# Heterologous Fibrils Shows Distinctive Kinetic and Conformational Specificity.

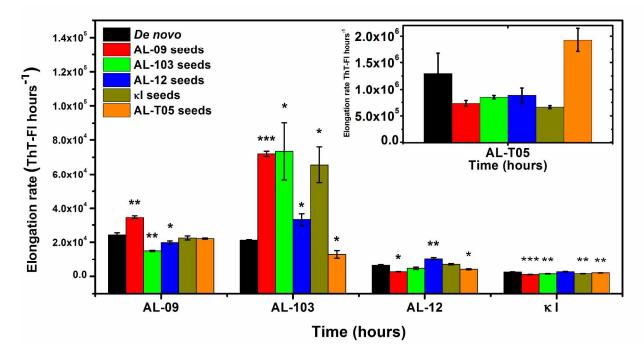
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KEYWORD: Seeding; fibril; monomer; nucleation; conformational specificity.

## Supporting Information for Publication

### **SUPPLEMENTAL FIGURE 1**



**FIGURE S1.** Effect of seeding on the rates of fibril elongation of V<sub>L</sub> proteins. Comparison of elongation rates as a function of the protein employed. Data are from fibril formation reactions conducted in triplicate. P-values where calculated with respect to *de novo* reaction.\*  $P \le 0.05$ ; \*\*  $P \le 0.01$ ; \*\*\*  $P \le 0.001$ .

#### **SUPPLEMENTAL FIGURE 2**

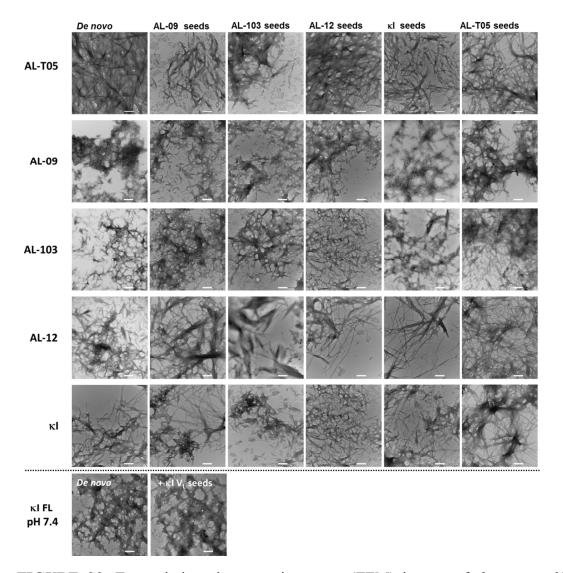
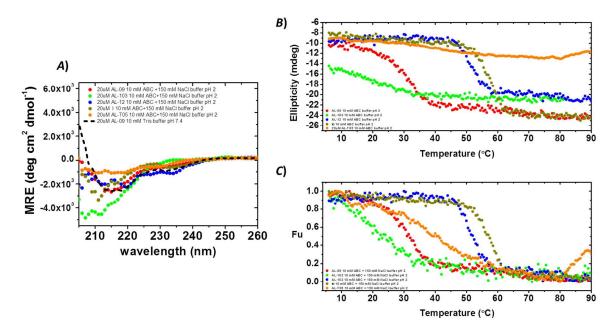


FIGURE S2. Transmission electron microscopy (TEM) images of *de novo*, self and crossseeding experiments at the endpoint of the reaction. All reactions were performed with 20  $\mu$ M protein and in presence of 1% of seeds. AL-09 Scale bar represent 200 nm.

#### **SUPPLEMENTAL FIGURE 3**



**FIGURE S3.** A) Far UV-CD spectra, (B) raw and (c) normalized thermal unfolding data of AL-09 (red), AL-103 (green), AL-12 (blue), AL-T05 (orange),  $\kappa I$  (green olive). 20  $\mu M$  protein samples were prepared in 10 mM ABC, pH 2.0. Far UV-CD spectra were acquired at 4°C. Thermal denaturation experiments were performed from 4–90°C at a rate of 0.5°C min<sup>-1</sup>.

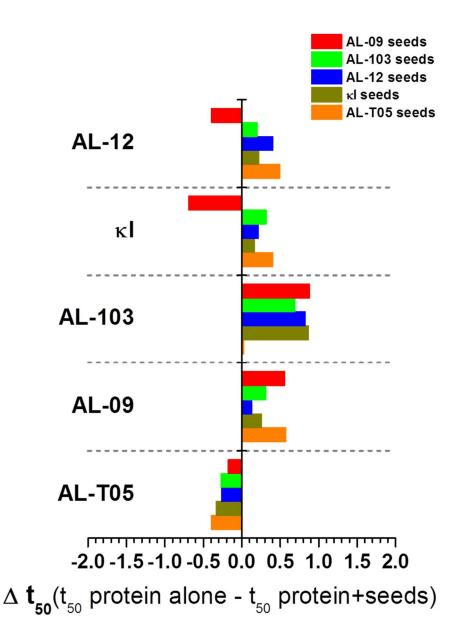


FIGURE S4. Comparison of normalized  $\Delta t_{50}$  values, as a function of the seed employed. Data are from fibril formation reactions conducted in triplicate. Reaction was considered positive when ThT fluorescence increased four-fold (>200,000 A.U). All proteins tested were able to form fibrils at pH 2.0.