## Supporting Information

# Environmentally Benign Synthesis of H-Phosphinic Acids Using a Water-Tolerant, Recyclable Polymer-Supported Catalyst 

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General Chemistry. ${ }^{1} \mathrm{H}$ NMR spectra were recorded on a Varian Mercury-300 spectrometer. Chemical shifts for ${ }^{1} \mathrm{H}$ NMR spectra are reported (in parts per million) relative to internal tetramethylsilane ( $\mathrm{Me}_{4} \mathrm{Si}, \delta=0.00 \mathrm{ppm}$ ) with $\mathrm{CDCl}_{3}$ as solvent. ${ }^{13} \mathrm{C}$ NMR spectra were recorded at 75 MHz . Chemical shifts for ${ }^{13} \mathrm{C}$ NMR spectra are reported (in parts per million) relative to $\mathrm{CDCl}_{3}(\delta=77.0 \mathrm{ppm}) .{ }^{31} \mathrm{P}$ NMR spectra were recorded at 121 MHz on a Varian Mercury-300, spectrometer and/or at 36 MHz on an Anasazi EFT-90 spectrometer, and chemical shifts reported (in parts per million) relative to external $85 \%$ phosphoric acid ( $\delta=0.0 \mathrm{ppm}$ ). Radial chromatography was carried out with a Harrison Associates Chromatotron using 1, 2, or 4 mm layers of silica gel $60 \mathrm{PF}_{254}$ containing gypsum (E. Merck). Silica gel (200-300 mesh, Natland International Corporation) was used for flash chromatography. Ethyl acetate/hexanes mixtures were used as the eluent for chromatographic purifications. TLC plates were visualized by immersion in anisaldehyde stain (by volume: $93 \%$ ethanol, $3.5 \%$ sulfuric acid, $1 \%$ acetic acid, and $2.5 \%$ anisaldehyde) followed by heating. Organic solutions of products were dried over anhydrous $\mathrm{MgSO}_{4}$.

Reagents and Solvents. Palladium catalysts and ligands were purchased from Strem or Aldrich. Polystyrene isocyanate was purchased from Advanced ChemTech or Aldrich, and used as received. Aqueous hypophosphorous acid ( $50 \mathrm{wt} . \%$ ), was obtained from Aldrich and used as received. Concentrated hypophosphorous acid was obtaining by concentrating the 50 wt . \% aqueous solution in vacuo on a rotary evaporator, at room temperature for 20-30 min before reaction. Anhydrous toluene was freshly distilled from $\mathrm{CaH}_{2}$ prior to use. All other reagents were used as received. Reagent or HPLC grade acetonitrile was used throughout this study and was not dried prior to use.
${ }^{31} \mathbf{P}$ NMR Yield Measurements. The NMR yields are determined by integration of all the resonances in the ${ }^{31} \mathrm{P}$ NMR spectra, an approach which is valid if no phosphorus-containing gas (ie. $\mathrm{PH}_{3}$ ) evolves, or if the precipitate in a heterogeneous mixture does not contain phosphorus. The yields determined by NMR are generally accurate within $\sim 10 \%$ of the value indicated, and are reproducible. Isolated yields are sometimes significantly lower because the acids are isolated by extraction, and therefore the corresponding isolated yields directly reflect water-solubility.

Octylphosphinic acid (Table 2, entry 1). References: (a) Deprèle, S.; Montchamp, J.-L. J. Org. Chem. 2001, 66, 6745; (b) Deprèle, S.; Montchamp, J.-L. J. Am. Chem. Soc. 2002, 124, 9386; (c) Karanewsky, D. S.; Badia, M. C.; Cushman, D. W.; DeForrest, J. M.; Dejneka, T.; Loots, M. J.; Perri, M. G.; Petrillo Jr.; E. W.; Powell, J. R. J. Med. Chem. 1988, 31, 204; (d) Nifant'ev, E. E.; Magdeeva, R. K.; Shchepet'eva, N. P. J. Gen. Chem. USSR 1980, 50, 1416; (e) Devedjiev, I.; Ganev, V.; Stefanova, R.; Borisov, G. Phosphorus Sulfur 1987, 31, 7. ${ }^{1} \mathrm{H}$ NMR ( $\mathrm{CDCl}_{3}$ )
$\delta 10.13$ (bs, 1H), $7.08(\mathrm{~d}, J=540 \mathrm{~Hz}, 1 \mathrm{H}), 1.70-1.80(\mathrm{~m}, 2 \mathrm{H}), 1.50-1.65(\mathrm{~m}, 2 \mathrm{H}), 1.25-1.40$ $(\mathrm{m}, 12 \mathrm{H}), 0.88(\mathrm{t}, J=7 \mathrm{~Hz}, 3 \mathrm{H}) ;{ }^{13} \mathrm{C}$ NMR $\left(\mathrm{CDCl}_{3}\right) \delta 31.6,30.3\left(\mathrm{~d}, J_{\mathrm{PCCC}}=16 \mathrm{~Hz}\right), 29.1\left(\mathrm{~d}, J_{\mathrm{PC}}\right.$ $=94 \mathrm{~Hz}), 29.0,28.9,22.5,20.6\left(\mathrm{~d}, J_{\mathrm{PCC}}=3 \mathrm{~Hz}\right), 14.0 ;{ }^{31} \mathrm{P} \operatorname{NMR}\left(\mathrm{CDCl}_{3}\right) \delta 38.9\left(\mathrm{dt}, J_{\mathrm{P}-\mathrm{H}}=540\right.$, 12 Hz ).

Decylphosphinic acid (Table 2, entry 2). References: : (a) Deprèle, S.; Montchamp, J.-L. J. Org. Chem. 2001, 66, 6745; (b) Karanewsky, D. S.; Badia, M. C.; Cushman, D. W.; DeForrest, J. M.; Dejneka, T.; Loots, M. J.; Perri, M. G.; Petrillo Jr.; E. W.; Powell, J. R. J. Med. Chem. 1988, 31, 204; (c) Nifant'ev, E. E.; Magdeeva, R. K.; Shchepet'eva, N. P. J. Gen. Chem. USSR 1980, 50, 1416. ${ }^{1} \mathrm{H}$ NMR $\left(\mathrm{CDCl}_{3}\right) \delta 11.58(\mathrm{bs}, 1 \mathrm{H}), 7.08(\mathrm{~d}, J=540 \mathrm{~Hz}, 1 \mathrm{H}), 1.20-1.75(\mathrm{~m}, 18 \mathrm{H})$, $0.88(\mathrm{t}, J=6 \mathrm{~Hz}, 3 \mathrm{H}) ;{ }^{13} \mathrm{C}$ NMR $\left(\mathrm{CDCl}_{3}\right) \delta 31.9,30.4\left(\mathrm{~d}, J_{\mathrm{PCCC}}=16 \mathrm{~Hz}\right), 29.9,29.2\left(\mathrm{~d}, J_{\mathrm{PC}}=94\right.$ $\mathrm{Hz}), 29.4,29.3,29.2,22.7,20.6\left(\mathrm{~d}, J_{\mathrm{PCC}}=3 \mathrm{~Hz}\right), 14.1 ;{ }^{31} \mathrm{P} \operatorname{NMR}\left(\mathrm{CDCl}_{3}\right) \delta 39.3\left(\mathrm{dm}, J_{\mathrm{P}-\mathrm{H}}=540\right.$ Hz ).

4-Phenylbutylphosphinic acid (Table 2, entries 3 \& 4). References: (a) Deprèle, S.; Montchamp, J.-L. J. Org. Chem. 2001, 66, 6745; (b) Karanewsky, D. S.; Badia, M. C.; Cushman, D. W.; DeForrest, J. M.; Dejneka, T.; Loots, M. J.; Perri, M. G.; Petrillo Jr.; E. W.; Powell, J. R. J. Med. Chem. 1988, 31, 204; (c) Anderson, N. G.; Coradetti, M. L.; Cronin, J. A.; Davies, M. L.; Gardineer, M. B.; Kotnis, A. S.; Lust, D. A.; Palaniswamy, V. A. Org. Process Res. Dev. 1997, $l, 315 .{ }^{1} \mathrm{H}$ NMR $\left(\mathrm{CDCl}_{3}\right) \delta 11.74(\mathrm{bs}, 1 \mathrm{H}), 7.15-7.25(\mathrm{~m}, 5 \mathrm{H}), 7.04(\mathrm{~d}, J=542 \mathrm{~Hz}, 1 \mathrm{H})$, $2.61(\mathrm{t}, J=7 \mathrm{~Hz}, 2 \mathrm{H}), 1.60-1.75(\mathrm{~m}, 6 \mathrm{H}) ;{ }^{13} \mathrm{C}$ NMR $\left(\mathrm{CDCl}_{3}\right) \delta 141.6,128.3,125.8,35.3,32.1$ $\left(\mathrm{d}, J_{\mathrm{PCCC}}=16 \mathrm{~Hz}\right), 29.0\left(\mathrm{~d}, J_{\mathrm{PC}}=94 \mathrm{~Hz}\right), 20.3\left(\mathrm{~d}, J_{\mathrm{PCC}}=3 \mathrm{~Hz}\right) ;{ }^{31} \mathrm{P} \operatorname{NMR}\left(\mathrm{CDCl}_{3}\right) \delta 38.2\left(\mathrm{dm}, J_{\mathrm{P}-\mathrm{H}}\right.$ $=542 \mathrm{~Hz}$ ).

7-Methyl-2-octenephosphinic acid (Table 2, entry 5). ${ }^{1} \mathrm{H}$ NMR $\left(\mathrm{CDCl}_{3}\right) \delta 10.30(\mathrm{bs}, 1 \mathrm{H}), 7.08$ (d, $J=540 \mathrm{~Hz}, 1 \mathrm{H}), 5.09(\mathrm{t}, J=1.5 \mathrm{~Hz}, 1 \mathrm{H}) ; 1.95-2.0(\mathrm{~m}, 2 \mathrm{H}), 1.55-1.80(\mathrm{~m}, 8 \mathrm{H}), 1.30-1.45$ $(\mathrm{m}, 6 \mathrm{H}) ;{ }^{13} \mathrm{C}$ NMR $\left(\mathrm{CDCl}_{3}\right) \delta 131.8,124.6,30.3\left(\mathrm{~d}, J_{\mathrm{PCCC}}=16 \mathrm{~Hz}\right), 29.5\left(\mathrm{~d}, J_{\mathrm{PC}}=94 \mathrm{~Hz}\right), 29.6$, 27.9, 25.9, $20.8\left(\mathrm{~d}, J_{\mathrm{PCC}}=3 \mathrm{~Hz}\right), 17.9 ;{ }^{31} \mathrm{P} \operatorname{NMR}\left(\mathrm{CDCl}_{3}\right) \delta 38.9\left(\mathrm{dm}, J_{\mathrm{P}-\mathrm{H}}=540 \mathrm{~Hz}\right)$.

1-Phenylethylphosphinic acid (Table 2, entry 6). Reference: Buckler, S. A.; Epstein, M. Tetrahedron 1962, 18, 1211. ${ }^{1} \mathrm{H}$ NMR $\left(\mathrm{CDCl}_{3}\right)$ d $9.08(\mathrm{bs}, 1 \mathrm{H}), 7.20-7.35(\mathrm{~m}, 5 \mathrm{H}), 6.90(\mathrm{~d}, J=$ $552 \mathrm{~Hz}, 1 \mathrm{H}), 3.10-3.20(\mathrm{~m}, 1 \mathrm{H}), 1.50-1.60(\mathrm{dd}, J=19,7 \mathrm{~Hz}, 3 \mathrm{H}) ;{ }^{31} \mathrm{P}$ NMR $\left(\mathrm{CDCl}_{3}\right) \mathrm{d} 41.2$ $\left(\mathrm{dm}, J_{\mathrm{P}-\mathrm{H}}=552 \mathrm{~Hz}\right)$.
2-Phenylethylphosphinic acid (Table 2, entry 6). References: (a) Arbuzova, S. N.; Gusarova, N. K.; Malysheva, S. F.; Brandsma, L.; Albanov, A. I.; Trofimov, B. A. Russ. J. Gen. Chem. 1996, 66, 54; (b) Jackson, P. F.; Tays, K. L.; Maclin, K. M.; Ko, Y.-S.; Li, W.; Vitharana, D.; Tsukamoto, T.; Stoermer, D.; Lu, X.-C. M.; Wozniak, K.; Slusher, B. S. J. Med. Chem. 2001, 44, 4170; (c) Lui, X.; Hu, E.; Tian, X.; Mazur, A.; Ebetino, F. H. J. Organomet. Chem. 2002, 646, 212; (d) Gusarova, N. K.; Shaikhudinova, S. I.; Sukhov, B. G.; Kazantseva, T. I.; Malysheva, S. F.; Smetannikov, Y. V.; Tarasova, N. P.; Kuimov, V. A.; Trofimov, B. A. Russ. Chem. Bull. 2003, 52, 511. ${ }^{1} \mathrm{H}$ NMR $\left(\mathrm{CDCl}_{3}\right)$ d $9.08(\mathrm{bs}, 1 \mathrm{H}), 7.20-7.35(\mathrm{~m}, 5 \mathrm{H}), 7.06(\mathrm{~d}, J=548 \mathrm{~Hz}, 1 \mathrm{H})$, 2.85-2.95 (m, 2 H ), 2.05-2.15 (m, 2H); ${ }^{31} \mathrm{P}$ NMR $\left(\mathrm{CDCl}_{3}\right) \mathrm{d} 37.6\left(\mathrm{dm}, J_{\mathrm{P}-\mathrm{H}}=549 \mathrm{~Hz}\right)$.
(1-Cyclohexyl) methylphosphinic acid (Table 2, entry 7). ${ }^{1} \mathrm{H}$ NMR $\left(\mathrm{CDCl}_{3}\right) \delta 10.65$ (bs, 1H), $7.18(\mathrm{~d}, J=540 \mathrm{~Hz}, 1 \mathrm{H}), 1.60-1.75(\mathrm{~m}, 7 \mathrm{H}), 1.40-1(\mathrm{~m}, 6 \mathrm{H}) ;{ }^{13} \mathrm{C}$ NMR $\left(\mathrm{CDCl}_{3}\right) \delta 37.0,36.7$, 34.5, 34.4, 31.7, 26.0, 25.9; ${ }^{31} \mathrm{P}$ NMR $\left(\mathrm{CDCl}_{3}\right) \delta 37.9\left(\mathrm{dm}, J_{\mathrm{P}-\mathrm{H}}=540 \mathrm{~Hz}\right)$.

## [2-(4-Chloro-phenyl)-propyl]-phosphinic acid (Table 2, entry 8).

${ }^{1} \mathrm{H}$ NMR $\left(\mathrm{CDCl}_{3}\right) \delta 10.0(\mathrm{bs}, 1 \mathrm{H}), 7.1-7.4(\mathrm{~m}, 4 \mathrm{H}), 6.85(\mathrm{~d}, J=550 \mathrm{~Hz}, 1 \mathrm{H}), 3.15-3.25(\mathrm{~m}, 1 \mathrm{H})$, 2.0-2.1 (m, 2 H$), \quad 1.34(\mathrm{t}, \quad J=6 \mathrm{~Hz}, \quad 3 \mathrm{H}) ;{ }^{13} \mathrm{C}$ NMR $\left(\mathrm{CDCl}_{3}\right)$ $\delta 143.9,143.8,132.4,128.8,128.3,128.07 .6\left(\mathrm{~d}, J_{\mathrm{PC}}=94 \mathrm{~Hz}\right), 33.3,23.6\left(\mathrm{~d}, J_{\mathrm{PCC}}=10 \mathrm{~Hz}\right)$, 14.0; ${ }^{31}$ P NMR $\left(\mathrm{CDCl}_{3}\right) \delta 39.7\left(\mathrm{dm}, J_{\text {P-H }}=550 \mathrm{~Hz}\right)$.

2-Phthalimidylethylphosphinic acid (Table 2, entry 9). ${ }^{1} \mathrm{H}$ NMR (DMSO) $\delta$ 7.80-7.85 (m, $4 \mathrm{H}), 7.0(\mathrm{~d}, J=531 \mathrm{~Hz}, 1 \mathrm{H}), 3.70-3.80(\mathrm{~m}, 2 \mathrm{H}), 1.95-2.05(\mathrm{~m}, 2 \mathrm{H}) ;{ }^{13} \mathrm{C}$ NMR (DMSO) $\delta 168.3,135.0,132.4,123.7,31.4,29.5\left(\mathrm{~d}, \mathrm{~J}_{\mathrm{PC}}=89 \mathrm{~Hz}\right) ;{ }^{31} \mathrm{P}$ NMR $(\mathrm{DMSO}) \delta 23.3\left(\mathrm{dm}, J_{\mathrm{P}-\mathrm{H}}=\right.$ 532 Hz ).
(1-Phenyl-vinyl)-phosphinic acid (Table 2, entry 10). Reference: Dumond, Y. R.; Montchamp, J.-L. J. Organomet. Chem. 2002, 653, 252. ${ }^{1} \mathrm{H}$ NMR $\left(\mathrm{CDCl}_{3}, \mathrm{SiMe}_{4}\right) \delta 12.37$ (br, 1 H), 7.4-7.5 (m, 2 H ), 7.2-7.4 (m, 3 H$), 7.30(\mathrm{~d}, J=571 \mathrm{~Hz}, 1 \mathrm{H}), 6.16(\mathrm{~d}, J=46 \mathrm{~Hz}, 1 \mathrm{H}), 6.14$ $(\mathrm{d}, J=25 \mathrm{~Hz}, 1 \mathrm{H}) ;{ }^{13} \mathrm{C}$ NMR $\left(\mathrm{CDCl}_{3}\right) \delta 142.3\left(\mathrm{~d}, J_{\mathrm{PC}}=124 \mathrm{~Hz}\right), 135.1\left(\mathrm{~d}, J_{\mathrm{PCC}}=13 \mathrm{~Hz}\right), 129.3$ $\left(\mathrm{d}, J_{\mathrm{PCC}}=13 \mathrm{~Hz}\right), 128.7(2 \mathrm{C}), 128.6,127.3\left(\mathrm{~d}, J_{\mathrm{PCCC}}=6 \mathrm{~Hz}, 2 \mathrm{C}\right) ;{ }^{31} \mathrm{P}$ NMR $\left(\mathrm{CDCl}_{3}\right) \delta 25.7(\mathrm{ddd}$, $J=571 \mathrm{~Hz}, J=46 \mathrm{~Hz}, J=24 \mathrm{~Hz}$ ).

Trans-Oct-1-enyl-phosphinic acid (Table 2, entry 11). References: (a) Dumond, Y. R.; Montchamp, J.-L. J. Organomet. Chem. 2002, 653, 252; (b) Nifant'ev, E. E.; Magdeeva, R. K.; Maslennikova, V. I.; Taber, A. M.; Kalechits, I. V. J. Gen. Chem. USSR 1982, 52, 2173; (c) Belakhov, V. V.; Yudelevich, V. I.; Fetter, A. P.; Ionin, B. I. J. Gen. Chem. USSR 1986, 56, 680. ${ }^{1} \mathrm{H}$ NMR $\left(\mathrm{CDCl}_{3}, \mathrm{SiMe}_{4}\right) \delta 10.64(\mathrm{br}, 1 \mathrm{H}), 7.2(\mathrm{~d}, J=562 \mathrm{~Hz}, 1 \mathrm{H}), 6.6-6.8(\mathrm{~m}, 1 \mathrm{H}), 5.82(\mathrm{dd}, J$ $=17 \mathrm{~Hz}, J=24 \mathrm{~Hz}, 1 \mathrm{H}), 2.2-2.3(\mathrm{~m}, 2 \mathrm{H}), 1.4-1.5(\mathrm{~m}, 2 \mathrm{H}), 1.2-1.4(\mathrm{~m}, 6 \mathrm{H}), 0.88(\mathrm{~d}, J=6 \mathrm{~Hz}$, $3 \mathrm{H}) ;{ }^{13} \mathrm{C}$ NMR $\left(\mathrm{CDCl}_{3}\right) \delta 153.7\left(\mathrm{~d}, J_{\mathrm{PCC}}=6 \mathrm{~Hz}\right), 120.5\left(\mathrm{~d}, J_{\mathrm{PC}}=134 \mathrm{~Hz}\right), 34.1\left(\mathrm{~d}, J_{\mathrm{PCCC}}=20 \mathrm{~Hz}\right)$, 31.5, 28.7, 27.6, 22.5, 14.0, ${ }^{31} \mathrm{P}$ NMR $\left(\mathrm{CDCl}_{3}\right) 23.9(\mathrm{~d}, J=562 \mathrm{~Hz})$.
(1-Hexyl-vinyl)-phosphinic acid (Table 2, entry 11). Reference: Dumond, Y. R.; Montchamp, J.-L. J. Organomet. Chem. 2002, 653, 252. ${ }^{1} \mathrm{H}$ NMR $\left(\mathrm{CDCl}_{3}, \mathrm{SiMe}_{4}\right) \delta 12.78$ (br, 1 H$), 7.15(\mathrm{~d}, \mathrm{~J}$ $=557 \mathrm{~Hz}, 1 \mathrm{H}), 5.92(\mathrm{~d}, J=25 \mathrm{~Hz}, 1 \mathrm{H}), 5.79(\mathrm{~d}, J=49 \mathrm{~Hz}, 1 \mathrm{H}), 2.30(\mathrm{~m}, 2 \mathrm{H}), 1.5-1.6(\mathrm{~m}, 2$ H), 1.2-1.4 (m, 6 H$), 0.88(\mathrm{t}, J=7 \mathrm{~Hz}, 3 \mathrm{H}) ;{ }^{13} \mathrm{C}$ NMR $\left(\mathrm{CDCl}_{3}\right) \delta 142.6\left(\mathrm{~d}, J_{\mathrm{PC}}=121 \mathrm{~Hz}\right), 127.4$ $\left(\mathrm{d}, J_{\mathrm{PCC}}=14 \mathrm{~Hz}\right), 31.46,30.5\left(\mathrm{~d}, J_{\mathrm{PCC}}=13 \mathrm{~Hz}\right), 28.7,27.7\left(\mathrm{~d}, J_{\mathrm{PCCC}}=5 \mathrm{~Hz}\right), 22.5,13.9 ;{ }^{31} \mathrm{P} \mathrm{NMR}$ $\left(\mathrm{CDCl}_{3}\right) \delta 27.4(\mathrm{dm}, J=557 \mathrm{~Hz})$.
(1-Butyl-hex-1-enyl) phosphinic acid (Table 2, entry 12). ${ }^{1} \mathrm{H}$ NMR $\left(\mathrm{CDCl}_{3}\right) \delta 12.77$ (bs, 1 H ), $7.10(\mathrm{~d}, J=550 \mathrm{~Hz}, 1 \mathrm{H}), 6.32(\mathrm{dt}, J=27 \mathrm{~Hz}, J=7 \mathrm{~Hz}, 1 \mathrm{H}), 2.15-2.3(\mathrm{~m}, 4 \mathrm{H}), 1.25-1.50(\mathrm{~m}, 8$ $\mathrm{H}), 0.89(\mathrm{~d}, J=7 \mathrm{~Hz}, 6 \mathrm{H}) ;{ }^{13} \mathrm{C} \operatorname{NMR}\left(\mathrm{CDCl}_{3}\right) \delta 145.3\left(\mathrm{~d}, J_{\mathrm{PCC}}=14 \mathrm{~Hz}\right), 132.8\left(\mathrm{~d}, J_{\mathrm{PC}}=127 \mathrm{~Hz}\right)$, $31.3,30.6,28.0,27.8,24.8\left(\mathrm{~d}, J_{\mathrm{PCCC}}=13 \mathrm{~Hz}\right), 22.7,22.3,13.7 ;{ }^{31} \mathrm{P} \operatorname{NMR}\left(\mathrm{CDCl}_{3}\right) 30.73(\mathrm{dm}, J=$ 550).

Preparation of supported-ligand 4. Dry toluene ( 15 mL ) was added to a mixture of polystyrene isocyanate resin ( $1.003 \mathrm{~g}, 2.4 \mathrm{mmol}$; Advanced Chemtech, 100-200 Mesh, 2.4 $\mathrm{mmol} / \mathrm{g}$ ), and 4,6-bis(diphenylphosphino)phenoxazine 2 ( 0.653 g , 1.185 mmol ; Strem, nixantphos). The suspension was stirred and heated at $115{ }^{\circ} \mathrm{C}$ overnight, under $\mathrm{N}_{2}$. To the resulting mixture was added anhydrous diisopropylamine ( $0.5 \mathrm{ml}, 3.56 \mathrm{mmol}$ ) in order to neutralize the unreacted isocyanate groups. The reaction mixture was stirred at room
temperature for 6 hours, under $\mathrm{N}_{2}$. After filtration, the collected resin ( 1.2 g ) was rinsed with dichloromethane and dried overnight under vacuum, over $\mathrm{P}_{2} \mathrm{O}_{5}$; while the filtrate was evaporated to recover unreacted nixantphos as an off-white powder. This amount was taken into account to calculate the loading of the resin. The supported ligand's loading was found to be $0.14 \mathrm{mmol} / \mathrm{g}$.

Representative hydrophosphinylation procedure with 50 \% aqueous $\mathbf{H}_{3} \mathbf{P O}_{2}$. Aqueous hypophosphorous acid ( $50 \mathrm{wt} . \%$ ) was first weighted in a round bottom flask, ( $0.2723 \mathrm{~g}, 2.06$ $\mathrm{mmol})$. Acetonitrile ( 2 mL ), was then added to the flask. Addition of 4-phenylbutene ( 0.15 mL , $0.99 \mathrm{mmol}), \mathrm{Pd}_{2} \mathrm{dba}_{3}(4.6 \mathrm{mg}, 1 \mathrm{~mol} \%)$, and supported nixantphos, $(0.14 \mathrm{mmol} / \mathrm{g}, 64.8 \mathrm{mg}, 1.08$ $\mathrm{mol} \%$ ) followed. In order to avoid any potential leak of solvent, the neck of the flask was washed with another portion of acetonitrile ( 2 mL ). The resulting mixture was stirred and refluxed overnight under nitrogen. After measuring the crude yield by ${ }^{31} \mathrm{P}$ NMR, the reaction mixture was cooled down to room temperature and then filtered to collect the resin. The filtrate was then extracted with hexane, once. The acetonitrile layer was concentrated down and the residue underwent a single extraction with ethyl acetate and brine. The organic layer was dried over $\mathrm{MgSO}_{4}$. After filtration and concentration the resulting yellow oil was dried overnight under vacuum, over $\mathrm{P}_{2} \mathrm{O}_{5}$. The reaction yielded 4-phenylbutylphosphinic acid ( $0.193 \mathrm{~g}, 97 \%$ ) as a yellow oil.

Hydrophosphinylation procedure with concentrated $\mathbf{H}_{\mathbf{3}} \mathbf{P O}_{\mathbf{2}}$. These runs were conducted as above with the 50 wt . \% solution except the solution was first concentrated in vacuo at room temperature for 20-30 min, prior to the addition of the substrate and catalyst.

Reaction in air and at room temperature. To a solution of $50 \%$ aq $\mathrm{H}_{3} \mathrm{PO}_{2}(0.211 \mathrm{~g}, 1.60$ mmol ) was added some HPLC grade $\mathrm{CH}_{3} \mathrm{CN}$, 4-phenylbutene ( $0.15 \mathrm{ml}, 0.99 \mathrm{mmol}$ ), $\mathrm{Pd}_{2} \mathrm{dba}_{3}$ ( 5 $\mathrm{mg}, 1.1 \mathrm{~mol} \%)$, and supported nixantphos ( $0.33 \mathrm{mmol} / \mathrm{g}, 33.9 \mathrm{mg}, 1.1 \mathrm{~mol} \%$ ). The reaction was stirred at room temperature in an open flask for 6 hours. After taking a ${ }^{31} \mathrm{P}$ NMR of the crude mixture, the yield of the desired phosphinic acid was $78 \%$. The corresponding isolated yield was $51 \%$ (after extraction as above with brine and ethyl acetate).

Reaction in air and at reflux. Aqueous hypophosphorous acid ( $50 \mathrm{wt} . \%, 0.356 \mathrm{~g}, 2.69 \mathrm{mmol}$ ) and $\mathrm{CH}_{3} \mathrm{CN}(10 \mathrm{~mL})$ were added to 1 -octene $(0.25 \mathrm{ml}, 1.59 \mathrm{mmol})$, along with some $\mathrm{Pd}_{2} \mathrm{dba}_{3}(8$ $\mathrm{mg}, 1.1 \mathrm{~mol} \%$ ) and supported nixantphos ( $0.33 \mathrm{mmol} / \mathrm{g}, 67 \mathrm{mg}, 1.4 \mathrm{~mol} \%$ ). The reaction was refluxed overnight open to air. After measuring the yield by ${ }^{31} \mathrm{P}$ NMR, concentration and partition between ethyl acetate and brine of the corresponding crude phosphinic acid was accomplished. Drying the residue overnight under vacuum over $\mathrm{P}_{2} \mathrm{O}_{5}$ gave the product $(0.188 \mathrm{~g}$, 89\%) as yellow oil.

Multi run reactions (Table 2, entry 6). To aqueous hypophosphorous acid (50 wt. \%, 0.490 g , 4 mmol ) was added styrene $(0.30 \mathrm{ml}, 2.62 \mathrm{mmol}), \mathrm{Pd}_{2} \mathrm{dba}_{3}(11.6 \mathrm{mg}, 0.96 \mathrm{~mol} \%)$ and supported Nixantphos ( $0.33 \mathrm{mmol} / \mathrm{g}, 95.6 \mathrm{mg}, 1.2 \mathrm{~mol} \%$ ). The reaction mixture was refluxed overnight under $\mathrm{N}_{2}$. The next day, after measuring the crude yield by ${ }^{31} \mathrm{P}$ NMR, the mixture was filtered in order to separate the resin from the filtrate. The resin was washed with $\mathrm{CH}_{3} \mathrm{CN}$ and kept in a dessicator, over $\mathrm{P}_{2} \mathrm{O}_{5}$, between the runs. The filtrate was kept at in a stoppered Erlenmeyer. The resin was then used in the next run without any further precautions. Aqueous hypophosphorous $\operatorname{acid}(50 \mathrm{wt} . \%, 0.490 \mathrm{~g}, 4 \mathrm{mmol})$, styrene $(0.3 \mathrm{ml}, 2.62 \mathrm{mmol})$ and the previously collected resin
were mixed together in $\mathrm{CH}_{3} \mathrm{CN}(10 \mathrm{~mL})$. The reaction mixture was again refluxed overnight under $\mathrm{N}_{2}$. The next day, after measuring the crude yield by ${ }^{31} \mathrm{P}$ NMR, the mixture was filtered in order to separate the resin and the filtrate. The resin was washed with $\mathrm{CH}_{3} \mathrm{CN}$, and collected for another run. The filtrate was kept in the same Erlenmeyer as before. For the third time, the reaction was run, with $50 \%$ aq $\mathrm{H}_{3} \mathrm{PO}_{2}(0.390 \mathrm{~g}, 3.17 \mathrm{mmol})$, styrene $(0.30 \mathrm{ml}, 2.62 \mathrm{mmol})$ and the previously collected resin. The same procedures were used to run the reaction, control the NMR yield and separate the resin from the desired product. The experiment was then run for one last time: $50 \%$ aq $\mathrm{H}_{3} \mathrm{PO}_{2}(0.499 \mathrm{~g}, 4 \mathrm{mmol})$, styrene $(0.30 \mathrm{ml}, 2.62 \mathrm{mmol})$ and the previously collected resin. At the end of the sequence, all the filtrates were in a single Erlenmeyer. After concentration, partition between EtOAc and brine, the product ( $2.00 \mathrm{~g}, 90 \%$ over 4 runs) was obtained as a yellow oil and a mixture of regioisomers.



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