

Supplementary Material

Protein Preparation, Crystallization, Data collection and Structure Resolution

Human estrogen receptor alpha-ligand binding domain (ER α -LBD, Ser301 to Thr553) with three Cys to Ser substitutions (C381S, C417S, C530S) was cloned and expressed in *Escherichia coli*. Protein purification was done as described previously.¹ The ER α -LBD in 50 mM Tris-HCl buffer (pH 8.0,) 50 mM NaCl, 2 mM DTT (1,4-Dithio-DL-threitol) and 2 μ M **29**, was concentrated to 10 mg/mL prior to crystallization.

Crystals were grown in a hanging drop setup (Linbro plate) at room temperature (20 °C). The reservoir buffer contained 100 mM Bicine buffer (pH = 9.0), 28% PEG-550 MME and 100 mM NaCl. The hanging drop was made by mixing 1 μ L protein solution with 1 μ L reservoir solution.

Crystals were cryoprotected by adding 1 μ L reservoir solution to the hanging drop (30 s equilibration time), then mounted in a cryo-loop and frozen directly at 100 K in the nitrogen stream. The first diffraction data set ("data set I", c.f. Table 1) was collected to 2.05 Å from a single frozen crystal on a MAR345 imaging plate system (150 μ m pixel size, 240 mm plate) at the Swiss Norwegian Beam line of the ESRF (Grenoble, F). 215 Images were collected with 1.0° rotation each, using an exposure time of 30 s per frame and a crystal-to-detector distance of 235 mm. Because of incompleteness (due to overloads) in the low resolution range of data set I, a second data set ("data set II", c.f. Table 1) using a second crystal was collected in house to 2.4 Å on an EnrafNonius FR591 rotating Cu-anode generator (operating at 45 kV, 90 mA) equipped with a MacScience DIP2030 imageplate system. A total of 326 diffraction

images were recorded at 100 K with an exposure time of 2000 s for 1.0° rotation per image at a crystal-to-detector distance of 195 mm. The two data sets were processed and subsequently merged with the HKL program suite version 1.6.1² and structure factor amplitudes were generated with the CCP4 4.0-package^{3,4}. Data processing and merging statistics are shown in Table 1. The space group is P2₁ with unit cell parameters $a = 55.1 \text{ \AA}$, $b = 156.6 \text{ \AA}$, $c = 59.0 \text{ \AA}$ and $\beta = 90.4^\circ$. There are four complexes of ER α LBD/**29** (forming two dimers) per asymmetric unit. The estimated B-factor by Wilson plot is 31.9 \AA^2 .

Table 1. Data processing and merging statistics for data sets I+II

X-ray source (data set I)	SNBL Grenoble ($\lambda = 0.8727\text{\AA}$)
X-ray source (data set II)	CuK α ($\lambda = 1.5418\text{\AA}$)
Temperature of data collection	100K
Number of crystals	2
Space group	P2 ₁
Unit cell dimensions	a = 55.1 \AA , b = 156.6 \AA , c = 59.0 \AA , $\beta = 90.4^\circ$
Number of monomers / asymmetric unit	4
Matthews coefficient (Vm)	2.2 $\text{\AA}^3/\text{Da}$
Solvent content	44 %
Overall B-factor from Wilson plot	31.9 \AA^2
Resolution range (last shell)	10.0 - 2.05 \AA (2.12 - 2.05 \AA)
Number of observations	403555
Number of unique reflection	59332
Mosaicity	0.61 $^\circ$
Data redundancy (last shell)	6.8 (4.9)
Data completeness (last shell)	95.2 % (89.9 %)
$\langle I / \sigma(I) \rangle$ (last shell)	22.3 (1.8)
R _{merge} on I (last shell)	9.5 % (42.2 %)

The structure was determined by molecular replacement using the coordinates of the ligand binding domain (LBD) of human ER α (PDB accession code 1UOM¹) as a starting model. Molecular replacement was performed with MOLREP⁴, using all measured data between 10 and 3 \AA . A clear solution was found with a correlation-coefficient of 54.3%, an R-factor of 46.5%, having 4 molecules in the asymmetric unit. The program REFMAC version 5.0⁵ was used for refinement. Bulk solvent

correction, an initial anisotropic B factor correction and restrained isotropic atomic B-factor refinement were applied. The refinement target was the maximum-likelihood target using amplitudes. No sigma cut-off was applied on the structure factor amplitudes and NCS-restraints were not applied. Cross-validation was used throughout refinement using a test set comprising 5.1% (2992) of the unique reflections. Water molecules were identified with the program ARP/wARP⁶ and selected based on difference peak height (greater than 3.0σ) and distance criteria. Water molecules with temperature factors greater than 70\AA^2 were rejected. The program O version 7.0⁷ was used for model rebuilding and the quality of the final refined model (Table 2) was assessed with the programs PROCHECK version 3.3⁸ and REFMAC version 5.0⁶. The final model contains residues 307-330, 341-461, 464-528, 535-548 for chains A, C and residues 307-330, 341-461, 464-525, 535-548 for chains B, D. For the superposition (using the lsq-option in O⁷) of the complex ER α -LBD/29 (chain identifiers A/L) with ER α -LBD/raloxifene (PDB accession code 1ERR⁹), the C α coordinates of the respective residues 307-330, 341-419, 421-459, 470-525 and 535-546 were used, yielding a root mean square deviation (rmsd)-value of 0.56 Å. A superposition with the complex of ER α -LBD containing estradiol (PDB accession code 1QKT¹⁰) using the same C α coordinates yielded an rmsd-value of 0.66 Å. A superposition with the complex of ER α -LBD containing estradiol (PDB accession code 1UOM¹⁰) using the same C α coordinates yielded an rmsd-value of 0.69 Å. Pictures were made with O⁷.

Structure factors and coordinates for the refined structure were deposited in the protein structure database (PDB ID code: 1XQC).

Table 2: Refinement statistics

<i>Data used in refinement:</i>	
- resolution range (last shell)	10.0 - 2.05 Å (2.10 - 2.05 Å)
- intensity cutoff (Sigma(F))	None
- number of reflections	55974
- completeness [working + test set] (last shell)	95.3 % (89.9 %)
- test set	5.1 % (2992 reflections)
<i>Fit to data used in refinement:</i>	
- overall R_{cryst} (last shell)	21.1 % (32.4 %)
- overall R_{free} (last shell)	25.4 % (35.2 %)
<i>Number of non-hydrogen atoms used in the refinement:</i>	
- protein atoms (total of chains A, B, C, D)	7116
- ligand atoms (total of chains L, M, N, O)	132
- water molecules (chain W)	374
Mean temperature factors for protein chains A, B, C, D	43.2 Å ² , 46.6 Å ² , 45.5 Å ² , 44.4 Å ²
Mean temperature factors for ligand chains L, M, N, O	33.2 Å ² , 35.1 Å ² , 32.6 Å ² , 36.6 Å ²
Mean temperature factors for water molecules chain W	47.0 Å ²
<i>Deviations from ideal geometry:</i>	
- bond length	0.018 Å
- bond angles	1.55°
- esu based on R-value	0.245 Å
- esu based on maximum likelihood	0.169 Å
- Ramachandran plot outliers	none
- G factor from PROCHECK	0.10

Acknowledgement:

The support of R.Cebe, M. Mahnke, E.Koch and P. Graff in cloning, expression, purification and mass spectrometry analysis of ER α -LBD is gratefully acknowledged. Experimental assistance from the staff of the Swiss-Norwegian Beam Line at the European Synchrotron Radiation Facilities in Grenoble is gratefully acknowledged.

References:

- (1) Renaud, J.; Bischoff, S. F.; Buhl, T.; Floersheim, P.; Fournier, B.; Halleux, C.; Kallen, J.; Keller, H.; Schlaeppli, J.-M.; Stark, W. *J. Med. Chem.* **2003**, *46*, 2945-2957.
- (2) Otwinowski, Z.; Minor, W. Processing of X-ray Diffraction Data Collected in Oscillation Mode. In *Methods in Enzymology, Macromolecular Crystallography, part A*; Carter, C. W. Jr., Sweet, R. M., Eds., Academic Press: San Diego, 1997, Vol. 276; pp 307-326.
- (3) French, G. S.; Wilson, K. S. On the Treatment of Negative Intensity Observations. *Acta Crystallogr., Sect. A* **1978**, *34*, 517-525.
- (4) Collaborative Computational Project, Number 4. The CCP4 Suite: Programs for Protein Crystallography. *Acta Crystallogr., Sect. D* **1994**, *50*, 760-763.
- (5) Murshudov, G.N.; Vagin, A.A.; Dodson, E.J. Refinement of Macromolecular Structures by the Maximum-Likelihood Method. *Acta Crystallogr., Sect.D* **1997**, *53*, 240-255.
- (6) Perrakis, A.; Sixma, T.K.; Wilson, K.S.; Lamzin, V.S. wARP: Improvement and Extension of Crystallographic Phases by Weighted Averaging of Multiple-Refined Dummy Atomic Models. *Acta Crystallogr., Sect.D* **1997**, *53*, 448-455.

- (7) Kleywegt, G. J.; Jones, T. A. Model Building and Refinement Practice. In *Methods in Enzymology, Macromolecular Crystallography, part B*; Carter, C. W. Jr., Sweet, R. M., Eds., Academic Press: San Diego, 1997, Vol. 277; pp 208-230.
- (8) Laskowski, R.A.; MacArthur M.W.; Moss, D. S.; Thornton, J. M. PROCHECK: a Program to Check the Stereochemical Quality of Protein Structures. *J. Appl. Crystallogr.*, **1993**, *26*, 283-291.
- (9) Brzozowski, A. M.; Pike, A. C. W.; Dauter, Z.; Hubbard, R. E.; Bonn, T.; Engström, O.; Ohman, L.; Greene, G. L.; Gustafsson, J. Å.; Carlquist, M. Molecular Basis of Agonism and Antagonism in the Oestrogen Receptor. *Nature* **1997**, *389*, 753-758.
- (10) Gangloff, M.; Ruff, M.; Eiler, S.; Duclaud, S.; Wurtz, J.-M.; Moras, D. Crystal Structure of a Mutant hER α Ligand-Binding Domain Reveals Key Structural Features for the Mechanism of Partial Agonism. *J. Biol. Chem.* **2001**, *276*, 15059-15065.