

Supporting Information

B(C₆F₅)₃-Catalyzed Allylation of Propargyl Acetates with Allylsilanes

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NMR spectra were recorded on Bruker Avance DPX-400 (400 MHz) and DRX-500 (500 MHz) instruments. GC/MS analyses were performed on a Hewlett Packard Model 6890 GC interfaced to a Hewlett Packard Model 5973 mass selective detector (15 m x 0.25 mm capillary column, HP-5MS). The same GC system with FID (30 m x 0.25 mm capillary column, HP-5) was used for capillary GLC analyses. HR EI MS analysis was performed on a JEOL GCmate II mass spectrometer. Elemental analyses were made by Midwest Microlab, Indiana.

All manipulations were conducted under an argon atmosphere using a combination of glovebox and standard Schlenk techniques. Anhydrous ether, THF, dichloromethane, triethylamine, and pyridine were purchased from Aldrich and stored over calcium hydride. B(C₃F₅)₃ is commercially available, but for our purpose it was prepared according to the known procedure.¹ Compounds **1a**,² **1c**,³ **4a**,⁴ were prepared according to the known procedures. Preparation of other substrates is provided below.

The spectral data for new compounds **1d-s**, **2c-m**, **4b**, **5b** are provided below, as well as for known compound **2a**,⁵ **2b**,⁶ **4a**,⁴ **5a**⁷ for which spectral data presented in literature are incomplete. (+) and (-) represent positive and negative intensities of signals in ¹³C DEPT-135 experiment.

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- (1) (a) Massey, A. G.; Park, A. J. *J. Organomet. Chem.* **1964**, *2*, 245. (b) Massey, A. G.; Park, A. J. *J. Organomet. Chem.* **1966**, *5*, 218.
 - (2) Spee, M. P. R.; Boersma, J.; Meijer, M. D.; Slagt, M. Q.; van Koten, G.; Geus, J. W. *J. Org. Chem.* **2001**, *66*, 1647.
 - (3) Fleming, I.; Higgins, D.; Lawrence, N. J.; Thomas, A. P. *J. Chem. Soc. Perkin Trans. I* **1992**, 3331.
 - (4) Mahrwald, R.; Quint, S. *Tetrahedron* **2000**, *56*, 7463.
 - (5) Luzung, M. R.; Toste, F. D. *J. Am. Chem. Soc.* **2003**, *125*, 15760.
 - (6) Ma, S.-M.; Wang, L.-S. *Chin. J. Chem.* **1999**, *17*, 531.
 - (7) Renard, D.; Rezaei, H.; Zard, S. Z. *Syn. Lett.* **2002**, 1257.

1d: To a stirred solution of 4-phenylbut-3-yn-2-ol (**1a**) (731 mg, 5.0 mmol), pyridine (15.0 mmol, 1.2 mL), and DMAP (15 mg) in anhydrous CH_2Cl_2 (10 mL) at 0°C was slowly added pivaloyl chloride (10 mmol, 1.2 mL). The mixture was warmed to room temperature, and stirred overnight, quenched with 2M aqueous HCl, and extracted with ether. The combined ethereal phases were washed (saturated aqueous NaHCO_3 , brine), dried (MgSO_4), and concentrated in vacuum. The residue was purified by preparative column chromatography on Silica gel, eluent hexane-EtOAc (30:1). Yield 1.15 g (5.0 mmol, 100%). ^1H NMR (500.13 MHz, CDCl_3) δ 7.44-7.42 (m, 2H), 7.30-7.27 (m, 3H), 5.67 (q, $J = 6.4$ Hz, 1H), 1.56 (d, $J = 6.4$ Hz, 3H), 1.23 (s, 9H); ^{13}C NMR (125.76 MHz, CDCl_3) δ 177.4, 131.8 (+), 128.5 (+, 2C), 128.3 (+, 2C), 122.5, 87.7, 84.3, 60.6 (+), 38.7, 27.1 (+, 3C), 21.4 (+); GC/MS m/z 230 (M^+ , 4.8%), 174 ($\text{M}^+ - \text{C}_4\text{H}_8$), 128 ($\text{M}^+ - (\text{CH}_3)_3\text{CCOOH}$, 100%).

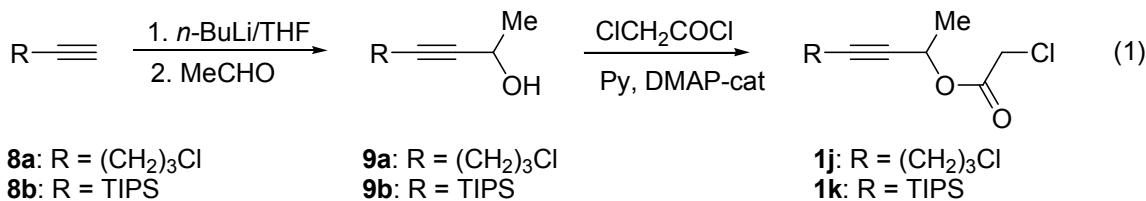
1e: To a stirred solution of **1a** (1.46 g, 10.0 mmol), triethylamine (16.0 mmol, 2.2 mL), and DMAP (15 mg) in anhydrous hexane (15 mL) at 0°C was slowly added trifluoroacetic anhydride (15.0 mmol, 2.1 mL). The mixture was warmed up to room temperature and quenched with ice-cold water and extracted with CH_2Cl_2 . The combined organic phases were (with ice-cold water, brine), dried (MgSO_4), filtered, and concentrated in vacuum. The residue was purified by distillation in vacuum, bp 62°C (1 mm Hg). Yield 1.40 g (5.78 mmol, 58%). ^1H NMR (500.13 MHz, CDCl_3) δ 7.48-7.46 (m, 2H), 7.37-7.32 (m, 3H), 5.83 (q, $J = 6.6$ Hz, 1H), 1.73 (d, $J = 6.6$ Hz, 3H); ^{13}C NMR (125.76 MHz, CDCl_3) δ 157.0 (q, $^2J_{\text{CF}} = 42.6$ Hz), 132.4 (+, 2C), 129.6 (+), 128.8 (+, 2C), 121.9, 114.8 (q, $^1J_{\text{CF}} = 284.9$ Hz), 87.3, 85.0, 66.0 (+), 21.6 (+); GC/MS m/z 242 (M^+ , 18%), 173 ($\text{M}^+ - \text{CF}_3$, 8.6%), 128 ($\text{M}^+ - \text{F}_3\text{CCOOH}$, 100%).

1f: To a stirred solution of **1a** (1.46 g, 10.0 mmol), pyridine (30.0 mmol, 2.4 mL), and DMAP (15 mg) in anhydrous CH_2Cl_2 (15 mL) at 0°C was slowly added trichloroacetic anhydride (15.0 mmol, 2.1 mL). The mixture was warmed to room temperature, and stirred overnight, quenched with diluted aqueous HCl, and extracted with CH_2Cl_2 . Combined organic phases were washed (10% aq. Na_2CO_3 , brine), dried (MgSO_4), filtered and concentrated in vacuum. The residue was purified by preparative column chromatography on Silica gel, eluent hexane-EtOAc (50:1). Yield 2.19 g (7.50 mmol, 75%). ^1H NMR (500.13 MHz, CDCl_3) δ 7.50-7.47 (m, 2H), 7.37-7.31 (m, 3H), 5.80 (q, $J = 7.0$ Hz, 1H), 1.75 (d, $J = 7.0$ Hz, 3H); ^{13}C NMR (125.76 MHz, CDCl_3) δ 160.9, 132.0 (+), 129.1 (+, 2C), 128.4 (+, 2C), 121.7, 89.8, 86.8, 85.0, 66.7 (+), 21.1 (+); GC/MS m/z 290 (M^+ , 1.4%), 255 ($\text{M}^+ - \text{Cl}$, 19%), 128 ($\text{M}^+ - \text{Cl}_3\text{CCOOH}$, 100%).

1g: To a stirred solution of **1a** (2.92 g, 20.0 mmol), pyridine (28.0 mmol, 2.3 mL), and DMAP (10 mg) in anhydrous CH_2Cl_2 (20 mL) at 0°C was slowly added chloroacetyl chloride (24.0 mmol, 1.9 mL). The mixture was warmed to room temperature, and stirred overnight, quenched with 2M aqueous HCl, and extracted with ether. The combined ethereal phases were washed (saturated aqueous NaHCO_3 , brine), dried (MgSO_4), and concentrated in vacuum. The residue was purified by preparative column chromatography on Silica gel, eluent hexane-EtOAc (30:1). Yield 3.44 g (15.4 mmol, 77%). ^1H NMR (400.13 MHz, CDCl_3) δ 7.46-7.43 (m, 2H), 7.33-7.28 (m, 3H), 5.75 (q, $J = 6.7$ Hz, 1H), 4.10 (s, 2H), 1.62 (d, $J = 6.7$ Hz, 3H); ^{13}C NMR (100.62 MHz, CDCl_3) δ

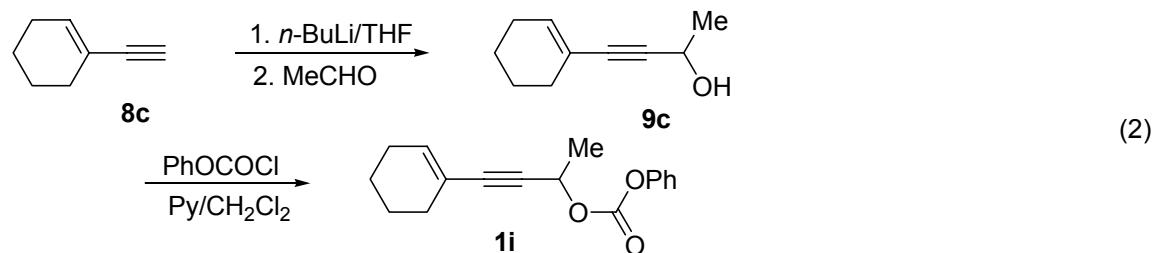
166.3, 131.9 (+, 2C), 128.9 (+), 128.3 (+, 2C), 121.9, 86.4, 85.5, 62.9 (+), 41.0 (-), 21.4 (+); GC/MS m/z 222 (M^+ , 5%), 187 (M^+-Cl , 45%), 128 (M^+-ClCH_2COOH , 100%).

1h: To a stirred solution of **1a** (731 mg, 5.0 mmol) and pyridine (10.0 mmol, 0.81 mL) in anhydrous CH_2Cl_2 (15 mL) at 0°C was slowly added phenyl chloroformate (10.0 mmol, 1.3 mL). Standard aqueous work-up (see procedure for preparation of **1g**), and purification by preparative column chromatography on Silica gel (eluent hexane-EtOAc 20:1) gave 1.26 g (4.73 mmol, 95%) of **1h**. 1H NMR (500.13 MHz, $CDCl_3$) δ 7.49-7.47 (m, 2H), 7.43-7.38 (m, 2H), 7.35-7.30 (m, 2H), 7.29-7.26 (m, 1H), 7.25-7.21 (m, 2H), 5.67 (q, $J = 6.6$ Hz, 1H), 1.73 (d, $J = 6.6$ Hz, 3H); ^{13}C NMR (125.76 MHz, $CDCl_3$) δ 153.2, 151.5, 132.4 (+, 2C), 129.9 (+, 2C), 129.3 (+), 128.7 (+, 2C), 126.5 (+), 122.4, 121.5 (+, 2C), 86.6, 86.4, 66.2 (+), 22.0 (+);

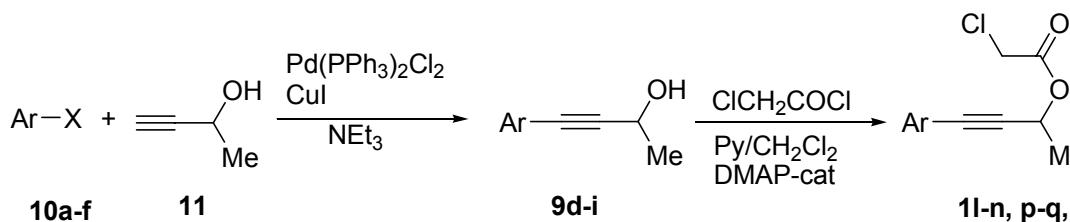


1j: To a stirred solution of 5-chloropentyne **8a** (524 μ L, 5.0 mmol) in anhydrous THF (10 mL) at -78°C was added n-BuLi (2.5M in hexane, 2.2 mL, 5.5 mmol). The mixture was stirred for 30 min at -78°C and quenched with ethanal (600 μ L, 10.6 mmol). Saturated aqueous NH_4Cl was added, and the mixture was extracted (ether), washed (water, brine), dried ($MgSO_4$), filtered, and concentrated in vacuum. The obtained crude propargylic alcohol **9a** was converted into chloroacetate **1j** according to procedure described above for the preparation of **1g**. Yield 981 mg (4.4 mmol, 88%). 1H NMR (500.13 MHz, $CDCl_3$) δ 5.49 (qt, $J = 6.6$ Hz, 1.5 Hz, 1H), 4.06 (s, 2H), 3.62 (t, $J = 6.3$ Hz, 2H), 2.40 (td, $J = 6.9$ Hz, 1.5 Hz, 2H), 1.95 (ps-quintet, $J = 6.6$ Hz, 2H), 1.50 (d, $J = 6.6$ Hz, 3H); ^{13}C NMR (125.76 MHz, $CDCl_3$) δ 166.7, 84.9, 79.2, 63.2 (+), 43.9 (-), 41.3 (-), 31.4 (-), 22.0 (+), 16.5 (-); GC/MS m/z 187 (M^+-Cl , 45%), 100 (75%), 77 (100%).

1k: Similar procedure starting from triisopropylsilylacetylene **8b** gave chloroacetate **1k**. Yield 1.29 g (4.26 mmol, 85%). 1H NMR (500.13 MHz, $CDCl_3$) δ 5.54 (q, $J = 6.6$ Hz, 1H); 4.07 (m, 2H), 1.54 (d, $J = 6.6$ Hz, 3H), 1.07-1.03 (m, 21H); ^{13}C NMR (125.76 MHz, $CDCl_3$) δ 166.2, 104.5, 87.0, 62.8 (+), 40.9 (-), 21.5 (+), 18.5 (+, 6C), 11.0 (+, 3C); GC/MS m/z 259 ($M^+-(CH_3)_2CH$, 11%), 207 (11%), 165 (18%), 123 (35%), 109 (47%), 95 (100%).

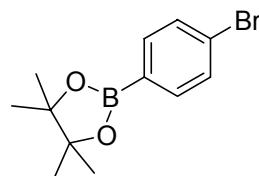


1i: To a stirred solution of 1-ethynylcyclohexene **8c** (0.6 mL, 5 mmol) in anhydrous THF (10 mL) at -78°C was added n-BuLi (2.5M in hexane, 2.2 mL, 5.5 mmol). The mixture was stirred for 30 min at -78°C and quenched with ethanal (0.55 mL, 10 mmol). Saturated aqueous NH₄Cl was added, and the mixture was extracted (ether), washed (water, brine), dried (MgSO₄), filtered, and concentrated in vacuum. The obtained crude alcohol **9c** was transformed into phenylcarbonate **1i** according to procedure described above for the preparation of **1h**. Yield 1.314 g (4.86 mmol, 97%). ¹H NMR (500.13 MHz, CDCl₃) δ 7.40-7.37 (m, 2H), 7.26-7.23 (m, 1H), 7.21-7.19 (m, 2H), 6.17 (tt, *J* = 4.0 Hz, 1.8 Hz, 1H), 5.55 (q, *J* = 6.6 Hz, 1H), 2.14-2.07 (m, 4H), 1.63 (d, *J* = 6.6 Hz, 3H), 1.65-1.61 (m, 2H), 1.61-1.55 (m, 2H); ¹³C NMR (125.76 MHz, CDCl₃) δ 153.2, 151.5, 136.9 (+), 129.9 (+, 2C), 126.4 (+), 121.3 (+, 2C), 120.1, 88.2, 83.9, 66.4 (+), 29.3 (-), 26.0 (-), 22.6 (-), 22.1 (+), 21.8 (-); GC/MS *m/z* 270 (M⁺, 2%), 226 (M⁺-CO₂, 5%), 133 (M⁺-PhOCO₂, 100%).



10a, 9d, 1l: Ar = *p*-PB-C₆H₄; **10a:** X = Br
10b, 9e, 1m: Ar = *p*-CF₃C₆H₄; **10b:** X = Br
10c, 9f, 1n: Ar = *o*-FC₆H₄; **10c:** X = I
10d, 9g, 1p: Ar = *p*-EtOCOC₆H₄; **10d:** X = Br
10e, 9h, 1q: Ar = *p*-O₂NC₆H₄; **10e:** X = Br
10f, 9i, 1t: Ar = *p*-MeC₆H₄; **10f:** X = I

1l: Ar = *p*-PB-C₆H₄



1l: Pd(PPh₃)₂Cl₂ (70 mg, 0.1 mmol), CuI (38mg, 0.2 mmol), and **10a**⁸ (1.42 g, 5 mmol) were dissolved in dry Et₃N (10 mL) and **11** (590 μL, 7.5 mmol) was added to the solution. The reaction was heated to 80°C and stirred overnight. When **10a** had been consumed, the reaction was cooled to room temperature and quenched with 2M aqueous HCl. The mixture was extracted (CH₂Cl₂), washed (sat. NaHCO₃, brine), dried (MgSO₄), filtered, and concentrated in vacuum. The crude alcohol **9d** was converted to the chloroacetate **1l** according to procedure described above for the preparation of **1g**. Yield 0.872 g (2.5 mmol, 50%). ¹H NMR (500.13 MHz, CDCl₃) δ 7.74 (d, *J* = 7.7 Hz, 2H), 7.43 (d, *J* = 7.7 Hz, 2H), 5.76 (q, *J* = 6.7 Hz, 1H), 4.12 (s, 2H), 1.63 (d, *J* = 6.7 Hz, 3H), 1.34 (s, 12H); ¹³C NMR (125.76 MHz, CDCl₃)⁹ δ 166.7, 134.9 (+, 2C), 131.4 (+, 2C), 124.9, 87.9, 86.0, 84.4 (2C), 63.3 (+), 41.4 (-), 25.3 (+, 4C), 21.8 (+); ¹¹B NMR (160.46

(8) Koolmeister, T.; Sodergren, M.; Scobie, M. *Tetrahedron Lett.* **2002**, 43, 5965.

(9) Signal of the aromatic quaternary carbon next to boron atom cannot be observed in the spectrum recorded at room temperature due to extreme broadening, caused by quick relaxation of the ¹³C nuclei via the ¹³C/¹¹B dipole/quadrupole interaction.

MHz, CDCl₃) δ 30.6; GC/MS *m/z* 348 (M⁺, <1%), 313 (M⁺-Cl, 7%), 254 (M⁺-ClCH₂COOH, 45%), 168 (50%), 154 (100%).

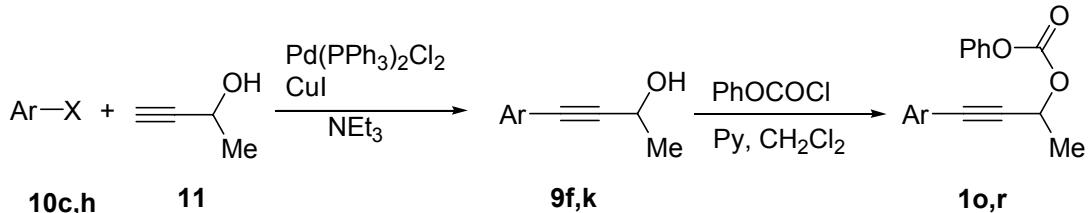
1m: Similar procedure starting from **10b** gave chloroacetate **1m**. Yield 1.15 g (3.95 mmol, 79%). ¹H NMR (500.13 MHz, CDCl₃) δ 7.57 (d, *J* = 8.4 Hz, 2H), 7.54 (d, *J* = 8.4 Hz, 2H), 5.75 (q, *J* = 6.6 Hz, 1H), 4.12 (s, 2H), 1.64 (d, *J* = 6.6 Hz, 3H); ¹³C NMR (125.76 MHz, CDCl₃) δ 166.3, 132.1 (+, 2C), 130.4, 125.7, 125.2 (+, q, ³J_{CF} = 3.7 Hz), 123.8 (q, ¹J_{CF} = 271.9 Hz), 88.7, 84.0, 62.6 (+), 40.8 (-), 21.2 (+); ¹⁹F NMR (470.59 MHz, CDCl₃) δ -64.5; GC/MS *m/z* 290 (M⁺, 8%), 271 (M⁺-F, 4%), 255 (M⁺-Cl), 241 (M⁺-CH₂Cl), 196 (M⁺-ClCH₂COOH, 100%), 177 (61%), 145 (F₃CC₆H₄⁺, 40%), 128 (70%).

1n: Similar procedure starting from **10c** gave chloroacetate **1n**. Yield 0.999 g (4.15 mmol, 83%). ¹H NMR (500.13 MHz, CDCl₃) δ 7.43 (td, *J* = 7.7 Hz, 1.5 Hz, 1H), 7.34-7.29 (m, 1H), 7.09 (t, *J* = 7.7 Hz, 1H), 7.06 (t, *J* = 8.5 Hz, 1H), 5.78 (q, *J* = 6.6 Hz, 1H), 4.12 (s, 2H), 1.64 (d, *J* = 6.9 Hz, 3H); ¹³C NMR (125.76 MHz, CDCl₃) δ 166.7, 163.4 (d, ¹J_{CF} = 251.5 Hz), 134.2 (+), 131.1 (+, d, ³J_{CF} = 7.4 Hz), 124.4 (+, d, ³J_{CF} = 3.7 Hz), 115.9 (+, d, ²J_{CF} = 22.2 Hz), 110.9 (d, ²J_{CF} = 14.8 Hz), 91.8 (d, ³J_{CF} = 2.8 Hz), 79.4, 63.3 (+), 41.3 (-), 21.7 (+); ¹⁹F NMR (470.59 MHz, CDCl₃) δ -111.2; GC/MS *m/z* 240 (M⁺, 7%), 205 (M⁺-Cl, 40%), 146 (M⁺-ClCH₂COOH, 100%).

1p: Similar procedure starting from **10d** gave chloroacetate **1p**. Yield 1.40 g (4.75 mmol, 95%). ¹H NMR (500.13 MHz, CDCl₃) δ 7.98 (d, *J* = 8.2 Hz, 2H), 7.49 (d, *J* = 8.2 Hz, 2H), 5.75 (q, *J* = 6.6 Hz, 1H), 4.36 (q, *J* = 7.0 Hz, 2H), 4.11 (br. s, 4H), 1.63 (d, *J* = 6.6 Hz, 3H), 1.38 (t, *J* = 7.0 Hz, 3H); ¹³C NMR (125.76 MHz, CDCl₃) δ 166.3, 165.9, 131.7 (+, 2C), 130.4, 129.4, 129.3 (+, 2C), 126.4, 89.0, 84.6, 62.7 (+), 61.2 (-), 40.8 (-), 21.2 (+), 14.2 (+); GC/MS *m/z* 294 (M⁺, 3%), 259 (M⁺-Cl, 81%), 249 (M⁺-CH₂Cl, 11%), 200 (M⁺-ClCH₂COOH, 46%), 155 (100%), 127 (50%).

1q: Similar procedure starting from **10e** gave chloroacetate **1q**. Yield 0.843 g (3.15 mmol, 63%). ¹H NMR (500.13 MHz, CDCl₃) δ 8.19 (d, *J* = 8.8 Hz, 2H), 7.59 (d, *J* = 8.8 Hz, 2H), 5.75 (q, *J* = 6.6 Hz, 1H), 4.13 (s, 2H), 1.65 (d, *J* = 6.6 Hz, 3H); ¹³C NMR (125.76 MHz, CDCl₃) δ 166.3, 147.4, 132.6 (+, 2C), 128.7, 123.5 (+, 2C), 91.4, 83.4, 62.4 (+), 40.8 (-), 21.0 (+); GC/MS *m/z* 267 (M⁺, 7%), 232 (M⁺-Cl, 100%), 173 (M⁺-ClCH₂COOH, 80%).

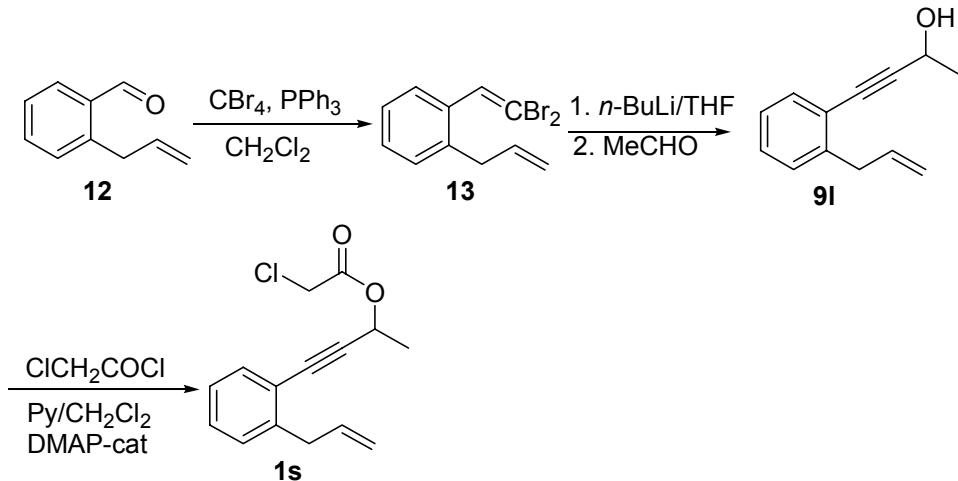
1t: Similar procedure starting from **10f** gave chloroacetate **1t**. Yield 1.152 g (4.86 mmol, 97%). ¹H NMR (500.13 MHz, CDCl₃) δ 7.34 (d, *J* = 7.9 Hz, 2H), 7.12 (d, *J* = 7.9 Hz, 2H), 5.76 (q, *J* = 6.6 Hz, 1H), 4.12 (d, *J* = 15.0 Hz, 1H), 4.10 (d, *J* = 15.0 Hz, 1H), 2.35 (s, 3H), 1.62 (d, *J* = 6.6 Hz, 3H); ¹³C NMR (125.76 MHz, CDCl₃) δ 166.3, 139.0, 131.8 (+, 2C), 129.0 (+, 2C), 118.8, 85.6, 63.0, 40.9 (-), 21.4 (+); GC/MS *m/z* 236 (M⁺, 3%), 201 (M⁺-Cl, 33%), 142 (M⁺-ClCH₂COOH, 100%), 115 (63%).



10c, 9f, 10: Ar = o-FC₆H₄; **10c:** X = I
10h, 9k, 1r, Ar = 2-naphthyl ; **10h:** X = Br

10: Pd(PPh₃)₂Cl₂ (70 mg, 0.1 mmol), CuI (38mg, 0.2 mmol), and **10c** (g, mmol) were dissolved in dry Et₃N (10 mL) and **11** (590 μ L, 7.5 mmol) was added to the solution. The reaction was heated to 60°C and stirred overnight. When **10ca** had been consumed, the reaction was cooled to room temperature and quenched with 2M aqueous HCl. The mixture was extracted (CH₂Cl₂), washed (sat. NaHCO₃, brine), dried (MgSO₄), filtered, and concentrated in vacuum. The crude alcohol **9f** was converted to the phenylcarbonate **10** according to procedure described above for the preparation of **1h**. Yield 1.42 g (5 mmol, 100%). ¹H NMR (500.13 MHz, CDCl₃) δ 7.47 (td, *J* = 7.3 Hz, 1.8 Hz, 1H), 7.40 (m, 2H), 7.35-7.31 (m, 1H), 7.26 (t, *J* = 7.3 Hz, 1H), 7.23 (m, 2H), 7.11 (td, *J* = 7.7 Hz, 0.7 Hz, 1H), 7.09 (ps-t, *J* = 8.4 Hz, 1H), 5.70 (q, *J* = 6.6 Hz, 1H), 1.75 (d, *J* = 6.6 Hz, 3H); ¹³C NMR (125.76 MHz, CDCl₃) δ 162.8 (d, ¹J_{CF} = 251.5 Hz), 152.7, 151.0, 133.7 (+), 130.6 (+, d, ²J_{CF} = 7.4 Hz), 129.4 (+, 2C), 126.1 (+), 123.9 (+, d, ³J_{CF} = 3.7 Hz), 121.0 (+, 2C), 115.5 (+, d, ²J_{CF} = 22.2 Hz), 110.5, 91.2 (d, ³J_{CF} = 3.7 Hz), 79.3, 65.6 (+), 21.3 (+); ¹⁹F NMR (470.59 MHz, CDCl₃) δ -111.1; GC/MS *m/z* 240 (M⁺-CO₂, <1%), 147 (M⁺-CO₃Ph, 100%), 127 (12%), 120 (16%), 94 (24%).

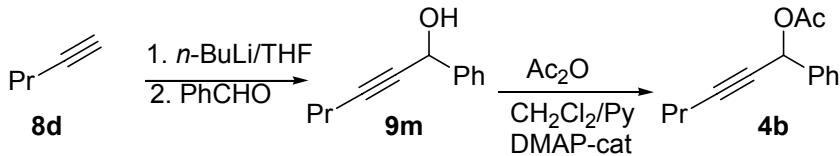
1r: Similar procedure starting from **10h** gave phenylcarbonate **1r**. Yield 1.49 g (4.7 mmol, 94%). ¹H NMR (500.13 MHz, CDCl₃) δ 8.02 (s, 1H), 7.83-7.79 (m, 3H), 7.52-7.50 (m, 3H), 7.43-7.39 (m, 2H), 7.30-7.24 (m, 3H), 5.72 (q, *J* = 6.6 Hz, 1H), 1.77 (d, *J* = 6.6 Hz, 3H); ¹³C NMR (125.76 MHz, CDCl₃) δ 152.8, 151.1, 133.0, 132.8, 132.1 (+), 129.5 (+, 2C), 128.4 (+), 128.0 (+), 127.82 (+), 127.76 (+), 126.9 (+), 126.6 (+), 126.1 (+), 121.0 (+, 2C), 119.3, 86.5, 86.3, 65.8 (+), 21.6 (+); GC/MS *m/z* 316 (M⁺, 3%), 179 (M⁺-CO₃Ph, 100%), 152 (32%).



1s: To a stirred solution of PPh₃ (5.25 g, 20 mmol) in dry CH₂Cl₂ (40 mL) at 0°C was added a solution of CBr₄ (3.28 g, 10 mmol) in dry CH₂Cl₂ (20 mL) *via* cannula. The subsequent yellow solution was stirred for 10 minutes, followed by the addition of **12**¹⁰ in dry CH₂Cl₂ (40 mL). The solution was stirred for 1 hour at 0°C. The reaction was quenched with brine, extracted (CH₂Cl₂), washed (H₂O), dried (MgSO₄), and concentrated. The crude product was diluted (hexanes) and filtered (Silica gel). The dibromide **13** was dissolved in THF (25 mL) and cooled to -78°C. To the reaction was added *n*-BuLi (2.5 M in hexane, 5 mL, 12.5 mmol) and the solution was stirred for 20 minutes before being quenched with ethanal (0.6 mL, 10 mmol). The solution was warmed to 0°C and 2M aqueous HCl was added and the mixture was extracted (ether), washed (water, brine), dried (MgSO₄), filtered, and concentrated in vacuum. The crude alcohol **9I** was converted to the chloroacetate **1s** in a manner described for **1g**. Yield 2.57 g (9.8 mmol, 98%). ¹H NMR (500.13 MHz, CDCl₃) δ 7.43 (d, *J* = 7.7 Hz, 1H), 7.29 (td, *J* = 7.5 Hz, 1.5 Hz, 1H), 7.21 (d, *J* = 7.7 Hz, 1H), 7.17 (td, *J* = 7.5 Hz, 1.1 Hz, 1H), 5.96 (ddt, *J* = 16.5 Hz, 10.6 Hz, 6.6 Hz, 1H), 5.78 (q, *J* = 6.6 Hz, 1H), 5.09 (m, *J* = 16.5 Hz, 1H), 5.08 (m, *J* = 10.6 Hz, 1H), 4.10 (s, 2H), 3.54 (d, *J* = 6.6 Hz, 2H), 1.64 (d, *J* = 6.6 Hz, 3H); ¹³C NMR (125.76 MHz, CDCl₃) δ 166.3, 142.4, 136.3 (+), 132.5 (+), 131.9, 129.1 (+), 128.8 (+), 128.3, 126.1 (+), 121.4, 116.1 (-), 90.2, 84.1, 63.0 (+), 40.9 (-), 38.6 (-), 21.4 (+); GC/MS *m/z* 262 (M⁺, <1%), 227 (M⁺-Cl, <1%), 213 (M⁺-CH₂Cl, 3.8%), 167 (76%), 153 (100%).

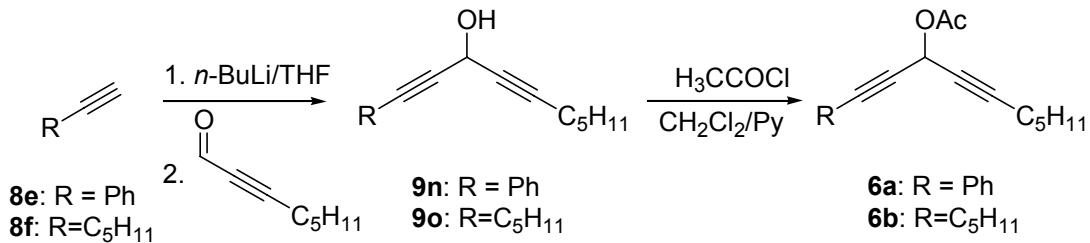
4a: Was prepared by method reported in literature⁴. ¹H NMR (500.13 MHz, CDCl₃) δ 7.67-7.63 (m, 2H), 7.53-7.50 (m, 2H), 7.45-7.40 (m, 3H), 7.35-7.31 (m, 3H), 6.75 (s, 1H), 2.15 (s, 3H); ¹³C NMR (125.76 MHz, CDCl₃) δ 170.3, 137.6, 132.4 (+, 2C), 129.4 (+), 129.3 (+), 129.2 (+, 2C), 129.0 (+, 2C), 128.8 (+, 2C), 128.3 (+, 2C), 122.5, 87.5, 86.1, 66.5 (+), 21.6 (+); GC/MS *m/z* 250 (M⁺, 45%), 208 (M⁺-CH₂CO, 85%), 191 (M⁺-CH₃CO₂, 93%), 189 (100%).

(10) Ashby, E. C.; Coleman, D.; Gamasa, M. *J. Org. Chem.* **1987**, 52, 4079.



4b: To a solution of **8d** (0.99 mL, 10 mmol) in anhydrous THF (10 mL) at -78°C was added *n*-BuLi (2.5M in hexane, 4.4 mL, 11 mmol). The solution was stirred for 30 minutes, warmed to 0°C for 1 hour, and cooled back to -78°C. After 10 minutes, the reaction was quenched with benzaldehyde (1.2 mL, 12 mmol). Upon warmup to room temperature, 2M aqueous HCl was added and the mixture was extracted with ether. Ethereal layers were combined and washed (sat. NaHCO₃, brine), brine (MgSO₄), and concentrated in vacuum to give **9m** of reasonably high purity. Yield 2.16 g (10 mmol, 100%). To a solution of **9m** (0.865 g, 4 mmol) and DMAP (10 mg, 0.08 mmol) in dry CH₂Cl₂ (10 mL) at 0°C was added pyridine (0.45mL, 5.6 mmol). After stirring for several minutes, acetic anhydride (0.5 mL, 4.8 mmol) was added slowly. Reaction was stirred overnight, quenched (2M aqueous HCl), extracted (CH₂Cl₂), washed (brine), dried (MgSO₄), and concentrated in vacuum. The crude oil was purified by preparative column chromatography (eluent hexane-EtOAc-CH₂Cl₂ 30:1:3) to give **4b**. Yield 0.631 g (2.9 mmol, 73%). ¹H NMR (500.13 MHz, CDCl₃) δ 7.55-7.52 (m, 2H), 7.40-7.32 (m, 3H), 6.47 (s, 1H), 2.27-2.24 (m, 2H), 2.09 (s, 3H), 1.58 (ps-sextet, *J* = 7.4 Hz, 2H), 0.99 (t, *J* = 7.4 Hz, 3H); ¹³C NMR (125.76 MHz, CDCl₃) δ 170.3, 138.1, 129.2 (+), 129.0 (+, 2C), 128.1 (+, 2C), 88.6, 77.3, 66.5 (+), 22.3 (-), 21.6 (+), 21.2 (-), 13.9 (+); GC/MS *m/z* 216 (M⁺, 20%), 174 (M⁺-CH₂CO, 45%), 173 (M⁺-C₃H₇, 45%), 156 (M⁺-CH₃CO₂H, 40%), 141 (90%), 128 (80%), 115 (100%).

4c: This compound was prepared from commercially available 1-phenyl-2-propyn-1-ol *via* standard procedure described previously for **1g**. Yield 0.58 g (2.78 mmol, 56%). ¹H NMR (500.13 MHz, CDCl₃) δ 7.57-7.55 (m, 2H), 7.44-7.41 (m, 3H), 6.52 (d, *J* = 1.8 Hz, 1H), 4.12 (d, *J* = 15.0 Hz, 1H), 4.08 (d, *J* = 15.0 Hz, 1H), 2.73 (d, *J* = 1.8 Hz, 1H); ¹³C NMR (125.76 MHz, CDCl₃) δ 166.1, 135.5, 129.5 (+), 128.8 (+, 2C), 127.8 (+, 2C), 79.2, 76.4 (+), 67.1 (+), 40.8 (-); GC/MS *m/z* 208 (M⁺, 4%), 173 (M⁺-Cl, 19%), 131 (M⁺-Ph, 32%), 114 (M⁺-ClCH₂COOH, 100%).



6a: To a stirred at -78°C solution of **8e** (1.1 mL, 10 mmol) in anhydrous THF (15 mL) was added *n*-BuLi (2.5M in hexane, 4.8 mL, 12 mmol). The solution was stirred for 20 minutes, followed by the addition of 2-octynal (1.6 mL, 11 mmol). Solution was warmed to room temperature and 2M aqueous HCl was added. The mixture was extracted (ether), washed (sat. NaHCO₃, brine), dried (MgSO₄), and concentrated in vacuum. The crude

alcohol **9n** was dissolved in anhydrous CH₂Cl₂ (15 mL) and cooled to 0°C. Pyridine (1.6 mL, 19.8 mmol) was added, followed by acetyl chloride (0.9 mL, 12.7 mmol). The reaction was warmed to room temperature and stirred for one hour. When reaction had completed, 2M aqueous HCl was added and mixture was extracted with ether, washed (sat. NaHCO₃, brine), dried (MgSO₄), and concentrated in vacuum. The crude oil was purified by preparative column chromatography on Silica gel (eluent hexane-EtOAc-CH₂Cl₂ 30:1:3) Yield 2.50 g (9.3 mmol, 93%). ¹H NMR (500.13 MHz, CDCl₃) δ 7.49-7.47 (m, 2H), 7.34-7.29 (m, 3H), 6.27 (t, *J* = 2.2 Hz, 1H), 2.25 (td, *J* = 7.3 Hz, 2.2 Hz, 2H), 2.15 (s, 3H), 1.55 (ps-quintet, *J* = 7.3 Hz, 2H), 1.40-1.30 (m, 4H), 0.90 (t, *J* = 7.2 Hz, 3H); ¹³C NMR (125.76 MHz, CDCl₃) δ 169.3, 132.0 (+, 2C), 128.9 (+), 128.2 (+, 2C), 121.7, 86.9, 84.5, 83.4, 74.1, 54.0 (+), 31.0 (-), 27.8 (-), 22.1 (-), 20.9 (+), 18.7 (-), 13.9 (+); GC/MS *m/z* 267 (M⁺-H, 6%), 211 (M⁺-Bu, 30%), 178 (46%), 165 (77%), 152 (100%).

6b: Similar procedure starting from **8f** gave acetate **6b**. Yield 2.62 g (10 mmol, 100%). ¹H NMR (500.13 MHz, CDCl₃) δ 6.01 (m, 1H), 2.22 (td, *J* = 7.2 Hz, 2.2 Hz, 4H), 2.11 (s, 3H), 1.52 (ps-quintet, *J* = 7.3 Hz, 4H), 1.38-1.28 (m, 8H), 0.89 (t, *J* = 7.0 Hz, 6H); ¹³C NMR (125.76 MHz, CDCl₃) δ 169.4, 86.1 (2C), 74.7 (2C), 54.0 (-), 31.0 (-, 2C), 27.9 (-, 2C), 22.1 (-, 2C), 21.0 (+), 18.7 (-, 2C), 13.9 (+, 2C); GC/MS *m/z* 262 (M⁺, 11%), 219 (M⁺-CH₃CO, 27%), 202 (M⁺-CH₃COOH, 25%), 91 (100%).

Procedure for B(C₆F₅)₃-Catalyzed Allylation of **2a** is representative

2a: To a solution of **1g** (223 mg, 1.0 mmol) and allyltrimethylsilane (240μL, 1.5 mmol) in anhydrous CH₂Cl₂ (0.5 mL) was added a solution of B(C₆F₅)₃ (26mg, 0.05 mmol) in anhydrous CH₂Cl₂ (0.5 mL). The reaction was stirred at room temperature for several hours. Upon consumption of **1g**, the reaction was filtered through Silica gel and concentrated. The crude product was purified by preparative column chromatography using Silica gel (eluent hexanes). Yield 162 mg (0.95 mmol, 95%). ¹H NMR (500.13 MHz, CDCl₃) δ 7.43-7.41 (m, 2H), 7.32-7.27 (m, 3H), 5.96 (ddt, *J* = 17.2 Hz, 10.3 Hz, 7.3 Hz, 1H), 5.16 (ddt, *J* = 17.2 Hz, 2.2 Hz, 1.5 Hz, 1H), 5.12 (ddt, *J* = 10.3 Hz, 1.8 Hz, 1.1 Hz, 1H), 2.75 (ps-sextet, *J* = 6.8 Hz, 1H), 2.37-2.27 (m, 2H), 1.28 (d, *J* = 7.0 Hz, 3H); ¹³C NMR (125.76 MHz, CDCl₃) δ 136.5 (+), 132.0 (+, 2C), 128.6 (+, 2C), 128.0 (+), 124.4, 117.1 (-), 94.5, 81.5, 41.6 (-), 26.9 (+), 20.9 (+); GC/MS *m/z* 170 (M⁺, 10%), 155 (M⁺-Me, 20%), 129 (M⁺-C₃H₅, 100%).

2b: ¹H NMR (500.13 MHz, C₆D₆) δ 7.40-7.38 (m, 2H), 7.30-7.27 (m, 3H), 4.84 (br. s 1H), 4.82 (br. s 1H), 2.84 (app. sextet, *J* = 7.0 Hz, 1H), 2.34 (dd, *J* = 13.7 Hz, 7.7 Hz, 1H), 2.20 (dd, *J* = 13.7 Hz, 7.2 Hz, 1H), 1.79 (br. s, 3H), 1.25 (d, *J* = 7.0 Hz, 3H); ¹³C NMR (125.76 MHz, CDCl₃) δ 143.2, 131.6 (+, 2C), 128.1 (+, 2C), 127.5 (+), 124.0, 112.5 (-), 94.5, 80.8, 45.3 (-), 24.9 (+), 22.3 (+), 20.7 (+); GC/MS *m/z* 184 (M⁺, 9%), 169 (M⁺-CH₃, 54%), 128 (M⁺-C₄H₈, 100%). HRMS Calcd for C₁₄H₁₆ 184.1252. Found 184.1262.

2c: ¹H NMR (500.13 MHz, CDCl₃) δ 6.01 (tt, *J* = 4.0 Hz, 1.8 Hz, 1H), 5.87 (ddt, *J* = 17.2, 10.3, 7.0 Hz, 1H), 5.07 (m, 1H), 5.05 (m, 1H), 2.64 (ps-sextet, *J* = 6.6 Hz, 1 H),

2.21 (m, 2 H), 2.08 (m, 4 H), 1.59 (m, 4 H), 1.17 (d, $J = 6.97$ Hz, 3 H); ^{13}C NMR (125.76 MHz, CDCl_3) δ 136.6 (+), 133.8 (+), 121.3, 116.9 (-), 91.6, 83.2, 41.7 (-), 30.1 (-), 26.7 (+), 26.0 (-), 22.8 (-), 22.0 (-), 21.1 (+); GC/MS m/z 174 (M^+ , 10%), 159 ($\text{M}^+ \text{-Me}$, 10%), 133 ($\text{M}^+ \text{-C}_3\text{H}_5$, 95%), 105 ($\text{C}_6\text{H}_9\text{C}\equiv\text{C}^+$, 100%). HRMS Calcd for $\text{C}_{13}\text{H}_{18}$ 174.1409. Found 174.1403.

2d: ^1H NMR (500.13 MHz, CDCl_3) δ 5.85 (ddt, $J = 17.2$ Hz, 10.3 Hz, 7.0 Hz, 1H), 5.07 (m, $J = 17.2$ Hz, 1H), 5.04 (m, $J = 10.3$ Hz, 1H), 3.65 (t, $J = 6.2$ Hz, 2H), 2.47 (m, $J = 7.0$ Hz, 2.2 Hz, 1H), 2.35 (td, $J = 6.8$ Hz, 2.2 Hz, 2H), 2.16 (ps-tq, $J = 7.0$ Hz, 1.5 Hz, 2H), 1.92 (ps-quintet, $J = 6.6$ Hz, 2H), 1.13 (d, $J = 6.6$ Hz, 3H); ^{13}C NMR (125.76 MHz, CDCl_3) δ 136.6 (+), 116.8 (-), 85.8, 79.0, 44.2 (-), 41.8 (-), 32.2 (-), 26.3 (+), 21.3 (+), 16.6 (-); GC/MS m/z 170 (M^+ , <1%), 155 ($\text{M}^+ \text{-Me}$, 1%), 135 ($\text{M}^+ \text{-Cl}$, 7%), 129 ($\text{M}^+ \text{-C}_3\text{H}_5$, 10%), 107 ($\text{M}^+ \text{-}(\text{CH}_2)_2\text{Cl}$, 20%), 93 ($\text{M}^+ \text{-}(\text{CH}_2)_3\text{Cl}$, 100%), 77 ($^+(\text{CH}_2)_3\text{Cl}$, 70%). HRMS Calcd for $\text{C}_{10}\text{H}_{16}\text{Cl}$ 171.0941. Found 171.0938.

2e: ^1H NMR (400.13 MHz, CDCl_3) δ 5.88 (ddt, $J = 17.0$ Hz, 9.9 Hz, 7.0 Hz, 1H), 5.07 (m, $J = 17.0$ Hz, 1H), 5.04 (m, $J = 9.9$ Hz, 1H), 2.54 (ps-sextet, $J = 7.0$ Hz, 1H), 2.21 (ps-tq, $J = 7.0$ Hz, 1.1 Hz, 2H), 1.17 (d, $J = 7.0$ Hz, 3H), 1.06 (m, 3H), 1.05 (m, 18H); ^{13}C NMR (100.61 MHz, CDCl_3) δ 136.0 (+), 116.5 (-), 113.1, 80.1, 41.3 (-), 26.9 (+), 20.8 (+), 18.6 (+, 6C), 11.2 (+, 3C); GC/MS m/z 250 (M^+ , <1%), 207 ($\text{M}^+ \text{-}(\text{CH}_3)_2\text{CH}$, 100%), 137 (53%), 123 (59%), 109 (54%), 59 (62%). HRMS Calcd for $\text{C}_{16}\text{H}_{30}\text{Si}$ 250.2117. Found 250.2133.

2f: ^1H NMR (500.13 MHz, CDCl_3) δ 7.71 (d, $J = 8.1$ Hz, 2H), 7.39 (d, $J = 8.1$ Hz, 2H), 5.93 (ddt, $J = 16.9$ Hz, 9.9 Hz, 7.0 Hz, 1H), 5.12 (m, $J = 16.9$ Hz, 1H), 5.09 (m, $J = 9.9$ Hz, 1H), 2.73 (ps-sextet, $J = 7.0$ Hz, 1H), 2.35-2.24 (m, 2H), 1.34 (s, 12H), 1.25 (d, $J = 7.0$ Hz, 3H); ^{13}C NMR (125.76 MHz, CDCl_3) δ 136.4 (+), 134.9 (+, 2C), 131.2 (+, 2C), 127.1, 117.2 (-), 95.9, 84.3 (2C), 81.7, 41.5 (-), 26.9 (+), 25.3 (+, 4C), 20.9 (+); ^{11}B NMR (160.46 MHz, CDCl_3) δ 30.9; GC/MS m/z 296 (M^+ , 7%), 255 ($\text{M}^+ \text{-C}_3\text{H}_5$, 100%). HRMS Calcd for $\text{C}_{19}\text{H}_{25}\text{O}_2\text{B}$ 296.1948. Found 296.1939.

2g: ^1H NMR (500.13 MHz, C_6D_6) δ 7.54 (d, $J = 8.4$ Hz, 2H), 7.49 (d, $J = 8.4$ Hz, 2H), 5.92 (ddt, $J = 17.2$ Hz, 10.3 Hz, 7.3 Hz, 1H), 5.15 (m, $J = 17.2$ Hz, 1H), 5.11 (m, $J = 10.3$ Hz, 1H), 2.75 (app. sextet, $J = 6.6$ Hz, 1H), 2.33 (dd, $J = 13.9$ Hz, 7.0 Hz, 1H), 2.29 (dd, $J = 13.9$ Hz, 7.0 Hz, 1H), 1.28 (d, $J = 7.0$ Hz, 3H); ^{13}C NMR (125.76 MHz, CDCl_3) δ 135.7 (+), 131.8 (+, 2C), 129.3 (q, $^2J_{\text{CF}} = 33.3$ Hz), 127.8, 125.1 (+, 2C), 123.7 (q, $^1J_{\text{CF}} = 271.9$ Hz), 116.9 (-), 96.8, 80.0, 40.9 (-), 26.5 (+), 20.3 (+); ^{19}F NMR (470.59 MHz, CDCl_3) δ -64.3; GC/MS m/z 238 (M^+ , 8.6%), 197 ($\text{M}^+ \text{-allyl}$, 100%), 177 (77%). HRMS Calcd for $\text{C}_{14}\text{H}_{13}\text{F}_3$ 238.0969. Found 238.0961.

2h: ^1H NMR (500.13 MHz, CDCl_3) δ 7.39 (td, $J = 7.7$ Hz, 1.8 Hz, 1H), 7.24 (m, 1H), 7.08-7.02 (m, 2H), 5.95 (ddt, $J = 17.2$ Hz, 10.3 Hz, 7.3 Hz, 1H), 5.14 (ddt, $J = 17.2$ Hz, 1.7 Hz, 1.5 Hz, 1H), 5.10 (ddt, $J = 10.3$ Hz, 2.2 Hz, 1.1 Hz, 1H), 2.78 (ps-sextet, $J = 6.6$ Hz, 1H), 2.38-2.27 (m, 2H), 1.28 (d, $J = 7.0$ Hz, 3H); ^{13}C NMR (125.76 MHz, CDCl_3) δ 163.2 (d, $^1J_{\text{CF}} = 251.6$ Hz), 136.2 (+), 134.0 (+), 129.6 (+, d, $^3J_{\text{CF}} = 7.4$ Hz), 124.2 (+, d, $^3J_{\text{CF}} = 3.7$ Hz), 117.2 (-), 115.7 (+, d, $^2J_{\text{CF}} = 20.3$ Hz), 112.8 (d, $^2J_{\text{CF}} = 16.6$ Hz), 99.9 (d,

$^3J_{\text{CF}} = 3.7$ Hz), 74.8, 41.4 (-), 27.1 (+), 20.8 (+); ^{19}F NMR (470.59 MHz, CDCl_3) δ - 112.4; GC/MS m/z 188 (M^+ , 7%), 173 ($\text{M}^+ \text{-Me}$, 20%), 147 ($\text{M}^+ \text{-C}_3\text{H}_5$, 100%). HRMS Calcd for $\text{C}_{13}\text{H}_{13}\text{F}$ 188.1001. Found 188.0985.

2i: ^1H NMR (500.13 MHz, CDCl_3) δ 7.95 (d, $J = 8.2$ Hz, 2H), 7.44 (d, $J = 8.2$ Hz, 2H), 5.92 (ddt, $J = 17.2$ Hz, 9.9 Hz, 7.0 Hz, 1H), 5.13 (m, $J = 17.2$ Hz, 1H), 5.10 (m, $J = 9.9$ Hz, 1H), 4.36 (q, $J = 7.3$ Hz, 2H), 2.75 (ps-sextet, $J = 7.0$ Hz, 1H), 2.35-2.25 (m, 2H), 1.39 (t, $J = 7.3$ Hz, 3H), 1.26 (d, $J = 7.0$ Hz, 3H); ^{13}C NMR (125.76 MHz, CDCl_3) δ 166.2, 135.7 (+), 131.4 (+, 2C), 129.3 (+, 2C), 129.2, 128.6, 116.9 (-), 97.3, 80.6, 61.0 (-), 40.9 (-), 26.5 (+), 20.3 (+), 14.3 (+); GC/MS m/z 242 (M^+ , 5%), 201 ($\text{M}^+ \text{-allyl}$, 100%), 169 (18%), 128 (45%). HRMS Calcd for $\text{C}_{16}\text{H}_{18}\text{O}_2$ 242.1307. Found 242.1296.

2j: ^1H NMR (500.13 MHz, CDCl_3) δ 8.15 (d, $J = 8.8$ Hz, 2H), 7.51 (d, $J = 8.8$ Hz, 2H), 5.90 (ddt, $J = 16.9$ Hz, 9.9 Hz, 7.0 Hz, 1H), 5.14 (m, $J = 16.9$ Hz, 1H), 5.11 (m, $J = 9.9$ Hz, 1H), 2.77 (ps-sextet, $J = 7.0$ Hz, 1H), 2.36-2.27 (m, 2H), 1.28 (d, $J = 7.0$ Hz, 3H); ^{13}C NMR (125.76 MHz, CDCl_3) δ 146.6, 135.5 (+), 132.3 (+, 2C), 131.0, 123.5 (+, 2C), 117.1 (-), 100.1, 79.8, 40.7 (-), 26.6 (+), 20.1 (+); GC/MS m/z 215 (M^+ , 7%), 174 ($\text{M}^+ \text{-allyl}$, 100%), 128 ($\text{M}^+ \text{-allyl-NO}_2$, 78%), 115 (53%). HRMS Calcd for $\text{C}_{13}\text{H}_{13}\text{NO}_2$ 215.0946. Found 215.0936.

2k: ^1H NMR (500.13 MHz, CDCl_3) δ 7.94 (s, 1H), 7.82-7.76 (m, 3H), 7.50-7.45 (m, 3H), 6.00 (ddt, $J = 17.2$ Hz, 10.3 Hz, 7.0 Hz, 1H), 5.19 (m, $J = 17.2$ Hz, 1H), 5.15 (m, $J = 10.3$ Hz, 1H), 2.81 (ps-sextet, $J = 6.6$ Hz, 1H), 2.42-2.31 (m, 2H), 1.33 (d, $J = 7.0$ Hz, 3H); ^{13}C NMR (125.76 MHz, CDCl_3) δ 136.0 (+), 133.1, 132.5, 131.1 (+), 128.8 (+), 127.8 (+), 127.7 (+), 127.6 (+), 126.3 (+), 126.2 (+), 121.3, 116.7 (-), 94.5, 81.5, 41.2 (-), 26.6 (+), 20.5 (+); GC/MS m/z 220 (M^+ , 17%), 179 ($\text{M}^+ \text{-allyl}$, 100%), 152 (17%). EA Calcd for $\text{C}_{17}\text{H}_{16}$: C 92.68, H 7.32. Found C 92.68, H 7.30.

2l: ^1H NMR (500.13 MHz, CDCl_3) δ 7.40 (d, $J = 7.3$ Hz, 1H), 7.22 (td, $J = 7.3$ Hz, 1Hz, 1H), 7.19 (d, $J = 7.3$ Hz, 1H), 7.14 (td, $J = 7.3$ Hz, 1.8 Hz, 1H), 6.00 (ddt, $J = 16.9$ Hz, 9.9 Hz, 6.6 Hz, 1H), 5.10 (m, $J = 16.9$ Hz, 1H), 5.07 (m, $J = 9.9$ Hz, 1H), 4.85 (br. s. 1H), 4.83 (br. s. 1H), 3.56 (d, $J = 6.6$ Hz, 2H), 2.90 (sextet, $J = 7.3$ Hz, 1H), 2.36 (dd, $J = 13.6$ Hz, 7.7 Hz, 1H), 2.22 (dd, $J = 13.6$ Hz, 6.6 Hz, 1H), 1.80 (s, 3H), 1.29 (d, $J = 7.0$ Hz, 3H); ^{13}C NMR (125.76 MHz, CDCl_3) δ 143.1, 141.8, 136.9 (+), 132.2 (+), 128.6 (+), 127.7 (+), 125.9 (+), 123.4, 115.7 (-), 112.5 (-), 98.3, 79.4, 45.4 (-), 38.8 (-), 25.1 (+), 22.3 (+), 20.9 (+); GC/MS m/z 224 (M^+ , 4%), 209 ($\text{M}^+ \text{-CH}_3$, 33%), 153 ($\text{M}^+ \text{-C}_5\text{H}_{11}$, 96%), 141 (100%). HRMS Calcd for $\text{C}_{17}\text{H}_{20}$ 224.1565. Found 224.1552.

2m: ^1H NMR (500.13 MHz, CDCl_3) δ 7.30 (d, $J = 7.9$ Hz, 1H), 7.09 (d, $J = 7.9$ Hz, 1H), 5.94 (ddt, $J = 16.9$ Hz, 9.9 Hz, 7.0 Hz, 1H), 5.13 (m, $J = 16.9$ Hz, 1H), 5.09 (m, $J = 9.9$ Hz, 1H), 2.72 (ps-sextet, $J = 7.0$ Hz, 1H), 2.34 (s, 3H), 2.32-2.27 (m, 2H), 1.26 (d, $J = 6.6$ Hz, 3H); ^{13}C NMR (125.76 MHz, CDCl_3) δ 137.5, 136.1, 131.4 (+, 2C), 128.9 (+, 2C), 120.8, 116.6 (-), 93.2, 81.0, 41.2 (-), 26.4 (+), 21.4 (+), 20.5 (+); GC/MS m/z 184 (M^+ , 12%), 169 ($\text{M}^+ \text{-CH}_3$, 10%), 143 ($\text{M}^+ \text{-allyl}$, 100%), 128 ($\text{M}^+ \text{-allyl-CH}_3$, 55%). HRMS Calcd for $\text{C}_{14}\text{H}_{16}$ 184.1252. Found 184.1247.

5a: ^1H NMR (500.13 MHz, CDCl_3) δ 7.48-7.42 (m, 4H), 7.37-7.34 (m, 2H), 7.32-7.29 (m, 3H), 7.29-7.25 (m, 1H), 5.93 (ddt, $J = 17.2$ Hz, 10.3 Hz, 7.3 Hz, 1H), 5.13 (m, 1H), 5.09 (m, 1H), 3.93 (t, $J = 7.0$ Hz, 1H), 2.61 (ps-t, $J = 7.0$ Hz, 2H); ^{13}C NMR (125.76 MHz, CDCl_3) δ 141.8, 135.9 (+), 132.1 (+, 2C), 128.9 (+, 2C), 128.6 (+, 2C), 128.2 (+), 128.0 (+, 2C), 127.2 (+), 124.1, 117.5 (-), 91.4, 84.2, 43.2 (-), 39.0 (+); GC/MS m/z 232 (M^+ , <1%), 191 ($\text{M}^+ - \text{C}_3\text{H}_5$, 100%).

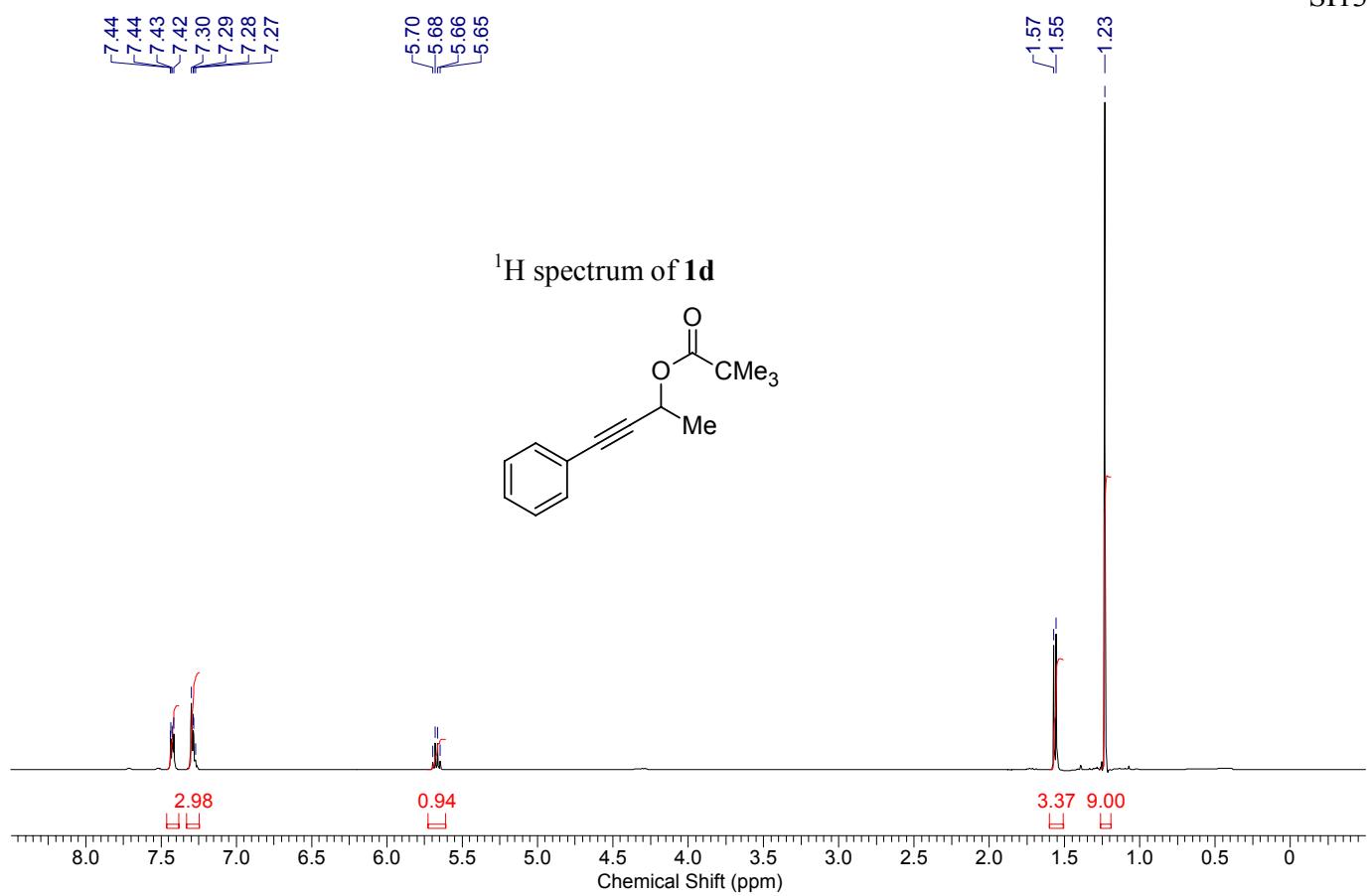
5b: ^1H NMR (500.13 MHz, CDCl_3) δ 7.38-7.36 (m, 2H), 7.34-7.30 (m, 2H), 7.26-7.21 (m, 1H), 5.87 (ddt, $J = 17.2$, 10.3, 7.0 Hz, 1H), 5.08-5.02 (m, 2H), 3.68 (m, $J = 7.3$ Hz, 2.2 Hz, 1H), 2.48 (t, $J = 7.2$ Hz, 2H), 2.22 (m, 2H), 1.56 (ps-sextet, $J = 7.3$ Hz, 2H), 1.01 (t, $J = 7.3$ Hz, 3H); ^{13}C NMR (125.76 MHz, CDCl_3) δ 142.6, 136.3 (+), 128.7 (+, 2C), 127.9 (+, 2C), 127.0 (+), 117.1 (-), 84.0, 81.7, 43.5 (-), 38.5 (+), 22.9 (-), 21.3 (-), 13.9 (+); GC/MS m/z 198 (M^+ , <1%), 157 ($\text{M}^+ - \text{C}_3\text{H}_5$, 100%). HRMS Calcd for $\text{C}_{15}\text{H}_{18}$ 198.1409. Found 198.1413.

5c: ^1H NMR (500.13 MHz, CDCl_3) δ 7.38 (d, $J = 8.1$ Hz, 2H), 7.35 (app. t, 2H), 7.26 (app. t, 1H), 5.87 (ddt, $J = 17.2$ Hz, 10.3 Hz, 7.0 Hz, 1H), 5.09 (m, $J = 17.2$ Hz, 1H), 5.08 (m, $J = 10.3$ Hz, 1H), 3.72 (td, $J = 7.2$ Hz, 2.2 Hz, 1H), 2.54 (ps-t, $J = 7.0$ -7.2 Hz, 2H); ^{13}C NMR (125.76 MHz, CDCl_3) δ 140.7, 135.1, 128.5 (+, 2C), 127.4 (+, 2C), 126.9 (+), 117.2 (-), 85.3, 71.4 (+), 42.4 (-), 37.7 (+); GC/MS m/z 156 (M^+ , 2%), 155 ($\text{M}^+ - \text{H}$, 6%), 141 (7%), 128 (7%), 115 (100%). HRMS Calcd for $\text{C}_{12}\text{H}_{12}$ 156.0939. Found 156.0934.

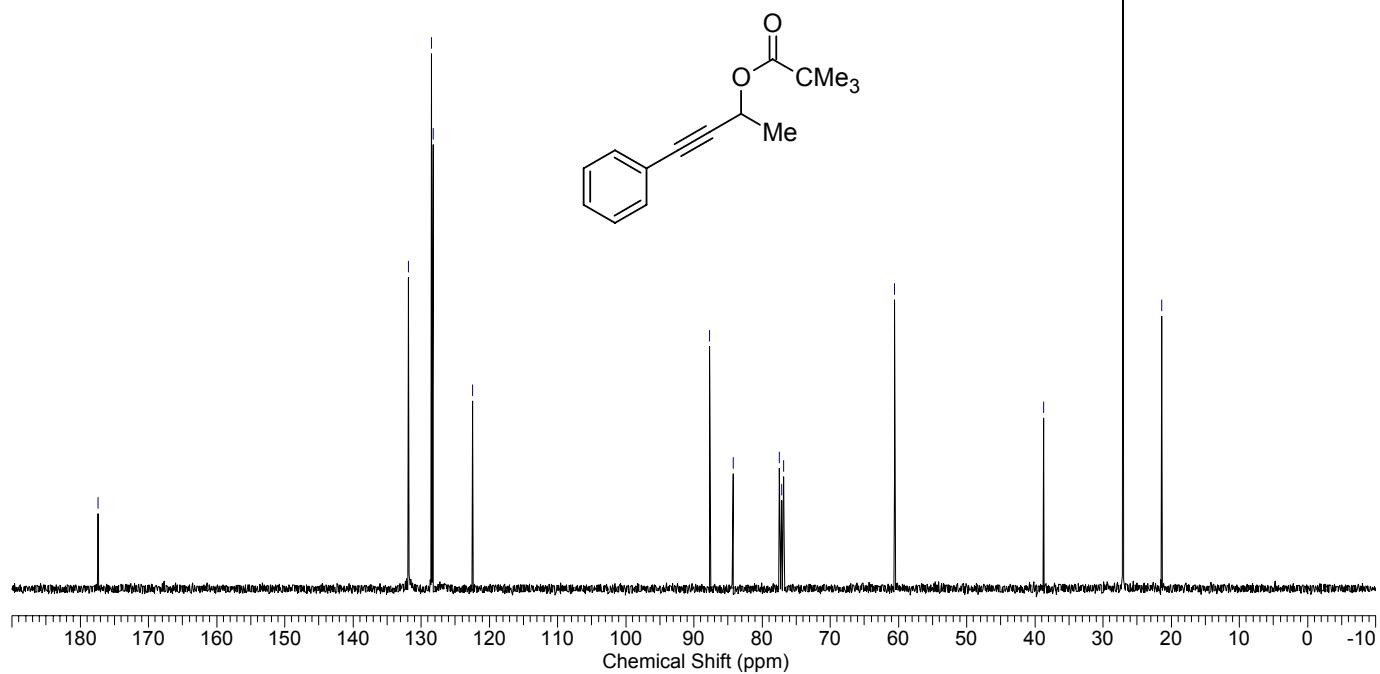
Procedure for isolation of acid-sensitive endiynes

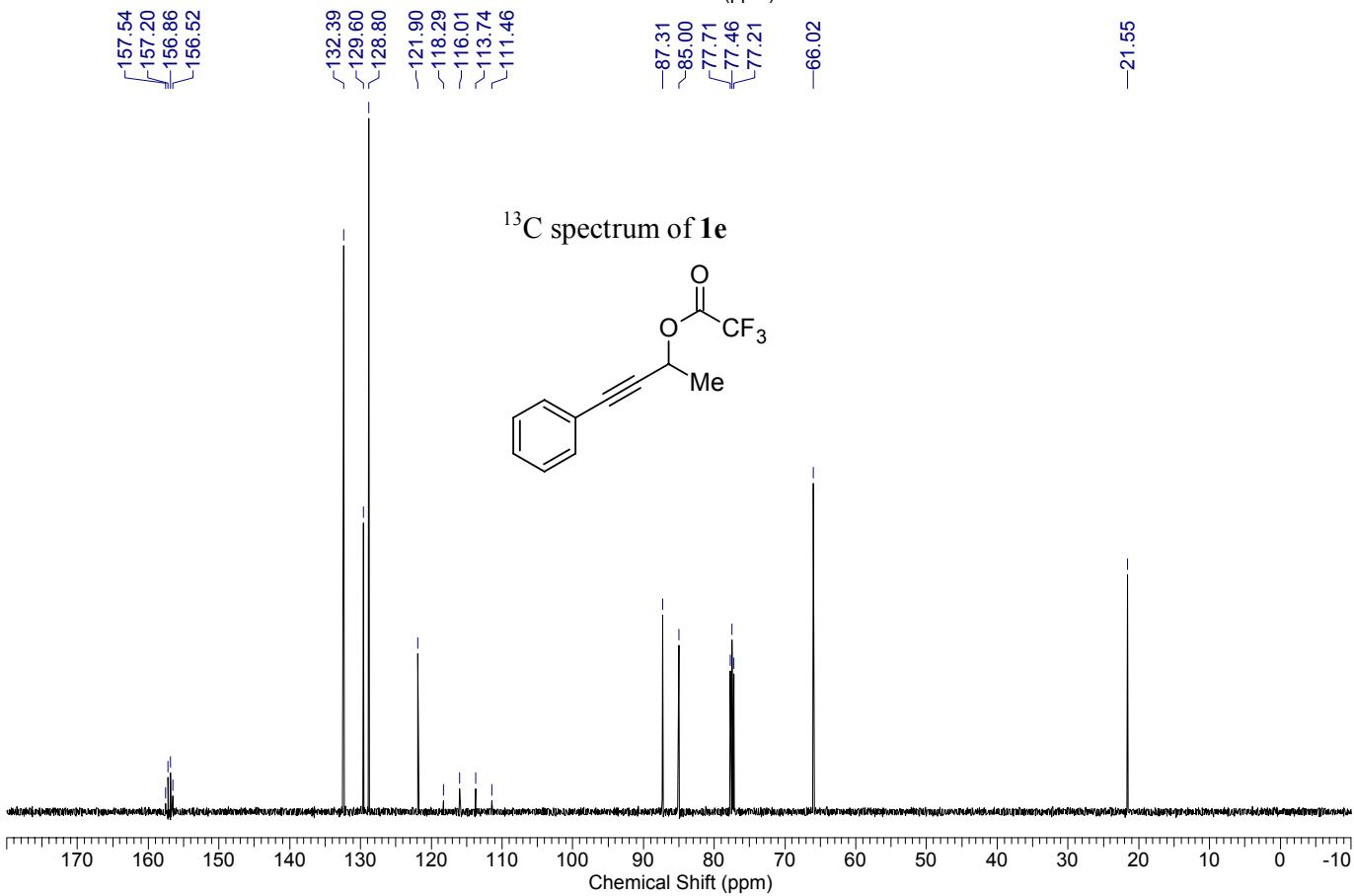
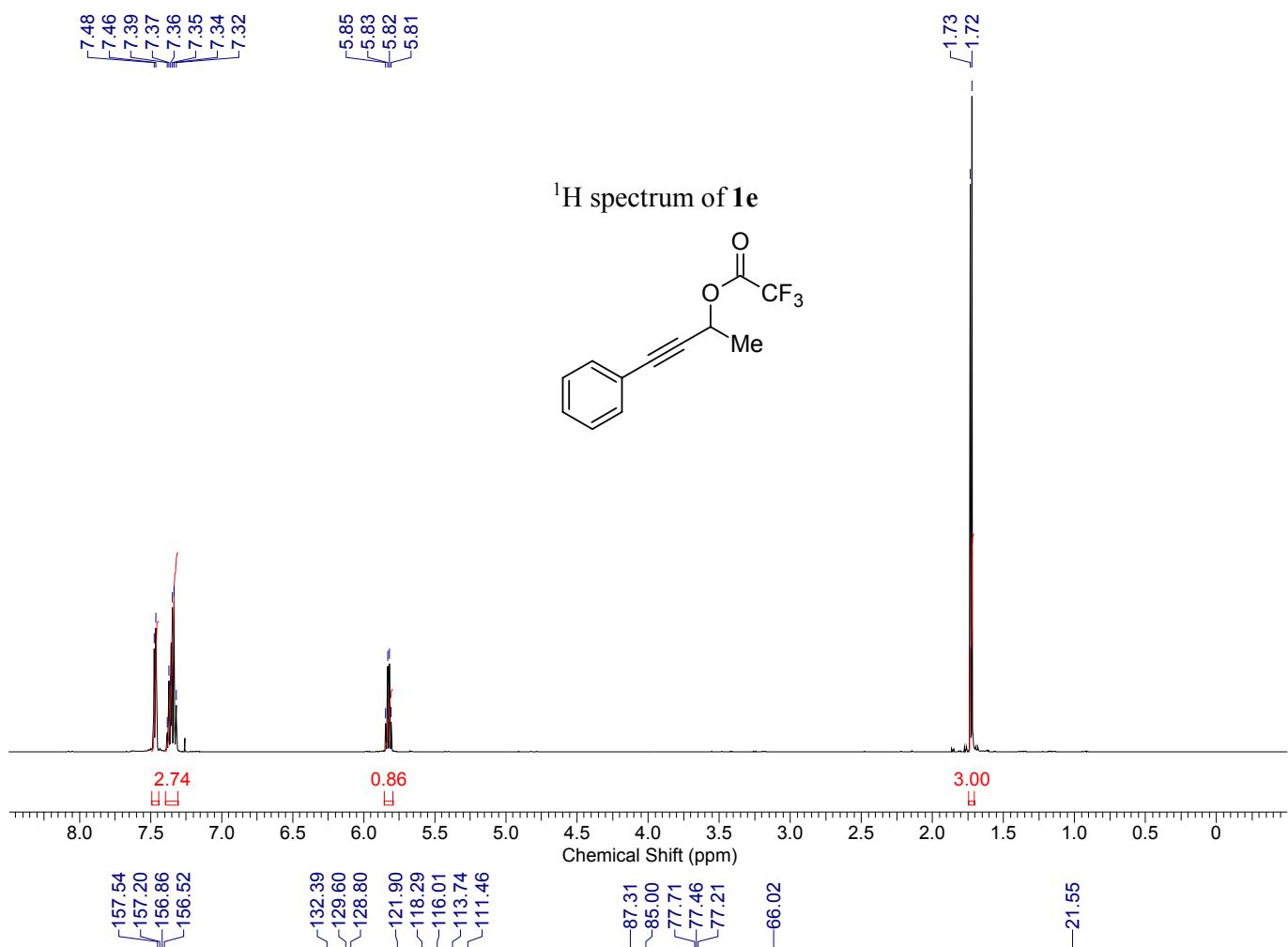
7a: Allylation was performed similarly as for **2a**. Reaction mixture was filtered through basic alumina and concentrated in vacuum. The crude oil was purified by preparative column chromatography using basic alumina (eluent pentane). Yield 198 mg (0.79 mmol, 79%). ^1H NMR (500.13 MHz, C_6D_6) δ 7.43-7.41 (m, 2H), 6.95-6.93 (m, 3H), 6.03 (ddt, $J = 17.2$ Hz, 10.3 Hz, 7.0 Hz, 1H), 5.12 (m, $J = 17.2$ Hz, 1H), 5.09 (m, $J = 10.3$ Hz, 1H), 3.62 (tt, $J = 6.6$ Hz, 2.2 Hz, 1H), 2.52 (app. t, $J = 7.0$ Hz, 2H), 2.04 (td, $J = 7.3$ Hz, 2.2 Hz, 2H), 1.37 (app. quintet, $J = 7.7$ Hz, 2H), 1.28-1.22 (m, 2H), 1.16 (app. sextet, $J = 7.3$ -7.7 Hz, 2H), 0.81 (t, $J = 7.3$ Hz, 3H); ^{13}C NMR (125.76 MHz, C_6D_6) δ 135.1 (+), 132.0 (+, 2C), 128.4 (+, 2C), 128.1 (+), 123.9, 117.6 (-), 88.8, 82.2, 81.9, 78.3, 40.9 (-), 31.3 (-), 28.8 (-), 24.8 (+), 22.5 (-), 19.0 (-), 14.1 (+); GC/MS m/z 244 (M^+ , 2%), 203 ($\text{M}^+ - \text{allyl}$, 2%), 105 (65%), 91 (100%). HRMS Calcd for $\text{C}_{19}\text{H}_{21}$ 249.1643. Found 249.1628.

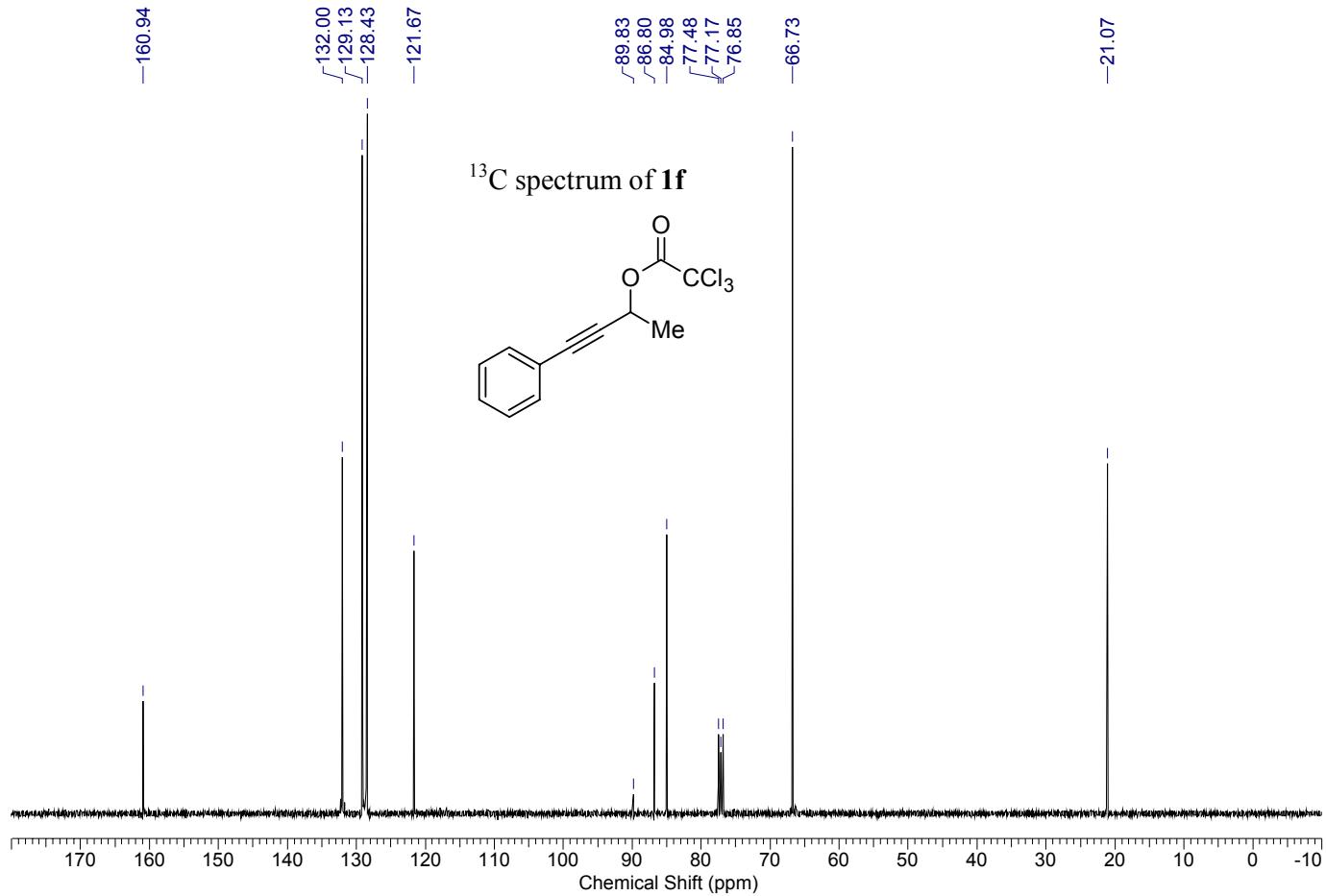
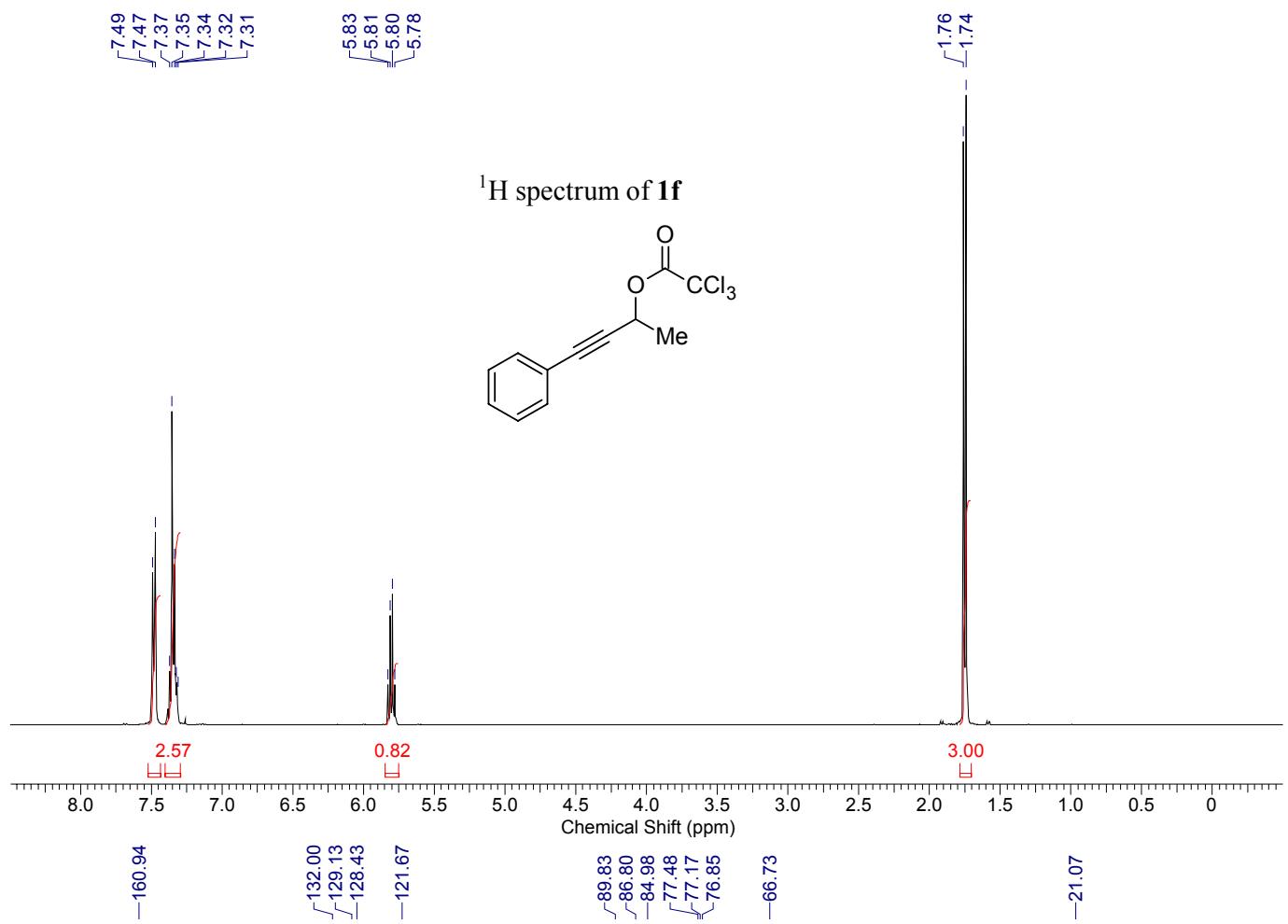
7b: ^1H NMR (500.13 MHz, C_6D_6) δ 6.05 (ddtd, $J = 17.2$ Hz, 10.3 Hz, 7.0 Hz, 1.5 Hz, 1H), 5.11 (m, $J = 17.2$ Hz, 1H), 5.09 (m, $J = 10.3$ Hz, 1H), 3.48 (triplet of quintets, $J = 8.9$ Hz, 2.2 Hz, 1H), 2.52-2.48 (m, 2H), 2.04 (td, $J = 7.0$ Hz, 2.2 Hz, 4H), 1.37 (app. quintet, $J = 7.0$ Hz, 4H), 1.28-1.22 (m, 4H), 1.16 (sextet, $J = 7.3$ Hz, 4H), 0.81 (t, $J = 7.2$ Hz, 6H); ^{13}C NMR (125.76 MHz, C_6D_6) δ 135.5 (+), 117.2 (-), 81.5 (2C), 79.2 (2C), 41.3 (-), 31.3 (-, 2C), 28.8 (-, 2C), 24.3 (+), 22.5 (-, 2C), 19.0 (-, 2C), 14.1 (+, 2C); GC/MS m/z 250 (M^+ , 4%), 209 ($\text{M}^+ - \text{allyl}$, 49%), 165 (100%), 152 (91%).

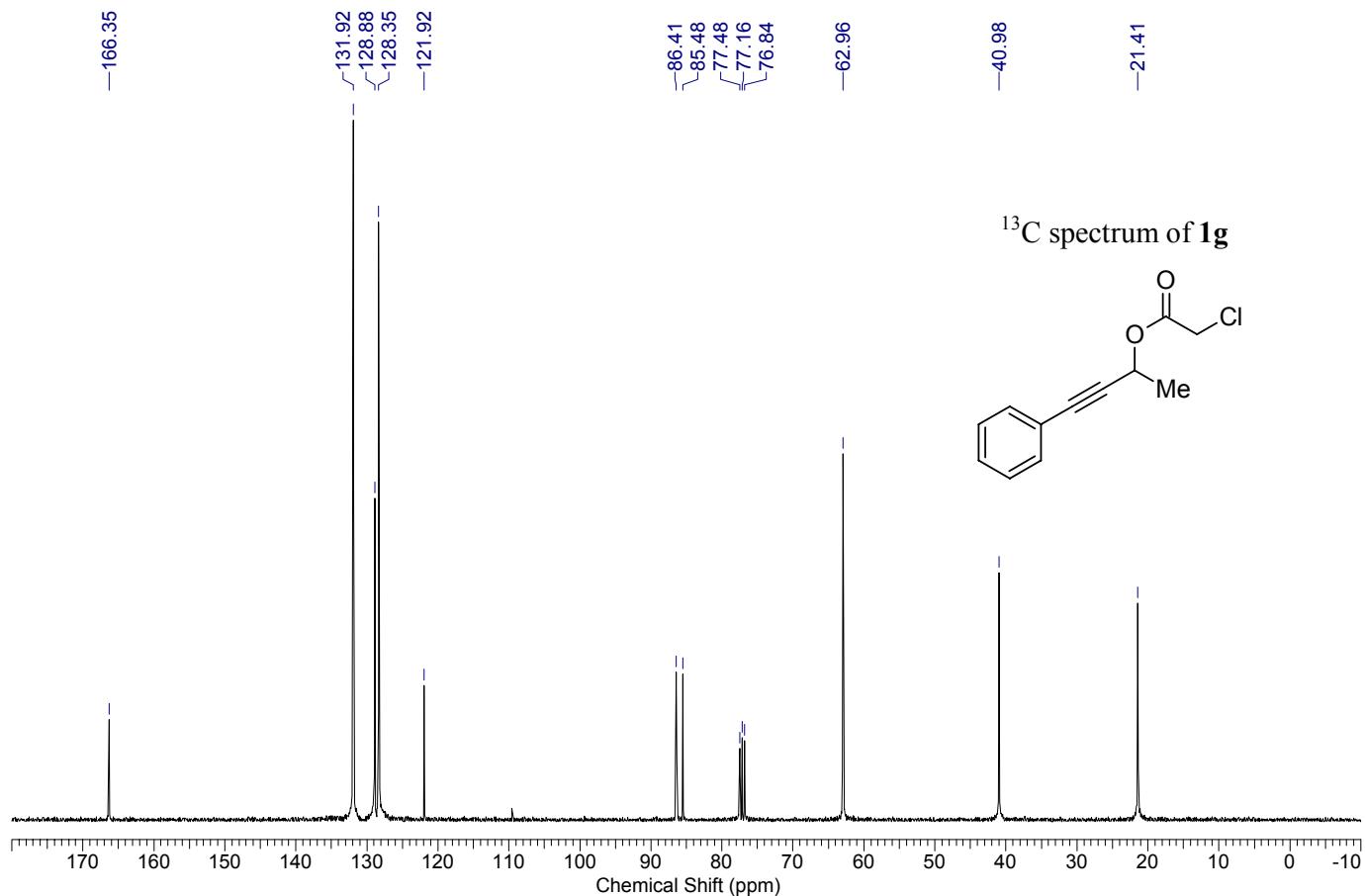
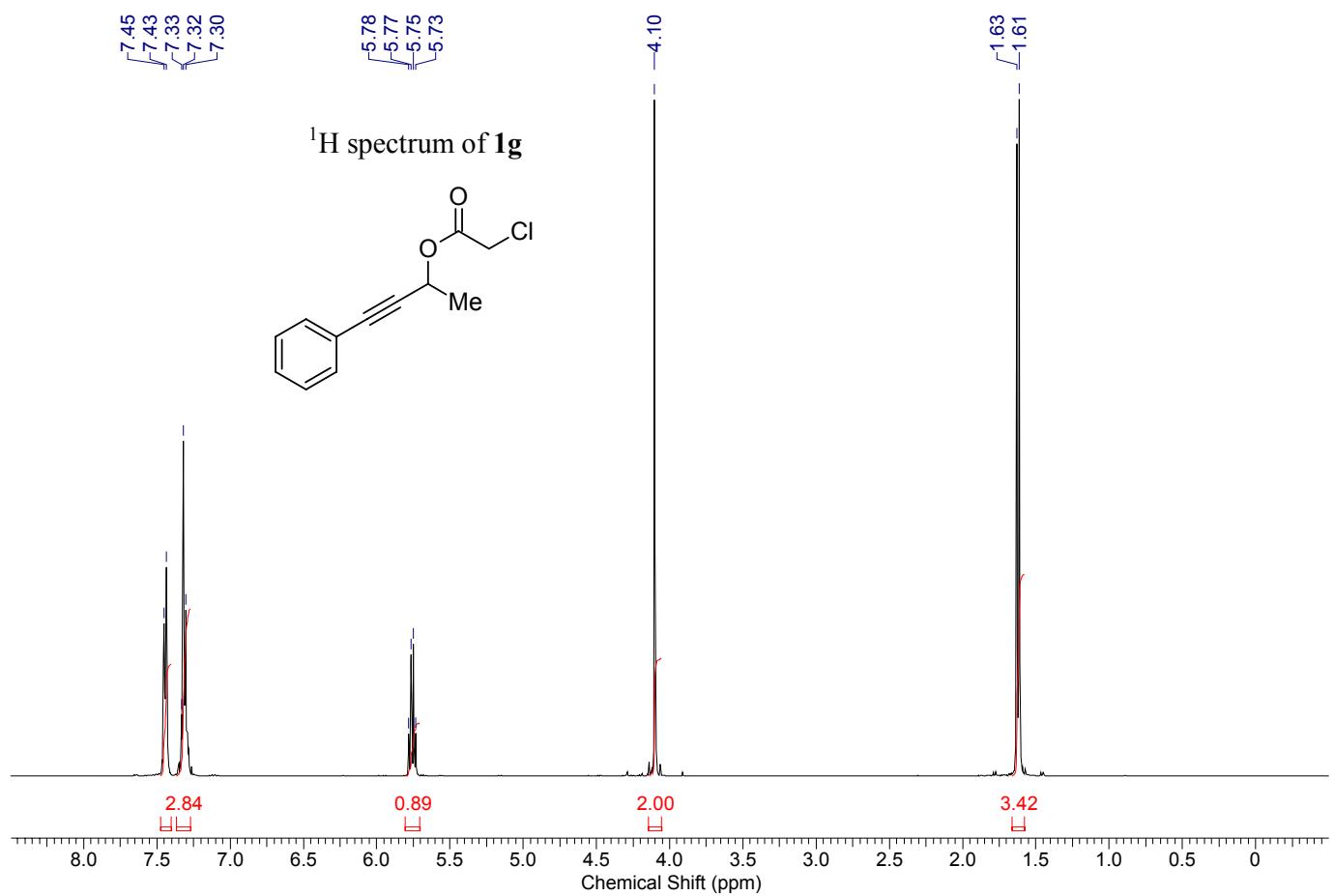


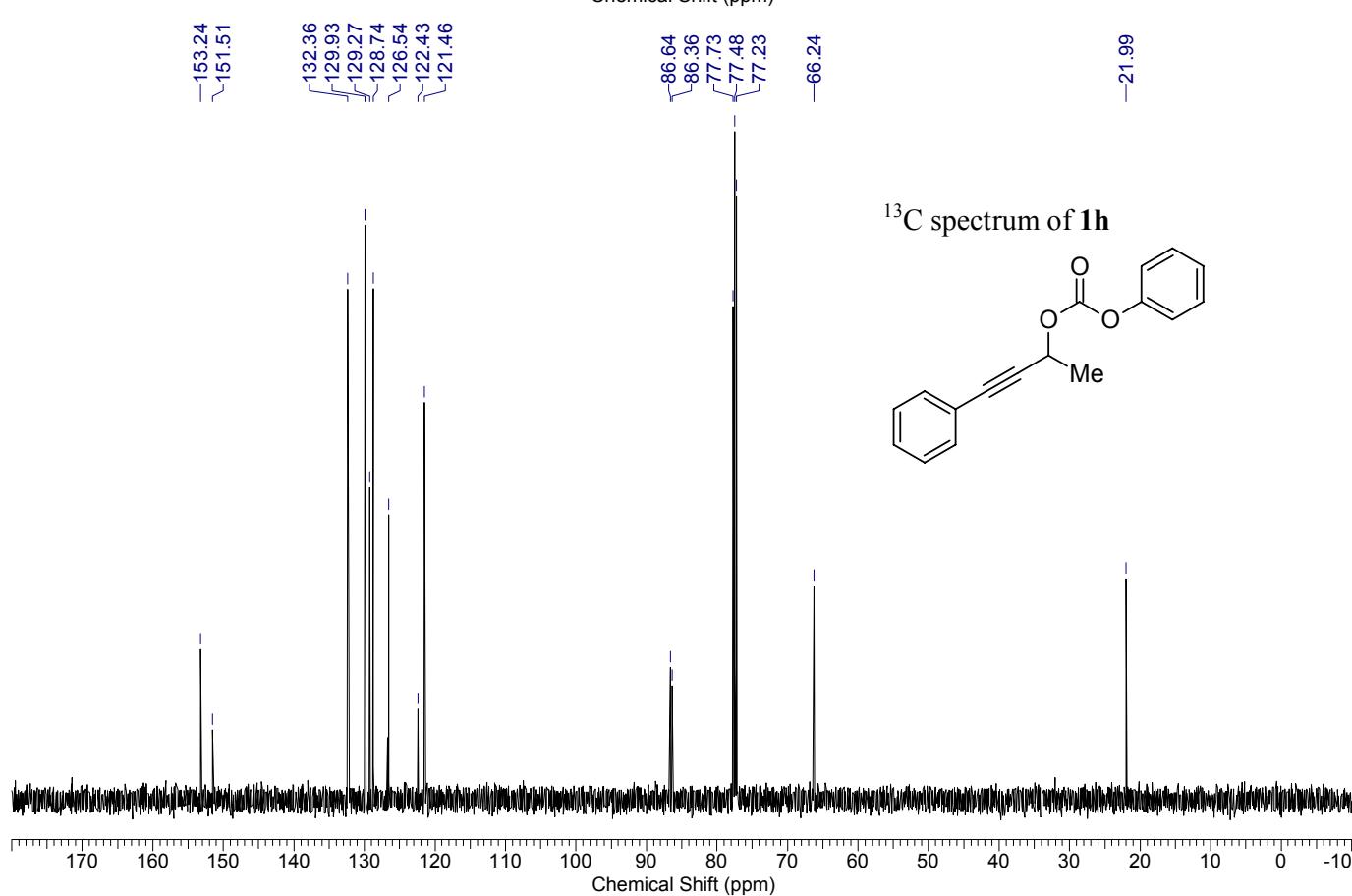
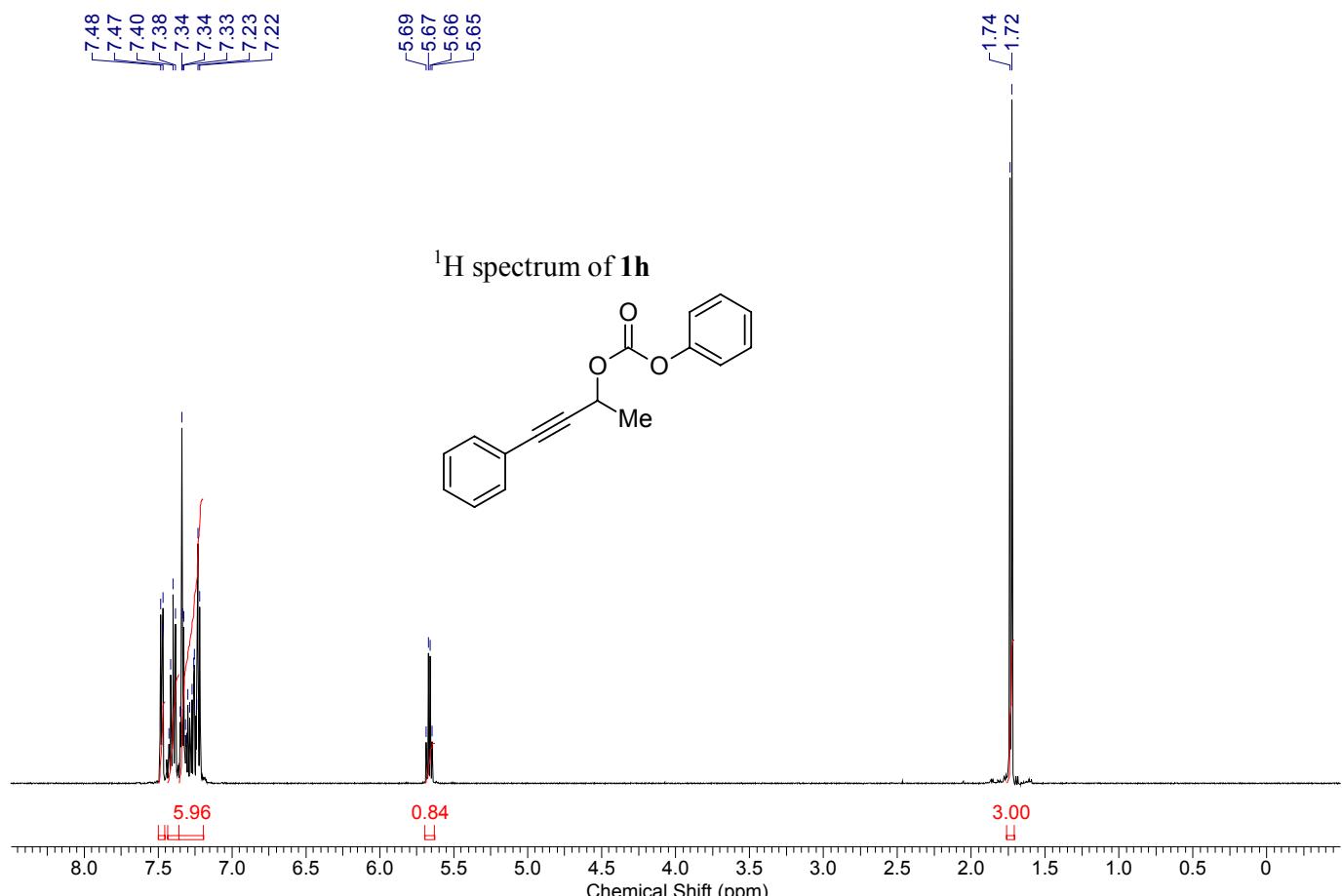
—177.35
—131.83
—128.51
—128.25
~—122.45
—87.69
—84.25
—77.44
—77.11
—76.81
—60.57
—38.68
—27.05
—21.35

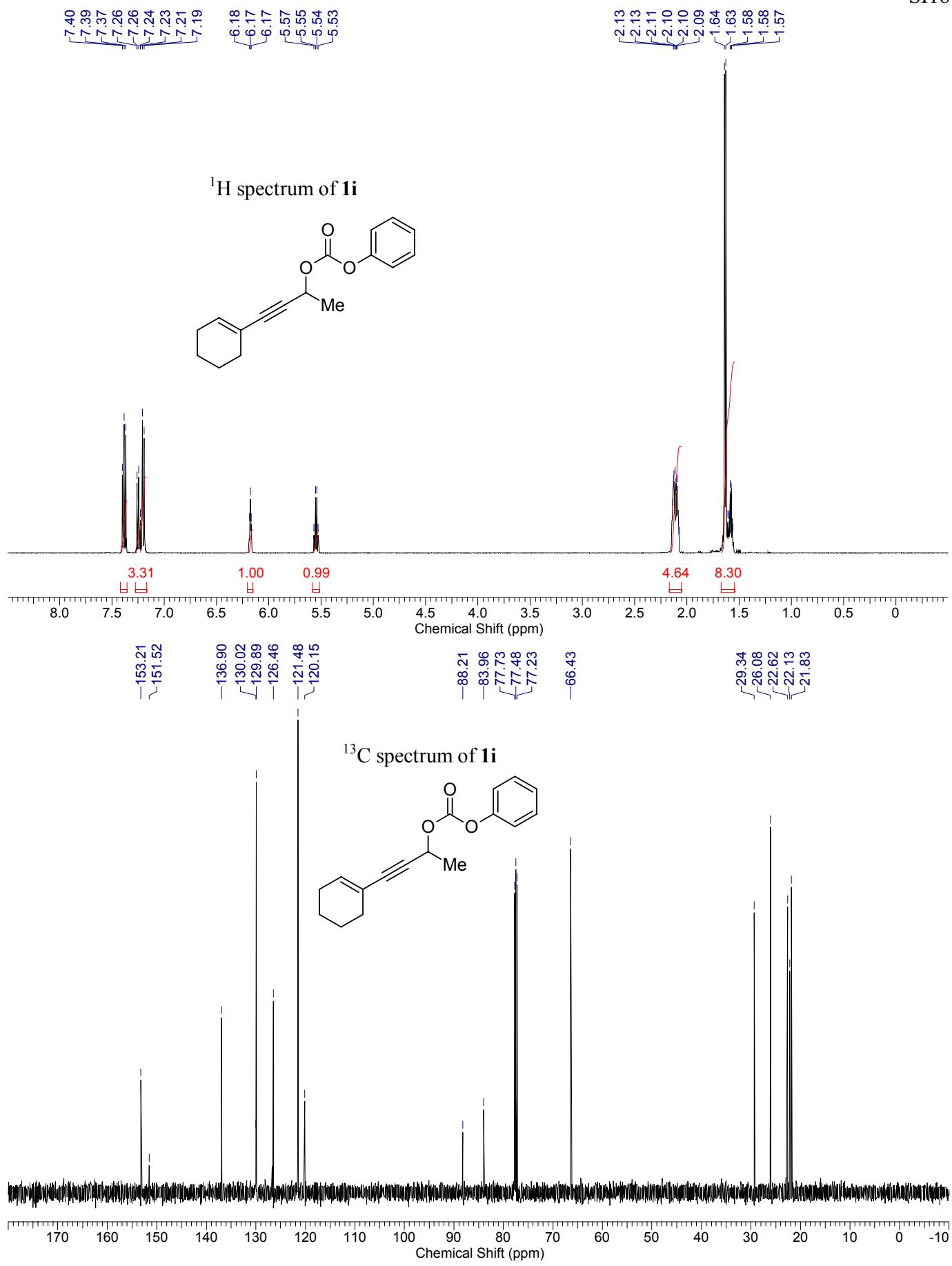
¹³C spectrum of **1d**

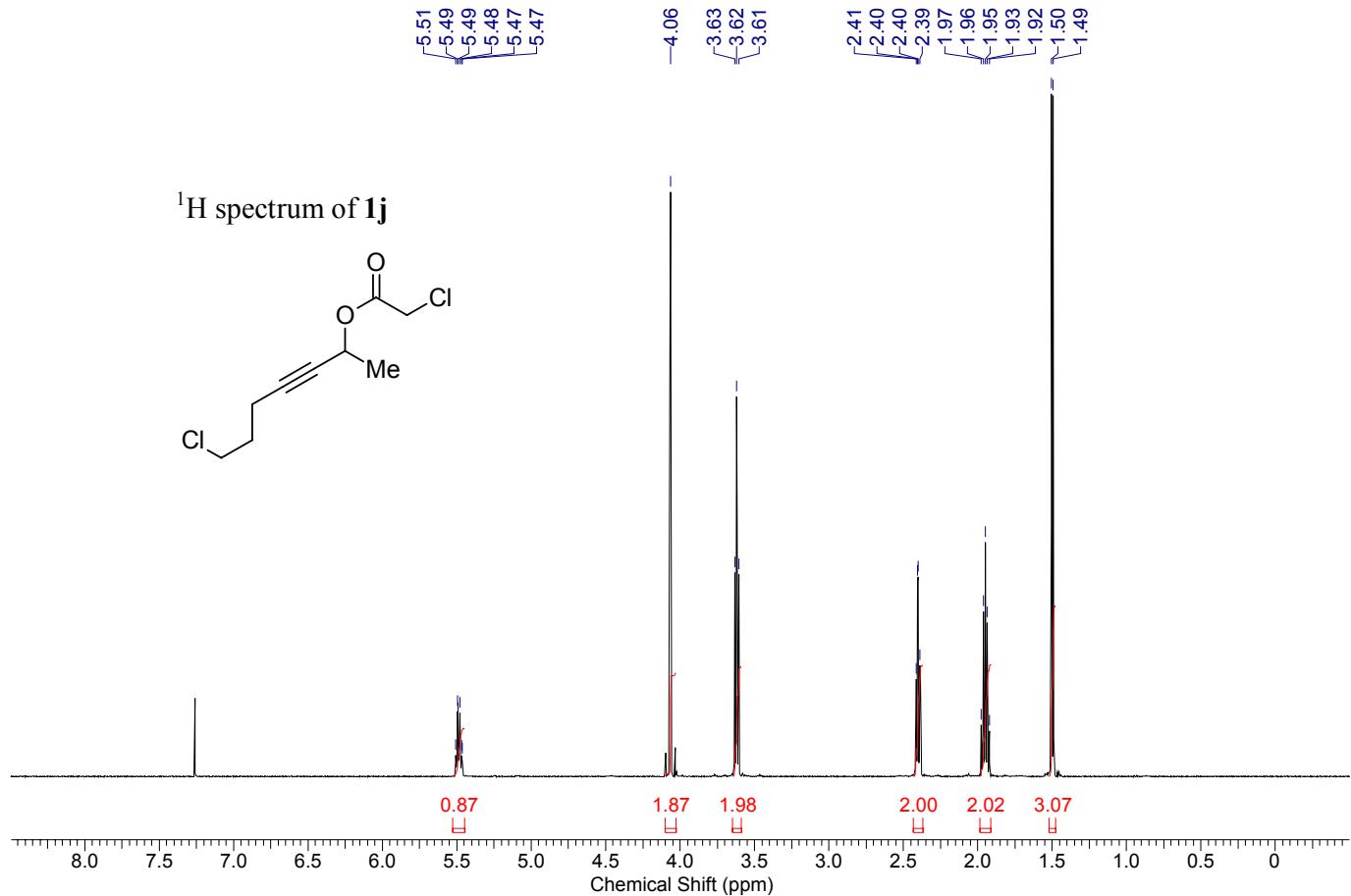
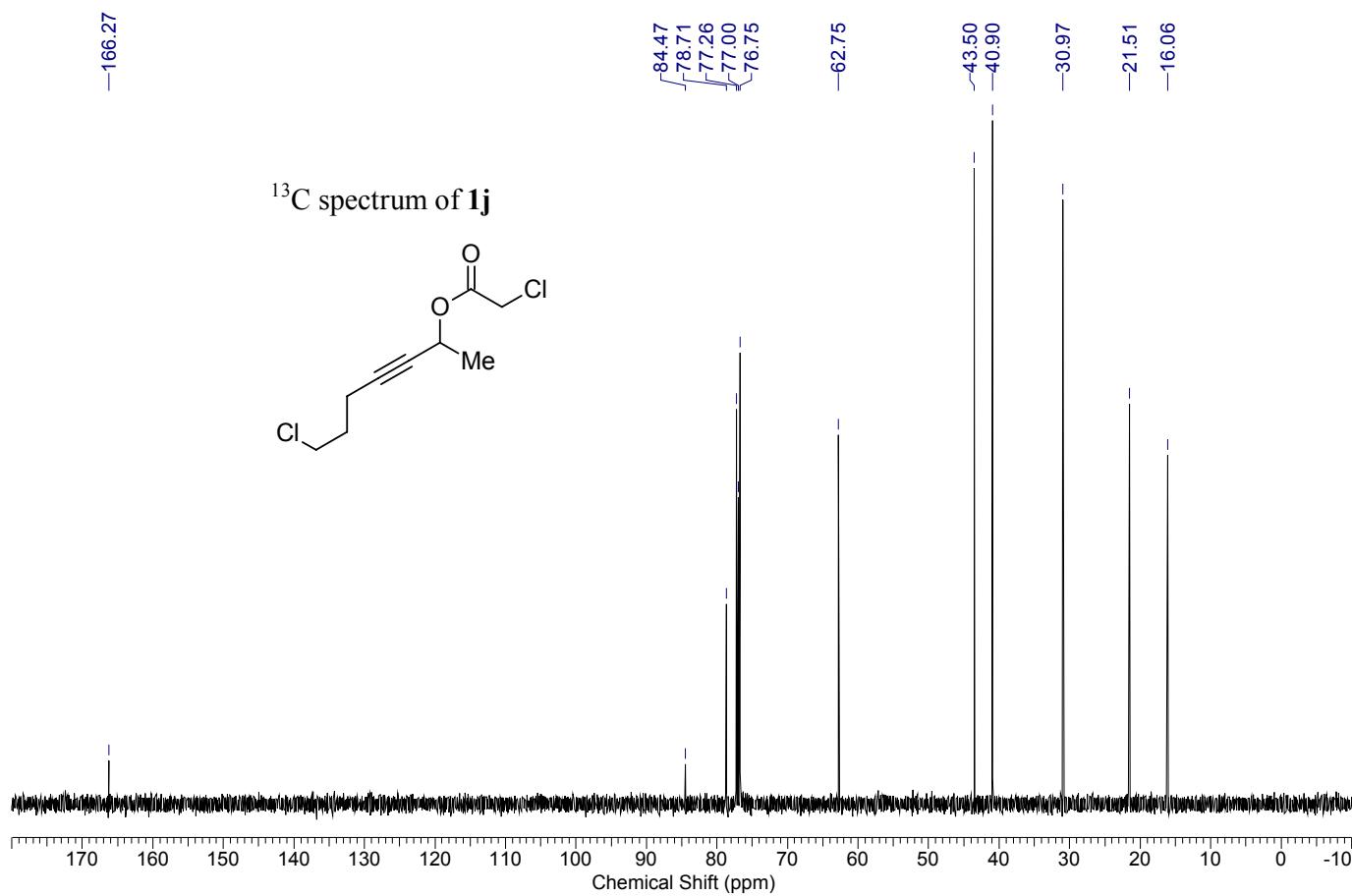


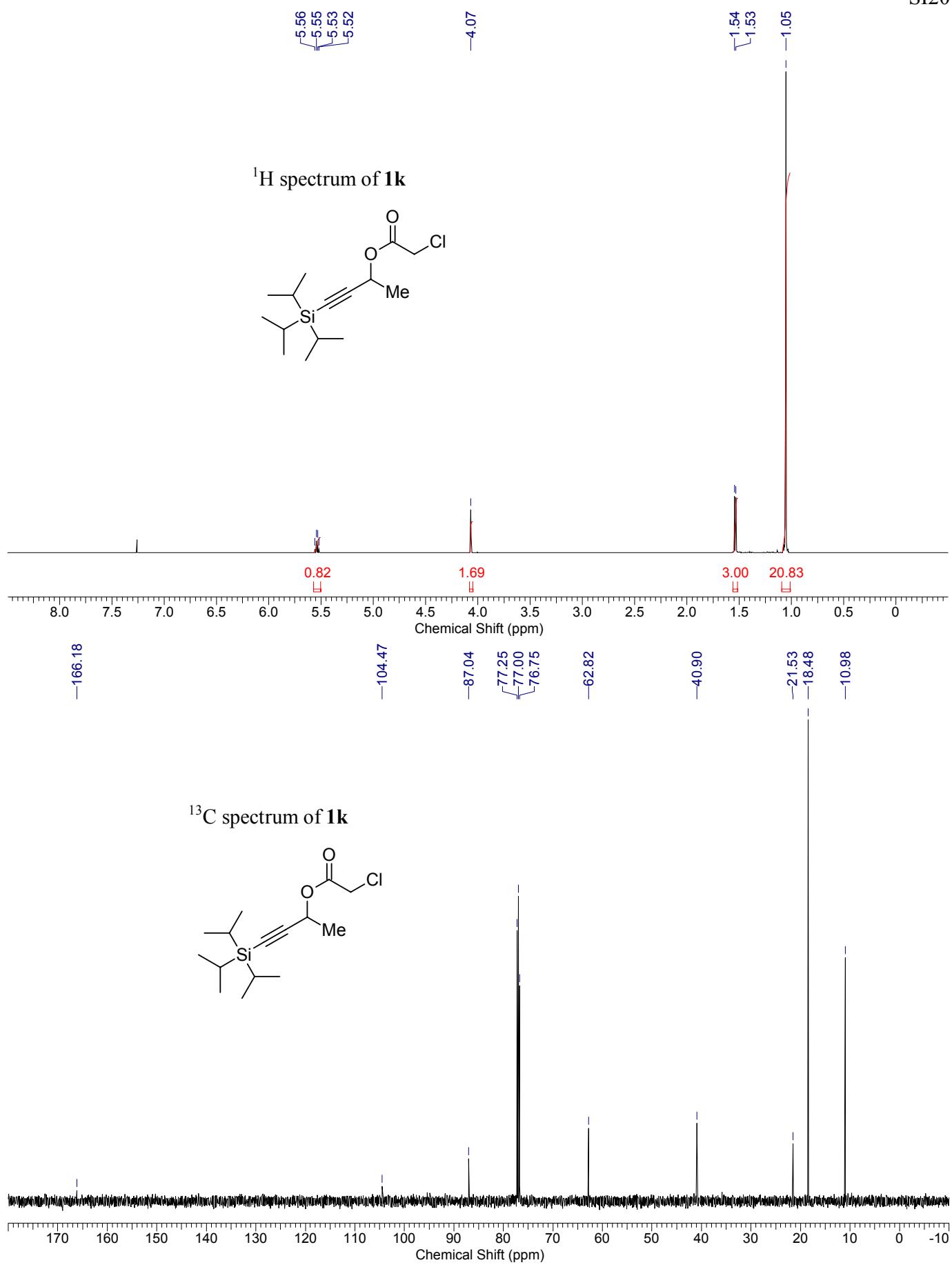


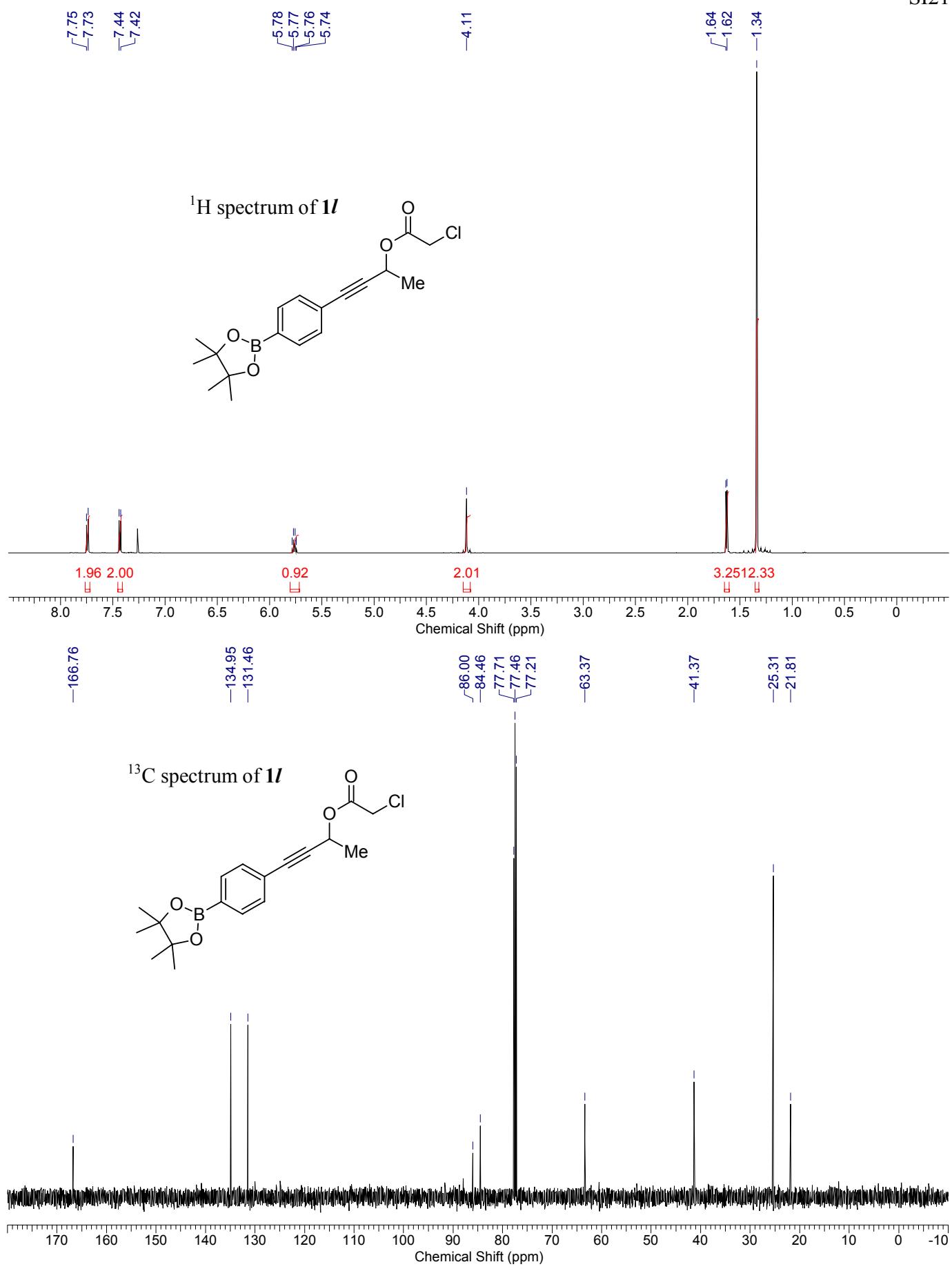


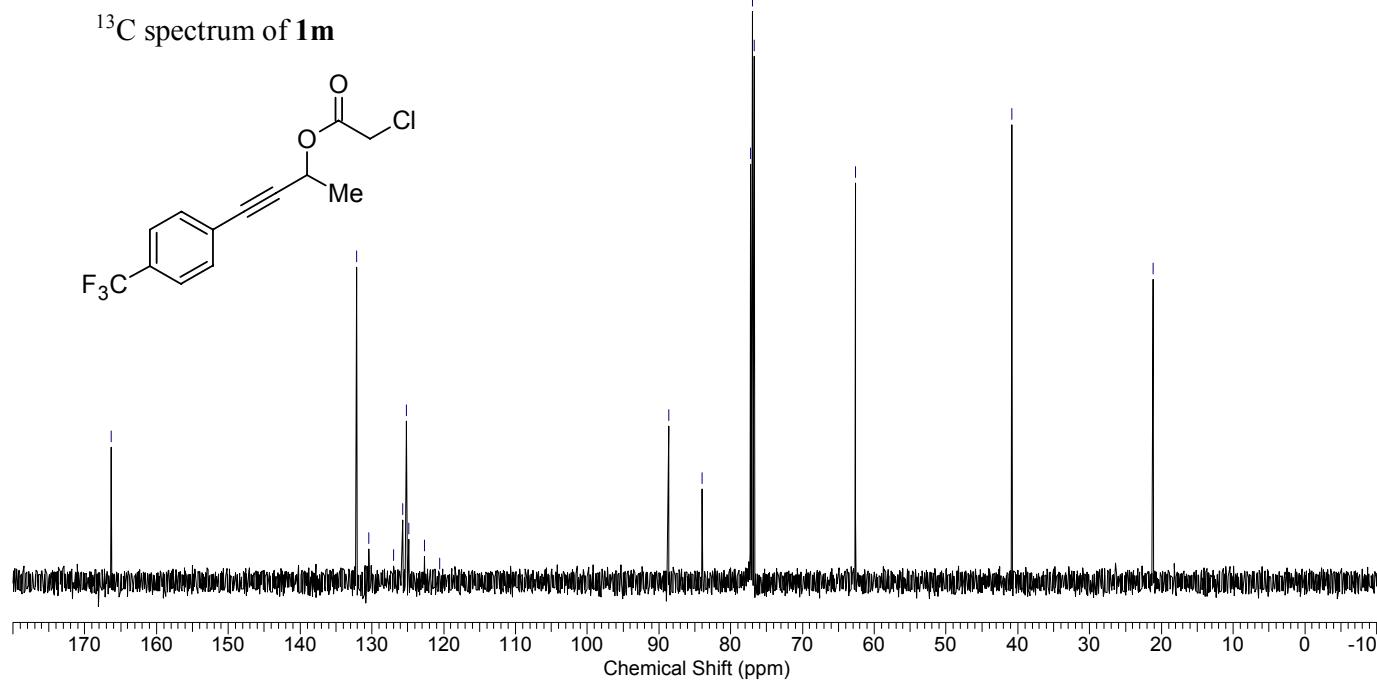
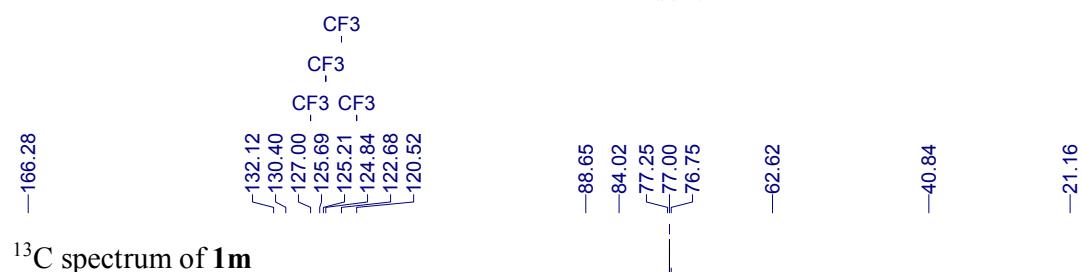
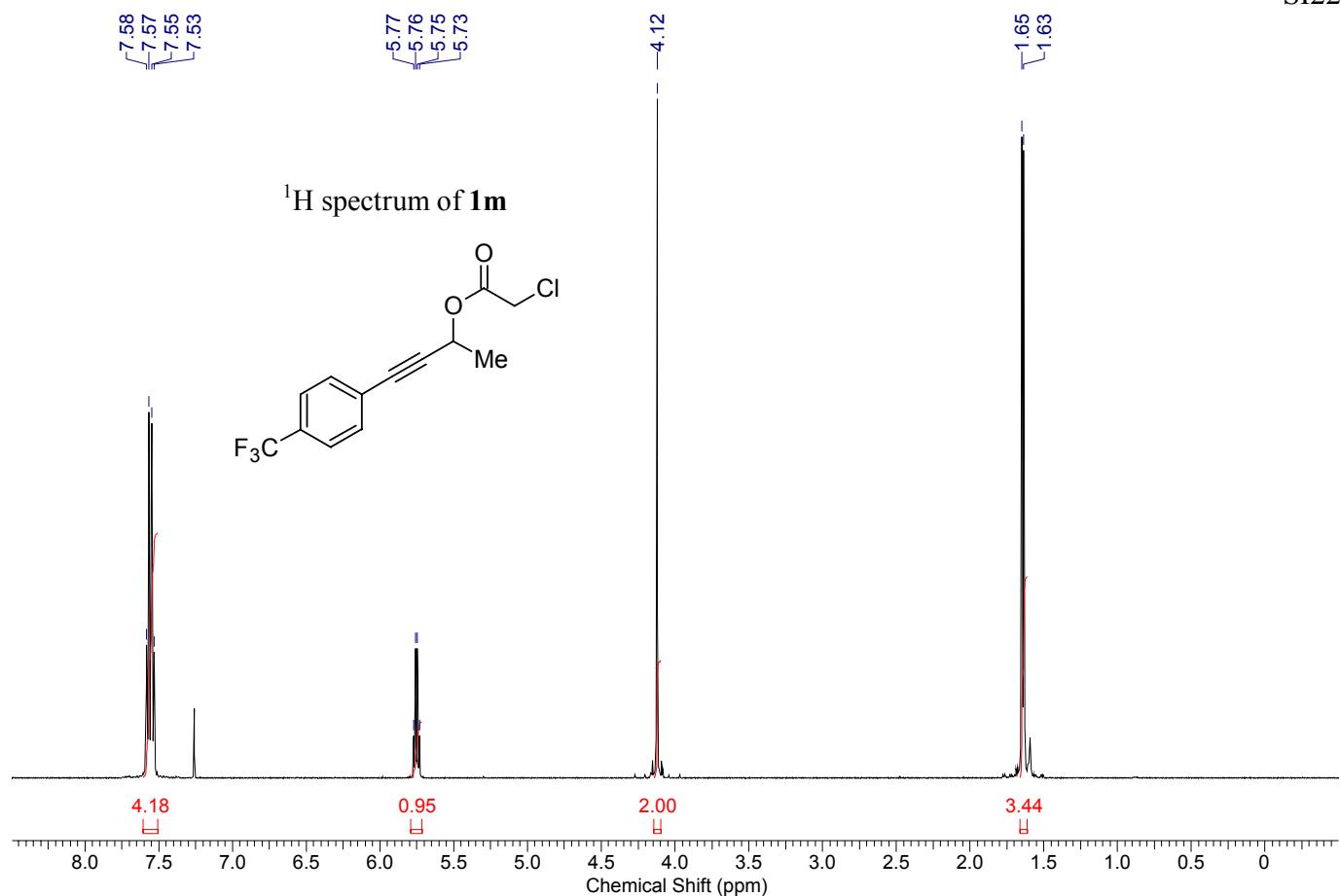


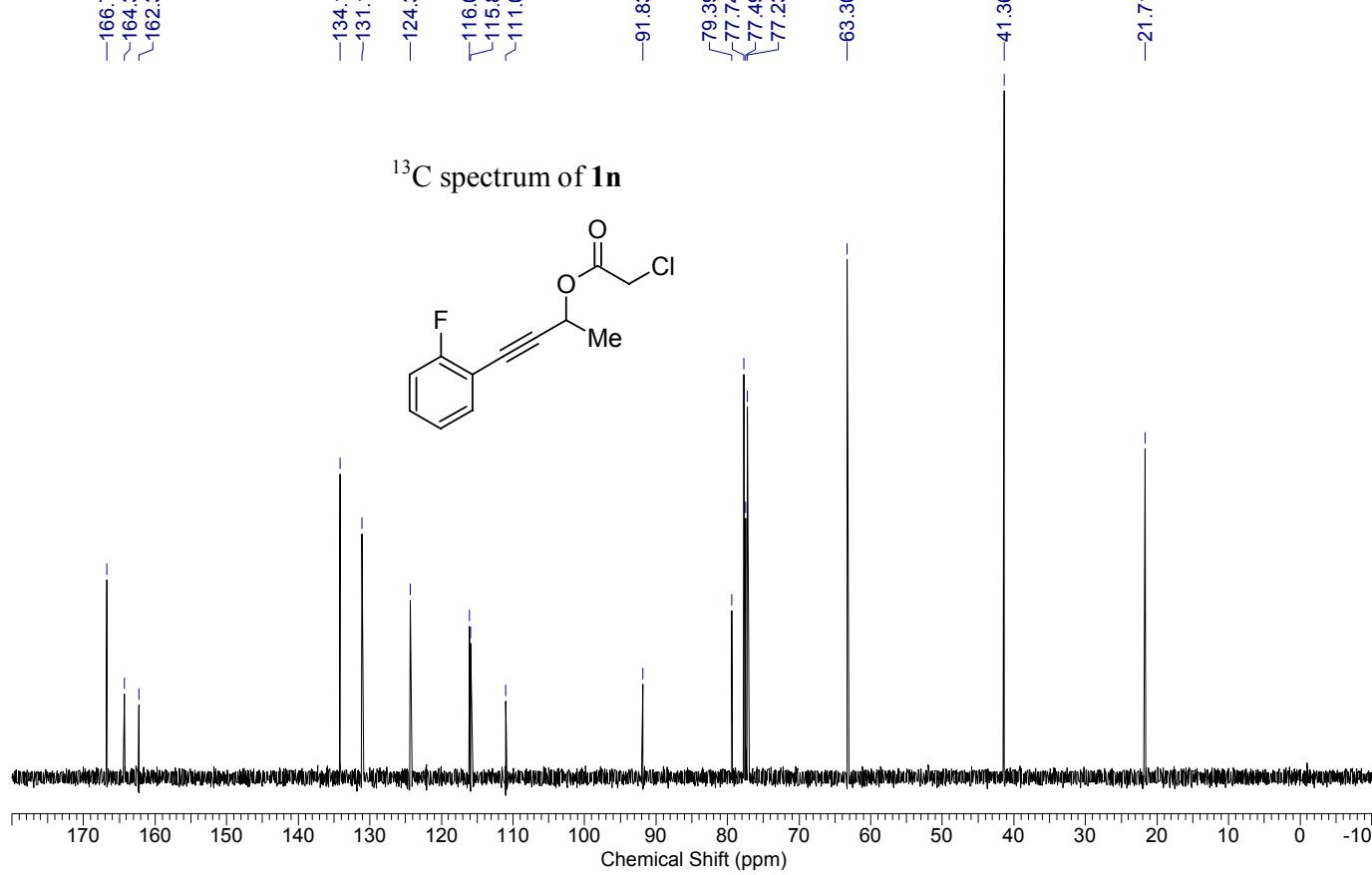
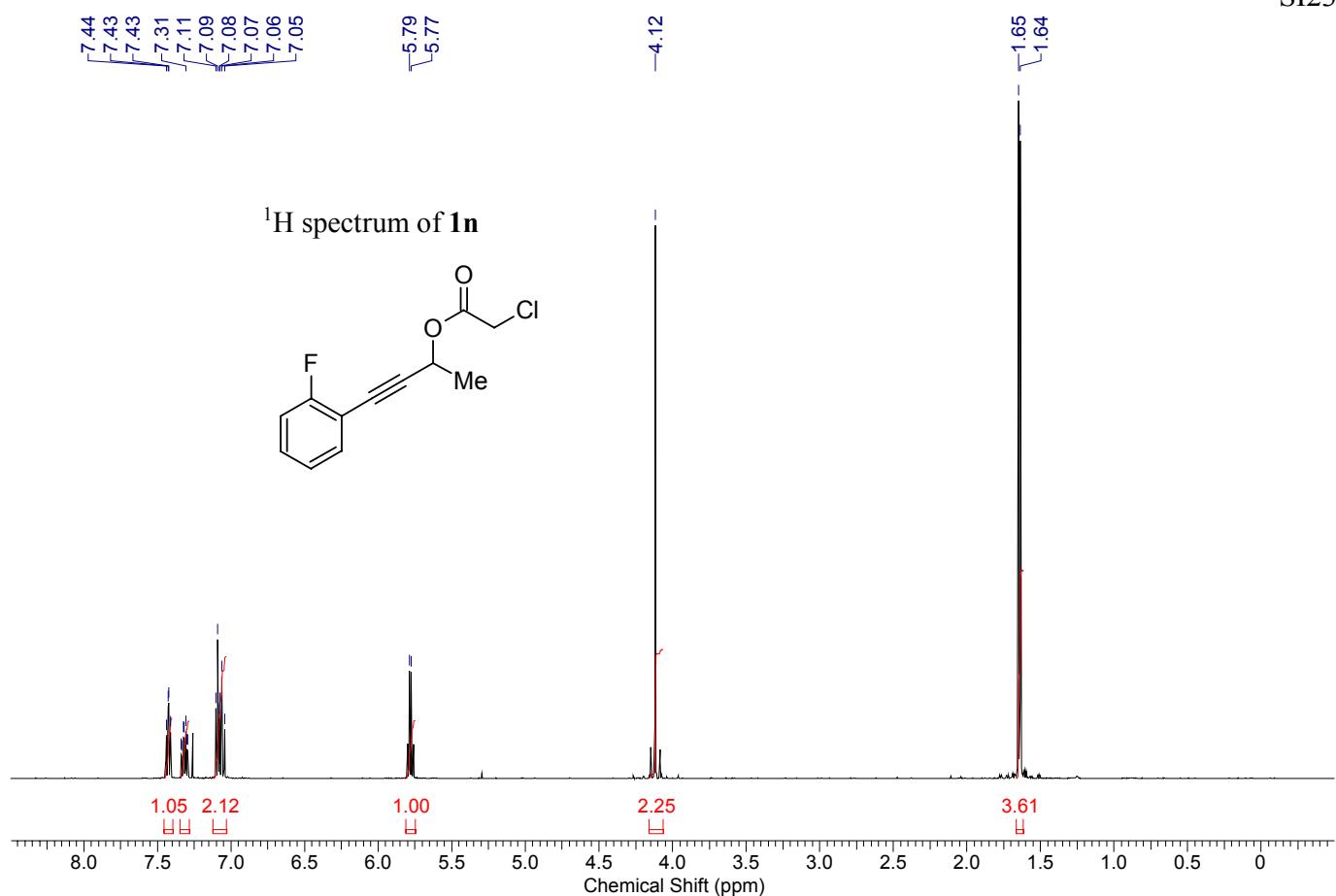


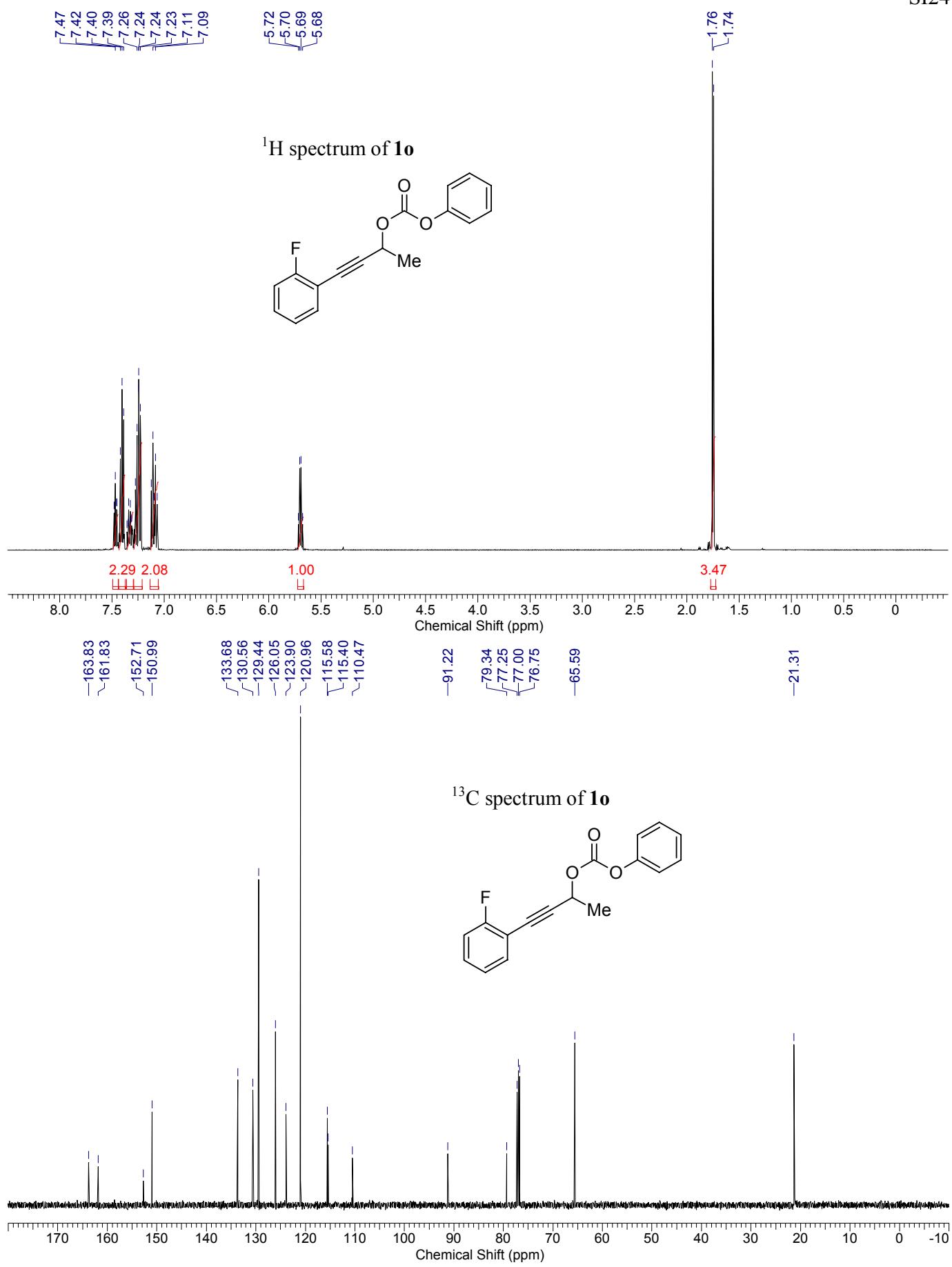
¹H spectrum of **1j**¹³C spectrum of **1j**

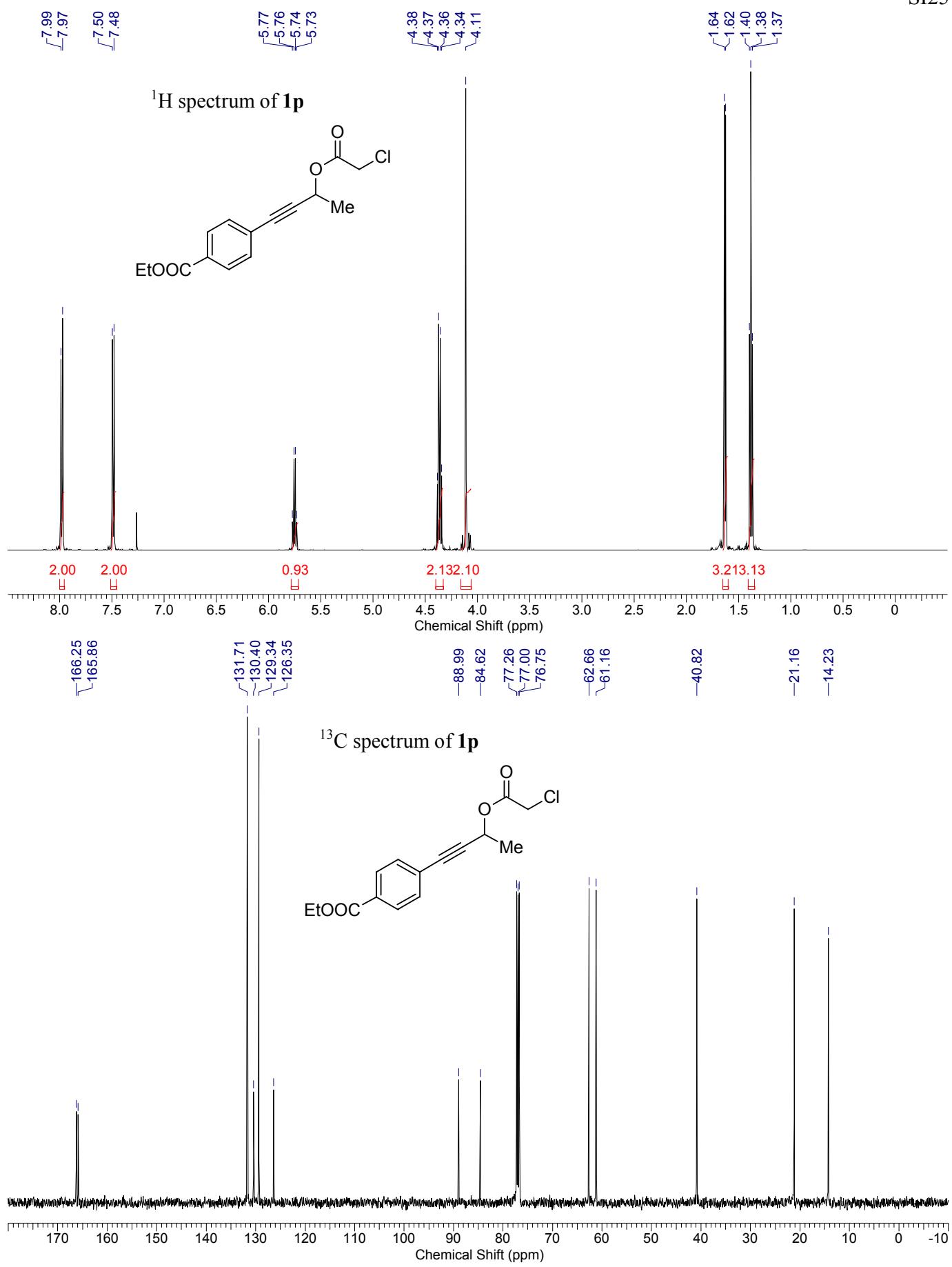


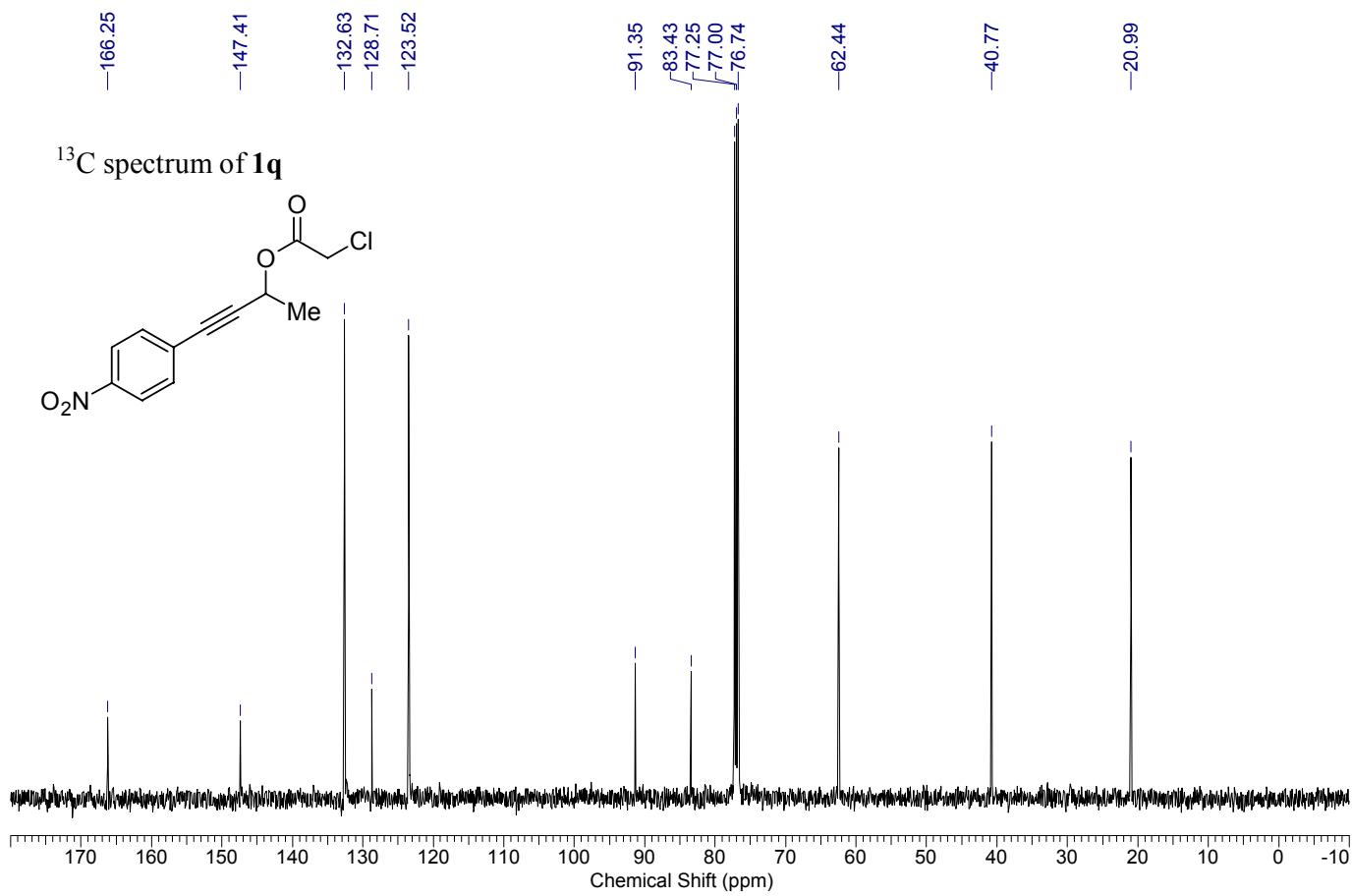
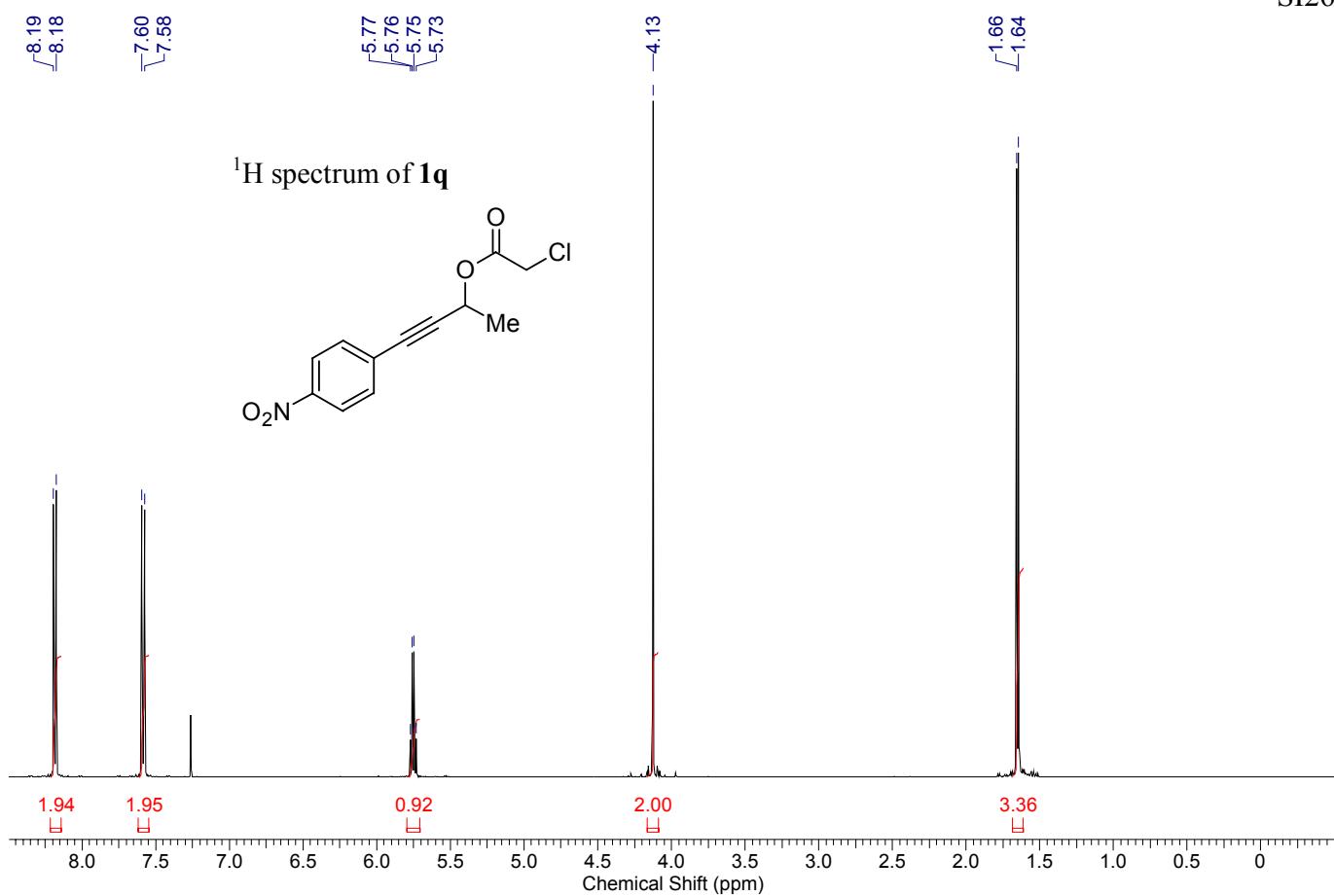


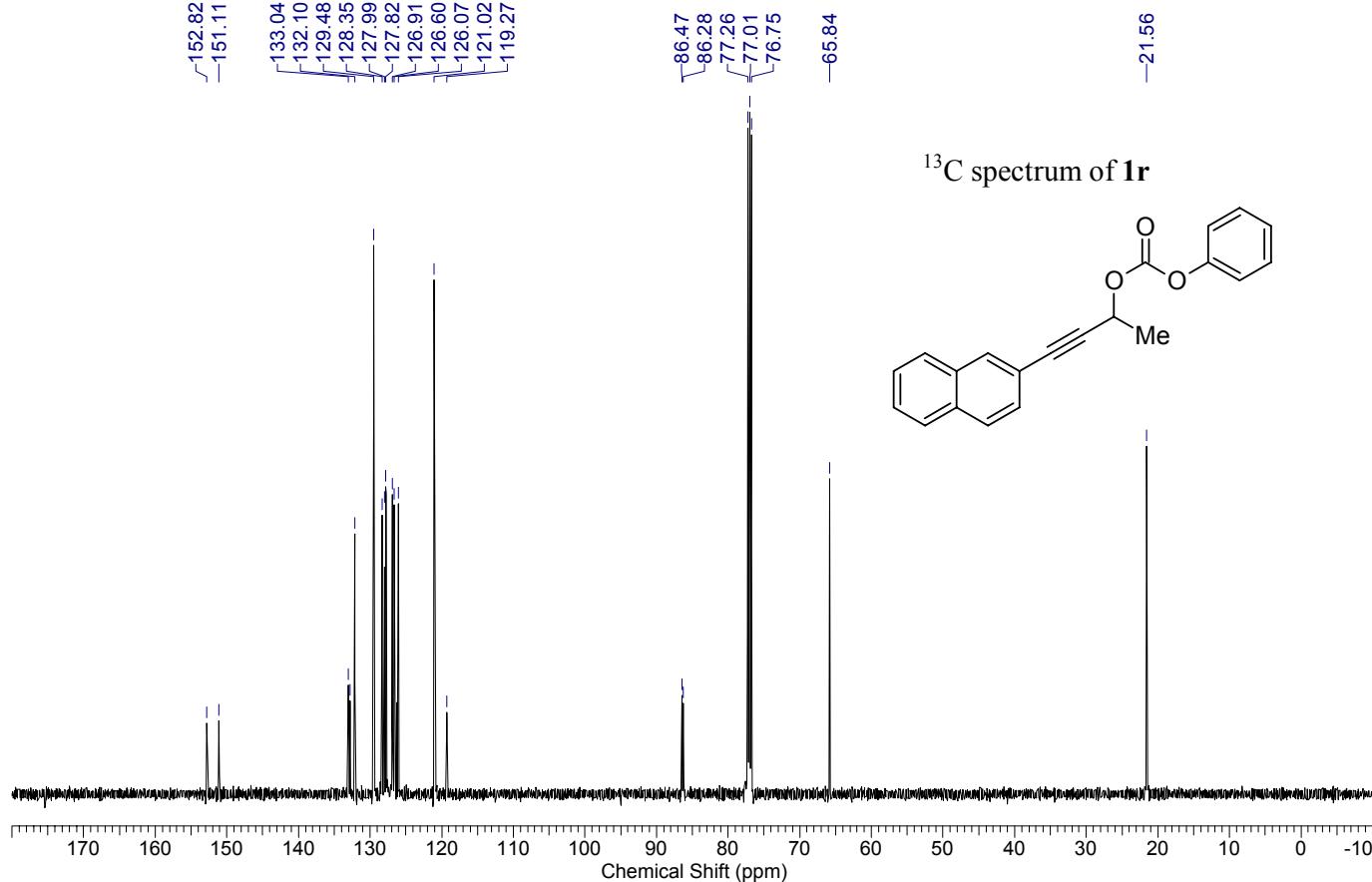
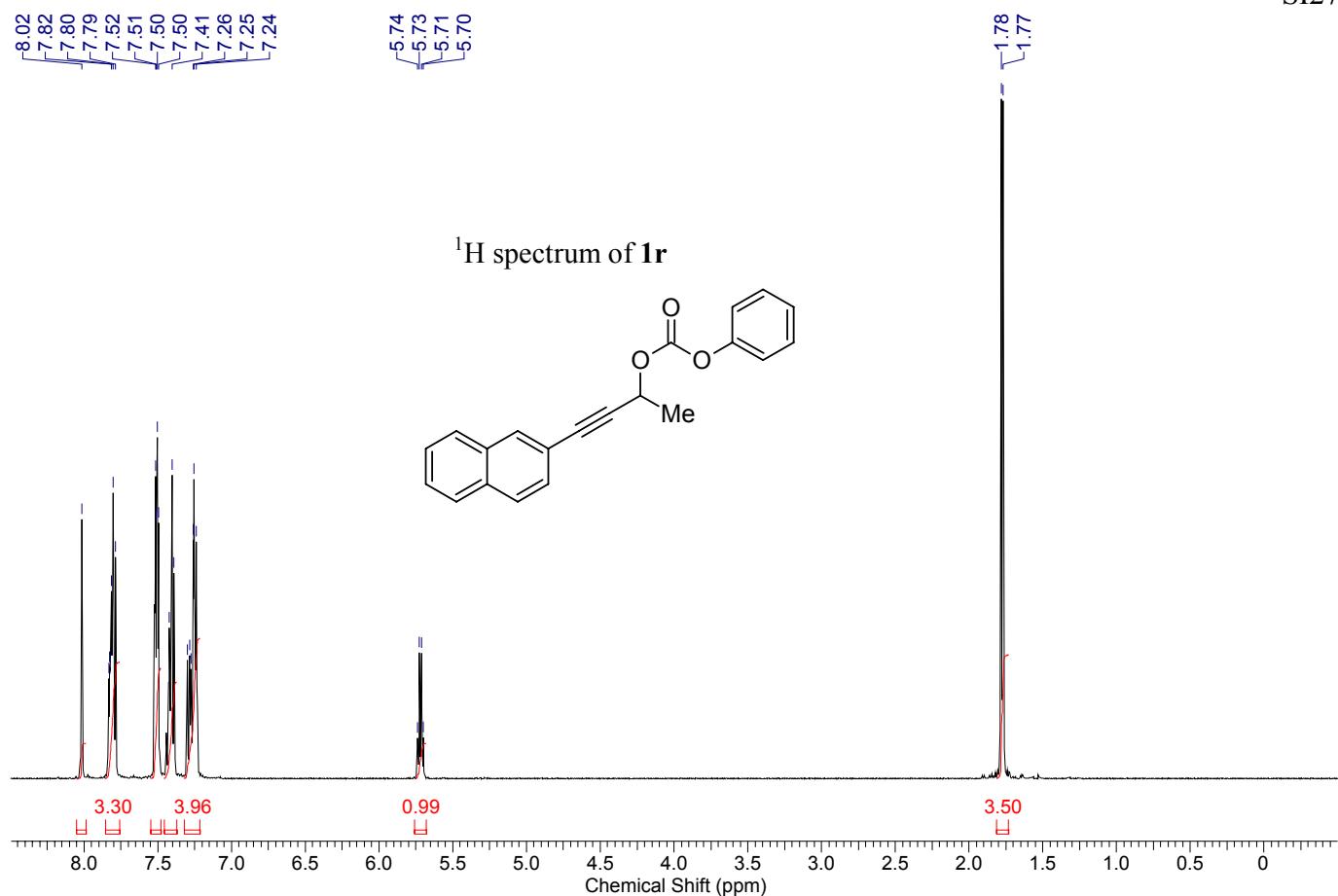


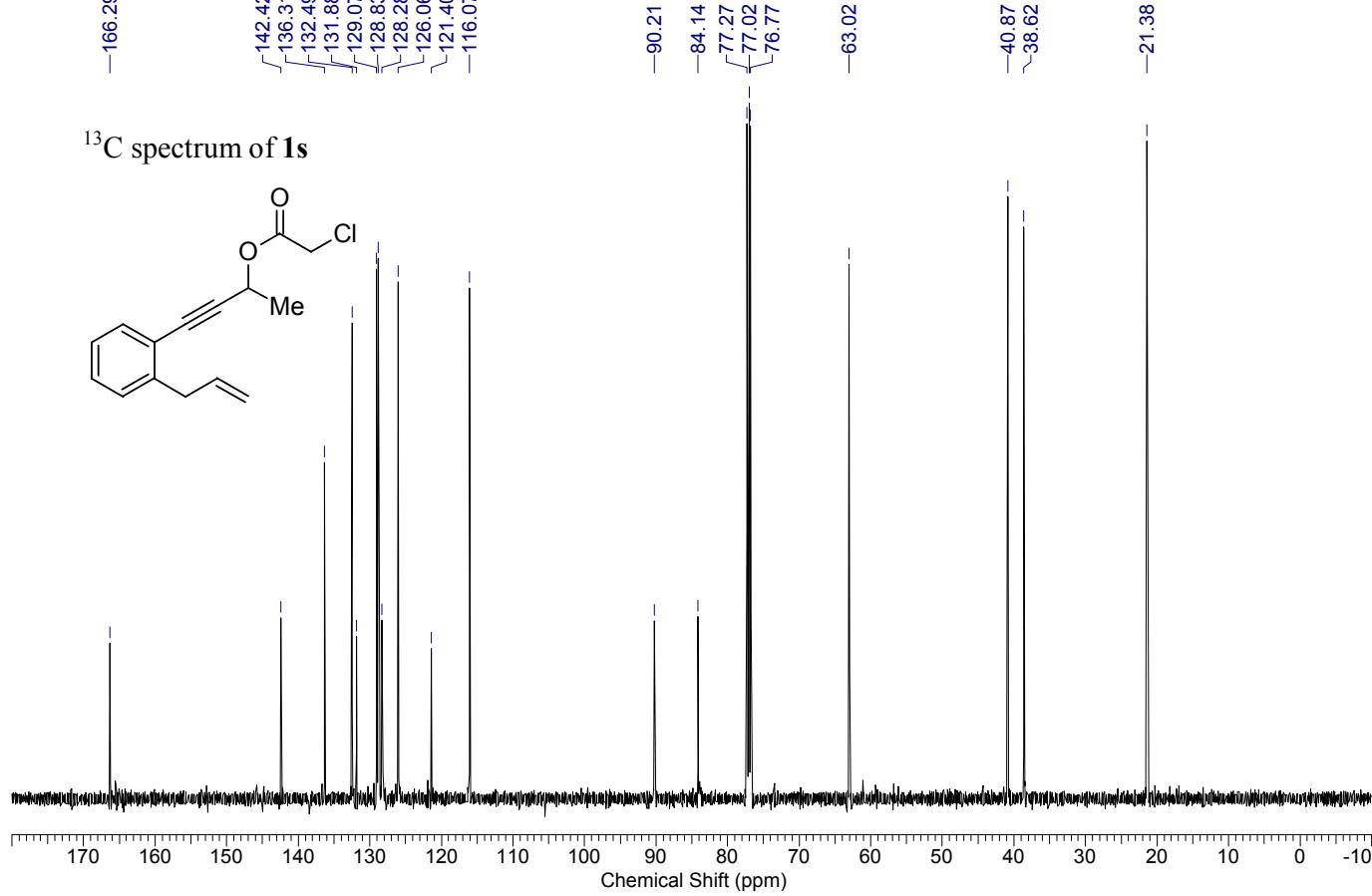
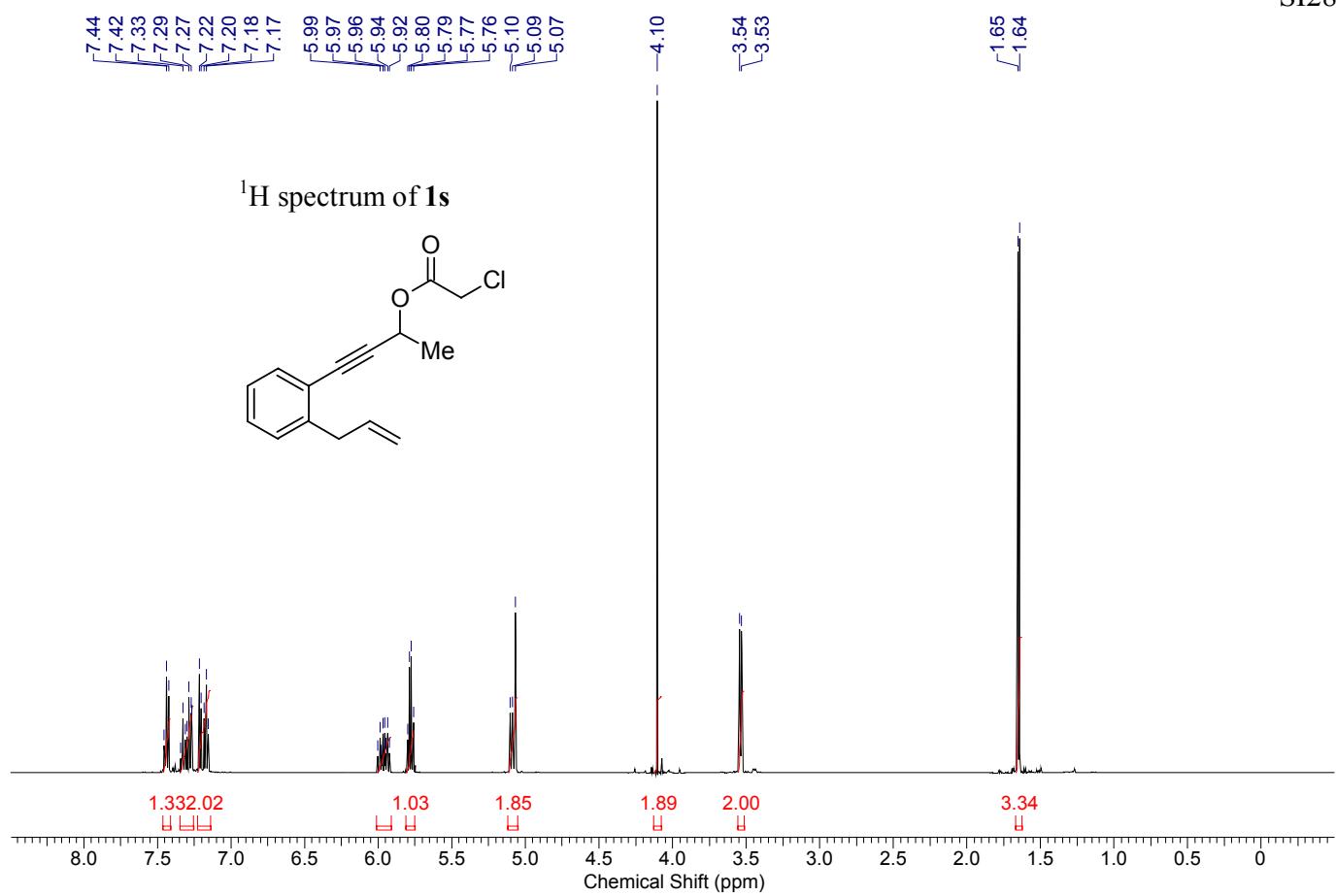


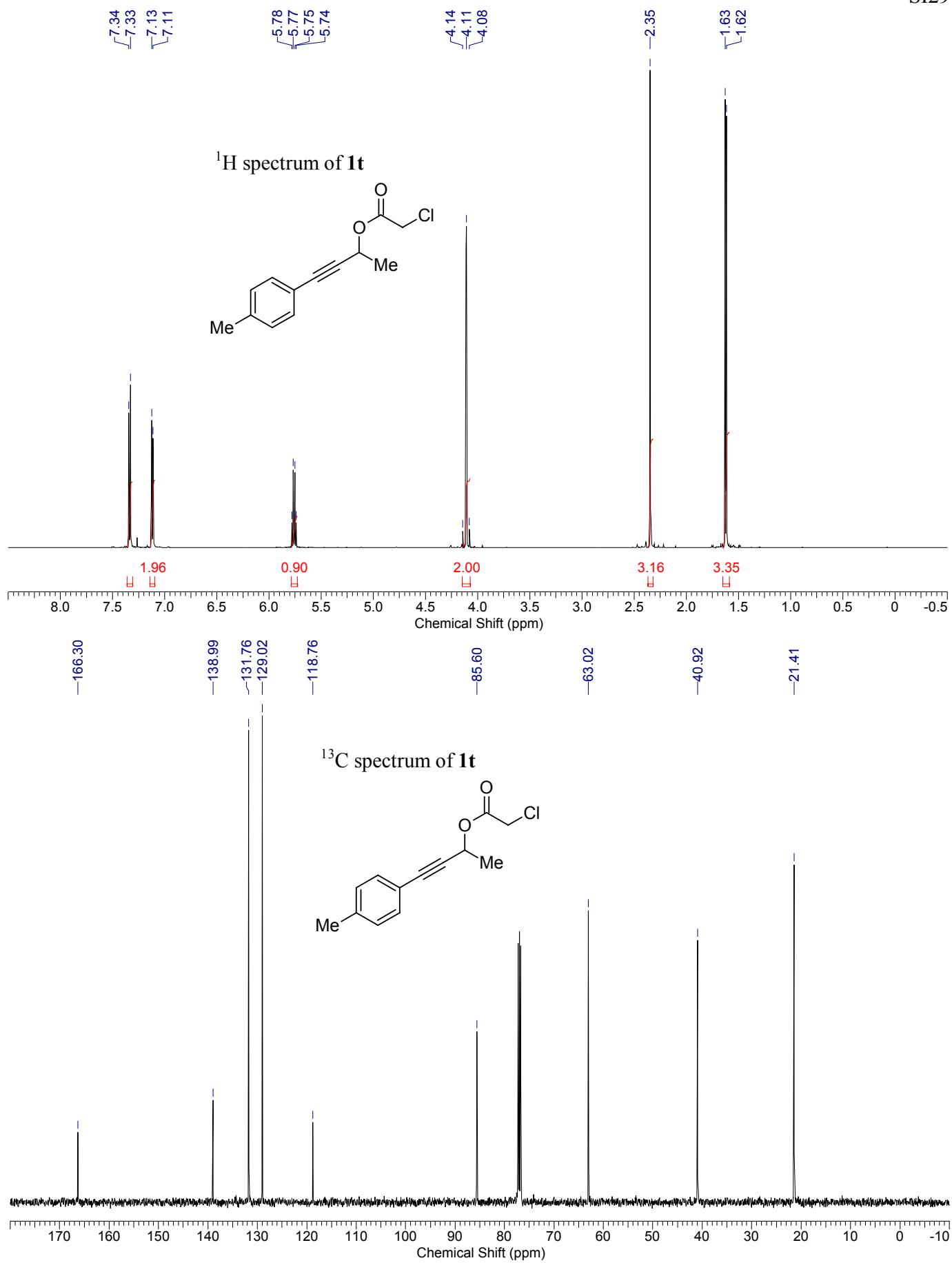


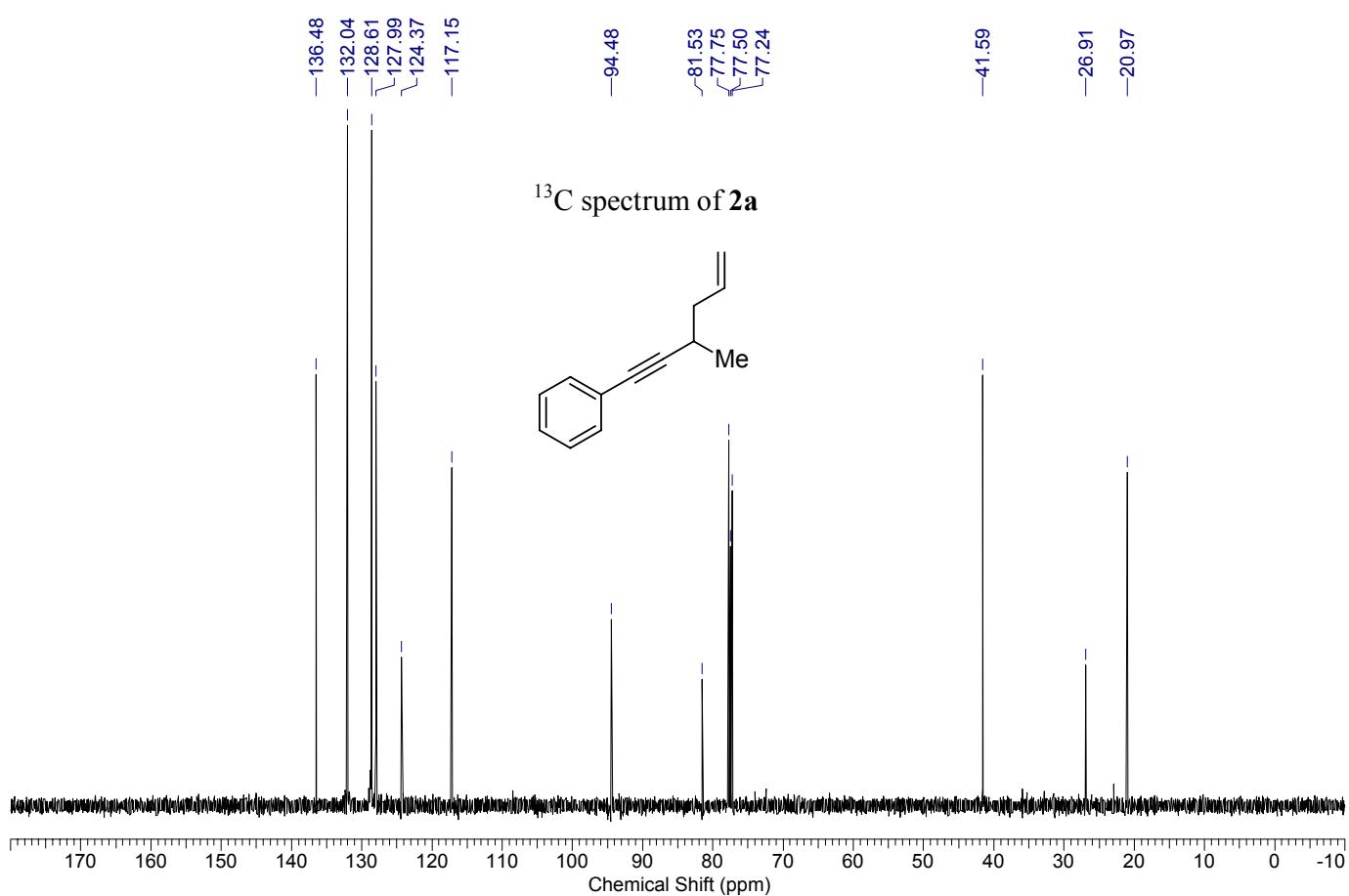
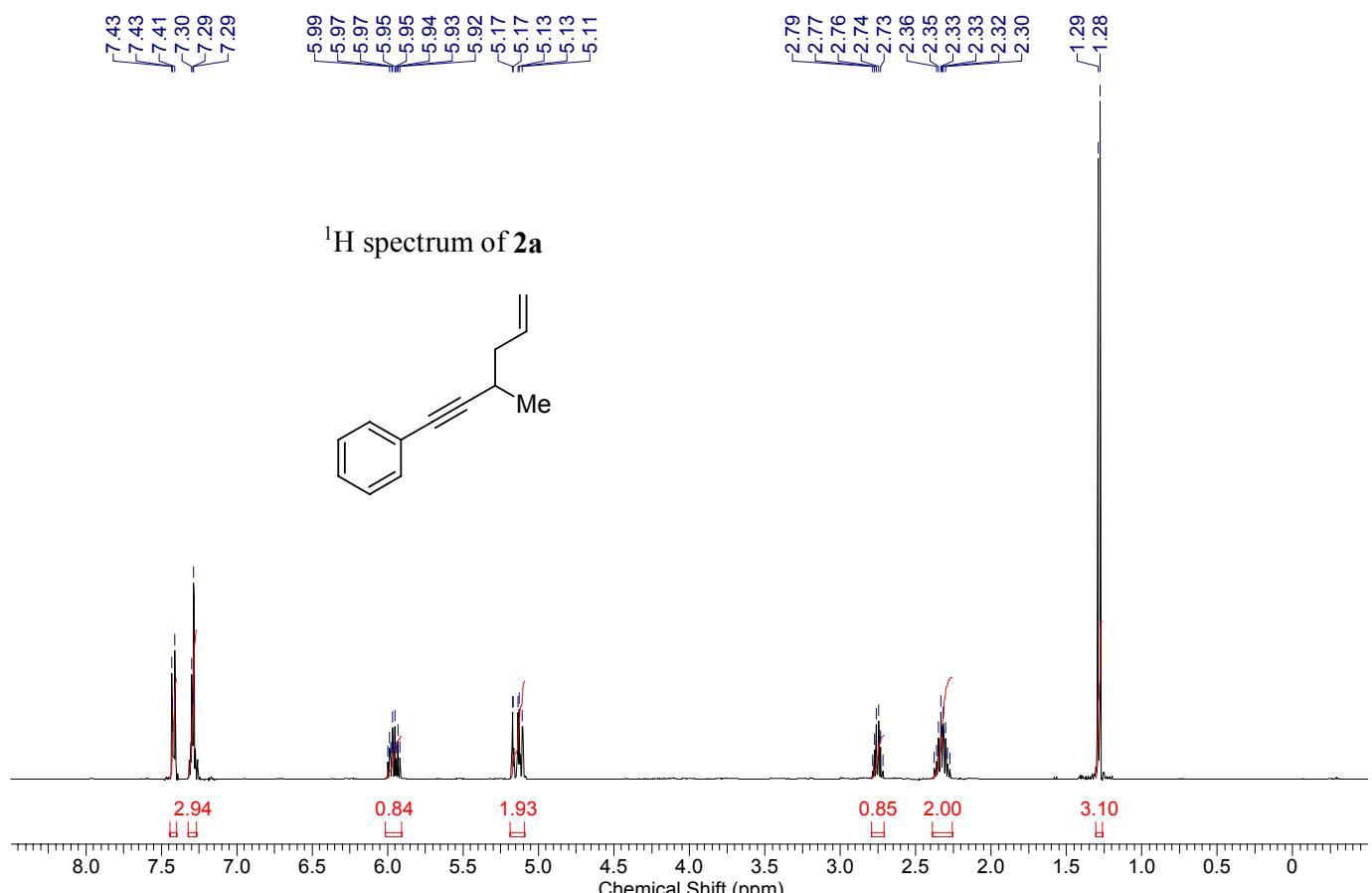


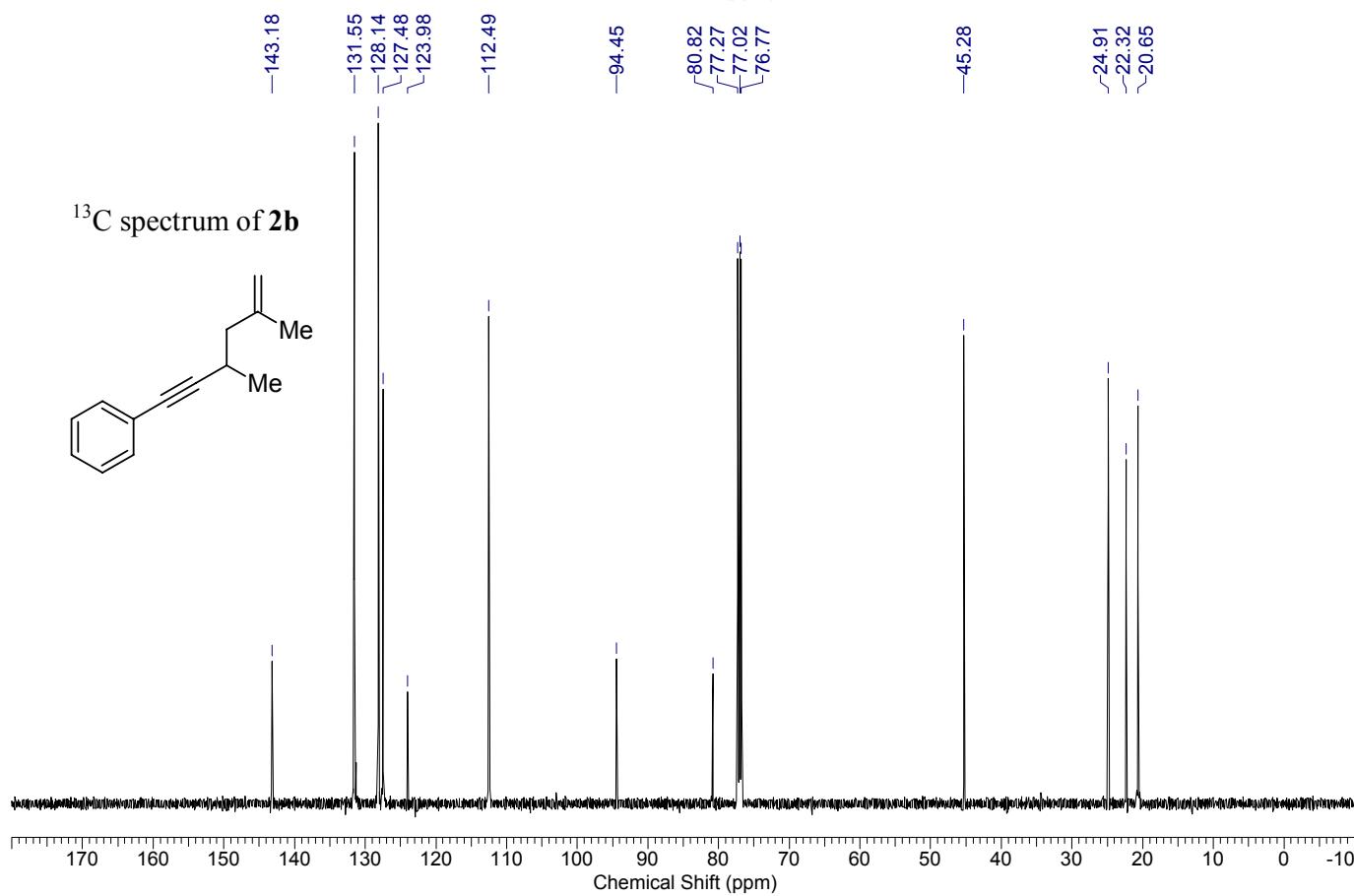
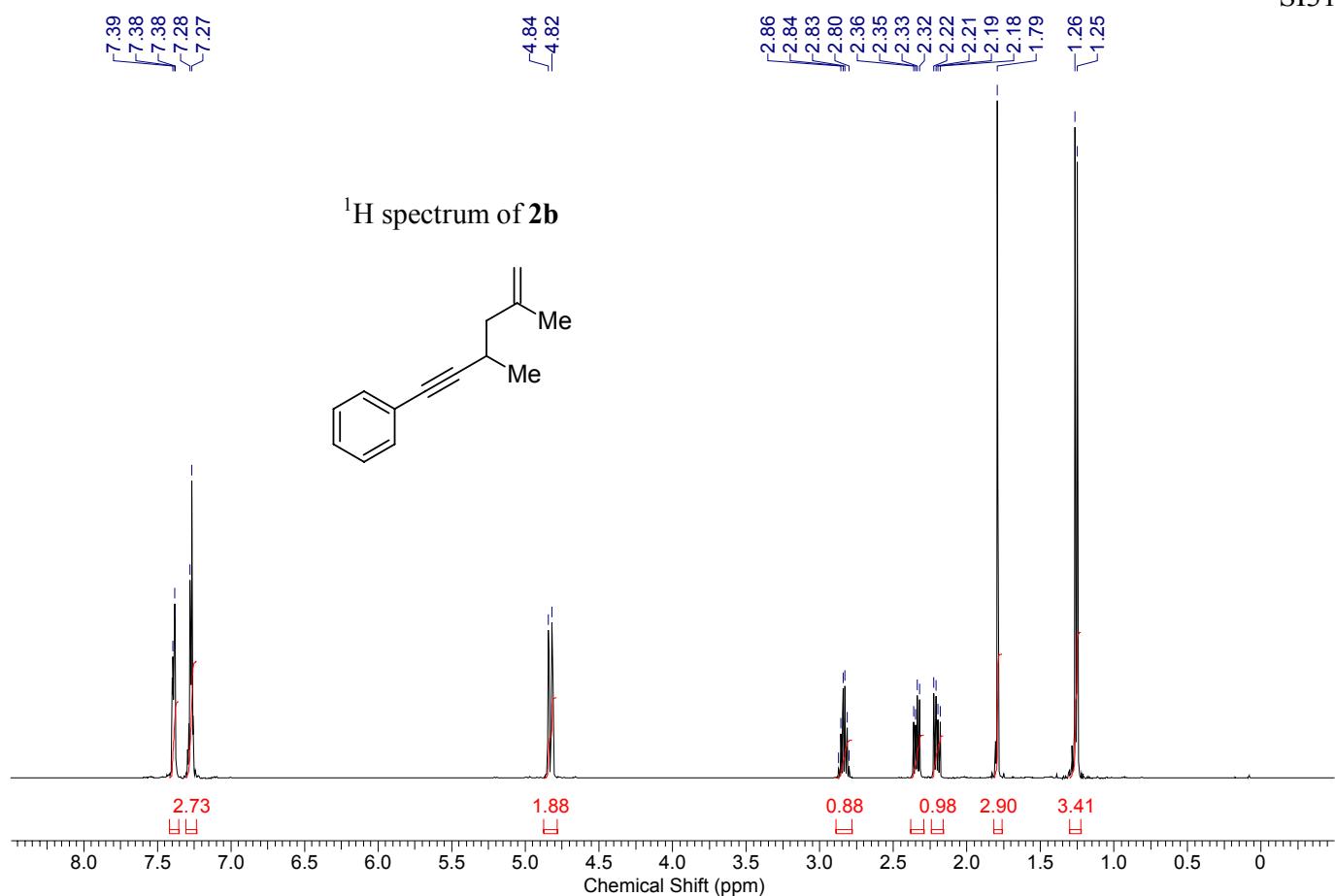


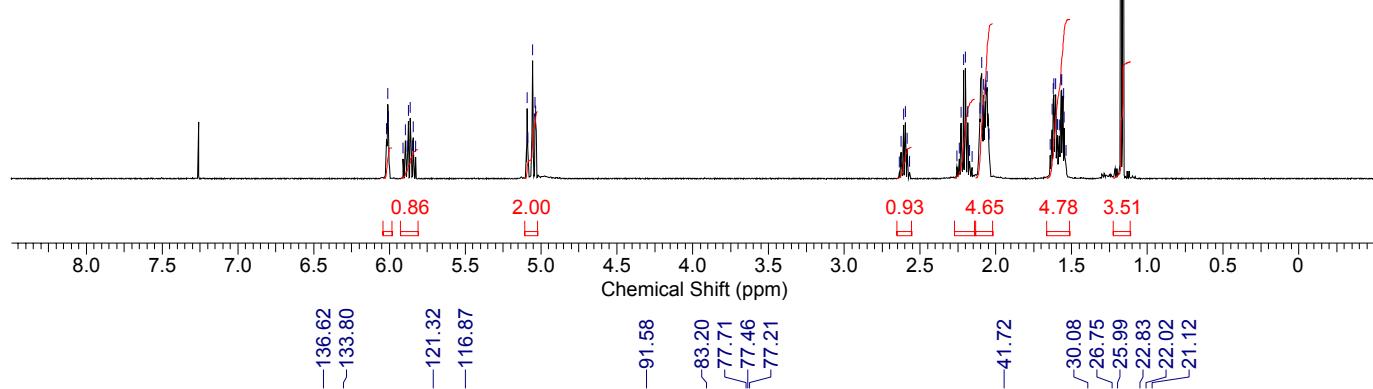
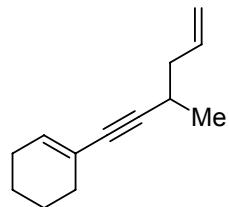
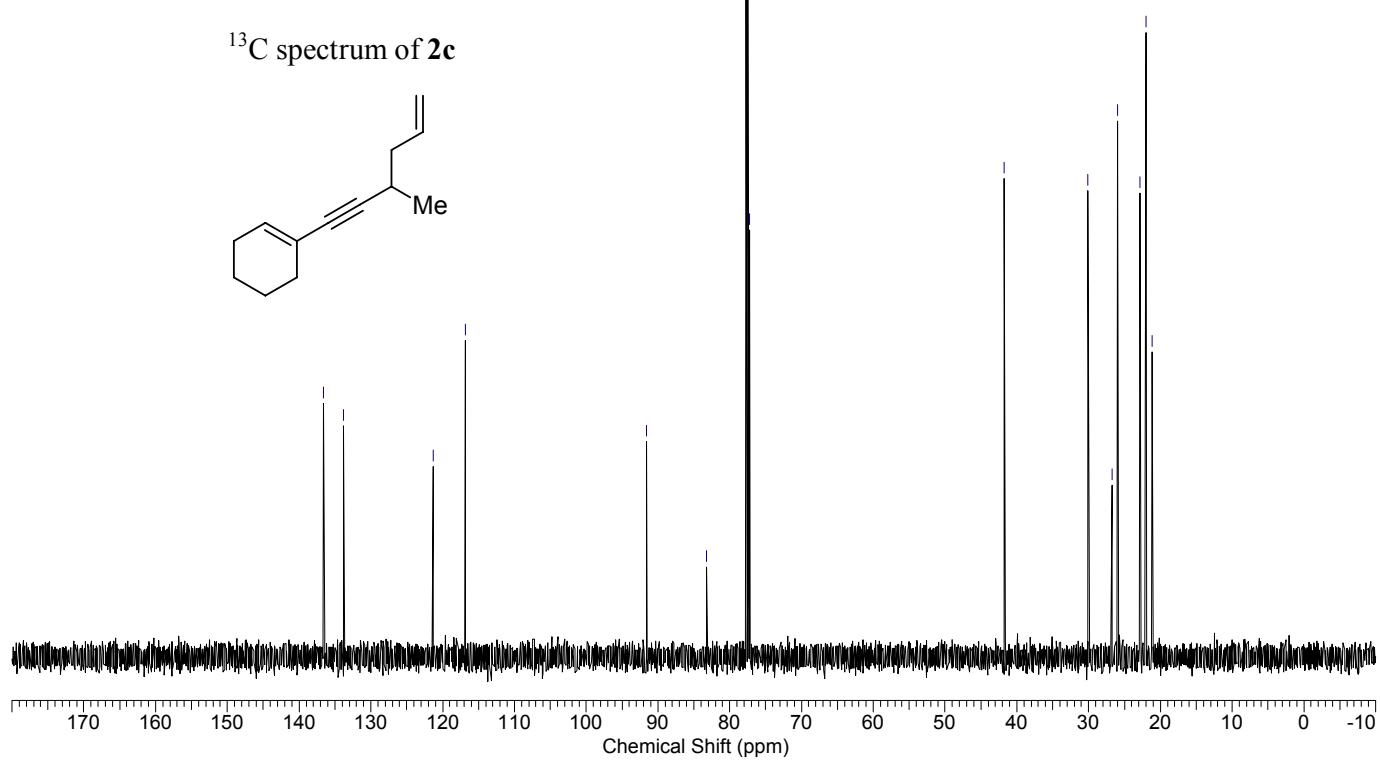
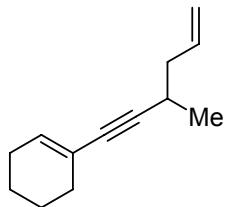


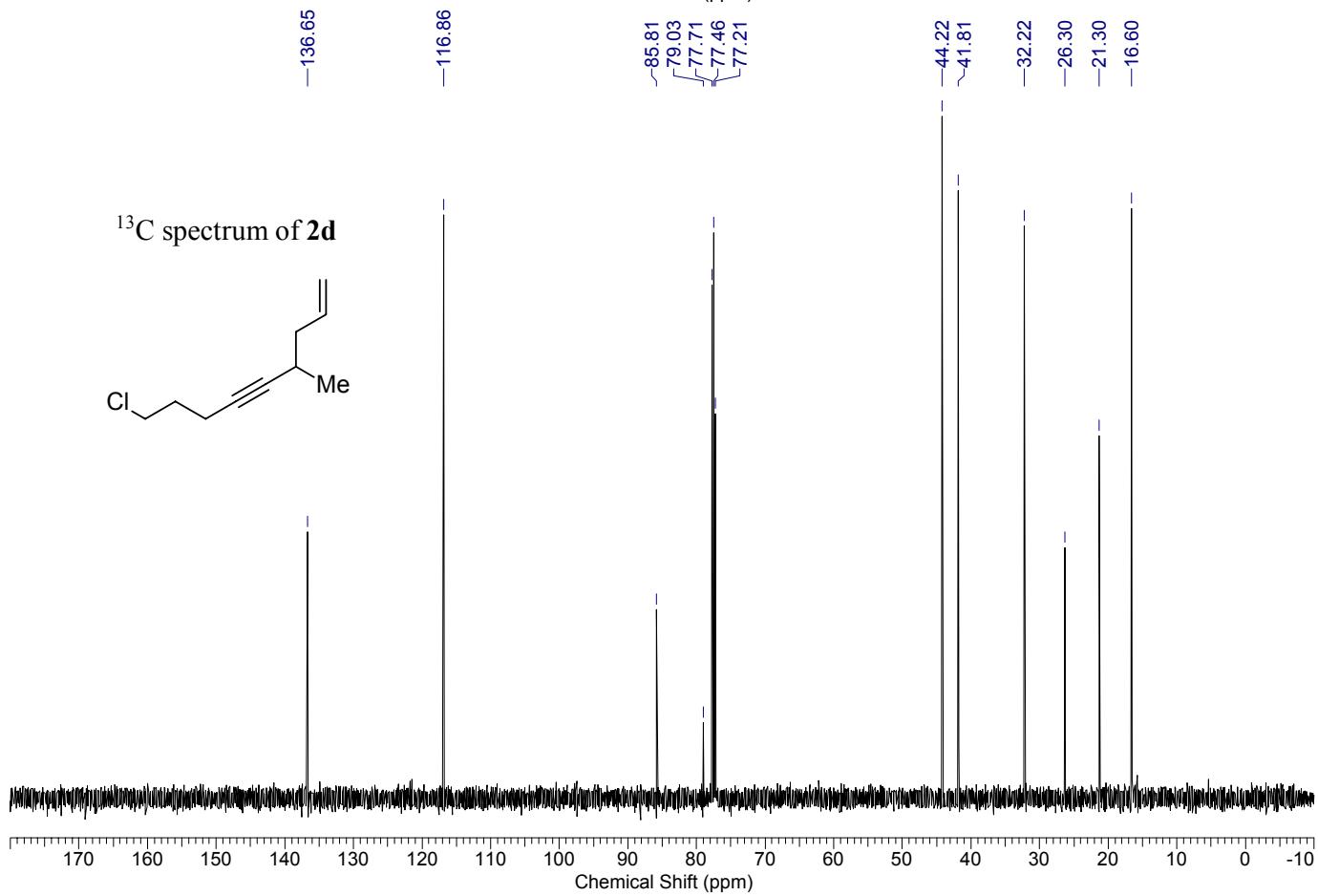
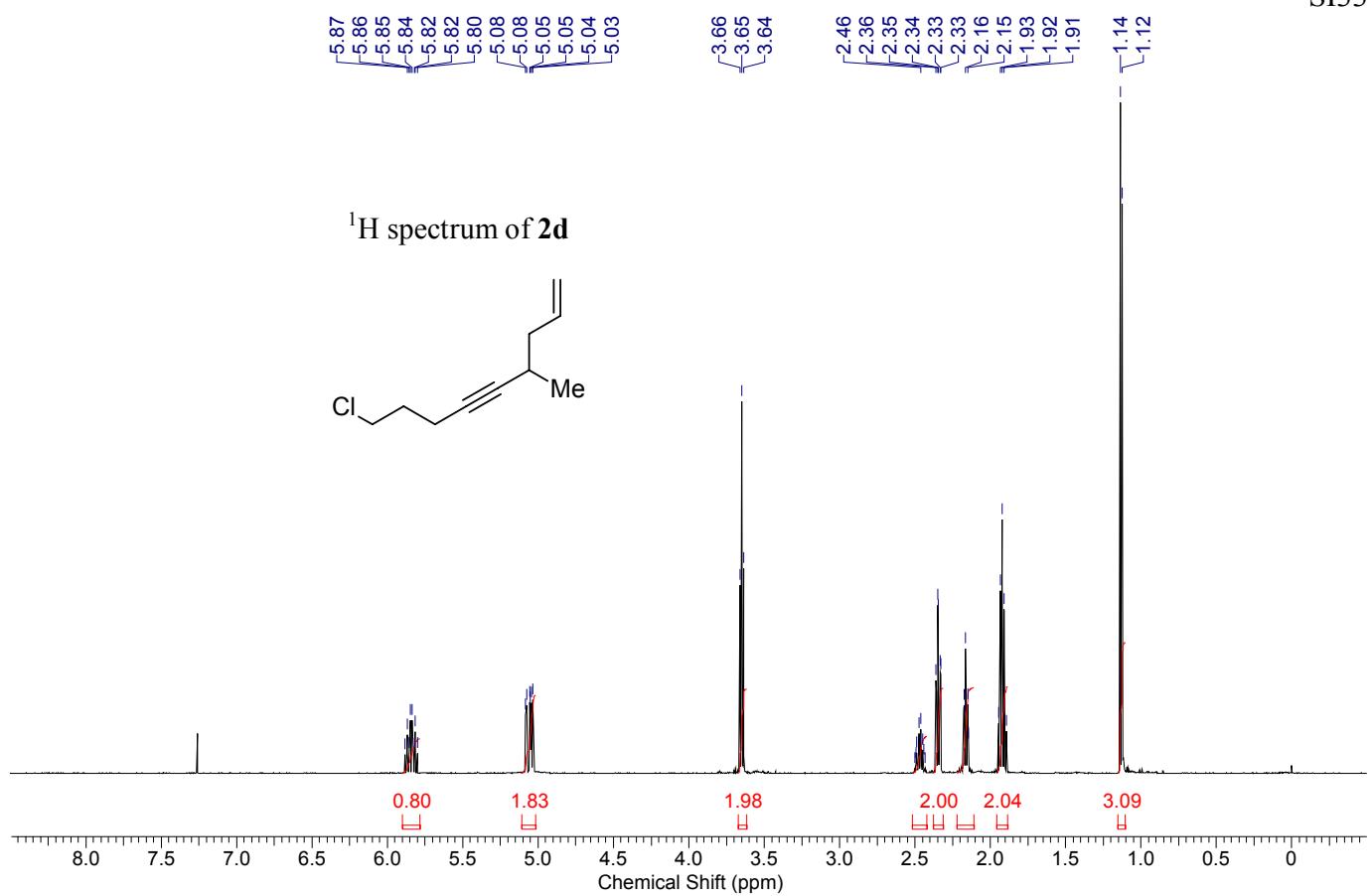


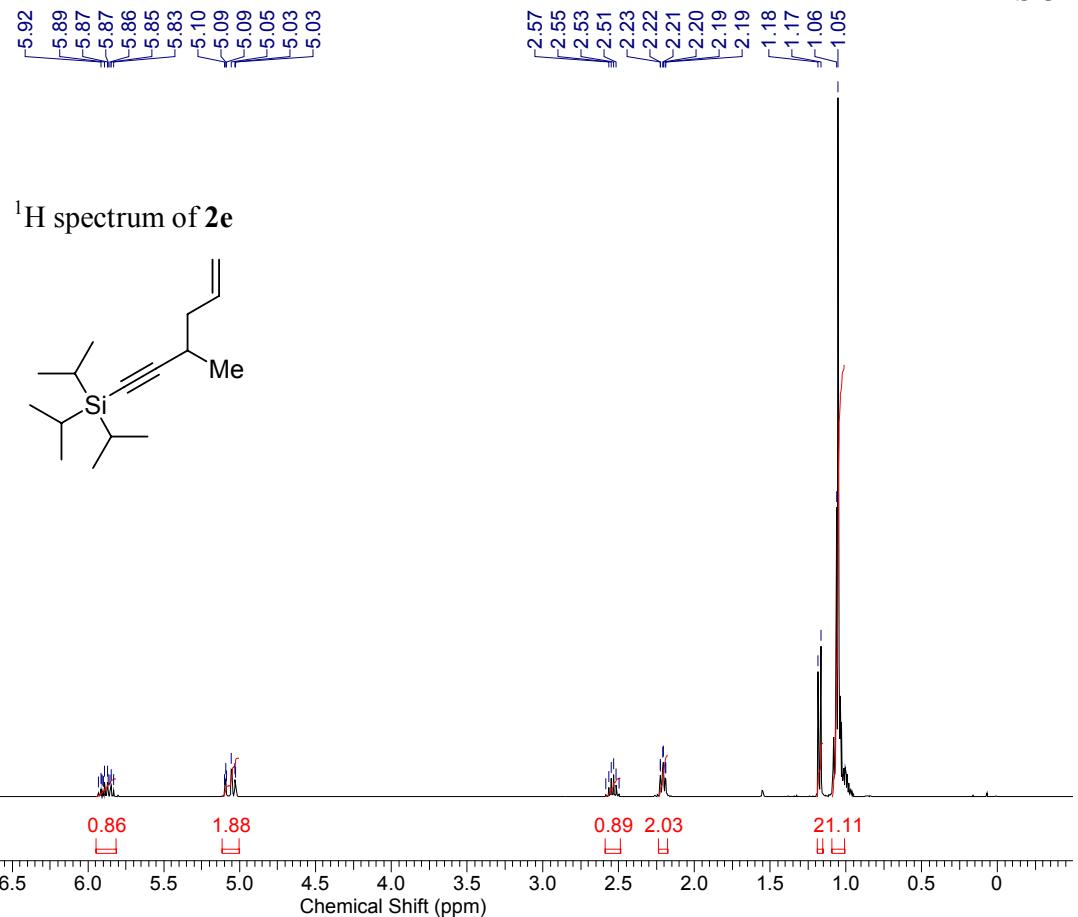






¹H spectrum of **2c**¹³C spectrum of **2c**



¹³C spectrum of **2e**

Chemical Shift (ppm)

Peaks labeled: -136.02, -116.48, -113.13, -113.13, -80.05, -77.32, -77.00, -76.68, -41.27, -26.85, -20.75, -18.61, -11.23

