A New Route to 2'-O-Alkyl-2-thiouridine Derivatives *via* 4-O-Protection of the Uracil Base and Hybridization Properties of Oligonucleotides Incorporating These Modified Nucleoside Derivatives

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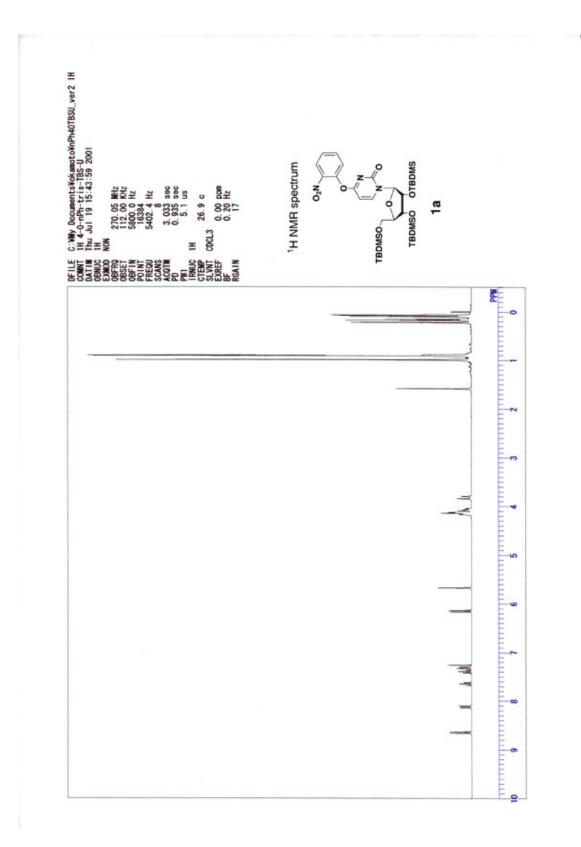
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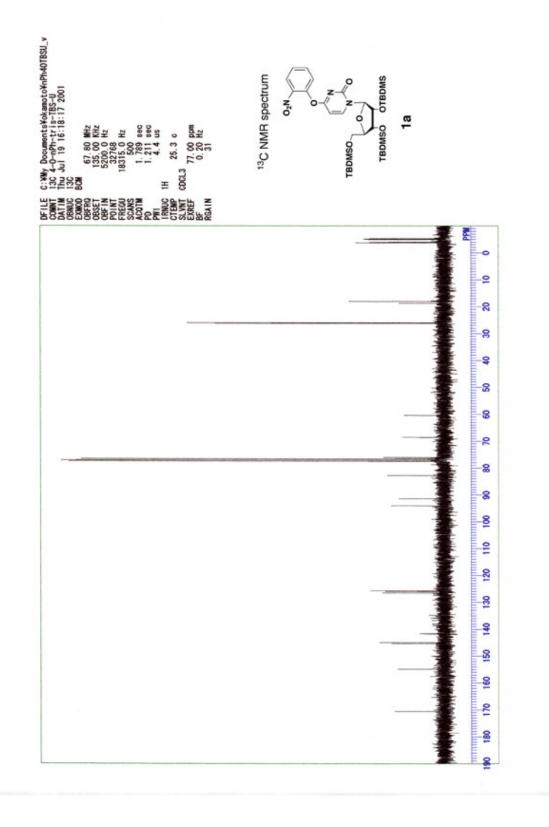
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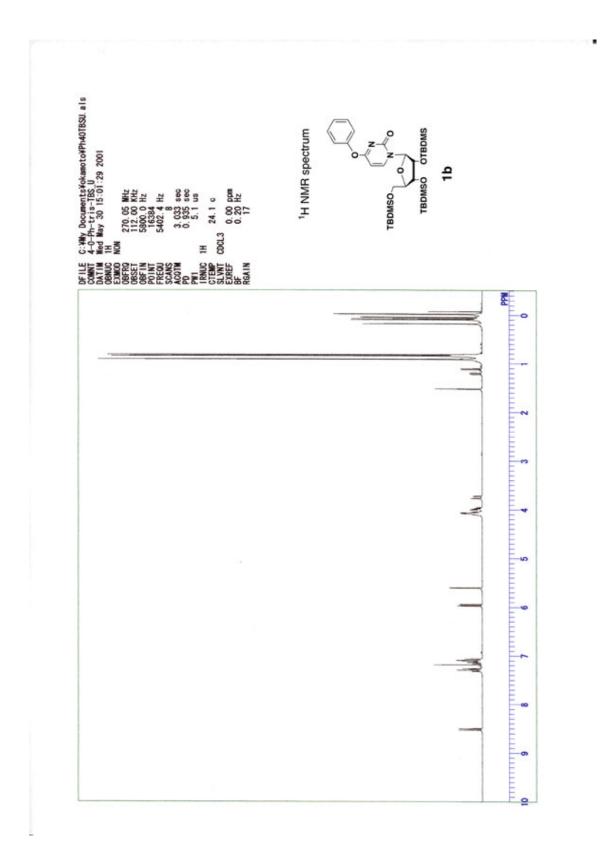
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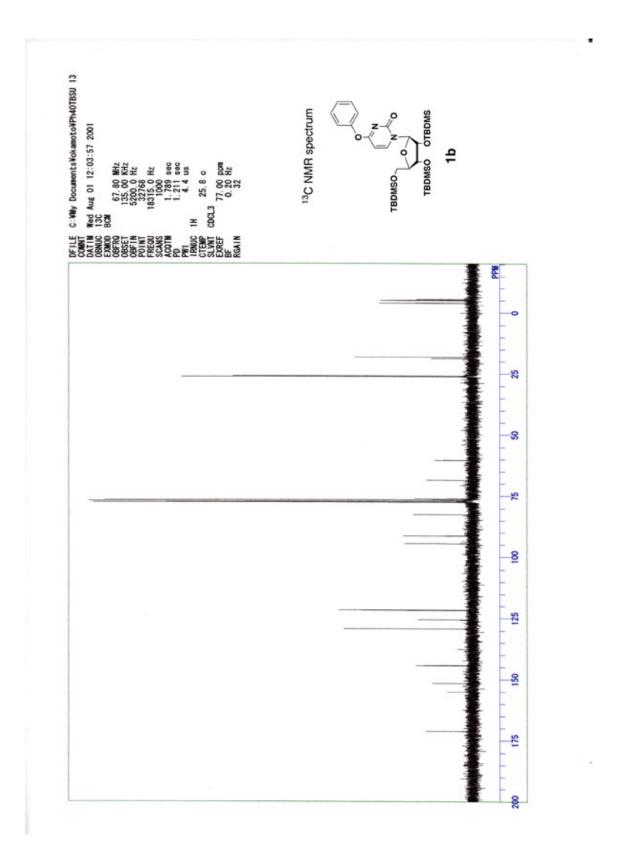
General. ¹H NMR spectra were recorded at 270 MHz and the chemical shifts were measured from tetramethylsilane (in CDCl₃) as an internal standard or DSS (in D₂O) as an external standard. ¹³C NMR spectra were recorded at 68 MHz and the chemical shifts were measured from $CDCl_3$ (77 ppm) as an internal standard or DSS (0 ppm, in D₂O) as the external standard. ³¹P NMR spectra were recorded at 109 MHz and the chemical shifts were measured from 85% H_3PO_4 as an external standard. The ¹H-¹H coupling constants were measured at 400 MHz at 25 °C. Pyridine was distilled twice from *p*-toluenesulfonyl chloride and CaH₂ after being refluxed for several hours and stored over molecular sieves The other dry solvents were purchased and stored over molecular sieves 4A. TLC 4A. was performed on precoated glass plates of silica gel. Column chromatography was carried out with silica gel C-200 and C-60. The solid-phase synthesis of oligonucleotides was carried out on a DNA/RNA synthesizer model 392 using normal phosphoramidite protocol. Reversed-phase HPLC was performed using a C18 column with a linear gradient (0-30%) starting from 0.1 M NH₄OAc (pH 7.0) and applying CH₃CN at a flow rate of 1.0 mL/min for 30 min at room temperature. Anion-exchange HPLC was performed on an HPLC apparatus using a FAX column with a linear gradient (0-25%) starting from 25 mM phosphate buffer (pH 6.0) and applying 25 mM phosphate buffer containing 1 M NaCl (pH 6.0) at a flow rate of 1.0 mL/min at 50 °C. The MALDI-TOF mass spectrometry was carried out.

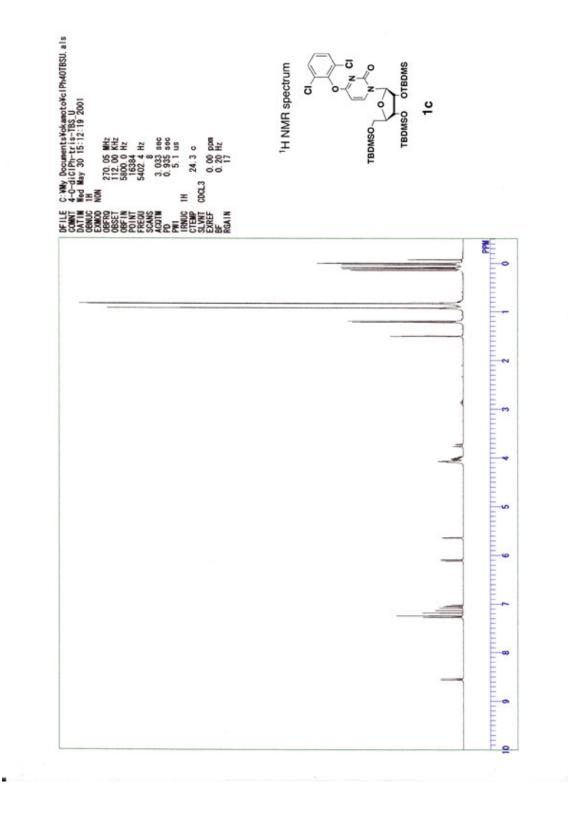


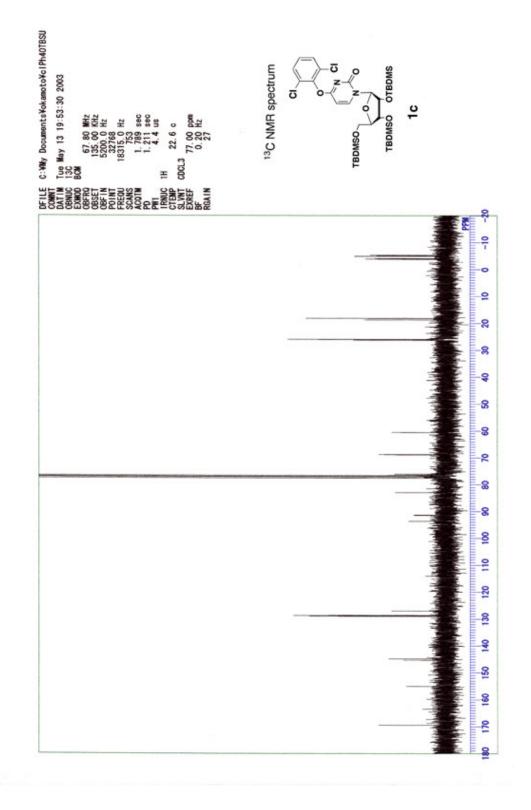


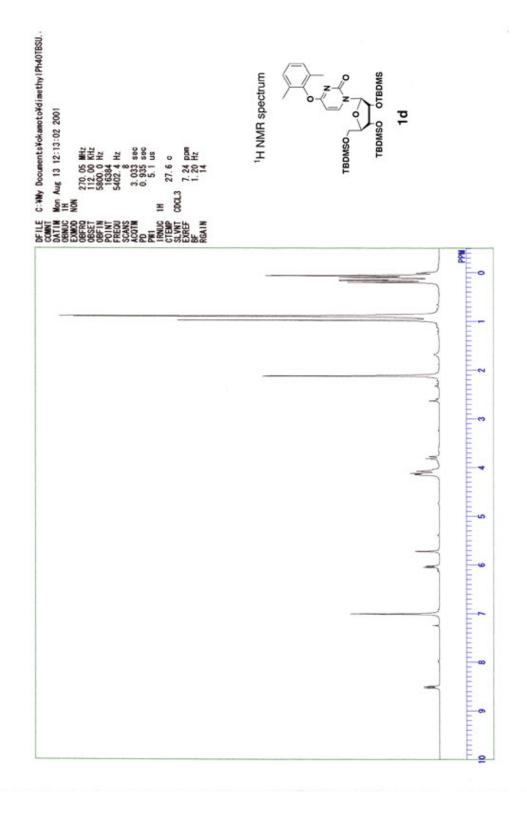
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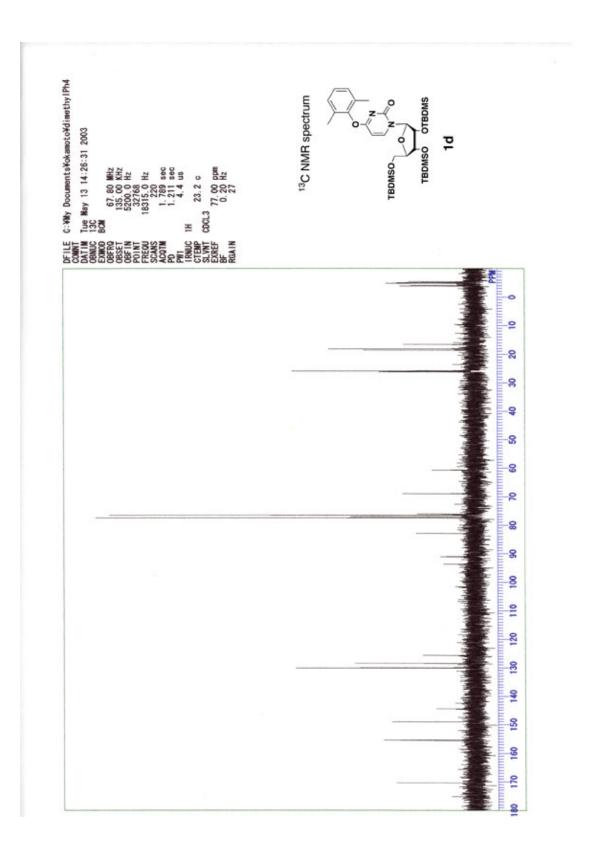


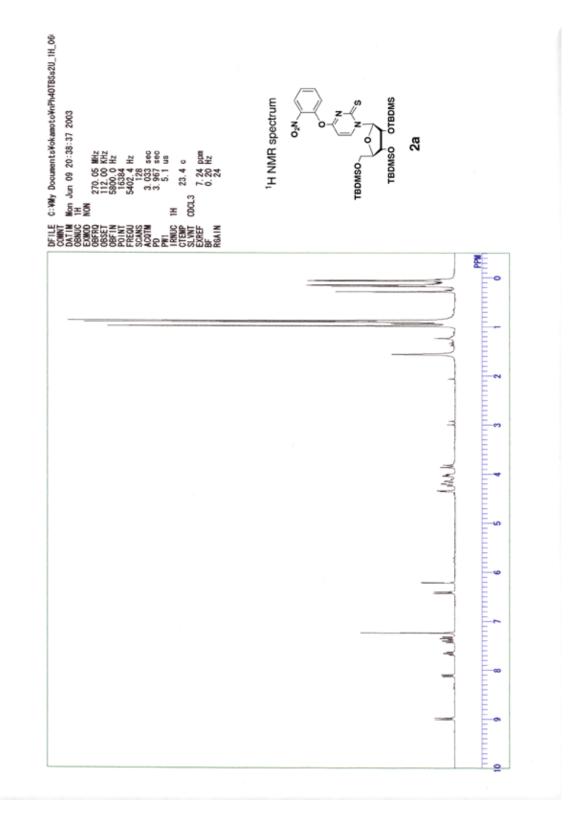


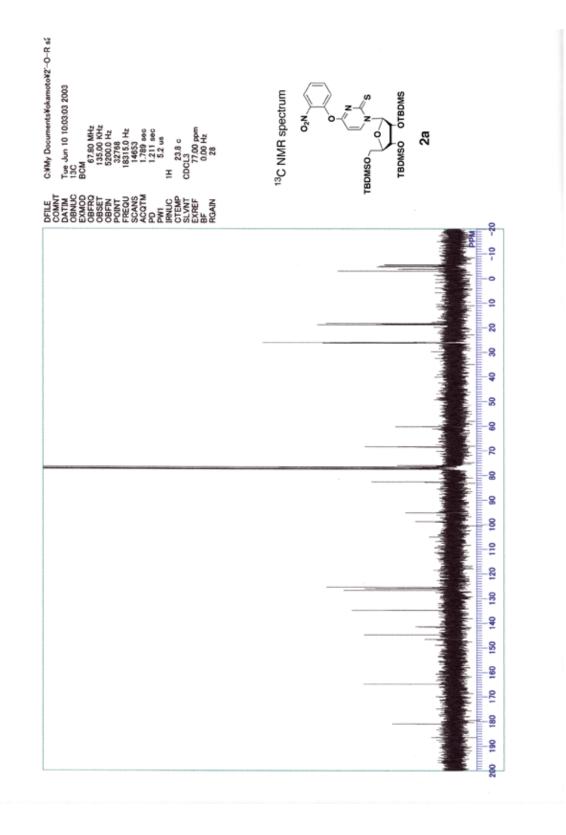


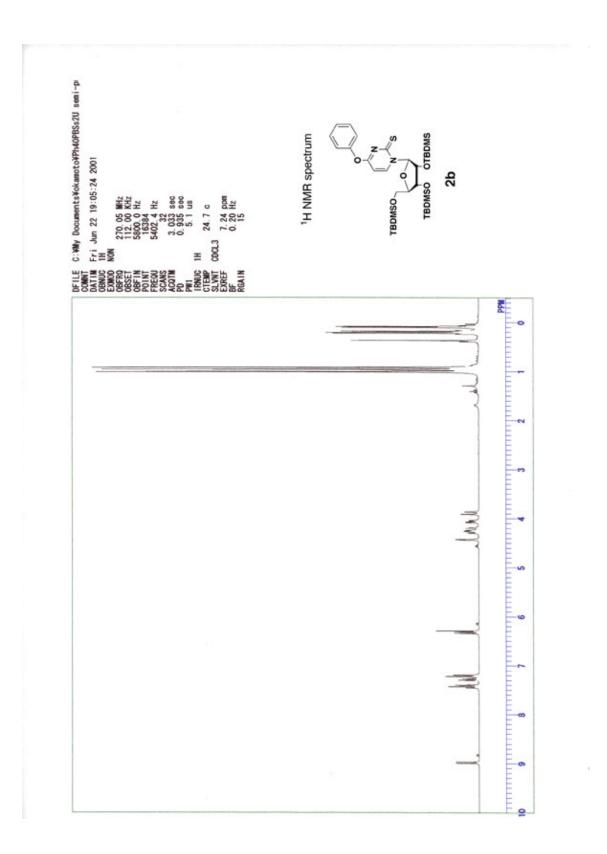


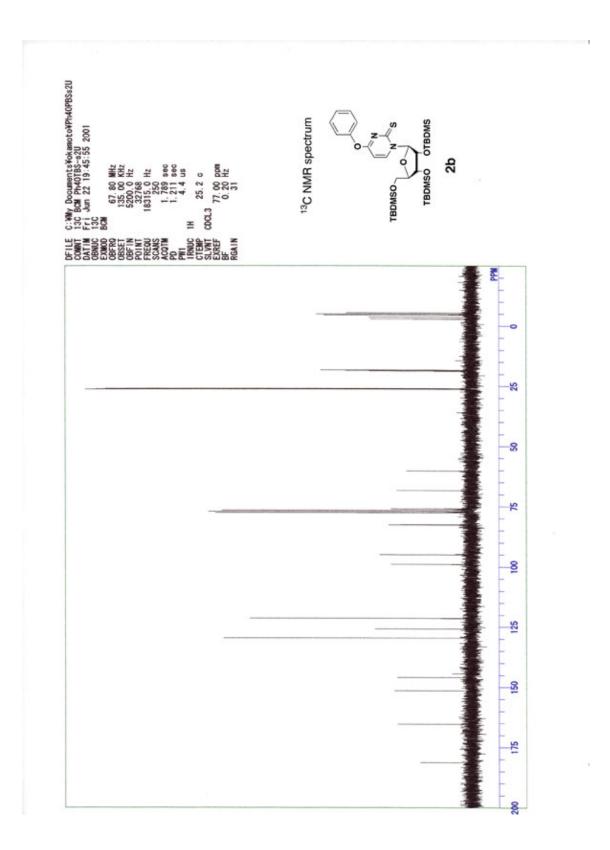
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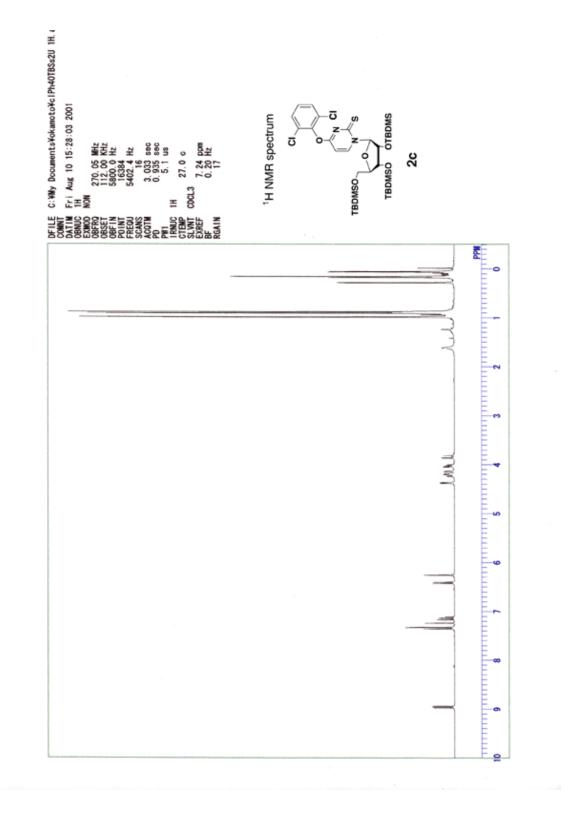


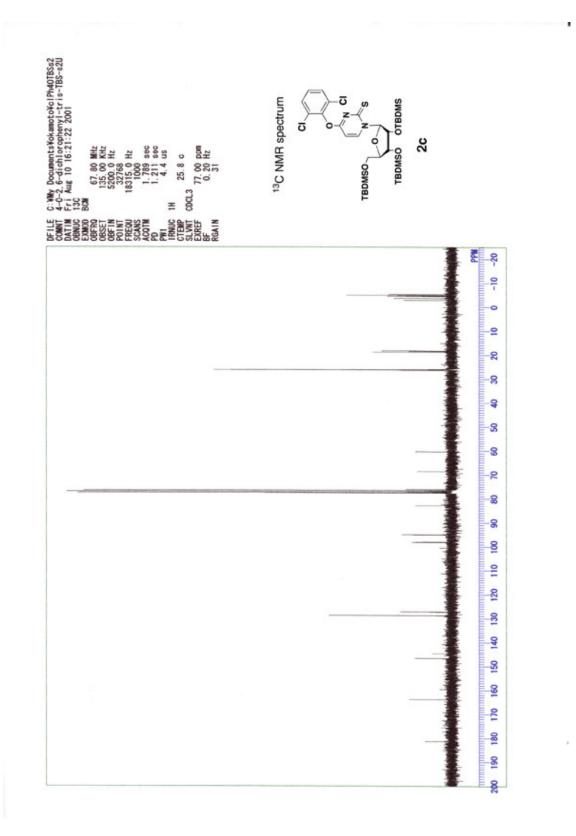


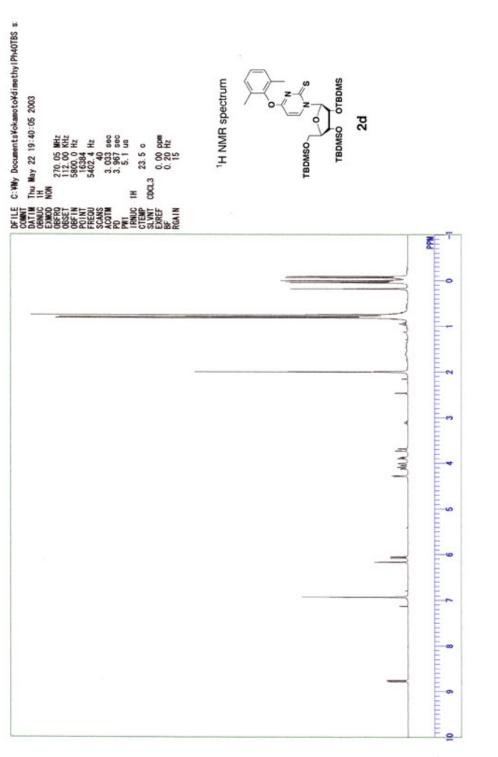


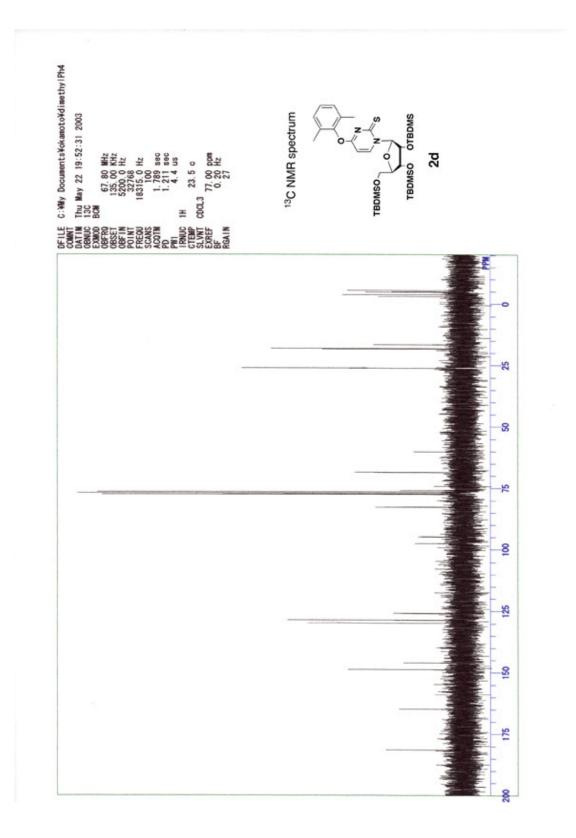


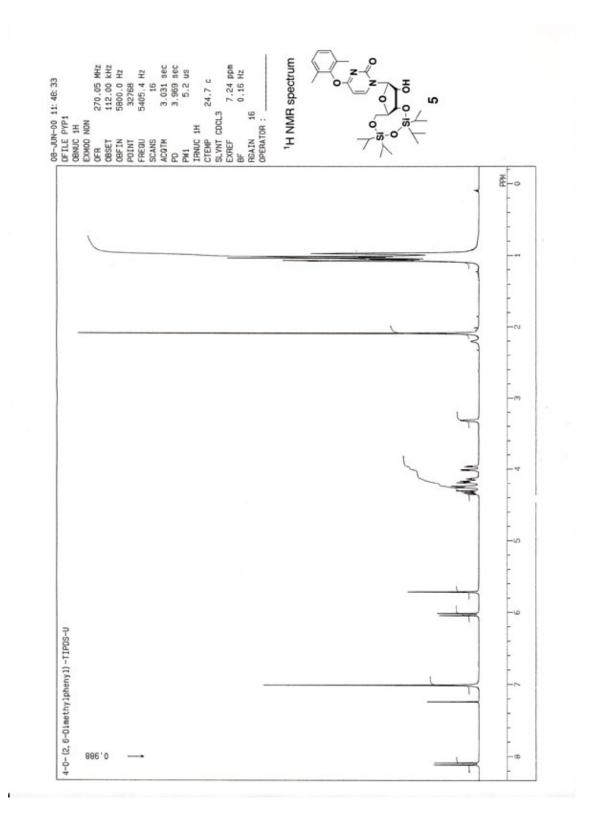


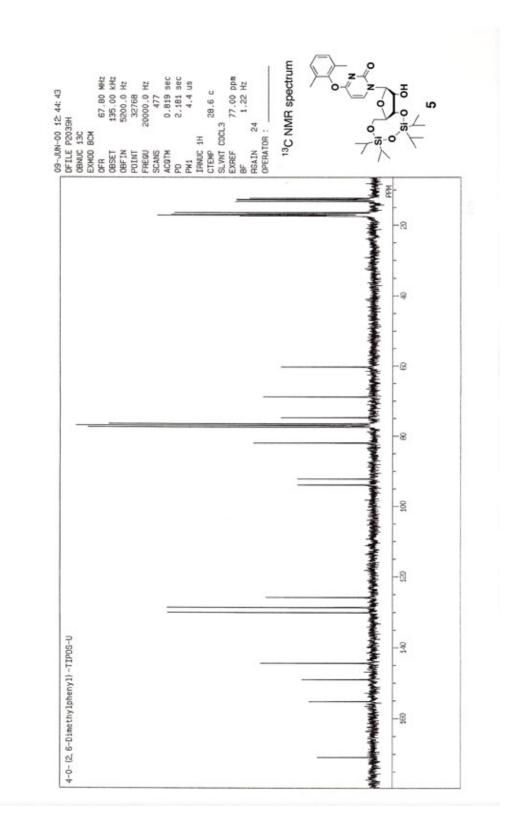


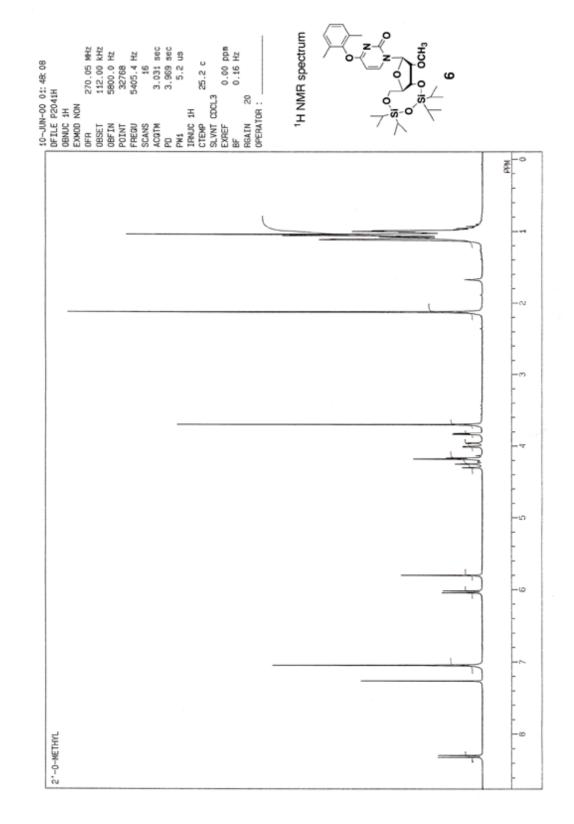


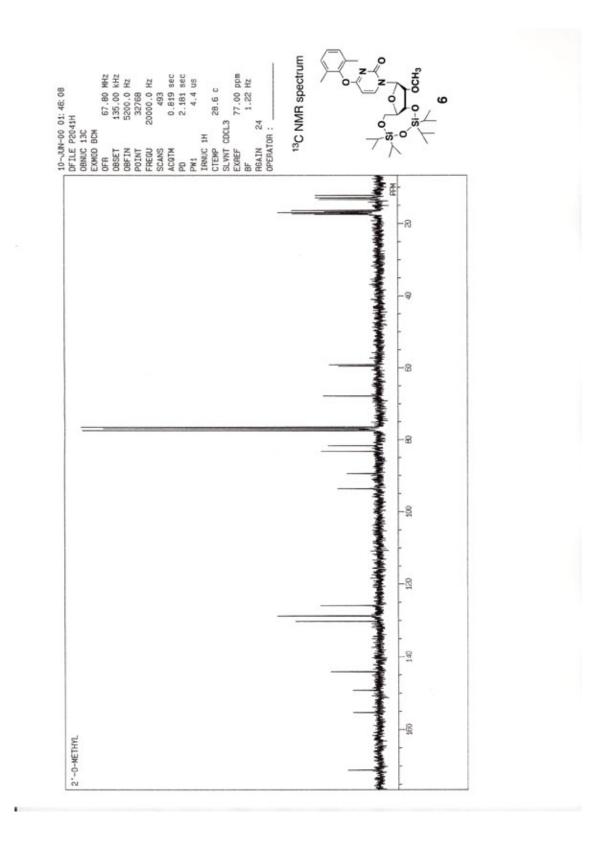


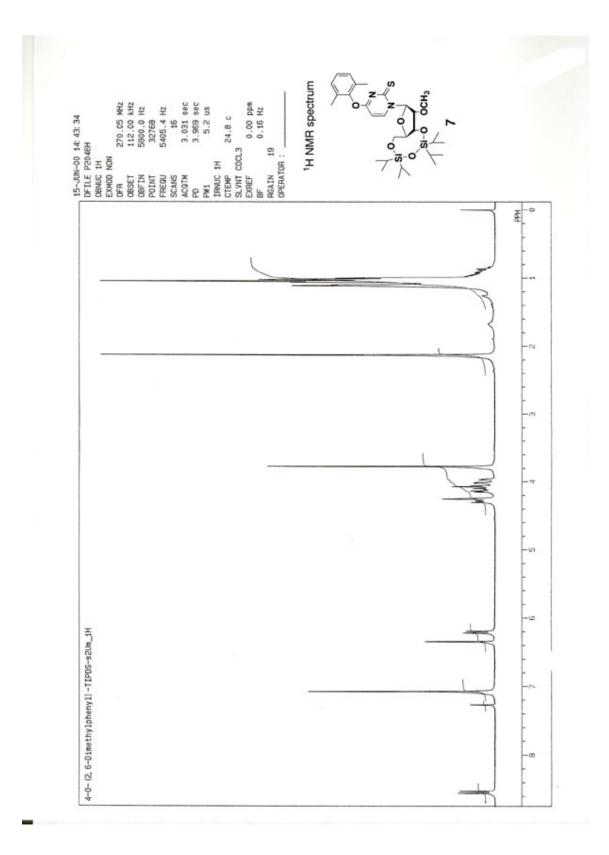


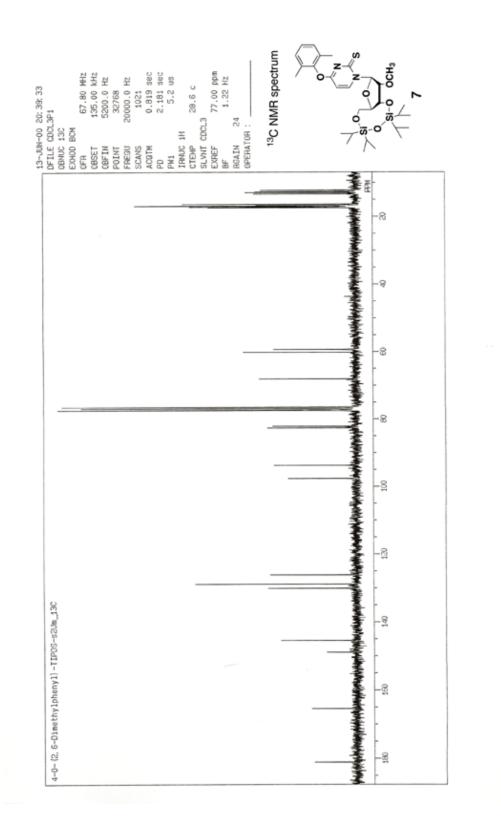


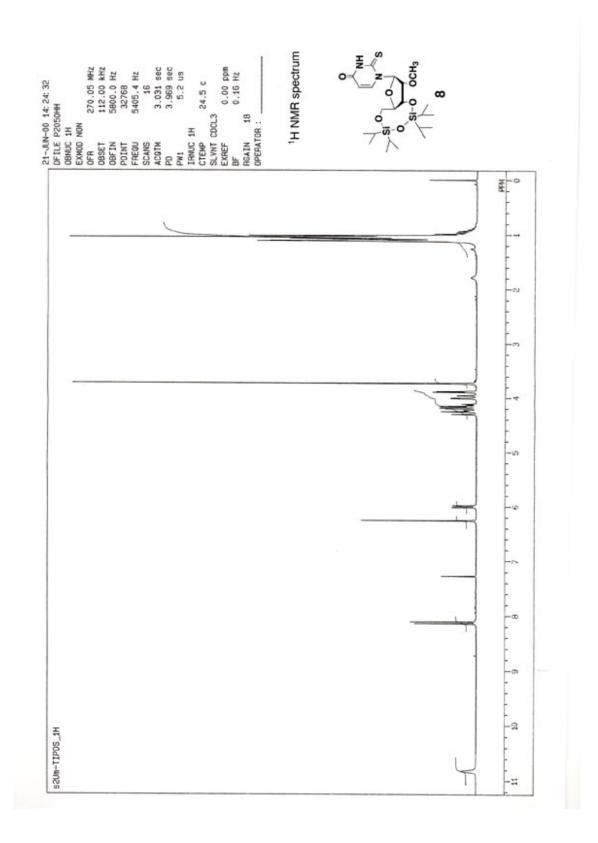


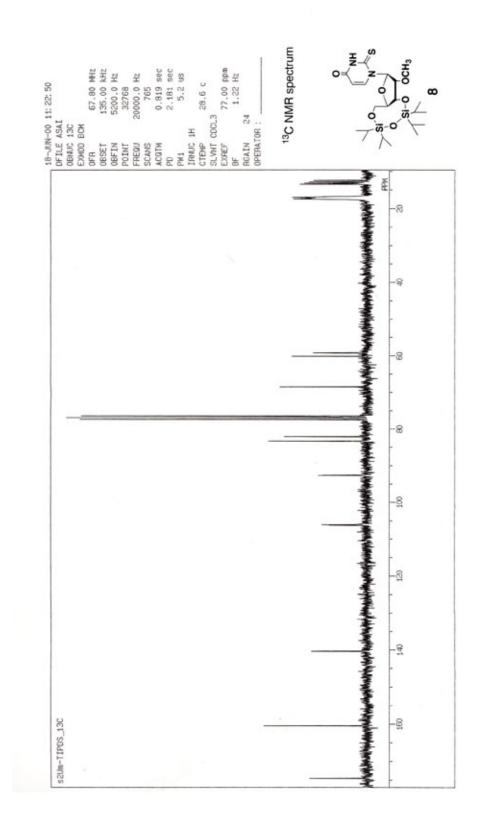


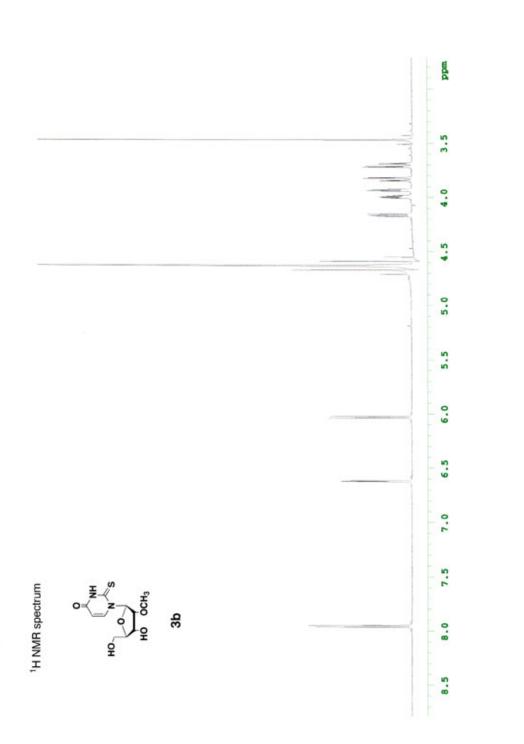


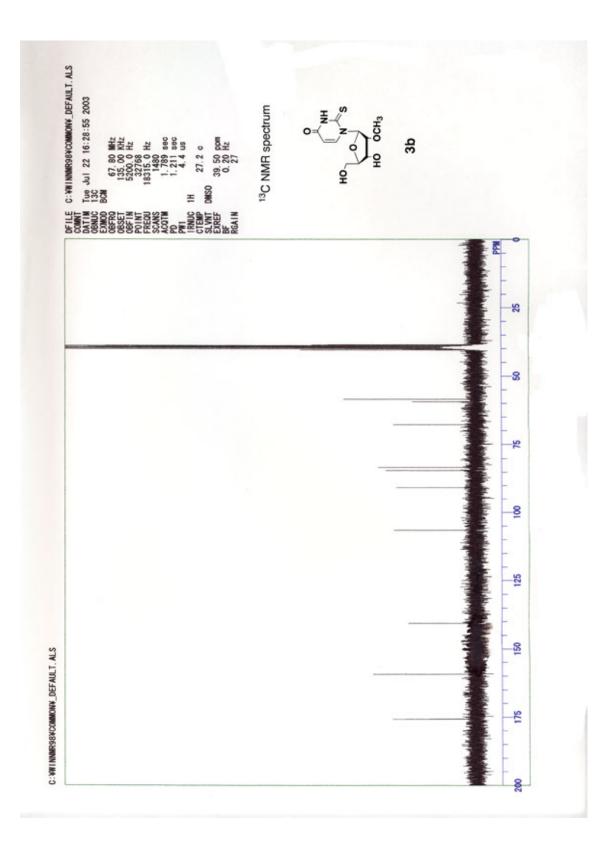




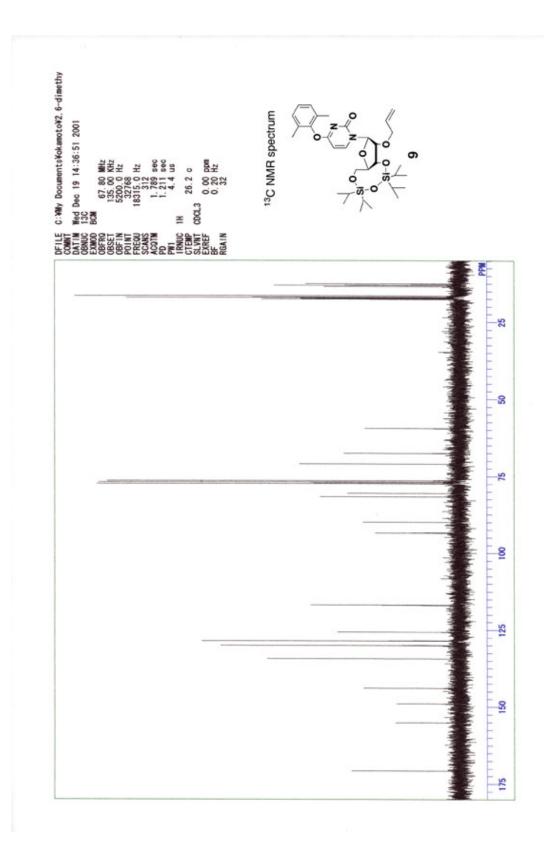


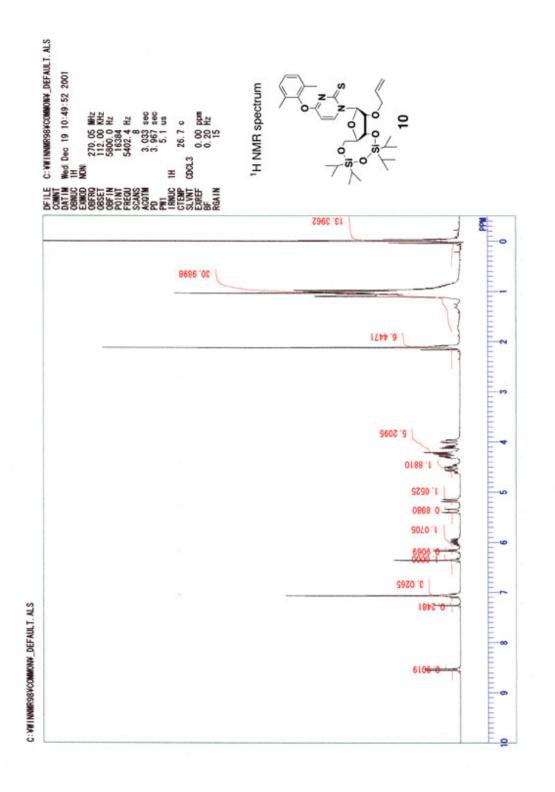


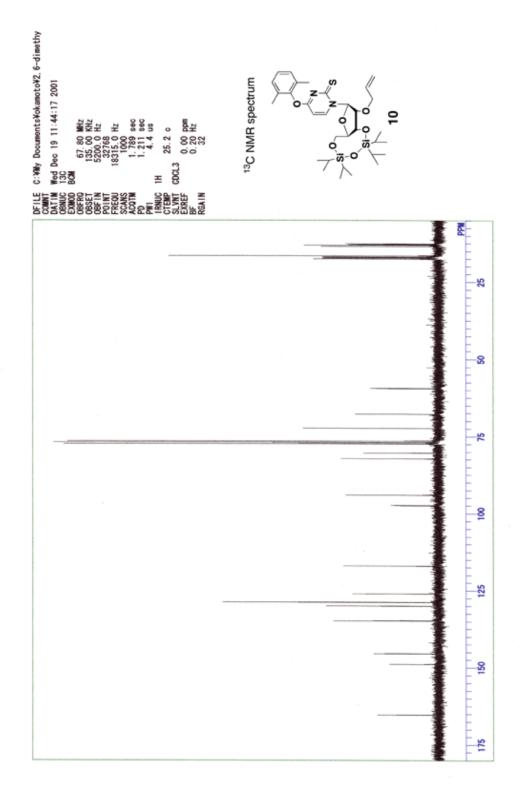


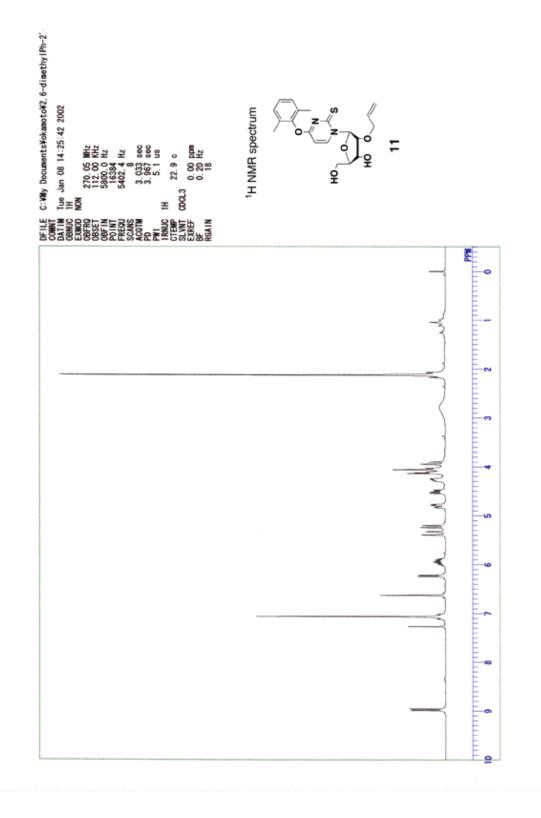


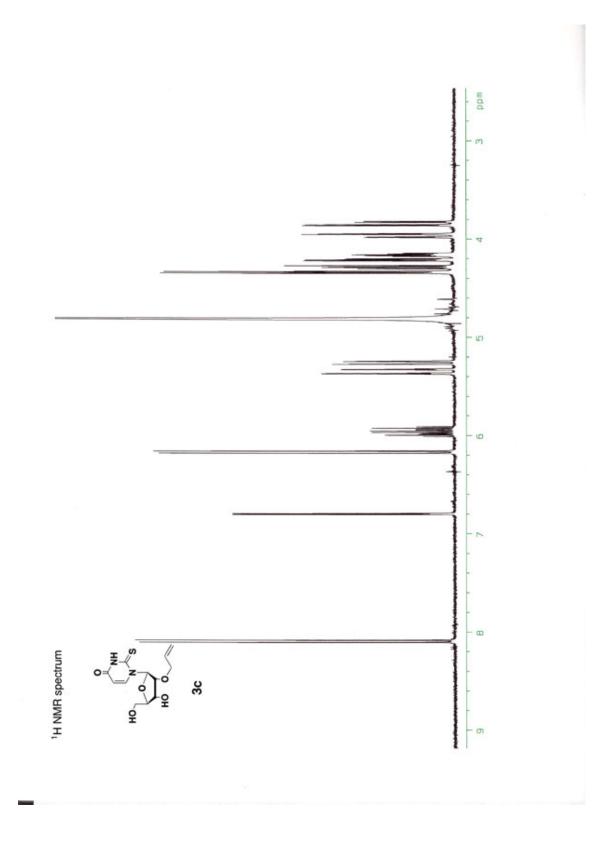


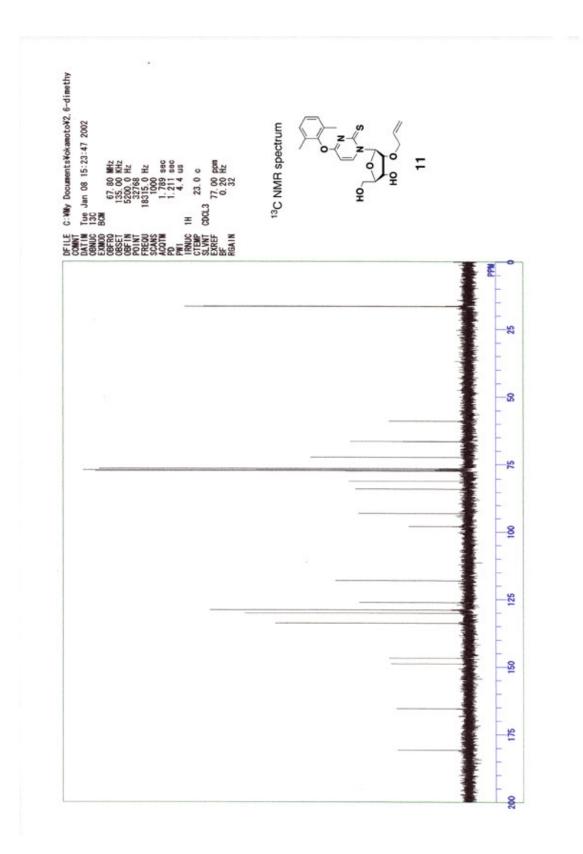


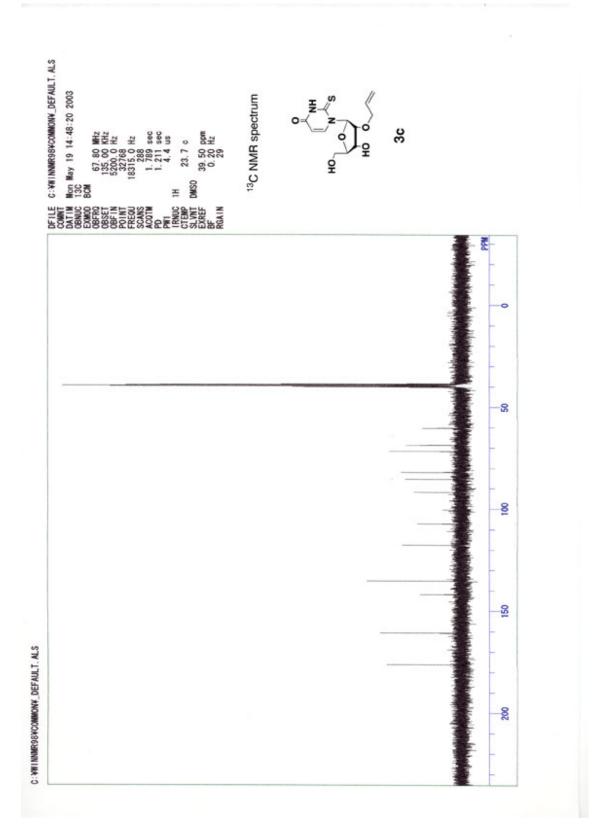


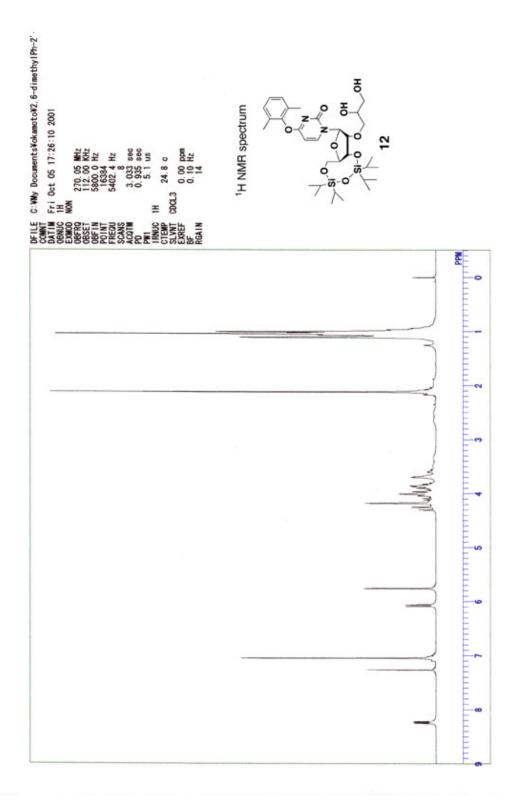




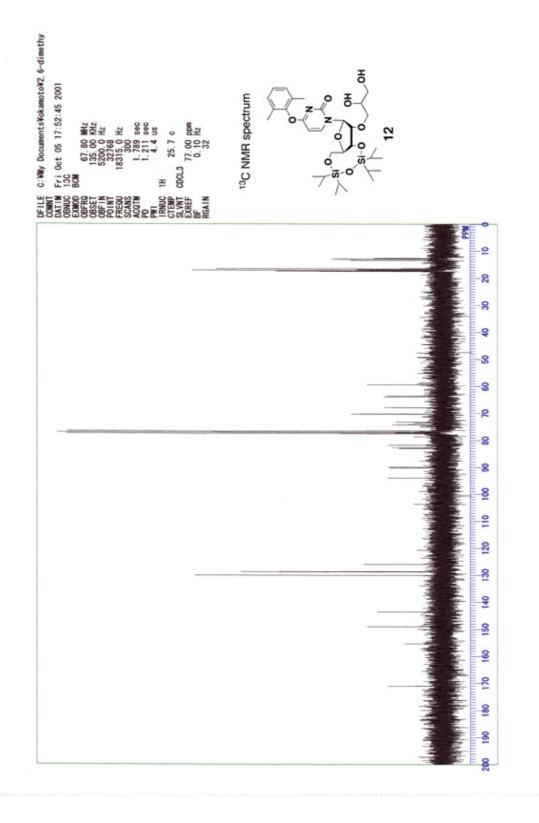


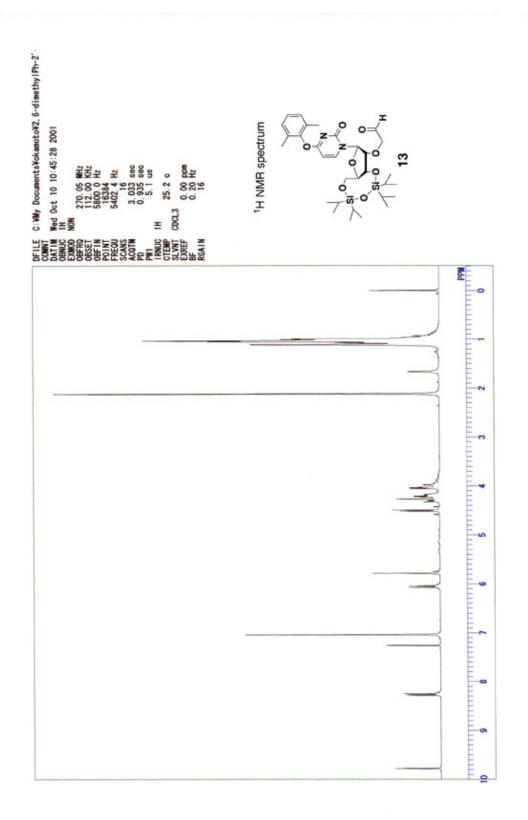


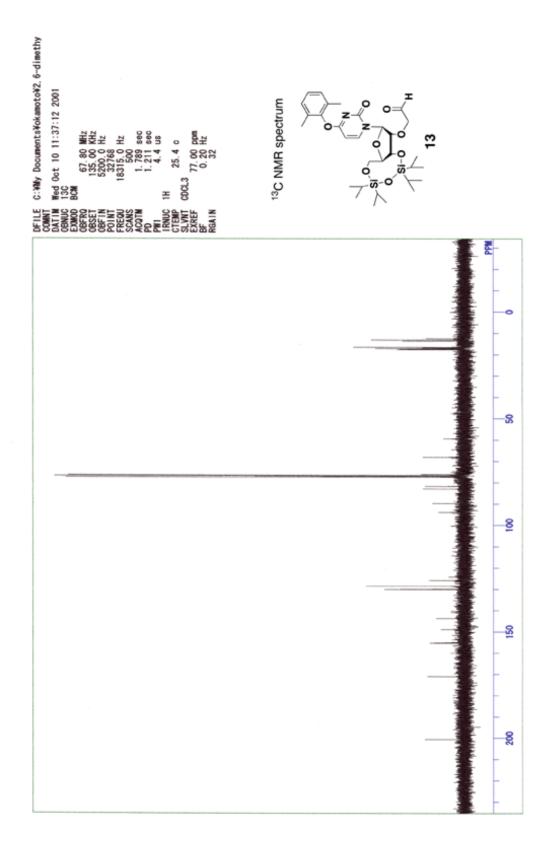


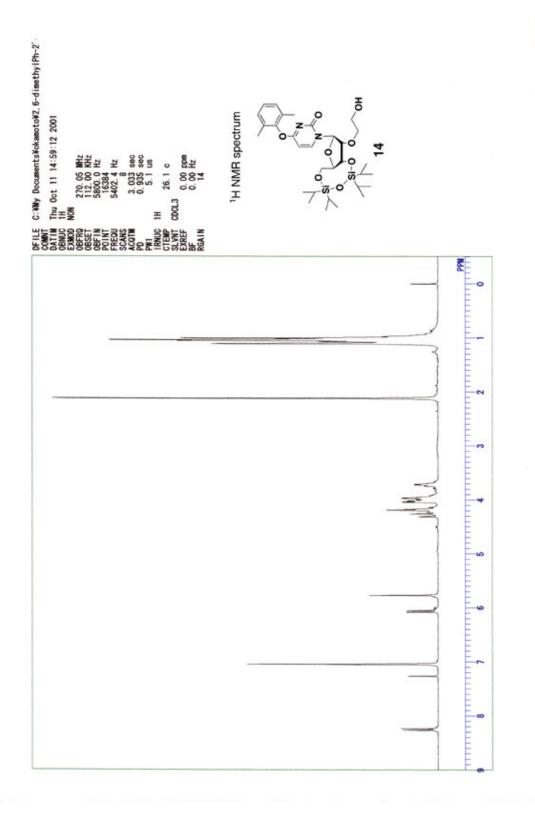


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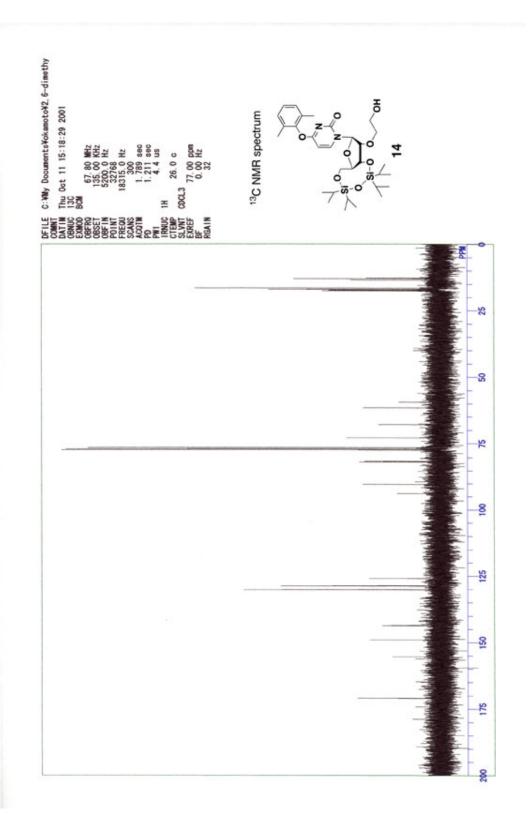


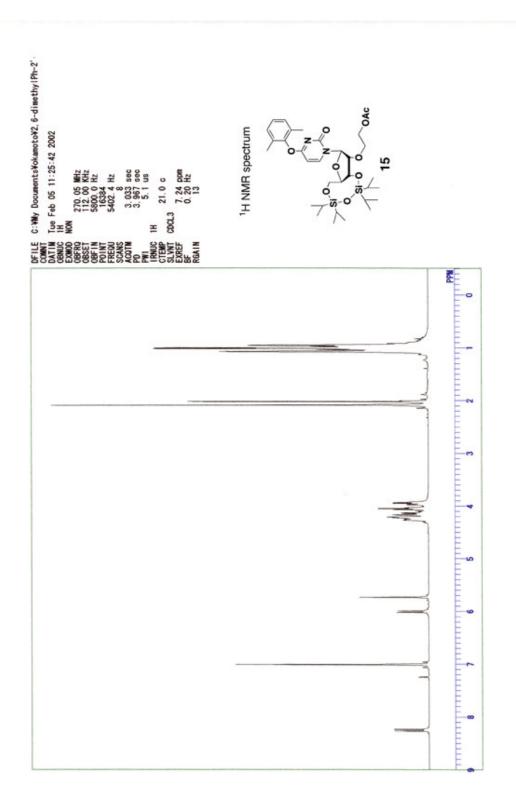


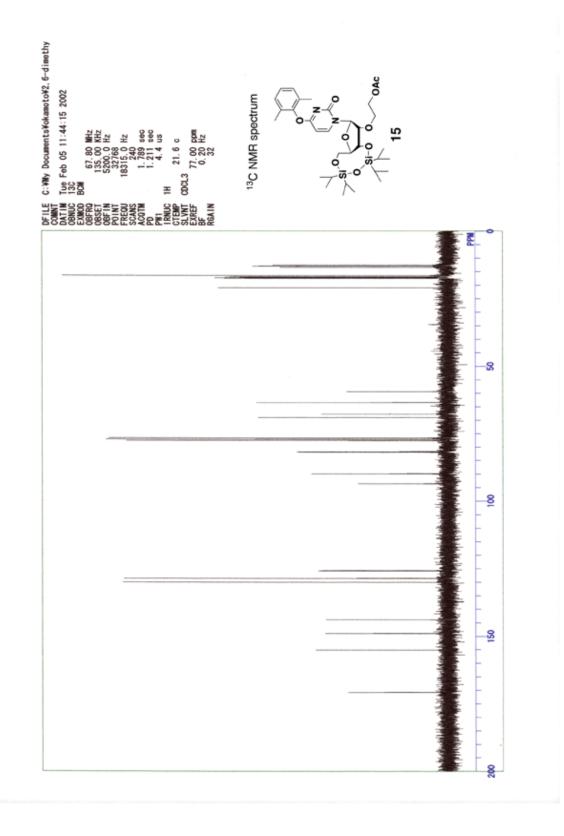


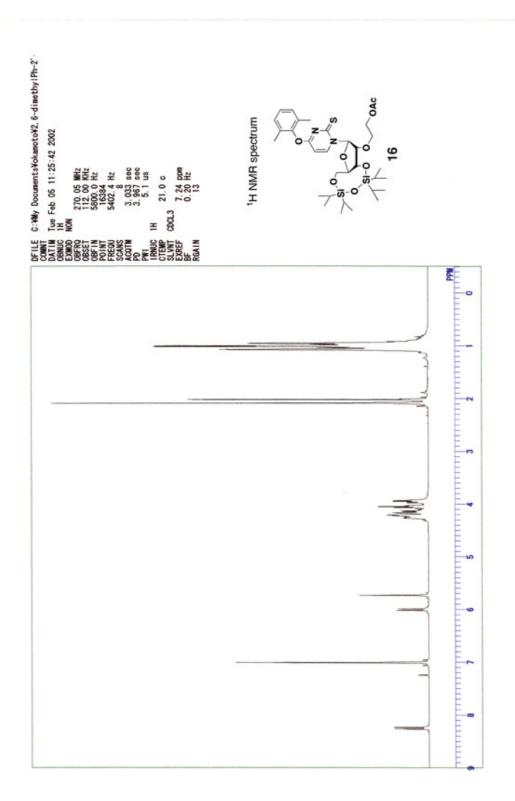


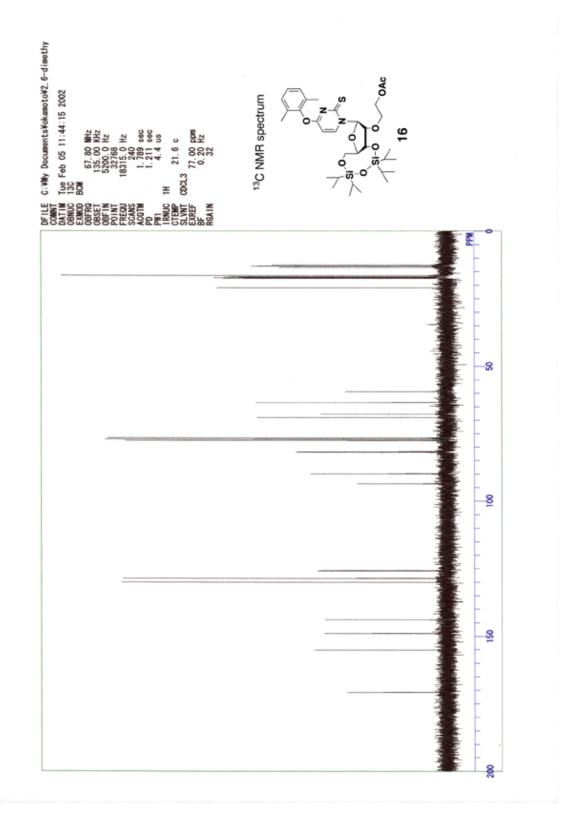
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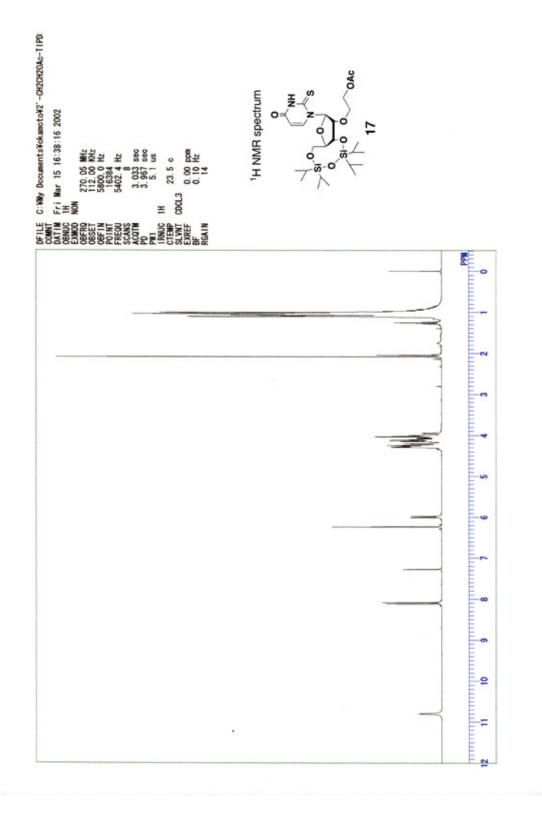


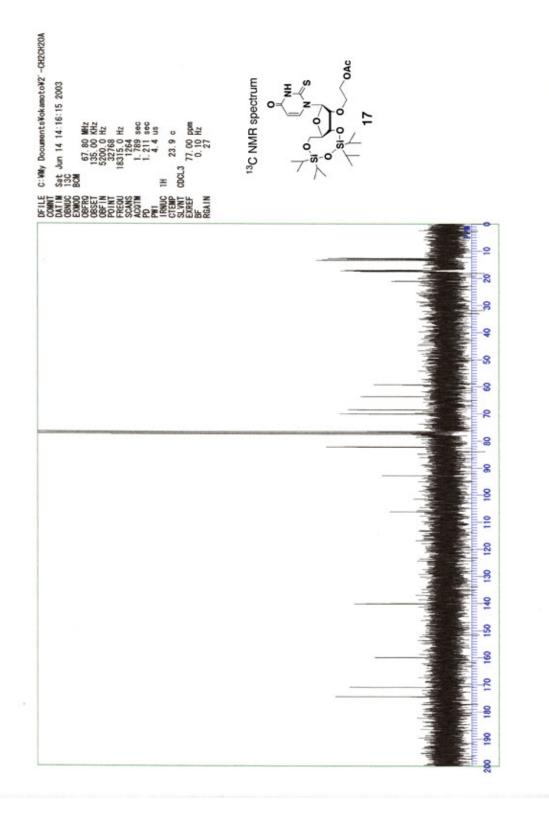




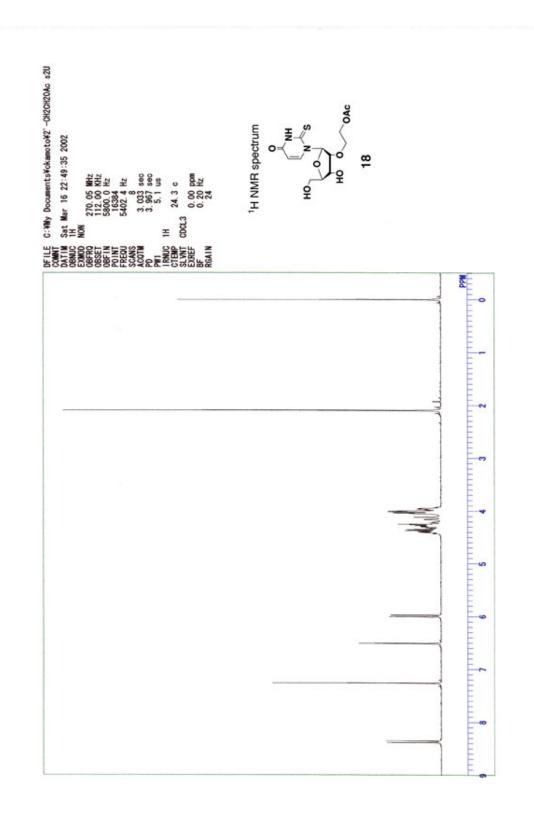


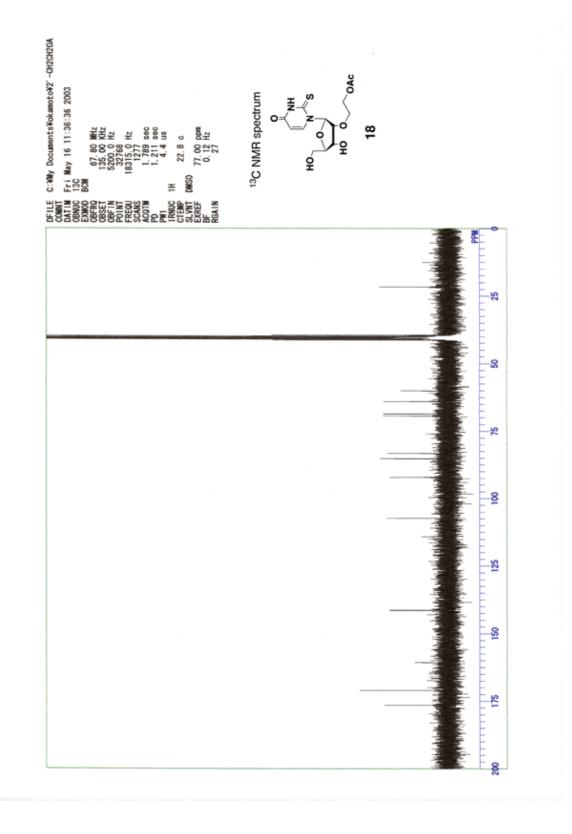


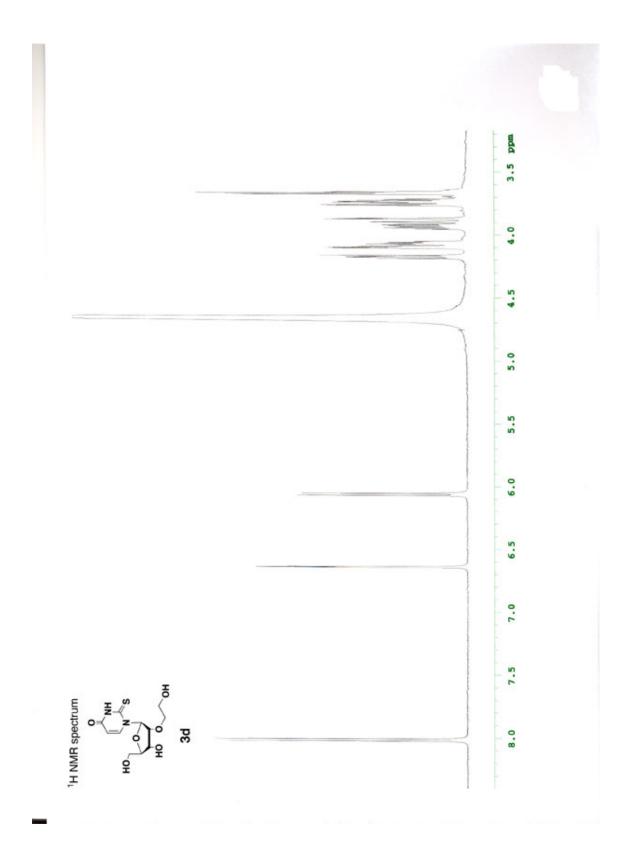


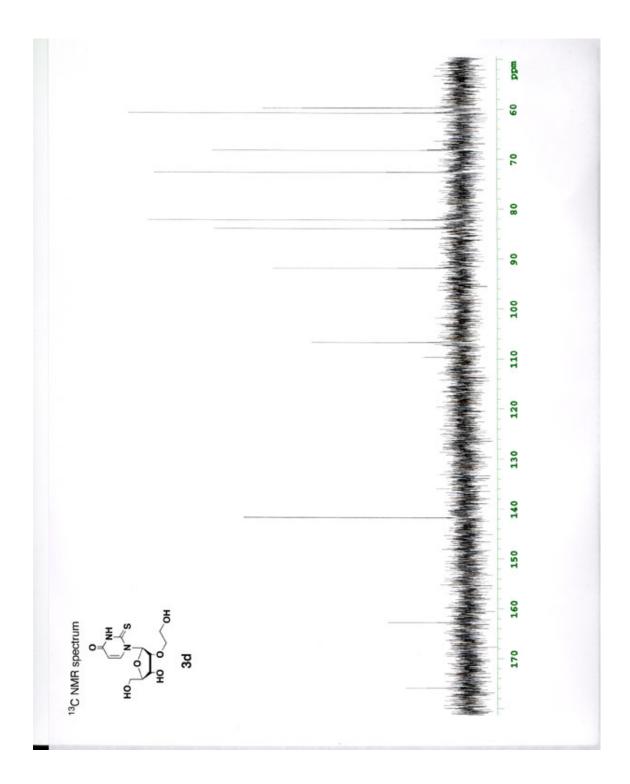


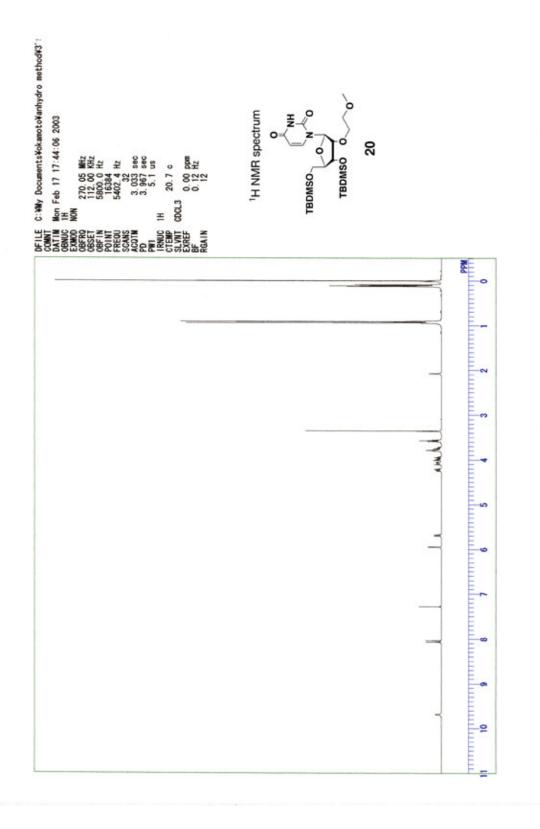
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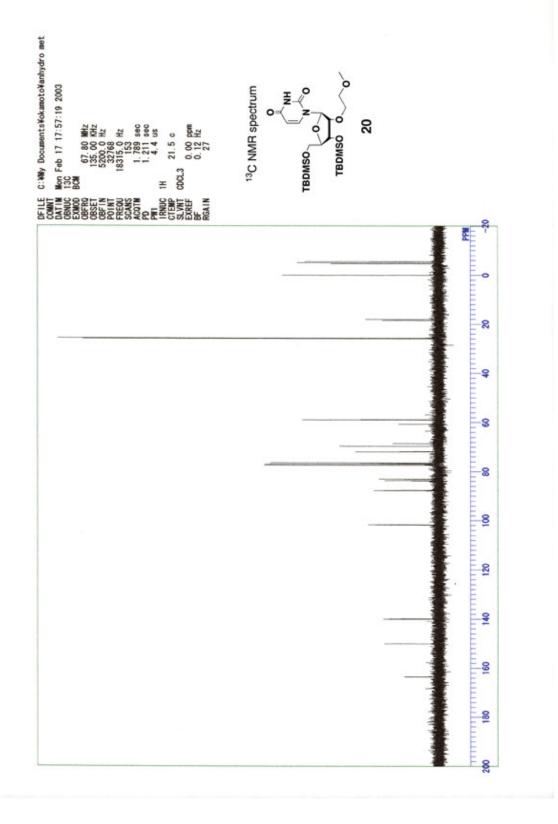




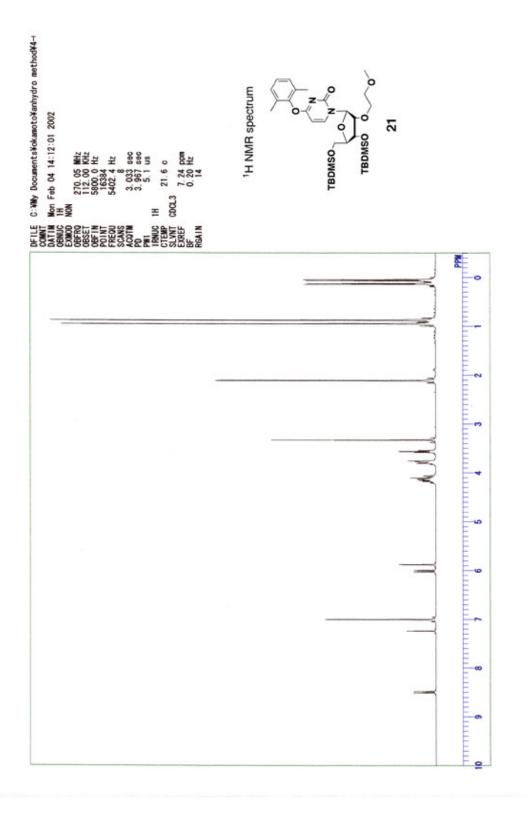


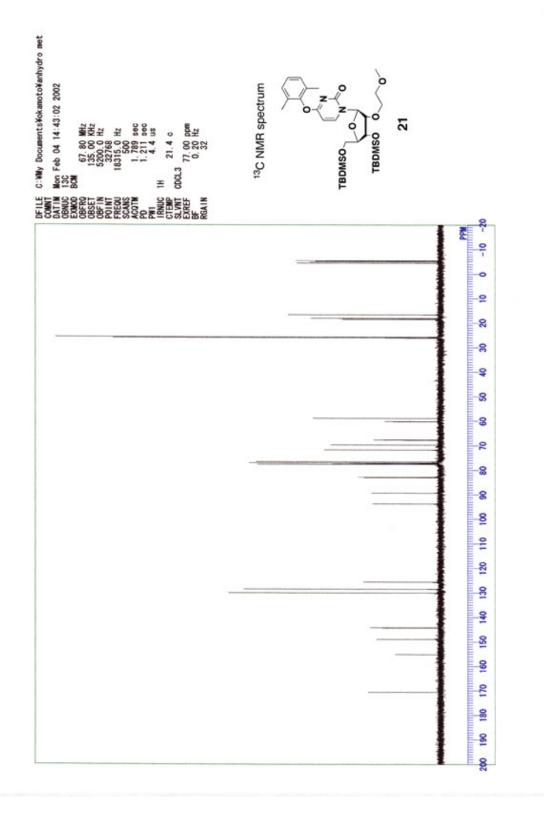


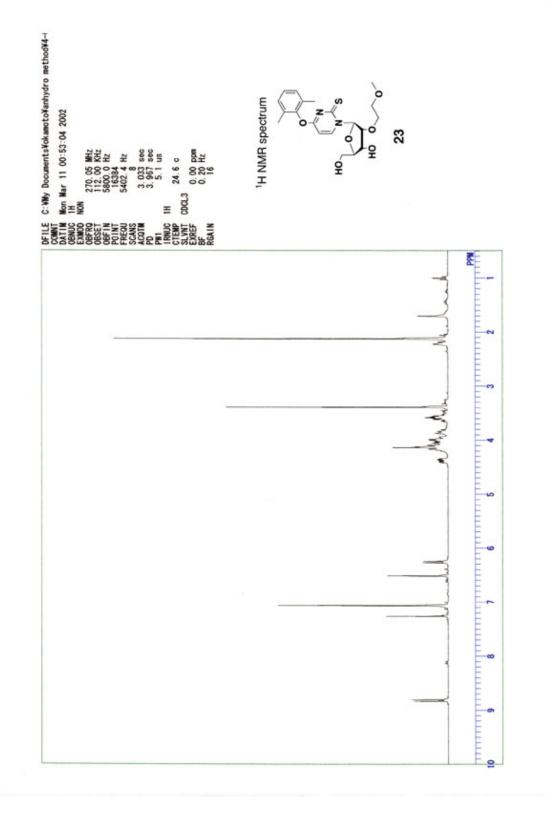




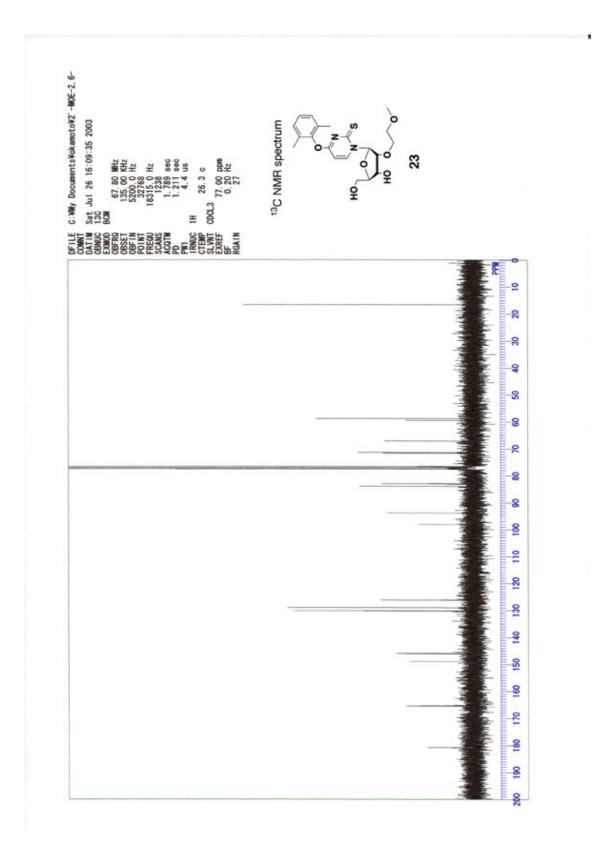
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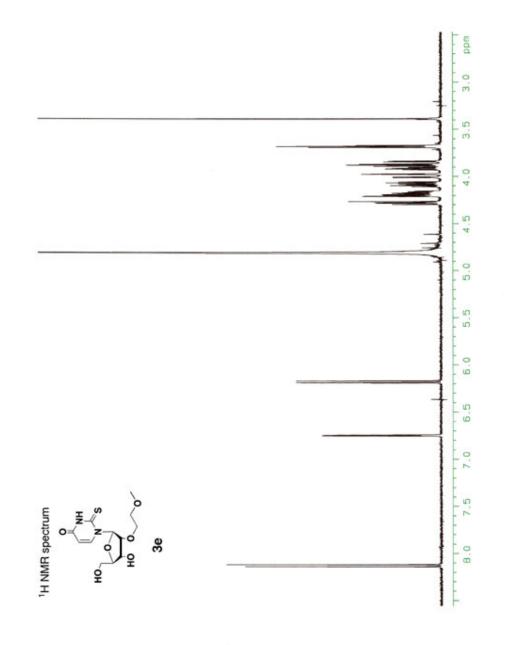




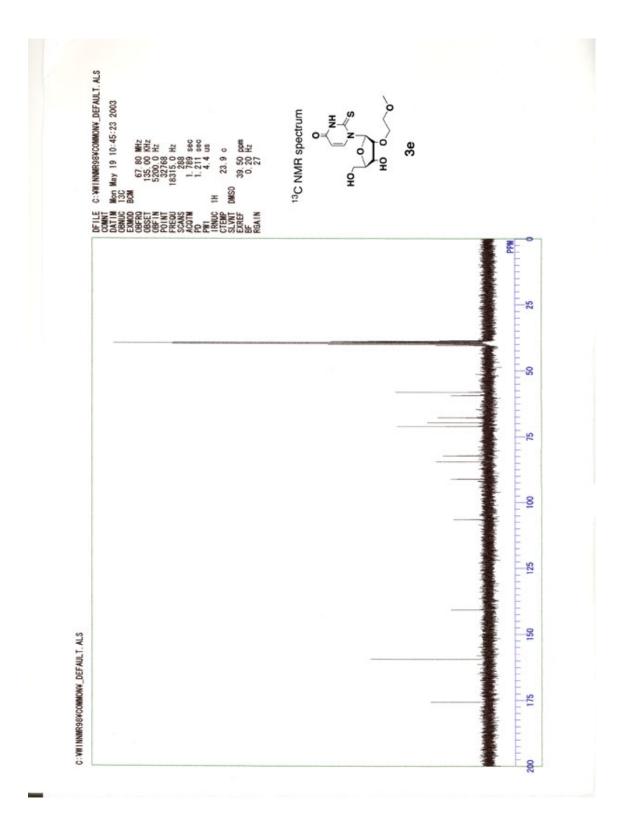


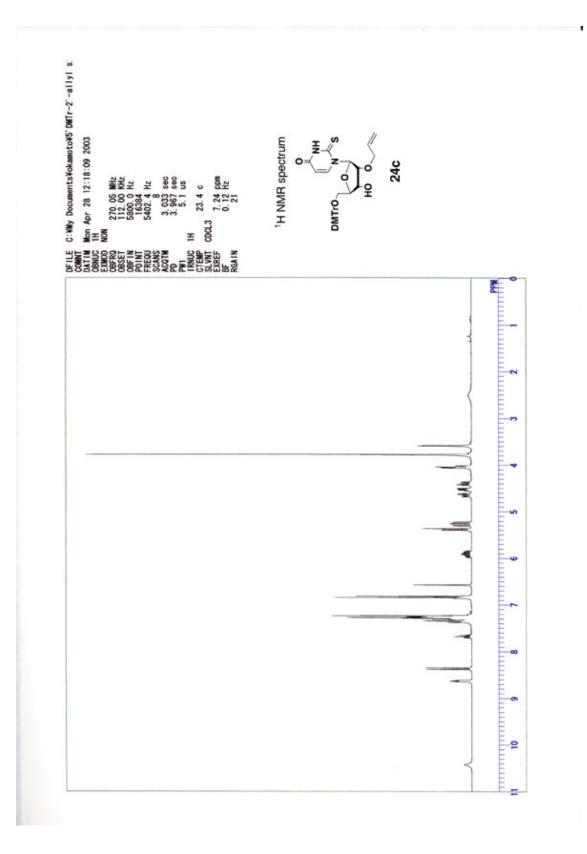
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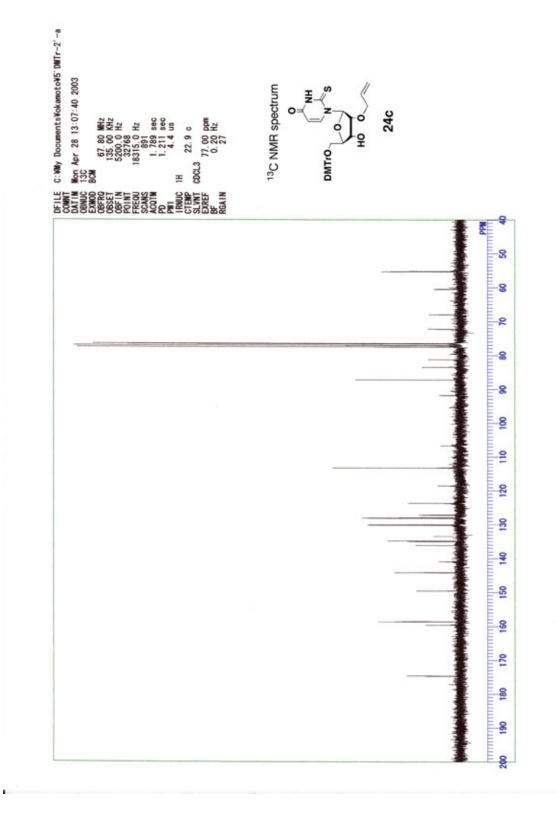


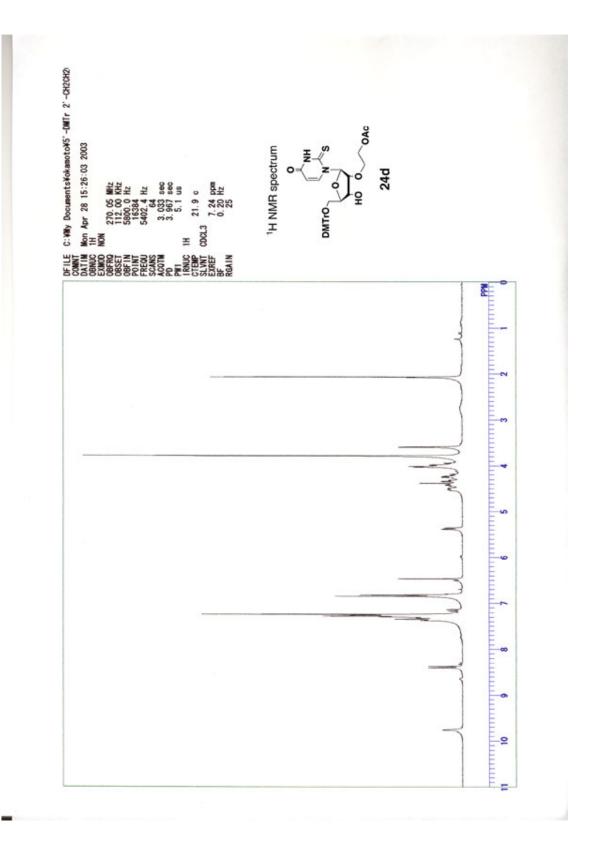


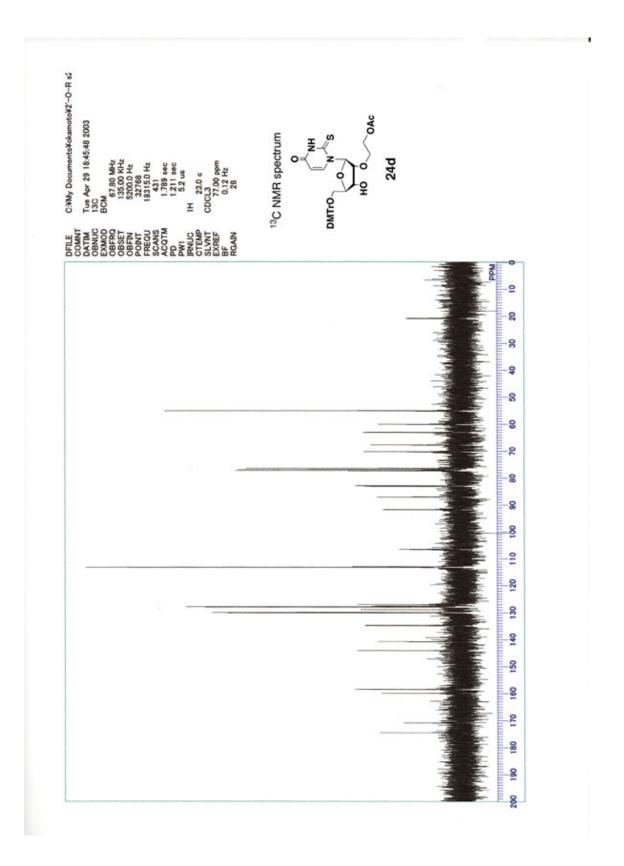
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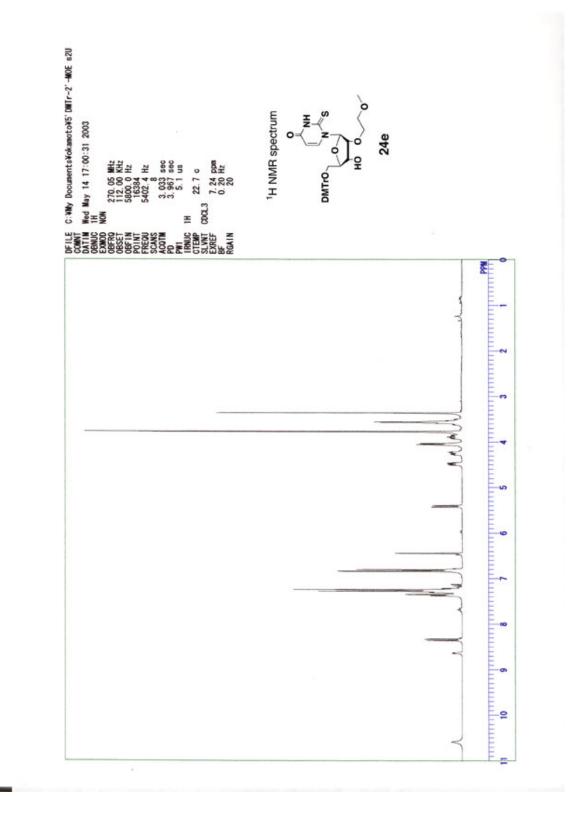


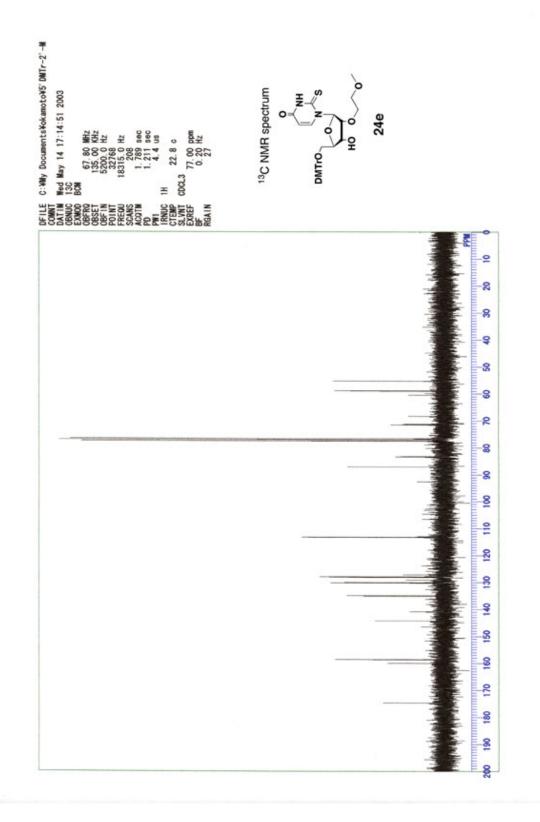


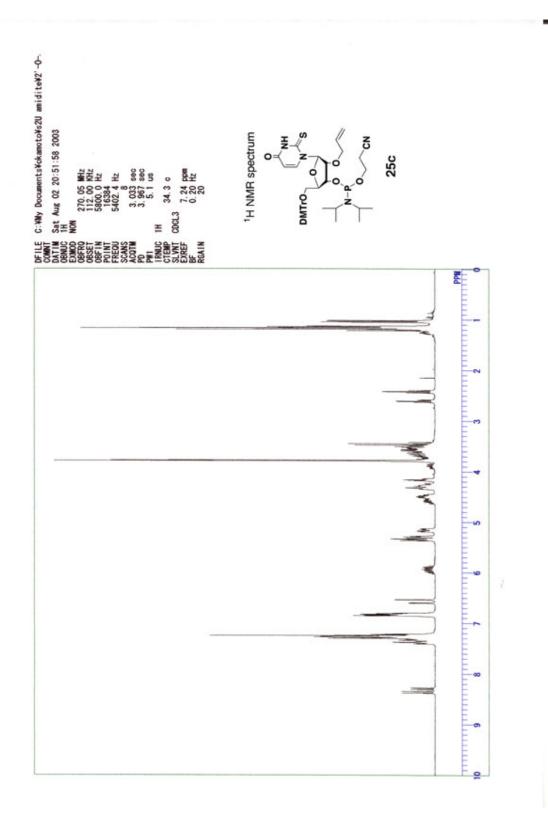


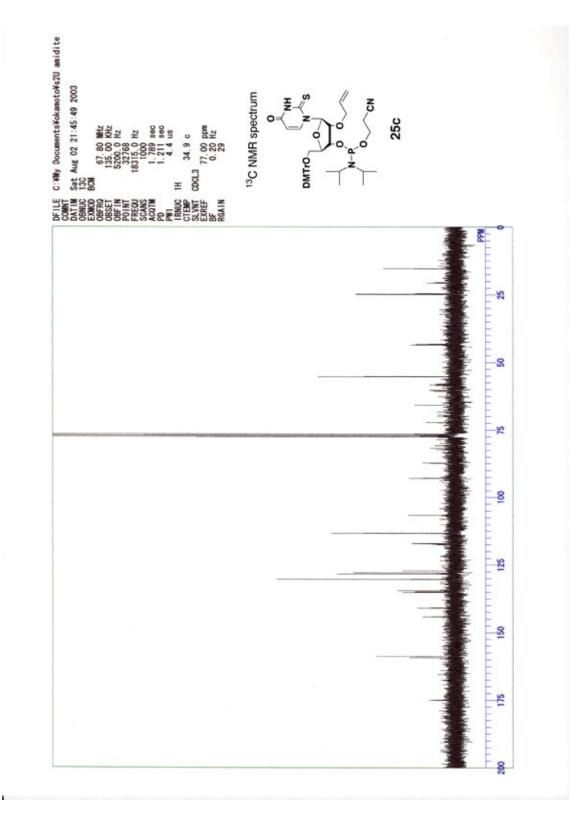


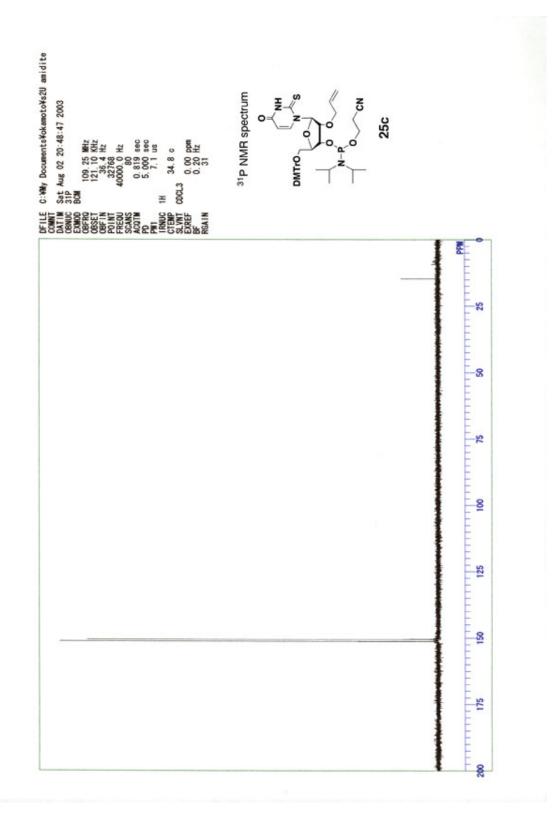


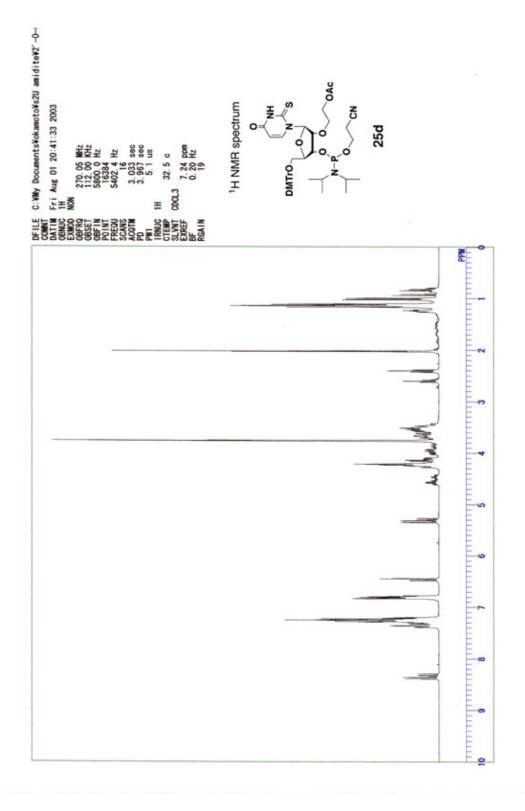




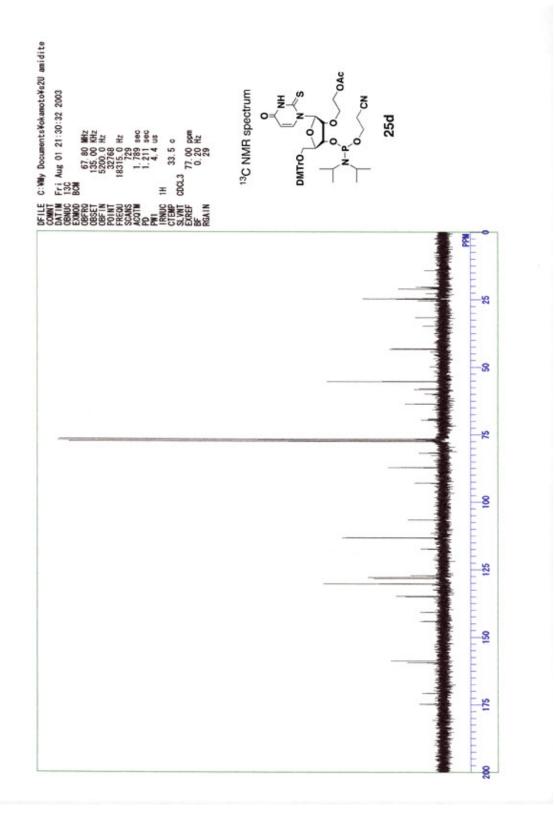




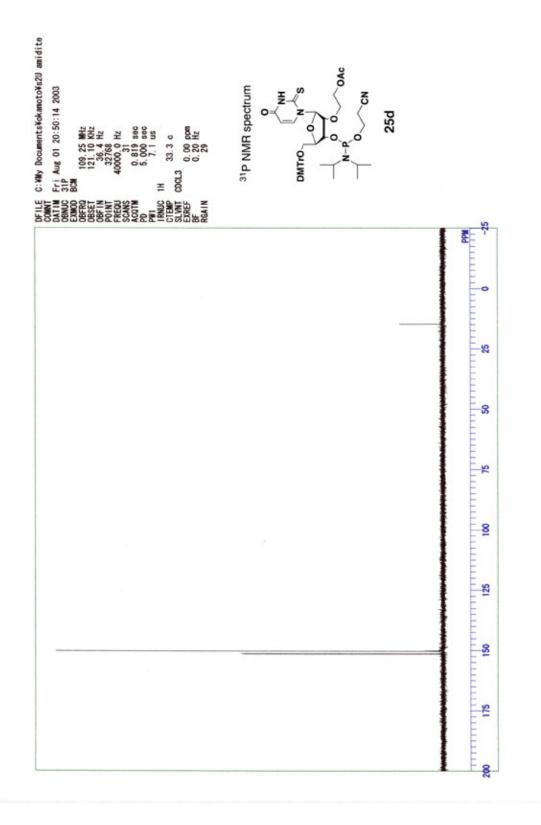




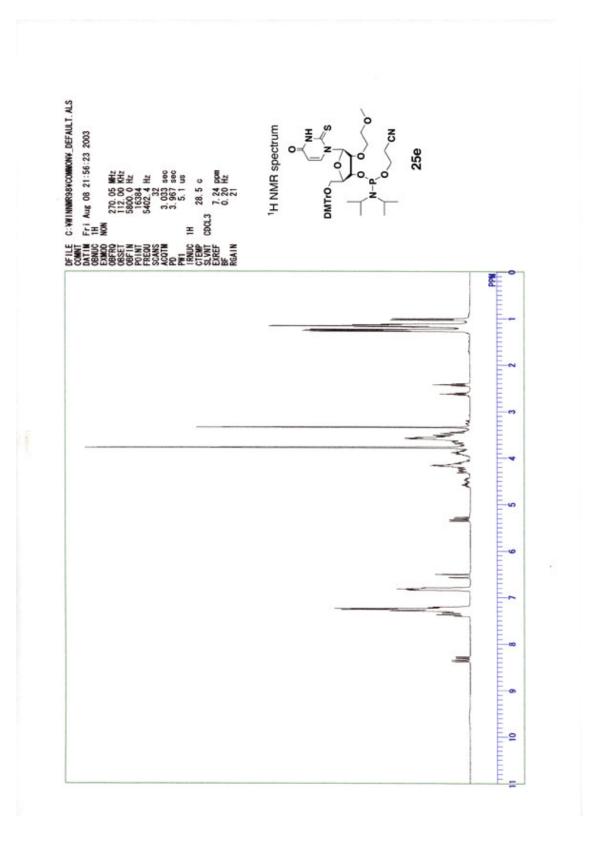
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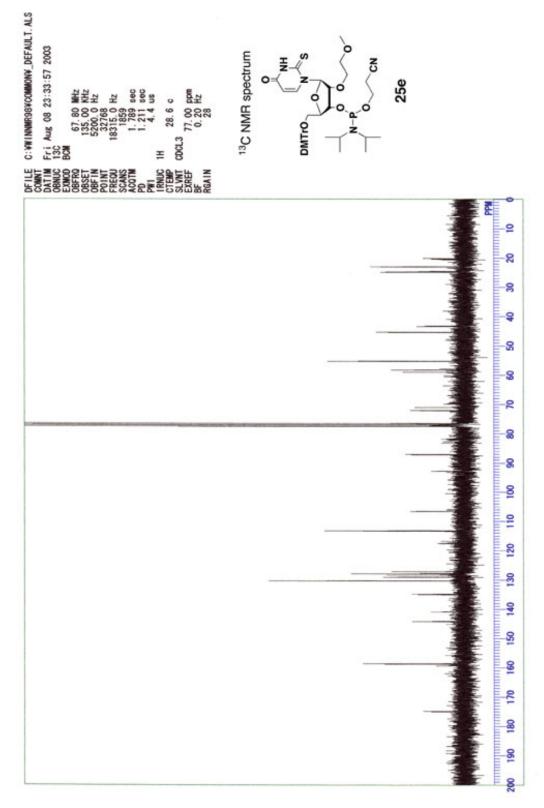


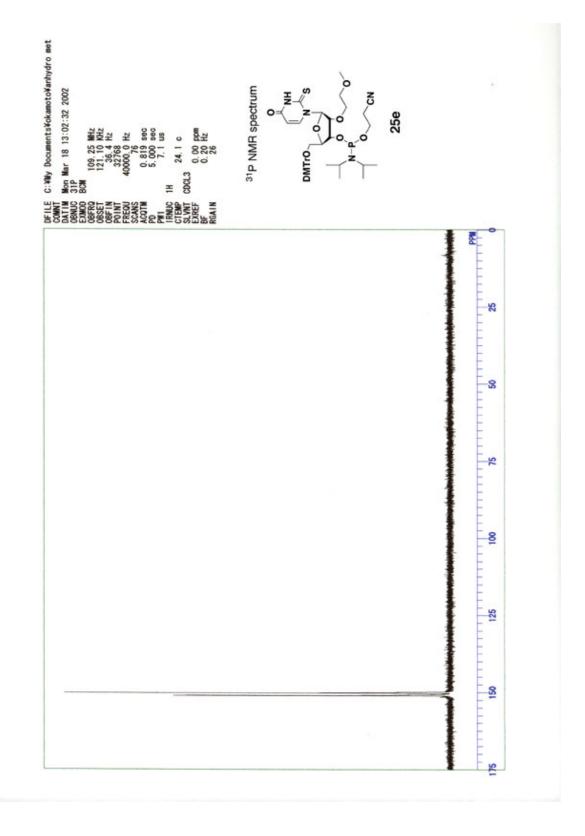
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