Self-Assembly of Concentric Hexagons and Hierarchical Self-Assembly of Supramolecular Metal-Organic Nanoribbons at Solid/Liquid Interface

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Table of Contents

1. Experimental section	4
2. Synthesis of ligands LA, LB, LC and LD	7
3. Synthesis of the ligands and complexes	4
4. ESI mass spectra data of $[\mathbf{Zn}_{12}\mathbf{LA}_6]$, $[\mathbf{Zn}_{12}\mathbf{LB}_6]$, $[\mathbf{Zn}_{12}\mathbf{LC}_6]$ are	ıd
[Zn ₁₂ LD ₆]	2
5. Energy-minimized structure from molecular modeling, ESI-MS and TWIM-M	S
plots of complex [Zn ₁₂ LA ₆]	3
6. Calibration of drift time scale	4
7. Molecular modeling results	7
8. ¹ H NMR, ¹³ C NMR, 2D COSY NMR, 2D NOESY NMR and MALDI-TOF.S48-6	3
9. ESI-MS of multiple macrocycles assembled by ditopic tpy ligand 6 with $Zn^{2+}S6$	4
10. STM images	6
11. TEM images	58
12. Reference	9

1. Experimental section

General Procedures. All reagents were purchased from Sigma-Aldrich, Matrix Scientific, Alfa Aesar and used without further purification. Column chromatography was conducted using basic Al₂O₃ (Brockman I, activity, 58 Å) or SiO₂ (VWR, 40-60 μ m, 60 Å) and the separated products were visualized by UV light. NMR spectra data were recorded on a 400-MHz and 500 MHz Bruker Avance NMR spectrometer in CDCl₃, CD₃CN or DMSO-*d*6 with TMS as reference. ESI-MS and TWIM-MS were recorded with a Waters Synapt G2 tandem mass spectrometer, using solutions of 0.01 mg sample in 1 mL of CHCl₃/CH₃OH (1:3, v/v) for ligands or 0.5 mg sample in 1 mL of MeCN/MeOH (3:1, v/v) for complexes. MALDI-TOF was carried out on Bruker AutoFlex TOF/TOF mass spectrometer.

TWIM-MS. The TWIM-MS experiments were performed under the following conditions: ESI capillary voltage, 3 kV; sample cone voltage, 30 V; extraction cone voltage, 3.5 V; source temperature 100 °C; desolvation temperature, 100 °C; cone gas flow, 10 L/h; desolvation gas flow, 700 L/h (N₂); source gas control, 0 mL/min; trap gas control, 2 mL/min; helium cell gas control, 100 mL/min; ion mobility (IM) cell gas control, 30 mL/min; sample flow rate, 5 μ L/min; IM traveling wave height, 25 V; and IM traveling wave velocity, 1000 m/s. Q was set in rf-only mode to transmit all ions produced by ESI into the triwave region for the acquisition of TWIM-MS data.

Collision cross-section calibration. The calibration procedure of Scrivens et al¹ was used to convert the drift time scale of the TWIM-MS experiments to a collision

cross-section (CCS) scale. The calibration curve was constructed by plotting the corrected CCSs of the molecular ions of myoglobin² against the corrected drift times of the corresponding molecular ions measured in TWIM-MS experiments at the same traveling wave velocity, traveling wave height and ion mobility gas flow settings *viz.*, 1000 m/s, 25 V, and 30 mL/min, respectively.

Molecular modeling. Energy minimization of the macrocycles was conducted with Materials Studio version 4.2, using the Anneal and Geometry Optimization tasks in the Forcite module (Accelrys Software, Inc.). All counterions are omitted. An initially energy-minimized structure was subjected to 70 - 100 annealing cycles with initial and mid-cycle temperatures of 300 and 1500 K, respectively, twenty heating ramps per cycle, one thousand dynamic steps per ramp, and one femtosecond per dynamic step. A constant volume/constant energy (NVE) ensemble was used and the geometry was optimized after each cycle. Geometry optimization used a universal force field with atom-based summation and cubic spline truncation for both the electrostatic and Van der Waals parameters. 70 - 100 energy-minimized structures were selected for the calculation of theoretical collision cross-sections using MOBCAL programs.

TEM: The sample was dissolved in CH_3CN at a concentration of 10^{-6} M. The solutions were drop cast on to a carbon-coated Cu grid and extra solution was absorbed by filter paper to avoid aggregation. The TEM images of the drop cast samples were taken with a JEOL 2010 transmission electron microscope.

STM: The sample was dissolved in DMF at a concentration of 5.0 mg/ml.

Solution (5ul) was dropped on HOPG surface. After 30 seconds, surface was washed slightly with water for three times and totally dried in R.T. in air. The STM images were taken with a PicoPlus SPM system with a PicoScan 3000 Controller. The obtained STM images were processed by WSxM software.³

2. Synthesis of ligands LA, LB, LC and LD



Scheme 1. Synthesis of ligands LA and LB.



Scheme 2. Synthesis of ligand LC.



Scheme 3. Synthesis of ligand LD.

3. Synthesis of the ligands and complexes



Compound **2**. The mixture of 4-Brom-terpyridine⁴ (3.12 g, 10 mmol), Bis-(pinacolato)diboron (2.54 g, 10 mmol), Pd(dppf)Cl₂ (326 mg, 0.4 mmol) and potassium acetate (2.9 g, 30 mmol) were added 35 mL anhydrous DMSO. The solution was heated at 85 °C for 8 h. The crude product was directly used in the following step. ¹H NMR (400 MHz, CDCl₃) δ 8.83 (s, 2H, tpy- $H^{3,5'}$), 8.74 (ddd, J =4.8, 1.8, 0.9 Hz, 2H, tpy- $H^{6,6''}$), 8.62 (dt, J = 8.0, 1.1 Hz, 2H, tpy- $H^{3,3''}$), 7.86 (ddd, J =8.0, 7.5, 1.8 Hz, 2H, tpy- $H^{4,4''}$), 7.34 (ddd, J = 7.5, 4.8, 1.2 Hz, 2H, tpy- $H^{5,5''}$), 1.40 (s, 6H, H^A). ¹³C NMR (100 MHz, CDCl₃) δ 156.49, 154.76, 149.15, 136.76, 126.27, 123.60, 121.22, 84.48, 24.95. ESI-MS (m/z): Calcd. for [C₂₁H₂₂BN₃O₂+H]⁺: 360.2. Found: 360.2.



Compound 5. To a solution of NaOH powder (4.8 g, 120 mmol) in EtOH (100 ml), 5-bromoisophthalaldehyde⁵ (2.1 g, 10 mmol) and 2-acetylpyridine (3.0 g, 25 mmol) was added. After stirring at room temperature for 24 h, aqueous $NH_3 \cdot H_2O$ (35 mL)

was added and the mixture was refluxed for 40 h. After cooling to room temperature, the precipitate was filtered. The crude was purified by column chromatography on silica gel with chloroform: ethanol (100:2) as eluent to afford the product as a white solid (2.8 g, 45%). ¹H NMR (400 MHz, CDCl₃) δ 8.80 (s, 4H, tpy- $H^{3',5'}$), 8.77 (ddd, *J* = 4.7, 1.7, 0.8 Hz, 4H, tpy- $H^{6.6''}$), 8.73 (d, *J* = 8.0 Hz, 4H, tpy- $H^{3,3''}$), 8.27 (t, *J* = 1.6 Hz, 1H, Ph- H^A), 8.13 (d, *J* = 1.6 Hz, 2H, Ph- H^B), 7.93 (td, *J* = 7.7, 1.8 Hz, 4H, tpy- $H^{4.4''}$), 7.40 (ddd, *J* = 7.4, 4.8, 1.2 Hz, 4H, tpy- $H^{5.5''}$). ¹³C NMR (100 MHz, CDCl₃) δ 156.17, 155.96, 149.15, 148.69, 141.45, 136.89, 130.75, 125.10, 123.98, 123.73, 121.44, 119.06. ESI-MS (m/z): Calcd. for [C₃₆H₂₃BrN₆+H]⁺: 619.1. Found: 619.1.



Compound 7. To a flask containing a degassed solution of compound 5 (620 mg, 1.0 mmol), 4-(4,4,5,5-tetramethyl-1,3,2-dioxaborolan-2-yl)aniline (330 mg, 1.5 mmol) in toluene (40 mL), an aqueous solution of 2 M K₂CO₃ (15 mL) was added. Pd(PPh₃)₂Cl₂ (70 mg, 0.1 mmol) was then added under N₂. The mixture was stirred at 90 °C for 24 h and then cooled to room temperature. The aqueous phase was extracted with CH₂Cl₂. The combined organic phase was washed with brine and dried with anhydrous Na₂SO₄. After removal of volatile under vacuum, the crude was purified by column chromatography on silica gel with chloroform: ethanol (100:1) as eluent to

afford the product as a white solid (505 mg, 80%). ¹H NMR (400 MHz, CDCl₃) δ 8.87 (s, 4H, tpy- $H^{3',5'}$), 8.76 (ddd, J = 4.8, 1.8, 0.9 Hz, 4H, tpy- $H^{6,6''}$), 8.73 (dt, J = 8.0, 1.1 Hz, 4H, tpy- $H^{3,3"}$), 8.23 (t, J = 1.7 Hz, 1H, Ph- H^A), 8.11 (d, J = 1.7 Hz, 2H, Ph- $H^{\rm B}$), 7.92 (ddd, J = 7.9, 7.5, 1.8 Hz, 4H, tpy- $H^{4,4"}$), 7.67 – 7.61 (m, 2H, Ph- $H^{\rm C}$), 7.38 (ddd, J = 7.5, 4.8, 1.2 Hz, 4H, tpy- $H^{5,5"}$), 6.88 – 6.82 (m, 2H, Ph- H^{D}), 3.78 (br, 2H, -*NH*₂). ¹³C NMR (100 MHz, CDCl₃) δ 156.26, 156.02, 150.50, 149.14, 146.35, 142.84, 140.11, 136.85, 130.72, 128.42, 126.19, 124.31, 123.81, 121.42, 119.39, 115.43. ESI-MS (m/z): Calcd. for $[C_{42}H_{29}N_7+H]^+$: 632.3. Found: 632.3.



Compound 9. Anhydrous sodium acetate (328 mg, 4.0 mmol) and the pyrylium salt 1^6 (704 mg, 1.0 mmol) were added to a solution of 7 (631 mg, 1.0 mmol) in anhydrous ethanol (25 mL). The mixture was reflux for 8 h and then cooled to room temperature. The precipitate was filtered and washed with water and diethyl ether to give the product as a white solid (1.07 g, 81%). ¹H NMR (400 MHz, DMSO- d_6) δ 8.72 (m, 8H, tpy- $H^{3',5'}$ and tpy- $H^{6,6''}$), 8.66 (s, 2H, Ph- H^{G}), 8.60 (d, J = 7.8 Hz, 4H, tpy- $H^{3,3''}$), 8.25

(m, 3H, Ph- H^{A} and Ph- H^{H}), 8.09 (s, 2H, Ph- H^{B}), 8.05 – 7.97 (m, 4H, tpy- $H^{4,4"}$), 7.84 (s, 2H, Ph- H^{C}), 7.79 (d, J = 8.3 Hz, 4H, Ph- H^{E}), 7.69 (d, J = 8.5 Hz, 2H, Ph- H^{J}), 7.62 (s, 2H, Ph- H^{D}), 7.51 (m, 4H, tpy- $H^{5,5"}$), 7.30 (d, J = 8.6 Hz, 4H, Ph- H^{F}), 1.36 (s, 9H, tert-butyl- H^{K}). ¹³C NMR (100 MHz, DMSO- d_{6}) δ 156.09, 155.85, 155.37, 150.82, 149.72, 149.30, 140.53, 140.20, 139.90, 139.02, 138.07, 137.92, 137.60, 132.93, 132.10, 130.95, 129.84, 129.75, 129.13, 127.81, 127.15, 126.59, 125.41, 125.00, 121.51, 119.07, 99.98, 98.26, 35.4, 31.3. ESI-MS (m/z): Calcd. for $[C_{69}H_{50}I_2N_7]^+$: 1230.2. Found: 1230.2.



Ligand LA: To a solution of compound **9** (120.0 mg, 91.1 μ mol), Pd(PPh₃)₂Cl₂ (14.0 mg, 20.0 μ mol) and compound **2** (81.8 mg, 227.8 μ mol) in DMSO (25 mL) under N₂, aqueous 2 M K₂CO₃ (8 mL) was added. The mixture was stirred at 80 °C for 30 h and then cooled to room temperature. The mixture was added into 200 ml water and was extracted with CHCl₃, and the combined organic phase was washed with brine and

dried over Na₂SO₄. After removal of solvent under vacuum, the residue was purified by column chromatography on Al₂O_{3.} The polarity of eluent was increased from chloroform/acetone 100/0 to 0/100, then a mixture of acetone/acetonitrile, 70/30 to 0/100 and finally a mixture of acetonitrile/water/NH₄BF₄ to afford LA as a white solid (61 % yield). ¹H NMR (400 MHz, CDCl₃) δ 8.55 – 8.51 (m, 4H, tpy-*H*^{a3,3"}), 8.45 -8.40 (m, 4H, tpy- $H^{b3,3"}$), 8.40 - 8.32 (m, 8H, tpy- $H^{a3',5'}$ and tpy- $H^{b3',5'}$), 8.28 (m, 8H, $tpy-H^{a6,6"}$ and $tpy-H^{b6,6"}$), 7.98-7.95 (m, 4H, Ph- H^{G} and Ph- H^{H}), 7.94-7.90 (m, 4H, Ph- H^{E}), 7.84 – 7.75 (m, 8H, Ph- H^{B} , Ph- H^{C} and Ph- H^{F}), 7.72 – 7.67 (m, 4H, tpy- $H^{b4,4"}$), 7.60-7.54 (m, 9H, Ph- H^A , Ph- H^D , Ph- H^I and tpy- $H^{a4,4"}$), 7.23 – 7.15 (m, 4H, tpy- $H^{b5,5"}$), 7.14 - 7.04 (m, 4H, tpy- $H^{a5,5"}$), 1.36 (s, 9H, tert-butyl- H^{J}). ¹³C NMR (100 MHz, CDCl₃) & 156.46, 156.45, 155.95, 155.91, 155.75, 155.71, 149.28, 149.08, 149.01, 148.95, 148.27, 141.62, 140.32, 140.00, 138.72, 136.77, 136.61, 133.65, 132.14, 131.92, 131.24, 130.80, 129.32, 128.43, 127.94, 127.47, 126.87, 126.36, 126.08, 123.82, 123.65, 121.35, 121.21, 121.11, 118.98, 118.74, 35.17, 31.10. MALDI-TOF MS (m/z): Calcd. for $[C_{105}H_{82}N_{13}O]^+$ 1440.6. Found: 1440.6.



S12

Complex [**Zn**₁₂**LA**₆]: To a solution of ligand **LA** (6.0 mg, 3.9 µmol) in CHCl₃ (3 mL), a solution of Zn(NO₃)₂•6H₂O (2.3 mg, 7.9 µmol) in MeOH (9 mL) was added. The mixture was stirred at 50 °C for 8 h and then cooled to room temperature. Upon addition of NH₄PF₆ (200 mg), a precipitate was formed and washed with water to give a white product (yield: 90%). ESI-MS (m/z): 1578.0 [M-8PF₆ –]⁸⁺ (calcd m/z: 1578.0), 1386.6 [M-9PF₆ –]⁹⁺ (calcd m/z: 1386.6), 1233.6 [M-10PF₆ –]¹⁰⁺ (calcd m/z: 1233.6), 1108.1 [M-11PF₆ –]¹¹⁺ (calcd m/z: 1108.1), 1003.6 [M-12PF₆ –]¹²⁺ (calcd m/z: 1003.6), 915.2 [M-13PF₆ –]¹³⁺ (calcd m/z: 915.2), 839.6 [M-14PF₆ –]¹⁴⁺ (calcd m/z: 839.6), 773.8 [M-15PF₆ –]¹⁵⁺ (calcd m/z: 773.8), 716.5 [M-16PF₆ –]¹⁶⁺ (calcd m/z: 716.5), 665.9 [M-17PF₆ –]¹⁷⁺ (calcd m/z: 665.9) and 620.8 [M-18PF₆ –]¹⁸⁺ (calcd m/z: 620.8).



Compound 4. A mixture of 4-bromo-2,6-bis(hydroxymethyl)phenol⁷ (6.0 g, 25.9 mmol), 1-bromohexane (5.1 g, 31.1 mmol), K_2CO_3 (7.2 g, 52.0 mmol) and methyl ethyl ketone (200 mL) was refluxed under N₂ for 24 h. The mixture was cooled to room temperature, filtrated and dried under vacuum. The white residue, PCC (11.8 g, 75 mmol) and celite (10 g) were dissolved in DCM (200 ml), and the mixture was stirred for 20 h at room temperature. The solution was filtered and poured onto silica gel column with DCM as eluent to afford

5-bromo-2-(hexyloxy)benzene-1,3-dialdehyde as a white solid (86% yield). ¹H NMR (400 MHz, CDCl₃) δ 10.36 (s, 2H, Ph- H^{B}), 8.20 (s, 2H, Ph- H^{A}), 4.14 (t, J = 6.4Hz, 2H), 1.95-1.87 (m, 2H), 1.56 – 1.45 (m, 2H), 1.40 – 1.35 (m, 4H), 0.97 – 0.89 (t, J = 6.4 Hz, 3H). ¹³C NMR (100 MHz, CDCl₃) δ 187.13, 139.03, 137.24, 131.76, 118.29, 81.04, 31.48, 29.85, 25.45, 22.49, 13.94.



Compound 6. To a solution of NaOH powder (6.24 g, 156.0 mmol) in EtOH (350 ml), 5-bromo-2-(hexyloxy)benzene-1,3-dialdehyde (4.0)g, 13.0 mmol) and 2-acetylpyridine (7.5 g, 62 mmol) were added. After stirring at room temperature for 20 h, aqueous NH₃•H₂O (150 mL) was added and the mixture was refluxed for 40 h. Upon cooling to room temperature, the precipitate was filtered and washed with cold ethanol to give 6 as a white solid (4.8 g, 51%). ¹H NMR (400 MHz, CDCl₃) δ 8.80 (s, 4H, tpy- $H^{3',5'}$), 8.77 (ddd, J = 4.8, 1.8, 0.9 Hz, 4H, tpy- $H^{6,6''}$), 8.70 (dt, J = 8.0, 1.0 Hz, 4H, tpy- $H^{3,3"}$), 7.91 (td, J = 7.7, 1.8 Hz, 4H, tpy- $H^{4,4"}$), 7.80 (s, 2H, Ph- H^{A}), 7.38 (ddd, J = 7.5, 4.8, 1.2 Hz, 4H, tpy- $H^{5,5"}$), 3.37 (t, J = 6.4 Hz, 2H), 1.24 – 1.17 (dd, J = 9.1, 6.2 Hz, 2H), 0.92 (s, 2H), 0.83 – 0.66 (m, 4H), 0.55 – 0.41 (t, J = 6.8 Hz, 3H). ¹³C NMR (100 MHz, CDCl₃) & 156.10, 155.58, 153.98, 149.20, 147.08, 136.80, 135.89, 133.66, 123.82, 121.45, 121.25, 117.05, 74.33, 31.29, 29.70, 25.39, 22.11, 13.77. ESI-MS (m/z): Calcd. for [C₄₂H₃₅BrN₆O+H]⁺: 719.2. Found: 719.2.



Compound 8. To a flask containing a degassed solution of compound 6 (720 mg, 1 mmol), 4-(4,4,5,5-tetramethyl-1,3,2-dioxaborolan-2-yl)aniline (330 mg, 1.5 mmol) in toluene (30 mL), an aqueous solution of 2 M K₂CO_{3 (15 mL)} was added. The Pd(PPh₃)₂Cl₂ (70 mg, 0.1 mmol) was added into the mixture under N₂. The mixture was stirred at 90 °C for 20 h and then cooled to room temperature. The aqueous phase was extracted with CH₂Cl₂, and the combined organic phase was washed with brine and dried with anhydrous Na₂SO₄. After removal of solvent under vacuum, the crude was purified by column chromatography on silica gel with chloroform: ethanol (100:1) as eluent to afford the product as a white solid (636 mg, 87%). ¹H NMR (400 MHz, CDCl₃) δ 8.85 (s, 4H, tpy- $H^{3',5'}$), 8.76 (ddd, J = 4.8, 1.6, 0.8 Hz, 4H, tpy- $H^{6,6''}$), 8.70 (d, J = 8.0 Hz, 4H, tpy- $H^{3,3"}$), 7.89 (td, J = 7.7, 1.8 Hz, 4H, tpy- $H^{4,4"}$), 7.78 (s, 2H, Ph- H^{A}), 7.52 (d, J = 8.5 Hz, 2H, Ph- H^{B}), 7.35 (ddd, J = 7.5, 4.8, 1.1 Hz, 4H, tpy- $H^{5,5"}$), 6.78 (d, J = 8.5 Hz, 2H, Ph- H^{C}), 3.76 (br, 2H, - NH_{2}), 3.40 (t, J = 6.1 Hz, 2H), 1.28 – 1.10 (m, 2H), 0.97 - 0.89 (m, 2H), 0.82 - 0.68 (m, 4H), 0.58 - 0.43 (t, J = 6.8 Hz, 3H). ¹³C NMR (100 MHz, CDCl₃) & 156.42, 155.41, 153.38, 149.19, 148.77, 145.95, 137.51, 136.73, 134.10, 130.41, 129.08, 128.12, 123.65, 121.82, 121.26, 115.41, 74.22, 31.36, 29.78, 25.47, 22.15, 13.79. ESI-MS (m/z): Calcd. for $[C_{48}H_{41}N_7O+H]^+$: 732.3. Found:



Compound **10**. Anhydrous sodium acetate (328 mg, 4.0 mmol) and the pyrylium salt **1** (704 mg, 1.0 mmol) were added to a solution of **8** (731 mg, 1.0 mmol) in dry ethanol (20 mL). The mixture was reflux for 10 h and then cooled to room temperature. The precipitate was filtered and washed with water and diethyl ether to give the product as a white solid (1.06 g, 75%). ¹H NMR (400 MHz, DMSO- d_6) δ 8.81 – 8.73 (m, 8H, tpy- $H^{3,5'}$ and tpy- $H^{6,6''}$), 8.69 (d, J = 7.9 Hz, 4H, tpy- $H^{3,3''}$), 8.65 (s, 2H, Ph- H^F), 8.27 (d, J = 8.6 Hz, 2H, Ph- H^G), 8.04 (td, J = 7.8, 1.7 Hz, 4H, tpy- $H^{4,4''}$), 7.94 (s, 2H, Ph- H^A), 7.88 (d, J = 8.5 Hz, 2H, Ph- H^B), 7.78 (d, J = 8.4 Hz, 4H, Ph- H^D), 7.67 (d, J = 8.6 Hz, 2H, Ph- H^F), 3.25 (t, J = 5.5 Hz, 2H), 1.35 (s, 9H, tert-butyl- H^J), 1.10 – 1.00 (m, 2H), 0.85 – 0.75 (m, 2H), 0.63 – 0.55 (m, 4H), 0.36 (t, J = 6.8 Hz, 3H). ¹³C NMR (100 MHz, DMSO- d_6) δ 156.38, 155.94, 155.88, 155.52, 154.80, 149.81, 147.78, 139.91, 138.71, 137.91, 137.56, 134.65, 133.00, 132.07, 131.98, 131.88, 131.01, 129.74, 129.44, 129.27, 129.15, 127.29, 127.10, 125.39, 124.92, 121.55, 121.35,

732.3.

98.22, 74.24, 35.37, 31.28, 31.08, 29.58, 25.38, 21.99, 13.96. ESI-MS (m/z): Calcd. for [C₇₅H₆₂I₂N₇O]⁺: 1330.3. Found: 1330.2.



Ligand LB: To a solution of compound 10 (150.0 mg, 105.9 µmol), Pd(PPh₃)₂Cl₂ (14.0 mg, 20.0 µmol) and compound 2 (95.0 mg, 264.6 µmol) in DMSO (30 mL) under nitrogen, aqueous 2 M K₂CO₃ (8 mL) was added. The mixture was stirred at 80 $\mathbb C$ for 20 h and then cooled to room temperature. The mixture was added into 250 ml water and was extracted with CHCl₃, and the combined organic phase was washed with brine and dried over Na₂SO₄. After removal of solvent under vacuum, the residue was purified by column chromatography on Al₂O_{3.} The polarity of eluent was chloroform/acetone increased from 100/0 0/100, then mixture of to а acetone/acetonitrile, 60/400/100 and finally mixture of to a acetonitrile/water/NH₄BF₄ to afford **LB** as a white solid (64 % yield). ¹H NMR (400 MHz, CDCl₃) δ 8.69 (s, 4H, tpy- $H^{a3',5'}$), 8.66 (s, 4H, tpy- $H^{b3',5'}$), 8.64-8.60 (d, J = 6.6

Hz, 12H, tpy- $H^{a6.6"}$, tpy- $H^{a3.3"}$ and tpy- $H^{b6.6"}$), 8.59 (d, J = 8.0 Hz, 4H, tpy- $H^{b3.3"}$), 8.26 (s, 2H, Ph- H^{F}), 7.98 (d, J = 8.5 Hz, 2H, Ph- H^{G}), 7.92 (d, J = 8.4 Hz, 4H, Ph- H^{D}), 7.88 – 7.78 (m, 14H, Ph- H^{E} , Ph- H^{B} , tpy- $H^{a4.4"}$ and tpy- $H^{b4.4"}$), 7.71 (s, 2H, Ph- H^{A}), 7.64 (d, J = 8.6 Hz, 2H, Ph- H^{H}), 7.60 (d, J = 8.1 Hz, 2H, Ph- H^{C}), 7.35 – 7.23 (m, 8H, tpy- $H^{a5.5"}$ and tpy- $H^{b5.5"}$), 3.27 (t, J = 6.0 Hz, 2H), 1.40 (s, 9H, tert-butyl- H^{I}), 1.19 – 1.05 (m, 2H), 0.90 – 0.80 (m, 2H), 0.79 – 0.62 (m, 4H), 0.45 (t, J = 6.8 Hz, 3H). ¹³C NMR (100 MHz, CDCl₃) δ 156.94, 156.48, 156.46, 156.17, 156.02, 155.89, 155.36, 155.07, 149.12, 149.10, 148.50, 147.94, 141.27, 140.37, 138.16, 136.70, 136.61, 134.37, 134.33, 133.57, 131.24, 130.66, 129.68, 129.20, 128.39, 127.54, 126.88, 126.12, 123.78, 123.68, 123.58, 121.56, 121.38, 121.21, 121.15, 118.90, 74.23, 67.09, 35.16, 31.27, 31.09, 29.63, 25.32, 22.07, 13.72. MALDI-TOF MS (m/z): Calcd. for [C₁₀₅H₈₂N₁₃O]⁺ 1540.7. Found: 1540.8.



Complex $[Zn_{12}LB_6]$: To a solution of ligand LB (6.8 mg, 4.2 µmol) in CHCl₃ (1 mL), a solution of $Zn(NO_3)_2$ •6H₂O (2.5 mg, 8.4 µmol) in MeOH (3 mL) was added. The mixture was stirred at 55 °C for 8 h and then cooled to room temperature. Upon

addition of NH₄PF₆ (200 mg), a precipitate was formed and washed with water to give a white product (yield: 92%). ¹H NMR (400 MHz, CD₃CN) δ 9.18 (s, 4H, tpy- $H^{a3',5'}$), 9.07 (s, 4H, tpy- $H^{b3',5'}$), 8.68 (m, tpy- $H^{a3,3''}$ and Ph- H^F), 8.60 (m, 4H, tpy- $H^{b3,3''}$), 8.42 $(m, 4H, Ph-H^{D})$, 8.27 $(m, 2H, Ph-H^{G})$, 8.22 $(s, 2H, Ph-H^{A})$, 8.02 $(m, 4H, Ph-H^{E})$, 7.94 (br, 4H, Ph- H^{B} and Ph- H^{C}), 7.87 (m, 2H, Ph- H^{H}), 7.79 (m, 8H, tpy- $H^{a4,4''}$ and tpy- $H^{a6,6"}$), 7.73 – 7.65 (m, 4H, tpy- $H^{b4,4"}$), 7.63 (m, 4H, tpy- $H^{b6,6"}$), 7.18 – 7.10 (m, 4H, tpy- $H^{a5,5"}$), 6.90 (s, 4H, tpy- $H^{b5,5"}$), 3.55 (br, 2H), 1.50 (s, 9H, tert-butyl- H^{I}), 1.28 - 1.20 (m, 2H), 1.13 - 1.05 (m, 2H), 0.75 - 0.66 (m, 2H), 0.57 - 0.42 (m, 2H), 0.16 -0.06 (m, 3H). ¹³C NMR (100 MHz, CD₃CN) δ 156.08, 154.69, 152.98, 149.81, 149.14, 148.07, 147.89, 147.72, 141.08, 138.00, 137.89, 135.62, 132.67, 132.25, 131.22, 131.09, 130.69, 129.32, 128.71, 128.39, 127.60, 127.18, 124.69, 123.50, 123.44, 123.09, 121.92, 35.01, 31.10, 30.31, 30.21, 25.91, 21.80, 12.75. ESI-MS (*m/z*): 1653.0 $[M-8PF_6^{-1}]^{8+}$ (calcd m/z: 1653.0), 1452.2 $[M-9PF_6^{-1}]^{9+}$ (calcd m/z: 1452.2), 1293.5 $[M-10PF_6^{-}]^{10+}$ (calcd *m/z*: 1293.5), 1162.7 $[M-11PF_6^{-}]^{11+}$ (calcd *m/z*: 1162.7), 1053.8 $[M-12PF_6^{-1}]^{12+}$ (calcd m/z: 1053.8), 961.5 $[M-13PF_6^{-1}]^{13+}$ (calcd m/z: 961.5), 882.5 $[M-14PF_6^{-1}]^{14+}$ (calcd m/z: 882.5), 814.0 $[M-15PF_6^{-1}]^{15+}$ (calcd m/z: 814.0), 754.1 $[M-16PF_6^{-1}]^{16+}$ (calcd m/z: 754.1), 701.2 $[M-17PF_6^{-1}]^{17+}$ (calcd m/z: 701.2), 654.2 $[M-18PF_6^{-1}]^{18+}$ (calcd m/z: 654.2), 612.2 $[M-19PF_6^{-1}]^{19+}$ (calcd m/z: 612.2), 574.3 $[M-20PF_6^{-1}]^{20+}$ (calcd *m/z*: 574.3) and 540.1 $[M-21PF_6^{-1}]^{21+}$ (calcd *m/z*: 540.1).



Compound **11**. Benzene-1,3,5-triol (3.78 g, 30 mmol) and K₂CO₃ (24.8 g 180 mmol) were degassed solution three times. 1-bromohexane (20 g, 120 mmol) and DMF 150 ml were added. The solution was stirred at 85 °C for 3d under N₂. After that, it was cooled to room temperature. Solution was added 500 ml water and extracted with CH₂Cl₂, and the combined organic phase was washed with brine and dried with anhydrous Na₂SO₄. The crude was purified by column chromatography on silica gel with DCM: hexane (1:4) as eluent to afford the product as a liquid (7.5 g, 66 %). ¹H NMR (400 MHz, CDCl₃) δ 6.09 (s, 1H, Ph-*H*^A), 3.93 (t, *J* = 6.6 Hz, 2H), 1.89 – 1.69 (m, 2H), 1.52 – 1.41 (m, 2H), 1.36 (m, 4H), 0.94 (t, *J* = 6.8 Hz, 3H). ¹³C NMR (100 MHz, CDCl₃) δ 160.98, 93.79, 68.01, 31.60, 29.24, 25.75, 22.62, 14.04.



Compound **12**. Compound **11** (4.0 g, 10.5 mmol) was dissolved in 70 ml CHCl₃. FeCl₃ (170 mg, 1.05 mmol) was added and solution was stirred at room temperature 10 min. A solution of Br₂ (6.7 g, 42 mmol) in 15 ml CHCl₃ was slowly added. Solution was stirred at room temperature 1 h and refluxed overnight. It was cooled down and added water 40 ml with Na₂S₂O₃ 4.0 g. Solvent was removed and purified with column chromatography on silica gel with hexane as eluent to afford a white solid (5.5 g, 85 %). ¹H NMR (400 MHz, CDCl₃) δ 4.00 (t, , *J* = 6.4 Hz, 2H), 1.96 – 1.85 (m, 2H), 1.64 – 1.49 (m, 2H), 1.47 – 1.34 (m, 4H), 0.94 (t, , *J* = 7.2 Hz, 3H). ¹³C NMR (100 MHz, CDCl₃) δ 154.27, 110.23, 73.66, 31.65, 29.94, 25.52, 22.62, 14.06.



Compound **13**. To a flask containing a degassed solution of compound **12** (6.0 g, 9.8 mmol), 4-(4,4,5,5-tetramethyl-1,3,2-dioxaborolan-2-yl)aniline (700 mg, 3.2 mmol) in toluene (60 mL), an aqueous solution of 2 M K₂CO₃ (25 mL) was added. Pd(PPh₃)₂Cl₂ (140 mg, 0.2 mmol) was then added under N₂. The mixture was stirred at 75 °C overnight and then cooled to room temperature. The aqueous phase was extracted with CH₂Cl₂. The combined organic phase was washed with brine and dried with anhydrous Na₂SO₄. The crude was purified by column chromatography on silica gel with DCM: hexane (1:2) as eluent to afford the product as a white solid (1.1 g, 55 %). ¹H NMR (400 MHz, CDCl₃) δ 7.24 (d, *J* = 8.5 Hz, 2H, Ph-*H*^A), 6.75 (d, *J* = 8.4 Hz, 2H, Ph-*H*^B), 4.05 (t, *J* = 6.6 Hz, 2H), 3.48 (t, *J* = 6.5 Hz, 4H), 1.97 – 1.86 (m, 2H), 1.62 – 1.52 (m, 2H), 1.50 – 1.38 (m, 8H), 1.32 – 1.10 (m, 12H), 0.97 – 0.82 (m, 9H). ¹³C NMR (100 MHz, CDCl₃) δ 154.82, 153.37, 145.43, 131.63, 128.08, 123.23, 114.63, 109.90, 73.48, 73.15, 31.70, 31.45, 30.02, 29.77, 25.59, 25.42, 22.64, 22.55, 14.09, 14.02. ESI-MS (m/z): Calcd. for [C₃₀H₄₅Br₂NO₃+H]⁺: 626.2. Found: 626.2.



Compound 14. To a solution of compound 13 (700 mg, 1.1 mmol), Pd(PPh₃)₂Cl₂ (80 mg, 114 µmol) and 4'-(4-Boronatophenyl)[2,2':6',2"]terpyridine (1.58 g, 4.5 mmol) in toluene (50 mL) and tert-butyl alcohol (8 ml) under N₂, aqueous 1 M K₂CO₃ (20 mL) was added. After refluxing for 48 h, the mixture was cooled to room temperature. The aqueous layer was extracted with CHCl₃ and the combined organic phase was washed with brine and dried over Na₂SO₄. After removal of solvent under vacuum, the residue was purified by column chromatography on silica gel with chloroform: ethanol (100:2) as eluent to afford a white solid in 82 % yield. $^1\!H$ NMR (400 MHz, CDCl_3) δ 8.83 (s, 4H, tpy- $H^{3',5'}$), 8.76 (ddd, J = 4.8, 1.8, 0.9 Hz, 4H, tpy- $H^{6,6''}$), 8.71 (dt, J = 8.0, 1.1 Hz, 4H, tpy- $H^{3,3"}$), 8.02 – 7.95 (d, J = 4.4 Hz, 4H, Ph- H^{D}), 7.90 (ddd, J = 7.9, 7.5, 1.8 Hz, 4H, tpy- $H^{4,4"}$), 7.74 – 7.69 (d, J = 4.4 Hz, 4H, Ph- H^{C}), 7.40 – 7.34 (m, 6H, tpy- $H^{5,5"}$ and Ph- H^{A}), 6.80 – 6.74 (d, J = 4.4 Hz, 2H, Ph- H^{B}), 3.28 (q, J = 6.6 Hz, 6H), 1.16 – 1.00 (m, 12H), 1.00 - 0.83 (m, 12H), 0.75 (t, J = 7.2 Hz, 6H), 0.66 (t, J = 7.2 Hz, 3H).¹³C NMR (100 MHz, CDCl₃) δ 156.35, 155.87, 155.66, 154.71, 150.41, 149.10, 145.07, 136.90, 136.65, 136.12, 132.15, 132.05, 131.62, 128.55, 128.43, 126.50, 123.78, 121.39, 118.89, 114.59, 73.57, 73.26, 31.42, 31.38, 30.90, 29.68, 25.35, 25.34,

22.53, 22.50, 13.97,13.91. ESI-MS (m/z): Calcd. for $[C_{72}H_{73}N_7O_3+H]^+$: 1084.6.

Found: 1084.6.



Compound **15**. Anhydrous sodium acetate (164 mg, 2.0 mmol) and the pyrylium salt **1** (352 mg, 0.5 mmol) were added to a solution of **14** (541 mg, 0.5 mmol) in dry ethanol (15 mL). The mixture was reflux for 8 h and then cooled to room temperature. The precipitate was filtered and washed with water and diethyl ether to give the product as a white solid (708 mg, 80%). ¹H NMR (400 MHz, DMSO- d_6) δ 8.79 (s, 4H, tpy- $H^{3,5'}$), 8.78 – 8.75 (m, 4H, tpy- $H^{6,6''}$), 8.70 (d, J = 8.0 Hz, 4H, tpy- $H^{3,3''}$), 8.61 (s, 2H, Ph- H^E), 8.28 (d, J = 8.7 Hz, 2H, Ph- H^F), 8.10 – 8.01 (m, 4H, tpy- $H^{4,4''}$), 7.97 (d, J = 8.4 Hz, 4H, Ph- H^D), 7.75 (d, J = 8.4 Hz, 4H, Ph- H^I), 7.70 – 7.64 (m, 4H, Ph- H^G and Ph- H^A), 7.62 (d, J = 8.3 Hz, 4H, Ph- H^C), 7.57 – 7.50 (m, 4H, tpy- $H^{5,5''}$), 7.44 (d, J = 8.6 Hz, 2H, Ph- H^B), 7.30 (d, J = 8.4 Hz, 4H, Ph- H^H), 3.33 (m, 2H), 2.90 (m, 4H), 1.36 (s, 9H, tert-butyl- H^I), 1.04 – 0.94 (m, 8H), 0.94 – 0.80 (m, 8H), 0.80 – 0.70 (m, 8H), 0.60 (t,

J = 7.2 Hz, 6H), 0.54 (t, J = 7.2 Hz, 3H). ¹³C NMR (100 MHz, DMSO- d_6) δ 156.59, 156.31, 156.06, 155.94, 155.84, 155.63, 155.41, 149.91, 149.76, 138.08, 137.83, 137.50, 136.98, 136.86, 135.98, 133.06, 132.24, 132.07, 131.52, 130.91, 129.17, 128.56, 127.04, 126.58, 126.05, 125.67, 124.88, 124.32, 121.39, 118.43, 97.98, 73.54, 73.32, 35.34, 31.43, 31.28, 31.10, 29.78, 29.48, 25.51, 25.12, 22.54, 22.30, 14.14, 13.96. ESI-MS (m/z): Calcd. for [C₉₉H₉₄I₂N₇O₃]⁺: 1682.5. Found: 1682.2.



Ligand LC: To a solution of compound 15 (177 mg, 100 μ mol), Pd(PPh₃)₂Cl₂ (10.5 mg, 15.0 μ mol) and 4'-(4-Boronatophenyl)[2,2':6',2"]terpyridine (141 mg, 400 μ mol) in DMSO (25 mL) under N₂, aqueous 2 M K₂CO₃ (7 mL) was added. The mixture was stirred at 80 °C for 2 d and then cooled to room temperature. The mixture was added into 200 ml water and was extracted with CHCl₃, and the combined organic phase was washed with brine and dried over Na₂SO₄. After removal of solvent under vacuum, the residue was purified by column chromatography on Al₂O₃. The polarity

of eluent was increased from chloroform/acetone 100/0 to 0/100, then a mixture of 70/30 acetone/acetonitrile, 0/100 finally mixture of to and a acetonitrile/water/NH₄BF₄ to afford ligand LC as a white solid (66 % yield). ¹H NMR (400 MHz, CDCl₃) δ 8.76 (s, 4H, tpy- $H^{a3',5'}$), 8.75 (s, 4H, tpy- $H^{b3',5'}$), 8.69 – 8.63 (m, 16H, tpy- $H^{a3,3"}$, tpy- $H^{a6,6"}$, tpy- $H^{b3,3"}$ and tpy- $H^{b6,6"}$), 8.23 (s, 2H, Ph- H^{E}), 8.00 – 7.95 (m, 6H, Ph- $H^{\rm D}$ and Ph- $H^{\rm F}$), 7.93 (d, J = 8.4 Hz, 4H, Ph- $H^{\rm K}$), 7.90 – 7.83 (m, 8H, tpy- $H^{a4,4"}$ and tpy- $H^{b4,4"}$), 7.79 (d, J = 8.4 Hz, 4H, Ph- H^{I}), 7.74 (d, J = 8.6 Hz, 2H, Ph- H^{A}), 7.69 – 7.63 (m, 8H, Ph- H^{C} and Ph- H^{H}), 7.62 (d, J = 5.5 Hz, 2H, Ph- H^{G}), 7.60 -7.54 (m, 6H, Ph- H^{B} and Ph- H^{J}), 7.35 - 7.29 (m, 8H, tpy- $H^{a5,5"}$ and tpy- $H^{b5,5"}$), 3.23 (t, J = 6.5 Hz, 2H), 2.95 (t, J = 6.0 Hz, 4H), 1.41 (s, 9H, tert-butyl- H^{L}), 1.03 (m, 6H), 0.94 - 0.78 (m, 10H), 0.78 - 0.70 (m, 4H), 0.68 - 0.59 (m, 7H), 0.55 (t, J = 7.6 Hz, 6H). ¹³C NMR (100 MHz, CDCl₃) δ 156.87, 156.65, 156.36, 156.24, 156.08, 156.05, 155.93, 155.81, 155.18, 150.11, 149.34, 149.07, 149.06, 141.84, 139.73, 138.28, 137.76, 137.09, 137.02, 136.84, 136.80, 135.46, 132.52, 131.84, 131.49, 131.40, 130.69, 128.40, 128.16, 127.97, 127.95, 127.42, 126.86, 126.63, 126.19, 126.16, 123.80, 123.71, 123.69, 121.30, 121.26, 118.86, 118.71, 73.68, 73.32, 35.15, 31.40, 31.32, 31.11, 29.74, 29.53, 25.40, 25.23, 22.59, 22.44, 13.95, 13.86. MALDI-TOF MS (m/z): Calcd. for $[C_{105}H_{82}N_{13}O]^+$ 2045.0. Found: 2045.1.

S25



Complex [$\mathbf{Zn_{12}LC_6}$]: To a solution of ligand LC (6.2 mg, 2.9 µmol) in CHCl₃ (1 mL), a solution of Zn(NO₃)₂•6H₂O (1.7 mg, 5.8 µmol) in MeOH (3 mL) was added. The mixture was stirred at 50 °C for 8 h and then cooled to room temperature. Upon addition of NH₄PF₆ (150 mg), a precipitate was formed and washed with water to give a white product (yield: 91%). ¹H NMR (400 MHz, CD₃CN) δ 8.98 (m, 8H, tpy- $H^{a3',5'}$ and tpy- $H^{b3',5'}$), 8.73 – 8.49 (m, 10H, tpy- $H^{a3,3''}$, tpy- $H^{b3,3''}$ and Ph- H^E), 8.33 (m, 4H, Ph- H^D), 8.27 (d, J = 7.0 Hz, 4H, Ph- H^K), 8.21 (d, J = 7.6 Hz, 2H, Ph- H^F), 8.15 (d, J =7.1 Hz, 4H, Ph- H^C), 8.02 (d, J = 7.5 Hz, 4H, Ph- H^I), 7.76 (m, 28H, tpy- $H^{a4,4''}$, tpy- $H^{b4,4''}$, tpy- $H^{a6,6''}$, tpy- $H^{b6,6''}$, Ph- H^H , Ph- H^G , Ph- H^A and Ph- H^J), 7.59 (m, 2H, Ph- H^B), 7.12 – 6.92 (m, 8H, tpy- $H^{a5,5''}$ and tpy- $H^{b5,5''}$), 3.35 (s, 2H), 3.12 (s, 4H), 1.47 (s, 9H, tert-butyl- H^L), 1.10 (m, 6H), 0.95 (m, 10H), 0.71– 0.50 (m, 17H). ¹³C NMR (100 MHz, CD₃CN) δ 157.06, 156.58, 155.72, 149.62, 147.84, 147.70, 141.23, 140.99, 140.67, 136.13, 134.80, 133.20, 132.30, 130.83, 128.99, 128.57, 127.94, 127.47, 127.29, 127.05, 126.88, 123.29, 123.10, 121.58, 121.33, 73.71, 73.62, 34.94, 31.23,

31.08, 30.29, 29.68, 29.30, 25.43, 25.10, 22.79, 22.35, 13.82, 13.40. ESI-MS (m/z): 1438.1 [M-11PF₆ –]¹¹⁺ (calcd m/z: 1438.1), 1306.2 [M-12PF₆ –]¹²⁺ (calcd m/z: 1306.2), 1194.5 [M-13PF₆ –]¹³⁺ (calcd m/z: 1194.5), 1098.8 [M-14PF₆ –]¹⁴⁺ (calcd m/z: 1098.8), 1015.9 [M-15PF₆ –]¹⁵⁺ (calcd m/z: 1015.9), 943.4 [M-16PF₆ –]¹⁶⁺ (calcd m/z: 943.4), 879.3 [M-17PF₆ –]¹⁷⁺ (calcd m/z: 879.3), 822.5 [M-18PF₆ –]¹⁸⁺ (calcd m/z: 822.5), 771.5 [M-19PF₆ –]¹⁹⁺ (calcd m/z: 771.5), 725.7 [M-20PF₆ –]²⁰⁺ (calcd m/z: 725.7), 684.3 [M-21PF₆ –]²¹⁺ (calcd m/z: 684.3), 646.6 [M-22PF₆ –]²²⁺ (calcd m/z: 646.6), 612.1[M-23PF₆ –]²³⁺ (calcd m/z: 612.1) and 580.6 [M-24PF₆ –]²⁴⁺ (calcd m/z: 580.6)



Compound containing 17. То flask degassed solution of a a 1,4-dibromo-2,5-bis(octyloxy)benzene⁸ (4.4 9.0 mmol). g, 4'-(4-Boronatophenyl)[2,2':6',2"]terpyridine (1.08 g, 3.0 mmol) in toluene (60 mL), an aqueous solution of 2 M K₂CO₃ (25 mL) was added. Pd(PPh₃)₂Cl₂ (105 mg, 0.15 mmol) was then added under N_2 . The mixture was stirred at 80 $\,$ \mathbb{C} for 20 h and then cooled to room temperature. The aqueous phase was extracted with CHCl₃. The combined organic phase was washed with brine and dried with anhydrous Na₂SO₄. After removal of volatile under vacuum, the crude was purified by column chromatography on silica gel with chloroform: ethanol (200:1) as eluent to afford the product as a white solid (1.08 g, 56%). ¹H NMR (400 MHz, CDCl₃) δ 8.72 (ddd, J =

4.8, 1.8, 0.9 Hz, 2H, tpy- $H^{6,6^{\circ}}$), 8.71 – 8.67 (m, 4H, tpy- $H^{3,3^{\circ}}$ and tpy- $H^{3^{\circ},5^{\circ}}$), 7.92 – 7.86 (m, 2H, tpy- $H^{4,4^{\circ}}$), 7.35 (ddd, J = 7.5, 4.8, 1.2 Hz, 2H, tpy- $H^{5,5^{\circ}}$), 7.23 (s, 1H, Ph- H^{A}), 7.13 (d, J = 4.7 Hz, 1H, Ph- H^{B}), 4.07 (t, J = 6.5 Hz, 2H), 3.95 (t, J = 6.3 Hz, 2H), 1.91 – 1.80 (m, 2H), 1.74 – 1.65 (m, 2H), 1.57 – 1.48 (m, 2H), 1.43 – 1.27 (m, 10H), 1.25 – 1.05 (m, 8H), 0.91 (t, J = 7.2 Hz, 3H), 0.82 (t, J = 7.1 Hz, 3H). ¹³C NMR (100 MHz, CDCl₃) δ 156.37, 155.21, 150.75, 150.00, 149.07, 147.73, 136.82, 128.49, 123.66, 121.66, 121.28, 118.23, 116.03, 113.08, 70.55, 69.70, 31.82, 31.76, 29.34, 29.33, 29.24, 29.21, 29.05, 26.10, 26.02, 22.67, 22.58, 14.11, 14.06. ESI-MS (m/z): Calcd. for [C₃₇H₄₆BrN₃O₂+H]⁺: 644.3. Found: 644.3.



Compound **18**. A mixture of compound **17** (1.28 g, 2.0 mmol), Bis-(pinacolato)diboron (508 mg, 2.0 mmol), Pd(dppf)Cl₂ (65 mg, 0.08 mmol) and potassium acetate (980 mg, 10 mmol) was degassed three times. After that, 30 ml anhydrous DMSO was added and then the mixture was stirred at 80 °C for 16 h and then cooled to room temperature. The aqueous phase was extracted with CHCl₃. The combined organic phase was washed with water. After removal of volatile under vacuum, the crude was directly used for next step.



Compound 20. To a solution of compound 19^9 (668 mg, 1.8 mmol), Pd(PPh_3)₂Cl₂ (100 mg, 143 μ mol) and compound **6** (1.08 g, 1.5 mmol) in toluene (40 mL) under N₂, aqueous 1.5 M K₂CO₃ (20 mL) was added. It was heated at 75 °C overnight. The mixture was cooled to room temperature. The aqueous layer was extracted with CHCl₃, and the combined organic phase was washed with brine and dried over Na₂SO₄. After removal of solvent under vacuum, the residue was purified by column chromatography on silica gel with chloroform: ethanol (100:3) as eluent to afford a white solid in 75 % yield. ¹H NMR (400 MHz, CDCl₃) δ 8.90 (s, 4H, tpy- $H^{3',5'}$), 8.79 (d, J = 4.0 Hz, 4H, tpy- $H^{6,6"}$), 8.74 (d, J = 8.0 Hz, 4H, tpy- $H^{3,3"}$), 7.97 – 7.88 (m, 6H, tpy- $H^{4,4"}$ and Ph- H^{G}), 7.83 (d, J = 8.4 Hz, 2H, Ph- H^{A}), 7.76 (d, J = 8.5 Hz, 2H, Ph- H^{B}), 7.72 (d, J = 8.5 Hz, 2H, Ph- H^{C}), 7.66 (d, J = 8.5 Hz, 2H, Ph- H^{D}), 7.50 (d, J = 8.5 Hz, 2H, Ph- H^{E}), 7.42 – 7.37 (m, 4H, tpy- $H^{5,5"}$), 6.81 (d, J = 8.4 Hz, 2H, Ph- H^{F}), 3.45 (t, J= 6.1 Hz, 2H), 1.25 (m, 2H), 0.96 (m, 2H), 0.78 (m, 4H), 0.53 (t, J = 6.7 Hz, 3H). ¹³C NMR (100 MHz, CDCl₃) & 156.28, 155.43, 154.28, 149.16, 148.59, 145.92, 140.20,

139.93, 139.15, 138.79, 138.53, 137.08, 136.90, 136.00, 134.35, 131.08, 131.07, 129.76, 127.94, 127.62, 127.32, 126.74, 123.77, 121.86, 121.35, 115.45, 74.35, 31.36, 29.81, 25.48, 22.16, 13.80. ESI-MS (m/z): Calcd. for $[C_{60}H_{49}N_7O+H]^+$: 884.4. Found: 884.4.



Compound **21**. Anhydrous sodium acetate (328 mg, 4.0 mmol) and the pyrylium salt **1** (704 mg, 1.0 mmol) were added to a solution of **20** (706 mg, 0.8 mmol) in dry ethanol (25 mL). The mixture was reflux for 16 h and then cooled to room temperature. The precipitate was filtered and washed with water and diethyl ether to give the product as a white solid (891 mg, 71%). ¹H NMR (400 MHz, CDCl₃) δ 8.89 (s, 4H, tpy- $H^{3',5'}$), 8.78 (ddd, J = 4.8, 1.8, 0.9 Hz, 4H, tpy- $H^{6,6''}$), 8.73 (dt, J = 8.0, 1.1 Hz, 4H, tpy- $H^{3,3''}$), 8.10 (s, 2H, Ph- H^{H}), 7.89 – 7.95 (m, 6H, tpy- $H^{4,4''}$ and Ph- H^{G}), 7.86 (d, J = 8.7 Hz, 2H,

Ph- H^{I}), 7.83 (d, J = 8.6 Hz, 2H, Ph- H^{A}), 7.71 – 7.77 (m, 4H, Ph- H^{B} and Ph- H^{C}), 7.70 – 7.66 (m, 4H, tpy- H^{L}), 7.64 – 7.60 (m, 4H, Ph- H^{J} and Ph- H^{D}), 7.52 (d, J = 8.8 Hz, 2H, Ph- H^{F}), 7.43 – 7.36 (m, 6H, tpy- $H^{5.5"}$ and tpy- H^{E}), 7.28 – 7.23 (m, 4H, tpy- H^{K}), 3.45 (t, J = 6.0 Hz, 2H), 1.49 (s, 9H, tert-butyl- H^{M}), 1.28 – 1.18 (m, 2H), 0.95 (s, 2H), 0.78 (m, 4H), 0.53 (t, J = 6.8 Hz, 3H). ¹³C NMR (100 MHz, DMSO-d6) δ 157.87, 156.57, 156.34, 155.83, 155.52, 154.42, 149.22, 148.48, 142.36, 140.98, 139.44, 139.19, 139.07, 137.82, 136.80, 134.44, 132.36, 131.29, 131.23, 129.80, 129.75, 128.87, 128.31, 127.74, 127.67, 127.47, 127.44, 126.92, 126.46, 126.34, 126.08, 123.75, 121.76, 121.30, 99.99, 97.38, 74.35, 35.17, 31.36, 31.08, 29.79, 25.47, 22.16, 13.79. ESI-MS (m/z): Calcd. for $[C_{87}H_{70}J_2N_7O]^+$: 1482.4. Found: 1482.2.



Ligand LD: To a solution of compound 21 (340 mg, 216 µmol), Pd(PPh₃)₂Cl₂ (15 mg,

21.6 μ mol) and compound **18** (599 mg, 867 μ mol) in DMSO (40 mL) under N₂, aqueous 2 M K₂CO₃ (10 mL) was added. The mixture was stirred at 80 °C for 2 d and then cooled to room temperature. The mixture was extracted with CHCl₃, and the combined organic phase was washed with brine and dried over Na₂SO₄. After removal of solvent under vacuum, the residue was purified by column chromatography on Al₂O₃. The polarity of eluent was increased from chloroform/acetone 100/0 to 0/100, then a mixture of acetone/acetonitrile, 70/30 to 0/100 and finally a mixture of acetonitrile/water/NH₄BF₄ to afford LD as a white solid (61 % yield). ¹H NMR (400 MHz, CDCl₃) δ 8.89 – 8.86 (s, 4H, tpy- $H^{a^{3',5'}}$), 8.76 (ddd, J = 4.8, 1.8, 0.9 Hz, 4H, tpy- $H^{a6,6''}$), 8.72 (dd, J = 5.5, 2.3 Hz, 12H, tpy- $H^{a3,3''}$, tpy- $H^{b3',5'}$ and tpy- $H^{b6,6''}$), 8.68 (d, J = 8.0 Hz, 4H, tpy- $H^{b3,3''}$), 8.29 (s, 2H, Ph- H^{H}), 8.01 (d, J = 8.8 Hz, 2H, Ph- H^{I}), 7.94 -7.85 (m, 14H, tpy- $H^{a4,4"}$, tpy- $H^{b4,4"}$ Ph- H^{F} and Ph- H^{G}), 7.79 (d, J = 8.4 Hz, 2H, Ph- H^{A}), 7.76 (d, J = 8.6 Hz, 4H, Ph- H^{L}), 7.71 (d, J = 8.1 Hz, 4H, Ph- H^{B} and Ph- H^{C}), 7.68 – 7.63 (m, 6H, Ph- H^{K} and Ph- H^{J}), 7.59 (d, J = 8.6 Hz, 2H, Ph- H^{D}), 7.56 – 7.52 (m, 2H, Ph- H^{E}), 7.39 – 7.31 (m, 8H, tpy- $H^{a5,5"}$ and tpy- $H^{b5,5"}$), 7.17 (s, 2H, Ph- H^{N}), 6.97 (s, 2H, Ph- H^{M}), 3.98 (t, J = 6.2 Hz, 4H), 3.90 (t, J = 6.7 Hz, 4H), 3.43 (t, J = 6.0Hz, 2H), 1.69 (m, 4H), 1.59 (m, 4H), 1.42 (s, 9H, tert-butyl- H^{0}), 1.35 (m, 4H), 1.31 – 1.20 (m, 22H), 1.20 - 1.12 (m, 8H), 1.07 (m, 8H), 0.97 (m, 2H), 0.86 (t, J = 6.9 Hz, 6H), 0.82 - 0.75 (m, 10H), 0.52 (t, J = 6.7 Hz, 3H). ¹³C NMR (100 MHz, CDCl₃) δ 156.98, 156.45, 156.41, 156.30, 155.46, 155.15, 154.36, 150.79, 150.26, 149.17, 149.06, 148.51, 148.02, 141.94, 140.59, 140.32, 139.30, 139.20, 138.50, 137.71, 136.88, 136.84, 134.38, 131.70, 131.37, 130.44, 129.75, 129.74, 129.60, 129.53,

129.10, 128.31, 127.68, 127.51, 127.42, 127.35, 127.08, 126.85, 125.81, 123.74, 123.60, 121.84, 121.79, 121.30, 115.98, 115.70, 100.00, 74.35, 69.79, 69.71, 35.18, 31.83, 31.76, 31.35, 31.12, 29.79, 29.38, 29.36, 29.28, 29.20, 29.18, 29.05, 26.17, 25.90, 25.46, 22.66, 22.58, 22.15, 14.13, 14.06, 13.79. MALDI-TOF MS (*m*/*z*): Calcd. for [C₁₀₅H₈₂N₁₃O]⁺ 2357.3. Found: 2357.4.



Complex [**Zn**₁₂**LD**₆]: To a solution of ligand **LD** (6.1 mg, 2.5 µmol) in CHCl₃ (1 mL), a solution of Zn(NO₃)₂•6H₂O (1.5 mg, 5.0 µmol) in MeOH (3 mL) was added. The mixture was stirred at 50 °C for 10 h and then cooled to room temperature. Upon addition of NH₄PF₆ (130 mg), a precipitate was formed and washed with water to give a white product (yield: 88%). ¹H NMR (400 MHz, CD₃CN) δ 9.35 (s, 4H, tpy-*H*^{a3',5'}), 9.05 (s, 4H, tpy-*H*^{b3',5'}), 8.84 (d, *J* = 8.2 Hz, 4H, tpy-*H*^{a3,3"}), 8.67 (d, *J* = 7.8 Hz, 4H, tpy-*H*^{b3,3"}), 8.61 (s, 2H, Ph-*H*^H), 8.52 (s, 2H, Ph-*H*^G), 8.21 (m, 12H, tpy-*H*^{a4,4"}, tpy-*H*^{b4,4"}, Ph-*H*^I and Ph-*H*^F), 8.02 (d, *J* = 7.8 Hz, 2H, Ph-*H*^J), 7.96 (m, 6H, tpy-*H*^{a6,6"} and Ph-*H*^A), 7.90 (d, *J* = 5.1 Hz, 4H, tpy-*H*^{b6,6"}), 7.82 (d, *J* = 6.7 Hz, 8H, Ph-*H*^L, Ph-*H*^E

and Ph- H^{B}), 7.73 (d, J = 8.3 Hz, 2H, Ph- H^{C}), 7.65 (d, J = 7.5 Hz, 4H, Ph- H^{K}), 7.55 (m, 4H, Ph- H^{D} and Ph- H^{N}), 7.51 – 7.41 (m, 8H, tpy- $H^{a5,5"}$ and tpy- $H^{b5,5"}$), 7.30 (s, 2H, Ph- H^{M}), 4.26 (t, J = 6.0 Hz, 4H), 4.12 (t, J = 6.3 Hz, 4H), 3.74 (s, 2H), 1.89 (m, 4H), 1.65 (m, 4H), 1.49 (s, 15H), 1.30 (m, 30H), 1.08 - 0.95 (m, 20H), 0.90 (t, J = 6.5 Hz, 6H), 0.62 (t, J = 6.8 Hz, 6H), 0.28 (t, J = 7.3 Hz, 3H). ¹³C NMR (100 MHz, CD₃CN) δ 157.00, 156.60, 154.76, 154.05, 153.52, 150.84, 150.59, 149.45, 149.00, 147.98, 141.47, 141.32, 140.08, 139.79, 139.65, 138.42, 138.26, 137.65, 132.88, 132.76, 132.55, 132.22, 130.98, 129.68, 129.62, 129.36, 129.29, 128.56, 127.89, 127.59, 127.52, 127.05, 125.90, 124.52, 124.17, 123.26, 123.07, 116.06, 115.69, 69.60, 69.57, 34.95, 31.67, 31.46, 31.26, 30.31, 29.32, 29.15, 29.13, 28.96, 28.92, 26.40, 26.11, 25.79, 22.47, 22.20, 21.97, 13.51, 13.24, 12.88. ESI-MS (*m*/*z*): 1608.5 [M-11PF₆ -]¹¹⁺ (calcd m/z: 1608.5), 1462.4 [M-12PF₆]¹²⁺ (calcd m/z: 1462.4), 1338.8 [M-13PF₆]¹³⁺ (calcd m/z: 1338.8), 1232.8 [M-14PF₆]¹⁴⁺ (calcd m/z: 1232.8), 1140.9 [M-15PF₆]¹⁵⁺ (calcd m/z: 1140.9), 1060.5 [M-16PF₆⁻]¹⁶⁺ (calcd m/z: 1060.5), 989.7 [M-17PF₆⁻]¹⁷⁺ (calcd m/z: 989.7), 926.6 [M-18PF₆^{-]¹⁸⁺} (calcd m/z: 926.6), 870.2 [M-19PF₆^{-]¹⁹⁺} (calcd m/z: 870.2), 819.5 [M-20PF₆]²⁰⁺ (calcd m/z: 819.5) and 773.5 [M-21PF₆]²¹⁺ (calcd *m/z*: 773.5)

4. ESI mass spectra data of complex $[Zn_{12}LA_6]$, $[Zn_{12}LB_6]$, $[Zn_{12}LC_6]$ and $[Zn_{12}LD_6]$ (PF₆⁻ as counterion).





Figure S1. Measured (bottom) and calculated (top) isotope patterns for different charge states observed from $[Zn_{12}LA_6]$ (PF₆⁻ as counterion).





Figure S2. Measured (bottom) and calculated (top) isotope patterns for different charge states observed from $[\mathbf{Zn}_{12}\mathbf{LB}_6]$ (PF₆⁻ as counterion).





Figure S3. Measured (bottom) and calculated (top) isotope patterns for different charge states observed from $[\mathbf{Zn}_{12}\mathbf{LC}_6]$ (PF₆⁻ as counterion).





Figure S4. Measured (bottom) and calculated (top) isotope patterns for different charge states observed from $[Zn_{12}LD_6]$ (PF₆⁻ as counterion).

5. Energy-minimized structure from molecular modeling, ESI-MS and TWIM-MS plots of complex [Zn₁₂LA₆].



Figure S5. Energy-minimized structure from molecular modeling of complex $[Zn_{12}LA_6]$. (B) ESI-MS and (C) 2D TWIM-MS plot (*m/z* vs drift time). The charge states of intact assemblies are marked.

6. Calibration of drift time scale

Corrected drift times (arrival times) were plotted against corrected published cross sections for the multiply charged ions arising from myoglobin. Drift times were measured at a traveling wave velocity of 1000 m/s and a traveling wave height of 25 V. This calibration plot was utilized to obtain the experimental collision cross sections (CCSs) listed in Table 1.



Figure S6. The calibration curve was constructed by plotting the corrected CCSs of the molecular ions of myoglobin at different corrected drift times.

7. Molecular modeling



Figure S7. Plot of collision cross-section (CCS) *vs.* relative energy for 70 candidate structures of $[Zn_{12}LB_6]$ generated by annealing simulations. CCSs were calculated by the TM method using the MOBCAL program. The average TM cross section area is 1907.7 ±29.8 Å².



Figure S8. Plot of collision cross-section (CCS) *vs.* relative energy for 100 candidate structures of $[Zn_{12}LC_6]$ generated by annealing simulations. CCSs were calculated by the TM method using the MOBCAL program. The average TM cross section area is 2554.2 ±44.1 Å².



Figure S9. Plot of collision cross-section (CCS) *vs.* relative energy for 100 candidate structures of $[Zn_{12}LD_6]$ generated by annealing simulations. CCSs were calculated by the TM method using the MOBCAL program. The average TM cross section area is $3018.9 \pm 58.3 \text{ Å}^2$.



8. ¹H NMR, ¹³C NMR, 2D COSY NMR, 2D NOESY NMR and MALDI-TOF.

Figure S11. ¹H NMR (400 MHz) spectrum of ligand LB.



Figure S13. 2D COSY NMR (400 MHz) spectrum of ligand LB.



Figure S14. 2D COSY NMR (400 MHz) spectrum of ligand LB (aromatic region).



Figure S15. MALDI-TOF mass spectrum of ligand LB.



Figure S17.¹³C NMR (400 MHz) spectrum of ligand LC.



Figure S18.2D COSY NMR (400 MHz) spectrum of ligand LC.



Figure S19. 2D COSY NMR (400 MHz) spectrum of ligand LC (aromatic region).



Figure S21. ¹H NMR (400 MHz) spectrum of ligand LD.



Figure S23.2D COSY NMR (400 MHz) spectrum of ligand LD.



Figure S24. 2D COSY NMR (400 MHz) spectrum of ligand LD (aromatic region).



Figure S25. MALDI-TOF mass spectrum of ligand LD.



Figure S27. ¹³C NMR (400 MHz) spectrum of complex [Zn₁₂LB₆].



Figure S28. 2D COSY NMR (400 MHz) spectrum of complex [Zn₁₂LB₆].



Figure S29. 2D COSY NMR (400 MHz) spectrum of complex [Zn₁₂LB₆] (aromatic

region).



Figure S30. 2D NOESY NMR (400 MHz) spectrum of complex $[Zn_{12}LB_6]$ (aromatic



Figure S31. ¹H NMR (400 MHz) spectrum of complex [Zn₁₂LC₆].



Figure S32. ¹³C NMR (400 MHz) spectrum of complex [$Zn_{12}LC_6$].



Figure S33. 2D COSY NMR (400 MHz) spectrum of complex $[Zn_{12}LC_6]$.



Figure S34. 2D COSY NMR (400 MHz) spectrum of complex [Zn₁₂LC₆] (aromatic

region).



Figure S35. 2D NOESY NMR (400 MHz) spectrum of complex $[Zn_{12}LC_6]$ (aromatic



Figure S37. ¹³C NMR (400 MHz) spectrum of complex [Zn₁₂LD₆].



Figure S38. 2D COSY NMR (400 MHz) spectrum of complex [Zn₁₂LD₆].



Figure S39. 2D COSY NMR (400 MHz) spectrum of complex $[\mathbf{Z}n_{12}\mathbf{L}\mathbf{D}_6]$ (aromatic

region).



Figure S40. 2D NOESY NMR (400 MHz) spectrum of complex $[Zn_{12}LD_6]$ (aromatic

region).



9. ESI-MS of multiple macrocycles assembled by ditopic tpy ligand 6 with Zn^{2+} .

Figure S41. ESI-MS of multiple macrocycles assembled by ditopic tpy ligand 6 with Zn^{2+} .

10. STM images.



Figure S42. STM images of complex $[Zn_{12}LC_6]$ on HOPG.



Figure S43. STM images of complex $[Zn_{12}LD_6]$ on HOPG.

11. TEM images.



Figure S44. TEM images of complex $[Zn_{12}LB_6]$.



Figure S45. TEM images of complex $[Zn_{12}LC_6]$.

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