## HIV-1 Capsid Function is Regulated by Dynamics: Quantitative Atomic-Resolution Insights by Integrating Magic-Angle-Spinning NMR, QM/MM, and MD

Huilan Zhang<sup>1,2,#</sup>, Guangjin Hou<sup>1,2,#</sup>, Manman Lu<sup>1,2</sup>, Jinwoo Ahn<sup>2,3</sup>, In-Ja L. Byeon<sup>2,3</sup>, Christopher J. Langmead<sup>4</sup>, Juan R. Perilla<sup>5</sup>, Ivan Hung<sup>6</sup>, Peter L. Gor'kov<sup>6</sup>, Zhehong Gan<sup>6</sup>, William W. Brey<sup>6</sup>, David A. Case<sup>7</sup>, Klaus Schulten<sup>5</sup>, Angela M. Gronenborn<sup>2,3\*</sup>, and Tatyana Polenova<sup>1,2\*</sup>

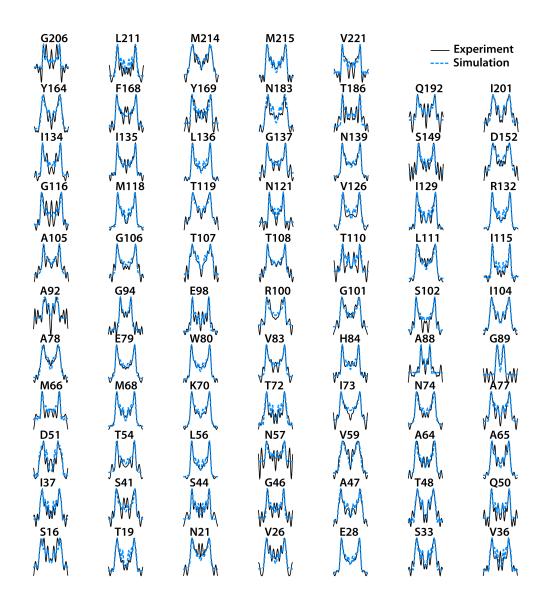
<sup>1</sup>Department of Chemistry and Biochemistry, University of Delaware, Newark, Delaware 19716, United States;
<sup>2</sup>Pittsburgh Center for HIV Protein Interactions, University of Pittsburgh School of Medicine, 1051 Biomedical Science Tower 3, 3501 Fifth Ave., Pittsburgh, PA 15261, United States; <sup>3</sup>Department of Structural Biology, University of Pittsburgh School of Medicine, 3501 Fifth Ave., Pittsburgh, PA 15261, United States; <sup>4</sup>Computer Science Department, Carnegie Mellon University, Gates Hillman Center, 5000 Forbes Avenue, Pittsburgh, PA, United States; ;
<sup>5</sup>Department of Physics and Beckman Institute for Advanced Science and Technology University of Illinois at Urbana-Champaign, Urbana, Illinois 61801; <sup>6</sup>National High Magnetic Field Laboratory, Florida State University, Tallahassee, FL, 32310, United States; <sup>7</sup>Department of Chemistry and Chemical Biology, Rutgers University, 174 Frelinghuysen Road, Piscataway, NJ 08854-8087, United States

<sup>#</sup>These authors have contributed equally

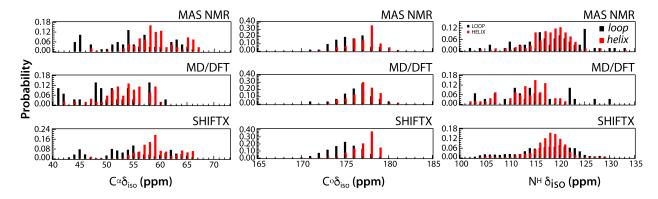
\*Corresponding authors: Tatyana Polenova, Department of Chemistry and Biochemistry, University of Delaware, Newark, DE, USA, Tel.: (302) 831-1968; Email: tpolenov@udel.edu; Angela M. Gronenborn, Department of Structural Biology, University of Pittsburgh School of Medicine, 3501 Fifth Ave., Pittsburgh, PA 15260, USA, Tel.: (412) 648-9959; Email: amg100@pitt.edu

Classification: Biological Sciences- Biophysics and Computational Biology

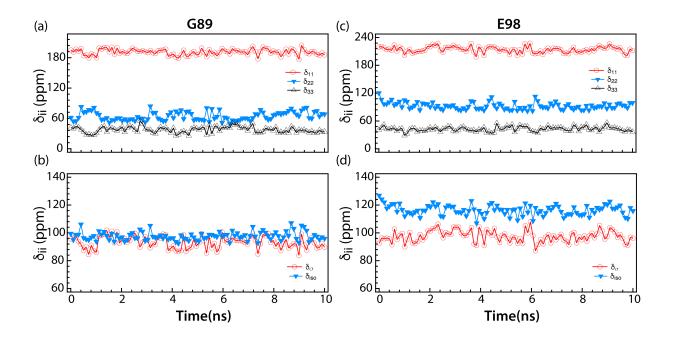
**Keywords:** magic-angle spinning NMR, HIV-1 capsid, CA protein assemblies, HIV-AIDS, conformational dynamics, chemical shift anisotropy, quantum mechanics/molecular mechanics



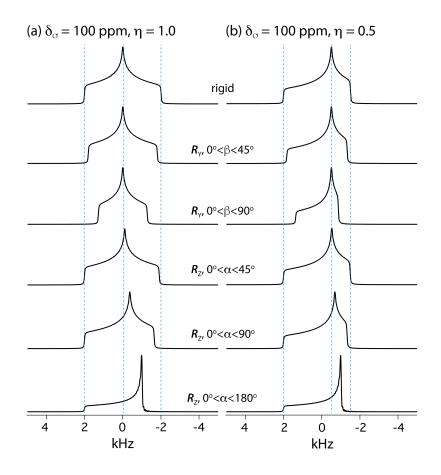
*Figure S1.* Experimental (solid black lines) and simulated (dashed blue lines)  $^{15}$ N CSA lineshapes for different residues in tubular assemblies of CA HXB2 extracted from the R8<sub>1</sub><sup>3</sup>-RNCSA 3D spectra, recorded at the magnetic field of 21.1 T and the MAS frequency of 14 kHz.



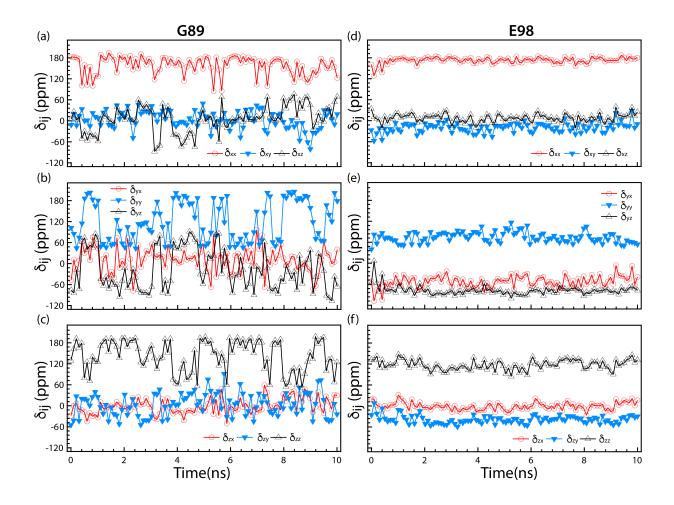
*Figure S2.* Distribution plots for the isotropic  ${}^{13}C^{\alpha}$  (left),  ${}^{13}C^{\circ}$  (middle), and  ${}^{15}N^{H}$  (right) chemical shifts in HIV-1 CA assemblies. Top: experimental MAS NMR; middle, calculated from MD/DFT; bottom, calculated by SHIFTX as the averaged values over the MD trajectory. The distributions for helical regions are shown in red, for loops- in black.



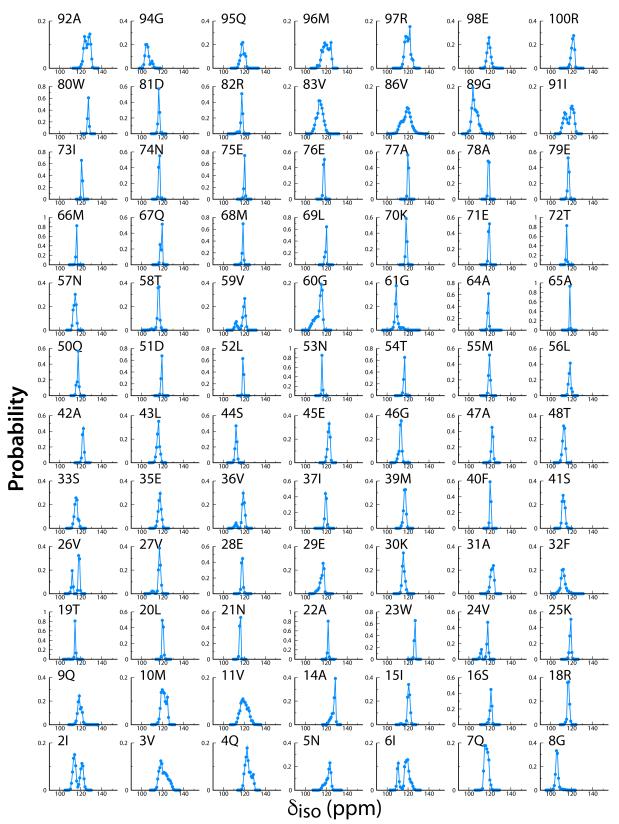
*Figure S3.* Principal components of <sup>15</sup>N CSA tensor,  $\delta_{i}$ ,  $\delta_{\sigma}$ , and  $\delta_{iso}$ , calculated along the MD trajectory, for selected CA residues: G89 (a, c) and E98 (b, d). For the calculations, 100 frames were used from the first 10 ns of the 100-ns MD trajectory.



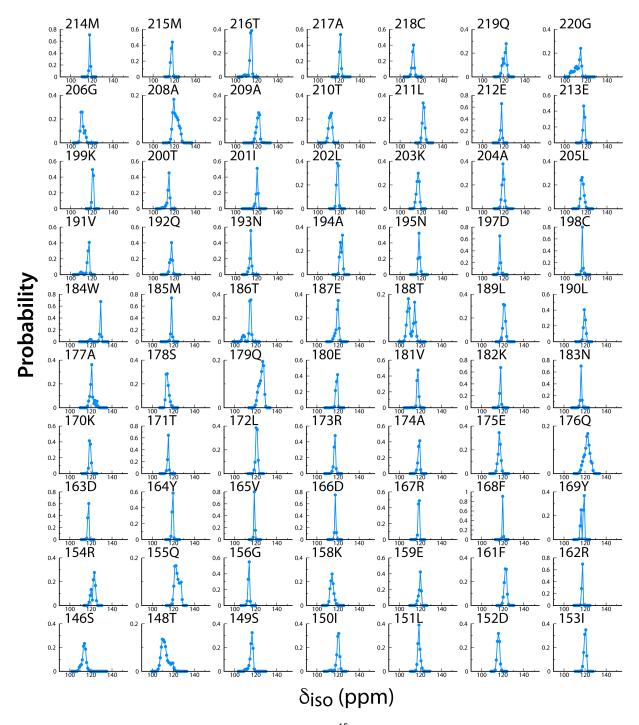
*Figure S4.* Simulated rigid and motionally reduced  $^{15}N$  CSA line shapes for the sites with the following CSA NMR parameters: (a)  $\delta_{\sigma}$  = 100 ppm and  $\eta$  = 1.0; (b)  $\delta_{\sigma}$  = 100 ppm and  $\eta$  = 0.5. The Euler angles are indicated next to each line shape.



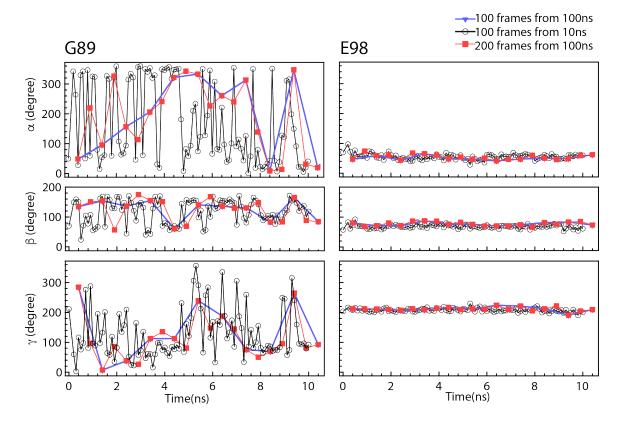
*Figure S5.* Individual components of <sup>15</sup>N CSA tensor  $\delta_{ij}$  (molecular fixed frame representation), calculated along the MD trajectory, for selected CA residues: G89 (a-c) and E98 (d-f). For the calculations, 100 frames were used from the first 10 ns of the 100-ns MD trajectory.



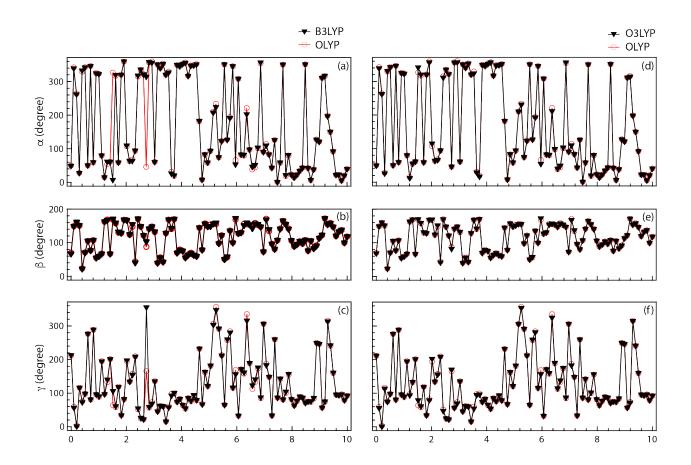
*Figure S6.* Probability distributions of <sup>15</sup>N isotropic chemical shifts of HIV-1 CA calculated by Shiftx based on 5000 frames extracted from 100 ns MD simulation.



*Figure S6.* (con'd) Probability distributions of <sup>15</sup>N isotropic chemical shifts of HIV-1 CA calculated by Shiftx based on 5000 frames extracted from 100 ns MD simulation.



*Figure* **S7.** Euler angles of the <sup>15</sup>N CSA tensors for G89 and E98 residues of CA, calculated by MD/DFT with different sampling schedules: 200 frames from 100-ns MD trajectory, red; 100 frames from 100-ns MD trajectory, blue; 100 frames from the first 10 ns of the 100-ns MD trajectory, black. The corresponding <sup>15</sup>N CSA parameters for G89 are:  $\delta_{\sigma}$  = 28.92 ppm,  $\eta_{\sigma}$  = 0.11;  $\delta_{\sigma}$  = 23.28 ppm,  $\eta_{\sigma}$  = 0.10;  $\delta_{\sigma}$  = 23.75 ppm,  $\eta_{\sigma}$  = 0.12. The corresponding <sup>15</sup>N CSA parameters for E98 are:  $\delta_{\sigma}$  = 93.83 ppm,  $\eta_{\sigma}$  = 0.52;  $\delta_{\sigma}$  = 93.00 ppm,  $\eta_{\sigma}$  = 0.53;  $\delta_{\sigma}$  = 92.94 ppm,  $\eta_{\sigma}$  = 0.54.



*Figure S8.* Euler angles of the <sup>15</sup>N CSA tensors for G89 residue in the molecular frame along the MD trajectory, calculated using (a-c) B3LYP and OLYP functionals (black and red symbols, respectively), and (d-f) O3LYP and OLYP functionals (black and red symbols, respectively). The angles were calculated using  $\delta_{\sigma}$  = 25.63, 23.75, and 24.66 ppm for B3LYP, OLYP, and O3LYP, respectively. Note that the differences are small for  $\delta_{\sigma}$  computed with the three functionals. For the calculations, 100 frames were used from the first 10 ns of the 100-ns MD trajectory.