## Supplementary Information for

# Plastic chip based magnetophoretic immunoassay for point-of-care diagnosis of tuberculosis 

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## Supplementary Methods

## Calculation of LOD

The LOD was calculated by the $3 \sigma$ criterion method ${ }^{1}$, where $\sigma$ stands for the standard deviation of negative control background data $(\mathrm{n}=5)$. In the analytical procedure, total signal $\left(\mathrm{S}_{\mathrm{t}}\right)$ can be explained by the sum of background signal $\left(\mathrm{S}_{\mathrm{b}}\right)$ and analyte signal $\left(\mathrm{S}_{\mathrm{x}}\right)$. And the analyte signal can be expressed as $\mathrm{gC}_{\mathrm{x}}$, where g is the sensitivity of instrument which is equal to the slope of calibration curve. According to this analytical method, LOD of signal ( $\mathrm{S}_{\text {LOD }}$ ) is calculated as equation S1,
$\mathrm{S}_{\mathrm{LOD}}=\mathrm{S}_{\mathrm{t}}-\mathrm{S}_{\mathrm{b}} \geq 3.3 \delta_{\mathrm{b}}$. (equation S 1 )

The $\mathrm{S}_{\text {LOD }}$ value must be higher than 3.3 times the $\delta_{\mathrm{b}}$ to have $99.9 \%$ of confidence limit, where the $\delta_{b}$ is standard deviation of the background signal. LOD of concentration ( $\mathrm{C}_{\mathrm{LOD}}$ ) can be obtained by substituting the calculated $\mathrm{S}_{\text {LOD }}$ value for Y in the regression equation for the signalconcentration calibration curve (Figure 5A).

Based on these considerations, the resulting $\mathrm{S}_{\mathrm{LOD}}$ and $\mathrm{C}_{\text {LOD }}$ are $2.3 \%$ and $1.8 \times 10^{-12} \mathrm{~g} \cdot \mathrm{~mL}^{-1}$, respectively.

## Supplementary Figures



Figure S1. Comparative time-based UV-vis absorbance kinetic plot for magnetophoresis time.


Figure S2. SEM image of bare magnetic micro particles used as the core material for MMP@Au probes.


Figure S3. Particle size distribution for MMPs and MMP@Au NPs.


Supplementary Movie S1. Video clip showing real-time monitoring of the magnetophoresis process before and after coating of the MMP cores with Au shells.

## Supplementary Tables

| Sputum number | Sex | $\begin{gathered} \mathrm{pcMPI}^{*} \\ (n \triangle A B S, \%) \end{gathered}$ | Gold standard |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: |
|  |  |  | $\begin{gathered} \mathrm{AFB} \\ \text { test** } \end{gathered}$ | MGIT growth unit*** | PCR |
| 1 | F | 9.25 | 2+ | ++ | MTB |
| 2 | F | 13.17 | $2+$ | ++ | MTB |
| 3 | F | 8.13 | $2+$ | ++ | MTB |
| 4 | M | 8.03 | $2+$ | ++ | MTB |
| 5 | F | 0.21 | $2+$ | ++ | MTB |
| 6 | M | 3.96 | $2+$ | + | MTB |
| 7 | M | -5.02 | $2+$ | ++ | MTB |
| 8 | M | -9.84 | $2+$ | ++ | MTB |
| 9 | F | 17.82 | $2+$ | ++ | MTB |
| 10 | M | 11.52 | 2+ | +++ | MTB |
| 11 | M | 6.84 | $2+$ | ++ | MTB |
| Sputum number | Sex | $\begin{gathered} \mathrm{pcMPI}{ }^{*} \\ (n \triangle A B S, \%) \end{gathered}$ | Gold standard |  |  |
|  |  |  | $\begin{gathered} \text { AFB } \\ \text { test** } \end{gathered}$ | MGIT growth unit*** | PCR |
| 12 | F | -0.32 | $2+$ | + | NTM <br> (M.intracellulare) |
| 13 | M | 3.90 | 2+ | ++ | NTM <br> (M.intracellulare) |
| 14 | M | -12.52 | $2+$ | ++ | NTM <br> (M.avium) |
| 15 | M | -7.35 | 2+ | ++ | NTM (M.avium) |
| 16 | M | 1.84 | $2+$ | ++ | NTM (M.avium) |

* Values for pcMPI analysis represent $n \triangle A B S(\%)$. Higher $n \triangle A B S(\%)$ value indicate higher concentration of secretory CFP-10 was monitored.
** Acid-Fast Bacilli (AFB) Smear test
*** Bacterial culture result from BACTEC MGIT 960 instrumentation. Detection signal levels: High (> 1,000 growth unit (GU)) for +++ ; medium ( 100 to $<1000 \mathrm{GU}$ ) for ++ ; and low ( 10 to $<100 \mathrm{GU}$ ) for + . Manufacturer-set threshold was 50 GU .

Table S1. Comparison of results from the pcMPI and conventional diagnostic systems (AFB test, MGIT bacterial culture, PCR) in clinical sputum samples.

## Supporting Information References

1. Apostol, I.; Miller, K. J.; Ratto, J.; Kelner, D. N. Comparison of Different Approaches for Evaluation of the Detection and Quantitation Limits of a Purity Method: A Case Study Using a Capillary Isoelectrofocusing Method for a Monoclonal Antibody. Anal. Biochem. 2009, 385, 101-106.
