## **Supporting Information**

## Use of Primary and Secondary Polyvinylamines for Efficient Gene Transfection

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Entry	Polymer	%C <sup>e</sup>	%N <sup>e</sup>	C/N <sub>exp</sub> <sup>e</sup>	C/N <sub>theor.</sub> f	% of hydrolysis <sup>g</sup>
1	PN700-Fs <sup>a</sup>	32.115	17.650	2.904	1.716	94
2	PN150-Cs <sup>a</sup>	n.d.	n.d.	n.d.	n.d.	n.d.
3	PN255-Cs <sup>a</sup>	33.105	17.675	1.873	1.716	91 <sup>h</sup>
4	PN660-Cs <sup>a</sup>	32.595	18.085	1.802	1.716	$94^{\rm h}$
5	PN940-Cs <sup>a</sup>	32.080	17.630	1.820	1.716	$95^{\rm h}$
6	PN1510-Cs <sup>a</sup>	32.485	17.930	1.812	1.716	$94^{\rm h}$
7	PN50-R <sup>b</sup>	32.420	17.920	1.809	1.716	99 <sup>i</sup>
8	PN170-R <sup>b</sup>	32.840	17.875	1.837	1.716	98 <sup>i</sup>
9	PN200-R <sup>b</sup>	31.825	18.150	1.753	1.716	$\geq 99^{i}$
10	PM100-Fh <sup>c</sup>	41.620	14.105	2.951	2.573	$78^{j}$
11	PM140-Fh <sup>c</sup>	38.945	13.410	2.904	2.573	81 <sup>j</sup>
12	PM165-Fh <sup>c</sup>	41.700	14.975	2.785	2.573	$88^{j}$
14	PM285-Fh <sup>c</sup>	46.165	15.060	3.065	2.573	71 <sup>j</sup>
15	PM110-Ch <sup>c</sup>	43.630	15.135	2.883	2.573	$82^{j}$
16	PM265-Ch <sup>c</sup>	43.895	15.680	2.799	2.573	87 <sup>j</sup>
17	PM310-Ch <sup>c</sup>	42.400	15.700	2.701	2.573	93 <sup>j</sup>
18	PM680-Ch <sup>c</sup>	43.610	16.010	2.724	2.573	91 <sup>j</sup>
19	PM155-Fm 23 <sup>d</sup>	52.480	13.450	3.902	2.573	23 <sup>j</sup>
20	PM155-Fm 37 <sup>d</sup>	50.800	13.935	3.645	2.573	37 <sup>j</sup>
21	PM155-Fm 44 <sup>d</sup>	48.570	13.740	3.535	2.573	44 <sup>j</sup>
22	PM155-Fm 54 <sup>d</sup>	46.665	13.940	3.348	2.573	54 <sup>j</sup>
23	PM155-Fm 64 <sup>d</sup>	44.345	13.895	3.191	2.573	64 <sup>j</sup>
24	PM155-Fm 76 <sup>d</sup>	43.760	14.670	2.983	2.573	76 <sup>j</sup>
25	PM155-Fm 94 <sup>d</sup>	42.005	15.695	2.676	2.573	94 <sup>j</sup>

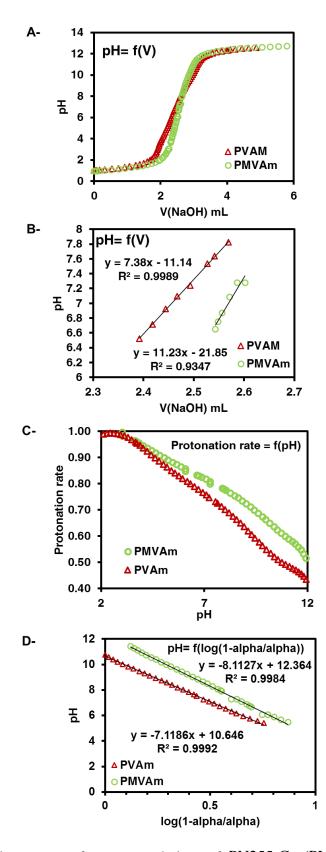
**<u>Table S1.</u>** Determination of the hydrolysis level of the poly(*N*-vinylamines) (PVAm) and poly(*N*-methylvinylamines) (PMVAm) by elemental analysis.

<sup>a</sup> Hydrolysis conditions: HCl 2 N at 120°C for 14h. <sup>b</sup> Hydrazinolysis conditions: [NVPi]/[hydrazine] = 1/24, in 1.4-dioxane/MeOH 1/2 at 65°C for one night. <sup>c</sup> Hydrolysis conditions: HCl 6 N at 120°C for 64 h. <sup>d</sup> Hydrolysis conditions: HCl 3 N at 100°C <sup>e</sup> Determined by elementary analysis. <sup>f</sup> Calculated for full hydrolysis of the amides moieties. <sup>g</sup> NVA hydrolysis level = 100 × (1- $f_{NVA residual}$ ) where  $f_{NVA residual}$  is the molar fraction of the residual non-hydrolyzed NVA units, and NMVA hydrolysis level = 100 × (1- $f_{NVA residual}$ ) where  $f_{NVA residual}$  where  $f_{NVA residual}$  is the molar fraction of the residual non-hydrolyzed NMVA units. <sup>h-j</sup>  $f_{NVA residual}$ ,  $f_{NWVA residual}$  and  $f_{NVPi residual}$  are determined based on formulas h-j (see below) established by taking into account the molar fraction of each comonomer in the copolymer precursor ( $F_{NVA}^0$ ,  $F_{NMVA}^0$  and  $F_{NVPi}^0$ ) and the respective numbers of carbon and nitrogen atoms in the hydrolyzed and non-hydrolyzed monomer units. MM<sub>c</sub> and MM<sub>N</sub> are the molar mass of C and N, respectively. n.d. = not determined.

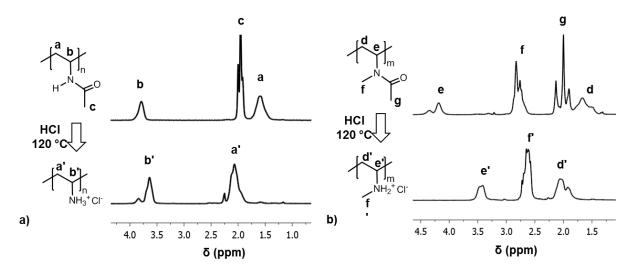
<sup>h</sup> 
$$F_{NVA\,residual} = \frac{[MM_N \times \frac{C}{N}] - [2 \times MM_C]}{2 \times MM_C}$$

<sup>i</sup> 
$$F_{residual NVPi} = \frac{\left[MM_N \times \frac{C}{N}\right] - [2 \times MM_C]}{8 \times MM_C}$$

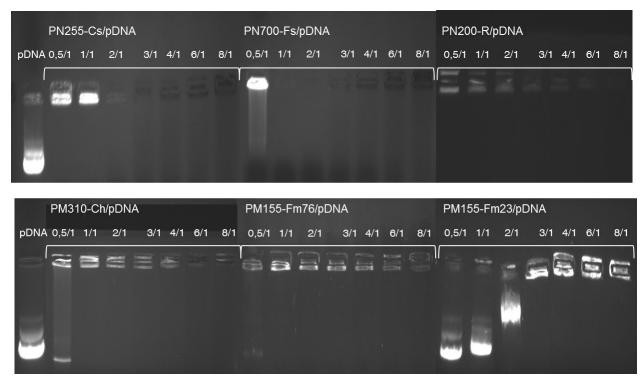
<sup>j</sup> 
$$F_{NMVA\,residual} = \frac{\left[MM_N \times \frac{C}{N}\right] - [3 \times MM_C]}{2 \times MM_C}$$



**Figure S1.** (A) Titration curves of aqueous solutions of PN255-Cs (PVAm) and PM140-Fh (PMVAm) (50 mg/ml in 10 mL of HCl 1M) with 0.5M NaOH, (B) linear regression of the titration curves of aqueous solutions of PN255-Cs and PM140-Fh in order to determine their buffer capacities between 6.5 < pH < 7.5, (C) the resulting protonation curves versus the pH and (D) determination of the pKa.



**Figure S2.** <sup>1</sup>H NMR analyses of **a**) PNVA ( $M_{n \text{ SEC-MALLS}} = 56300 \text{ g/mol}$ , D = 1.18) and **b**) PNMVA ( $M_{n \text{ SEC-MALLS}} = 30800 \text{ g/mol}$ , D = 1.12) samples before and after acid hydrolysis (6 N HCl/120°C). Spectra were recorded at 298K in D<sub>2</sub>O.



**Figure S3.** Agarose gel electrophoresis retardation assays of polyplexes made with different polyvinylamines (PN) (top) and poly(*N*-methylvinylamines) (PM) (bottom). Polyplexes were formed with various polymer/pDNA weight ratios (WR).

**<u>Table S2.</u>** Characteristics of polyvinylamine (PVAm) synthesized by RAFT polymerization of *N*-vinylphthalimide and successive hydrazinolysis.

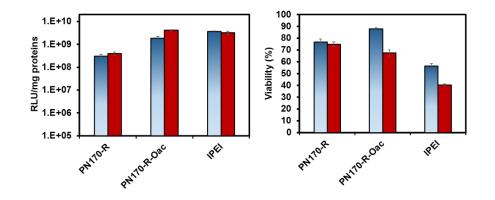
			PNVPi <sup>a</sup>					PVAm <sup>b</sup>		
Entry	Name	$\overline{M_{ m n,th}}^{ m c}$	$M_{\rm n}^{\rm LS}$	<i>DP</i> <sup>e</sup> <sub>n</sub>	${oldsymbol{ heta}}^{ m f}$	Conv.	% of	M <sub>n</sub>		
			(kg/mol) <sup>d</sup>	DI n		(%)	hydrazinolysis <sup>g</sup>	(kg/mol) <sup>h</sup>		
	1	PN50-R	5	8	46	1.43	> 99	99	2.0	
	2	PN170-R	21	29	167	1.52	81	98	7.2	
	3	PN200-R	23	35	200	1.61	59	99	8.6	

<sup>a</sup> Conditions for PN50-R, PN170-R and PN200-R are respectively: [NVPi]/[AIBN]/[CTA] = 25/0.25/1, 150/0.25/1 and 227/0.25/1, for 12 h, 24 h and 72 h. <sup>b</sup> Conditions of the hydrazinolysis: [PNVPi]/[hydrazine] = 1/24, in 1.4-dioxane/MeOH 1/2 at 65°C for one night. <sup>c</sup>  $M_{n,th} = DP_n^{th} x$  conversion x MM<sub>monomer</sub> <sup>d</sup>  $M_n^{LS}$  determined by SEC in DMF equipped with a MALLS detector,  $dn/dc_{PNVPi} = 0.131$ . <sup>e</sup> Calculated using the following formula:  $DP_n = M_n/MM_{monomer}$ . <sup>f</sup> Determined by SEC in DMF using a PMMA calibration. <sup>g</sup> Determined by elemental analysis (SI Table S1 for crude EA analysis and calculations). <sup>h</sup> Number-average molar mass calculated by the molar mass of the precursor.

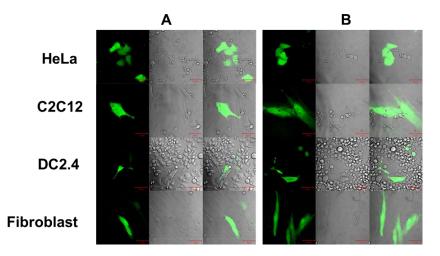
Entry	Polyplexes	polymer/pDNA WR <sup>a</sup>	N/P <sup>b</sup>	$\mathbf{D_h}^{\mathbf{c}}$	$\zeta^{\mathbf{d}}$	
	_ •- <i>5</i> <b>F</b> ••••			( <b>nm</b> )	( <b>mV</b> )	
1	PN50-R-plex	1	7	3490	+27	
		3	22	150	+37	
2	PN170-R-plex	1	7	460	+40	
		3	21	140	+44	
3	PN200-R-plex	1	7	90	+23	
		3	22	130	+40	
2'	PN170-R-OAc-plex <sup>e</sup>	1	2	185	+27	
		3	5	140	+33	

Table S3. Characteristics of pDNA complexes made with PVAm polymers prepared via RAFT polymerization

<sup>a</sup> WR = polymer/pDNA weight ratio. <sup>b</sup> amine/phosphate molar ratio calculated as described in experimental part. <sup>c</sup> Hydrodynamic diameters  $D_h$  of the polyplexes at 298K in HEPES 10 mM, pH 7.4. <sup>d</sup>  $\zeta$  potential of polyplexes at 298K in HEPES 10 mM, pH 7.4. <sup>e</sup> Obtained by acetylation of PN170-R (degree of acetylation = 50 %).



**Figure S4.** (A) Transfection efficiency and (B) cell viability of HeLa cells. Transfection was performed with PVAm (made by RAFT) before (PN170-R) and after 50 % acetylation (PN170-R-OAc) polyplexes at two polymer/pDNA ratios (ratio 1 (blue) and ratio 2 (red): lower and higher amount of polymer, Table S3). The luciferase activity was measured 48 h after the transfection and expressed as RLU/mg of protein. The cell viability was evaluated by MTT assay 48 h after transfection and expressed as percentage relative to untreated cells.



**Figure S5.** HeLa, C2C12, DC2.4 cells and fibroblasts were transfected with PM140-Fh polyplexes at N/P = 3 (A) and 6 (B) containing pDNA encoding EGFP. EGFP-positive cells were analyzed by fluorescent confocal microscopy. Fluorescence images (left), phase contrast images (middle) and their merge (right).