Supporting Information

Novel Dolabellane-type Diterpene Alkaloids with Lipid Metabolism Promoting Activities from the Seeds of *Nigella sativa*

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The following experimental conditions were used for chromatography: normal-phase silica gel column chromatography, Silica gel BW-200 (Fuji Silysia Chemical, 150–350 mesh); reversed-phase silica gel column chromatography, Chromatorex ODS DM1020T (Fuji Silysia Chemical, 100–200 mesh); HPLC column, YMC-Pack ODS-A (YMC, 250 \times 20 mm i.d.); TLC, pre-coated TLC plates with Silica gel 60F₂₅₄ (Merck, 0.25 mm) (normal-phase) and Silica gel RP-18 F_{254S} (Merck, 0.25 mm) (reversed-phase); reversed-phase HPTLC, pre-coated TLC plates with Silica gel RP-18 WF_{254S} (Merck, 0.25 mm); detection was achieved by spraying with 1% Ce(SO₄)₂–10% aqueous H₂SO₄ followed by heating.

Extraction and Isolation. The seeds of *N. sativa* (10.0 kg, collected in Egypt) were extracted with methanol 3 times under reflux for 3 h. Evaporation of the solvent under reduced pressure provided the methanolic extract (1740 g, 17.4% from this natural medicine). The methanolic extract (1245 g) was partitioned into an EtOAc and water mixture to give an EtOAc-soluble fraction (723 g, 10.1%) and an aqueous phase (522 g, 7.3%). The EtOAcsoluble fraction (285 g) was subjected to ordinary-phase silica gel column chromatography $[3.0 \text{ kg}, n-\text{hexane}-\text{EtOAc} (20:1-10:1-5:1-2:1-1:2)-\text{CHCl}_3-\text{MeOH}-\text{H}_2\text{O} (20:3:1, \text{lower})$ layer-6:4:1)-MeOH] to give 12 fractions [Fr. 1 (12.36 g), 2 (55.91 g), 3 (74.90 g), 4 (18.51 g), 5 (61.82 g), 6 (2.55 g), 7 (0.97 g), 8 (9.68 g), 9 (6.69 g), 10 (8.13 g), 11 (15.82 g), and 12 Fraction 7 (0.97 g) was subjected to reversed-phase silica gel column (16.57 g)]. chromatography [30 g, MeOH-H₂O (50:50-70:30-85:15, v/v)-MeOH] to give seven fractions [Fr. 7-1 (112 mg), 7-2 (105 mg), 7-3 (68 mg), 7-4 (315 mg), 7-5 (87 mg), 7-6 (152 mg), and 7-7 (120 mg)]. Fraction 7-4 (315 mg) was further separated by HPLC [YMC-Pack ODS-5-A, 250×20 mm i.d., MeOH-H₂O (85:15, v/v)] to give nigellamine A₁ (1, 271 mg, 0.0096% from the natural medicine). Fraction 7-6 (152 mg) was purified by HPLC [MeOH-H₂O (80:20, v/v)] to give nigellamine B₁ (3, 33 mg, 0.0012%). Fraction 9 (6.69 g) was subjected to reversed-phase silica gel column chromatography [200 g, MeOH-H₂O (40:60-60:40-80:20, v/v)-MeOH] to give ten fractions [Fr. 9-1 (1.150 g), 9-2 (943 mg), 9-3 (627 mg), 9-4 (1.312 g), 9-5 (142 mg), 9-6 (750 mg), 9-7 (150 mg), 9-8 (241 mg), 9-9 (805 mg), and 9-10 (428 mg)]. Fraction 9-6 (750 mg) was purified by HPLC [MeOH-H₂O (75:25, v/v)] to give nigellamine B₂ (4, 102 mg, 0.0036%). Fraction 9-9 (805 mg) was purified by HPLC $[MeOH-H_2O (80:20, v/v)]$ to give nigellamine A₂ (2, 221 mg, 0.0078%).

Treatment of Nigellamine A₁ (1) with 0.1% NaOMe–MeOH. A solution of **1** (10.0 mg) in 0.1% NaOMe–MeOH (2.0 mL) was stirred at room temperature for 8 h. From the reaction mixture, methyl nicotinate (i) and methyl benzoate (ii) were identified by HPLC analyses [t_R (i): 5.58, (ii): 15.80 min, detection: UV (254 nm), column: YMC-Pack ODS-5-A, 250×4.6 mm i.d., mobile phase: MeOH–H₂O (60:40, v/v); flow rate 0.7 ml/min (standard samples were obtained by diazomethane methylation of commercial nicotinic acid and benzoic acid)].

Then the reaction mixture was neutralized with Dowex HCR-W2 (H⁺ form) and the resin was removed by filtration. Evaporation of the solvent from the filtrate under reduced pressure furnished a residue, which was purified by HPLC [detection: RI, column: YMC-Pack ODS-5-A, 250×20 mm i.d., mobile phase: MeOH–H₂O (60:40, v/v)] to give **1a** (4.0 mg, 77%).

On the other hand, a solution of **1** (15.0 mg) in 0.1% NaOMe–MeOH (2.0 mL) was stirred at 0°C for 3.5 h. The reaction mixture was poured into ice-water and the whole was extracted with EtOAc. The EtOAc extract was successively washed with brine then dried over MgSO₄ powder and filtrated. Evaporation of the solvent from the filtrate under reduced pressure furnished a residue, which was purified by HPLC [detection: RI, column: YMC-Pack ODS-5-A, 250×20 mm i.d., mobile phase: MeOH–H₂O (80:20, v/v)] to give **1b** (4.1 mg, 40%) and 10-desacyl derivative (**1c**, 4.8 mg, 38%).

1a: A white powder, $[a]_D^{23} + 33.9^{\circ}$ (*c*=0.20, MeOH). IR (KBr): 3304, 3240, 1655, 1561, 1509, 1375, 1122, 1038, 1017, 953, 777, 718 cm⁻¹. ¹³C NMR (CD₃OD): δ_C 59.5 (C-1), 71.9 (C-2), 129.4 (C-3), 136.5 (C-4), 38.8 (C-5), 23.9 (C-6), 67.4 (C-7), 60.9 (C-8), 46.5 (C-9), 74.4 (C-10), 49.8 (C-11), 141.5 (C-12), 28.7 (C-13), 29.4 (C-14), 62.7 (C-15), 16.4 (C-16), 18.8 (C-17), 124.5 (C-18), 22.3 (C-19), 22.4 (C-20).

1b: A white powder, $[a]_D^{24} + 36.3^{\circ}$ (*c*=0.40, CHCl₃). High resolution positive-ion FAB-MS: Calcd for C₂₇H₃₇O₅ (M+H)⁺: 441.2641. Found: 441.2650. UV (MeOH, log ε): 230 (4.10), 273 (2.91) nm. IR (KBr): 3325, 1717, 1636, 1603, 1541, 1509, 1451, 1271, 1111, 1071, 1026, 953, 754, 714 cm⁻¹. ¹³C NMR (CDCl₃): δ_C 57.6 (C-1), 73.7 (C-2), 123.3 (C-3), 139.1 (C-4), 37.9 (C-5), 22.8 (C-6), 65.9 (C-7), 59.4 (C-8), 44.8 (C-9), 73.6 (C-10), 48.4 (C-11), 138.4 (C-12), 28.3 (C-14), 27.5 (C-14), 61.5 (C-15), 16.2 (C-16), 18.5 (C-17), 124.9 (C-18), 21.9 (C-19), 22.3 (C-20), 130.2 (C-1'), 129.6 (C-2',6'), 128.5 (C-3',5'), 133.1 (C-4'), 165.9 (C-7'). Positive-ion FAB-MS: *m/z* 441 (M+H)⁺.

1c: A white powder, $[a]_D^{24} + 29.8^{\circ}$ (*c*=0.50, CHCl₃). High resolution positive-ion FAB-MS: Calcd for C₃₄H₄₁O₆ (M+H)⁺: 545.2903. Found: 545.2906. UV (MeOH, log *ε*): 228 (4.46), 274 (3.30) nm. IR (KBr): 3517, 1717, 1647, 1636, 1603, 1541, 1509, 1451, 1281, 1113, 1069, 1026, 953, 756, 712 cm⁻¹. ¹H NMR (CDCl₃): *δ*1.40, 1.74, 1.79 (3H each, all s, H₃-17, 19, 20), 1.52 (1H, dd, *J* = 12.8, 13.5 Hz, H*β*-9), 1.66 (1H, m, H*α*-6), 1.82 (3H, d, *J* = 0.9 Hz, H₃-16), 1.97 (1H, m, H*β*-6), 2.20 (2H, m, H₂-14), 2.31 (2H, m, H₂-13), 2.37 (1H, dd, *J* = 5.5, 13.5 Hz, H*α*-9), 2.39 (1H, m, H*α*-5), 2.43 (1H, ddd, *J* = 5.2, 12.8, 12.8 Hz, H*β*-5), 2.51 (1H, br s, H-11), 3.02 (1H, br d, *J* = *ca*. 9 Hz, H-7), 4.21 (1H, m, H-10), 4.76, 5.25 (1H each, both d, *J* = 11.0 Hz, H₂-15), 5.40 (1H, d, *J* = 10.1 Hz, H-2), 5.62 (1H, dd, *J* = 0.9, 10.1 Hz, H-3), 7.19 (2H, dd, *J* = 7.6, 8.3 Hz, H-3',5'), 7.42 (2H, dd, *J* = 7.6, 8.3 Hz, H-3''',5'''), 7.45 (1H, tt, *J* = 1.2, 7.6 Hz, H-4'), 7.58 (1H, tt, *J* = 1.2, 7.6 Hz, H-4'''), 7.87 (2H, dd, *J* = 1.2, 8.3 Hz, H-2',6'), 8.07 (2H, dd, *J* = 1.2, 8.3 Hz, H-2''',6'''). ¹³C NMR (CDCl₃): δ_{C} 56.2 (C-1), 73.2 (C-2), 123.7 (C-3), 139.4 (C-4), 37.8 (C-5), 22.9 (C-6), 65.6 (C-7), 58.9 (C-8), 45.4 (C-9), 73.5 (C-10), 49.1 (C-11), 137.8 (C-12), 28.5 (C-13), 30.5 (C-14), 66.5 (C-15), 16.4 (C-16), 18.5 (C-17), 125.8 (C-18), 21.9 (C-19), 22.5 (C-20), 130.0 (C-1'), 129.6 (C-2',6'), 128.2 (C-3',5'), 132.8 (C-4'), 166.3 (C-7'), 130.3 (C-1''), 129.8 (C-2''',6'''), 128.5 (C-3''',5'''), 133.1 (C-4'''), 166.3 (C-7''). Positive-ion FAB-MS: *m/z* 545 (M+H)⁺.

Treatment of Nigellamine A₂ (2) with 0.1% NaOMe–MeOH. A solution of **2** (10.0 mg) in 0.1% NaOMe–MeOH (2.0 mL) was stirred at room temperature for 8 h. From the reaction mixture, methyl nicotinate and methyl benzoate were identified by HPLC analyses and **1a** (3.8 mg, 74%) was purified by the similar procedure.

On the other hand, a solution of **2** (5.0 mg) in 0.1% NaOMe–MeOH (1.0 mL) was stirred at 0°C for 2 h. The reaction mixture was poured into ice-water and the whole was extracted with EtOAc. The EtOAc extract was successively washed with brine then dried over MgSO₄ powder and filtrated. Evaporation of the solvent from the filtrate under reduced pressure furnished a residue, which was purified by HPLC [detection: RI, column: YMC-Pack ODS-5-A, 250×20 mm i.d., mobile phase: MeOH–H₂O (80:20, v/v)] to give **2a** (0.1 mg, 3%), 10-desacyl derivative (**2b**, 1.8 mg, 43%), 2-desacyl derivative (**2c**, 1.0 mg, 24%), 2,15-desacyl derivative (**2d**, 0.2 mg, 6%), 2,10-desacyl derivative (**2e**, 0.4 mg, 12%), and **1a** (0.3 mg, 9%).

2a: A white powder.

2b: A white powder. High resolution EI-MS: Calcd for $C_{33}H_{39}NO_6$ (M⁺): 545.2777. Found: 545.2781. ¹H NMR (CD₃OD): δ 1.41, 1.76, 1.83 (3H each, all s, H₃-17, 19, 20), 1.45 (1H, dd, J = 12.5, 13.5 Hz, H β -9), 1.76 (1H, m, H α -6), 1.79 (3H, d, J = 1.2 Hz, H₃-16), 1.92 (1H, m, H β -6), 2.24 (2H, m, H₂-14), 2.32 (2H, m, H₂-13), 2.39, 2.57 (1H each, both m, H₂-5), 2.45 (1H, br s, H-11), 2.48 (1H, dd, J = 5.8, 13.5 Hz, H α -9), 3.09 (1H, d-like, H-7), 4.22 (1H, br dd, J = ca. 6, 13 Hz, H-10), 4.77, 5.33 (1H each, both d, J = 10.7 Hz, H₂-15), 5.49 (1H, d, J = 10.4 Hz, H-2), 5.77 (1H, d, J = ca. 10 Hz, H-3), 7.10 (1H, dd, J = 4.8, 7.9 Hz, H-5'), 7.42 (2H, dd, J = 7.7, 8.4 Hz, H-3''', 5'''), 7.61 (1H, br t, J = ca. 8 Hz, H-4'''), 7.97 (1H, ddd, J = 1.2, 2.1, 7.9 Hz, H-6'), 8.04 (2H, dd, J = 1.7, 8.4 Hz, H-2''', 6'''), 8.59 (1H, br s, H-4'), 8.92 (1H, br s, H-2'). EI-MS (%): m/z 545 (M⁺, 2), 423 (3), 300 (72), 121 (59), 105 (100).

2c: A white powder. High resolution EI-MS: Calcd for $C_{33}H_{39}NO_6$ (M⁺): 545.2777. Found: 545.2772. ¹H NMR (CD₃OD): δ 1.50, 1.65 1.86 (3H each, all s, H₃-17, 19, 20), 1.62 (1H, dd, J = 12.2, 13.2 Hz, H β -9), 1.72 (3H, d, J = 1.2 Hz, H₃-16), 1.76, 1.91 (1H each, both m, H₂-6), 2.27, 2.48 (1H, each, both m, H₂-14), 2.32 (2H, m, H₂-13), 2.47 (1H, dd, J = 5.5, 13.2 Hz, H α -9), 2.48, 2.53 (1H each, both m, H₂-5), 2.60 (1H, br s, H-11), 3.08 (1H, br d, J = ca. 9 Hz, H-7), 4.04 (1H, d, J = 10.7 Hz, H-2), 4.97, 5.02 (1H each, both d, J = 11.3 Hz, H₂-15), 5.68 (1H, dd, J = 5.5, 11.0 Hz, H-10), 5.69 (1H, br d, J = ca. 11 Hz, H-3), 7.37 (2H, dd, J = 7.8, 8.5 Hz, H-3^{'''}, 5^{'''}), 7.53 (1H, m, H-5^{''}), 7.53 (1H, tt, J = 1.2, 7.8 Hz, H-4^{'''}), 8.08 (2H, dd, J = 1.2, 8.5 Hz, H-2^{'''}, 6^{'''}), 8.38 (1H, ddd, J = 1.9, 2.2, 8.0 Hz, H-6^{''}), 8.71 (1H, br s, H-4^{''}), 9.11 (1H, br s, H-2^{''}). EI-MS (%): m/z 545 (M⁺, 2), 527 (M⁺-H₂O, 1), 423 (29), 300 (45), 121 (100), 105 (95).

2d: A white powder. High resolution EI-MS: Calcd for $C_{26}H_{35}NO_5$ (M⁺): 441.2515. Found: 441.2520. ¹H NMR (CD₃OD): δ 1.42 (1H, dd, J = 13.1, 15.3 Hz, H β -9), 1.45, 1.63, 1.82 (3H

each, all s, H₃-17, 19, 20), 1.67 (3H, d, J = 1.2 Hz, H₃-16), 1.75, 1.88 (1H each, both m, H₂-6), 1.99, 2.23 (1H, each, both m, H₂-14), 2.34 (2H, m, H₂-13), 2.34, 2.63 (1H each, both m, H₂-5), 2.45 (1H, dd-like, H α -9), 2.48 (1H, br s, H-11), 3.06 (1H, br d, J = ca. 9 Hz, H-7), 3.97, 4.06 (1H each, both d, J = 10.7 Hz, H₂-15), 4.49 (1H, d, J = 10.4 Hz, H-2), 5.58 (1H, dd, J = 5.8, 13.5 Hz, H-10), 5.67 (1H, br d, J = ca. 10 Hz, H-3), 7.60 (1H, m, H-5"), 8.38 (1H, br d, J = ca. 8 Hz, H-6"), 8.79 (1H, br s, H-4"), 9.13 (1H, br s, H-2"). EI-MS (%): *m/z* 441 (M⁺, 1), 423 (M⁺-H₂O, 6), 300 (25), 121 (100).

2e: A white powder. High resolution positive-ion FAB-MS: Calcd for $C_{27}H_{37}O_5$ (M+H)⁺: 441.2641. Found: 441.2642. ¹H NMR (CD₃OD): δ 1.37, 1.73, 1.77 (3H each, all s, H₃-17, 19, 20), 1.66 (3H, d, J = 1.2 Hz, H₃-16), 2.52 (1H, br s, H-11), 3.03 (1H, br d, J = ca. 10 Hz, H-7), 3.98 (1H, d, J = 10.3 Hz, H-2), 4.14 (1H, m, H-10), 4.70, 4.98 (1H each, both d, J = 10.7 Hz, H₂-15), 5.59 (1H, br d, J = ca. 10 Hz, H-3), 7.46 (2H, dd, J = 7.8, 8.2 Hz, H-3",5"), 7.58 (1H, t-like, H-4"'), 8.07 (2H, dd, J = 1.2, 8.2 Hz, H-2",6"'). Positive-ion FAB-MS: m/z 441 (M+H)⁺.

Triphenylphosphine (PPh₃) Reduction of Nigellamines B_1 (3) and B_2 (4). A solution of 3 (20.0 mg) or 4 (15.0 mg) in CH_2Cl_2 (1.0 mL) was treated with PPh₃ (20.0 mg), and the mixture was stirred at 0 °C for 30 min. Removal of the solvent from the filtrate under reduced pressure furnished a residue, which was purified by HPLC [detection: RI, column: YMC-Pack ODS-5-A, 250×20 mm i.d., mobile phase: MeOH–H₂O (80:20 or 75:25, v/v)] to give 3a (13.2 mg, 68% from 3) or 4a (9.4 mg, 64% from 4), respectively.

3a: A white powder, $[\alpha]^{24}_{D}$ +22.8° (*c*=0.20, CHCl₃). High resolution EI-MS: Calcd for C₄₀H₄₃NO₈ (M⁺): 665.2988. Found: 665.2995. CD (MeOH, $\Delta \varepsilon$): +8.01 (216 nm), -1.54 (236 nm). UV (MeOH, log ε): 226 (4.52), 264 (3.68) nm. IR (KBr): 3517, 1717, 1647, 1636, 1592, 1453, 1279, 1115, 1026, 941, 756, 712 cm⁻¹. ¹³C NMR (CDCl₃): δ_{C} 54.6 (C-1), 73.3 (C-2), 122.6 (C-3), 141.1 (C-4), 38.1 (C-5), 23.0 (C-6), 66.8 (C-7), 59.0 (C-8), 41.4 (C-9), 73.9 (C-10), 52.0 (C-11), 149.0 (C-12), 125.2 (C-13), 38.2 (C-14), 66.2 (C-15), 17.4 (C-16), 17.1 (C-17), 71.5 (C-18), 32.1 (C-19), 32.5 (C-20), 129.8 (C-1'), 129.4 (C-2',6'), 127.9 (C-3',5'), 132.5 (C-4'), 166.2 (C-7'), 125.5 (C-1''), 150.7 (C-2''), 153.5 (C-4''), 123.3 (C-5''), 137.0 (C-6''), 164.9 (C-7''), 130.0 (C-1'''), 129.9 (C-2''',6'''), 128.2 (C-3''',5'''), 132.9 (C-4'''), 166.3 (C-7'''). EI-MS (%): *m/z* 665 (M⁺, 13), 122 (100).

4a: A white powder, $[\alpha]^{26}_{D}$ +22.8° (*c*=0.40, CHCl₃). High resolution Positive-ion FAB-MS: Calcd for C₃₉H₄₃N₂O₈ (M+H)⁺: 667.3019. Found: 667.3022. CD (MeOH, $\Delta \varepsilon$): +7.37 (221 nm), -0.41 (256 nm). UV (MeOH, log ε): 222 (4.58), 264 (3.94) nm. IR (KBr): 3415, 1717, 1647, 1636, 1592, 1456, 1283, 1113, 1026, 745, 714 cm⁻¹. ¹³C NMR (CDCl₃): δ_{C} 54.8 (C-1), 74.1 (C-2), 122.4 (C-3), 141.9 (C-4), 38.2 (C-5), 23.0 (C-6), 66.9 (C-7), 59.1 (C-8), 41.5 (C-9), 74.0 (C-10), 52.1 (C-11), 149.5 (C-12), 125.2 (C-13), 38.3 (C-14), 66.2 (C-15), 17.4 (C-16), 17.1 (C-17), 71.6 (C-18), 32.2 (C-19), 32.7 (C-20), 126.0 (C-1'), 150.9 (C-2'), 153.2 (C-4'), 122.9 (C-5'), 136.9 (C-6'), 165.2 (C-7'), 125.6 (C-1''), 150.9 (C-2''), 153.6 (C-4''), 123.8 (C-5''), 137.3 (C-6''), 165.1 (C-7''), 130.1 (C-1'''), 129.9 (C-2''',6'''), 128.6 (C-3''',5'''), 133.3 (C-4'''). Positive-ion FAB-MS: *m/z* 667 (M+H)⁺.

Treatment of 3a with 0.1% NaOMe–MeOH. A solution of **3a** (5.0 mg) in 0.1% NaOMe–MeOH (2.0 mL) was stirred at room temperature for 8 h. From the reaction mixture, methyl nicotinate and methyl benzoate were identified by HPLC analyses and **3b** (1.9 mg, 72%) was purified by the similar procedure.

On the other hand, a solution of **3a** (8.0 mg) in 0.1% NaOMe–MeOH (2.0 mL) was stirred at 0°C for 3.5 h. The reaction mixture was poured into ice-water and the whole was extracted with EtOAc. The EtOAc extract was successively washed with brine then dried over MgSO₄ powder and filtrated. Evaporation of the solvent from the filtrate under reduced pressure furnished a residue, which was purified by HPLC [detection: RI, column: YMC-Pack ODS-5-A, 250×20 mm i.d., mobile phase: MeOH–H₂O (80:20, v/v)] to give **3c** (2.3 mg, 42%) and 10-desacyl derivative (**3d**, 2.5 mg, 37%).

3b: A white powder, $[\alpha]^{24}_{D}$ +35.6° (*c*=0.10, MeOH). High resolution EI-MS: Calcd for $C_{20}H_{30}O_4$ (M⁺–H₂O): 334.2144. Found: 334.2140. IR (KBr): 3346, 1653, 1559, 1507, 1387, 1262, 1120, 1044, 938, 756, 718 cm⁻¹. ¹³C NMR (CDCl₃): δ_C 56.3 (C-1), 72.3 (C-2), 127.6 (C-3), 137.3 (C-4), 38.1 (C-5), 23.0 (C-6), 66.6 (C-7), 59.5 (C-8), 43.3 (C-9), 70.1 (C-10), 52.4 (C-11), 149.1 (C-12), 126.2 (C-13), 36.4 (C-14), 63.6 (C-15), 16.8 (C-16), 17.5 (C-17), 71.6 (C-18), 33.0 (C-19), 33.2 (C-20). EI-MS (%): *m/z* 334 (M⁺–H₂O, 1), 120 (100).

3c: A white powder, $[\alpha]^{24}_{D}$ +32.9° (*c*=0.20, CHCl₃). High resolution positive-ion CI-MS: Calcd for C₂₇H₃₇O₆ (M+H)⁺: 457.2590. Found: 457.2587. UV (MeOH, log ε): 230 (3.93), 273 (2.84) nm. IR (KBr): 3368, 1717, 1636, 1601, 1584, 1453, 1279, 1098, 1026, 938, 756, 714 cm⁻¹. ¹³C NMR (CDCl₃): δ_{C} 56.6 (C-1), 74.5 (C-2), 122.8 (C-3), 140.0 (C-4), 38.0 (C-5), 23.0 (C-6), 66.8 (C-7), 59.4 (C-8), 43.1 (C-9), 70.2 (C-10), 52.4 (C-11), 148.5 (C-12), 126.3 (C-13), 34.7 (C-14), 61.1 (C-15), 16.8 (C-16), 17.7 (C-17), 71.6 (C-18), 32.7 (C-19), 33.1 (C-20), 130.2 (C-1'), 129.6 (C-2',6'), 128.5 (C-3',5'), 133.1 (C-4'), 166.0 (C-7'). Positive-ion CI-MS: *m/z* 457 (M+H)⁺.

3d: A white powder, $[\alpha]^{24}_{D}$ +18.2° (c=0.20, CHCl₃). High resolution positive-ion CI-MS: Calcd for $C_{34}H_{41}O_7$ (M+H)⁺: 561.2852. Found: 561.2869. UV (MeOH, log ε): 228 (4.11), 274 (3.03) nm. IR (KBr): 3517, 1717, 1647, 1603, 1559, 1451, 1283, 1117, 1026, 941, 756, 712 cm⁻¹. ¹H NMR (CDCl₃): δ 1.38, 1.54, 1.56 (3H each, all s, H₃-17, 19, 20), 1.62 (1H, m, $H\beta$ -9), 1.66 (1H, m, H\alpha-6), 1.89 (3H, d, J = 0.9 Hz, H_3 -16), 1.96 (1H, m, $H\beta$ -6), 2.32 (1H, m, $H\alpha$ -5), 2.41 (1H, dd, J = 5.5, 13.5 Hz, $H\alpha$ -9), 2.44 (1H, ddd, J = 4.9, 12.5, 12.5 Hz, $H\beta$ -5), 2.64 (1H, dd, J = 3.1, 17.7 Hz, H α -14), 2.73 (1H, br d, J = ca. 18 Hz, H β -14), 2.75 (1H, br s, H-11), 2.95 (1H, br d, J = ca. 10 Hz, H-7), 4.62 (1H, br dd, J = ca. 6, 13 Hz, H-10), 4.87, 5.29 (1H each, both d, J = 11.0 Hz, H₂-15), 5.46 (1H, d, J = 10.4 Hz, H-2), 5.67 (1H, dd, J = 0.9, 10.4 Hz, H-3), 5.73 (1H, br s, H-13), 7.12 (2H, dd, J = 7.7, 8.3 Hz, H-3',5'), 7.42 (1H, tt, J = 1.3, 7.7 Hz, H-4'), 7.43 (2H, dd, J = 7.7, 8.3 Hz, H-3", 5"'), 7.58 (1H, tt, J = 1.3, 7.7 Hz, H-4"'), 7.82 (2H, dd, J = 1.3, 8.3 Hz, H-2',6'), 8.07 (2H, dd, J = 1.3, 8.3 Hz, H-2'',6''). ¹³C NMR (CDCl₃): δ_{C} 54.3 (C-1), 73.9 (C-2), 123.1 (C-3), 140.6 (C-4), 38.1 (C-5), 22.9 (C-6), 66.6 (C-7), 59.4 (C-8), 43.8 (C-9), 70.0 (C-10), 53.2 (C-11), 149.6 (C-12), 125.5 (C-13), 37.9 (C-14), 65.7 (C-15), 17.0 (C-16), 17.5 (C-17), 71.7 (C-18), 33.1 (C-19), 33.2 (C-20), 130.1 (C-1'), 129.6 (C-2',6'), 128.1 (C-3',5'), 132.8 (C-4'), 166.4 (C-7'), 130.4 (C-1'''), 129.8 (C-2''',6'''), 128.5 (C-3"',5"'), 133.0 (C-4"'), 166.4 (C-7"'). Positive-ion CI-MS: m/z 561 (M+H)+.

Treatment of 4a with 0.1% NaOMe–MeOH. A solution of **4a** (6.0 mg) in 0.1% NaOMe–MeOH (2.0 mL) was stirred at room temperature for 8 h. From the reaction mixture, methyl nicotinate and methyl benzoate were identified by HPLC analyses and **3b** (2.1 mg, 66%) was purified by the similar procedure.















