Synthesis of [Gly-1]RA-VII, [Gly-2]RA-VII and [Gly-4]RA-VII. Glycine-containing Analogues of RA-VII, an Antitumor Bicyclic Hexapeptide from *Rubia* Plants

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Supporting Information

Experimental details	S2 - S22
¹ H NMR spectra of 4–6, 10, 14–16, and 18–36	S23 – S48
X-ray data of 14	S49 – S59

Experimental

General. Melting points were recorded uncorrected. NMR spectra were recorded at 300 K unless otherwise specified. ¹H Chemical shifts in CDCl₃, CD₃OD or C₅D₅N were referenced to the residual CHCl₃ (7.26 ppm), CD₂HOD (3.31 ppm) or C₅D₄HN (7.21 ppm); ¹³C chemical shifts were referenced to the solvent (CDCl₃, 77.03 ppm; CD₃OD, 49.0 ppm; C₅D₅N, 135.5 ppm) unless otherwise stated. Preparative HPLC was performed by using a pre-packed ODS column (20 × 250 mm, 10 μm) and a UV detector.

[N-Methyl-Ala-2]RA-VII (9). Aqueous NaOH (50%, 3.5 mL) was added to a solution of 1 (1.02 g, 1.32 mmol), iodomethane (0.41 mL, 6.6 mmol) and tetrabutylammonium bromide (85 mg, 0.26 mmol) in dichloromethane (15 mL). After vigorous stirring for 8 h at room temperature, water (20 mL) was added to the reaction mixture. The organic layer was separated and the aqueous layer was extracted with dichloromethane (2 × 10 mL). The extracts and the organic layer were combined, washed successively with H₂O (5 mL), hydrochloric acid (2 M, 2 × 5 mL) and brine (5 mL), dried over Na₂SO₄ and filtered. The solvent was removed *in vacuo*, and the residue was subjected to column chromatography (silica gel, 15:2:1 CHCl₃/EtOAc/MeOH) to give 9^{18,21} (1.01 g, 97%) as a crystalline powder.

Bis(thioamide) 10. Davy-Reagent-Methyl (7) (187 mg, 0.658 mmol) was added to a solution of 9 (421 mg, 0.536 mmol) in dioxane (4 mL), and the mixture was stirred at room temperature for 4 days. Saturated aqueous NaHCO₃ (10 mL) was added to the mixture, and the whole was stirred at room temperature for 10 min, and then the mixture was extracted with CHCl₃ (3 × 30 mL). The combined CHCl₃ extracts were washed with brine (20 mL), dried over Na₂SO₄ and filtered, and the solvent removed *in vacuo*. Chromatography of the residue on alumina eluting with 15:2:1 CHCl₃/EtOAc/MeOH gave a mixture of thioamides, which was subjected to HPLC (3:1 MeOH/H₂O) to provide 10 (399 mg, 91%) as a crystalline powder: mp >300 °C; $[\alpha]^{23}_{\rm D}$ –132 (c = 0.22, CHCl₃); IR (KBr) $\nu_{\rm max}$ 3290, 3266, 2999, 2966,

2937, 1663, 1645, 1632, 1513, 1500, 1441, 1413, 1390, 1263, 1245, 1215, 1128, 1092, 1080, 1028, 971, 840 cm⁻¹; ¹H NMR (500 MHz, CDCl₃, mixture of conformers) major conformer δ 8.52 (d, 1H, J = 7.4 Hz), 8.17 (d, 1H, J = 7.0 Hz), 7.45 (dd, 1H, J = 8.4, 2.1 Hz), 7.28 (dd, 1H, J = 8.4, 2.1 Hz), 7.23 (dd. 1H, J = 8.4, 2.4 Hz), 7.07 (d-like, 2H, J = 8.6 Hz), 6.88 (dd. 1H, J =8.4, 2.4 Hz), 6.85 (d-like, 2H, J = 8.6 Hz), 6.79 (d, 1H, J = 8.4 Hz), 6.55 (dd, 1H, J = 8.4, 1.8 Hz), 5.42 (dd, 1H, J = 11.3, 2.7 Hz), 5.39 (m, 1H), 5.32 (m, 1H), 5.30 (m, 1H), 4.81 (dd, 1H, J= 11.8, 3.6 Hz), 4.34 (d, 1H, J = 1.8 Hz), 3.94 (s, 3H), 3.84 (dd, 1H, J = 13.6, 4.5 Hz), 3.80 (s, 3H), 3.78 (dd, 1H, J = 11.2, 4.5 Hz), 3.69 (dd, 1H, J = 11.3, 11.3 Hz), 3.53 (dd, 1H, J = 13.6, 11.2 Hz), 3.16 (s, 3H), 3.14 (dd, 1H, J = 18.3, 11.8 Hz), 3.08 (s, 3H), 2.89 (s, 3H), 2.81 (dd, 1H, J = 18.3, 3.6 Hz), 2.78 (s, 3H), 2.61 (dd, 1H, J = 11.3, 2.7 Hz), 1.41 (d, 3H, J = 7.1 Hz), 1.31 (d, 3H, J = 6.7 Hz), 1.09 (d, 3H, J = 6.7 Hz); ¹³C NMR (125 MHz, CDCl₃, mixture of conformers) major conformer δ 202.9, 198.0, 171.8, 171.4, 170.9, 169.3, 158.5, 158.0, 153.1, $146.4, 135.3, 132.6, 131.4, 130.9, 130.4 \times 2, 128.3, 126.0, 124.0, 120.7, 114.0 \times 2, 113.3,$ $112.2, 73.5, 63.9, 56.1, 55.3, 54.7, 51.2 \times 2, 47.8, 39.9, 38.4, 37.4, 35.9, 30.7, 30.5 \times 2, 16.5,$ 16.4, 14.6; FABMS m/z 817 [M+H]⁺; HR-FABMS m/z 817.3389 [M+H]⁺ (calcd for C₄₂H₅₃N₆O₇S₂, 817.3417). Anal. Calcd for C₄₂H₅₂N₆O₇S₂, C, 61.74; H, 6.42; N, 10.29. Found C, 61.62; H, 6.44; N, 10.18.

Cycloisodityrosine Thioester 14. Iodomethane (190 μ L, 3.05 mmol) and K₂CO₃ (212 mg, 1.53 mmol) were added to a solution of 10 (121.5 mg, 0.149 mmol) in acetone (3 mL), and the mixture was stirred at room temperature for 6 h. CHCl₃ (30 mL) was added to the mixture, and the insoluble matter was filtered off. After removal of the solvent under reduced pressure, the resulting residue was dissolved in MeCN (0.25 mL). After addition of hydrochloric acid (6 M, 0.25 mL), the solution was stirred at room temperature for 2 h, and then neutralized with aqueous K₂CO₃ (1 M) at 0 °C, to which phenyl isothiocyanate (210 μ L, 1.76 mmol) was added. The solution was stirred at room temperature for 2 h. The solution was extracted with CHCl₃ (3 × 10 mL), and the combined CHCl₃ extracts were washed with brine (5 mL), dried

over Na₂SO₄ and filtered, and the solvent removed in vacuo to give a residue, which was separated from the unreacted phenyl isothiocyanate by silica gel column chromatography eluting with CHCl₃. The 12:2:1 CHCl₃/EtOAc/MeOH eluate was evaporated to dryness, and the residue was dissolved in MeCN (0.3 mL). Hydrochloric acid (6 M, 0.25 mL) was added to the solution, and the mixture was stirred at room temperature for 5 h. The solution was neutralized with aqueous K₂CO₃ (1 M) at 0 °C, and then di-tert-butyl dicarbonate (145 mg, 0.664 mmol) was added to the solution. The solution was stirred at room temperature for 4 h. The solution was extracted with CHCl₃ (3 × 10 mL). The combined CHCl₃ extracts were washed with brine (5 mL), dried over Na₂SO₄ and filtered, and the solvent removed in vacuo. The residue was subjected to MPLC (silica gel, 7:7:1 hexane/CHCl₃/acetone) and then HPLC (3:2 MeCN/H₂O) to provide 14 (60.0 mg, 78%) as a crystalline powder, which was recrystallized from isopropyl ether/CHCl₃ to give colorless prisms: mp 182–184 °C; $[\alpha]^{23}$ _D -239 (c = 0.26, CHCl₃); IR (KBr) v_{max} 2977, 2927, 2835, 1692, 1683, 1643, 1513, 1498, 1438, 1335, 1312, 1265, 1259, 1227, 1206, 1173, 1149, 1126, 1034, 988, 802 cm⁻¹; ¹H NMR (500 MHz, CDCl₃, mixture of conformers) major conformer δ 7.47 (dd, 1H, J = 8.4, 2.2 Hz), 7.26 (dd, 1H, J = 8.4, 2.2 Hz), 7.17 (dd, 1H, J = 8.4, 2.2 Hz), 6.88 (dd, 1H, J = 8.4, 2.2 Hz), 6.80 (d, 1H, J = 8.3 Hz), 6.59 (br d, 1H, J = 8 Hz), 4.91 (dd, 1H, J = 11.2, 2.6 Hz), 4.76 (dd, 1H, J = 11.2, 4.76 (dd, 1H, J = 11.2, 4.76 (dd, 1H, J = 11.2), 4.76 (dd, 1 12.2, 3.7 Hz), 4.36 (br s, 1H), 3.94 (s, 3H), 3.63 (t, 1H, J = 11.2 Hz), 3.33 (dd, 1H, J = 17.8, 3.7 Hz), 2.91 (m, 1H), 2.87 (s, 3H), 2.72 (dd, 1H, J = 11.2, 2.6 Hz), 2.61 (s, 3H), 2.20 (s, 3H), 1.43 (s, 9H); EIMS m/z 514 [M]⁺; HR-EIMS m/z 514.2124 [M]⁺ (calcd for $C_{27}H_{34}N_2O_6S$, 514.2138).

Methyl Ester 15. A mixture of aqueous H₂O₂ (35%, 0.14 mL) and a LiOH solution (LiOH·H₂O 10.1 mg, 0.241 mmol in H₂O 0.3 mL) was slowly added to a cooled (0 °C) solution of 14 (30.9 mg, 0.060 mmol) in a mixture of 3:1 THF/H₂O (1.5 mL), and the solution was stirred at this temperature for 20 min. Saturated aqueous Na₂SO₃ (0.3 mL) was added to the solution, and after stirring at 0 °C for 20 min, aqueous citric acid (10%, 0.75 mL) was added to the mixture, and the mixture was extracted with CHCl₃ (3 × 5 mL). The combined CHCl₃ extracts were washed with brine (5 mL), dried over Na₂SO₄ and filtered, and the

solvent removed in vacuo. The residue was dissolved in a mixture of 10:1 MeCN/MeOH (0.9 mL), to which a solution of (trimethylsilyl)diazomethane in hexanes (2 M, 0.3 mL, 0.6 mmol) was added, and the whole was stirred at room temperature for 3 h. After addition of AcOH (0.28 mL) to the solution, the solvent was removed in vacuo. The residue was subjected to MPLC (silica gel, 20:20:1 hexane/CHCl₃/MeOH) to provide 15 (28.9 mg, 97%) as a colorless gummy solid: $[\alpha]^{18}_{D}$ -198 (c = 0.18, CHCl₃); IR (film) v_{max} 2975, 2931, 1747, 1685, 1650, 1585, 1518, 1500, 1445, 1367, 1313, 1266, 1210, 1143, 1130, 1094, 1028, 839 cm⁻¹; ¹H NMR (500 MHz, CDCl₃, mixture of conformers) major conformer δ 7.45 (dd. 1H, J = 8.4, 1.9 Hz). 7.26 (dd, 1H, J = 8.4, 1.9 Hz), 7.17 (dd, 1H, J = 8.4, 2.3 Hz), 6.88 (dd, 1H, J = 8.4, 2.3 Hz), 6.80 (d, 1H, J = 8.3 Hz), 6.59 (br d, 1H, J = 8 Hz), 4.90 (dd, 1H, J = 11.2, 2.8 Hz), 4.69 (dd, 1H, J = 12.2, 3.8 Hz), 4.42 (s, 1H), 3.94 (s, 3H), 3.67 (s, 3H), 3.64 (m, 1 H), 3.32 (dd. 1H, J =18.0, 3.8 Hz), 2.92 (s, 3 H), 2.92 (m, 1 H), 2.72 (dd, 1 H, J = 11.3, 2.8 Hz), 2.56 (s, 3H), 1.47 (s, 9H); ¹³C NMR (125 MHz, CDCl₃, mixture of conformers) major conformer δ 171.4, 171.0, 157.8, 155.6, 153.0, 146.4, 136.2, 132.7, 131.1, 128.2, 126.0, 124.0, 121.0, 113.5, 112.2, 80.2, 57.2, 56.9, 56.2, 52.6, 37.4, 32.9, 29.8, 28.5 \times 3, 28.2; EIMS m/z 498 [M]⁺; HR-EIMS m/z $498.2363 \text{ [M]}^+ \text{ (calcd for C}_{27}\text{H}_{34}\text{N}_2\text{O}_7, 498.2366).$

Benzyl Ester 16. To a cooled (0 °C) solution of 14 (30.0 mg, 0.0583 mmol) in 3:1 THF/H₂O (1.5 mL) was slowly added a mixture of aqueous H₂O₂ (35%, 0.14 mL) and a LiOH solution (LiOH·H₂O 10.8 mg, 0.257 mmol in H₂O 0.3 mL), and the solution was stirred at this temperature for 20 min. Saturated aqueous Na₂SO₃ (0.3 mL) was added to the solution, and the mixture was stirred at 0 °C for 20 min. Aqueous citric acid (10%, 0.75 mL) was added to the mixture, and the whole was extracted with CHCl₃ (3 × 5 mL). The combined CHCl₃ extracts were washed with brine (5 mL) dried over Na₂SO₄ and filtered, and the solvent removed *in vacuo*. The residue, triphenylphosphine (30.7 mg, 0.117 mmol) and benzyl alcohol (15.0 μL, 0.145 mmol) were dissolved in THF (0.9 mL), to which diethyl azodicarboxylate (20.0 μL, 0.127 mmol) was added slowly at 0 °C under an atmosphere of argon, and the solution was stirred at this temperature for 2 h. The solvent was removed *in vacuo*. The residue was subjected to MPLC (silica gel, 60.1 CHCl₃/acetone) to provide 16

(30.2 mg, 90%) as a colorless gummy solid: $[\alpha]^{18}_{D}$ –202 (c = 0.10, CHCl₃); IR (film) v_{max} 2975, 2932, 1744, 1683, 1651, 1585, 1518, 1500, 1445, 1368, 1314, 1266, 1218, 1163, 1143, 1130, 1093, 1028, 982, 840 cm⁻¹; ¹H NMR (500 MHz, CDCl₃, mixture of conformers) major conformer δ 7.46 (dd, 1H, J = 8.3, 1.8 Hz), 7.37–7.23 (m, 6H), 7.18 (dd, 1H, J = 8.3, 2.2 Hz), 6.88 (dd, 1H, J = 8.4, 2.2 Hz), 6.78 (d, 1H, J = 8.3 Hz), 6.56 (br d, 1H, J = 8 Hz), 5.13 (d, 1H, J = 12.3 Hz), 5.02 (d, 1H, J = 12.3 Hz), 4.94 (dd, 1H, J = 11.3, 2.7 Hz), 4.75 (dd, 1H, J = 12.2, 3.6 Hz), 4.42 (s, 1H), 3.93 (s, 3H), 3.63 (t, 1H, J = 11.3 Hz), 3.32 (dd, 1H, J = 17.9, 3.6 Hz), 2.90 (s, 3H), 2.89 (dd, 1H, J = 17.9, 12.2 Hz), 2.73 (dd, 1H, J = 11.3, 2.7 Hz), 2.54 (s, 3H), 1.48 (s, 9H); ¹³C NMR (125 MHz, CDCl₃, mixture of conformers) major conformer δ 171.0, 170.8, 157.9; 155.6, 153.0, 146.4, 136.1, 135.3, 132.7, 131.1, 128.6 × 2, 128.4, 128.2, 128.1 × 2, 126.0, 124.1, 121.0, 113.6, 112.2, 80.1, 67.3, 57.3, 56.8, 56.2, 37.4, 32.9, 29.8, 28.6 × 3, 28.3; EIMS m/z 574 [M]⁺; HR-EIMS m/z 574.2695 [M]⁺ (calcd for C₃₃H₃₈N₂O₇, 574.2679).

Cbz-N-Methyl-Tyr-Ala-OMe (18). Cbz-N-Methyl-Tyr-OH (17)^{12a,b} (1.00 g. 3.04 mmol). H-Ala-OMe·HCl (775 mg, 5.55 mmol) and HOBt (585 mg, 4.33 mmol) were dissolved in CH₂Cl₂ (15 mL) to prepare a solution, to which triethylamine (0.78 mL, 5.60 mmol) and a solution of EDC (776 mg, 4.05 mmol) in CH₂Cl₂ (15 mL) were slowly added. The mixture was stirred at room temperature for 16 h, and then hydrochloric acid (1 M, 30 mL) was added. The whole was extracted with CHCl₃ (3 × 30 mL), and the combined CHCl₃ extracts were washed successively with saturated aqueous NaHCO₃ (30 mL) and brine (30 mL), dried over Na₂SO₄ and filtered, and the solvent removed in vacuo. The residue was subjected to column chromatography (silica gel, 2:1 EtOAc/hexane) and then MPLC (silica gel, 2:1 EtOAc/hexane) to afford 18 (894 mg, 71%) as an amorphous solid: $\left[\alpha\right]_{D}^{28}$ -58.0 (c = 1.35, $CHCl_3$); IR (film) v_{max} 3323, 3017, 2954, 1744, 1669, 1615, 1596, 1518, 1455, 1402, 1311. 1220, 1182, 1147, 1104, 1060, 998, 828, 811, 756 cm⁻¹; ¹H NMR (400 MHz, CDCl₃, 297 K, mixture of rotamers) δ 7.37–7.20 (br m, 5H), 7.06–6.90, 6.70–6.58 (each br m, total 4H), 6.30, 6.00 (each br m, total 1H), 5.11 (br s, 2H), 5.10–4.68 (br m, 2H), 4.51 (br quintet, 1H, J = 7Hz), 3.71 (s, 3H), 3.28–3.18 (m, 1H), 2.97–2.82 (m, 1H), 2.86 (s, 3H), 1.33 (br d, 3H, J = 7Hz); EIMS m/z 397 [M-OH]⁺; FABMS m/z 415 [M+H]⁺; HR-EIMS m/z 397.1749 [M-OH]⁺

(calcd for C₂₂H₂₅N₂O₅, 397.1763).

Cbz-*N*,*O*-Dimethyl-Tyr-Ala-OMe (19). A solution of 18 (817 mg, 1.97 mmol) in MeOH (5 mL) was treated with diazomethane in diethyl ether solution and was left standing until the reaction completed. The solvent was removed *in vacuo*, and the residue was subjected to column chromatography (silica gel, 1:1 hexane/EtOAc) to afford 19 (844 mg, 100%) as an amorphous solid: $[\alpha]^{29}_{D}$ –57.7 (c = 1.0, CHCl₃); IR (film) v_{max} 3328, 2953, 2837, 1745, 1675, 1613, 1515, 1455, 1401, 1306, 1249, 1213, 1180, 1146, 1035, 1000, 825, 798, 754 cm⁻¹; ¹H NMR (400 MHz, CDCl₃, 297 K, mixture of rotamers) δ 7.37–6.98, 6.83–6.72, 6.55, 6.27 (each br m, total 10H), 5.17–4.85, 4.74 (each br m, total 3H), 4.52 (quintet, 1H, J = 7.2 Hz), 3.77 (s, 3H), 3.72 (s, 3H), 3.33–3.20 (br m, 1H), 3.01–2.87 (br m, 1H), 2.84 (s, 3H), 1.34 (br d, 3H, J = 7 Hz); EIMS m/z 397 [M–OMe]⁺; HR-EIMS m/z 397.1749 [M–OMe]⁺ (calcd for $C_{22}H_{25}N_2O_5$, 397.1763).

Boc-Ala-N,O-dimethyl-Tyr-Ala-OMe (20). A solution of 19 (612 mg, 1.43 mmol) in MeOH (10 mL) was stirred at room temperature under an atmosphere of hydrogen for 2 h in the presence of 10% Pd/C (53 mg) and hydrochloric acid (0.1 mL). The catalyst was filtered off, and the filtrate was concentrated to dryness. The residue was dissolved in CH₂Cl₂ (10 mL) together with Boc-Ala-OH (540 mg, 2.85 mmol) and PyBOP (1.49 g, 2.86 mmol), to which N,N-diisopropylethylamine (1.0 mL, 5.74 mmol) was slowly added at -20 °C under an atmosphere of argon. The mixture was stirred at this temperature for 1 h and then at room temperature for 5 days. Aqueous citric acid (10%, 10 mL) was added to the mixture, and the whole was extracted with CHCl₃ (3 × 30 mL). The combined CHCl₃ extracts were washed successively with saturated aqueous NaHCO₃ (10 mL) and brine (10 mL), dried over Na₂SO₄ and filtered, and the solvent removed in vacuo. The residue was subjected to MPLC (silica gel, 10:10:1, hexane/CHCl₃/MeOH) to afford 20 (587 mg, 88%) as a crystalline powder, which was recrystallized from isopropyl ether to give colorless prisms: mp 127–128 °C; [α]²⁰_D –119 $(c = 1.01, CHCl_3)$; IR (KBr) v_{max} 3355, 3295, 2985, 2951, 1746, 1687, 1663, 1535, 1516, 1458, 1366, 1296, 1249, 1214, 1177, 1167, 1087, 1073, 1023, 821 cm⁻¹; ¹H NMR (400 MHz, CDCl₃, mixture of two rotamers) major rotamer δ 8.22 (d, 1H, J = 7.3 Hz), 7.02 (d-like, 2H, J = 8.6 Hz), 6.79 (d-like, 2H, J = 8.6 Hz), 4.99 (d, 1H, J = 6.9 Hz), 4.78 (dd, 1H, J = 11.2, 3.4 Hz), 4.53 (quintet, 1H, J = 7.3 Hz), 4.13 (quintet, 1H, J = 6.7 Hz), 3.72 (s, 3H), 3.71 (s, 3H), 3.12 (dd, 1H, J = 14.7, 3.4 Hz), 2.96 (dd, 1H, J = 14.7, 11.2 Hz), 2.87 (s, 3H), 1.38 (d, 3H, J = 7.0 Hz), 1.35 (s, 9H), 0.34 (d, 3H, J = 6.7 Hz); minor rotamer δ 7.08 (d-like, 2H, J = 8.6 Hz), 6.77 (d-like, 2H, J = 8.6 Hz), 6.56 (d, 1H, J = 7.1 Hz), 5.26 (d, 1H, J = 7.9 Hz), 5.14 (t-like, 1H, J = 7.8 Hz), 4.53 (quintet, 1H, J = 7.3 Hz), 4.46 (quintet, 1H, J = 7.2 Hz), 3.73 (s, 3H), 3.68 (s, 3H), 3.23 (dd, 1H, J = 14.5, 7.8 Hz), 2.95 (dd, 1H, J = 14.5, 8.4 Hz), 2.93 (s, 3H), 1.35 (s, 9H), 1.31 (d, 3H, J = 7.1 Hz), 1.21 (d, 3H, J = 6.7 Hz); ¹³C NMR (100 MHz, CDCl₃, mixture of two rotamers) major rotamer δ 174.1, 172.9, 169.0, 158.6, 156.0, 130.4 × 2, 129.6, 114.3 × 2, 80.3, 62.3, 55.2, 52.1, 48.3, 44.9, 33.1, 29.0, 28.2 × 3, 17.4, 16.4; minor rotamer δ 173.7, 172.7, 169.2, 158.3, 154.9, 129.8 × 2, 128.7, 113.9 × 2, 79.5, 58.6, 55.1, 52.3, 48.0, 46.6, 32.7, 31.6, 28.3 × 3, 18.3, 18.0; EIMS m/z 465 [M]⁺; HR-EIMS m/z 465.2460 [M]⁺ (calcd for C₂₃H₃₅N₃O₇, 465.2475). Anal. Calcd for C₂₃H₃₅N₃O₇, C, 59.34; H, 7.58; N, 9.03. Found C, 59.14; H, 7.56; N, 8.98.

Boc-Gly-Ala-*N*,*O*-dimethyl-Tyr-Ala-OMe (21). A solution of 20 (353 mg, 0.758 mmol) in TFA (1.5 mL) was stirred at room temperature for 2 h. TFA was removed *in vacuo*, and the residue was dissolved in CHCl₃ (30 mL). The solution was washed successively with saturated aqueous NaHCO₃ (10 mL) and brine (10 mL), dried over Na₂SO₄ and filtered, and the solvent removed *in vacuo*. The residue was dissolved in CH₂Cl₂ (5 mL) together with Boc-Gly-OH (201 mg, 1.15 mmol) and HOBt (154 mg, 1.14 mmol), to which EDC (218 mg, 1.14 mmol) was added at -20 °C. The mixture was stirred at this temperature for 1 h, and then at room temperature for 24 h. Saturated aqueous NaHCO₃ (10 mL) was added to the solution, and the mixture was extracted with CHCl₃ (3 × 10 mL). The combined CHCl₃ extracts were washed with brine (10 mL), dried over Na₂SO₄ and filtered, and the solvent removed *in vacuo*. The residue was subjected to MPLC (silica gel, 5:5:1 hexane/CHCl₃/MeOH) to provide 21 (376 mg, 95%) as an amorphous solid: [α]²³_D –103 (c = 1.84, CHCl₃); IR (film) v_{max} 3280, 2980, 2937, 1747, 1711, 1667, 1631, 1540, 1515, 1456, 1367, 1284, 1248, 1175, 1033, 755 cm⁻¹; ¹H NMR (400 MHz, CDCl₃, mixture of two rotamers) major rotamer δ 8.21 (d, 1H, J =

6.7 Hz), 7.45 (br s, 1H), 7.00 (d-like, 2H, J = 8.6 Hz), 6.78 (d-like, 2H, J = 8.6 Hz), 5.46 (br m, 1H), 4.92 (br d, 1H, J = 9 Hz), 4.42 (quintet, 1H, J = 7.2 Hz), 4.35 (quintet, 1H, J = 6.4Hz), 3.71 (m, 1H), 3.71 (s, 3H), 3.70 (s, 3H), 3.64 (dd, 1H, J = 17.2, 5.8 Hz), 3.08 (dd, 1H, J = 17.2, 5.8 Hz), 3.0 = 14.6, 3.2 Hz), 2.95 (dd, 1H, J = 14.6, 11.4 Hz), 2.84 (s, 3H), 1.36 (s, 9H), 1.35 (d, 3H, J = 14.6, 11.4 Hz) 7.0 Hz), 0.42 (d, 3H, J = 6.7 Hz); ¹³C NMR (100 MHz, CDCl₃, mixture of two rotamers) major rotamer δ 173.6, 173.1, 170.2, 168.8, 158.7, 155.9, 130.3 \times 2, 129.4, 114.3 \times 2, 79.8, 62.4, 55.3, 52.2, 48.5, 44.2, 43.1, 33.2, 29.1, 28.2 \times 3, 17.2, 16.2; EIMS m/z 522 [M]⁺; FABMS m/z 523 [M+H]⁺; HR-EIMS m/z 522.2682 [M]⁺ (calcd for $C_{25}H_{38}N_4O_8$, 522.2690). Boc-Gly-Ala-N,O-dimethyl-Tyr-Ala-OH (22). To a solution of 21 (224 mg, 0.429 mmol) in a mixture of 3:1:1 THF/MeOH/H₂O (4.2 mL) was added LiOH·H₂O (35.9 mg, 0.856 mmol), and the mixture was stirred at room temperature for 3 h. The mixture was acidified with aqueous citric acid (10%, 2 mL) and extracted with CHCl₃ (3 × 10 mL). The combined CHCl₃ extracts were washed with brine (10 mL), dried over Na₂SO₄ and filtered, and the solvent removed in vacuo. The residue was subjected to column chromatography (silica gel, 5:1 CHCl₃/MeOH) to provide **22** (216 mg, 99%) as an amorphous solid: $[\alpha]^{28}$ _D -97.1 (c =0.15, MeOH); IR (film) v_{max} 3288, 2980, 2936, 2838, 1708, 1663, 1634, 1514, 1456, 1411, 1367, 1300, 1285, 1249, 1176, 1033, 943, 756 cm⁻¹; ¹H NMR (500 MHz, CD₃OD, mixture of two rotamers) major rotamer δ 7.14 (d-like, 2H, J = 8.6 Hz), 6.87 (d-like, 2H, J = 8.6 Hz), 5.03 (dd, 1H, J = 11.3, 3.2 Hz), 4.44 (q, 1H, J = 6.8 Hz), 4.34 (q, 1H, J = 7.1 Hz), 3.75 (s, 3H), 3.69 (d, 1H, J = 17.2 Hz), 3.64 (d, 1H, J = 17.2 Hz), 3.15 (dd, 1H, J = 14.5, 3.2 Hz), 2.97 (dd, 1H, J = 17.2 Hz), 3.64 (d, 1H, J = 17.2 Hz), 3.64 (d, 1H, J = 17.2 Hz), 3.64 (d, 1H, J = 17.2 Hz), 3.65 (dd, 1H, J = 17.2 Hz),1H, J = 14.5, 11.3 Hz), 2.87 (s, 3H), 1.45 (d, 3H, J = 7.1 Hz), 1.42 (s, 9H), 0.47 (d, 3H, J = 7.1 Hz), 1.42 (s, 9H), 1.4 6.8 Hz); minor rotamer δ 7.13 (d-like, 2H, J = 8.6 Hz), 6.83 (d-like, 2H, J = 8.6 Hz,), 4.96 (m, 1H), 4.71 (q, 1H, J = 6.8 Hz), 4.32 (q, 1H, J = 7.2 Hz), 3.76 (s, 3H), 3.69 (d, 1H, J = 17.2 Hz), 3.64 (d, 1H, J = 17.2 Hz), 3.26 (dd, 1H, J = 14.5, 5.8 Hz), 3.04 (dd, 1H, J = 14.5, 10.2 Hz), 2.97 (s, 3H), 1.45 (s, 9H), 1.37 (d, 3H, J = 7.2 Hz), 1.24 (d, 3H, J = 6.8 Hz); HR-ESIMS m/z

 $509.2653 \, [M+H]^{+} \, (calcd for C_{24}H_{37}N_4O_8, 509.2611).$

Boc-Gly-*N*, *O*-dimethyl-Tyr-Ala-OMe (23). When 19 was treated as described for the preparation of 20 by using Boc-Gly-OH, 23 was obtained in 96% yield as an amorphous solid: $[\alpha]^{20}_D$ –55.7 (c = 0.87, CHCl₃); IR (film) v_{max} 3320, 2979, 1744, 1714, 1651, 1515, 1456, 1367, 1250, 1174, 1052, 1034, 756 cm⁻¹; ¹H NMR (400 MHz, CDCl₃, mixture of two rotamers) major rotamer δ 7.08 (d-like, 2H, J = 8.6 Hz), 6.79 (d-like, 2H, J = 8.6 Hz), 6.54 (d, 1H, J = 7.2 Hz), 5.39 (br s, 1H), 5.27 (dd, 1H, J = 9.2, 6.9 Hz), 4.47 (quintet, 1H, J = 6.2 Hz), 3.92 (dd, 1H, J = 17.2, 4.5 Hz), 3.76 (s, 3H), 3.75 (dd, 1H, J = 17.2, 4.7 Hz), 3.70 (s, 3H), 3.22 (dd, 1H, J = 14.6, 6.9 Hz), 2.93 (dd, 1H, J = 14.6, 9.2 Hz), 2.87 (s, 3H), 1.42 (s, 9H), 1.35 (d, 3H, J = 7.2 Hz); ¹³C NMR (100 MHz, CDCl₃, mixture of two rotamers) major rotamer δ 172.7, 169.8, 169.4, 158.2, 155.6, 129.6 × 2, 128.5, 113.8 × 2, 79.5, 57.9, 55.0, 52.2, 48.0, 42.4, 33.1, 28.2 × 3, 22.7, 17.7; EIMS m/z 451 [M]⁺; HR-EIMS m/z 451.2309 [M]⁺ (calcd for C₂₂H₃₃N₃O₇, 451.2319).

Boc-D-Ala-Gly-*N*,*O*-dimethyl-Tyr-Ala-OMe (24). When 23 was treated as described for the preparation of 21 by using Boc-D-Ala-OH, 24 was obtained in 84% yield as an amorphous solid: $[α]^{23}_D$ –47.1 (c = 0.99, CHCl₃); IR (film) v_{max} 3318, 2980, 2936, 2838, 1744, 1646, 1515, 1455, 1367, 1301, 1249, 1217, 1168, 1111, 1032, 797 cm⁻¹; ¹H NMR (400 MHz, CDCl₃, mixture of two rotamers) major rotamer δ 7.06 (d-like, 2H, J = 8.6 Hz), 7.03 (br m, 1H), 6.78 (d-like, 2H, J = 8.6 Hz), 6.72 (br d, 1H, J = 7 Hz), 5.31 (dd, 1H, J = 9.5, 6.5 Hz), 5.16 (d, 1H, J = 7.5 Hz), 4.49 (quintet, 1H, J = 7.2 Hz), 4.26 (br m, 1H), 4.06 (dd, 1H, J = 17.7, 4.3 Hz), 3.81 (dd, 1H, J = 17.7, 2.7 Hz), 3.76 (s, 3H), 3.70 (s, 3H), 3.22 (dd, 1H, J = 14.5, 6.5 Hz), 2.94 (dd, 1H, J = 14.5, 9.5 Hz), 2.93 (s, 3H), 1.43 (s, 9H), 1.36 (d, 3H, J = 7.1 Hz), 1.34 (d, 3H, J = 6.9 Hz); ¹³C NMR (100 MHz, CDCl₃, mixture of two rotamers) major rotamer δ 173.0, 172.8, 169.6, 169.3, 158.4, 155.3, 129.7 × 2, 128.6, 114.0 × 2, 80.0, 57.9, 55.2, 52.4, 50.0, 48.1, 41.5, 33.5, 30.5, 28.3 × 3, 19.0, 17.8; EIMS m/z 522 [M]⁺; FABMS m/z 523 [M+H]⁺; HR-EIMS m/z 522.2696 [M]⁺ (calcd for C₂₅H₃₈N₄O₈, 522.2690).

Boc-D-Ala-Gly-*N*, *O*-dimethyl-Tyr-Ala-OH (25). When 24 was processed as described for the preparation of 22, 25 was obtained in 98% yield as an amorphous solid: $[\alpha]^{28}_{D}$ –15.7 (c = 0.31, MeOH); IR (film) v_{max} 3320, 2979, 2936, 2838, 1647, 1514, 1456, 1367, 1301, 1249,

1168, 1032, 756 cm⁻¹; ¹H NMR (500 MHz, CD₃OD, mixture of two rotamers) major rotamer δ 7.14 (d-like, 2H, J = 8.6 Hz), 6.82 (d-like, 2H, J = 8.6 Hz), 5.23 (dd, 1H, J = 10.6, 5.5 Hz), 4.37 (q, 1H, J = 7.3 Hz), 4.09 (m, 1H), 4.06 (d, 1H, J = 17.2 Hz), 3.78 (d, 1H, J = 17.2 Hz), 3.75 (s, 3H), 3.25 (dd, 1H, J = 14.5, 5.5 Hz), 2.97 (dd, 1H, J = 14.5, 10.6 Hz), 2.91 (s, 3H), 1.44 (s, 9H), 1.39 (d, 3H, J = 7.3 Hz), 1.31 (d, 3H, J = 7.2 Hz); minor rotamer δ 7.15 (d-like, 2H, J = 8.6 Hz), 6.85 (d-like, 2H, J = 8.6 Hz), 4.65 (m, 1H), 4.40 (q, 1H, J = 7.2 Hz), 4.06 (m, 1H), 4.05 (d, 1H, J = 16.3 Hz), 3.76 (s, 3H), 3.22 (dd, 1H, J = 14.5, 4.1 Hz), 3.10 (br d, 1H, J = 16.3 Hz), 2.97 (dd, 1H, J = 14.5, 10.6 Hz), 2.92 (s, 3H), 1.44 (d, 3H, J = 7.2 Hz), 1.43 (s, 9H), 1.27 (d, 3H, J = 7.2 Hz); HR-ESIMS m/z 509.2653 [M+H]⁺ (calcd for C₂₄H₃₇N₄O₈, 509.2611).

Cbz-N-Methyl-Tyr-Gly-OMe (26). When 17 was treated as described for the preparation of 18 by using H-Gly-OMe·HCl, 26 was obtained in 79% yield as an amorphous solid: $[\alpha]^{28}_{D}$ -69.7 (c = 0.98, CHCl₃); IR (film) v_{max} 3341, 3029, 2953, 1752, 1670, 1615, 1596, 1518, 1452, 1403, 1369, 1319, 1217, 1182, 1141, 1028, 971, 756 cm⁻¹; ¹H NMR (400 MHz, CDCl₃, mixture of rotamers) δ 7.37–7.16, 7.07–6.90, 6.74–6.60, 6.53–6.30 (each br m, total 11H), 5.18–4.76 (br m, 3H), 4.14–3.98 (br m, 1H), 3.96–3.82 (br m, 1H), 3.71 (s, 3H), 3.34–3.21 (br m, 1H), 2.96–2.80 (br m, 1H), 2.89 (s, 3H); FABMS m/z 401 [M+H]⁺; EIMS m/z 400 [M]⁺; HR-EIMS m/z 400.1606 [M]⁺ (calcd for $C_{21}H_{24}N_{2}O_{6}$, 400.1634).

Cbz-*N*,*O*-Dimethyl-Tyr-Gly-OMe (27). When 26 was treated as described for the preparation of 19, 27 was obtained in 100% yield as an amorphous solid: $[\alpha]^{30}_{D}$ –68.7 (c = 1.0, CHCl₃); IR (film) v_{max} 3346, 2953, 2837, 1753, 1681, 1613, 1515, 1454, 1402, 1369, 1308, 1248, 1210, 1180, 1140, 1033, 738 cm⁻¹; ¹H NMR (400 MHz, CDCl₃, 297 K, mixture of rotamers) δ 7.36–7.00, 6.82–6.73 (each br m, total 9H), 6.44, 6.35 (each br m, total 1H), 5.17–4.94, 4.82 (each br m, total 3H), 4.16–4.00 (m, 1H), 3.88 (dd, 1H, J = 18.1, 4.7 Hz), 3.77 (s, 3H), 3.72 (s, 3H), 3.37–3.25 (m, 1H), 2.98–2.83 (m, 1H), 2.87 (s, 3H); EIMS m/z 383 [M–OMe]⁺; HR-EIMS m/z 383.1625 [M–OMe]⁺ (calcd for C₂₁H₂₃N₂O₅, 383.1607).

Boc-Ala-N,O-dimethyl-Tyr-Gly-OMe (28). When 27 was treated as described for the

preparation of 20, 28 was obtained in 73% yield as an amorphous solid: $[\alpha]^{28}$ D -130 (c = 0.38, CHCl₃); IR (film) v_{max} 3306, 2979, 2936, 2836, 1754, 1680, 1641, 1514, 1457, 1411, 1367, 1300, 1249, 1208, 1176, 1033 cm⁻¹; ¹H NMR (400 MHz, CDCl₃, mixture of two rotamers) major rotamer δ 8.23 (br t, 1H, J = 6 Hz), 7.04 (d-like, 2H, J = 8.6 Hz), 6.87 (d-like, 2H, J =8.6 Hz), 5.04 (d, 1H, J = 7.4 Hz), 4.82 (dd, 1H, J = 11.2, 3.5 Hz), 4.19 (dq, 1H, J = 7.4, 6.7 Hz), 4.05 (dd, 1H, J = 17.5, 6.0 Hz), 3.92 (dd, 1H, J = 17.5, 5.8 Hz), 3.74 (s, 3H), 3.70 (s, 3H), 3.15 (dd, 1H, J = 14.6, 3.5 Hz), 3.00 (dd, 1H, J = 14.6, 11.2 Hz), 2.93 (s, 3H), 1.35 (s, 9H),0.43 (d, 3H, J = 6.7 Hz); minor rotamer δ 7.10 (d-like, 2H, J = 8.6 Hz), 6.78 (d-like, 2H, J =8.6 Hz), 6.63 (br t, 1H, J = 6 Hz), 5.24 (br d, 1H, J = 8 Hz), 5.18 (br t, 1H, J = 8 Hz), 4.52 (m, 1H), 4.09 (dd, 1H, J = 18.1, 6.5 Hz), 3.78 (dd, 1H, J = 18.1, 4.8 Hz), 3.75 (s, 3H), 3.69 (s, 3H), 3.27 (dd, 1H, J = 14.4, 7.3 Hz), 2.97 (s, 3H), 2.95 (m, 1H), 1.41 (s, 9H), 1.26 (d, 3H, J = 6.8Hz); ¹³C NMR (100 MHz, CDCl₃, mixture of two rotamers) major rotamer δ 174.0, 169.8, $169.8, 158.7, 156.4, 130.3 \times 2, 129.5, 114.4 \times 2, 80.6, 62.4, 55.3, 52.1, 44.8, 41.1, 33.1, 29.1,$ 28.2×3 , 16.6; minor rotamer δ 174.1, 170.1, 169.9, 158.4, 155.1, 129.9 \times 2, 128.8, 113.9 \times 2. 79.6, 58.6, 55.1, 52.2, 46.7, 41.0, 32.6, 31.7, 28.3 \times 3, 18.3; EIMS m/z 451 [M]⁺; HR-EIMS m/z 451.2321 [M]⁺ (calcd for C₂₂H₃₃N₃O₇, 451.2319).

Boc-D-Ala-Ala-N,O-dimethyl-Tyr-Gly-OMe (29). When **28** was treated as described for the preparation of **21** by using Boc-D-Ala-OH, **29** was obtained in 79% yield as an amorphous solid: $[\alpha]^{23}_{D}$ –96.1 (c = 0.90, CHCl₃); IR (film) v_{max} 3298, 2980, 2937, 1755, 1683, 1641, 1515, 1454, 1368, 1301, 1249, 1210, 1177, 1034, 756 cm⁻¹; ¹H NMR (400 MHz, CDCl₃, mixture of two rotamers) major rotamer δ 8.23 (br t, 1H, J = 5.3 Hz), 7.04 (d-like, 2H, J = 8.6 Hz), 6.81 (d-like, 2H, J = 8.6 Hz), 5.31 (br d, 1H, J = 6.6 Hz), 4.93 (dd, 1H, J = 11.1, 3.5 Hz), 4.34 (quintet, 1H, J = 6.6 Hz), 4.14 (br m, 1H), 4.10 (dd, 1H, J = 17.7, 5.3 Hz), 3.95 (dd, 1H, J = 17.7, 5.3 Hz), 3.74 (s, 3H), 3.71 (s, 3H), 3.14 (dd, 1H, J = 14.6, 3.5 Hz), 2.99 (dd, 1H, J = 14.6, 11.1 Hz), 2.91 (s, 3H), 1.40 (s, 9H), 1.26 (d, 3H, J = 7.1 Hz), 0.48 (d, 3H, J = 6.7 Hz); ¹³C NMR (100 MHz, CDCl₃, mixture of two rotamers) major rotamer δ 173.5, 173.2, 170.0, 169.6, 158.7, 155.4, 130.4 × 2, 129.5, 114.4 × 2, 80.2, 62.4, 55.3, 52.1, 49.9, 44.2, 41.4, 33.3, 29.3, 28.3 × 3, 17.8, 16.4; EIMS m/z 522 [M]⁺; FABMS m/z 523 [M+H]⁺; HR-EIMS m/z

522.2681 [M]⁺ (calcd for C₂₅H₃₈N₄O₈, 522.2690).

Boc-D-Ala-Ala-N,O-dimethyl-Tyr-Gly-OH (30). When 29 was processed as described for the preparation of 22, 30 was obtained in 93% yield as an amorphous solid: $[α]^{28}_{D}$ –90.8 (c = 1.3, MeOH); IR (film) v_{max} 3303, 2979, 2936, 2837, 1638, 1514, 1454, 1412, 1367, 1301, 1248, 1176, 1034, 756 cm⁻¹; ¹H NMR (500 MHz, CD₃OD, mixture of two rotamers) major rotamer δ 7.14 (d-like, 2H, J = 8.6 Hz), 6.86 (d-like, 2H, J = 8.6 Hz), 5.02 (br m, 1H), 4.42 (q, 1H, J = 6.8 Hz), 4.04 (m, 1H), 3.92 (d, 1H, J = 17.6 Hz), 3.85 (d, 1H, J = 17.6 Hz), 3.74 (s, 3H), 3.19 (br d, 1H, J = 14 Hz), 2.98 (m, 1H), 2.93 (s, 3H), 1.41 (s, 9H), 1.25 (d, 3H, J = 7.4 Hz), 0.50 (br d, 3H, J = 6.8 Hz); minor rotamer δ 7.13 (d-like, 2H, J = 8.6 Hz), 6.82 (d-like, 2H, J = 8.6 Hz), 5.02 (br m, 1H), 4.73 (q, 1H, J = 6.9 Hz), 4.05 (d, 1H, J = 17.6 Hz), 4.04 (m, 1H), 3.92 (d, 1H, J = 17.6 Hz), 3.74 (s, 3H), 3.26 (dd, 1H, J = 14.2, 6.2 Hz), 3.00 (m, 1H), 2.98 (s, 3H), 1.44 (s, 9H), 1.26 (d, 3H, J = 6.9 Hz), 1.25 (d, 3H, J = 7.4 Hz); HR-ESIMS m/z 509.2653 [M+H]⁺ (calcd for C₂₄H₃₇N₄O₈, 509.2611).

Hexapeptide 31. A solution of cycloisodityrosine 16 (23.6 mg, 0.0411 mmol) in TFA (0.5 mL) was stirred at room temperature for 3 h. TFA was removed *in vacuo*. CHCl₃ (5 mL) and saturated aqueous NaHCO₃ (5 mL) were added to the residue, and the whole was extracted with CHCl₃ (3 × 10 mL). The combined CHCl₃ extracts were washed with brine (10 mL), dried over MgSO₄ and filtered, and the solvent removed *in vacuo*. The residue was dissolved in THF (0.5 mL), to which Boc-Gly-Ala-*N*,*O*-dimethyl-Tyr-Ala-OH (22) (39.8 mg, 0.0783 mmol), HOObt (13.4 mg, 0.0821 mmol) and EDC (12.6 mg, 0.0657 mmol) were added, and the solution was stirred at room temperature for 48 h. Saturated aqueous NaHCO₃ (10 mL) was added to the solution, and the mixture was extracted with CHCl₃ (3 × 10 mL). The combined CHCl₃ extracts were washed with brine (5 mL), dried over MgSO₄ and filtered, and the solvent removed *in vacuo*. The residue was subjected to MPLC (silica gel, 10:10:1 hexane/CHCl₃/MeOH) to provide 31 (38.4 mg; 97%) as an amorphous solid: [α]²⁴_D –132 (c = -14.5 mc)

0.12, CHCl₃); IR (film) v_{max} 3294, 2979, 2933, 1719, 1641, 1515, 1453, 1412, 1367, 1249, 1220, 1177, 1129, 1094, 1032 cm⁻¹; ¹H NMR (500 MHz, CDCl₃, mixture of rotamers) δ 7.83 (br m), 7.54–6.71 (m), 6.61 (m), 6.55 (dd, J = 8.3, 1.8 Hz), 6.49 (d, J = 7.3 Hz), 6.32 (br m), 5.88 (dd, J = 12.0, 4.7 Hz), 5.76 (br m), 5.38 (br m), 5.32 (dd, J = 11.4, 2.2 Hz), 5.23–4.68 (m), 4.65 (dd, J = 12.3, 3.6 Hz), 4.40 (br m), 4.38 (d, J = 1.9 Hz), 4.32 (quintet, J = 7.0 Hz), 3.93 (s), 3.78 (s), 3.76 (s), 3.75–3.57 (m), 3.50–3.41 (m), 3.35–3.14 (m), 3.28 (s), 3.18 (s), 3.08 (s), 3.02 (s), 3.01–2.84 (m), 2.94 (s), 2.89 (s), 2.80 (dd, J = 11.4, 3.1 Hz), 2.74–2.69 (m), 2.56 (s), 2.55 (s), 1.457 (s), 1.453 (s), 1.404 (s), 1.397 (s), 1.348 (s), 1.341 (s), 1.353 (s), 1.33–1.19 (m), 1.16 (d, J = 6.7 Hz), 0.93 (m), 0.61 (d, J = 6.6 Hz), 0.48 (d, J = 6.6 Hz); FABMS m/z 965 [M+H]⁺; HR-ESIMS m/z 987.4484 [M+Na]⁺ (calcd for C₅₂H₆₄N₆O₁₂Na, 987.4480).

Hexapeptide 32. A solution of cycloisodityrosine 16 (30.0 mg, 0.0522 mmol) in TFA (0.5 mL) was stirred at room temperature for 3 h. TFA was removed *in vacuo*. CHCl₃ (5 mL) and saturated aqueous NaHCO₃ (5 mL) were added to the residue, and the whole was extracted with CHCl₃ (3 × 10 mL). The combined CHCl₃ extracts were washed with brine (10 mL), dried over MgSO₄ and filtered, and the solvent removed *in vacuo*. The residue was dissolved in THF (0.5 mL), to which Boc-D-Ala-Gly-N,O-dimethyl-Tyr-Ala-OH (25) (31.3 mg, 0.0615 mmol), HOObt (17.0 mg, 0.104 mmol) and EDC (16.0 mg, 0.0835 mmol) were added, and the solution was stirred at room temperature for 48 h. Saturated aqueous NaHCO₃ (10 mL) was added to the solution, and the mixture was extracted with CHCl₃ (3 × 10 mL). The combined CHCl₃ extracts were washed with brine (5 mL), dried over MgSO₄ and filtered, and the solvent removed *in vacuo*. The residue was subjected to MPLC (silica gel, 10:10:1 hexane/CHCl₃/MeOH) to provide 32 (39.6 mg, 79%) as an amorphous solid: [α]²⁴_D –141 (c = 0.13, CHCl₃); IR (film) v max 3307, 2975, 2934, 1742, 1711, 1641, 1515, 1500, 1456, 1412, 1367, 1266, 1249, 1178, 1129, 1094, 1030 cm⁻¹; H NMR (500 MHz, CDCl₃, mixture of

rotamers) δ 7.56–6.70 (m), 6.63–6.53 (m), 5.88 (dd, J = 12.1, 4.8 Hz), 5.38–4.63 (m), 4.38 (d, J = 2.0 Hz), 4.25–3.82 (m), 3.93 (s), 3.77 (s), 3.74 (s), 3.71 (s), 3.68–3.59 (m), 3.47 (m), 3.37–3.15 (m), 3.20 (s), 3.18 (s), 3.09–2.63 (m), 2.93 (s), 2.85 (s), 2.55 (s), 2.54 (s), 2.53 (s), 1.44 (s), 1.43 (s), 1.38–1.19 (m); FABMS m/z 965 [M+H]⁺; HR-ESIMS m/z 987.4484 [M+Na]⁺ (calcd for $C_{52}H_{64}N_6O_{12}Na$, 987.4480).

Hexapeptide 33. A solution of cycloisodityrosine 16 (9.5 mg, 0.0165 mmol) in TFA (0.5 mL) was stirred at room temperature for 3 h. TFA was removed in vacuo. CHCl₃ (5 mL) and saturated aqueous NaHCO3 (5 mL) were added to the residue, and the whole was extracted with CHCl₃ (3 × 10 mL). The combined CHCl₃ extracts were washed with brine (10 mL), dried over MgSO₄ and filtered, and the solvent removed in vacuo. The residue was dissolved in THF (0.5 mL), to which Boc-D-Ala-Ala-N,O-dimethyl-Tyr-Gly-OH (30) (12.8 mg, 0.0252 mmol), HOAt (5.0 mg, 0.037 mmol) and EDC (6.4 mg, 0.033 mmol) were added, and the solution was stirred at room temperature for 3 h. Saturated aqueous NaHCO3 (10 mL) was added to the solution, and the mixture was extracted with CHCl₃ (3 × 10 mL). The combined CHCl₃ extracts were washed with brine (5 mL), dried over MgSO₄ and filtered, and the solvent removed in vacuo. The residue was subjected to MPLC (silica gel, 10:10:1 hexane/CHCl₃/MeOH) to provide 33 (14.7 mg, 92%) as an amorphous solid: $\left[\alpha\right]^{24}$ D -94.6 (c =0.13, CHCl₃); IR (film) v_{max} 3307, 2976, 2932, 1740, 1711, 1646, 1515, 1454, 1413, 1367, 1265, 1249, 1218, 1178, 1129, 1094, 1031 cm⁻¹; ¹H NMR (500 MHz, CDCl₃, mixture of rotamers) δ 7.90 (br m), 7.51–6.52 (m), 5.85 (dd, J = 12.5, 4.4 Hz), 5.37 (br m), 5.31 (dd, J = 12.5, 4.4 Hz), 5.37 (dd, J = 12.5, 4.5 (dd, J = 12.5), 5.37 (dd, J = 111.3, 2.7 Hz), 5.23-4.63 (m), 4.42 (br s), 4.18-4.03 (m), 3.94 (s), 3.93 (s), 3.92-3.74 (m), 3.78 (s), 3.76 (s), 3.62 (t, J = 10 Hz), 3.50 (m), 3.36 (dd, J = 18.3, 3.2 Hz), 3.32–3.13 (m), 3.09-2.72 (m), 3.06 (s), 2.94 (s), 2.84 (s), 2.81 (s), 2.69 (dd, J = 11.2, 2.6 Hz), 2.57 (s), 1.45(s), 1.43 (s), 1.41–1.23 (m), 1.19 (d, J = 6.7 Hz), 0.62 (d, J = 6.5 Hz), 0.42 (d, J = 6.6 Hz); FABMS m/z 965 $[M+H]^+$; HR-ESIMS m/z 987.4484 $[M+Na]^+$ (calcd for $C_{52}H_{64}N_6O_{12}Na$,

987.4480).

Amine 34. A solution of 31 (10.0 mg, 0.0104 mmol) in TFA (0.5 mL) was stirred at room temperature for 30 min, and then TFA was removed *in vacuo*. CHCl₃ (5 mL) and saturated aqueous NaHCO₃ (5 mL) were added to the residue, and the whole was extracted with 97:3 CHCl₃/MeOH (3 × 10 mL). The combined extracts were washed with brine (10 mL), dried over MgSO₄ and filtered, and the solvent removed *in vacuo*. The residue was subjected to column chromatography (silica gel, 10:1 CHCl₃/MeOH) to provide 34 (8.5 mg, 95%) as an amorphous solid: $[\alpha]^{24}_D$ –185 (c = 0.16, pyridine); ¹H NMR (400 MHz, C₃D₅N, mixture of rotamers) δ 9.02 (d, J = 6.7 Hz), 8.95 (d, J = 6.6 Hz), 8.69 (d, J = 7.3 Hz), 8.63 (d, J = 7.5 Hz), 7.65 (d, J = 7.2 Hz), 7.57 (d, J = 5.7 Hz), 7.50–6.90 (m), 6.88 (d, J = 8.5 Hz), 6.63 (d-like, J = 8.3 Hz), 5.80 (dd, J = 11.4, 3.2 Hz), 5.71–5.64 (m), 5.43 (dd, J = 10.9, 3.4 Hz), 5.35 (d, J = 13 Hz), 5.24 (d, J = 13 Hz), 5.21–5.07 (m), 5.01 (m), 4.81 (m), 4.71 (s), 3.84 (s), 3.75 (t, J = 11.3 Hz), 3.65 (s), 3.62 (m), 3.60 (s), 3.42 (s), 3.37 (s), 3.28 (s), 3.23 (s), 3.21 (s), 3.20–3.00 (m), 2.95–2.87 (m), 2.78 (s), 2.75 (s), 2.66 (dd, J = 11.3, 2.8 Hz), 1.63 (d, J = 7.0 Hz), 1.51 (d, J = 6.8 Hz), 1.36 (d, J = 6.8 Hz), 0.74 (d, J = 6.7 Hz); FABMS m/z 865 [M+H]⁺.

Amine 35. When **32** was treated in the same manner as described for the preparation of **34**, **35** was obtained in 98% yield as an amorphous solid: $[\alpha]^{24}_{D}$ –137 (c = 0.29, pyridine); ¹H NMR (400 MHz, C₅D₅N, mixture of rotamers) δ 9.11 (d, J = 6.9 Hz), 9.01 (d, J = 7.3 Hz), 8.84 (d, J = 7.0 Hz), 8.79 (br m), 8.59 (br t, J = 4 Hz), 7.63–7.53 (m), 7.51 (d, J = 7.2 Hz), 7.48–7.00 (m), 6.98 (dd, J = 8.4, 2.3 Hz), 6.91 (d, J = 8.4 Hz), 6.86 (d, J = 8.6 Hz), 6.84–6.78 (m), 6.73 (dd, J = 8.1, 1.9 Hz), 6.62 (dd, J = 8.3, 2.0 Hz), 5.89 (dd, J = 9.1, 6.8 Hz), 5.68 (dd, J = 11.4, 3.1 Hz), 5.41–5.07 (m), 5.01 (dd, J = 12.2, 3.8 Hz), 4.71 (d, J = 2.0 Hz), 4.49 (dd, J = 16.2, 4.7 Hz), 4.28 (dd, J = 17.4, 4.6 Hz), 4.15 (dd, J = 17.4, 4.7 Hz), 3.84 (s), 3.78–3.60 (m), 3.59 (s), 3.57 (s), 3.57–3.50 (m), 3.47–3.37 (m), 3.32 (s), 3.29 (s), 3.27–3.20 (m), 3.20 (s), 3.20–3.11 (m), 3.10 (s), 3.10–2.96 (m), 2.87 (s), 2.74 (s), 2.69 (dd, J = 11.4, 3.0 Hz), 1.52 (d, J = 6.8 Hz), 1.50–1.40 (m), 1.43 (d, J = 6.9 Hz), 1.39 (d, J = 6.9 Hz), 1.33-(d, J = 7.0 Hz),

1.32-1.18 (m); FABMS m/z 865 [M+H]⁺.

Amine 36. When 33 was treated in the same manner as described for the preparation of 34, 36 was obtained in 94% yield as an amorphous solid: $[\alpha]^{24}_D$ –169 (c = 0.13, pyridine); ¹H NMR (400 MHz, C₅D₅N, mixture of rotamers) δ 8.85 (d, J = 5.6 Hz), 8.75 (t, J = 5.4 Hz), 8.37 (d, J = 7.9 Hz), 8.26–8.07 (m), 7.51 (d, J = 7.3 Hz), 7.45 (d, J = 7.2 Hz), 7.28–6.87 (m), 6.80 (dd, J = 8.8, 2.3 Hz), 6.78–6.65 (m), 6.49–6.42 (m), 5.54–4.80 (m), 4.23–4.10 (m), 4.01–3.90 (m), 3.64 (s), 3.44 (s), 3.41 (s), 3.35–3.20 (m), 3.07 (s), 2.99 (s), 2.92 (s), 2.85 (s), 2.77–2.66 (m), 2.55 (s), 2.54 (s), 2.48–2.39 (m), 1.21 (d, J = 6.9 Hz), 1.19 (d, J = 6.8 Hz), 1.15 (d, J = 6.8 Hz), 0.54 (d, J = 6.7 Hz); FABMS m/z 865 $[M+H]^+$.

[Gly-2]RA-VII (5). In the presence of 10% Pd/C (12.8 mg), a solution of 35 (8.8 mg, 0.0102 mmol) in EtOH (3 mL) was stirred at room temperature under an atmosphere of hydrogen for 40 min. The catalyst was filtered off, and the filtrate was concentrared to give a

residue, which was dissolved in DMF (7.8 mL). To this solution were added HOObt (13.3 mg, 0.0815 mmol) and EDC (15.5 mg, 0.0809 mmol), and the mixture was stirred at room temperature for 4 days. The solvent was removed by distillation under reduced pressure. Water (10 mL) was added to the residue, and the whole was extracted with 97:3 CHCl₃/MeOH (3 × 10 mL). The combined extracts were washed with brine (10 mL), dried over MgSO₄ and filtered, and the solvent removed *in vacuo*. The residue was subjected to MPLC (silica gel, 20:2:1 CHCl₃/EtOAc/MeOH) to provide 5 (0.8 mg, 10%) as an amorphous powder, which was recrystallized from MeOH to give a white crystalline powder: mp 263–264 °C; $[\alpha]^{26}_D$ –217 (c = 0.32, CHCl₃); IR (KBr) v_{max} 3387, 3314, 3067, 2934, 2837, 1675, 1664, 1656, 1648, 1638, 1628, 1543, 1514, 1499, 1459, 1448, 1411, 1264, 1249, 1128, 1094, 1032, 800 cm⁻¹; ¹H NMR (500 MHz, CDCl₃, selected data) δ 3.94 (s, 3H), 3.78 (s, 3H), 1.46 (d, 3H, J = 6.9 Hz); FABMS m/z 757 [M+H]⁺; HR-FABMS m/z 757.3519 [M+H]⁺ (calcd for C₄₀H₄₉N₆O₉, 757.3561).

[Gly-4]RA-VII (6). A solution of 36 (8.5 mg, 0.0098 mmol) in EtOH (3.4 mL) was stirred in the presence of 10% Pd/C (14.4 mg) at room temperature under an atmosphere of hydrogen for 25 min. The catalyst was filtered off, and the filtrate was concentrated to dryness. The residue was dissolved in DMF (7.6 mL), to which were added HOObt (12.8 mg, 0.0785 mmol) and EDC (15.0 mg, 0.0782 mmol), and the mixture was stirred at room temperature for 4 days. The solvent was removed by distillation under reduced pressure. Water (10 mL) was added to the residue, and the whole was extracted with 97:3 CHCl₃/MeOH (3 × 10 mL). The combined extracts were washed with brine (10 mL), dried over MgSO₄ and filtered, and the solvent removed *in vacuo*. The residue was subjected to MPLC (silica gel, 20:2:1 CHCl₃/EtOAc/MeOH) to provide 6 (4.0 mg, 54%) as an amorphous solid: [α]²⁶_D –150 (c = 0.4, MeOH); IR (film) v_{max} 3294, 2934, 2837, 1656, 1637, 1513, 1500, 1445, 1411, 1265, 1248, 1216, 1127, 1094, 1031, 753 cm⁻¹; ¹H and ¹³C NMR: refer to Tables S2 and S3; FABMS m/z 757 [M+H]⁺; HR-FABMS m/z 757.3550 [M+H]⁺ (calcd for C₄₀H₄₉N₆O₉, 757.3561).

 $\begin{array}{ccc} & R^1 & R^2 \\ \text{[Gly-1]RA-VII (4)} & H_a & \text{Me} \\ \text{[Gly-4]RA-VII (6)} & \text{Me} & H_b \end{array}$

Table S1 ¹H NMR Chemical Shifts for [Gly-1]RA-VII (4) in CDCl₃ at 300 K

		major conformer	minor conformer
Gly-1	α_a	3.48 (d, 17.7)	3.39 (m)
	α_b	4.29 (dd, 17.7, 7.0)	4.35 (m)
	NH	6.22 (d, 7.0)	6.14 (d, 7.2)
Ala-2	α	4.82 (dq, 8.1, 6.9)	4.85 (m)
	β	1.35 (d, 6.9)	0.98 (d, 6.4)
	NH	6.41 (d, 8.1)	6.43 (*)
Tyr-3	α	3.58 (dd, 10.9, 4.8)	4.68 (m)
	β_a	3.33 (dd, 14.0, 10.9)	2.98-3.07 (m)
	βь	3.38 (dd, 14.0, 4.8)	2.98-3.07 (m)
	δ	7.04 (d-like, 8.6)	7.09 (d-like, 8.6)
	ε	6.83 (d-like, 8.6)	6.84 (d-like, 8.6)
	NMe	2.87 (s)	2.99 (s)
	OMe	3.79 (s)	3.77 (s)
Ala-4	α	4.75 (dq, 7.6, 6.7)	4.59 (m)
	β	1.13 (d, 6.7)	1.20 (d, 6.8)
	NH	6.76 (d, 7.6)	6.88 (*)
Tyr-5	α	5.44 (dd, 11.4, 3.1)	5.42 (dd, 11.3, 3.0)
	β_a	2.65 (dd, 11.3, 3.1)	2.74 (dd, 11.3, 3.0)
	β_b	3.68 (dd, 11.4, 11.3)	3.70 (dd, 11.3, 11.3)
	δ_a	7.26 (dd, 8.4, 2.2)	7.26 (*)
	δ_b	7.42 (dd, 8.4, 2.2)	7.44 (*)
	εa	6.88 (dd, 8.4, 2.4)	6.89 (*)
	ϵ_b	7.20 (dd, 8.4, 2.4)	7.24 (*)
	NMe	3.13 (s)	3.13 (s)
Tyr-6	α	4.63 (dd, 12.0, 3.8)	4.71 (m)
	β_a	3.11 (dd, 17.8, 12.0)	3.02-3.10 (m)
	β_b	2.98 (dd, 17.8, 3.8)	3.02-3.10 (m)
	δ_a	6.57 (dd, 8.4, 1.9)	6.57 (*)
	δ_b	4.34 (d, 1.9)	4.39 (d, 1.8)
	ε	6.79 (d, 8.4)	6.83 (*)
	NMe	2.67 (s)	2.65 (s)
	OMe	3.93 (s)	3.93 (s)

J-values in parentheses are given in Hz.

^{*} Multiplicity was not determined due to overlapping of the resonances.

Table S2 ¹H NMR Chemical Shifts for [Gly-4]RA-VII (6) in CD₃OD at 300 K

****	<u> </u>	major conformer	minor conformer
D-Ala-1	α	4.14 (q, 7.0)	4.42 (q, 7.0)
	β	1.20 (d, 7.0)	1.28 (d, 7.0)
	NH	*	*
Ala-2	α	4.56 (q, 6.5)	4.54 (q, 6.6)
	β	0.88 (d, 6.5)	1.29 (d, 6.6)
	NH	*	*
Tyr-3	α	4.83 (dd, 11.1, 4.2)	3.96 (br m)
	β_a	3.00 (dd, 14.4, 11.1)	3.25-3.32 (m)
	β_b	3.22 (dd, 14.4, 4.2)	3.25-3.32 (m)
	δ	7.20 (d-like, 8.6)	7.10 (d-like, 8.6)
	ε	6.89 (d-like, 8.6)	6.88 (d-like, 8.6)
	NMe	2.95 (s)	2.90 (s)
	OMe	3.77 (s)	3.77 (s)
Gly-4	α_a	4.12 (d, 17.3)	4.50 (br d, 18)
	α_b	3.80 (d, 17.3)	3.50 (br d, 18)
	NH	*	*
Tyr-5	α	5.35 (dd, 11.3, 3.1)	5.28 (dd, 11.1, 2.8)
	β_a	2.76 (dd, 11.3, 3.1)	2.75 (11.7, 2.8)
	β_b	3.57 (dd, 11.3, 11.3)	3.52 (dd, 11.7, 11.1)
	δ_{a}	7.23 (dd, 8.4, 2.3)	7.22 (dd, 8.4, 2.3)
	δ_b	7.50 (dd, 8.4, 2.3)	7.50 (dd, 8.4, 2.3)
	εa	6.80 (dd, 8.4, 2.4)	6.79 (dd, 8.4, 2.4)
	ϵ_{b}	7.26 (dd, 8.4, 2.4)	7.24 (dd, 8.4, 2.4)
	NMe	2.99 (s)	2.95 (s)
Tyr-6	α	4.72 (dd, 12.1, 4.0)	4.65 (m)
	β_a	3.00 (dd, 17.6, 12.1)	2.99-3.07 (m)
	β_b	3.17 (dd, 17.6, 4.0)	2.99-3.07 (m)
	δ_a	6.68 (dd, 8.3, 2.0)	6.67 (dd, 8.3, 2.0)
	δ_b	4.62 (d, 2.0)	4.58 (d, 2.0)
	ε	6.91 (d, 8.3)	6.90 (d, 8.3)
	NMe	2.57 (s)	2.63 (s)
	OMe	3.90 (s)	3.89 (s)

J-values in parentheses are given in Hz.

^{*} Not detected due to chemical exchange with deuterium.

Table S3 13 C NMR Chemical Shifts for [Gly-1]RA-VII (4) and [Gly-4]RA-VII (6) at 300 K

		4 ^a	6 ^b
		major conformer	major/minor ^c
Gly-1	α	41.4	49.5/*
(D-Ala-1)	β	_	20.6/20.2
	CO	168.6	172.8/174.0
Ala-2	α	44.8	45.4/46.8
	β	16.4	17.5/15.9
	CO	172.5	173.7/175.6
Tyr-3	α	68.5	63.2/69.0
J • •	β	32.7	35.1/34.1
	γ	130.7	130.2/130.3
	δ	130.3	131.2/131.5
	ε	114.0	115.3/115.0
	ζ	158.4	160.3/160.1
	CO	168.1	172.3/172.3#
	NCH_3	39.9	30.9/40.3
	OCH_3	55.3	55.7/55.7
Ala-4	α	46.5	42.7/42.8
(Gly-4)	β	18.6	_
	CO	171.8	169.1/170.1
Tyr-5	α	54.2	56,5/57.4
	β	37.1	37.5/37.6
	γ	135.1	137.3/137.5
	δ_a	132.8	134.0/134.0
	δ_b	131.0	132.0/132.0
	ϵ_a	124.3	124.9/124.9
	ϵ_b	125.9	127.2/127.2
	ζ	158.3	159.6/159.5
	CO	169.6	171.7/171.2
	NCH ₃	30.6	29.9/30.1
Tyr-6	α	57.5	58.9/58.4
	β	35.3	35.8/35.8
	γ	128.1	130.5/130.5
	δ_a	121.0	122.6/122.6
	δ_b	113.4	115.4/115.2
	ϵ_a	112.2	114.2/114.2
	ϵ_{b}	153.1	154.4/154.4
	ζ	146.5	148.0/148.0
	CO	171.3	172.8/172.2#
	NCH_3	29.2	29.9/30.0
	O <i>C</i> H₃	56.2	56.9/56.9

^a Data are recorded in CDCl₃. ^b Data are recorded in CD₃OD. ^c Major and minor conformers.

^{*} Not assigned due to overlapping to the solvent resonance. * The values may be reversed.



































