

Supporting Information

Homo- and Heterobimetallic Ruthenium(II) and Osmium(II) Complexes Based on a Pyrene-Biimidazolate Spacer as Efficient DNA- Binding Probes in the Near-Infrared Domain

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Physical Measurements. Elemental analyses of the compounds were performed with a Vario-Micro V2.0.11 elemental (CHNSO) analyzer. NMR spectra were collected on either a Bruker 300 or Bruker 500 spectrometer in DMSO-*d*₆, and high resolution mass spectroscopy was performed on a Waters Xevo G2 QTOF mass spectrometer. The UV/vis absorption spectra were recorded with a Shimadzu UV 1800 spectrometer. A matched pair of quartz cuvettes (path length 1 cm) was employed. Steady state luminescence spectra were obtained either by a Perkin–Elmer LS55 or Spex fluorolog-2 spectrofluorometer equipped with DM3000F software. Luminescence quantum yields were determined by using literature method taking [Ru(bpy)₃]²⁺ as the standard. Luminescence lifetime measurements were carried out by using time–correlated single photon counting set up from Horiba Jobin-Yvon. The luminescence decay data were collected on a Hamamatsu MCP photomultiplier (R3809) and were analyzed by using IBH DAS6 software. Cyclic and square-wave voltammetric experiments were performed in deaerated acetonitrile with a BAS epsilon electrochemistry system and a three-electrode set up consisting of a platinum or glassy carbon working electrode, a platinum counter electrode, and Ag/AgCl reference electrode. Tetraethylammonium perchlorate (TEAP) was used as background electrolyte. The potentials reported in this study were referenced against the Ag/AgCl electrode, which under the given experimental conditions gave a value of 0.36 V for the Fc/Fc⁺ couple.

DNA Binding Experiments

Absorption and Emission Spectral Experiments. UV-vis absorption and emission titrations were carried out by maintaining a constant metal complex concentration (20 μM) and adding incrementally the CT-DNA solution of appropriate concentration in 5 mM Tris-HCl/ NaCl buffer with the ionic strength of 50 .0 × 10⁻³ m (pH 7.30) at room temperature. After each addition of DNA to the solution of the metal complexes, the resulting solution was allowed to equilibrate at 25 °C for 5 min, after which the absorption and emission readings were noted. Proper corrections were made to the absorbance of CT-DNA. The equilibrium binding constant (K_b) and the binding site size (s, per base pair) of the complexes to CT-DNA were calculated by non-linear least square analysis of the isotherm using the expression of Bard and co-workers based on the McGhee-von Hippel (MvH) model.^{S1-S2}

$$(\epsilon_a - \epsilon_f)/(\epsilon_b - \epsilon_f) = (b - (b^2 - 2K_b^2 C_t [DNA]/s)^{1/2})/2K_b C_t \quad (S1)$$

$$(I_a - I_f)/(I_b - I_f) = (b - (b^2 - 2K_b^2 C_t [DNA]/s)^{1/2})/2K_b C_t \quad (S2)$$

where $b = 1 + K_b C_t + K_b [DNA]/2s$, ϵ_a is the extinction coefficient observed for the spectral band at a given DNA concentration, ϵ_f is the extinction coefficient of the free complex in solution, ϵ_b is the extinction coefficient of the complex when fully bound to DNA. Similarly I_a , I_f and I_b are the luminescence intensities of the complex at a given DNA concentration, free complex and fully bound to DNA respectively. K_b is the equilibrium binding constant, C_t is the total complex concentration, $[DNA]$ is the DNA concentration in nucleotides and s is the binding site size of the complexes in base pairs. The non-linear least-square fit analysis was done using Origin Lab software. The lifetimes of the complexes were also recorded as a function of CT-DNA in 5 mM Tris-HCl/ NaCl buffer with the ionic strength of 50.0×10^{-3} m (pH 7.30) at room temperature.

Competitive Binding Fluorescence Experiments. The apparent binding constants (K_{app}) of the complexes have been evaluated by using ethidium bromide (EB)-bound CT-DNA solution in 5 mM Tris-HCl/ NaCl buffer with the ionic strength of 50.0×10^{-3} m (pH 7.30) at room temperature. The luminescence intensities at 602 nm (546 nm excitation) of EB-DNA conjugate with increasing concentration of **1**, **2** and **3** were recorded. The K_{app} values were calculated by using the equation $K_{EB} \times [EB] = K_{app} \times [\text{complex}]$, where K_{EB} is the binding constant of EB ($K_{EB} = 1.25 \times 10^6 \text{ M}^{-1}$), $[EB]$ is the concentration of EB (13 μM) and $[\text{complex}]$ is the concentration of the complex at 50% reduction of the initial fluorescence emission intensity.^{S3}

Circular Dichroism Experiments. The Circular dichroism (CD) spectroscopy was studied on a JASCO J-815 CD spectropolarimeter between 400 and 200 nm in continuous scanning mode at 25°C. Experiments were performed by adding progressively increasing amounts of complexes (**1**, **2** and **3**) to the solutions of CT-DNA [40 μM] in 5 mM Tris-HCl/ NaCl buffer with the ionic strength of 50.0×10^{-3} m (pH 7.30).

X-ray Crystallographic Analyses. The crystallographic data, details of data collection, and refinement parameter for the complexes **1** and **2** were summarized in Table S1. In each case, single crystals of suitable size were obtained by diffusing toluene to 1:1 acetonitrile-dichloromethane solution of the complexes. The crystals were immersed in paratone oil and then mounted on the tip of a glass fiber and cemented using epoxy resin. Intensity data for these crystals were collected using MoK α ($\lambda = 0.7107 \text{ \AA}$) radiation on a Bruker SMART APEX II diffractometer equipped with CCD area detector at room temperature. The data integration and reduction were processed with SAINT^{S4} software provided with the software package of SMART APEX II. An empirical absorption correction was applied to the collected reflections with SADABS.^{S4} The structures were solved by direct

methods using SHELXTL^{S5} and were refined on F² by the full-matrix least-squares technique using the SHELXL-97^{S6} program package. Graphics were generated using PLATON.^{S7} In both cases, the non-hydrogen atoms were refined anisotropically until the convergence. All the hydrogen atoms were geometrically positioned and treated as riding atoms. In both complexes **1** and **2**, the hydrogen atoms of the lattice water molecules could not be located from the difference Fourier map.

CCDC reference numbers are 1439503 for **1** and 1439504 for **2**.

Theoretical Computational Methods. Quantum chemical calculations were performed with the Gaussian 09 program^{S8} employing the DFT method with Becke's three-parameter hybrid functional and Lee-Yang-Parr's gradient corrected correlation functional B3LYP level of theory.^{S9-S10} The 6-31G(d) basis set was employed for the C, H and N while SDD basis set was used for Ru and Os atoms.^{S11} Geometries were fully optimized using the criteria of the respective programs. To compute the UV-vis transition of the compounds, the singlet excited state geometries corresponding to the vertical excitations were optimized using the time-dependent DFT (TDDFT) scheme starting with the ground state geometries optimized in solution phase.^{S12-S15} The excitation energies, computed within the acetonitrile solvent simulated by the CPCM model,^{S16} has been determined by using the so-called nonequilibrium approach, which has been designed for the study of the absorption process.^{S17-S18} Only singlet-singlet transitions, that is, the spin-allowed transitions, have been taken into account. UKS calculations were also performed directly on triplet state of the complexes to calculate singlet-triplet energy gap in CH₃CN using the CPCM model. Orbital analysis was completed with Gauss View^{S19} and Gauss sum 2.2.^{S20}

Table S1. Crystallographic Data for Complex **1** and **2**

Crystallographic table		
	1	2
formula	C ₆₀ H ₄₂ N ₁₂ Ru ₂ Cl ₂ O ₈	C ₆₇ H ₅₀ Cl ₂ N ₁₂ O ₈ Os ₂
fw	1332.10	1602.55
T (K)	273(2)	273(2)
Cryst syst	Monoclinic	Monoclinic
Space group	P2(1)/n	P2(1)/n
<i>a</i> (Å)	11.7951(6)	11.8080(4)
<i>b</i> (Å)	27.9283(13)	27.8673(9)
<i>c</i> (Å)	23.3331(12)	23.4456(8)
α (deg)	90	90
β (deg)	100.952(3)	101.537(2)
γ (deg)	90	90
<i>V</i> (Å ³)	7546.3(6)	7559.1(4)
<i>Dc</i> (g cm ⁻³)	1.172	1.367
Z	4	4
μ (mm ⁻¹)	0.522	3.483
F(000)	2688.0	3068.0
θ range (deg)	2.24-25.00	2.25-27.00
data/restraints/params	13287/0/757	16473/0/809
GOF on F ²	1.029	0.941
R1 ^a [<i>I</i> > 2σ(<i>I</i>)], wR2 ^b (all data)	0.0831 0.2363	0.0616 0.1438
$\Delta\rho_{\max}/\Delta\rho_{\min}$ (e Å)	2.362/-0.394	1.004/-1.061
^a R1(<i>F</i>) = [$\sum \ F_0 - F_C\ / \sum F_0 $], ^b wR2 (<i>F</i> ²) = [$\sum w(F_0^2 - F_C^2)^2 / \sum w(F_0^2)^2$] ^{1/2}		

Table S2. Selected Calculated Bond Distances (\AA) for **1**, **2** and **3** in Ground State and UKS State Along with Available X-ray Crystal Data

	1				2				3		
	Expt	Soln.(1)	uks.(1)		Expt	Soln.(2)	uks.(2)		Soln.(3)	uks(3)	
Ru1-N7	2.022(7)	2.072	2.104	Os1-N7	2.033(7)	2.084	2.071	Ru1-N7	2.080	2.082	
Ru1-N8	2.032(6)	2.054	2.070	Os1-N8	2.014(6)	2.063	2.068	Ru1-N8	2.060	2.058	
Ru1-N5	2.043(6)	2.081	2.081	Os1-N5	2.033(7)	2.087	2.109	Ru1-N5	2.088	2.084	
Ru1-N6	2.046(6)	2.087	2.084	Os1-N6	2.039(7)	2.097	2.128	Ru1-N6	2.093	2.096	
Ru1-N1	2.118(6)	2.137	2.108	Os1-N1	2.11196)	2.153	2.092	Ru1-N1	2.145	2.155	
Ru1-N3	2.198(6)	2.298	2.242	Os1-N3	2.191(6)	2.293	2.284	Ru1-N3	2.306	2.317	
Ru2-N12	2.013(6)	2.054	2.059	Os2-N12	2.010(6)	2.063	2.062	Os1-N12	2.065	2.070	
Ru2-N10	2.030(6)	2.087	2.097	Os2-N10	2.050(7)	2.097	2.099	Os1-N10	2.097	2.127	
Ru2-N11	2.042(6)	2.072	2.082	Os2-N11	2.046(7)	2.084	2.087	Os1-N11	2.084	2.072	
Ru2-N9	2.045(6)	2.081	2.087	Os2-N9	2.037(7)	2.087	2.084	Os1-N9	2.089	2.111	
Ru2-N2	2.110(6)	2.137	2.147	Os2-N2	2.093(7)	2.153	2.161	Os1-N2	2.146	2.086	
Ru2-N4	2.205(6)	2.298	2.310	Os2-N4	2.223(7)	2.293	2.302	Os1-N4	2.284	2.273	

Table S3. Selected Calculated Bond Angles (\AA) for **1**, **2** and **3** in Ground State and UKS State Along with Available X-ray Crystal Data

	1				2				3		
	Expt	Soln.(1)	uks(1)		Expt	Soln.(2)	uks(2)		Soln.(3)	uks.(3)	
N7-Ru1-N8	79.1(3)	78.8	78.2	N7-Os1-N8	79.3(3)	78.0	79.1	N7-Ru1-N8	78.6	78.7	
N7-Ru1-N5	98.1(3)	97.7	98.5	N7-Os1-N5	98.4(3)	98.6	97.0	N7-Ru1-N5	97.9	97.8	
N8-Ru1-N5	92.5(2)	88.9	85.9	N8-Os1-N5	89.0(3)	88.8	85.7	N8-Ru1-N5	89.0	89.3	
N7-Ru1-N6	176.8(3)	175.4	175.8	N7-Os1-N6	177.1(3)	175.6	173.7	N7-Ru1-N6	175.4	175.4	
N8-Ru1-N6	99.1(3)	98.7	98.2	N8-Os1-N6	100.5(3)	99.6	99.9	N8-Ru1-N6	98.8	98.8	
N5-Ru1-N6	79.2(3)	78.2	78.8	N5-Os1-N6	78.7(3)	77.5	76.6	N5-Ru1-N6	78.0	78.1	
N7-Ru1-N1	88.2(2)	88.9	87.3	N7-Os1-N1	87.5(3)	88.4	89.5	N7-Ru1-N1	88.8	88.6	
N8-Ru1-N1	87.5(2)	90.7	92.4	N8-Os1-N1	89.2(3)	91.4	91.4	N8-Ru1-N1	90.9	91.1	
N5-Ru1-N1	173.6(3)	173.0	173.4	N5-Os1-N1	173.4(3)	172.7	172.1	N5-Ru1-N1	173.0	173.4	
N6-Ru1-N1	94.4(3)	94.8	95.1	N6-Os1-N1	95.4(3)	95.3	96.6	N6-Ru1-N1	95.0	95.3	
N7-Ru1-N3	95.7(3)	94.7	96.9	N7-Os1-N3	95.1(3)	95.4	96.8	N7-Ru1-N3	95.1	95.5	
N8-Ru1-N3	167.4(2)	168.5	171.8	N8-Os1-N3	168.2(2)	168.3	170.2	N8-Ru1-N3	168.8	169.3	
N5-Ru1-N3	99.7(2)	101.4	101.3	N5-Os1-N3	102.2(2)	101.7	103.6	N5-Ru1-N3	101.0	100.4	
N6-Ru1-N3	86.7(2)	88.2	86.8	N6-Os1-N3	85.6(2)	87.4	85.0	N6-Ru1-N3	87.8	87.4	
N1-Ru1-N3	80.8(2)	79.5	80.7	N1-Os1-N3	80.1(2)	78.6	79.6	N1-Ru1-N3	79.5	79.5	
N12-Ru2-N10	99.0(2)	101.6	98.7	N12-Os2-N10	100.8(3)	99.6	99.5	N12-Os1-N10	99.6	99.9	
N12-Ru2-N11	80.2(3)	78.8	78.7	N12-Os2-N11	78.5(3)	78.0	78.0	N12-Os1-N11	78.0	79.0	
N10-Ru2-N11	177.3(3)	175.4	175.3	N10-Os2-N11	177.3(3)	175.6	175.5	N10-Os1-N11	175.5	173.8	
N12-Ru2-N9	88.5(2)	88.9	88.9	N12-Os2-N9	93.6(2)	88.8	89.0	N12-Os1-N9	88.6	85.5	
N10-Ru2-N9	79.4(3)	78.2	78.1	N10-Os2-N9	78.4(3)	77.5	77.5	N10-Os1-N9	77.4	76.6	
N11-Ru2-N9	98.0(3)	97.7	97.8	N11-Os2-N9	99.1(3)	98.6	98.5	N11-Os1-N9	98.6	97.2	
N12-Ru2-N2	89.5(2)	90.7	91.1	N12-Os2-N2	87.1(2)	91.4	91.6	N12-Os1-N2	91.4	91.5	
N10-Ru2-N2	94.6(2)	94.8	95.4	N10-Os2-N2	95.3(3)	95.3	95.6	N10-Os1-N2	95.3	96.6	
N11-Ru2-N2	88.0(2)	88.9	88.5	N11-Os2-N2	87.2(2)	88.4	88.2	N11-Os1-N2	88.5	89.4	
N9-Ru2-N2	173.3(2)	173.0	173.5	N9-Os2-N2	173.7(3)	172.7	173.1	N9-Os1-N2	172.6	172.0	
N12-Ru2-N4	168.4(2)	168.5	168.9	N12-Os2-N4	167.3(2)	168.3	168.8	N12-Os1-N4	168.1	170.1	
N10-Ru2-N4	86.7(2)	88.2	87.7	N10-Os2-N4	85.8(3)	87.4	87.1	N10-Os1-N4	87.4	85.1	
N11-Ru2-N4	94.6(3)	94.7	95.4	N11-Os2-N4	95.4(3)	95.4	95.8	N11-Os1-N4	95.4	96.8	
N9-Ru2-N4	102.5(2)	101.4	101.2	N9-Os2-N4	98.4(2)	101.7	101.2	N9-Os1-N4	102.2	103.9	
N2-Ru2-N4	79.9(2)	79.5	79.2	N2-Os2-N4	81.5(2)	78.6	78.6	N2-Os1-N4	78.4	79.4	

Table S4. Fractional Intensities of Short and Long Components of **1** and **3**

		Fractional Intensities(α)	
Compound		α_1	α_2
Ru-Ru(1)		0.71	0.28
Ru-Os(3)	Ru-center	0.78	0.21
	Os-center	0.79	0.20

Table S5. Selected Molecular Orbital Along with their Energies and Compositions for **1- 3** in Solution Phase

	Energy/ eV	(%) Composition			
MO	1	1(%) Composition			
		Ru ^{II}	pyrene imida	imida	bpy
LUMO+3	-2.35	7.65	0.31	0.05	91.96
LUMO+2	-2.38	7.65	0.37	0.09	91.87
LUMO+1	-2.42	4.23	0.21	0.26	95.29
LUMO	-2.44	3.70	0.28	0.35	95.64
HOMO	-4.97	8.93	69.18	20.80	1.07
HOMO-1	-5.48	74.35	0.69	13.24	11.70
HOMO-2	-5.58	65.64	24.81	0.06	9.46
HOMO-3	-5.63	72.89	10.97	2.25	13.87
MO	2	2(%) Composition			
		Os ^{II}	pyrene imida	imida	bpy
LUMO+3	-2.28	9.78	0.33	0.09	89.77
LUMO+2	-2.31	9.68	0.43	0.17	89.70
LUMO+1	-2.39	5.67	0.39	0.32	93.60
LUMO	-2.42	4.81	0.47	0.52	94.17
HOMO	-4.97	19.51	58.89	18.57	3.01
HOMO-1	-5.24	74.51	0.24	11.18	14.05
HOMO-2	-5.37	69.19	17.56	0.27	12.96
HOMO-3	-5.43	67.21	13.51	2.41	16.85
MO	3	3(%) Composition			
		Ru ^{II}	Os ^{II}	pyrene imida	imida
LUMO+3	-2.28	0.56	9.03	0.38	0.11
LUMO+2	-2.34	6.45	0.74	0.32	0.08
LUMO+1	-2.38	1.94	2.80	0.29	0.29
LUMO	-2.41	1.64	2.48	0.36	0.41
HOMO	-4.95	2.80	13.33	61.88	19.56
HOMO-1	-5.31	11.13	58.13	7.73	9.99
HOMO-2	-5.43	9.11	58.75	18.27	1.25
HOMO-3	-5.55	27.67	44.97	3.50	3.32

Table S6. Phosphorescence Emission of the Complexes **1-3** in Acetonitrile According to UKS-Calculations and Associated Experimental Values

	Energy for singlet state (E1/Hartree)	Energy for triplet state (E2/Hartree)	Emission energy ΔE=(E2- E1)/(eV)	λemission (theo)/nm	λemission (expt)/nm
1	-3158.6240	-3158.5586	1.77	696	719
2	-3150.2064	-3150.1487	1.57	790	827
3	-3154.4161	-3154.3591	1.55	799	820

Table S7. Selected Molecular Orbital Along with their Energies and Compositions for **1- 3** in UKS Optimised State

	Energy/ eV	(%) Composition			
MO	1	1(%) Composition			
		Ru ^{II}	pyrene imida	imida	bpy
LUMO+3	-2.33	6.69	0.26	0.06	92.97
LUMO+2	-2.36	5.51	0.05	0.19	94.23
LUMO+1	-2.39	3.54	0.23	0.26	95.95
LUMO	-2.66	6.31	0.52	0.11	93.03
HOMO	-4.91	9.72	67.36	21.79	1.11
HOMO-1	-5.47	75.54	2.12	11.98	10.34
HOMO-2	-5.56	65.43	23.44	1.19	9.91
HOMO-3	-5.63	72.12	13.08	2.24	12.54
MO	2	2(%) Composition			
		Os ^{II}	pyrene imida	imida	bpy
LUMO+3	-2.29	9.53	0.34	0.12	89.99
LUMO+2	-2.32	8.30	0.57	0.14	90.97
LUMO+1	-2.41	5.19	0.40	0.39	93.99
LUMO	-2.67	7.76	0.17	0.52	91.53
HOMO	-4.94	25.17	52.46	17.90	4.44
HOMO-1	-5.18	67.94	6.49	12.04	13.52
HOMO-2	-5.38	69.77	17.14	0.52	12.55
HOMO-3	-5.42	65.53	16.00	2.70	15.76
MO	3	3(%) Composition			
		Ru ^{II}	Os ^{II}	pyrene imida	imida
LUMO+3	-2.30	0.60	7.56	0.53	0.10
LUMO+2	-2.34	6.37	0.61	0.31	0.08
LUMO+1	-2.40	3.46	0.09	0.22	0.26
LUMO	-2.66	0.06	7.78	0.17	0.52
HOMO	-4.92	1.59	23.10	52.11	18.73
HOMO-1	-5.23	9.58	47.82	19.60	11.33
HOMO-2	-5.45	9.50	58.84	20.00	1.03
HOMO-3	-5.55	28.42	44.18	2.52	3.91
					20.94

Table S8. Changes in Lifetime Values of **1-3** in presence of Increasing DNA Concentration

Compound	Lifetime Changes in Presence of DNA
1	$\tau_1 = 5.6 \rightarrow 5.5$ $\tau_2 = 13.6 \rightarrow 26.5$
2	$\tau_1 = 0.1 \rightarrow 0.4$ $\tau_2 = 1.1 \rightarrow 3.6$
3	$\tau_1 = 0.4 \rightarrow 1.1$ $\tau_2 = 2.8 \rightarrow 5.1$

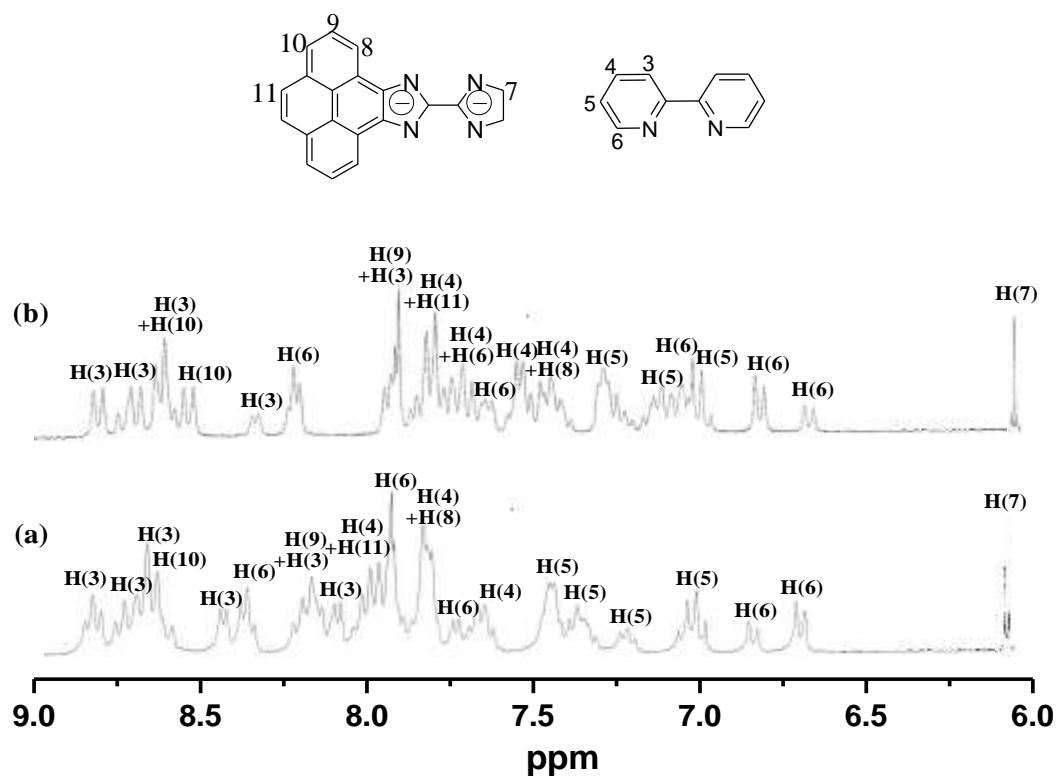


Figure S1. ¹H NMR spectra of complex **1** (a) and **2** (b) (300 MHz) in DMSO-*d*₆.

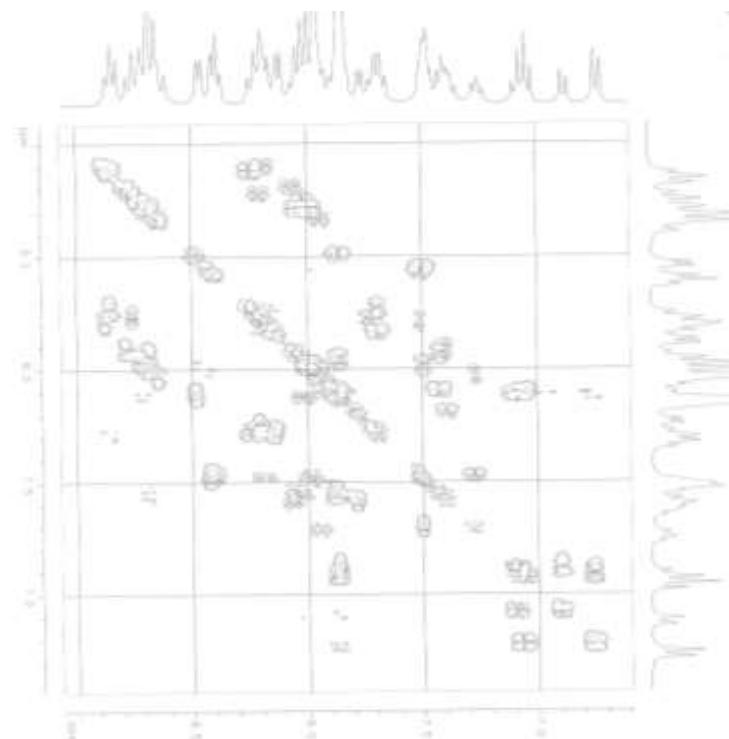


Figure S2. ¹H-¹H COSY NMR spectrum of **1** in DMSO-*d*₆.

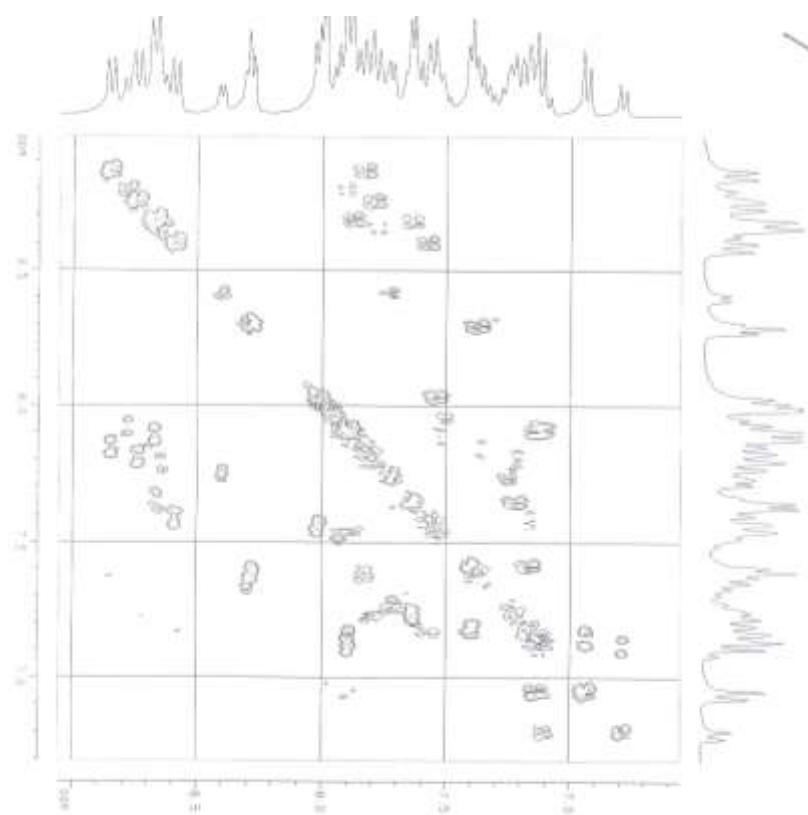


Figure S3. ^1H - ^1H COSY NMR spectrum of **2** in $\text{DMSO}-d_6$.

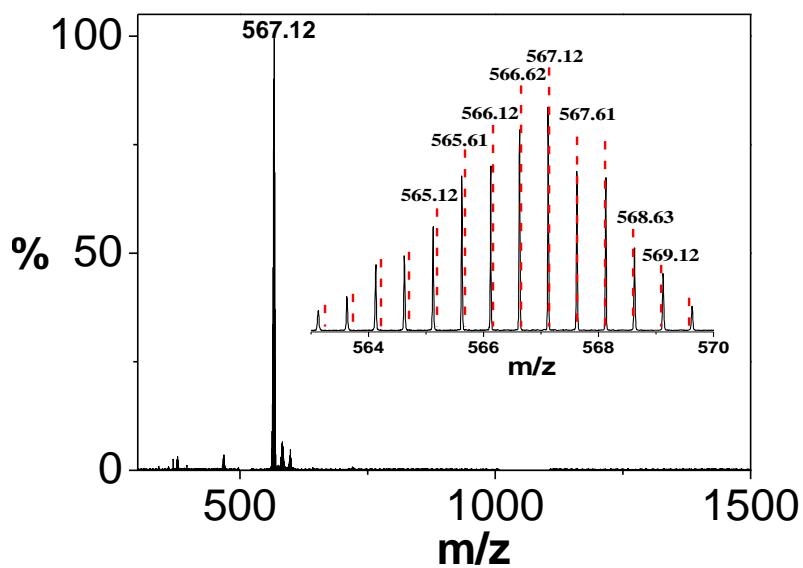


Figure S4. ESI-MS (positive) for the complex cations $[(\text{bpy})_2\text{Ru}(\text{Py-biimz})\text{Ru}(\text{bpy})_2]^{2+}$ ($m/z = 567.12$) in acetonitrile showing the observed and isotopic distribution patterns.

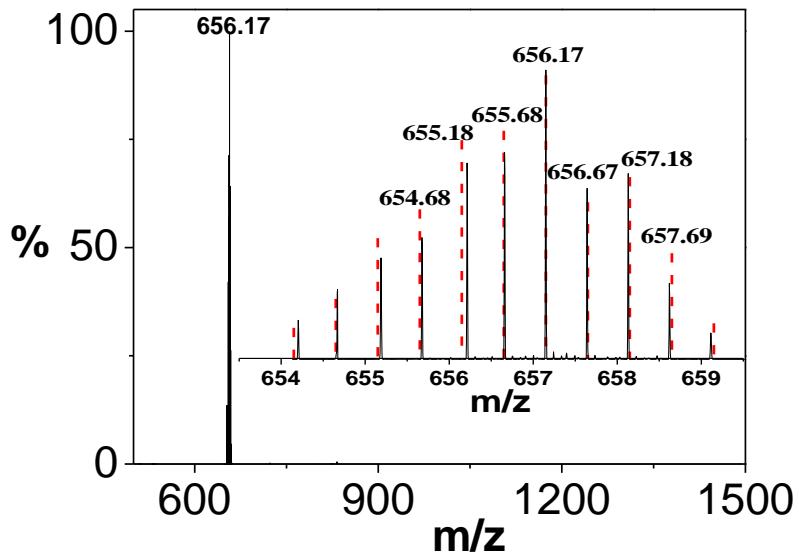


Figure S5. ESI-MS (positive) for the complex cations $[(\text{bpy})_2\text{Os}(\text{Py}-\text{biimz})\text{Os}(\text{bpy})_2]^{2+}$ ($m/z = 656.17$) in acetonitrile showing the observed and isotopic distribution patterns.

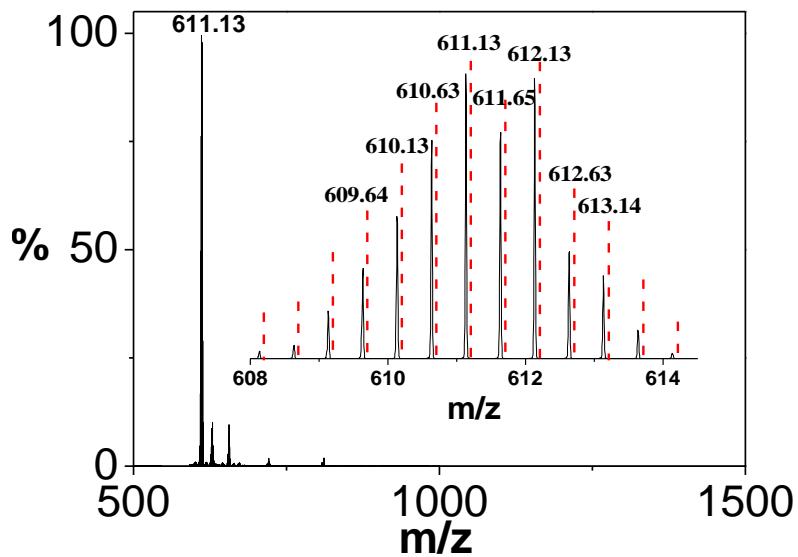


Figure S6. ESI-MS (positive) for the complex cations $[(\text{bpy})_2\text{Ru}(\text{Py}-\text{biimz})\text{Os}(\text{bpy})_2]^{2+}$ ($m/z = 611.13$) in acetonitrile showing the observed and isotopic distribution patterns.

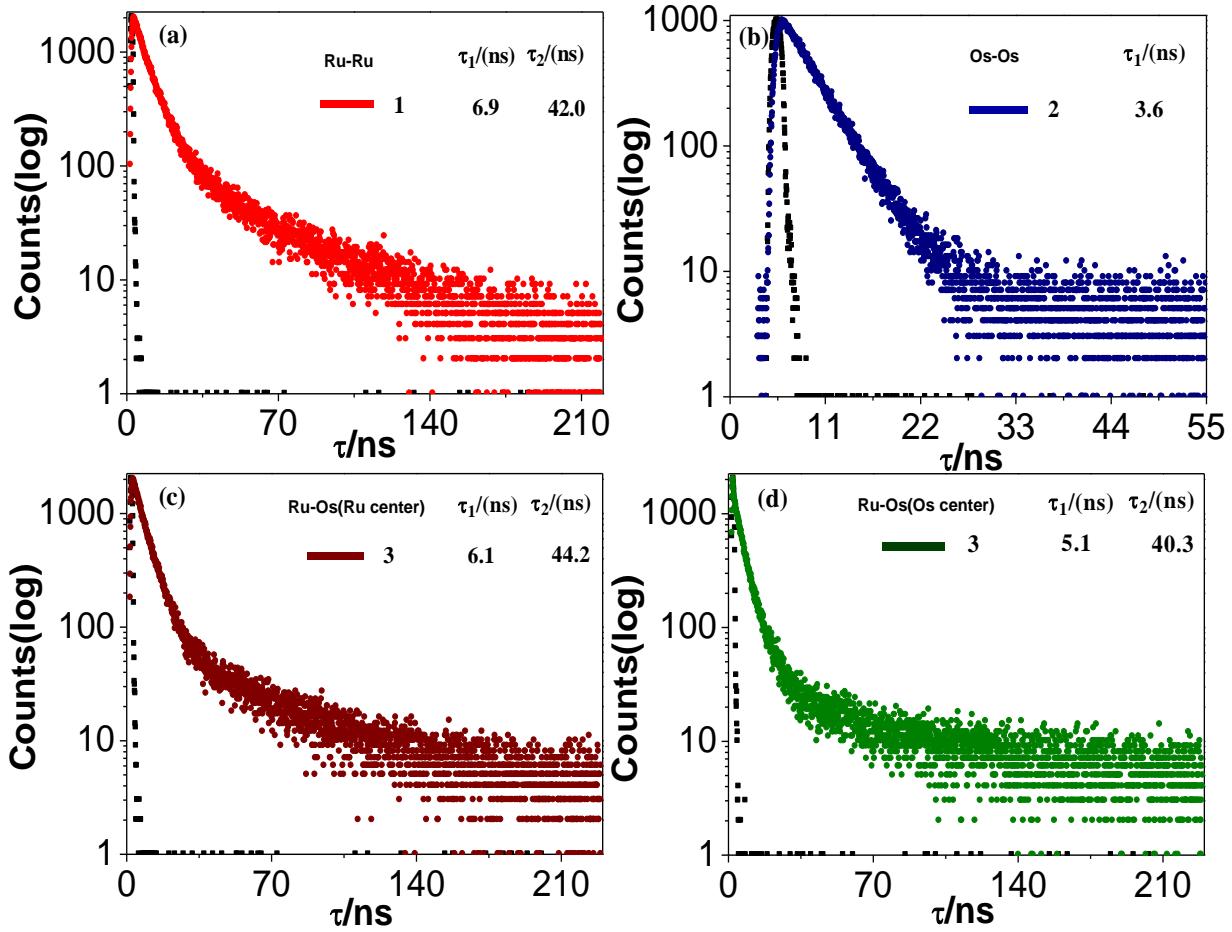


Figure S7. Time resolved emission spectra of **1-3** in their free state in acetonitrile solution.

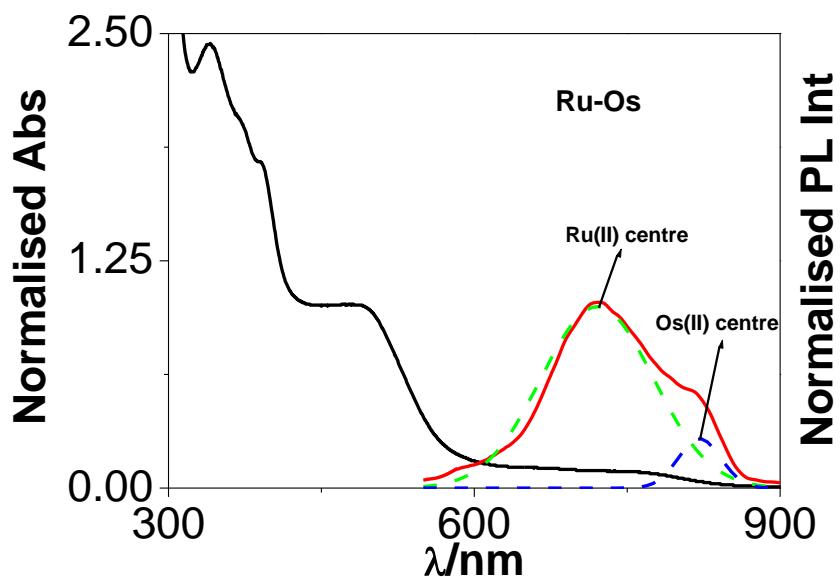


Figure S8. Overlay of normalized absorption and emission spectra of the hetero dimmer **3**. Dotted lines indicate the deconvoluted emission spectra of **3**.

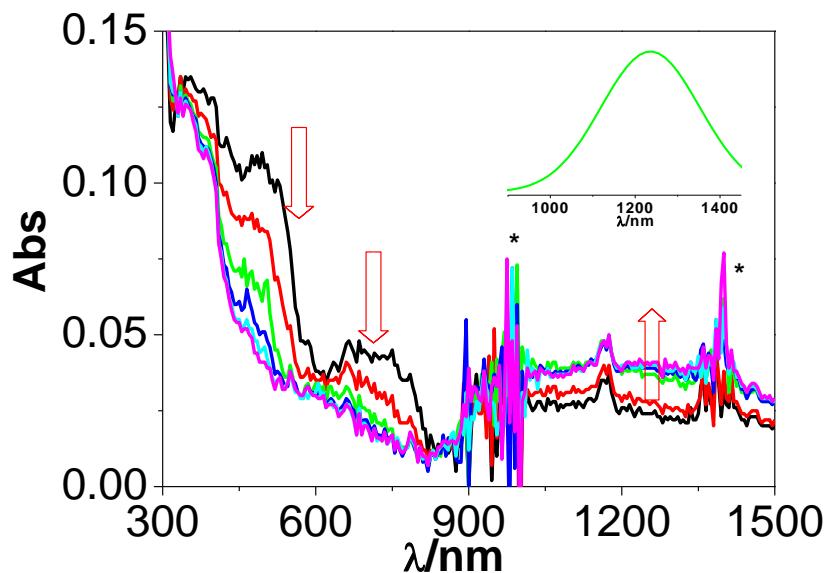


Figure S9. Spectroelectrochemical changes during the oxidation of $[(\text{bpy})_2\text{Os}(\text{Py}-\text{Biimz})\text{Os}(\text{bpy})_2]^{2+}$ (**2**) in acetonitrile. The IVCT band obtained from spectral deconvolution is shown in the inset. Asterisks denote artefacts due to non-perfect background compensation.

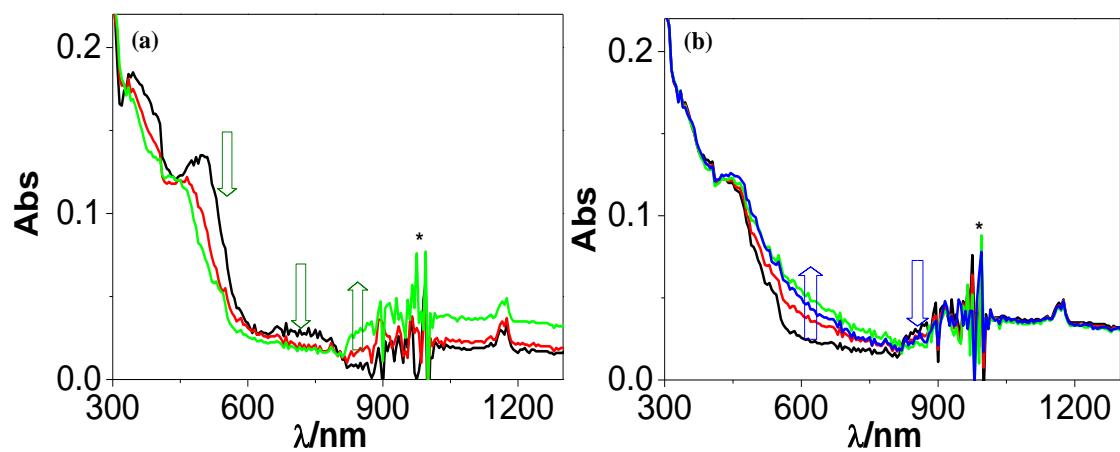


Figure S10. Spectroelectrochemical changes during the oxidation of $[(\text{bpy})_2\text{Ru}(\text{Py}-\text{Biimz})\text{Os}(\text{bpy})_2]^{2+}$ (**3**) in acetonitrile. Asterisks denote artefacts due to non-perfect background compensation.

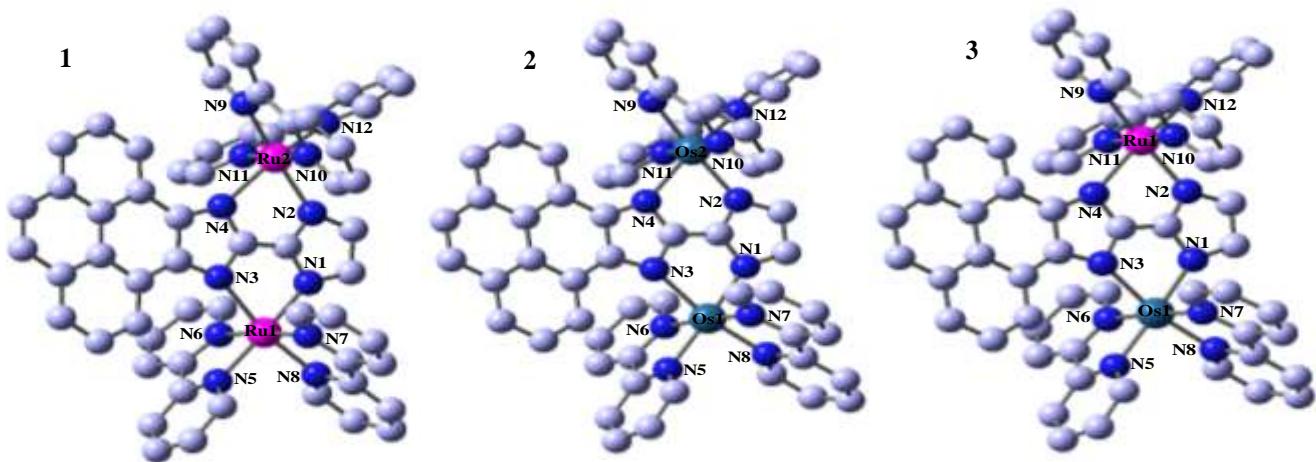


Figure S11. Optimized geometries and labelling scheme for $[(\text{bpy})_2\text{Ru}(\text{Py-Biimz})\text{Ru}(\text{bpy})_2]^{2+}$ (**1**), $[(\text{bpy})_2\text{Os}(\text{Py-Biimz})\text{Os}(\text{bpy})_2]^{2+}$ (**2**) and $[(\text{bpy})_2\text{Ru}(\text{Py-Biimz})\text{Os}(\text{bpy})_2]^{2+}$ (**3**) in solution phase.

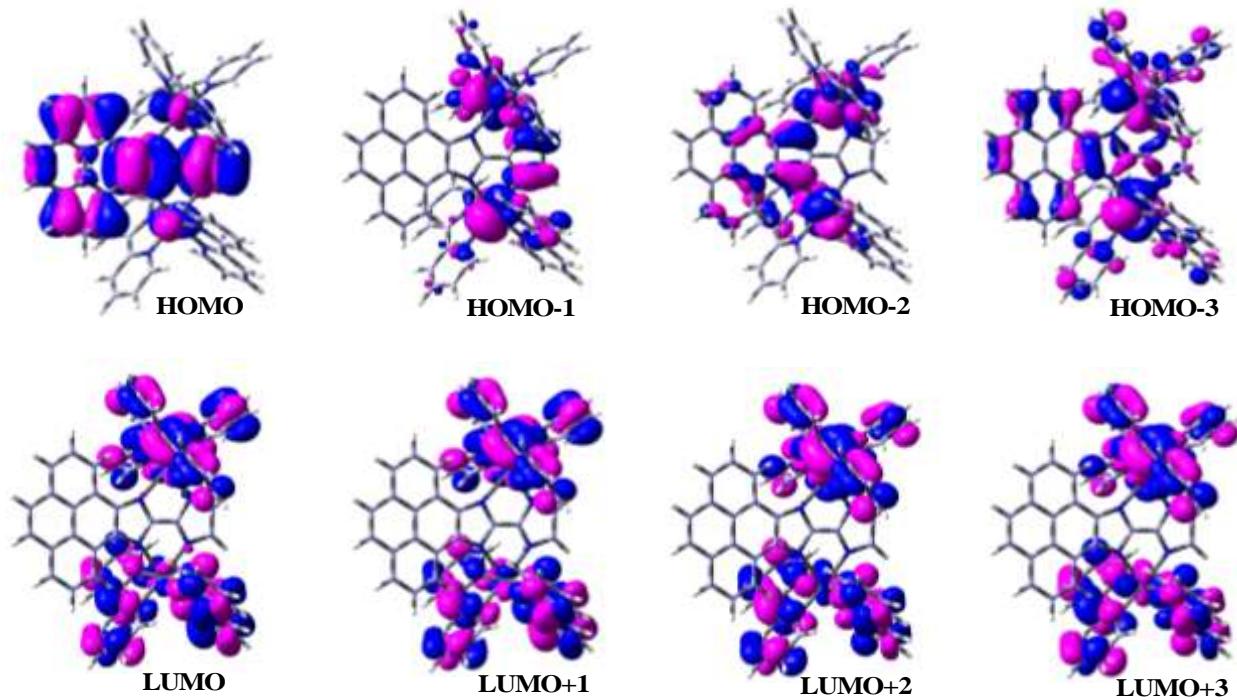


Figure S12. Schematic drawings of the selective frontier molecular orbitals for $[(\text{bpy})_2\text{Os}(\text{Py-Biimz})\text{Os}(\text{bpy})_2]^{2+}$ (**2**) in acetonitrile.

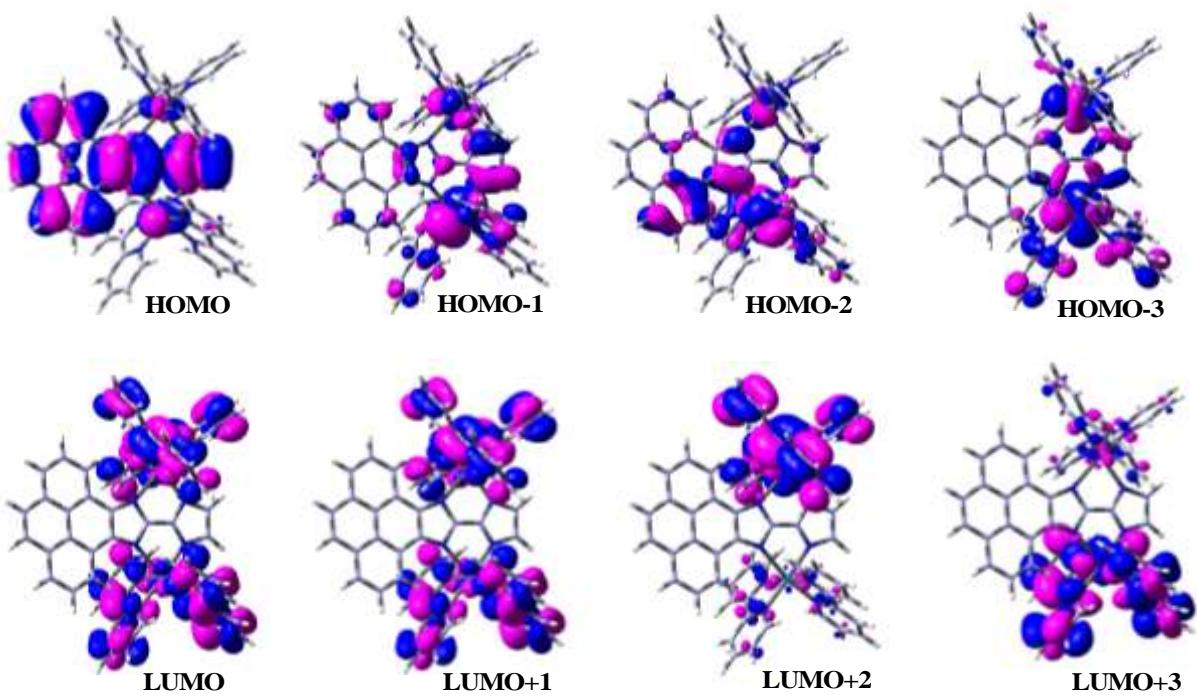


Figure S13. Schematic drawings of the selective frontier molecular orbitals for $[(\text{bpy})_2\text{Ru}(\text{Py-Biimz})\text{Os}(\text{bpy})_2]^{2+}$ (**3**) in acetonitrile.

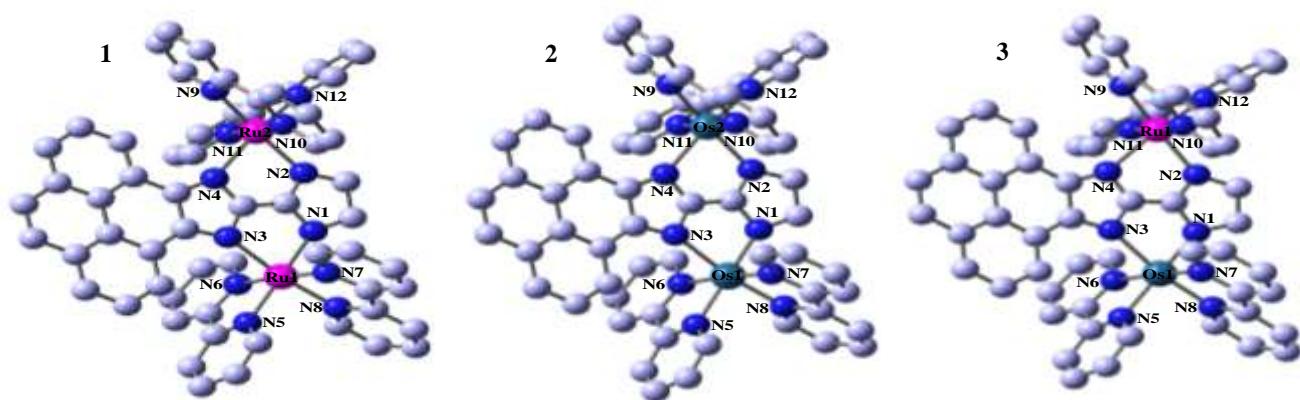


Figure S14. UKS optimized geometries and labelling scheme for $[(\text{bpy})_2\text{Ru}(\text{Py-Biimz})\text{Ru}(\text{bpy})_2]^{2+}$ (**1**), $[(\text{bpy})_2\text{Os}(\text{Py-Biimz})\text{Os}(\text{bpy})_2]^{2+}$ (**2**) and $[(\text{bpy})_2\text{Ru}(\text{Py-Biimz})\text{Os}(\text{bpy})_2]^{2+}$ (**3**) in acetonitrile.

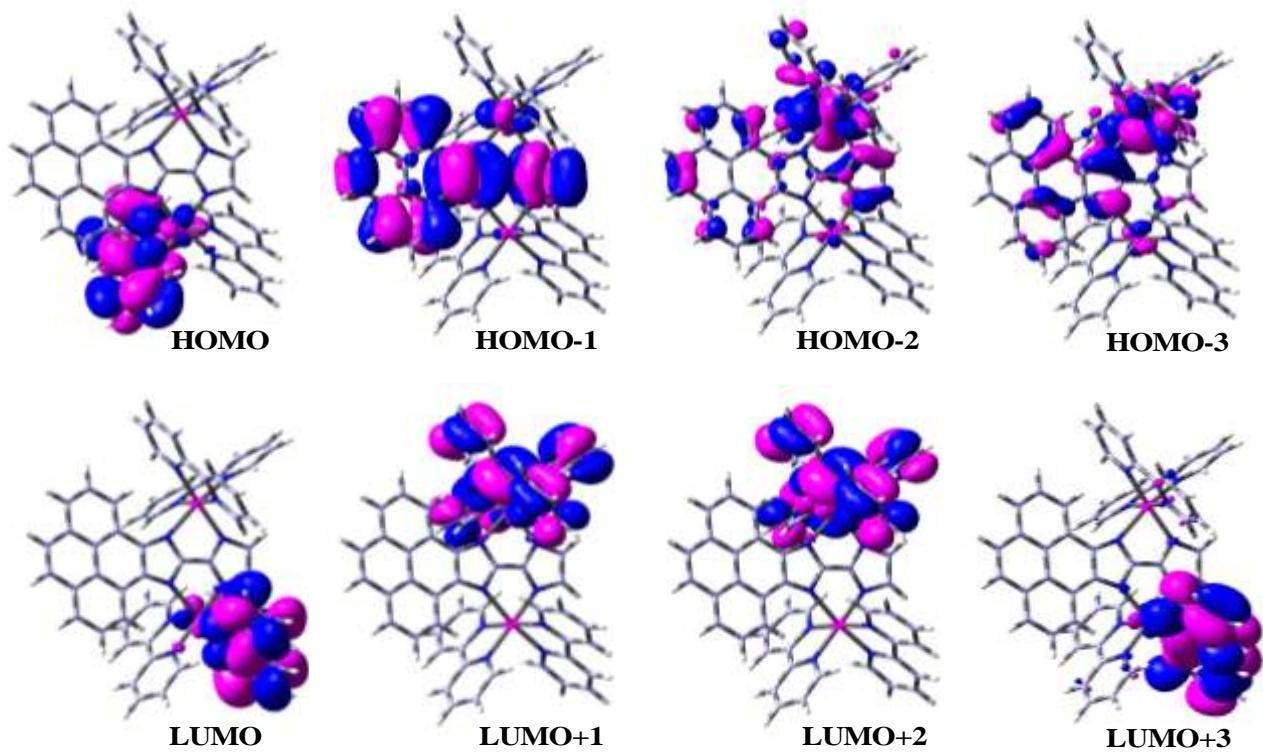


Figure S15. Schematic drawings of the selective frontier molecular orbitals for $[(bpy)_2Ru(Py\text{-}Biimz)Ru(bpy)_2]^{2+}$ (**1**) in UKS optimized state.

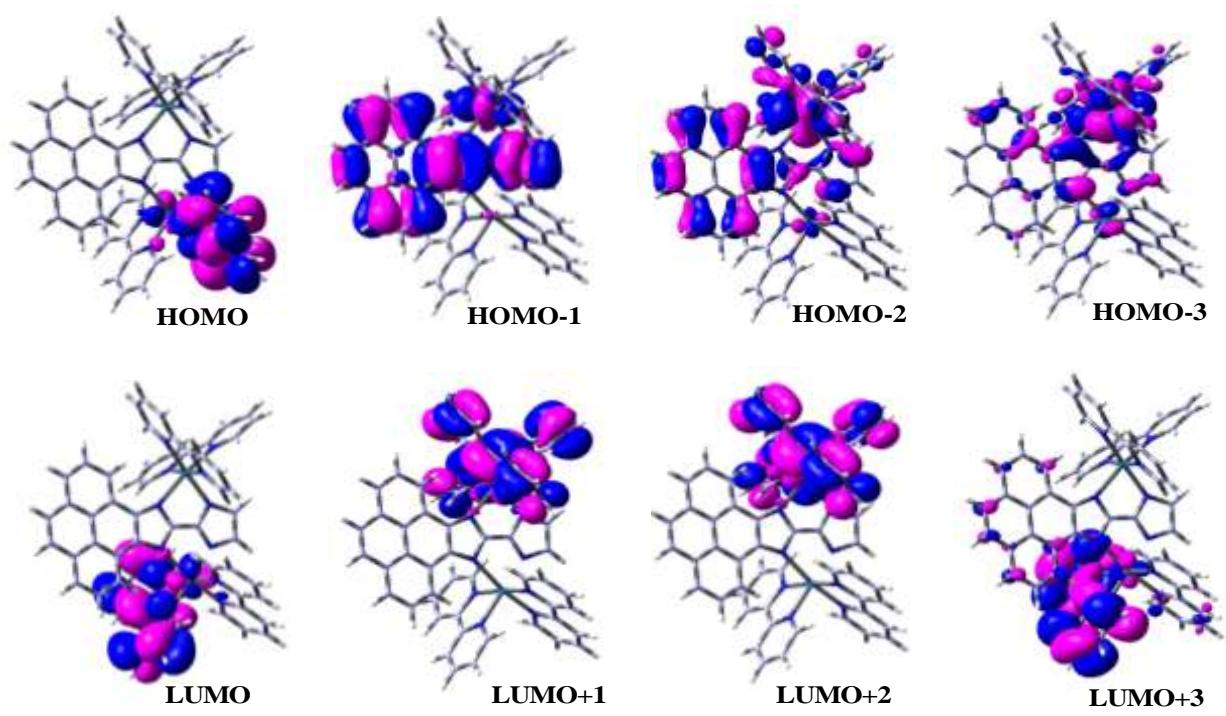


Figure S16. Schematic drawings of the selective frontier molecular orbitals for $[(\text{bpy})_2\text{Os}(\text{Py}-\text{Biimz})\text{Os}(\text{bpy})_2]^{2+}$ (**2**) in UKS optimized state.

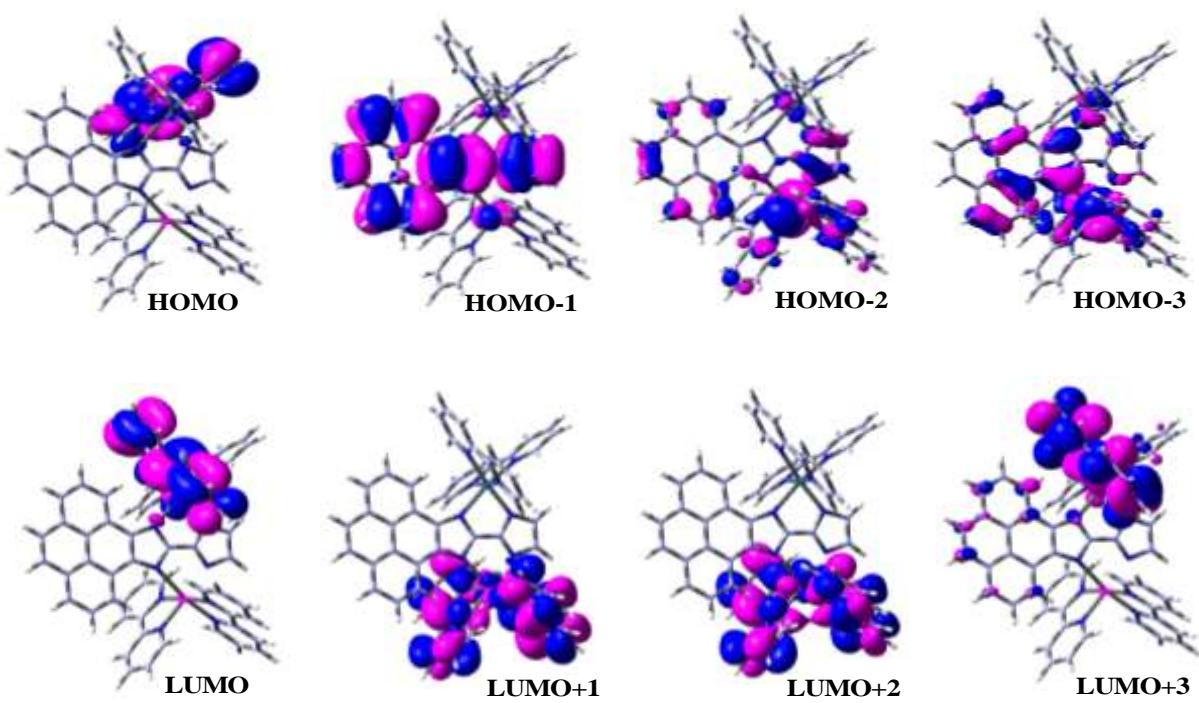


Figure S17. Schematic drawings of the selective frontier molecular orbitals for $[(\text{bpy})_2\text{Ru}(\text{Py}-\text{Biimz})\text{Os}(\text{bpy})_2]^{2+}$ (**3**) in UKS optimized state.

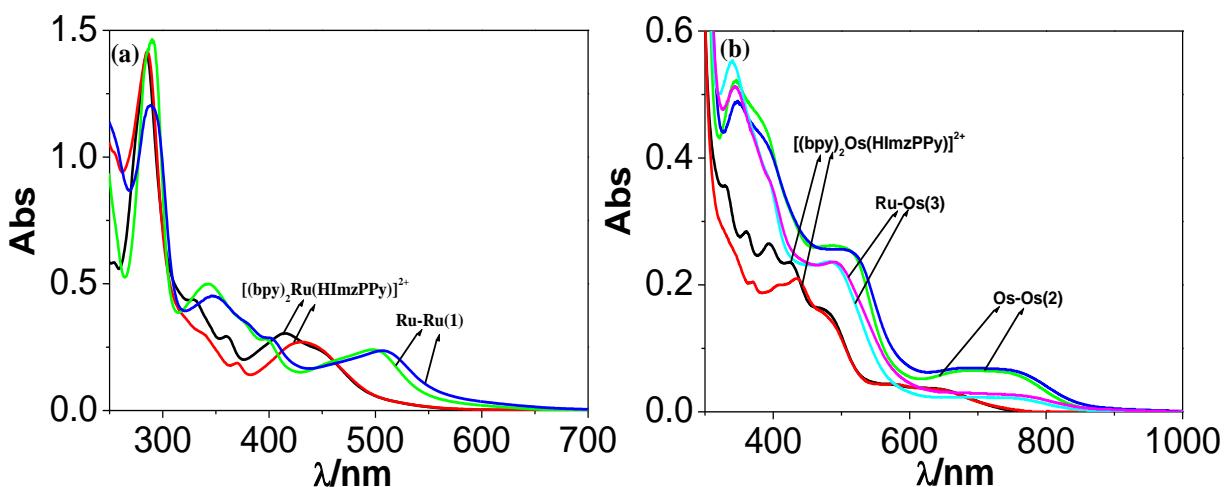


Figure S18. Overlayed absorption spectra of $[(bpy)_2M(HImzPPy)]^{2+}$ and $[(bpy)_2M/M_1(Py-Biimz)M/M_2(bpy)_2]^{2+}$ systems in presence of DNA.

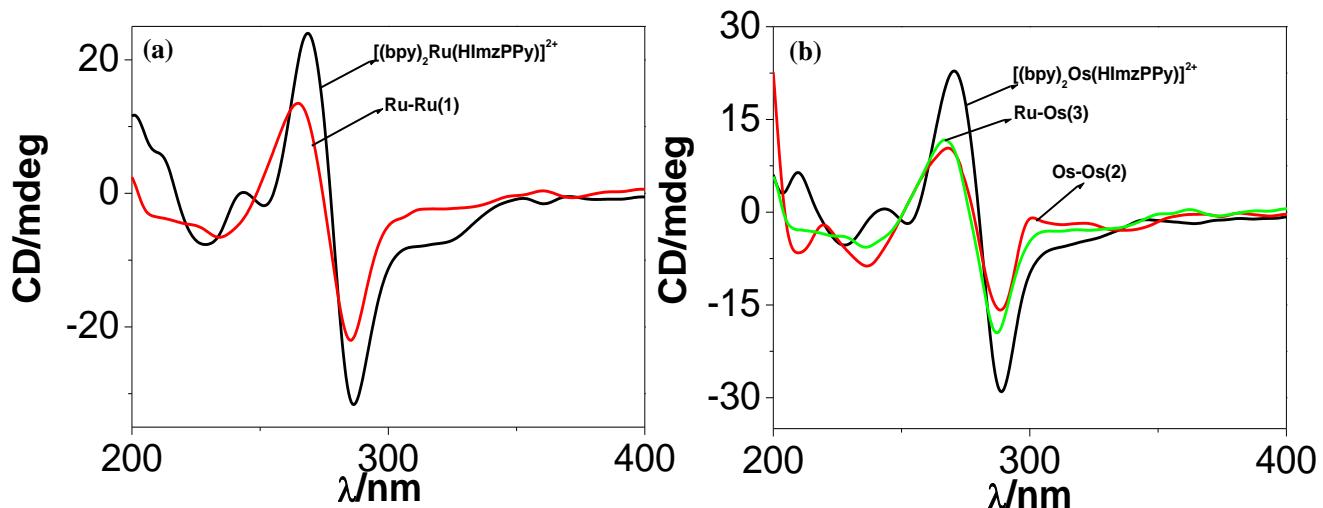


Figure S19. Overlayed CD spectra of $[(bpy)_2M(HImzPPy)]^{2+}$ and $[(bpy)_2M/M_1(Py-Biimz)M/M_2(bpy)_2]^{2+}$ systems in presence of DNA.

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