## Model peptides based on the binding loop of the copper metallochaperone Atx1: selectivity of the consensus sequence MxCxxC for metal ions Hg(II), Cu(I), Cd(II), Pb(II) and Zn(II).

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## NMR data for P<sup>L</sup>.

$$\begin{array}{c} \mathbf{P}^{\mathbf{L}}\\ & \bigcirc\\ & \parallel\\ & \mathsf{CH}_3\mathsf{C}\text{-}\mathsf{Met}\text{-}\mathsf{Thr}\text{-}\mathsf{Cys}\text{-}\mathsf{Ser}\text{-}\mathsf{Gly}\text{-}\mathsf{Cys}\text{-}\mathsf{Ser}\text{-}\mathsf{Arg}\text{-}\mathsf{Pro}\text{-}\mathsf{Gly}\text{-}\mathsf{NH}_2\\ & 2 & 3 & 4 & 5 & 6 & 7 & 8 & 9 & 10 & 11 \end{array}$$

**Table S1.** <sup>1</sup>H NMR (500 MHz) chemical shifts ( $\delta$  ppm) for P<sup>L</sup> in H<sub>2</sub>O/D<sub>2</sub>O 9:1 at 298 K

Residue	HN	Нα	Нβ	others
Met 2	8.05	4.50	1.91,2.05	CH <sub>2</sub> (γ) 2.42, 2.50 CH <sub>3</sub> (ε) 1.90
Thr 3	8.21	4.30	4.15	CH <sub>3</sub> (γ)1.12
Cys 4	8.28	4.42	2.82	
Ser 5	8.38	4.42	3.80	
Gly 6	8.15	3.82		
Cys 7	8.40	4.52	2.90	
Ser 8	8.42	4.40	3.78	
Arg 9	8.15	4.6	1.65, 1.75	$CH_2(\gamma) 1.60, CH_2(\delta) 3.31 \text{ HN}(\epsilon) 7.12$
Pro 10	-	4.30	1.82, 2.20	CH <sub>2</sub> (γ) 1.92 , 2.10
				CH <sub>2</sub> (δ) 3.52, 3.70
Gly 11	8.50	3.90		

**Table S2.**  ${}^{3}J_{\text{HN, H}\alpha}$  coupling constants (Hz) for P<sup>L</sup> in H<sub>2</sub>O/D<sub>2</sub>O 9:1 at 298 K

Met 2	Thr 3	Cys 4	Ser 5	Cys 7	Ser 8	Arg 9
7.2	8.0	7.2	7.2	7	6.5	7.2

Residue	HN	Ηα	Нβ	Others
Gly 1	8.65	3.63, 4.09		
Met 2	7.75	4.63	1.96, 2.11	CH <sub>2</sub> (γ): 2.36, 2.46; CH <sub>3</sub> (ε): 2.06
Thr 3	8.55	4.49	4.28	CH <sub>3</sub> (γ): 1.16
Cys 4	7.81	4.62	3.32	
Ser 5	8.87	4.29	3.80	
Gly 6	9.03	3.73, 3.98		
Cys 7	7.72	4.38	3.20, 3.27	
Ser 8	8.62	4.43	3.70, 3.84	
Arg 9	7.59	4.60	1.65, 1.76	CH <sub>2</sub> (γ): 1.56, 1.65; CH <sub>2</sub> (δ): 3.14; HN(ε): 7.10
Pro 10	-	4.27	2.23 ( <i>pro-S</i> ),	CH <sub>2</sub> (γ): 1.94 ( <i>pro-R</i> ), 2.02 ( <i>pro-S</i> );
			1.82 ( <i>pro-R</i> )	CH <sub>2</sub> (δ): 3.54( <i>pro-S</i> ), 3.84 ( <i>pro-R</i> )

**Table S3.** <sup>1</sup>H NMR (500 MHz) chemical shifts ( $\delta$  ppm) for P<sup>C</sup><sub>SS</sub> in H<sub>2</sub>O/D<sub>2</sub>O 9:1 at 298 K.

**Table S4.**  ${}^{3}J_{\text{HN},\text{H}\alpha}$  coupling constants (Hz) for P<sup>C</sup><sub>SS</sub> in H<sub>2</sub>O/D<sub>2</sub>O 9:1 at 298 K.

Met 2	Thr 3	Cys 4	Ser 5	Cys 7	Ser 8	Arg 9
8.2	9.2	5.9	4.5	6.9	8.5	7.2

**Table S5** ROESY-derived distances (Å) and distances calculated using X-PLOR for  $\mathbf{P}^{C}_{ss}$  in H<sub>2</sub>O/D<sub>2</sub>O 9:1 at 298 K.  $d(H1,H2)_{min}$  and  $d(H1,H2)_{max}$  are the lower and upper distance limits used for X-PLOR calculations.

H1	H2	<i>d</i> (H1,H2)	$d(H1,H2)_{min}$	$d(H1,H2)_{max}$
GLY 1 HN	MET 2 HN	2.71	2.44	2.98
GLY 1 HN	PRO 10 HA	2.08	1.87	2.29
GLY 1 HN	GLY 1 HA1	2.81	2.53	3.09
GLY 1 HN	GLY 1 HA2	2.29	2.06	2.52
GLY 1 HN	PRO 10 HB1	3.63	3.27	3.99
GLY 1 HN	PRO 10 HB2	3.53	3.18	3.88
MET 2 HN	THR 3 HN	3.42	3.08	3.76
MET 2 HN	PRO 10 HA	3.48	3.13	3.83

MET 2 HN	GLY 1 HA1	2.91	2.62	3.20
MET 2 HN	GLY 1 HA2	3.14	2.83	3.46
MET 2 HN	MET 2 HG*	2.91	2.62	3.90
MET 2 HN	MET 2 HB*	2.69	2.42	3.66
THR 3 HN	CYS 4 HN	2.53	2.27	2.78
THR 3 HN	MET 2 HA	2.19	1.97	2.41
THR 3 HN	THR 3 HB	2.84	2.56	3.12
THR 3 HN	CYS 4 HB*	4.73	4.26	5.91
THR 3 HN	THR 3 HG2*	3.40	3.06	4.14
THR 3 HN	MET 2 HB*	3.16	2.85	4.18
THR 3 HN	MET 2 HG*	3 98	3 58	5.08
THR 3 HA	THR 3 HB	3.08	2.77	3.38
THR 3 HA	THR 3 HG2*	2.88	2.59	3.56
THR 3 HR	THR 3 HG2*	2.00	2.39 2.20	3.09
CVS 4 HN	SER 5 HN	3.85	2.20 3.46	4 23
CVS 4 HN	CVS 4 HA	2.69	2 42	2.96
CVS 4 HN	$\frac{C13411A}{THR 3HA}$	2.09	2.42	2.90
CVS 4 HN	THE 3 HE	2.40	2.22	2.71
C I S 4 I I N C V S 4 U N		2.00	3.29 2.60	4.02
$C$ I S 4 $\Pi$ N	$CIS4\Pi D^{+}$	2.99	2.09	5.99
CIS4HN		3.01	2.71	4.01
CYS 4 HN	$ME I 2 HB^*$	5.24	2.92	4.27
CIS4HN	$\frac{1}{1} \frac{1}{1} \frac{1}$	4.41	3.97	5.25
CIS4HA	$CYS 4 HB^*$	2.33	2.09	3.20
SEK 5 HN	CYS4HA	2.18	1.96	2.40
SER 5 HN	SER 5 HA	2.70	2.43	2.97
SER 5 HN	SER 5 HB*	2.54	2.28	3.49
SER 5 HN	CYS 4 HB*	2.59	2.33	3.55
SER 5 HA	SER 5 HB*	2.37	2.13	3.30
GLY 6 HN	CYS 7 HN	2.59	2.33	2.85
GLY 6 HN	SER 5 HA	2.13	1.91	2.34
GLY 6 HN	GLY 6 HA*	2.46	2.22	3.41
GLY 6 HN	SER 5 HB*	3.93	3.54	5.02
CYS 7 HN	SER 5 HA	3.38	3.04	3.71
CYS 7 HN	CYS 7 HA	2.81	2.53	3.09
CYS 7 HN	GLY 6 HA*	3.19	2.87	4.21
CYS 7 HN	CYS 7 HB*	2.58	2.32	3.54
CYS 7 HA	CYS 7 HB*	2.66	2.39	3.62
SER 8 HN	ARG 9 HN	2.42	2.18	2.66
SER 8 HN	SER 8 HA	2.51	2.26	2.76
SER 8 HN	CYS 7 HA	2.21	1.99	2.43
SER 8 HN	SER 8 HB*	2.85	2.57	3.84
SER 8 HN	CYS 7 HB*	3.21	2.89	4.23
SER 8 HA	SER 8 HB*	2.41	2.17	3.35
ARG 9 HN	ARG 9 HA	2.70	2.43	2.97
ARG 9 HN	SER 8 HA	2.88	2.59	3.17
ARG 9 HN	SER 8 HB*	3.77	3.40	4.85
ARG 9 HN	CYS 7 HB*	3.46	3.12	3.81
ARG 9 HA	PRO 10 HD1	2.09	1.88	2.30
ARG 9 HA	PRO 10 HD2	2.49	2.24	2.74

**Figure S1.** Calibration curve and experimental results for the determination of molecular weights of the peptides by size exclusion chromatography in aqueous buffer at pH 7.



**Figure S2.** UV titration of P<sup>L</sup> (54  $\mu$ M) with mercury chloride at pH 7 (20 mM phosphate buffer). Spectra shown are difference spectra ( $\Delta \varepsilon = \varepsilon HgP^L - \varepsilon P^L$ ) and correspond to samples with 0-1.3 equiv of Hg (II) per P<sup>L</sup>. The insert shows the  $\Delta \varepsilon$  at 220 and 250 nm *vs* equiv of Hg(II) per P<sup>L</sup>.



**Figure S3.** UV titration of  $P^L$  (54  $\mu$ M) with cadmium chloride at pH 7 (20 mM phosphate buffer). Spectra shown are difference spectra ( $\Delta \varepsilon = \varepsilon C dP^L - \varepsilon P^L$ ) and correspond to samples with 0-1.4 equiv of Cd (II) per P<sup>L</sup>. The insert shows the absorbance at 225 and 240 nm *vs* equiv of Cd(II) per P<sup>L</sup>.

