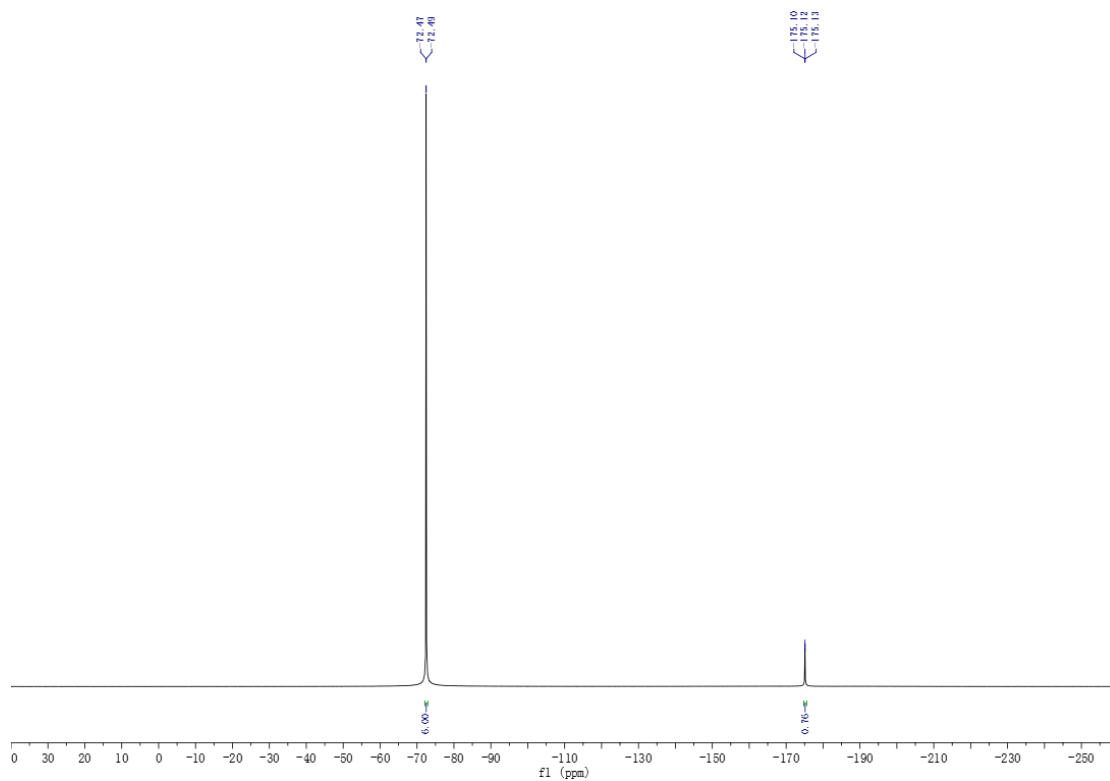
^{19}F NMR of compound **14**



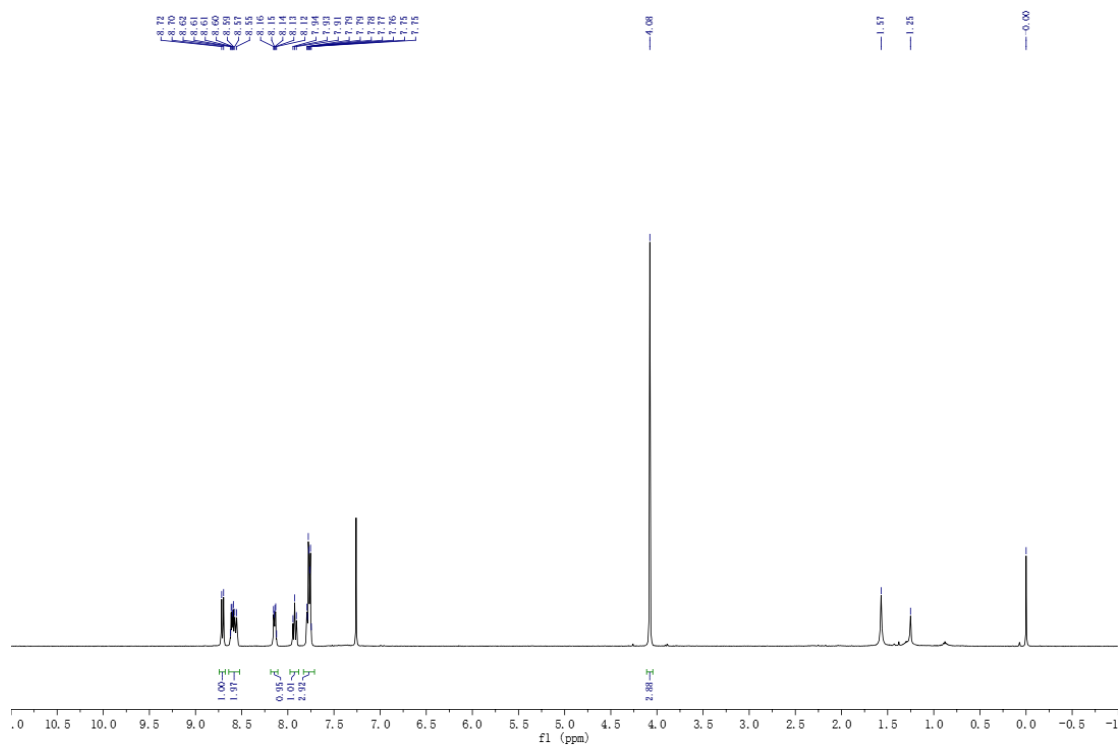
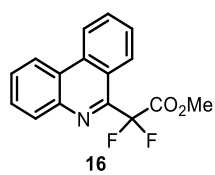
¹H NMR of compound **15**



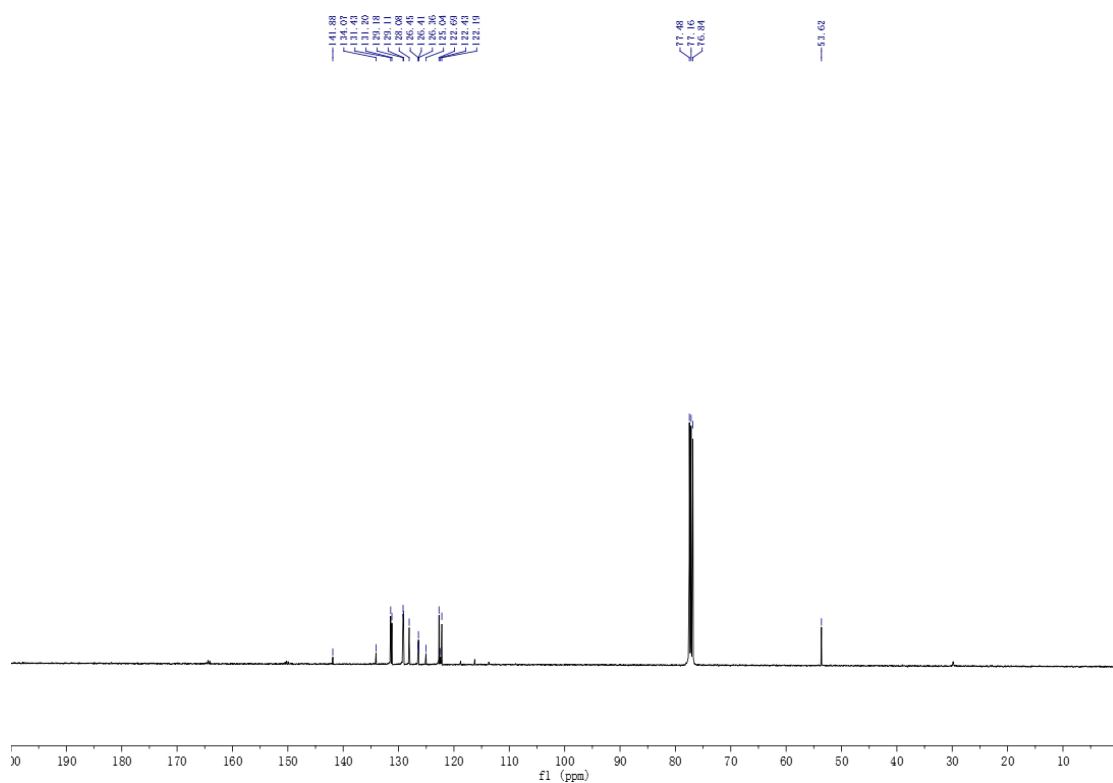
¹³C NMR of compound **15**



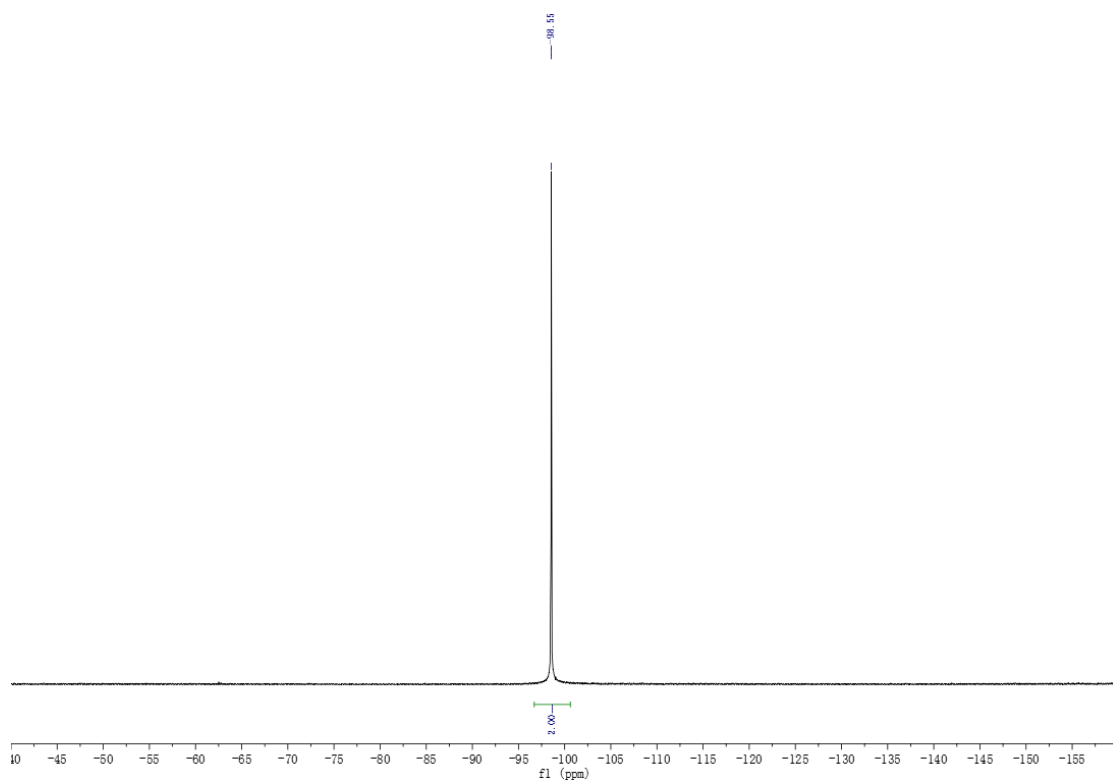
^{19}F NMR of compound **15**



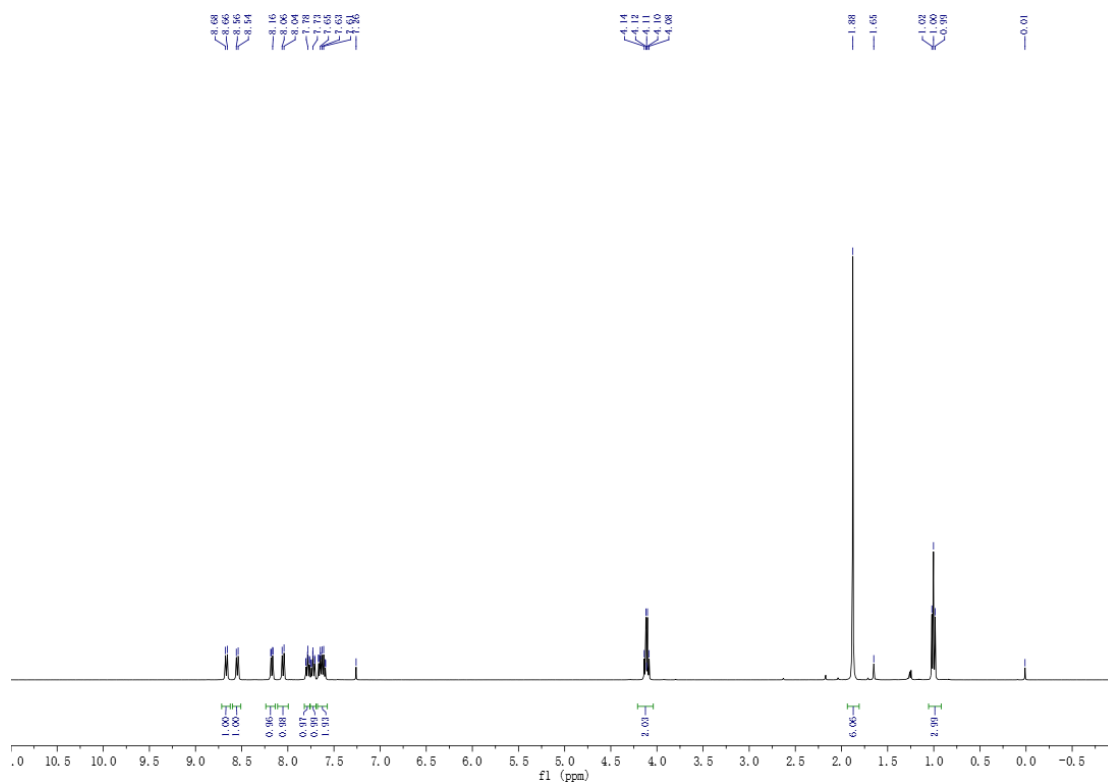
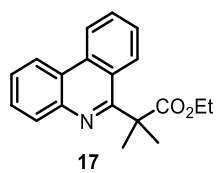
^1H NMR of compound **16**



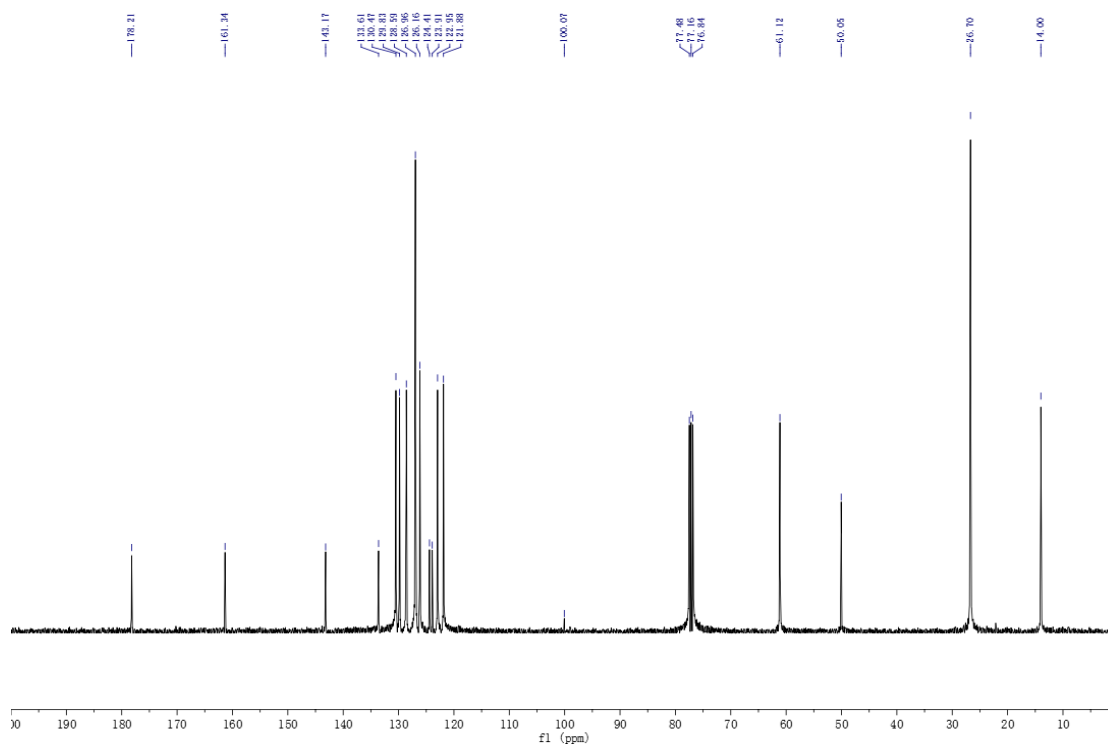
^{13}C NMR of compound **16**



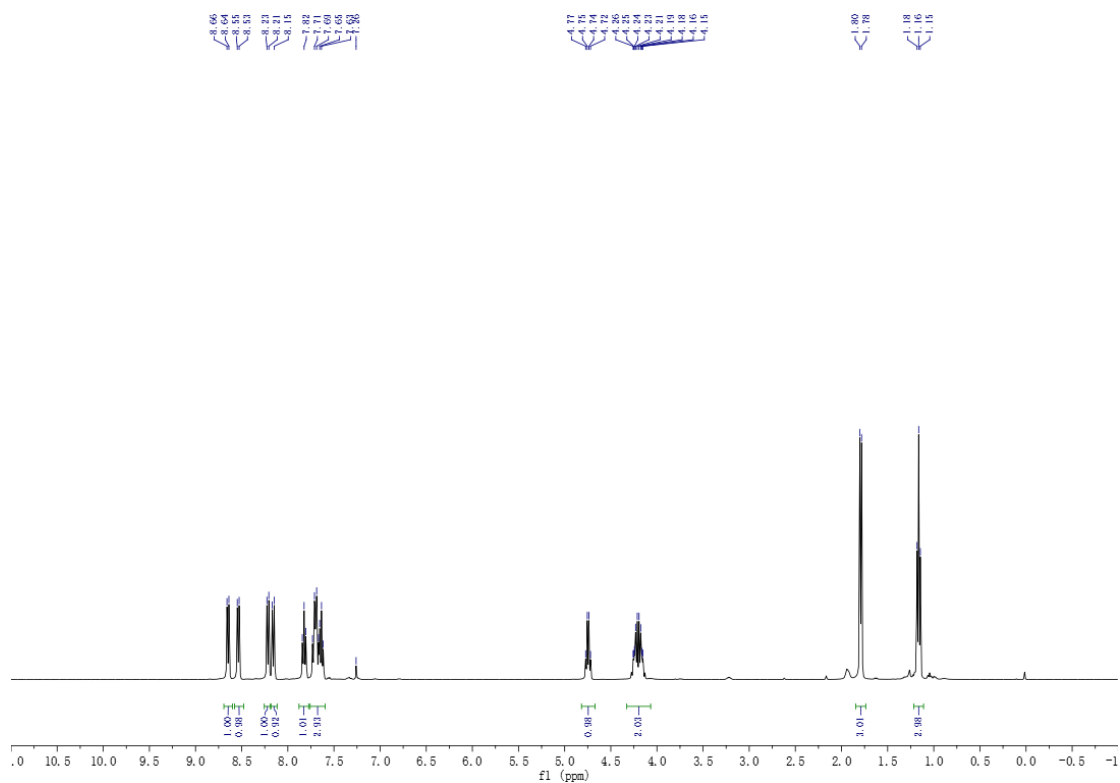
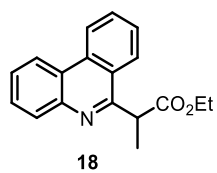
^{19}F NMR of compound **16**



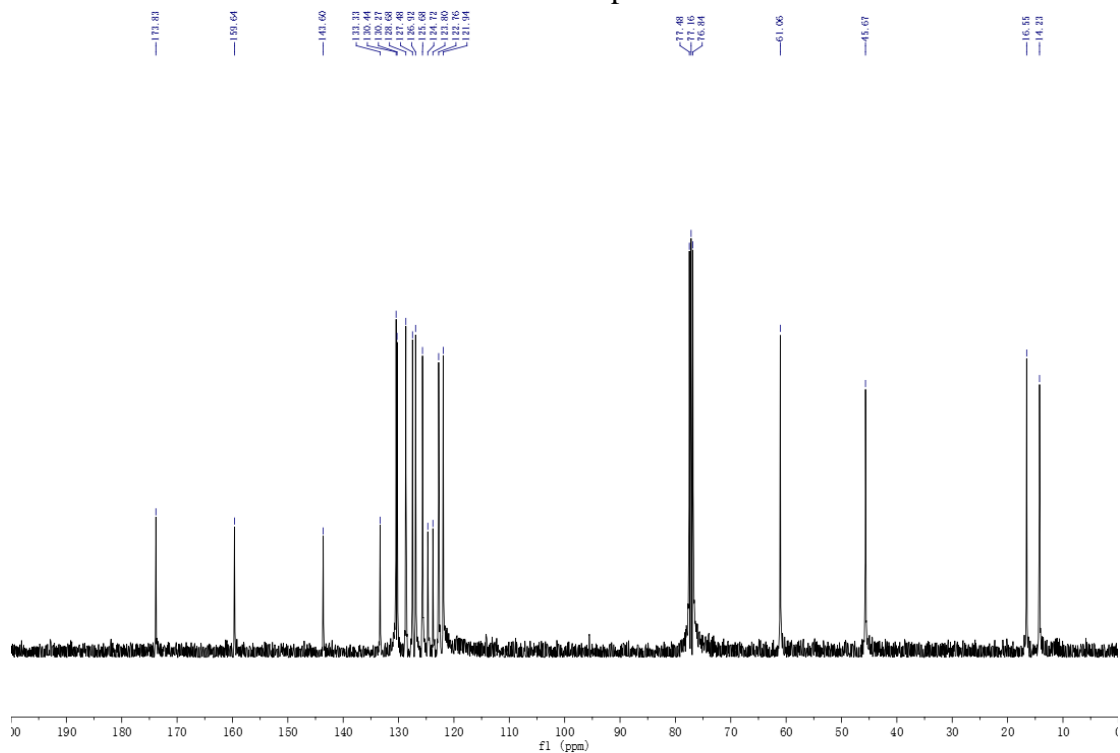
¹H NMR of compound **17**



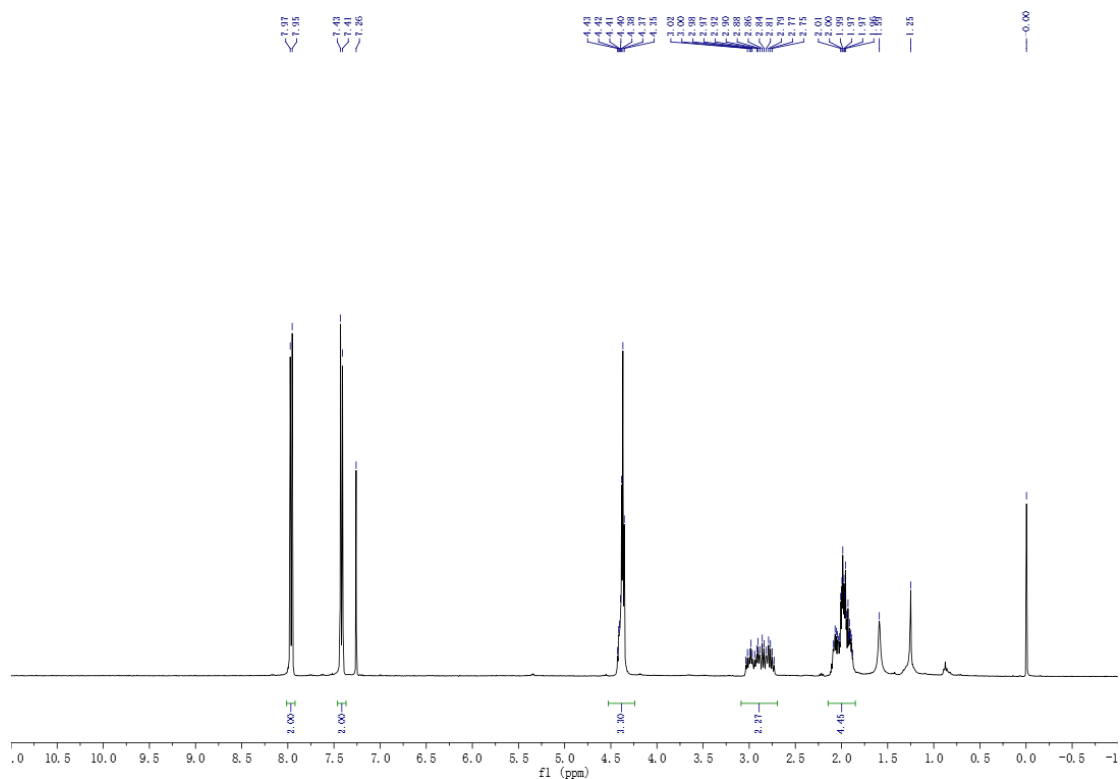
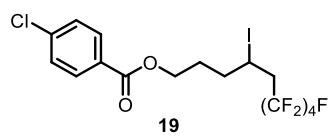
¹³C NMR of compound **17**



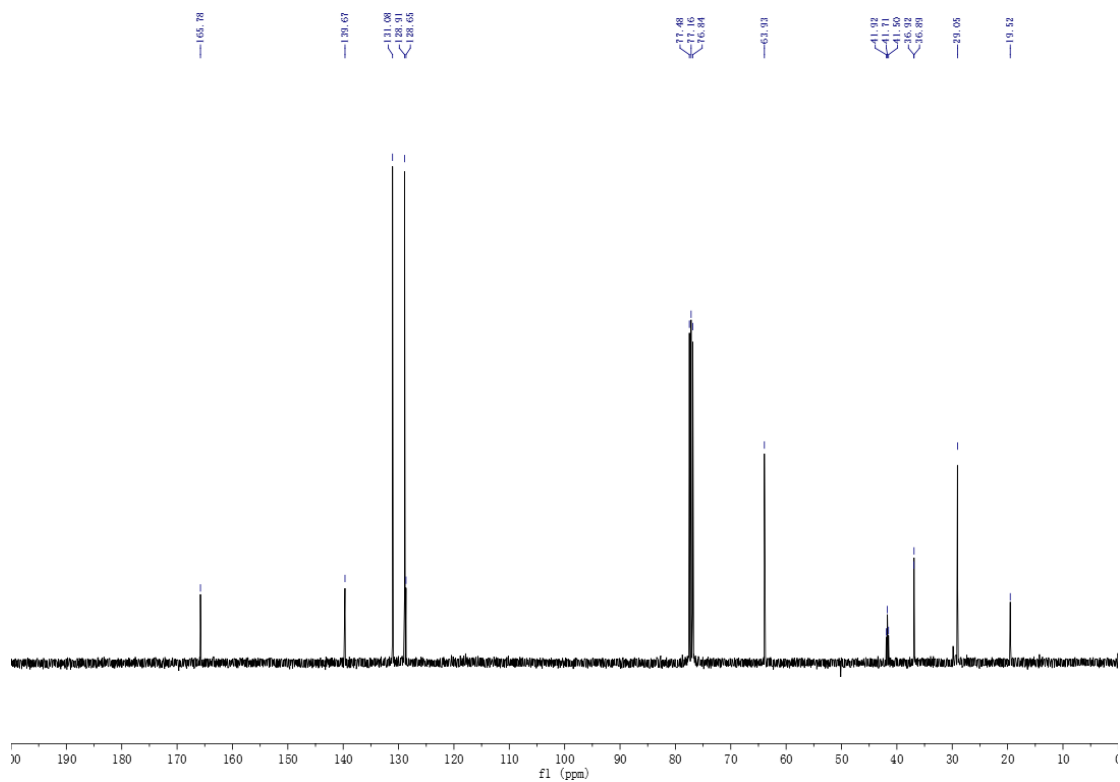
¹H NMR of compound **18**



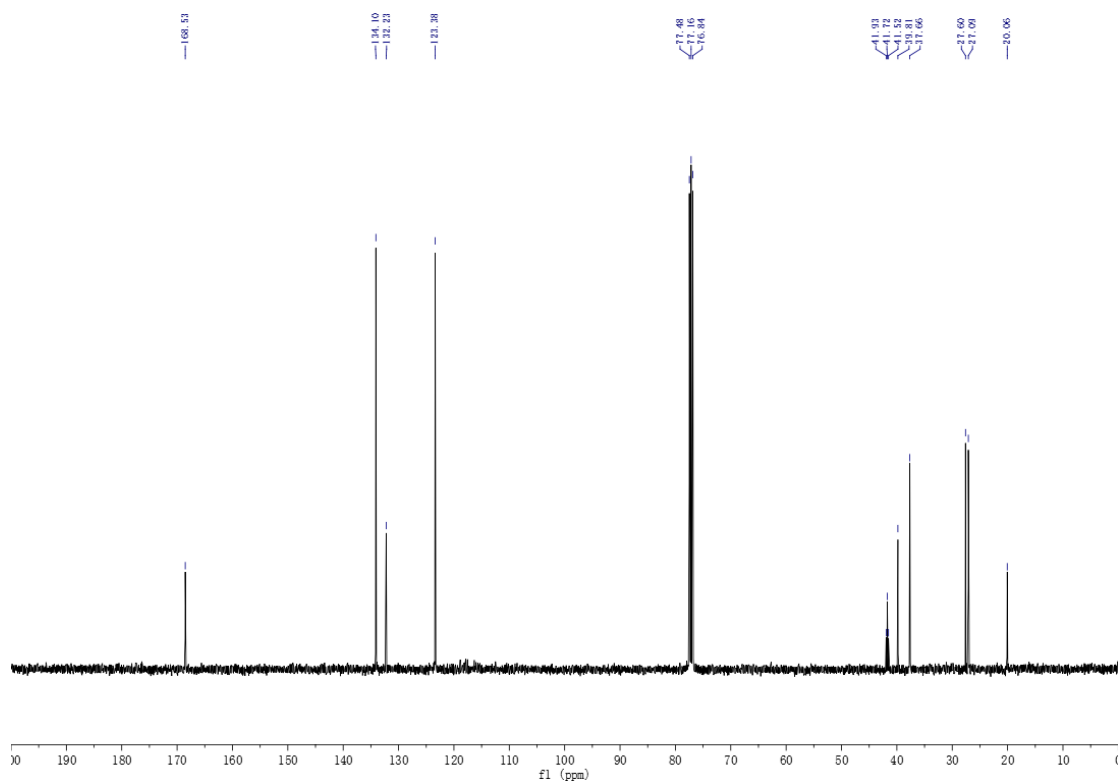
¹³C NMR of compound **18**



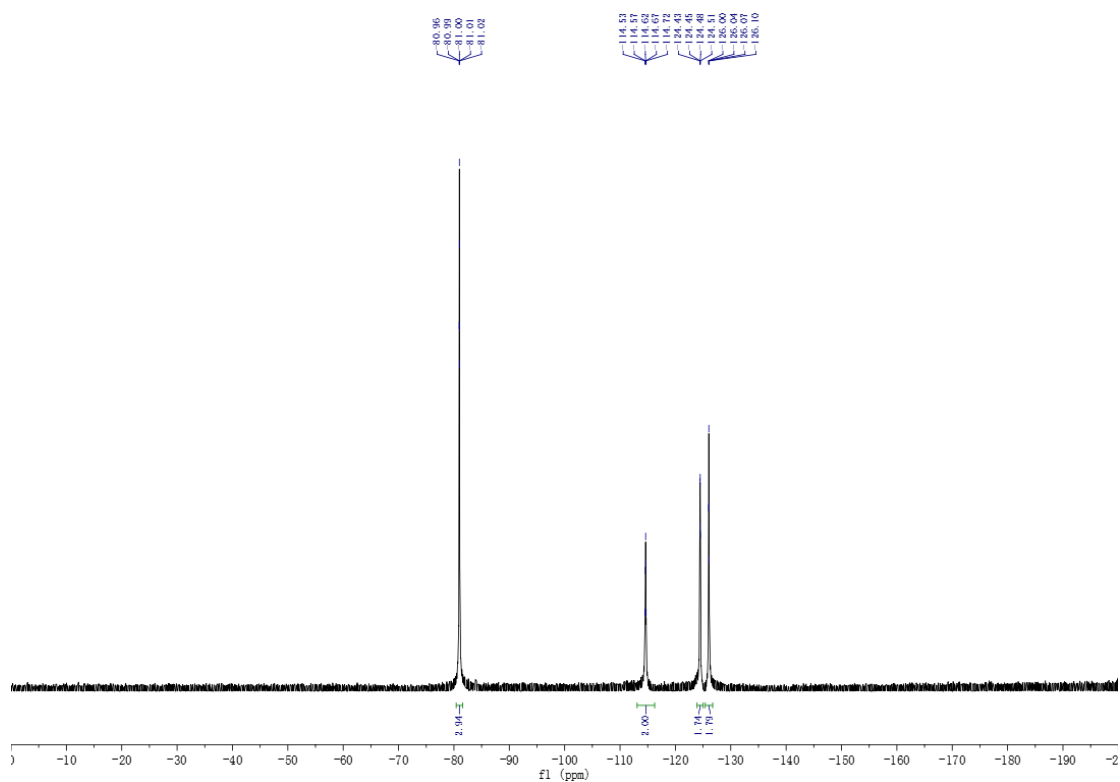
¹H NMR of compound **19**



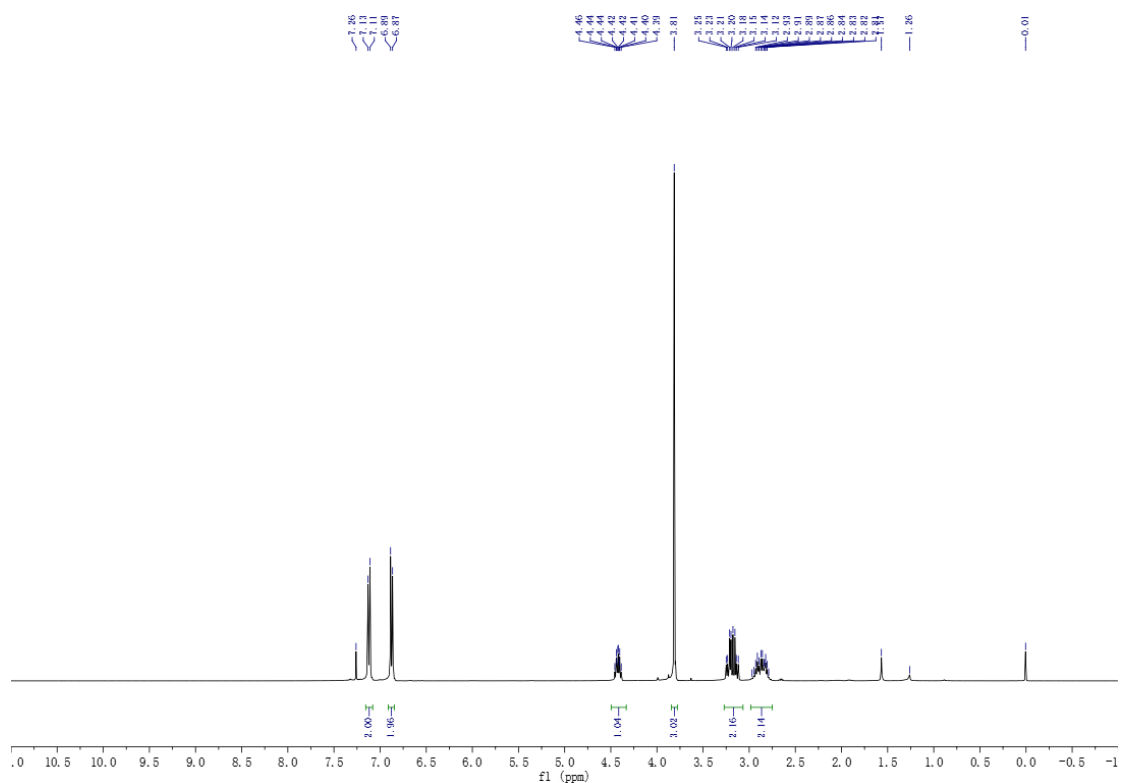
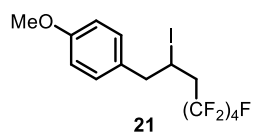
¹³C NMR of compound **19**



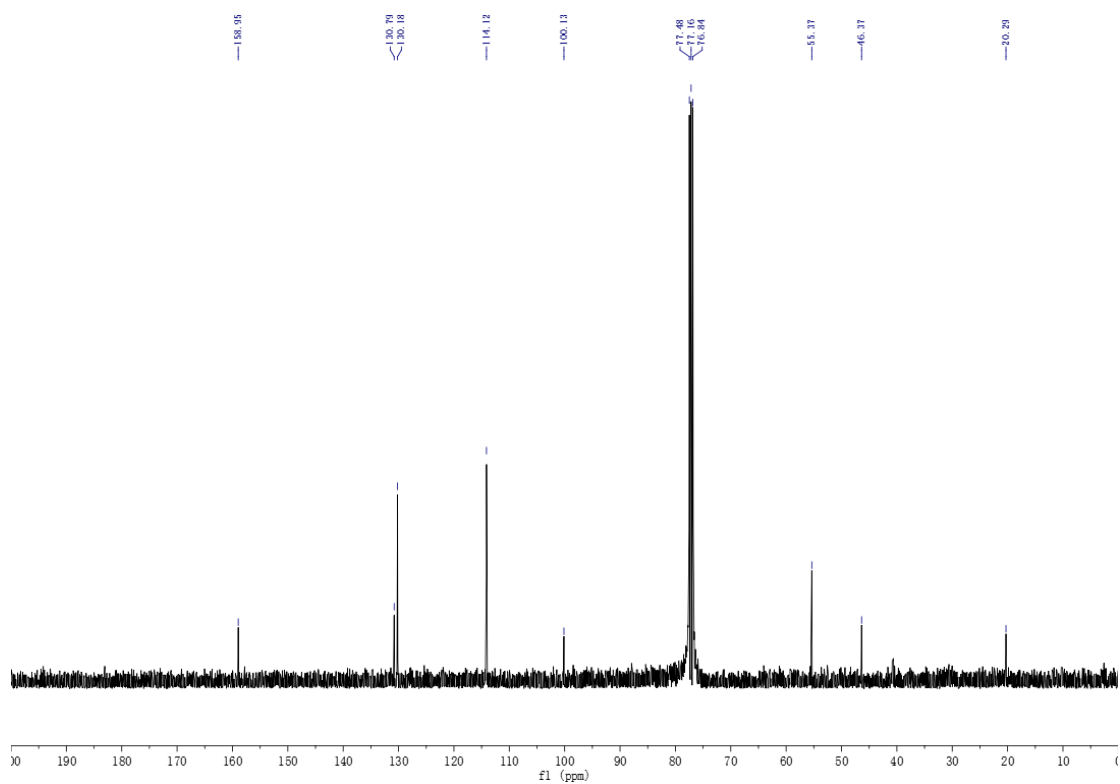
¹³C NMR of compound **20**



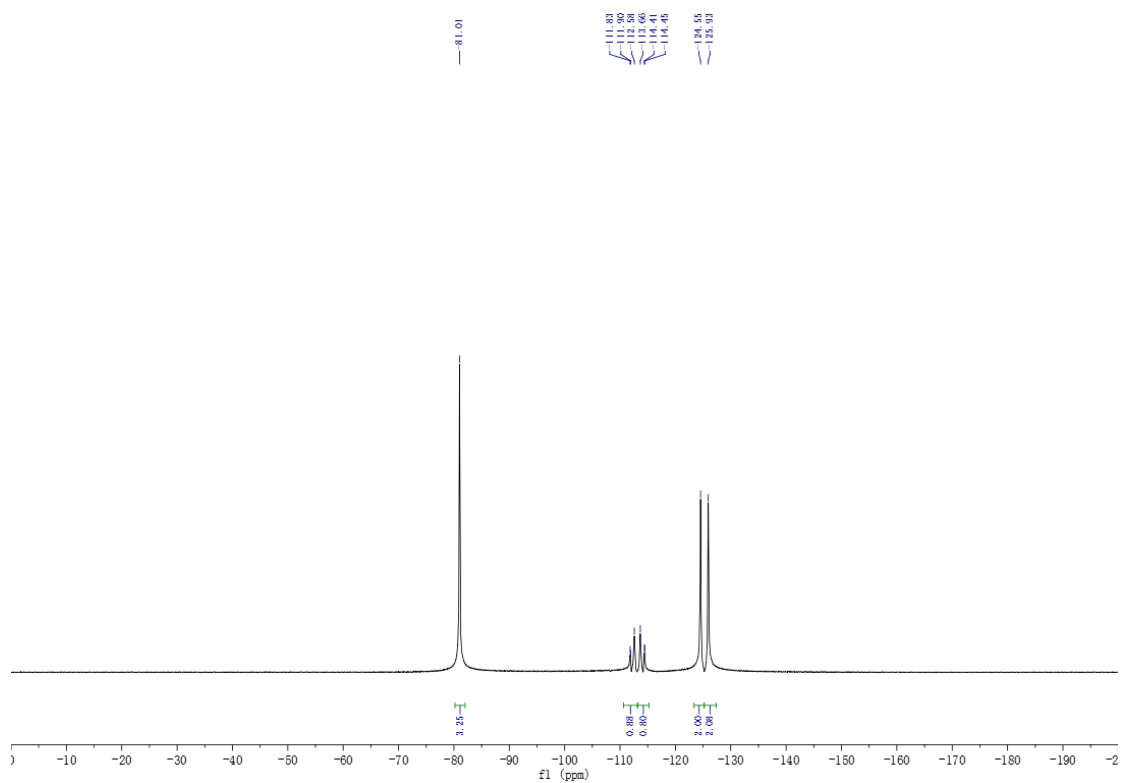
¹⁹F NMR of compound **20**



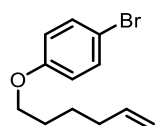
¹H NMR of compound **21**



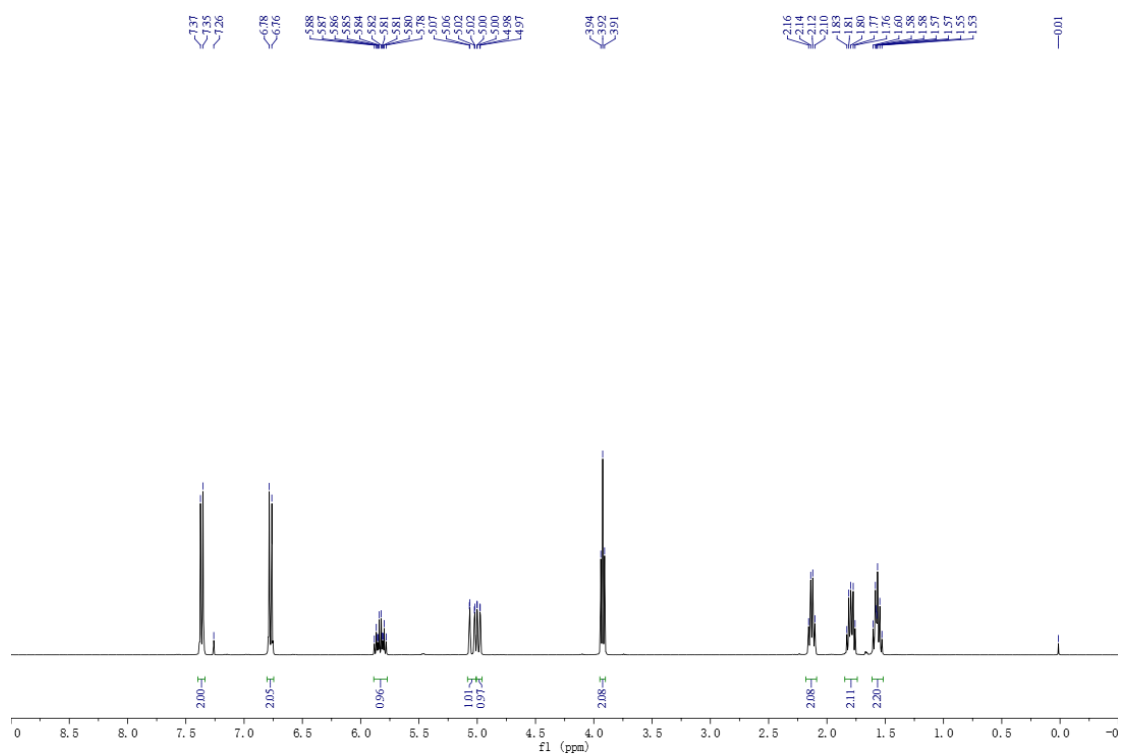
¹³C NMR of compound **21**



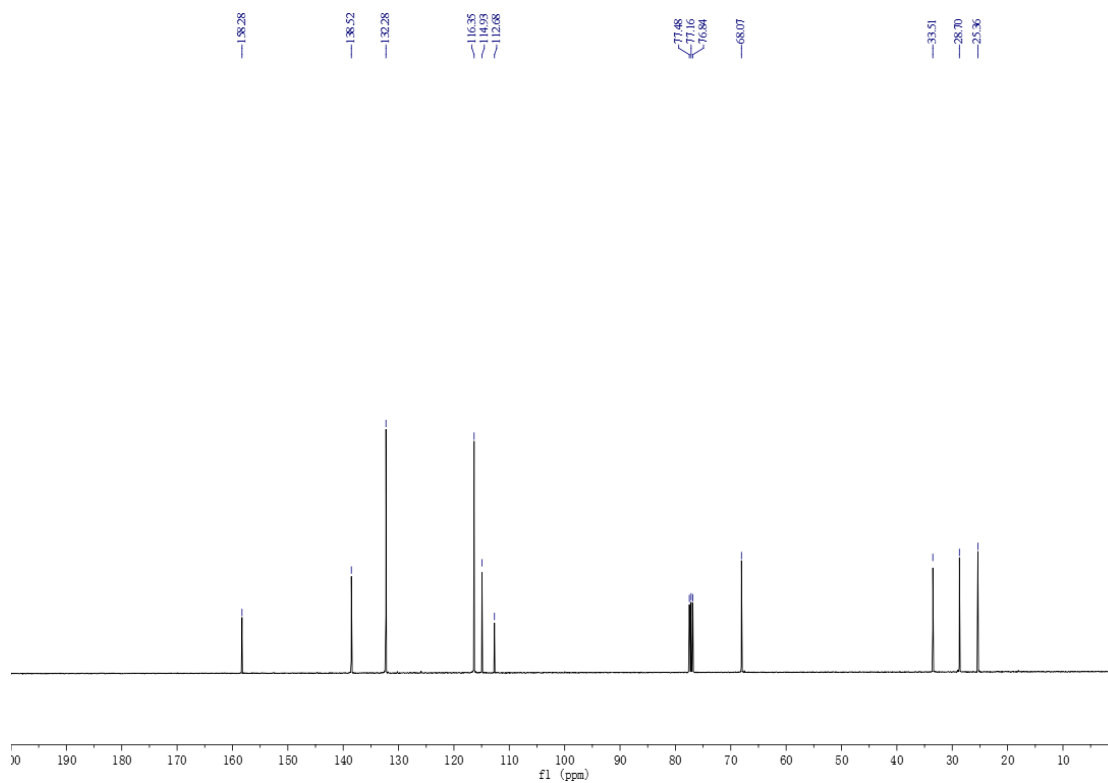
¹⁹F NMR of compound **21**



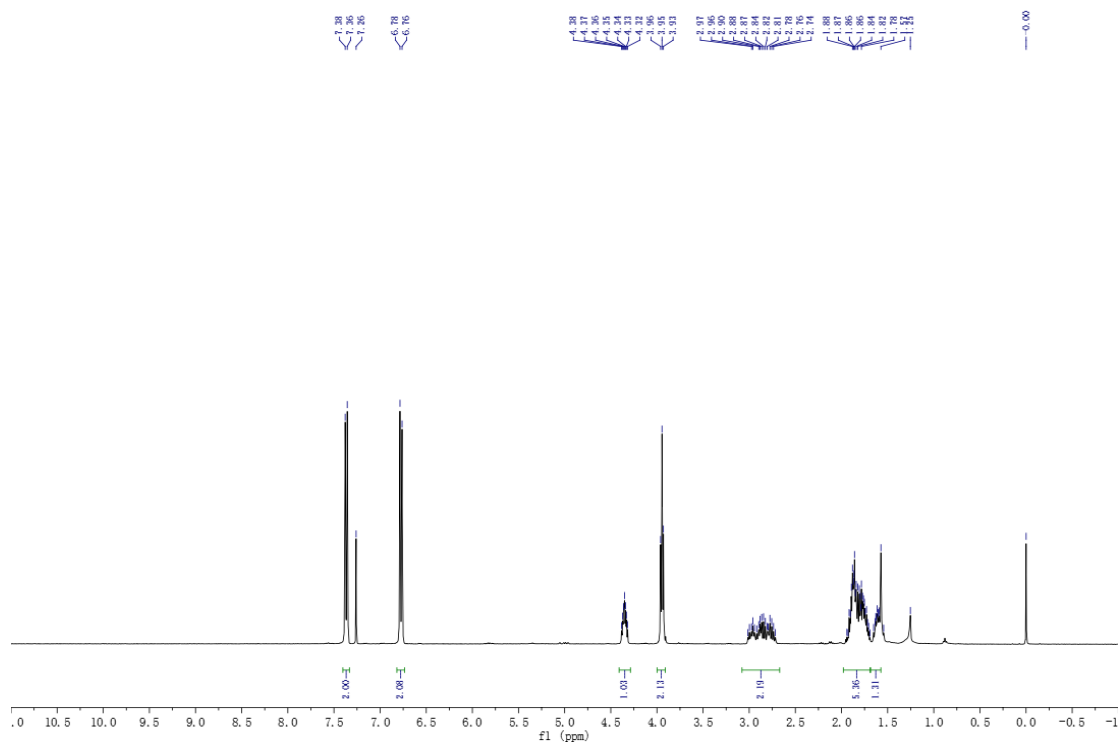
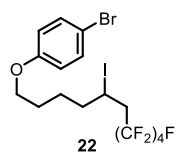
22-1



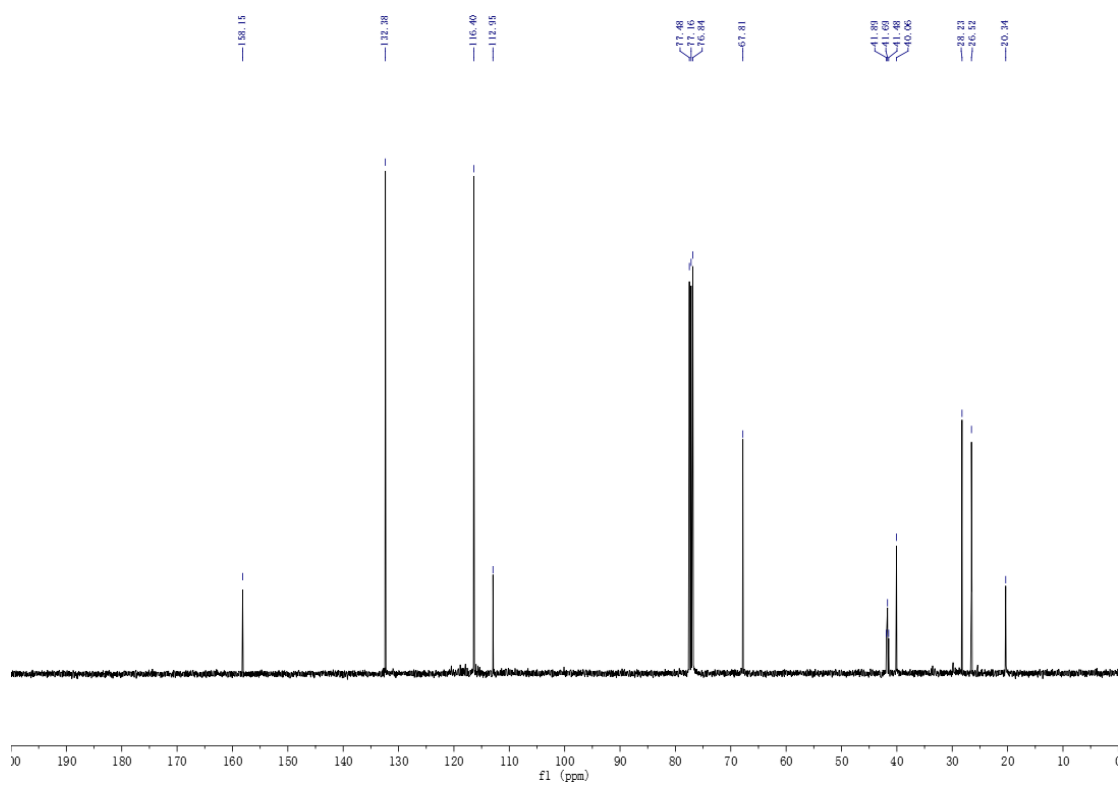
¹H NMR of compound **22-1**



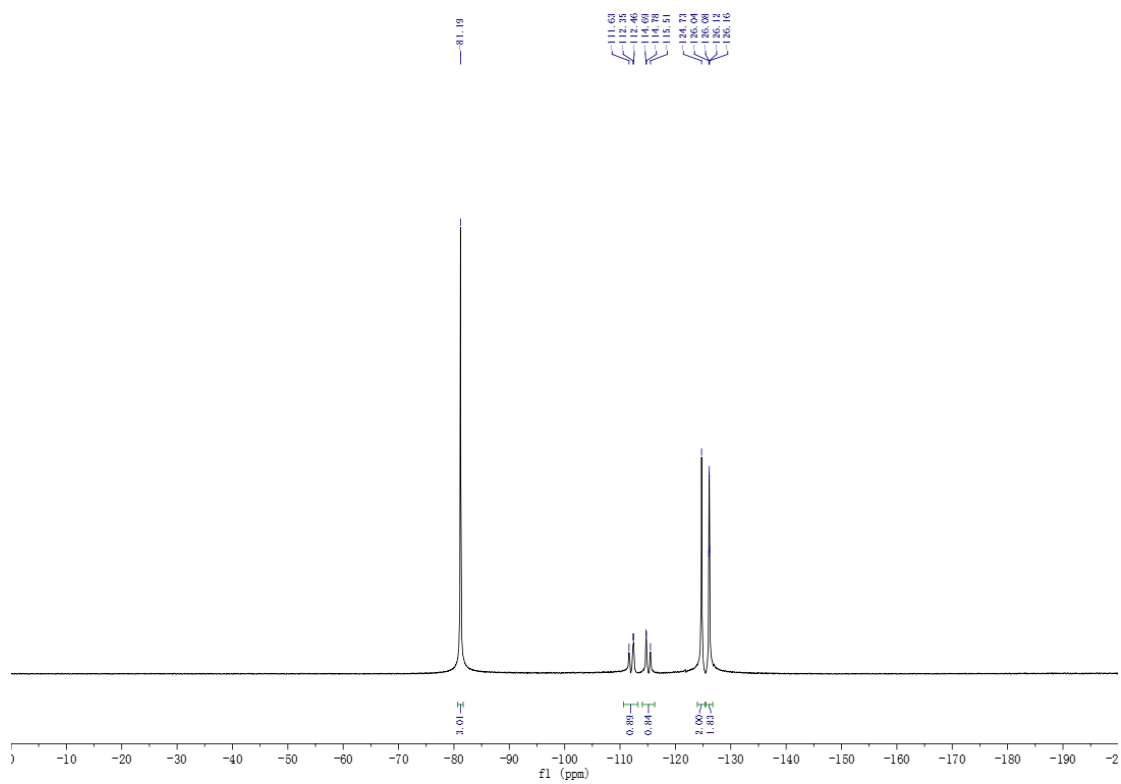
¹³C NMR of compound 22-1



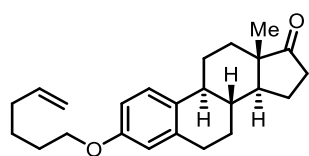
¹H NMR of compound 22



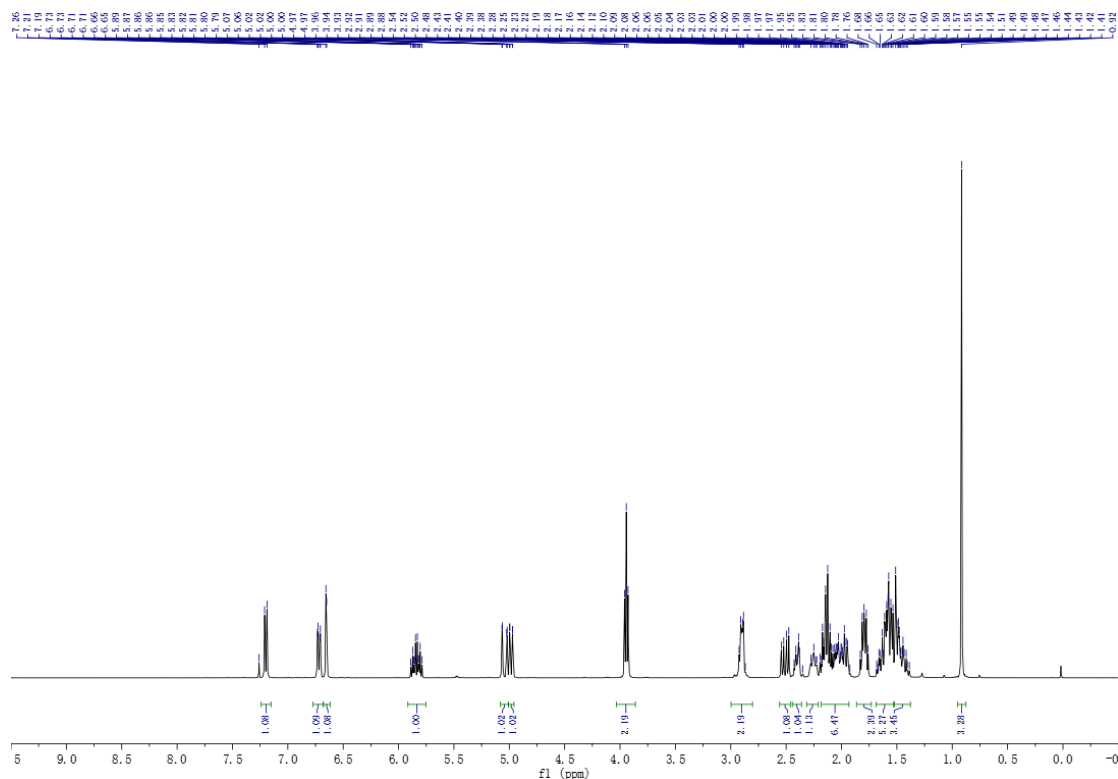
¹³C NMR of compound **22**



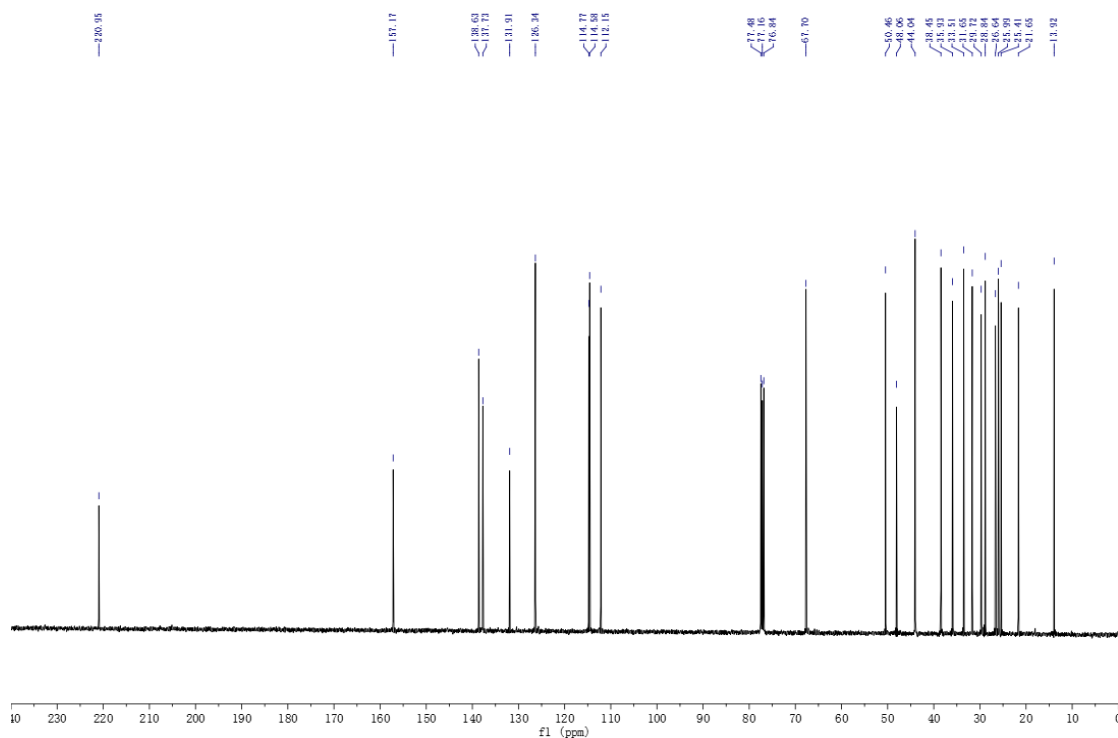
¹⁹F NMR of compound **22**



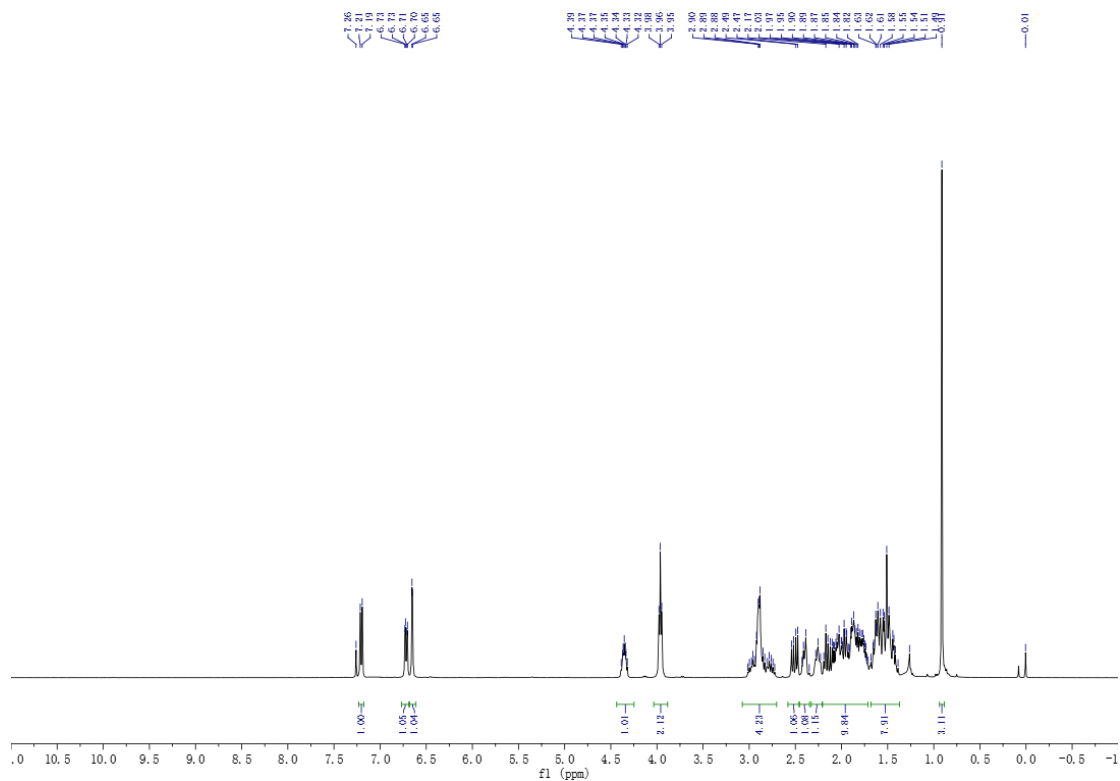
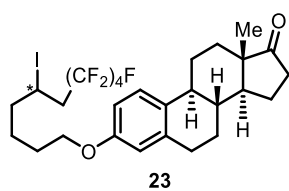
23-1



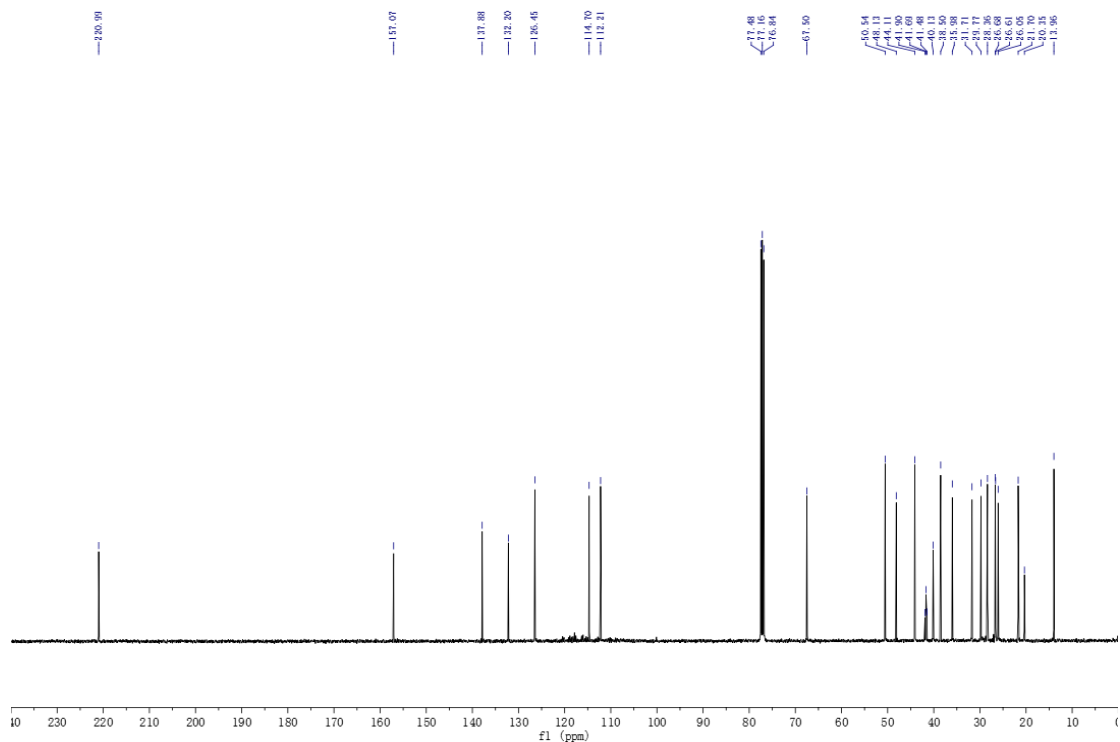
^1H NMR of compound **23-1**



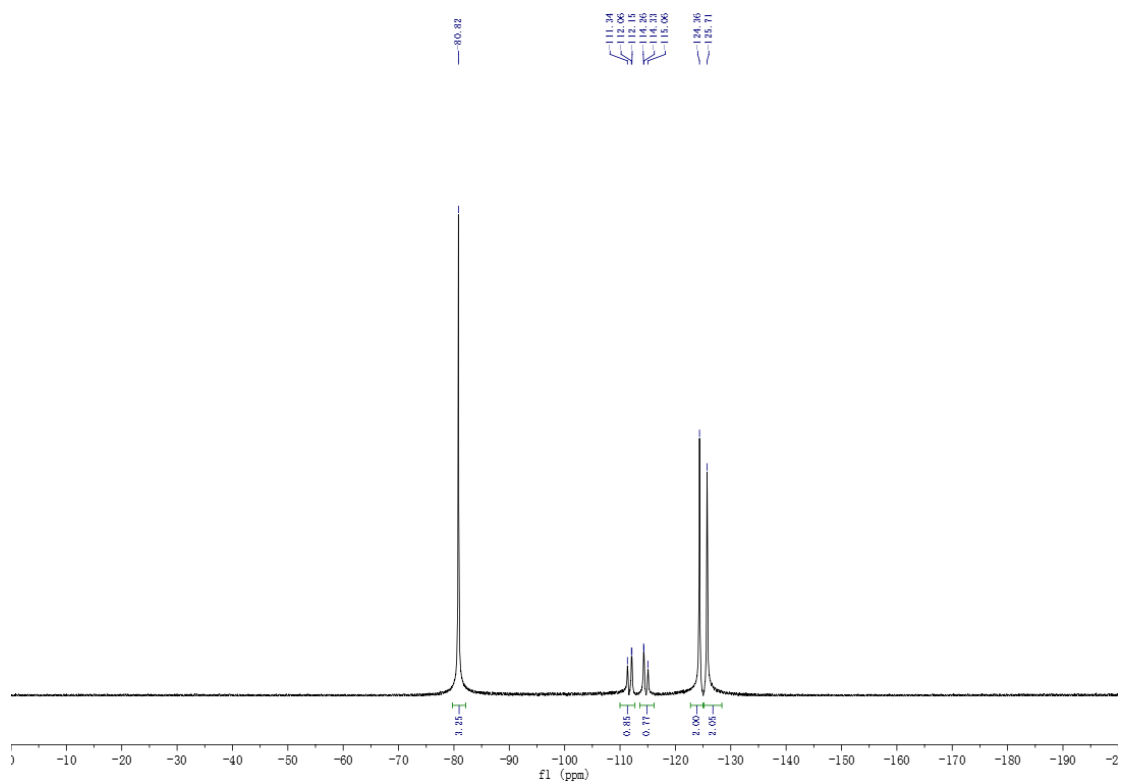
^{13}C NMR of compound **23-1**



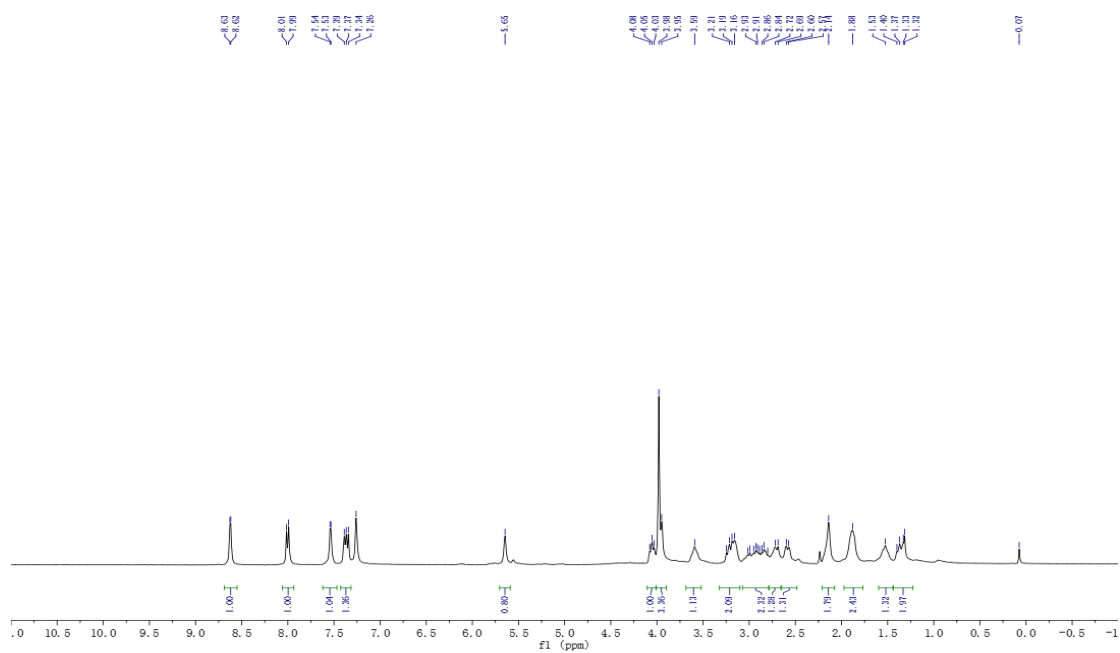
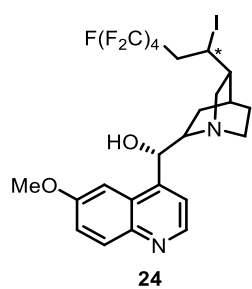
¹H NMR of compound **23**



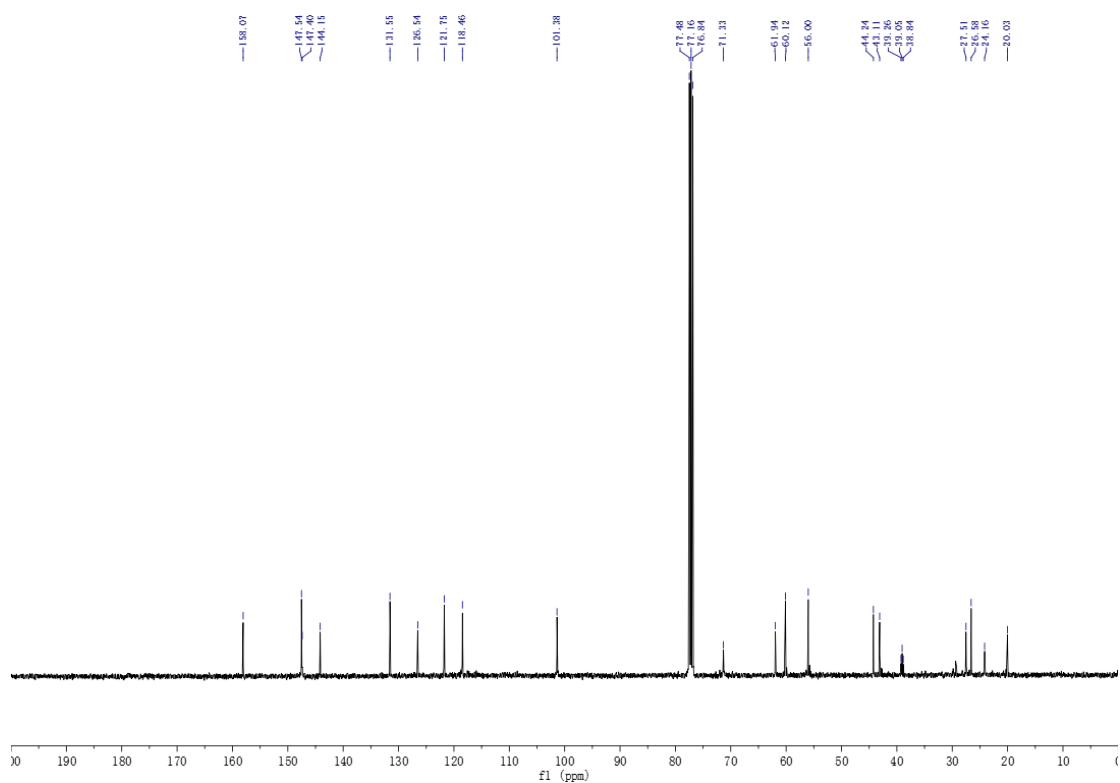
¹³C NMR of compound **23**



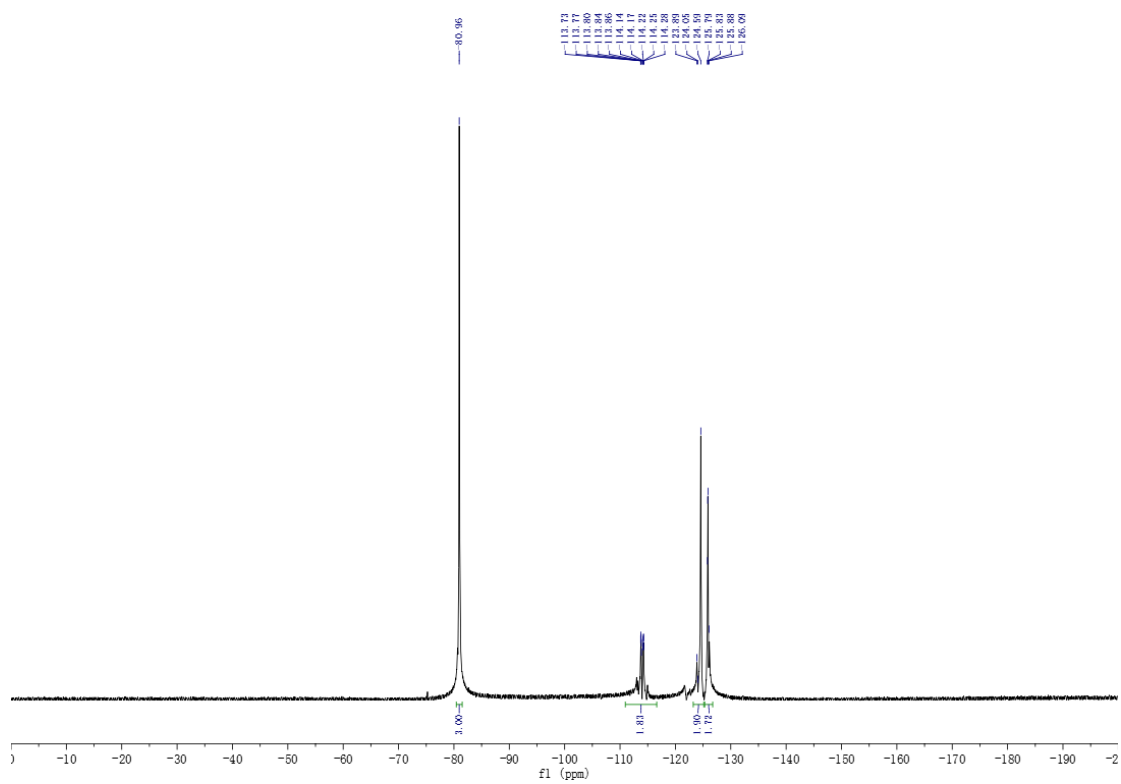
^{19}F NMR of compound **23**



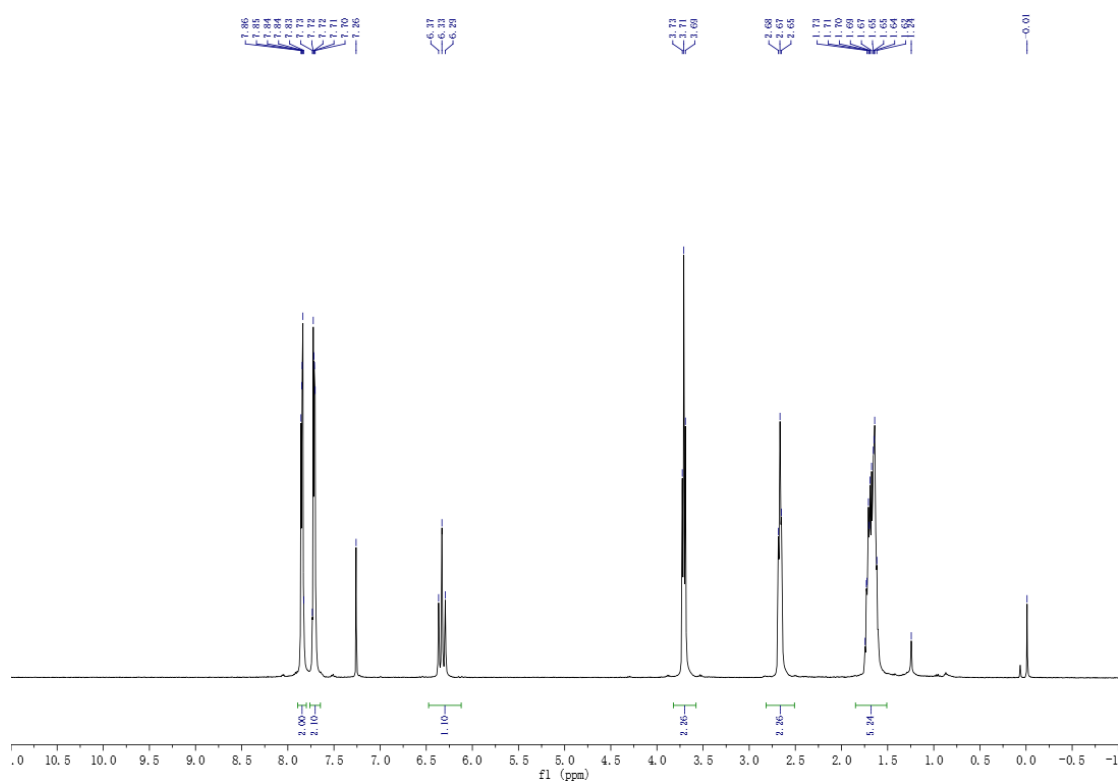
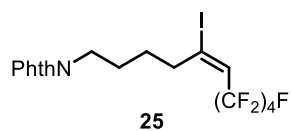
^1H NMR of compound **24**



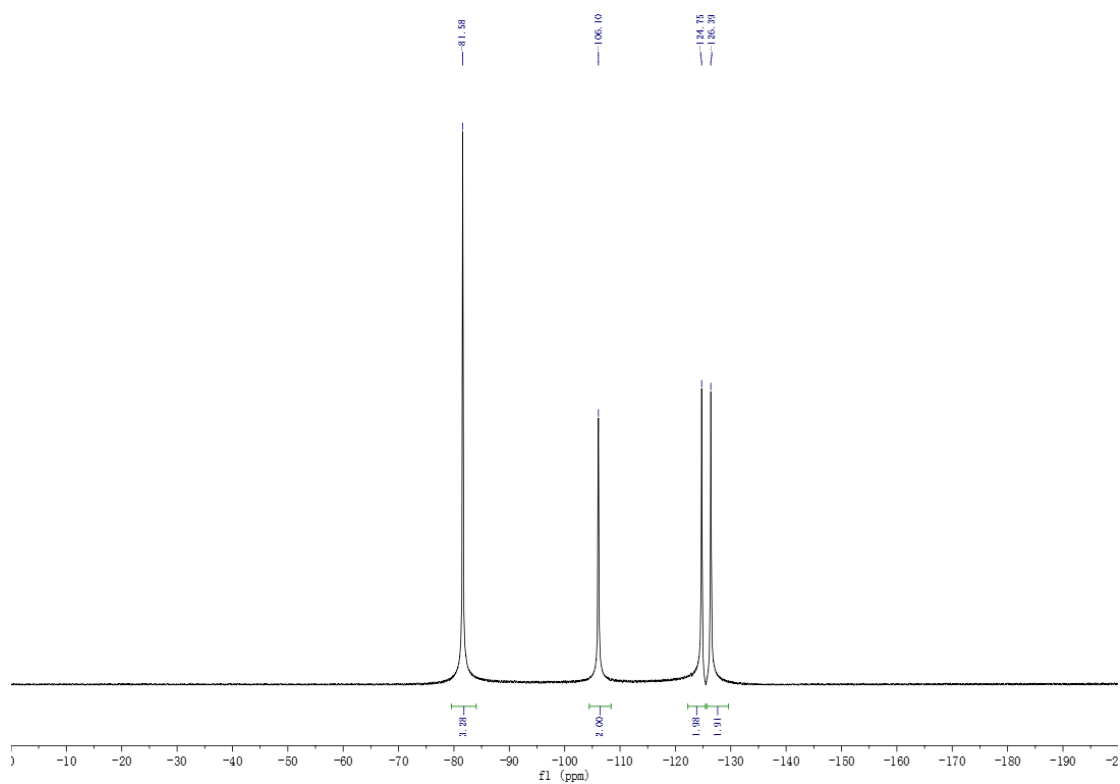
^{13}C NMR of compound **24**



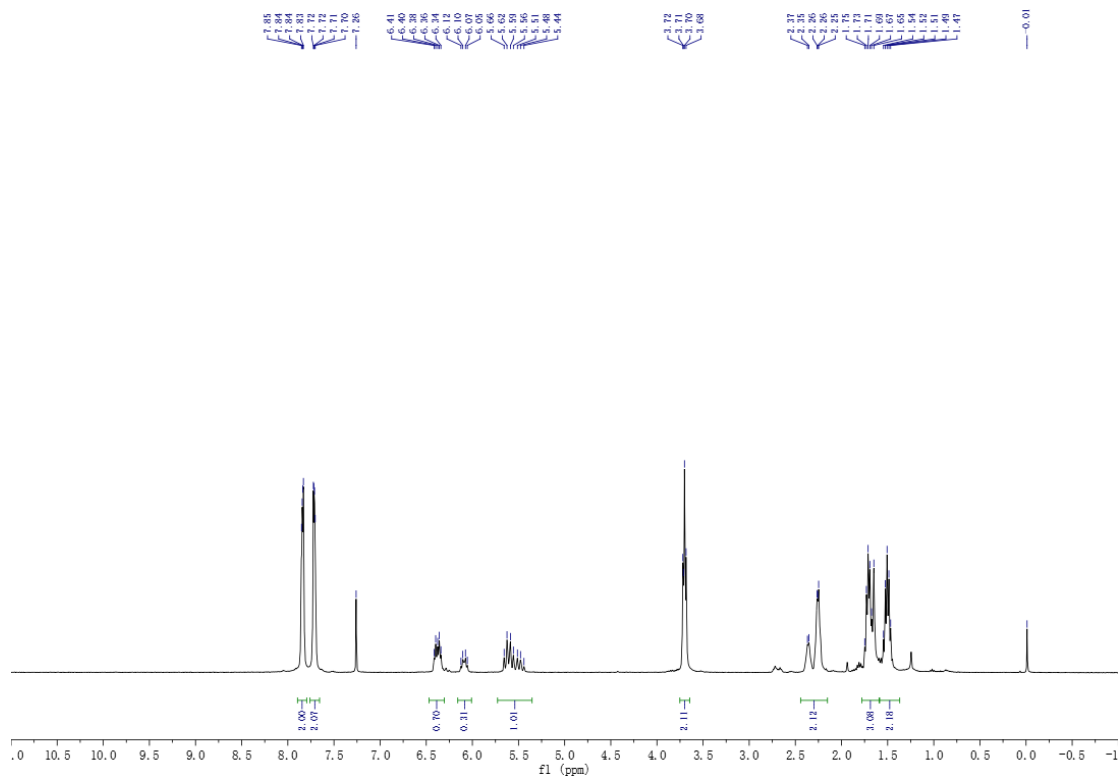
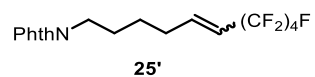
^{19}F NMR of compound **24**



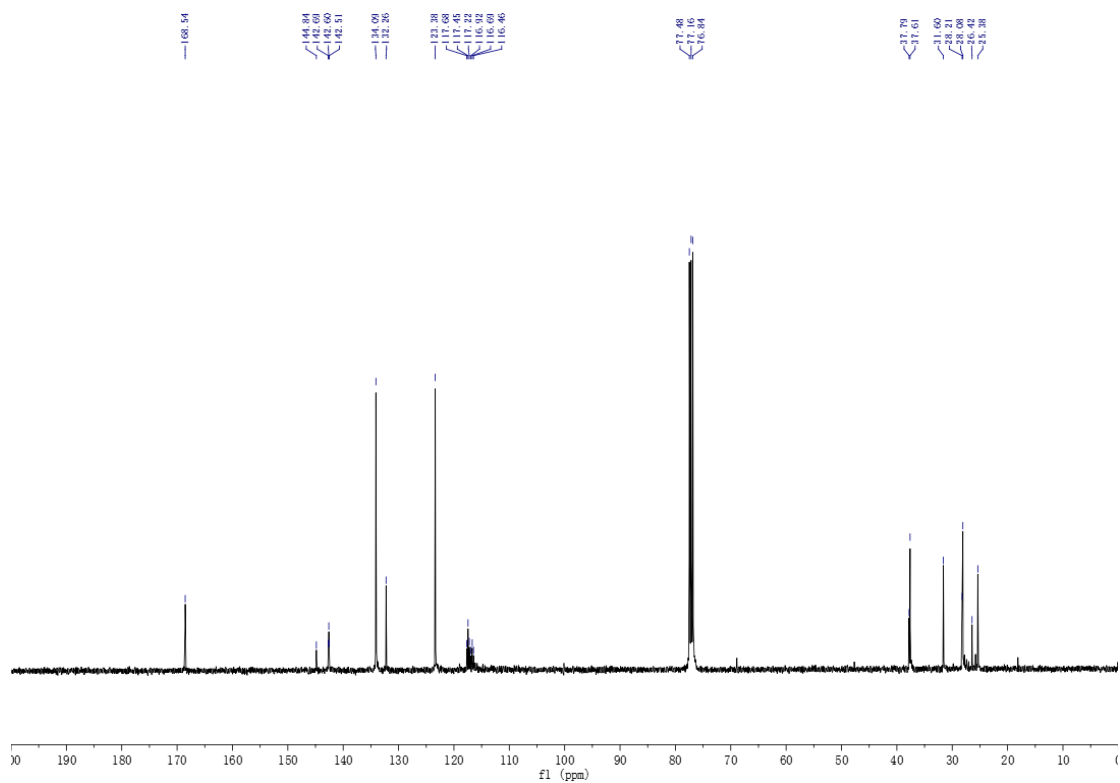
^1H NMR of compound **25**



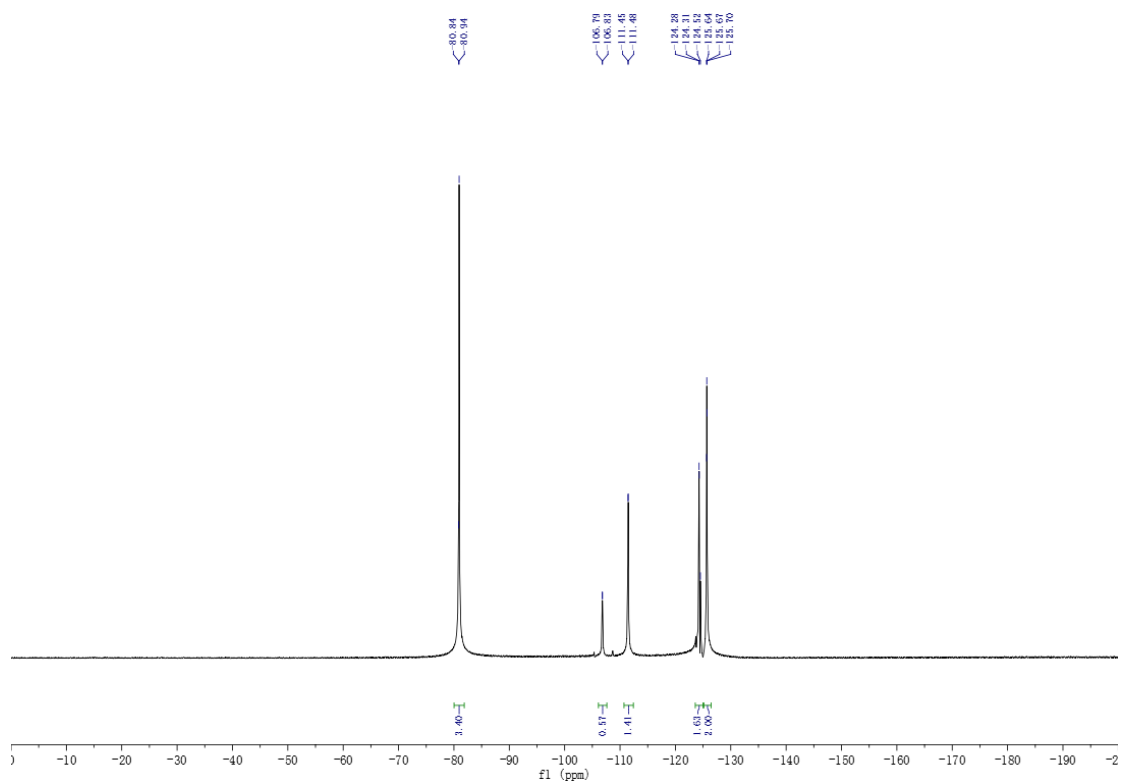
^{19}F NMR of compound **25**



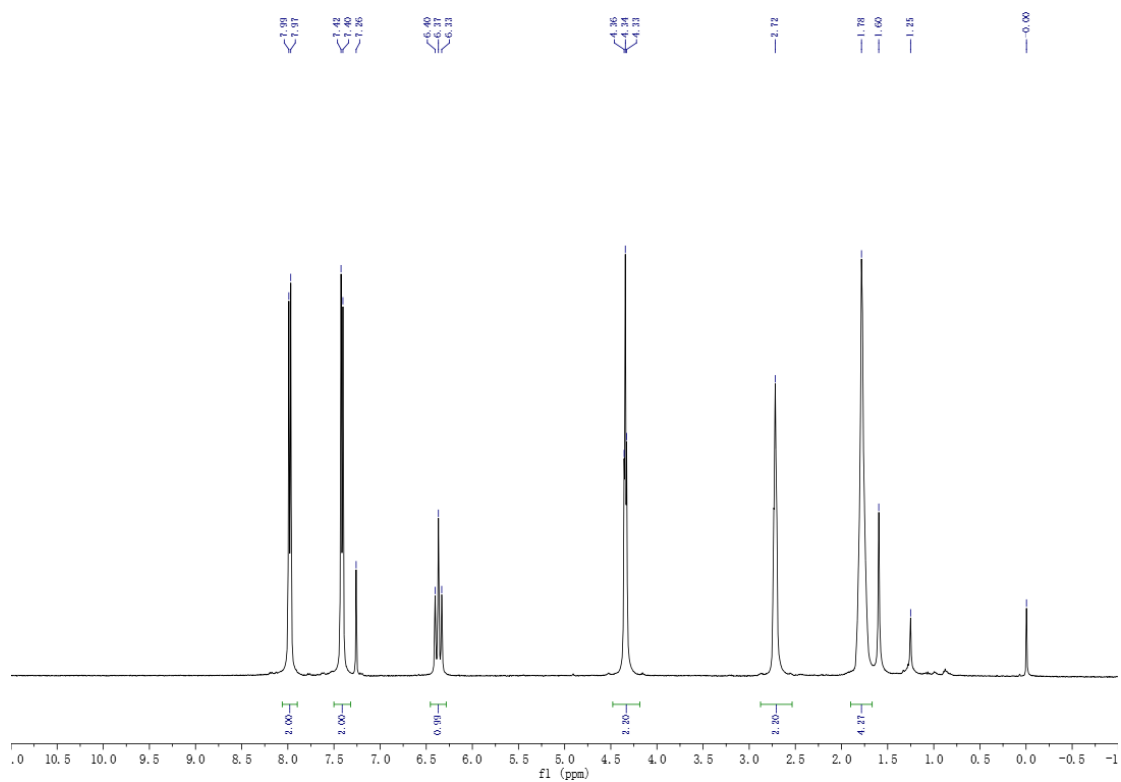
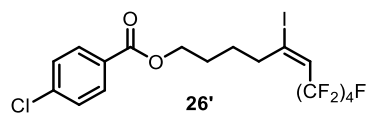
¹H NMR of compound **25'**



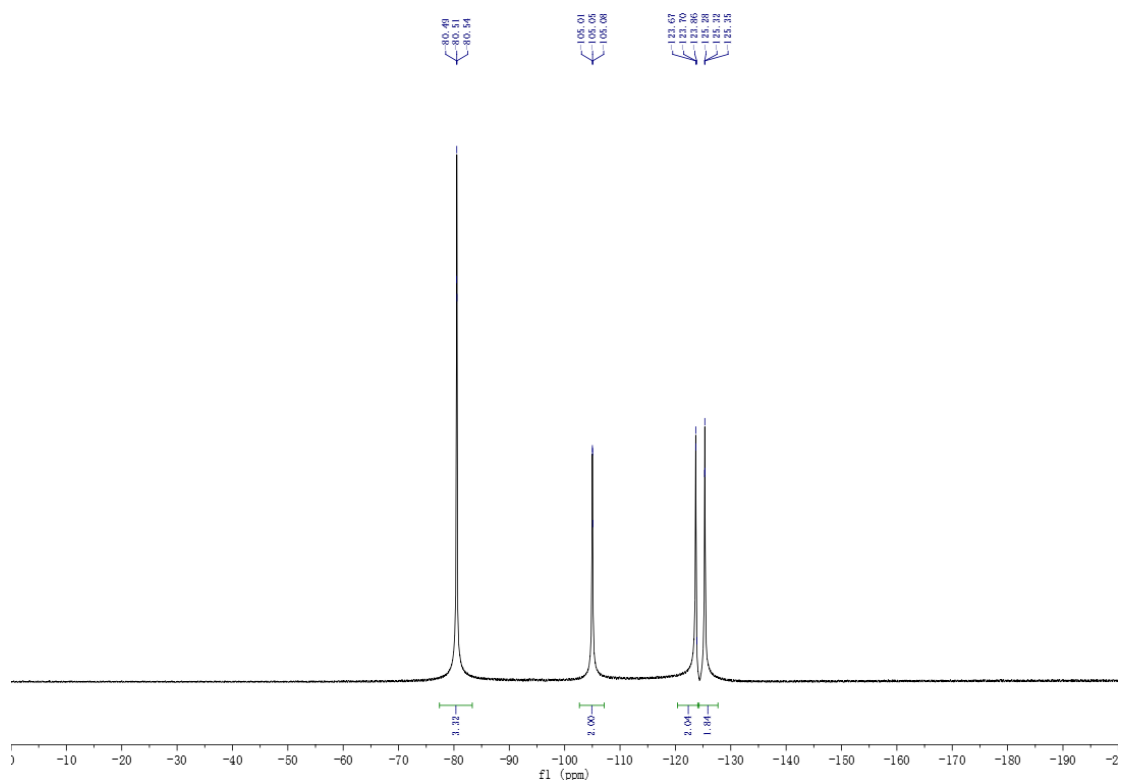
¹³C NMR of compound **25'**



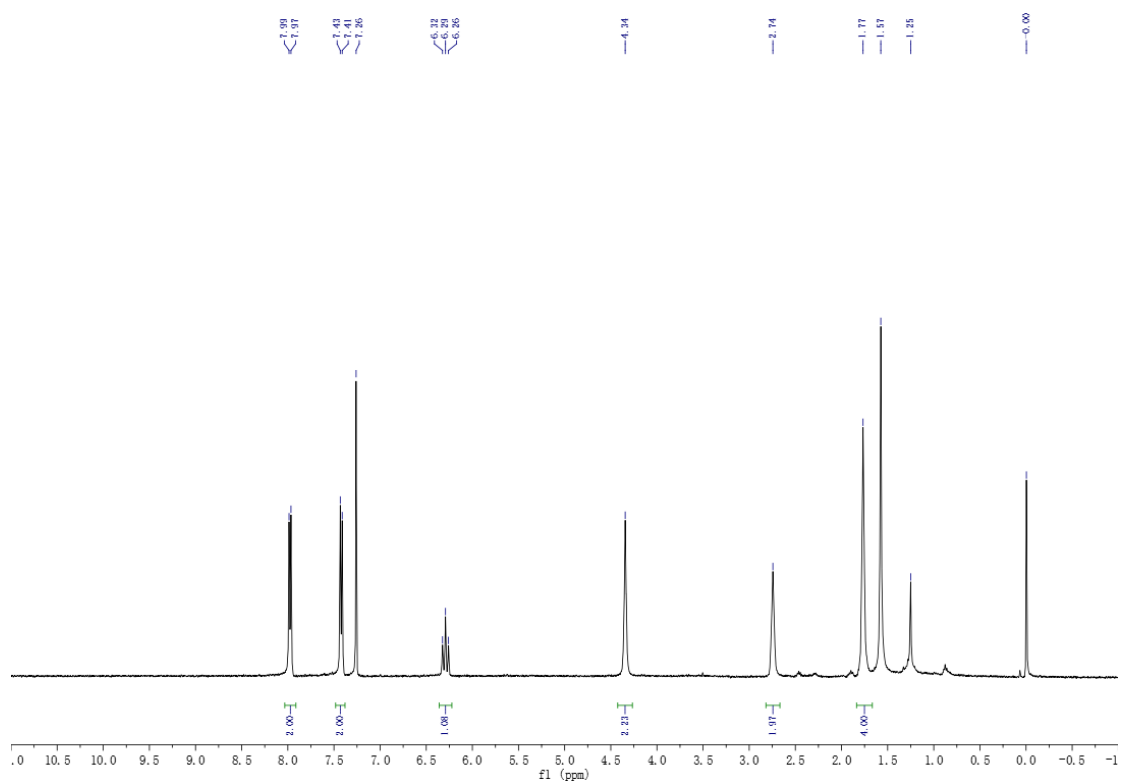
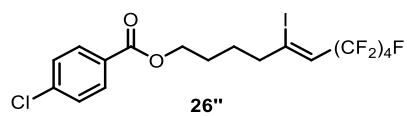
¹⁹F NMR of compound **25'**



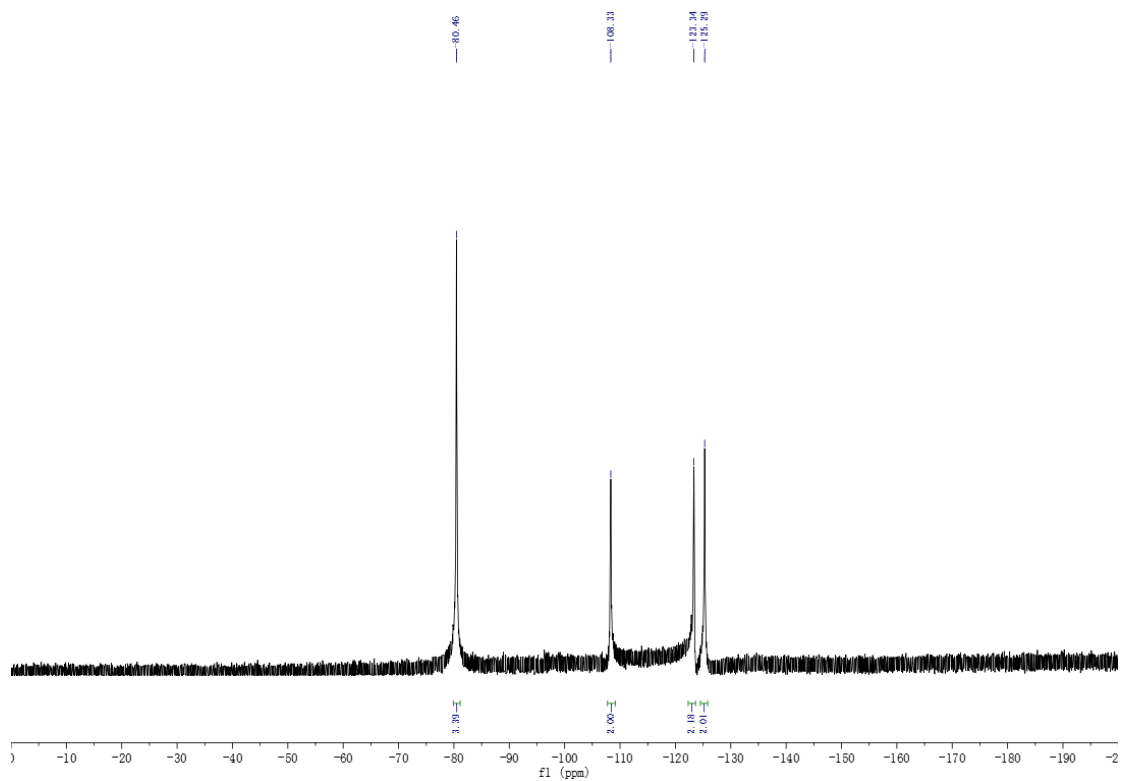
¹H NMR of compound **26'**



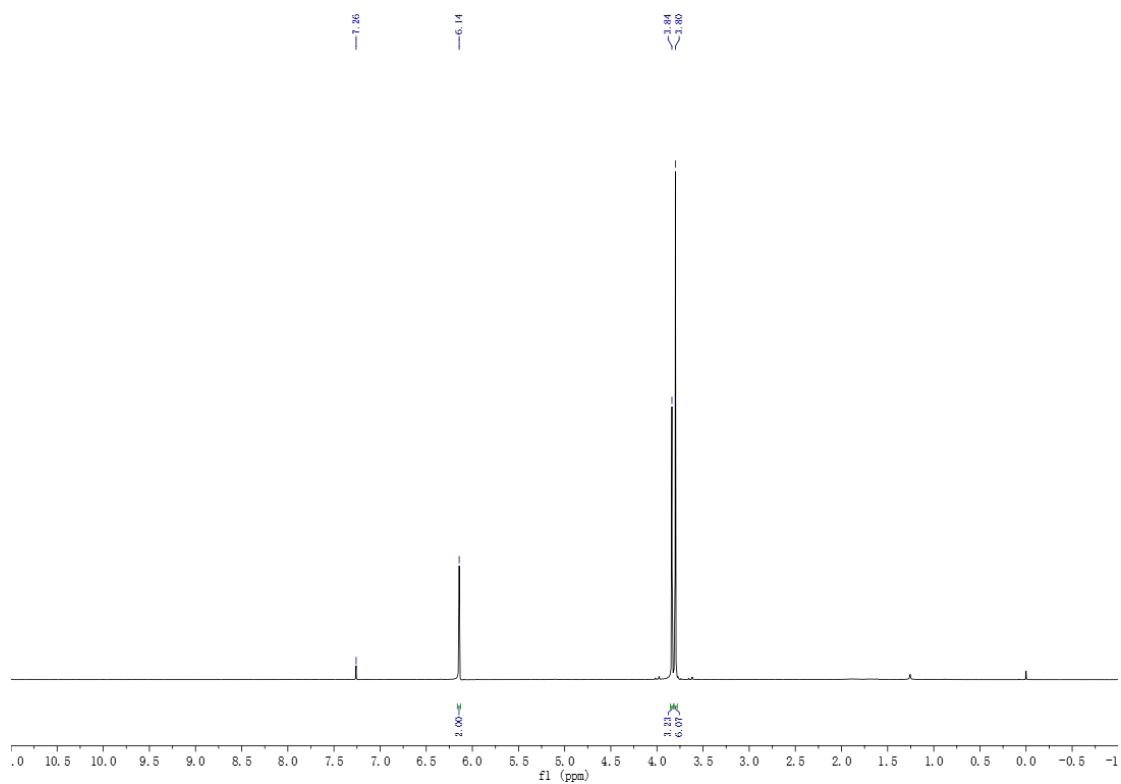
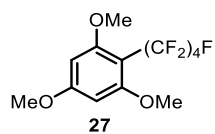
¹⁹F NMR of compound **26'**



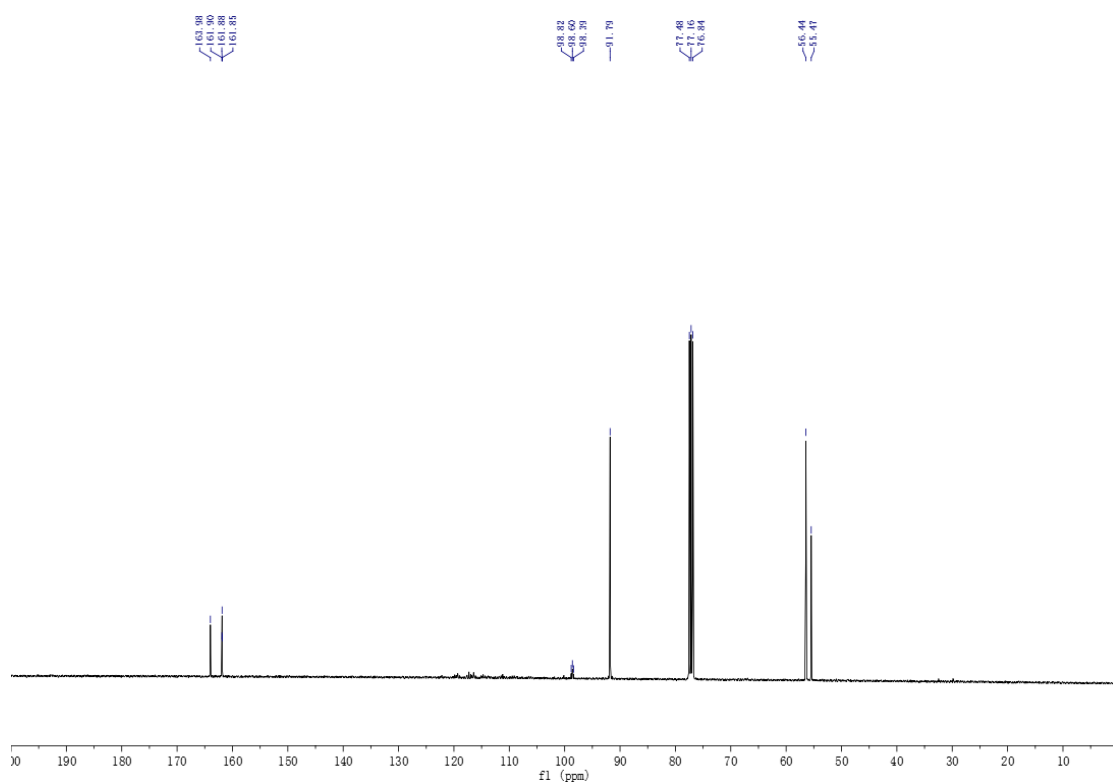
¹H NMR of compound **26''**



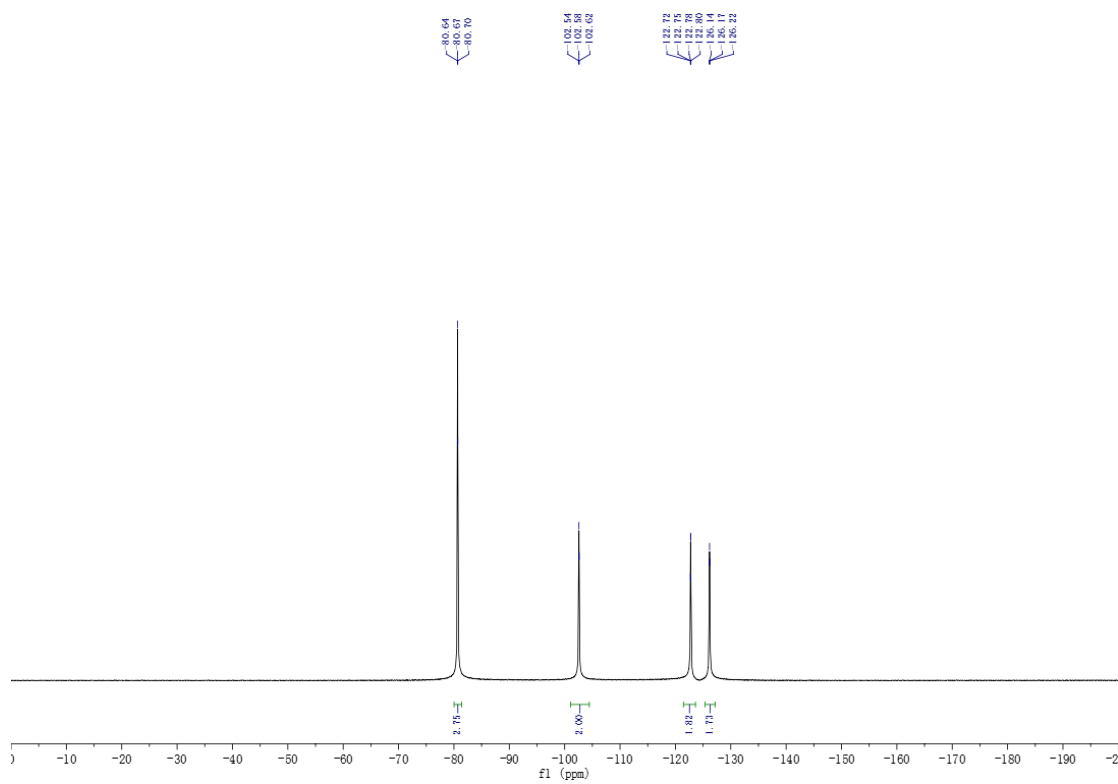
¹⁹F NMR of compound 26



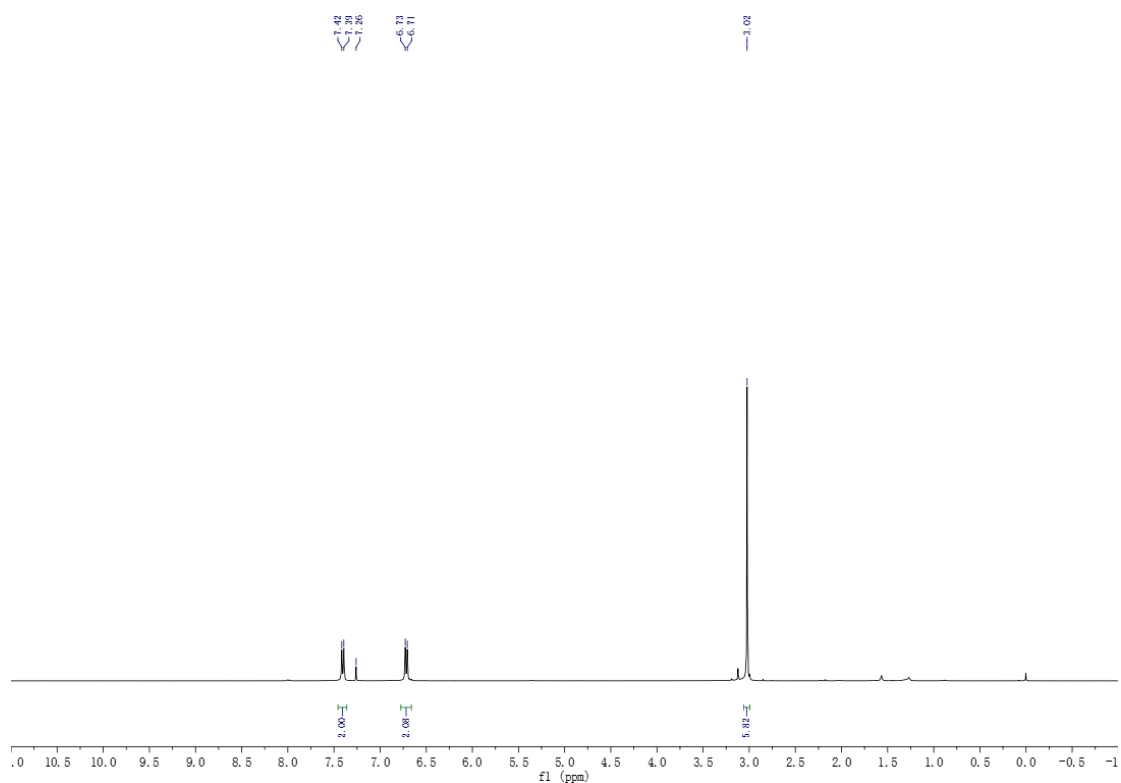
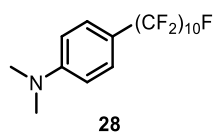
¹H NMR of compound 27



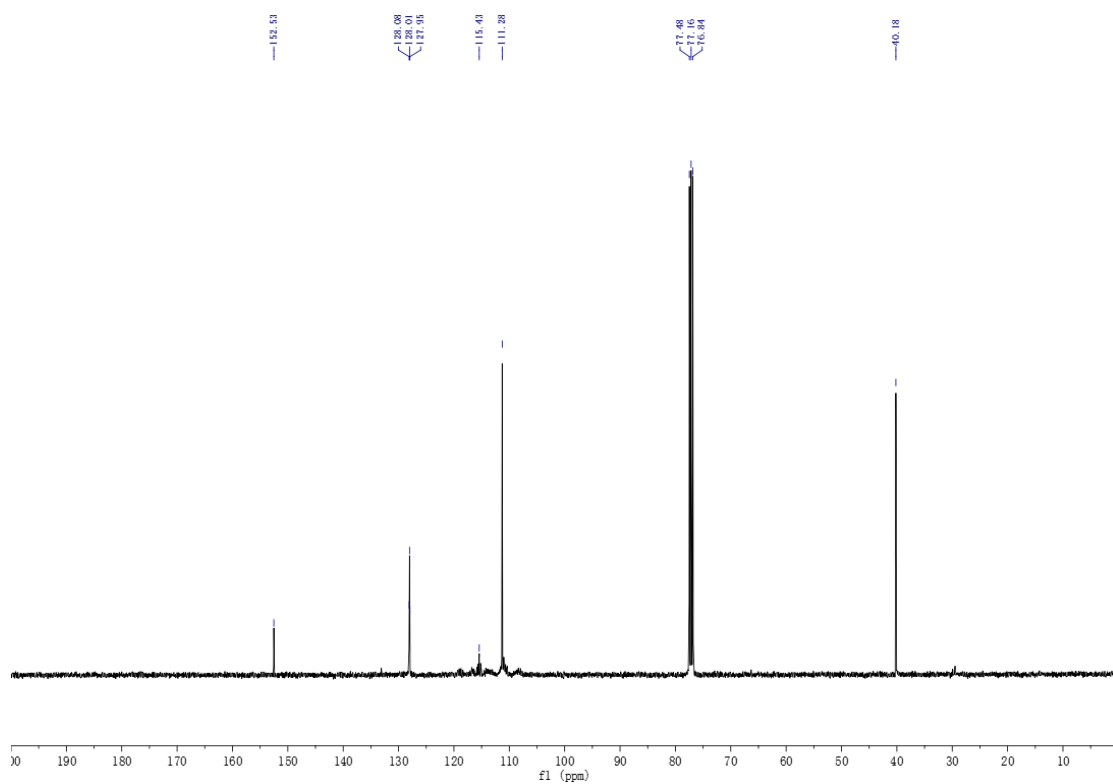
¹³C NMR of compound **27**



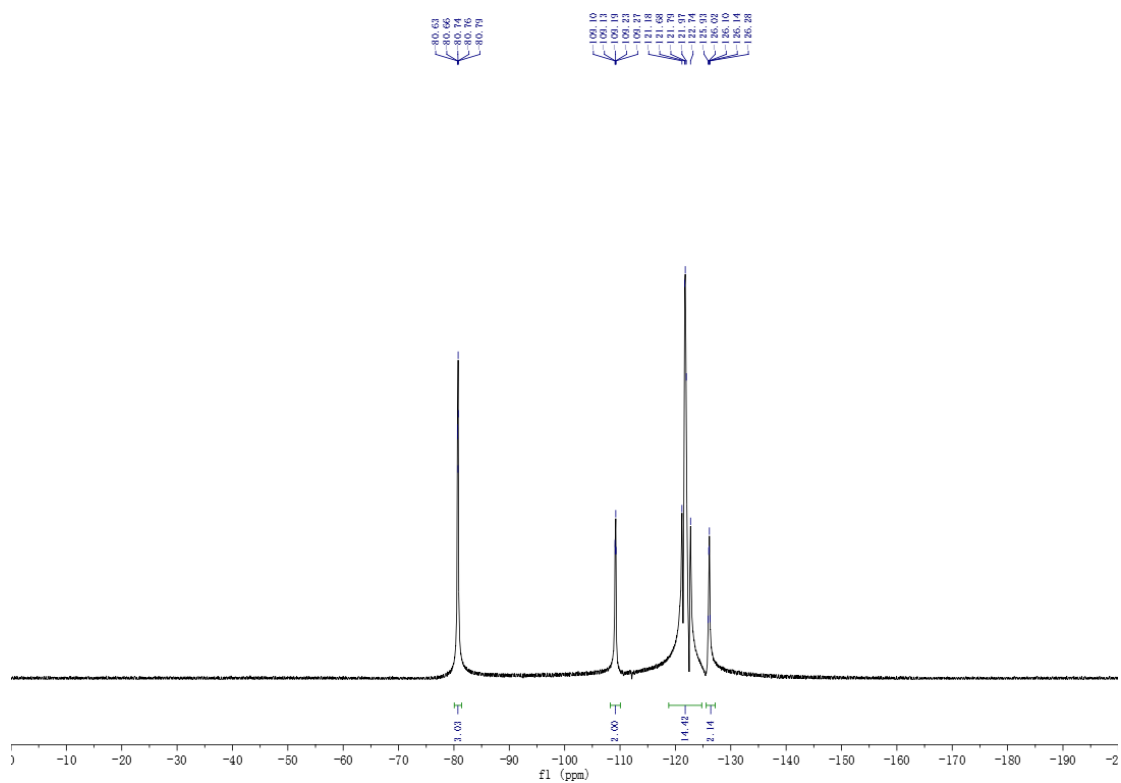
¹⁹F NMR of compound **27**



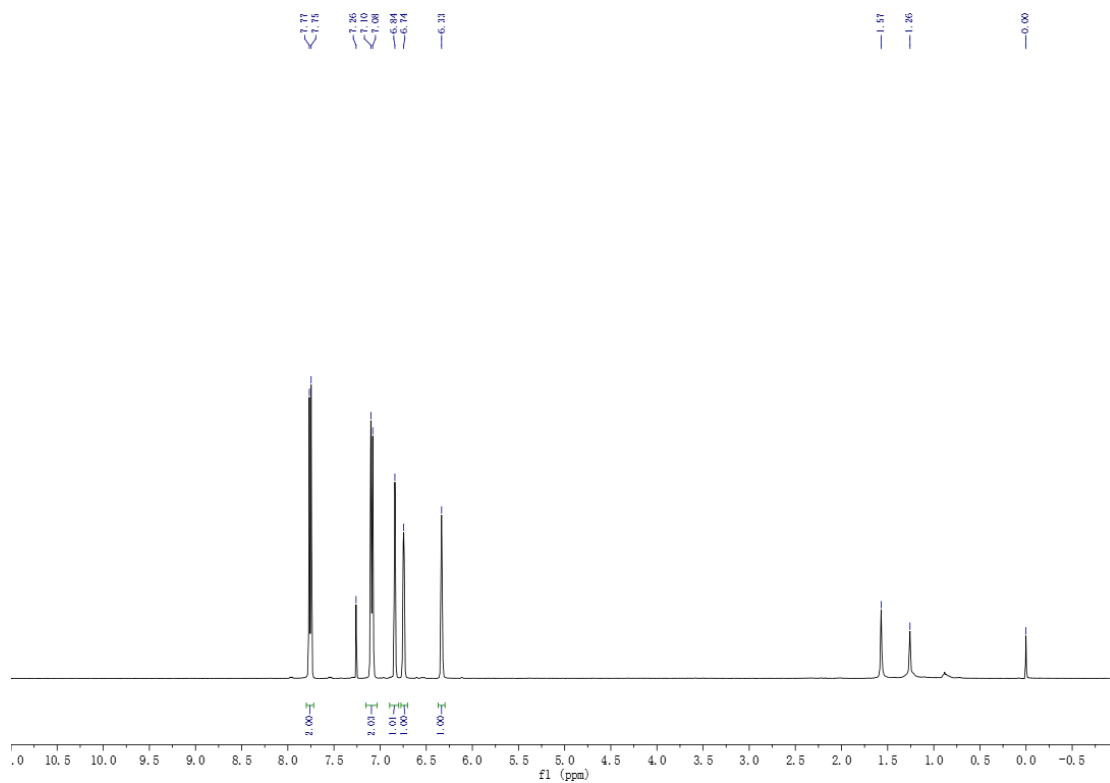
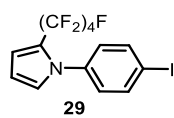
¹H NMR of compound **28**



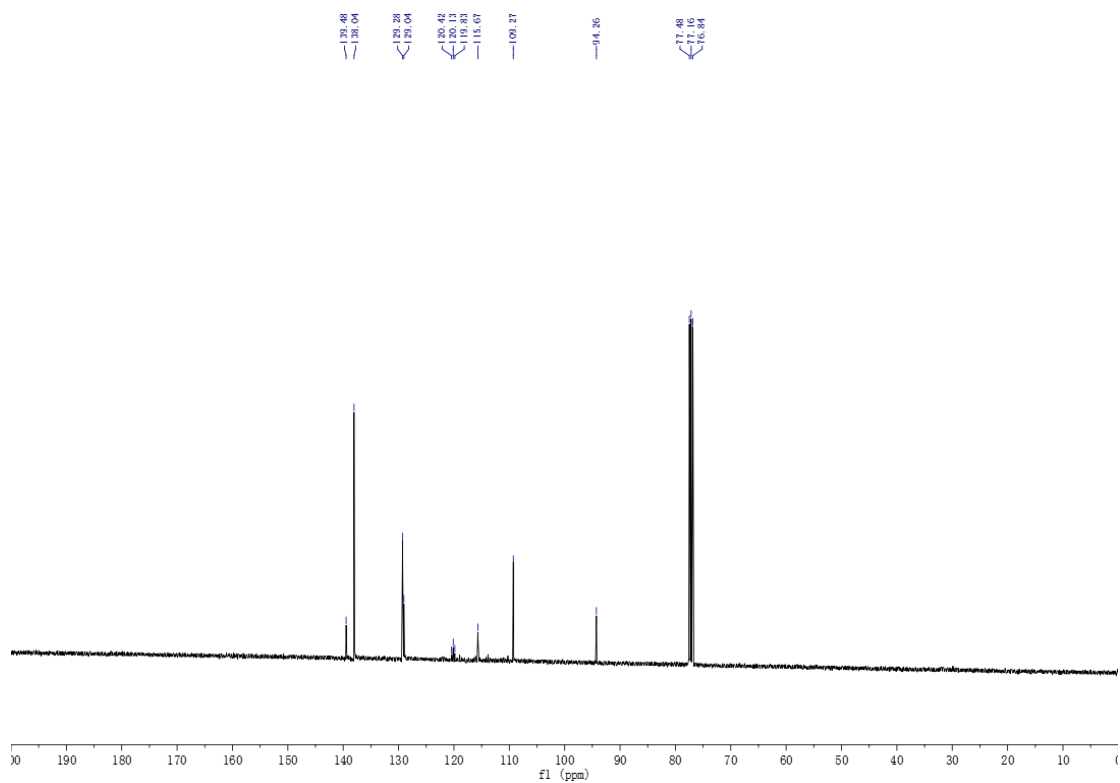
¹³C NMR of compound **28**



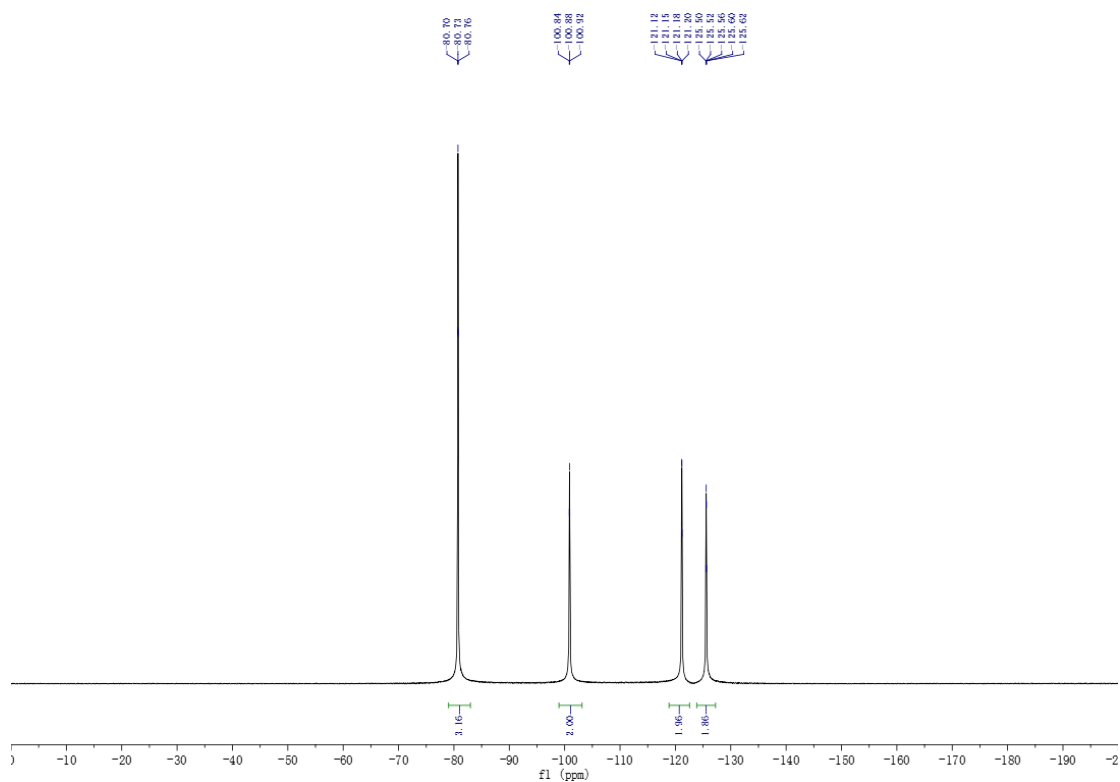
¹⁹F NMR of compound **28**



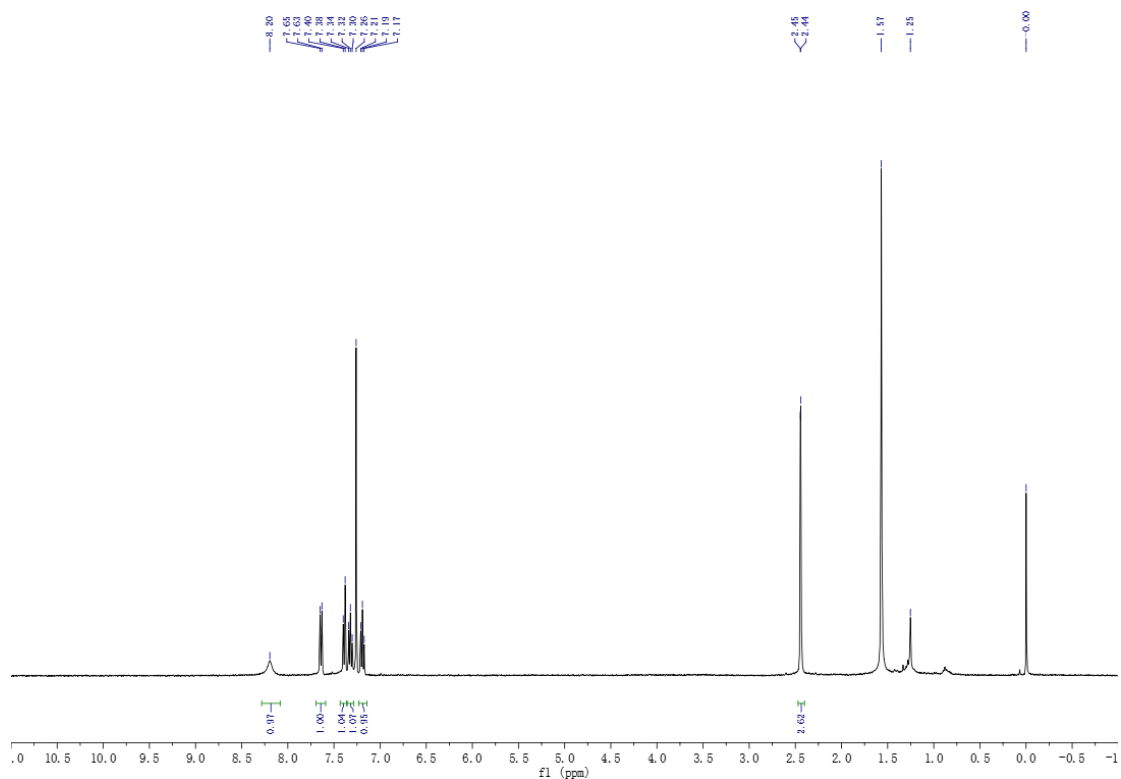
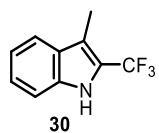
¹H NMR of compound **29**



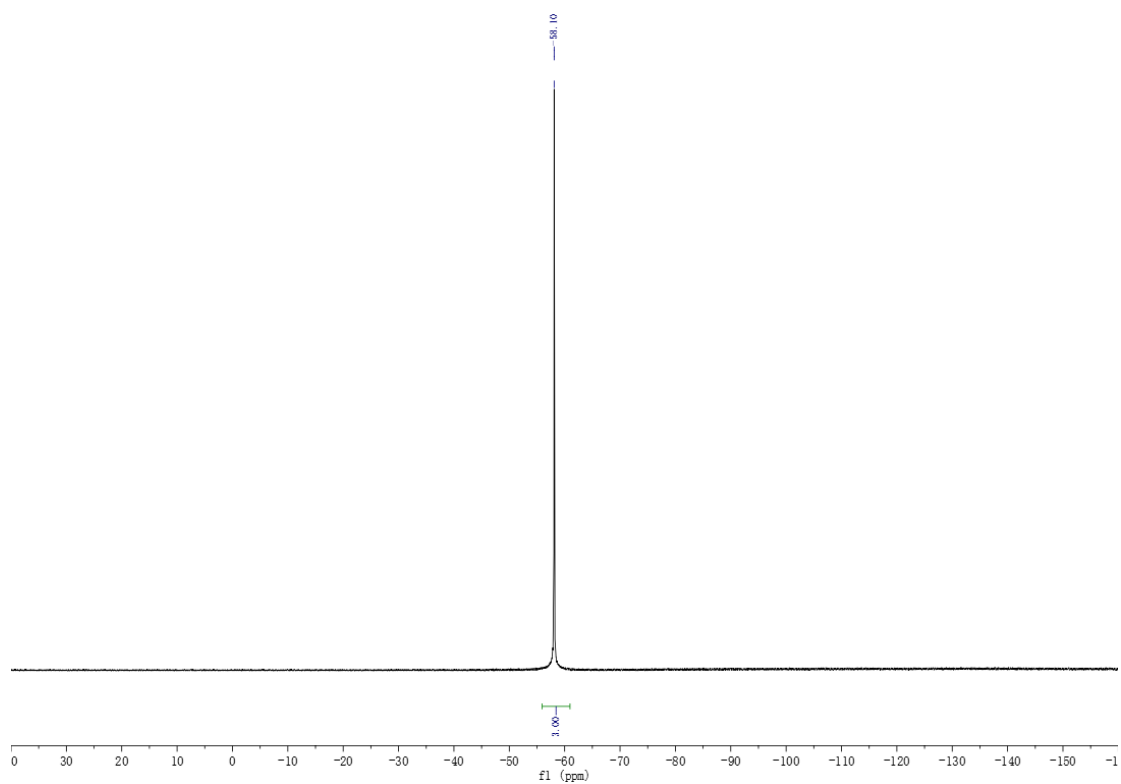
^{13}C NMR of compound **29**



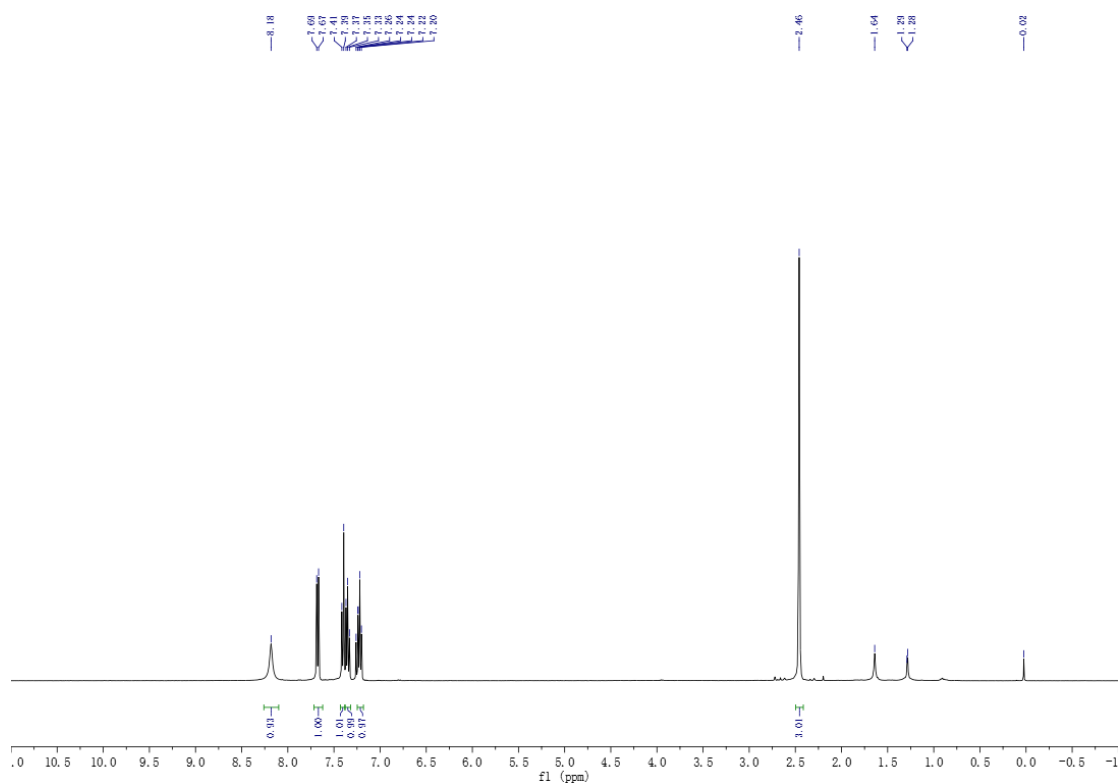
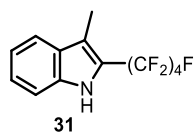
^{19}F NMR of compound **29**



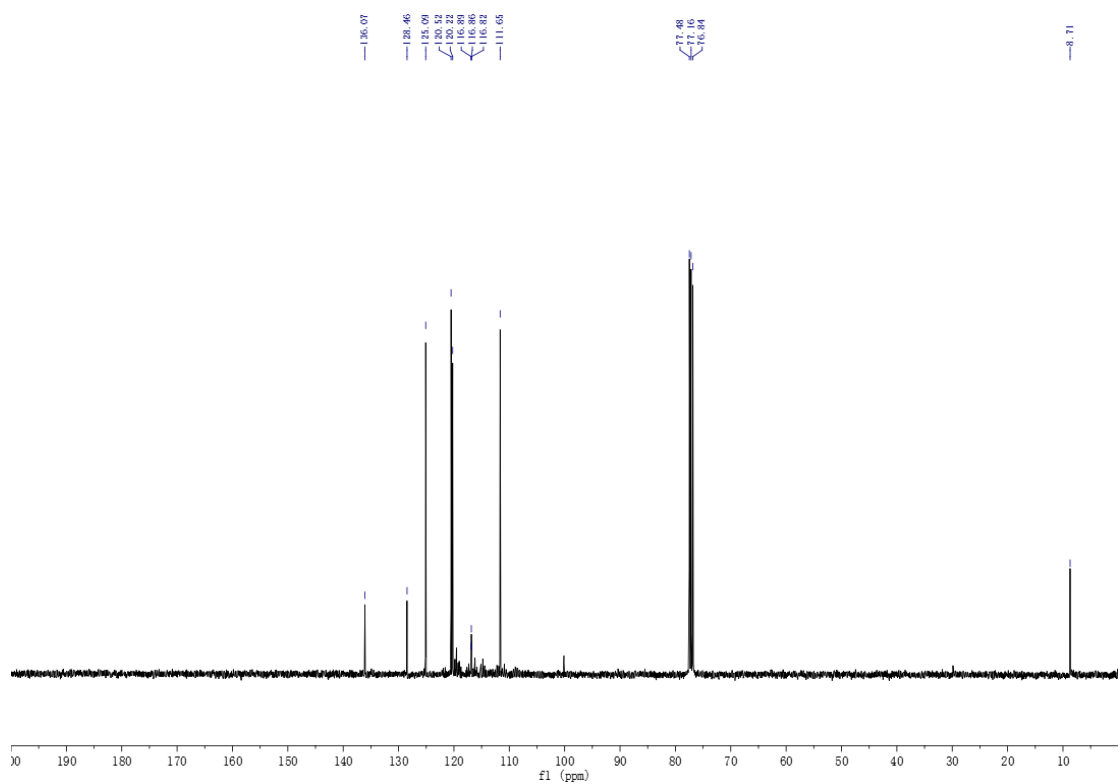
¹H NMR of compound **30**



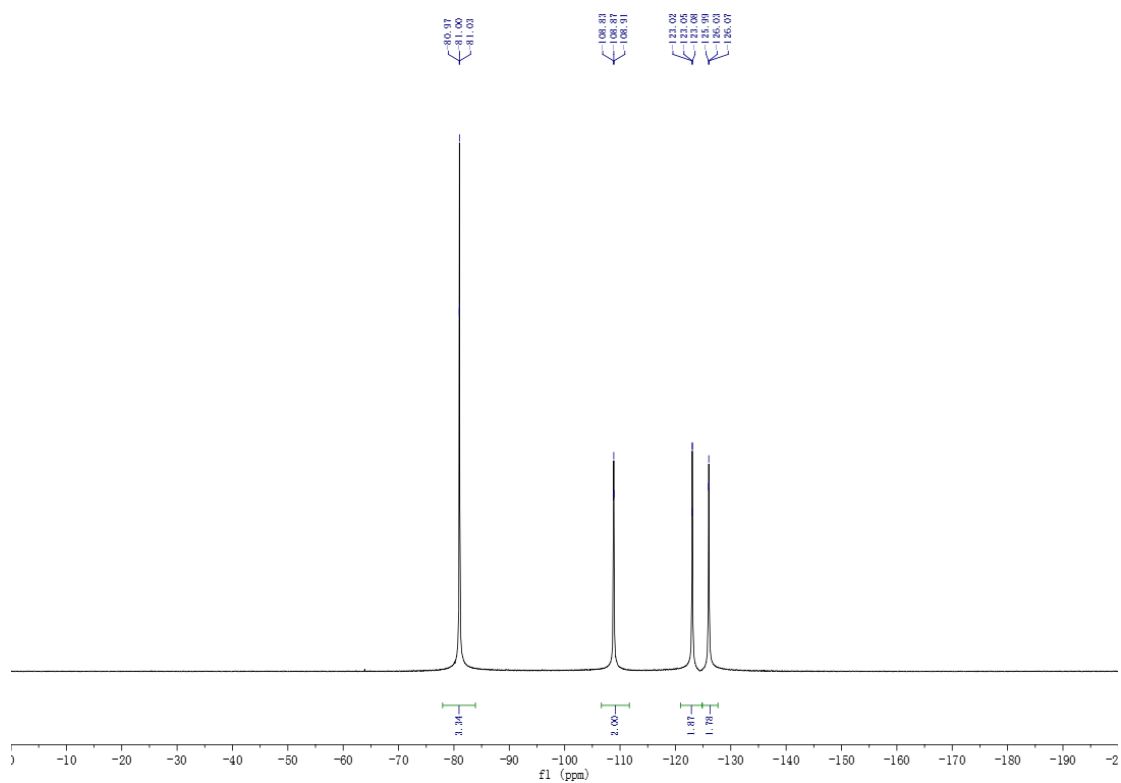
¹⁹F NMR of compound **30**



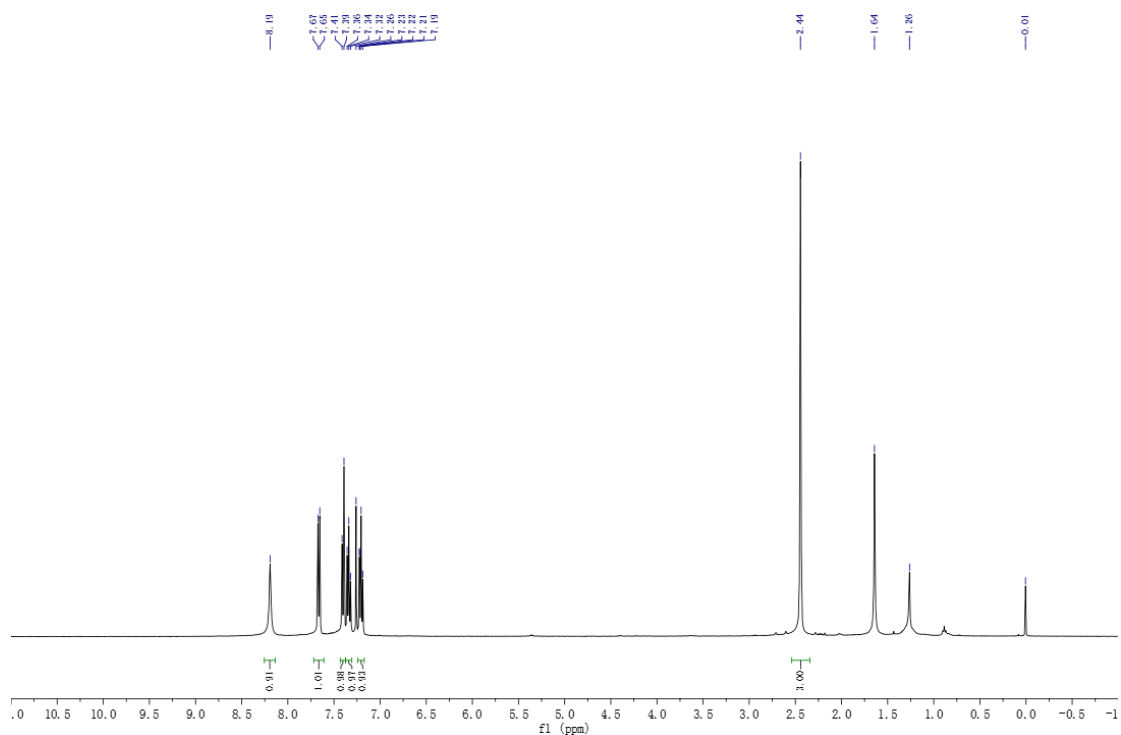
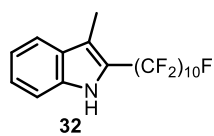
¹H NMR of compound **31**



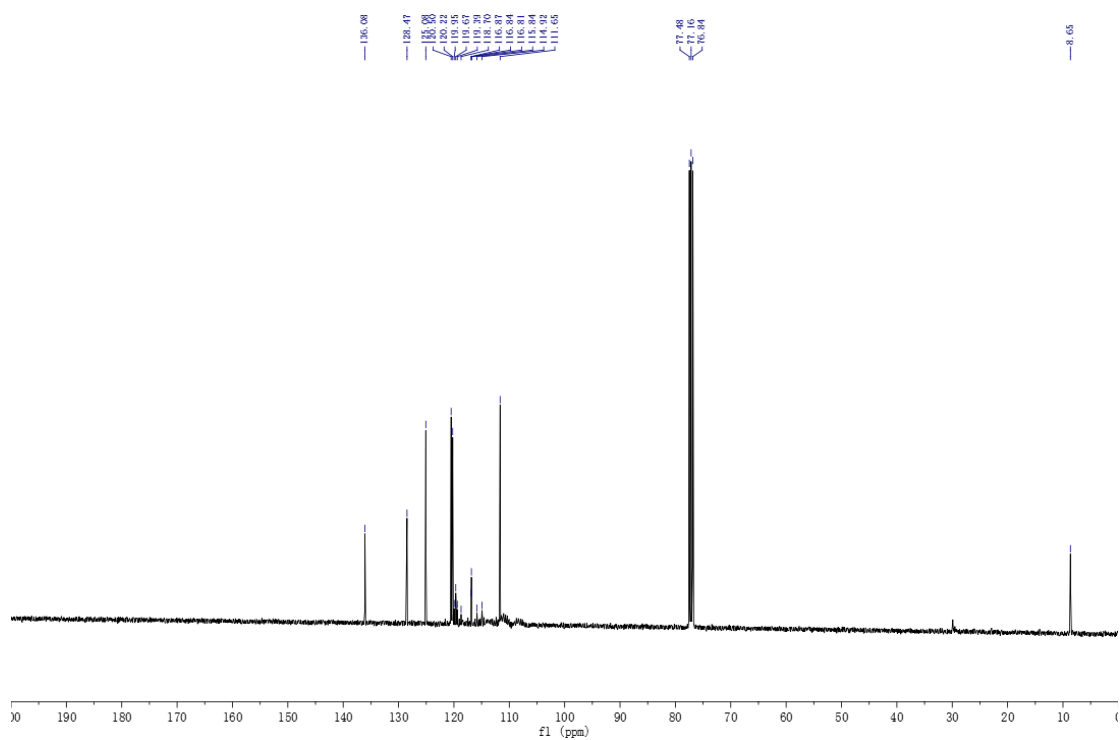
¹³C NMR of compound **31**



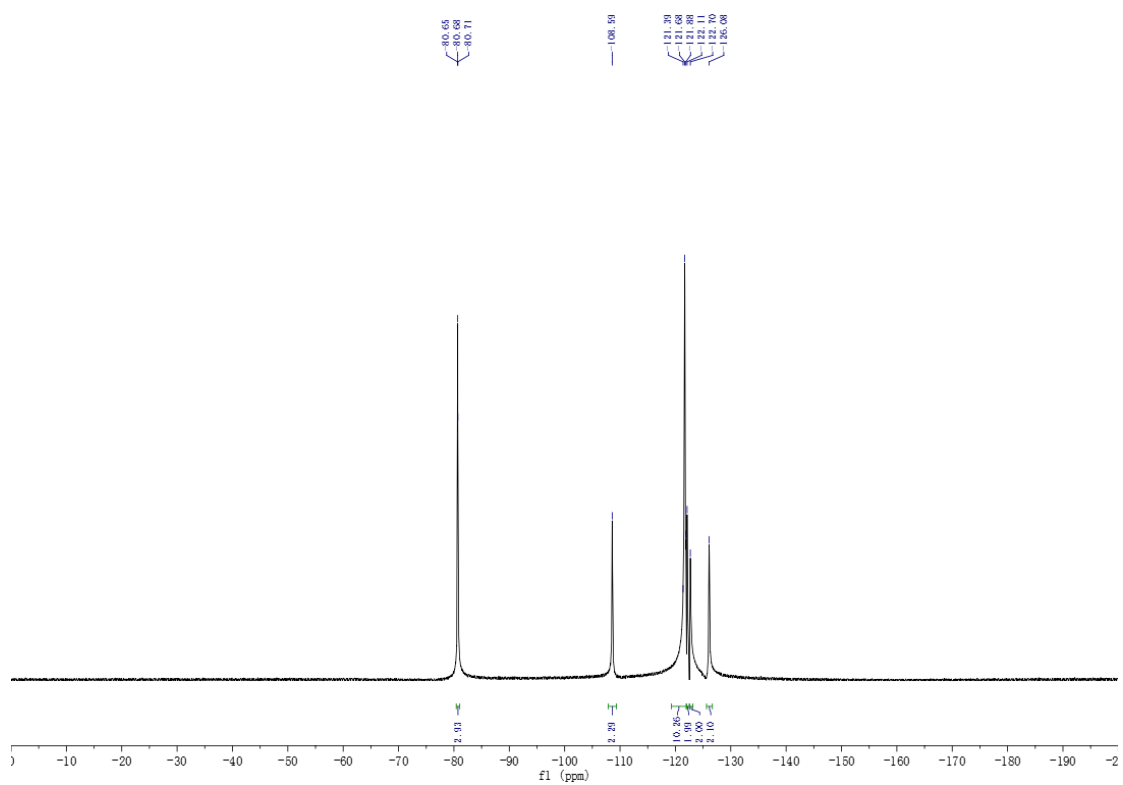
¹⁹F NMR of compound **31**



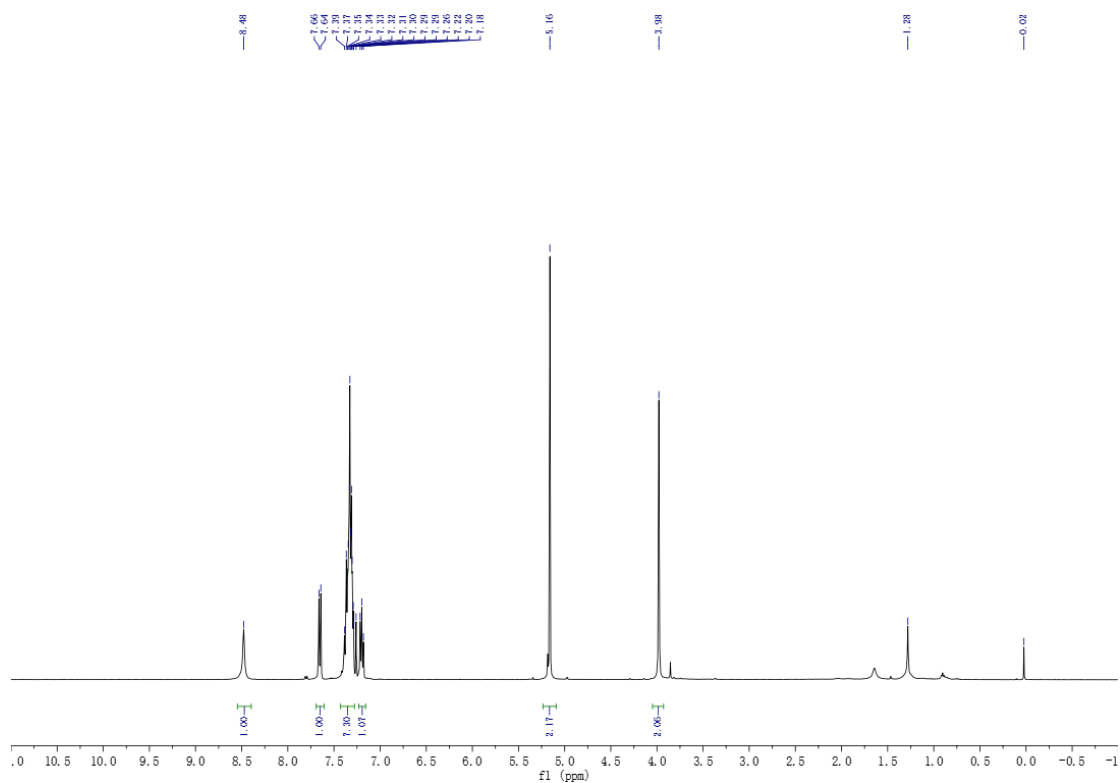
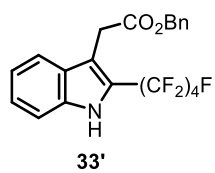
¹H NMR of compound **32**



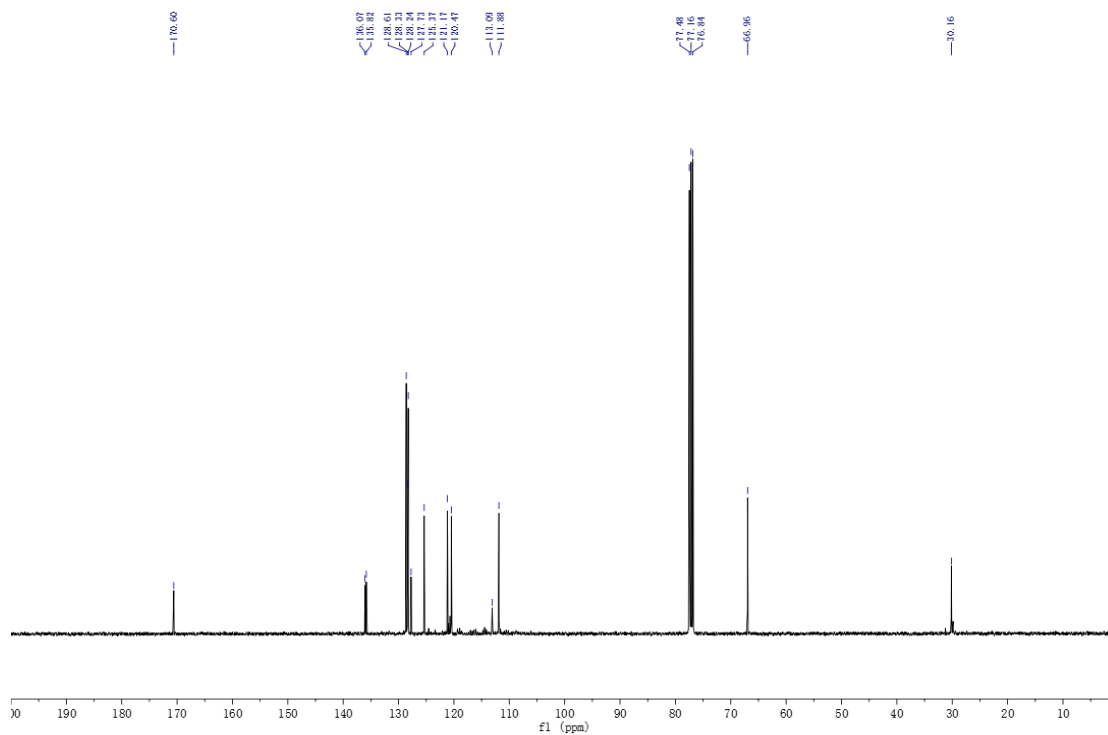
¹³C NMR of compound **32**



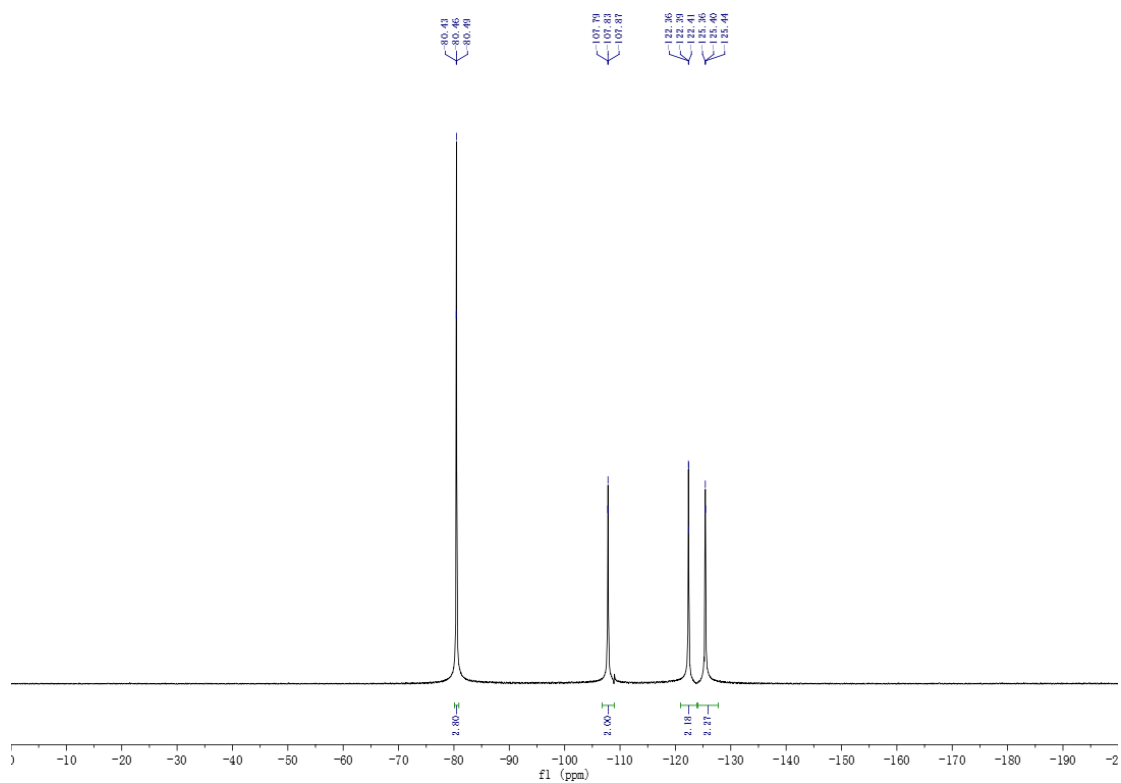
¹⁹F NMR of compound **32**



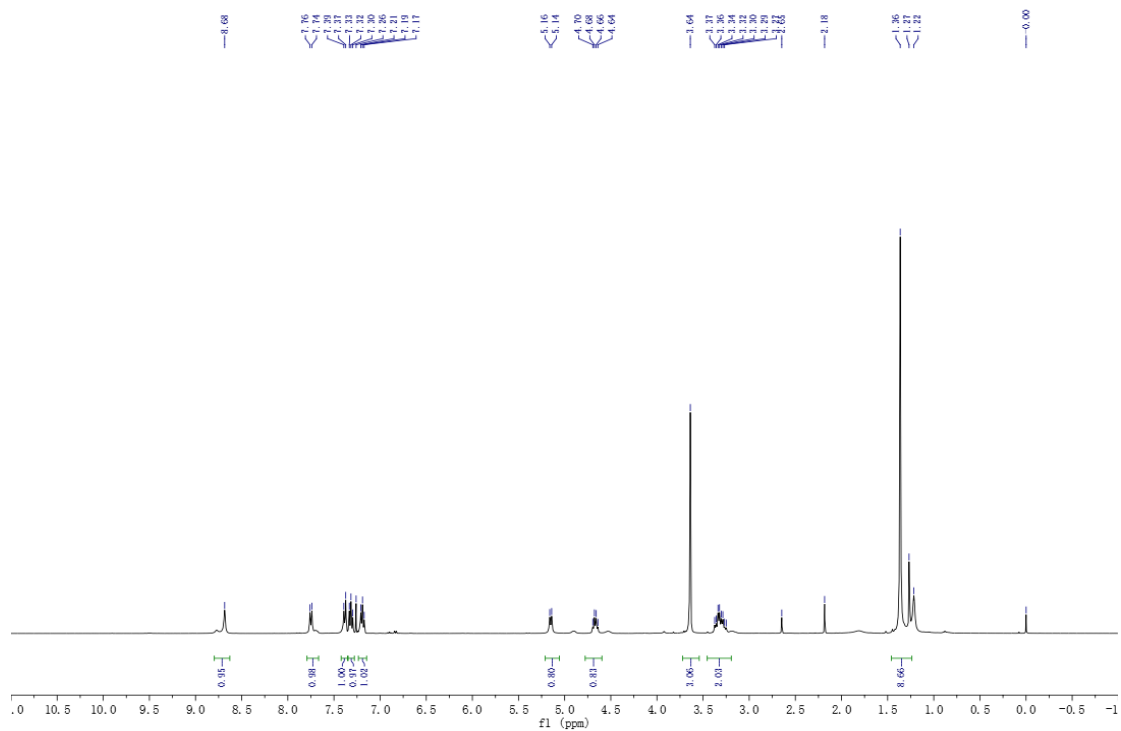
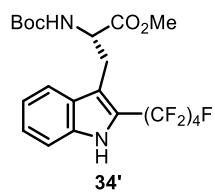
¹H NMR of compound **33'**



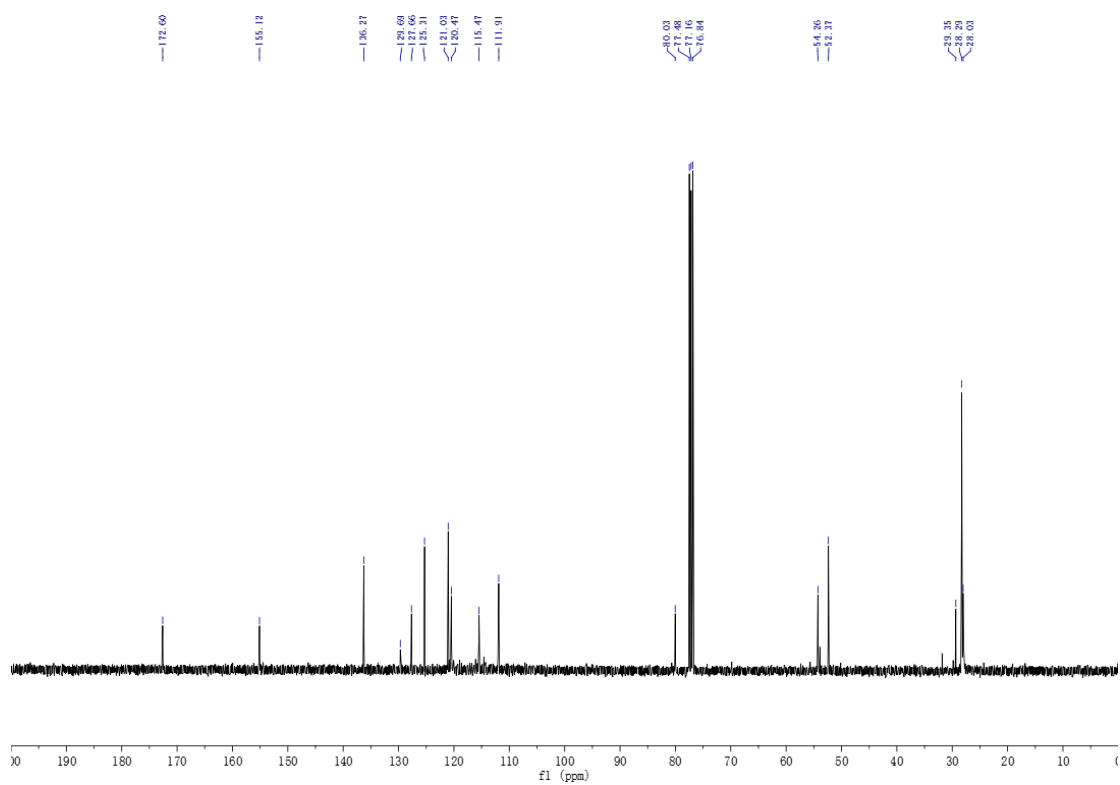
¹³C NMR of compound **33'**



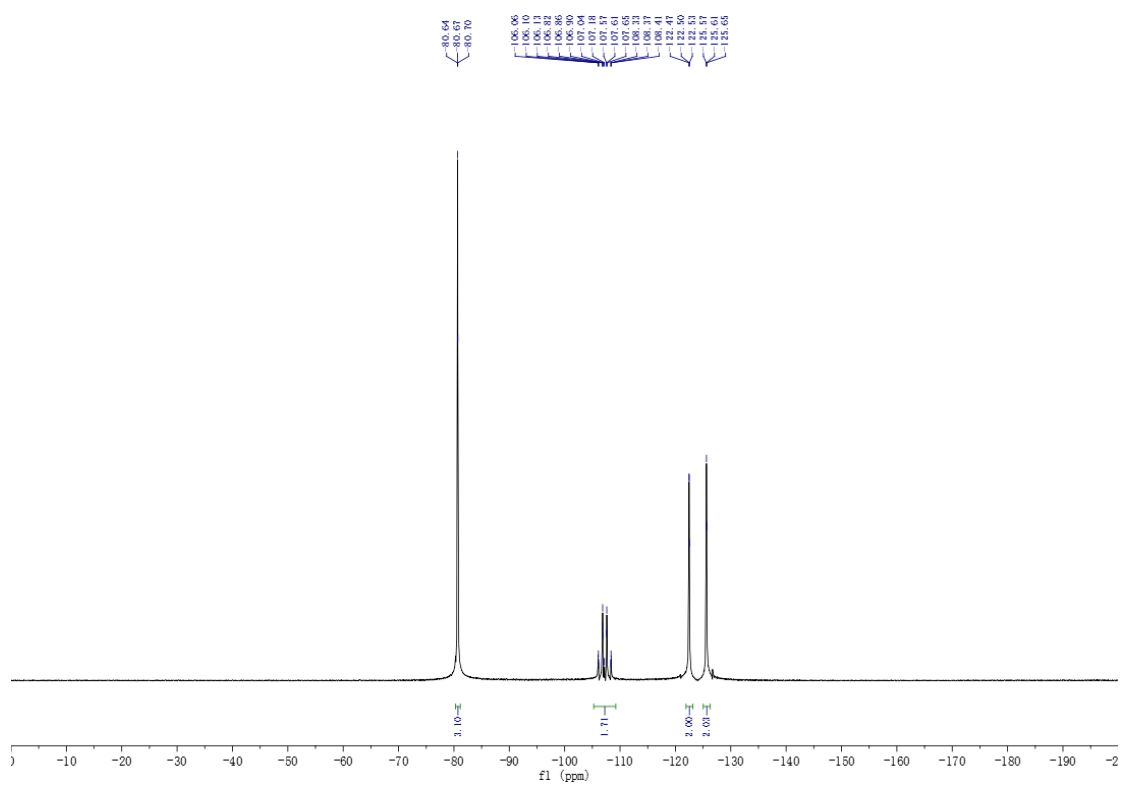
¹⁹F NMR of compound **33'**



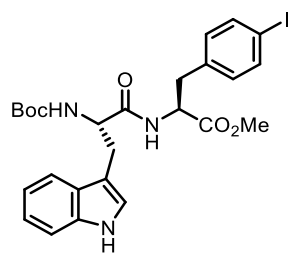
¹H NMR of compound **34'**



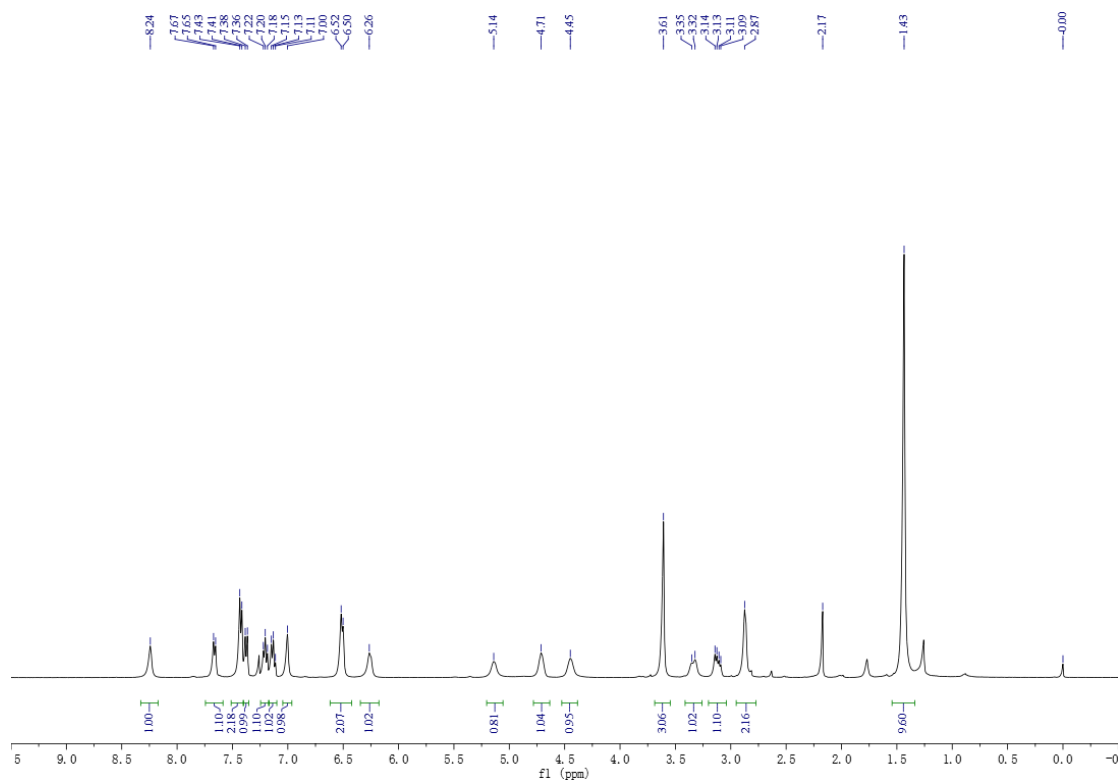
^{13}C NMR of compound **34'**



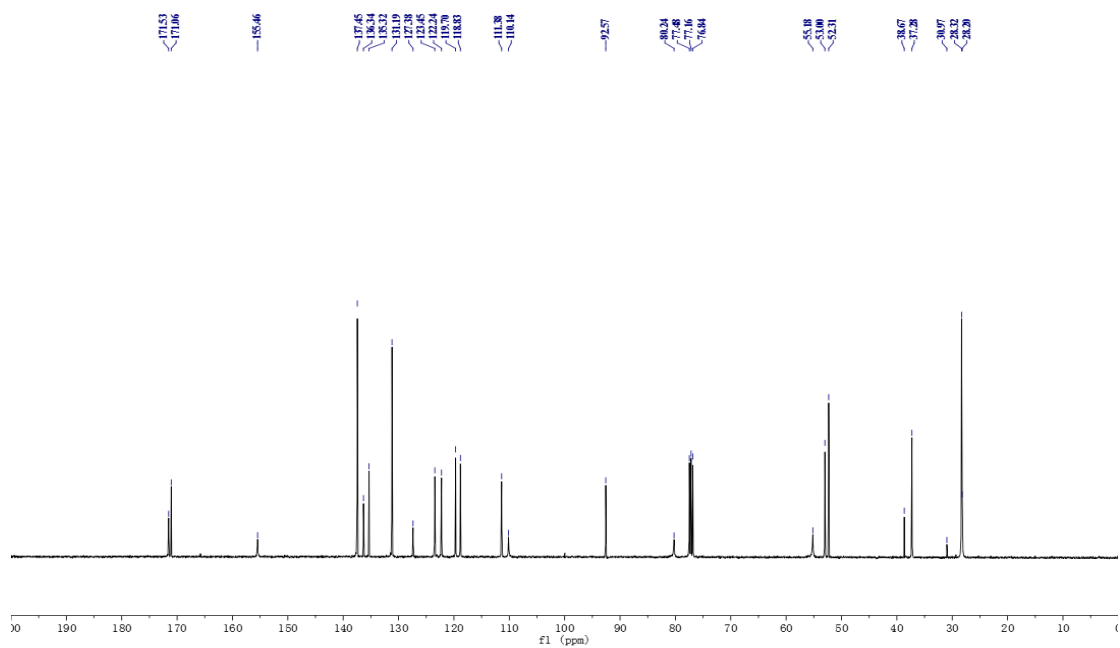
^{19}F NMR of compound **34'**



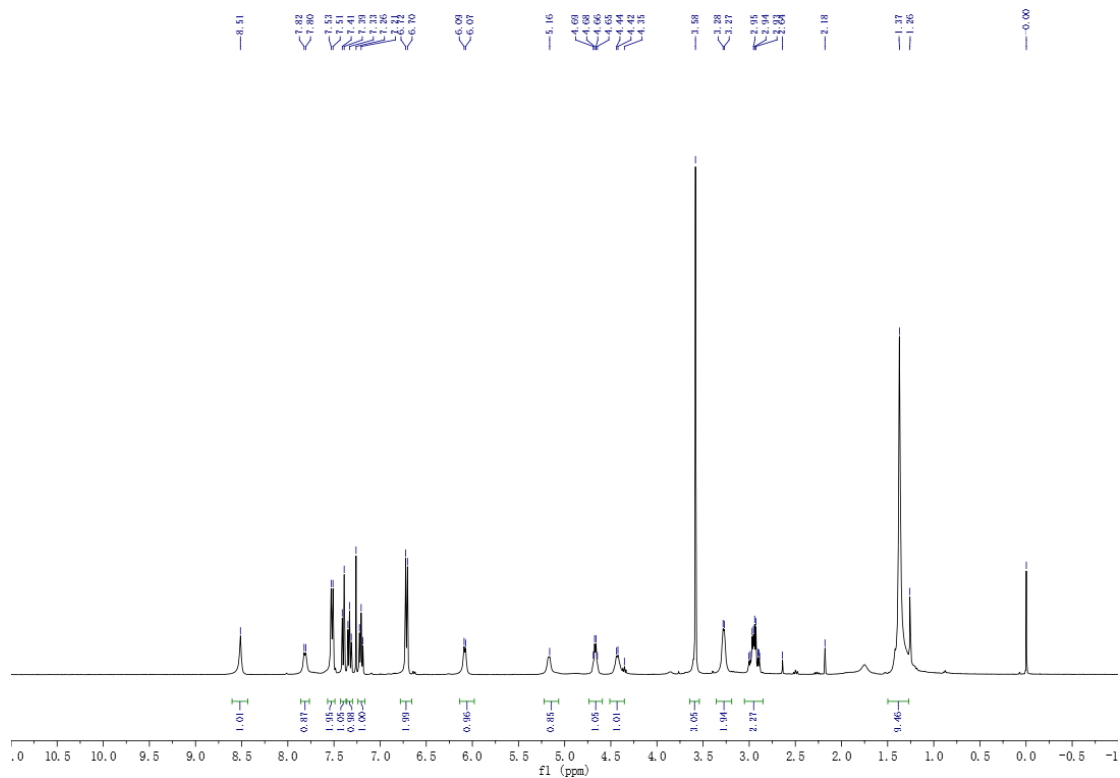
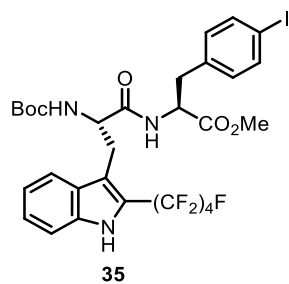
35-1



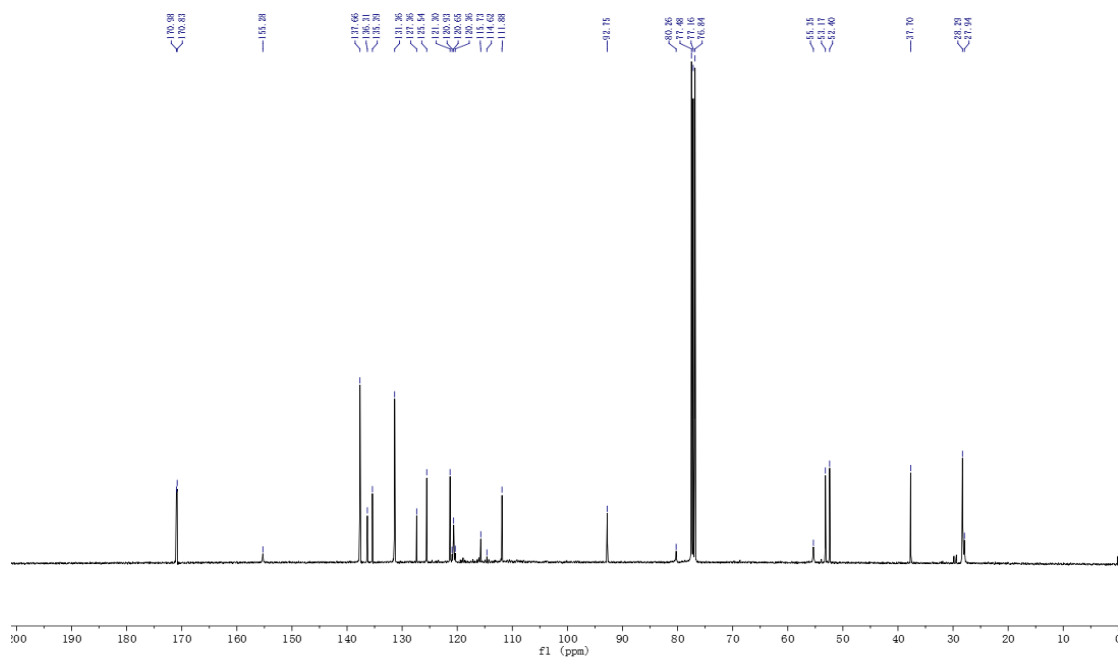
¹H NMR of compound 35-1



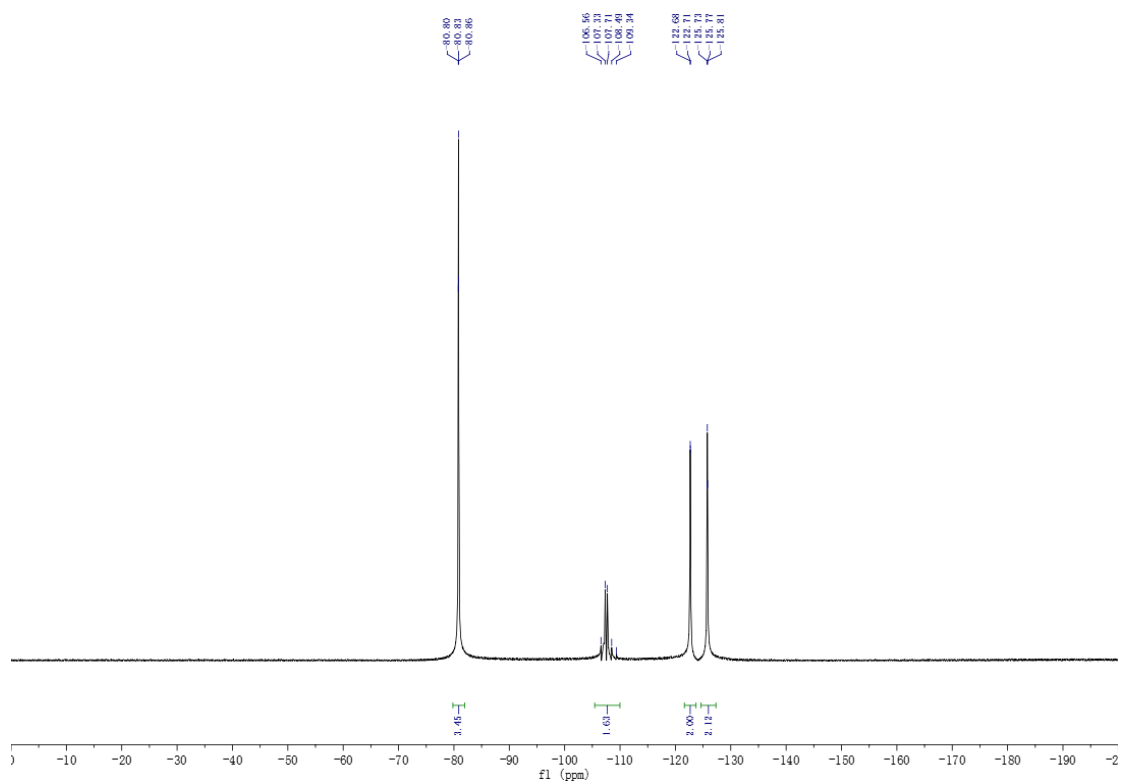
¹³C NMR of compound 35-1



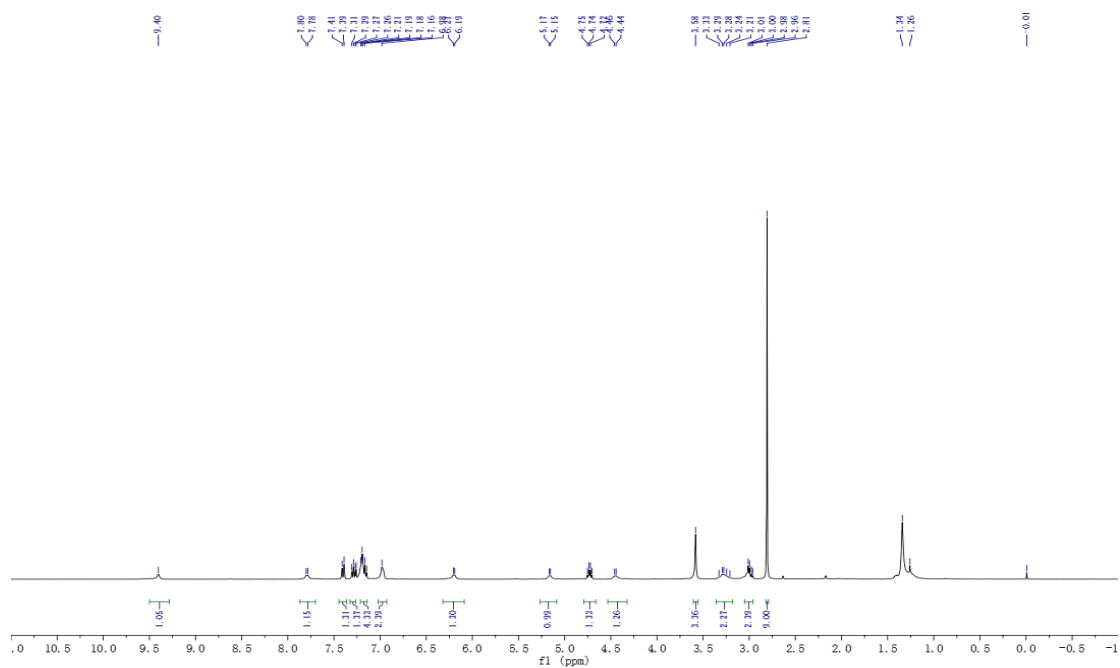
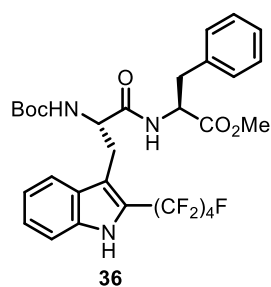
¹H NMR of compound **35**



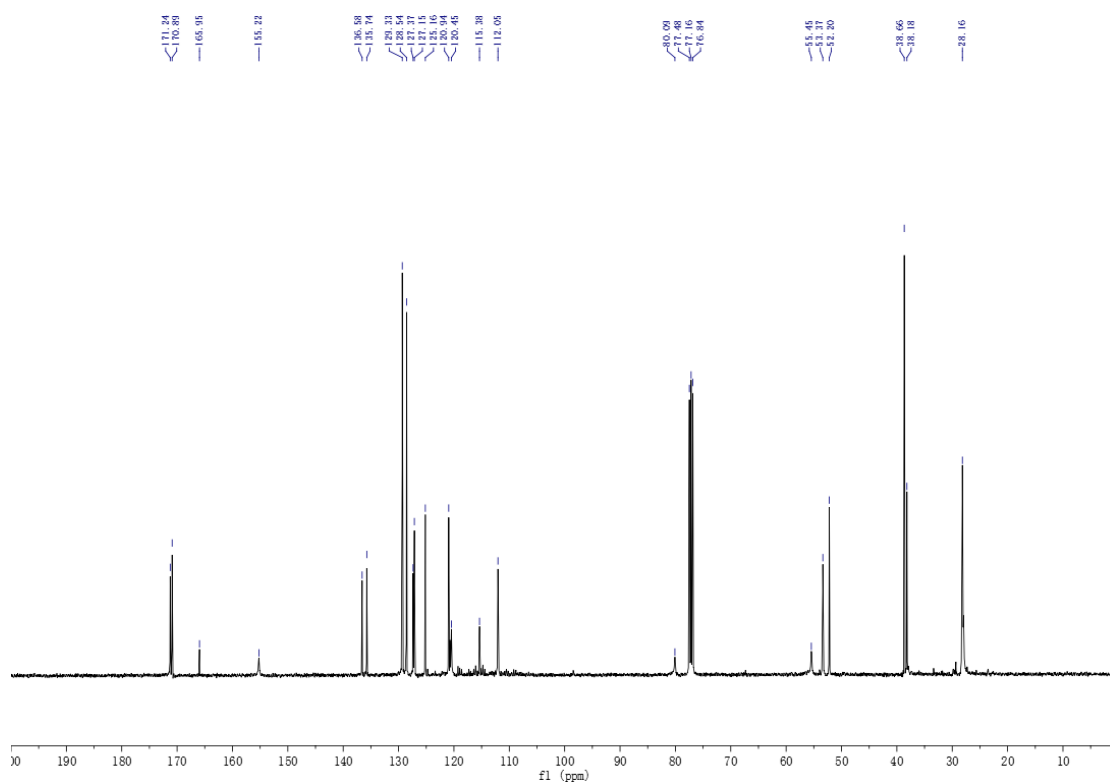
¹³C NMR of compound **35**



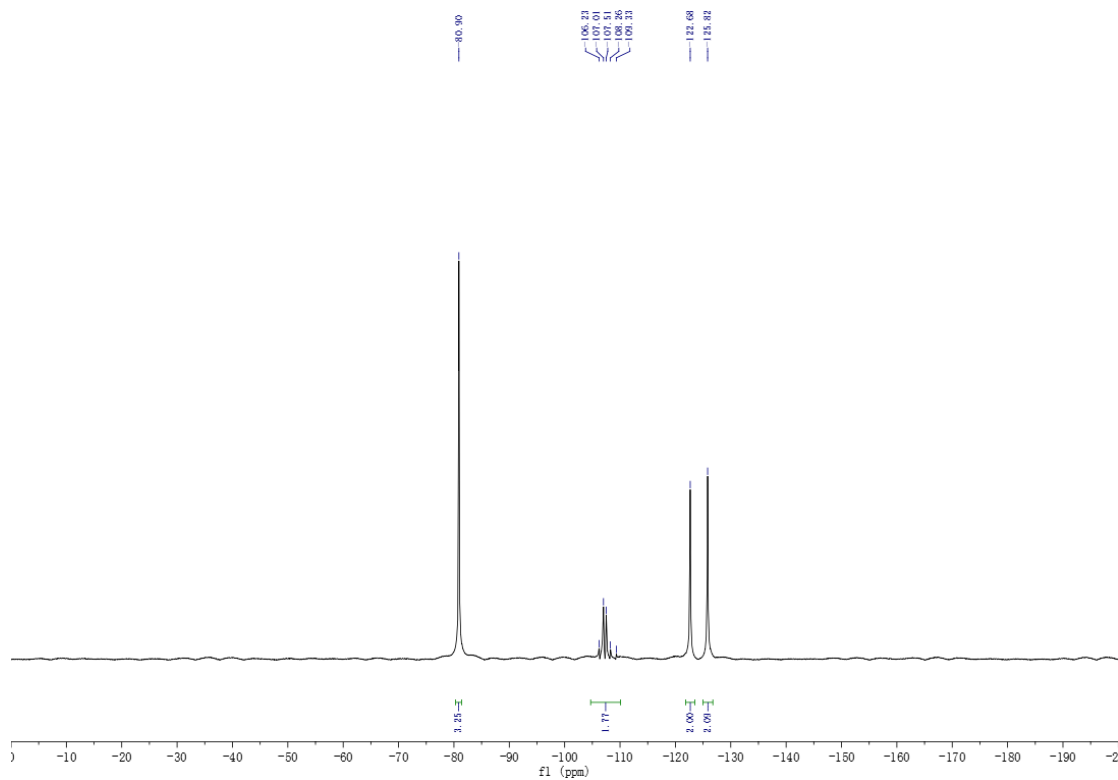
¹⁹F NMR of compound **35**



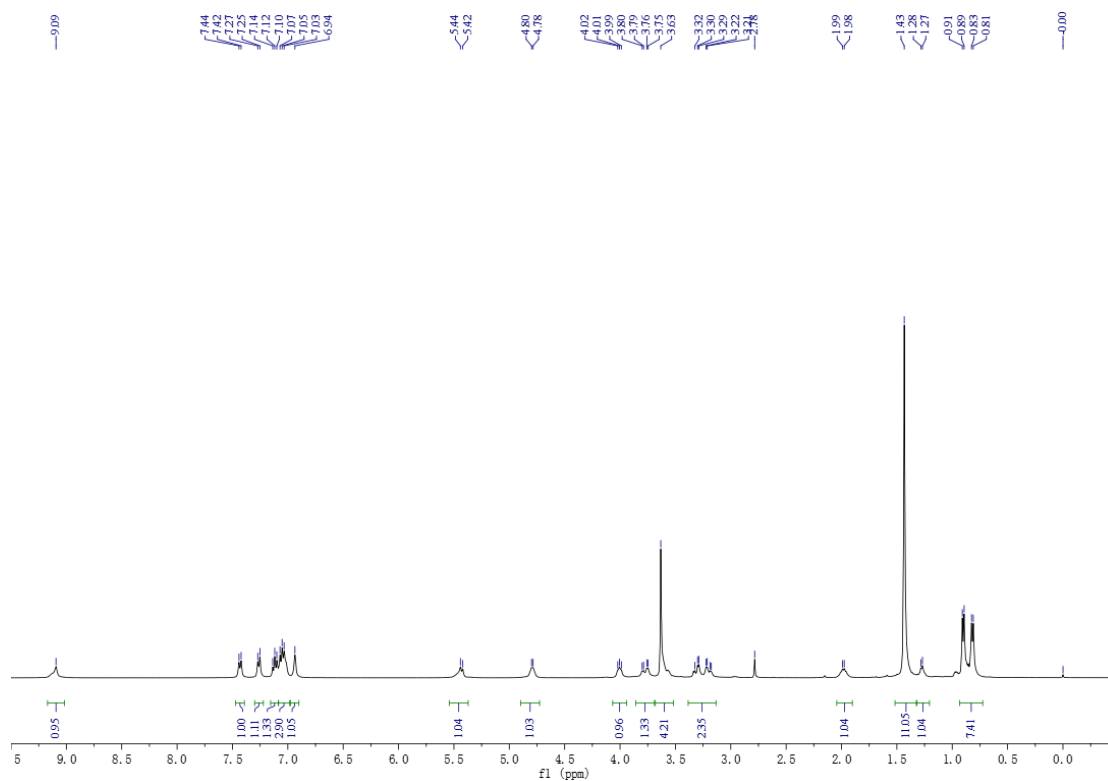
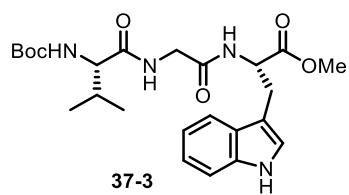
¹H NMR of compound **36**



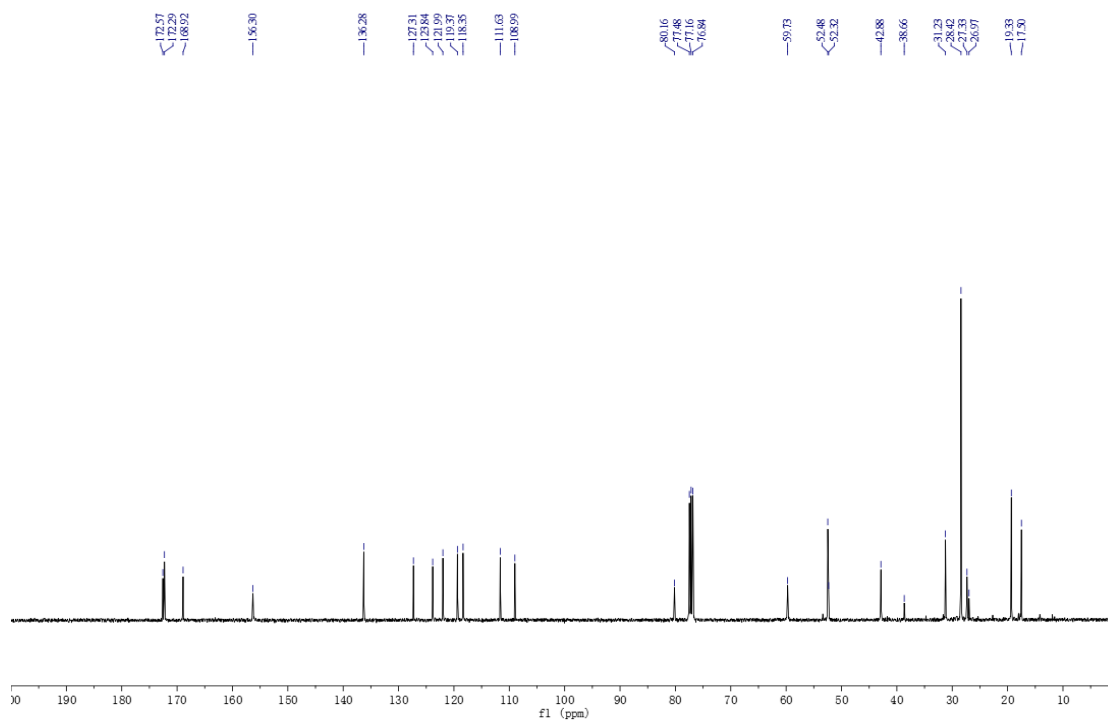
¹³C NMR of compound **36**



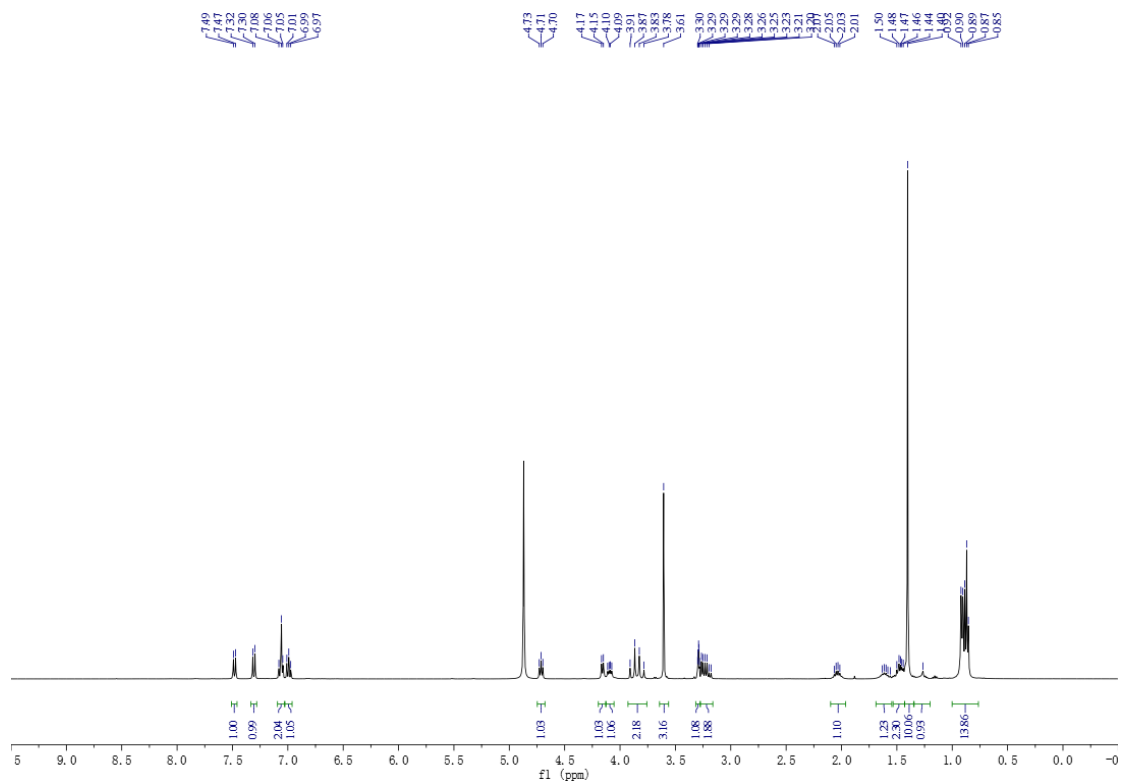
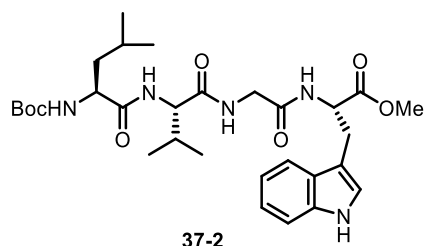
¹⁹F NMR of compound **36**



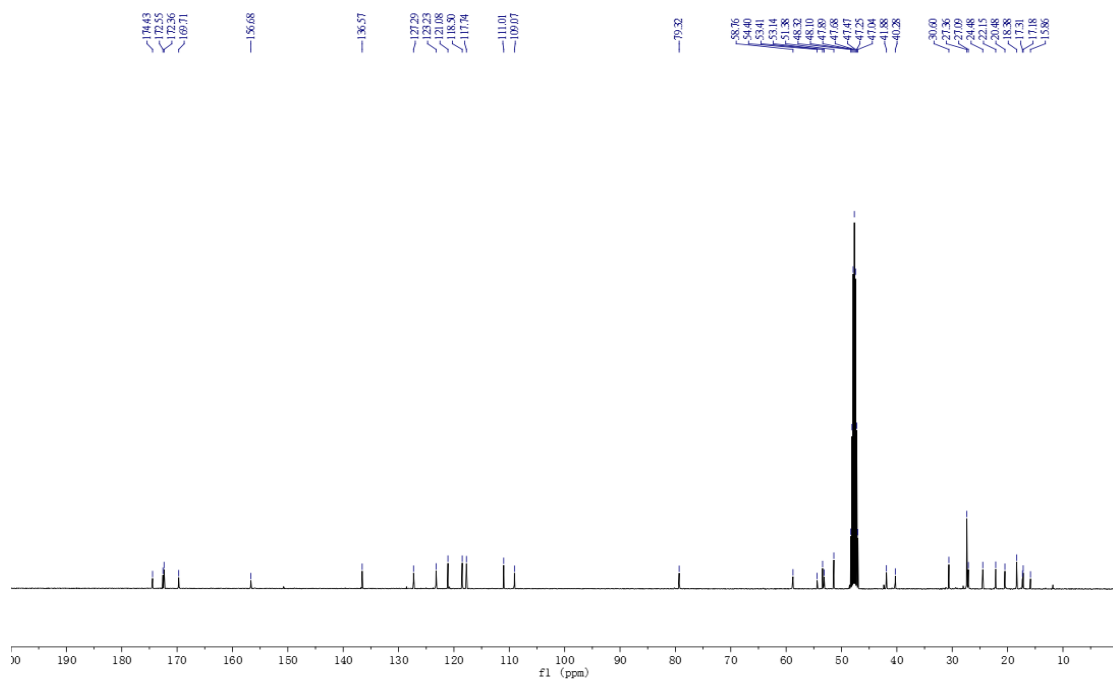
¹H NMR of compound **37-3**



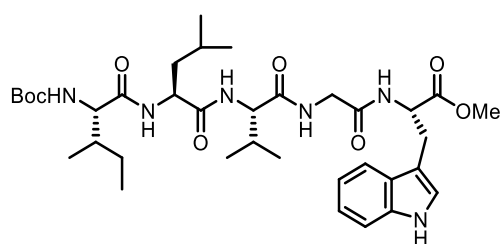
¹³C NMR of compound **37-3**



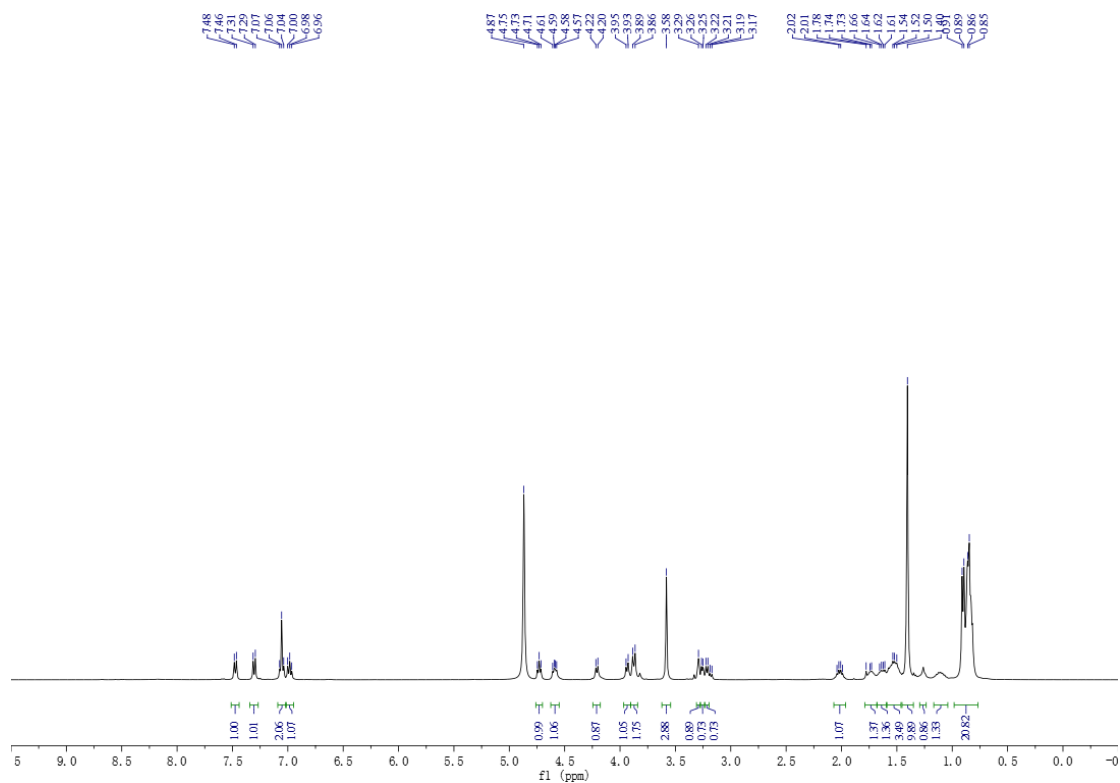
¹H NMR of compound **37-2**



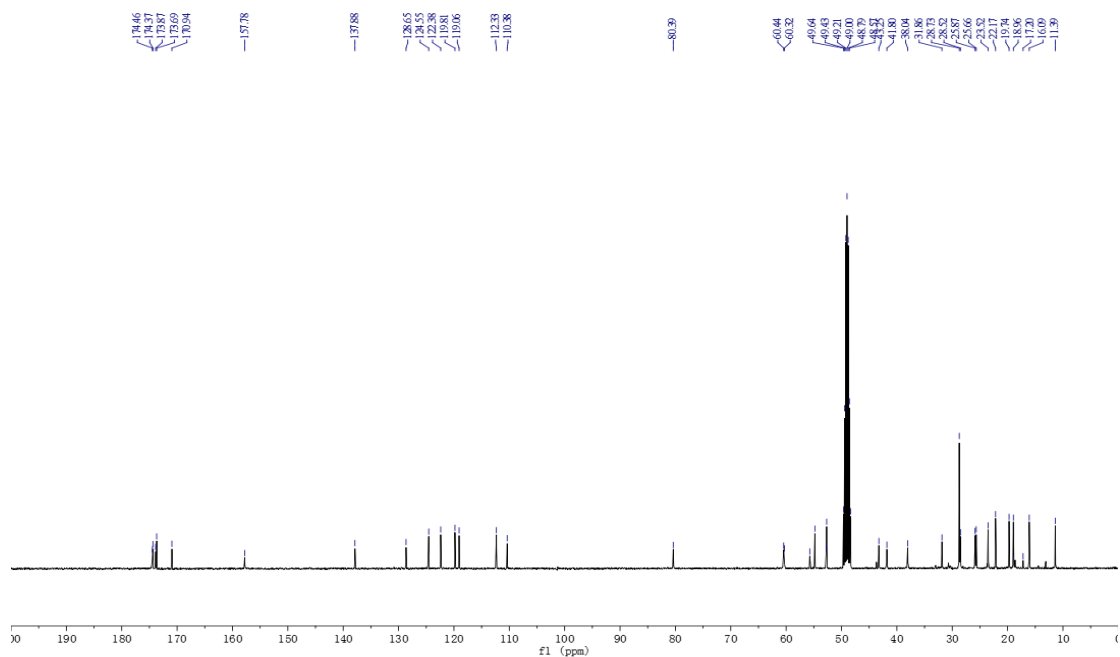
¹³C NMR of compound **37-2**



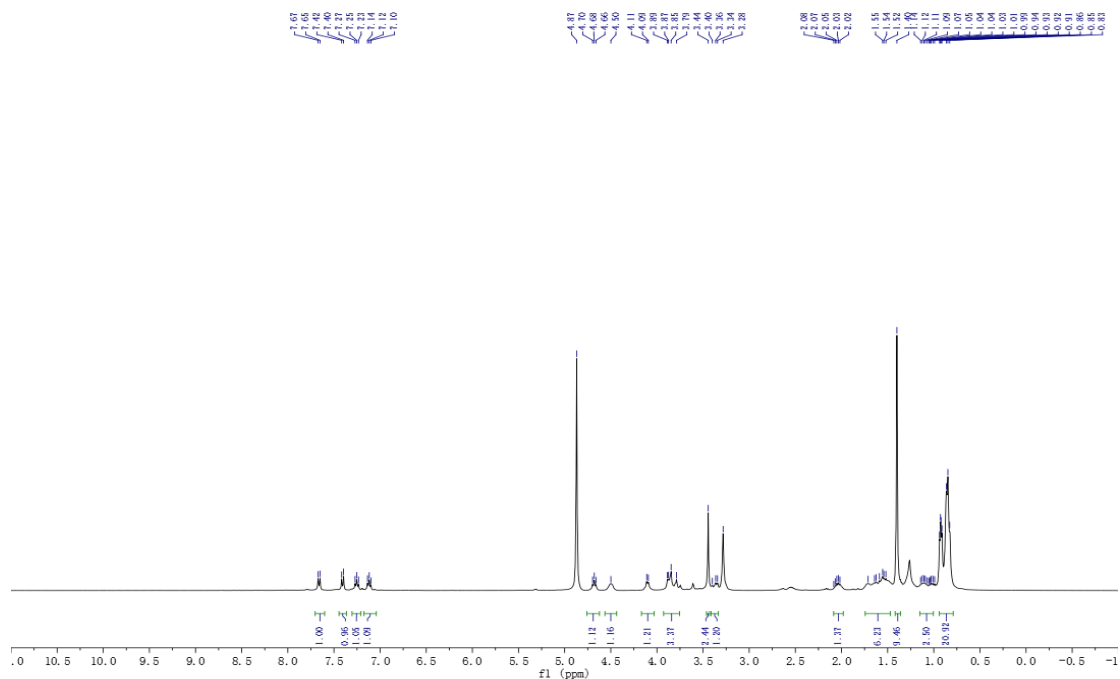
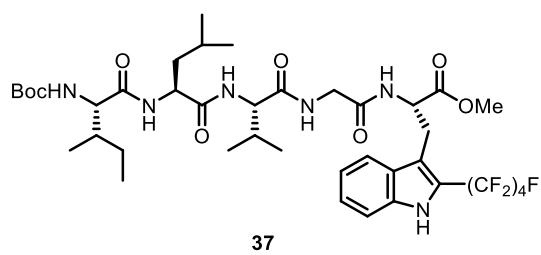
37-1



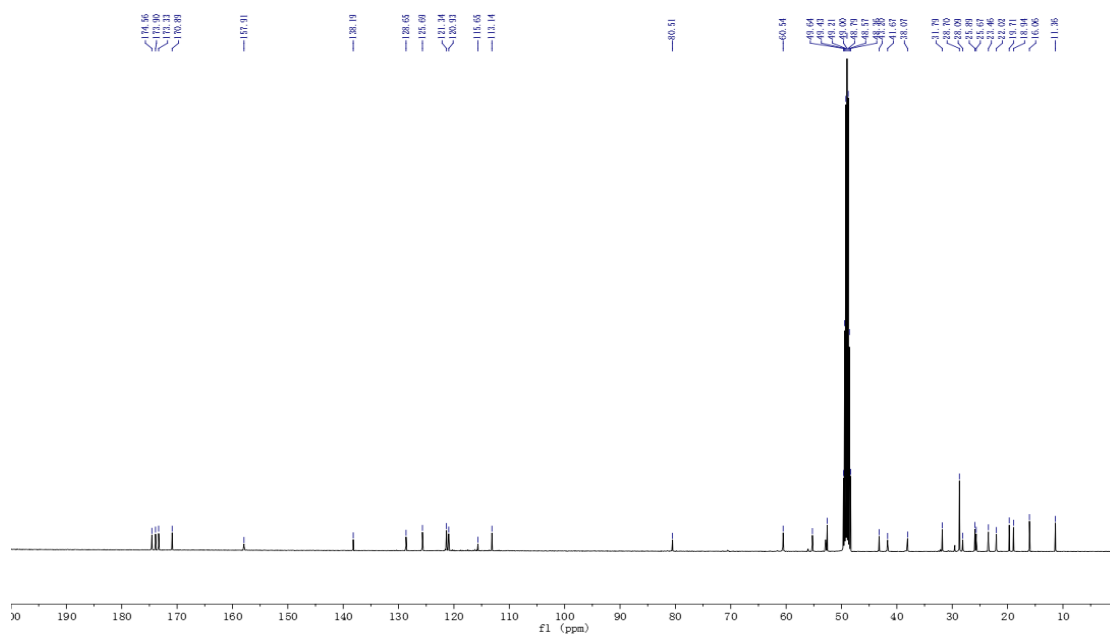
¹H NMR of compound **37-1**



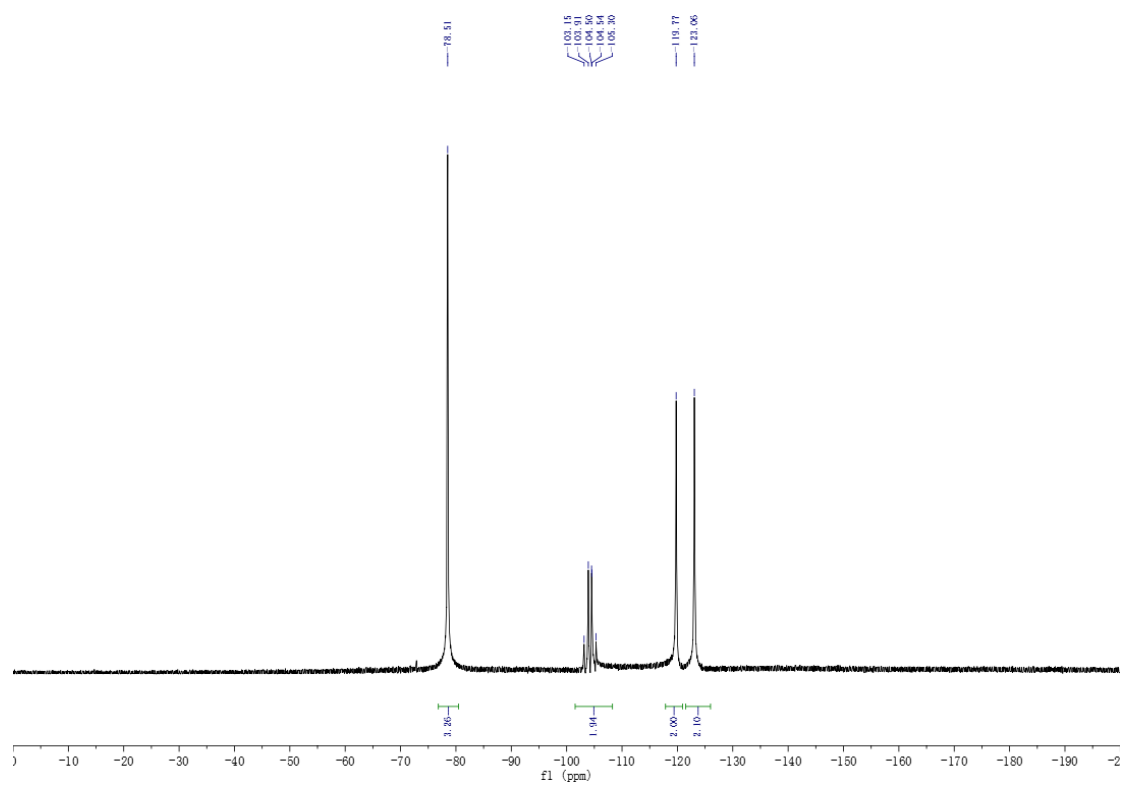
¹³C NMR of compound **37-1**



¹H NMR of compound **37**



¹³C NMR of compound **37**



^{19}F NMR of compound **37**