Pseudotrienic Acid A and B, two Bioactive Metabolites from *Pseudomonas* sp. MF381-IODS

Anton Pohanka, *† Anders Broberg, † Maria Johansson, [‡] Lennart Kenne, [†] and Jolanta Levenfors^{$\$, \perp}$ </sup>

Department of Chemistry, Swedish University of Agricultural Sciences, P.O. Box 7015, SE-750 07 Uppsala, Sweden, Department of Plant Pathology and Biological Control Unit, Swedish University of Agricultural Sciences, P.O. Box 7035, SE-750 07 Uppsala, Sweden, and Agrivir AB,

P.O. Box 914, SE-751 09 Uppsala, Sweden

NMR and MS data for DDR methanol adduct and pyrrolnitrine		Page S2
NMR spectra for 2:	DQF-COSY	Page S3
	TOCSY	Page S4
	NOESY	Page S5
	DEPT-HSQC	Page S6
	HMBC	Page S7
Experimental data for lactone formation of 2 to 2a		Page S8
¹ H NMR spectra of lactone formation analysis		Page S9

Co-authors are listed in alphabetical order. ^{*}To whom correspondence should be addressed. Tel: +46-18-671549. Fax: +46-18-673476. E-mail: Anton.Pohanka@kemi.slu.se.

[†]Department of Chemistry, Swedish University of Agricultural Sciences

[‡]Plant Pathology and Biological Control Unit, Swedish University of Agricultural Sciences

[§]Agrivir AB

[⊥]Present address: The MASE Laboratories, P.O. Box 148, SE-751 04 Uppsala, Sweden

DDR methanol adduct (3a): White powder; ¹H NMR (CDCl₃, 600 MHz) δ 7.53 (1H, s, H-25), 6.77 (1H, ddd, J = 15.6, 6.5, 6.5 Hz, H-3), 6.60 (1H, dd, J = 15.0 Hz, 10.8, H-20), 6.38 (1H, d, J = 15.0 Hz, H-21), 6.26 (1H, s, H-23), 6.10 (1H, d, J = 10.8 Hz, H-19), 5.70 (1H, d, J = 15.6 Hz, H-2), 5.51 (1H, dd, J = 15.6, 9.0 Hz, H-9), 5.15 (1H, dd, J = 15.6, 8.4 Hz, H-10), 4.80 (1H, dd, J = 10.2, 3 Hz, H-15), 3.71 (3H, s, H-36), 3.25 (1H, d, J = 10.8 Hz, H-17), 3.18 (1H, m, H-7), 3.17 (3H, s, H-32), 3.14 (1H, d, J = 8.4 Hz, H-11), 3.04 (1H, dd, J = 9.6, ~1.5 Hz, H-13), 2.54 (1H, dd, J = 9.6), ~1.5 Hz, H-13), 2.54 (1H, dd, J = 9.6), ~1.5 Hz, H-13), 2.54 (1H, dd, J = 9.6), ~1.5 Hz, H-13), 2.54 (1H, dd, J = 9.6), ~1.5 Hz, H-13), 2.54 (1H, dd, J = 9.6), ~1.5 Hz, H-13), 2.54 (1H, dd, J = 9.6), ~1.5 Hz, H-13), 2.54 (1H, dd, J = 9.6), ~1.5 Hz, H-13), 2.54 (1H, dd, J = 9.6), ~1.5 Hz, H-13), 2.54 (1H, dd, J = 9.6), ~1.5 Hz, H-13), 2.54 (1H, dd, J = 9.6), ~1.5 Hz, H-13), 2.54 (1H, dd, J = 9.6), ~1.5 Hz, H-13), 2.54 (1H, dd, J = 9.6), ~1.5 Hz, H-13), ~1.5 HJ = 15.6, 7.2 Hz, H-27a), 2.46 (3H, s, H-35), 2.44 (1H, m, H-4a), 2.40 (1H, dd, J = 15.6, 7.2 Hz, H-27b), 2.23 (1H, bm, H-5), 2.15 (3H, s, H-34), 2.10 (1H, m, H-16), 2.08 (1H, m, H-4b), 2.05 (1H, m, H-8), 1.98 (1H, m, H-14a), 1.86 (3H, s, H-33), 1.80 (1H, m, H-14b), 1.78 (1H, M, H-6a), 1.35 (3H, s, H-30), 1.07 (1H, m, H-6b), 1.05 (3H, d, J = 6.6 Hz, H-29), 1.00 (3H, d, J = 6.6 Hz, H-31); ¹³C NMR (CDCl₃, 150 MHz) δ173.9 (C, C-28), 165.5 (C, C-1), 161.4 (C, C-26), 146.5 (CH, C-3), 141.3 (CH, C-9), 138.4 (C, C-24), 137.6 (CH, C-21), 137.0 (C, C-22), 136.7 (C, C-18), 135 (CH, C-25), 129.7 (CH, C-19), 125.6 (CH, C-10), 124.9 (CH, C-2), 124.3 (CH, C-20), 120.7 (CH, C-23), 89.4 (CH, C-17), 78.7 (CH, C-13), 74.4 (CH, C-7), 73.5 (CH, C-15), 66.0 (C, C-12), 63.9 (CH, C-11), 56.8 (CH₃, C-32), 51.8 (CH₃, C-36), 45.7 (CH, C-8), 40.8 (CH₂, C-27), 39.5 (CH, C-16), 38.0 (CH₂, C-6), 37.7 (CH₂, C-4), 32.2 (CH, C-5), 32.0 (CH₂, C-14), 17.3 (CH₃, C-29), 14.6 (CH₃, C-34), 13.9 (CH₃, C-35), 11.7 (CH₃, C-33), 11.4 (CH₃, C-30), 10.4 (CH₃, C-31); ESIMS m/z 664.3 [M+Na]⁺.

Pyrrolnitrine (**4**): White powder; ¹H NMR (CDCl₃, 600 MHz) δ8.29 (1H, br s, NH), 7.53 (1H, m, H-4'), 7.45 (1H, m, H-5), 7.43 (1H, m, H-6'), 6.85 (1H, m, H-2), 6.84 (1H, m, H-5); ¹³C NMR (CDCl₃, 150 MHz) δ 149.6 (C, C-2'), 130.6 (CH, C-4'), 130.2 (CH, C-5'), 129.0 (CH, C-6'), 128.2 (C, C-1'), 125.3 (C, C-3'), 117.5 (CH, C-5), 116.7 (CH, C-2), 115.7 (C, C-4), 112.5 (C, C-3); ESIMS *m*/*z* 279 [M+Na]⁺.

DQFCOSY spectrum of 1 (CD₃OD, 600 MHz)





S4





HMBC spectrum of 1 (CD₃OD, 150/600 MHz)



Lactone formation of 2 to 2a:

About 1 mg of **2** was dissolved in 950 μ L 1% TFA in 50/50 MeCN/H₂O and analyzed at 30 min intervals by RP-HPLC with UV-detection at 310 nm (41% A for 15 min, 41% to 100% A in 5 min, 100% A for 10 min; A: aqueous 10 mM ammonium acetate at pH 4.5, B: CH₃CN). Compound **2** eluted at *t*_R=14.1 min and a peak at *t*_R=22.7 min appeared immediately at t=0 and reached maximum area after 60 min. This also correlated with disappearance of the peak at *t*_R=14.1 min. When subjected to LC-ESIMS in negative mode, the peak at *t*_R=22.7 min correlated to *m*/z of 499.5 [M-H]⁻, *i.e.* corresponding to the lactone **2a.** The peak of **2a** was isolated and subjected to ¹H NMR and COSY in CD₃CN/D₂O 50/50 (see page S9, spectrum C). To verify the lactone formation, ¹H NMR and COSY was run on 0.5 mg of **2** dissolved in 500 μ L CD₃CN/D₂O 50/50 (see page S9, spectrum A). Then TFA was added to a concentration of 5% and the mixture was monitored by ¹H NMR and COSY. After five min about 60% of the material had formed lactones (see page S9, spectrum B). Although the high TFA content influenced the HDO resonance the chemical shifts and connectivities could be verified by comparison with the isolated lactone sample from the HPLC experiment and the control spectra acquired prior to addition of TFA.

¹H NMR spectra (600 MHz) of lactone formation analysis with the H-20 (**2**) and H-16 (**2a**) resonance marked. A: ¹H NMR of **2** (CD₃CN/D₂O 50/50); B: ¹H NMR of **2** five min after addition of 5% TFA (CD₃CN/D₂O 50/50, 5% TFA); C: ¹H NMR of **2a** (CD₃CN/D₂O 50/50)

