

Terms & Conditions

Electronic Supporting Information files are available without a subscription to ACS Web Editions. The American Chemical Society holds a copyright ownership interest in any copyrightable Supporting Information. Files available from the ACS website may be downloaded for personal use only. Users are not otherwise permitted to reproduce, republish, redistribute, or sell any Supporting Information from the ACS website, either in whole or in part, in either machine-readable form or any other form without permission from the American Chemical Society. For permission to reproduce, republish and redistribute this material, requesters must process their own requests via the RightsLink permission system. Information about how to use the RightsLink permission system can be found at <http://pubs.acs.org/page/copyright/permissions.html>



ACS Publications

MOST TRUSTED. MOST CITED. MOST READ.

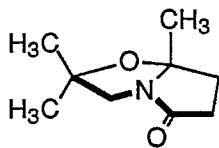
Copyright © 1996 American Chemical Society

Experimental Section

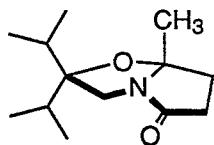
General Experimental: All reactions were carried out under argon in septum stoppered flasks. Unless otherwise noted, reagents were added by syringe, organic extracts were dried over anhydrous Na₂SO₄, filtered through a fritted glass funnel and concentrated with a rotary evaporator at aspirator pressure (20-40 mm). Flash chromatography was performed with Amicon (200-400 mesh) silica gel. Thin layer chromatography (TLC) was performed with Merck F-254 silica gel plates.

Unless otherwise noted, starting materials were obtained from commercial suppliers and used without further purification. Tetrahydrofuran (THF) and ether were distilled under N₂ from sodium/benzophenone immediately prior to use. Dichloromethane (CH₂Cl₂) and toluene were distilled from CaH₂ immediately prior to use.

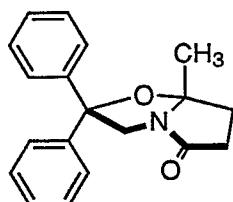
¹H NMR spectra were recorded with a Bruker AC-300 spectrometer. ¹³C NMR spectra were recorded with the same instrument at 75 MHz. All NMR spectra were recorded in CDCl₃ and chemical shifts are expressed in ppm relative to internal CHCl₃ (7.27 ppm). Significant NMR data are tabulated in order: multiplicity (s, singlet; d, doublet; t, triplet; m, multiplet), coupling constants in hertz, and number of protons. Infrared spectra were recorded on a Perkin-Elmer 1600 Fourier Transform Infrared Spectrometer. Microanalyses were performed by Atlantic Microlab, Inc., Norcross, Georgia.



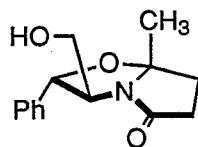
General Procedure for Bicyclic Lactam Formation: Lactam (5a, R = Me). 2-Amino-1-hydroxy-1-methyl propane (2.45 g, 27.0 mmol) and levulinic acid (7) (3.13 g, 27.0 mmol) were dissolved in toluene (50 mL). The flask was equipped with Dean-Stark trap and the solution was heated to reflux. After 16h the solution was cooled, washed with saturated aqueous NaHCO₃, dried and concentrated. Chromatography on silica gel with ethyl acetate afforded 4.06 g (89%) of lactam (5a) as a light yellow oil. IR (neat) 2974, 1713 cm⁻¹. ¹H NMR (300 MHz) δ 0.98 (s, 3H), 1.24 (s, 3H), 1.30 (s, 3H), 2.06 (m, 2H), 2.37 (m, 1H), 2.53 (m, 1H), 2.81 (d, *J* = 11.6, 1H), 3.74 (d, *J* = 11.6, 1H). ¹³C NMR (75 MHz) δ 26.6, 27.7, 28.0, 32.3, 34.8, 52.8, 81.3, 99.4, 177.4. Anal. Calcd for C₉H₁₅NO₂: C, 63.88; H, 8.93. Found: C, 63.95; H, 8.97.



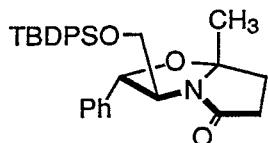
Lactam (5b, R = i-Pr). Light yellow oil. IR (neat) 2974, 1713 cm⁻¹. ¹H NMR (300 MHz) δ 0.79 (d, *J* = 6.7, 3H), 0.83 (d, *J* = 6.7, 3H), 0.90 (d, *J* = 6.7, 3H), 0.95 (d, *J* = 6.7, 3H), 1.42 (s, 3H), 1.76 (m, 1H), 2.03 (m, 1H), 2.15 (m, 2H), 2.36 (m, 1H), 2.61 (m, 1H), 2.95 (d, *J* = 12.2, 1H), 3.72 (d, *J* = 12.2, 1H). ¹³C NMR (75 MHz) δ 17.9, 18.0, 18.4, 18.6, 27.0, 32.7, 33.3, 33.8, 35.8, 44.0, 92.2, 99.4, 175.0.



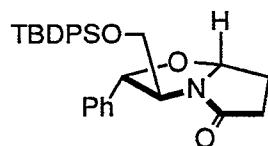
Lactam (5c, R = Ph). White solid, mp. 95-96 °C (hexane/EtOAc). IR (neat) 2977, 1716 cm⁻¹. ¹H NMR (300 MHz) δ 1.47 (s, 3H), 2.09 (m, 3H), 2.48 (m, 1H), 3.41 (d, *J* = 12.3, 1H), 4.90 (d, *J* = 12.3, 1H), 7.11-7.33 (m, 10H). ¹³C NMR (75 MHz) δ 27.1, 32.8, 34.6, 53.0, 89.0, 101.0, 126.1, 126.8, 127.7, 128.0, 128.80, 128.82, 144.1, 145.3, 177.7. Anal. Calcd for C₁₉H₁₉NO₂: C, 77.79; H, 6.53. Found: C, 77.66; H, 6.61.



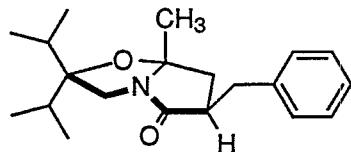
Lactam (9). Colorless crystals. mp. 89-90 °C (hexanes/EtOAc). [α]_D²³ +65.3° (c 1.4, EtOH). IR (neat) 3407, 2978, 1686 cm⁻¹. ¹H NMR (300 MHz) δ 1.65 (s, 3H), 2.39 (m, 2H), 2.60 (m, 1H), 2.85 (m, 2H), 3.70 (m, 1H), 3.76 (m, 2H), 5.14 (d, *J* = 7.6, 1H), 7.35 (m, 5H). ¹³C NMR (75 MHz) δ 24.8, 33.4, 35.3, 63.1, 65.0, 82.9, 100.1, 126.3, 128.7, 128.8, 178.6.



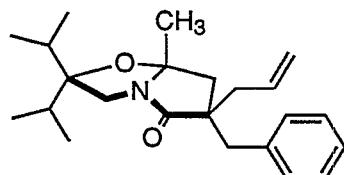
General Silylation Procedure: Lactam (10). To a CH₂Cl₂ solution (100 mL) of lactam **9** (8.0 g, 33.7 mmol) and TBDPS-Cl (11.1 g, 40.5 mmol) was added neat imidazole (2.8 g, 41.0 mmol). After stirring for 24h at 25 °C the slurry was filtered through celite and concentrated. Recrystallization from hexanes/ethyl acetate afforded lactam **10** (13.9 g, 28.6 mmol) in 85% yield as colorless crystals. Mp. 127-128 °C (hexanes/EtOAc). [α]_D²³ +38.7° (c 2.0, EtOH). IR (neat) 3409, 2976, 1687 cm⁻¹. ¹H NMR (300 MHz) δ 1.00 (s, 9H), 1.53 (s, 3H), 2.23 (m, 2H), 2.49 (m, 1H), 2.74 (m, 1H), 3.75 (m, 1H), 3.87 (m, 2H), 5.30 (d, J = 6.7, 1H), 7.30 (m, 9H), 7.61 (m, 6H). ¹³C NMR (75 MHz) δ 19.5, 25.3, 26.8, 27.1, 33.6, 34.9, 63.9, 64.0, 83.1, 100.5, 126.8, 127.9, 128.0, 128.5, 128.8, 129.8, 130.0, 130.1, 133.1, 133.3, 135.0, 135.8, 135.9, 139.3, 178.3. Anal. Calcd for C₁₉H₁₉NO₂: C, 74.19; H, 7.26. Found: C, 74.04; H, 7.31.



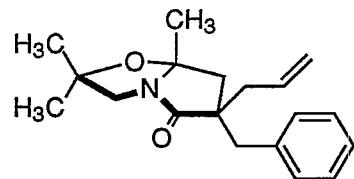
Lactam (12).⁸ Colorless oil. [α]_D²³ +28.2° (c 1.5, EtOH). IR (neat) 2930, 1719 cm⁻¹. ¹H NMR (300 MHz) δ 1.06 (s, 9H), 2.17 (m, 1H), 2.37 (m, 1H), 2.60 (m, 2H), 3.78 (m, 1H), 3.95 (m, 2H), 5.15 (d, J = 5.8, 2H), 5.36 (dd, J = 6.1, 2.1, 1H), 7.16-7.42 (m, 11H), 7.61 (m, 4H). ¹³C NMR (75 MHz) δ 19.2, 24.3, 26.9, 31.3, 63.5, 63.9, 82.6, 93.1, 126.3, 127.8, 127.9, 128.3, 128.6, 129.8, 129.9, 133.0, 135.6, 135.7, 139.1, 179.3.



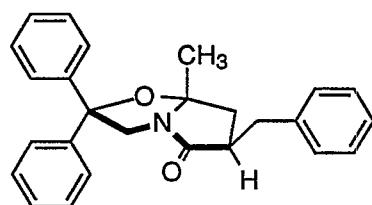
General Procedure for Bicyclic Lactam Alkylation: Lactam (6b, R=i-Pr, R¹=H, R²=Bn). Lactam **5b** (400 mg, 1.80 mmol) was dissolved in THF (20 mL) and cooled to -78 °C. s-BuLi (1.65 mL, 1.1M in hexane) was added dropwise to the solution over 5 min. After 2h the solution was placed in a 0 °C bath for 15 min and subsequently recooled to -78 °C. To the stirring -78 °C solution was added benzyl bromide (0.34 g, 1.97 mmol) dropwise over 5 min. After 2h the solution was quenched by addition of saturated aqueous NH₄Cl solution (4 mL). Ethyl acetate (20 mL) was added to the mixture and the resulting solution was separated, dried and concentrated. Purification on silica (hexanes/EtOAc, 1:1) afforded 516 mg (91%) of **6b** as a clear oil. IR (neat) 2965, 1707 cm⁻¹. ¹H NMR (300 MHz) δ 0.88 (app t, *J* = 7.0, 6H), 0.99 (app t, *J* = 7.0, 6H), 1.23 (s, 3H), 1.81 (m, 1H), 2.05 (m, 2H), 2.43 (dd, *J* = 14.0, 10.1, 1H), 3.01 (m, 4H), 3.89 (d, *J* = 12.2, 1H), 7.27 (m, 5H). ¹³C NMR (75 MHz) δ 17.9, 18.1, 18.5, 18.6, 28.8, 33.4, 33.6, 37.5, 38.7, 45.3, 46.9, 91.3, 98.5, 126.5, 128.4, 129.1, 138.6, 178.1.



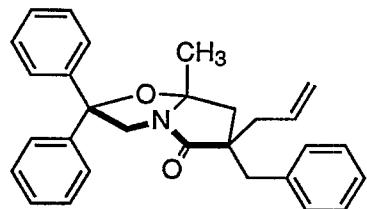
Lactam (6b, R=i-Pr, R¹=Bn, R²=Allyl). White solid, mp. 65-66 °C (hexanes). IR (neat) 2962, 1709 cm⁻¹. ¹H NMR (300 MHz) δ 0.53 (d, *J* = 6.7, 3H), 0.74 (d, *J* = 6.7, 3H), 0.95 (m, 6H), 1.50 (s, 3H), 2.02 (m, 2H), 2.32 (d, *J* = 13.4, 1H), 2.47 (m, 2H), 2.74 (d, *J* = 13.7, 1H), 2.96 (d, *J* = 11.9, 1H), 3.05 (d, *J* = 13.7, 1H), 3.84 (d, *J* = 11.9, 1H), 5.20 (m, 2H), 5.93 (m, 1H), 7.25 (m, 5H). ¹³C NMR (75 MHz) δ 17.0, 19.1, 19.2, 28.2, 31.8, 34.6, 40.3, 42.8, 44.0, 45.2, 52.9, 90.5, 96.2, 118.8, 126.4, 128.0, 131.2, 133.6, 137.3, 179.0.



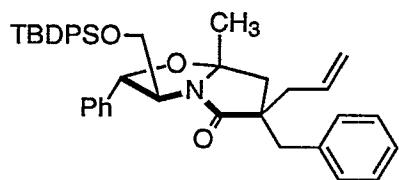
Lactam (6a, R=Me, R¹=Bn, R²=Allyl). IR (neat) 2976, 1707 cm⁻¹. ¹H NMR (300 MHz) δ 0.61 (s, 3H), 1.30 (s, 3H), 1.44 (s, 3H), 1.98 (d, *J* = 13.7, 1H), 2.40 (m, 2H), 2.75 (d, *J* = 13.7, 1H), 2.94 (d, *J* = 11.5, 1H), 3.04 (d, *J* = 13.4, 1H), 3.81 (d, *J* = 11.7, 1H), 5.17 (m, 2H), 5.84 (m, 1H), 7.21 (m, 5H). ¹³C NMR (75 MHz) δ 27.7, 28.6, 42.0, 42.3, 43.4, 52.8, 53.2, 80.7, 96.9, 119.2, 126.7, 128.3, 131.4, 133.8, 137.4, 180.4.



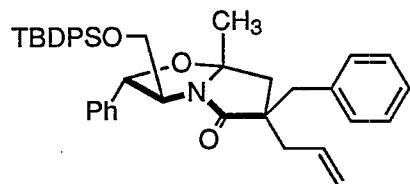
Lactam (6c, R=Ph, R¹=H, R²=Ph). Yellow oil. IR (neat) 3026, 2978, 1714 cm⁻¹. ¹H NMR (300 MHz) δ 1.24 (s, 3H), 1.91 (dd, *J* = 14.3, 4.8, 1H), 2.40 (m, 1H), 2.62 (m, 1H), 2.79 (m, 1H), 2.95 (dd, *J* = 13.5, 4.8, 1H), 3.57 (d, *J* = 12.2, 1H), 4.98 (d, *J* = 11.9, 1H), 7.09-7.38 (m, 15H). ¹³C NMR (75 MHz) δ 27.3, 37.0, 37.2, 44.9, 53.9, 87.7, 99.3, 125.5, 126.0, 126.4, 127.1, 127.3, 128.1, 128.3, 129.0, 138.3, 144.0, 145.0, 180.3.



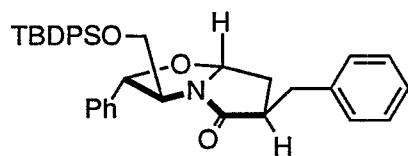
Lactam (6c, R=Ph, R¹=Bn, R²=Allyl). Yellow oil. IR (neat) 3027, 2977, 1712 cm⁻¹. ¹H NMR (300 MHz) δ 1.54 (s, 3H), 1.79 (d, *J* = 11.8, 1H), 1.86 (d, *J* = 12.0, 1H), 2.15 (m, 1H), 2.40 (dd, *J* = 13.5, 6.4, 1H), 2.52 (d, *J* = 14.0, 1H), 2.67 (d, *J* = 13.7, 1H), 3.58 (d, *J* = 11.9, 1H), 5.11 (m, 2H), 5.76 (m, 1H), 7.03 (m, 2H), 7.19-7.48 (m, 13H). ¹³C NMR (75 MHz) δ 27.3, 41.5, 41.8, 42.8, 51.3, 52.9, 87.8, 97.8, 118.8, 125.5, 126.4, 126.6, 127.2, 127.5, 128.0, 128.30, 128.33, 130.5, 133.9, 137.2, 143.7, 145.3, 181.4.



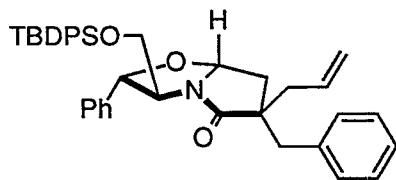
Lactam (11, R¹=Bn, R²=Allyl). Clear oil. [α]_D²³ +63.8° (c 1.9, EtOH). IR (neat) 3067, 2965, 1709 cm⁻¹. ¹H NMR (300 MHz) δ 1.12 (s, 9H), 1.60 (s, 3H), 2.09 (d, *J* = 13.8, 1H), 2.60 (m, 4H), 3.24 (d, *J* = 13.4, 1H), 3.81 (m, 2H), 3.94 (m, 1H), 5.15 (d, *J* = 6.7, 1H), 5.23 (m, 2H), 5.93 (m, 1H), 6.81 (d, *J* = 8.0, 2H), 7.19-7.44 (m, 14H), 7.71 (m, 4H). ¹³C NMR (75 MHz) δ 19.3, 26.3, 27.0, 41.2, 41.7, 44.1, 53.5, 64.0, 64.9, 81.6, 97.3, 119.3, 126.5, 127.9, 128.0, 128.2, 128.5, 128.6, 129.9, 130.0, 131.3, 133.0, 133.2, 133.7, 135.7, 135.8, 135.9, 137.7, 139.3, 181.7.



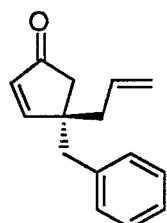
Lactam (11, R¹=Allyl, R²=Bn). Clear oil. $[\alpha]_D^{23} +11.8^\circ$ (c 1.3, EtOH). IR (neat) 3067, 2965, 1709 cm^{-1} . ¹H NMR (300 MHz) δ 0.87 (s, 3H), 1.05 (s, 9H), 2.06 (d, $J = 14.4$, 1H), 2.27 (m, 1H), 2.44 (d, $J = 14.3$, 1H), 2.59 (m, 1H), 2.65 (d, $J = 13.5$, 1H), 3.08 (d, $J = 13.4$, 1H), 3.71-3.98 (m, 3H), 5.05 (d, $J = 7.0$, 1H), 5.15 (m, 2H), 5.73 (m, 1H), 7.19 (m, 10H), 7.35 (m, 6H), 7.68 (m, 4H). ¹³C NMR (75 MHz) δ 19.1, 24.8, 26.7, 40.7, 43.2, 44.0, 53.2, 63.8, 64.9, 80.9, 97.3, 119.3, 126.2, 126.7, 127.7, 128.0, 128.1, 128.3, 129.7, 130.4, 132.8, 133.9, 135.5, 137.0, 139.3, 182.1.



Lactam (13, R=H, R'=Bn). White solid, mp. 109-110 °C. $[\alpha]_D^{23} = +8.2^\circ$ (c 0.8, EtOH). IR (neat) 3029, 2930, 1716 cm^{-1} . ¹H NMR (300 MHz) δ 1.06 (s, 9H), 2.07 (m, 1H), 2.25 (m, 1H), 2.80 (dd, $J = 13.7$, 9.1, 1H), 3.04 (m, 1H), 3.19 (dd, $J = 13.7$, 4.5, 1H), 3.78 (m, 1H), 3.90 (m, 1H), 4.05 (m, 1H), 5.06 (d, $J = 6.1$, 1H), 5.14 (dd, $J = 4.6$, 1.2, 1H), 7.20 (m, 10H), 7.34 (m, 6H), 7.64 (d, $J = 6.5$, 4H). ¹³C NMR (75 MHz) δ 19.1, 26.8, 29.3, 37.7, 44.2, 63.5, 64.5, 82.3, 91.5, 126.2, 126.5, 127.7, 128.1, 128.4, 128.5, 128.9, 129.7, 129.8, 132.8, 135.5, 138.3, 139.1, 181.4.

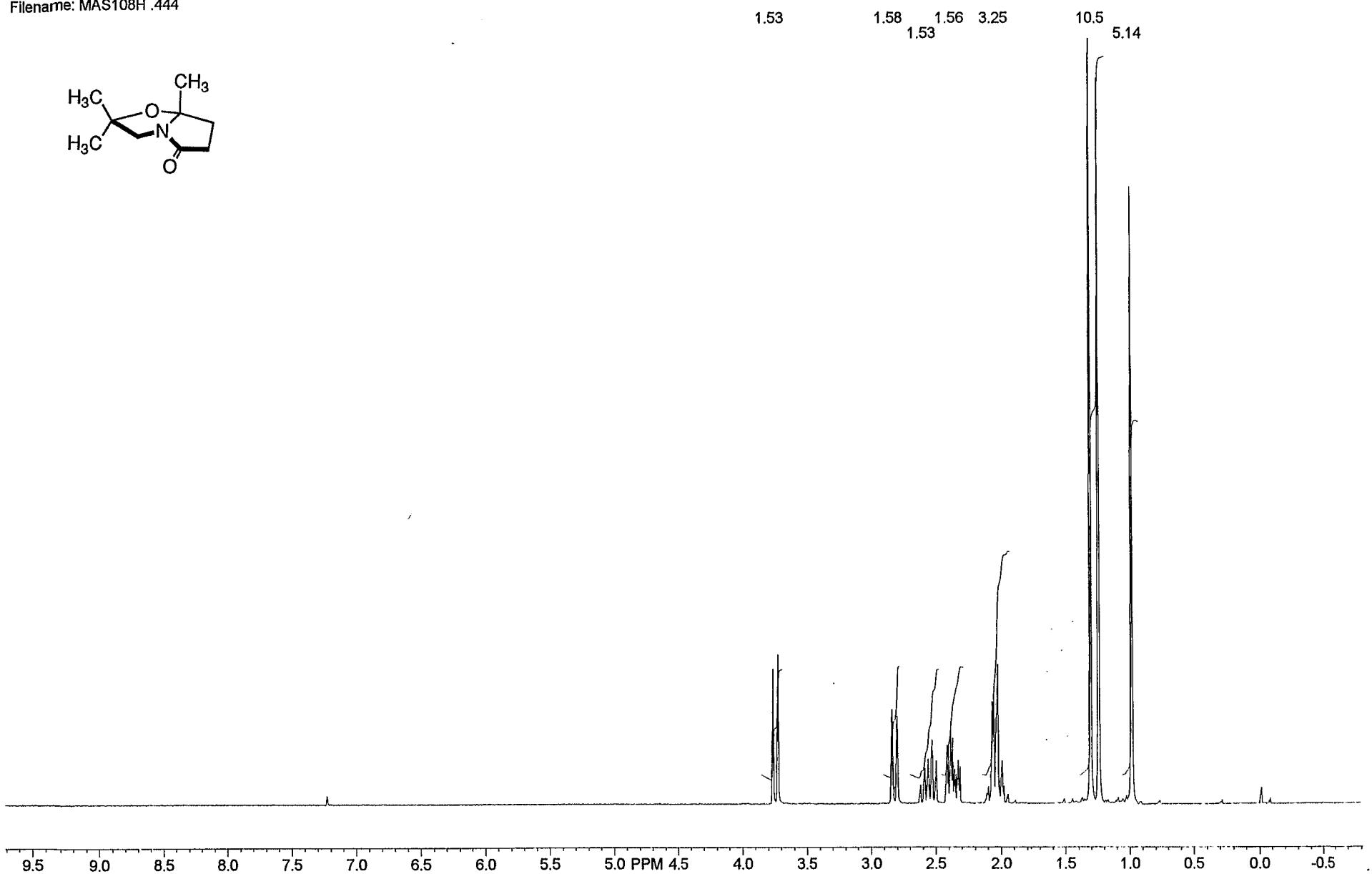
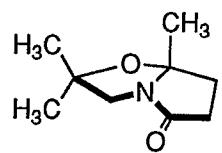


Lactam (13, R=Bn, R'=Allyl). Yellow oil. $[\alpha]_D^{23} = +60.0^\circ$ (c 1.0, EtOH). IR (neat) 3069, 2929, 1706 cm^{-1} . ^1H NMR (300 MHz) δ 1.02 (s, 9H), 2.20 (m, 3H), 2.50 (dd, $J = 13.4, 6.4, 1\text{H}$), 2.63 (d, $J = 13.4, 1\text{H}$), 3.15 (d, $J = 13.4, 1\text{H}$), 3.65 (m, 2H), 3.91 (dd, $J = 10.7, 3.6, 1\text{H}$), 5.05 (d, $J = 6.5, 1\text{H}$), 5.11 (m, 2H), 5.81 (m, 1H), 6.73 (dd, $J = 7.9, 2.1$), 7.10-7.38 (m, 16H), 7.60 (m, 4H). ^{13}C NMR (75 MHz) δ 19.3, 26.9, 33.9, 42.5, 44.0, 53.7, 62.9, 63.7, 82.5, 90.3, 119.1, 126.3, 126.5, 127.8, 127.9, 128.2, 128.4, 128.5, 129.9, 130.0, 131.1, 132.9, 133.4, 135.6, 135.7, 137.6, 139.0, 179.9.

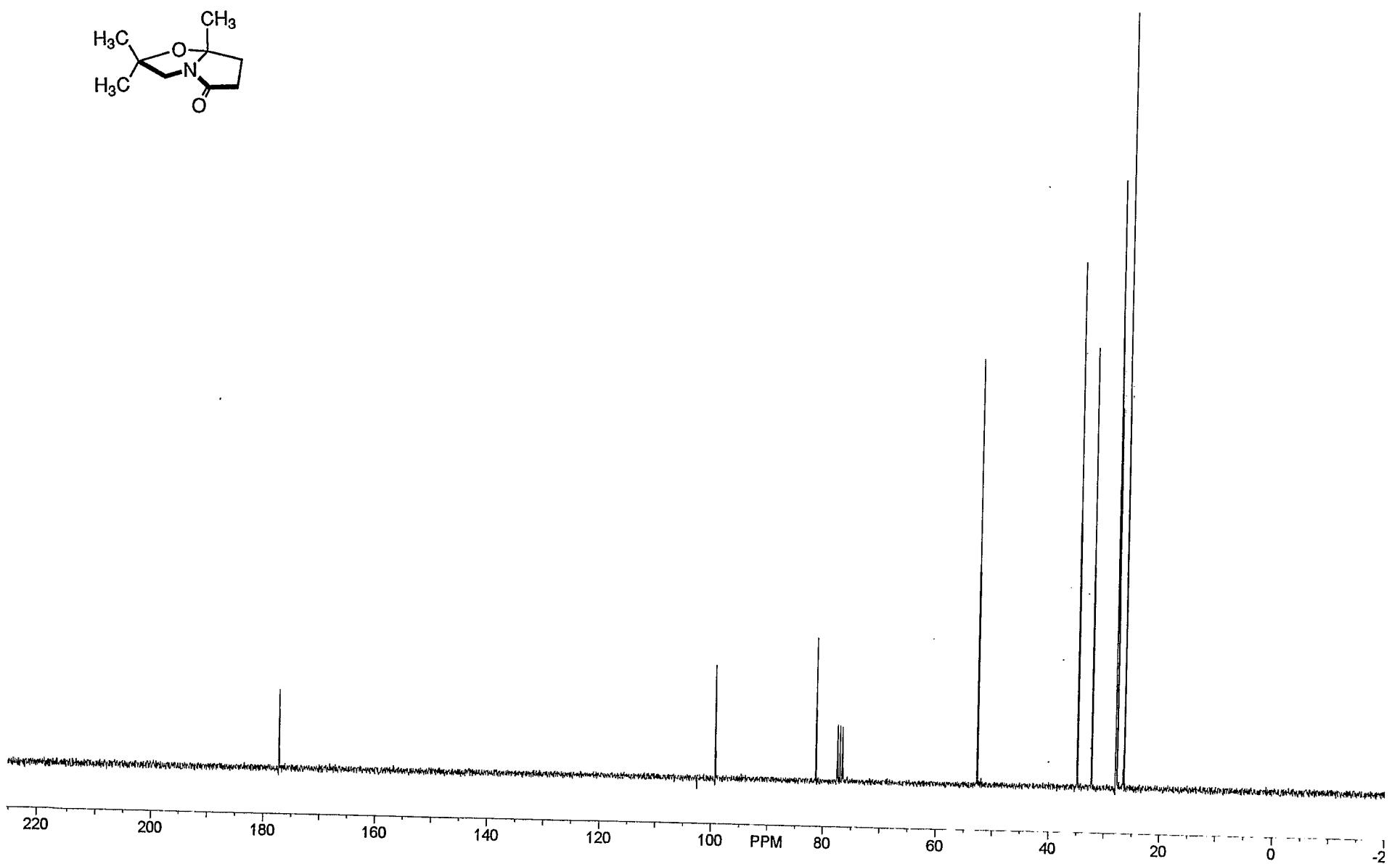


Cyclopentenone (4, R¹=Allyl, R²=Bn). Clear oil. $[\alpha]_D^{23} -62.6^\circ$ (c 1.7, EtOH), (lit.⁵ $[\alpha]_D^{23} -64.1^\circ$). IR (neat) 3028, 2916, 1714 cm^{-1} . ^1H NMR (300 MHz) δ 2.27 (d, $J = 1.5, 2\text{H}$), 2.30 (m, 2H), 2.77 (d, $J = 3.4, 1\text{H}$), 2.90 (d, $J = 3.4, 1\text{H}$), 5.06 (m, 2H), 5.65 (m, 1H), 6.04 (d, $J = 5.8, 1\text{H}$), 7.07 (m, 2H), 7.25 (m, 3H), 7.44 (d, $J = 5.8, 3\text{H}$). ^{13}C NMR (75 MHz) δ 42.8, 44.6, 45.3, 49.1, 119.3, 126.7, 128.2, 130.3, 133.2, 133.4, 136.7, 170.2, 213.0.

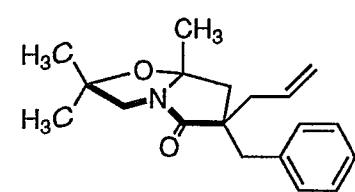
Filename: MAS108H .444



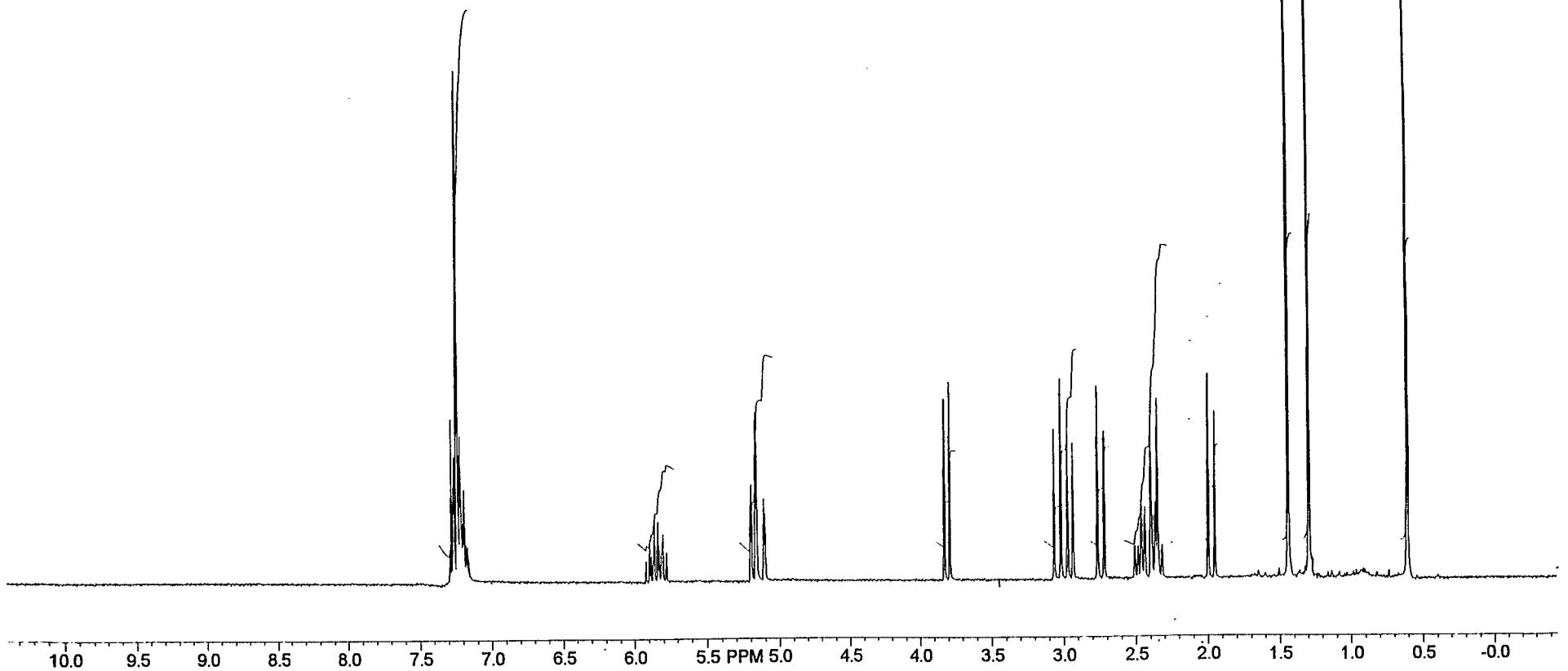
Filename: MAS108C .444



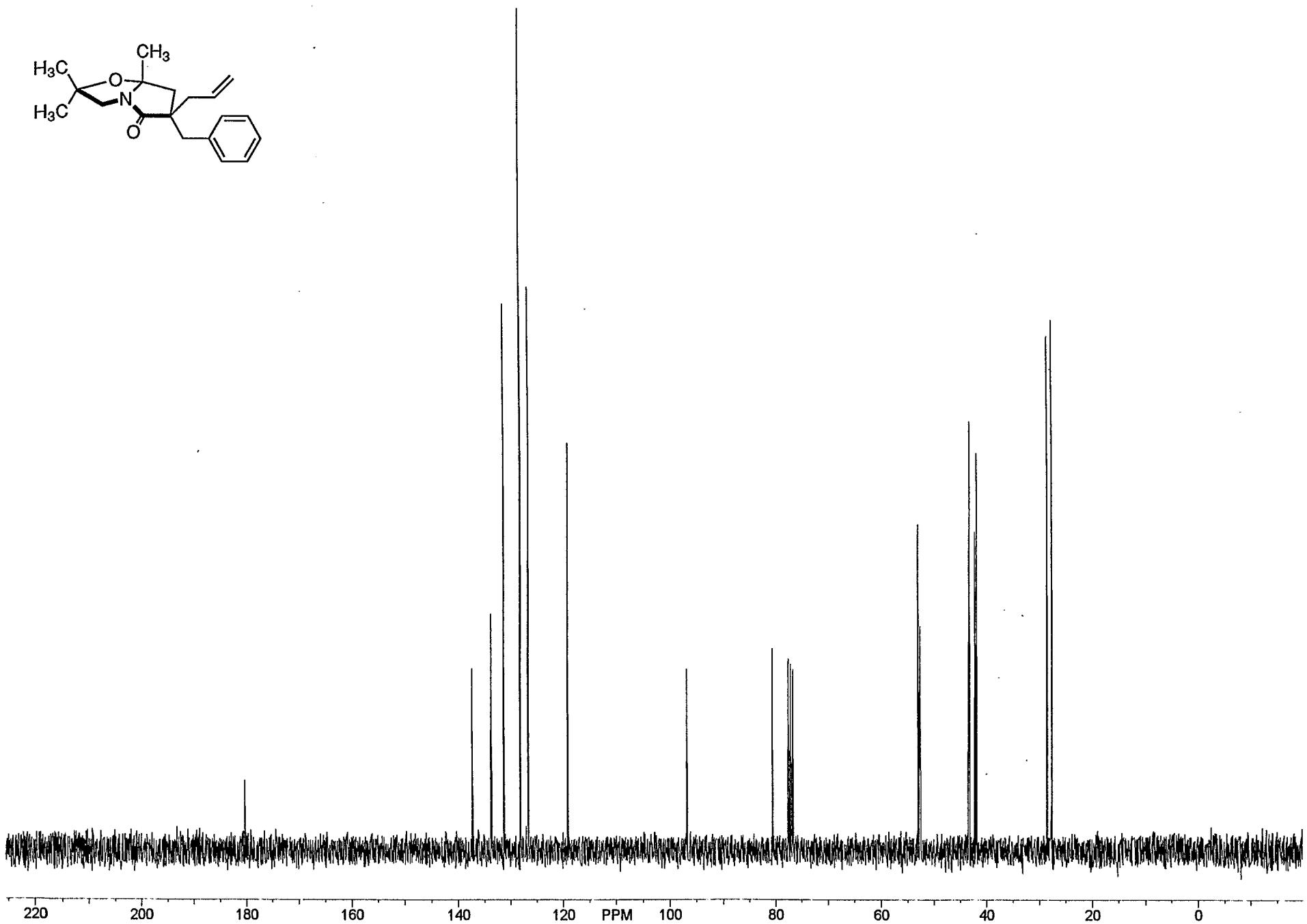
Filename: MAS155CR.555



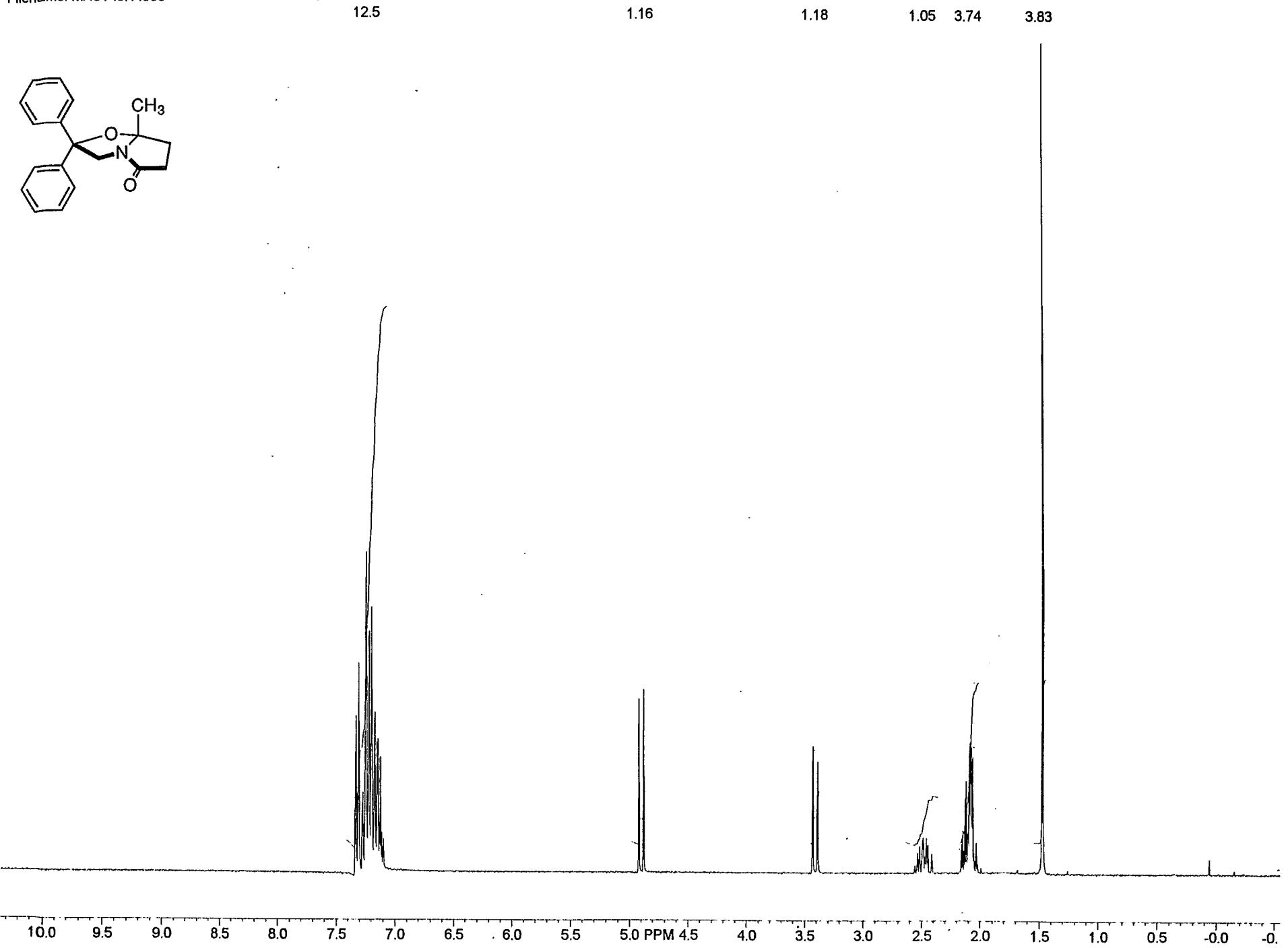
4.1 0.563 1.41 0.69 1.46
0.718 2.26 0.726 2.35
2.51 2.29



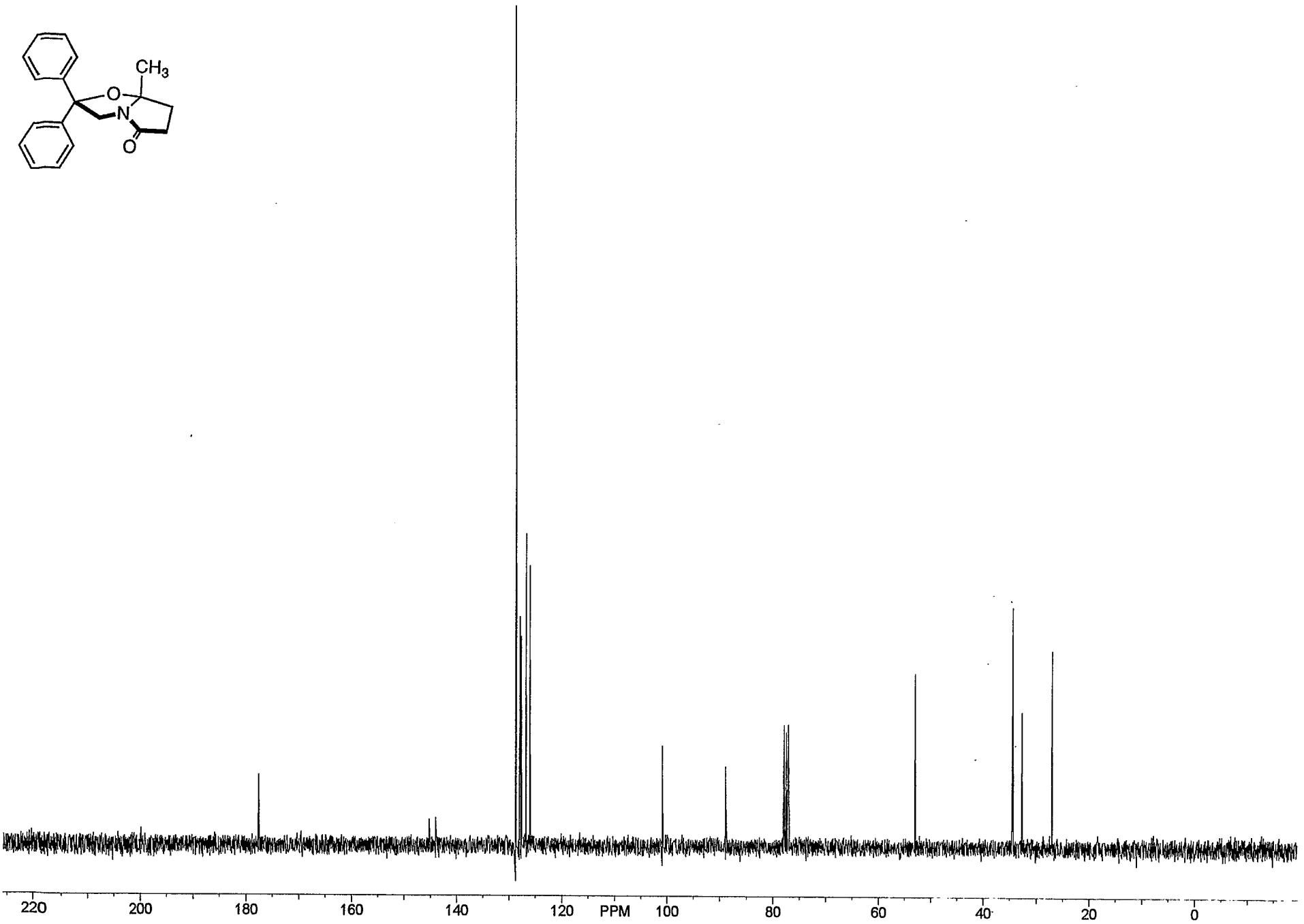
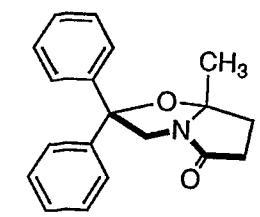
Filename: MAS155C .333



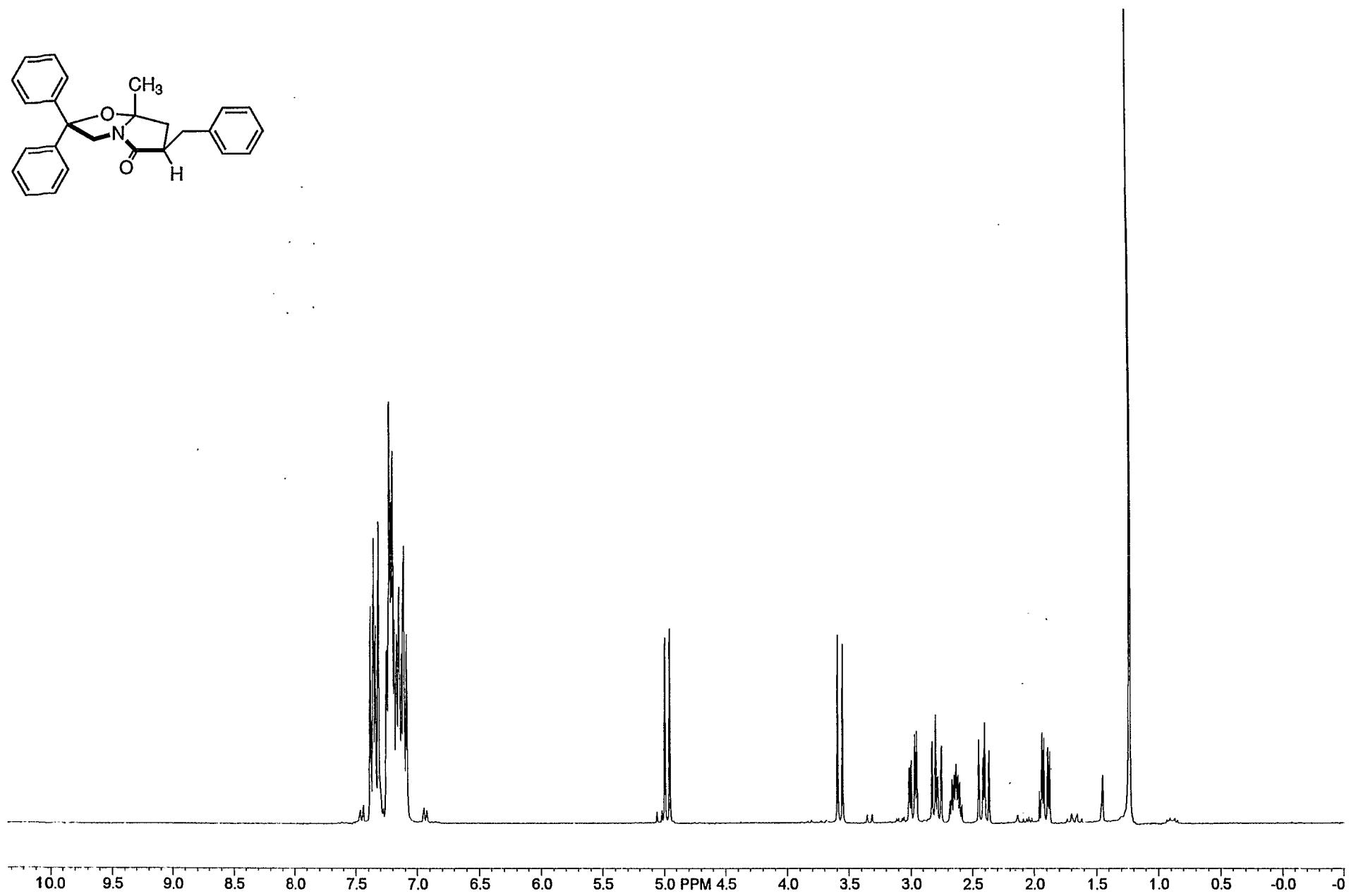
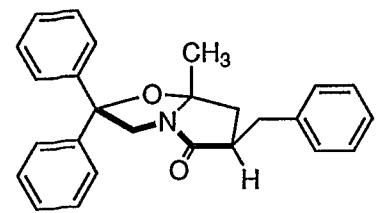
Filename: MAS143H .333



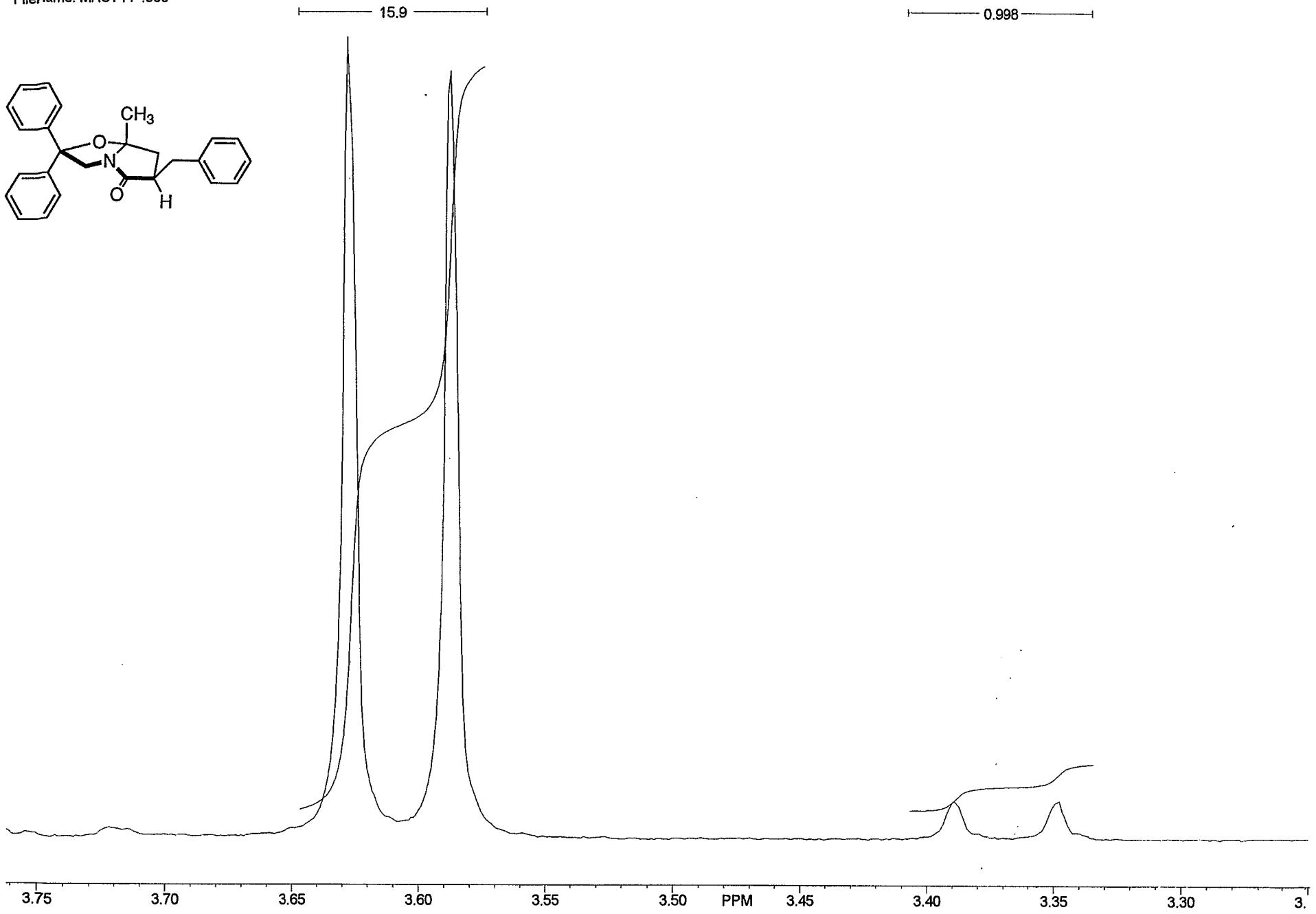
Filename: MAS143C .444



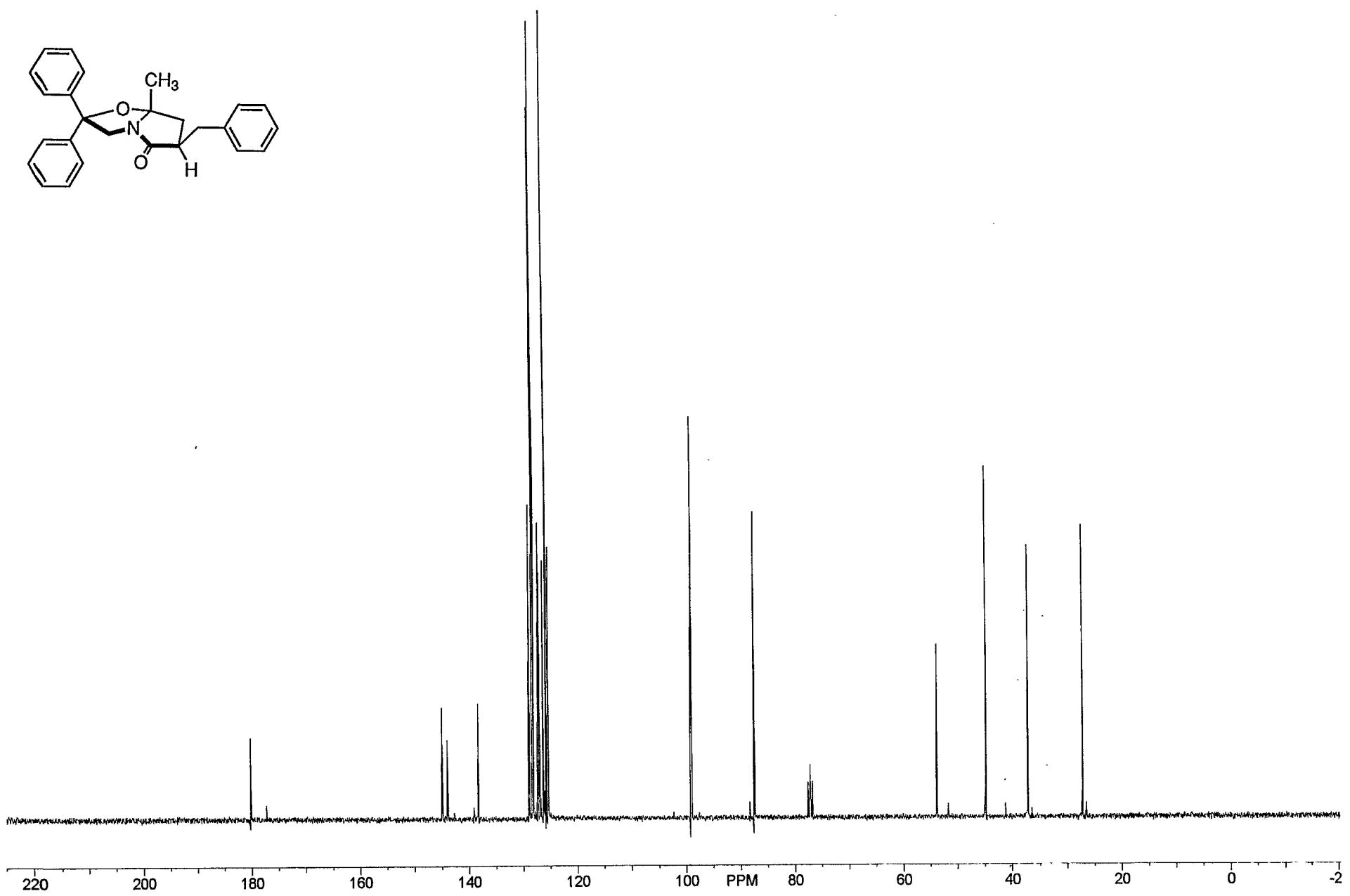
Filename: MAS144 .000

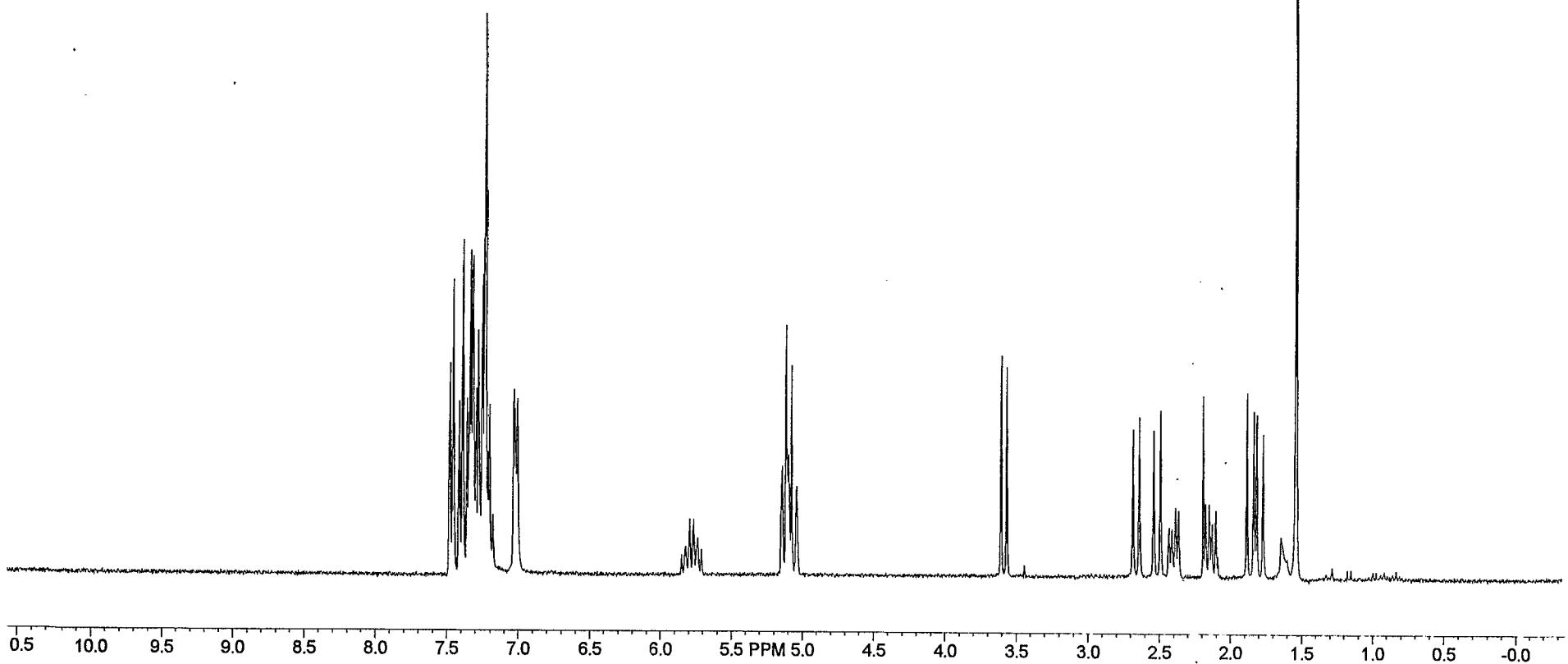
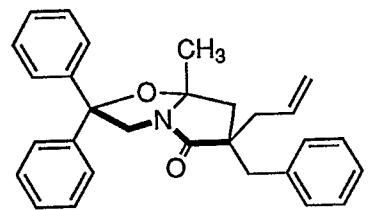


Filename: MAS144 .000

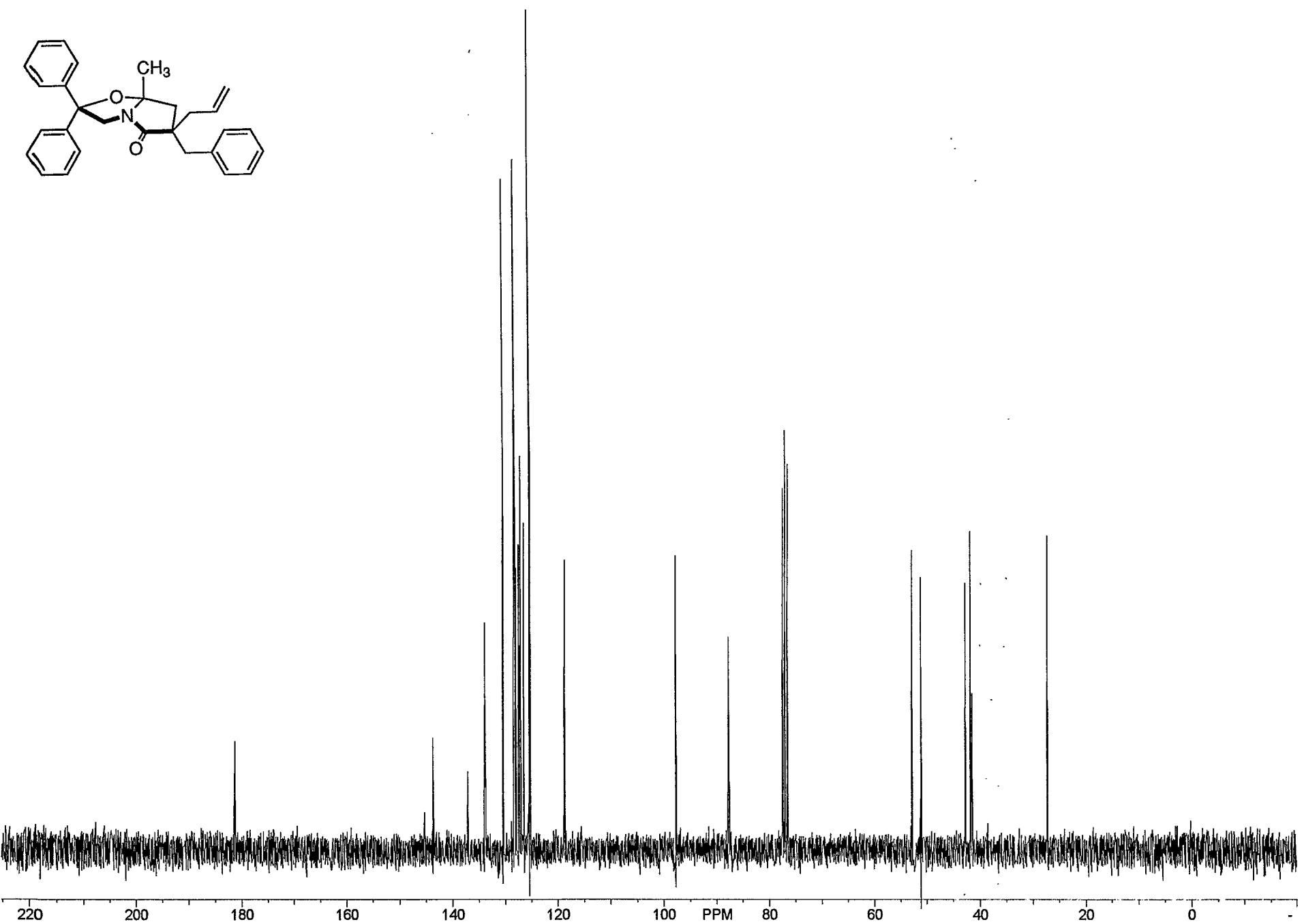


Filename: MAS144C.333



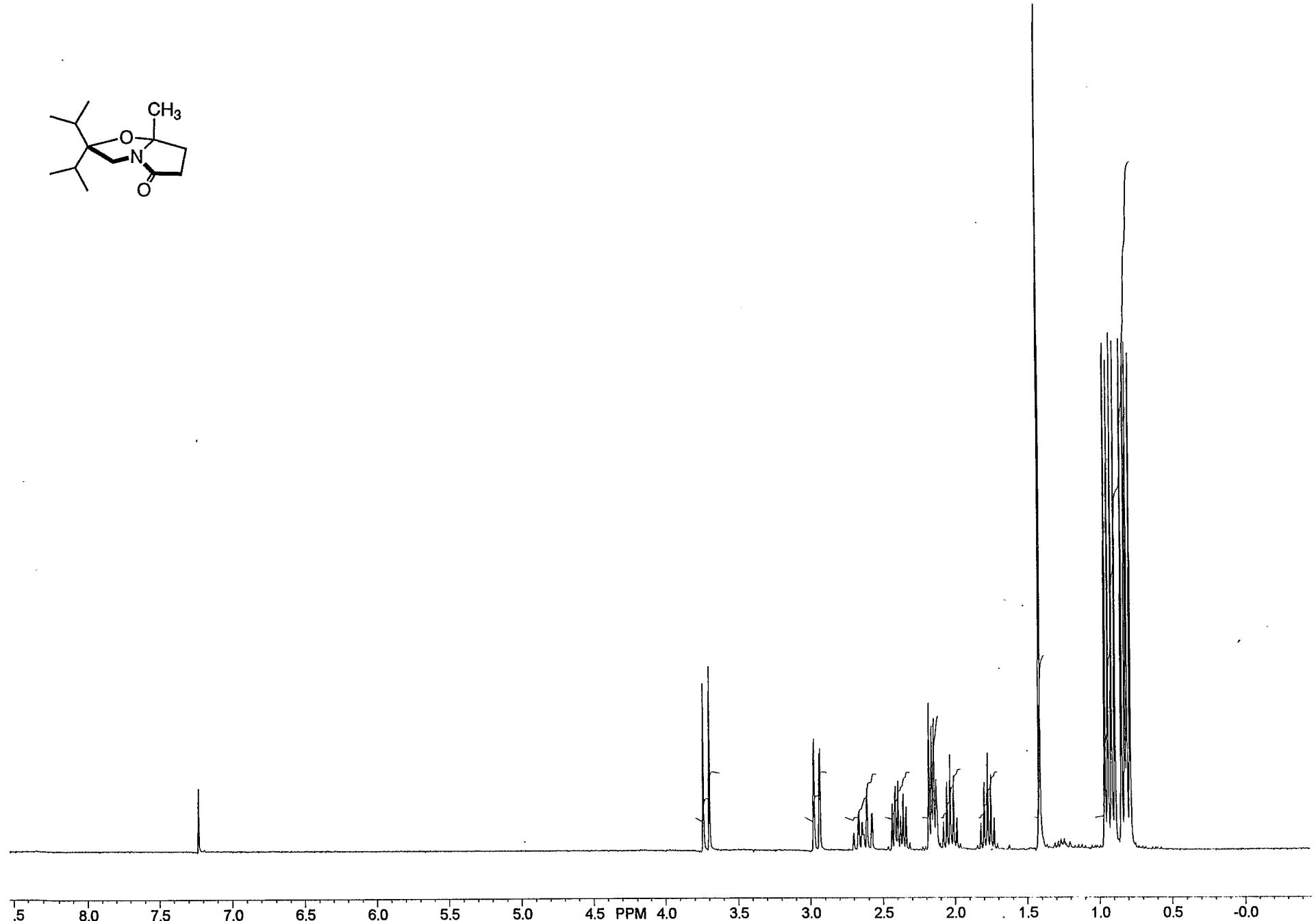


Filename: MAS152H .999

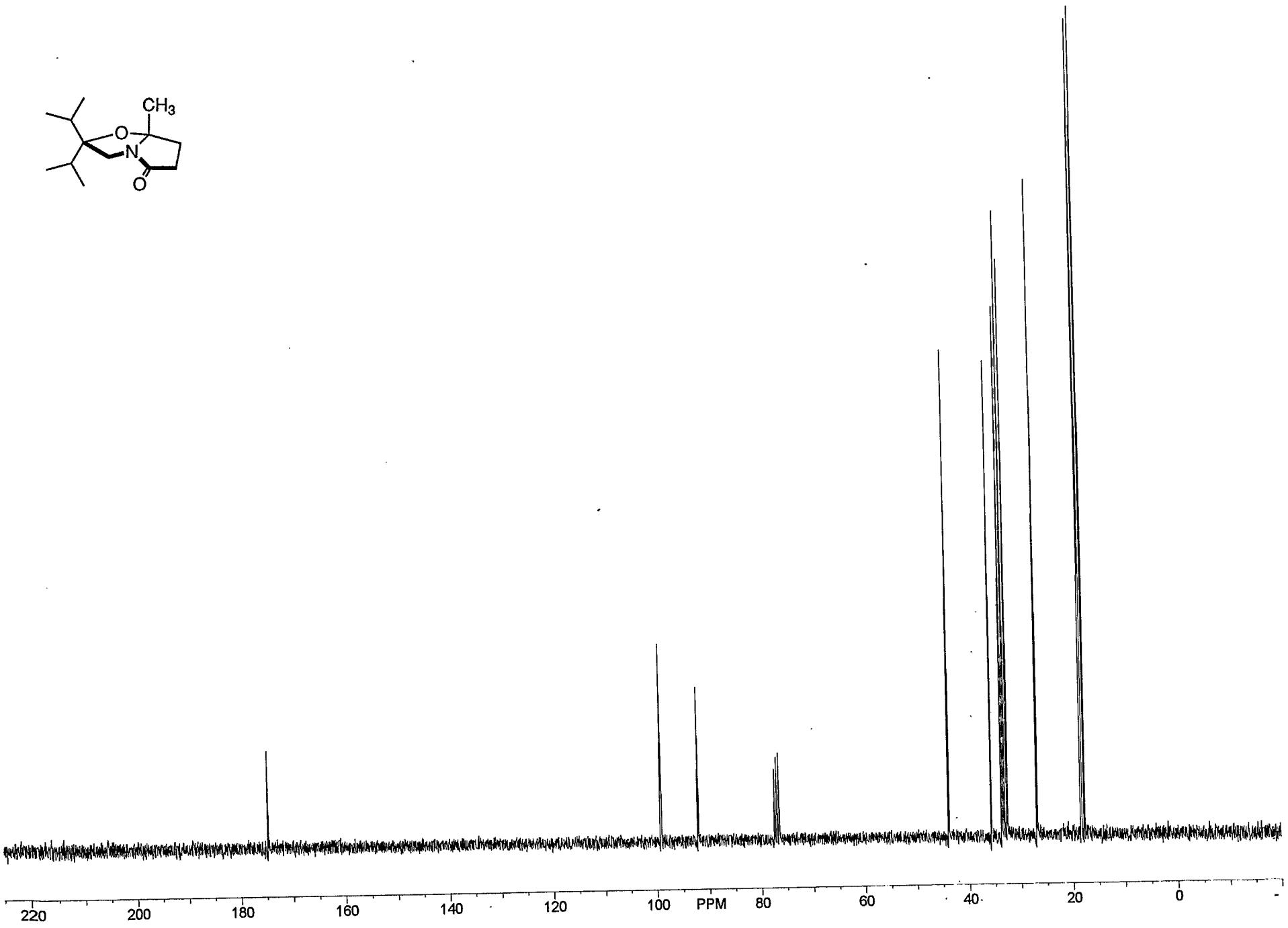
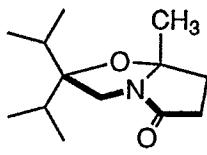


Filename: MAS119 .999

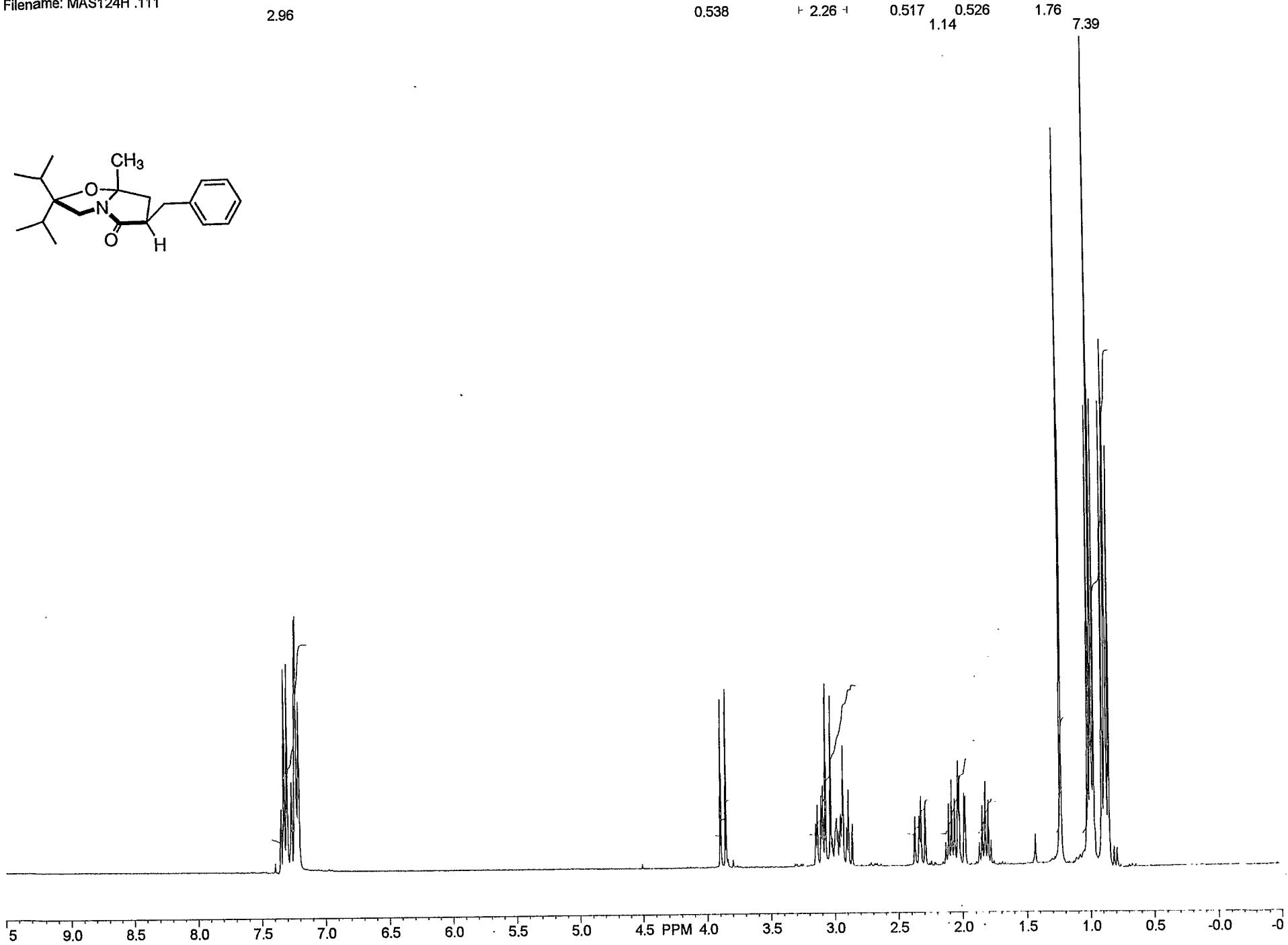
0.663 0.667 0.644 1.52 0.681 2.41 9.78
 0.678 0.726



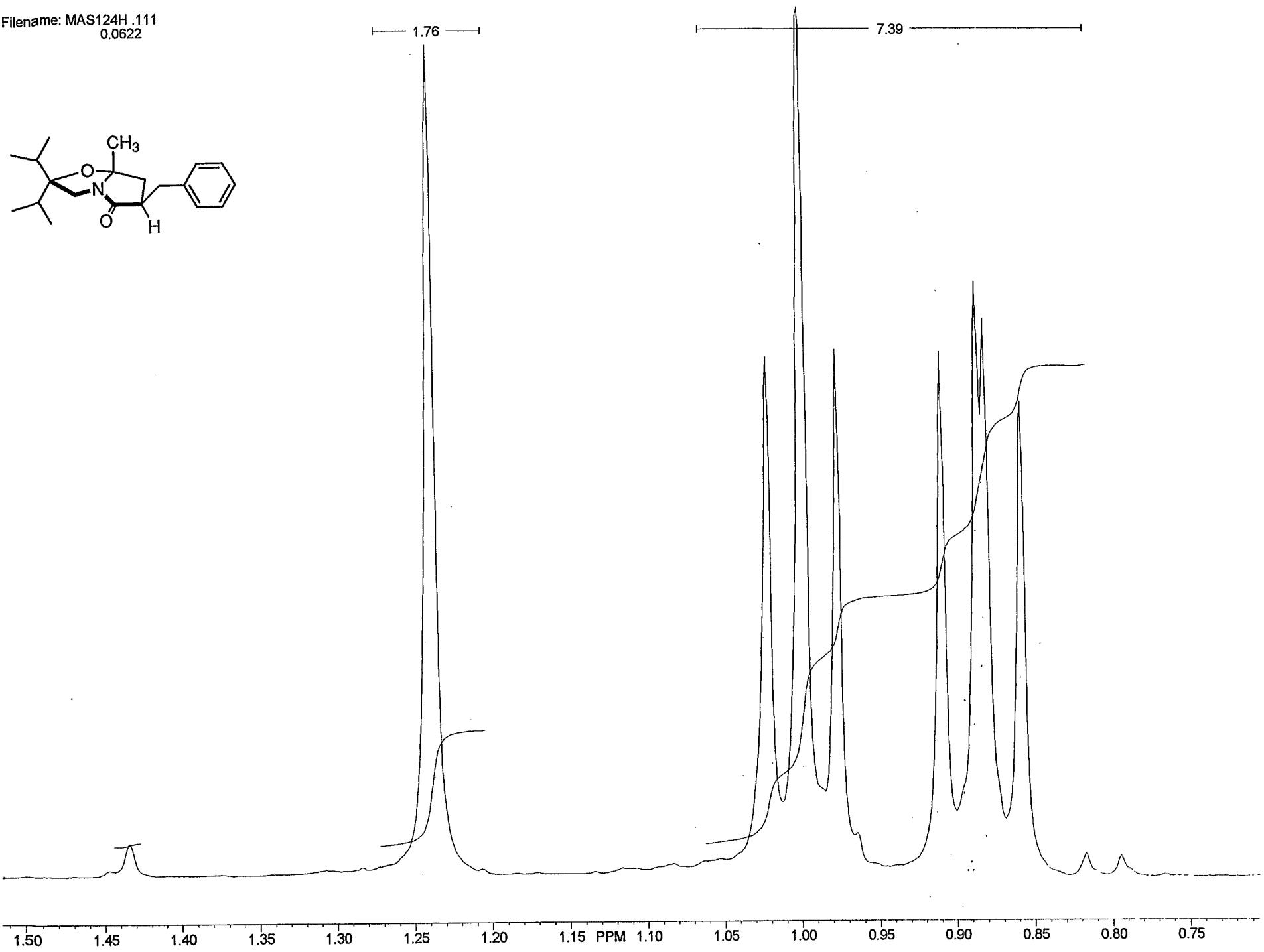
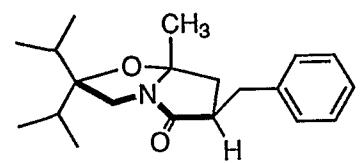
Filename: MAS119C .555



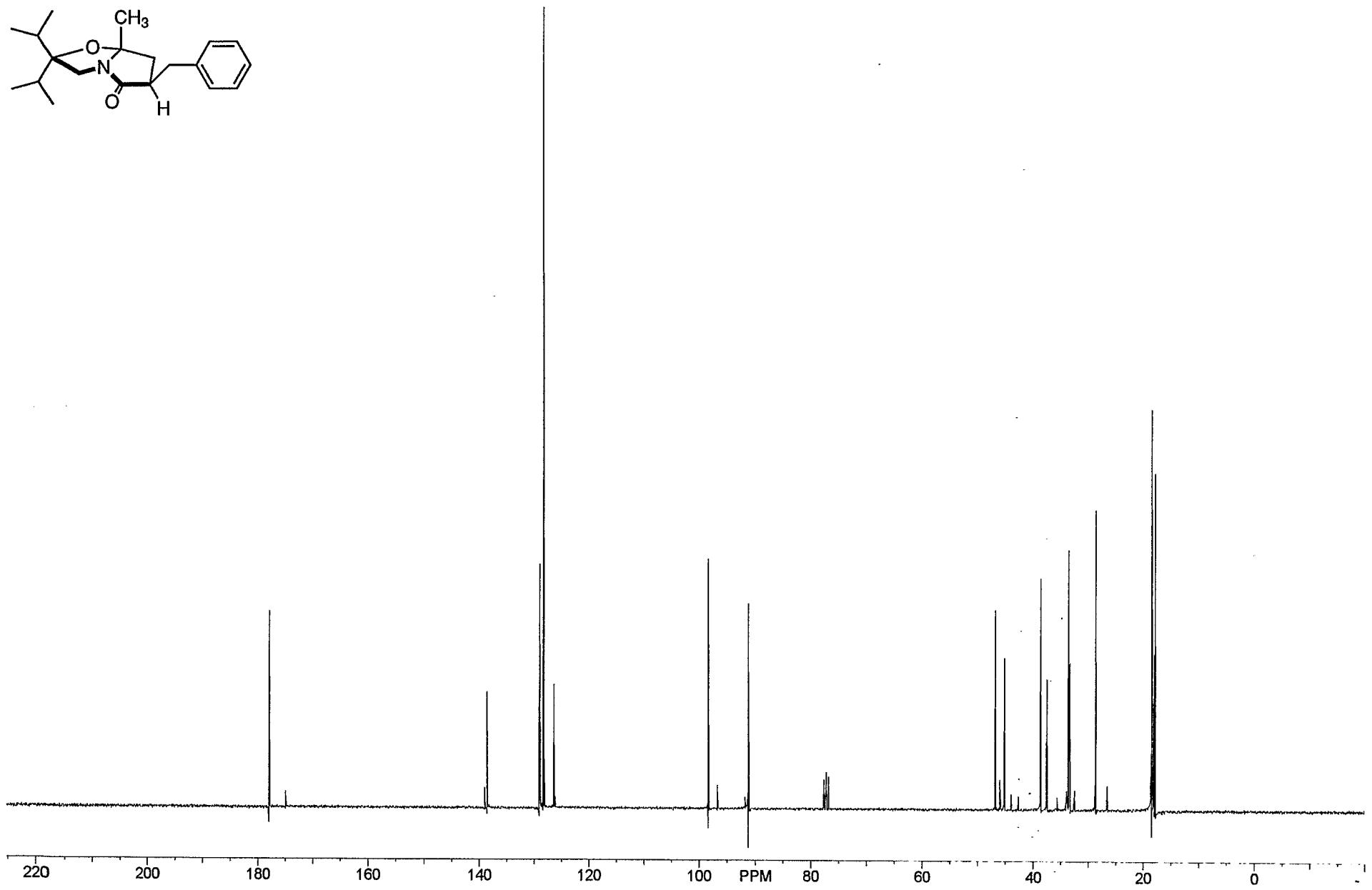
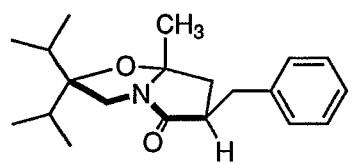
Filename: MAS124H.111



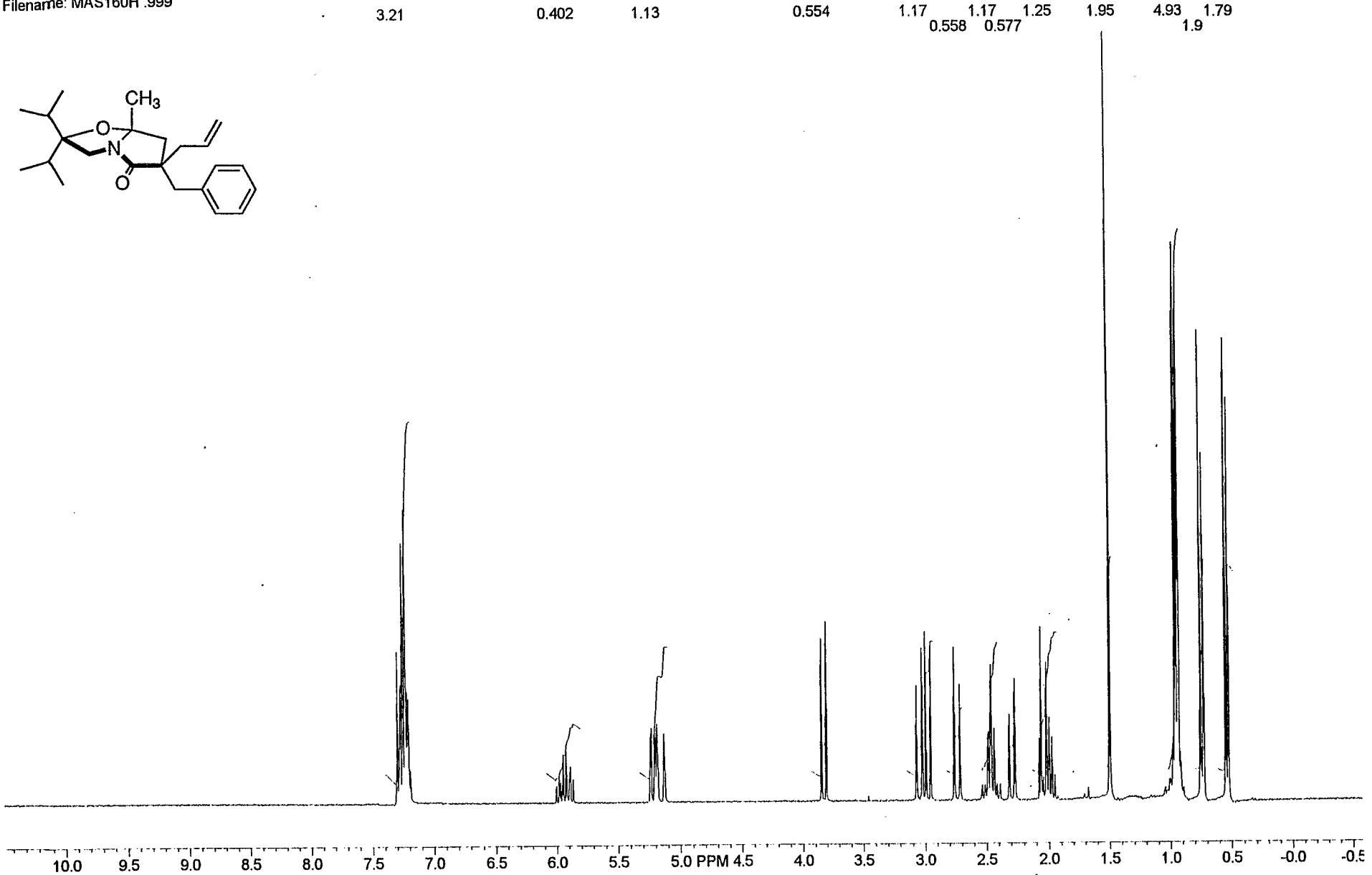
Filename: MAS124H.111
0.0622



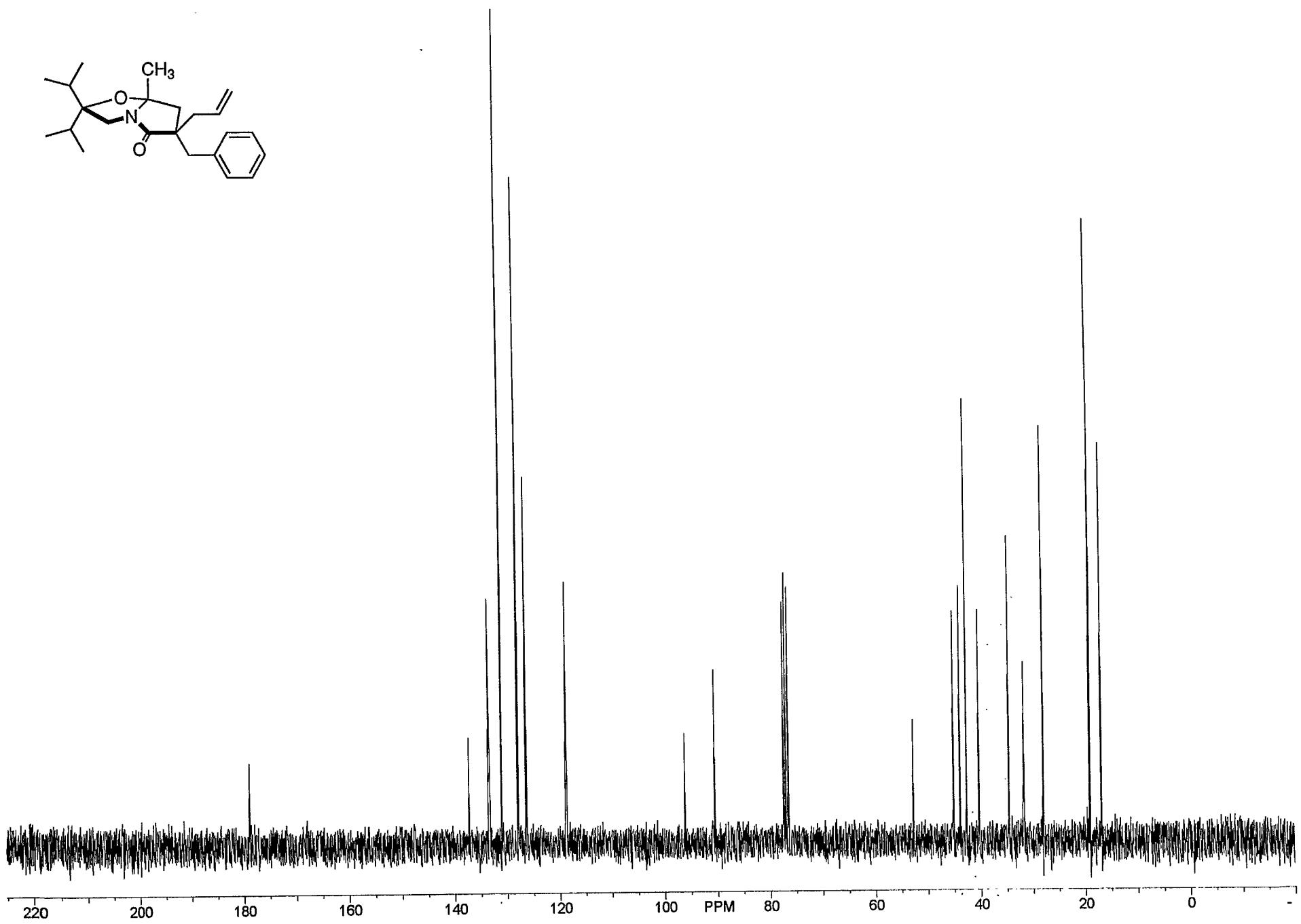
Filename: MAS124C.111



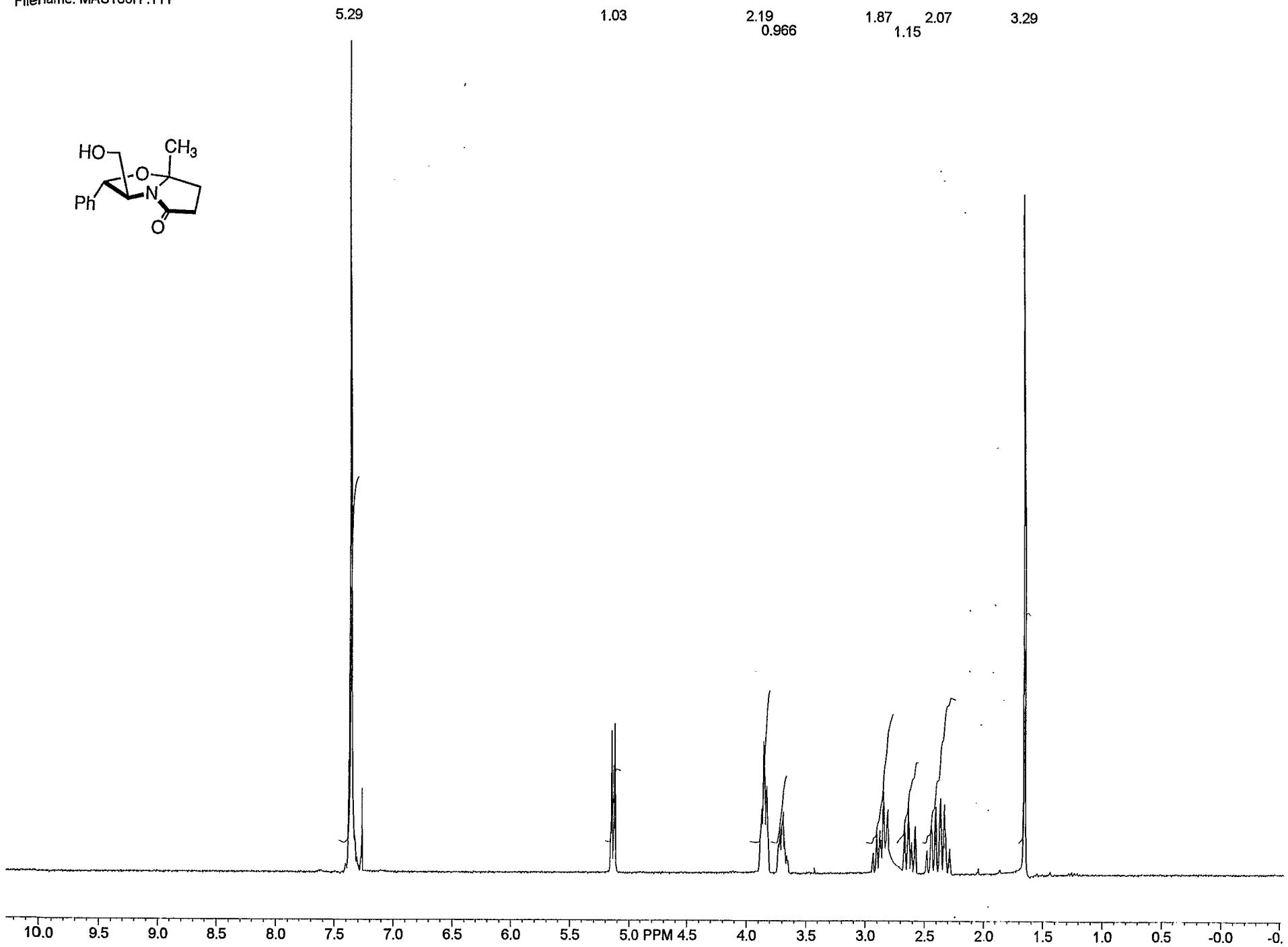
Filename: MAS160H.999



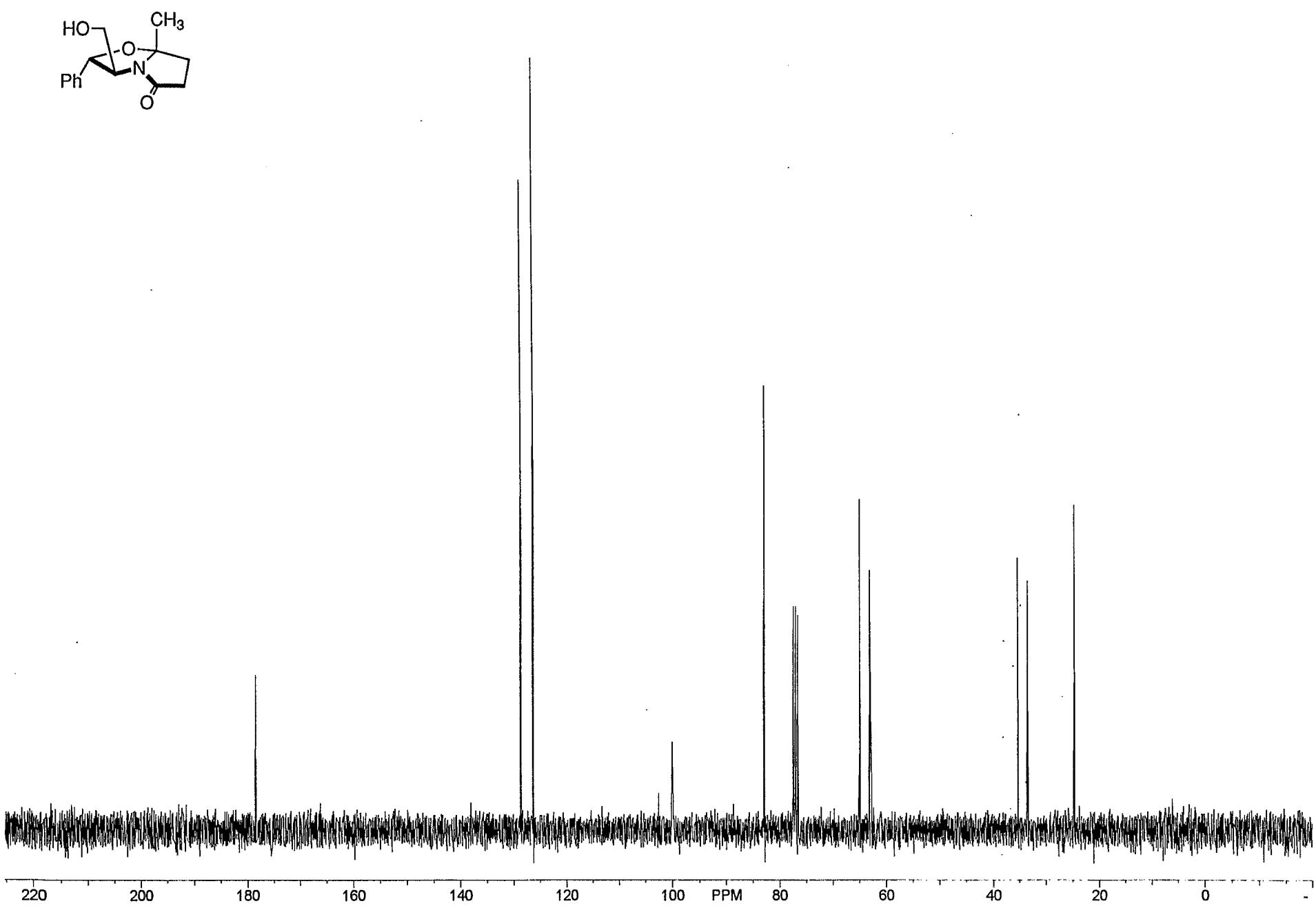
Filename: MAS160C .123



Filename: MAS156H.111



Filename: MAS156C .222



Filename: MASTBDPS.111

— 8.27 —

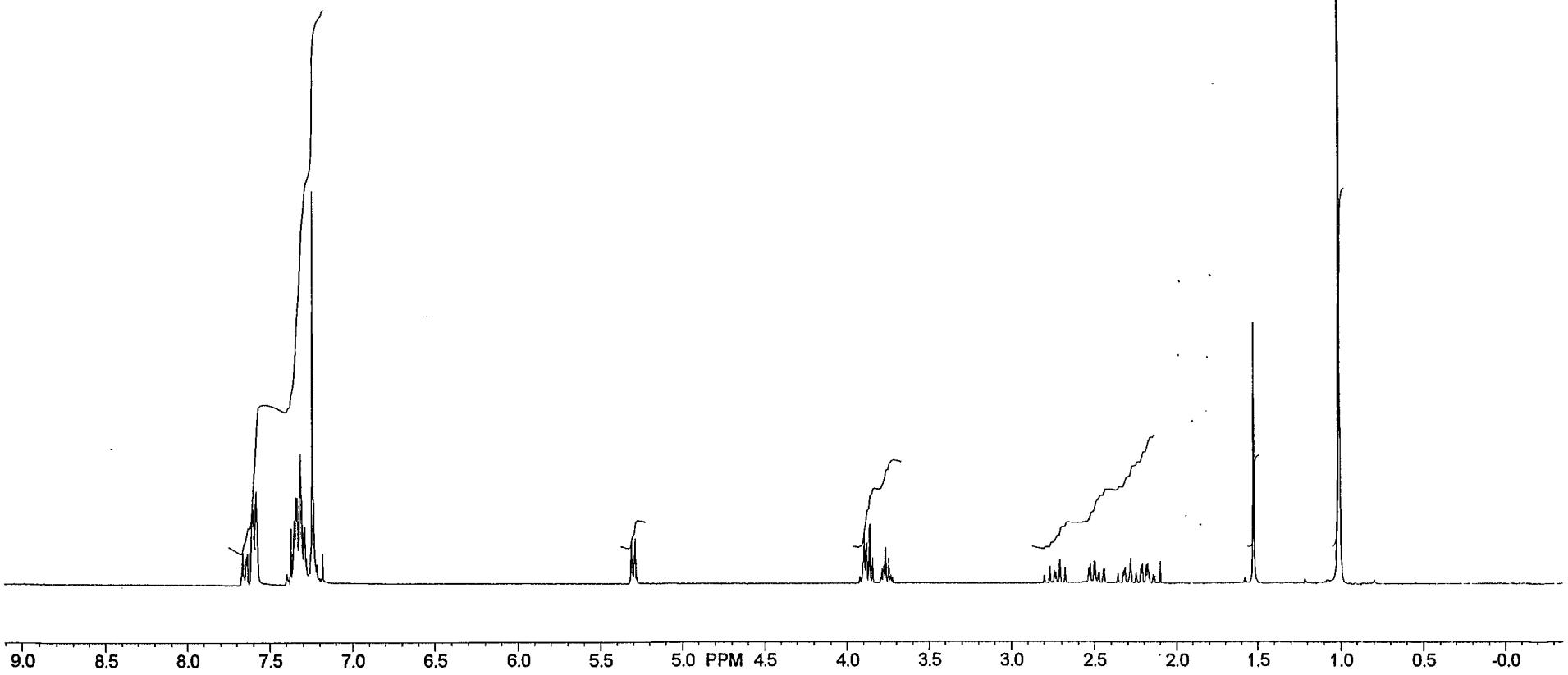
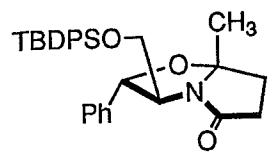
0.381

1.31

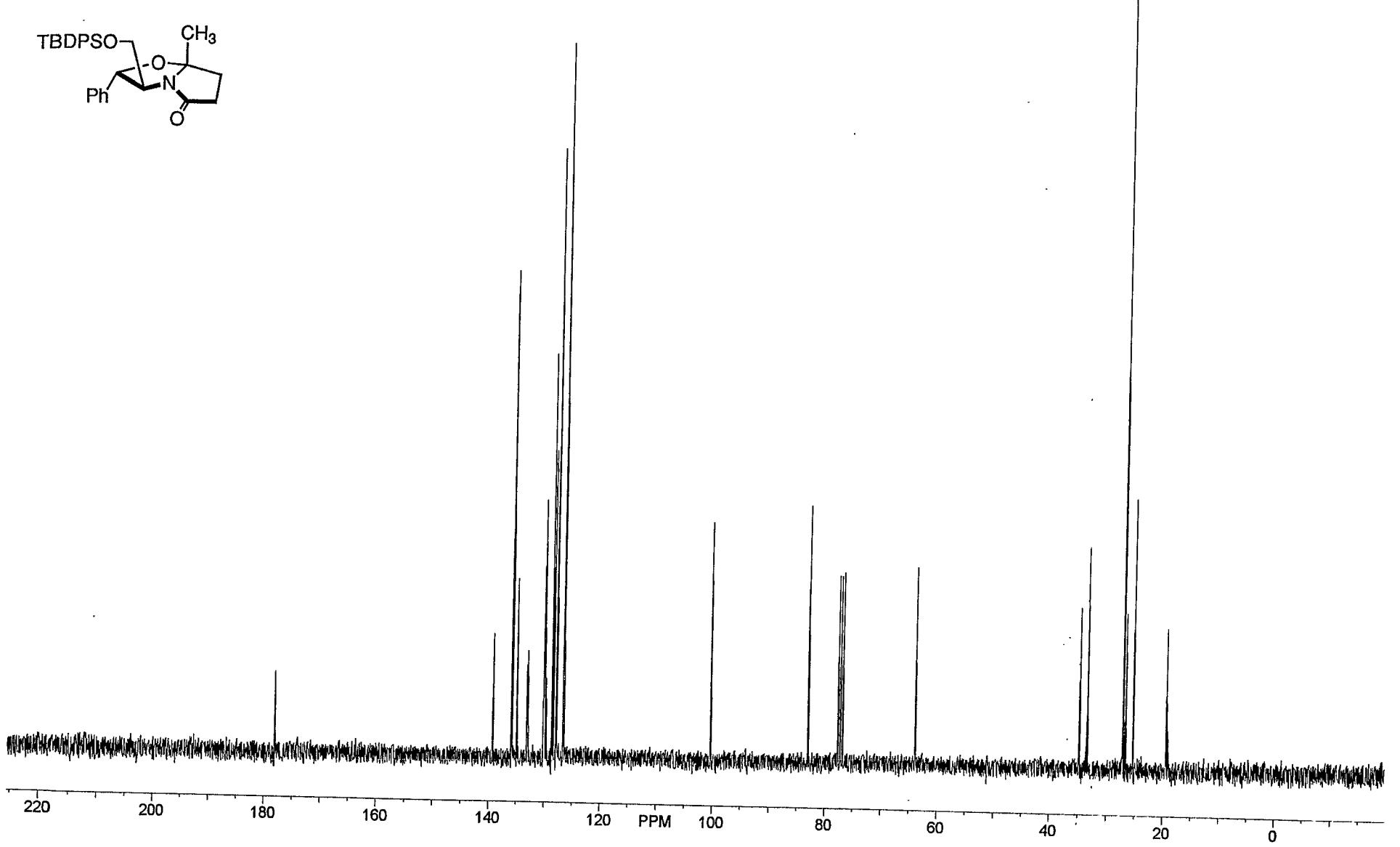
— 1.73 —

1.41

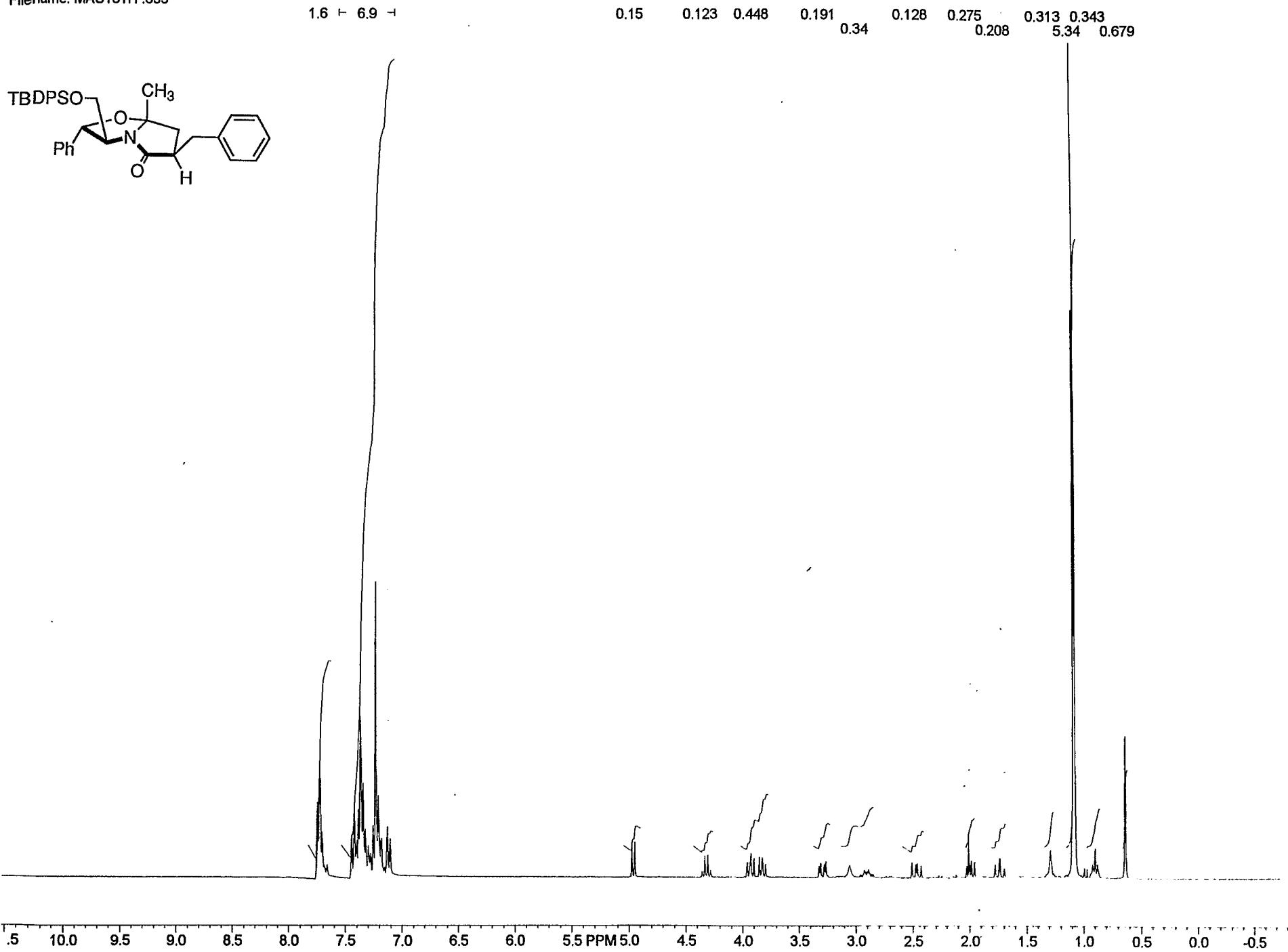
5.53

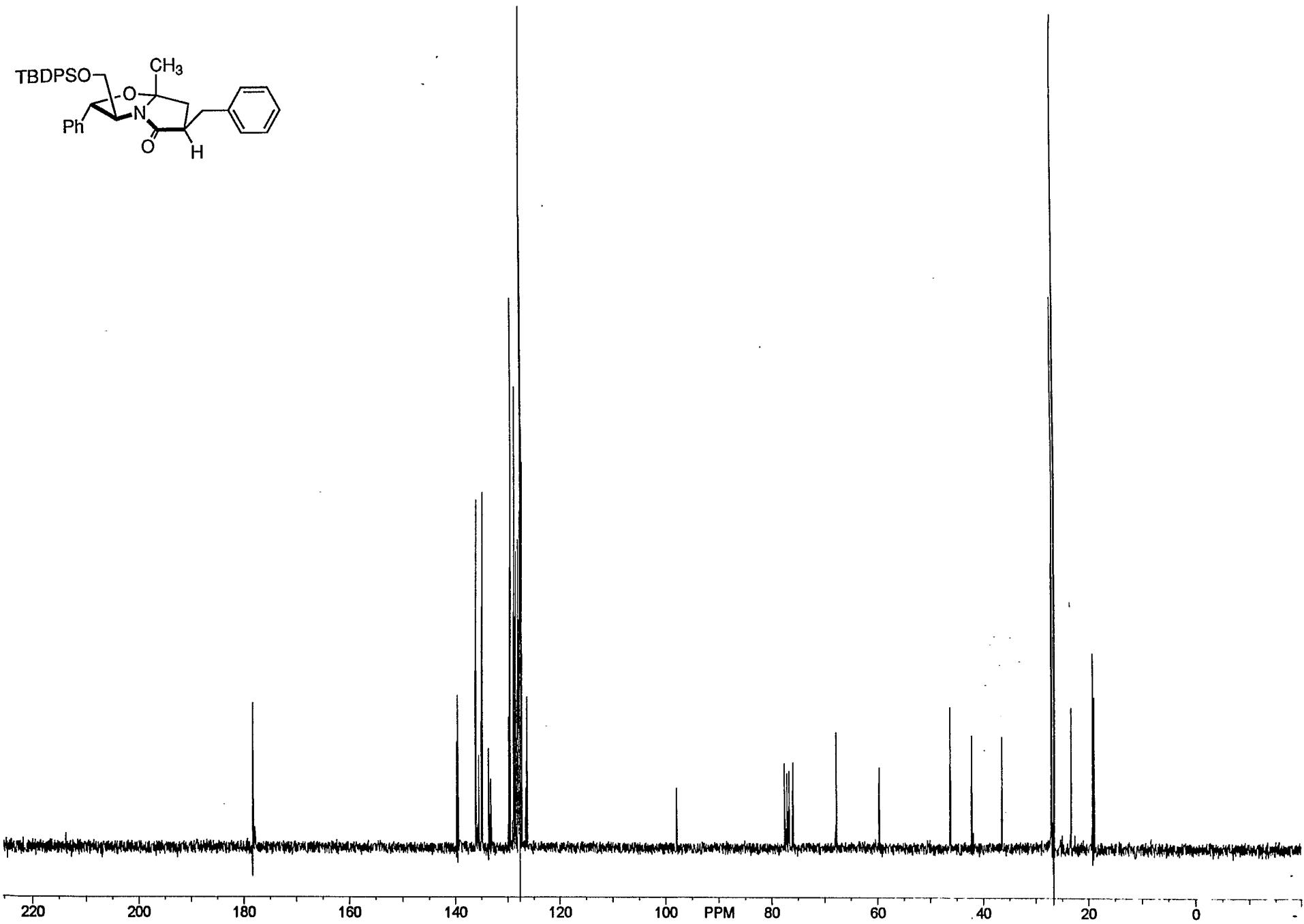
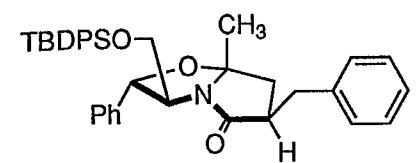


Filename: MAS120C .555



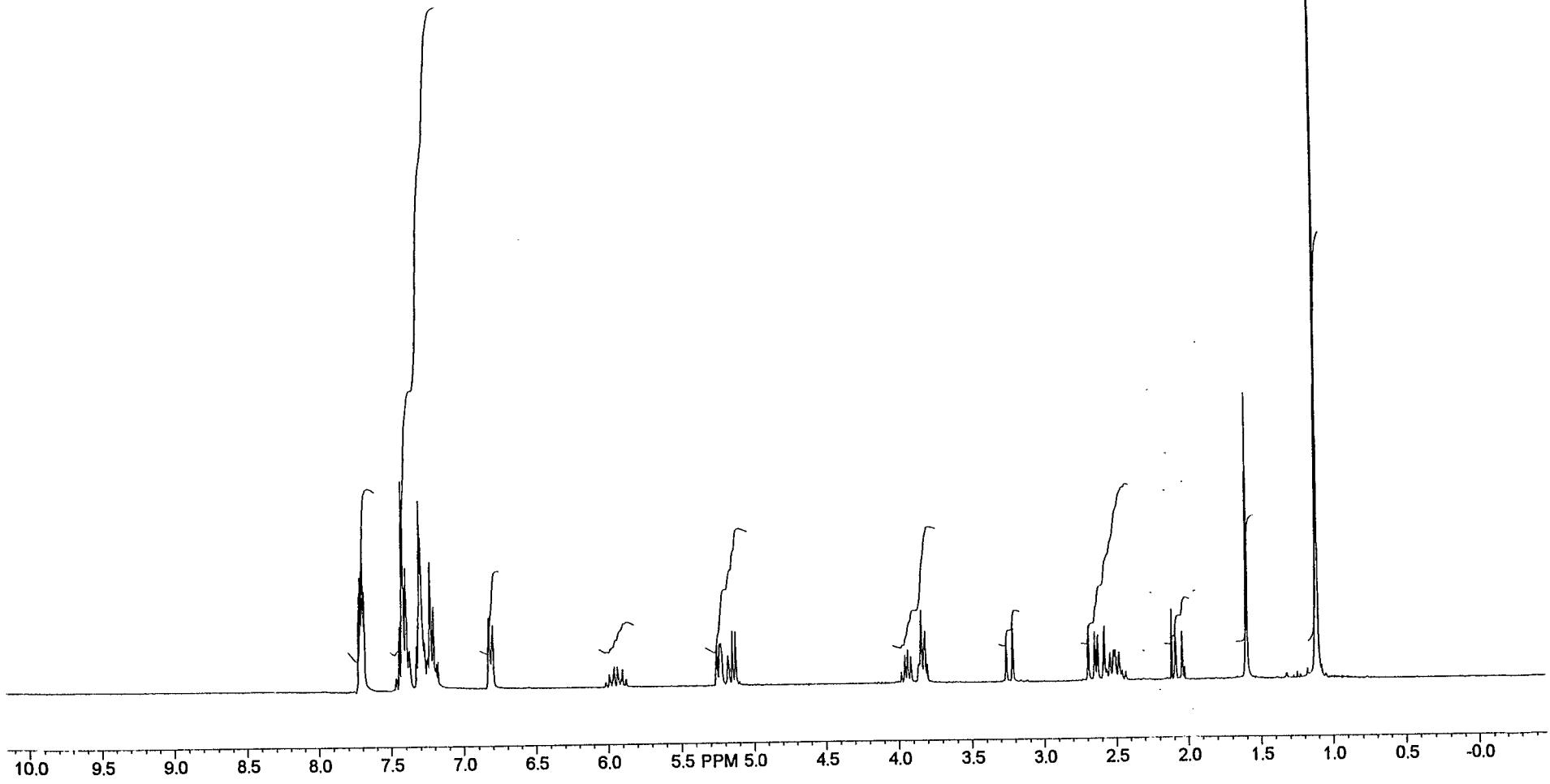
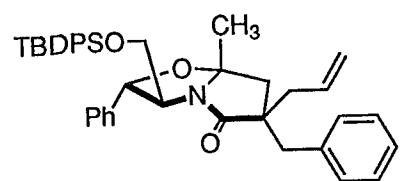
Filename: MAS131H.555



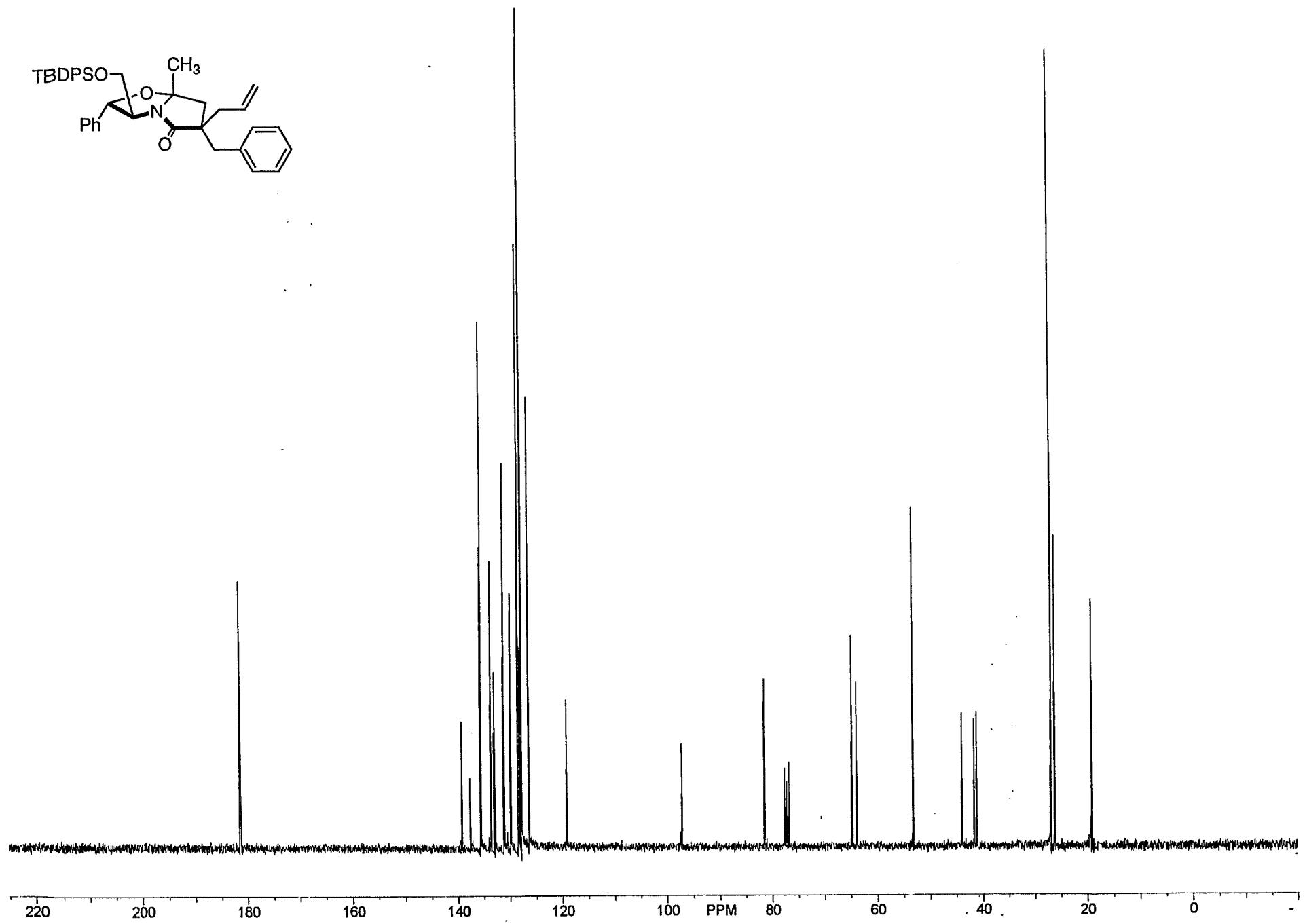


Filename: MAS147H.123

2.6 10.5 1.29 0.398 1.89 1.9 0.534 2.59 0.711 2.06 6.7



Filename: MAS147C.123

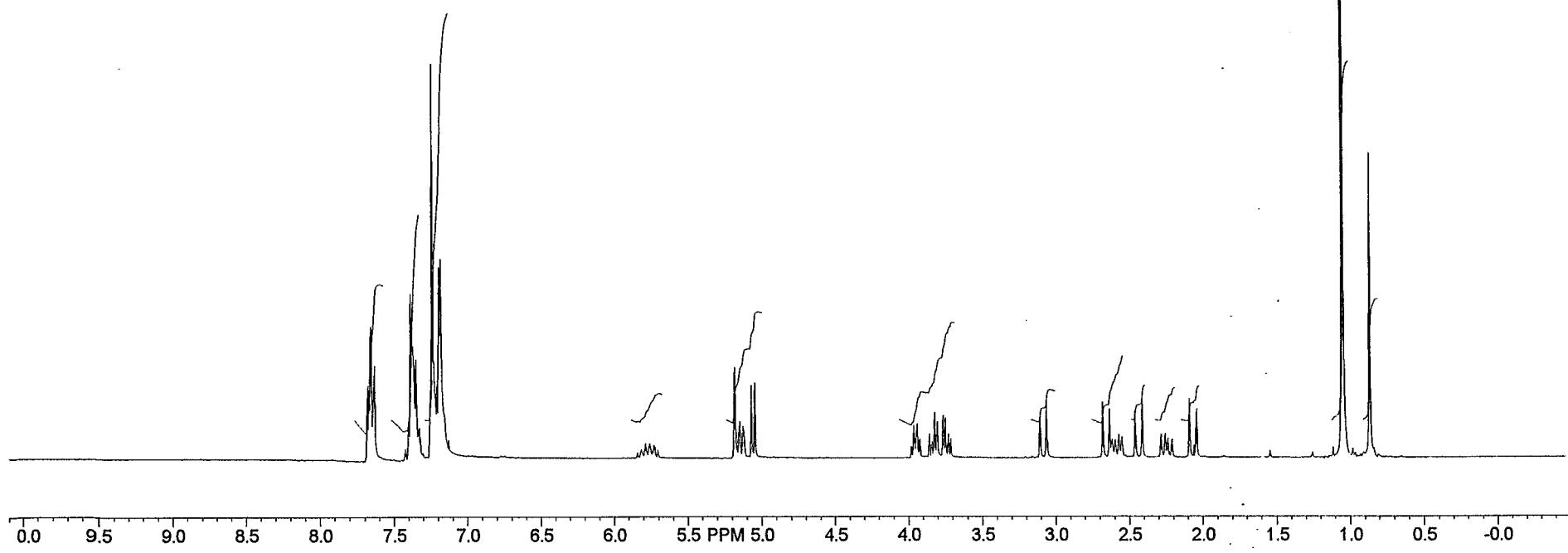
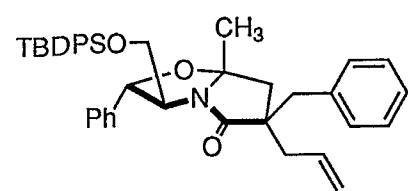


1.47
2.27

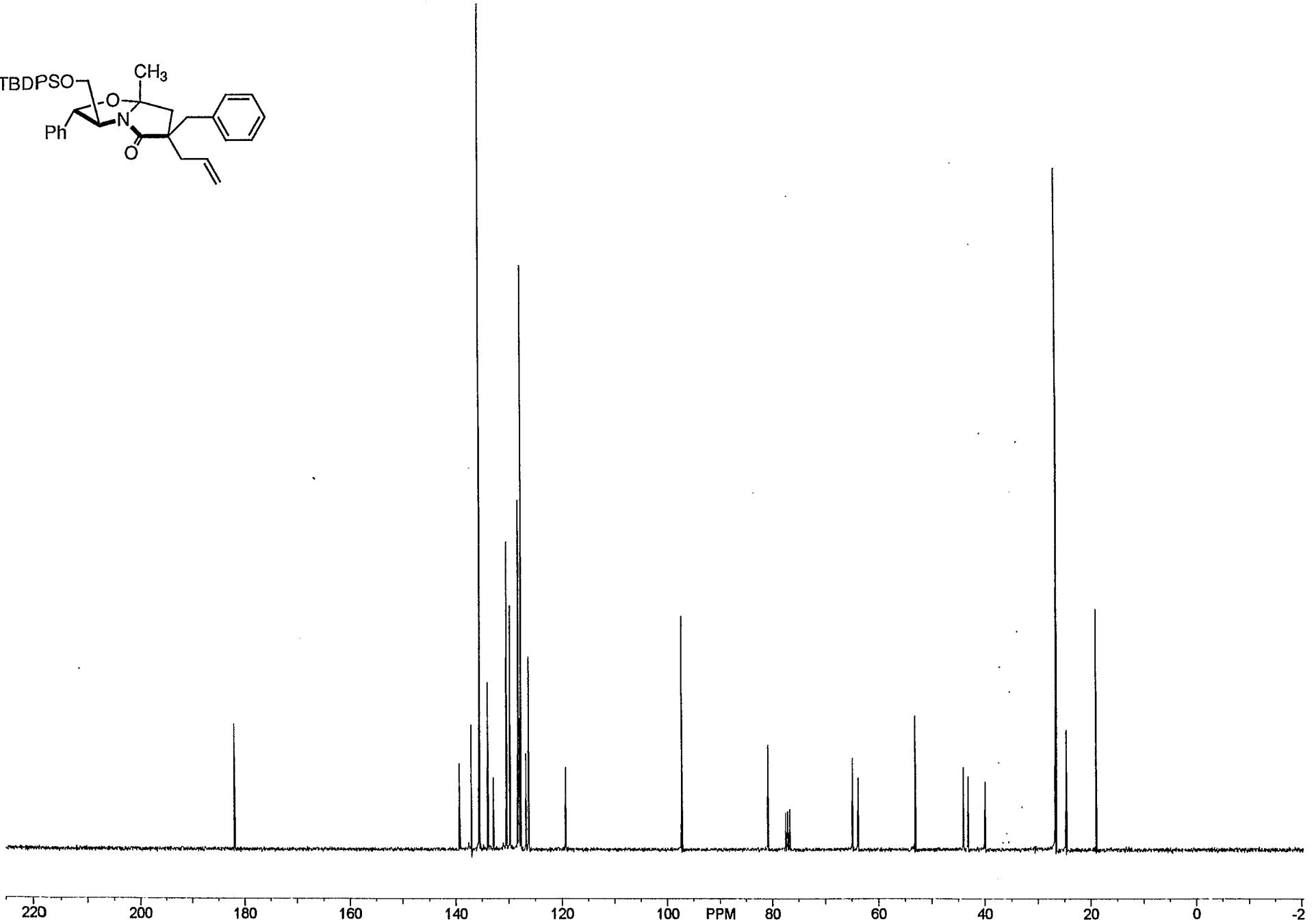
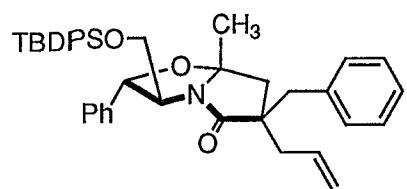
0.282

1.18

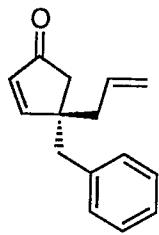
1.06

0.314
0.381
0.712
0.349
0.3733.94
1.33

Filename: MASALBNC.000



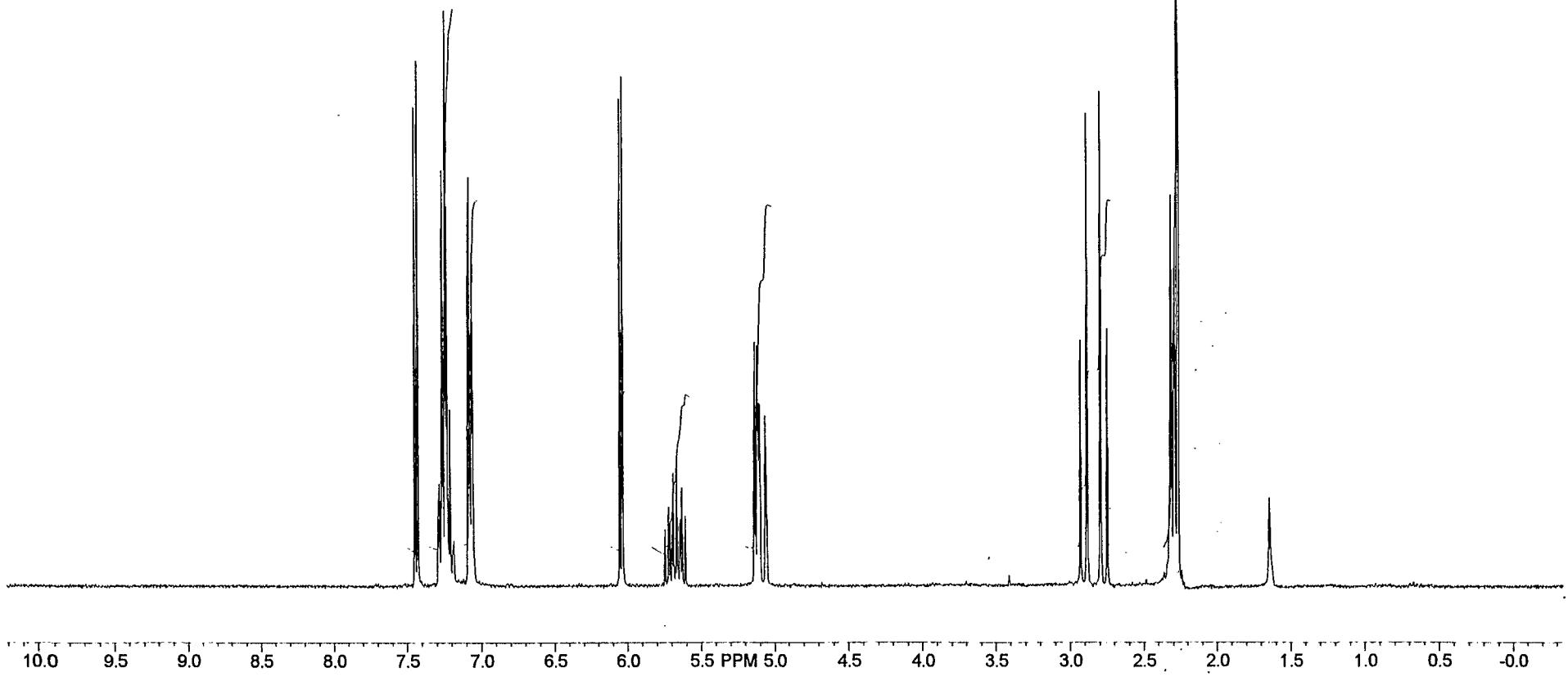
Filename: MAS149H .001



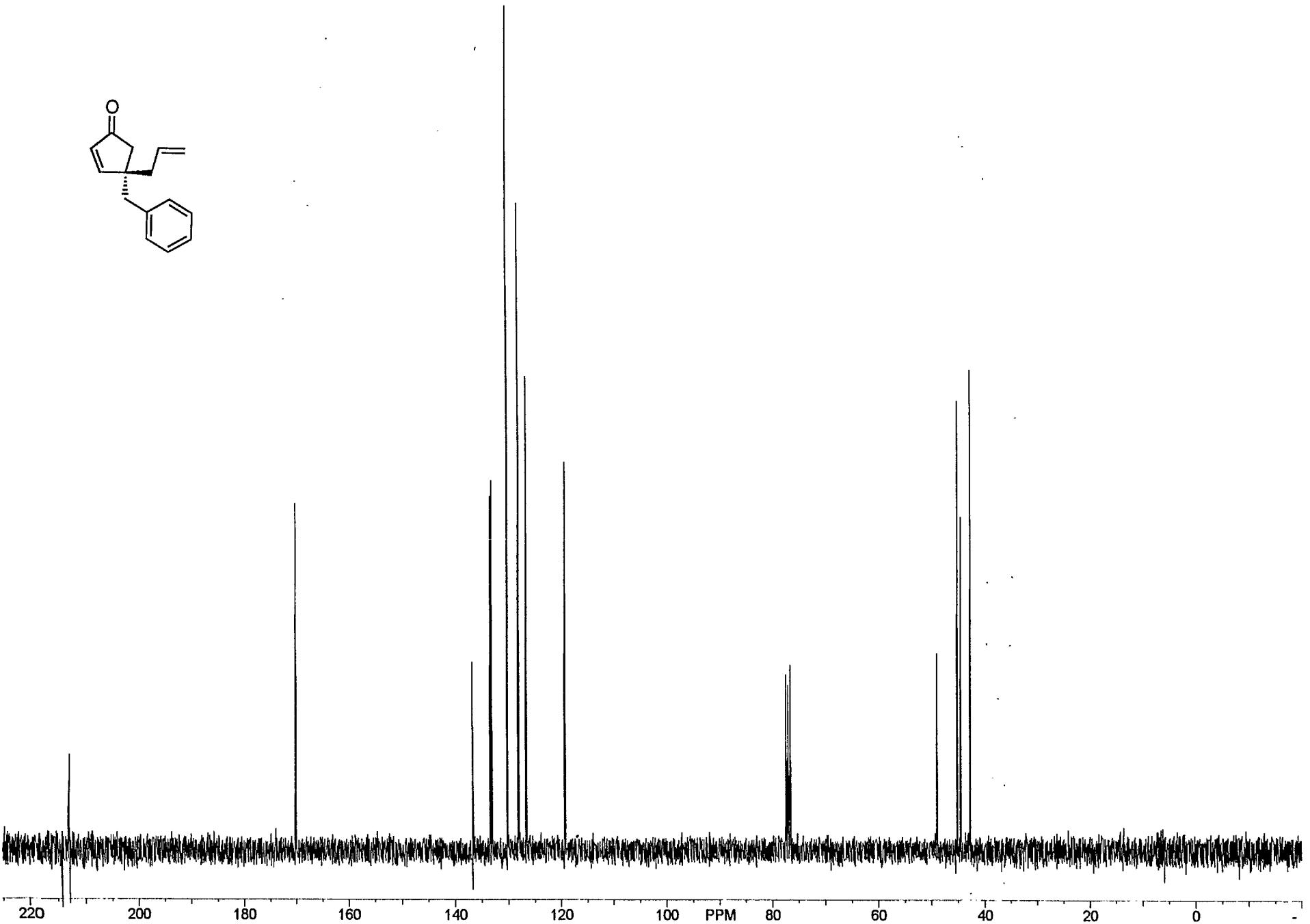
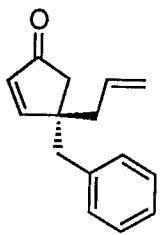
1.24 2.69
4.18

1.2 1.16 2.64

2.69 5.55



Filename: MAS149C .222



Filename: MAS167H .333

