Supporting Information for Analytical Chemistry

Pinpoint the Positions of Single Nucleotide Polymorphisms by a Nanocluster Dimer

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DNA-template AgNCs probes and target DNA hybridization experiments

For NCD-1, NC-probe-0 AgNCs solution (49 uL) was mixed with NC-probe-1 AgNCs (49 uL) solution, then DPBS (2.94 uL) was quickly added into the mixed probes solution to make sure the volume of solution same as that of NCD-4 after hybridization. NC-probe-0 and NC-probe-1 hybridized with each other. The solutions were kept in 37 °C water bath for 30 mins before the following fluorescence measurements. The experimental conditions of NCD-2 were same as NCD-1 except that NC-probe-1 AgNCs solution was replaced by NC-probe-2 AgNCs solution.

For NCD-4, NC-probe-0 AgNCs solution (49 uL) was mixed with 49 uL NC-probe-4 AgNCs solution (49 uL), then perfectly-matched (PM) DNA target (2.94 uL 250 uM, dissolved in DPBS) was quickly added into the mixed probes solution. The solutions were kept in 37 °C water bath for 30 mins before the following fluorescence measurements. The experimental conditions of NCD-3 were same as NCD-4 except that NC-probe-4 AgNCs solution was replaced by NC-probe-3 AgNCs solution.

The fluorescence excitation and emission spectral curves of Nanocluster Dimers (NCD-1, NCD-2, NCD-3 and NCD-4) after hybridization were shown in Figure S1-S4. The fluorescence emission spectral curves (excited at the maximum excitation wavelength) of Nanocluster Dimers (NCD-1, NCD-2, NCD-3 and NCD-4) before and after hybridization were also shown in Figure S1-S4.

The sensitivity of NCD-4 for target DNA detection

Under the optimal conditions, we investigated the sensitivity of the NCD-4 probe for target DNA detection. We obtained a linear response of the fluorescence intensity with respect to the concentration of PM DNA target over the range 50 nM-7.28 uM ($R^2 = 0.99$). Finally, we obtained the limit of detection (LOD) to be 27.4 nM. The linear equation is y = 30.543x + 9.599 (Figure S10).

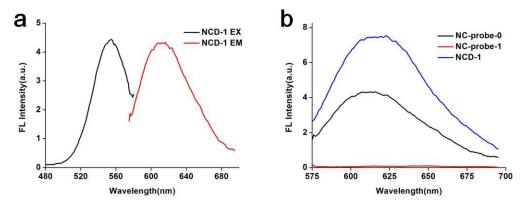


Figure S1. (a) Fluorescence excitation (in black) and emission (in red) spectra of NCD-1 after hybridization; (b) Fluorescence emission spectra of NC-probe-0 (in black), NC-probe-1 (in red), and NCD-1 (in blue) after hybridization excitated at 555 nm (Maximum emission of NCD-1 is at 613 nm).

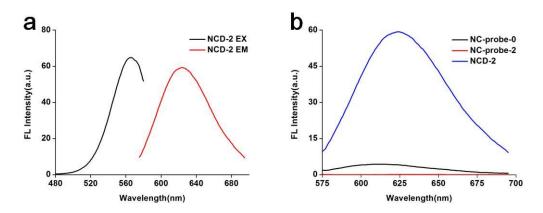


Figure S2. (a) Fluorescence excitation (in black) and emission (in red) spectra of NCD-2 after hybridization; (b) Fluorescence emission spectra of NC-probe-0 (in black), NC-probe-2 (in red), and NCD-2 (in blue) after hybridization excitated at 565 nm (Maximum emission of NCD-2 is at 623 nm).

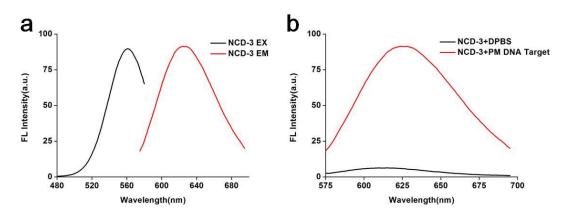


Figure S3. (a) Fluorescence excitation (in black) and emission (in red) spectra of NCD-3 after hybridization; (b) Fluorescence emission spectra of mixed solutions of NC-probe-0 and NC-probe-3 alone (in black), or hybridized with PM DNA target (in red) excitated at 560nm (Maximum emission of NCD-3 is at 625 nm).

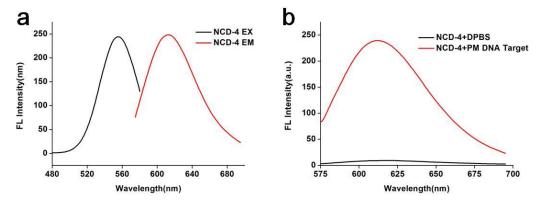


Figure S4. (a) Fluorescence excitation (in black) and emission (in red) spectra of NCD-4 after hybridization; (b) Fluorescence emission spectra of mixed solutions of NC-probe-0 and NC-probe-4 alone (in black), or hybridized with PM DNA target (in red) excitated at 555nm (Maximum emission of NCD-4 is at 613 nm).

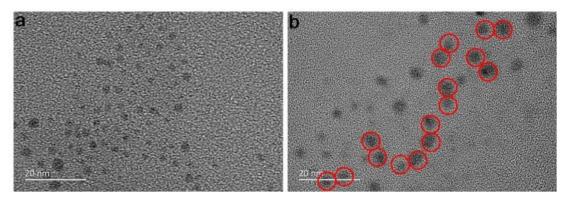


Figure S5. HRTEM micrographs of (a) AgNCs of NC probe-0 and NC probe-4 without target DNA, (b) The close proximity arrangement of two AgNCs of NC probe-0 and NC probe-4 with PM DNA. Scale bars = 20 nm.

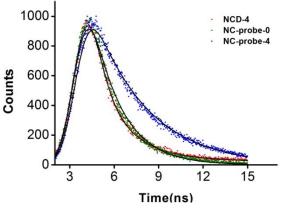


Figure S6. The fluorescence decay of NC-probe-0 AgNCs (in green), NC-probe-4 AgNCs (in blue) and NCD-4 after hybridization (in red), and their fitting lines (in black, NC-probe-0 and NC-probe-4 with a mono-exponential function, NCD-4 with a dual-exponential function)

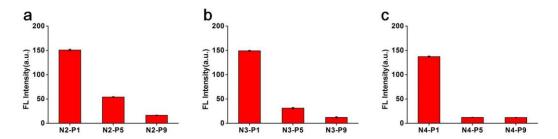


Figure S7. (a) Fluorescence intensities of NCD-4 for N2-P1, N2-P5 and N2-P9; (b) Fluorescence intensities of NCD-4 for N3-P1, N3-P5 and N3-P9; (c) Fluorescence intensities of NCD-4 for N4-P1, N4-P5 and N4-P9; (maximum emission 613nm, excited at 555nm, all the fluorescence intensities were measured at least three times)

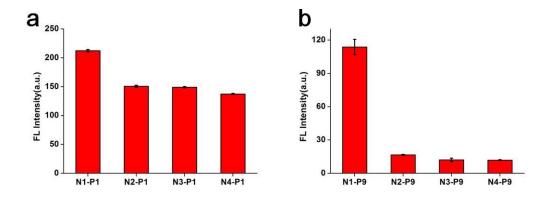


Figure S8. (a) Fluorescence intensities of NCD-4 for N1-P1, N2-P1, N3-P1 and N4-P1; (b) Fluorescence intensities of NCD-4 for N1-P9, N2-P9, N3-P9 and N4-P9; (maximum emission 613nm, excited at 555nm, all the fluorescence intensities were measured at least three times)

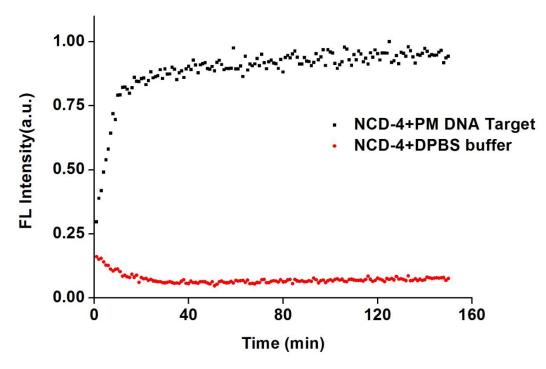


Figure S9. The kinetics of the mixed solutions of NC-probe-0 and NC-probe-4 alone, or hybridized with PM DNA target at 37 $^{\circ}$ C

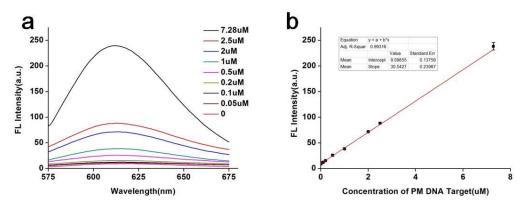


Figure S10. (a) Fluorescence emission spectra (excited at 555nm) of the solution with various concentrations of PM DNA target: $0, 5.0 \times 10^{-8}, 1.0 \times 10^{-7}, 2.0 \times 10^{-7}, 5.0 \times 10^{-7}, 1.0 \times 10^{-6}, 2.0 \times 10^{-6}, 2.5 \times 10^{-6}, 7.28 \times 10^{-6} \text{ mol} \cdot \text{L}^{-1}$; (b) Linear relationship between the fluorescence intensity (maximum emission 613nm) and the concentration of PM DNA (each error bar indicates the standard deviation).

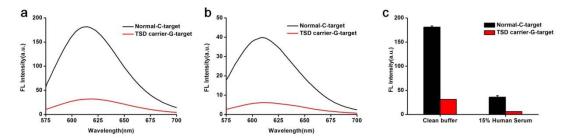


Figure S11. Fluorescence emission spectra of mixed solutions of NC-probe-0-tsd and NC-probe-4-tsd hybridized with normal-C-target (in black), or TSD carrier-G-target (in red) excitated at 555nm (Maximum emission of NCD-4 is at 613 nm) in clean buffer (a); and in 15% diluted human serum (b); (c) Fluorescence intensities of emission spectra in Figure S11a-b at 613nm (all the fluorescence intensities were measured at least three times).

Table S1. All the Sequences of oligonucleotides used in this work. NC-nucleation sequences of NC-probes in italic and underlined. Hybridization sequences of NC-probes in boldface. The SNPs sites and the nucleotides of target DNA are shown in red and boldface. T nucleotides spacers of target DNA are shown in red and underlined. The SNP information of Tay-Sachs disease mutations is obtained from http://www.snpedia.com.

Name of Sequences	Sequences from 5' to 3'					
NC probe-0	<u>CCCTAACTCCCC</u> AGATACAGCAAGAAGTT					
NC probe-1	$\underline{\mathit{CCTCCTTCCTCC}}\mathbf{AACTTCTTGCTGTATCT}$					
NC probe-2	${\bf AACTTCTTGCTGTATCT}\underline{CCTCCTTCCTCC}$					
NC probe-3	${\bf TTGAAGAACGACATAGA} \underline{CCTCCTTCCTCC}$					
NC-probe-4	TTGAAGAACGACATAGA <u>CCCTAACTCCCC</u>					
PM DNA	AACTTCTTGCTGTATCTTCTATGTCGTTCTTCAA					
N1-P1	${\sf AACTTCTTGCTGTATC} {\color{red}{\bf C}} {\tt TCTATGTCGTTCTTCAA}$					
N1-P5	${\sf AACTTCTTGCTG}^{\bf A}{\sf ATCTTCTATGTCGTTCTTCAA}$					
N1-P9	${\sf AACTTCTT}{\color{red}{\bf A}{\tt CTGTATCTTCTATGTCGTTCTTCAA}}$					
N2-P1	$AACTTCTTGCTGTAT \\ \textcolor{red}{GA}TCTATGTCGTTCTTCAA$					
N2-P5	$AACTTCTTGCT \color{red}{\textbf{CA}} ATCTTCTATGTCGTTCTTCAA$					
N2-P9	${\sf AACTTCT}{\color{red}{\bf AC}}{\color{blue}{\bf CTGTATCTTCTATGTCGTTCTTCAA}$					
N3-P1	AACTTCTTGCTGTA <mark>AGA</mark> TCTATGTCGTTCTTCAA					
N3-P5	AACTTCTTGC <mark>ACA</mark> ATCTTCTATGTCGTTCTTCAA					
N3-P9	AACTTCAACCTGTATCTTCTATGTCGTTCTTCAA					
N4-P1	$AACTTCTTGCTGT \color{red}{\textbf{TAGA}} TCTATGTCGTTCTTCAA$					
N4-P5	AACTTCTTGGACAATCTTCTATGTCGTTCTTCAA					
N4-P9	AACTT GAAC CTGTATCTTCTATGTCGTTCTTCAA					
P1	${\sf AACTTCTTGCTGTATC} {\color{red}{\bf C}} {\tt TCTATGTCGTTCTTCAA}$					
P2	$AACTTCTTGCTGTAT \textcolor{red}{\mathbf{T}} TTCTATGTCGTTCTTCAA$					
P3	$AACTTCTTGCTGTA{\color{red}GCTTCTATGTCGTTCTTCAA}$					
P4	$AACTTCTTGCTGT \\ \\ GTCTTCTATGTCGTTCTTCAA$					
P5	${\sf AACTTCTTGCTG}^{\bf A}{\sf ATCTTCTATGTCGTTCTTCAA}$					
P6	$AACTTCTTGCT {\color{red}A} TATCTTCTATGTCGTTCTTCAA$					
P7	$AACTTCTTGC{\color{red}{\bf C}}GTATCTTCTATGTCGTTCTTCAA$					
P8	$AACTTCTTG{\color{red}G}TGTATCTTCTATGTCGTTCTTCAA$					
P9	${\sf AACTTCTT}{\color{red}{\bf A}CTGTATCTTCTATGTCGTTCTTCAA}$					
P10	${\sf AACTTCT}{\color{red}{\bf A}{\sf GCTGTATCTTCTATGTCGTTCTTCAA}}$					
P11	${\sf AACTTC}{\textbf{G}}{\sf TGCTG}{\sf TATCTTCTATGTCGTTCTTCAA}$					
P12	AACTTGTTGCTGTATCTTCTATGTCGTTCTTCAA					
P13	AACTGCTTGCTGTATCTTCTATGTCGTTCTTCAA					
P14	AACATCTTGCTGTATCTTCTATGTCGTTCTTCAA					
P15	AATTTCTTGCTGTATCTTCTATGTCGTTCTTCAA					
P16	ACCTTCTTGCTGTATCTTCTATGTCGTTCTTCAA					
P17	CACTTCTTGCTGTATCTTCTATGTCGTTCTTCAA					
N1-P4 T-T	${\sf AACTTCTTGCTGT}{\color{red}{\bf T}{\sf TCTTCTATGTCGTTCTTCAA}}$					
N1-P4 T-C	${\sf AACTTCTTGCTGT}{\color{red}{\bf C}}{\color{blue}{\bf TCTTCTATGTCGTTCTTCAA}}$					

Name of Sequences	Sequences from 5' to 3'
N1-P4 T-G	AACTTCTTGCTGTGTCTTCTATGTCGTTCTTCAA
NC probe-0-tsd	<u>CCCTAACTCCCC</u> CCAATCACCAGAGCCTT
NC-probe-4-tsd	ACATACAAGCCTCTCCA <u>CCCTAACTCCCC</u>
Normal-C-target	$AAGGCT {\color{red}C} TGGTGATTGGTGGAGAGGCTTGTATGT$
TSD carrier-G-target	$AAGGCT {\color{red}G} TGGTGATTGGTGGAGAGGCTTGTATGT$
PM DNA-spacer-1T	$AACTTCTTGCTGTATCT\underline{\mathbf{T}}TCTATGTCGTTCTTCAA$
PM DNA-spacer-5T	$AACTTCTTGCTGTATCT\underline{-(T)}_{\underline{5}}\underline{-}TCTATGTCGTTCTTCAA$
PM DNA-spacer-10T	${\sf AACTTCTTGCTGTATCT}\underline{\text{-(T)}_{10}}\underline{\text{-}}TCTATGTCGTTCTTCAA$
PM DNA-spacer-15T	${\tt AACTTCTTGCTGTATCT\underline{-(T)}_{15\underline{-}}}{\tt TCTATGTCGTTCTTCAA}$
PM DNA-spacer-20T	${\sf AACTTCTTGCTGTATCT\underline{-(T)_{20}}\underline{-}}{\sf TCTATGTCGTTCTTCAA}$
PM DNA-spacer-25T	$AACTTCTTGCTGTATCT\underline{-(T)_{25}}\underline{-}TCTATGTCGTTCTTCAA$

Table S2. Gibbs binding free energy (ΔG) for the NC-Probe-0 and DNA target with single-base substitutions at different positions ("P0" means perfectly matched target). The values of ΔG shown in Table S2, are referred to the OligoAnalyzer 3.1 from the IDT Integrated DNA Technologies. http://sg.idtdna.com/calc/analyzer.

Mismat position		P0	P1	P2	P3	P4	P5	P6	P7
ΔG/(kc	al/mole)	-27.5	4 -25.95	-24.37	-22.9	-21.93	-20.59	-18.64	-17.04
P8	P9	P10	P11	P12	P13	P14	P15	P16	P17
-13.9	-11.95	-13.64	-15.6	-17.54	-19.14	-20.71	-22.66	-24.26	-25.6