

Supporting Information

Can Simulations and Modeling Decipher NMR Data for Conformational Equilibria? Arginine-Vasopressin

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Computational details

Long-scale molecular dynamics simulation

Table S1 Parameters for the unrestrained long-scale molecular-dynamics simulation of Arg⁸-vasopressin

Force field	ff99SB ¹ (Amber 10, Amber 14 CUDA) ²
Initial conformation	open (PDB ID: 1YF4), ³ neutralized with 2 Cl ⁻ (ions08.lib, frcmod.ionsjc_tip4pew ^{4,5})
Solvation	explicit, TIP4PEw, ⁶ truncated octahedral box
Temperature and Pressure	T= 300K, p= 1 bar (Berendsen coupling, ⁷ 1.0 ps external heat bath)
Minimization	8,945 steps: 500 steps steepest-descent followed by conjugate-gradient method
Molecular dynamics	2 fs time steps, SHAKE algorithm, ⁸ 8.0 Å non-bonded cut-off, Particle Mesh Ewald method, ⁹ periodic boundary conditions
Simulation time	23,000 ns

Representative conformations of Arg⁸-vasopressin

Analysis of the conformational space of 11 μs MD simulation revealed 4 main ring conformations for AVP: *open*, *saddle*, *clinched open*, *twisted saddle*. A detailed description of the conformations has been published previously.¹⁰

The most populated overall state of each cluster (ring-state type, main ring conformation) was chosen as representative for further DFT and NMR calculations; *saddle* and *clinched open* each of with *extended* and *folded* tail. The mean torsions of the representatives are given in Table S2.

Table S2 Mean backbone torsions and standard deviations (±) of representative conformations of Arg⁸-vasopressin

		<i>saddle_{ext}</i>		<i>saddle_{fold}</i>		<i>cl.open_{ext}</i>		<i>cl.open_{fold}</i>		<i>tw.saddle</i>		<i>open</i>	
		state3 ^a	state6	state3 ^a	state6	state12	state14	state19	state27	state19	state27	state19	state27
Tyr ²	phi	-80.9	±21.4	-78.5	±18.5	-101.2	±28.8	-86.8	±25.4	-84.6	±29.0	-112.0	±37.9
	psi	144.2	±12.8	143.2	±11.6	-16.4	±23.3	-22.9	±21.1	162.1	±13.1	134.6	±18.8
Phe ³	phi	-63.0	±9.5	-62.7	±9.1	-99.9	±32.8	-106.6	±26.0	-52.0	±16.8	54.9	±12.0
	psi	-20.7	±13.6	-22.5	±12.5	157.4	±13.5	154.5	±16.0	127.3	±15.1	4.4	±31.3
Gln ⁴	phi	-86.5	±17.4	-89.2	±16.3	-66.4	±18.0	-69.4	±11.3	55.0	±8.4	-135.9	±23.8
	psi	-7.6	±17.8	-4.7	±16.5	-18.2	±25.7	-21.8	±18.7	12.4	±21.1	151.6	±19.2
Asn ⁵	phi	-113.3	±21.2	-113.8	±21.0	-112.1	±27.8	-109.9	±30.4	-106.1	±29.4	-75.4	±19.8
	psi	-27.4	±21.8	-26.4	±23.3	74.5	±62.7	92.8	±57.7	-8.7	±47.3	124.7	±31.9
Cys ⁶	phi	-126.3	±20.0	-126.6	±21.0	-111.1	±34.9	-117.3	±38.9	-120.8	±28.3	-128.8	±30.4
	psi	132.7	±34.5	139.8	±23.1	145.9	±19.2	145.6	±20.1	143.8	±24.2	149.0	±22.7
Pro ⁷	phi	-67.8	±11.3	-64.6	±11.1	-66.3	±11.1	-64.8	±11.5	-67.7	±11.2	-67.5	±11.5
	psi	-63.0	±11.3	-62.7	±11.3	-99.9	±32.8	-106.6	±26.0	-52.0	±16.8	53.3	±12.0
Arg ⁸	phi	-20.8	±13.8	150.3	±12.5	150.3	±16.0	150.3	±16.0	127.3	±15.1	4.4	±31.3
	psi	-86.5	±17.4	-89.2	±16.3	-69.4	±18.0	-69.4	±11.3	55.0	±8.4	-135.9	±23.8

^aPDB files of the representative states are available under these designations from the SI (ESM4) of our previous publication (<http://dx.doi.org/10.1007/s00894-014-2485-0>); Abbreviations: clinched (cl.), twisted (tw.); ext (extended); fold (folded)

After extending the MD-simulation to 23 μ s, the conformational space was again clustered using DASH¹¹. The resulting representatives were assigned to the former representatives already defined for the first 11 μ s of the MD simulation via circular similarity of backbone torsions in order to ensure consistency. From the 23 μ s MD simulation the representatives of the five most populated ring state types were chosen in order to calculate their free energies and populations. The first four clusters were the already identified main conformations from the 11 μ s MD; the additional conformation (*variants*) was seen for the first time after 11 μ s. *Variants* is a *clinched open* variant; a rotamer of the peptide-bond between residue Gln⁴ and Asn⁵ of the *clinched open* conformation (Figure S1). *Variants* was added to the selection, although low populated in the simulation, because it occurred only once and at the end of the 23 μ s simulation with no following interconversion. Thus, it could not be ruled out that this cluster might be another main cluster of AVP. However, the thermodynamic calculations (metadynamics) showed that the conformation is the least stable (*cf.* main text) and it was not considered further for DFT-NMR-calculations.

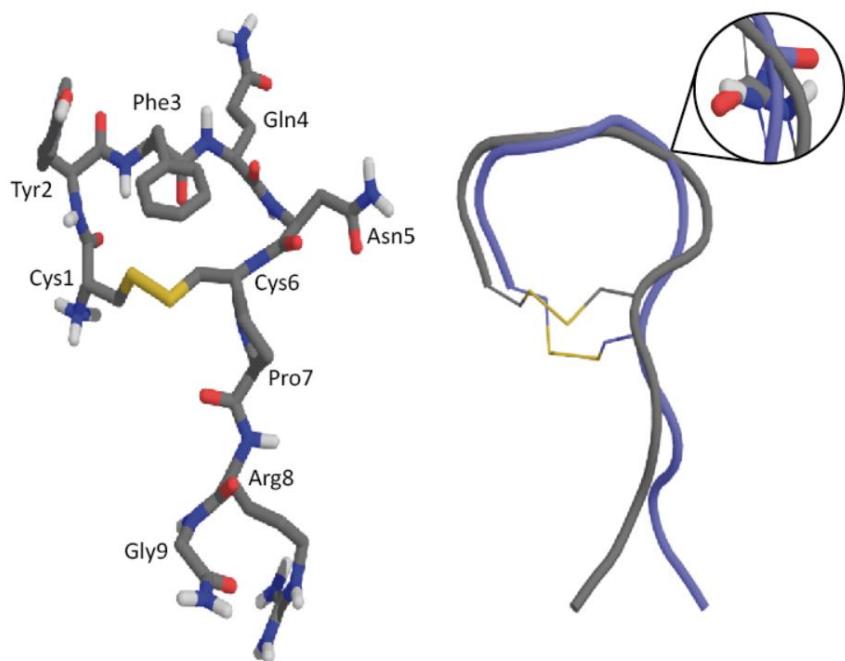


Figure S1 Representative for the ring-state type *variants* (grey). Left: stick depiction of *variants*. Right: cartoon depiction of *variants* and ring alignment with the representative for *clinched open* (blue).

Variants is a 4,5 peptide-bond rotamer of *clinched open* as illustrated in the zoom. The conformation *variants* occurred for the first time at the end of 23 μ s MD simulation of Arg⁸-vasopressin and is expected to be populated insignificantly in aqueous solution according to thermodynamic calculations.

Cartesian coordinates of the B3LYP/6-31G(d) optimized geometries are given below as Gaussian Archive Entries.

Metadynamics simulations

We used a combination of metadynamics,^{12, 13} in its well-tempered variant (WT),¹⁴ the multiple-walker technique¹⁵ and the path collective variable (PCV)^{16, 17} to determine the free-energy differences between AVP conformations (identified from the 23 μs trajectory) in water. Four main ring conformers identified previously¹⁰ and one new found in the extended 23 μs MD simulation were used as starting geometries for the metadynamics simulations. The PCV used was a numerical assignment to the most similar AVP conformer based on the RMSD of the backbone atoms of the ring residues. An analysis of the unbiased 23 μs MD trajectory using this PCV showed that 90% of the frames can be uniquely assigned to one of the five ring conformers. We were therefore able to use this single PCV for the metadynamics simulation.

The simulation boxes and topologies used for the AMBER MD simulations were converted to GROMACS¹⁸ format. The simulation configuration used the same water model, temperature and thermodynamic ensemble as the reference unbiased simulation. Particle mesh Ewald (PME) was used to treat electrostatic interactions, using a cut-off distance of 1.0 nm and each of the five models were equilibrated for 20 ns. Gaussian hills with an initial height of 0.6 kcal mol⁻¹ applied every ps were used. The Gaussian functions were rescaled in the WT scheme using a bias factor of 10. The metadynamics simulations were performed using GROMACS with the PLUMED plug-in.¹⁹

Calculation of chemical shifts and correlation with experimental values

Representative structures were taken from the 23 μ s MD simulation and optimized using the B3LYP hybrid density functional,²⁰ the 6-31G(d) basis set²¹ and the default polarizable continuum model (PCM) for water solvent using Gaussian09.²² These optimized structures were then used to calculate ¹H and ¹³C chemical shifts using the GIAO formalism,²³ again with PCM-water. The chemical shifts were obtained from the calculated magnetic shielding using the correlation formulae given in the main text. These formulae were obtained by linear regression of a training set of experimental chemical shifts²⁴ and the corresponding shielding values at level B3LYP/6-31G* with PCM-water (Figure S2).

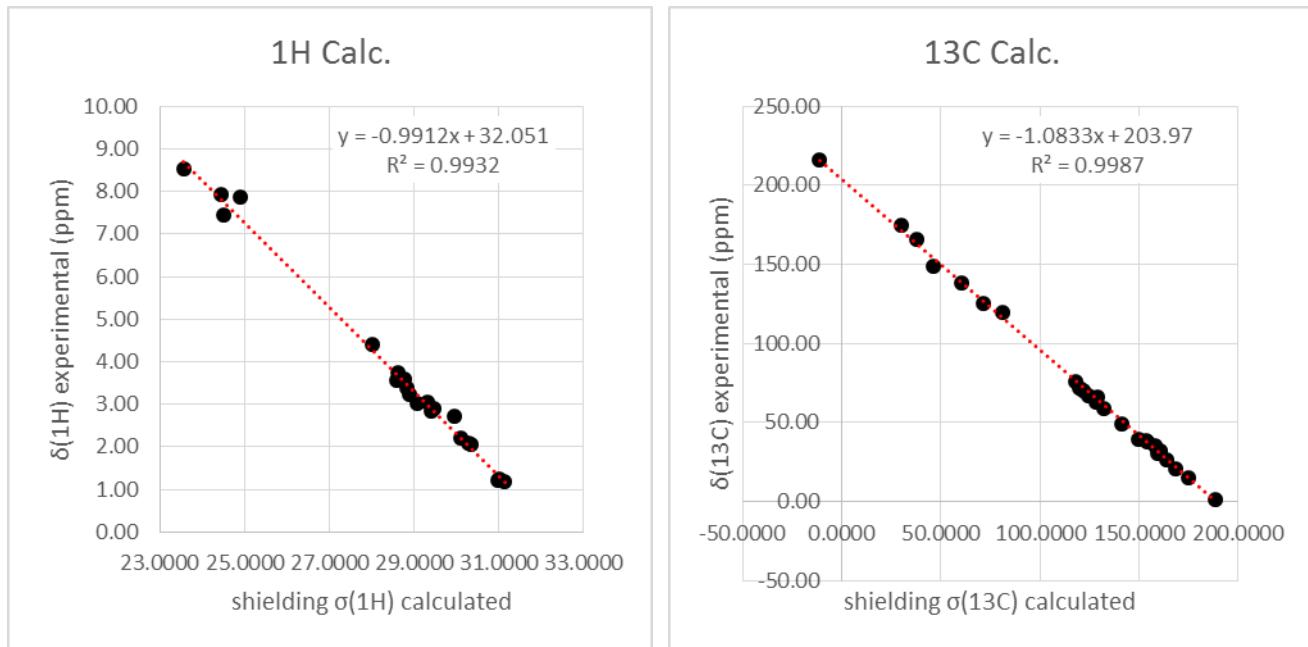


Figure S2 Linear regression of magnetic shielding at level B3LYP/6-31G(d) with PCM water and a training set of experimental chemical shifts (ppm) for ¹H and ¹³C

Table S3-4 show the calculated ¹³C and ¹H chemical shifts (δ ppm) for the individual conformations and the equilibrium mixtures (Eq. 1-4) calculated from the metadynamics free-energy differences, both assuming a single (*extended*) tail conformation and using the *extended:folded* equilibrium determined from the 23 μ s MD simulation for each representative conformation.

$$\delta_{saddle_{eq.}} = 0.7314 \times \delta_{saddle_{ext}} + 0.2686 \times \delta_{saddle_{fold}} \quad (1)$$

$$\delta_{cl.open_{eq.}} = 0.6263 \times \delta_{cl.open_{ext}} + 0.3737 \times \delta_{cl.open_{fold}} \delta_{open} \quad (2)$$

$$\delta_{equilibrium_{ext}} = 0.6865 \times \delta_{saddle_{ext}} + 0.2951 \times \delta_{cl.open_{ext}} + 0.0043 \times \delta_{tw.saddle} + 0.0141 \times \delta_{open} \quad (3)$$

$$\delta_{equilibrium_{eq.}} = 0.6865 \times (0.7314 \times \delta_{saddle_{ext}} + 0.2686 \times \delta_{saddle_{fold}}) + 0.2951 \times (0.6263 \times \delta_{cl.open_{ext}} + 0.3737 \times \delta_{cl.open_{fold}}) + 0.0043 \times \delta_{tw.saddle} + 0.0141 \times \delta_{open} \quad (4)$$

Table S3 Calculated ^{13}C chemical shifts (B3LYP/6-31G(d), PCM water) of Arg⁸-vasopressin

Calculated ^{13}C chemical shifts (ppm)													
Individual conformations													
res	atom	<i>Saddle</i>			<i>Clinched open</i>			<i>Twisted saddle</i>		<i>Open</i>		Metadynamics equilibrium	
		<i>ext</i>	<i>fold</i>	<i>equil^a</i>	<i>ext</i>	<i>fold</i>	<i>equil^b</i>	<i>ext</i>	<i>ext</i>	<i>ext</i>	<i>ext^c</i>	<i>Equilibrium</i>	<i>Equilibrium^d</i>
Cys ¹	C ^α	57.90	57.27	57.73	56.81	58.08	57.28	57.23	58.08	57.58	57.58	57.60	
Cys ¹	C ^β	43.53	43.38	43.49	49.46	49.98	49.66	46.52	45.04	45.32	45.32	45.34	
Tyr ²	C ^α	63.12	63.82	63.30	65.73	65.80	65.76	58.02	63.64	63.87	63.87	64.01	
Tyr ²	C ^β	37.72	39.58	38.22	40.42	40.46	40.44	39.97	42.43	38.59	38.59	38.94	
Tyr ²	C ^{δ¹}	133.17	131.80	132.80	134.21	134.62	134.37	135.89	134.23	133.51	133.51	133.30	
Tyr ²	C ^{δ²}	131.08	133.13	131.63	133.02	132.52	132.83	133.95	133.37	131.70	131.70	132.02	
Tyr ²	C ^{ε¹}	118.14	117.08	117.85	115.46	114.56	115.13	115.79	115.83	117.31	117.31	117.01	
Tyr ²	C ^{ε²}	118.09	116.37	117.63	115.90	115.61	115.79	117.39	116.41	117.42	117.42	117.07	
Phe ³	C ^α	62.93	62.34	62.77	54.22	56.81	55.19	62.29	62.98	60.36	60.36	60.54	
Phe ³	C ^β	37.98	38.02	37.99	41.37	44.82	42.66	40.51	39.87	39.02	39.02	39.40	
Phe ³	C ^{δ¹}	131.31	131.04	131.23	134.45	133.75	134.19	131.16	130.93	132.23	132.23	132.10	
Phe ³	C ^{δ²}	131.63	131.33	131.55	133.42	131.86	132.84	133.24	134.32	132.20	132.20	131.98	
Phe ³	C ^{ε¹}	131.22	131.40	131.27	130.95	131.08	131.00	131.95	130.27	131.13	131.13	131.18	
Phe ³	C ^{ε²}	131.67	131.34	131.58	130.01	131.21	130.46	131.21	129.58	131.15	131.15	131.22	
Phe ³	C ^ζ	129.93	129.83	129.90	129.57	128.92	129.32	128.48	128.24	129.79	129.79	129.70	
Gln ⁴	C ^α	57.93	56.76	57.61	61.15	58.78	60.26	61.77	55.39	58.86	58.86	58.38	
Gln ⁴	C ^β	30.23	27.62	29.53	28.74	27.92	28.44	25.60	40.73	29.92	29.92	29.35	
Gln ⁴	C ^γ	34.65	29.18	33.18	29.41	29.45	29.42	33.47	33.86	33.09	33.09	32.08	
Asn ⁵	C ^α	58.15	56.35	57.67	53.20	52.10	52.79	53.70	53.33	56.61	56.61	56.15	
Asn ⁵	C ^β	41.48	43.06	41.90	36.42	35.29	36.00	42.79	40.79	39.98	39.98	40.15	
Cys ⁶	C ^α	55.48	54.06	55.10	53.99	55.27	54.47	55.96	52.63	55.00	55.00	54.88	
Cys ⁶	C ^β	45.80	49.45	46.78	50.39	48.02	49.50	46.15	52.71	47.25	47.25	47.67	
Pro ⁷	C ^α	66.11	66.08	66.10	66.59	66.73	66.64	65.76	66.07	66.25	66.25	66.26	
Pro ⁷	C ^β	31.87	32.15	31.95	31.93	32.19	32.03	31.61	31.67	31.88	31.88	31.96	
Pro ⁷	C ^γ	26.37	27.70	26.73	27.51	27.54	27.52	26.92	26.81	26.72	26.72	26.97	
Pro ⁷	C ^δ	51.28	51.17	51.25	50.72	51.23	50.91	49.90	50.72	51.10	51.10	51.13	
Arg ⁸	C ^α	56.93	58.60	57.38	56.10	58.71	57.08	56.21	55.59	56.66	56.66	57.26	
Arg ⁸	C ^β	36.06	32.65	35.14	36.20	33.57	35.22	36.46	35.94	36.10	36.10	35.18	
Arg ⁸	C ^γ	30.23	26.10	29.12	28.70	27.17	28.13	27.37	28.20	29.74	29.74	28.81	
Arg ⁸	C ^δ	44.58	44.90	44.66	46.36	45.84	46.16	44.45	46.34	45.13	45.13	45.13	
Gly ⁹	C ^α	42.47	44.61	43.04	42.59	46.31	43.98	42.80	42.54	42.51	42.51	43.31	

^aEq. 1; ^bEq. 2; ^cEq. 3; ^dEq. 4; Abbreviations: extended (ext); folded (fold); equilibrium (equil)

Table S4 Calculated ^1H chemical shifts (B3LYP/6-31G(d), PCM water) of Arg⁸-vasopressin

Calculated ^1H chemical shifts (ppm)													
Individual conformations													
res	atom	Saddle			Clinched open			Twisted saddle		Open		Metadynamics equilibrium	
		ext	fold	equil ^a	ext	fold	equil ^b	ext	ext	ext	ext	Equilibrium	Equilibrium
Cys ¹	H ^{α}	3.87	3.91	3.88	3.63	4.26	3.86	4.40	4.10	3.80	3.88		
Cys ¹	H ^{βa}	3.39	3.69	3.47	3.37	2.65	3.10	2.84	3.10	3.38	3.35		
Cys ¹	H ^{βb}	3.05	2.74	2.97	3.32	4.20	3.65	3.49	2.85	3.13	3.17		
Tyr ²	H ^{α}	4.52	4.26	4.45	3.95	3.81	3.90	4.87	4.40	4.35	4.29		
Tyr ²	H ^{βa}	3.14	3.20	3.16	3.11	2.87	3.02	3.12	3.12	3.13	3.12		
Tyr ²	H ^{βb}	3.99	2.75	3.66	2.95	2.64	2.84	2.75	2.98	3.66	3.40		
Tyr ²	H ^{δ1}	7.37	7.24	7.33	7.33	5.73	6.73	7.48	7.18	7.36	7.15		
Tyr ²	H ^{δ2}	7.25	6.95	7.17	7.25	7.09	7.19	7.54	7.37	7.25	7.18		
Tyr ²	H ^{ϵ1}	7.10	6.41	6.91	6.72	6.32	6.57	6.74	6.71	6.98	6.81		
Tyr ²	H ^{ϵ2}	6.52	6.42	6.49	6.56	6.49	6.54	6.94	6.73	6.54	6.51		
Phe ³	H ^{α}	4.64	5.02	4.75	4.48	4.71	4.57	4.03	3.52	4.58	4.67		
Phe ³	H ^{βa}	3.68	3.71	3.69	3.12	2.62	2.94	2.72	2.47	3.49	3.44		
Phe ³	H ^{βb}	2.73	2.74	2.73	3.19	3.20	3.19	2.63	3.90	2.88	2.88		
Phe ³	H ^{δ1}	7.08	7.10	7.09	7.15	7.41	7.25	7.49	7.10	7.11	7.14		
Phe ³	H ^{δ2}	7.07	7.17	7.09	7.57	7.56	7.57	7.26	7.61	7.23	7.24		
Phe ³	H ^{ϵ1}	7.37	7.42	7.38	7.66	7.56	7.63	7.56	7.34	7.46	7.45		
Phe ³	H ^{ϵ2}	7.43	7.47	7.44	7.52	7.60	7.55	7.51	7.30	7.46	7.47		
Phe ³	H ^{ζ}	7.39	7.40	7.39	7.48	7.64	7.54	7.44	7.24	7.41	7.43		
Gln ⁴	H ^{α}	4.45	4.79	4.54	4.12	4.27	4.17	3.28	4.26	4.34	4.42		
Gln ⁴	H ^{βa}	2.05	1.38	1.87	2.60	1.55	2.21	1.76	1.25	2.20	1.96		
Gln ⁴	H ^{βb}	2.26	2.52	2.33	1.84	2.40	2.05	2.37	2.37	2.14	2.25		
Gln ⁴	H ^{γa}	2.00	1.81	1.95	2.12	2.31	2.19	1.51	2.19	2.03	2.02		
Gln ⁴	H ^{γb}	2.25	2.20	2.23	2.23	2.38	2.29	0.50	2.66	2.24	2.25		
Asn ⁵	H ^{α}	4.73	4.93	4.78	4.71	4.61	4.68	4.97	5.10	4.73	4.76		
Asn ⁵	H ^{βa}	2.75	2.36	2.64	2.50	2.32	2.44	2.71	2.63	2.67	2.58		
Asn ⁵	H ^{βb}	2.63	2.84	2.68	3.31	2.84	3.14	2.39	2.50	2.83	2.81		
Cys ⁶	H ^{α}	5.09	5.05	5.08	5.03	4.72	4.91	4.66	4.43	5.06	5.02		
Cys ⁶	H ^{βa}	2.78	2.48	2.70	3.18	2.96	3.10	3.23	2.86	2.90	2.82		
Cys ⁶	H ^{βb}	2.97	3.19	3.03	3.41	3.41	3.41	3.72	3.34	3.11	3.15		
Pro ⁷	H ^{α}	4.16	4.07	4.14	4.20	4.00	4.12	4.19	4.16	4.17	4.13		
Pro ⁷	H ^{βa}	1.95	2.12	1.99	2.05	2.16	2.09	1.93	2.03	1.98	2.02		
Pro ⁷	H ^{βb}	2.23	2.37	2.27	2.37	2.36	2.37	2.23	2.34	2.27	2.30		
Pro ⁷	H ^{γa}	2.26	2.47	2.32	2.39	2.50	2.43	2.25	2.17	2.30	2.35		
Pro ⁷	H ^{γb}	2.03	2.09	2.05	2.11	2.12	2.11	1.99	2.01	2.06	2.07		
Pro ⁷	H ^{δa}	3.95	3.80	3.91	3.90	4.10	3.97	3.66	3.68	3.93	3.92		
Pro ⁷	H ^{δb}	3.79	3.69	3.76	3.74	4.14	3.89	3.54	3.60	3.77	3.80		
Arg ⁸	H ^{α}	4.31	3.86	4.19	4.33	3.80	4.13	4.41	4.32	4.32	4.18		
Arg ⁸	H ^{βa}	1.88	2.38	2.02	1.46	2.29	1.77	1.68	1.39	1.75	1.93		
Arg ⁸	H ^{βb}	2.51	1.71	2.29	2.21	1.98	2.13	2.05	2.20	2.41	2.24		
Arg ⁸	H ^{γa}	1.86	2.00	1.90	1.80	1.97	1.86	1.98	1.84	1.84	1.89		
Arg ⁸	H ^{γb}	2.19	2.28	2.22	2.06	2.23	2.12	2.19	1.83	2.15	2.18		
Arg ⁸	H ^{δa}	3.38	3.17	3.32	3.40	3.55	3.45	3.16	3.28	3.38	3.36		

Arg ⁸	H ^{δb}	3.27	3.09	3.22	3.33	3.18	3.27	3.32	3.40	3.29	3.24
Gly ⁹	H ^{αa}	4.37	4.57	4.43	4.43	3.28	4.00	4.38	4.39	4.39	4.30
Gly ⁹	H ^{αb}	3.26	4.34	3.55	3.29	3.81	3.48	3.27	3.24	3.27	3.53
<hr/>											
Tyr ²	H ^N	6.03	5.52	5.90	5.21	5.48	5.31	8.92	6.62	5.81	5.75
Phe ³	H ^N	4.93	5.17	4.99	5.40	6.28	5.73	4.90	5.90	5.08	5.22
Gln ⁴	H ^N	4.97	4.62	4.88	5.26	5.30	5.28	5.59	6.03	5.08	5.02
Gln ⁴	H ^{ε21}	4.52	4.45	4.50	4.39	4.41	4.39	4.25	4.44	4.48	4.47
Gln ⁴	H ^{ε22}	5.18	4.82	5.08	4.92	4.81	4.88	4.48	6.34	5.12	5.04
Asn ⁵	H ^N	6.49	6.97	6.62	6.39	5.58	6.09	6.40	5.53	6.44	6.44
Asn ⁵	H ^{δ21}	4.54	4.40	4.50	4.50	4.40	4.47	4.86	5.03	4.54	4.50
Asn ⁵	H ^{δ22}	5.20	5.04	5.16	4.98	4.86	4.93	5.17	5.39	5.14	5.10
Cys ⁶	H ^N	6.98	7.32	7.07	5.98	6.36	6.12	6.85	6.12	6.67	6.78
Arg ⁸	H ^N	5.75	6.01	5.82	5.69	5.92	5.78	5.80	5.62	5.73	5.81
Arg ⁸	H ^ε	3.83	4.07	3.90	4.62	4.00	4.39	4.39	4.67	4.08	4.05
Gly ⁹	H ^N	4.88	6.60	5.35	4.99	7.12	5.79	4.97	4.95	4.92	5.47
(Gly ⁹)NH ₂	H ^{N1}	4.76	7.23	5.42	4.77	6.67	5.48	4.76	4.78	4.76	5.43
(Gly ⁹)NH ₂	H ^{N2}	4.43	4.88	4.55	4.43	4.29	4.38	4.41	4.43	4.43	4.50

^aEq. 1; ^bEq. 2; ^cEq. 3; ^dEq. 4; Abbreviations: extended (ext); folded (fold); equilibrium (equil)

Calculation of interatomic distances and correlation with experimental values

To calculate interatomic distances, the longest sections of the 23 μs MD trajectory of Arg⁸-vasopressin that were occupied entirely by a distinct ring state were chosen (278 ns *saddle_{ext}*; 212 ns *saddle_{fold}*; 136 ns *clinched open_{ext}*; 67 ns *clinched open_{fold}*; 191 ns *twisted saddle* and 220 ns *open*). The individual distance-trajectories corresponding to the experimental NOE distances were extracted from each representative MD-section. The equilibrium distances are calculated as the 1/6 power means of the distances within each state, weighted according to the distribution given by the metadynamics simulations (Eq. 5-8).

$$r_{equilibrium_{ext}} = (0.6865 \times r_{saddle_{ext}}^{1/6} + 0.2951 \times r_{cl.open_{ext}}^{1/6} + 0.0043 \times r_{tw.saddle}^{1/6} + 0.0141 \times r_{open}^{1/6})^6 \quad (5)$$

$$r_{equilibrium_{eq.}} = [0.6865 \times (0.7314 \times r_{saddle_{ext}}^{1/6} + 0.2686 \times r_{saddle_{fold}}^{1/6}) + 0.2951 \times (0.6263 \times r_{cl.open_{ext}}^{1/6} + 0.3737 \times r_{cl.open_{fold}}^{1/6}) + 0.0043 \times r_{tw.saddle}^{1/6} + 0.0141 \times r_{open}^{1/6}]^6 \quad (6)$$

The distances for the main states (*saddle* and *clinched open*) were refined by taking the relative populations of *extended* and *folded* tail conformations (Eq. 5-6) into consideration.

$$r_{saddle_{eq.}} = (0.7314 \times r_{saddle_{ext}}^{1/6} + 0.2686 \times r_{saddle_{fold}}^{1/6})^6 \quad (7)$$

$$r_{cl.open_{eq.}} = (0.6263 \times r_{cl.open_{ext}}^{1/6} + 0.3737 \times r_{cl.open_{fold}}^{1/6})^6 \quad (8)$$

The results of the statistical analysis of the correlation between calculated and experimental distances are given in the main text. Figure S3 shows the plot of calculated vs. experimental interatomic distances at pH 6.0 and pH 4.7 and calculated distances are listed in Table S5. Mean unsigned errors (MUE) and root mean square deviations (RMSD) are the same order of magnitude as the experimental error limits (pH 6.0 \pm 0.5 Å, pH 4.7 \pm 0.7 Å) for all individual conformations and equilibrium distances calculated from metadynamics. The results are discussed in the main text. At pH 6.0, the number of experimental distances is decreased due to proton exchange. As long as only a small number of experimental distances are available and if the experimental error limits are relatively higher than the differences between representative conformations, the linear regression remains insignificant.

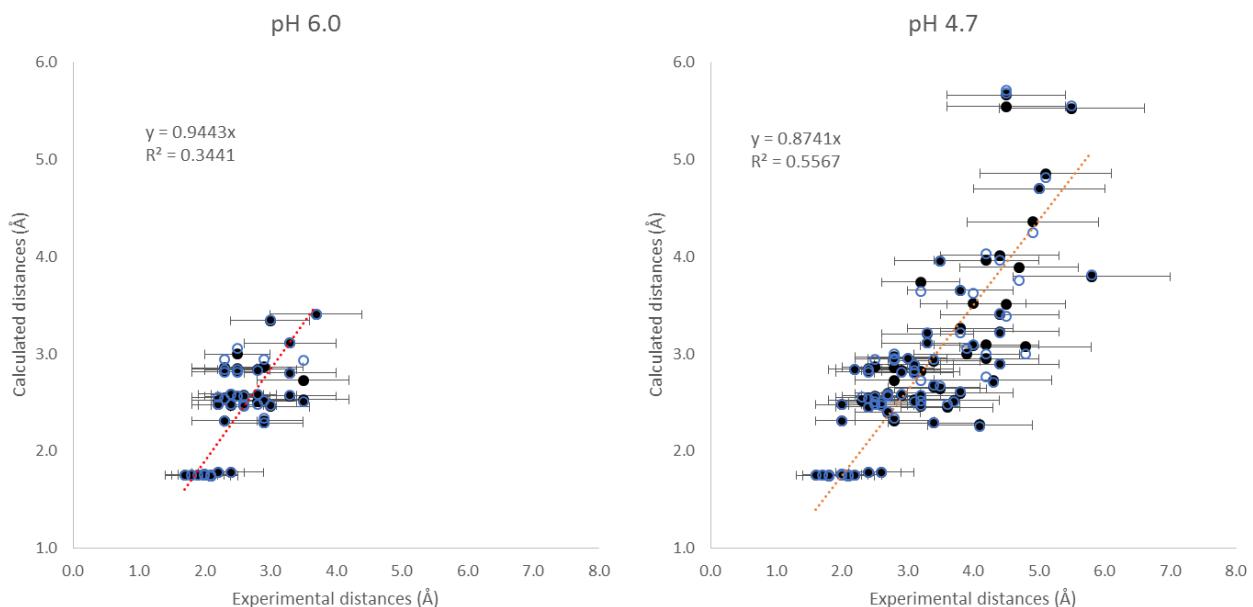


Figure S3 Linear regression of calculated equilibrium and experimental NOE distances at pH 6.0 (left) and pH 4.7 (right). Open blue circles indicate the equilibrium conformations with extended tail. The arrow bars show the error limits of the experimental NOE constraints.

Table S5 Calculated (r^6 weighted) interatomic distances of conformations of Arg⁸-vasopressin in solution (TIP4PEw water)

res	atom	res	atom	Interatomic distances (Å)				
				Individual conformations			Metadynamics equilibrium	
				Saddle ^a equilibrium	Clinched open ^b equilibrium	Twisted saddle extended	Open extended	Equilibrium ^c extended
Cys ¹	H ^α	Cys ¹	H ^{βa}	2.49	2.66	2.61	2.98	2.53
Cys ¹	H ^α	Cys ¹	H ^{βb}	2.94	2.69	2.55	2.51	2.94
Cys ¹	H ^{βa}	Cys ¹	H ^{βb}	1.76	1.75	1.75	1.75	1.76
Cys ¹	H ^α	Tyr ²	H	2.26	2.28	2.33	2.30	2.26
Cys ¹	H ^α	Phe ³	H	6.30	4.00	6.39	6.16	5.55
Cys ¹	H ^{βa}	Cys ⁶	H	5.44	5.88	4.76	4.30	5.68
Cys ¹	H ^{βa}	Cys ⁶	H ^α	5.94	5.09	4.95	5.22	5.71
Tyr ²	H	Tyr ²	H ^α	2.88	2.93	2.89	2.93	2.89
Tyr ²	H ^α	Tyr ²	H ^{βa}	2.53	2.50	2.49	2.59	2.56
Tyr ²	H ^α	Tyr ²	H ^{βb}	2.90	2.64	3.01	2.78	2.81
Tyr ²	H ^α	Tyr ²	H ^{δ*(H^{δ1})}	4.03	3.83	4.25	3.46	4.03
Tyr ²	H ^{βa}	Tyr ²	H ^{βb}	1.75	1.74	1.75	1.75	1.75
Tyr ²	H ^{βa}	Tyr ²	H ^{δ*(H^{δ1})}	3.24	3.22	3.57	2.91	3.22
Tyr ²	H ^{βb}	Tyr ²	H ^{δ*(H^{δ1})}	2.66	2.88	2.41	3.17	2.70
Tyr ²	H ^{δ1}	Tyr ²	H ^{ε*(H^{ε1})}	2.48	2.47	2.48	2.49	2.48
Tyr ²	H	Phe ³	H	4.63	2.30	4.57	4.36	3.81
Tyr ²	H ^α	Phe ³	H	2.28	3.38	2.38	2.20	2.57
Tyr ²	H ^{βa}	Phe ³	H	3.97	4.12	3.90	3.94	3.96
Phe ³	H	Phe ³	H ^α	2.77	2.91	2.78	2.23	2.80
Phe ³	H	Phe ³	H ^{βa}	3.18	2.89	2.49	3.30	3.00
Phe ³	H	Phe ³	H ^{βb}	2.47	2.88	2.48	3.78	2.62
Phe ³	H ^α	Phe ³	H ^{βa}	2.40	2.57	2.45	2.77	2.47
Phe ³	H ^α	Phe ³	H ^{βb}	2.60	2.78	2.98	2.66	2.68
Phe ³	H ^α	Phe ³	H ^{δ*(H^{δ1})}	4.04	3.59	3.18	3.41	3.75
Phe ³	H ^{βa}	Phe ³	H ^{βb}	1.74	1.75	1.75	1.75	1.74
Phe ³	H ^{βa}	Phe ³	H ^{δ*(H^{δ1})}	2.95	3.01	2.88	3.06	3.00
Phe ³	H ^{βb}	Phe ³	H ^{δ*(H^{δ1})}	3.10	3.05	3.28	3.00	3.10
Phe ³	H ^{δ1}	Phe ³	H ^{ε*(H^{ε1})}	2.47	2.47	2.47	2.48	2.47
Phe ³	H ^{ε1}	Phe ³	H ^ζ	2.48	2.48	2.48	2.48	2.48
Phe ³	H ^α	Gln ⁴	H	3.49	2.38	2.15	3.06	3.11
Phe ³	H ^{βa}	Gln ⁴	H	4.11	3.61	4.13	4.17	3.96
Phe ³	H ^{βb}	Gln ⁴	H	3.86	3.44	4.19	4.12	3.65
Gln ⁴	H	Gln ⁴	H ^α	2.92	2.82	2.23	2.94	2.88
Gln ⁴	H	Gln ⁴	H ^{βa}	2.58	2.76	3.49	3.11	2.67
Gln ⁴	H	Gln ⁴	H ^{βb}	3.39	2.97	3.94	3.27	3.22
Gln ⁴	H	Gln ⁴	H ^{γ*(H^{γa})}	3.49	3.21	3.37	3.75	3.41
Gln ⁴	H ^α	Gln ⁴	H ^{βa}	2.89	2.70	2.95	2.68	2.81
Gln ⁴	H ^α	Gln ⁴	H ^{βb}	2.55	2.58	2.52	2.67	2.57
Gln ⁴	H ^α	Gln ⁴	H ^{γ*(H^{γa})}	2.90	3.24	2.68	3.12	3.05
Gln ⁴	H ^{βa}	Gln ⁴	H ^{βb}	1.76	1.75	1.76	1.76	1.76
Gln ⁴	H ^{βa}	Gln ⁴	H ^{γ*(H^{γa})}	2.84	2.86	2.92	2.94	2.84
Gln ⁴	H ^{ε1}	Gln ⁴	H ^{ε2}	1.75	1.75	1.75	1.75	1.75
Gln ⁴	H ^α	Cys ⁶	H	4.62	4.85	4.02	5.80	4.70

res	atom	res	atom	Interatomic distances (Å)					Metadynamics equilibrium		
				Individual conformations				Equilibrium ^c		Equilibrium ^d	
				Saddle ^a equilibrium	Clinched open ^b equilibrium	Twisted saddle extended	Open extended	extended	equilibrium	equilibrium	
Asn ⁵	H	Asn ⁵	H ^α	2.96	2.94	2.94	2.86	2.95	2.95	2.95	
Asn ⁵	H	Asn ⁵	H ^{β*} (H ^{βa})	2.86	3.10	2.81	2.63	2.94	2.94	2.93	
Asn ⁵	H ^α	Asn ⁵	H ^{β*} (H ^{βa})	2.54	2.49	2.50	2.49	2.52	2.52	2.52	
Asn ⁵	H ^{δ1}	Asn ⁵	H ^{δ2}	1.75	1.75	1.75	1.75	1.75	1.75	1.75	
Asn ⁵	H	Cys ⁶	H	2.17	3.49	2.49	4.30	2.51	2.51	2.53	
Asn ⁵	H ^α	Cys ⁶	H	3.53	2.58	3.32	2.26	3.22	3.22	3.21	
Cys ⁶	H	Cys ⁶	H ^α	2.97	2.92	2.94	2.93	2.95	2.95	2.95	
Cys ⁶	H	Cys ⁶	H ^{βa}	2.83	2.80	3.19	2.70	2.73	2.73	2.82	
Cys ⁶	H	Cys ⁶	H ^{βb}	3.68	3.17	3.25	3.65	3.62	3.62	3.52	
Cys ⁶	H ^α	Cys ⁶	H ^{βa}	2.93	2.72	2.61	2.96	2.94	2.94	2.86	
Cys ⁶	H ^α	Cys ⁶	H ^{βb}	2.44	2.70	2.57	2.54	2.48	2.48	2.51	
Cys ⁶	H ^{βa}	Cys ⁶	H ^{βb}	1.75	1.75	1.75	1.75	1.75	1.75	1.75	
Cys ⁶	H	Pro ⁷	H ^{δa}	4.78	5.03	5.02	4.96	4.81	4.81	4.86	
Cys ⁶	H	Pro ⁷	H ^{δb}	4.20	4.74	4.72	4.62	4.25	4.25	4.36	
Cys ⁶	H ^α	Pro ⁷	H ^{δa}	2.50	2.58	2.66	2.63	2.51	2.51	2.52	
Cys ⁶	H ^α	Pro ⁷	H ^{δb}	2.49	2.36	2.34	2.36	2.48	2.48	2.45	
Pro ⁷	H ^α	Pro ⁷	H ^{βa}	2.84	2.84	2.84	2.84	2.84	2.84	2.84	
Pro ⁷	H ^α	Pro ⁷	H ^{βb}	2.31	2.31	2.31	2.31	2.31	2.31	2.31	
Pro ⁷	H ^α	Pro ⁷	H ^{γ*}	3.66	3.65	3.65	3.65	3.65	3.65	3.65	
Pro ⁷	H ^{βa}	Pro ⁷	H ^{βb}	1.78	1.78	1.78	1.79	1.78	1.78	1.78	
Pro ⁷	H ^{βa}	Pro ⁷	H ^{γ*}	2.45	2.46	2.46	2.46	2.45	2.45	2.45	
Pro ⁷	H ^{βb}	Pro ⁷	H ^{γ*}	2.50	2.49	2.49	2.49	2.50	2.50	2.50	
Pro ⁷	H ^{γ*}	Pro ⁷	H ^{δ*}	2.29	2.29	2.29	2.29	2.29	2.29	2.29	
Pro ⁷	H ^{δb}	Pro ⁷	H ^{δb}	1.78	1.78	1.79	1.78	1.78	1.78	1.78	
Pro ⁷	H ^α	Arg ⁸	H	2.33	2.26	2.38	2.36	2.34	2.34	2.31	
Arg ⁸	H	Arg ⁸	H ^α	2.75	2.68	2.93	2.94	2.93	2.93	2.73	
Arg ⁸	H	Arg ⁸	H ^{βa}	2.94	2.98	2.77	2.75	2.77	2.77	2.95	
Arg ⁸	H	Arg ⁸	H ^{βb}	3.50	3.55	3.34	3.34	3.39	3.39	3.51	
Arg ⁸	H ^α	Arg ⁸	H ^{βa}	2.84	2.86	2.81	2.81	2.83	2.83	2.84	
Arg ⁸	H ^α	Arg ⁸	H ^{βb}	2.57	2.57	2.60	2.59	2.59	2.59	2.57	
Arg ⁸	H ^{βa}	Arg ⁸	H ^{βb}	1.75	1.75	1.76	1.76	1.75	1.75	1.75	
Arg ⁸	H ^{βa}	Arg ⁸	H ^{γ*}	2.57	2.56	2.57	2.58	2.56	2.56	2.57	
Arg ⁸	H ^{βb}	Arg ⁸	H ^{γ*}	2.60	2.59	2.60	2.61	2.59	2.59	2.60	
Arg ⁸	H ^{γ*}	Arg ⁸	H ^{δ*}	2.40	2.40	2.40	2.39	2.40	2.40	2.40	
Gly ⁹	H	Gly ⁹	H ^{α1,2}	2.49	2.50	2.50	2.50	2.50	2.50	2.50	
Gly ⁹	H ^{α1,2}	Gly ⁹	H ^{N1,2}	3.07	3.09	3.00	3.00	3.00	3.00	3.07	
Gly ⁹	H ^{N1}	Gly ⁹	H ^{N2}	1.75	1.75	1.75	1.75	1.75	1.75	1.75	

^aEq. 7; ^bEq. 8; ^cEq. 5; ^dEq. 6

Experimental details

Sample preparation for NMR

Arg⁸-vasopressin was obtained from Bachem (UK) Ltd as the trifluoroacetate salt of the chemically synthesized peptide, having a purity (by HPLC) of >96%. Mass spectrometry of the synthesized material gave a molecular mass of 1084.55 Da, in close agreement to the calculated molecular mass of 1086.26 Da for the reduced form of the peptide.

Samples of 5.0 mg dry weight were dissolved in 320 µl of 90% H₂O/ 10% D₂O to give a peptide concentration of 14.4 mM. The pH of the sample was measured to be 4.7 and NMR spectra were recorded without adjustment. In addition the sample was dried by lyophilization, then redissolved in 320 µl of 20 mM potassium phosphate buffer (pH 6.5) in 90% H₂O/ 10% D₂O and NMR spectra were recorded at a pH measured as 6.0. NMR spectra of Arg⁸-vasopressin in D₂O at both pH 4.7 and pH 6.0 were recorded at least 2h after redissolving the extensively dried samples in 99.9% D₂O (Sigma Aldrich).

NMR experiments

NMR spectroscopy was performed on a Varian Inova 600 MHz spectrometer, equipped with 5-channels, a 5 mm triple resonance (¹H/¹³C/¹⁵N) coldprobe and actively shielded pulse field z-axis gradients.

Proton resonance assignments were achieved using a combination of 2D ¹H-¹H total chemical shift correlation spectroscopy (TOCSY),²⁵ and ¹H-¹H nuclear Overhauser effect spectroscopy (NOESY) NMR spectra.²⁶ Spectra were acquired as 2048 complex points, with 32 transients for each of 512 increments and a spectral width of 10.0 ppm in both dimensions. Mixing times of 60 and 75 ms for the TOCSY experiment and 200 and 300 ms for the NOESY experiment were used. Water suppression was achieved through use of the watergate 3919 sequence.²⁷

Resonance assignments for carbon and nitrogen at natural abundance were obtained through the use of gradient heteronuclear single quantum coherence (gHSQC) experiments. A standard ¹³C-¹H gHSQC NMR spectrum,^{28, 29} was acquired as 1024 complex points in t2 (observe ¹H dimension) and 280 increments in t1 (indirect ¹³C dimension) using 32 transients over spectral widths of 6000.60 Hz (10.0 ppm) and 21114.68 Hz (140.0 ppm) respectively. The transmitter offsets were initially set to the water position in the ¹H and to 70 ppm in the ¹³C dimension, but other combinations of offset and sweep width were later used to focus on the aliphatic and aromatic regions. A 2D sensitivity enhanced ¹⁵N-¹H gHSQC NMR spectrum^{28, 30, 31} was acquired as 2048 complex points in t2 (observe ¹H dimension) and 128 increments in t1 (indirect ¹⁵N dimension) using 32 transients over spectral widths of 6000.60 Hz (10.0 ppm) and 2431.06 Hz (40.0 ppm) respectively. The transmitter offsets were set to the water position in the ¹H and to 120 ppm in the ¹⁵N dimension. States-TPPI quadrature detection was employed in the ¹⁵N-dimension.³²

Spectral processing and format conversion was performed using NMRPipe³³ and visualized with NMRView³⁴. Arg⁸-vasopressin spectra were assigned using Analysis v2.0.7 from the CcpNMR software suite.³⁵ Proton and ¹³C chemical shifts were referenced to 3-trimethyl silyl propane sulfonic acid (DSS) and ¹⁵N chemical shifts were referenced to an external liquid ammonia. The ¹H, ¹³C and ¹⁵N chemical shifts of the major populated *trans*-Pro⁷ isomer of Arg⁸-vasopressin in H₂O/pH 4.7, D₂O/pH 4.7, H₂O/pH 6.0, D₂O/pH 6.0 are given in Table S6-9. The volumes of assigned peaks were determined using the box sum method in Analysis with an r⁶ distance calibration against the fixed distance between the Tyr² H⁶ and H^ε atoms. A 20% change (the default) in the calculated target distance was taken. These experimentally derived distances are listed in Table S10.

Experimental ^1H , ^{13}C and ^{15}N chemical shifts

Table S6 Experimental NMR chemical shifts (δ ppm) of Arg⁸-vasopressin in H₂O at pH 6.0/ 298 K

Residue	H ^N	N	H ^{α}	C ^{α}	Others
Cys ¹	-	-	3.97	56.18	3.27:H ^{βa} ; 3.12:H ^{βb} 44.23:C ^{β}
Tyr ²	8.57 [?]	123.79 [?]	4.64	58.02 [?]	2.80:H ^{βa} ; 2.96:H ^{βb} ; 7.05:H ^{δ*} ; 6.83:H ^{ϵ*} 39.10:C ^{β} ; 133.34:C ^{δ*} ; 118.46:C ^{ϵ*}
Phe ³	8.04	122.55 [?]	4.54	58.37	3.01:H ^{βa} ; 3.31:H ^{βb} ; 7.23:H ^{δ*} ; 7.40:H ^{ϵ*} ; 7.37:H ^{ζ} 39.42:C ^{β} ; 131.99:C ^{δ*} ; 131.81:C ^{ϵ*} ; 130.17:C ^{ζ}
Gln ⁴	8.32	120.80	4.12	57.82	2.05:H ^{βa} ; 2.12:H ^{βb} ; 2.29:H ^{γ*} ; 6.89:H ^{ϵa} ; 7.53:H ^{ϵb} 28.66:C ^{β} ; 33.99:C ^{γ} ; 114.24:N ^{ϵ}
Asn ⁵	8.30	118.20	4.77	-	2.86:H ^{β*} ; 6.92:H ^{δa} ; 7.63:H ^{δb} 38.74:C ^{β} ; 114.53:N ^{δ}
Cys ⁶	8.15	122.39	4.89	-	3.18:H ^{βa} ; 2.93:H ^{βb} 41.69:C ^{β}
trans-Pro ⁷	-	-	4.45	63.52	1.93:H ^{βa} ; 2.31:H ^{βb} ; 2.06:H ^{γ*} ; 3.73:H ^{δa} ; 3.83:H ^{δb} 32.16:C ^{β} ; 27.60:C ^{γ} ; 50.79:C ^{δ}
Arg ⁸	8.63	123.97	4.32	56.45	1.80:H ^{βa} ; 1.90:H ^{βb} ; 1.67:H ^{γ*} ; 3.22:H ^{δ*} ; 7.22:H ^{ϵ} 30.76:C ^{β} ; 27.28:C ^{γ} ; 43.46:C ^{δ} ; 86.76:N ^{ϵ}
Gly ⁹	8.45	113.01	3.93*	45.06	
NH ₂ ¹⁰	-	-	-	-	7.09:H ^{N1} ; 7.48:H ^{N2} ; 109.16:N

[?]tentative assignment; *degenerate atoms

Table S7 Experimental NMR chemical shifts (δ ppm) of Arg⁸-vasopressin in D₂O at pH 6.0/ 298 K

Residue	H ^N	N	H ^{α}	C ^{α}	Others
Cys ¹	-	-	3.98	56.01	3.28:H ^{βa} ; 3.12:H ^{βb} 44.20:C ^{β}
Tyr ²	-	-	4.64	58.02	2.81:H ^{βa} ; 2.97:H ^{βb} ; 7.06:H ^{δ*} ; 6.83:H ^{ϵ*} 39.11:C ^{β} ; 133.35:C ^{δ*} ; 118.38:C ^{ϵ*}
Phe ³	8.05	-	4.54	58.33	3.01:H ^{βa} ; 3.31:H ^{βb} ; 7.23:H ^{δ*} ; 7.40:H ^{ϵ*} ; 7.37:H ^{ζ} 39.37:C ^{β} ; 131.99:C ^{δ*} ; 131.81:C ^{ϵ*} ; 130.18 C ^{ζ}
Gln ⁴	-	-	4.12	57.75	2.05:H ^{βa} ; 2.13:H ^{βb} ; 2.29:H ^{γ*} ; 6.89:H ^{ϵa} ; 7.53:H ^{ϵb} 28.60:C ^{β} ; 33.93:C ^{γ}
Asn ⁵	8.31	-	4.78	53.08	2.86:H ^{β*} ; 6.92:H ^{δa} ; 7.63:H ^{δb} 38.66:C ^{β}
Cys ⁶	8.15	-	4.90	54.27	3.19:H ^{βa} ; 2.93:H ^{βb} 41.67:C ^{β}
trans-Pro ⁷	-	-	4.46	63.51	1.94:H ^{βa} ; 2.32:H ^{βb} ; 2.07:H ^{γ*} ; 3.74:H ^{δa} ; 3.84:H ^{δb} 32.16:C ^{β} ; 27.61:C ^{γ} ; 50.80:C ^{δ}
Arg ⁸	8.62	-	4.32	56.36	1.81:H ^{βa} ; 1.91:H ^{βb} ; 1.68:H ^{γ*} ; 3.23:H ^{δ*} 30.72:C ^{β} ; 27.28:C ^{γ} ; 43.34:C ^{δ}
Gly ⁹	8.46	-	3.92*	44.93	
NH ₂ ¹⁰	-	-	-	-	-

*degenerate atoms

Table S8 Experimental NMR chemical shifts (δ ppm) of Arg⁸-vasopressin in H₂O at pH 4.7/ 298 K

Residue	H ^N	N	H ^a	C ^a	Others
Cys ¹	-	-	4.29	55.39	3.46:H ^{Ba} ; 3.25:H ^{Bb} 42.87:C ^B
Tyr ²	8.90	125.26	4.67	-	2.85:H ^{Ba} ; 2.95:H ^{Bb} ; 7.07:H ^{δ*} ; 6.84:H ^{ε*} 39.25:C ^B ; 133.36:C ^{δ*} ; 118.49:C ^{ε*}
Phe ³	8.15	123.04	4.48	58.63	3.01:H ^{Ba} ; 3.30:H ^{Bb} ; 7.22:H ^{δ*} ; 7.40:H ^{ε*} ; 7.38:H ^ζ 39.31:C ^B ; 131.96:C ^{δ*} ; 131.81:C ^{ε*} ; 130.17:C ^ζ
Gln ⁴	8.32	121.11	4.12	57.93	2.05:H ^{Ba} ; 2.13:H ^{Bb} ; 2.30:H ^{γ*} ; 6.90:H ^{εa} ; 7.53:H ^{εb} 28.64:C ^B ; 33.96:C ^γ ; 114.18:N ^ε
Asn ⁵	8.33	118.14	4.80	-	2.88:H ^{B*} ; 6.93:H ^{δa} ; 7.63:H ^{δb} 38.83:C ^B ; 114.55:N ^δ
Cys ⁶	8.14	122.09	4.92	-	3.21:H ^{Ba} ; 2.94:H ^{Bb} 41.28:C ^B
trans-Pro ⁷	-	-	4.46	63.48	1.94:H ^{Ba} ; 2.32:H ^{Bb} ; 2.07:H ^{γ*} ; 3.75:H ^{δa} ; 3.85:H ^{δb} 32.18:C ^B ; 27.60:C ^γ ; 50.80:C ^δ
Arg ⁸	8.65	124.06	4.32	56.50	1.81:H ^{Ba} ; 1.91:H ^{Bb} ; 1.69:H ^{γa} ; 1.68:H ^{γb} ; 3.23:H ^{δ*} ; 7.21:H ^ε 30.75:C ^B ; 27.28:C ^γ ; 43.46:C ^δ ; 86.76:N ^ε
Gly ⁹	8.44	112.97	3.94*	45.04	
NH ₂ ¹⁰	-	-	-	-	7.09:H ^{N1} ; 7.48:H ^{N2} ; 109.17:N

²tentative assignment; *degenerate atomsTable S9 Experimental NMR chemical shifts (δ ppm) of Arg⁸-vasopressin in D₂O at pH 4.7/ 298 K

Residue	H ^N	N	H ^a	C ^a	Others
Cys ¹	-	-	4.29	55.22	3.46:H ^{Ba} ; 3.25:H ^{Bb} 42.83:C ^B
Tyr ²	-	-	4.66	58.12	2.85:H ^{Ba} ; 2.96:H ^{Bb} ; 7.07:H ^{δ*} ; 6.84:H ^{ε*} 39.27:C ^B ; 133.37:C ^{δ*} ; 118.42:C ^{ε*}
Phe ³	-	-	4.47	58.59	3.02:H ^{Ba} ; 3.30:H ^{Bb} ; 7.21:H ^{δ*} ; 7.40:H ^{ε*} ; 7.37:H ^ζ 39.28:C ^B ; 131.96:C ^{δ*} ; 131.81:C ^{ε*} ; 130.18:C ^ζ
Gln ⁴	8.33	-	4.12	57.85	2.05:H ^{Ba} ; 2.13:H ^{Bb} ; 2.30:H ^{γ*} ; 6.90:H ^{εa} ; 7.54:H ^{εb} 28.58:C ^B ; 33.89:C ^γ
Asn ⁵	8.30	-	4.81	53.04	2.89:H ^{B*} ; 6.94:H ^{δa} ; 7.63:H ^{δb} 38.75:C ^B
Cys ⁶	-	-	4.92	53.97	3.21:H ^{Ba} ; 2.93:H ^{Bb} 41.26:C ^B
trans-Pro ⁷	-	-	4.46	63.49	1.94:H ^{Ba} ; 2.33:H ^{Bb} ; 2.07:H ^{γ*} ; 3.75:H ^{δa} ; 3.85:H ^{δb} 32.18:C ^B ; 27.61:C ^γ ; 50.81:C ^δ
Arg ⁸	8.65	-	4.31	56.41	1.82:H ^{Ba} ; 1.91:H ^{Bb} ; 1.68:H ^{γ*} ; 3.23:H ^{δ*} 30.71:C ^B ; 27.28:C ^γ ; 43.34:C ^δ
Gly ⁹	8.45	-	3.93*	44.91	
NH ₂ ¹⁰	-	-	-	-	7.48:H ^{N*}

*degenerate atoms

Experimental NOE distances

Table S10 Experimental NOE distances of Arg⁸-vasopressin

res	atom	res	atom	NOE distances (Å)					
				pH 4.7			pH 6.0		
				Constraint	Error limits		Constraint	Error limits	
				r _{exp}	+	-	r _{exp}	+	-
Cys ¹	H ^α	Cys ¹	H ^{βa}	2.4	0.5	0.5	2.3	0.5	0.5
Cys ¹	H ^α	Cys ¹	H ^{βb}	2.8	0.6	0.6	2.3	0.5	0.5
Cys ¹	H ^{βa}	Cys ¹	H ^{βb}	2.0	0.3	0.4	2.0	0.2	0.4
Cys ¹	H ^α	Tyr ²	H	4.1	0.8	0.8	-	-	-
Cys ¹	H ^α	Phe ³	H	5.5	1.1	1.1	-	-	-
Cys ¹	H ^{βa}	Cys ⁶	H	4.5	0.9	0.9	-	-	-
Cys ¹	H ^{βa}	Cys ⁶	H ^α	4.5	0.9	0.9	-	-	-
Tyr ²	H	Tyr ²	H ^α	4.4	0.9	0.9	-	-	-
Tyr ²	H ^α	Tyr ²	H ^{βa,b}	2.3	0.5	0.5	2.2	0.4	0.4
Tyr ²	H ^α	Tyr ²	H ^{βb,a}	2.4	0.5	0.5	2.3	0.5	0.5
Tyr ²	H ^α	Tyr ²	H ^{δ*}	4.2	0.8	0.8	-	-	-
Tyr ²	H ^{βa}	Tyr ²	H ^{βb}	1.8	0	0.4	2.1	0.3	0.4
Tyr ²	H ^{βa}	Tyr ²	H ^{δ*}	4.4	0.9	0.9	-	-	-
Tyr ²	H ^{βb}	Tyr ²	H ^{δ*}	4.3	0.9	0.9	-	-	-
Tyr ²	H ^{δ1}	Tyr ²	H ^{ε*}	2.5	0.5	0.5	2.6	0.5	0.5
Tyr ²	H	Phe ³	H	5.8	1.2	1.2	-	-	-
Tyr ²	H ^α	Phe ³	H	3.2	0.6	0.6	3.3	0.7	0.7
Tyr ²	H ^{βa}	Phe ³	H	4.4	0.9	0.9	-	-	-
Phe ³	H	Phe ³	H ^α	3.1	0.6	0.6	3.3	0.7	0.7
Phe ³	H	Phe ³	H ^{βa}	4.2	0.8	0.8	-	-	-
Phe ³	H	Phe ³	H ^{βb}	3.8	0.8	0.8	-	-	-
Phe ³	H ^α	Phe ³	H ^{βa}	3.2	0.6	0.6	3.0	0.6	0.6
Phe ³	H ^α	Phe ³	H ^{βb}	3.4	0.7	0.7	-	-	-
Phe ³	H ^α	Phe ³	H ^{δ*}	4.7	0.9	0.9	-	-	-
Phe ³	H ^{βa}	Phe ³	H ^{βb}	2.1	0.4	0.4	2.1	0.4	0.4
Phe ³	H ^{βa}	Phe ³	H ^{δ*}	2.8	0.6	0.6	-	-	-
Phe ³	H ^{βb}	Phe ³	H ^{δ*}	4.0	0.8	0.8	-	-	-
Phe ³	H ^{δ1}	Phe ³	H ^{ε*}	2.6	0.5	0.5	2.6	0.5	0.5
Phe ³	H ^{ε1}	Phe ³	H ^ζ	2.0	0.3	0.4	2.2	0.4	0.4
Phe ³	H	Gln ⁴	H	-	-	-	3.7	0.7	0.7
Phe ³	H ^α	Gln ⁴	H	3.3	0.7	0.7	3.3	0.7	0.7
Phe ³	H ^{βa}	Gln ⁴	H	3.5	0.7	0.7	-	-	-
Phe ³	H ^{βb}	Gln ⁴	H	3.2	0.6	0.6	-	-	-
Gln ⁴	H	Gln ⁴	H ^α	3.1	0.6	0.6	-	-	-
Gln ⁴	H	Gln ⁴	H ^{βa}	3.5	0.7	0.7	-	-	-
Gln ⁴	H	Gln ⁴	H ^{βb}	3.8	0.8	0.8	-	-	-
Gln ⁴	H	Gln ⁴	H ^{γ*} (H ^{γa})	4.4	0.9	0.9	-	-	-
Gln ⁴	H ^α	Gln ⁴	H ^{βa}	2.9	0.6	0.6	2.5	0.5	0.5
Gln ⁴	H ^α	Gln ⁴	H ^{βb}	2.7	0.5	0.5	2.5	0.5	0.5
Gln ⁴	H ^α	Gln ⁴	H ^{γ*} (H ^{γa})	3.9	0.8	0.8	2.5	0.5	0.5
Gln ⁴	H ^{βa}	Gln ⁴	H ^{βb}	2.1	0.3	0.4	-	-	-

res	atom	res	atom	NOE distances (Å)					
				pH 4.7			pH 6.0		
				Constraint	Error limits		Constraint	Error limits	
				r _{exp}	+	-	r _{exp}	+	-
Gln ⁴	H ^{βa}	Gln ⁴	H ^{γ*} (H ^{γa})	2.4	0.5	0.5	2.5	0.5	0.5
Gln ⁴	H ^{ε1}	Gln ⁴	H ^{ε2}	1.8	0.0	0.4	1.8	0.1	0.4
Gln ⁴	H ^α	Cys ⁶	H	5.0	1.0	1.0	-	-	-
Gln ⁴	H ^α	Asn ⁵	H	-	-	-	3.0	0.6	0.6
Asn ⁵	H	Asn ⁵	H ^α	3.0	0.6	0.6	-	-	-
Asn ⁵	H	Asn ⁵	H ^{β*}	3.4	0.7	0.7	-	-	-
Asn ⁵	H ^α	Asn ⁵	H ^{β*}	3.2	0.6	0.6	2.9	0.6	0.6
Asn ⁵	H ^{δ1}	Asn ⁵	H ^{δ2}	1.7	0.1	0.3	1.7	0	0.3
Asn ⁵	H	Cys ⁶	H	3.1	0.6	0.6	3.5	0.7	0.7
Asn ⁵	H ^α	Cys ⁶	H	3.3	0.7	0.7	-	-	-
Cys ⁶	H	Cys ⁶	H ^α	2.8	0.6	0.6	-	-	-
Cys ⁶	H	Cys ⁶	H ^{βa}	3.2	0.6	0.6	-	-	-
Cys ⁶	H	Cys ⁶	H ^{βb}	4.0	0.8	0.8	-	-	-
Cys ⁶	H ^α	Cys ⁶	H ^{βa}	2.5	0.5	0.5	2.9	0.6	0.6
Cys ⁶	H ^α	Cys ⁶	H ^{βb}	2.5	0.5	0.5	2.8	0.6	0.6
Cys ⁶	H ^{βa}	Cys ⁶	H ^{βb}	2.0	0.3	0.4	2.0	0.3	0.4
Cys ⁶	H	Pro ⁷	H ^{δa}	5.1	1.0	1.0	-	-	-
Cys ⁶	H	Pro ⁷	H ^{δb}	4.9	1.0	1.0	-	-	-
Cys ⁶	H ^α	Pro ⁷	H ^{δa}	3.7	0.7	0.7	-	-	-
Cys ⁶	H ^α	Pro ⁷	H ^{δb}	3.6	0.7	0.7	-	-	-
Pro ⁷	H ^α	Pro ⁷	H ^{βa}	2.2	0.4	0.4	2.3	0.5	0.5
Pro ⁷	H ^α	Pro ⁷	H ^{βb}	2.0	0.2	0.4	2.3	0.5	0.5
Pro ⁷	H ^α	Pro ⁷	H ^{γ*}	3.8	0.8	0.8	-	-	-
Pro ⁷	H ^{βa}	Pro ⁷	H ^{βb}	2.6	0.5	0.5	2.4	0.5	0.5
Pro ⁷	H ^{βa}	Pro ⁷	H ^{γ*}	2.4	0.5	0.5	2.4	0.5	0.5
Pro ⁷	H ^{βb}	Pro ⁷	H ^{γ*}	2.6	0.5	0.5	2.6	0.5	0.5
Pro ⁷	H ^{γ*}	Pro ⁷	H ^{δ*}	3.4	0.7	0.7	2.9	0.6	0.6
Pro ⁷	H ^{δb}	Pro ⁷	H ^{δb}	2.4	0.5	0.5	2.2	0.4	0.4
Pro ⁷	H ^α	Arg ⁸	H	2.8	0.6	0.6	2.9	0.6	0.6
Arg ⁸	H	Arg ⁸	H ^α	2.8	0.6	0.6	3.5	0.7	0.7
Arg ⁸	H	Arg ⁸	H ^{βa}	4.2	0.8	0.8	-	-	-
Arg ⁸	H	Arg ⁸	H ^{βb}	4.5	0.9	0.9	-	-	-
Arg ⁸	H ^α	Arg ⁸	H ^{βa}	3.1	0.6	0.6	2.8	0.6	0.6
Arg ⁸	H ^α	Arg ⁸	H ^{βb}	2.9	0.6	0.6	2.8	0.6	0.6
Arg ⁸	H ^{βa}	Arg ⁸	H ^{βb}	2.2	0.4	0.4	2.1	0.4	0.4
Arg ⁸	H ^{βa}	Arg ⁸	H ^{γ*}	2.5	0.5	0.5	2.4	0.5	0.5
Arg ⁸	H ^{βb}	Arg ⁸	H ^{γ*}	2.7	0.5	0.5	2.6	0.5	0.5
Arg ⁸	H ^{γ*}	Arg ⁸	H ^{δ*}	2.7	0.5	0.5	-	-	-
Gly ⁹	H	Gly ⁹	H ^{α1,2}	2.5	0.5	0.5	-	-	-
Gly ⁹	H ^{α1,2}	Gly ⁹	H ^{N1,2}	4.8	1.0	1.0	-	-	-
Gly ⁹	H ^{N1}	Gly ⁹	H ^{N2}	1.6	0.1	0.3	1.9	0.1	0.4

Gaussian archive entries for the B3LYP/6-31G(d)-optimized geometries.

Saddle, folded:

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Saddle, extended:

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Clinched open, folded:

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 -14.342446348, 10.4418269488, 14.2228168529\c, -12.7686729154, 9.704517182
 8, 12.9976678454\o, -12.5320672351, 8.6842689788, 13.6463701879\n, -12.3833
 909922, 9.8871476241, 11.7159987309\h, -12.6016210048, 10.722883909, 11.178
 6756797\h, -11.8827108197, 9.1376263655, 11.2579064837\h, \Version=ES64L-G09
 RevD.01\State=1-A\HF=-4313.5754649\RMSD=7.546e-09\RMSF=2.142e-06\Diopol
 e=-9.3182369, -3.6441124, -8.9227906\Quadrupole=207.4827637, -101.2244415
 ,-106.2583222, -49.7858048, 259.7104929, 20.5083893\PG=C01 [X(C46H67N1501
 2S2)]\\@\\

Clinched open, extended:

```
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pr-2015\0\#\ b3lyp/6-31g(d) opt name=clark scrfpcm\AVP_10us_T16_12_c
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4,-13.1779261134,1.8211119347\H,-10.137759484,-11.9119649439,2.7978609
187\H,-8.9856452812,-12.9745120635,3.3534778048\C,-8.2632699003,-11.69
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.4253666584\H,-6.1235026876,-11.6700761369,1.6497814328\S,-6.485045714
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,-1.9716805145\C,-7.661840712,-9.3504767688,-2.3978935499\H,-7.9734032
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Twisted saddle, extended:

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Open, extended:

1\1\FAU-CCC-CCDH173\FOpt\RB3LYP\6-31G(d)\C46H67N15O12S2(2+)\CLARK\24-A
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