

Experimental Section

Chemistry. Melting points were determined on a Kofler hot stage apparatus and are uncorrected. Elemental analyses (C, H, N) were carried out on a C. Erba Model 1106 Elemental Analyzer and the results were within $\pm 0.4\%$ of the theoretical values. Merck silica gel 60 F₂₅₄ plates were used for TLC. ¹H NMR spectra were measured with a Varian Gemini 300 spectrometer in CDCl₃ with TMS as internal standard; chemical shifts are expressed in δ (ppm) and coupling constants (J) in hertz.

General Procedure for 2,3-Diaryl-1,3-thiazolidin-4-ones 4-34. To a stirred solution of aromatic amine (1) (8 mmol) in dry toluene (50 mL), 2-mercaptopropanoic acid (2) (16 mmol) and the appropriate aromatic aldehyde (3) (8 mmol) were added. The reaction mixture was refluxed for 48 h and then neutralized by a solution of sodium hydrogen carbonate. After removal of the solvent under reduced pressure, the oily residue was powdered by treatment with a mixture of ethanol and diethyl ether. All compounds were recrystallized from ethanol.

Physical and spectral data of compounds 4-34

2-(2,6-Dichlorophenyl)-3-phenyl-1,3-thiazolidin-4-one (4). Mp: 128-129 °C, yield 78%. ¹H NMR: 3.96 (d, 1H, $J = 15.4$, 5-H_A), 4.09 (dd, 1H, $J = 2.2$ and 15.4, 5-H_B), 7.06-7.35 (m, 9H, ArH and H-2). Anal. (C₁₅H₁₁Cl₂NOS) C, H, N.

2-(2,6-Difluorophenyl)-3-phenyl-1,3-thiazolidin-4-one (5). Mp: 144-145 °C, yield 65%. ¹H NMR: 3.87 (d, 1H, $J = 15.4$, 5-H_A), 4.14 (dd, 1H, $J = 1.9$ and 15.4, 5-H_B), 6.58 (d, 1H, $J = 1.9$, H-2), 6.82 (m, 2H, H-3',5'), 7.17-7.34 (m, 6H, ArH). Anal. (C₁₅H₁₁F₂NOS) C, H, N.

2-(2,6-Dichlorophenyl)-3-pyridin-2-yl-1,3-thiazolidin-4-one (6). Mp: 135-138 °C, yield 8%. ¹H NMR: 3.94 (d, 1H, $J = 15.7$, 5-H_A), 4.17 (dd, 1H, $J = 2.2$ and 15.7, 5-H_B), 6.98-7.40 (m, 4H, ArH and H_{5-Py}), 7.63 (d, 1H, $J = 2.2$, H-2), 7.67 (m, 1H, H_{4-Py}), 8.13 (d, 1H, $J = 8.5$, H_{3-Py}), 8.20 (m, 1H, H_{6-Py}). Anal. (C₁₄H₁₀Cl₂N₂OS) C, H, N.

2-(2-Chloro,6-fluorophenyl)-3-pyridin-2-yl-1,3-thiazolidin-4-one (7). Mp: 112-114 °C, yield 45%. ¹H NMR: 3.84 (d, 1H, $J = 15.9$, 5-H_A), 4.19 (dd, 1H, $J = 1.4$ and 15.9, 5-H_B), 6.88-7.35 (m, 5H, ArH, H-2 and H_{5-Py}), 7.68 (m, 1H, H_{4-Py}), 8.23 (m, 2H, H_{3,6-Py}). Anal. (C₁₄H₁₀ClFN₂OS) C, H, N.

2-(2,6-Difluorophenyl)-3-pyridin-2-yl-1,3-thiazolidin-4-one (8). Mp: 115-118 °C, yield 18%. ^1H NMR: 3.84 (d, 1H, $J = 15.9$, 5-H_A), 4.22 (dd, 1H, $J = 1.1$ and 15.9, 5-H_B), 6.81 (m, 2H, H-3',5'), 7.0 (m, 1H, H₅-Py), 7.16 (m, 1H, H-4'), 7.20 (d, 1H, $J = 1.1$, H-2), 7.67 (m, 1H, H₄-Py), 8.20 (d, 1H, $J = 8.5$, H₆-Py), 8.24 (m, 1H, H₃-Py). Anal. (C₁₄H₁₀F₂N₂OS) C, H, N.

2-(2,6-Dichlorophenyl)-3-pyridin-3-yl-1,3-thiazolidin-4-one (9). Mp: 129-130 °C, yield 10%. ^1H NMR: 4.00 (d, 1H, $J = 15.7$, 5-H_A), 4.13 (dd, 1H, $J = 2.2$ and 15.7, 5-H_B), 7.13-7.31 (m, 5H, ArH, H-2 and H₅-Py), 7.81 (m, 1H, H₄-Py), 8.44 (dd, 1H, $J = 1.6$ and 4.8, H₆-Py), 8.64 (m, 1H, $J = 2.5$, H₂-Py). Anal. (C₁₄H₁₀Cl₂N₂OS) C, H, N.

2-(2,6-Difluorophenyl)-3-pyridin-3-yl-1,3-thiazolidin-4-one (10). Mp: 171-173 °C, yield 40%. ^1H NMR: 3.91 (d, 1H, $J = 15.7$, 5-H_A), 4.17 (dd, 1H, $J = 1.4$ and 15.7, 5-H_B), 6.65 (d, 1H, $J = 1.4$, H-2), 6.87 (m, 2H, H-3',5'), 7.21-7.32 (m, 2H, H-4' and H₅-Py), 7.72 (m, 1H, H₄-Py), 8.46 (dd, 1H, $J = 1.4$ and 4.7, H₆-Py), 8.58 (d, 1H, $J = 2.4$, H₂-Py). Anal. (C₁₄H₁₀F₂N₂OS) C, H, N.

2-(2,6-Dichlorophenyl)-3-pyridin-4-yl-1,3-thiazolidin-4-one (11). Mp: 137-138 °C, yield 14%. ^1H NMR: 3.95 (d, 1H, $J = 15.9$, 5-H_A), 4.12 (dd, 1H, $J = 2.1$ and 15.9, 5-H_B), 7.11-7.31 (m, 4H, ArH, H-2), 7.39 (d, 2H, $J = 6.4$, H_{3,5}-Py), 8.51 (d, 2H, $J = 6.4$, H_{2,6}-Py). Anal. (C₁₄H₁₀Cl₂N₂OS) C, H, N.

2-(2,6-Difluorophenyl)-3-pyridin-4-yl-1,3-thiazolidin-4-one (12). Mp: 114-117 °C, yield 15%. ^1H NMR: 3.85 (d, 1H, $J = 15.9$, 5-H_A), 4.20 (dd, 1H, $J = 1.4$ and 15.9, 5-H_B), 6.67 (d, 1H, $J = 1.4$, H-2), 6.86 (m, 2H, H-3',5'), 7.25 (m, 1H, H-4'), 7.41 (d, 2H, $J = 6.4$, H_{3,5}-Py), 8.52 (d, 2H, $J = 6.4$, H_{2,6}-Py). Anal. (C₁₄H₁₀F₂N₂OS) C, H, N.

3-(5-Chloropyridin-2-yl)-2-(2,6-dichlorophenyl)-1,3-thiazolidin-4-one (13). Mp: 138-141 °C, yield 53%. ^1H NMR: 3.93 (d, 1H, $J = 15.9$, 5-H_A), 4.16 (dd, 1H, $J = 2.2$ and 15.9, 5-H_B), 7.05-7.31 (m, 3H, ArH), 7.55 (d, 1H, $J = 2.2$, H-2), 7.64 (dd, 1H, $J = 2.5$ and 8.7, H₄-Py), 8.14 (d, 1H, $J = 2.5$, H₆-Py), 8.17 (d, 1H, $J = 8.7$, H₃-Py). Anal. (C₁₄H₉Cl₃N₂OS) C, H, N.

2-(2-Chloro-6-fluorophenyl)-3-(5-chloropyridin-2-yl)-1,3-thiazolidin-4-one (14). Mp: 91-94 °C, yield 41%. ^1H NMR: 3.84 (d, 1H, $J = 15.9$, 5-H_A), 4.18 (dd, 1H, $J = 2.2$ and 15.9, 5-H_B), 6.87-7.16 (m, 4H, ArH and H-2), 7.64 (dd, 1H, $J = 2.5$ and 8.9, H₄-Py), 8.17 (d, 1H, $J = 2.5$, H₆-Py), 8.26 (d, 1H, $J = 8.9$, H₃-Py). Anal. (C₁₄H₉Cl₂FN₂OS) C, H, N.

3-(5-Chloropyridin-2-yl)-2-(2,6-difluorophenyl)-1,3-thiazolidin-4-one (15). Mp: 95-98 °C, yield 66%. ^1H NMR: 3.84 (d, 1H, $J = 15.9$, 5-H_A), 4.22 (dd, 1H, $J = 1.1$ and 15.9, 5-H_B), 6.83 (m,

2H, H-3',5'), 7.12 (d, 1H, $J = 1.1$, H-2), 7.18 (m, 1H, H-4'), 7.64 (dd, 1H, $J = 2.8$ and 9.1, H₄-Py), 8.19 (d, 1H, $J = 2.8$, H₆-Py), 8.26 (d, 1H, $J = 9.1$, H₃-Py). Anal. (C₁₄H₉ClF₂N₂OS) C, H, N.

3-(5-Bromopyridin-2-yl)-2-(2,6-dichlorophenyl)-1,3-thiazolidin-4-one (16). Mp: 148-151 °C, yield 79%. ¹H NMR: 3.93 (d, 1H, $J = 15.9$, 5-H_A), 4.16 (dd, 1H, $J = 2.2$ and 15.9, 5-H_B), 7.06-7.31 (m, 3H, ArH), 7.54 (d, 1H, $J = 2.2$, H-2), 7.77 (dd, 1H, $J = 2.6$ and 8.8, H₄-Py), 8.13 (d, 1H, $J = 8.8$, H₃-Py), 8.24 (d, 1H, $J = 2.6$, H₆-Py). Anal. (C₁₄H₉BrCl₂N₂OS) C, H, N.

3-(5-Bromopyridin-2-yl)-2-(2-chloro-6-fluorophenyl)-1,3-thiazolidin-4-one (17). Mp: 90-91 °C, yield 41%. ¹H NMR: 3.84 (d, 1H, $J = 15.9$, 5-H_A), 4.18 (dd, 1H, $J = 1.6$ and 15.9, 5-H_B), 6.88-7.25 (m, 4H, ArH and H-2), 7.78 (dd, 1H, $J = 2.5$ and 9.1, H₄-Py), 8.23 (d, 1H, $J = 9.1$, H₃-Py), 8.27 (d, 1H, $J = 2.5$, H₆-Py). Anal. (C₁₄H₉BrClFN₂OS) C, H, N.

3-(5-Bromopyridin-2-yl)-2-(2,6-difluorophenyl)-1,3-thiazolidin-4-one (18). Mp: 88-90 °C, yield 68%. ¹H NMR: 3.83 (d, 1H, $J = 15.8$, 5-H_A), 4.21 (dd, 1H, $J = 1.4$ and 15.8, 5-H_B), 6.83 (m, 2H, H-3',5'), 7.12 (d, 1H, $J = 1.4$, H-2), 7.19 (m, 1H, H-4'), 7.77 (dd, 1H, $J = 2.4$ and 8.2, H₄-Py), 8.22 (d, 1H, $J = 8.2$, H₃-Py), 8.29 (d, 1H, $J = 2.4$, H₆-Py). Anal. (C₁₄H₉BrF₂N₂OS) C, H, N.

3-(6-Bromopyridin-2-yl)-2-(2,6-dichlorophenyl)-1,3-thiazolidin-4-one (19). Mp: 162-164 °C, yield 10%. ¹H NMR: 3.94 (d, 1H, $J = 15.9$, 5-H_A), 4.16 (dd, 1H, $J = 2.2$ and 15.9, 5-H_B), 7.07-7.36 (m, 4H, ArH and H₅-Py), 7.53 (dd, 1H, $J = 7.8$ and 8.2, H₄-Py), 7.54 (d, 1H, $J = 2.2$, H-2), 8.19 (d, 1H, $J = 8.2$, H₃-Py). Anal. (C₁₄H₉BrCl₂N₂OS) C, H, N.

3-(6-Bromopyridin-2-yl)-2-(2-chloro-6-fluorophenyl)-1,3-thiazolidin-4-one (20). Mp: 104-106 °C, yield 74%. ¹H NMR: 3.85 (d, 1H, $J = 15.9$, 5-H_A), 4.19 (dd, 1H, $J = 2.2$ and 15.9, 5-H_B), 6.89-7.19 (m, 5H, ArH, H-2 and H₅-Py), 7.52 (dd, 1H, $J = 7.8$ and 8.2, H₄-Py), 8.26 (d, 1H, $J = 8.2$, H₃-Py). Anal. (C₁₄H₉BrClFN₂OS) C, H, N.

3-(6-Bromopyridin-2-yl)-2-(2,6-difluorophenyl)-1,3-thiazolidin-4-one (21). Mp: 126-130 °C, yield 60%. ¹H NMR: 3.84 (d, 1H, $J = 15.9$, 5-H_A), 4.23 (dd, 1H, $J = 1.4$ and 15.9, 5-H_B), 6.86 (m, 2H, H-3',5'), 7.08 (d, 1H, $J = 1.4$, H-2), 7.16-7.25 (m, 2H, H-4' and H₅-Py), 7.52 (dd, 1H, $J = 7.9$ and 8.2, H₄-Py), 8.29 (d, 1H, $J = 8.2$, H₃-Py). Anal. (C₁₄H₉BrF₂N₂OS) C, H, N.

2-(2,6-Dichlorophenyl)-3-(3-methylpyridin-2-yl)-1,3-thiazolidin-4-one (22). Mp: 159-160 °C, yield 20%. ¹H NMR: 2.26 (s, 3H, Me), 3.94 (d, 1H, $J = 15.6$, 5-H_A), 4.14 (dd, 1H, $J = 2.2$ and 15.6, 5-H_B), 7.05-7.33 (m, 4H, ArH and H₅-Py), 7.49 (dd, 1H, $J = 1.4$ and 7.6, H₄-Py), 7.65 (d, 1H, $J = 2.2$, H-2), 8.25 (dd, 1H, $J = 1.4$ and 4.7, H₃-Py). Anal. (C₁₅H₁₂Cl₂N₂OS) C, H, N.

2-(2,6-Difluorophenyl)-3-(3-methylpyridin-2-yl)-1,3-thiazolidin-4-one (23). Mp: 131-133 °C, yield 21%. ^1H NMR: 2.17 (s, 3H, Me), 3.84 (d, 1H, 15.4, 5-H_A), 4.18 (dd, 1H, J = 1.9 and 15.4, 5-H_B), 6.81 (m, 2H, H-3',5'), 7.05 (d, 1H, J = 1.9, H-2), 7.09 (dd, 1H, J = 4.7 and 7.7, H_{5-Py}), 7.20 (m, 1H, H-4'), 7.49 (dd, 1H, J = 1.4 and 7.7, H_{4-Py}), 8.27 (dd, 1H, J = 1.7 and 4.7, H_{5-Py}). Anal. (C₁₅H₁₂F₂N₂OS) C, H, N.

2-(2,6-Dichlorophenyl)-3-(4-methylpyridin-2-yl)-1,3-thiazolidin-4-one (24). Mp: 192 °C, yield 71%. ^1H NMR: 2.38 (s, 3H, Me), 3.97 (d, 1H, J = 15.6, 5-H_A), 4.20 (dd, 1H, J = 1.9 and 15.6, 5-H_B), 6.85 (d, 1H, J = 4.9, H_{5-Py}), 7.08-7.33 (m, 3H, ArH), 7.67 (d, 1H, J = 1.9, H-2), 7.96 (s, 1H, H_{3-Py}), 8.09 (d, 1H, J = 4.9, H_{6-Py}). Anal. (C₁₅H₁₂Cl₂N₂OS) C, H, N.

2-(2-Chloro-6-fluorophenyl)-3-(4-methylpyridin-2-yl)-1,3-thiazolidin-4-one (25). Mp: 130-133 °C, yield 40%. ^1H NMR: 2.34 (s, 3H, Me), 3.84 (d, 1H, J = 15.4, 5-H_A), 4.18 (dt, 1H, J = 1.6 and 15.4, 5-H_B), 6.82 (d, 1H, J = 5.2, H_{5-Py}), 6.86-7.34 (m, 4H, ArH and H-2), 8.02 (s, 1H, H_{3-Py}), 8.08 (d, 1H, J = 5.2, H_{6-Py}). Anal. (C₁₅H₁₂ClFN₂OS) C, H, N.

2-(2,6-Difluorophenyl)-3-(4-methylpyridin-2-yl)-1,3-thiazolidin-4-one (26). Mp: 108-110 °C, yield 42%. ^1H NMR: 2.33 (s, 3H, Me), 3.83 (d, 1H, 15.9, 5-H_A), 4.21 (dd, 1H, J = 1.6 and 15.9, 5-H_B), 6.70-6.83 (m, 3H, H-3',5' and H_{5-Py}), 7.14 (m, 1H, H-4'), 7.20 (d, 1H, J = 1.6, H-2), 8.00 (s, 1H, H_{3-Py}), 8.08 (d, 1H, J = 5.0, H_{6-Py}). Anal. (C₁₅H₁₂F₂N₂OS) C, H, N.

2-(2,6-Dichlorophenyl)-3-(5-methylpyridin-2-yl)-1,3-thiazolidin-4-one (27). Mp: 157-159 °C; yield 85%. ^1H NMR: 2.22 (s, 3H, Me), 3.93 (d, 1H, J = 15.7, 5-H_A), 4.15 (dd, 1H, J = 2.2 and 15.7, 5-H_B), 7.02-7.29 (m, 3H, ArH), 7.47 (dd, 1H, J = 1.9 and 8.5, H_{4-Py}), 7.61 (d, 1H, J = 2.2, H-2), 7.97 (d, 1H, J = 8.5, H_{3-Py}), 8.03 (d, 1H, J = 1.9, H_{6-Py}). Anal. (C₁₅H₁₂Cl₂N₂OS) C, H, N.

2-(2,6-Difluorophenyl)-3-(5-methylpyridin-2-yl)-1,3-thiazolidin-4-one (28). Mp: 99-101 °C, yield 18%. ^1H NMR: 2.21 (s, 3H, Me), 3.83 (d, 1H, 15.6, 5-H_A), 4.20 (dd, 1H, J = 1.4 and 15.6, 5-H_B), 6.80 (m, 2H, H-3',5'), 7.15 (m, 1H, H-4'), 7.17 (d, 1H, J = 1.4, H-2), 7.47 (dd, 1H, J = 2.2 and 8.5, H_{4-Py}), 8.03 (d, 1H, J = 8.5, H_{3-Py}), 8.06 (d, 1H, J = 2.2, H_{6-Py}). Anal. (C₁₅H₁₂F₂N₂OS) C, H, N.

2-(2,6-Dichlorophenyl)-3-(6-methylpyridin-2-yl)-1,3-thiazolidin-4-one (29). Mp: 142-144 °C, yield 38%. ^1H NMR: 2.29 (s, 3H, Me), 3.94 (d, 1H, J = 15.9, 5-H_A), 4.16 (d, 1H, J = 2.2 and 15.9, 5-H_B), 6.82 (d, 1H, J = 7.7, H_{5-Py}), 7.02-7.30 (m, 3H, ArH), 7.54 (dd, 1H, J = 7.7 and 8.2, H_{4-Py}), 7.63 (d, 1H, J = 2.2, H-2), 7.94 (d, 1H, J = 8.2, H_{3-Py}). Anal. (C₁₅H₁₂Cl₂N₂OS) C, H, N.

2-(2-Chloro-6-fluorophenyl)-3-(6-methylpyridin-2-yl)-1,3-thiazolidin-4-one (30). Mp: 100-103 °C, yield 56%. ^1H NMR: 2.32 (s, 3H, Me), 3.85 (d, 1H, $J = 15.6$, 5-H_A), 4.20 (d, 1H, $J = 15.6$, 5-H_B), 6.83 (d, 1H, $J = 7.4$, H_{5-Py}), 6.87-7.36 (m, 4H, ArH and H-2), 7.55 (dd, 1H, $J = 7.4$ and 8.0, H_{4-Py}), 8.03 (d, 1H, $J = 8.0$, H_{3-Py}). Anal. (C₁₅H₁₂ClFN₂OS) C, H, N.

2-(2,6-Difluorophenyl)-3-(6-methylpyridin-2-yl)-1,3-thiazolidin-4-one (31). Mp: 88-89 °C, yield 46%. ^1H NMR: 2.32 (s, 3H, Me), 3.84 (d, 1H, $J = 15.9$, 5-H_A), 4.23 (dd, 1H, $J = 1.6$ and 15.9, 5-H_B), 6.81 (t, 2H, $J = 8.8$, H-3',5'), 6.83 (d, 1H, $J = 7.4$, H_{5-Py}), 7.15 (m, 1H, H-4'), 7.20 (d, 1H, $J = 1.6$, H-2), 7.54 (dd, 1H, $J = 7.4$ and 8.2, H_{4-Py}), 8.03 (d, 1H, $J = 8.2$, H_{3-Py}). Anal. (C₁₅H₁₂F₂N₂OS) C, H, N.

2-(2,6-Dichlorophenyl)-3-(4,6-dimethylpyridin-2-yl)-1,3-thiazolidin-4-one (32). Mp: 156-158 °C, yield 87%. ^1H NMR: 2.25 and 2.82 (2s, 6H, Me), 3.93 (d, 1H, $J = 15.9$, 5-H_A), 4.15 (dd, 1H, $J = 2.2$ and 15.9, 5-H_B), 6.66 (s, 1H, H_{5-Py}), 7.02-7.28 (m, 3H, ArH), 7.63 (d, 1H, $J = 2.2$, H-2), 7.75 (s, 1H, H_{3-Py}). Anal. (C₁₆H₁₄Cl₂N₂OS) C, H, N.

2-(2-Chloro-6-fluorophenyl)-3-(4,6-dimethylpyridin-2-yl)-1,3-thiazolidin-4-one (33). Mp: 143-145 °C, yield 65%. ^1H NMR: 2.28 (s, 6H, Me), 3.84 (d, 1H, $J = 15.7$, 5-H_A), 4.19 (d, 1H, $J = 15.7$, 5-H_B), 6.67 (s, 1H, H_{5-Py}), 6.84-7.36 (m, 4H, ArH and H-2), 7.84 (s, 1H, H_{3-Py}). Anal. (C₁₆H₁₄ClFN₂OS) C, H, N.

2-(2,6-Difluorophenyl)-3-(4,6-dimethylpyridin-2-yl)-1,3-thiazolidin-4-one (34). Mp: 122 °C, yield 80%. ^1H NMR: 2.27 (s, 6H, Me), 3.84 (d, 1H, $J = 15.9$, 5-H_A), 4.23 (dd, 1H, $J = 1.2$ and 15.9, 5-H_B), 6.67 (s, 1H, H_{5-Py}), 6.80 (t, 2H, $J = 8.5$, H-3',5'), 7.15 (m, 1H, H-4'), 7.20 (d, 1H, $J = 1.2$, H-2), 7.84 (s, 1H, H_{3-Py}). Anal. (C₁₆H₁₄F₂N₂OS) C, H, N.

Computational studies.

Structures: **1. Protein.** The X-ray structure of RT complexed with the inhibitor Nevirapine was retrieved from the Brookhaven Protein Data Bank (code 1VRT, 2.20 Å resolution) and used as target for our modeling studies. The ligand and water molecules were discarded and missing side chains of the unbound enzyme were manually completed using the InsightII/Biopolymer module.¹ Using the Biopolymer utility within the software package Sybyl 6.7,² “Essential_only” hydrogen atoms were added to the protein and later “KOLLUA” partial charges were assigned. **2. Ligands.** Coordinates for Nevirapine were taken directly from the X-ray structure of its complex with the

HIV-1 RT. The structures of the ligands **21** and **29** were constructed using standard bond lengths and angles from the Sybyl fragment library and fully optimized by semiempirical quantum mechanical method AM1. The flexible torsions identified by the docking procedure for the two inhibitors correspond to those defined by the two aromatic nuclei.

GRID studies. The calculations of the main intermolecular interactions involved in the RT inhibition were performed using the GRID software.³ This approach is useful for determining energetically favourable binding sites for small chemical groups (probes) and a macromolecule target of known three-dimensional structure (i.e. RT enzyme). The probe groups are small, distinct chemical entities and correspond to the functional groups of the inhibitors, as methyl, hydroxyl or carbonyl groups. In the GRID method, the probe is moved through a regular grid of points in a region of interest around the target molecule and, at each point, the interaction energy between the probe and the target molecule is calculated using a classical empirical energy function. All GRID calculations were performed with several probes in a box with dimensional equal 32 Å and a grid spacing of 1 Å. The interaction energies were contoured at an energy level of -0.3 Kcal/mol for the hydrophobic DRY probe, and -3.5 Kcal/mol for both aliphatic methyl and aromatic probes

Docking calculations. Docking studies to RT were carried out using the package AutoDock 3.0, a program that allows torsional flexibility in the ligand, while the protein is kept rigid.⁴ The dimensions of the grid were 60Å×60Å×60Å with grid points separated by 0.375 Å. The grid was centered on the allosteric site, using the Nevirapine crystallographic position as reference. The AutoGrid program generated separate grid maps for all atom types of the ligand structures plus one for electrostatic interactions. Our inhibitors present fluorine, chlorine and bromine atoms, but the AutoDock force field is based on a version of the AMBER force field⁵ which lacks parameters for halogens. However, AutoDock provides the actual values of the non-bond parameters for halogens derived from the Merck force field, MMFF.⁶ In order to allow the software to use these atom type parameters, we edit the atom names of the appropriate HETATM records in our PDBQ file, so that the first letter corresponded to the AutoDock single-letter code (i.e. "F" for fluorine, "c" for chlorine and "b" for bromine). We used the so-called Lamarckian genetic algorithm (LGA), which combines a traditional genetic algorithm (GA) with a local search method. For both ligands, the docking simulation was composed of 100 independent runs, starting each time from a different position and orientation chosen randomly. Clustering of the docked conformations was performed

with a tolerance of 1.5 Å. All the other values were used as default settings and the results were ranked by the lowest energy representative of each cluster combined with the root mean square deviation.

In vitro anti-HIV assay. The antiviral experiments using MT-4 cell cultures and HIV-1 (III_B) and HIV-2 (ROD) or CEM cell cultures and NNRTI-resistant mutant HIV-1 strains were performed following previously described procedures.⁷⁻⁹

RT assay. The RT assay using recombinant HIV-1 RT was essentially performed as described.¹⁰

Supporting References

- (1) InsightII 1999. Accelrys, Inc., San Diego, CA, <http://www.accelrys.com>.
- (2) SYBYL 6.7 2000, Tripos Associates Inc., St. Louis, Missouri, USA.
- (3) Goodford P. J. A computational procedure for determining energetically favorable binding sites on biologically important macromolecules. *J. Med. Chem.*, **1985**, *28*, 849-857.
- (4) Morris, G. M.; Goodsell, D.S.; Halliday, R. S.; Huey, R.; Hart, W. E., Belew R. K.; Olson, A.J. Automated docking using a Lamarckian genetic algorithm and an empirical binding free energy function. *J. Comp. Chem.*, **1998**, *19*, 1639-1662.
- (5) Weiner, S. J.; Kollman, P. A.; Nguyen, D. T.; and Case, D.A. An all atom force field for simulations of proteins and nucleic acids. *J. Comp. Chem.* **1986**, *7*, 230-252.
- (6) Halgren, T. A. Representation of van der Waals (vdW) Interactions in Molecular Mechanics Force Fields: Potential Form, Combination Rules, and vdW Parameters. *J. Am. Chem. Soc.*, **1992**, *114*, 7827-7843.
- (7) Pauwels, R.; Balzarini, J.; Baba, M.; Snoeck, R.; Schols, D.; Herdewijin, P.; Desmyter, J.; De Clercq, E. Rapid and automated tetrazolium-based colorimetric assay for the detection of anti-HIV compounds. *J. Virol. Methods* **1988**, *20*, 309-321.
- (8) Witvrouw, M.; Schols, D.; Andrei, G.; Snoeck, R.; Hosoya, M.; Pauwels, R.; Balzarini, J.; De Clercq, E. Antiviral activity of low-MW dextran sulfate (derived from dextran MW 1000) compared to dextran sulfate samples of higher MW. *Antiviral Chem. Chemoth.* **1991**, *2*, 171-179.

- (9) Balzarini, J.; Karlsson, A.; Pérez-Pérez, M. J.; Vrang, L.; Walberg, J.; Zhang, H.; Öberg, B.; Vandamme, A. M.; Bathurst, I. C.; Barr, P. J.; Camarasa, M. J.; De Clercq, E. HIV-1-specific reverse transcriptase inhibitors show differential activity against HIV-1 mutant strains containing different amino acid substitution in the reverse transcriptase. *Virology* 1993, 192, 246-253.
- (10) Balzarini, J.; Pérez-Pérez, M. J.; San-Félix, A.; Camarasa, M. J.; Bathurst, I. C.; Barr, P. J.; De Clercq, E. Kinetics of inhibition of human immunodeficiency virus type 1 (HIV-1) reverse transcriptase by the novel HIV-1-specific nucleoside analogue [2',5'-bis-O-(tert-butyldimethylsilsil)-beta-D-ribofuranosyl]-3'-spiro-5''-(4''-amino-1'',2''-oxathiole-2'',2''-dioxide)thymine (TSAO-T). *J. Biol. Chem.* 1992, 267, 11831-11838.