New Tools for Molecular Imaging of Redox Metabolism: Development of a Fluorogenic probe for 3α-Hydroxysteroid Dehydrogenases

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Supporting Information

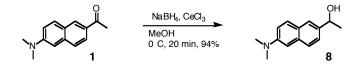
Materials and General methods

¹H and ¹³C NMR spectra were recorded on Bruker 300 or 400 Fourier transform NMR spectrometers. Spectra were recorded in CDCl₃ solutions referenced to TMS or the solvent residual peak unless otherwise indicated. IR spectra were taken as neat for liquids on NaCl plates or as KBr pellets for solids using a Perkin-Elmer 1600 FTIR spectrometer. High Resolution Mass Spectra were obtained on a JOEL JMS-HX110 HF mass spectrometer. Flash chromatography was performed on SILICYCLE silica gel (230-400 mesh). All chemicals were purchased from Aldrich and used as received. All reactions were monitored by Thin Layer Chromatography.

Ultraviolet spectra were measured on a Cary 100 UV-Visible spectrophotometer and recorded in EtOH solutions. Recorded λ_{max} is that of the longest wavelength transition. Fluorescence measurements were taken on a Jobin Yvon Fluorolog fluorescence spectrofluorometer in potassium phosphate pH 7.0 buffer unless otherwise indicated. Quantum yields were measured relative to 9, 10 diphenylanthracene in EtOH¹ for probes 1-4 and alcohols 8, 11, 13, and 15, or Coumarin 6 in EtOH² for probes 5-7 and alcohols 19, 21, and 22. Reported quantum efficiencies are the average of at least three independent preparations of the probes and their cognate alcohols.

Synthesis of Probes 1-7 and the Corresponding Alcohols

Synthesis of probe 1



1-(6-Dimethylamino-naphthalen-2-yl)-ethanone (1).

This compound was prepared by a literature procedure and spectral data are consistent with those previously published³.

1-(6-Dimethylamino-naphthalen-2-yl)-ethanol (8).

CeCl₃.7H₂O (116 mg, 0.31 mmol) was added to a solution of **1** (50 mg, 0.23 mmol) in MeOH (10 ml) at 0 C, followed by addition of NaBH₄ (46 mg, 1.22 mmol). After 20 minutes, the reaction was quenched with a saturated aqueous solution of NH₄Cl and extracted with CHCl₃. Organic layer was dried over MgSO₄, evaporated and the crude product was purified by column chromatography on silica gel (CH₂Cl₂-EtOAc 98:2) to provide pure alcohol (47 mg, 94%).

NMR ¹**H** (300 MHz, CDCl₃) δ ppm:

7.67 (d, 1H, J1=9.0 Hz); 7.63 (bs, 1H); 7.63 (d, 1H, J1=8.5 Hz); 7.37 (dd, 1H, J1=8.5 Hz, J2=1.7 Hz); 7.15 (dd, 1H, J1=9.0 Hz, J2=2.5 Hz); 6.90 (d, 1H, J1=2.5 Hz); 4.99 (m, 1H); 3.03 (s, 6H); 1.79 (d, 1H, J1=3.5 Hz), 1.59 (d, 3H, J1=6.4 Hz).

NMR ¹³C (300 MHz, CDCl₃) δ ppm:

148.7; 139.3; 134.5; 128.7; 126.6; 126.5; 124.2; 123.6; 116.7; 106.5; 70.6; 40.9; 24.9.

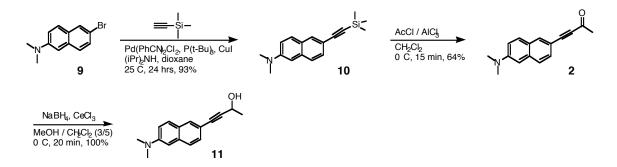
IR (NaCl, cm⁻¹): 3358, 2969, 2875, 1632, 1606, 1507, 1444, 1382, 1334, 1171, 1069, 968, 845, 804, 676.

HRMS (FAB): 215.1308 (C₁₄H₁₇ON, M; calc 215.1310).

UV (EtOH): $\lambda_{\text{max}} = 348$ nm.

Fluorescence (potassium phosphate pH 7.0): $\lambda_{em} = 429 \text{ nm}, \Phi_f = 0.07$.

Synthesis of probe 2



Dimethyl-(6-trimethylsilanylethynyl-naphthalen-2-yl)-amine (10).

This compound was prepared by the procedure of Buchwald and Fu⁴ from bromide **9**, which was obtained from 2-bromo-6-naphthol according to literature⁵. Pd(PhCN)₂Cl₂ (4.6 mg, 0.012 mmol), CuI (1.5 mg, 0.008 mmol), **9** (100 mg, 0.400 mmol), dioxane (1 ml), diisopropylamine (68 μ l, 0.024 mmol) and (trimethylsilyl)acetylene (110 μ l, 0.800 mmol) were mixed in a vial under argon and allow to stir 24 hrs at room temperature. The resultant mixture was diluted with EtOAc, washed with brine and dried over MgSO₄. Following solvent evaporation and product purification by column chromatography using silica gel and hexanes-EtOAc 98:2, **10** was yielded (99 mg, 93%).

NMR ¹**H** (300 MHz, CDCl₃) δ ppm:

7.81 (bs, 1H); 7.61 (d, 1H. J1=9.1 Hz); 7.52 (d, 1H, J1=8.5 Hz); 7.36 (dd, 1H, J1=8.5 Hz, J2=1.6 Hz); 7.11 (dd, 1H, J1=9.1 Hz, J2=2.5 Hz); 6.83 (d, 1H, J1=2.5 Hz); 3.05 (s, 6H); 0.27 (s, 9H).

NMR ¹³C (300 MHz, acetone-d) δ ppm:

150.4; 135.8; 132.3; 129.5; 129.3; 127.0; 126.8; 117.6; 116.5; 107.4; 106.5; 92.9; 40.6; 0.1.

IR (NaCl, cm⁻¹): 2960, 2901, 2812, 2147, 1629, 1598, 1247, 894, 850, 838, 809. **HPMS** (FAB): 267, 1442 (C, H, NSi, M: calc 267, 1443)

HRMS (FAB): 267.1442 (C₁₇H₂₁NSi, M; calc 267.1443).

4-(6-Dimethylamino-naphthalen-2-yl)-but-3-yn-2-one (2).

AcCl (13 μ l, 0.18 mmol) was added to a solution of **10** (43 mg, 0.16 mmol) in CH₂Cl₂ (2 ml) at 0 C, followed by addition of AlCl₃ (107 mg, 0.80 mmol). After 15 minutes, the reaction was quenched with H₂O and extracted with EtOAc. After the organic layer was dried over MgSO₄, the solvent was removed, and the residue was purified by column chromatography on silica gel (hexanes-EtOAc 98:2) to yield ketone **2** (55 mg, 64%).

NMR ¹**H** (300 MHz, CDCl₃) δ ppm:

7.96 (bs, 1H); 7.66 (d, 1H, J1=9.1 Hz); 7.56 (d, 1H, J=8.5 Hz); 7.41 (dd, 1H, J1=8.5 Hz, J2=1.6 Hz); 7.14 (dd, 1H, J1=9.1 Hz, J2=2.5 Hz); 6.82 (d, 1H, J1=2.5 Hz); 3.09 (s, 6H); 2.46 (s, 3H).

NMR ¹³**C** (300 MHz, CDCl₃) δ ppm:

184.6; 149.8; 135.9; 134.6; 129.3; 129.1; 126.3; 125.5; 116.5; 111.9; 105.4; 93.1; 88.5; 40.4; 32.7.

IR (NaCl, cm⁻¹): 2892, 2817, 2180, 1667, 1625, 1507, 1354, 1280, 1190, 1168, 896, 851, 810.

HRMS (FAB): 237.1138 (C₁₆H₁₅ON, M; calc 237.1154).

UV (EtOH): $\lambda_{max} = 389$ nm.

Fluorescence (potassium phosphate pH 7.0): 448 nm, $\Phi_f = 0.00$.

4-(6-Dimethylamino-naphthalen-2-yl)-but-3-yn-2-ol (11).

Reduction of **2** (20 mg, 0.084 mmol) in MeOH-CH₂Cl₂ 3:5 (5ml) followed a procedure analogous to that used for the preparation of **8.** Column chromatography on silica gel (CH₂Cl₂) afforded alcohol **11** (20 mg, 100%).

NMR ¹**H** (300 MHz, $CDCl_3$) δ ppm:

7.77 (bs, 1H); 7.61 (d, 1H, J1=9.1 Hz); 7.54 (d, 1H, J1=8.5 Hz); 7.33 (dd, 1H, J1=8.5 Hz, J2=1.5 Hz); 7.12 (dd, 1H; J1=9.1 Hz, J2=2.4 Hz); 6.83 (d, 1H, J1=2.4 Hz); 4.78 (m, 1H); 3.05 (s, 6H); 1.88 (d, 1H, J1=4.8 Hz); 1.57 (d, 3H, J1=6.5 Hz). NMR ¹³C (300 MHz, CDCl₃) δ ppm:

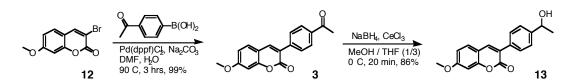
149.1; 134.5; 131.4; 128.8; 128.7; 126.1; 126.0; 116.6; 115.3; 105.9; 89.9; 85.0; 59.0; 40.6; 24.5.

IR (NaCl, cm⁻¹): 3346, 2982, 2930, 2882, 1628, 1598, 1505, 1389, 1101, 1072, 1035, 893, 848, 809.

HRMS (FAB): 239.1305 (C₁₆H₁₇ON, M; calc 239.1310).

UV (EtOH): $\lambda_{\text{max}} = 361$ nm.

Fluorescence (potassium phosphate pH 7.0): 440 nm, $\Phi_f = 0.08$.



3-(4-Acetyl-phenyl)-7-methoxy-coumarin (3).

Bromide **12** (400 mg, 1.57 mmol), obtained by bromination of 7-methoxycoumarin, was mixed with 4-acetylphenylboronic acid (283 mg, 1.72 mmol), PdCl₂dppf (40 mg, 0.047 mmol), Na₂CO₃ (831 mg, 7.84 mmol), H₂O (3.92 ml) and DMF (16 ml) under argon. The resulting mixture was heated to 90 C and stirred until completion (3 hrs). The cooled mixture was then diluted with water and extracted with CH₂Cl₂. Combined organic fractions were dried over MgSO₄. Following the evaporation of solvent, the residue was purified by column chromatography on silica gel (CH₂Cl₂) to afford desired product **3** (456 mg, 99%).

NMR ¹**H** (300 MHz, CDCl₃) δ ppm:

8.00 (m, 2H); 7.83 (m, 2H); 7.77 (bs, 1H); 7.46 (d, 1H, J1=8.4 Hz); 6.88 (m, 2H); 3.90 (s, 3H); 2.64 (s, 3H).

NMR ¹³**C** (300 MHz, CDCl₃) δ ppm:

197.6; 163.1; 160.5; 155.6; 141.0; 139.6; 136.6; 129.2; 128.5; 128.4; 123.5; 113.1; 113.1; 100.4; 55.9; 26.7.

IR (NaCl, cm⁻¹): 3070, 2962, 1710, 1670, 1613, 1505, 1442, 1360, 1275, 1198, 1122, 1022, 929, 859, 829, 776.

HRMS (FAB): 295.0967 (C₁₈H₁₅O₄, M+1; calc 295.0970).

UV (EtOH): $\lambda_{\text{max}} = 348$ nm.

Fluorescence (potassium phosphate pH 7.0): 462 nm, $\Phi_f = 0.00$.

3-[4-(1-Hydroxy-ethyl)-phenyl]-7-methoxy-coumarin (13).

Reduction of **3** (42 mg, 0.14 mmol) in MeOH-THF 1:3 (15ml) proceeded as described for the preparation of **8**. Column chromatography on silica gel (eluent gradient: CH_2Cl_2 to CH_2Cl_2 -EtOAc 8:2) afforded alcohol **13** (36 mg, 86%).

NMR ^{$\mathbf{\tilde{1}}}H (300 \text{ MHz}, \text{CDCl}_3) \delta \text{ ppm}$:</sup>

7.75 (s, 1H); 7.67 (m, 2H); 7.44 (m, 3H); 4.95 (m, 1H); 3.89 (s, 3H); 1.85 (d, 1H, J1=3.4 Hz); 1.52 (d, 3H, J1=6.4 Hz).

NMR ¹³**C** (300 MHz, CDCl₃) δ ppm:

162.5; 160.9; 155.2; 146.1; 139.8; 134.0; 128.8; 128.4; 125.4; 124.4; 113.3; 112.7; 100.3; 70.0; 55.7; 25.1.

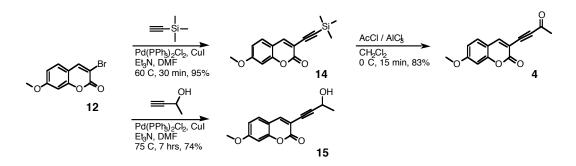
IR (NaCl, cm⁻¹): 3415, 2971, 1719, 1611, 1057, 1443, 1364, 1271, 1202, 1163, 1120, 1089, 1026, 832.

HRMS (FAB): 297.1112 (C₁₈H₁₇O₄, M+1; calc 297.1127).

UV (EtOH): $\lambda_{\text{max}} = 342$ nm.

Fluorescence (potassium phosphate pH 7.0): 429 nm, $\Phi_f = 0.12$.

Synthesis of probe 4



7-Methoxy-3-trimethylsilanylethynyl-coumarin (14).

PdCl₂(PPh₃)₂ (28 mg, 0.04 mmol), CuI (8 mg, 0.04 mmol), Et₃N (278 μ l, 2.00 mmol) and (trimethylsilyl)acetylene (138 μ l, 1.50 mmol) were added to a solution of bromide **12** (255 mg, 1.00 mmol) in dry DMF (10 ml) under argon. The resulting solution was heated to 60 C and allowed to react 30 minutes. The mixture was then cooled, diluted with water, and extracted with CH₂Cl₂. The organic fractions were then combined and dried over MgSO₄. Removal of solvent *in vacuo* and purification of the residue by column chromatography on silica gel (CH₂Cl₂) afforded product **14** (259 mg, 95%).

NMR ¹**H** (300 MHz, $CDCl_3$) δ ppm:

7.82 (s, 1H); 7.32 (d, 1H, J1=8.6 Hz); 6.83 (dd, 1H, J1=8.6 Hz, J2=2.4 Hz); 6.78 (d, 1H, J1=2.4 Hz); 3.86 (s, 3H); 0.26 (s, 9H).

NMR ¹³**C** (300 MHz, acetone-d) δ ppm:

164.6; 159.5; 156.5; 147.5; 130.4; 113.8; 113.2; 109.3; 101.3; 100.2; 99.8; 56.5; -0.2.

IR (NaCl, cm⁻¹): 3040, 2961, 2840, 1721, 1600, 1441, 1368, 1272, 1247, 1034, 973, 831, 807, 765.

HRMS (FAB): 272.0869 (C₁₅H₁₆O₃Si, M; calc 272.0869).

7-Methoxy-3-(3-oxo-but-1-ynyl)-coumarin (4).

Compound 14 (103 mg, 0.38 mmol) was converted into ketone 4 by the procedure used for the preparation of 2. Column chromatography of the crude product on silica gel (CH_2Cl_2) provided 4 (76 mg, 83%).

NMR ¹**H** (300 MHz, CDCl₃) δ ppm:

8.00 (bs, 1H); 7.40 (d, 1H, J1=8.7 Hz); 6.88 (dd, 1H, J1=8.7 Hz, J2=2.3 Hz); 6.81 (d, 1H, J1=2.3 Hz); 3.90 (s, 3H); 2.47 (s, 3H).

NMR ¹³C (300 MHz, CDCl₃) δ ppm:

184.1, 164.7; 158.9; 156.3; 149.8; 129.7; 113.8; 112.1; 106.0; 100.9; 92.2; 84.1; 56.0; 32.6.

IR (NaCl, cm⁻¹): 3046, 2197, 1725, 1664, 1617, 1596, 1557, 1504, 1368, 1273, 1250, 1152, 1116, 1019, 836.

HRMS (FAB): 242.0572 ($C_{14}H_{10}O_4$, M+1; calc. 242.0579).

UV (EtOH): $\lambda_{max} = 368$ nm.

Fluorescence (potassium phosphate pH 7.0): 416 nm, $\Phi_f = 0.00$.

3-(3-Hydroxy-but-1-ynyl)-7-methoxy-coumarin (15).

Alcohol **15** was prepared by Sonogashira coupling of bromide **12** (100 mg, 0.39 mmol) and but-3-yn-2-ol (32 μ l, 0.43 mmol) under conditions similar to that used for the

preparation of **14**. After 7 hours at 75 C, the reaction was complete. The crude alcohol was purified by column chromatography on silica gel (CH_2Cl_2 -EtOAc 95:5) to afford product **15** (96 mg, 74%).

NMR ¹**H** (300 MHz, CDCl₃) δ ppm:

7.81 (bs, 1H); 7.35 (d, 1H, J1=8.6 Hz); 6.86 (dd, 1H, J1=8.6 Hz, J2=2.4 Hz); 6.81 (d, 1H, J1=2.4 Hz); 4.79 (m, 1H); 3.88 (s, 3H); 2.26 (d, 1H, J1=5.2 Hz); 1.56 (d, 3H, J1=6.6 Hz).

NMR ¹³**C** (300 MHz, CDCl₃) δ ppm:

163.3; 160.1; 155.2; 145.5; 128.8; 113.2; 112.4; 108.6; 100.7; 96.7; 77.9; 58.7; 55.8; 24.0.

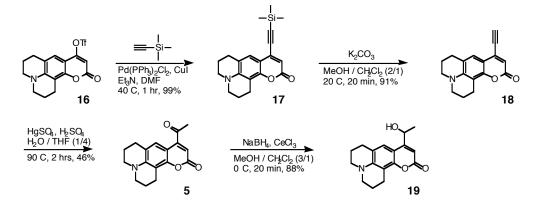
IR (NaCl, cm⁻¹): 3414, 2983, 2939, 2843, 1733, 1618, 1506, 1365, 1269, 1121, 1024, 768.

HRMS (FAB): 244.0744 (C₁₄H₁₂O₄, M; calc 244.0736).

UV (EtOH): $\lambda_{\text{max}} = 346$ nm.

Fluorescence (potassium phosphate pH 7.0): 420 nm, $\Phi_f = 0.18$.

Synthesis of probe 5



8-Trimethylsilanylethynyl-2,3,5,6-tetrahydro-1*H*, 4*H*-11-oxa-3a-azabenzo[*de*]anthracen-10-one (17).

Triflate **16** (707mg, 1.82 mmol), obtained from 8-hydroxyjulolidine according to the literature⁶, was coupled with (trimethylsilyl)acetylene (377 μ l, 2.72 mmol) under conditions described for the preparation of **14**. The reaction was complete after 1 hr at 40 C. Column chromatography on silica gel (CH₂Cl₂) provided desired product **17** (607 mg, 99%).

NMR ¹**H** (300 MHz, CDCl₃) δ ppm:

7.16 (s, 1H); 6.11 (s, 1H); 3.26 (m, 4H); 2.83 (m, 4H); 1.97 (m, 4H); 0.31 (s, 9H). NMR ¹³C (300 MHz, CDCl₃) δ ppm:

161.8; 151.1; 146.1; 137.0; 123.5; 118.3; 110.8; 107.6; 106.6; 106.3; 98.8; 49.9; 49.4; 27.6; 21.4; 20.4; 20.2; -0.4.

IR (NaCl, cm⁻¹): 2946, 2848, 1701, 1612, 1546, 1511, 1421, 1367, 1310, 1245, 1184, 843.

HRMS (FAB): 338.1574 (C₂₀H₂₄O₂NSi, M+1; calc 338.1576).

8-Ethynyl-2,3,5,6-tetrahydro-1*H*, 4*H*-11-oxa-3a-aza-benzo[*de*]anthracen-10-one (18).

Powdered K_2CO_3 (600 mg) was added to a solution of **17** (580 mg, 1.72 mmol) in MeOH-CH₂Cl₂ 2:1 (30 ml). The mixture was stirred at room temperature until the reaction was complete (20 min). Reaction mixture was diluted with CHCl₃, filtered, and washed with brine. The resultant organic layers were combined and dried over MgSO₄, after which the solvent was removed *in vacuo*. Purification by column chromatography on silica gel (eluent gradient: CH₂Cl₂ to CH₂Cl₂-EtOAc 95:5) afforded terminal alkyne **18** (416 mg, 91%).

NMR ¹**H** (300 MHz, CDCl₃) δ ppm:

7.19 (s, 1H); 6.16 (s, 1H); 3.58 (s, 1H); 3.27 (m, 4H); 2.87 (m, 2H); 2.78 (m, 2H); 1.97 (m, 4H).

NMR ¹³**C** (300 MHz, CDCl₃) δ ppm:

161.6; 151.1; 146.3; 136.3; 123.4; 118.5; 111.7; 107.6; 106.4; 87.5; 78.0; 49.9; 49.5; 27.5; 21.3; 20.4; 20.2.

IR (NaCl, cm⁻¹): 3221, 2931, 2838, 2103, 1699, 1616, 1519, 1428, 1371, 1311, 1176, 826.

HRMS (FAB): 266.1193 (C₁₇H₁₆O₂N, M+1; calc 266.1181).

8-Acetyl-2,3,5,6-tetrahydro-1H, 4H-11-oxa-3a-aza-benzo[de]anthracen-10-one (5).

HgSO₄ (112 mg, 0.38 mmol) was added to a solution of **18** (100 mg, 0.38 mmol) in THF (8 ml), followed by addition of conc. H₂SO₄ (105 μ l, 1.88 mmol) in H₂O (2ml). The reaction mixture was heated in a sealed tube at 90 C for 2 hrs. After cooling to room temperature, a spatula tip of NaHCO₃ was added and the mixture was evaporated to dryness. MgSO₄ was added and the residual solids were washed thoroughly with CHCl₃. The solvent was the evaporated and the residue purified by column chromatography on silica gel (CH₂Cl₂-Et₂O 95:5) yielding ketone **5** (49 mg, 46%).

NMR ¹**H** (300 MHz, CDCl₃) δ ppm:

7.18 (s, 1H); 6.13 (s, 1H); 3.27 (m, 4H); 2.88 (m, 2H); 2.74 (m, 2H); 2.55 (s, 3H); 1.96 (m, 4H).

NMR ¹³**C** (300 MHz, CDCl₃) δ ppm:

200.4; 162.1; 152.1; 150.8; 146.2; 123.2; 118.7; 106.8; 106.8; 103.7; 49.9; 49.4; 29.7; 27.6; 21.3; 20.4; 20.3.

IR (NaCl, cm⁻¹): 2933, 2844, 1694, 1611, 1544, 1525, 1434, 1373, 1352, 1311, 1232, 1170, 1148.

HRMS (FAB): 283.1195 ($C_{17}H_{17}O_3N$, M; calc 283.1208).

UV (EtOH): $\lambda_{max} = 418$ nm.

Fluorescence (potassium phosphate pH 7.0): 520 nm, $\Phi_f = 0.00$.

8-(1-Hydroxy-ethyl)-2,3,5,6-tetrahydro-1*H*, 4*H*-11-oxa-3a-aza-benzo[*de*]anthracen-10-one (19).

Reduction of **5** (16 mg, 0.056 mmol) in MeOH-CH₂Cl₂ 3:1 (5ml) proceeded by previously described procedures (used for preparation of **8**). Column chromatography on silica gel (eluent gradient: CH_2Cl_2 to CH_2Cl_2 -EtOAc 9:1) afforded alcohol **19** (14 mg, 88%).

NMR ¹**H** (300 MHz, CDCl₃) δ ppm:

7.01 (s, 1H); 6.24 (s, 1H); 5.14 (m, 1H); 3.26 (m, 4H); 2.87 (m, 2H); 2.77 (m, 2H); 2.07 (d, 1H, J1=3.8 Hz); 2.10 (m, 4H); 1.55 (d, 3H, J1=6.5 Hz).

NMR ¹³**C** (300 MHz, CDCl₃) δ ppm:

163.0; 159.4; 151.4; 145.6; 121.0; 118.0; 107.1; 105.9; 103.8; 65.9; 49.9; 49.5; 27.8; 23.6; 21.5; 20.6; 20.5.

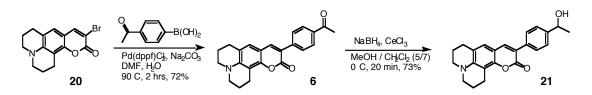
IR (NaCl, cm⁻¹): 3396, 2936, 2843, 1688, 1611, 1554, 1520, 1433, 1372, 1311, 1183, 1133.

HRMS (FAB): 286.1437 (C₁₇H₂₀O₃N, M+1; calc 286.1443).

UV (EtOH): $\lambda_{\text{max}} = 398$ nm.

Fluorescence (potassium phosphate pH 7.0): 509 nm, $\Phi_f = 0.21$.

Synthesis of probe 6



9-(4-Acetyl-phenyl)-2,3,5,6-tetrahydro-1*H*, 4*H*-11-oxa-3a-aza-benzo[*de*]anthracen-10-one (6).

Bromide **20** (100 mg, 0.31 mmol), obtained by bromination of coumarin 6H, was coupled with 4-acetylphenylboronic acid (77 mg, 0.46 mmol), under similar conditions as those used for preparation of **3**. Reaction was complete after 2 hrs at 90 C. Column chromatography on silica gel (eluent gradient: CH_2Cl_2 to CH_2Cl_2 -EtOAc 95:5) provided desired ketone **6** (81 mg, 72%).

NMR ¹**H** (300 MHz, $CDCl_3$) δ ppm:

7.96 (m, 2H); 7.81 (m, 2H); 7.68 (s, 1H); 6.91 (s, 1H); 3.29 (m, 4H); 2.93 (m, 2H); 2.77 (m, 2H); 2.61 (s, 3H); 1.98 (m, 4H).

NMR¹³C (300 MHz, CDCl₃) δ ppm:

197.7; 161.4; 151.5; 146.3; 141.7; 141.0; 135.6; 128.3; 128.0; 125.4; 118.7; 118.0; 108.7; 106.1; 50.0; 49.6; 27.4; 26.6; 21.4; 20.4; 20.2.

IR (NaCl, cm⁻¹): 2941, 2845, 1699, 1677, 1616, 1594, 1563, 1518, 1360, 1306, 1269, 1213, 1171.

HRMS (FAB): 359.1527 (C₂₃H₂₁O₃N, M; calc 359.1521).

UV (EtOH): $\lambda_{\text{max}} = 435$ nm.

Fluorescence (potassium phosphate pH 7.0): 511 nm, $\Phi_f = 0.01$.

9-[4-(1-Hydroxy-ethyl)-phenyl]-2,3,5,6-tetrahydro-1*H*, 4*H*-11-oxa-3a-aza-benzo[*de*]anthracen-10-one (21).

Reduction of **6** (15 mg, 0.041 mmol) in MeOH-CH₂Cl₂ 5:7 (6ml) by the procedure used for preparation of **8** and recrystallization from CHCl₃-hexanes afforded alcohol **21** (11 mg, 73%).

NMR ¹**H** (300 MHz, CDCl₃) δ ppm:

7.66 (m, 2H); 7.58 (s, 1H); 7.40 (m, 2H); 6.88 (s, 1H); 4.92 (q, 1H, J1=6.4 Hz); 3.28 (m, 4H); 2.92 (m, 2H); 2.76 (m, 2H); 1.98 (m, 4H); 1.81 (bs, 1H); 1.51 (d, 3H, J1=6.4 Hz).

NMR ¹³**C** (300 MHz, CDCl₃) δ ppm:

161.9; 151.2; 145.8; 145.1; 140.8; 135.3; 128.3; 125.3; 125.1; 119.6; 118.5; 109.0; 106.3; 70.2; 50.0; 49.6; 27.5; 25.1; 21.5; 20.6; 20.3.

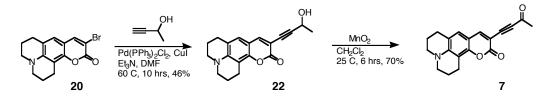
IR (NaCl, cm⁻¹): 3408, 2930, 2844, 1694, 1615, 1599, 1564, 1519, 1309, 1209, 1170, 839, 748.

HRMS (FAB): 361.1673 (C₂₃H₂₃O₃N, M; calc 361.1678).

UV (EtOH): $\lambda_{\text{max}} = 422 \text{ nm.}$

Fluorescence (potassium phosphate pH 7.0): 509 nm, $\Phi_f = 0.14$.

Synthesis of probe 7



9-(3-Hydroxy-but-1-ynyl)-2,3,5,6-tetrahydro-1*H*, 4*H*-11-oxa-3a-aza-benzo[*de*]anthracen-10-one (22).

Alcohol **22** was prepared by Sonogashira coupling of bromide **20** (100 mg, 0.31 mmol) and but-3-yn-2-ol (26 μ l, 0.34 mmol) as described for the preparation of **14**. The reaction was stopped after 10 hrs at 60 C. Column chromatography on silica gel (eluent gradient: CH₂Cl₂ to CH₂Cl₂-EtOAc 9:1) provided **22** (45 mg, 46%).

NMR ¹**H** (300 MHz, $CDCl_3$) δ ppm:

7.60 (s, 1H); 6.78 (s, 1H); 4.77 (m, 1H); 3.28 (m, 4H); 2.87 (m, 2H); 2.75 (m, 2H); 2.14 (d, 1H, J1=4.9 Hz); 1.97 (m, 4H); 1.54 (d, 3H, J1=6.6 Hz).

NMR ¹³**C** (300 MHz, CDCl₃) δ ppm:

161.6; 151.3; 146.6; 146.4; 125.0; 118.9; 108.1; 106.4; 102.8; 94.7; 79.3; 58.9; 50.1; 49.7; 27.4; 24.1; 21.3; 20.4; 20.2.

IR (NaCl, cm⁻¹): 3397, 2934, 2849, 1709, 1692, 1616, 1594, 1518, 1360, 1309, 1290, 1169, 765.

HRMS (FAB): 309.1365 ($C_{19}H_{19}O_3N$, M; calc 309.1365).

UV (EtOH): $\lambda_{\text{max}} = 429$ nm.

Fluorescence (potassium phosphate pH 7.0): 508 nm, $\Phi_f = 0.35$.

9-(3-Oxo-but-1-ynyl)-2,3,5,6-tetrahydro-1*H*, 4*H*-11-oxa-3a-aza-benzo[*de*]anthracen-10-one (7).

To alcohol **22** (30 mg, 0.097 mmol) dissolved in dry CH_2Cl_2 (3 ml) was added powdered MnO₂ (150 mg) at room temperature. The resulting suspension was stirred until the reaction was complete (6 hrs). The subsequent mixture was filtered through Celite, dried *in vacuo*, and purified by column chromatography on silica gel (eluent gradient: CH_2Cl_2 to CH_2Cl_2 -EtOAc 98:2) to afford **7** (21 mg, 70%). **NMR** ¹**H** (300 MHz, CDCl₃) δ ppm: 7.77 (s, 1H); 6.82 (s, 1H); 3.33 (m, 4H); 2.86 (m, 2H); 2.75 (m, 2H); 2.44 (s, 3H); 1.97 (m, 4H).

NMR ¹³**C** (300 MHz, CDCl₃) δ ppm:

184.3; 160.6; 152.3; 150.2; 148.2; 125.9; 119.4; 108.1; 106.2; 98.8; 92.4; 87.9; 50.2; 49.8; 32.5; 27.3; 21.0; 20.1; 20.0.

IR (NaCl, cm⁻¹): 2937, 2844, 2170, 1714, 1657, 1620, 1586, 1520, 1358, 1295, 1154, 760.

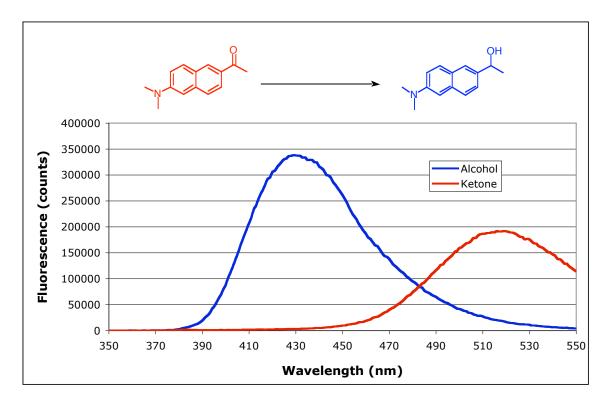
HRMS (FAB): 308.1295 ($C_{19}H_{18}O_3N$, M+1; calc 308.1287).

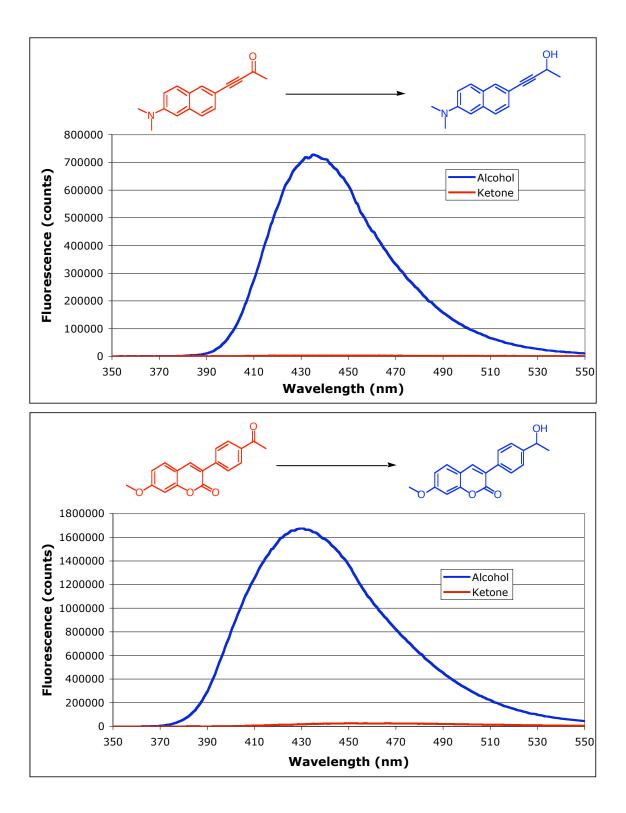
UV (EtOH): $\lambda_{\text{max}} = 464$ nm.

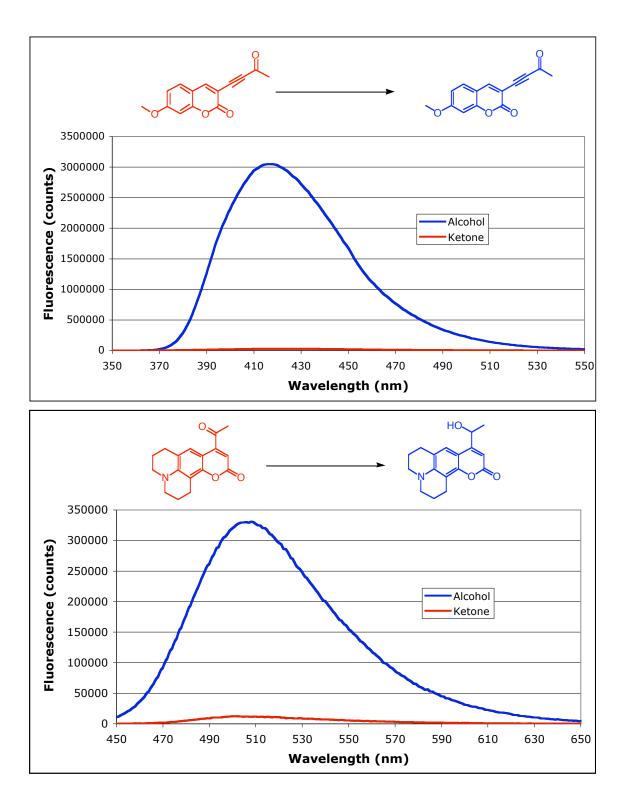
Fluorescence (potassium phosphate pH 7.0): 512 nm, $\Phi_f = 0.01$.

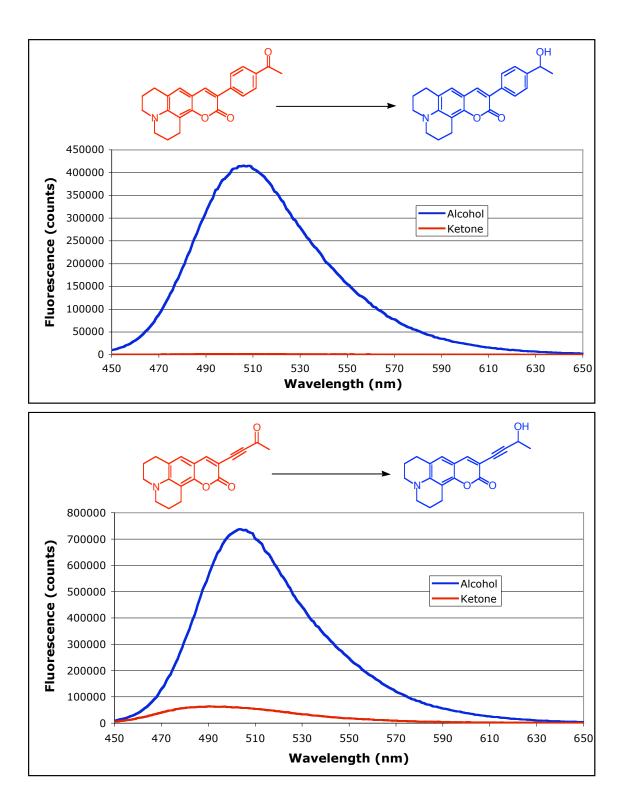
Fluorescence Spectra of Selected Probes 1-7:

Compounds 1-4 were excited at 340 nm, while compounds 5-7 were excited at 440 nm. Fluorescence emission spectra were recorded with 10 μ M solutions (<1% DMSO v/v) in 100 mM potassium phosphate buffer (pH 7.0).









S13

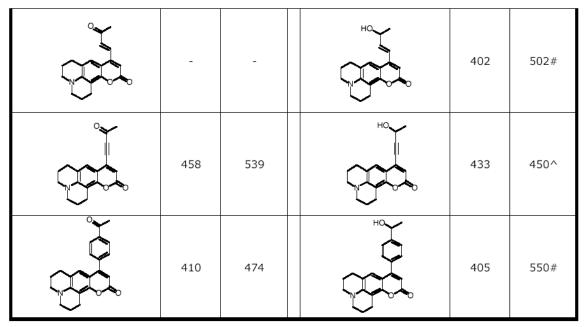
Ketone	Abs. Max (nm)	Fluor. Max (nm)	Alcohol	Abs. Max (nm)	Fluor. Max (nm)
	312	349		274	366
	316	354		279	380
	334	454	U C C C C C C C C C C C C C C C C C C C	346	411#
y C L	360	410		295	404
	346	392		318	474
y Cord	368	521	N COL	348	429

Photochemical Characterization of All Synthesized Compounds:

The second secon	361	-	W COH	316	498
	396	416	W Contraction of the second se	347	512#
	377	516	When the second	335	461
y Correl	395	-		318	453
Y	389	448	ОН	361	440
W COLOR	364	-	W COL	315	447

North Contraction	392	452	он N 348 5	510#
	378	-	ЭН 331	-
HOLOGIC	333	668	но от 329	461
	359	429	он 320	400
	322	443	он 278	455
	368	416	он 346	420

	348	462		342	429
J. J	449	504	C C C C C C C C C C C C C C C C C C C	378	501*
June 1	465	587	OH OH	416	519^
	464	512	OH NOTOTO	429	508
	435	511	CH CH	422	509
	418	520		398	509



low quantum yield, ^ reactivity with cellular reductants, * no change in wavelength of emission

Protocols for Enzymatic Assays

<u>Procedure for Enzymatic Screening of Selected Probes 1-7:</u>

Horse Liver alcohol dehydrogenase (Lot Number 51K7520), *Thermoanaerobium* brockii NADP⁺ dependent alcohol dehydrogenase (Lot Number 033K4093), *Pseudomonas testosteroni* 3α -hydroxysteroid dehydrogenase (Lot Number 053K8624), and *Bacillus sphaericus* 12 α -hydroxysteroid dehydrogenase (Lot Number 70K16621) were purchased from Sigma. Yeast alcohol dehydrogenase (Lot Number 93122920), glycerol dehydrogenase (Lot Number 92110122), (D)-lactate dehydrogenase (Lot Number 92419236), (L)-lactate dehydrogenase (Lot Number 92801821), NAD⁺, NADP⁺, NADH, and NADPH were purchased from Roche. Enzyme activity was confirmed by compliance to supplier's quality control assays prior to usage. Rat and human 3α -hydroxysteroid dehydrogenases were provided by Professor Trevor Penning (University of Pennsylvania School of Medicine) and human amyloid- β peptide binding alcohol dehydrogenase was supplied by Professor Shi Du Yan (Columbia University School for Physicians and Surgeons).

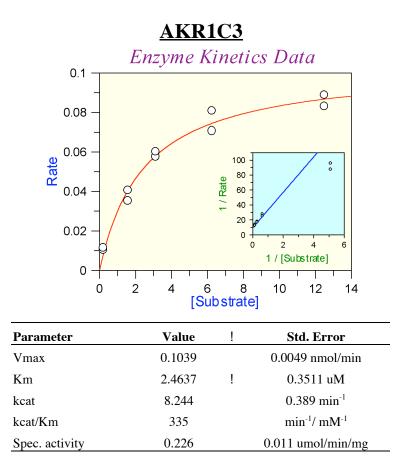
Enzymatic assays were performed in triplicate on selected fluorogenic substrates according to the following protocol. To each well of a FALCON 96-well black flat bottom plate was added (1) 40 μ L of 500 mM potassium phosphate buffer pH 7.0, (2) 113 μ L of double deionized water, (3) 25 μ L of 2 mM NADH (except for *Pseudomonas testosteroni* 3 α -hydroxysteroid dehydrogenase, rat 3 α -hydroxysteroid dehydrogenase, in which cases 2 mM of NADPH was used), (4) 2 μ L of a 3-5 mM solution of substrate in DMSO, and (5) 20 μ L of a 40-50 μ g/mL solution of enzyme. Reaction volumes were mixed thoroughly after addition of cofactor, substrate, and enzyme and allowed to react 12 hours at 25 C. Scanning of the 96-well plate was performed by the MicroMax 384 connected to a Jobin Yvon Fluorolog through F-3000 fiber optic cables.

Determination of Kinetic Parameters for AKR1C3

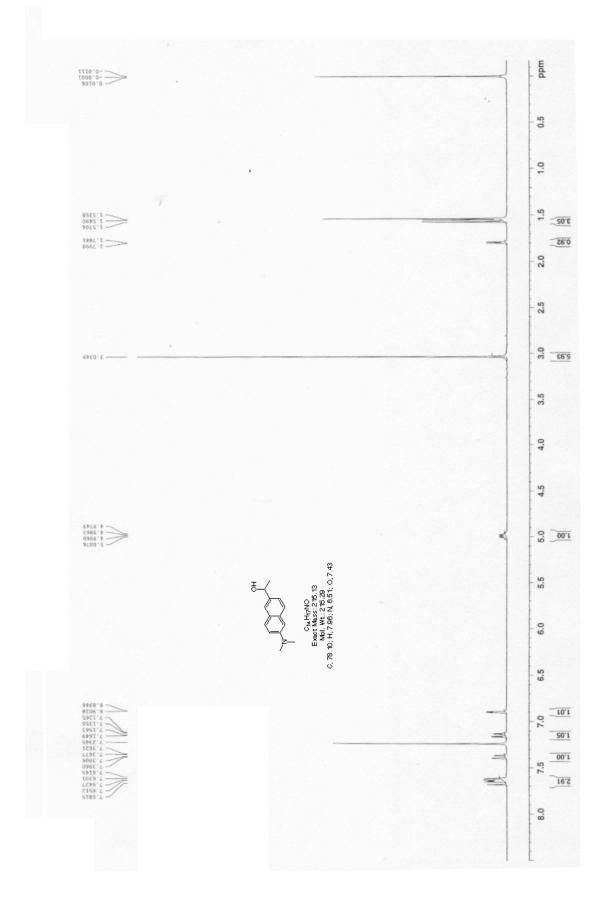
Fluorogenic substrate **5** reduction was monitored on a Hitachi F-4500 fluorimeter in Starna quartz cuvettes fluorometrically in 1 mL systems containing 100 mM potassium phosphate pH 6.0 containing excess of NADPH cofactor (250 μ M) and various amounts of the substrate (0.1953-50 μ M) dissolved in 4% acetonitrile. Aqueous assay components were added first, followed by addition of 20 μ L of acetonitrile as a cosolvent, and then addition of 20 μ L of the substrate in acetonitrile (total acetonitrile in the assay did not exceed 4%). Cuvettes were mixed thoroughly after addition of cofactor, cosolvent, and substrate. Reactions were initiated by the addition of 4 μ L of dilute AKR1C3 (115 μ g/mL) and were corrected for nonenzymatic rates. All reactions were followed by monitoring the increase in fluorescence of the product alcohol for 5 minutes at λ_{em} 510 nm with λ_{ex} 440 nm (Excitation and emission band pass slits both at 2.5 nm, lamp 900 V) at 37°C. The initial velocities, expressed in units of nanomoles per minute, were calculated according to previously published procedures⁷:

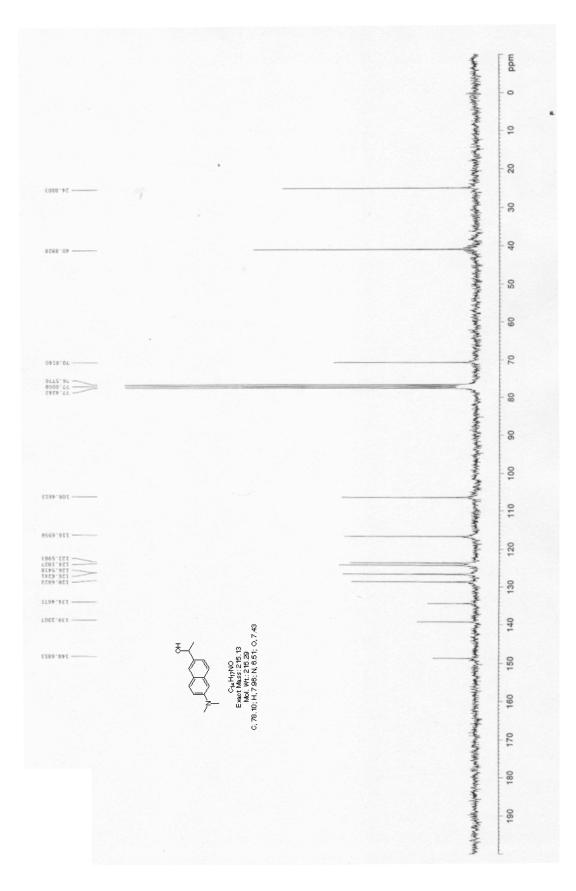
initial rate =
$$[n_{st} \times (F_t - F_0)/(F_{st})] / t$$
 (1)

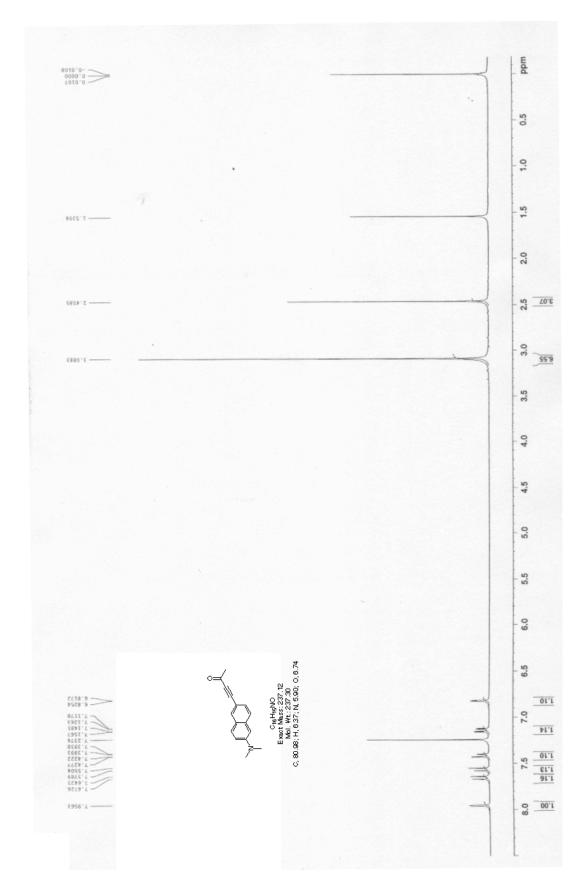
where F_t and F_0 represent the fluorescence at time t and 0, n_{st} is the nanomoles of the product standard, and F_{st} is the fluorescence resulting from n_{st} of product. Kinetic constants were approximated using the GraFit (Erithacus Software, Surrey, UK) non-linear regression analysis program to fit the untransformed data to a hyperbolic function as originally described⁹, yielding estimated values of k_{cat} , K_m , and their associated standard errors.

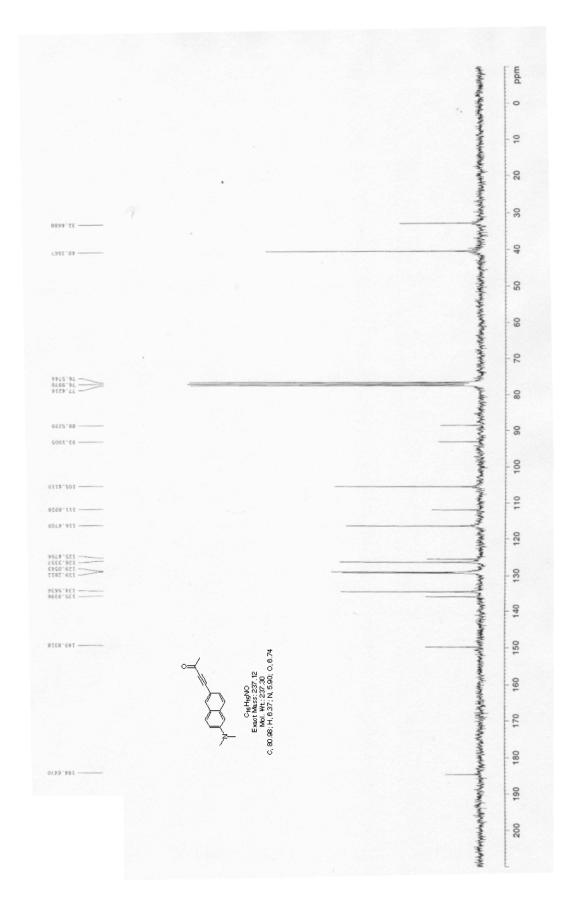


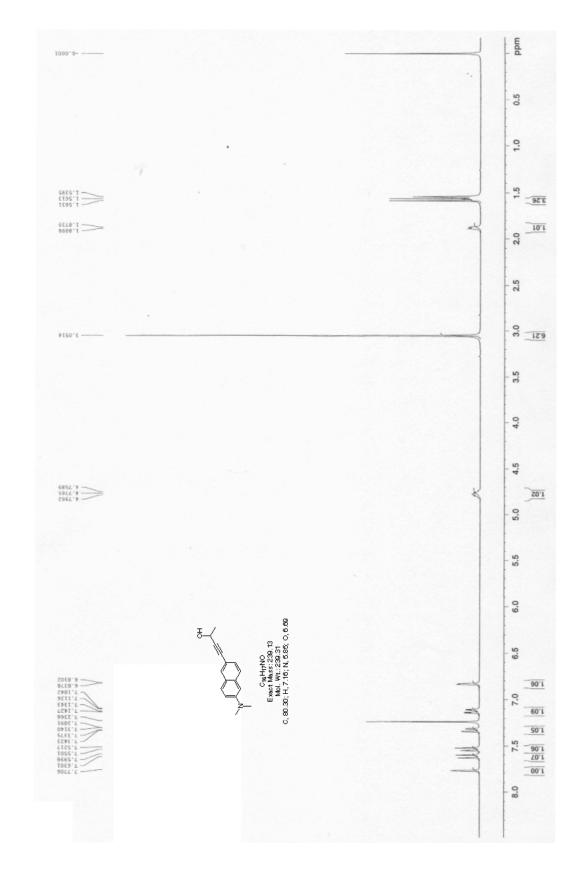
AKR1C3 kinetic data was also performed by HPLC separation of the fluorogenic substrate and its product alcohol and measurement of ketone to alcohol ratios. This data was found to correlate well with kinetic parameters determined fluorometrically¹⁰.

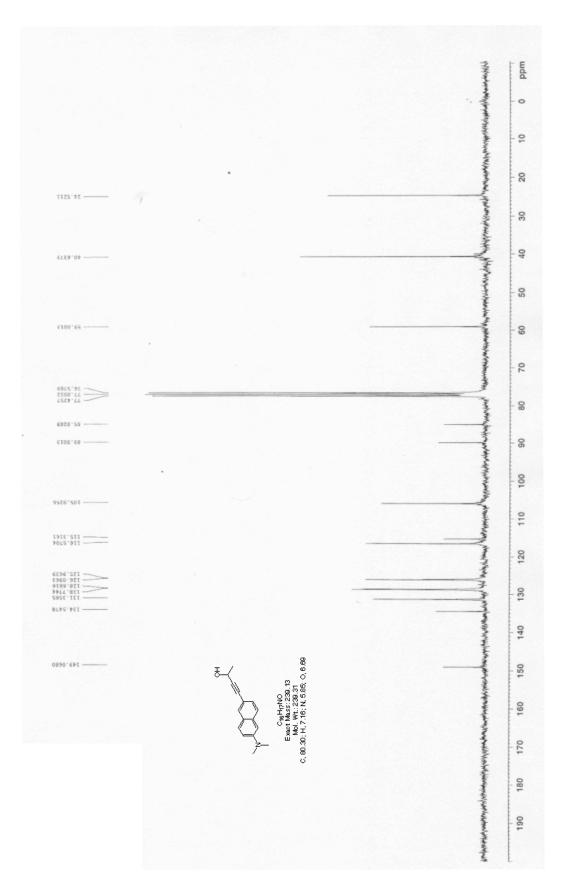


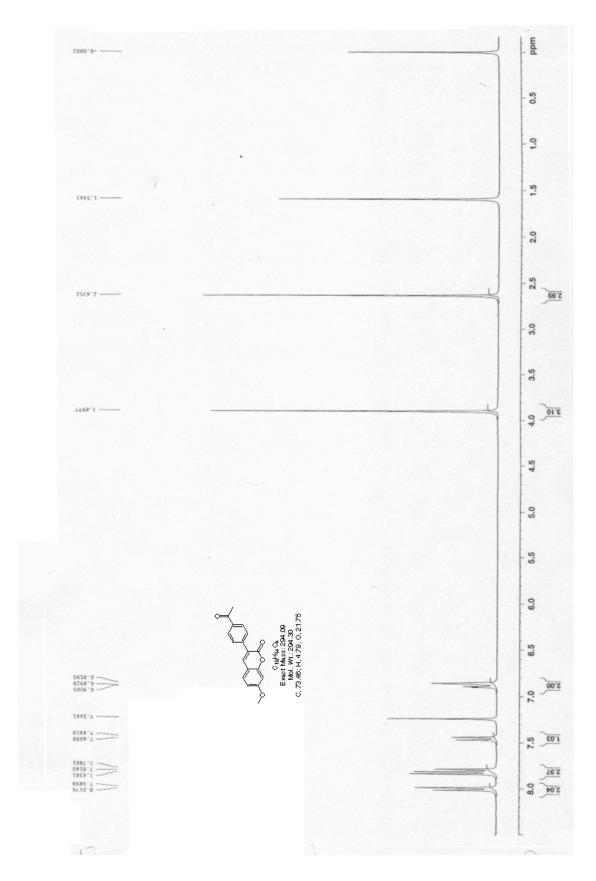


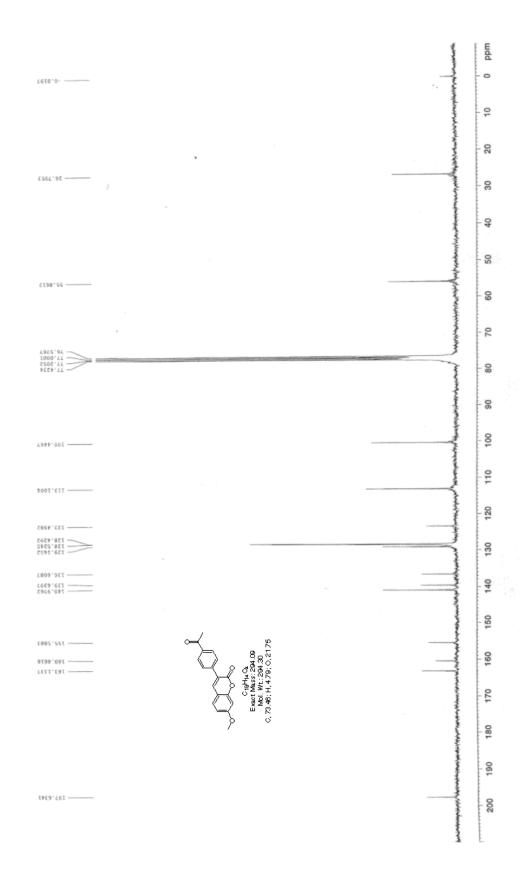


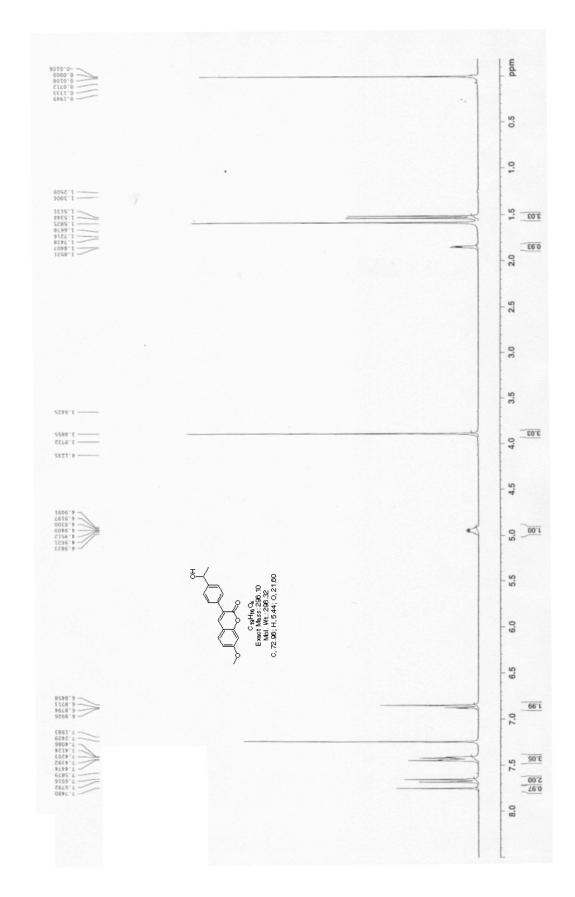


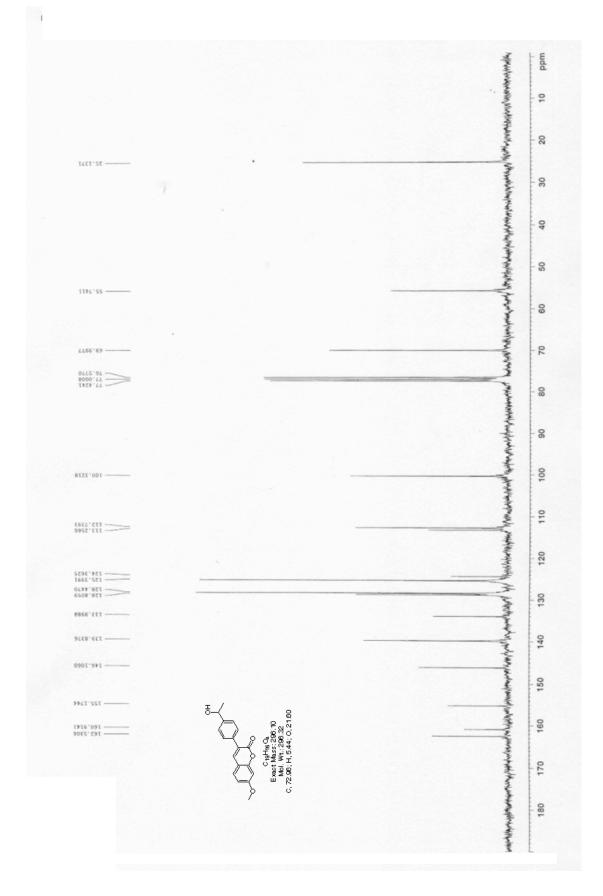


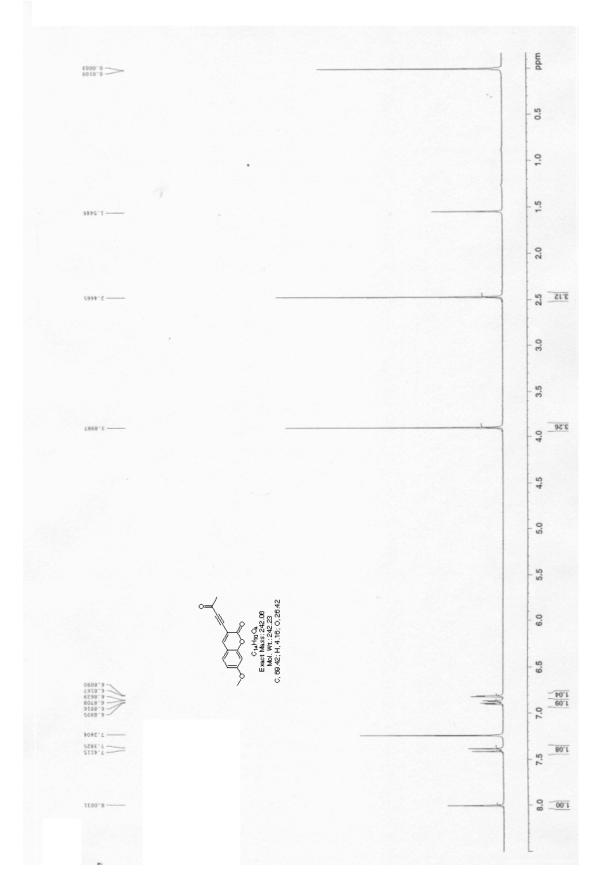


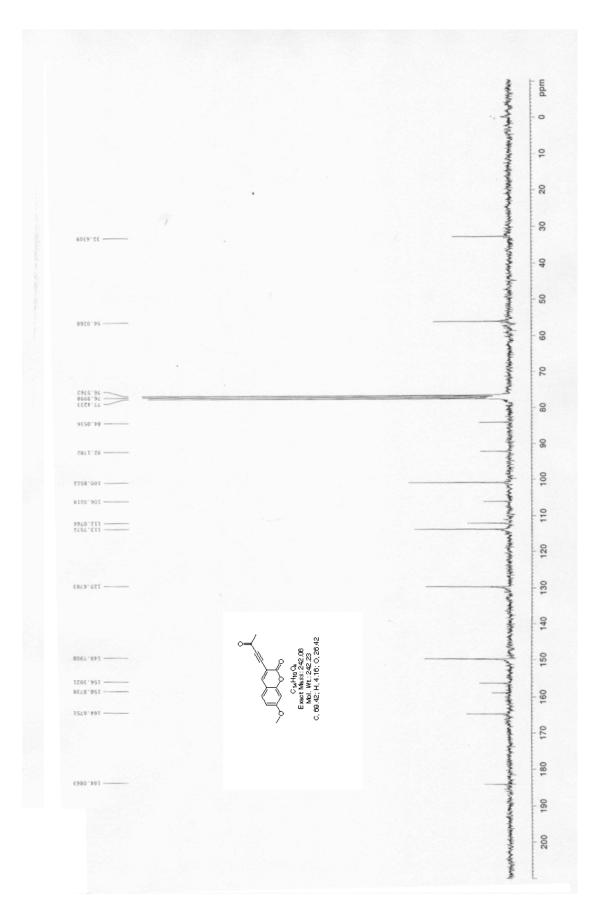


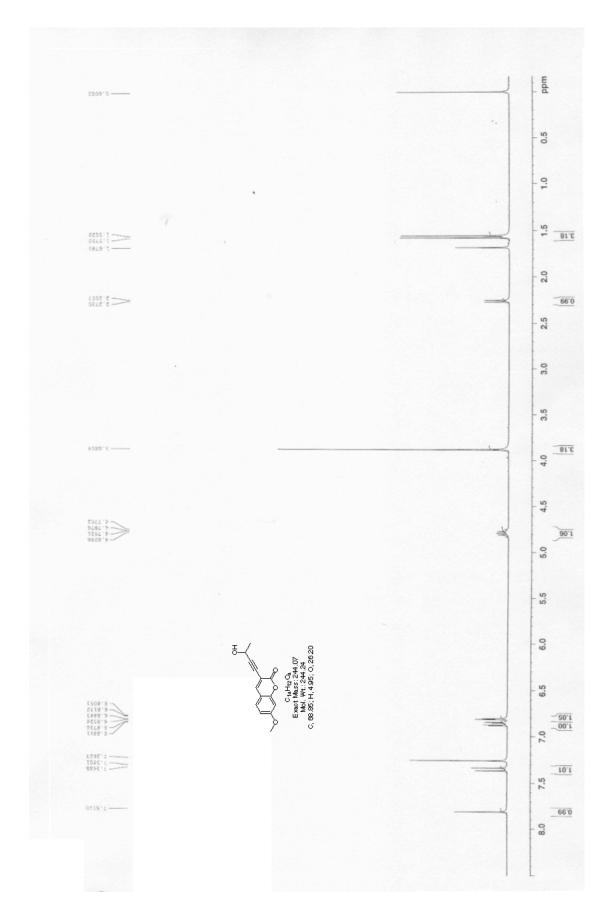


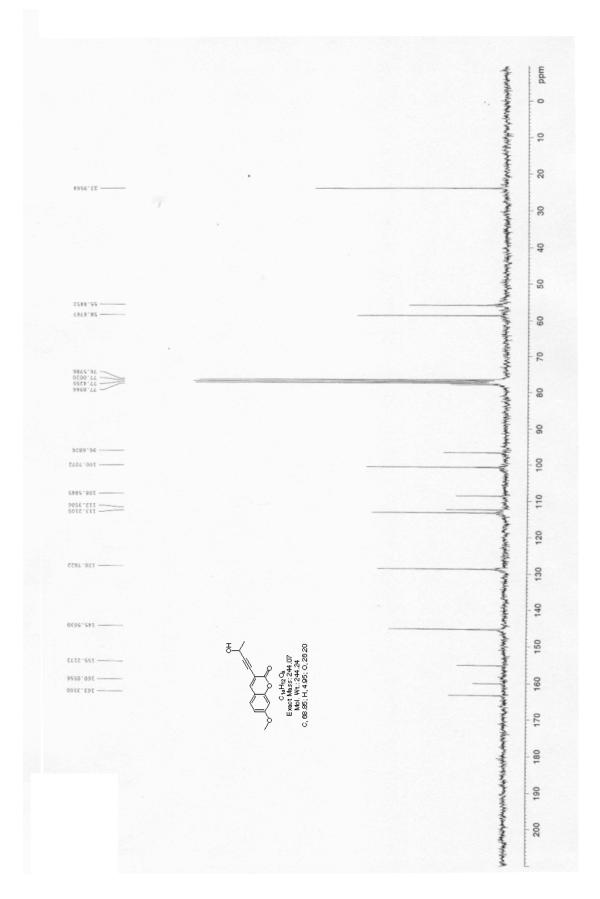




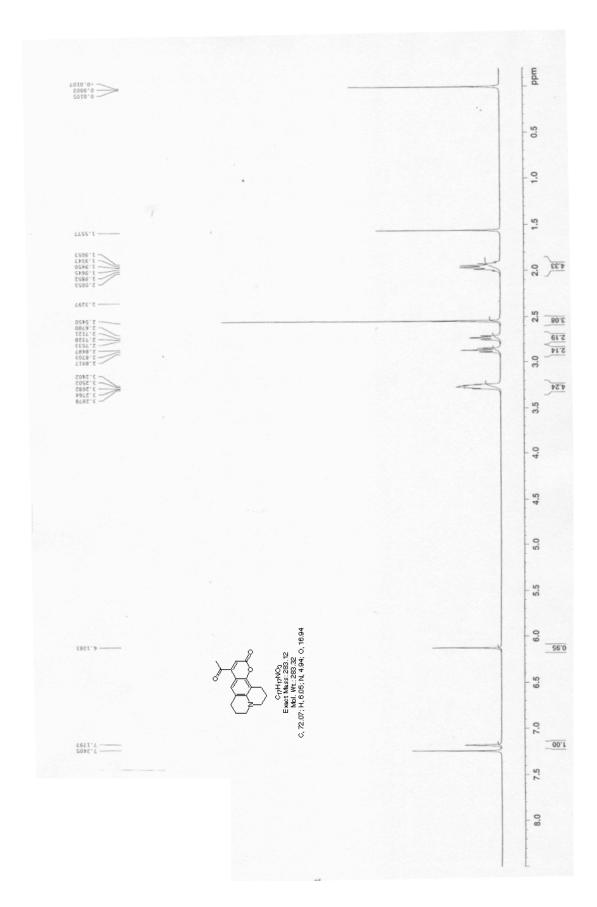


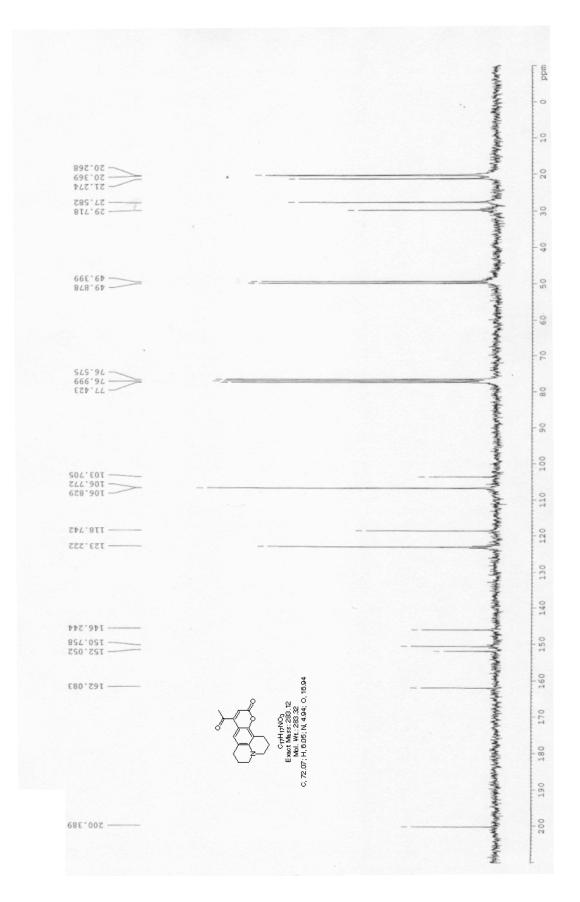


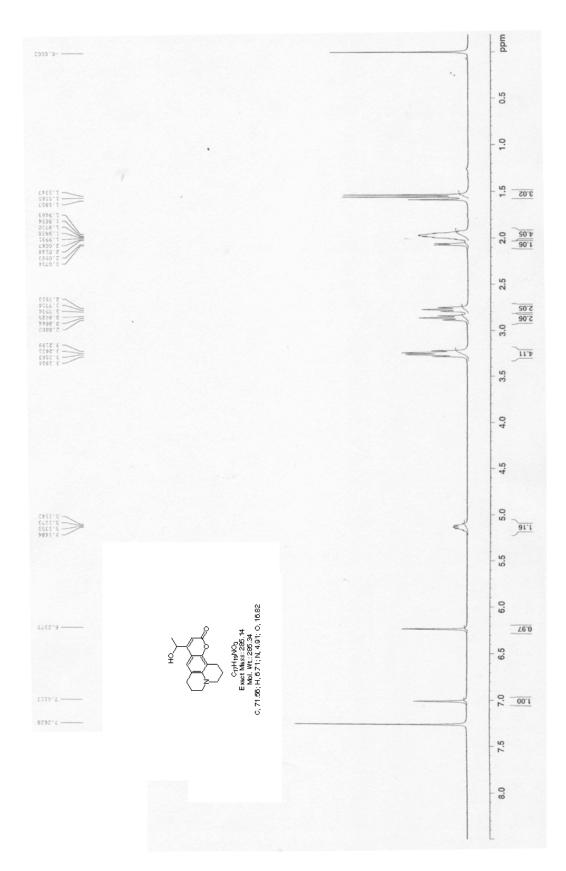


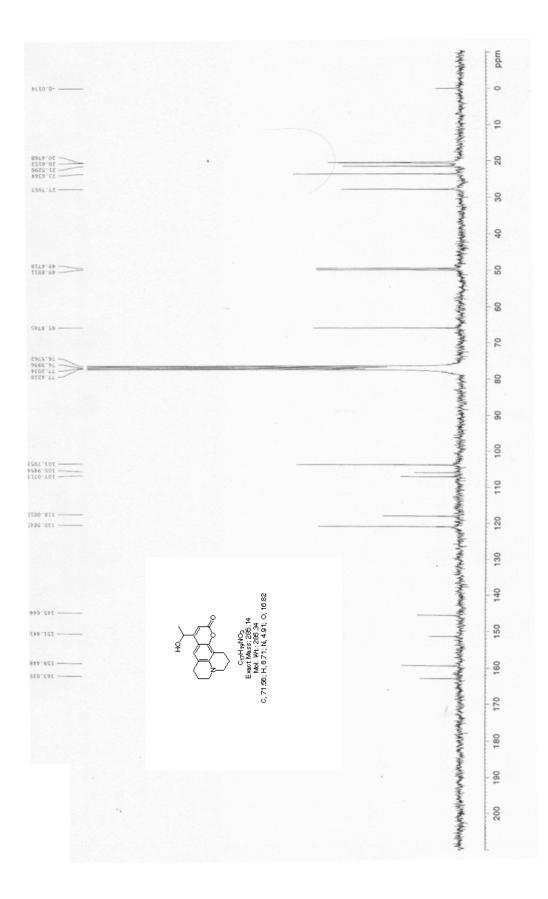


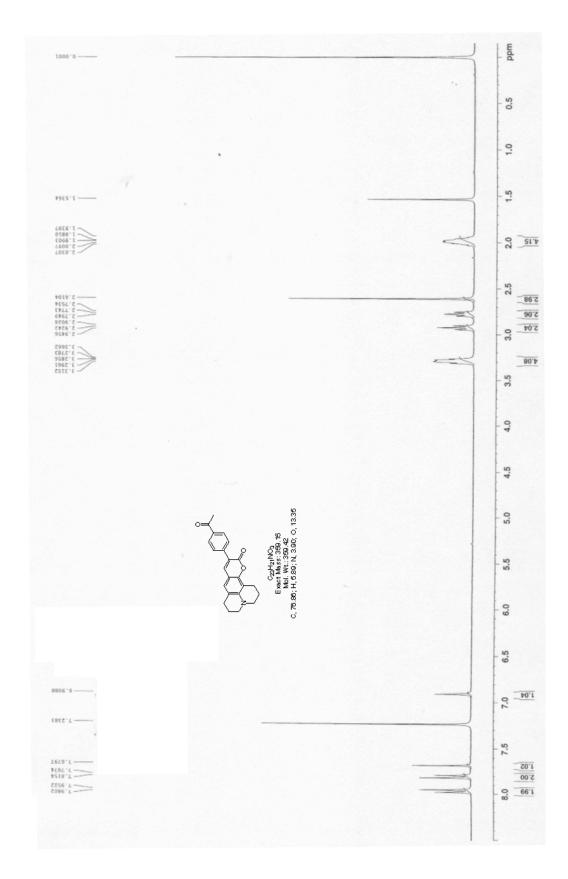
S34

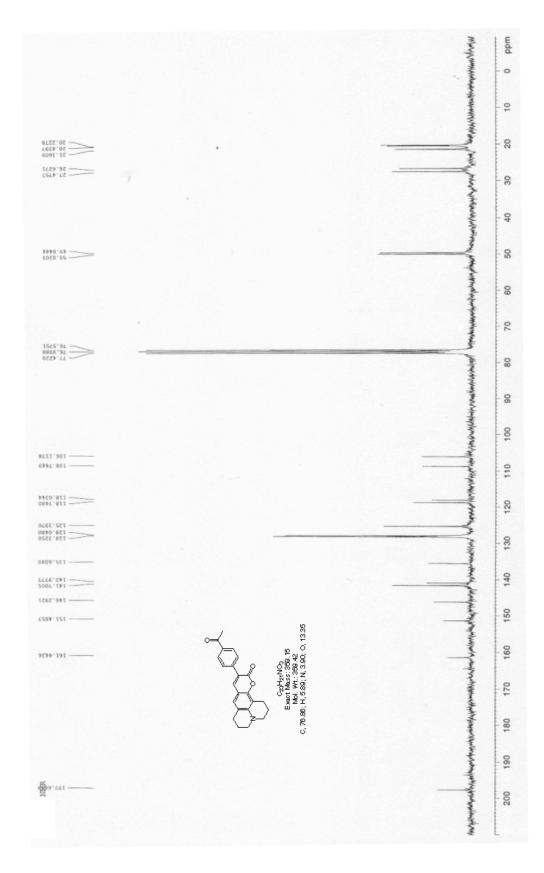




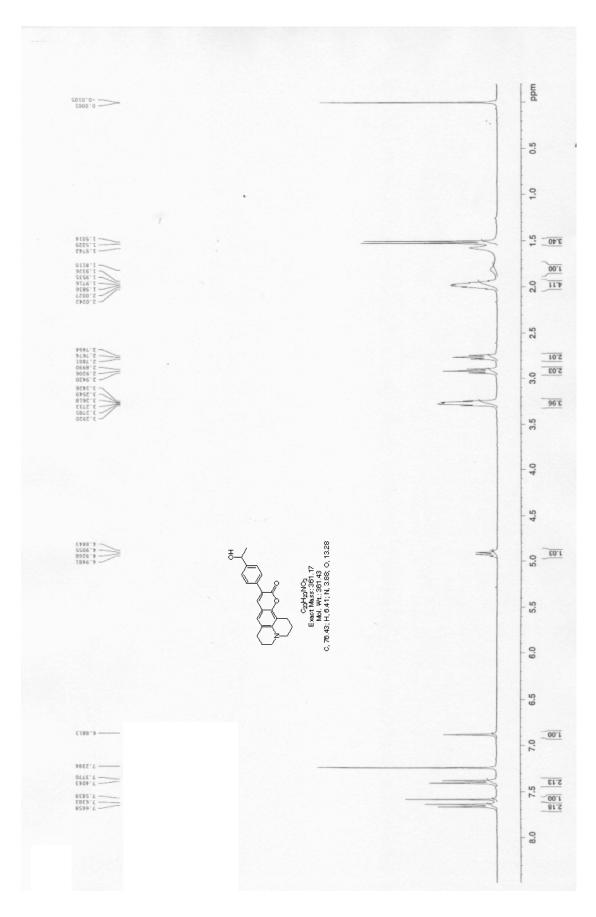


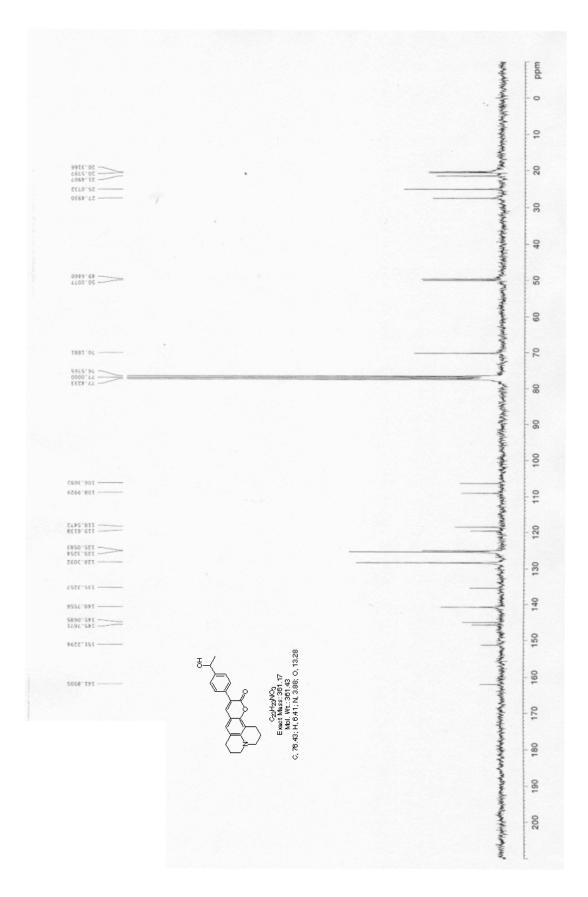


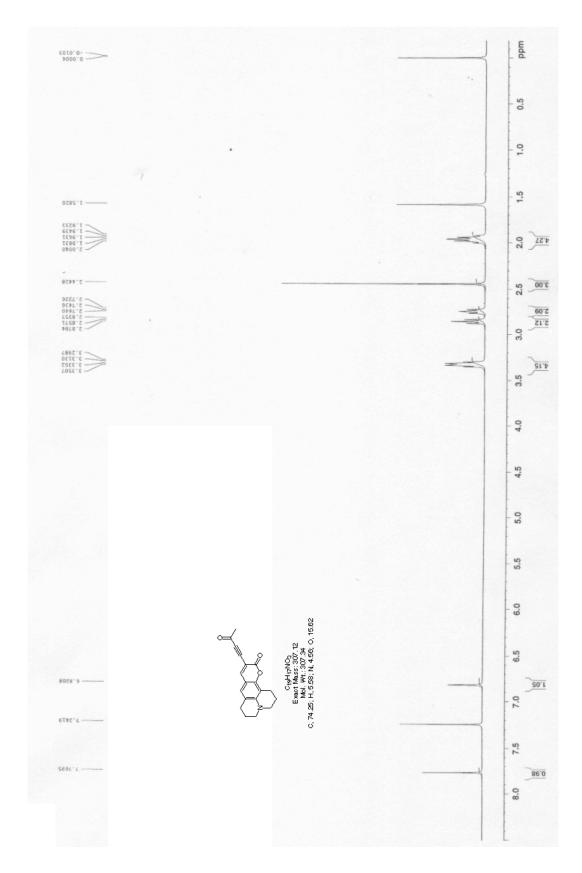


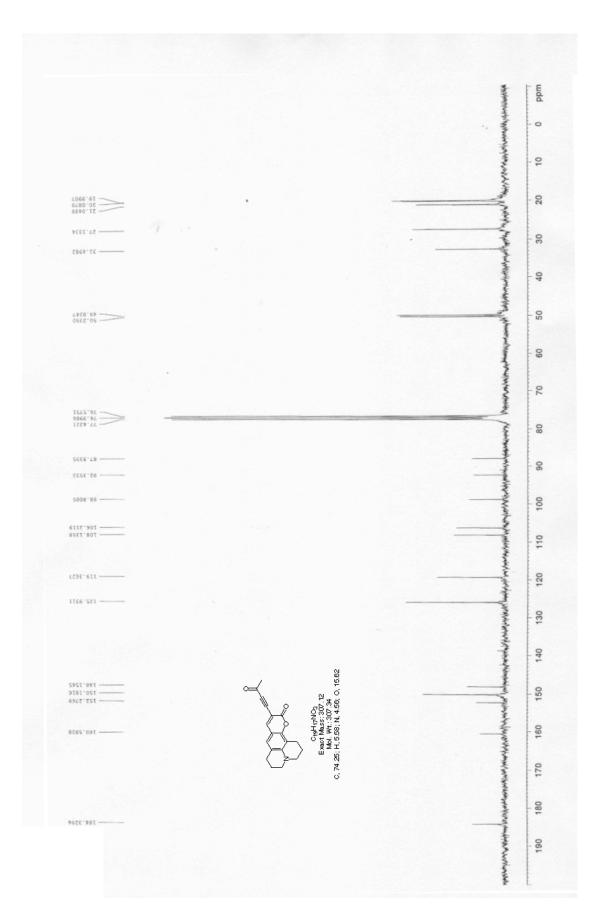


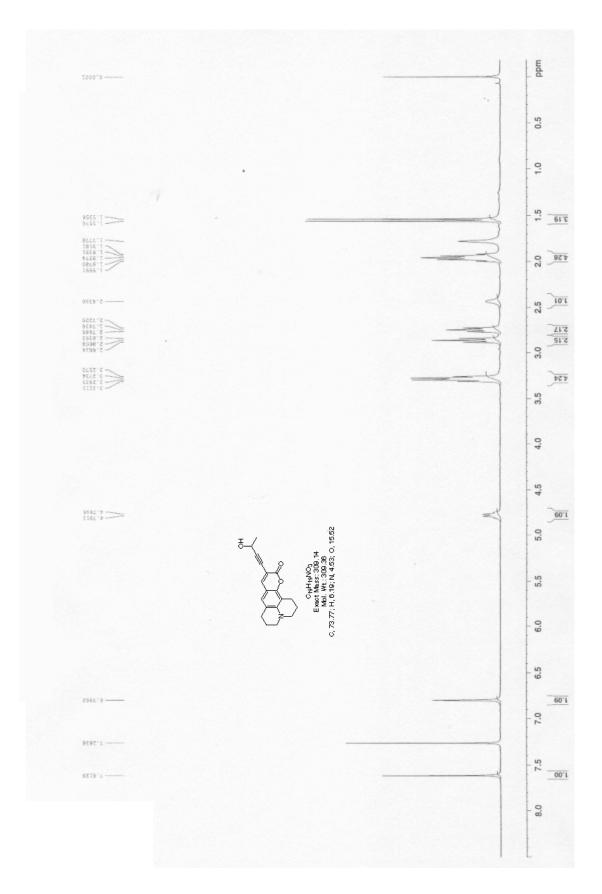
S40

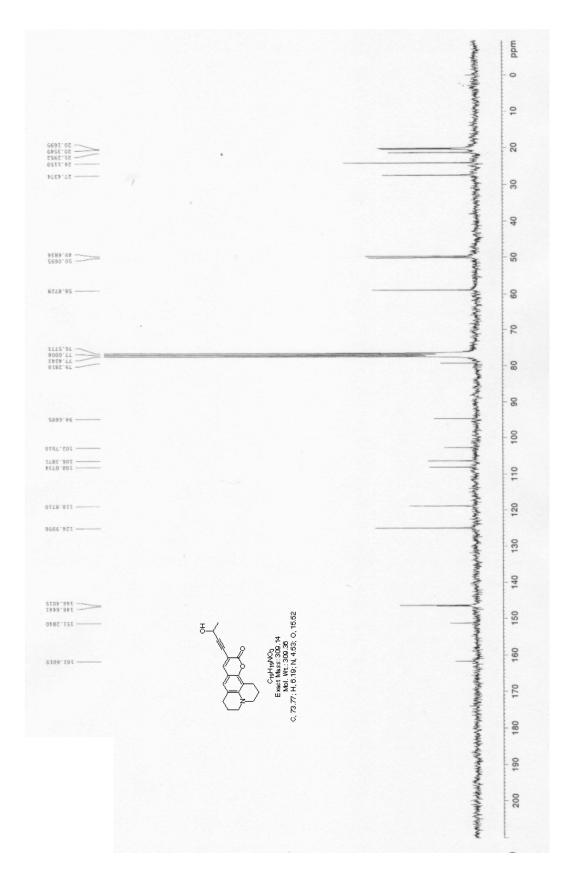












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