

Supplemental Information

Instrument-free synthesizable fabrication of label-free optical biosensing paper strips for the early detection of infectious keratoconjunctivitis

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■ Raman spectrum of 2-NAT molecule

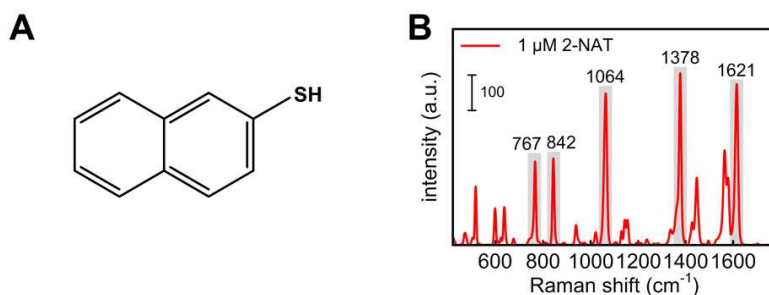


Figure S1. (A) The molecular structure of 2-naphthalenethiol (2-NAT) and (B) Raman spectra of a 1 μM 2-NAT molecule for GNPs deposited on paper with optimal SILAR condition. The prominent Raman peaks of the 2-NAT molecule were obtained at 767 cm⁻¹ (C-H wag), 842 cm⁻¹ (C-H twist), 1064 cm⁻¹ (symmetric C-H bend), 1378 cm⁻¹ (ring stretch), and 1621 cm⁻¹ (ring stretch).

■ Distribution of GNPs with number of SILAR cycles (10 mM SILAR reagents)

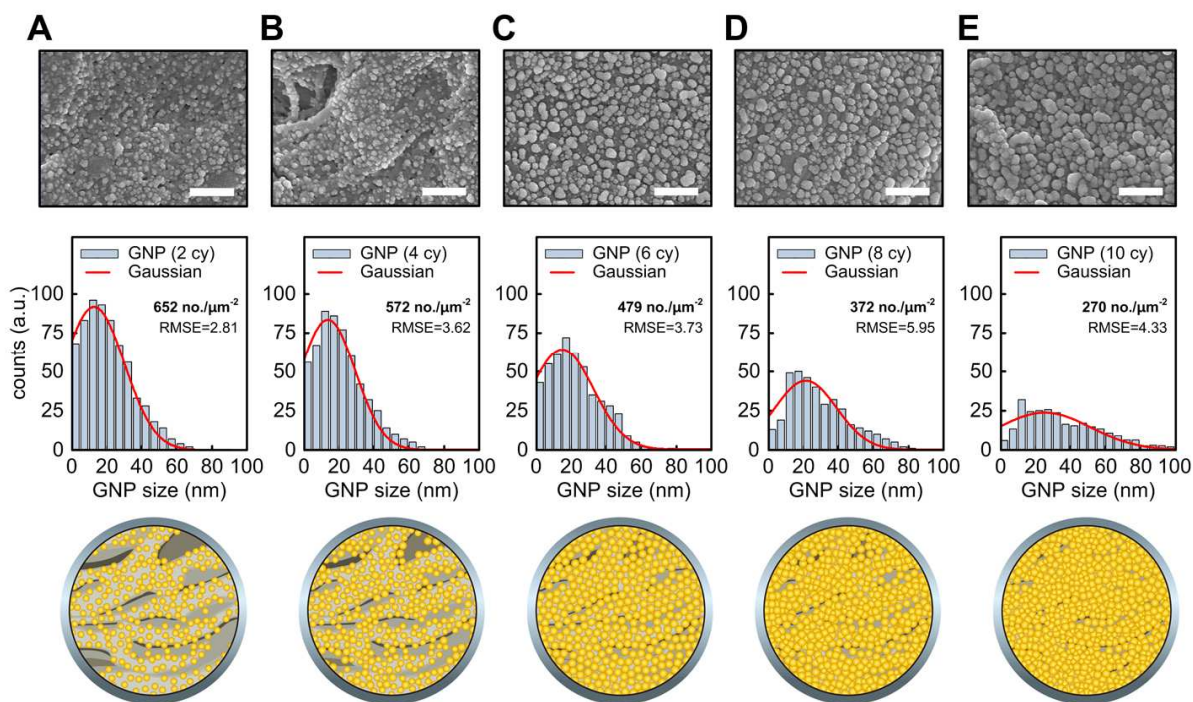


Figure S2. Distributions of SILAR-synthesized GNPs deposited on paper with (A) two, (B) four, (C) six, (D) eight, and (E) 10 SILAR cycles (top: SEM. Scale bar=250 nm; middle: GNP distribution; bottom: schematic distribution). The surface density of GNP was represented by the number of GNP per unit area (no./μm²). RMSE indicates the root mean square error between the Gaussian-predicted data and experimental data.

■ Distribution of SILAR reagent-concentrated GNPs with number of SILAR cycles

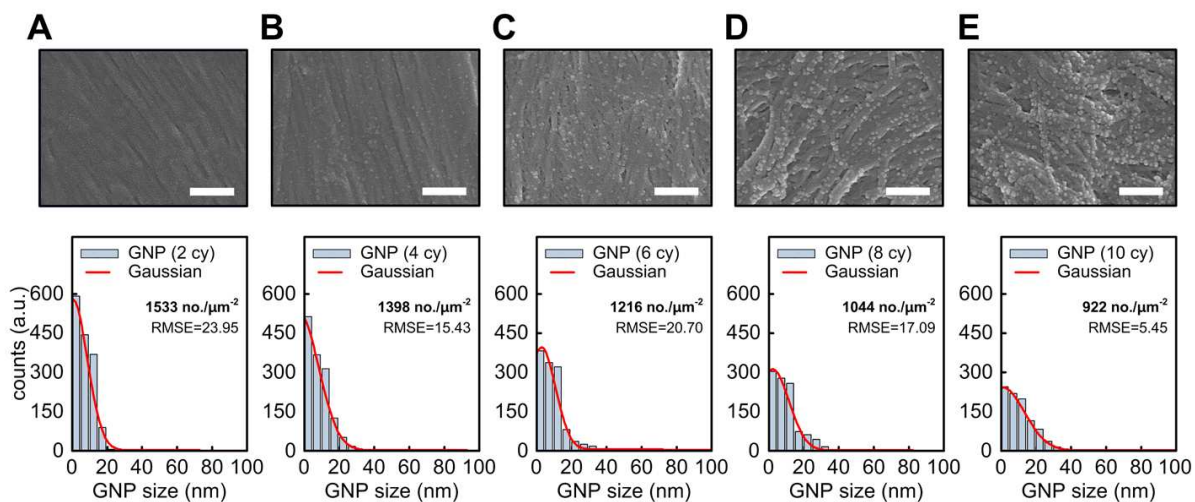


Figure S3. Distributions of SILAR-synthesized GNPs deposited on paper with (A) two, (B) four, (C) six, (D) eight, and (E) 10 SILAR using 1 mM HAuCl₄ and NaBH₄ SILAR reagents. (top: SEM. Scale bar=250 nm; bottom: GNP distribution). The surface density of GNP was represented by the number of GNP per unit area (no./μm²). RMSE indicates the root mean square error between the Gaussian-predicted data and experimental data.

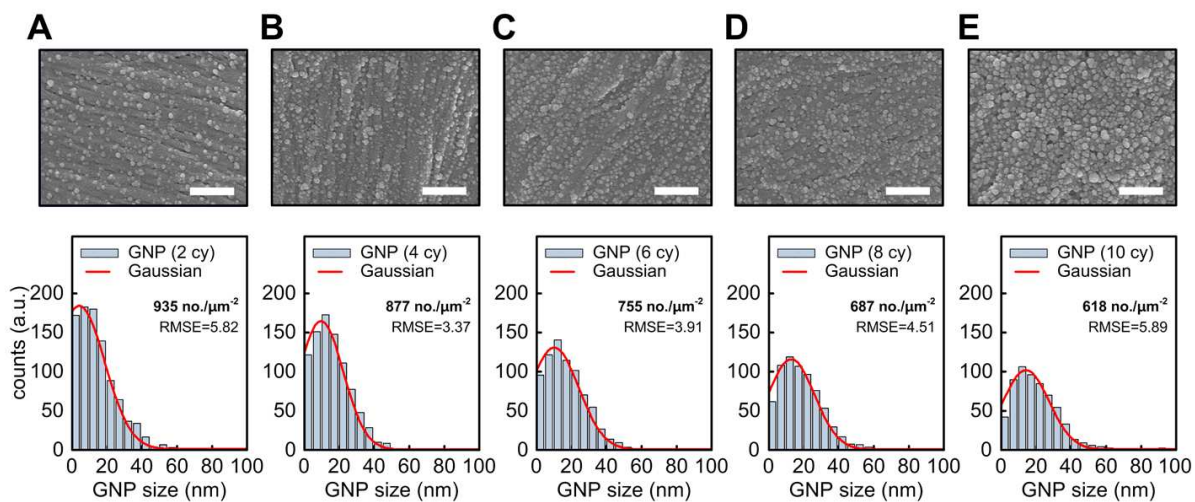


Figure S4. Distributions of SILAR-synthesized GNPs deposited on paper with (A) two, (B) four, (C) six, (D) eight, and (E) 10 SILAR using 5 mM HAuCl₄ and NaBH₄ SILAR reagents. (top: SEM. Scale bar=250 nm; bottom: GNP distribution). The surface density of GNP was represented by the number of GNP per unit area (no./μm²). RMSE indicates the root mean square error between the Gaussian-predicted data and experimental data.

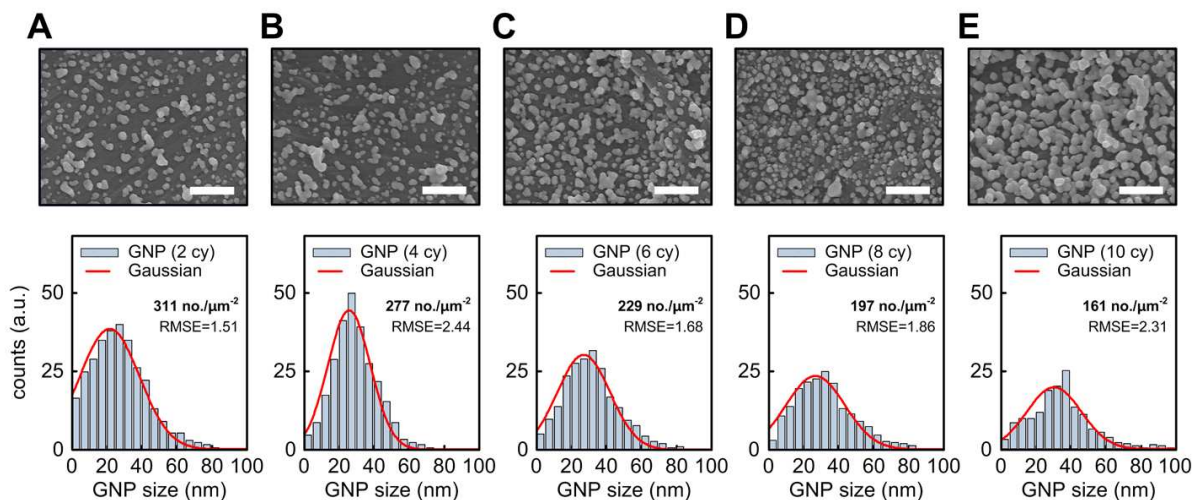


Figure S5. Distributions of SILAR-synthesized GNPs deposited on paper with (A) two, (B) four, (C) six, (D) eight, and (E) 10 SILAR using 20 mM SILAR HAuCl₄ and NaBH₄ reagents. (top: SEM. Scale bar=250 nm; bottom: GNP distribution). The surface density of GNP was represented by the number of GNP per unit area (no./μm²). RMSE indicates the root mean square error between the Gaussian-predicted data and experimental data.

■ Computational modeling

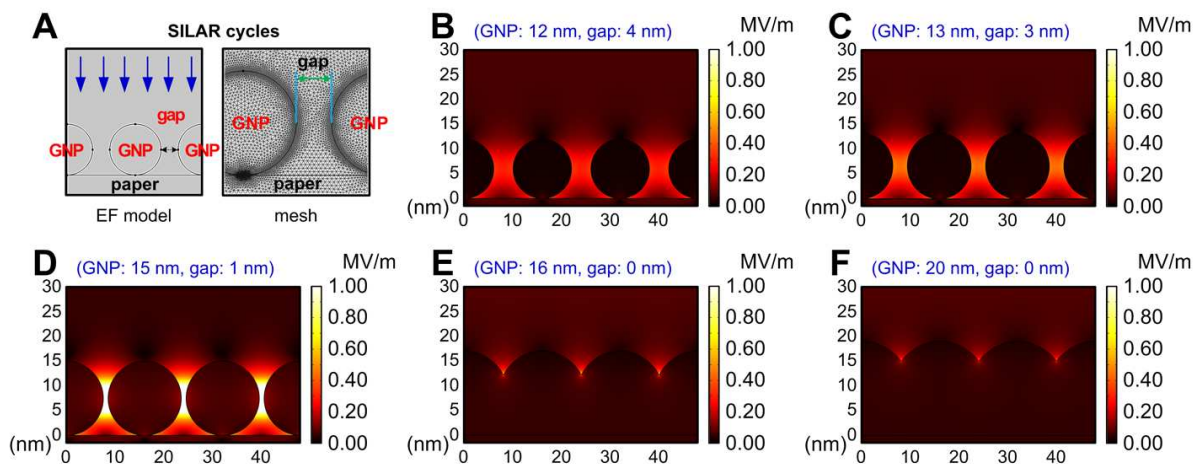


Figure S6. (A) Finite element model of GNPs with SILAR cycles and computational results of GNP diameter and interparticle gap distance-dependent LSPR effect with (B) two, (C) four, (D) six, (E) eight, and (F) 10 SILAR cycles on the electromagnetic field (EF).

■ Sensitivity

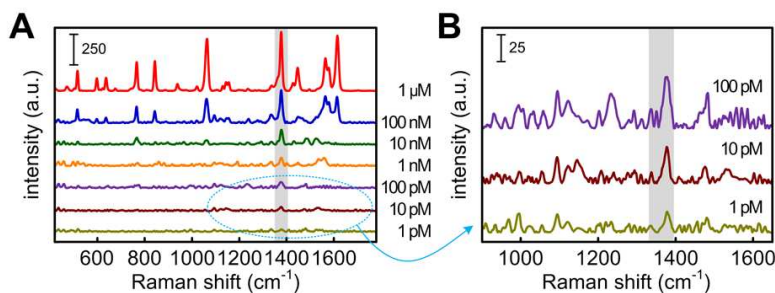


Figure S7. Representative SERS spectra with (A) different 2-NAT concentrations ($10^{-12} \sim 10^{-6}$ M) and (B) low concentrations of 2-NAT on SILAR-synthesized GNPs deposited on SERS paper. Gray indicates a 2-NAT molecule-characterized peak at 1378 cm^{-1} .

■ Enhancement factor

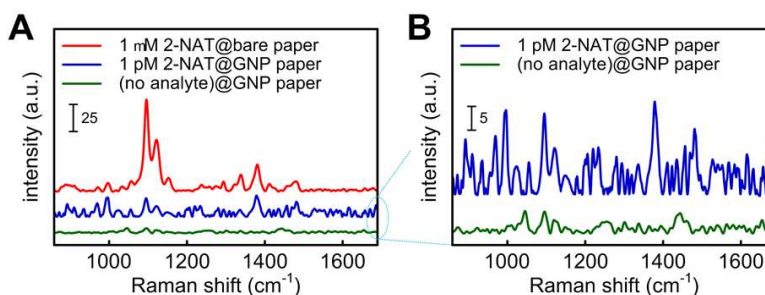


Figure S8. Raman spectra of 1 mM 2-NAT on bare paper and 1 pM 2-NAT and no analyte on SILAR-synthesized GNP paper strip.

The enhancement factor (EF) was calculated as the difference in Raman intensity between two different substrates as

$$EF = \left(\frac{I_{\text{SERS}}}{I_{\text{bare}}} \right) \left(\frac{N_{\text{bare}}}{N_{\text{SERS}}} \right), \quad (\text{S1})$$

where I_{SERS} and I_{bare} are the Raman intensity of the molecule on the SERS and bare papers, respectively, and N_{SERS} and N_{bare} are the average number of adsorbed molecules in the scattering volume for SERS and non-SERS areas, respectively.¹ Assuming that the probe molecules were uniformly distributed on the substrates, the number of adsorbed molecules can be estimated as

$$N = \left(N_A \cdot c \cdot \frac{V_{\text{droplet}}}{A_{\text{spot}}} \right) A_{\text{laser}}, \quad (\text{S2})$$

where N_A is Avogadro's constant, c is the concentration of the probe molecule, V is the volume of the molecule droplet, A_{spot} is the size of the substrate, and A_{laser} is the size of the laser spot.² Since the same methods for assessing the Raman measurement were applied to two substrates, the parameters of N_A for 2-NAT, V , A_{spot} , and A_{laser} were the same.

Therefore, Eq. (S2) can be written as

$$EF = \left(\frac{I_{SERS}}{I_{bare}} \right) \left(\frac{c_{bare}}{c_{SERS}} \right), \quad (S3)$$

where c_{SERS} and c_R are the concentration of 2-NAT molecule on the GNP and bare papers, respectively.

■ Bio-applications

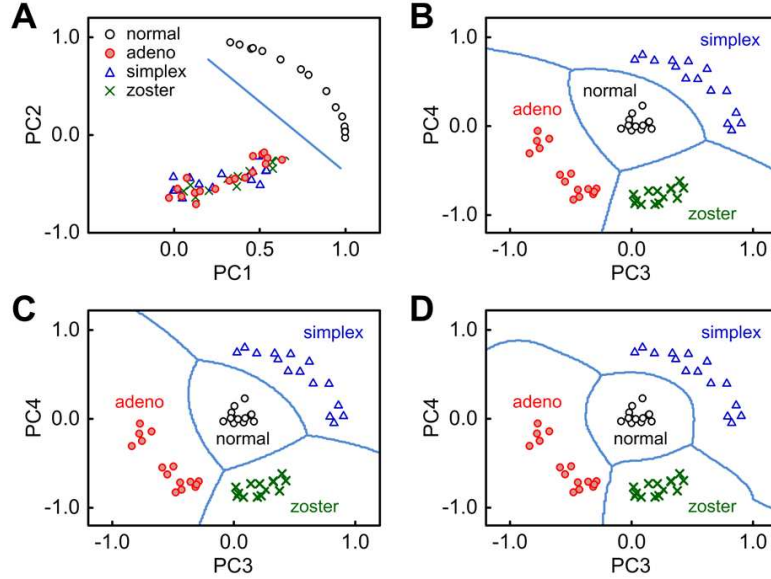


Figure S9. PCA-SVM scores (n=15, each): (A) to detect the presence of keratoconjunctivitis using SVM classifier with linear kernel and to classify the normal eye and keratoconjunctivides using SVM classifier with (B) the second-order polynomial kernel ($d=2$, Eq. S4), (C) Gaussian kernel ($\sigma=2$, Eq. S5), and (D) Hilbert transform radial basis function kernel ($\rho=2$, $a=1$, $b=2$, Eq. S6).^{3,4}

$$K_{poly}(\mathbf{x}_1, \mathbf{x}_2) = (\langle \mathbf{x}_1 \cdot \mathbf{x}_2 \rangle + 1)^d \quad (S4)$$

$$K_{Gaussian}(\mathbf{x}_1, \mathbf{x}_2) = \exp\left(-\|\mathbf{x}_1 - \mathbf{x}_2\|^2 / 2\sigma^2\right) \quad (S5)$$

$$K_{HiBRF}(\mathbf{x}_1, \mathbf{x}_2) = \exp\left(-\rho \sum_i |\mathbf{x}_{1i}^a - \mathbf{x}_{2i}^a|^b\right) \quad (S6)$$

■ References

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- (3) Choi, S.; Jiang, Z. *Comput. Biol. Med.* **2010**, *40* (1), 8–20.
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