

# Supporting Information

## Duplex and Quadruplex DNA Binding and Photocleavage by Trioxatriangulenium Ion<sup>†</sup>

*Arti Pothukuchy,<sup>1</sup> Carolyn L. Mazzitelli,<sup>2</sup> Mireya L. Rodriquez,<sup>1</sup> Bodin Tuesuwan,<sup>1</sup>*

*Miguel Salazar,<sup>1</sup> Jennifer S. Brodbelt<sup>2\*</sup> and Sean M. Kerwin<sup>1\*</sup>*

<sup>1</sup>Division of Medicinal Chemistry and Institute for Cellular and Molecular Biology, The University of Texas at Austin, Austin, TX 78712. <sup>2</sup>Department of Chemistry and Biochemistry, The University of Texas at Austin, Austin, TX 78712

[skerwin@mail.utexas.edu](mailto:skerwin@mail.utexas.edu)

<sup>†</sup> This work was support by grants from the Robert A. Welch Foundation (F-1298 to SMK and F-1155 to JSB) and the National Institutes of Health (RO1 GM65956 and ES 07784)

\*correspondence can be addressed to either author: Sean M. Kerwin (E-mail: [skerwin@mail.utexas.edu](mailto:skerwin@mail.utexas.edu); Tel: 512-471-5074; FAX: 512-232-2606) and Jennifer S. Brodbelt (E-mail: [jbrodbelt@mail.utexas.edu](mailto:jbrodbelt@mail.utexas.edu); Tel: 512-471-0028).

*DMS footprinting.* DMS footprinting reactions were carried out essentially as described (1S) with the exception that parallel reactions were carried out in the presence of 10  $\mu$ M TOTA. Briefly, reaction mixtures (20  $\mu$ L) containing 3  $\mu$ L 5'-labeled DNA substrates in the presence or absence of 10  $\mu$ M TOTA perchlorate at room temperature in 50 mM sodium phosphate buffer containing 10 or 100 mM KCl, pH 7 were treated with 1  $\mu$ L of a 20% DMS solution in water. After a period of 5 to 30 min, the reaction was quenched by the addition of 100  $\mu$ L stop buffer (1.0 M  $\beta$ -mercaptoethanol, 1.5 M sodium acetate and 1  $\mu$ g glycogen). The samples were precipitated by cold EtOH, and washed twice with 70 % EtOH. Samples were subjected to piperidine heat treatment (1 M piperidine, 90 °C, 15 min), dried in a vacuum concentrator, and subjected to electrophoresis on a 12% denaturing polyacrylamide gel.

## Reference

1S. Tuntiwechapikul, W., Lee, J. T., and Salazar, M. (2001) Design and synthesis of the G-quadruplex-specific cleavage reagent perylene-EDTA•Iron(II), *J. Am. Chem. Soc.* 123, 5606–5607.

## Figure Legends

**Figure 1S.** Phosphorimage of DMS footprinting of the 5'-labeled TAG strand of the duplex (D) and quadruplex (DQ) substrates in the presence and absence of 10  $\mu$ M TOTA. Each group of lanes corresponds to a time-course exposure to DMS for 5, 10, 20, and 30 min. Lanes 1: TAG•T6 quadruplex (DQ) in 100 mM KCl after heat treatment (90 °C, 5

min) and rapid annealing; Lanes 2: TAG•T6 quadruplex (DQ) in 100 mM KCl after annealing overnight; Lanes 3: TAG•T6 quadruplex (DQ) in 100 mM KCl in the presence of 10  $\mu$ M TOTA; Lane 4: TAG•T6 quadruplex (DQ) in 10 mM KCl; Lanes 5: TAG•T6 quadruplex (DQ) in 10 mM KCl in the presence of 10  $\mu$ M TOTA; Lanes 6: TAG•CTA duplex (D) in 100 mM KCl. Lanes G and GA: Maxam-Gilbert G and GA sequencing.

**Figure 2S.** Phosphorimage of DMS footprinting of the 5'-labeled TG strand of the duplex (D) and quadruplex (DQ) substrates in the presence and absence of 10  $\mu$ M TOTA. Each group of lanes corresponds to a time-course exposure to DMS for 5, 10, and 20 min. Lanes 1: TG•T6 quadruplex (DQ) in 100 mM KCl after heat treatment (90 °C, 5 min) and rapid annealing; Lanes 2: TG•T6 quadruplex (DQ) in 100 mM KCl after annealing overnight; Lanes 3: TG•T6 quadruplex (DQ) in 100 mM KCl in the presence of 10  $\mu$ M TOTA; Lane 4: TG•T6 quadruplex (DQ) in 10 mM KCl; Lanes 5: TG•T6 quadruplex (DQ) in 10 mM KCl in the presence of 10  $\mu$ M TOTA; Lanes 6: TG•CT duplex (D) in 100 mM KCl. Lanes G and GA: Maxam-Gilbert G and GA sequencing.

**Figure 3S.** Phosphorimage of the cleavage product for duplex (D) and quadruplex (DQ) substrates in the presence of 10  $\mu$ M TOTA. A. Photocleavage of 5'-labeled TAG duplex (lanes 6 and 12) and quadruplex (all other lanes) substrates: Lanes 2, 4–6, 8, and 10–12 were irradiated (350 nm, 30 min), all other lanes are dark controls; Lanes 3–6 and 9–12 are in the presence of 10  $\mu$ M TOTA; Lanes 7–12 were subjected to piperidine heat treatment prior to electrophoresis, Lanes 5 and 11 are in 10 mM KCl, all other lanes are in 100mM KCl phosphate buffer pH 7.0. B. Photocleavage of 5'-labeled TG duplex (lanes 6 and 12) and quadruplex (all other lanes) substrates: Lanes 2, 4–6, 8, and 10–12 were irradiated (350 nm, 30 min), all other lanes are dark controls; Lanes 3–6 and 9–12

are in the presence of 10  $\mu$ M TOTA; Lanes 7–12 were subjected to piperidine heat treatment prior to electrophoresis, Lanes 5 and 11 are in 10 mM KCl, all other lanes are in 100mM KCl phosphate buffer pH 7.0. Lanes G and GA: Maxam-Gilbert G and GA sequencing.

**Figure 4S:** Histogram of phosphorimage in Figure 3S-A. A. TOTA photocleavage of TAG•CTA duplex (D), lane 12; B. TOTA photocleavage of TAG•T6 quadruplex (DQ) in 10 mM KCl, lane 11; C. TOTA photocleavage of TAG•T6 quadruplex (DQ) in 100 mM KCl, lane 10.

**Figure 5S.** Histogram of phosphorimage in Figure 3S-B. A. TOTA photocleavage of TG•CA duplex (D), lane 12; B. TOTA photocleavage of TG•T6 quadruplex (DQ) in 10 mM KCl, lane 11; C. TOTA photocleavage of TG•T6 quadruplex (DQ) in 100 mM KCl, lane 10.

Figure 1S.

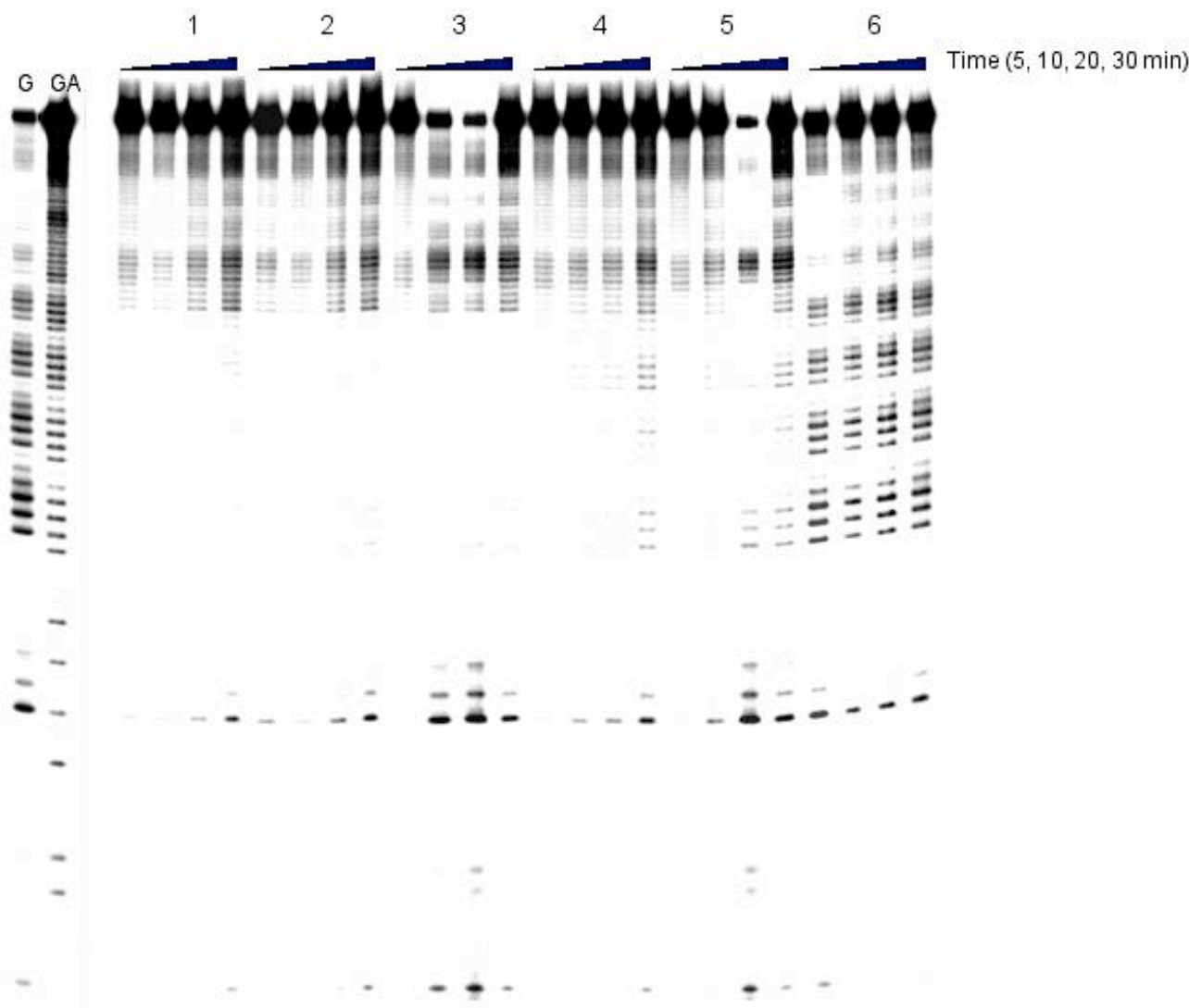


Figure 2S.

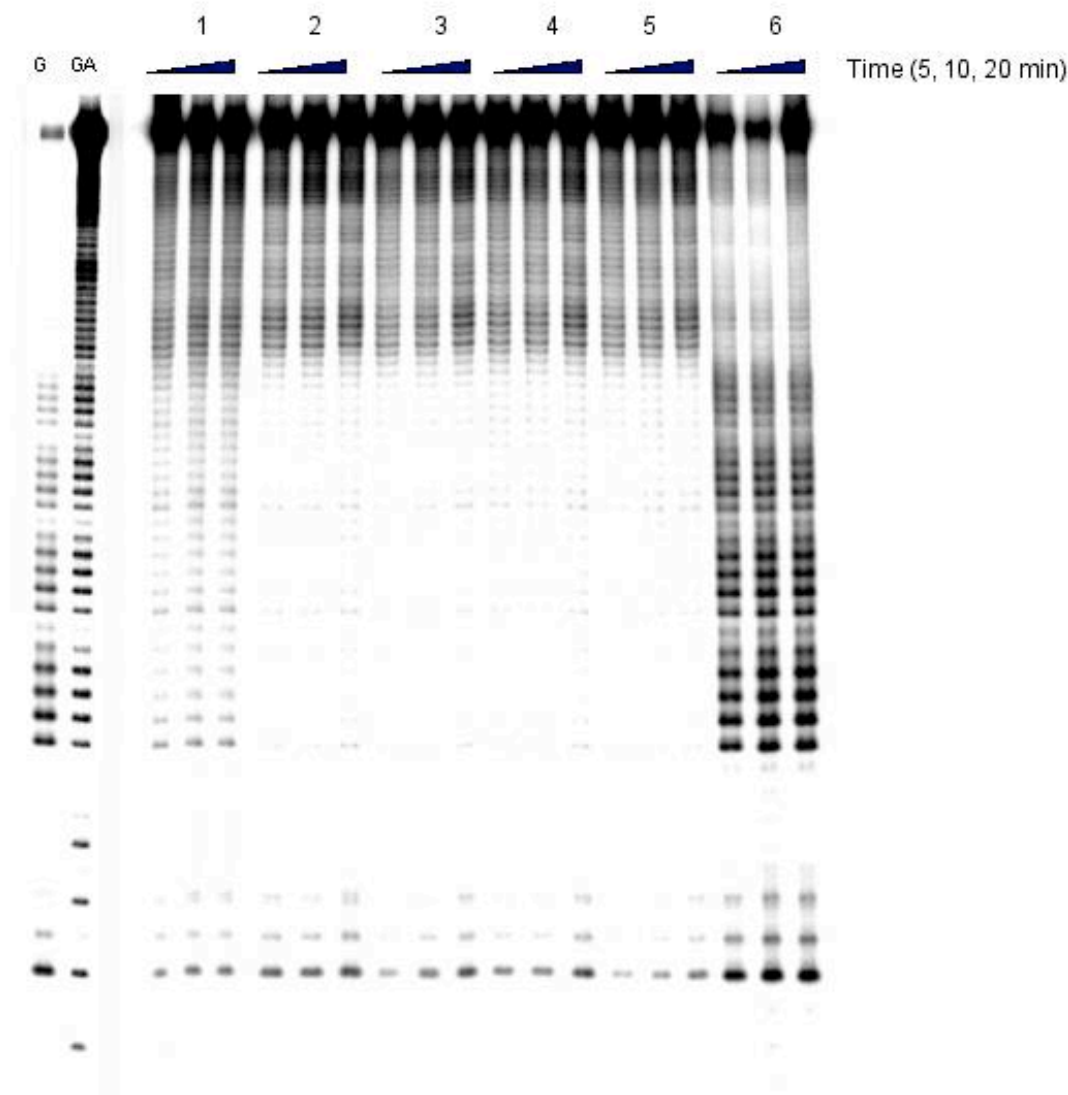


Figure 3S.

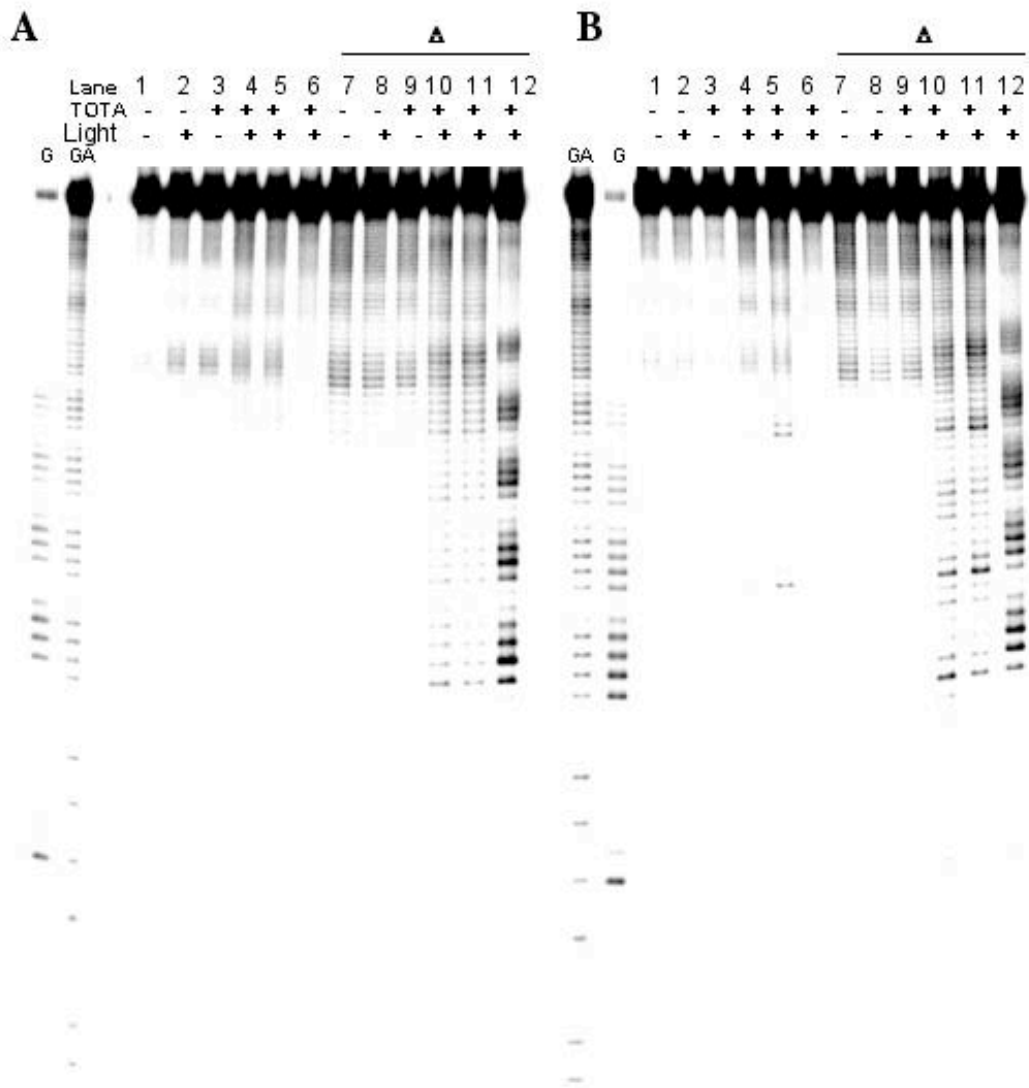


Figure 4S.

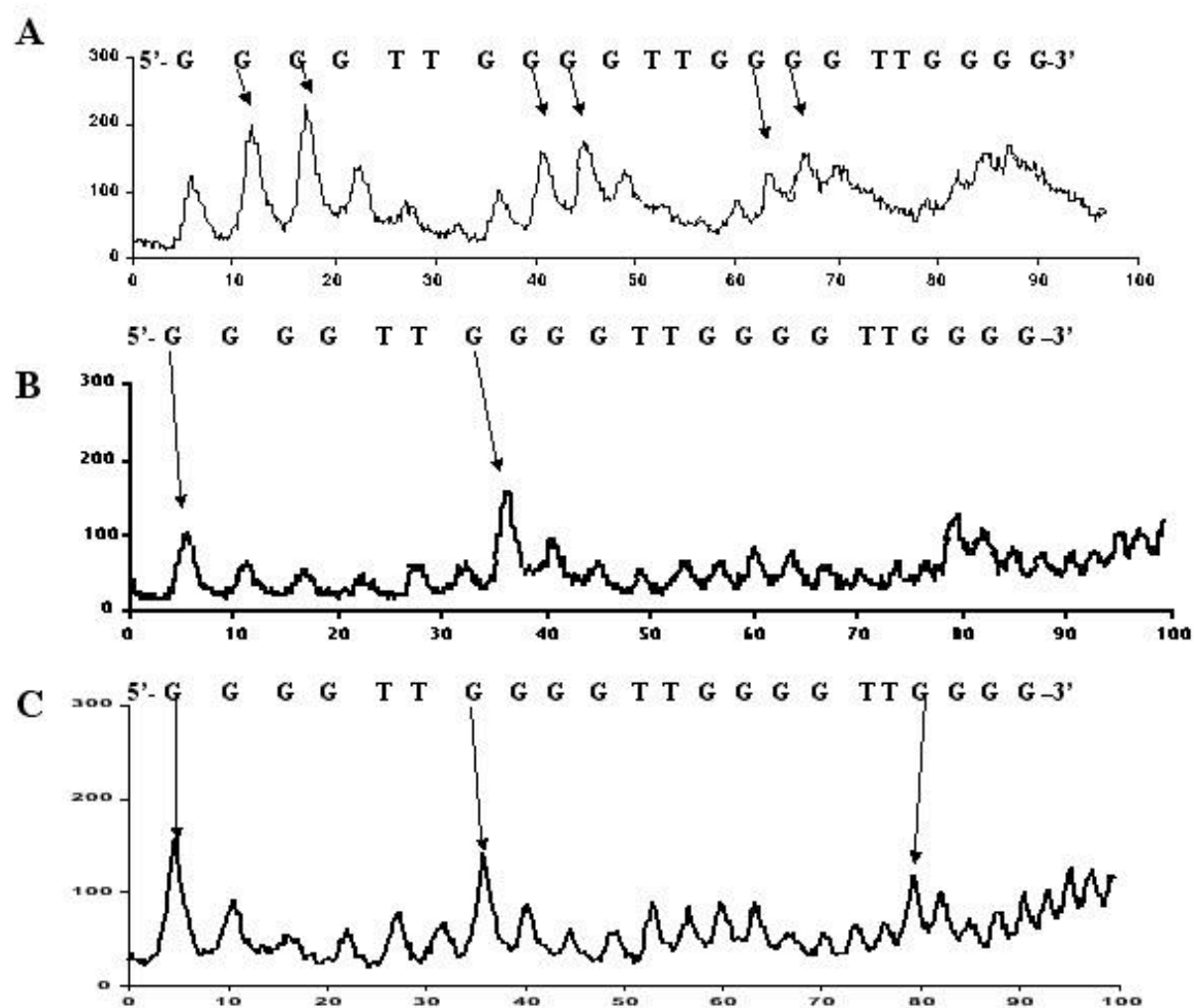




Figure 5S.

