

Supporting information

Core-Shell-Shell NaYbF₄:Tm@CaF₂@NaDyF₄ Nanocomposites for Upconversion/T₂-weighted MRI/ Computed Tomography Lymphatic Imaging

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Synthesis of NaDyF₄:Yb,Tm nanoparticles capped with oleic acid (OA-NaDyF₄:Yb,Tm). 0.327 mmol Yb(TFA)₃, 3 μ mol Tm(TFA)₃, 0.67 mmol Dy(TFA)₃ and 1 mmol NaTFA were added to a 100 mL flask containing 2.84 g oleic acid (OA), 2.67 g oleylamine (OM) and 5.04 g 1-octadecene (ODE). The resulting mixture was heated to 110 °C with magnetic stirring to form a transparent solution. The flask was sealed and degassed for 30 min with N₂ gas. The solution was slowly heated to 300 °C and maintained for 30 min under the nitrogen environment. Then, the solution was cooled down to room temperature. The NaDyF₄:Yb,Tm nanoparticles were collected by centrifugation at 15000 rpm for 10 min and washed several times with ethanol/cyclohexane, after that dispersed in cyclohexane.

Synthesis of core-shell NaYbF₄:Tm@NaDyF₄ nanoparticles capped with oleic acid (OA-NaYbF₄:Tm@NaDyF₄). The core-shell NaYbF₄:Tm@NaDyF₄ nanoparticles were synthesized similarly to that of NaYbF₄:Tm@CaF₂. 5 mL as-prepared solution of NaYbF₄:Tm nanoparticles dispersed in cyclohexane, and 0.5 mmol Dy(TFA)₃ and 0.5 mmol NaTFA were add in a 100 mL flask containing 2.84 g oleic acid (OA), 2.68 g oleylamine (OM) and 5.04 g 1-octadecene (ODE). The mixture was then heated to 110 °C for 30 min to remove the cyclohexane. After the cyclohexane was evaporated, the flask was sealed and degassed for 30 min. Subsequently, the reaction was slowly heated to 300 °C under the nitrogen environment and kept at it for 8 min, then cooling down to room temperature. The NaYbF₄:Tm@CaF₂ nanoparticles were collected by centrifugation at 15000 rpm and washed several times with ethanol/cyclohexane, then dispersed in cyclohexane.

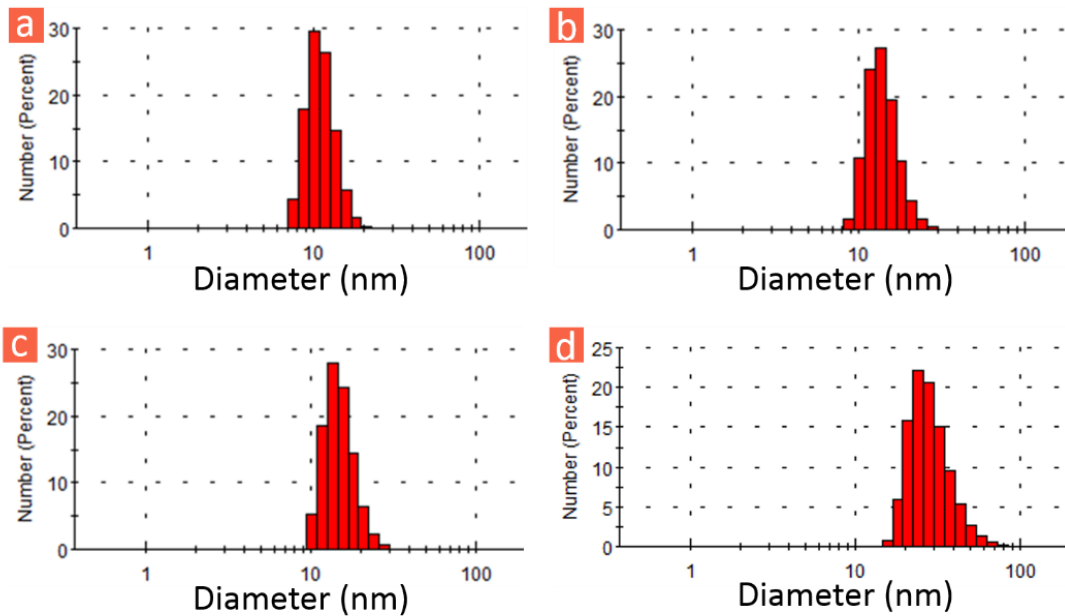


Figure S1. DLS data of these nanoparticles OA-NaYbF₄:Tm (a), OA-NaYbF₄:Tm@CaF₂ (b), OA-NaYbF₄:Tm@CaF₂@NaDyF₄ and Cit-NaYbF₄:Tm@CaF₂@NaDyF₄ (d).

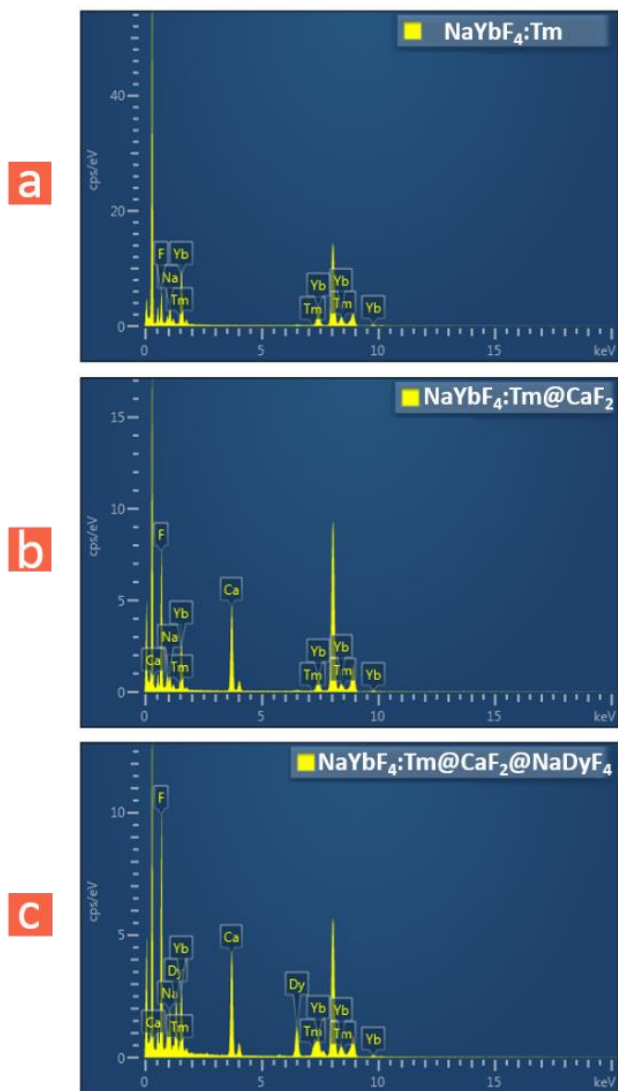


Figure S2. EDXA pattern of these nanoparticles OA-NaYbF₄:Tm (a), OA-NaYbF₄:Tm@CaF₂ (b), and OA-NaYbF₄:Tm@CaF₂@NaDyF₄ (c).

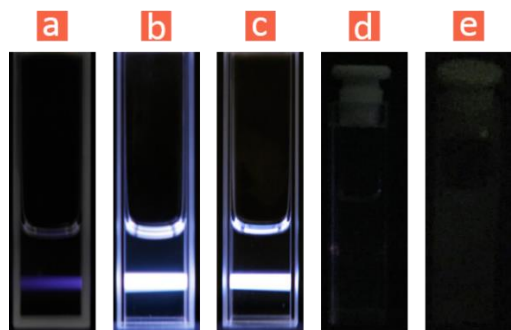


Figure S3. Photograph of the nanoparticles OA-NaYbF₄:Tm (a) , OA-NaYbF₄:Tm@CaF₂, (b) OA-NaYbF₄:Tm@CaF₂@NaDyF₄ (c), OA-NaDyF₄:Yb,Tm (d), OA-NaYbF₄:Tm@NaDyF₄:Tm (e) in cyclohexane solution under excitation with a 980 nm laser. The Tm³⁺ concentration of all samples was fixed at 1 mg/mL.

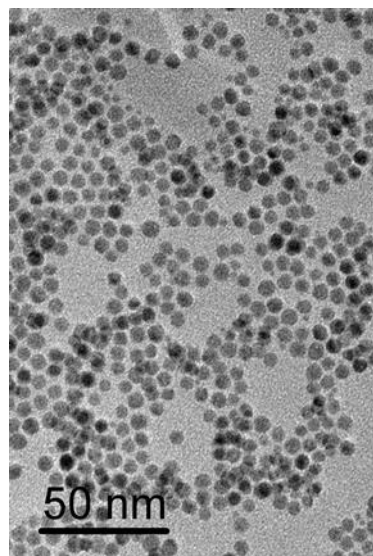


Figure S4. TEM image of the OA-NaDyF₄:Yb,Tm nanoparticles.

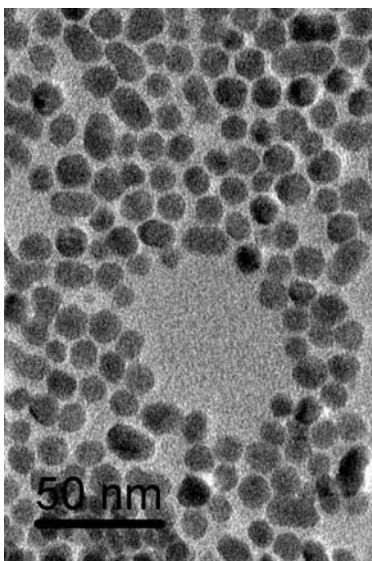


Figure S5. TEM image of the OA-NaYbF₄:Tm@NaDyF₄ nanoparticles.

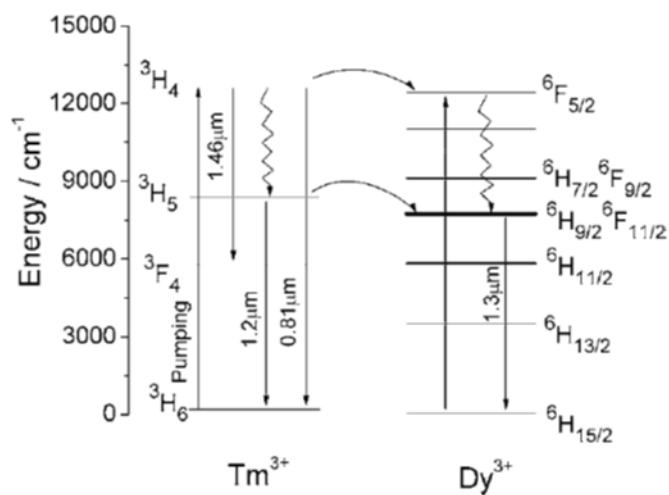


Figure S6. The energy level diagram of Tm³⁺ and Dy³⁺ showing their energy transfer process.

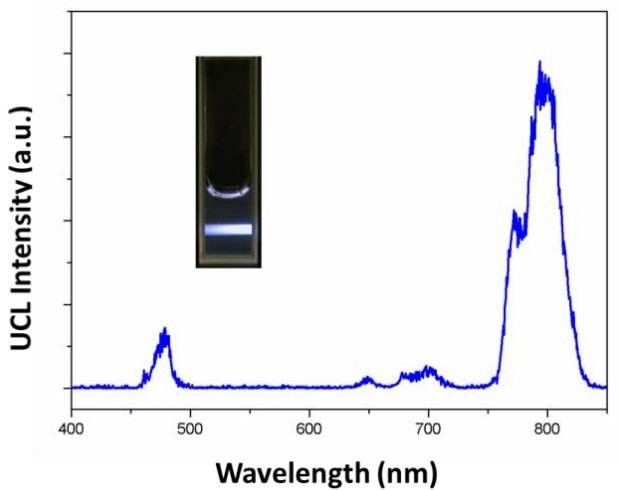


Figure S7. UCL emission spectrum of the nanoparticles Cit-NaYbF₄:Tm@CaF₂@NaDyF₄ dispersed in deionized water under excitation at 980 nm.



Figure S8. The photo showing the stability of Cit-NaYbF₄:Tm@CaF₂@NaDyF₄ (30 mg/mL) dispersed in water for more than one month.

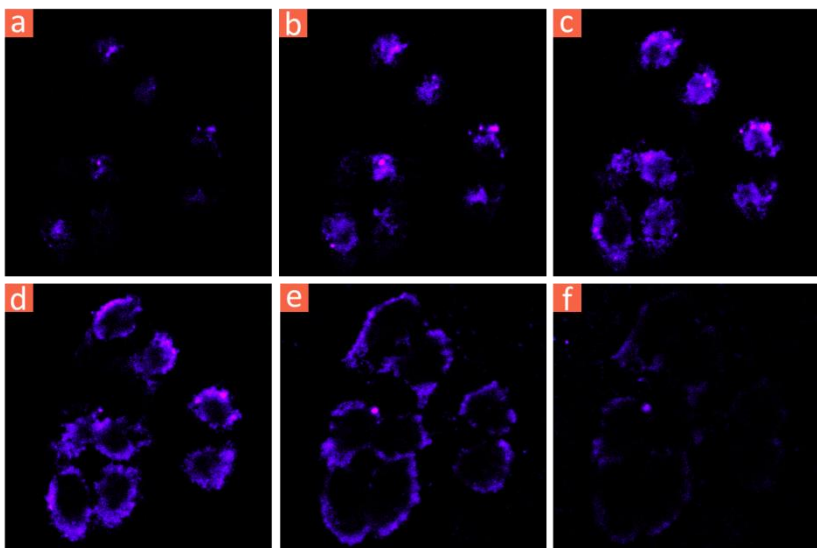


Figure S9. The overlap Z-scan confocal UCL image of the living HeLa cells incubated with 20 $\mu\text{g/mL}$ Cit-NaYbF₄:Tm@CaF₂@NaDyF₄ for 90 min at 37 °C ($\lambda_{\text{ex}} = 980 \text{ nm}$).

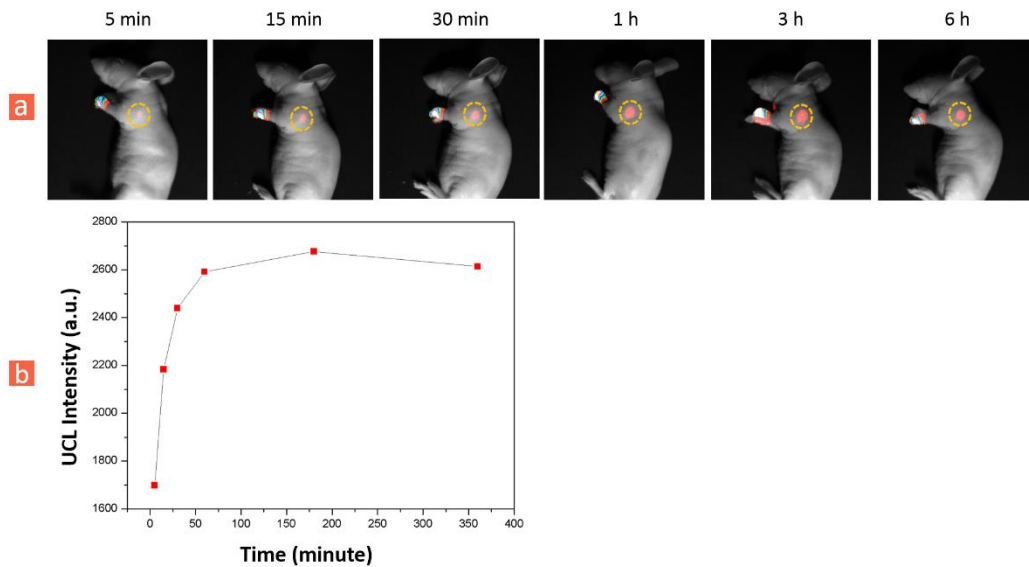


Figure S10. (a) *In vivo* lymphatic drainage of bright-field and UCL imaging overlay images after injection with 50 μL of Cit-NaYbF₄:Tm@CaF₂@NaDyF₄ nanocomposites (30 mg/mL) on the left forepaw. All images were acquired under the same instrumental conditions. (Power density: 50 mW cm^{-2} . $\lambda_{\text{ex}} = 980 \text{ nm}$, $\lambda_{\text{UCL}} = 800 \pm 12 \text{ nm}$). (b) Average UCL intensity measured in region of interest (ROI) at 5, 15, 30, 60, 180 and 360 minutes.