

Synthesis, Nicotinic Acetylcholine Receptor Binding, and Antinociceptive Properties of 2-*exo*-2-(2'-Substituted-3'-phenyl-5'-pyridinyl)-7-azabicyclo-[2.2.1]heptanes. Novel Nicotinic Antagonist

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Supporting Information

| Analysis Appendix | | | | | | | |
|-------------------|--|--------|------|-------|-------|------|-------|
| compd | formula | Calcd. | | | Found | | |
| | | C | H | N | C | H | N |
| (±) 5a | C ₁₇ H ₁₈ ClFN ₂ •H ₂ O | 63.25 | 6.25 | 8.67 | 63.06 | 6.25 | 8.80 |
| (-) 5a | C ₁₇ H ₁₈ ClFN ₂ •0.5H ₂ O | 65.07 | 5.94 | 8.93 | 65.10 | 6.02 | 8.91 |
| (+) 5a | C ₁₇ H ₁₈ ClFN ₂ •0.33H ₂ O | 65.70 | 6.05 | 9.01 | 65.61 | 5.95 | 8.86 |
| 5b | C ₁₇ H ₁₇ Cl ₂ N ₂ •1.25H ₂ O | 59.40 | 6.01 | 8.15 | 59.28 | 5.86 | 8.12 |
| 5c | C ₁₇ H ₂₄ N ₃ O•2.5HCl•1.25H ₂ O | 53.87 | 6.38 | 11.09 | 53.95 | 6.35 | 10.68 |
| 5d | C ₁₉ H ₂₅ Cl ₂ N ₃ •2.25H ₂ O | 56.09 | 7.31 | 10.33 | 55.96 | 7.40 | 9.85 |
| 7 | C ₁₆ H ₂₂ BrN ₃ O ₂ | 52.18 | 6.02 | 11.41 | 52.23 | 6.11 | 11.35 |
| 8 | C ₂₂ H ₂₇ N ₃ O | 72.30 | 7.45 | 11.50 | 71.74 | 7.45 | 11.26 |

7-*tert*-Butoxycarbonyl-2-*exo*-(2'-amino-3'-bromo-5'-pyridinyl)-7-azabicyclo[2.2.1]heptane (7). To a stirred solution of 968 mg (3.30 mmol) of 7-*tert*-Butoxycarbonyl-2-*exo*-(2'-amino-5'-pyridinyl)-7-azabicyclo[2.2.1]heptane (**6**) in methylene chloride (8 mL) and acetic acid (7 mL) under nitrogen at 0 °C was added bromine (0.260 mL, 5.05 mmol) followed by triethylamine (0.260 mL). After stirring the reaction for 16 h, the mixture was poured into 100 mL of NH₄OH:H₂O (1:2) solution and extracted 3 × with chloroform. The combined organic extracts were dried with magnesium sulfate and concentrated. The residue was purified by flash chromatography using 4:1 ether triethylamine to give 1.04 g (85%) of **7** as a colorless solid: mp 129–130 °C; ¹H NMR (CDCl₃) δ (ppm) 1.44 (s, 9H), 1.40–1.55 (m, 2H), 1.70–1.84 (m, 3H), 1.90 (dd, *J* = 9.0, 12.3 Hz, 1H), 2.70 (dd, *J* = 4.8, 8.8 Hz, 1H), 4.08 (br s, 1H), 4.33 (br s, 1H), 7.62 (s, 1H, pyridyl CH), 7.83 (s, 1H, pyridyl CH); ¹³C NMR (CDCl₃) δ (ppm) 28.3 (3C), 28.7, 29.7, 40.3, 44.6, 55.7, 62.0, 79.7, 104.6, 132.9, 138.8, 145.5, 154.0, 154.9. Anal. (C₁₆H₂₂BrN₃O₂) C, H, N.

7-*tert*-Butoxycarbonyl-2-*exo*-(2'-amino-3'-phenyl-5'-pyridinyl)-7-azabicyclo[2.2.1]heptane (8). To a resealable reaction tube under nitrogen was added 403 mg (1.08 mmol) of 7-*tert*-butoxycarbonyl-2-*exo*-(2'-amino-3'-bromopyridinyl)-7-azabicyclo[2.2.1]heptane (**7**), Pd(OAc)₂ (25 mg, 0.011 mmol), P(*o*-tolyl)₃ (60 mg, 0.02 mmol), sodium carbonate (230 mg, 2.17 mmol), phenylboronic acid (210 mg, 1.72 mmol), degassed water (0.800 mL), and DME (4 mL). The reaction was heated at 80 °C for 1.5 h. The reaction mixture was poured into saturated sodium bicarbonate and extracted 3 × with ethyl acetate. The organic layers were dried with sodium sulfate, and concentrated. The residue was purified by flash chromatography using hexane:ethyl acetate (1:2) as eluent to provide 347 mg (88%) of **8** as a colorless solid: ¹H NMR (CDCl₃) δ (ppm) 1.38 (br s, 9H), 1.38–1.65 (m, 2H), 1.75–2.0 (m, 4H), 2.78 (dd, *J* = 5.2, 8.6 Hz, 1H), 4.16 (s, 1H), 4.35 (s, 1H), 4.60 (br s, 2 NH), 7.3–7.45 (m, 6H), 7.92 (d, *J* = 2.2 Hz, 1H); ¹³C NMR (CDCl₃) δ (ppm) 28.2, 28.8, 29.7, 40.2, 44.8, 55.5, 62.1, 79.3, 121.6, 127.5, 128.6(2C), 128.8(2), 131.7, 136.5, 138.2, 145.6, 154.3, 154.8. Anal. (C₂₂H₂₇N₃O₂) C, H, N.

2-*exo*-(2'-Fluoro-3'-phenyl-5'-pyridinyl)-7-azabicyclo[2.2.1]heptane (5a) Hydrochloride. A solution of 150 mg (0.410 mmol) of 7-*tert*-butoxycarbonyl-2-*exo*-(2'-amino-3'-phenyl-5'-pyridinyl)-7-azabicyclo[2.2.1]heptane (**8**) in concentrated hydrofluoric acid/pyridine (0.6 mL) was prepared in a plastic vessel. Sodium nitrite (110 mg, 1.6 mmol) was added and the reaction mixture stirred for 45 min at room temperature followed by heating to 100 °C for 1 h. The mixture was poured into a solution of 50 mL of NH₄OH:H₂O (1:1) and extracted with ethyl acetate. The combined organic layers were dried with magnesium sulfate and concentrated. The

residue was purified via flash chromatography using $\text{CHCl}_3:\text{CH}_3\text{OH}:\text{NH}_4\text{OH}$ (45:9:1) to give 91 mg (83%) of **5a** as a colorless oil: ^1H NMR (CDCl_3) δ (ppm) 1.45–1.76 (m, 4H), 1.93 (dd, J = 9.3, 12.3 Hz, 2H), 2.04 (s, 1H), 2.83 (dd, J = 6.0, 9.3 Hz, 1H), 3.62 (br s, 1H), 3.80 (br s, 1H), 7.33–7.60 (m, 5H), 7.98 (dd, J_F = 2.4, 9.6 Hz, 1H), 8.07 (t, J_F = 1.5 Hz, 1H); ^{13}C NMR (CDCl_3) δ (ppm) 29.97, 31.23, 40.32, 44.35, 56.36, 62.74, 123.01(d, J_{CF} = 28.5 Hz), 128.5 (m, 4C), 134.15 (d, J_{CF} = 5.1 Hz), 139.70 (d, J_{CF} = 4.2 Hz), 140.34 (d, J_{CF} = 18.9 Hz), 144.59 (d, J_{CF} = 57 Hz), 157.39, 160.55.

To a stirred solution of 91 mg (0.339 mmol) of **5a** in methylene chloride (2.5 mL) was added 1M HCl in ether (1.6 mL). After 30 min at room temperature, the solvent was removed under reduced pressure and the remaining solid was pumped overnight to give 90 mg (81%) of **5a**•HCl as a colorless solid. Anal. ($\text{C}_{17}\text{H}_{18}\text{ClFN}_2\cdot\text{H}_2\text{O}$) C, H, N.

(+)- and (–)-2-*exo*-(2'-Fluoro-3'-phenyl-5'-pyridinyl)-7-azabicyclo[2.2.1] heptane (+)- and (–)-5a. To a mixture of 520 mg (1.94 mmol) of racemic 2-*exo*-(2'-fluoro-3'-phenyl-5'-pyridinyl)-7-azabicyclo[2.2.1] heptane (**5a**) and 0.80 g (0.0020 mol) of di-*p*-toluoyl-D-tartaric acid was added 45 mL of (4:1) isopropanol/water. The resulting mixture was warmed to dissolution and allowed to stand at room temperature overnight. The resulting crystals were filtered to give 0.83 g of the di-*p*-toluoyl salt of **5a**. This salt was recrystallized twice from isopropanol/water (4:1) mixtures to give 0.27 g of the tartrate salt. This salt was neutralized with aqueous Na_2CO_3 and extracted with CH_2Cl_2 to give 100 mg of the free base. The free base was converted to the hydrochloride salt and recrystallized from EtOAc/MeOH mixtures to give 42 mg of (–)-2-*exo*-(2'-fluoro-3'-phenyl-5'-pyridinyl)-7-azabicyclo[2.2.1] heptane hydrochloride: mp 239–242 °C; $[\alpha]_D = -11.4$ (c 0.35, MeOH). Anal. ($\text{C}_{17}\text{H}_{17}\text{FN}_2\text{HCl}$) C, H, N.

The mother liquors from above were combined and evaporated to dryness, dissolved in water, basified with Na_2CO_3 , and extracted with CH_2Cl_2 . The dried extracts (Na_2SO_4) were evaporated to give 0.25 g of solid. To the solids was added 0.378 g of di-*p*-toluoyl-L-tartaric acid and 20 mL of (4:1) isopropanol/water and crystallized as above to afford 0.32 g of the tartrate salt of **5a**. The salt was converted to its free base as above to afford 0.13 g. The free base was converted to the hydrochloride salt and recrystallized from EtOAc/MeOH mixtures to afford 56 mg of (+)- 2-*exo*-(2'-fluoro-3'-phenyl-5'-pyridinyl)-7-azabicyclo[2.2.1] heptane hydrochloride hydrate: mp 240–242 °C; $[\alpha]_D = +11.1$ (c = 0.28, MeOH). Anal. ($\text{C}_{17}\text{H}_{18}\text{ClFN}_2\cdot 0.33\text{H}_2\text{O}$) C, H, N.

2-*exo*-(2'-Amino-3'-phenyl-5'-pyridinyl)-7-azabicyclo[2.2.1]-heptane (5c) Hydrochloride. A solution of 165 mg (0.451 mmol) of 7-*tert*-butoxycarbonyl-2-*exo*-(2'-amino-3'-phenyl-5'-pyridinyl)-7-azabicyclo[2.2.1]heptane (**8**) in methylene chloride (1.0 mL) and trifluoroacetic acid (1.0 mL) was allowed to stir at room temperature for 1 h. The reaction was decanted into a saturated NaHCO₃ solution and extracted 3 × with chloroform. The combined organic extracts were dried with sodium sulfate and concentrated. The residue was purified by flash chromatography using CHCl₃:CH₃OH:NH₄OH (45:9:1) as eluent to give 116 mg (97%) of **5c** as a colorless oil: ¹H NMR (CDCl₃) δ (ppm) 1.38–1.82 (m, 4H), 1.87 (dd, *J* = 9.0, 12.2 Hz, 1H), 2.75 (dd, *J* = 5.1, 8.67 Hz, 1H), 3.54 (br s, 1H), 3.73 (br s, 1H), 4.61 (br s, 2 H), 7.30–7.47 (m, 6H), 7.93 (s, 1H, pyridyl CH); ¹³C NMR (CDCl₃) δ (ppm) 29.71, 30.82, 39.91, 44.68, 56.28, 62.85, 121.55, 127.48, 128.61(2C), 128.82(2C), 132.28, 136.85, 138.23, 145.51, 154.15.

To a stirred solution of 96 mg (0.362 mmol) of **5c** in methylene chloride (1.5 mL) was added 1M HCl in ether (3 mL). After 1 h. at room temperature, the solvent was removed under reduced pressure, and the remaining solid was pumped overnight to give 127 mg (92%) of **5c**•HCl as a colorless solid: mp decomposed >200 °C. Anal. (C₁₇H₂₄N₃O•2.5HCl•1.25H₂O) C, H, N.

2-*exo*-(2'-Chloro-3'-phenyl-5'-pyridinyl)-7-azabicyclo[2.2.1]heptane (5b) Hydrochloride. To a solution of 217 mg (0.594 mmol) of 7-*tert*-butoxycarbonyl-2-*exo*-[5'-(3'-phenyl-2'-aminopyridinyl)]-7-azabicyclo[2.2.1]heptane (**8**) in concentrated hydrochloric acid (1.5 mL) was added sodium nitrite (800 mg, 11.6 mmol). Copper (I) chloride (800 mg, 8.1 mmol) was then added in small portions and stirring continued for 30 min at 0 °C. The mixture was poured into a solution of 50 mL of NH₄OH:H₂O (1:1) and extracted 3 × with ethyl acetate. The combined organic layers were dried with magnesium sulfate and concentrated. The residue was purified via flash chromatography using CHCl₃:CH₃OH:NH₄OH (45:9:1) to give 100 mg (59%) of **5b** as a colorless oil: ¹H NMR (CDCl₃) δ (ppm) 1.45–1.78 (m, 5H), 1.93 (dd, *J* = 9.0, 12.1 Hz, 1H), 2.81 (dd, *J* = 4.9, 8.9 Hz, 1H), 3.61 (br s, 1H), 3.78 (br s, 1H), 7.36–7.48 (m, 5H), 7.76 (s, pyridyl 1 CH), 8.29 (s, pyridyl 1 CH); ¹³C NMR (CDCl₃) δ (ppm) 30.07, 31.30, 40.27, 44.45, 56.28, 62.64, 127.98, 128.11(2C), 129.23(2C), 136.19, 137.69, 138–54, 141.41, 146.92, 147.33.

To a stirred solution of 70 mg (0.246 mmol) of **5b** in ether (1.5 mL) was added 1M HCl in ether (1.5 mL). After 30 min at room temperature, the solvent was removed under reduced pressure and the remaining solid was pumped overnight to give 80 mg (97%) of **5b**•HCl as a colorless solid: mp 144–147 °C; Anal. (C₁₇H₁₇Cl₂N₂•1.25H₂O) C, H, N.

7-tert-Butoxycarbonyl-2-*exo*-(2'-dimethylamino-3'-phenyl-5-pyridinyl)-7-

azabicyclo[2.2.1]heptane (9). A mixture of 123 mg (0.337 mmol) of 7-*tert*-butoxycarbonyl-2-*exo*-(2'-amino-3'-phenyl-5'-pyridinyl)-7-azabicyclo[2.2.1]heptane (**8**), acetonitrile (9 mL), 37% formaldehyde (1.2 mL), and NaCNBH₃ (380 mg, 7.1 mmol) was allowed to stir for 3 h at room temperature. Glacial acetic acid (0.5 mL) was added and stirring continued overnight. After 24 h the reaction mixture was decanted into NH₄OH:H₂O (1:1) and extracted 3 × with chloroform. The combined organic extracts were dried with sodium sulfate and concentrated. The residue was purified by flash chromatography using hexane:ethyl acetate (1:2) as eluent to give 99 mg (75%) of **9** as a colorless oil: ¹H NMR (CDCl₃) δ (ppm) 1.38 (s, 9H), 1.5–1.6 (m, 2H), 1.7–2.0 (m, 4H), 2.68 (s, 6H), 2.82 (dd, *J* = 6.0, 9.0 Hz, 1H), 4.18 br s, 1H), 4.34 (br s, 1H), 7.20–7.55 (m, 6H), 8.05 (d, *J* = 2.4 Hz, 1H); ¹³C NMR (CDCl₃) δ (ppm) 28.22 (3C), 28.72, 29.75, 40.23, 41.28 (2C), 44.77, 55.81, 62.03, 79.35, 125.43, 126.73 127.94 (2C), 128.45 (2C), 132.52, 138.28, 140.88, 144.51, 154.5, 158.74.

2-*exo*-(2'-Dimethylamino-3'-phenyl-5'-pyridinyl)-7-azabicyclo[2.2.1]heptane (5d)

Dihydrochloride. A solution of 98 mg (0.249 mmol) of 7-*tert*-butoxycarbonyl-2-*exo*-(2'-dimethylamino-3'-phenyl-5'-pyridinyl)-7-azabicyclo[2.2.1]heptane (**9**) in methylene chloride (1.5 mL) was stirred at 0 °C for 15 min. Trifluoroacetic acid (1.5 mL) was added and the reaction mixture was allowed to stir at room temperature for 30 min. The reaction mixture was decanted into a NH₄OH:H₂O (1:1) solution and extracted 3 × with chloroform. The combined organic extracts were dried with sodium sulfate and concentrated. The residue was purified by flash chromatography using CHCl₃:CH₃OH:NH₄OH (80:18:2) as eluent to give (64 mg, 88%) of **5d** as a colorless oil: ¹H NMR (CDCl₃) δ (ppm) 1.3–1.8 (m, 5H), 1.88 (dd, *J* = 9.0, 12.3 Hz, 1H), 2.04 (br s, 1H), 2.68 (s, 6H), 2.80 (dd, *J* = 6.0, 9.0 Hz, 1H), 3.57 (br s, 1H), 3.74 (br s, 1H), 7.25–7.55 (m, 6H), 8.07 (d, *J* = 2.4 Hz, 1H); ¹³C NMR (CDCl₃) δ (ppm) 29.75, 30.77, 39.84, 41.27 (2C), 44.78, 56.38; 62.78, 125.40, 126.72, 127.96 (2C), 128.49 (2C), 133.05, 138.67 140.91, 144.40, 158.56.

To a stirred solution of 61 mg (0.208 mmol) of **5d** in methylene chloride (2.0 mL) was added 1M HCl in ether (2.0 mL). After 30 min at room temperature, the solvent was removed under reduced pressure and the remaining solid was pumped overnight to give 76 mg (99%) of **5d**•2HCl as a colorless solid: mp 176–181 °C. Anal. (C₁₉H₂₅Cl₂N₃•2.25H₂O) C, H, N.

[³H]Epibatidine Binding Assay. Adult male rat cerebral cortices (Pelfreeze Biological, Rogers, AK) were homogenized in 39 volumes of ice-cold 50 mM Tris buffer (pH 7.4 at 4 °C)

containing 120 mM NaCl, 5 mM KCl, 2 mM CaCl₂, and 1 mM MgCl₂ and sedimented at 37,000 g for 10 min at 4 °C. The supernatant was discarded, the pellet resuspended in the original volume of buffer, and the wash procedure repeated twice more. After the last centrifugation, the pellet was resuspended in 1/10 its original homogenization volume and stored at -80 °C until needed. In a final volume of 0.5 mL, each assay tube contained 3 mg wet weight male rat cerebral cortex homogenate (added last), 0.5 nM [³H]epibatidine (NEN Life Science Products, Wilmington, DE) and one of 10–12 different concentrations of test compound dissolved in buffer (pH 7.4 at room temperature) containing 10% DMSO resulting in a final DMSO concentration of 1%. Total and nonspecific bindings were determined in the presence of vehicle and 300 uM (–)-nicotine, respectively. After a 4-h incubation at room temperature, the samples were vacuum-filtered over GF/B filter papers presoaked in 0.03% polyethylenimine using a Brandel 48-well harvester and washed with 6 mL of ice-cold buffer. The amount of radioactivity trapped on the filter was determined by standard liquid scintillation techniques in a TriCarb 2200 scintillation counter (Packard Instruments, Meriden, CT) at approximately 50% efficiency. The binding data were fit using the nonlinear regression analysis routines in Prism (Graphpad, San Diego, CA). The K_i values for the test compounds were calculated from their respective IC₅₀ values using the Cheng-Prusoff equation.

[¹²⁵I]iodo-MLA Binding Assay. Adult male rat cerebral cortices (Pel-Freez Biologicals, Rogers, AK) were homogenized (polytron) in 39 volumes of ice-cold 50 mM Tris buffer (assay buffer; pH 7.4 at 4 °C) containing 120 mM NaCl, 5 mM KCl, 2 mM CaCl₂, and 1 mM MgCl₂. The homogenate was centrifuged at 35,000 g for 10 min at 4 °C and the supernatant discarded. The pellet was resuspended in the original volume of buffer and the wash procedure repeated twice more. After the last centrifugation step, the pellet was resuspended in one-tenth the original homogenization volume and stored at -80 °C until needed. Triplicate samples were run in 1.4-mL polypropylene tubes (Matrix Technologies Corporation, Hudson, NH). Briefly, in a final volume of 0.5 mL, each assay sample contained 3 mg wet weight rat cerebral cortex (added last), 40–50 pM [¹²⁵I]MLA and 50 nM final concentration of test compound dissolved in buffer containing 10% DMSO, giving a final DMSO concentration of 1%. Total and nonspecific binding were determined in the presence of vehicle and 300 uM (–)-nicotine, respectively. After a 2-h incubation on ice, the samples were vacuum-filtered using a Multimate 96-well harvester (Packard Instruments, Meriden, CT) onto GF/B filters presoaked for at least 30 min in assay buffer containing 0.15% bovine serum albumin. Each well was then washed with approximately 3.0 mL of ice-cold buffer. The filter plates were dried, and 30 uL of Microscint20 (Packard) was added to each well. The amount of radioligand remaining on each filter was determined using a

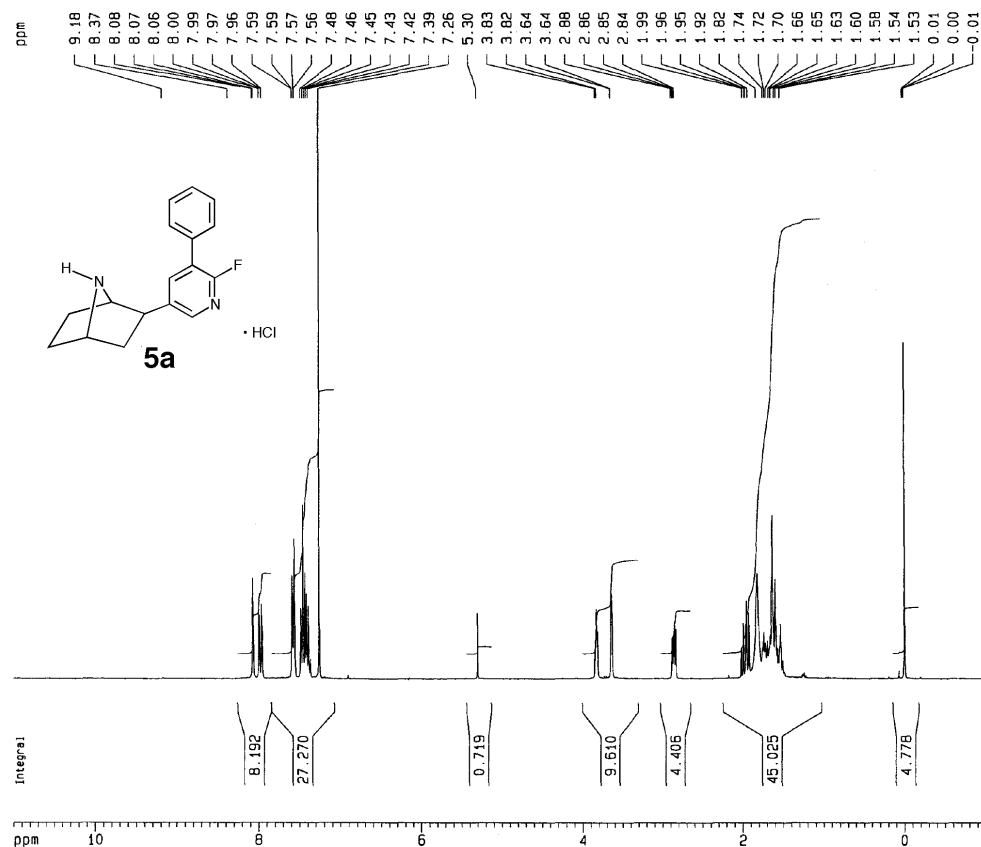
TopCount 12-detector (Packard) microplate scintillation counter at approximately 70% efficiency.

Tail-flick test. Antinociception was assessed by the tail-flick method of D'Amour and Smith.²² A control response (2–4 sec) was determined for each mouse before treatment, and a test latency was determined after drug administration. In order to minimize tissue damage, a maximum latency of 10 sec was imposed. Antinociceptive response was calculated as percent maximum possible effect (% MPE), where $\%MPE = [(test-control)/(10-control)] \times 100$. Groups of eight to twelve animals were used for each dose and for each treatment. The mice were tested 5 min after i. t. injections of epibatidine analogs for the dose-response evaluation. Eight to twelve mice were treated per dose and a minimum of 4 doses were performed for dose-response curve determination.

Hot-plate Test. Mice were placed into a 10 cm wide glass cylinder on a hot plate (Thermojust Apparatus) maintained at 55.0 C. Two control latencies at least ten min apart were determined for each mouse. The normal latency (reaction time) was 8 to 12 seconds. Antinociceptive response was calculated as percent maximum possible effect (% MPE), where $\%MPE = [(test-control)/40-control] \times 100$. The reaction time was scored when the animal jumped or licked its paws. Eight mice per dose were injected s.c. with epibatidine analogs and tested 5 min thereafter in order to establish a dose-response curve.

Locomotor activity. Mice were placed into individual Omnitech photocell activity cages (28 x 16.5 cm) 5 min after s.c. administration of either 0.9% saline or epibatidine analogs. Interruptions of the photocell beams (two banks of eight cells each) were then recorded for the next 10 min. Data were expressed as number of photocell interruptions.

Body temperature. Rectal temperature was measured by a thermistor probe (inserted 24 mm) and digital thermometer (Yellow Springs Instrument Co., Yellow Springs, OH). Readings were taken just before and at different times after the s.c. injection of either saline or epibatidine analogs. The difference in rectal temperature before and after treatment was calculated for each mouse. The ambient temperature of the laboratory varied from 21–24°C from day to day.



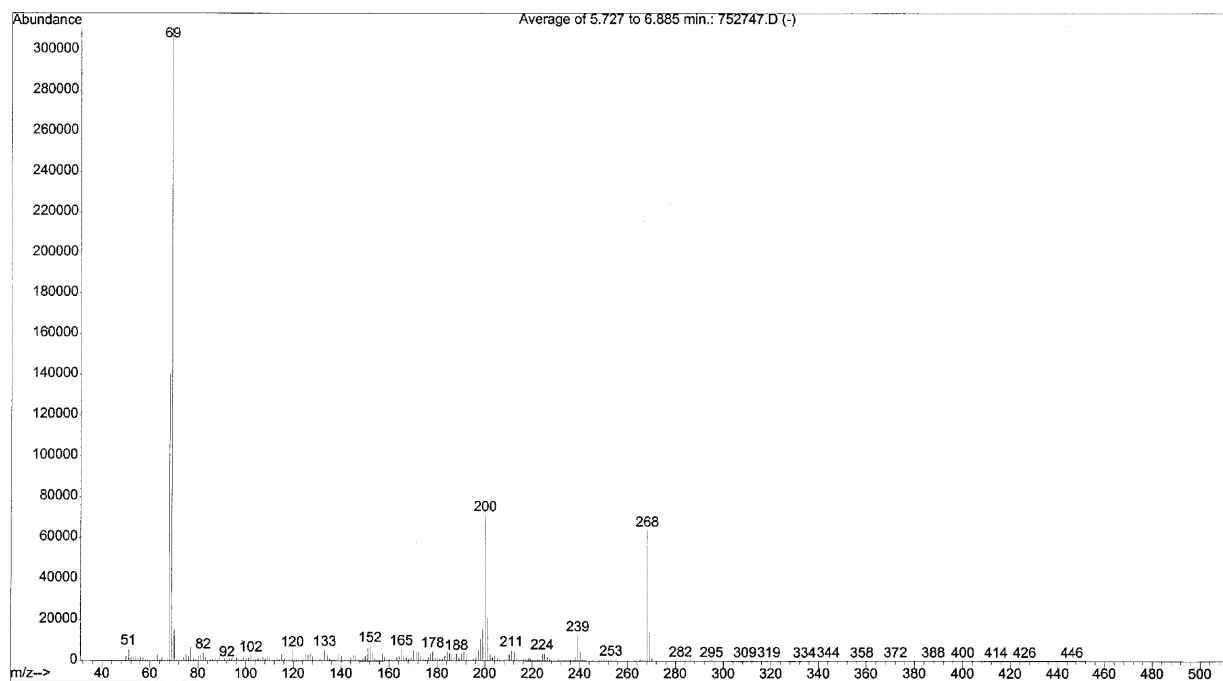
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5a

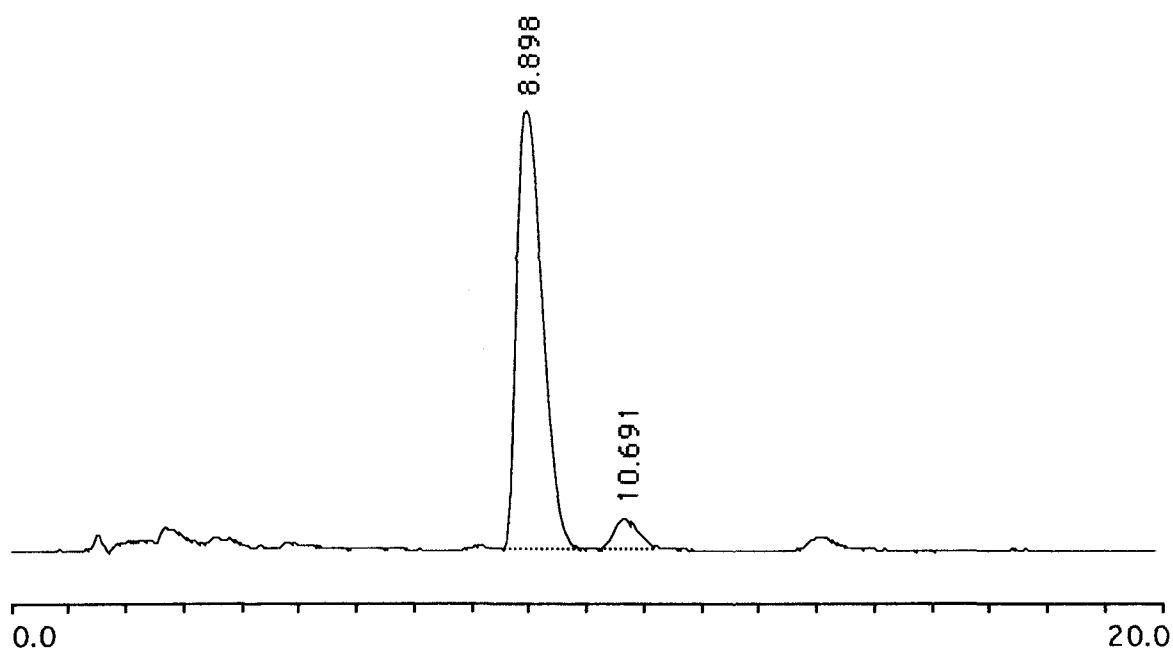
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5a

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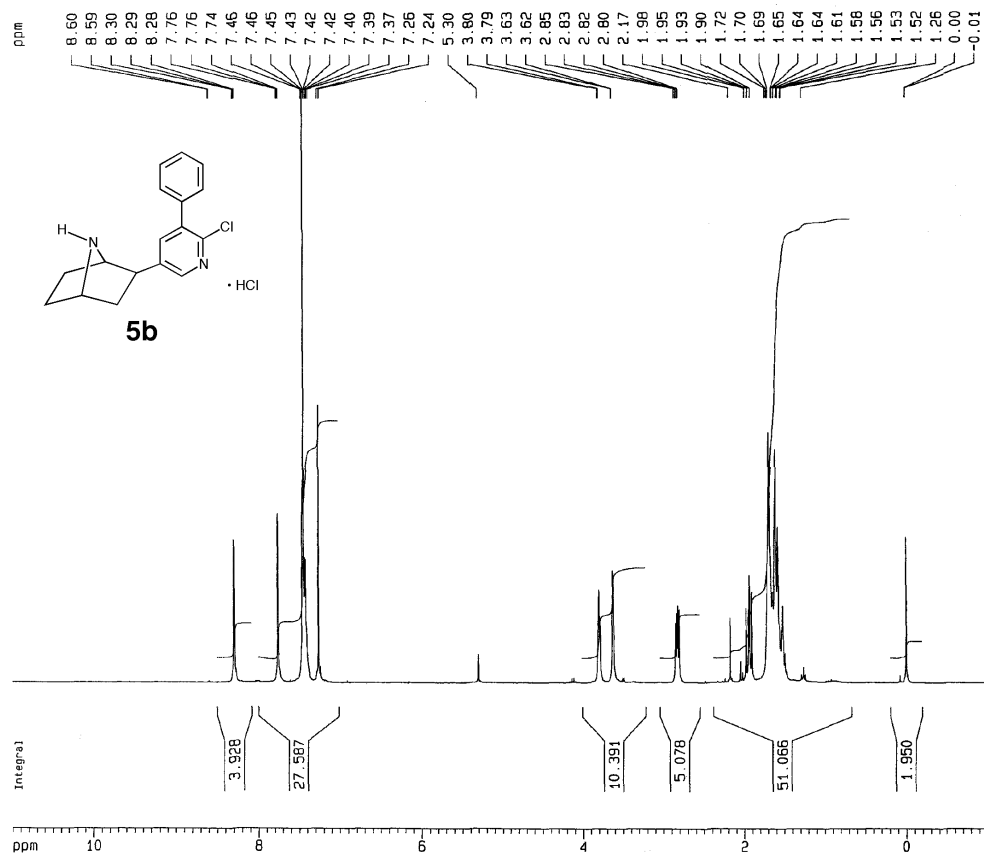
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Data:



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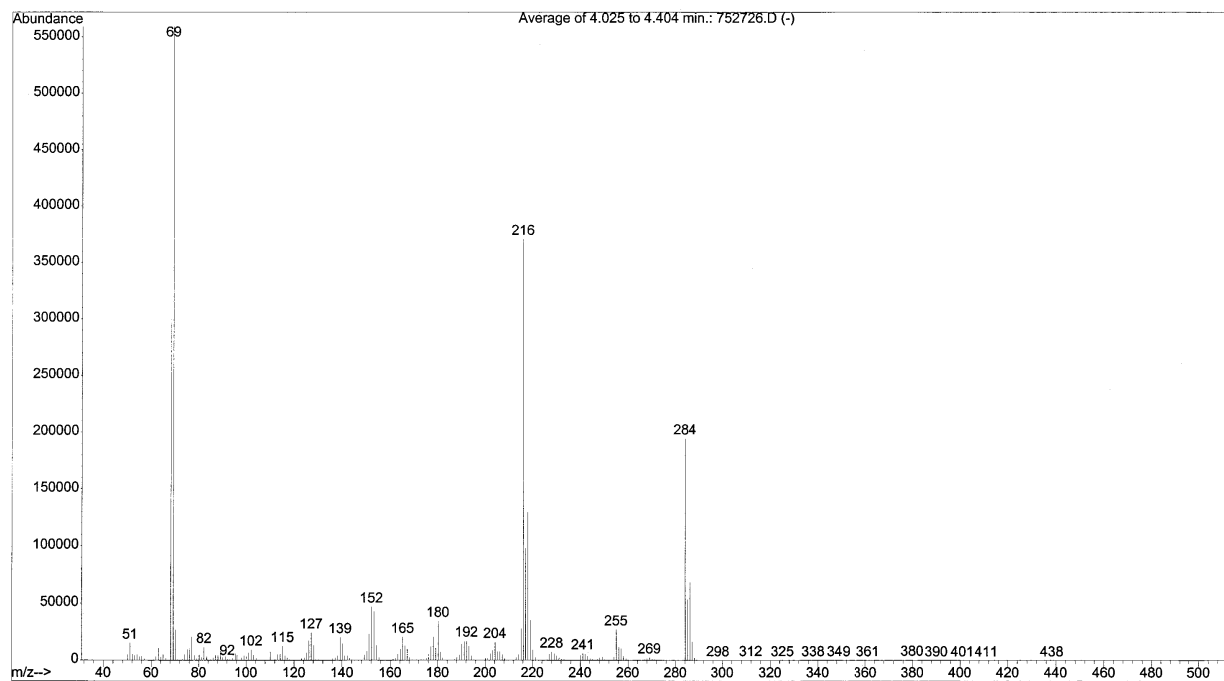
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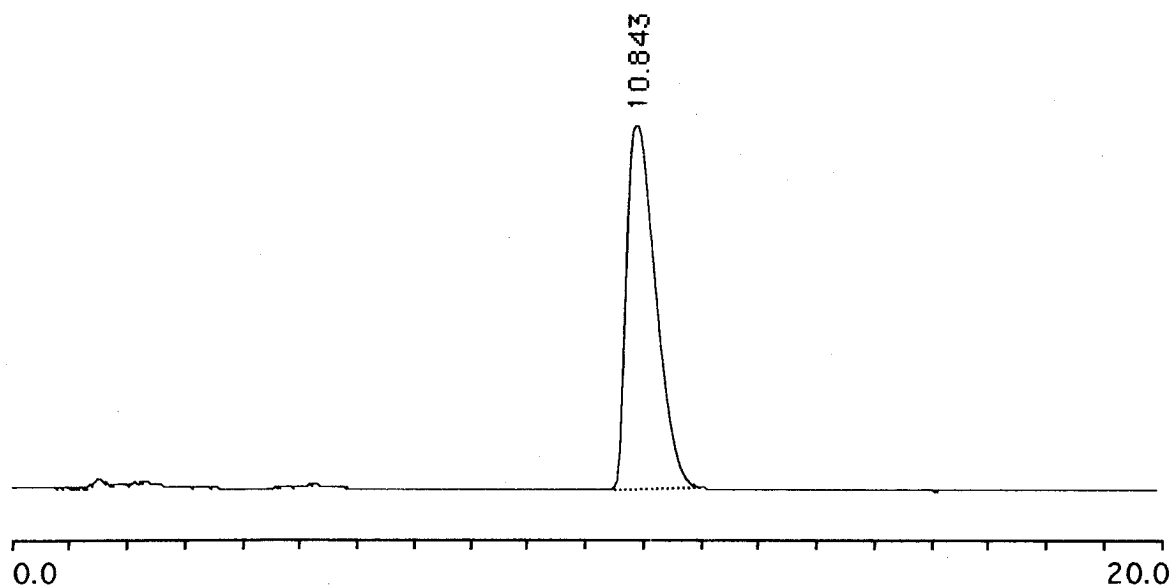
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Data: Epibatidine-17JUL101-008

5b

Sample: RTI-7527-26 (about 1 mg/ml CH₃OH), A: [TFA-H₂O], B: CH₃CN, 25% B,
0.6 ml/min, 225 nm, Phenomenex Synergi 4 μ Hydro-RP 80A (3*150 mm,
4 μ m)

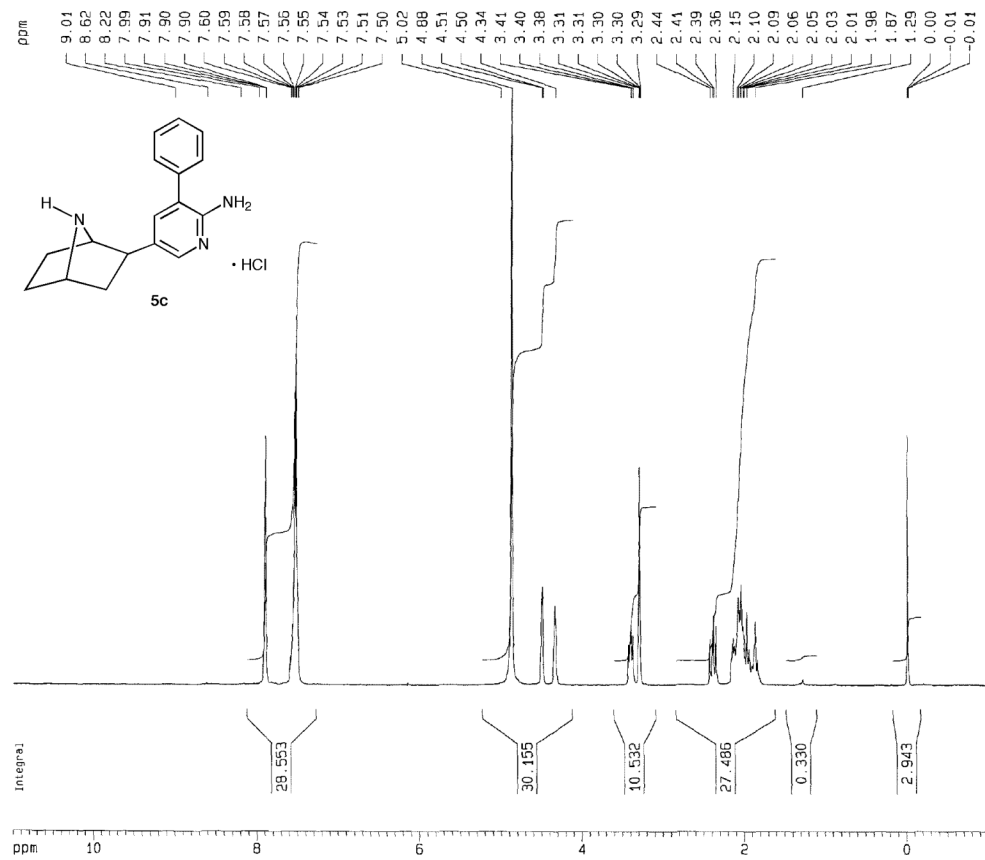
Method: Epibatidine
Inject Vol: 1
Sampling Int: 0.1 Seconds

Data:



Analysis: Channel A

| Peak No. | Time | Type | Height(μ V) | Area(μ V-sec) | Area% |
|------------|--------|------|------------------|--------------------|----------------|
| 1 | 10.843 | N | 84079 | <u>2763672</u> | <u>100.000</u> |
| Total Area | | | | 2763672 | 100.000 |



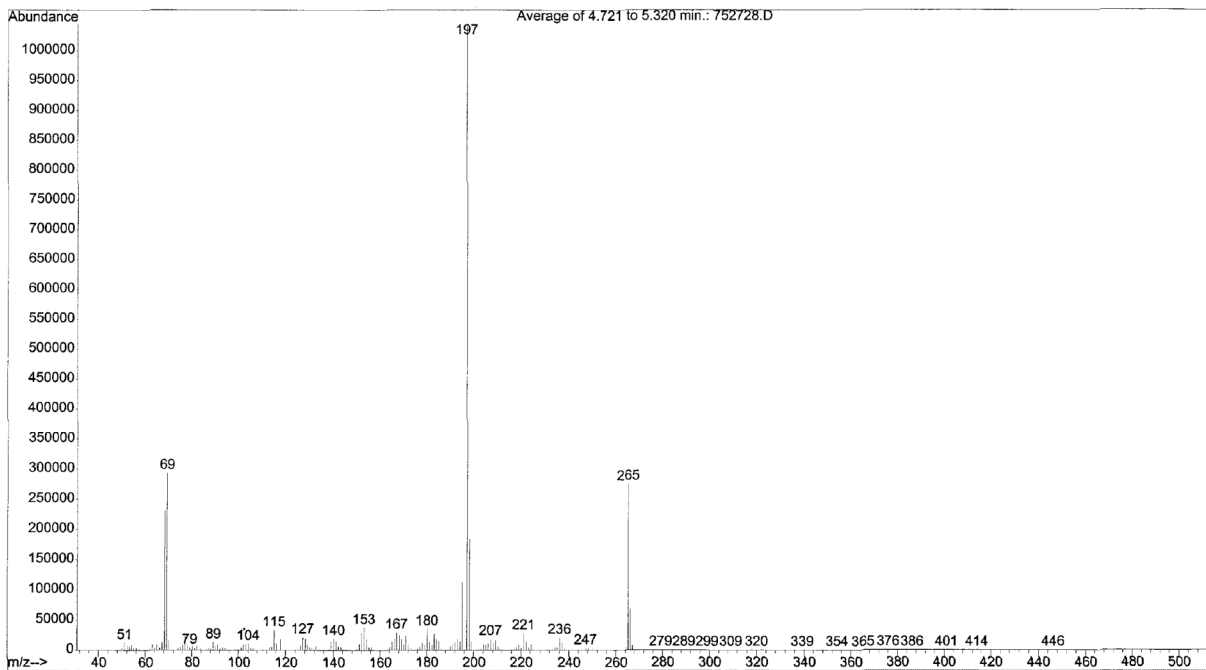
Current Data Parameters
 NAME RT1752728
 EXPNO 190
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20010715
 Time 15.19
 INSTRUM spect
 PROBHD 5 mm QNP 1H/
 PULPROG zg
 TD 32768
 SOLVENT MeOH
 NS 16
 QS 2
 SWH 6172.838 Hz
 FIDRES 0.188380 Hz
 AQ 2.6542580 sec
 RG 456.1
 DW 81.000 usec
 DE 6.00 usec
 TE 300.0 K
 D1 1.00000000 sec
 P1 5.50 usec
 SFO1 300.1318534 MHz
 NUC1 1H
 PL1 -6.00 dB

F2 - Processing parameters
 SI 16384
 SF 300.1300055 MHz
 WDM EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00

1D NMR plot parameters
 CX 20.00 cm
 F1P 11.000 ppm
 F1 3301.43 Hz
 F2P -1.000 ppm
 F2 -300.13 Hz
 PPMCM 0.60000 ppm/cm
 HZCM 180.07800 Hz/cm

File : C:\HPCHEM\1\DATA\SAMPLES\752728.D
Operator : R. Mitchell
Acquired : 17 Jul 2001 8:57 am using AcqMethod EIDIPHED
Instrument : GC/MS Ins
Sample Name: RTI-7527-28
Misc Info : Abraham
Vial Number: 1

5c

Date: Wed, Jul 18, 2001 9:20 AM

Data: Epibatidine-18JUL101-002

5c

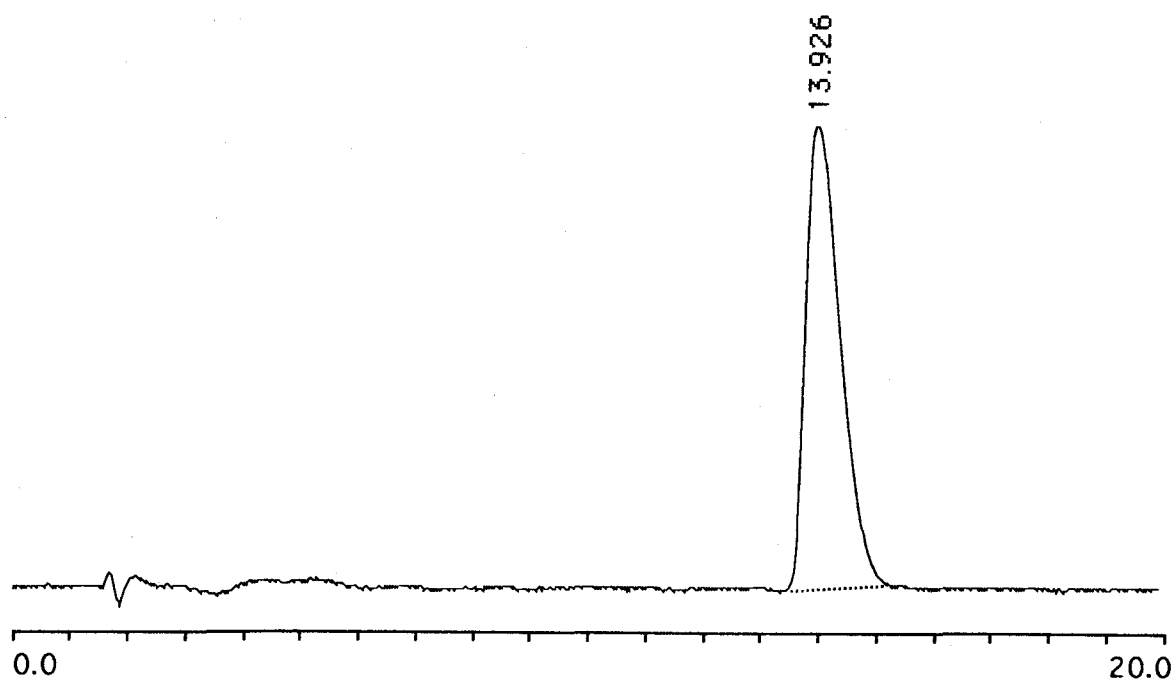
Sample: RTI-7527-28 (about 1 mg/ml CH₃OH), A: [TFA-H₂O], B: CH₃CN, 10% B,
0.6 ml/min, 225 nm, Phenomenex Synergi 4 μ Hydro-RP 80A (3*150 mm,
4 μ m)

Method: Epibatidine

Inject Vol: 1

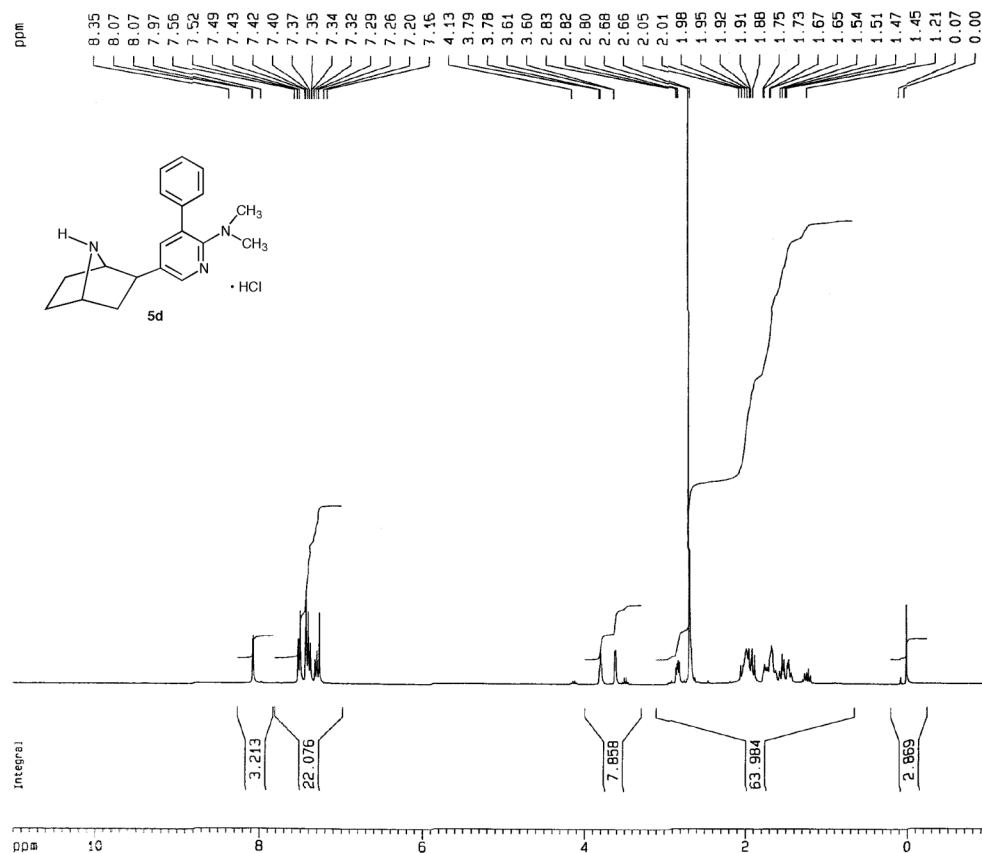
Sampling Int: 0.1 Seconds

Data:



Analysis: Channel A

| Peak No. | Time | Type | Height(μ V) | Area(μ V-sec) | Area% |
|------------|--------|------|------------------|--------------------|---------|
| 1 | 13.926 | N | 34311 | 1326864 | 100.000 |
| Total Area | | | | 1326864 | 100.000 |



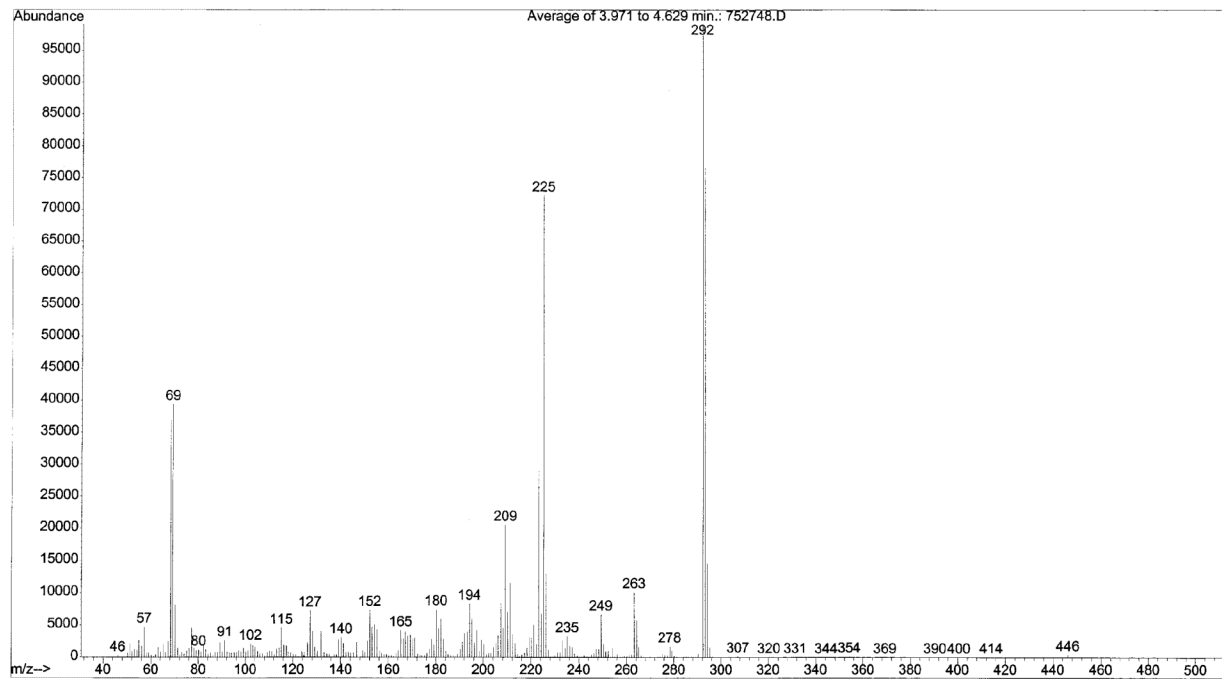
Current Data Parameters
 NAME 1eb
 EXPNO 180
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20001206
 Time 12.12
 INSTRUM spect
 PROBHD 5 mm QNP 1H/
 PULPROG zg
 TD 32768
 SOLVENT CDCl3
 NS 16
 DS 2
 SMH 6172.839 Hz
 FIDRES 0.188380 Hz
 AQ 2.6542580 sec
 RG 456.1
 DN 81.000 usec
 DE 6.00 usec
 TE 300.0 K
 D1 1.0000000 sec
 P1 5.50 usec
 SF01 300.1318534 MHz
 NUC1 1H
 PL1 -6.00 dB

F2 - Processing parameters
 SI 16384
 SF 300.130051 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00

1D NMR plot parameters
 CX 20.00 cm
 F1P 11.000 ppm
 F1 3301.43 Hz
 F2P -1.000 ppm
 F2 -300.13 Hz
 PPMCM 0.60000 ppm/cm
 HZCM 180.07800 Hz/cm

File : C:\HPCHEM\1\DATA\SAMPLES\752748.D
Operator : R. Mitchell
Acquired : 17 Jul 2001 10:01 am using AcqMethod EIDIPHED
Instrument : GC/MS Ins
Sample Name: RTI-7527-48
Misc Info : Abraham
Vial Number: 1

5d

Date: Wed, Jul 18, 2001 10:07 AM

5d

Data: Epibatidine-18JUL101-003

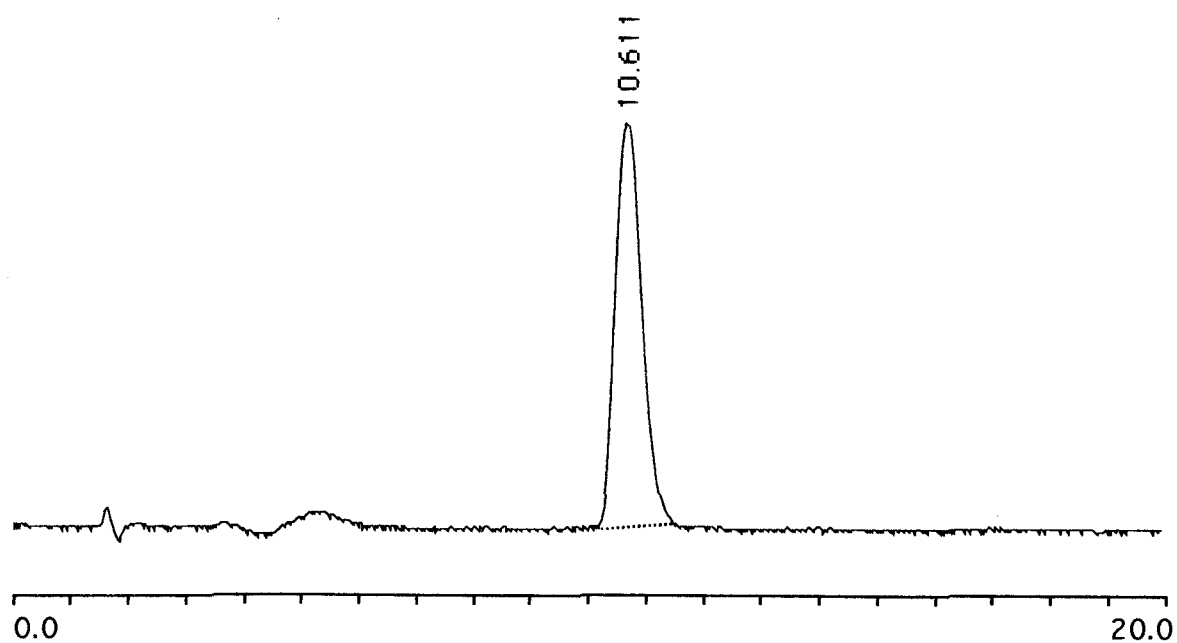
Sample: RTI-7527-48 (about 1 mg/ml CH₃OH), A: [TFA-H₂O], B: CH₃CN, 15% B,
0.6 ml/min, 225 nm, Phenomenex Synergi 4 μ Hydro-RP 80A (3*150 mm,
4 μ m)

Method: Epibatidine

Inject Vol: 1

Sampling Int: 0.1 Seconds

Data:



Analysis: Channel A

| Peak No. | Time | Type | Height(μ V) | Area(μ V-sec) | Area% |
|------------|--------|------|------------------|--------------------|---------|
| 1 | 10.611 | N | 16462 | 484919 | 100.000 |
| Total Area | | | | 484919 | 100.000 |