

Supporting Information

CO-Trapping Reaction under Thermolysis of Alkoxyamines.

Application to the Synthesis of 3,4-Cyclopenta-1-Tetralones

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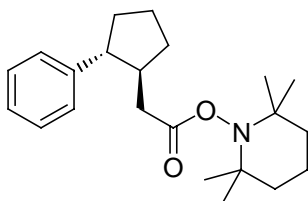
Corrensstrasse 40, 48149 Münster, Germany

General information. ¹H NMR spectra were recorded with a JEOL JMN ECP-500 (500 MHz) spectrometer in CDCl₃. Chemical shifts are reported in parts per million (δ) downfield from internal TMS at 0.00. ¹³C NMR spectra were recorded with a JEOL JMN ECP-500 (125 MHz) spectrometer and referenced to the solvent peak at 77.00 ppm. Infrared spectra were obtained on a JASCO FT/IR 4100 spectrometer; absorptions are reported in reciprocal centimeters. Both conventional and high resolution mass spectra were recorded with a JEOL MS700 spectrometer. GPC was performed on a JAI LC-908 chromatograph with JAIGEL-1H + JAIGEL-2H columns. Alkoxyamines **1a-1h** were prepared from the corresponding benzyl bromides and TEMPO (2,2,6,6-tetramethylpiperidine-1-oxyl) according the procedure described in reference 2. Alkoxyamine **1i** was prepared from 6-bromo-6-phenyl-1-hexene¹ and the corresponding nitroxide according to the similar procedure. Products were purified by flash chromatography on silica gel (Kanto Chemical Co., Inc., Silica Gel 60N, 70-230 mesh).

Experimental procedure for carbonylation of 1a (entry 1 of Table 1).

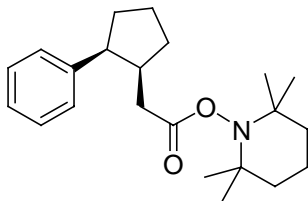
A magnetic stirring bar, 2,2,6,6-tetramethyl-1-(1-phenylhex-5-enoyloxy)piperidine **1a** (157.7 mg, 0.50 mmol) and ^tBuOH (50 mL) were placed in a 100 mL stainless steel autoclave. The autoclave was closed, purged three times with carbon monoxide, pressurized to 85 atm of CO and then heated at 130 °C. After 12 h, excess CO was discharged at room temperature. And the solvent was removed under reduced pressure. After adding ether (5 mL) and 1%-NaOHaq (5 mL) to the residue, the aqueous phase was then separated, washed twice with ether, and acidified with 2M-HCl. This solution was extracted with ether twice. The ether phase was washed with brine, dried over Na₂SO₄, and concentrated *in vacuo*, to give carboxylic acid **4a** as the product (14.2 mg, 14%), which was essentially pure by NMR that there was no need for further purification. On the other hand, the organic phase was washed with brine, dried over Na₂SO₄ and then filtrated. After evaporation, tetrachloroethane (0.121 mmol) was added to the residue as a standard and the yield of **2a** was determined to be 7% by NMR. After determining the NMR yield, the crude mixture was purified by column chromatography on silica gel (eluent: hexane/EtOAc = 10/1) to give **3a** (41.5 mg, 24%). The separation of the *cis*- and *trans*-isomers of **3a** was achieved using a preparative HPLC.

trans-1-Phenyl-2-(2,2,6,6-tetramethylpiperidin-1-yloxy-carbonylmethyl)cyclopentane (trans-3a)



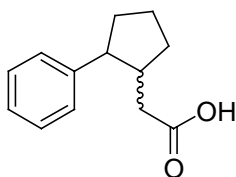
IR (neat) 1763 cm⁻¹; ¹H NMR (500 MHz, CDCl₃) δ 0.98 (s, 3.00H, CH₃), 1.01 (s, 3.00H, CH₃), 1.10 (s, 3.00H, CH₃), 1.11 (s, 3.00H, CH₃), 1.32-1.82 (m, 10.00H, CH₂), 2.04-2.22 (m, 3.00H, CH₂CO, CH₂), 2.32-2.41 (m, 1.00H, CHCH₂CO), 2.45 (dd, *J* = 15.12, 3.67 Hz, 1.00H, CH₂CO), 2.53-2.59 (m, 1.00H, PhCH), 7.19-7.32 (m, 5.00H, ArH); ¹³C NMR (125 MHz, CDCl₃) δ 17.03, 20.60, 23.79, 32.05, 32.32, 35.19, 37.17, 39.03, 44.30, 52.86, 59.96, 126.36, 127.64, 128.57, 144.02, 172.81; HRMS (EI) *m/z* calcd for C₂₂H₃₃NO₂ (M⁺) 343.2511, found 343.2513.

**cis-1-Phenyl-2-(2,2,6,6-tetramethylpiperidin-1-yloxy-carbonylmethyl)cyclopentane
(cis-3a)**



^1H NMR (500 MHz, CDCl_3) δ 0.97 (s, 3.00H, CH_3), 1.00 (s, 3.00H, CH_3), 1.08 (s, 3.00H, CH_3), 1.10 (s, 3.00H, CH_3), 1.35-2.03 (m, 13.00H, CH_2CO , CH_2), 2.05-2.22 (m, 1.00H, CH_2CO), 2.64-2.72 (m, 1.00H, CHCH_2CO), 3.30-3.45 (m, 1.00H, PhCH), 7.17-7.32 (m, 5.00H, ArH); ^{13}C NMR (125 MHz, CDCl_3) δ 17.03, 20.60, 23.55, 30.38, 31.45, 34.64, 39.04, 40.54, 44.30, 48.32, 59.97, 126.12, 128.27, 128.53, 142.94, 173.28.

2-Phenylcyclopentylacetic acid (4a)

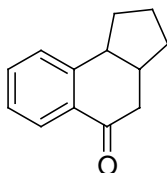


IR (neat) 1707, 2400-3600 cm^{-1} ; ^1H NMR (500 MHz, CDCl_3) δ 1.17-2.15 (m, 7.32H, CH_2CO , trans, CH_2CO , cis, CH_2), 2.20-2.32 (m, 0.68H, CHCH_2CO , trans), 2.41 (dd, J = 15.12, 4.12 Hz, 0.68H, CH_2CO , trans), 2.48-2.66 (m, 1.00H, PhCH , trans, CHCH_2CO , cis), 3.26-3.32 (m, 0.32H, PhCH , cis), 7.10-7.38 (m, 5.00H, ArH); ^{13}C NMR (125 MHz, CDCl_3) δ 23.50 (cis), 23.76 (trans), 30.05 (cis), 31.08 (cis), 32.16 (trans), 35.01 (trans), 35.88 (cis), 38.36 (trans), 40.42 (cis), 44.42 (trans), 48.18 (cis), 52.54 (trans), 126.24 (cis), 126.40 (trans), 127.61 (trans), 128.30 (cis), 128.45 (cis), 128.55 (trans), 142.59 (cis), 143.95 (trans), 179.64 (trans), 180.01 (cis); HRMS (EI) m/z calcd for $\text{C}_{13}\text{H}_{16}\text{O}_2$ (M^+) 204.1150, found 204.1146.

Typical Procedure for 3,4-Cyclopenta-1-tetralones 2. A magnetic stirring bar, 2,2,6,6-tetramethyl-1-(1-phenylhex-5-enoyloxy)piperidine **1a** (156.8 mg, 0.5 mmol) and $t\text{BuOH}$ (50 mL) were placed in a 100 mL stainless steel autoclave. The autoclave was closed, purged three times with carbon monoxide, pressurized to 75 atm of CO, then

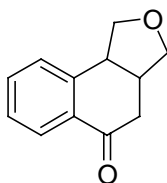
heated 130 °C (bath temp.) for 20 h. Excess CO was then discharged at room temperature. The solvent was removed under reduced pressure. CF₃SO₃H (3 mL) was added to the residue, followed by stirring for 2 h at 50°C (bath temp). After quenching with ice, the solution was extracted with ether three times, the organic layer was washed with brine and then dried over Na₂SO₄. After evaporation, the residue was purified by flash chromatography on silica gel (eluent: hexane/EtOAc = 30/1) to give **2a** (46.6 mg, 51%).

1,2,3,3a,4,9b-Hexahydro-cyclopenta[*a*]naphthalen-5-one (**2a**)



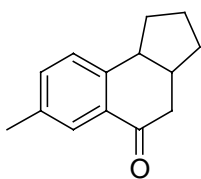
Obtained as a diastereomer mixture. mp 89-90 °C; IR (neat) 1682 cm⁻¹; ¹H NMR (500 MHz, CDCl₃) δ 1.20-2.08 (m, 5.65H, CH₂), 2.15-2.26 (m, 0.35H, CH₂, cis), 2.31-2.46 (m, 1.30H, CH₂CO, CHCH₂CO, trans), 2.53-2.80 (m, 1.70H, CH₂CO, cis, CHCH₂CO, cis, CHPh, trans), 3.00 (dd, *J* = 16.96, 3.67 Hz, 0.65H, CH₂CO, trans), 3.23-3.30 (m, 0.35H, CHPh, cis), 7.20-7.38 (m, 3.00H, ArH), 7.96 (d, *J* = 7.33 Hz, 0.35H, ArH, cis), 8.03 (d, *J* = 7.79 Hz, 0.65H, ArH, trans); ¹³C NMR (125 MHz, CDCl₃) δ 22.21 (trans), 23.75 (cis), 28.41 (trans), 31.00 (trans), 31.41 (cis), 33.18 (cis), 38.61 (cis), 40.85 (cis), 42.86 (cis), 44.24 (trans), 45.51 (trans), 46.59 (trans), 126.24 (trans), 126.37 (cis), 126.50 (trans), 126.66 (cis), 127.64 (trans), 129.23 (cis), 131.61 (cis), 132.33 (trans), 133.54 (trans), 133.74 (cis), 146.34 (cis), 146.96 (trans), 199.03 (cis), 199.22 (trans); HRMS (EI) *m/z* calcd for C₁₃H₁₄O (M⁺) 186.1045, found 186.1044;

1,3,3a,9b-Tetrahydronaphtho[1,2-*c*]furan-5(4H)-one (**2b**)³



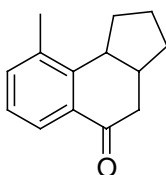
Obtained as a diastereomer mixture. IR (neat) 1686 cm^{-1} ; ^1H NMR (500 MHz, CDCl_3) δ 2.72-2.79 (m, 1.18H, CH_2CO , cis), 2.99-3.08 (m, 0.59H, CHCH_2CO , cis), 3.00-3.04 (m, 0.82H, CH_2CO , trans), 3.15-3.20 (m, 0.41H, CHCH_2CO , trans), 3.62-3.70 (m, 1.00H, PhCH), 3.72-3.92 (m, 2.00H, CH_2O), 4.10 (dd, $J = 8.71, 5.96$ Hz, 0.59 H, CH_2O , cis), 4.18 (t, $J = 7.33$ Hz, 0.41H, CH_2O , trans), 4.33 (t, $J = 8.25$ Hz, 0.59H, CH_2O , cis), 4.56 (t, $J = 7.33$ Hz, 0.41H, CH_2O , trans), 7.07 (d, $J = 7.79$ Hz, 0.41H, ArH , trans), 7.26 (d, $J = 7.79$ Hz, 0.59H, ArH , cis), 7.36 (t, $J = 7.79$ Hz, 0.59H, ArH , cis), 7.40 (d, $J = 7.79$ Hz, 0.41H, ArH , trans), 7.52-7.57 (m, 1.00H, ArH), 8.00 (d, $J = 7.79$ Hz, 0.59H, ArH , cis), 8.07 (d, $J = 7.79$, 0.41H, ArH , trans); ^{13}C NMR (125 MHz, CDCl_3) δ 38.47, 38.74, 41.39, 41.91, 43.40, 45.57, 70.06, 71.76, 73.39, 73.71, 125.74, 127.06, 127.36, 127.47, 128.21, 129.38, 131.60 (cis), 132.27 (trans), 133.84 (trans), 134.13 (cis), 141.63 (cis), 142.24 (trans), 197.16 (trans), 197.38 (cis); HRMS (EI) m/z calcd for $\text{C}_{12}\text{H}_{12}\text{O}_2$ (M^+) 188.0837, found 188.0830.

7-Methyl-1,2,3,3a,4,9b-hexahydro-cyclopenta[*a*]naphthalen-5-one (2c)



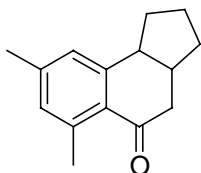
Obtained as a diastereomer mixture. mp 81-82 $^{\circ}\text{C}$; IR (neat) 1682 cm^{-1} ; ^1H NMR (500 MHz, CDCl_3) δ 1.18-2.05 (m, 5.67H, CH_2), 2.11-2.23 (m, 0.33H, CH_2 , cis), 2.25-2.41 (m, 1.34H, CH_2CO , CHCH_2CO , trans), 2.35 (s, 2.01H, PhCH_3 , cis), 2.37 (s, 0.99H, PhCH_3 , trans), 2.50-2.80 (m, 1.66H, CH_2CO , cis, CHCH_2CO , cis, CHPh , trans), 2.96 (dd, $J = 16.96, 3.67$ Hz, 0.67H, CH_2CO , trans), 3.18-3.27 (m, 0.33H, CHPh , cis), 7.12-7.38 (m, 2.00H, ArH), 7.77 (s, 0.33H, ArH , cis), 7.84 (s, 0.67H, ArH , trans); ^{13}C NMR (125 MHz, CDCl_3) δ 20.09 (cis), 21.10 (trans), 22.23 (trans), 23.76 (cis), 28.51 (trans), 30.95 (trans), 31.23 (cis), 33.21 (cis), 38.72 (cis), 40.92 (cis), 42.51 (cis), 44.39 (trans), 45.53 (trans), 46.30 (trans), 126.17 (trans), 126.73 (cis), 127.86 (trans), 129.15 (cis), 131.40 (cis), 132.11 (trans), 134.37 (trans), 134.77 (cis), 136.02 (cis), 136.14 (trans), 143.44 (cis), 144.17 (trans), 199.26 (cis), 199.46 (trans); HRMS (EI) m/z calcd for $\text{C}_{14}\text{H}_{16}\text{O}$ (M^+) 200.1201, found 200.1194;

9-Methyl-1,2,3,3a,4,9b-hexahydro-cyclopenta[*a*]naphthalen-5-one (2d)



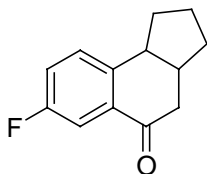
Obtained as a diastereomer mixture. mp 105-107 °C; IR (neat) 1682 cm⁻¹; ¹H NMR (500 MHz, CDCl₃) δ 1.18-2.05 (m, 5.53H, CH₂), 2.22-2.58 (m, 1.53H, CH₂, cis, CH₂CO, trans, CHCH₂CO, trans), 2.36 (s, 1.41H, PhCH₃, cis), 2.43 (s, 1.59H, PhCH₃, trans), 2.60-2.82 (m, 1.94H, CH₂CO, cis, CHCH₂CO, cis, CHPh, trans), 2.95 (dd, *J* = 16.04, 3.21 Hz, 0.53H, CH₂CO, trans), 3.20-3.24 (m, 0.47H, CHPh, cis), 7.17-7.40 (m, 2.00H, ArH), 7.87 (d, *J* = 7.79 Hz, 0.47H, ArH, cis), 7.97 (s, *J* = 7.79 Hz 0.53H, ArH, trans); ¹³C NMR (125 MHz, CDCl₃) δ 19.35 (cis), 21.78 (trans), 22.43 (trans), 23.27 (cis), 29.39 (cis), 30.17 (trans), 30.88 (trans), 31.65 (cis), 39.34 (cis), 40.09 (cis), 40.76 (cis), 44.02 (trans), 44.41 (trans), 46.81 (trans), 124.73 (cis), 126.01 (cis), 126.08 (trans), 126.27 (trans), 131.50 (cis), 133.59 (trans), 135.45 (cis), 136.14 (trans), 136.80 (cis), 136.84 (trans), 144.78 (cis), 145.03 (trans), 199.11 (cis), 199.36 (trans); HRMS (EI) *m/z* calcd for C₁₄H₁₆O (M⁺) 200.1201, found 200.1199;

6,8-Dimethyl-1,2,3,3a,4,9b-hexahydro-cyclopenta[*a*]naphthalen-5-one (2e)



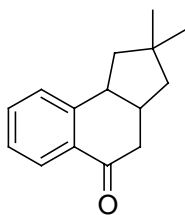
Obtained as a diastereomer mixture. mp 82-84 °C; IR (neat) 1673 cm⁻¹; ¹H NMR (500 MHz, CDCl₃) δ 1.20-2.02 (m, 5.74H, CH₂), 2.12-2.24 (m, 0.26H, CH₂, cis), 2.26-2.40 (m, 1.48H, CH₂CO, CHCH₂CO, trans), 2.31 (s, 0.78H, PhCH₃, cis), 2.34 (s, 2.22H, PhCH₃, trans), 2.50-2.73 (m, 1.52H, CH₂CO, cis, CHCH₂CO, cis, CHPh, trans), 2.56 (s, 0.78H, PhCH₃, cis), 2.61 (s, 2.22H, PhCH₃, trans), 2.93 (dd, *J* = 17.41, 4.58 Hz, 0.74H, CH₂CO, trans), 3.17-3.22 (m, 0.26H, CHPh, cis), 6.68-6.92 (m, 2.00H, ArH); ¹³C NMR (125 MHz, CDCl₃) δ 21.53, 21.66, 22.44, 22.85, 23.15, 23.89, 28.97, 31.05, 31.88, 33.97, 38.15, 43.39, 43.66, 44.71, 47.00, 47.12, 124.66 (trans), 127.85 (cis), 128.14 (cis), 128.51 (trans), 130.96 (cis), 131.33 (trans), 140.18 (cis), 141.64 (trans), 142.87 (trans), 143.30 (cis), 147.65 (cis), 148.13 (trans), 200.68 (trans), 200.90 (cis); HRMS (EI) *m/z* calcd for C₁₅H₁₈O (M⁺) 214.1358, found 214.1358;

7-Fluoro-1,2,3,3a,4,9b-hexahydro-cyclopenta[a]naphthalen-5-one (2f)



Obtained as a diastereomer mixture. mp 54-56 °C; IR (neat) 1690 cm⁻¹; ¹H NMR (500 MHz, CDCl₃) δ 1.35-2.08 (m, 5.59H, CH₂), 2.10-2.23 (m, 0.41H, CH₂, cis), 2.28-2.43 (m, 1.18H, CH₂CO, CHCH₂CO, trans), 2.51-2.78 (m, 1.82H, CH₂CO, cis, CHCH₂CO, cis, CHPh, trans), 3.00 (dd, *J* = 17.41, 3.67 Hz, 0.59H, CH₂CO, trans), 3.18-3.26 (m, 0.41H, CHPh, cis), 7.17-7.32 (m, 2.00H, ArH), 7.61 (dd, *J* = 9.17, 2.75 Hz, 0.41H, ArH, cis), 7.68 (dd, *J* = 9.17, 2.75 Hz, 0.59H, ArH, trans); ¹³C NMR (125 MHz, CDCl₃) δ 22.12 (trans), 23.69 (cis), 28.61 (trans), 30.84 (trans), 31.24 (cis), 33.19 (cis), 38.64 (cis), 40.52 (cis), 42.27 (cis), 44.35 (trans), 45.20 (trans), 46.07 (trans), 112.52 (d, ²*J*_{C-F} = 22.07 Hz, cis), 113.79 (d, ²*J*_{C-F} = 22.07 Hz, trans), 120.53 (d, ²*J*_{C-F} = 22.07 Hz, trans), 121.03 (d, ²*J*_{C-F} = 22.07 Hz, cis), 128.12 (d, ³*J*_{C-F} = 7.68 Hz, trans), 131.07 (d, ³*J*_{C-F} = 7.68 Hz, cis), 133.19 (d, ³*J*_{C-F} = 6.72 Hz, cis), 134.06 (d, ³*J*_{C-F} = 5.76 Hz, trans), 142.00 (d, ⁴*J*_{C-F} = 2.88 Hz, cis), 142.75 (d, ⁴*J*_{C-F} = 2.88 Hz, trans), 161.56 (d, ¹*J*_{C-F} = 245.68 Hz, trans), 161.77 (d, ¹*J*_{C-F} = 246.64 Hz, cis), 197.92 (cis), 198.12 (trans); HRMS (EI) *m/z* calcd for C₁₃H₁₃FO (M⁺) 204.0950, found 204.0947

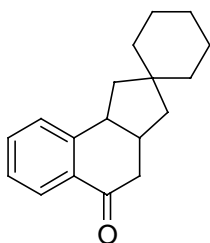
2,2-Dimethyl-1,2,3,3a,4,9b-hexahydro-cyclopenta[a]naphthalen-5-one (2g)



Obtained as a diastereomer mixture. IR (neat) 1684 cm⁻¹; ¹H NMR (500 MHz, CDCl₃) δ 1.05 (s, CH₃), 1.12 (s, CH₃), 1.13 (s, CH₃), 1.20 (s, CH₃), 1.34-1.43 (m, CH₂), 1.52 (dd, *J* = 11.91, 11.91 Hz, CH₂), 1.79-1.85 (m, CH₂), 2.00 (dd, *J* = 12.83, 7.33 Hz, CH₂), 2.10-2.23 (m, 0.54H, CHCH₂CO, trans), 2.20 (dd, *J* = 11.91, 6.87 Hz, CH₂), 2.36 (dd, *J* = 16.96, 13.29 Hz, 0.54H, CH₂CO, trans), 2.60-2.69 (m, 0.92H, CH₂CO, cis), 2.80-2.89 (m, 1.00H, CHCH₂CO, cis, PhCH, trans), 2.92 (dd, *J* = 16.96, 3.67 Hz, 0.54H, CH₂CO, trans), 3.43-3.48 (m, 0.46H, PhCH, cis), 7.18-7.50 (m, 3.00H, ArH), 7.93 (d, *J* = 7.79 Hz, 0.46H,

ArH, cis), 8.02 (d, $J = 7.79$ Hz, 0.54H, ArH, trans); ^{13}C NMR (125 MHz, CDCl_3) δ 31.33, 31.46, 32.08, 32.21, 37.26, 38.08, 38.79, 41.77, 42.05, 43.73, 44.93, 45.43, 46.07, 47.09, 47.58, 48.99, 126.08 (trans), 126.26 (cis), 126.50 (trans), 126.58 (cis), 127.64 (trans), 129.14 (cis), 131.80 (cis), 132.37 (trans), 133.51 (trans), 133.69 (cis), 146.30 (cis), 147.07 (trans), 198.98 (cis), 199.02 (trans); HRMS (EI) m/z calcd for $\text{C}_{15}\text{H}_{18}\text{O}$ (M^+) 214.1358, found 214.1357.

**Spiro[cyclohexane-1,2'-(1',2',3',3a',4',9b'-hexahydro-cyclopenta[a]naphthalen-5'-one)]
(2h)**



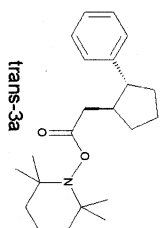
Obtained as a diastereomer mixture. IR (neat) 1684 cm^{-1} ; ^1H NMR (500 MHz, CDCl_3) δ 1.20-1.62 (m, CH_2), 1.73-1.94 (m, CH_2), 2.02-2.14 (m, 0.42H, CHCH_2CO , trans), 2.08 (dd, $J = 13.29, 7.79$ Hz, CH_2), 2.29 (dd, $J = 13.27, 6.87$ Hz, CH_2), 2.33 (dd, $J = 16.96, 13.29$ Hz, 0.42H, CH_2CO , trans), 2.58-2.68 (m, 1.16H, CH_2CO , cis), 2.74-2.84 (m, 1.00H, PhCH , trans, CHCH_2CO , cis), 2.93 (dd, $J = 16.96, 3.67$ Hz, 0.42H, CH_2CO , trans), 3.35-3.39 (m, 0.58H, PhCH , cis), 7.23-7.56 (m, 3.00H, ArH), 7.95 (d, $J = 7.79$ Hz, 0.58H, ArH, cis), 8.02 (d, $J = 7.79$ Hz, 0.42H, ArH, trans); ^{13}C NMR (125 MHz, CDCl_3) δ 23.44, 23.63, 23.73, 24.78, 26.03, 26.06, 37.80, 39.69, 40.48, 40.73, 41.08, 41.50, 41.60, 41.76, 42.83, 42.88, 45.12, 45.51, 126.09 (trans), 126.30 (cis), 126.46 (trans), 126.67 (cis), 127.63 (trans), 129.15 (cis), 131.73 (cis), 132.39 (trans), 133.49 (trans), 133.68 (cis), 146.33 (cis), 147.21 (trans), 199.01 (cis), 199.09 (trans); HRMS (EI) m/z calcd for $\text{C}_{18}\text{H}_{22}\text{O}$ (M^+) 254.1671, found 254.1672.

References.

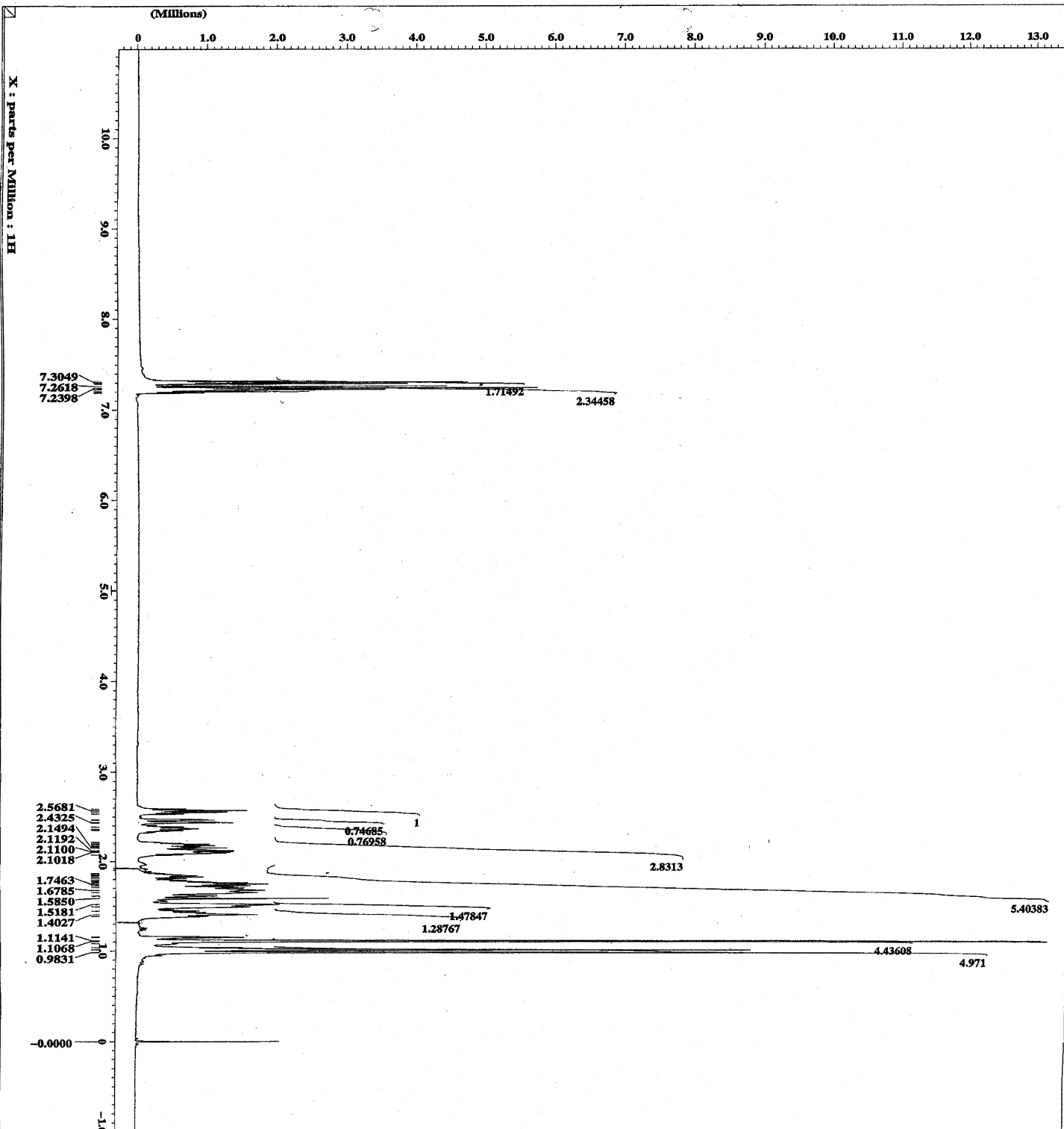
- (1) Ashby, E. C.; DePriest, R. N.; Goel, A. B.; Wenderoth, B.; Pham, T. N. *J. Org. Chem.* **1984**, *49*, 3545.
- (2) (a) Hawker, C. J.; Barclay, G. G.; Orellana, A.; Dao, J.; Devonport, W.

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- (3) Oh, S.-H; Sato, T. *J. Org. Chem.*, **1994**, *59*, 3744.

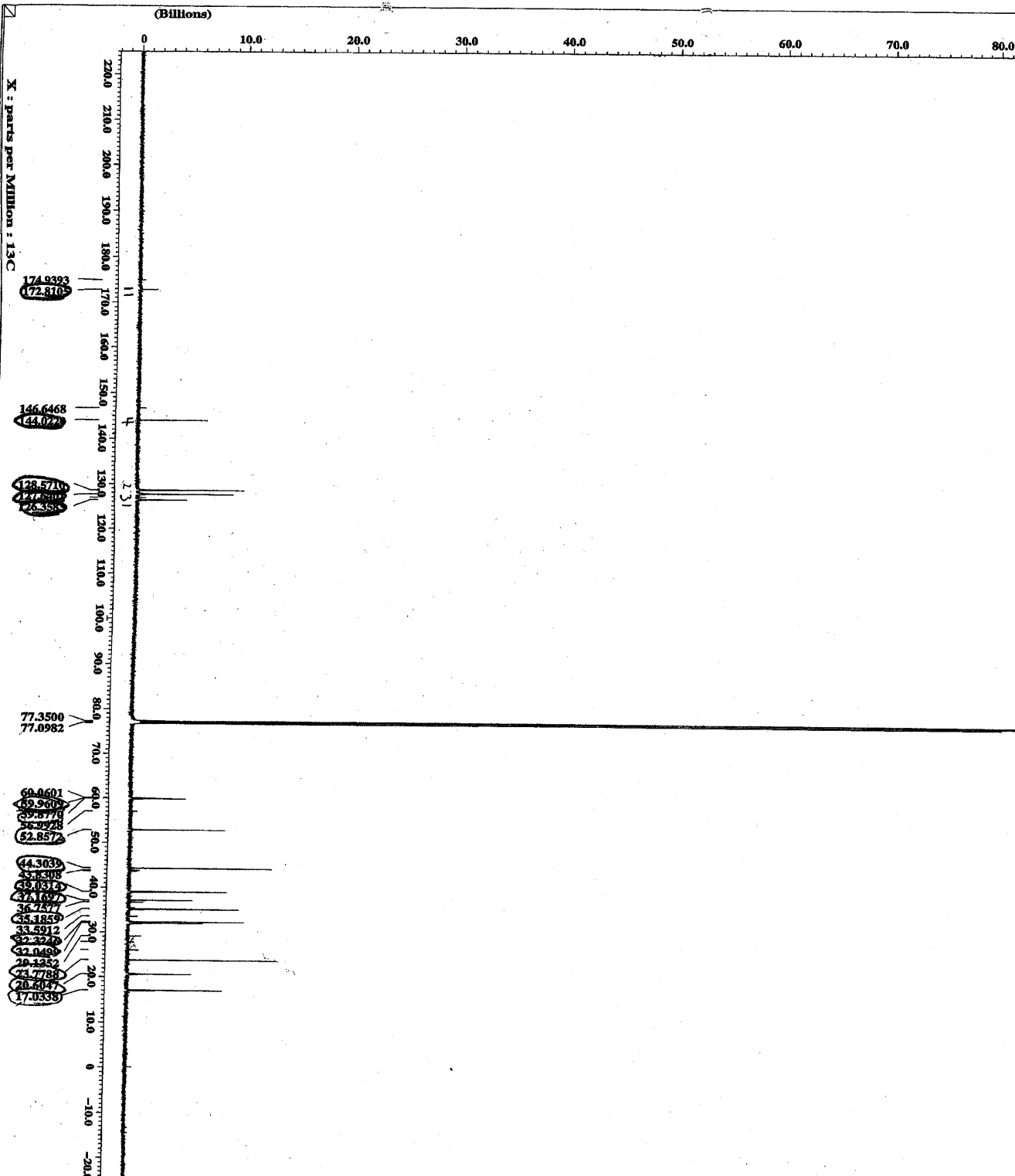
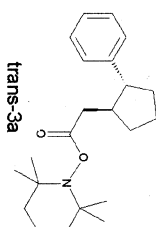
----- ACQUISITION PARAMETERS -----
 File Name = 14_spectrum.141
 Author =
 Sample ID = MAS-7500-017 HPLC
 Sample Name = S10
 Creation Date = 31-AUG-2004 11:50:17
 Revision Date = 31-AUG-2004 09:05:30
 Spec file = KCP500
 Spec Type = NMR
 Data Format = 1D COMPLEX
 Dimensions = 1
 Num Fids = 1
 Num Sols = 1
 Num Units = 1
 Sols = 1
 Mod Return = 1
 X Name = 1H
 X Freq = 500.1624602 [MHz]
 X Sweep = 7.50750731 [kHz]
 Solvent = CHLOROFORM-D
 Spat prec = 16 [Hz]
 Recvz gain = 19 [dB]
 Field strength = 11.7473579 [T]
 Filter mode = HYPERNOISE
 Filter width = 3.7510936 [kHz]



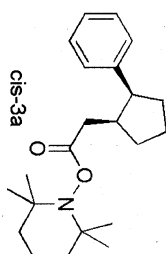
S10



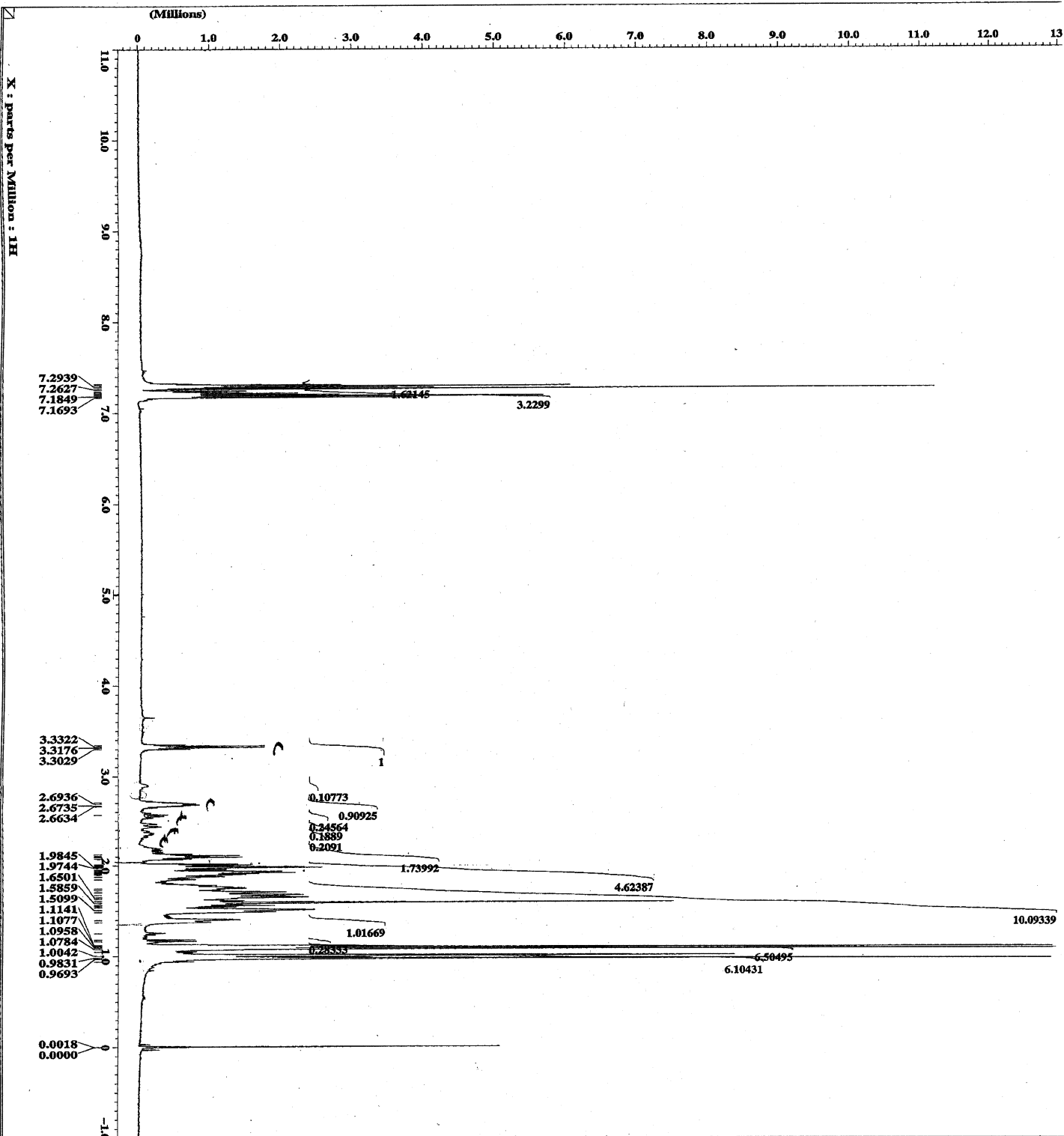
Acquisition Parameters
 File Name = 14_116_spectrum_copy.9
 Author ID = NIS-7780-017 HP/C-C
 Comment = Second
 Creation Date = 11-30-2004 09:28:17
 Revision Date = 1-SEP-2004 07:03:50
 Spec File = 809500
 Spec Type = 1D NMR
 Data Format = ID COMPLEX
 Data File = 1
 Data Size = 12766
 Data Units = [ppm]
 Acq. Method = 13C
 X offset = 100 [ppm]
 X freq = 125.77787547 [MHz]
 X presat = 31.44654088 [Hz]
 Solvent = CDCl3
 NS/DS = 24 [sec]
 Recyc. gain = 30
 Field strength = 11.7473579 [T]
 Filter name = HUYSENHOFER
 Filter value = 15.72065221 [Hz]



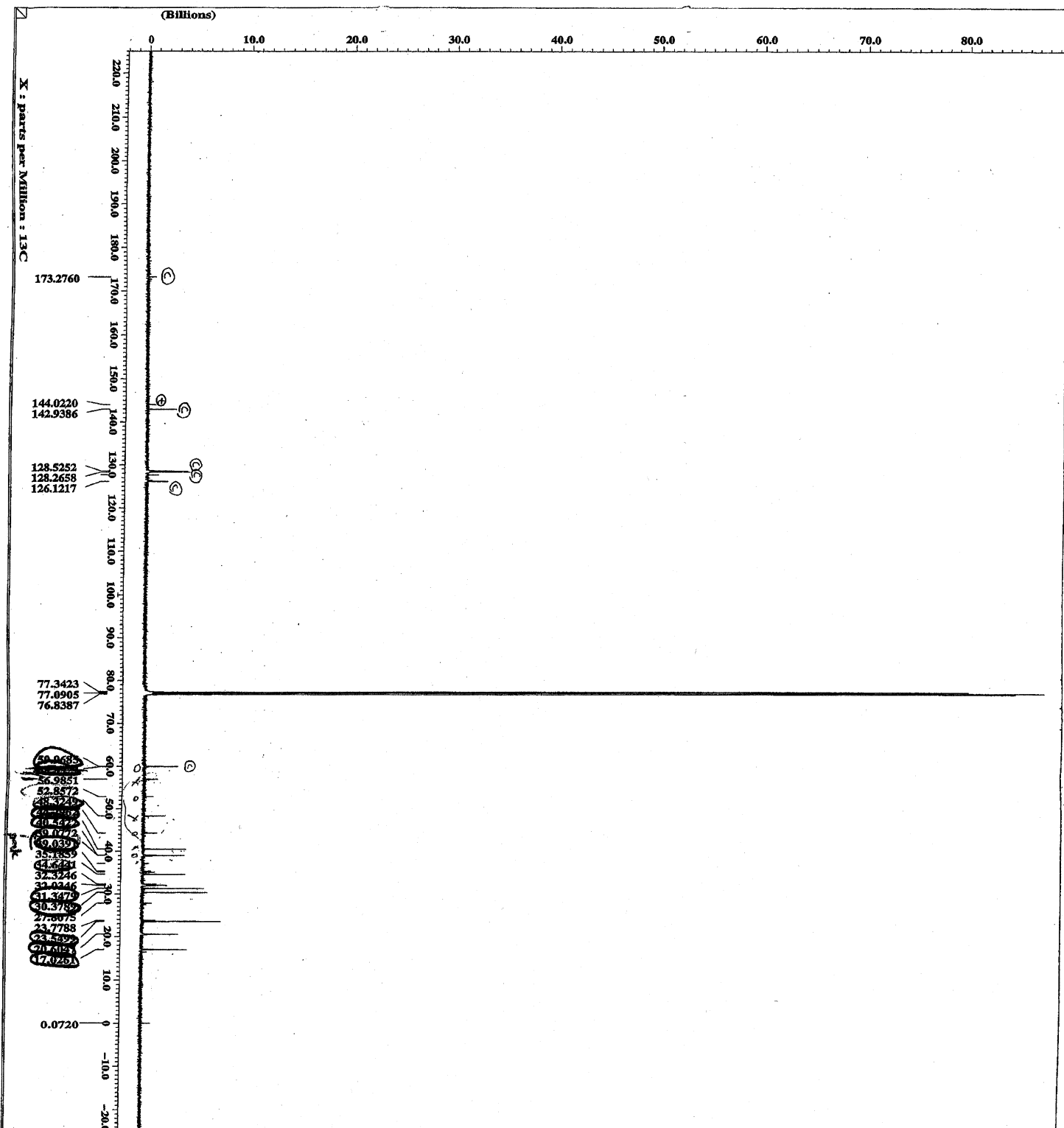
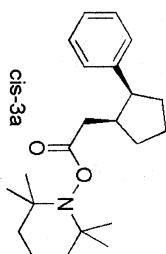
Acquisition Parameters
 File Name = id_spectrum_550
 Author ID = 110-2004-017 HNC-2
 Comment = Sample Plate Spectrum
 Creation Date = 30-AUG-2004 11:49:56
 Revision Date = 31-AUG-2004 09:24:33
 Spec Site = HCN500
 Spec Type = 1D/2D NMR
 Data Format = ID COMPLEX
 Dimensions = 1
 Data Size = 16384
 Data Date = [ppm]
 Scan = 8
 Recv. Gain = 11
 X Offset = 5 [ppm]
 X Freq = 500.13241602 [MHz]
 X Fwamp = 7.50750751 [Hz]
 Solvent = DMSO-D6
 Temp. deg = 22
 Recv. Gain = 11.743579 [V]
 Filter Mode = HETEROMODE
 Filter Width = 5.751395 [kHz]



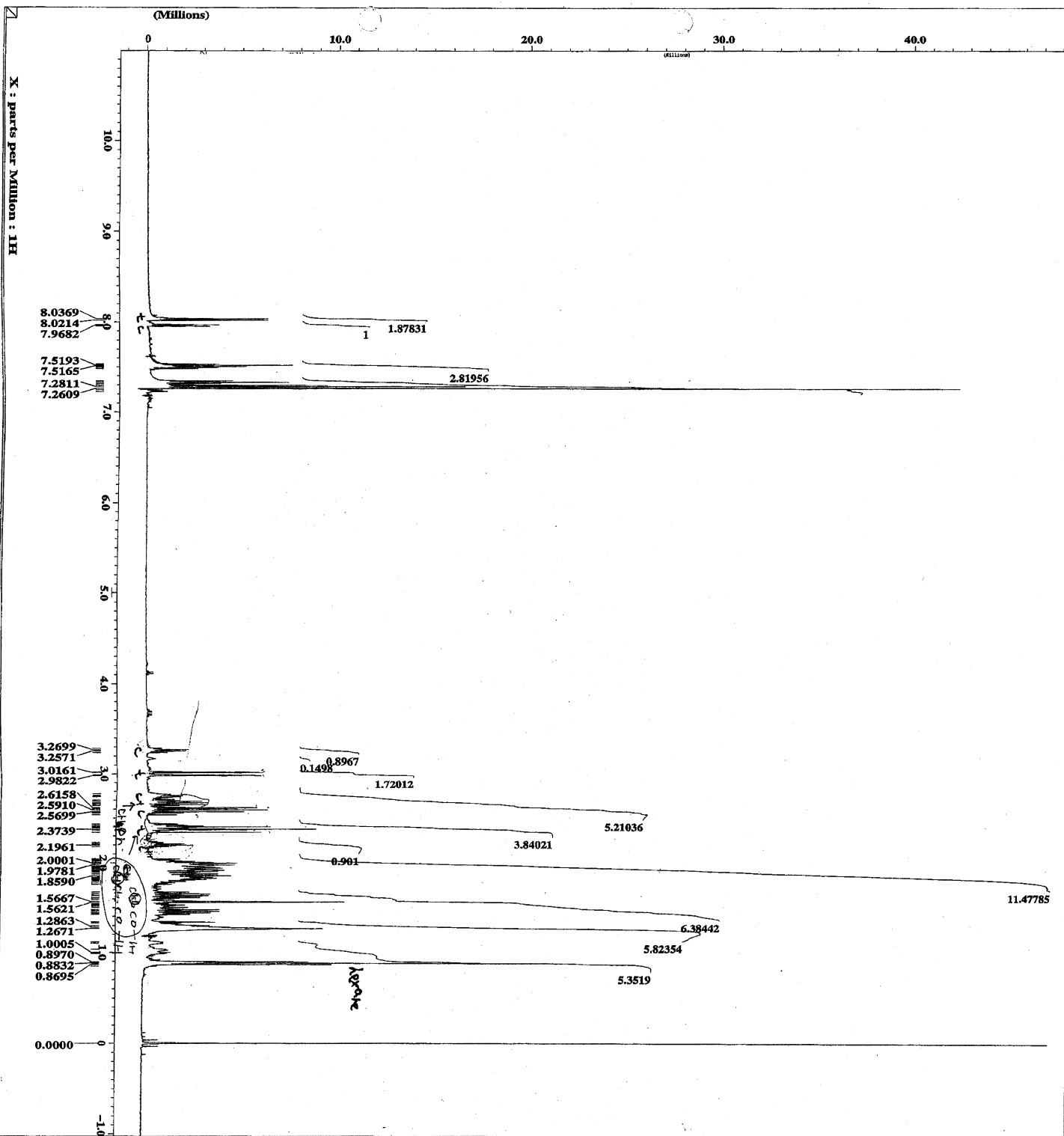
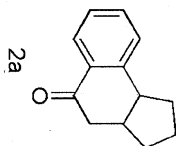
S12



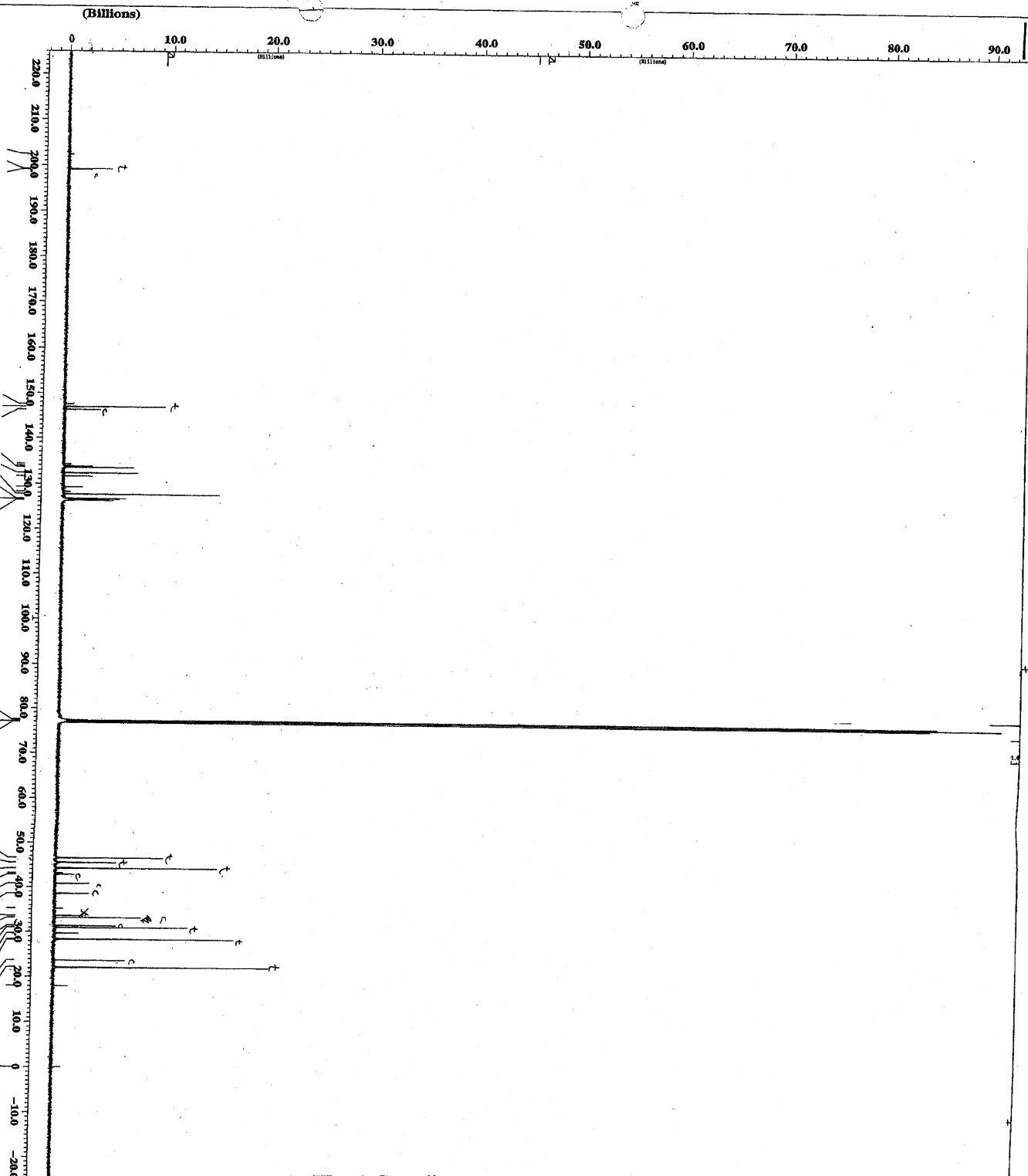
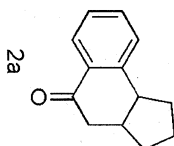
ACQUISITION PARAMETERS
 File Name = id_13c_spectrum.194
 Sample ID = 13C-PMPO-017 HPC-P
 Comment = 13C-PMPO-017 HPC-P
 Creation Date = 1-SEP-2004 09:21:01
 Revision Date = 2-SEP-2004 06:57:58
 Spec Date = 20040901
 Spec Time = 06:57:58
 Data Type = 1D
 Data Format = ID
 Dimensions = 1
 Num Points = 32768
 Num Slices = 1
 Num Vials = 1
 Solvent = CDCl3
 Ref. Temp. = 300.2
 X (ppm) = 100.625
 X (offset) = 100.625
 X (freq) = 125.77787547 [MHz]
 X (ampl) = 31.4454081 [kHz]
 Solvent = CDCl3
 Ref. Temp. = 30.0
 Heavy Atoms = 11.7473579 [e]
 Field Strength = 11.7473579 [e]
 File Name = 13C-PMPO-017 HPC-P
 File Path = 15:72066221 [kHz]



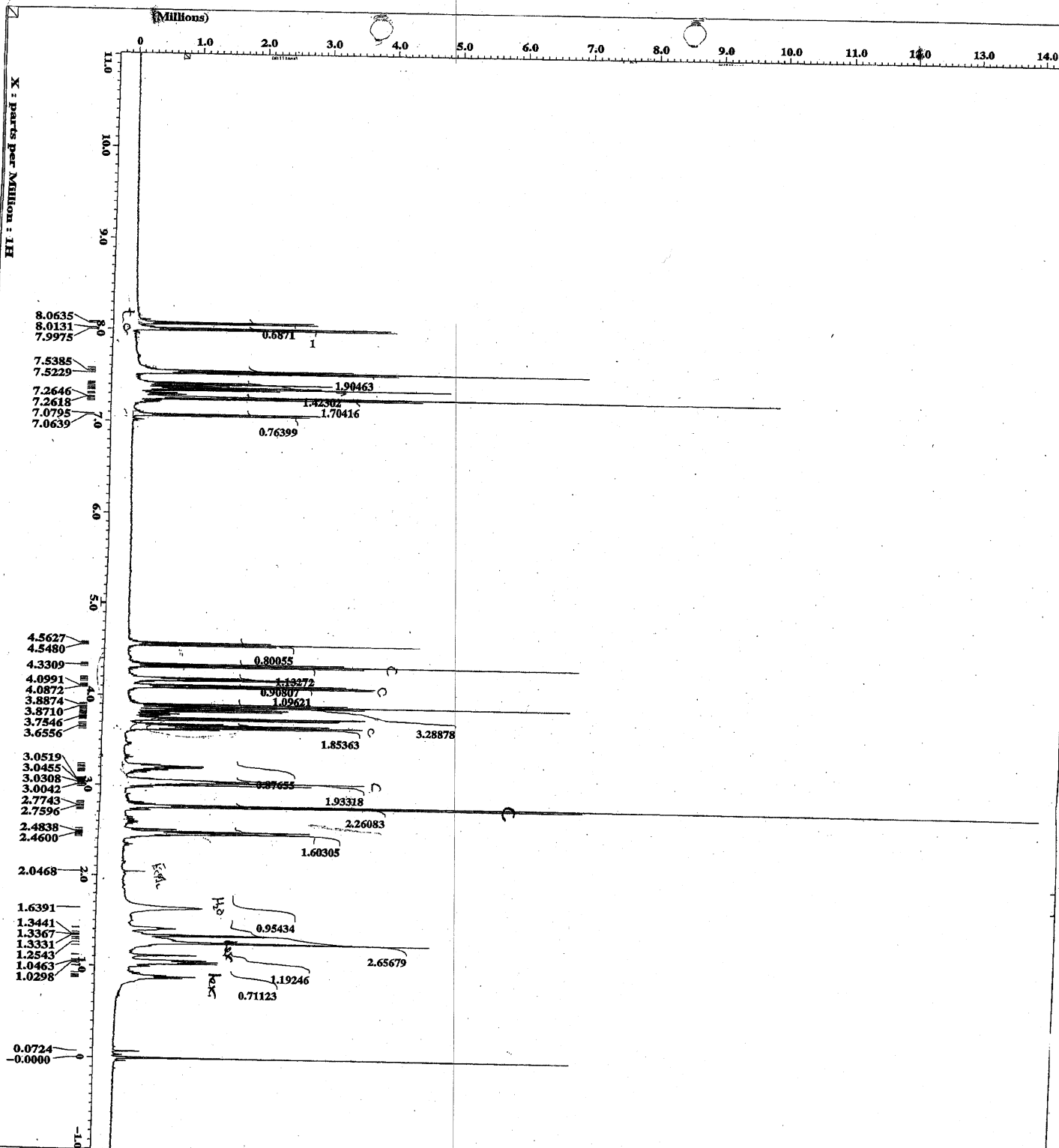
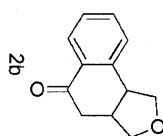
----- ACQUISITION PARAMETERS -----
 File Name = 1d_spectrum_1314
 Author ID =
 Sample ID = NMR-2004-029 4-4-11
 Concentration = 50.01 mg/ml
 Creation Date = 28-SEP-2004 15:25:41
 Revision Date = 29-SEP-2004 14:34:54
 Spec Site = ZCP500
 Spec Type = 1D NMR
 Data Format = 1D COMPLEX
 Data Name =
 Data Site = 16384
 Data Units = [ppm]
 Scale = 15
 Scale Factor = 1
 X Offset = 51.000
 X Freq = 500.16341602 [MHz]
 X Sweep = 7.50750751 [KHz]
 Solvent = CDCl3
 Temp Set = 23.81 [C]
 Recv Gain = 24.743391 [V]
 Field Strength = 11.743391 [T]
 Pulse Program = zgpg30
 Filter Width = 3.731535 [Hz]



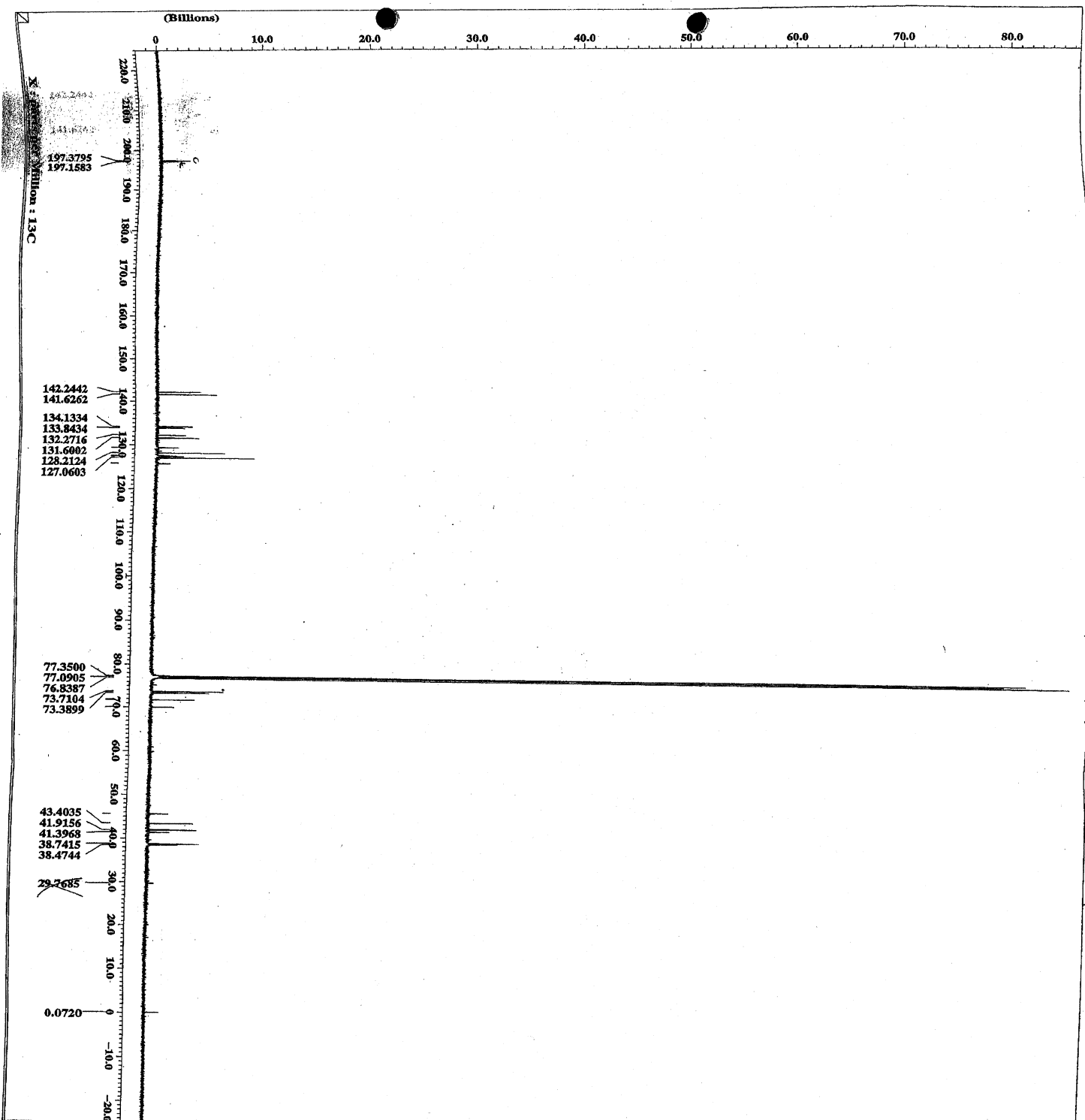
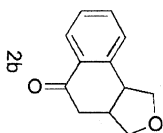
--- ACQUISITION PARAMETERS ---
 File Name = 14_13c_spectrum_241
 Author =
 Sample ID = MG-1280-022 fcd-7
 Sample Name =
 Sample Notes =
 Creation Date = 7-SEP-2004 09:27:43
 Revision Date = 8-SEP-2004 07:09:34
 Spec Site = MS500
 Spec Type = 13C NMR
 Data Format = ID COMPLEX
 Dimensions = 1
 Dimensions = 13C
 Dim Size = 128K
 Dim Units = [ppm]
 Secs = 2061
 Mod_Petusa = 1
 X_Offset = 13C
 X_Freq = 125.77787547 [MHz]
 X_Pwmp = 11.44540881 [kHz]
 SOLVENT = CDCl3
 Temp = 17 [degC]
 Temp Set = 30 [degC]
 Recvry Gain = 11.7472579 [V]
 Field Strength = 125.77787547 [MHz]
 Pulse Width = 15.7206221 [kHz]

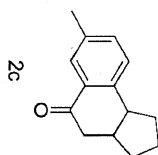


ACQUISITION PARAMETERS
 File Name = 1d spectrum.1302
 Author ID = NIS-REMO-028 8221-40
 Sample ID = 1302
 Content = 21-SEP-2004 11:28:10
 Creation Date = 21-SEP-2004 11:28:10
 Acquisition Date = 21-SEP-2004 11:27:03
 Spec File = ECH510
 Spec Type = 1H NMR
 Data Format = ID COMPLEX
 Data File = 1302
 Data Size = 16184
 Data Units = [ppm]
 Scale Factor = 1
 X Offset = 0
 X Gain = 500.16241602 [Hz/1]
 X Resol = 7.50730751 [Hz/1]
 Solvent = CDCl3
 Temp. Unit = 23.1 [C]
 Recv. Gain = 11.747379 [V]
 Field Strength = 500.13 [MHz]
 Pulse Width = 3.7511936 [Hz]

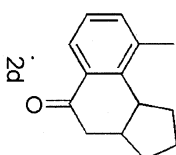
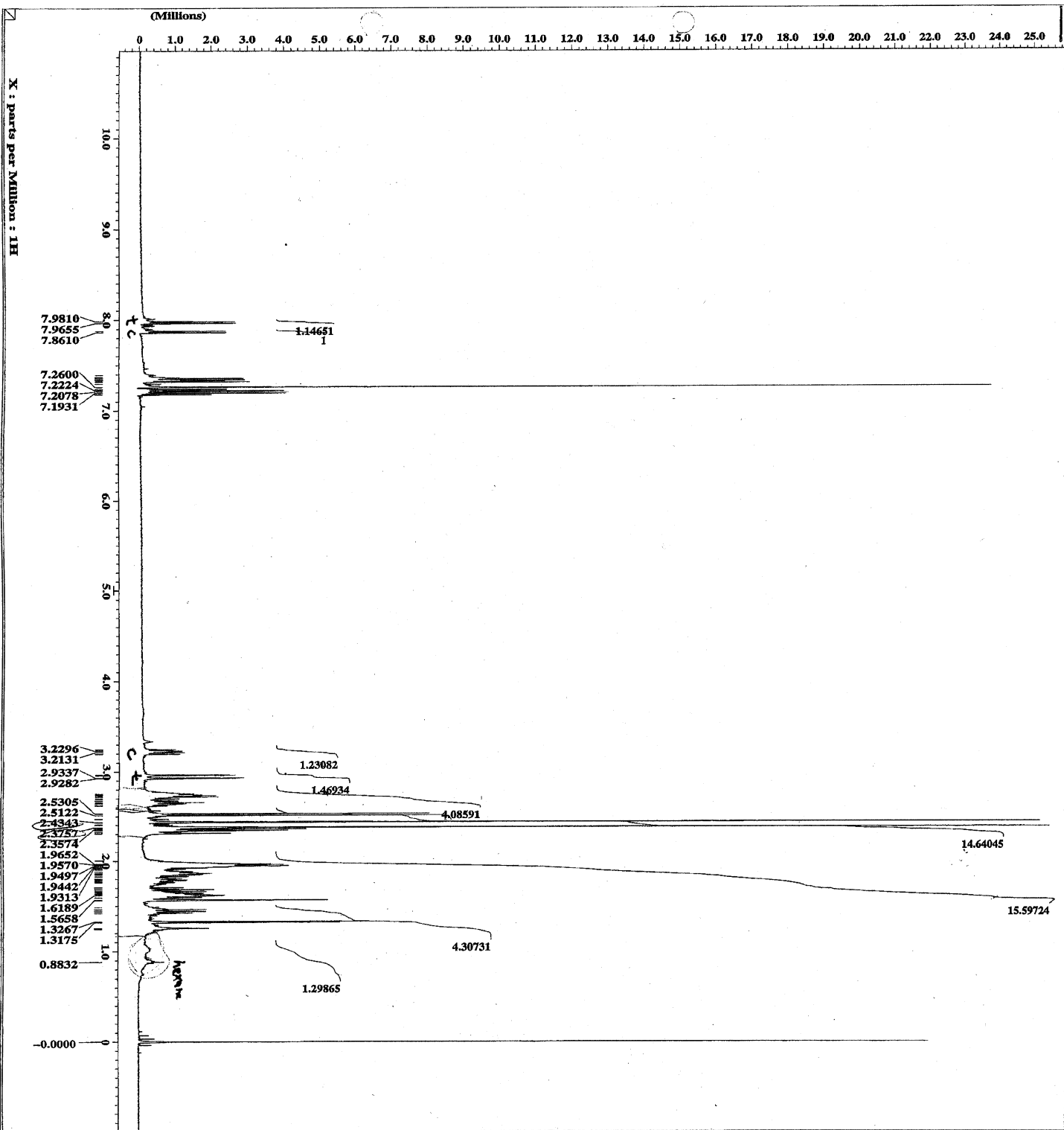


Acquisition Parameters
 File Name = 14_13c_spectrum_173
 Author =
 Sample ID = MS-2004-026 6-23-04
 Content = Single Pulse with Broad
 Creation Date = 22-Sep-2004 09:22:58
 Revision Date = 23-Sep-2004 07:23:08
 Spec Site = KCP500
 Spec Type =
 Data Source =
 Dimensions =
 Dim File =
 Dim File =
 Dim File =
 Dim File =
 Dim File =
 Mod Path =
 K. Acq =
 K. Offset =
 K. Freq =
 K. Freq =
 Solvent =
 Spill Set =
 Temp Set =
 Temp Set =
 Temp Set =
 Filter Mode =
 Filter Width = 15.72065231 [Hz]



[illegible]





S22

ACQUISITION PARAMETERS

File Name = 10_spectra.1167

Author =

Sample ID = MS-TRBO-027 #9-21

Content = Single Pulse Experiment

Creation Date = 16-SEP-2004 12:40:01

Revision Date = 17-SEP-2004 10:33:42

Spec Site = EXP500

Spec Type = 1H

Data Format = 1D

Dimensions = 1H

Dim F1/2 = 16384

Dim F2/2 = 16384

Scans = 1

Mod Return = 1

X Domain = 1H

X Offset = 5199.1

X Freq = 500.136011MHz

X Sweep = 7.507507511Hz

Solvent = CDCl3

Solvent = CHLOROFORM-D

Solvent = 17Hz

Temp Unit = 24.0C

Temp = 24.0C

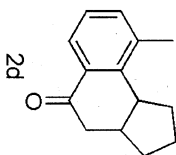
Field Strength = 11.747379T

Pulse Mode = HYPERNOISE

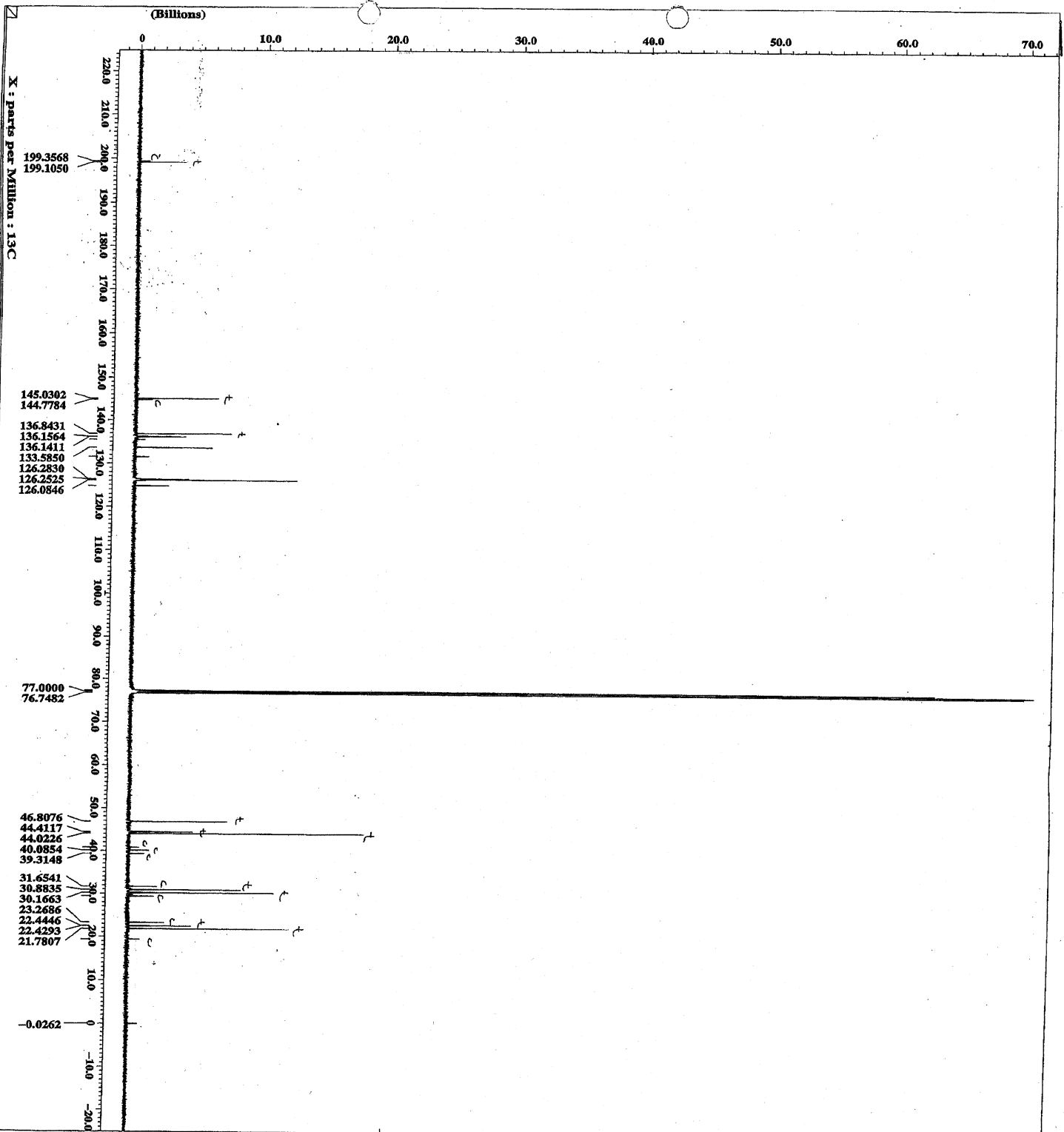
Pulse Width = 3.7511936Hz



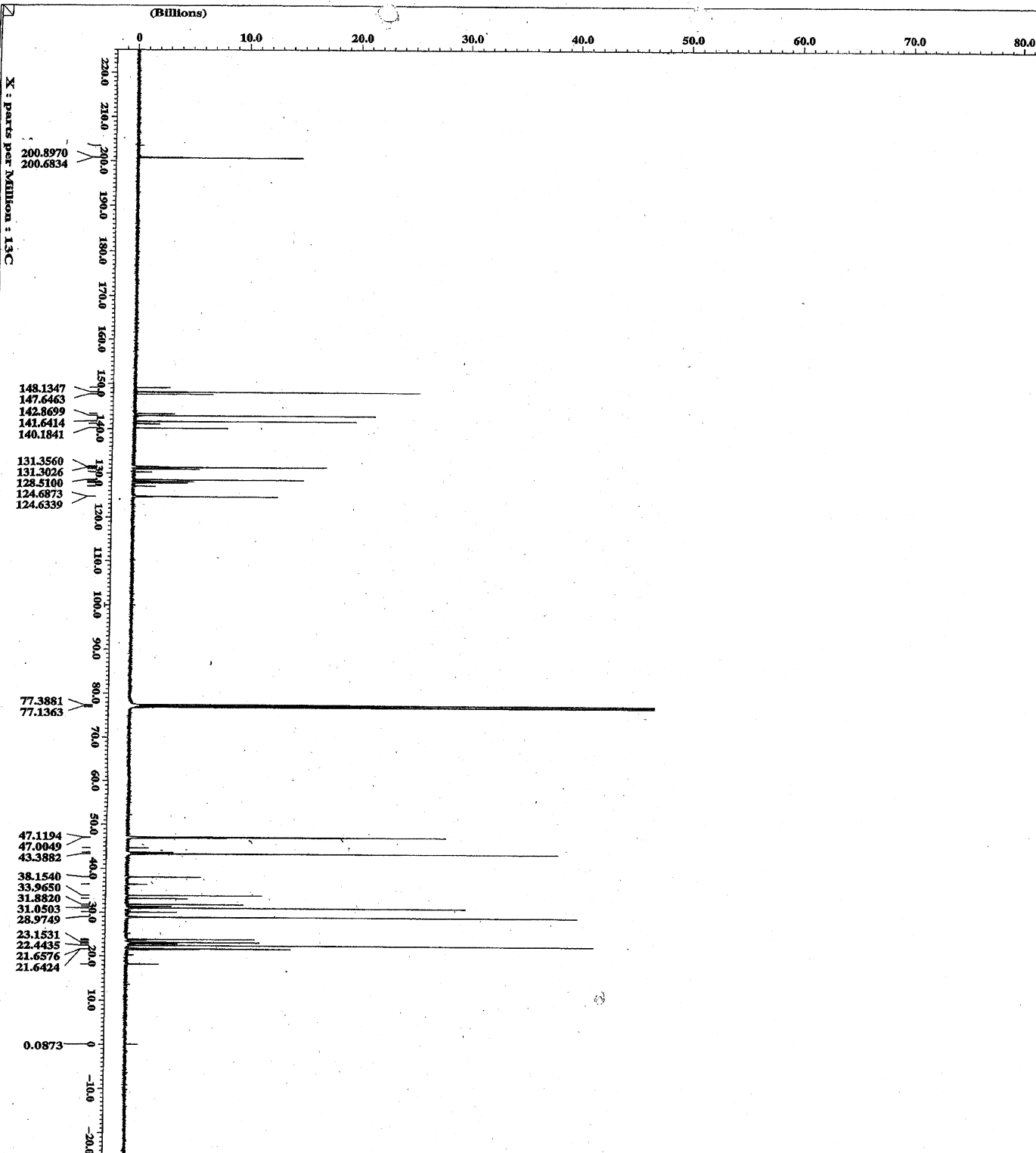
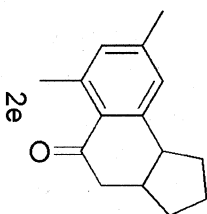
Acquisition Parameters
 File Name = 14_13c_spectrum_copy.4
 Author =
 Sample ID = MAS-7880-027 f2-9-21 ps
 Sample Name = Sample Polym with Record
 Creation Date = 10-08-2004 03:15:12K
 Revision Date = 10-08-2004 07:42:25
 Spec Site = K29500
 Spec Type =
 Data Format = ID COMPLEX
 Parameters =
 Pulse Program =
 Dm Scales = 1306
 Dm Units = 1397
 Scan = 1397
 Root Prefix =
 X Offset = 1306
 X Fixed = 129.77757547 [Hz]
 X Sweep = 31.44654088 [Hz]
 Solvent = CH2Cl2
 Name_001 = 2446 [Hz]
 Name_002 = 30 [Hz]
 Field Strength = 11.747375 [T]
 Filter Mode = HETZSCHMIDT
 Filter Width = 15.7706521 [Hz]



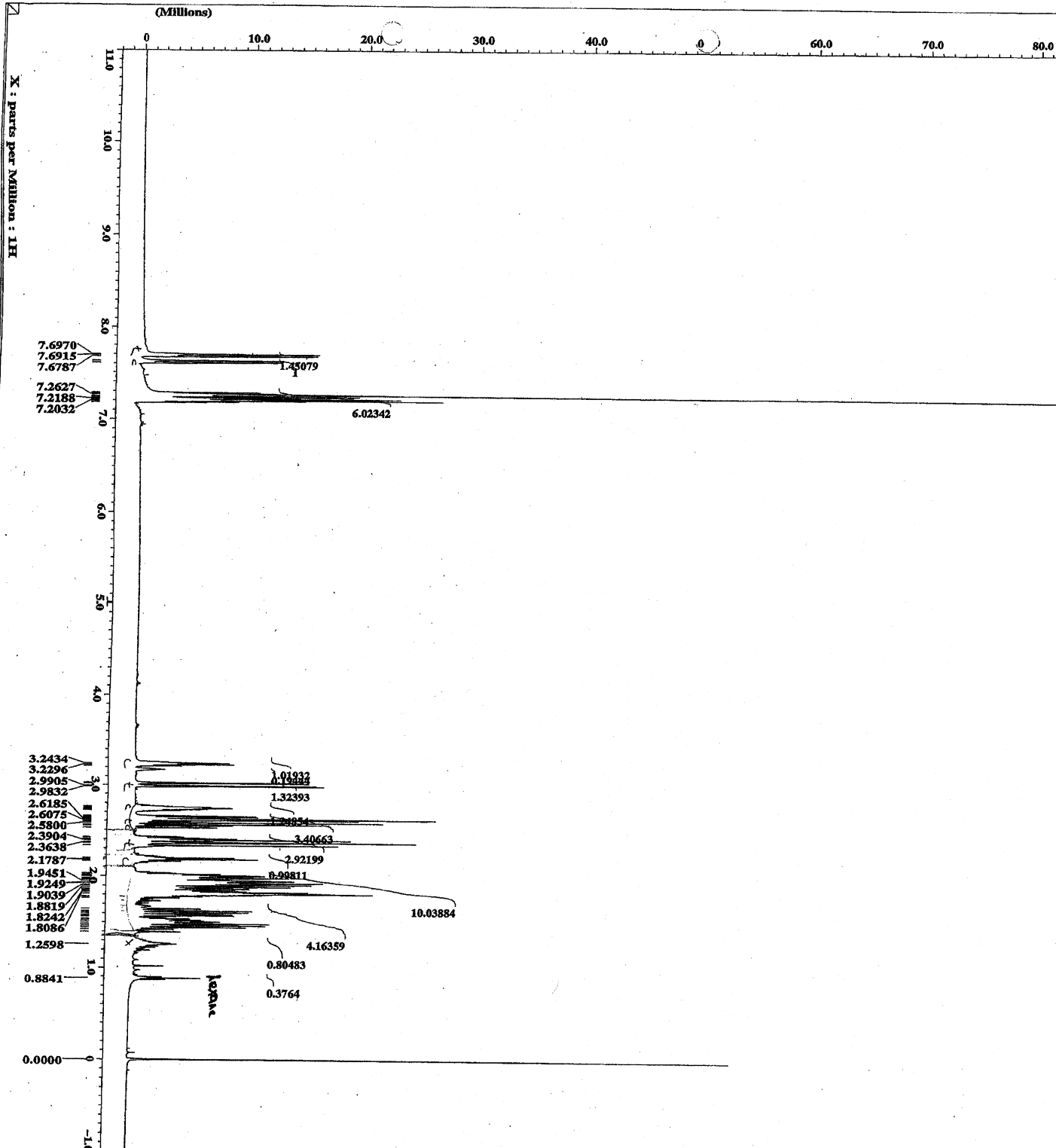
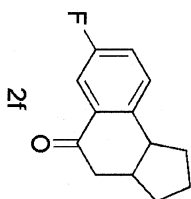
S23



Acquisition Parameters
 File Name = 14_13c_spectrum.196
 Sample ID = N45-PMO-013 3H-C-3
 Content = Single Pulse with Broad
 Creation Date = 25-Nov-2004 07:37:17
 Acquisition Date = 25-Nov-2004 06:47:23
 Spec Site = KCP500
 Spec Type = 13C NMR
 Solvent = CDCl3
 Dimensions = 13C
 Num Fids = 13C
 Num Slices = 12768
 Num Orbits = 10996
 Num Scans = 1
 Mod Return = 13C
 X Offset = 100.625
 X Freq = 125.7778147 MHz
 X F2 = 125.7778147 MHz
 Solvent = CDCl3
 Solvent = 13C
 Spins per = 23147
 Temp Set = 30.74187873
 Probe Gain = 15.7066221 MHz
 Filter Mode = 15.7066221 MHz
 Filter Width = 15.7066221 MHz



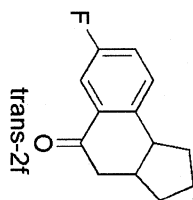
--- ACQUISITION PARAMETERS ---
 File Name = 14.spectrum.654
 Sample ID = NMR-7500-037 E5-10
 Compound = Single Pulse Experiment
 Creation Date = 15-DEC-2004 14:25:38
 Revision Date = 14-DEC-2004 12:54:37
 Spec Site = KCP500
 Spec Type = NMR 1H
 Data Format = 1D COMPLEX
 Dimensions = 1H
 Num F2s = 1
 Num F1s = 16384
 Num Sols = 1
 Num Scans = 32
 Num Solvent = 1
 Num Acq = 1
 Num Relax = 1
 Num Delay = 1
 Num Dec = 1
 Num Rec = 1
 Num Recycle = 1
 Num Sweep = 1
 Num Solvent = CHLOROFORM-D
 Spin rate = 27 [Hz]
 Acq. gain = 25.4 [dB]
 Yield strength = 11.743579 [V]
 Filter mode = HYPERCUT
 Filter width = 3.7511936 [Hz]



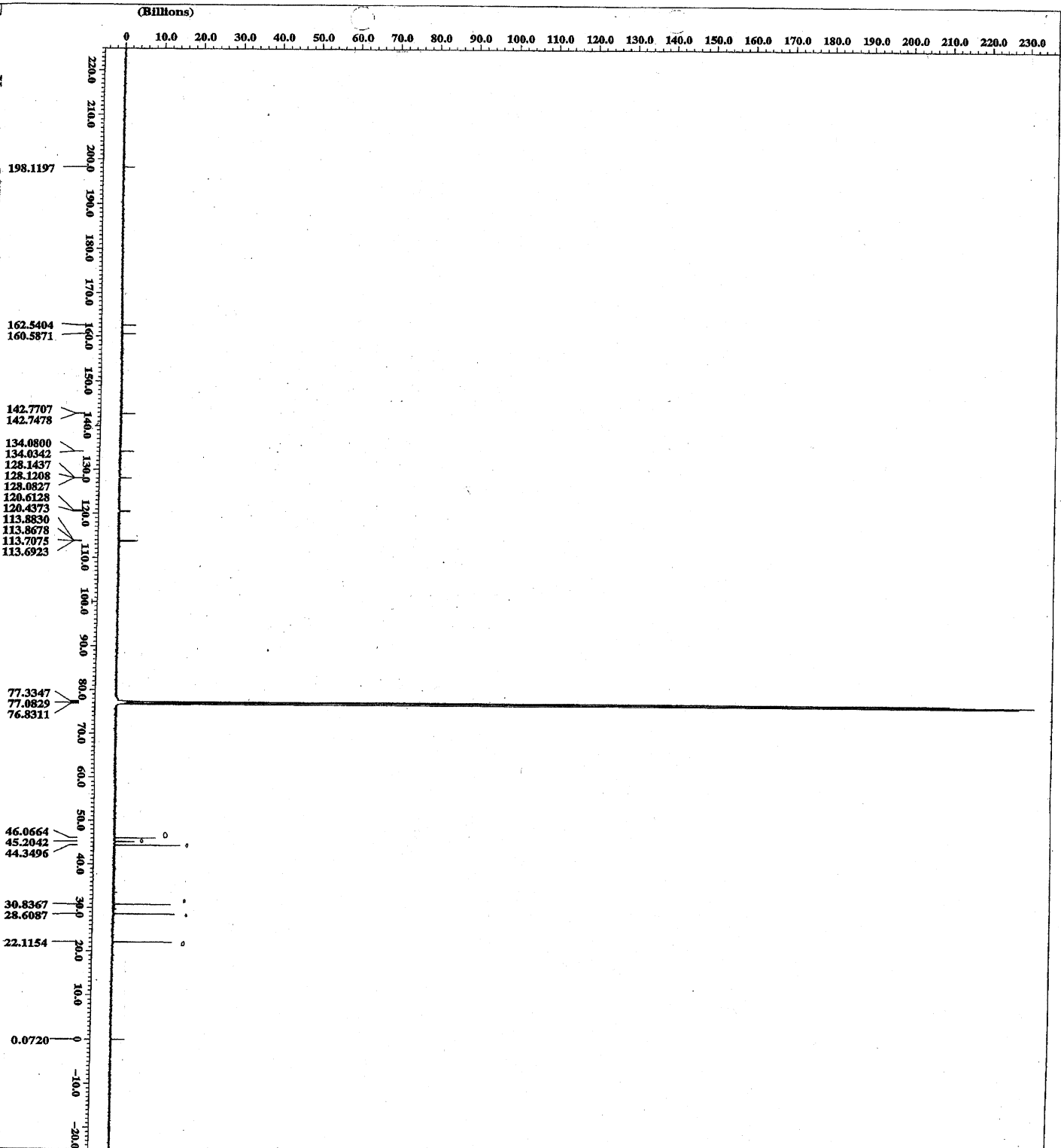
```

===== ACQUISITION PARAMETERS =====
File Name      = 14_11c_spectra.108
Author         =
Sample ID      = MAG-7280-036-2-E4-5
Comment        = Single Pulse with Broad
Creation Date   = 13-SEP-2004 09:18:56
Revision Date   = 14-SEP-2004 07:08:18
Spec Site      = ECE500

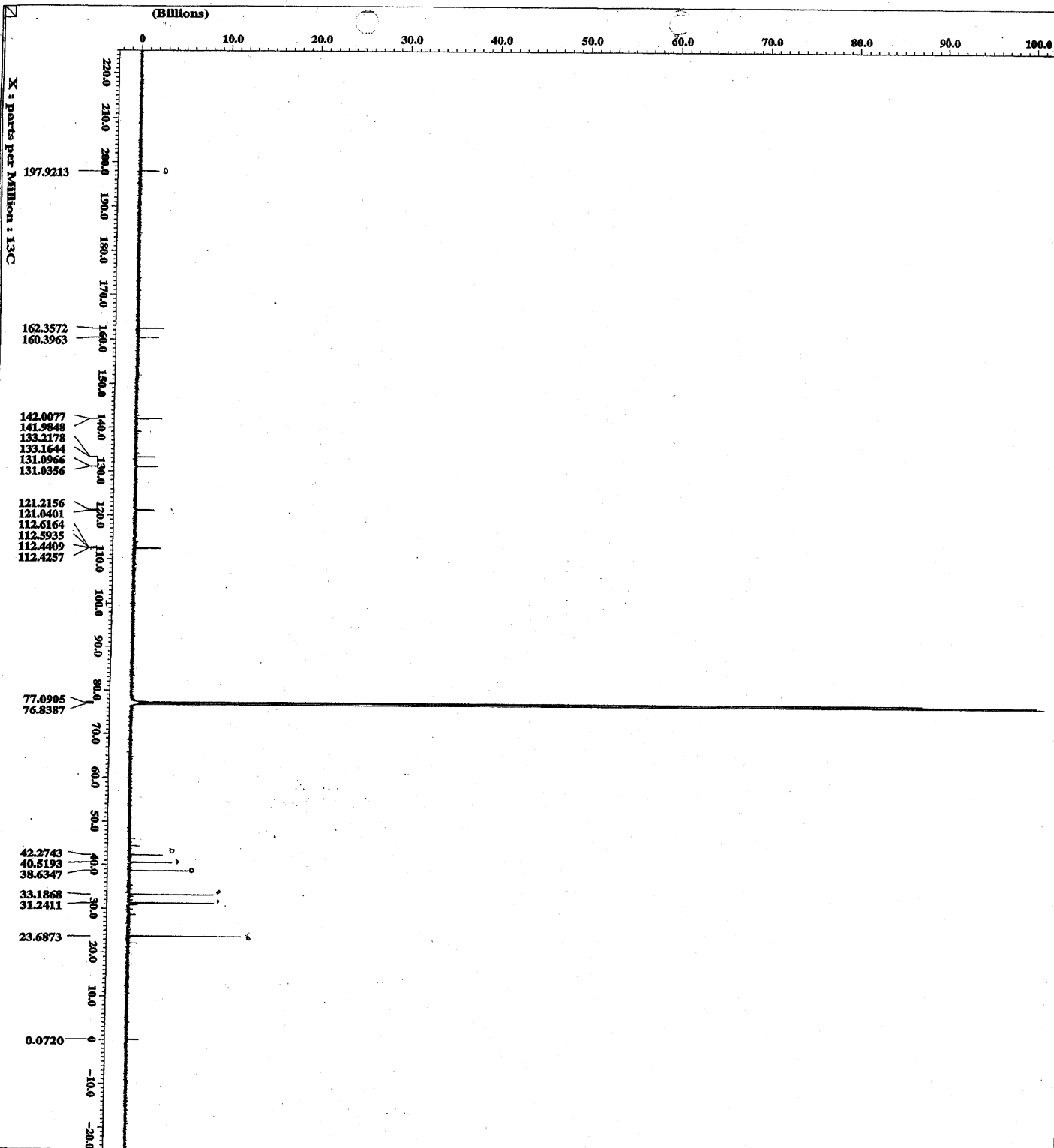
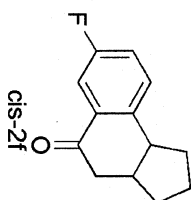
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Spec Type      = 1H/13C NMR
Data Format     = 1D COMPARE
Dimensions     = 1
Dir File       = 13C
Dir File       = 3278
Dir File       = 13C
Dir File       = 41863
Mod. return    = 1
X. domain      = 13C
X. offset      = 100.625[13C]
X. sweep       = 11.4465088[Hz]
Solvent        = CHLOROFORM-D
Solv. proc     = 16[Hz]
Temp. proc     = 28.6[degC]
Field strength = 11.747379[T]
Pulse mode     = BPPHPPHPPH
Pulse width    = 15.72065221[Hz]
  
```



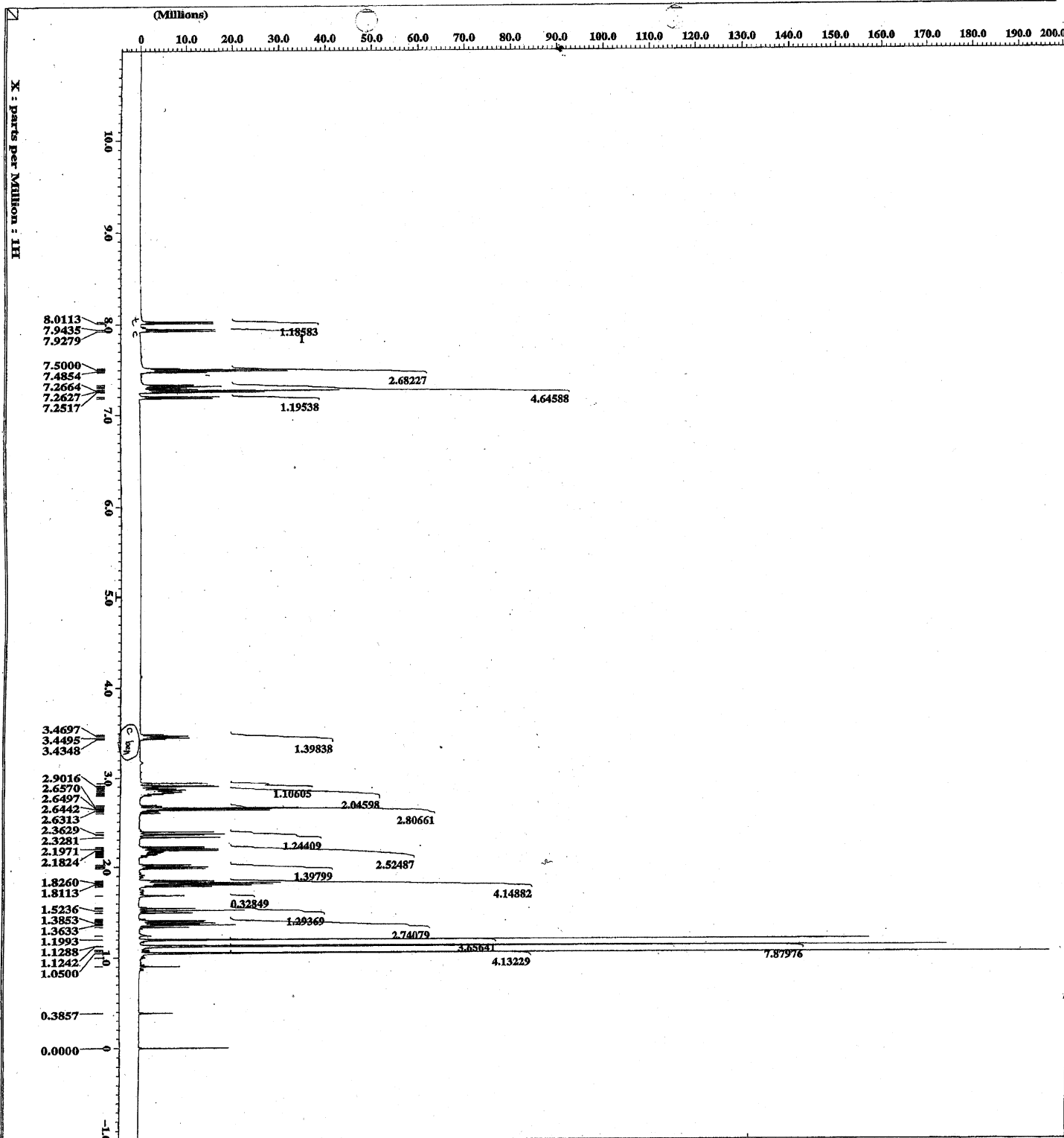
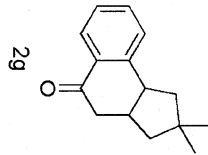
X: parts per Million: 13C



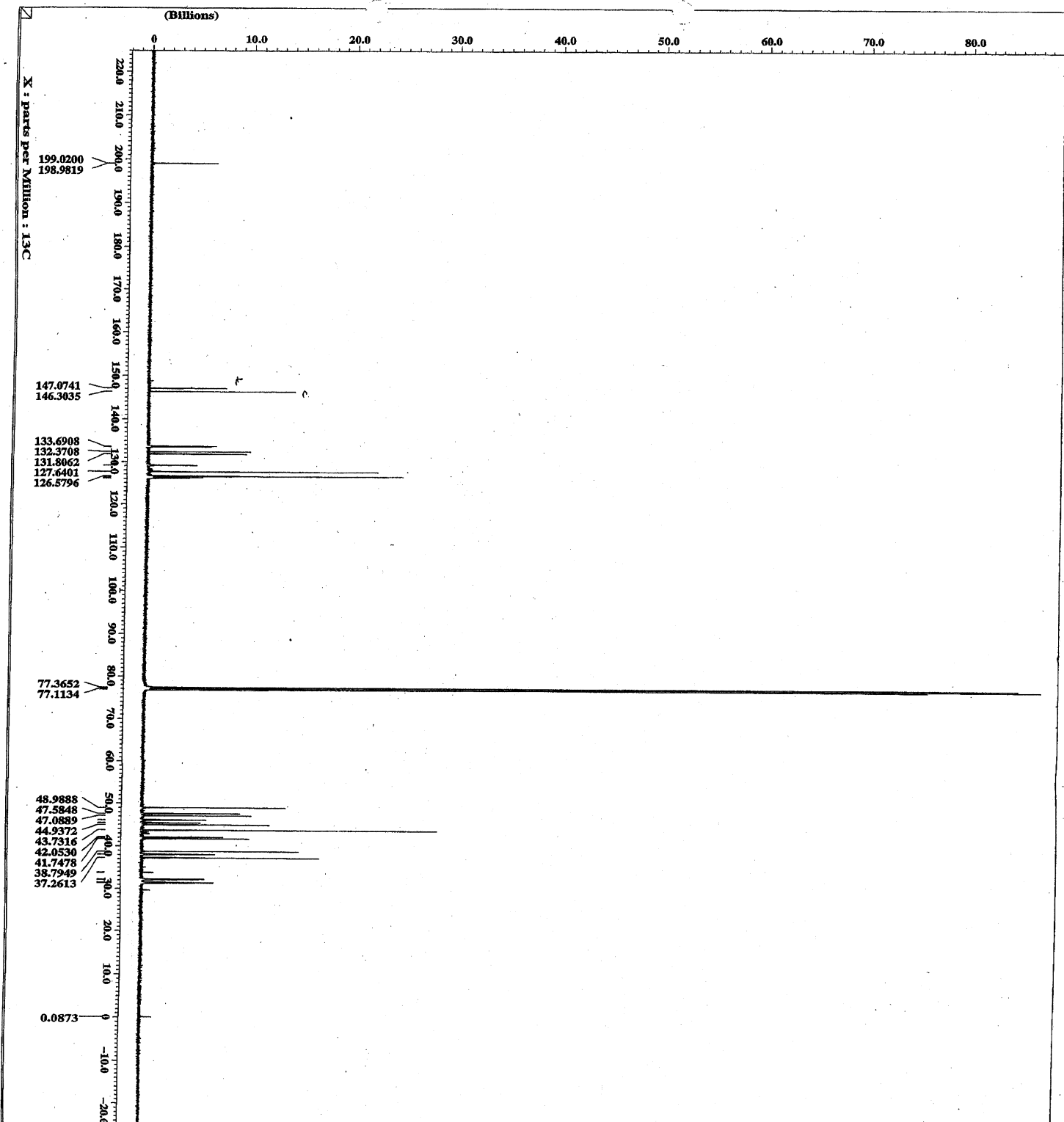
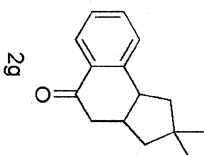
ACQUISITION PARAMETERS
 File Name = 14_13c_spectrum_138
 Sample ID = NUS-7890-013 PFC-C
 Content = Single Pulse with Broad
 Creation Date = 16-Jul-2004 08:53:01
 Acquisition Date = 17-Jul-2004 06:12:15
 Spec Site = 82580
 Spec Type = 13C
 Spec Name = 13C
 Data File = 13C
 Data Size = 32768
 Data Units = [ppm]
 Mod Return = 20000
 X Constant = 13C
 X Offset = 106 [ppm]
 X Freq = 125.77787547 [MHz]
 X Name = 13C
 Solvent = CDCl3
 Spin Seq = 13 [Hz]
 Temp Seq = 23.1 [deg]
 Acq. Start = 10.7673379 [s]
 Acq. End = 15.7206521 [s]
 Filter Width = 15.7206521 [Hz]



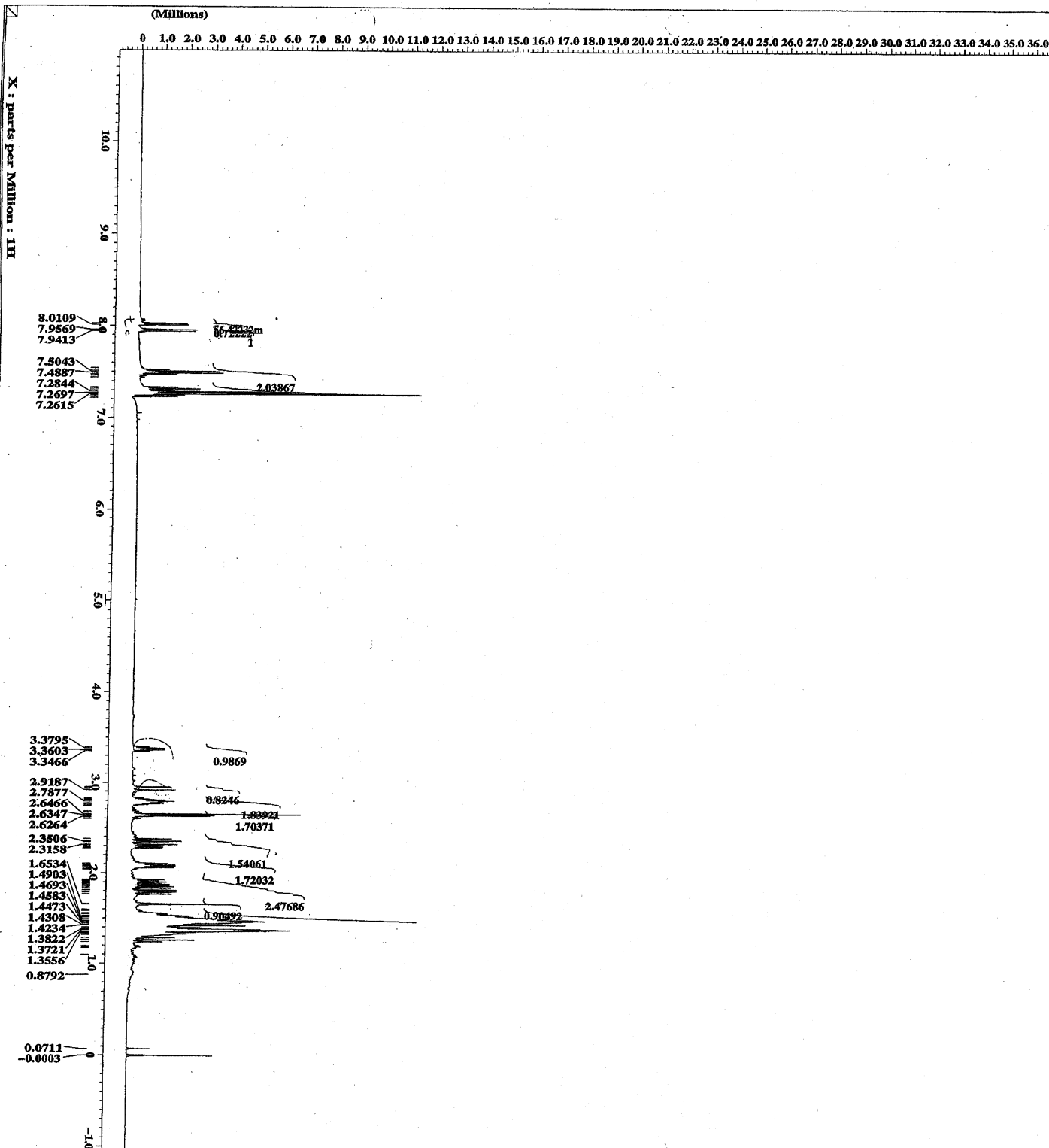
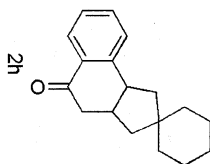
Acquisition Parameters
 File Name: 10_00000001.d
 Sample ID: NMR-2004-035 HPLC-B
 Content: Single Pulse Experiment
 Creation Date: 2004-03-16 16:45:50
 Revision Date: 2004-03-16 16:45:50
 Spec File: NMR2004
 Spec Type: 1D
 Date Acquired: 2004-03-16
 Dimensions: 1H
 Data File: 10_00000001.d
 Data Size: 15384
 Name: 10_00000001.d
 Mod Path: 1
 X Offset: 1H
 X Range: 0.00000000 [Hz]
 X Center: 7.00000000 [Hz]
 Solvent: CHLOROFORM-D
 Spin Set: 14 [Hz]
 Temp Set: 22.9 [C]
 Acq. Time: 11.741579 [s]
 FID File: 10_00000001.fid
 Filter Mode: NOTREMOVED
 Filter Width: 3.7511936 [Hz]



Acquisition Parameters
 File Name = 10_130_spectrum.326
 Sample ID = 10_130-015 HPLC-8
 Content = Single Pulse with Broad
 Creation Date = 3-DEC-2004 09:11:55
 Revision Date = 4-DEC-2004 07:33:01
 Spec File = 10_130-015
 Data Format = 1D COSY
 Data Title = 10_130-015
 Data File = 10_130-015
 Data Path = 10_130-015
 Mod. Name = 10_130-015
 X Domain = 100 [ppm]
 X Offset = 100 [ppm]
 X Freq = 125.76 [MHz]
 Solvent = CHLOROFORM-D
 Spin. Set = 15 [Hz]
 Temp. Set = 24 [C]
 Field Strength = 11.747379 [T]
 Filter Mode = HETCOR
 Filter Width = 15.7206221 [Hz]



ACQUISITION PARAMETERS
 File Name = 14_Spectrum_531
 Author =
 Sample ID = NMR-GENO-032 HYLC-8
 Content = Single Pulse Experiment
 Creation Date = 11-NOV-2004 12:26:41
 Revision Date = 12-NOV-2004 11:30:12
 Spec File = EXPS00
 Spec Type =
 Data Format =
 Dimensions =
 Dia Size =
 Dia Size =
 Dia Size =
 Mod. return =
 X. domain =
 X. offset =
 X. speed =
 Solvent =
 Spin proc =
 Temp. proc =
 Field strength =
 Field mode =
 Filter mode =
 Filter width =



Acquisition Parameters
 File Name = 14_13c_spectrum.114
 Sample ID = NIS-TEMP-012 HPC-2
 Content = Single Pulse with Broad
 Creation Date = 12-NOV-2004 08:24:15
 Revision Date = 12-NOV-2004 07:23:35
 Spec File = EXP500
 Spec Type = 1D NMR
 Spec Name = 1D COMPLEX
 Dimension = 1
 Dim File = 13c
 Dim Size = 32768
 Dim Units = [ppm]
 Ref. File = 13c
 Ref. Units = [ppm]
 X Offset = 100 [ppm]
 X Range = 125.7778547 [MHz]
 X Center = 125.7778547 [MHz]
 Solvent = CDCl3 (non-D)
 Spins_per = 16 [Hz]
 Temp_set = 23.6 [deg]
 Heavy Spin = 30.7473579 [Hz]
 P1 = 12.00 [sec]
 P2 = 15.72066221 [Hz]
 Filter Width = 15.72066221 [Hz]

