Supporting Information

The Inverted Cucurbit[*n*]uril Family

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Experimental Section.

General. The guests used in this study were purchased from commercial suppliers and were used without further purification. The crude cucurbit[*n*]uril (CB[*n*]) mixture was prepared according to the literature procedures.^{1,2} Gel permeation chromatography was performed using Sephadex G-15 or Superdex 30. Melting points were measured on a Meltemp apparatus in open capillary tubes and are uncorrected. IR spectra were recorded on a Nicolet Magna or on a Perkin Elmer Spectrum GX FT-IR spectrophotometers as KBr pellets and are reported in cm⁻¹. NMR spectra were measured on a Bruker AM400 or on a Bruker DRX500 spectrometers operating at 400 or 500 MHz for ¹H and 100 or 125 MHz for ¹³C. 2D NMR experiments were performed using the standard pulse sequences supplied by the manufacturer. Mass spectrometry was performed on a JEOL AccuTOF electrospray instrument, or on an ABI 4700 Proteomics Analyzer MALDI-TOF instrument. The formation constants for the inclusion of several guests in *i*CB[7] were determined by isothermal titration calorimetry using a VP-ITC instrument from MicroCal. All solutions were prepared in purified water (Milli-Q, Millipore). A solution (0.2 mM) of *i*CB[7] was placed in the sample cell. As 5 mM solution of guests was added in a series of fifty injections (4 μ L), the heat evolved was recorded at 30 °C.

Purification of iCB[6] Fractional Recrystallization. A crude CB[n] reaction mixture prepared from 80 g of glycoluril by the literature method² was used in subsequent purification steps.

Initial processing. The reaction mixture which contains a large amount of solid was evaporated to a minimum volume. This slurry was poured into water (250 mL). The solid was collected by filtration to give the first crop (Crop 1) (contains: CB[6], CB[7], CB[8], and some *i*CB[6]). The filtrate was evaporated to about 60 mL and then slowly poured into a mixture of MeOH (300 mL) and water (20 mL) with vigorous stirring. After stirring overnight, the precipitate was obtained by filtration to give a second crop (Crop 2 contains CB[7], CB[6], and CB[5]).

Subsequent purification. The separation of each component (CB[5], CB[6], CB[7], CB[8], and *i*CB[6]) from Crop 1 and Crop 2 was enabled due to their differential solubility in HCl solutions.

• *CB*[5] and *CB*[7]. CB[5] and CB[7] were isolated in pure form using the literature procedure¹ which relies on the solubility of both CB[5] and CB[7] in water and the moderate solubility of CB[5] in 50% aqueous MeOH (v/v).

• *CB[8]*. CB[6] and *i*CB[6] have appreciable solubility in 3.5 M HCl solution whereas CB[8] is substantially less soluble. By washing the crude mixture of CB[6], CB[8], and *i*CB[6] with 3.5 M HCl it is possible to isolate CB[8] as an insoluble solid.

• *iCB[6] and CB[6]*. CB[6] and *i*CB[6] were separated by fractional crystallization from different concentration HCl solutions. For example, the initial CB[6] / *i*CB[6] mixture was recrystallized from a minimum volume of conc. HCl. The filtrate is enriched in *i*CB[*n*]; adding the filtrate to MeOH gives the precipitate which is filtered and dried. The solid now enriched in *i*CB[6] is recrystallized from 17.5% HCl which gives CB[6] as a solid and filtrate further enriched in *i*CB[6]. In this manner, the ratio of *i*CB[6]:CB[6] is raised to \approx 80:20. At this point, the mixture is dissolved in a minimum of conc. HCl. To this solution is added enough

 $H_2N(CH_2)_6NH_2$ to complex all of the CB[6] ($\approx 25\%$). The solution is then diluted 5-fold with H_2O . The precipitate is isolated by centrifugation and washed several times with H_2O which yields *i*CB[6] as a white solid in 2% overall yield.

Purification of iCB[7] by Gel Permeation Chromatography.

Initial processing. A partially purified CB[*n*] mixture (20 g) was stirred in aqueous 0.15 M NH₄HCO₃ solution (500 mL) for 3 h. The insoluble solid (mostly CB[6] and *i*CB[6] (~86 %, 5:4), and some CB[8] (7 %)) was filtered off, and methanol (1 L) was added in small portions to the filtrate. The fine precipitate was collected by filtration to give the first crop (contains: CB[7], *i*CB[7] (~94%, 4:1), and some CB[6] and *i*CB[6] (~6%)). This procedure was repeated once to enrich the solid in *i*CB[7] (up to CB[7]:*i*CB[7] \approx 3:2).

Purification of iCB[7] by GPC. A sample enriched in *i*CB[7] (~2 g) dissolved in 10 mL of 0.15 M NH₄HCO₃ solution was injected on a SuperdexTM 30 column (HiLoadTM Prep Grade, 26 x 600 mm). Elution with 0.15 M NH₄HCO₃ solution in flow rate of 2 mL/min, while monitoring at 184 nm, provided baseline separation between CB[7] (retention volume, $R_v = 270$ ~530 mL) and *i*CB[7] ($R_v = 660$ ~890 mL). After evaluating the purity of selected fractions by ¹H NMR spectra, fractions were combined and lyophilized to obtain pure *i*CB[7] (69 mg).

Characterization of iCB[6] and iCB[7].

*i*CB[6]. White solid. M.p. > 300 °C. IR (KBr, cm⁻¹): 3446s, 2994w, 2927w, 2850w, 1735s, 1478s, 1417m, 1377m, 1328m, 1238s, 1192m, 966m, 803s. ¹H NMR (400 MHz, 35% DCl / D₂O): 5.60 (d, J = 8.5, 2H), 5.48 (d, J = 8.5, 2H), 4.50 – 4.45 (ABq, 4H), 5.42 (s, 2H), 5.32 (d, J = 15.7, 4H), 5.30 (d, J = 15.7, 4H), 5.13 (d, J = 14.0, 4H), 5.02 (s, 2H), 4.34 (d, J = 14.0, 4H), 4.32 (d, J = 15.7, 4H), 4.20 (d, J = 15.7, 4H). ¹³C NMR (100 MHz, 35% DCl / D₂O): 157.1, 156.9, 156.3, 156.0, 70.9, 70.6, 70.3, 70.1, 69.8, 62.8, 52.0, 51.7, 51.4. MS (ES): *m/z* 997 (100, [M + H]⁺). HR-MS (ES): *m/z* 997.3219 ([M + H]⁺, C₃₆H₃₇N₂₄O₁₂, calcd 997.3023). Anal. Calcd for C₃₆H₃₆N₂₄O₁₂•(H₂O)₆ : C 39.13, H 4.37, N 30.42. Found: C 39.38, H 4.38, N 30.24.

*i*CB[7]. White solid. M.p. > 350 °C. IR (KBr, cm⁻¹): 3445s, 2994w, 2928w, 1734s, 1474s, 1420m, 1377m, 1325m, 1233m, 1192m, 967m, 807s. ¹H NMR (500 MHz, D₂O/NaCl, TSP): 5.82 (d, J = 15.2, 6H), 5.81 (d, J = 15.6, 4H), 5.71 (d, J = 8.5, 2H), 5.65 – 5.55 (m, 10H), 5.48 (d, J = 13.7, 4H), 5.39 (br, 2H), 4.49 (d, J = 13.7, 4H), 4.37 (d, J = 15.6, 4H), 4.30 (d, J = 15.2, 4H), 4.28 (d, J = 14.0, 2H). ¹³C NMR (125 MHz, D₂O): 157.5, 157.0, 156.9, 156.7, 71.9, 71.8, 71.7, 71.4, 71.0, 64.4, 53.3, 53.1, 52.7, 52.5. MS (MALDI-TOF): m/z 1163 ([M + H]⁺). HR-MS (MALDI-TOF): m/z 1163.3186 (100, [M + H]⁺, C₄₂H₄₃N₂₈O₁₄, calcd 1163.3508). Anal. Calcd for C₄₂H₄₂N₂₈O₁₄•(H₂O)₉ : C 38.06, H 4.56, N 29.59. Found: C 38.22, H 4.82, N 29.69.

References:

- (1) Kim, J.; Jung, I. S.; Kim, S.-Y.; Lee, E.; Kang, J.-K.; Sakamoto, S; Yamaguchi, K.; Kim, K. *J. Am. Chem. Soc.* **2000**, *122*, 540-541.
- (2) Day, A. I.; Arnold, A. P.; Blanch, R. J.; Snushall, B. J. Org. Chem. 2001, 66, 8094-8100.

X-ray crystallography.

A colorless plate of *i*CB[6] with approximate dimensions $0.022 \times 0.230 \times 0.380 \text{ mm}^3$, was used for the X-ray crystallographic analysis. The X-ray intensity data were measured at 213(2) K on a three-circle diffractometer system equipped with a Bruker Smart1000 CCD area detector using a graphite monochromator and a MoK α fine-focus sealed tube (λ = 0.71073 Å). Data were corrected for absorption effects with the semi-empirical method using SADABS. The structure was solved and refined using the SHELXS-97 and SHELXL-97 software. The final anisotropic full-matrix least-squares refinement on F^2 converged to the R values listed in Table S1. The diffraction data from a colorless block-shaped crystal of *i*CB[7] measuring 0.21 \times 0.19 \times 0.06 mm³ mounted on the loop were collected at 100 K on a ADSC Quantum 210 CCD diffractometer with synchrotron radiation ($\lambda = 1.00000$ Å) at Macromolecular Crystallography Wiggler Beamline 4A, Pohang Accelerator Laboratory (PAL), Pohang, Korea. The crystal was rotated through a total of 180°. The autoindexing procedure performed with DENZO indicated that the crystals belong to a rhombohedral space group, with unit-cell parameters a = 32.200(5)Å, c = 32.581(7) Å, $\gamma = 120^{\circ}$. The raw data were processed and scaled using the program HKL2000. The space group was determined to be R-3. A total of 15222 measured reflections were merged into 6769 independent reflections. The structure was solved by directed methods and refined by full-matrix least-squares method implemented in SHELXTL program package. All the non-hydrogen atoms were refined anisotropically except included THF. Hydrogen atoms were added to their geometrically ideal positions. The crystallographic data are summarized in Table S2.



Figure S1. Another views of iCB[6] (left) and iCB[7] (right). Anisotropic atomic displacement ellipsoids for the non-hydrogen atoms are shown at the 50% probability level. A THF molecule is encapsulated in the cavity of iCB[7]. Water molecules and Cl⁻ are omitted for clarity.

Empirical formula	C36 H56.40 Cl N24 O21.70	
Formula weight	1208.10	
Temperature	213(2) K	
Wavelength	0.71073 Å	
Crystal system	Monoclinic	
Space group	P 2 ₁	
Unit cell dimensions	$a = 12.434(2)$ Å $\alpha = 90^{\circ}$	
	$b = 16.122(3)$ Å $\beta = 104.923(4)^{\circ}$	
	$c = 12.667(2) \text{ Å}$ $\gamma = 90^{\circ}$	
Volume	$2453.6(8) \text{ Å}^{3}$	
Z	2	
Density, ρ_{calc}	1.635 g/cm^3	
Absorption coefficient, µ	0.188 mm^{-1}	
F(000)	1262	
Crystal size	$0.380 \ 0.230 \times 0.022 \ \text{mm}^3$	
Index ranges	$-14 \le h \le 10, -18 \le k \le 19, -14 \le l \le 14$	
Reflections collected	8493	
Independent reflections	7318 [R(int) = 0.0193]	
Observed reflection, $I \ge 2\sigma(I)$	5794	
Data / restraints / parameters	7318 / 34 / 850	
Goodness-of-fit on F^2	0.998	
Final R indices [I>2sigma(I)]	$R_1 = 0.0431$, $wR_2 = 0.0998$	
R indices (all data)	$R_1 = 0.0649, wR_2 = 0.1129$	
Largest diff. peak and hole	0.400 and -0.311 e^{-1}/A^{3}	

Table S1. X-ray crystal data for $iCB[6] \cdot H_3O^+ \cdot Cl^- \cdot 8.7H_2O$

 $R_1 = \Sigma ||F_o| - |F_c|| / \Sigma |F_o|, \quad wR2 = [\Sigma w (F_o^2 - F_c^2)^2 / \Sigma w (F_o^2)^2]^{1/2}$

Empirical formula Formula weight Temperature Wavelength Crystal system Space group	C46 H78 N28 O29 1487.36 100(2) K 1.00000 Å ^a Rhombohedral R-3	
Unit cell dimensions	a = 32.200(5) Å b = 32.200(5) Å	$\alpha = 90^{\circ}$ $\beta = 90^{\circ}$
	c = 32.581(7) Å	$\gamma = 120^{\circ}$
Volume Z	29255(8) Å ³ 18	
Density (calculated)	1.520 g/cm^3	
Absorption coefficient F(000)	0.301 mm ⁻¹ 14076	
Crystal size	$0.21 \times 0.19 \times 0.06 \text{ mm}^3$	
Index ranges	$-18 \le h \le 32, -32 \le k \le 21, -32 \le l \le 27$	
Reflections collected	15222	
Independent reflections	6769 [R(int) = 0.0992]	
Observed reflection, $I \ge 2\sigma(I)$	5794	
Data / restraints / parameters	6769 / 217 / 1002	
Goodness-of-fit on F^2	1.613	
Final R indices [I>2sigma(I)]	$R_1 = 0.0967, wR_2 = 0.317$	9
R indices (all data) Extinction coefficient	$R_1 = 0.0979, wR_2 = 0.322$ 0.00046(9)	6
Largest diff. peak and hole	1.173 and -0.797 $e^{-}/\text{\AA}^{3}$	

Table S2. X-ray crystal data for *i*CB[7]•THF•14H₂O.

$$\begin{split} &R_1 = \Sigma ||F_o| - |F_c|| / \Sigma |F_o|, \ wR2 = [\Sigma w (F_o^2 - F_c^2)^2 / \Sigma w (F_o^2)^2]^{1/2} \\ ^a \ The x-ray \ data \ were \ collected \ with \ synchrotron \ radiation \ at \ Macromolecular \ Crystallography \ Wiggler \ Beamline \ 4A, \ Pohang \ Accelerator \ Laboratory \ (PAL). \end{split}$$



Figure S2. ES-MS spectrum of *i*CB[6].



Figure S3. ¹³C NMR spectrum (100 MHz, RT, 35% DC1 / D₂O) for *i*CB[6].



Figure S5. ¹H NMR spectra of *i*CB[6] complexed with **1**.



Figure S6. Complexation induced shifts of *i*CB[6] protons upon complexation with **1**.



Figure S7. DQF-COSY and ROESY spectra of *i*CB[6]•1.



Figure S8. Intermolecular NOEs between *i*CB[6] and **1** in *i*CB[6]•**1**.



Figure S9. ${}^{1}\text{H}{}^{-13}\text{C}$ HSQC spectra of *i*CB[6]•1 and *i*CB[6].



Figure S10. MALDI-TOF spectrum of *i*CB[6]•1.



Figure S11. ¹H NMR spectra (0.1 M Na₂SO₄ / D_2O) for: *i*CB[6] alone (bottom) and with excess 1,6-hexanediamine (top).



Figure S13. MALDI-TOF spectrum of *i*CB[7]. CHCA = α -cyano-4-hydroxycinnamic acid.



Figure S14. DQF-COSY and ROESY spectra of *i*CB[7].



Figure S15. ¹H NMR of *i*CB[7]•1.



Figure S16. Complexation induced shifts of *i*CB[7] protons upon complexation with **1**.



Figure S17. ¹H NMR spectra of CB[7] (top) and *i*CB[7] (bottom) with a slight excess of **2**. In the presence of CB[7] (top), there are two sets of NMR signals corresponding to free **2** and complexed **2** with CB[7], which indicates that the exchange of **2** in and out of CB[7] is slow on the NMR time scale whereas the spectrum of *i*CB[7] (bottom) with a slight excess of **2** shows only one set of NMR signals of **2**, indicating the fast exchange of **2** in and out of CB[7].



Figure S18. Thermogram (top) and binding isotherm (bottom) of (ferrocenemethyl)trimethylammonium ion complexing with *i*CB[7] at 303K.