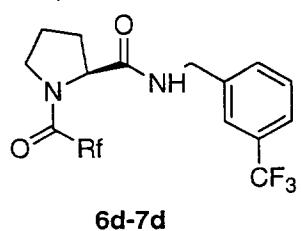
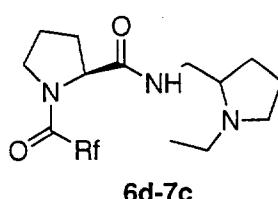
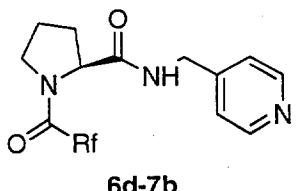
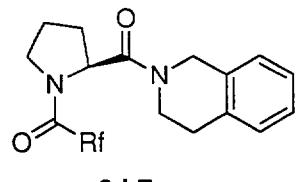
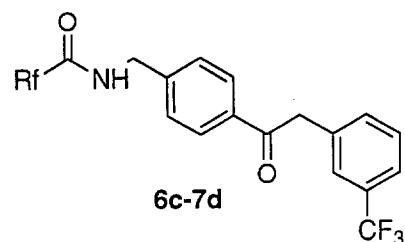
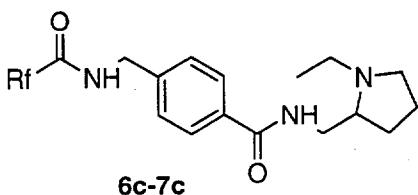
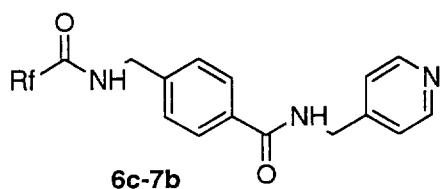
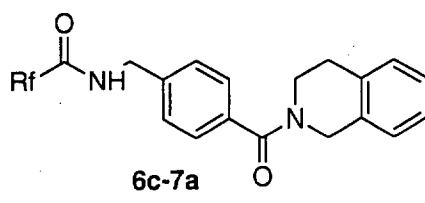
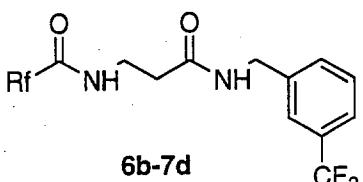
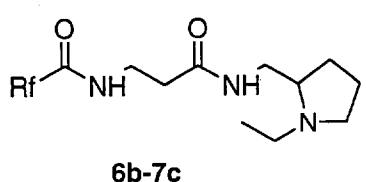
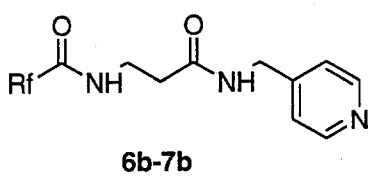
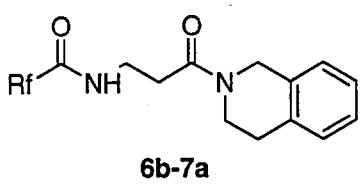
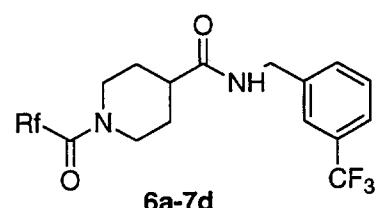
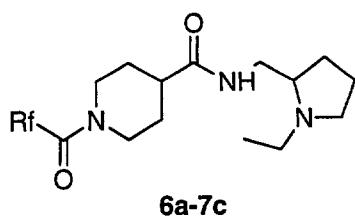
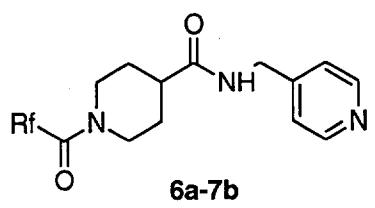
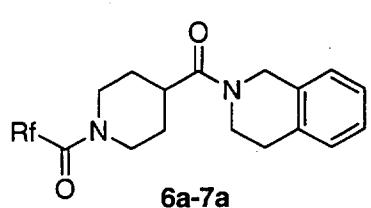


Structures of the products of 4 x 4 amide library



General: Unless otherwise noted, all reagents and solvents were used from commercial sources as received. Anhydrous solvents were obtained as follows: ethyl ether was distilled from sodium/benzophenone under argon; dichloromethane was distilled from calcium hydride. NMR spectra were recorded at 300 MHz for ¹H, 75 MHz for ¹³C and 282 MHz for ¹⁹F. The chemical shift values are in parts per million (ppm) using the CDCl₃ as reference unless otherwise specified. Infrared spectra were obtained from an IBM IR/32 system and samples were run as thin film. Low resolution mass spectra (LRMS) were recorded on a Hewlett Packard-9000 GC-MS system. High resolution mass spectroscopy were taken on a Varian MATCH-5 DF spectrometer. Melting points were measured on a MEL-TEMP II apparatus without correction.

(3,4-Dihydro-1*H*-isoquinolin-2-yl)-piperidin-4-yl-methanone (1).ⁱ *N*-Trifluoroacetyl isonipecotic acidⁱⁱ (2.56 g, 11.4 mmol), tetrahydroisoquinoline (1.82 g, 13.7 mmol), EDCI (2.63 g, 13.7 mmol), HOBT (1.85 g, 13.7 mmol) and triethylamine (1.38 g, 13.7 mmol) were stirred at 25 °C in dry dichloromethane for 6 h. The mixture was quenched with water. The organic phase was dried over MgSO₄ and purified by column chromatography (40/60 EtOAc/hexanes). The solid obtained was stirred with excess K₂CO₃ in MeOH at 25 °C overnight. After evaporation of MeOH, the residue was partitioned between dichloromethane and basic water. Evaporation of the organic layer after drying over MgSO₄ gives pure product. Yield 2.12 g (76% for two steps). ¹H NMR (CDCl₃) (mixture of two rotamers) δ 7.23–7.16 (m, 4 H), 4.73 (s, 1 H), 4.67 (s, 1 H), 3.83 (t, *J* = 5.9 Hz, 1 H), 3.74 (t, *J* = 5.8 Hz, 1 H), 3.21–3.16 (m, 2 H), 2.92 (t, *J* = 5.7 Hz, 1 H), 2.85 (t, *J* = 5.7 Hz, 1 H), 2.76–2.67 (m, 3 H), 2.29 (s, 1 H), 1.80–1.73 (m, 4 H); ¹³C NMR (CD₃OD-CDCl₃) δ 175.5, 175.3, 135.8, 135.1, 134.0, 133.8, 129.6, 129.3, 127.9, 127.6, 127.4, 127.3, 127.0, 48.2, 45.8, 45.4, 44.2, 41.4, 40.2, 39.6, 39.5, 30.5, 29.5, 29.4, 29.1; LRMS 244 (M⁺, 37), 188 (100), 132 (74); HRMS calcd 244.1576, found 244.1574. MP 75–76 °C.

General Procedures for the Preparation of Compounds 1a-1i.

Perfluorodecanoyl Chloride. Perfluorodecanoic acid (3.50 g, 6.81 mmol) was refluxed in thionyl chloride with 45 mg pyridine for 4 h. The mixture was partitioned between dichloromethane and FC-72. The dichloromethane layer was further washed with FC-72 for three times. The combined FC-72 phase was evaporated at 10 °C to give 3.66 g of colorless oil. Yield 100%. IR 1801.2, 1335.5, 1240.0, 1151.3 cm⁻¹.

Compounds 1a-1i. (C₃F₇CO)₂O, C₄F₉COCl,ⁱⁱⁱ C₅F₁₁COCl, C₆F₁₃COCl, C₇F₁₅COCl, C₈F₁₇COCl, C₉F₁₉COCl, C₁₀F₂₁COCl, or C₇H₁₅COCl was added to a solution of triethylamine and compound **1** in dry dichloromethane at 0 °C. The reaction mixture was further stirred at 25°C for 1 h before quenching with water. The dichloromethane layer was dried over MgSO₄ and purified by column chromatography to give pure compounds **1a-1i**. Characterization data for **1f** are representative: ¹H NMR (CDCl₃) (mixture of rotamers) δ 7.23–7.14 (m, 4 H), 4.74 (s, 1 H), 4.68 (s, 1 H), 4.48 (d, *J* = 8.8 Hz, 1 H), 4.19 (d, *J* = 11.4 Hz, 1 H), 3.89 –3.81 (m, 1 H), 3.75 (t, *J* = 5.8 Hz, 1 H), 3.35–3.27 (m, 1 H), 3.09–3.02 (m, 1 H), 2.97–2.86 (m, 3 H), 1.95–1.88 (m, 4 H). ¹⁹F NMR (CDCl₃, relative to CCl₃F) δ –81.2 (3 F), –111.3 (2 F), –120.9 (2 F), –121.5 (2 F), –122.2 (6 F), –123.2 (2 F), –126.6 (2 F); LRMS 740 (M⁺, 45), 188 (100), 132 (58); HRMS calcd 740.1143, found 740.1177.

Fluorous HPLC Experiments.

A mixture of compounds **1a-1i** was injected onto a FluofixTM 120E column (Keystone Scientific) and eluted with a linear gradient of 20/80 H₂O-MeOH to pure MeOH over 30 min. Each individual peak in the HPLC chromatogram was identified by injecting the authentic sample and eluting with the same solvent gradient.

General Procedure for the Preparation of *N*-Perfluorodecanoyl Amino Acids.

The *N*-perfluoroacylated amino acids were prepared from the corresponding amino acids in a three-step sequence of benzyl esterification-acylation-hydrogenolysis if the benzyl ester of the amino acid was not commercially available. Otherwise, the benzyl esters of amino acids were acylated with perfluorodecanoyl chloride followed by hydrogenolysis. Thus, perfluorodecanoyl chloride (1 equiv) was added slowly to an ice-cooled mixture of amino acid benzyl esters (1 equiv) and triethylamine (1 equiv) in dry dichloromethane. The reaction mixture was quenched with water after stirring at 25°C for 1 h. The dichloromethane phase was dried over MgSO₄ and purified by column chromatography. The resulting *N*-perfluorodecanoyl amino acid benzyl ester was hydrogenated with 10 % Pd/C in methanol for 5 h. The catalyst was removed by filtration through celite and the filtrate was evaporated to dryness to give pure *N*-perfluorodecanoyl amino acid which was used without further purification.

***N*-Perfluorodecanoyl Isonipeptic Acid Benzyl Ester.** ¹H NMR (CDCl₃) δ 7.41–7.34 (m, 5 H), 5.16 (s, 2 H), 4.32 (d, *J* = 13.3 Hz, 1 H), 4.04 (d, *J* = 13.1 Hz, 1 H), 3.30 (t, *J* = 11.2 Hz, 1 H), 3.08 (t, *J* = 10.8 Hz, 1 H), 2.74–2.64 (m, 1 H), 2.03 (**br**, 2 H), 1.87–1.75 (m, 2 H); ¹³C NMR (CDCl₃) δ 173.2, 155.7 (t), 135.8, 128.5, 128.2, 128.0, 115.2 –107.2 (m), 66.4, 44.8, 43.0, 40.3, 28.3, 27.5; ¹⁹F NMR (CDCl₃, relative to CCl₃F) δ –81.2 (3 F), –111.4 (2 F), –120.9 (2 F), –121.5 (2 F), –122.1 (6 F), –123.2 (2 F), –126.6 (2 F); LRMS 715 (M⁺, 10), 624 (100), 578 (65), 91 (47); HRMS calcd 715.0830, found 715.0827. MP 65 – 66 °C.

***N*-Perfluorodecanoyl Isonipeptic Acid.** ¹H NMR (CDCl₃) δ 4.35 (d, *J* = 13.1 Hz, 1 H), 4.06 (d, *J* = 13.7 Hz, 1 H), 3.34 (t, *J* = 11.0 Hz, 1 H), 3.13 (t, *J* = 10.7 Hz, 1 H), 2.76–2.67 (m, 1 H), 2.09 (**br**, 2 H), 1.88–1.76 (m, 2 H); ¹³C NMR (CD₃OD) δ 177.5, 157.3 (t), 116.7–108.8 (m), 46.5, 44.6, 41.5, 30.0, 29.1; ¹⁹F NMR (CDCl₃, relative to

CCl_3F) δ -81.2 (3 F), -111.0 (2 F), -119.0 (2 F), -120.3 (2 F), -121.6 (6 F), -123.2 (2 F), -126.6 (2 F); LRMS 625 (M^+ , 31), 606 (24), 128 (100), 69 (36); HRMS calcd 625.0357, found 625.0345. MP 124–127 °C.

N-Perfluorodecanoyl L-Proline Benzyl Ester. ^1H NMR (CDCl_3) δ 7.4–7.35 (m, 5 H), 5.20 (d, J = 3.9 Hz, 2 H), 4.69–4.65 (m, 1 H), 3.91–3.78 (m, 2 H), 2.27–2.20 (m, 2 H), 2.12–2.00 (m, 3 H); ^{13}C NMR (CDCl_3) δ 170.4, 156.7 (t), 135.2, 129.6, 128.5, 128.3, 128.1, 122.9 –107.0 (m), 67.1, 61.1, 59.7, 48.8, 47.5, 31.6, 28.0, 25.0, 20.7; ^{19}F NMR (CDCl_3 , relative to CCl_3F) δ -81.2 (3 F), -115.6 (2 F), -120.9 (2 F), -121.9 (8 F), -123.2 (2 F), -126.6 (2 F); LRMS 701 (M^+ , 18), 566 (100), 91 (49), 69 (37); HRMS calcd 701.0670, found 701.0699. MP 71–72 °C.

N-Perfluorodecanoyl L-Proline. ^1H NMR (CDCl_3) δ 4.65–4.61 (m, 1 H), 3.88–3.79 (m, 2 H), 2.34–2.03 (m, 4 H); ^{13}C NMR (CD_3OD) δ 174.9, 174.2, 157.9 (t), 120.5–108.1 (m), 62.8, 32.9, 29.4, 26.3, 22.0; ^{19}F NMR (CDCl_3 , relative to CCl_3F) δ -81.2 (3 F), -115.6 (2 F), -121.6 (2 F), -122.1 (8 F), -123.3 (2 F), -126.7 (2 F); LRMS 611 (M^+ , 20), 592 (37), 566 (100), 131 (36), 96 (57), 69 (87); HRMS calcd 611.0201, found 611.0216. MP 113–116 °C.

N-Perfluorodecanoyl β -Alanine Benzyl Ester. ^1H (CDCl_3) δ 7.41–7.35 (m, 5 H), 7.12 (s, 1 H), 5.17 (s, 2 H), 3.67 (q, J = 5.9 Hz, 2 H), 2.67 (t, J = 5.8 Hz, 2 H); ^{19}F NMR (CDCl_3 , relative to CCl_3F) δ -81.2 (3 F), -120.5 (2 F), -122.3 (8 F), -123.9 (4 F), -126.6 (2 F); ^{13}C NMR (CDCl_3) δ 172.1, 157.8 (t), 135.4, 128.9, 128.5, 127.5, 119.2–107.1 (m), 67.2, 38.5, 35.7; LRMS 675 (M^+ , 42), 568 (85), 526 (63), 199 (87), 108 (100), 91 (97); HRMS calcd 675.0514, found 675.0531. MP 83–84 °C.

N-Perfluorodecanoyl β -Alanine. ^1H NMR (CD_3OD) δ 3.55 (t, $J = 6.8$ Hz, 2 H), 2.58 (t, $J = 6.8$ Hz, 2 H); ^{13}C NMR (CD_3OD) δ 174.9, 159.5, 37.4, 34.1; ^{19}F NMR (CD_3OD , relative to CCl_3F) δ -81.2 (3 F), -119.5 (2 F), -121.7 (8 F), -123.2 (4 F), -126.2 (2 F); LRMS 585 (M^+ , 10), 539 (23), 98 (100), 70 (56); HRMS calcd 585.0044, found 585.0064.

N-Perfluorodecanoyl 4-Aminomethyl Benzoic Acid Benzyl Ester. ^1H NMR (CDCl_3) δ 8.03 (d, $J = 8.3$ Hz, 2 H), 7.44–7.31 (m, 7 H), 6.74 (s, 1 H), 5.35 (s, 2 H), 4.60 (d, $J = 5.9$ Hz, 2 H).

N-Perfluorodecanoyl 4-Aminomethyl Benzoic Acid. ^1H (CD_3OD) δ 8.00 (d, $J = 8.2$ Hz, 2 H), 7.40 (d, $J = 8.1$ Hz, 2 H), 4.55 (s, 2 H). MP 200 – 203 °C.

General Procedure for the Synthesis of the Library of Amides.

To sixteen test tubes were added *N*-perfluorodecanoyl amino acid (0.05 mmol), and stock solutions of amines (0.20 mmol), EDCI (0.07 mmol), Et_3N (0.07 mmol) and HOBT (0.07 mmol). The reaction mixtures were stirred at 25°C for 15 h. The reaction mixtures were quenched with water and the dichloromethane layers were transferred to another sixteen tubes, which were evaporated to dryness simultaneously with a vacuum centrifuge. The residue were taken up in minimum amount of methanol and charged onto sixteen short columns with about 2 g of fluorous reverse phase silica, which was previously conditioned with ether and 20:80 H_2O -MeOH sequentially. The short columns were first eluted with 20:80 H_2O -MeOH (8 ml) to remove excess amines and coupling reagents, and then with acetonitrile (8 ml). The acetonitrile fractions were evaporated on a vacuum centrifuge to dryness to give the sixteen-membered library of *N*-perfluorodecanoyl amides. Raw data for all the products are shown in the following pages. The data for the amide formed from **6a** and **7a** are representative: ^1H NMR (CDCl_3) δ 7.23–7.14 (m, 4 H), 4.85 (s, 1 H), 4.74 (s, 1 H), 4.50–4.43 (m, 1 H), 4.22–4.18 (m, 1 H), 3.89–3.81 (m, 1 H),

3.75 (t, $J = 5.9$ Hz, 1 H), 3.32 (t, $J = 12.5$ Hz, 1 H), 3.09–3.02 (m, 1 H), 2.97–2.86 (m, 3 H), 1.88 (br, 4 H); ^{19}F NMR (CDCl_3 , relative to CCl_3F), δ –81.2 (3 F), –111.3 (2 F), –120.9 (2 F), –121.5 (2 F), –122.1 (6 F), –123.2 (2 F), –126.6 (2 F); LRMS 740 (M^+ , 41), 721 (12), 188 (100), 132 (58); HRMS calcd 740.1143, found 740.1177.

Procedures for the Deprotection of the Library of Fluorous Amides.

For amides made from isonipecotic acid: The amides were stirred with K_2CO_3 in MeOH at 25 °C overnight. After evaporation of methanol, the residue was partitioned between water and ethyl acetate. The organic phase was dried over MgSO_4 and passed through a pipette column eluted with mixture of methanol and ethyl acetate.

For amides made from β -alanine: The amides were heated with LiOH in methanol at 70 °C overnight. After evaporated to dryness, the residue was suspended in ethyl acetate, to which triethylamine and acetic anhydride was added. The reaction mixture was quenched with water after stirring at 25 °C for 30 min. The organic layer was dried over MgSO_4 and the crude product was purified by column chromatography.

For amides made from L-proline: The amides were heated with NaBH_4 in ethanol at 60 °C until TLC showed completion of the reaction (from 1 h to overnight). Then they were treated in the same way as those amides made from β -alanine.

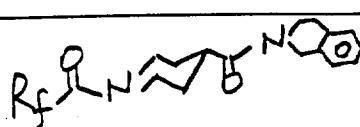
i. Patent, Pfizer Corp. DD 143613, 1979; DE 2919800. *Chem. Abstr.* **1980**, 92, 181222.

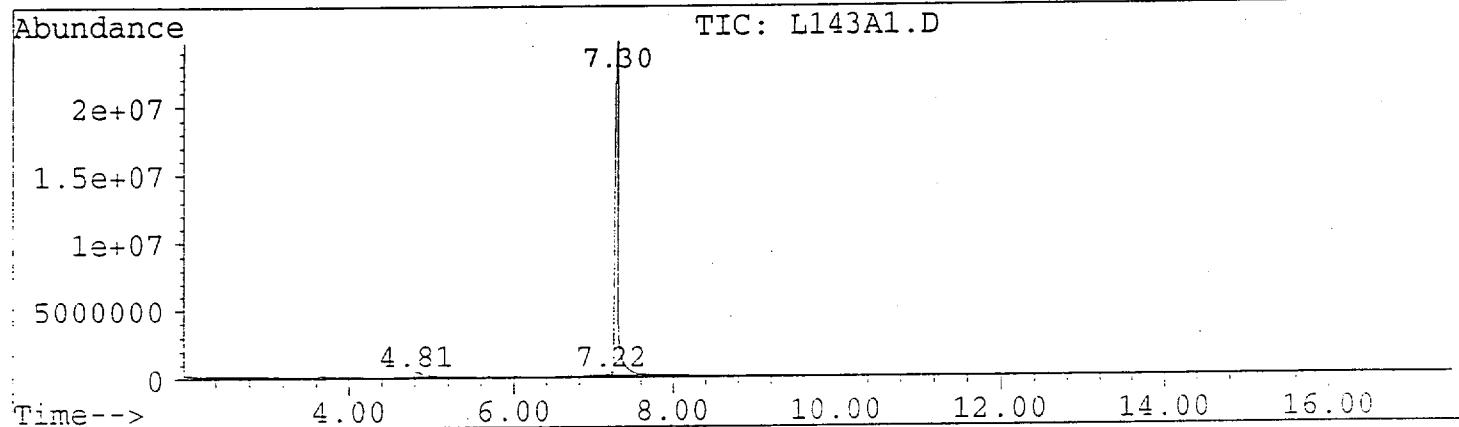
ii. Hibert, M. F.; Hoffmann, R.; Miller, R. C.; Carr, A. A. *J. Med. Chem.* **1990**, 1594.

iii. Simons, J. H.; Black, W. T.; Clark, R. R. *J. Am. Chem. Soc.* **1953**, 5621.

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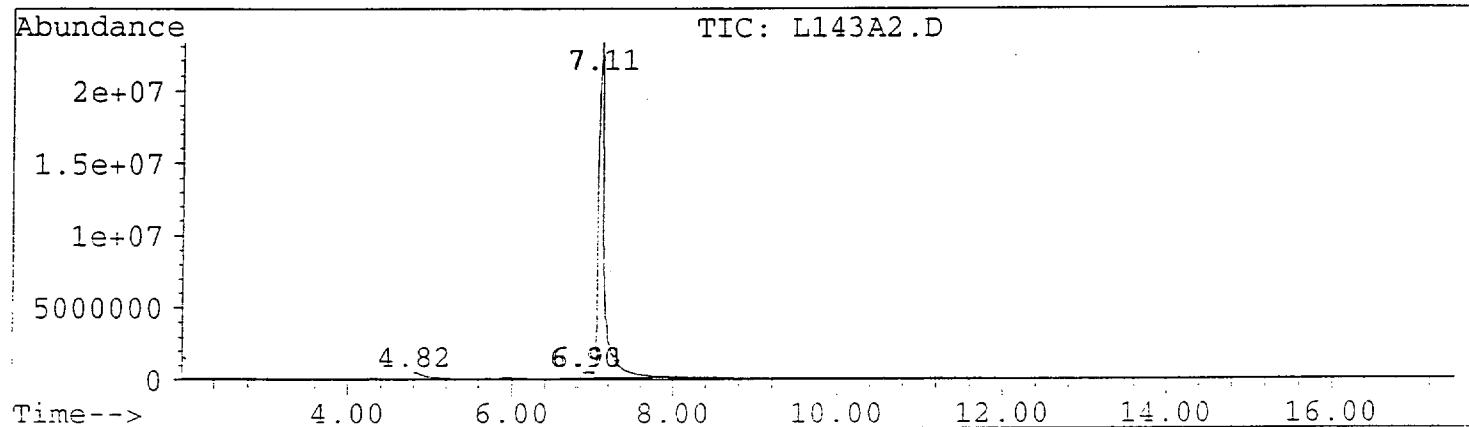
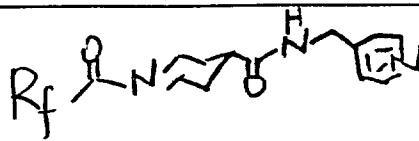


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Area Percent Report -- Sorted by Signal

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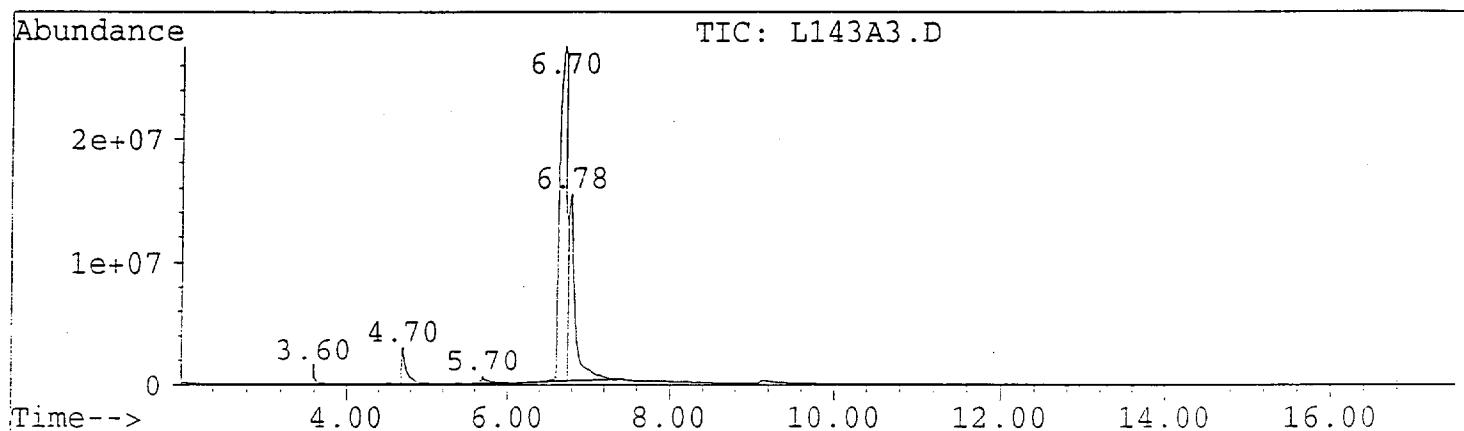
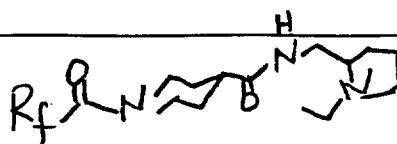
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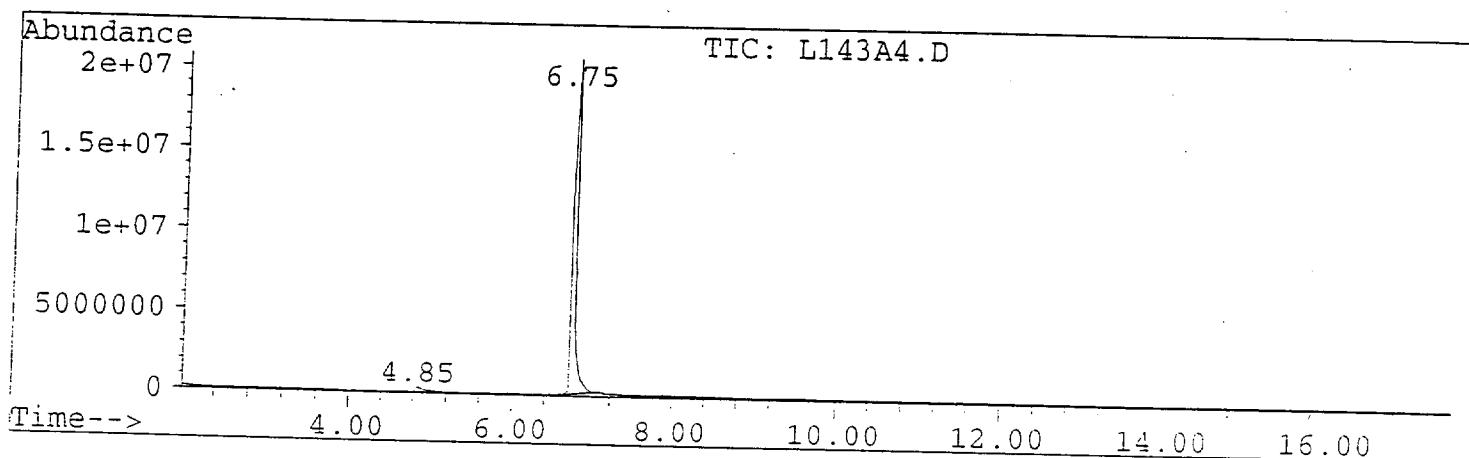
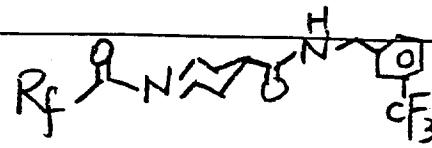


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Area Percent Report -- Sorted by Signal

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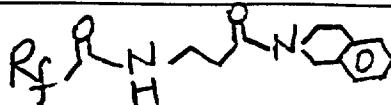
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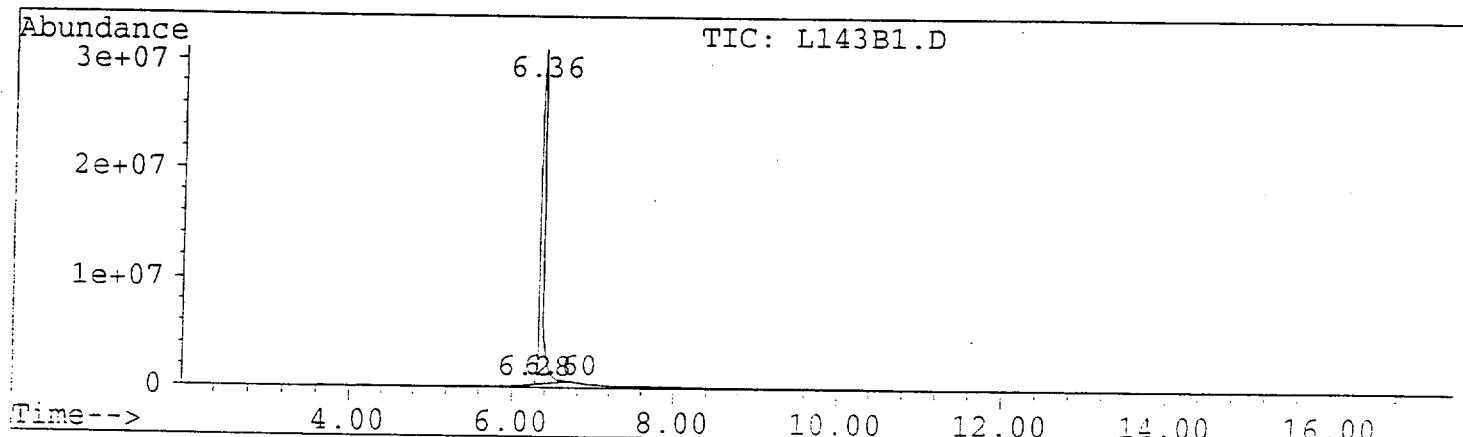


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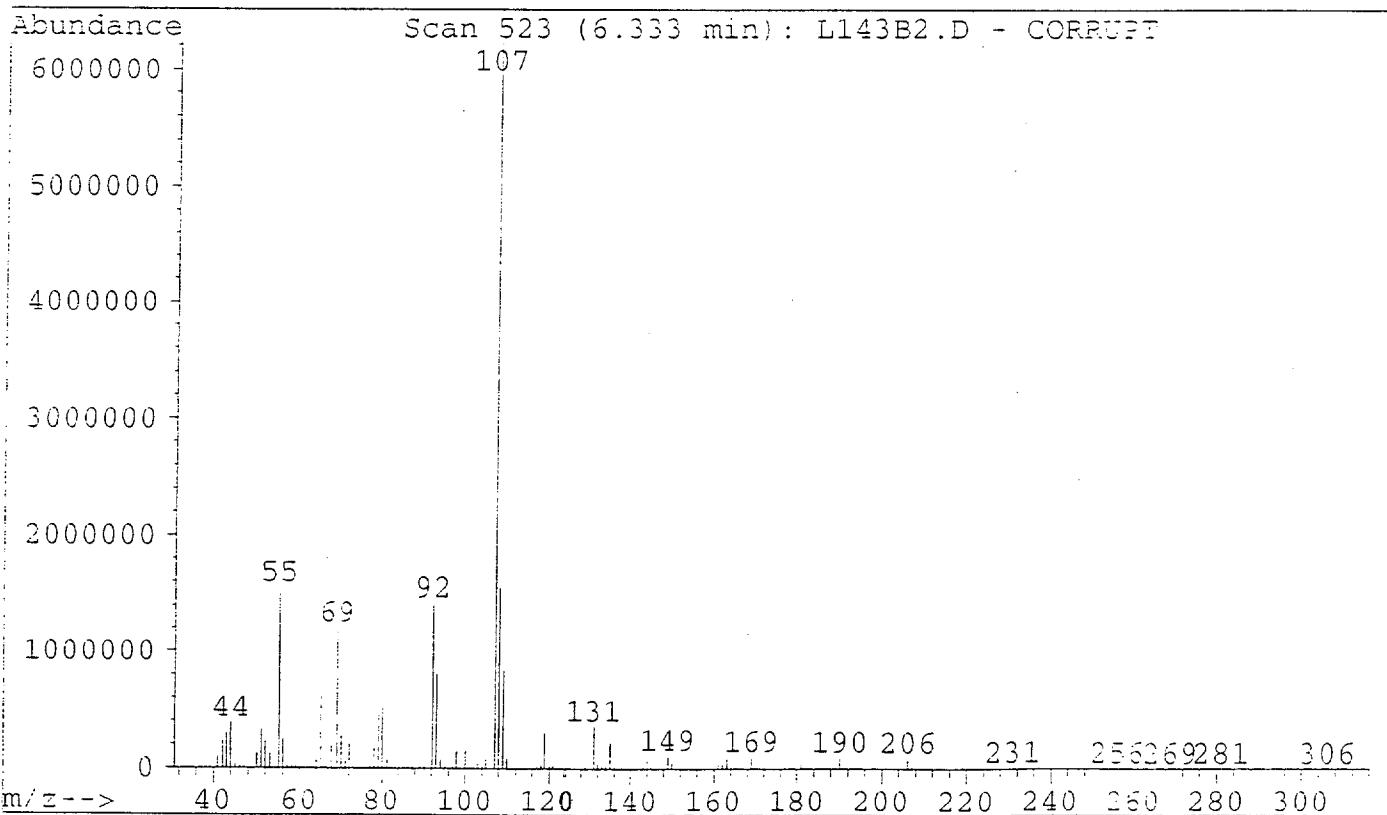
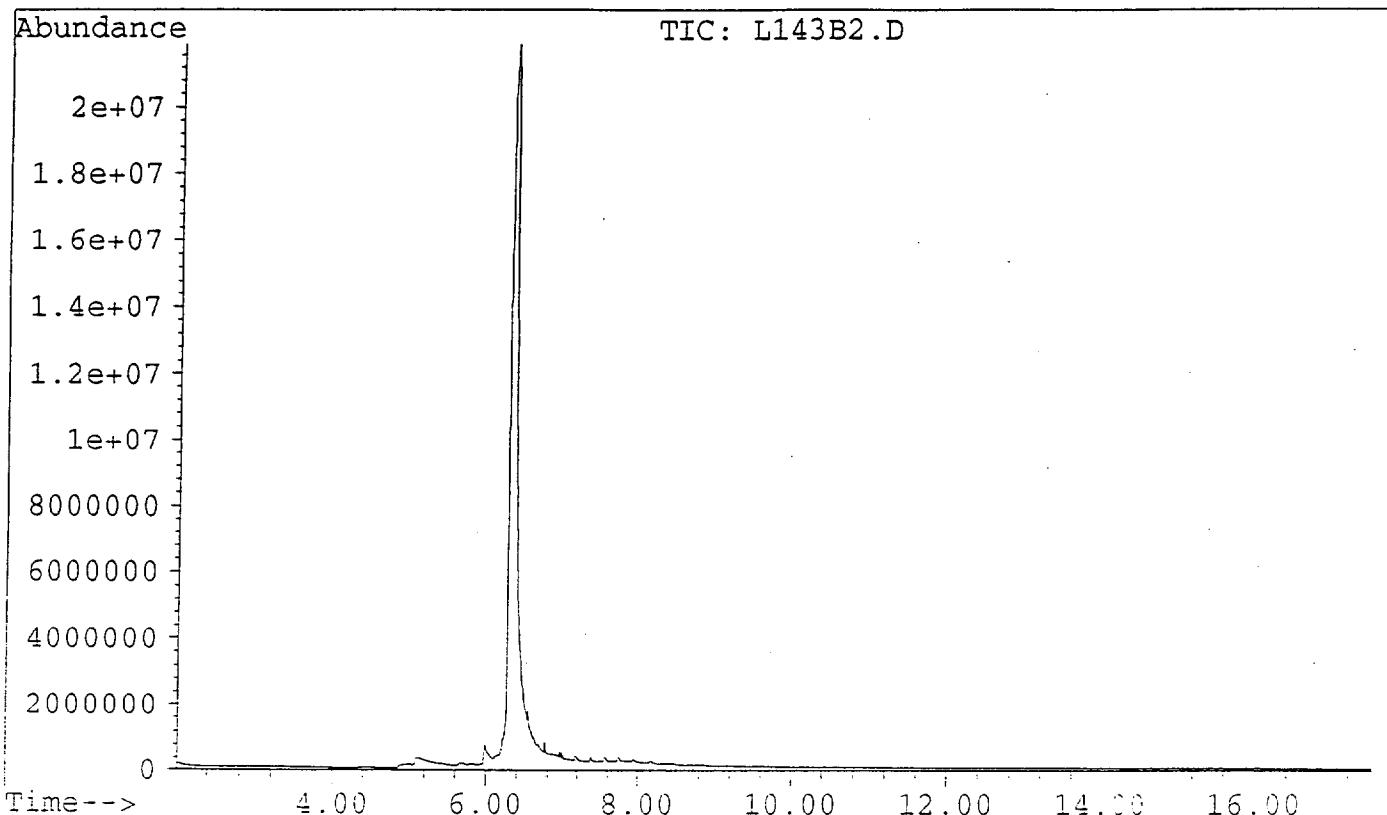
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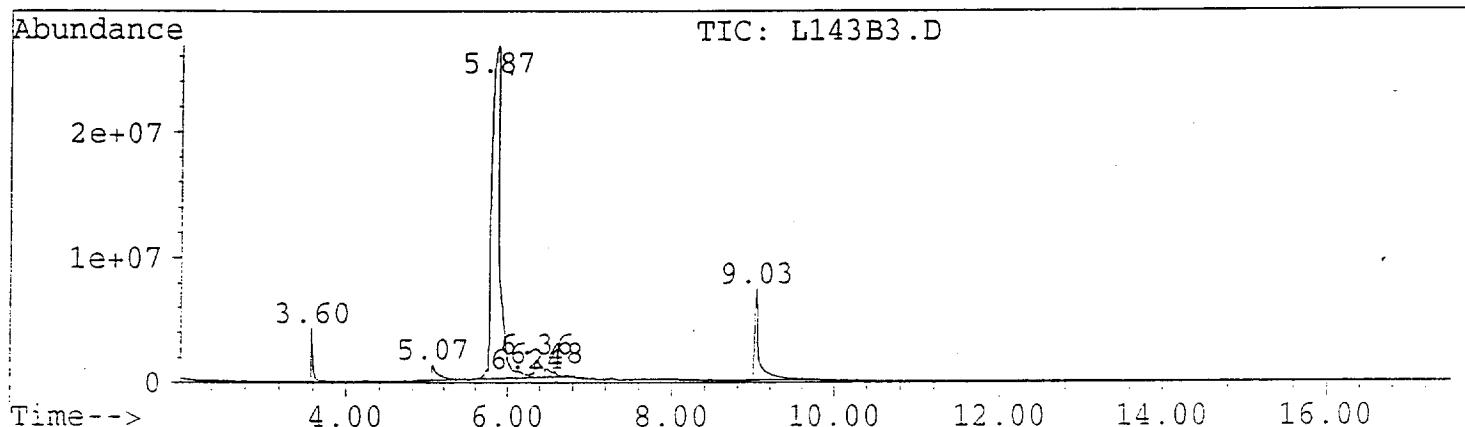
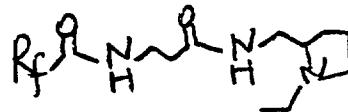
Acquired : 28 Jan 99 5:30 pm using AcqMethod ZYLUO

Sample Name: 4 x 4 Library of amides B3

Misc Info :

Vial Number: 1

CurrentMeth: C:\HPCHEM\1\METHODS\ZYLUO.M

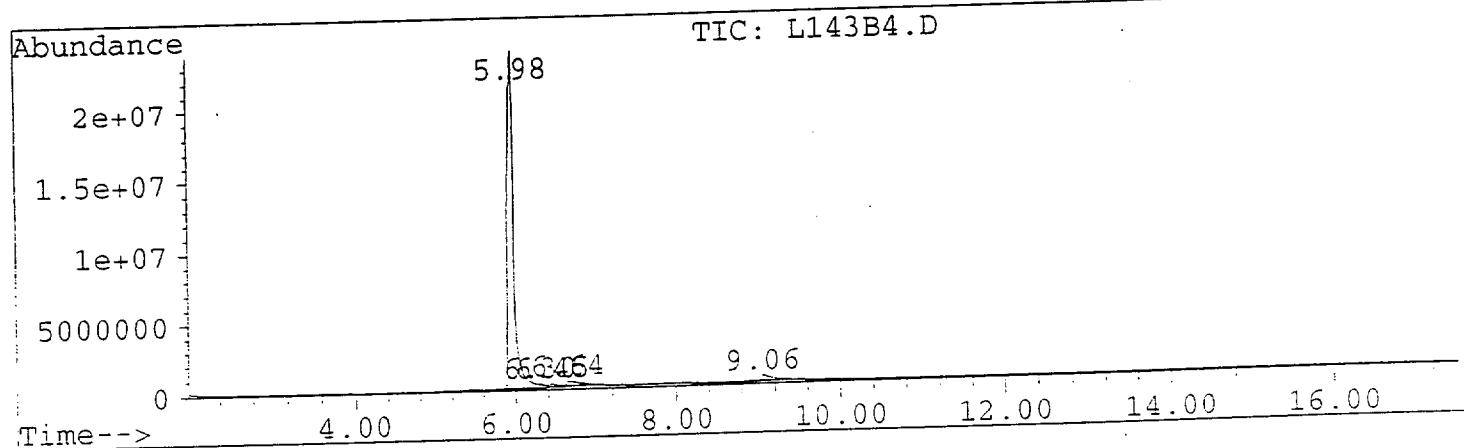
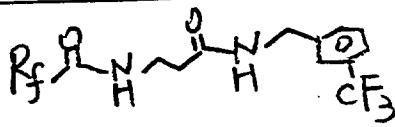


Retention Time	Area	Area %	Ratio %
<hr/>			
Total Ion Chromatogram			
3.596	56835354	2.220	2.831
5.071	76633794	2.993	3.817
5.865	2007581147	78.419	100.000
6.241	7016547	0.274	0.350
6.356	88929795	3.474	4.430
6.483	45123681	1.763	2.248
9.032	277958785	10.857	13.845

Area Percent Report -- Sorted by Signal

Information from Data File:

File : C:\HPCHEM\1\DATA\L143B4.D
 Operator :
 Acquired : 28 Jan 99 5:52 pm using AcqMethod ZYLUO
 Sample Name: 4 x 4 Library of amides B4
 Misc Info :
 Vial Number: 1
 CurrentMeth: C:\HPCHEM\1\METHODS\ZYLUO.M



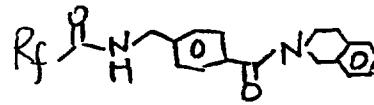
Retention Time	Area	Area %	Ratio %
Total Ion Chromatogram			
5.976	1249872396	92.178	100.000
6.304	11137286	0.821	0.891
6.448	15975903	1.178	1.278
6.640	38835374	2.864	3.107
9.059	40118273	2.959	3.210

Area Percent Report -- Sorted by Signal

Information from Data File:

File : C:\HPCHEM\1\DATA\L143C1.D

Operator :



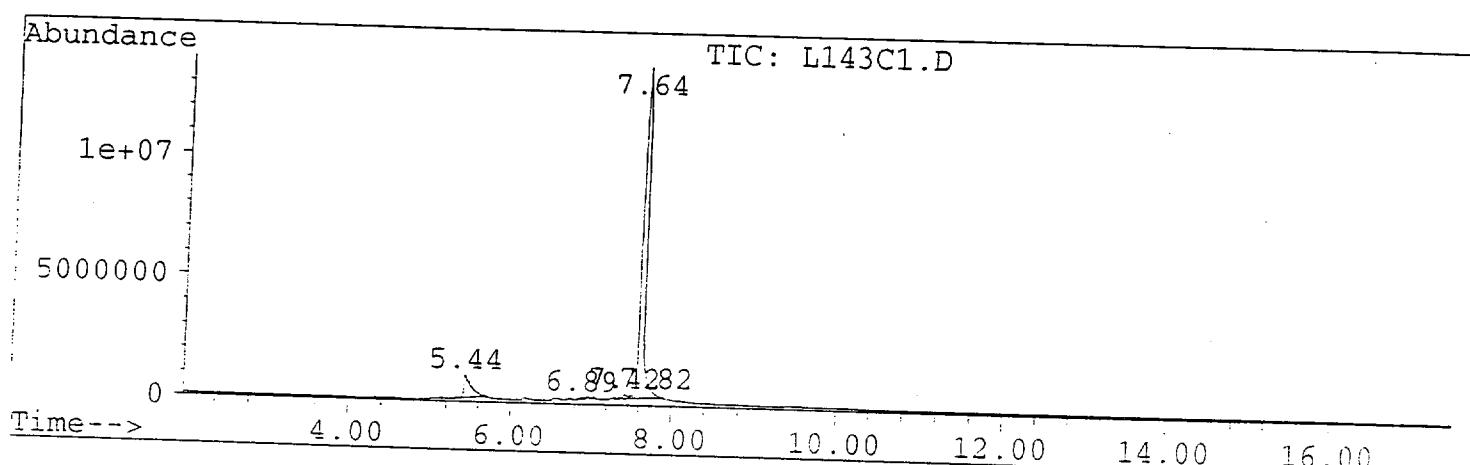
Acquired : 28 Jan 99 6:16 pm using AcqMethod ZYLUO

Sample Name: 4 x 4 Library of amides C1

Misc Info :

Vial Number: 1

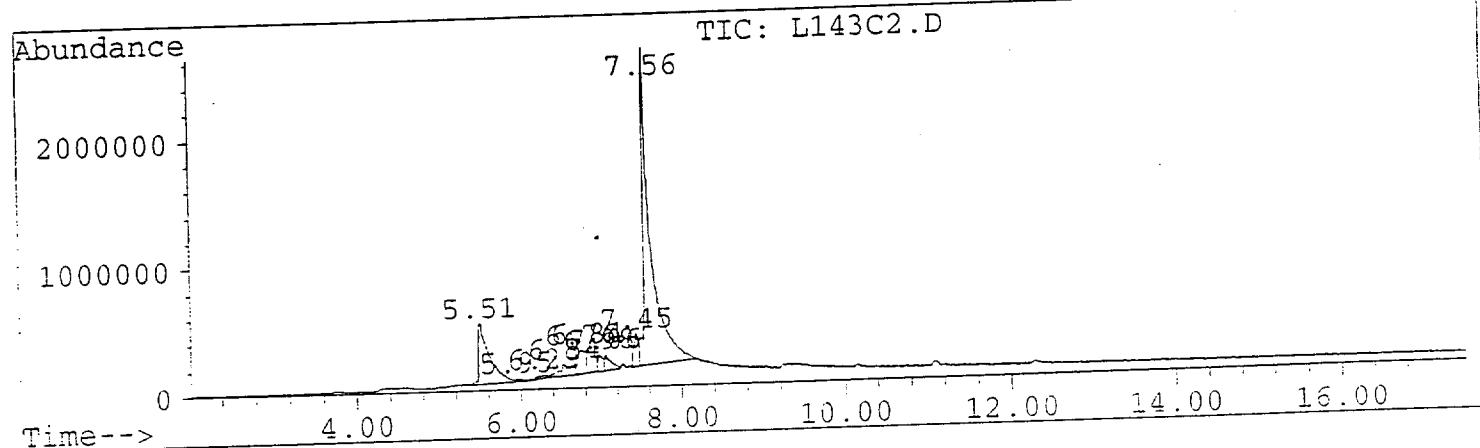
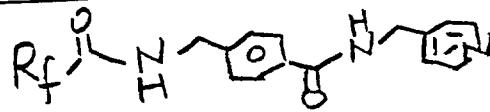
CurrentMeth: C:\HPCHEM\1\METHODS\ZYLUO.M



Retention Time	Area	Area %	Ratio %
Total Ion Chromatogram			
5.438	56943212	9.514	10.812
6.888	3101934	0.518	0.589
7.424	8566019	1.431	1.627
7.636	526645338	87.988	100.000
7.822	3283087	0.549	0.623

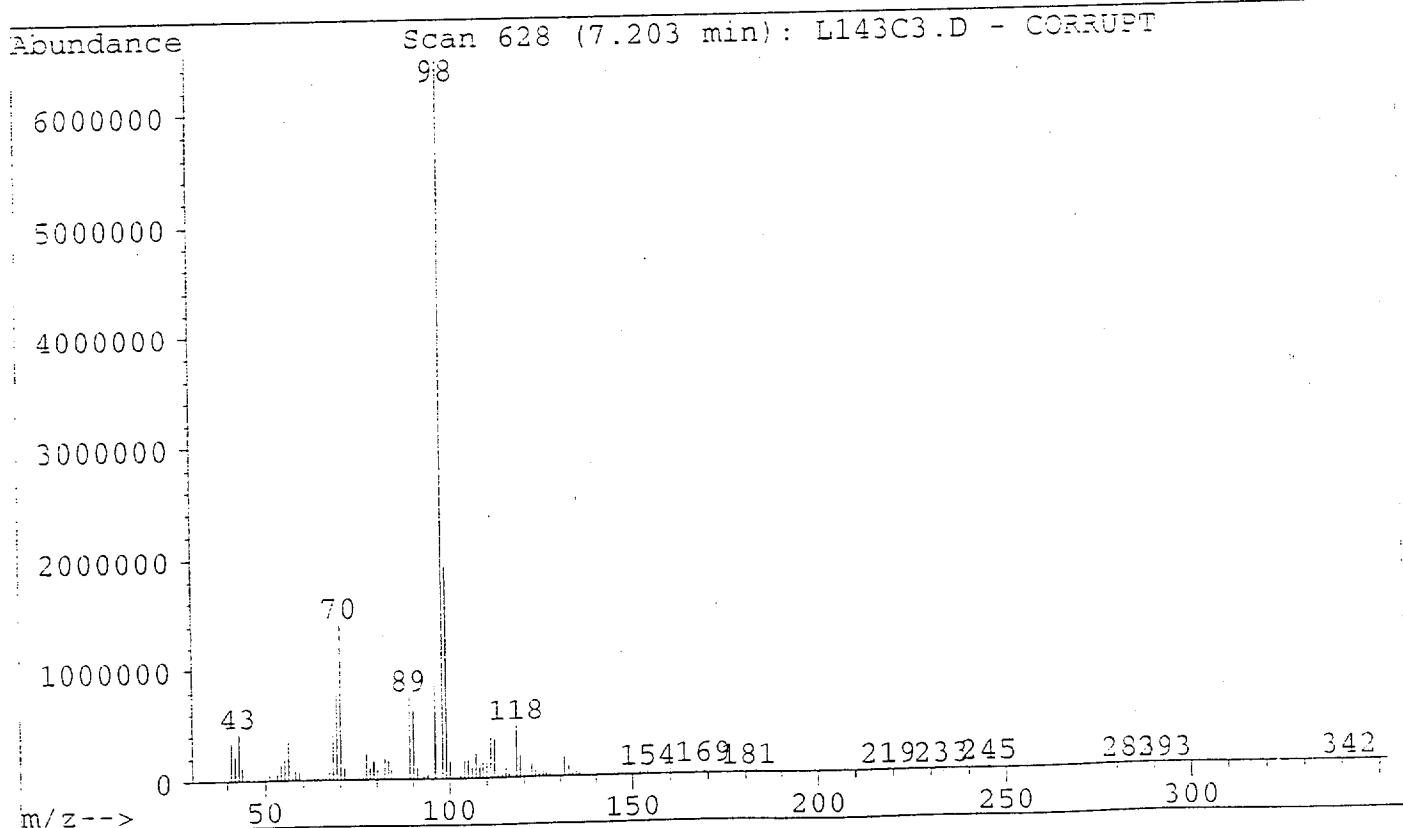
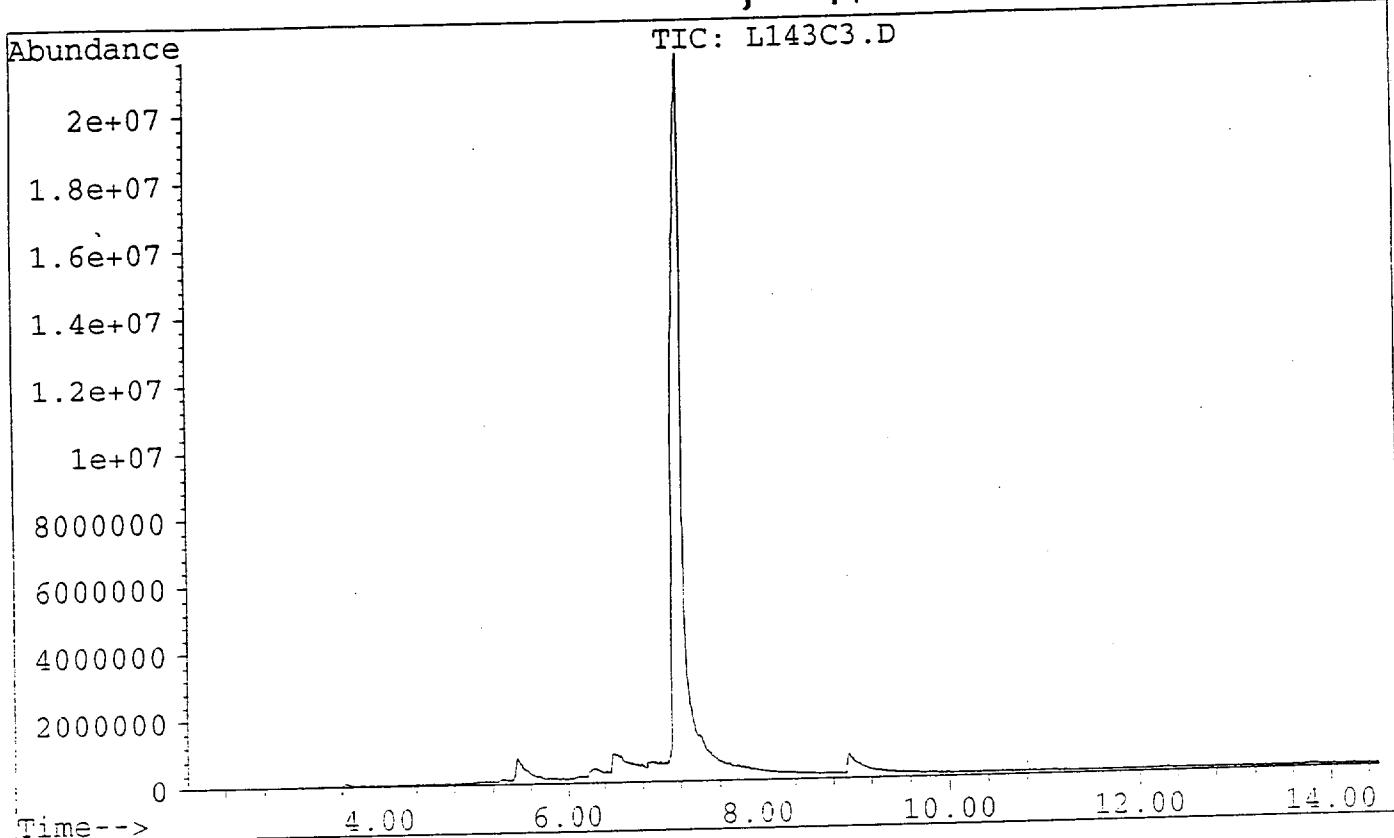
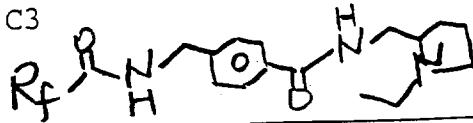
Area Percent Report -- Sorted by Signal

Information from Data File:
 File : C:\HPCHEM\1\DATA\L143C2.D
 Operator :
 Acquired : 28 Jan 99 7:01 pm using AcqMethod ZYLUO
 Sample Name: 4 x 4 Library of amides C2
 Misc Info :
 Vial Number: 1
 CurrentMeth: C:\HPCHEM\1\METHODS\ZYLUO.M



Retention Time	Area	Area %	Ratio %
Total Ion Chromatogram			
5.505	43945145	15.950	24.958
5.950	795771	0.289	0.452
6.287	1888205	0.685	1.072
6.538	9460671	3.434	5.373
6.757	11802596	4.284	6.703
6.837	10303389	3.740	5.852
6.981	5138153	1.865	2.918
7.058	7122994	2.585	4.045
7.451	8986603	3.262	5.104
7.565	176074267	63.907	100.000

File : C:\HPCHEM\1\DATA\L143C3.D
 Operator :
 Acquired : 28 Jan 99 7:22 pm using AcqMethod ZYLUO
 Instrument : 5970B - I
 Sample Name: 4 x 4 Library of amides C3
 Misc Info :
 Vial Number: 1



File : C:\HPCHEM\1\DATA\L143C4.D

Operator :

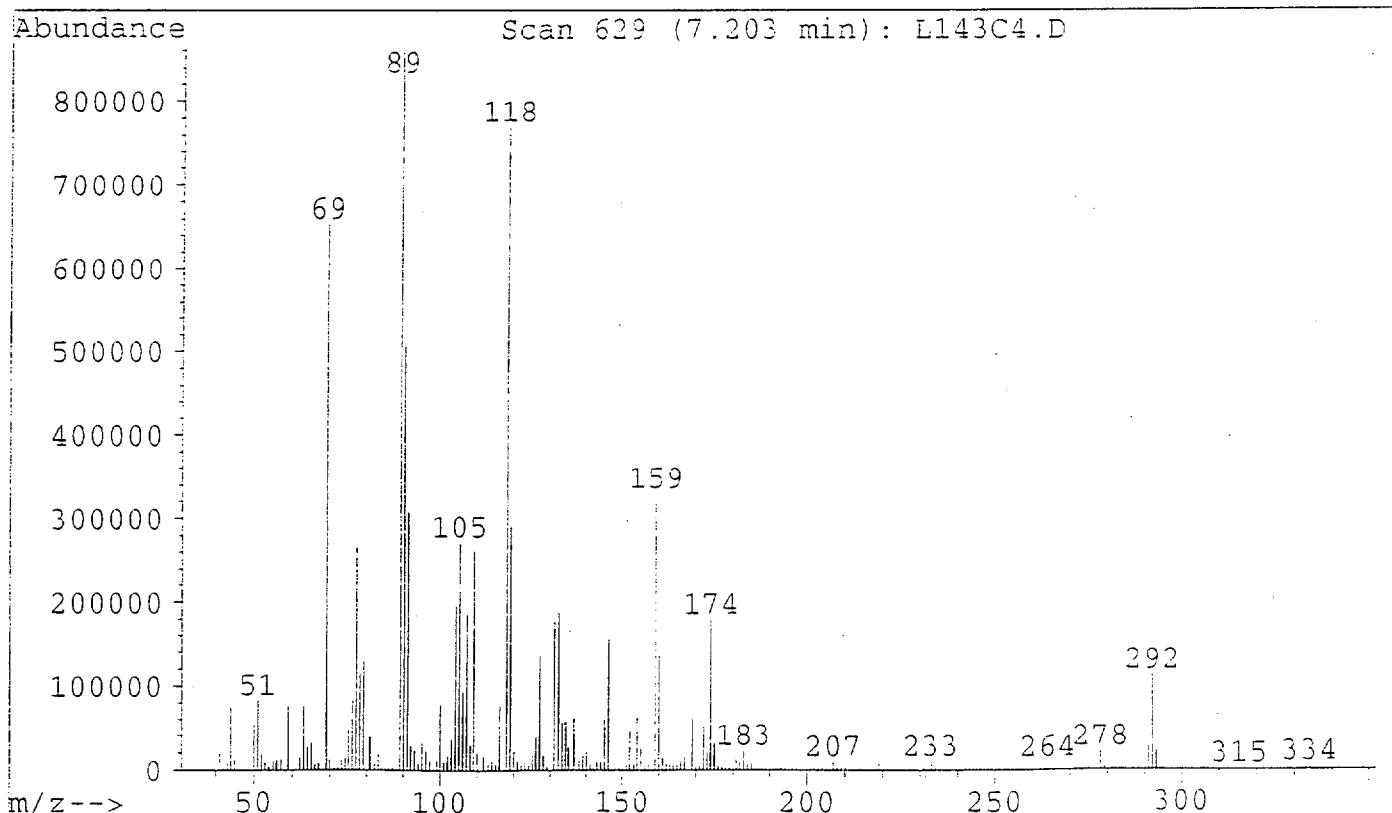
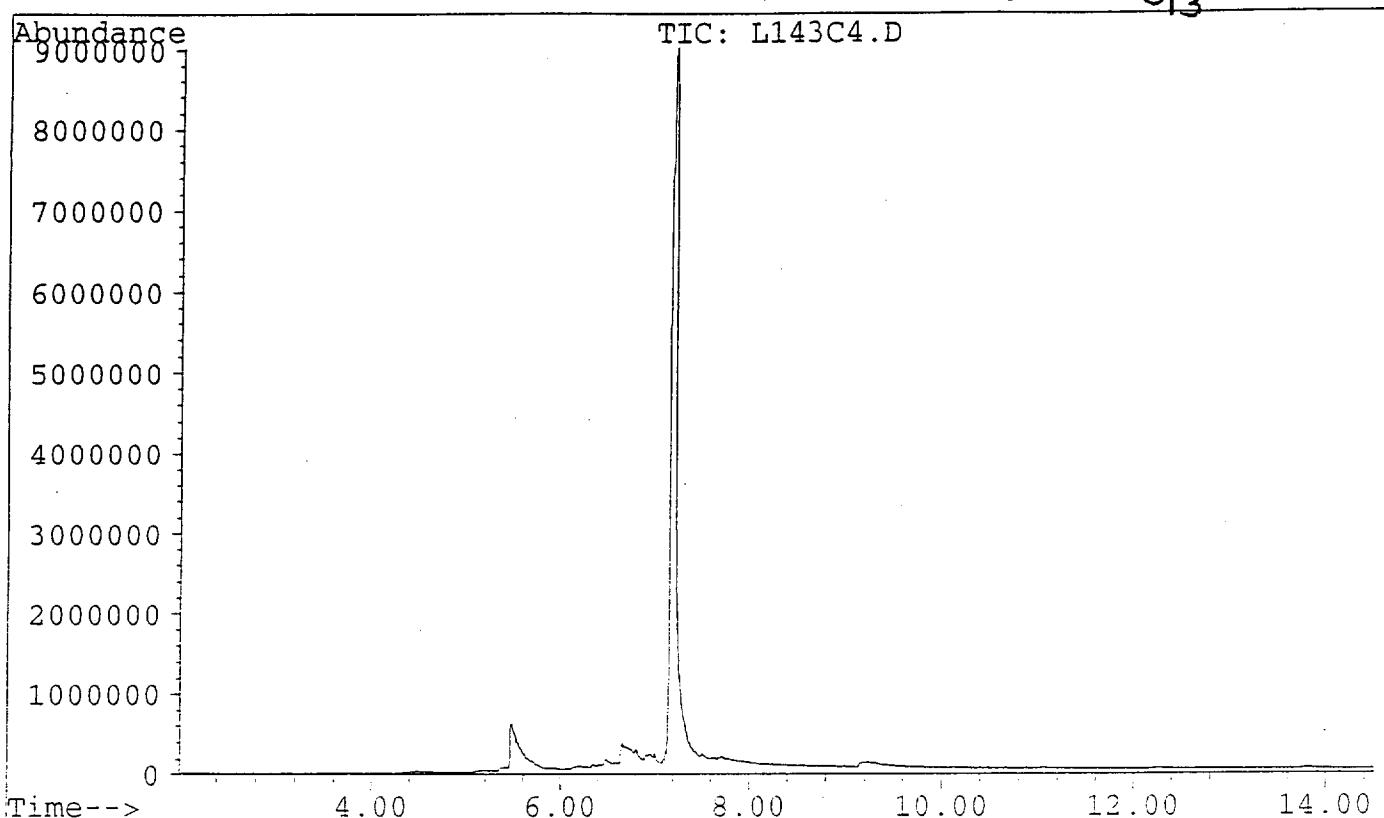
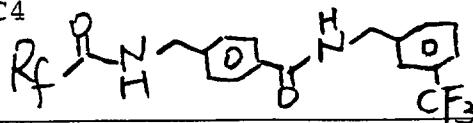
Acquired : 28 Jan 99 7:41 pm using AcqMethod ZYLUO

Instrument : 5970B - I

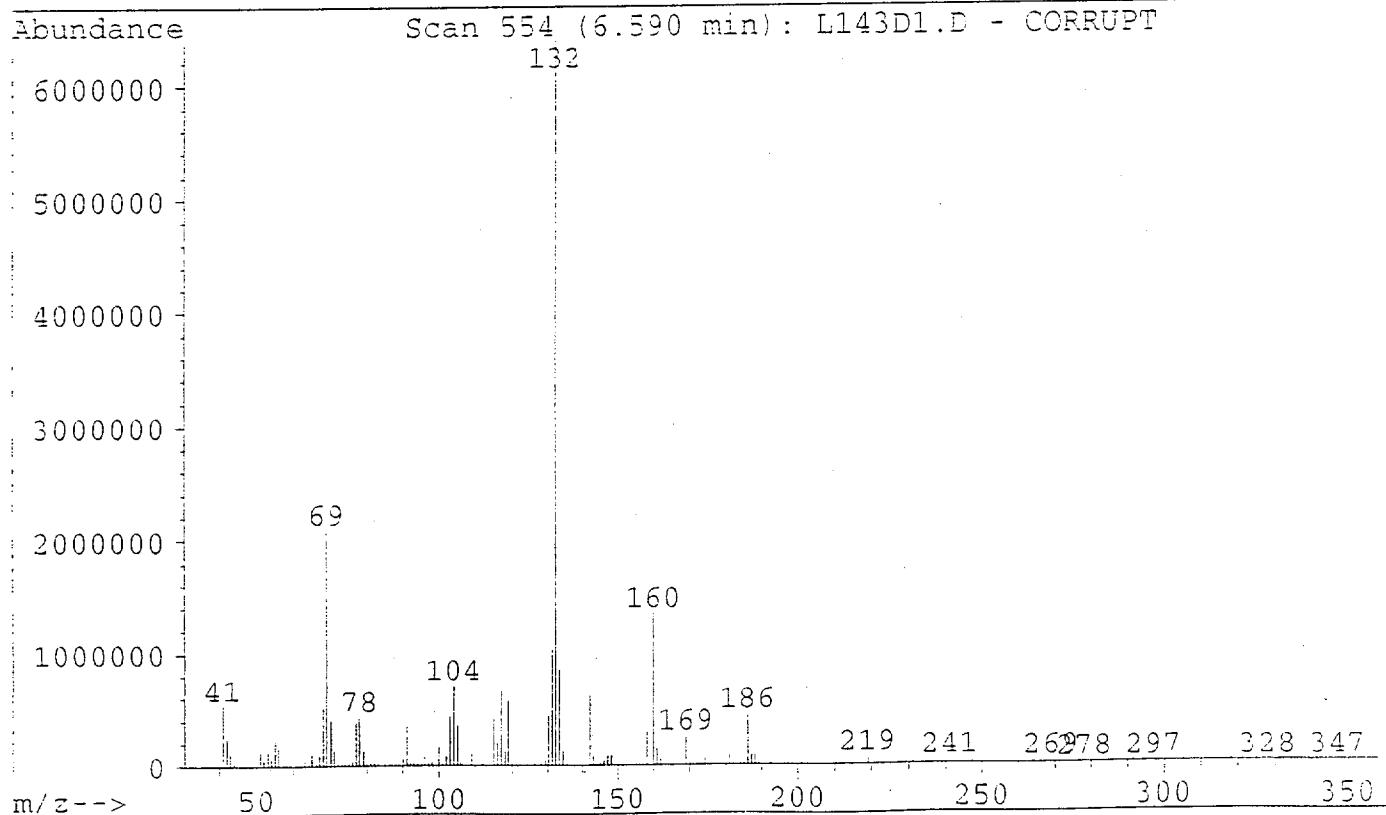
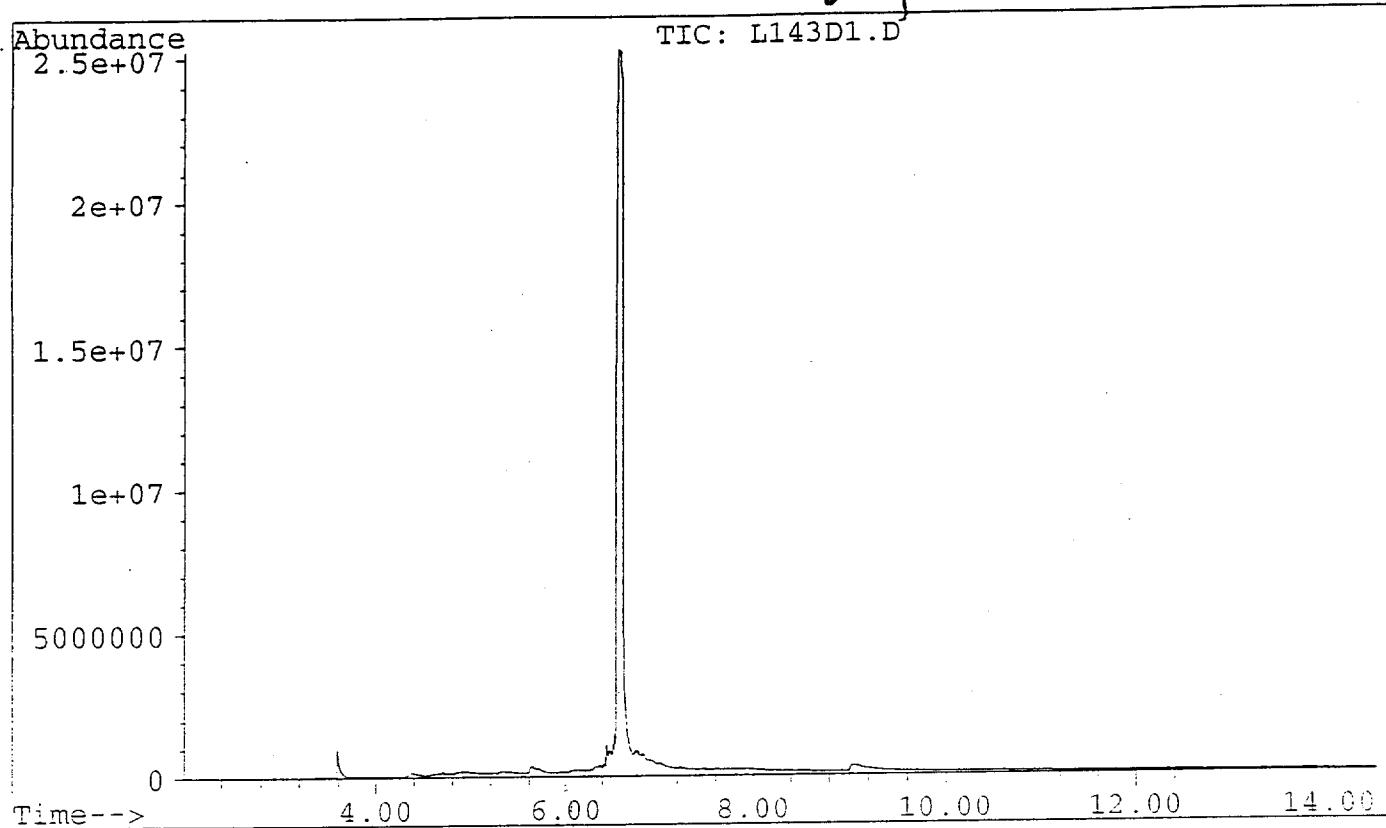
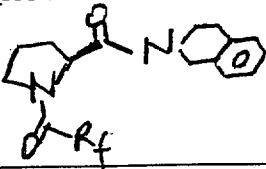
Sample Name: 4 x 4 Library of amides C4

Misc Info :

Vial Number: 1



File : C:\HPCHEM\1\DATA\L143D1.D
 Operator :
 Acquired : 28 Jan 99 7:59 pm using AcqMethod ZYLUO
 Instrument : 5970B - I
 Sample Name: 4 x 4 Library of amides D1
 Misc Info :
 Vial Number: 1



File : C:\HPCHEM\1\DATA\L143D2.D

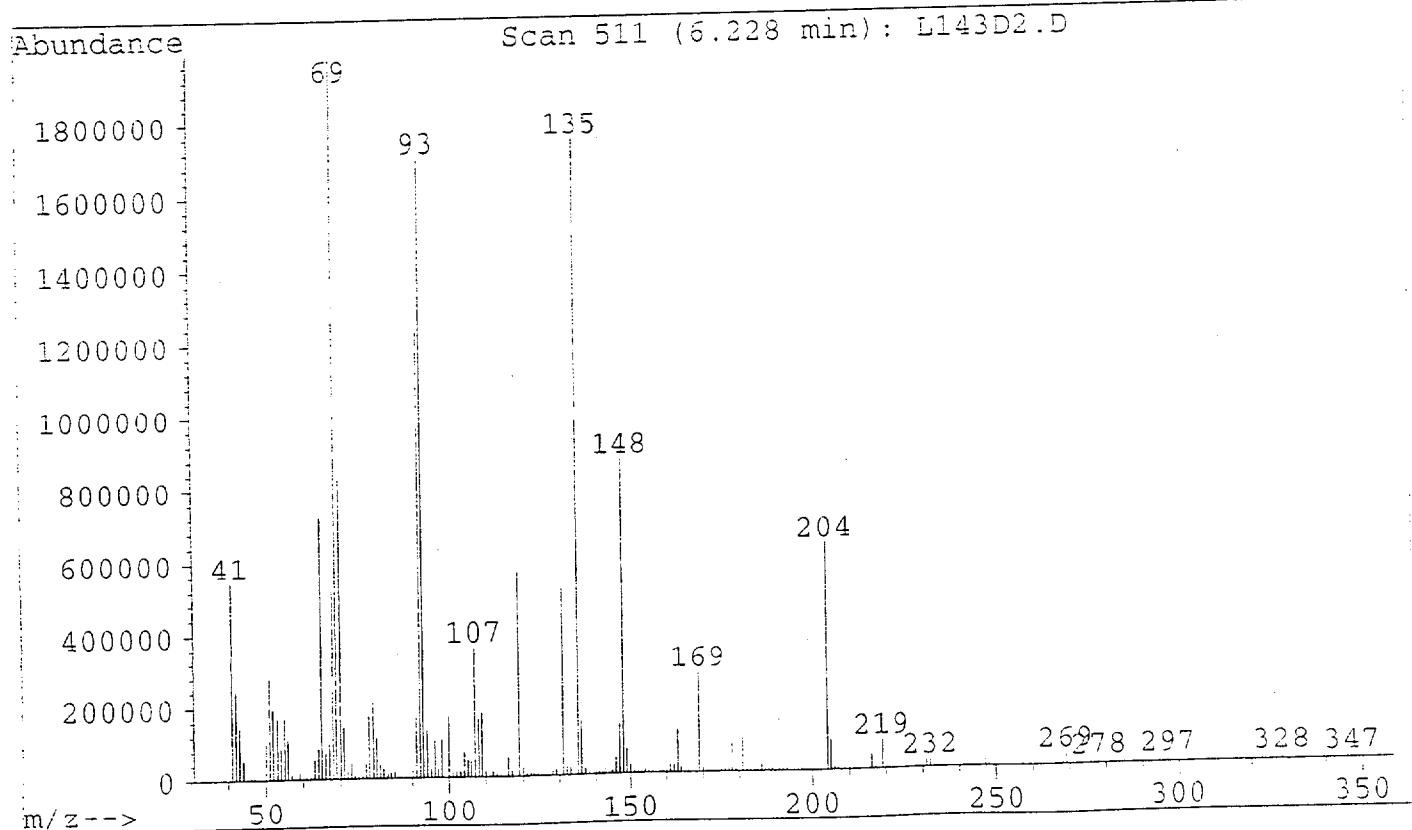
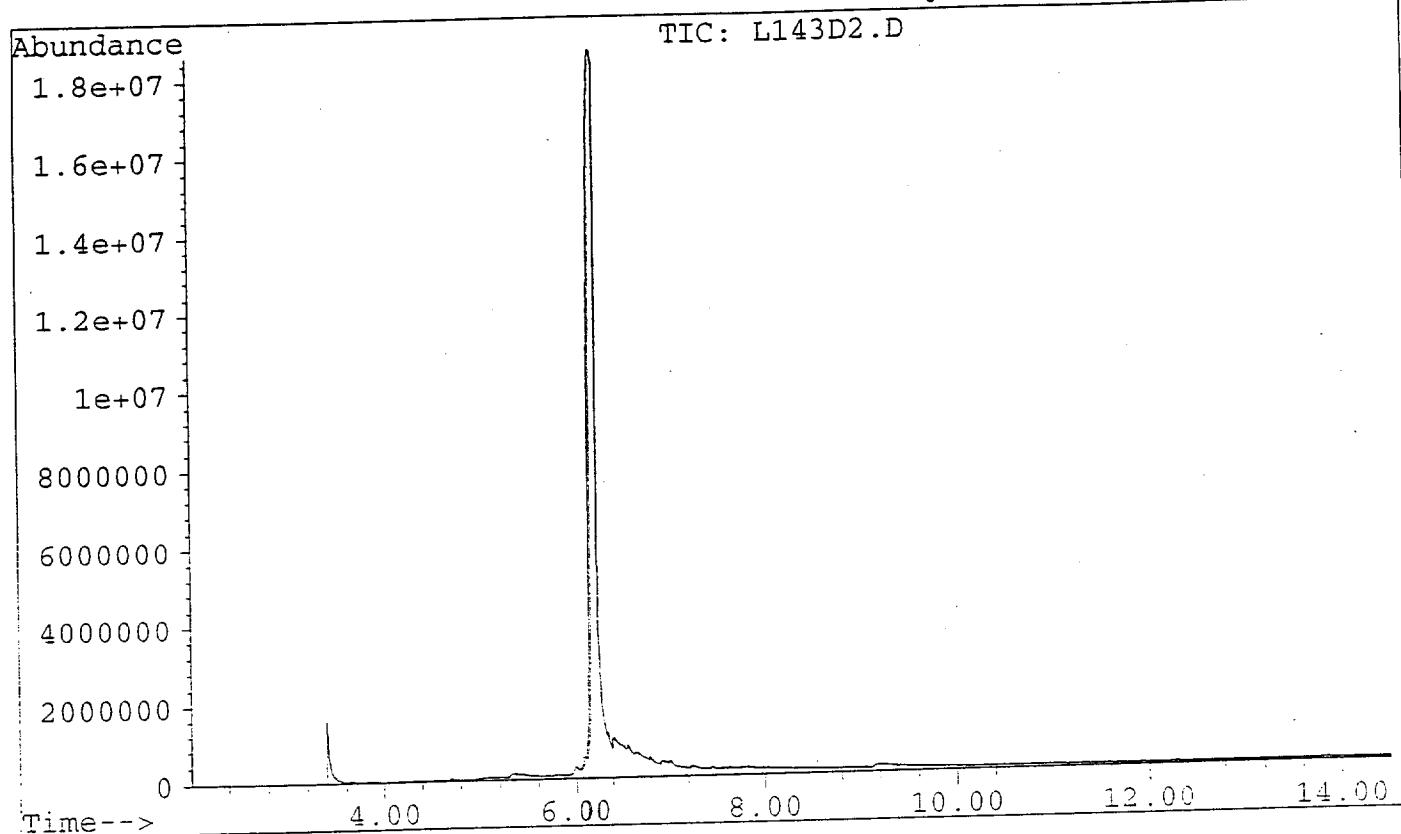
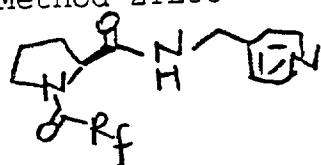
Operator :
Acquired : 28 Jan 99 8:21 pm using AcqMethod ZYLUO

Instrument : 5970B - I

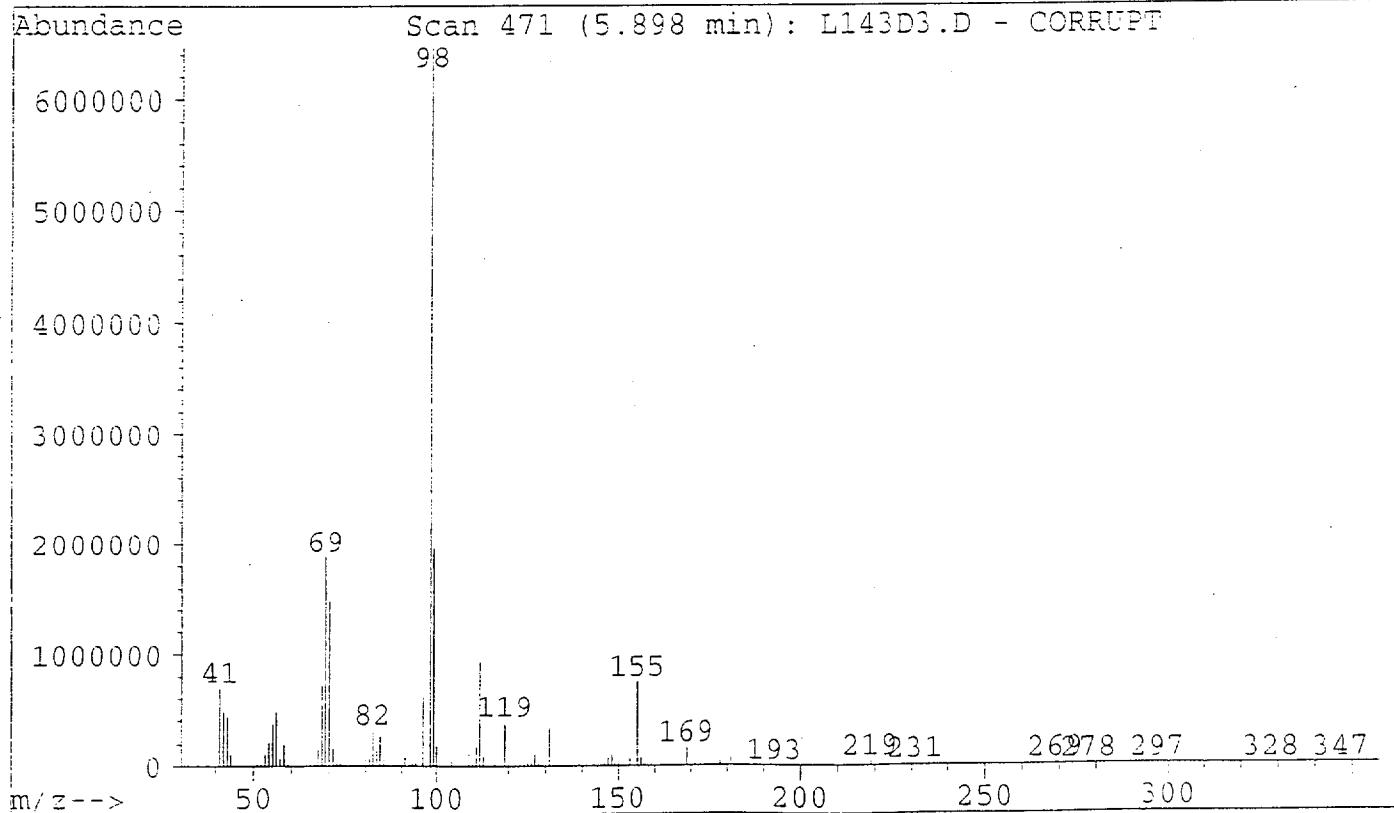
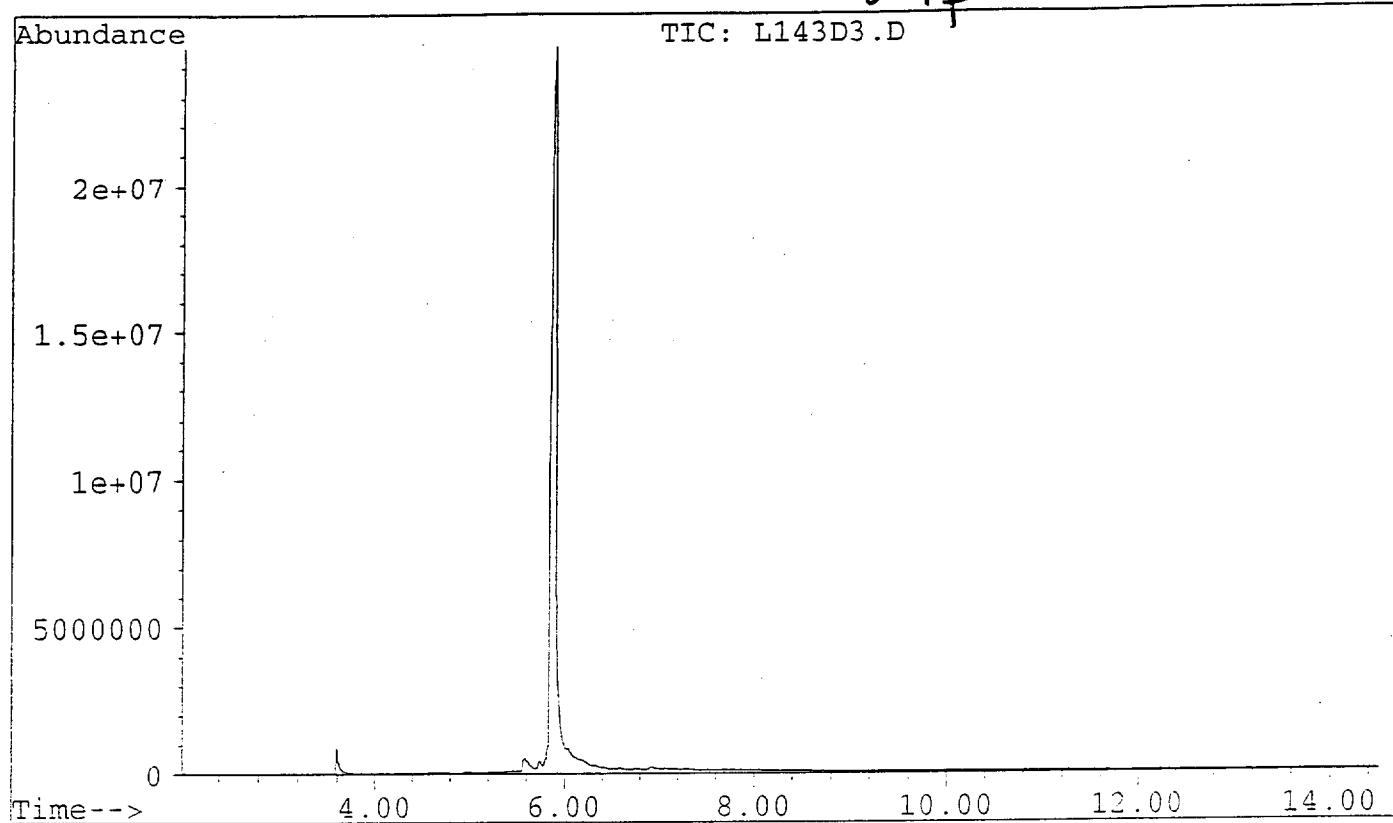
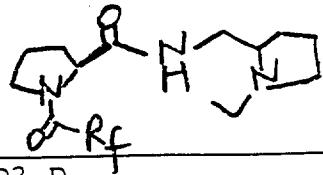
Sample Name: 4 x 4 Library of amides D2

Misc Info :

Vial Number: 1



File : C:\HPChem\1\DATA\L143D3.D
 Operator :
 Acquired : 28 Jan 99 8:40 pm using AcqMethod ZYLUO
 Instrument : 5970B - I
 Sample Name: 4 x 4 Library of amides D3
 Misc Info :
 Vial Number: 1



File : C:\HPCHEM\1\DATA\L143D4.D

Operator :

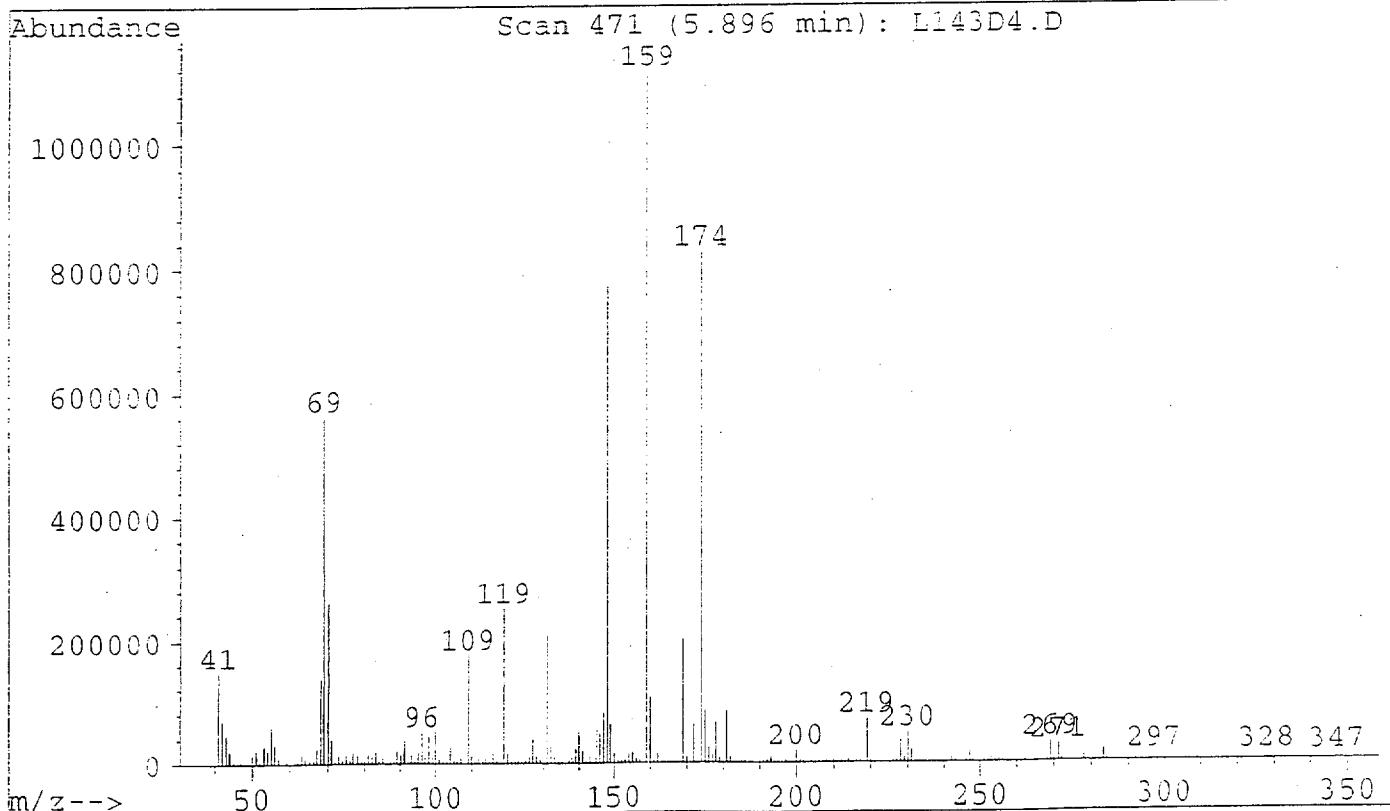
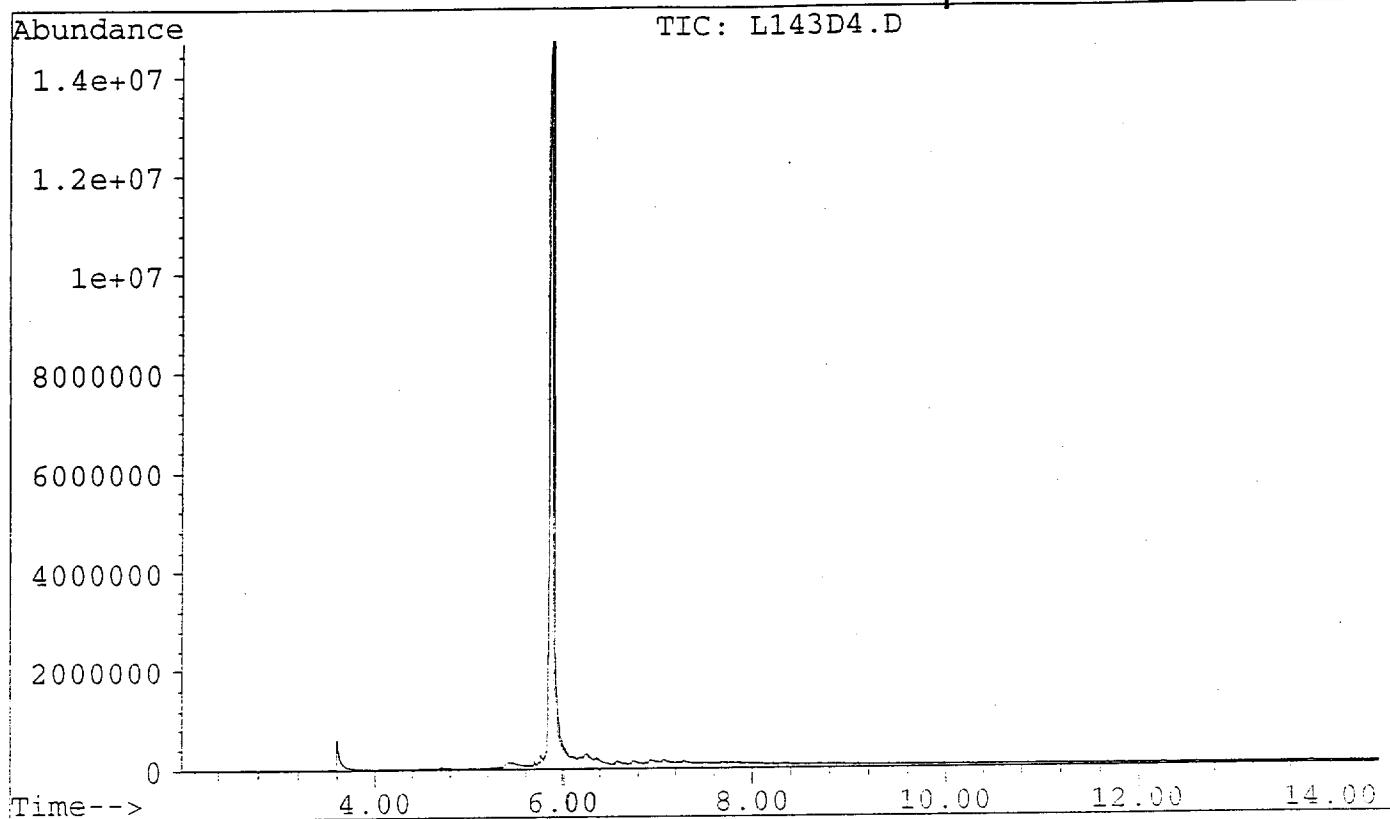
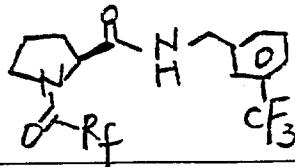
Acquired : 28 Jan 99 8:58 pm using AcqMethod ZYLUO

Instrument : 5970B - I

Sample Name: 4 x 4 Library of amides D4

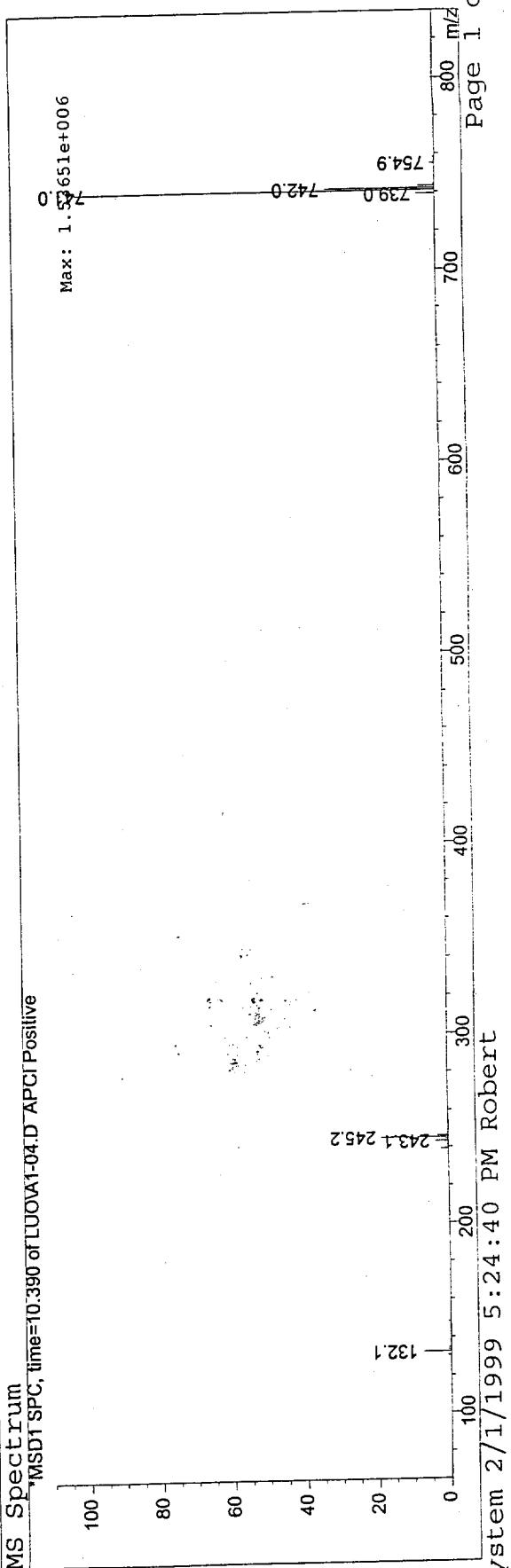
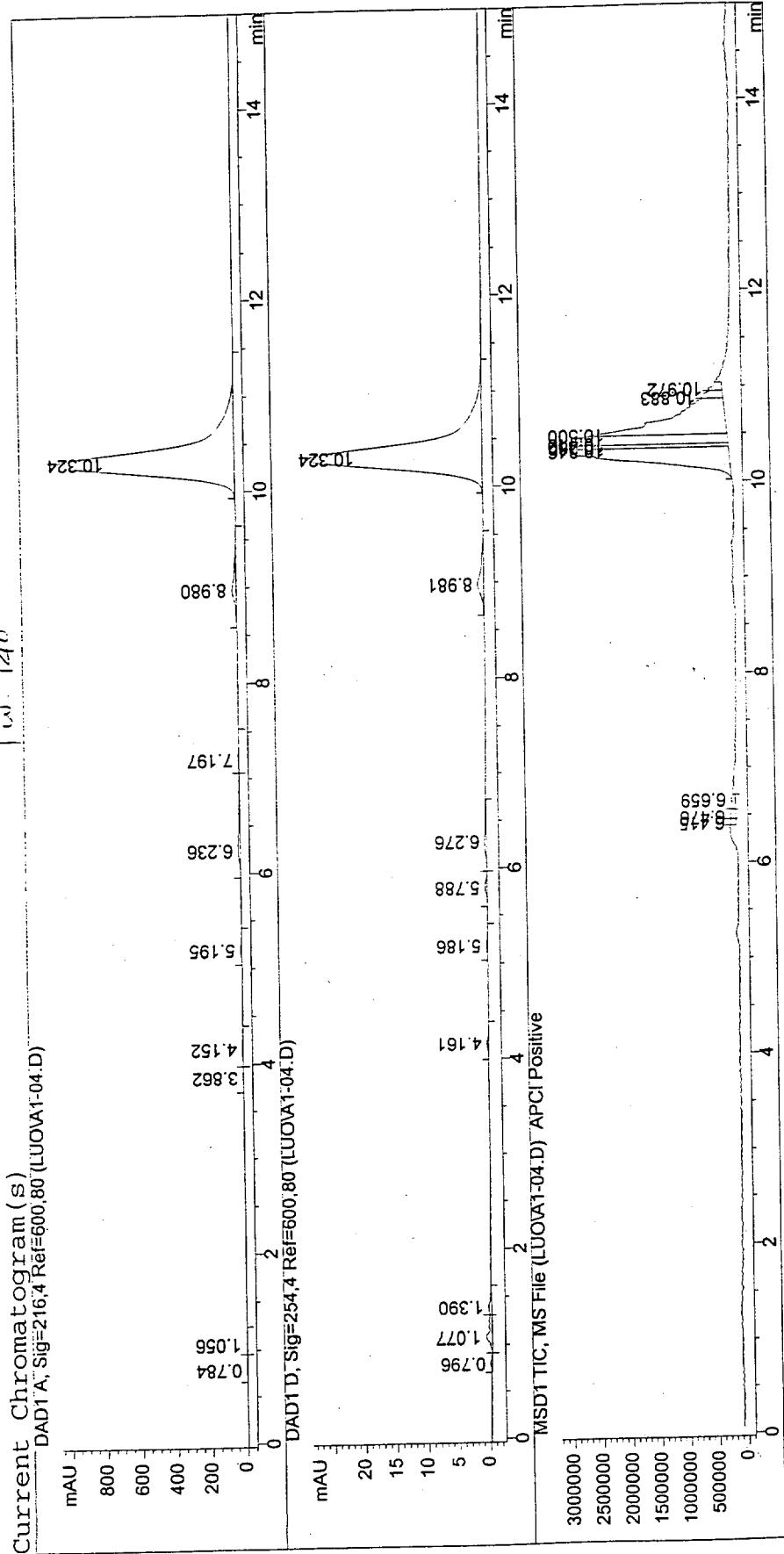
Misc Info :

Vial Number: 1



Print or all graphic windows

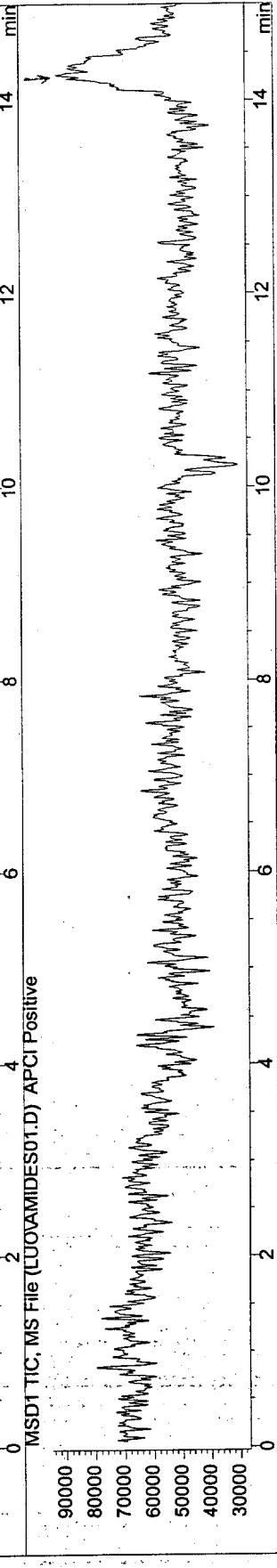
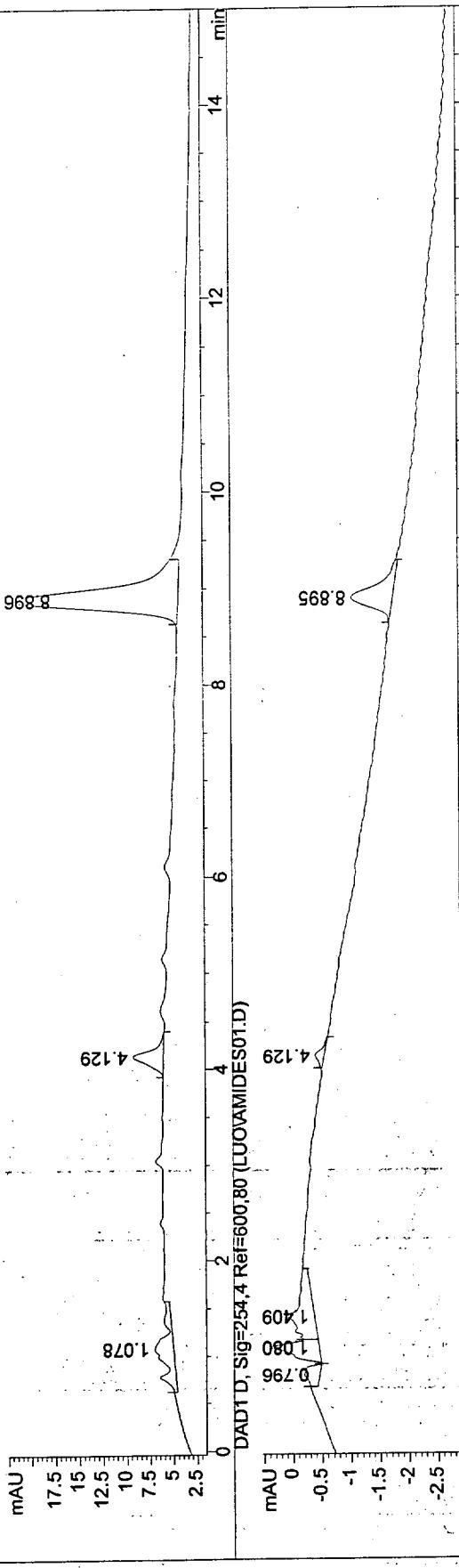
All Ca²⁺-Zn



Print of all graphic windows A2 : CgFig_115.msp Fw: 115

Current Chromatogram (s)

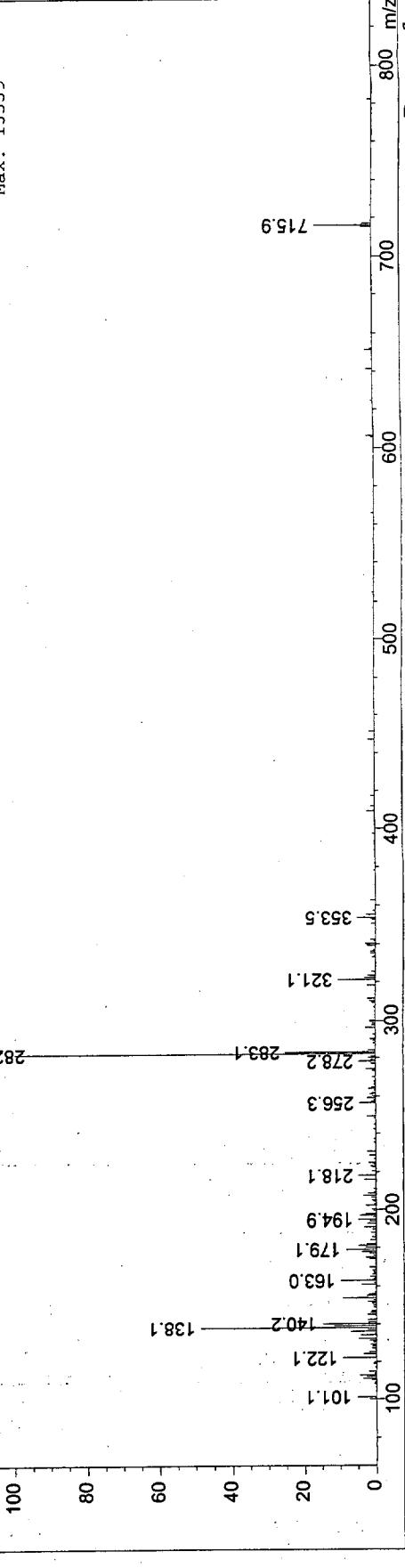
DAD1A, Sig=216.4 Ref=600.80 (LUOAMIDES01.D)



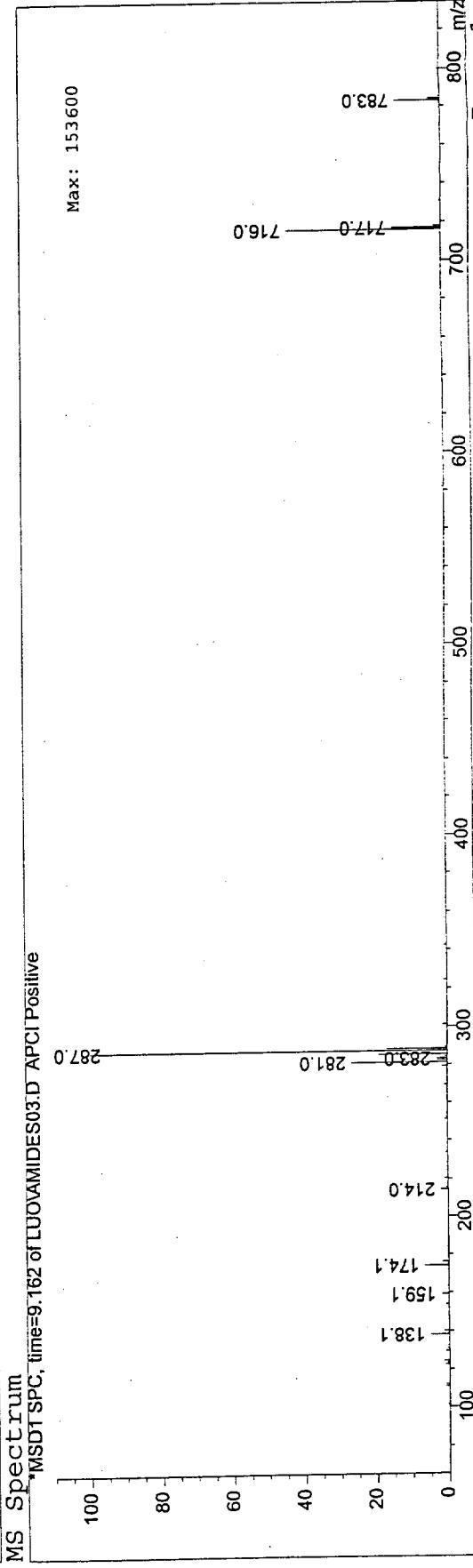
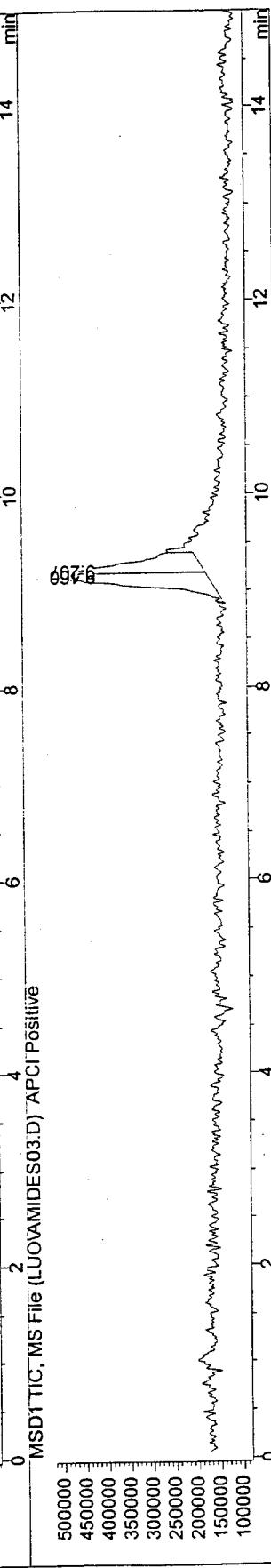
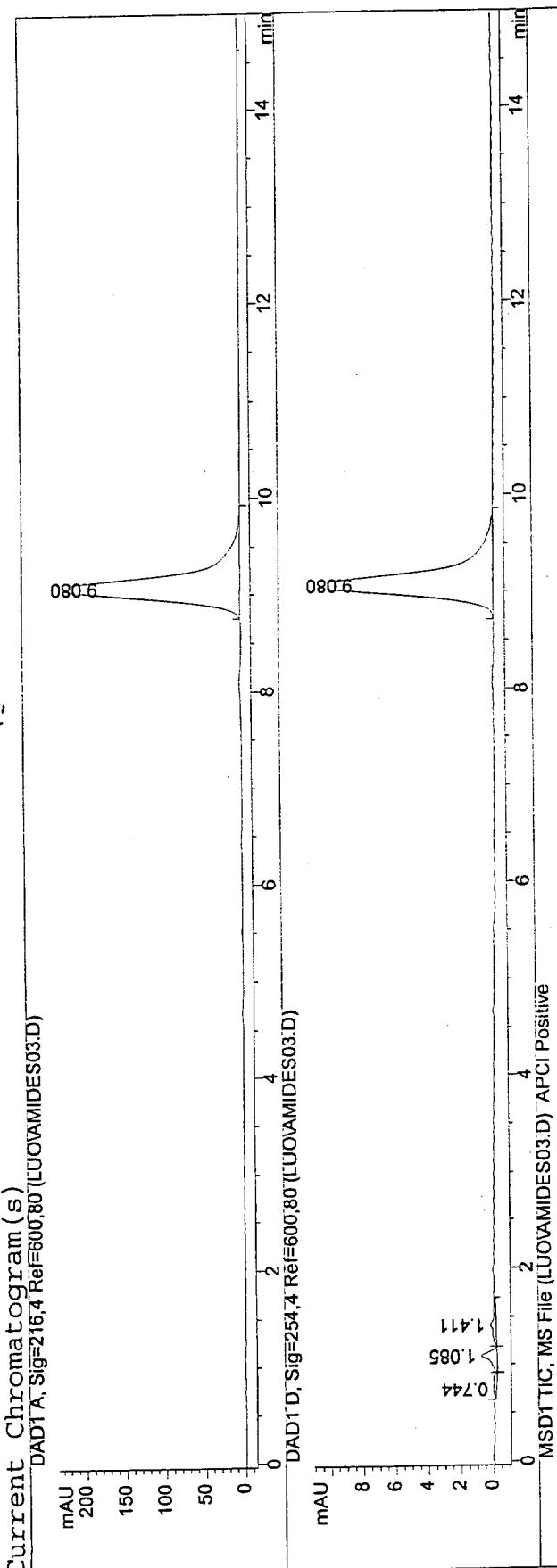
MS Spectrum

MSD1 SPC, time=14.284 of LUOAMIDES01.D APCI Positive

Max: 15359

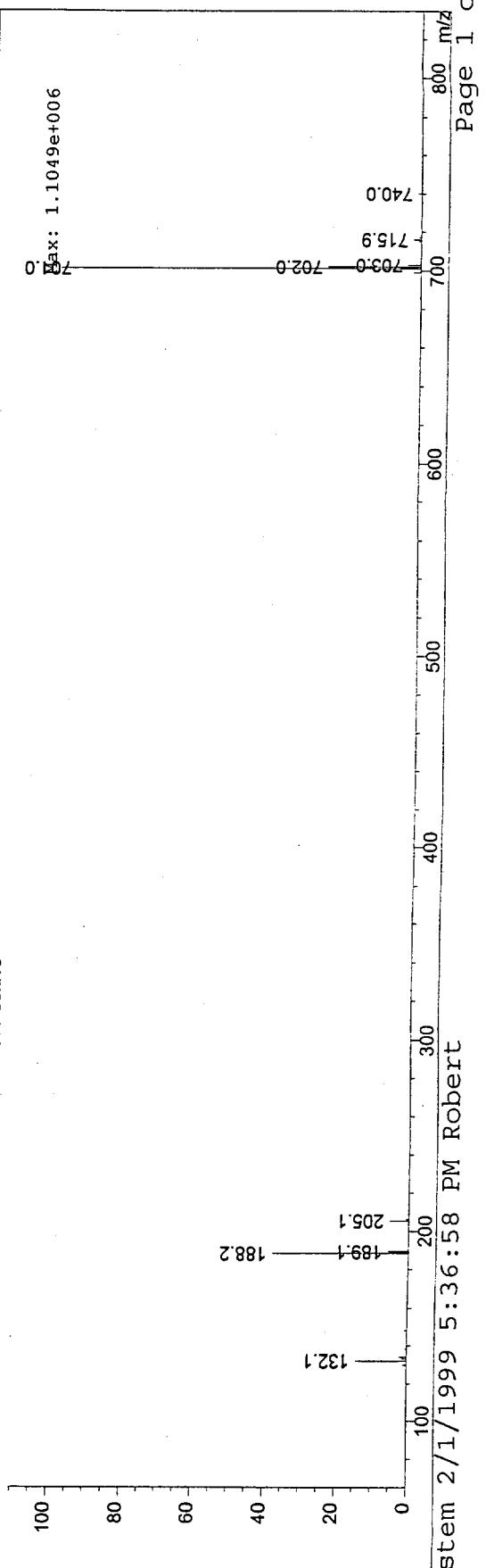
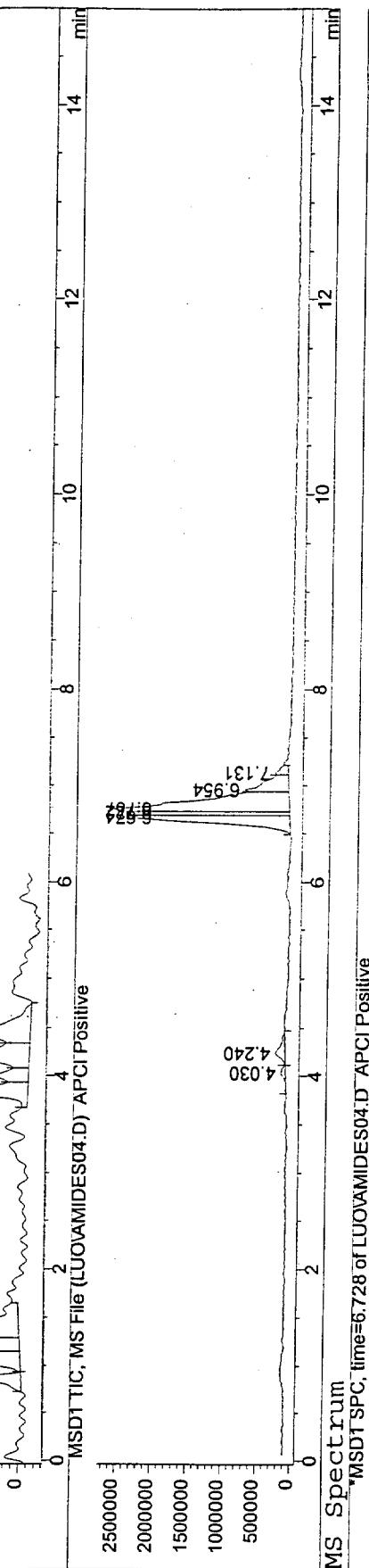
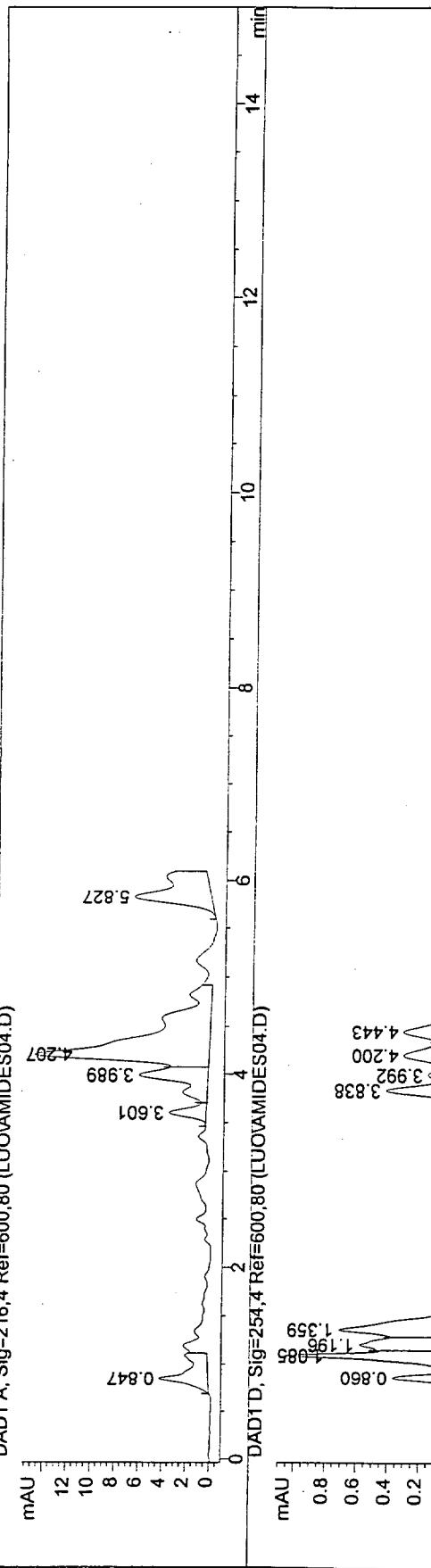


Print or all graphic windows A4 :   

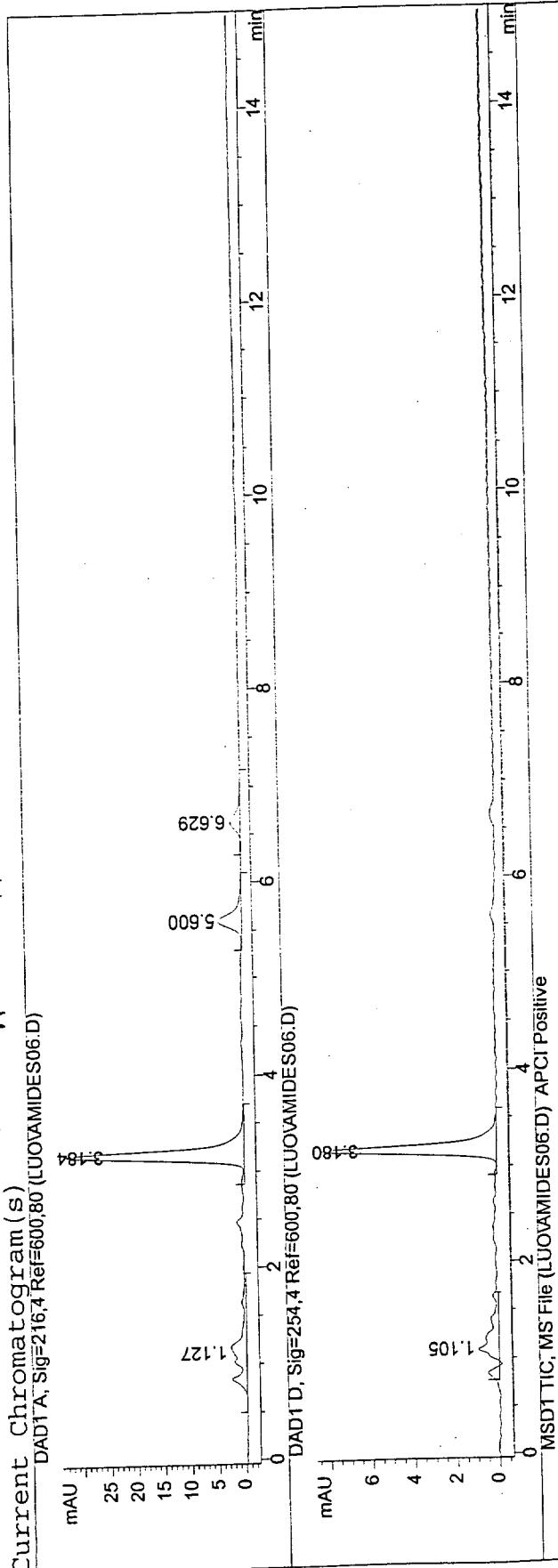


- 537

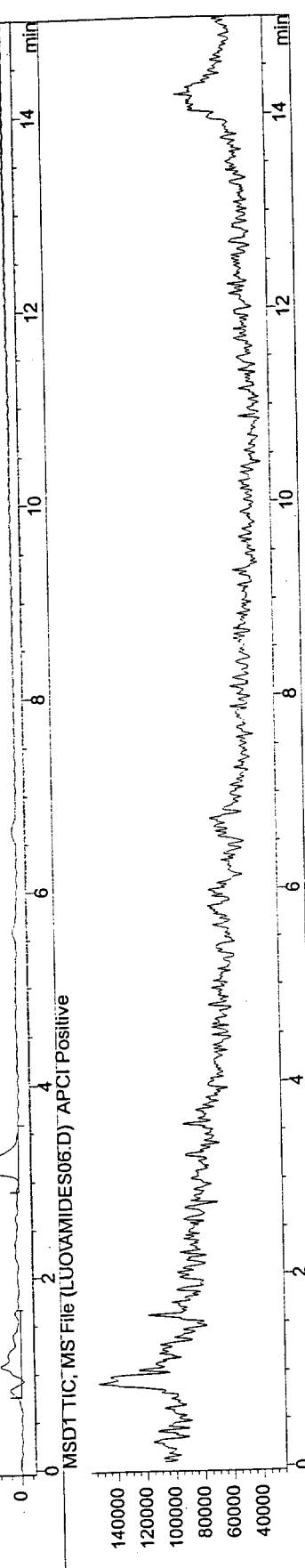
Print or all graphic windows [x]

Current Chromatogram (S)
DAD1A, Sig=216.4 Ref=600.80 (LUOAMIDES04.D)

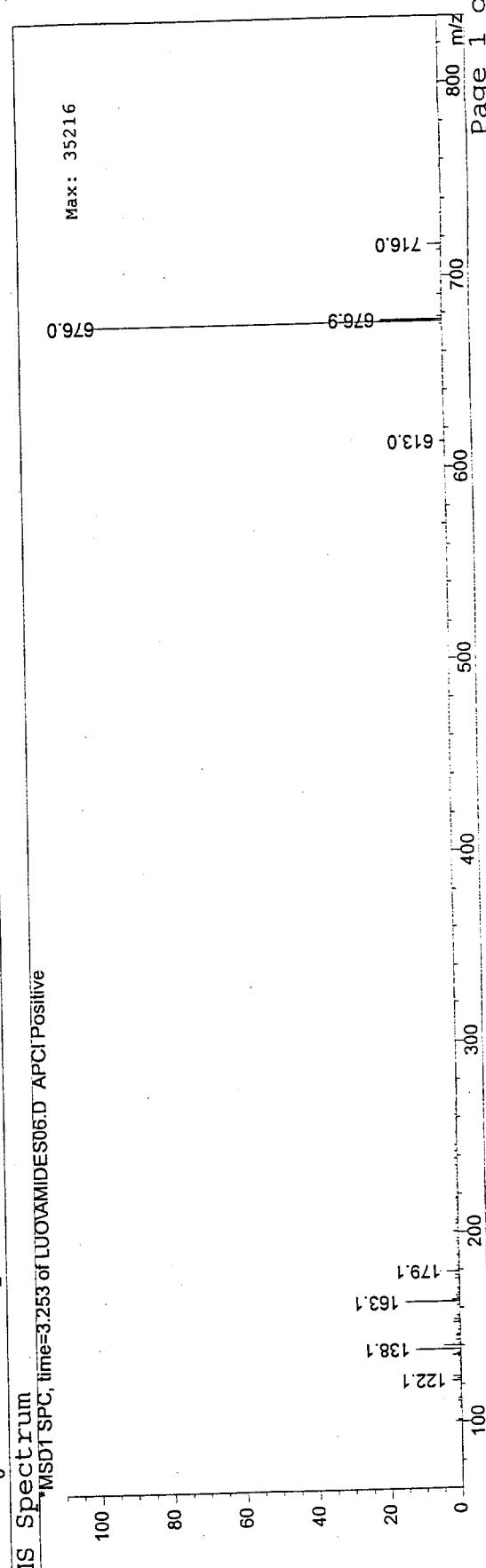
Print of all graphic windows (S)
DAD1A, Sig=216.4 Ref=600.80 (LUOVAMIDES06.D)



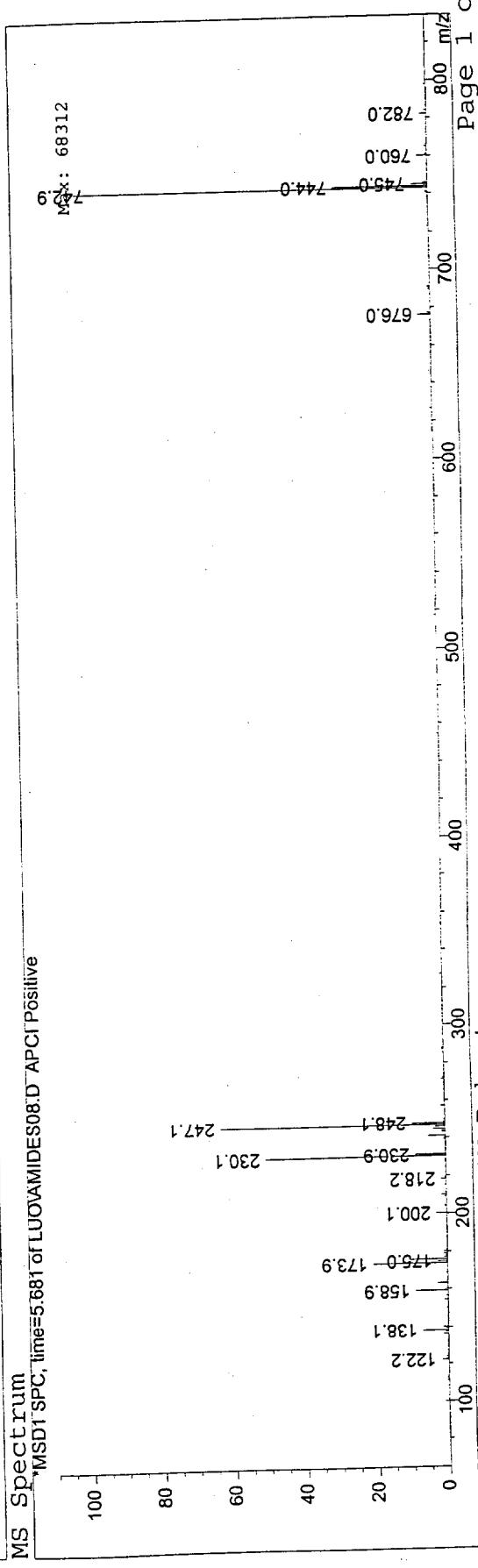
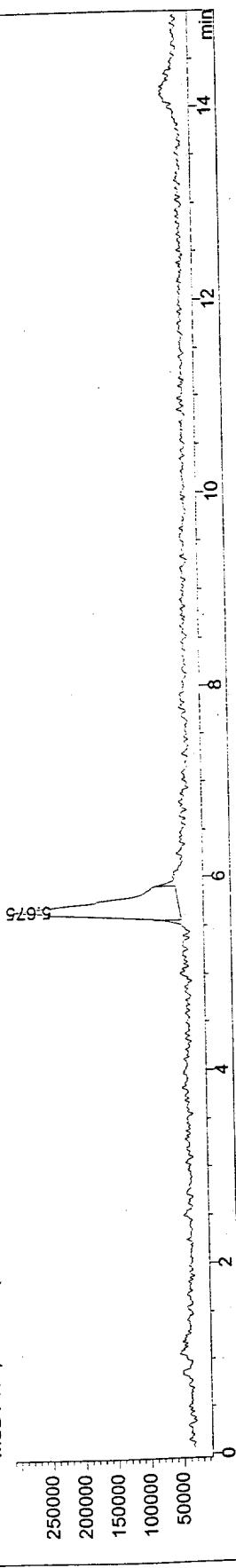
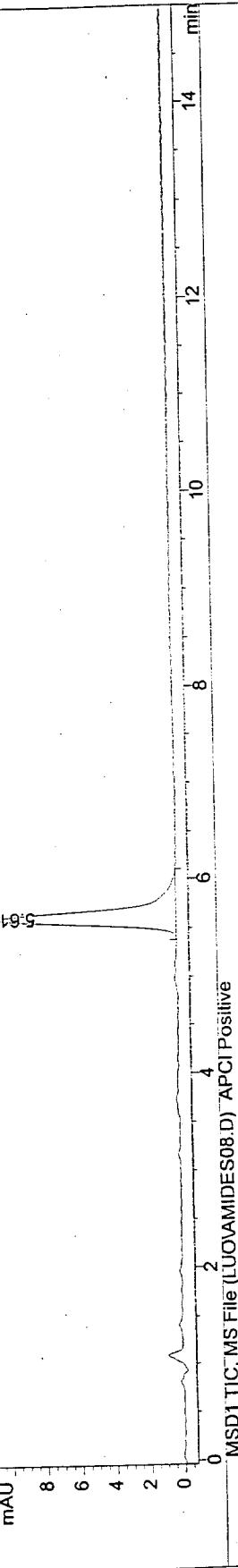
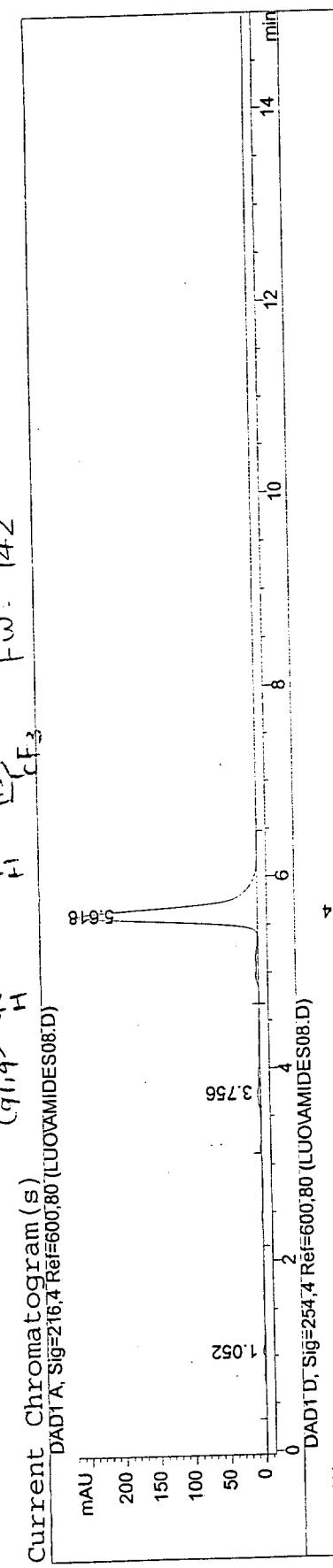
MSD1TIC, MS File (LUOVAMIDES06.D) APCI Positive



MS Spectrum
MSD1TSPC, time=3.253 of LUOVAMIDES06.D APCI Positive

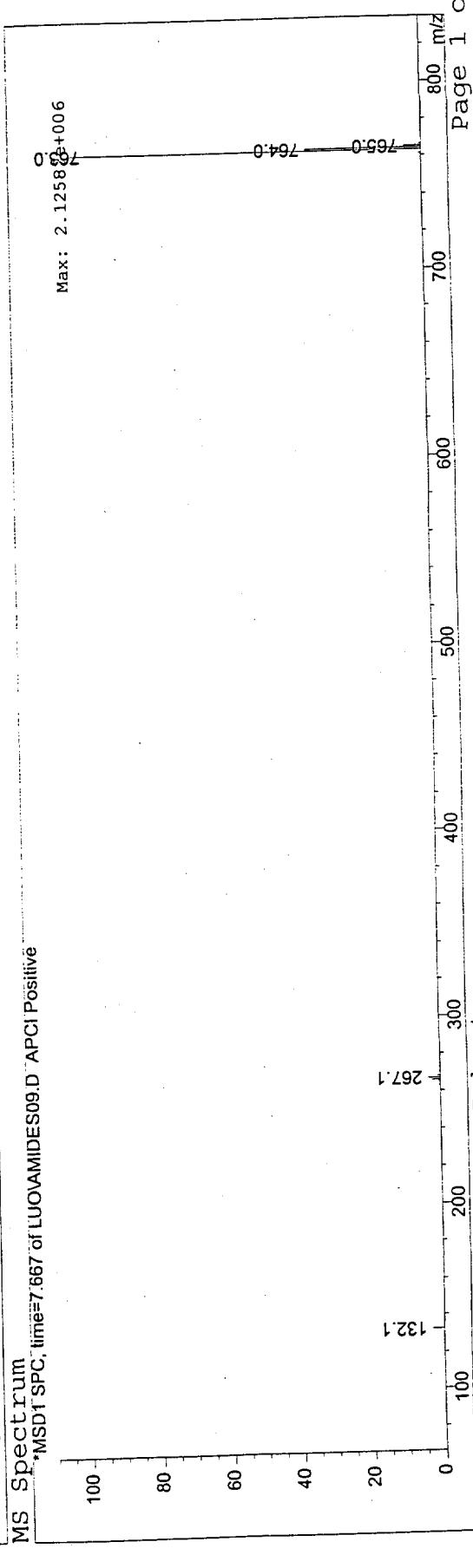
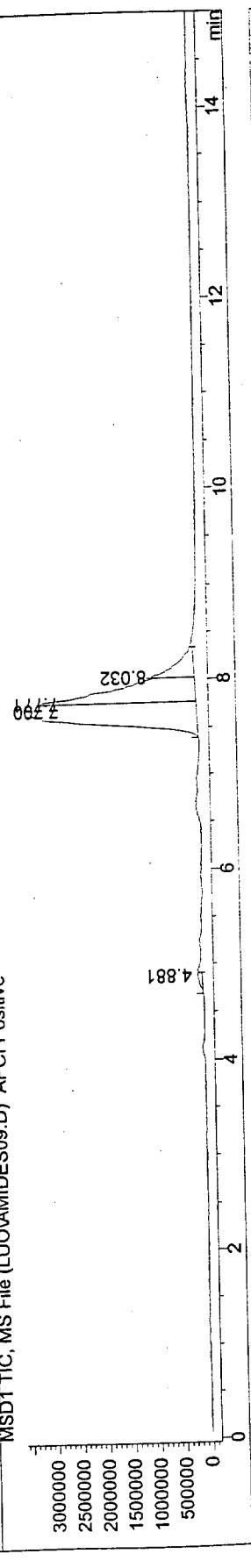
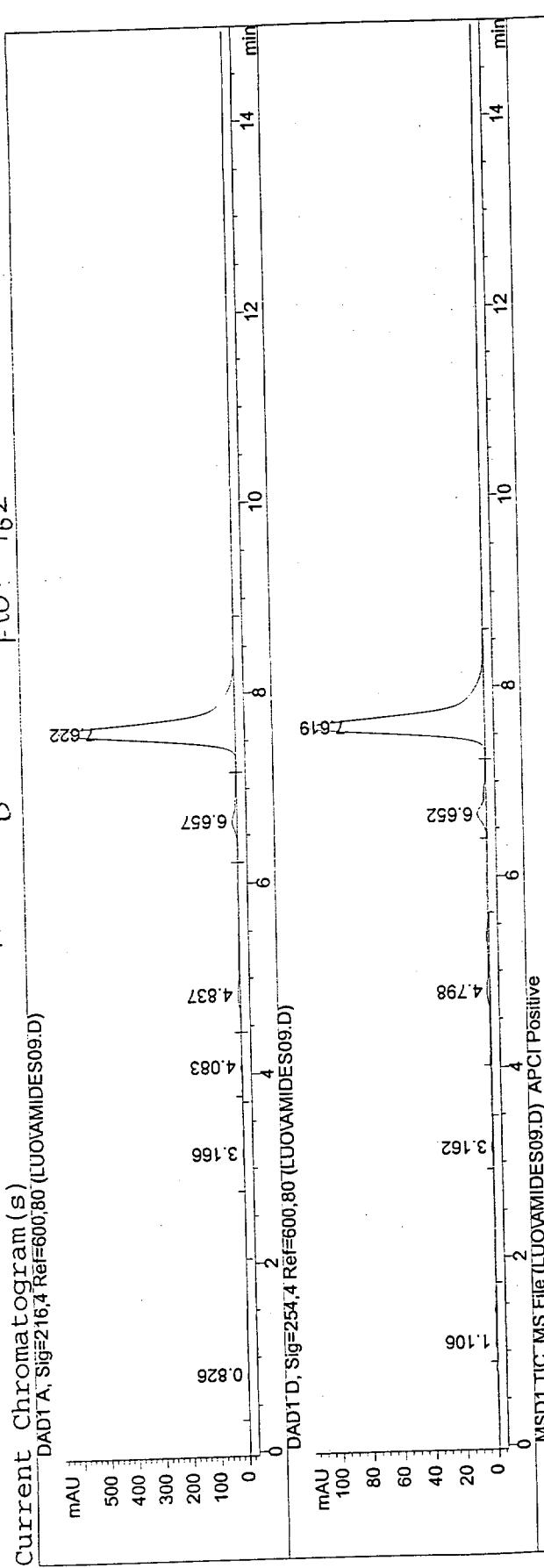


Print of all graphic windows

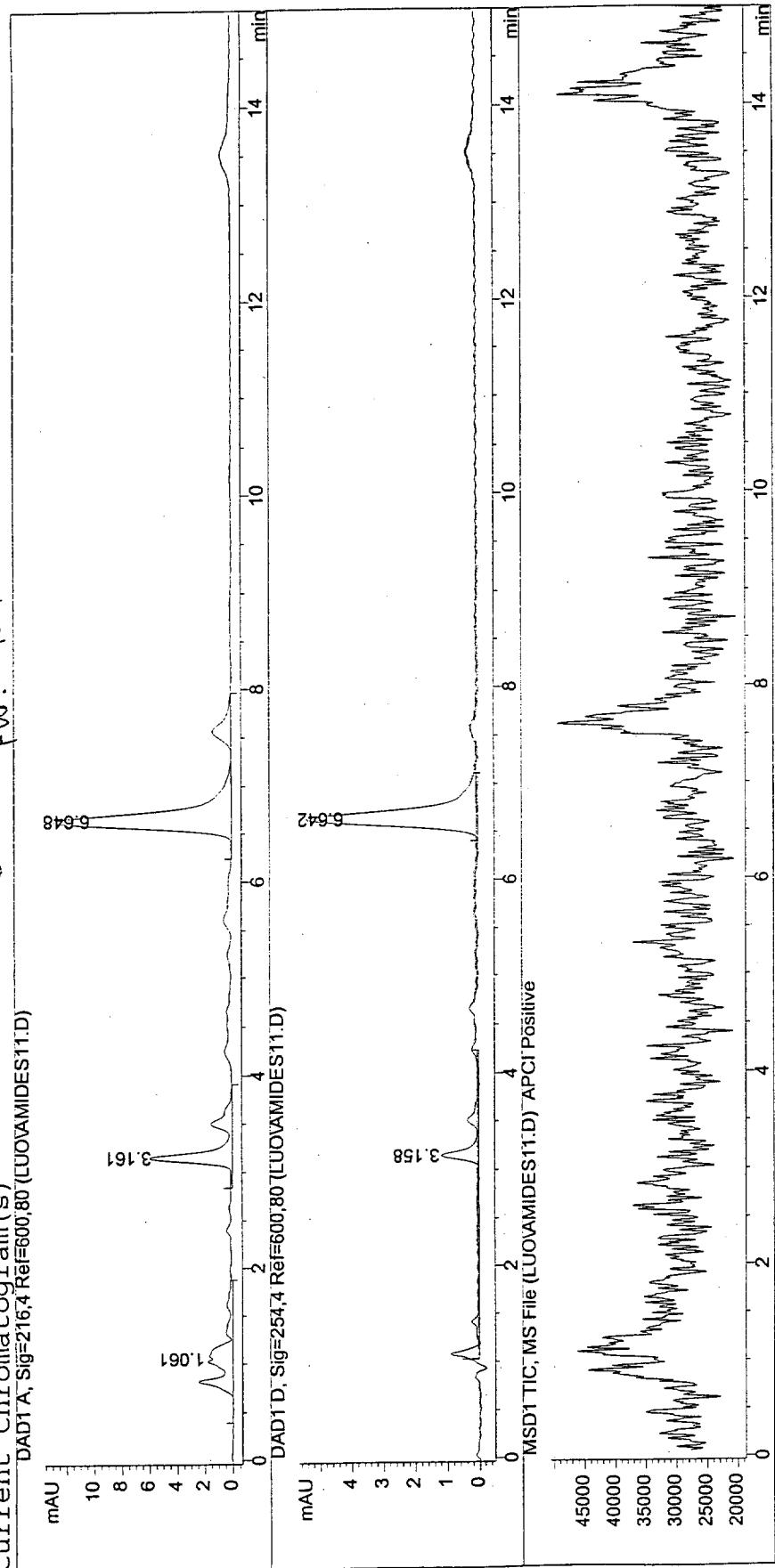


Print of all graphic windows

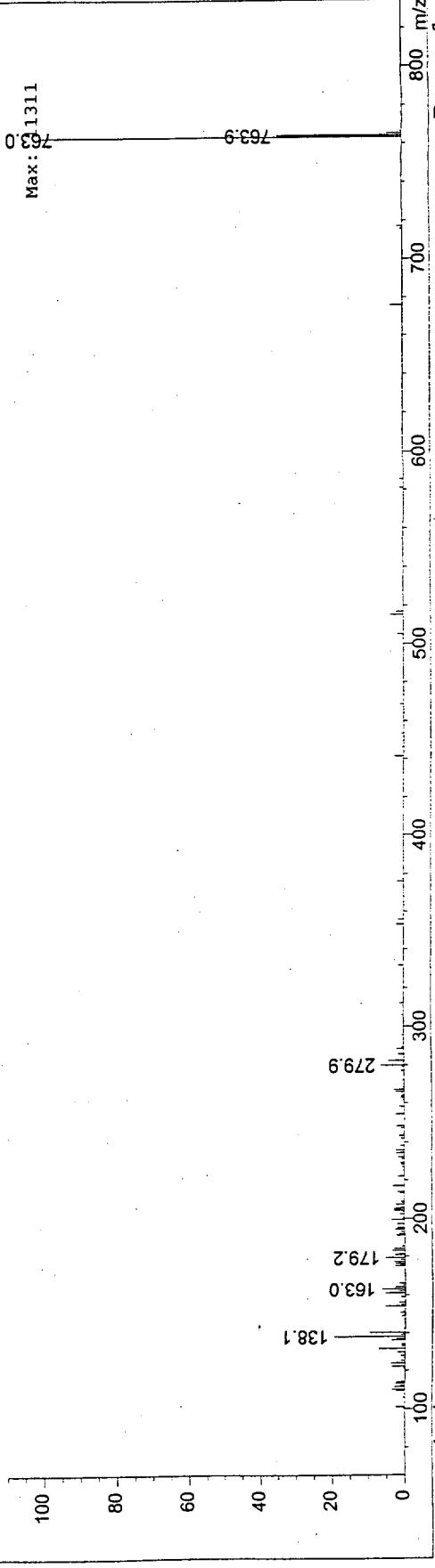
Current Chromatogram (S)
DAD1A, Sig=251.4 Ref=600.80 (LUOVAMIDES09.D)



Print of all graphic windows (2) C9Fig
 Current Chromatogram (S)
 DAD1A, Sig=216.4, Ref=600,80 (LUOAMIDES11.D)

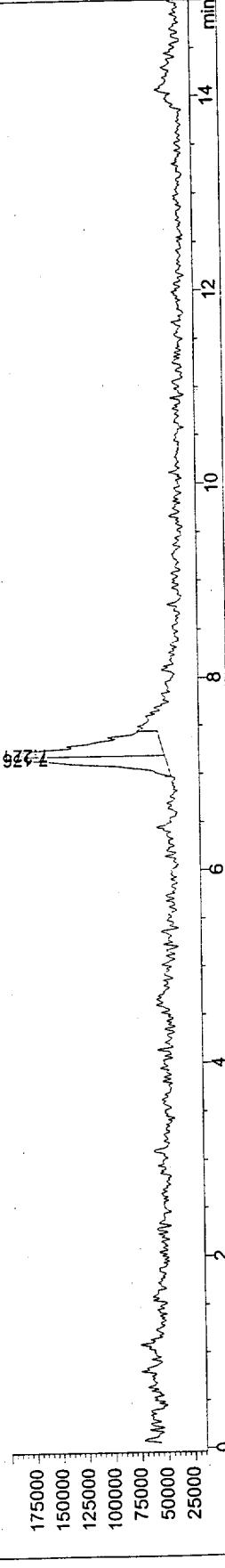
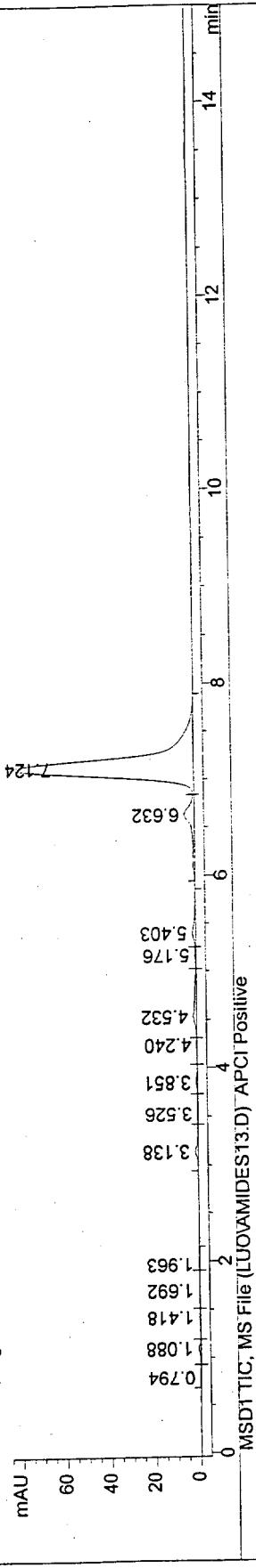
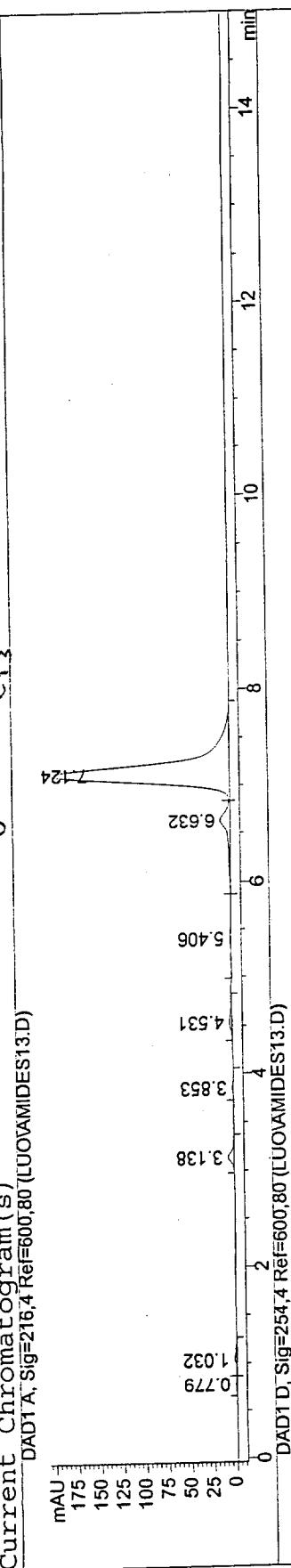


MS Spectrum, MSD1 SPC, time=7.610 of LUOAMIDES11.D APCI Positive



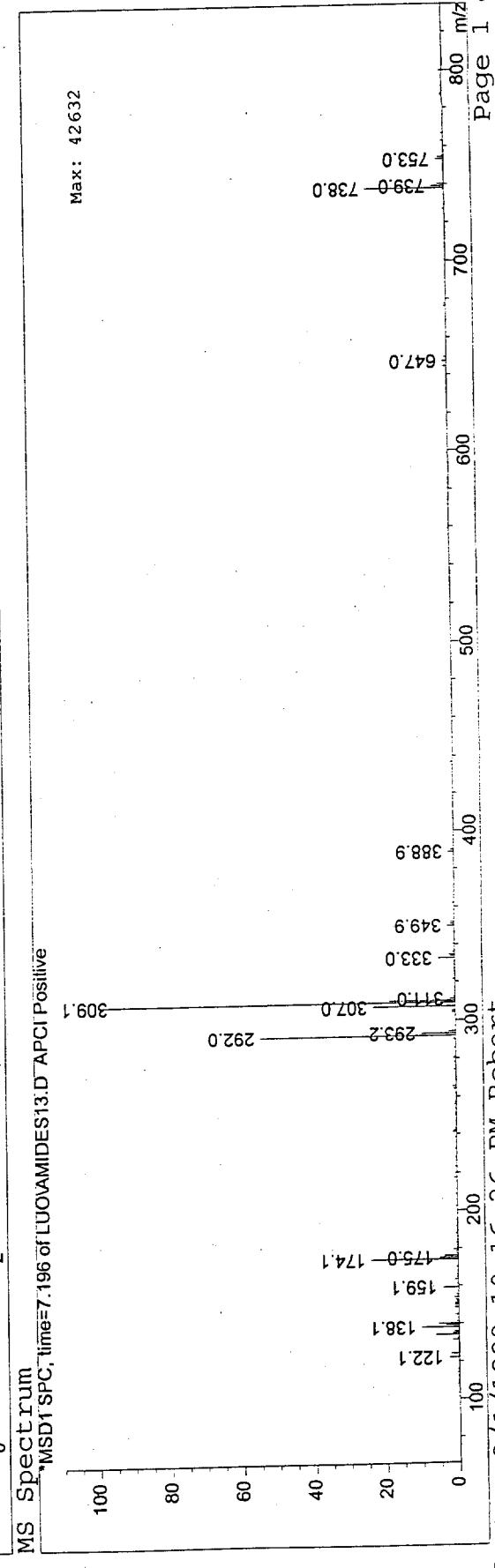
Print or all graphic windows

Print of all graphic windows

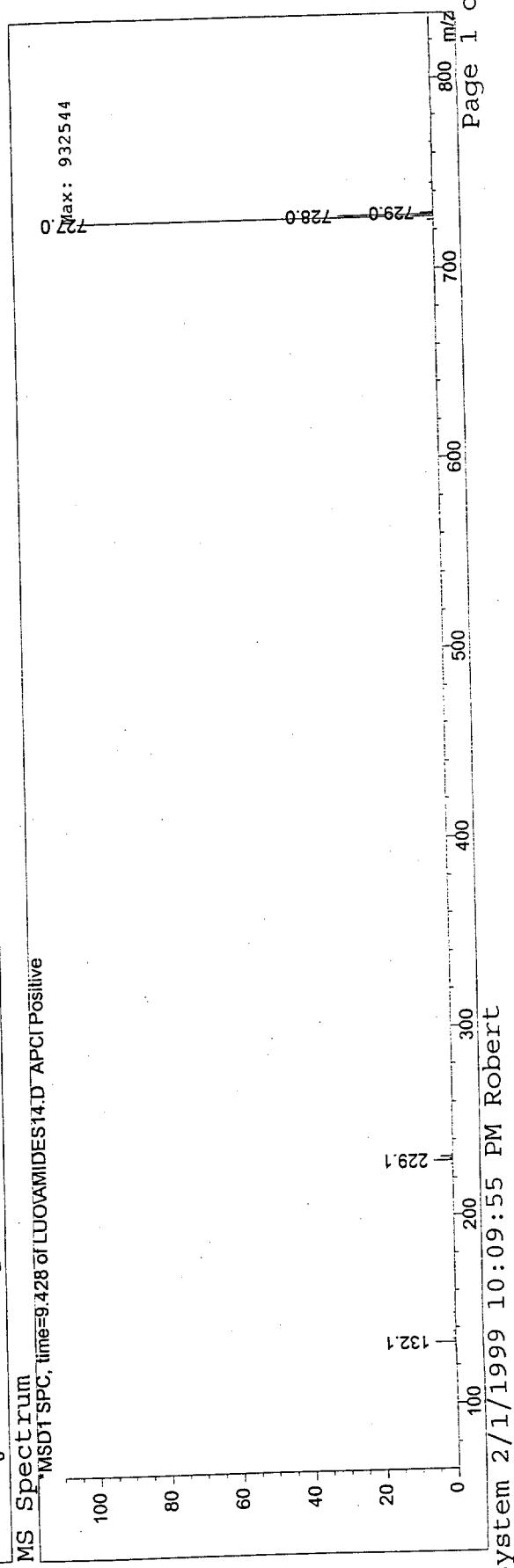
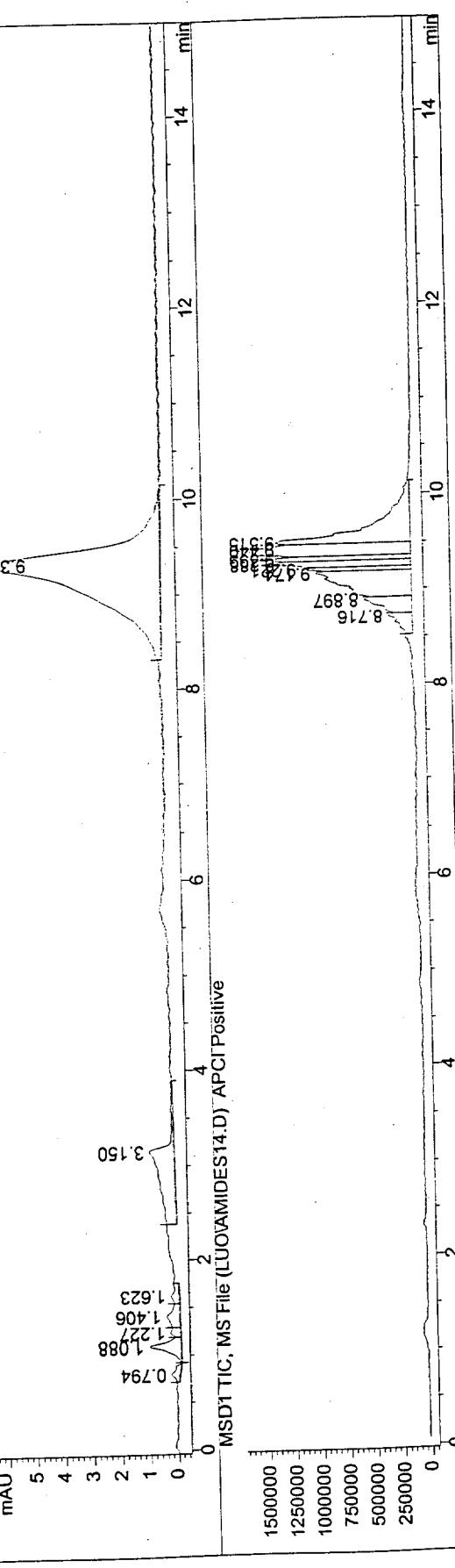
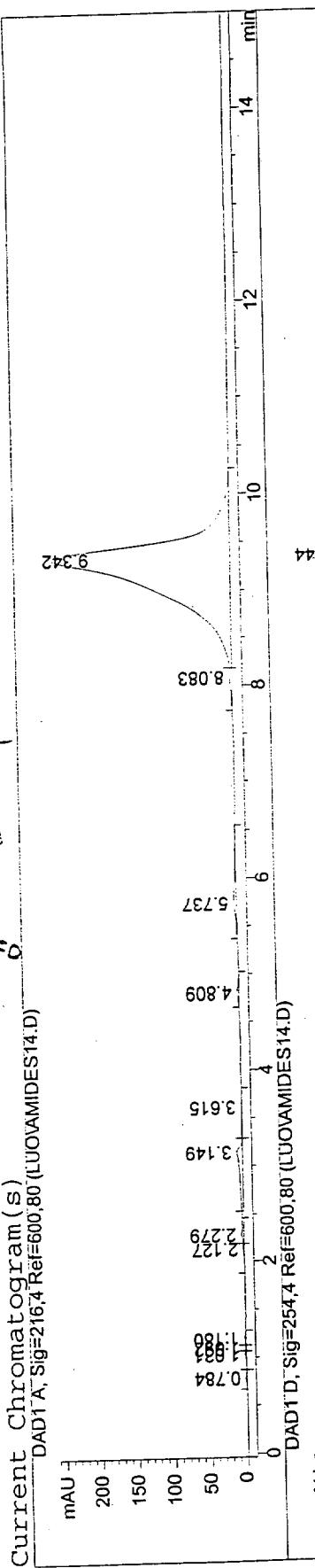


MS Spectrum *MSP1 SPC Time=7.196 of LUQAMIDES13.D APCI Positive

Max: 42632



Print or all graphic windows



Print of all graphic windows
D4Current Chromatogram(s)
DAD1A, sig=216.4 Ref=600.80 (LUOAMIDES18:D)