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Supporting Information

Environmentally Benign Synthesis of H-Phosphinic Acids Using a Water-Tolerant, Recyclable Polymer-Supported Catalyst

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General Chemistry. ¹H NMR spectra were recorded on a Varian Mercury-300 spectrometer. Chemical shifts for ¹H NMR spectra are reported (in parts per million) relative to internal tetramethylsilane (Me₄Si, $\delta = 0.00$ ppm) with CDCl₃ as solvent. ¹³C NMR spectra were recorded at 75 MHz. Chemical shifts for ¹³C NMR spectra are reported (in parts per million) relative to CDCl₃ ($\delta = 77.0$ ppm). ³¹P NMR spectra were recorded at 121 MHz on a Varian Mercury-300, spectrometer and/or at 36 MHz on an Anasazi EFT-90 spectrometer, and chemical shifts reported (in parts per million) relative to external 85% phosphoric acid ($\delta = 0.0$ ppm). Radial chromatography was carried out with a Harrison Associates Chromatotron using 1, 2, or 4 mm layers of silica gel 60 PF₂₅₄ containing gypsum (E. Merck). Silica gel (200-300 mesh, Natland International Corporation) was used for flash chromatography. Ethyl acetate/hexanes mixtures were used as the eluent for chromatographic purifications. TLC plates were visualized by immersion in anisaldehyde stain (by volume: 93% ethanol, 3.5% sulfuric acid, 1% acetic acid, and 2.5% anisaldehyde) followed by heating. Organic solutions of products were dried over anhydrous MgSO₄.

Reagents and Solvents. Palladium catalysts and ligands were purchased from Strem or Aldrich. Polystyrene isocyanate was purchased from Advanced ChemTech or Aldrich, and used as received. Aqueous hypophosphorous acid (50 wt. %), was obtained from Aldrich and used as received. Concentrated hypophosphorous acid was obtaining by concentrating the 50 wt. % aqueous solution in vacuo on a rotary evaporator, at room temperature for 20-30 min before reaction. Anhydrous toluene was freshly distilled from CaH₂ prior to use. All other reagents were used as received. Reagent or HPLC grade acetonitrile was used throughout this study and was not dried prior to use.

³¹**P NMR Yield Measurements.** The NMR yields are determined by integration of all the resonances in the ³¹**P** NMR spectra, an approach which is valid if no phosphorus-containing gas (ie. PH₃) evolves, or if the precipitate in a heterogeneous mixture does not contain phosphorus. The yields determined by NMR are generally accurate within ~10% of the value indicated, and are reproducible. Isolated yields are sometimes significantly lower because the acids are isolated by extraction, and therefore the corresponding isolated yields directly reflect water-solubility.

Octylphosphinic acid (Table 2, entry 1). References: (a) Deprèle, S.; Montchamp, J.-L. J. Org. Chem. 2001, 66, 6745; (b) Deprèle, S.; Montchamp, J.-L. J. Am. Chem. Soc. 2002, 124, 9386; (c) Karanewsky, D. S.; Badia, M. C.; Cushman, D. W.; DeForrest, J. M.; Dejneka, T.; Loots, M. J.; Perri, M. G.; Petrillo Jr.; E. W.; Powell, J. R. J. Med. Chem. 1988, 31, 204; (d) Nifant'ev, E. E.; Magdeeva, R. K.; Shchepet'eva, N. P. J. Gen. Chem. USSR 1980, 50, 1416; (e) Devedjiev, I.; Ganev, V.; Stefanova, R.; Borisov, G. Phosphorus Sulfur 1987, 31, 7. ¹H NMR (CDCl₃)

δ 10.13 (bs, 1H), 7.08 (d, J = 540 Hz, 1 H), 1.70-1.80 (m, 2 H), 1.50-1.65 (m, 2 H), 1.25-1.40 (m, 12 H), 0.88 (t, J = 7 Hz, 3 H); ¹³C NMR (CDCl₃) δ 31.6, 30.3 (d, $J_{PCCC} = 16$ Hz), 29.1 (d, $J_{PC} = 94$ Hz), 29.0, 28.9, 22.5, 20.6 (d, $J_{PCC} = 3$ Hz), 14.0; ³¹P NMR (CDCl₃) δ 38.9 (dt, $J_{P-H} = 540$, 12 Hz).

Decylphosphinic acid (Table 2, entry 2). References: : (a) Deprèle, S.; Montchamp, J.-L. J. Org. Chem. **2001**, 66, 6745; (b) Karanewsky, D. S.; Badia, M. C.; Cushman, D. W.; DeForrest, J. M.; Dejneka, T.; Loots, M. J.; Perri, M. G.; Petrillo Jr.; E. W.; Powell, J. R. J. Med. Chem. **1988**, 31, 204; (c) Nifant'ev, E. E.; Magdeeva, R. K.; Shchepet'eva, N. P. J. Gen. Chem. USSR **1980**, 50, 1416. ¹H NMR (CDCl₃) δ 11.58 (bs, 1H), 7.08 (d, J = 540 Hz, 1 H), 1.20-1.75 (m, 18 H), 0.88 (t, J = 6 Hz, 3 H); ¹³C NMR (CDCl₃) δ 31.9, 30.4 (d, $J_{PCCC} = 16$ Hz), 29.9, 29.2 (d, $J_{PC} = 94$ Hz), 29.4, 29.3, 29.2, 22.7, 20.6(d, $J_{PCC} = 3$ Hz), 14.1; ³¹P NMR (CDCl₃) δ 39.3 (dm, $J_{P-H} = 540$ Hz).

4-Phenylbutylphosphinic acid (Table 2, entries 3 & 4). References: (a) Deprèle, S.; Montchamp, J.-L. *J. Org. Chem.* **2001**, *66*, 6745; (b) Karanewsky, D. S.; Badia, M. C.; Cushman, D. W.; DeForrest, J. M.; Dejneka, T.; Loots, M. J.; Perri, M. G.; Petrillo Jr.; E. W.; Powell, J. R. *J. Med. Chem.* **1988**, *31*, 204; (c) Anderson, N. G.; Coradetti, M. L.; Cronin, J. A.; Davies, M. L.; Gardineer, M. B.; Kotnis, A. S.; Lust, D. A.; Palaniswamy, V. A. *Org. Process Res. Dev.* **1997**, *1*, 315. ¹H NMR (CDCl₃) δ 11.74 (bs, 1H), 7.15-7.25 (m, 5H), 7.04 (d, *J* = 542 Hz, 1 H), 2.61 (t, *J* = 7 Hz, 2 H), 1.60-1.75 (m, 6 H); ¹³C NMR (CDCl₃) δ 141.6, 128.3, 125.8,35.3, 32.1 (d, *J*_{PCCC} = 16 Hz), 29.0 (d, *J*_{PC} = 94 Hz), 20.3 (d, *J*_{PCC} = 3 Hz); ³¹P NMR (CDCl₃) δ 38.2 (dm, *J*_{P-H} = 542 Hz).

7-Methyl-2-octenephosphinic acid (Table 2, entry 5). ¹H NMR (CDCl₃) δ 10.30 (bs, 1H), 7.08 (d, *J* = 540 Hz, 1 H), 5.09 (t, *J* = 1.5 Hz, 1 H); 1.95-2.0 (m, 2 H), 1.55-1.80 (m, 8 H), 1.30-1.45 (m, 6 H); ¹³C NMR (CDCl₃) δ 131.8, 124.6, 30.3 (d, *J*_{PCCC} = 16 Hz), 29.5 (d, *J*_{PC} = 94 Hz), 29.6, 27.9, 25.9, 20.8 (d, *J*_{PCC} = 3 Hz), 17.9; ³¹P NMR (CDCl₃) δ 38.9 (dm, *J*_{P-H} = 540 Hz).

1-Phenylethylphosphinic acid (Table 2, entry 6). Reference: Buckler, S. A.; Epstein, M. *Tetrahedron* **1962**, *18*, 1211. ¹H NMR (CDCl₃) d 9.08 (bs, 1H), 7.20-7.35 (m, 5H), 6.90 (d, J = 552 Hz, 1 H), 3.10-3.20 (m, 1 H), 1.50-1.60 (dd, J = 19, 7 Hz, 3 H); ³¹P NMR (CDCl₃) d 41.2 (dm, $J_{P-H} = 552$ Hz).

2-Phenylethylphosphinic acid (Table 2, entry 6). References: (a) Arbuzova, S. N.; Gusarova, N. K.; Malysheva, S. F.; Brandsma, L.; Albanov, A. I.; Trofimov, B. A. *Russ. J. Gen. Chem.* **1996**, *66*, 54; (b) Jackson, P. F.; Tays, K. L.; Maclin, K. M.; Ko, Y.-S.; Li, W.; Vitharana, D.; Tsukamoto, T.; Stoermer, D.; Lu, X.-C. M.; Wozniak, K.; Slusher, B. S. *J. Med. Chem.* **2001**, *44*, 4170; (c) Lui, X.; Hu, E.; Tian, X.; Mazur, A.; Ebetino, F. H. *J. Organomet. Chem.* **2002**, *646*, 212; (d) Gusarova, N. K.; Shaikhudinova, S. I.; Sukhov, B. G.; Kazantseva, T. I.; Malysheva, S. F.; Smetannikov, Y. V.; Tarasova, N. P.; Kuimov, V. A.; Trofimov, B. A. *Russ. Chem. Bull.* **2003**, *52*, 511. ¹H NMR (CDCl₃) d 9.08 (bs, 1H), 7.20-7.35 (m, 5H), 7.06 (d, *J* = 548 Hz, 1 H), 2.85-2.95 (m, 2 H), 2.05-2.15 (m, 2H); ³¹P NMR (CDCl₃) d 37.6 (dm, $J_{P-H} = 549$ Hz).

(1-Cyclohexyl) methylphosphinic acid (Table 2, entry 7). ¹H NMR (CDCl₃) δ 10.65 (bs, 1H), 7.18 (d, J = 540 Hz, 1 H), 1.60-1.75 (m, 7 H), 1.40-1 (m, 6 H); ¹³C NMR (CDCl₃) δ 37.0, 36.7, 34.5, 34.4, 31.7, 26.0, 25.9; ³¹P NMR (CDCl₃) δ 37.9 (dm, $J_{P-H} = 540$ Hz).

[2-(4-Chloro-phenyl)-propyl]-phosphinic acid (Table 2, entry 8).

¹H NMR (CDCl₃) δ 10.0 (bs, 1H), 7.1-7.4 (m, 4H), 6.85 (d, J = 550 Hz, 1 H), 3.15-3.25 (m, 1 H), 2.0-2.1 (m, 2 H), 1.34 (t, J = 6 Hz, 3 H); ¹³C NMR (CDCl₃) δ 143.9, 143.8, 132.4, 128.8, 128.3, 128.37.6 (d, $J_{PC} = 94$ Hz), 33.3, 23.6(d, $J_{PCC} = 10$ Hz), 14.0; ³¹P NMR (CDCl₃) δ 39.7 (dm, $J_{P-H} = 550$ Hz).

2-Phthalimidylethylphosphinic acid (Table 2, entry 9). ¹H NMR (DMSO) δ 7.80-7.85 (m, 4H), 7.0 (d, J = 531 Hz, 1 H), 3.70-3.80 (m, 2 H), 1.95-2.05 (m, 2H); ¹³C NMR (DMSO) δ 168.3, 135.0, 132.4, 123.7, 31.4, 29.5 (d, $J_{PC} = 89$ Hz); ³¹P NMR (DMSO) δ 23.3 (dm, $J_{P-H} = 532$ Hz).

(1-Phenyl-vinyl)-phosphinic acid (Table 2, entry 10). Reference: Dumond, Y. R.; Montchamp, J.-L. J. Organomet. Chem. 2002, 653, 252. ¹H NMR (CDCl₃, SiMe₄) δ 12.37 (br, 1 H), 7.4-7.5 (m, 2 H), 7.2-7.4 (m, 3 H), 7.30 (d, J = 571 Hz, 1 H), 6.16 (d, J = 46 Hz, 1 H), 6.14 (d, J = 25 Hz, 1 H); ¹³C NMR (CDCl₃) δ 142.3 (d, $J_{PC} = 124$ Hz), 135.1 (d, $J_{PCC} = 13$ Hz), 129.3 (d, $J_{PCC} = 13$ Hz), 128.7 (2 C), 128.6, 127.3 (d, $J_{PCCC} = 6$ Hz, 2 C); ³¹P NMR (CDCl₃) δ 25.7 (ddd, J = 571 Hz, J = 46 Hz, J = 24 Hz).

Trans-Oct-1-enyl-phosphinic acid (Table 2, entry 11). References: (a) Dumond, Y. R.; Montchamp, J.-L. *J. Organomet. Chem.* 2002, *653*, 252; (b) Nifant'ev, E. E.; Magdeeva, R. K.; Maslennikova, V. I.; Taber, A. M.; Kalechits, I. V. *J. Gen. Chem. USSR* 1982, *52*, 2173; (c) Belakhov, V. V.; Yudelevich, V. I.; Fetter, A. P.; Ionin, B. I. *J. Gen. Chem. USSR* 1986, *56*, 680. ¹H NMR (CDCl₃, SiMe₄) δ 10.64 (br, 1 H), 7.2 (d, *J* = 562 Hz, 1 H), 6.6-6.8 (m, 1 H), 5.82 (dd, *J* = 17 Hz, *J* = 24 Hz, 1 H), 2.2-2.3 (m, 2 H), 1.4-1.5 (m, 2 H), 1.2-1.4 (m, 6 H), 0.88 (d, *J* = 6 Hz, 3 H); ¹³C NMR (CDCl₃) δ 153.7 (d, *J*_{PCC} = 6 Hz), 120.5 (d, *J*_{PCC} = 134 Hz), 34.1 (d, *J*_{PCCC} = 20 Hz), 31.5, 28.7, 27.6, 22.5, 14.0; ³¹P NMR (CDCl₃) 23.9 (d, *J* = 562 Hz).

(1-Hexyl-vinyl)-phosphinic acid (Table 2, entry 11). Reference: Dumond, Y. R.; Montchamp, J.-L. *J. Organomet. Chem.* 2002, 653, 252. ¹H NMR (CDCl₃, SiMe₄) δ 12.78 (br, 1 H), 7.15 (d, *J* = 557 Hz, 1 H), 5.92 (d, *J* = 25 Hz, 1 H), 5.79 (d, *J* = 49 Hz, 1 H), 2.30 (m, 2 H), 1.5-1.6 (m, 2 H), 1.2-1.4 (m, 6 H), 0.88 (t, *J* = 7 Hz, 3 H); ¹³C NMR (CDCl₃) δ 142.6 (d, *J*_{PC} = 121 Hz), 127.4 (d, *J*_{PCC} = 14 Hz), 31.46, 30.5 (d, *J*_{PCC} = 13 Hz), 28.7, 27.7 (d, *J*_{PCCC} = 5 Hz), 22.5, 13.9; ³¹P NMR (CDCl₃) δ 27.4 (dm, *J* = 557 Hz).

(1-Butyl-hex-1-enyl) phosphinic acid (Table 2, entry 12). ¹H NMR (CDCl₃) δ 12.77 (bs, 1 H), 7.10 (d, J = 550 Hz, 1 H), 6.32 (dt, J = 27 Hz, J = 7 Hz, 1 H), 2.15-2.3 (m, 4 H), 1.25-1.50 (m, 8 H), 0.89 (d, J = 7 Hz, 6 H); ¹³C NMR (CDCl₃) δ 145.3 (d, $J_{PCC} = 14$ Hz), 132.8 (d, $J_{PC} = 127$ Hz), 31.3, 30.6, 28.0, 27.8, 24.8 (d, $J_{PCC} = 13$ Hz), 22.7, 22.3, 13.7; ³¹P NMR (CDCl₃) 30.73 (dm, J = 550).

Preparation of supported-ligand 4. Dry toluene (15 mL) was added to a mixture of polystyrene isocyanate resin (1.003 g, 2.4 mmol; Advanced Chemtech, 100-200 Mesh, 2.4 mmol/g), and 4,6-bis(diphenylphosphino)phenoxazine **2** (0.653 g, 1.185 mmol; Strem, nixantphos). The suspension was stirred and heated at 115 °C overnight, under N₂. To the resulting mixture was added anhydrous diisopropylamine (0.5 ml, 3.56 mmol) in order to neutralize the unreacted isocyanate groups. The reaction mixture was stirred at room

temperature for 6 hours, under N₂. After filtration, the collected resin (1.2 g) was rinsed with dichloromethane and dried overnight under vacuum, over P_2O_5 ; while the filtrate was evaporated to recover unreacted nixantphos as an off-white powder. This amount was taken into account to calculate the loading of the resin. The supported ligand's loading was found to be 0.14 mmol/g.

Representative hydrophosphinylation procedure with 50 % aqueous H₃PO₂. Aqueous hypophosphorous acid (50 wt. %) was first weighted in a round bottom flask, (0.2723 g, 2.06 mmol). Acetonitrile (2 mL), was then added to the flask. Addition of 4-phenylbutene (0.15 mL, 0.99 mmol), Pd₂dba₃ (4.6 mg, 1 mol %), and supported nixantphos, (0.14 mmol/g, 64.8 mg, 1.08 mol %) followed. In order to avoid any potential leak of solvent, the neck of the flask was washed with another portion of acetonitrile (2 mL). The resulting mixture was stirred and refluxed overnight under nitrogen. After measuring the crude yield by ³¹P NMR, the reaction mixture was cooled down to room temperature and then filtered to collect the resin. The filtrate was then extracted with hexane, once. The acetonitrile layer was concentrated down and the residue underwent a single extraction with ethyl acetate and brine. The organic layer was dried over MgSO₄. After filtration and concentration the resulting yellow oil was dried overnight under vacuum, over P₂O₅. The reaction yielded 4-phenylbutylphosphinic acid (0.193 g, 97%) as a yellow oil.

Hydrophosphinylation procedure with concentrated H_3PO_2 . These runs were conducted as above with the 50 wt. % solution except the solution was first concentrated in vacuo at room temperature for 20-30 min, prior to the addition of the substrate and catalyst.

Reaction in air and at room temperature. To a solution of 50 % aq H_3PO_2 (0.211 g, 1.60 mmol) was added some HPLC grade CH₃CN, 4-phenylbutene (0.15 ml, 0.99 mmol), Pd₂dba₃ (5 mg, 1.1 mol %), and supported nixantphos (0.33 mmol/g, 33.9 mg, 1.1 mol %). The reaction was stirred at room temperature in an open flask for 6 hours. After taking a ³¹P NMR of the crude mixture, the yield of the desired phosphinic acid was 78 %. The corresponding isolated yield was 51% (after extraction as above with brine and ethyl acetate).

Reaction in air and at reflux. Aqueous hypophosphorous acid (50 wt. %, 0.356 g, 2.69 mmol) and CH₃CN (10 mL) were added to 1-octene (0.25 ml, 1.59 mmol), along with some Pd₂dba₃ (8 mg, 1.1 mol %) and supported nixantphos (0.33 mmol/g, 67 mg, 1.4 mol %). The reaction was refluxed overnight open to air. After measuring the yield by ³¹P NMR, concentration and partition between ethyl acetate and brine of the corresponding crude phosphinic acid was accomplished. Drying the residue overnight under vacuum over P₂O₅ gave the product (0.188 g, 89%) as yellow oil.

Multi run reactions (Table 2, entry 6). To aqueous hypophosphorous acid (50 wt. %, 0.490 g, 4 mmol) was added styrene (0.30 ml, 2.62 mmol), Pd_2dba_3 (11.6 mg, 0.96 mol %) and supported Nixantphos (0.33 mmol/g, 95.6 mg, 1.2 mol %). The reaction mixture was refluxed overnight under N₂. The next day, after measuring the crude yield by ³¹P NMR, the mixture was filtered in order to separate the resin from the filtrate. The resin was washed with CH₃CN and kept in a dessicator, over P₂O₅, between the runs. The filtrate was kept at in a stoppered Erlenmeyer. The resin was then used in the next run without any further precautions. Aqueous hypophosphorous acid (50 wt. %, 0.490 g, 4 mmol), styrene (0.3 ml, 2.62 mmol) and the previously collected resin

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were mixed together in CH₃CN (10 mL). The reaction mixture was again refluxed overnight under N₂. The next day, after measuring the crude yield by ³¹P NMR, the mixture was filtered in order to separate the resin and the filtrate. The resin was washed with CH₃CN, and collected for another run. The filtrate was kept in the same Erlenmeyer as before. For the third time, the reaction was run, with 50 % aq H₃PO₂ (0.390 g, 3.17 mmol), styrene (0.30 ml, 2.62 mmol) and the previously collected resin. The same procedures were used to run the reaction, control the NMR yield and separate the resin from the desired product. The experiment was then run for one last time: 50 % aq H₃PO₂ (0.499 g, 4 mmol), styrene (0.30 ml, 2.62 mmol) and the previously collected resin. At the end of the sequence, all the filtrates were in a single Erlenmeyer. After concentration, partition between EtOAc and brine, the product (2.00 g, 90% over 4 runs) was obtained as a yellow oil and a mixture of regioisomers.













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