

Supporting Information

Ring Expansion of Functionalized Octahydroindoles to Enantiopure *cis*-Decahydroquinolines

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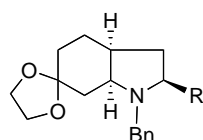
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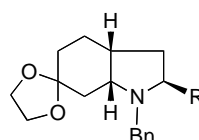
Table 1. ^{13}C NMR Chemical shifts of octahydroindoles 3-12^a

	3	4	5	6	7	8	9a	10a	11a
C2	66.2	62.9	66.2	61.7	74.9	70.8	71.0	72.5	66.1
C3	33.6	31.7	32.1	30.5	33.3	30.6	30.6	32.2	25.4
C3a	35.7	35.2	35.0	35.6	35.6	35.6	34.6	34.8	35.6
C4	24.0	23.3	24.3	22.8	24.2	22.8	23.9	22.7	22.9
C5	31.0	29.9	31.0	28.9	31.0	29.2	27.5	29.3	28.9
C6	109.1	109.3	108.9	109.4	108.8	109.2	108.9	109.0	109.4
C7	37.5	31.9	37.8	30.6	38.2	30.9	38.2	38.8	30.4
C7a	62.5	59.5	63.0	59.3	63.0	59.4	62.9	63.3	58.9
NCH₂	58.8	53.4	58.8	51.7	60.0	53.4	58.9	63.2	51.4
ips-Ar	139.1	139.0	139.2	139.3	139.2	138.5	140.0	140.0	139.4
o-Ar	129.1	128.8	128.9	128.4	129.4	128.7	129.1	128.3	128.4
m-Ar	127.9	128.1	128.3	128.3	128.1	128.2	128.4	128.3	128.2
p-Ar	126.8	126.9	127.2	127.0	127.1	127.1	127.2	127.0	127.0
C-1'	175.0	174.2	61.2	62.4	213.2	216.4	63.5	71.9	64.7
Me	51.6	51.6	---	---	24.9	24.9	18.2	20.8	18.2
OCH₂	64.3	64.3	64.1	64.3	64.0	64.3	64.0	64.1	64.1
	64.0	64.0	63.9	63.9	64.0	63.9	64.0	63.9	63.8

^a Values for compounds **3**, **5**, **9a**, **10a** and **11a** were assigned on the basis of gHSQC spectra.



3 R = CO₂Me
5 R = CH₂OH
7 R = COCH₃
9a R = (*R*)-CHOHCH₃
10a R = (*S*)-CHOHCH₃

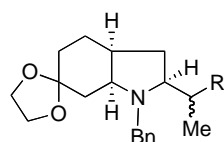


4 R = CO₂Me
6 R = CH₂OH
8 R = COCH₃
11a R = (*S*)-CHOHCH₃

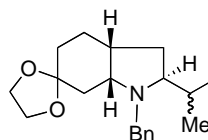
Table 1 (continued). ^{13}C NMR Chemical shifts of octahydroindoles 3-12^b

	12a	9b	10b	11b	12b	9c^c	10c^c	11c^c	12c^c
C2	66.5	72.1	71.6	66.7	66.4	69.1	68.8	64.4	62.9
C3	30.3	29.7	30.4	28.2	27.1	30.1	31.0	27.7	27.4
C3a	35.3	34.5	34.5	34.6	35.0	34.4	34.6	34.8	35.2
C4	22.7	23.0	23.3	22.8	22.8	23.3	23.2	22.9	22.8
C5	29.0	29.5	29.9	29.7	29.0	29.8	29.7	29.1	29.0
C6	109.4	109.2	109.1	109.5	109.4	109.2	109.1	109.5	109.5
C7	31.2	39.2	39.1	31.0	30.9	39.1	39.3	30.2	30.6
C7a	58.8	62.5	63.7	58.9	58.3	62.2	63.8	58.2	58.4
NCH₂	53.9	61.4	61.3	52.2	52.7	60.4	61.8	52.0	52.5
ips-Ar	139.5	140.6	140.7	139.9	139.8	139.2	141.2	139.8	140.0
o-Ar	128.2	128.3	128.6	128.2	128.2	129.4	128.2	128.1	128.1
m-Ar	127.8	128.1	128.2	128.2	128.1	128.1	128.0	128.0	128.0
p-Ar	126.7	126.6	126.8	126.7	126.8	127.1	126.5	126.6	126.6
C-1'	71.1	63.4	60.6	60.9	59.8	71.1	75.2	70.7	72.3
Me	20.6	22.9	19.8	22.3	17.7	17.1	16.5	16.7	14.5
OCH₂	64.2	64.1	64.0	64.2	64.3	63.9	64.0	64.1	64.2
	63.8	63.8	63.9	63.9	63.9	63.8	63.8	63.8	63.9

^b Values for compounds **12a**, **10b**, **11b**, **9c**, **11c**, and **12c** were assigned on the basis of gHSQC spectra. ^c OAc: 170.6 / 170.7 and 21.3 / 21.5.



9b R = Cl (1'*R*)
10b R = Cl (1'*S*)
9c R = OAc (1'*R*)
10c R = OAc (1'*S*)

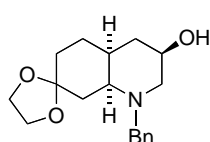


12a R = OH (1'*R*)
11b R = Cl (1'*S*)
12b R = Cl (1'*R*)
11c R = OAc (1'*S*)
12c R = OAc (1'*R*)

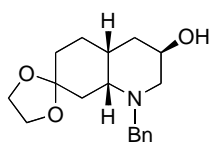
Table 2. ^{13}C NMR Chemical shifts of decahydroquinolines **13-21**^a

	13	14	15	16 ^b	17	18	19 ^c	21
C2	52.0	50.6	57.1	56.0	53.0	51.9	53.2	55.7
C3	68.1	65.4	69.7	61.4	75.5	73.2	73.1	73.6
C4	33.1	30.5	26.8	33.1	29.6	27.0	24.2	33.3
C4a	32.8	28.8	28.3	35.5	31.4	33.6	28.4	32.0
C5	26.9	26.6	26.4	26.7	26.6	25.0	26.8	26.9
C6	29.9	29.8	30.2	32.4	29.7	30.2	30.0	29.9
C7	109.9	109.8	109.3	109.6	109.8	109.4	109.3	109.9
C8	27.0	25.6	33.5	28.9	28.1	34.1	34.7	27.6
C8a	57.0	57.7	56.6	56.2	55.8	56.1	56.9	56.5
Me	---	---	14.9	18.4	16.8	11.5	15.7	16.9
NCH₂	58.3	58.5	55.3	52.7	52.6	55.7	55.9	52.8
Ar	126.8	127.2	127.1	126.5	126.5	126.9	140.2	126.5
	128.1	128.4	128.3	127.6	127.7	128.2	128.3	127.9
	129.5	128.7	128.5	128.1	128.2	128.2	127.9	128.1
	139.3	139.0	n.o.	141.1	141.2	139.8	126.6	140.6
OCH₂	64.2	64.3	63.9	64.0	64.1	64.2	64.2	64.1
	64.1	64.1	64.2	63.7	64.0	63.9	63.9	64.0
Other					170.6	170.4	170.7	
					21.3	21.3	21.4	

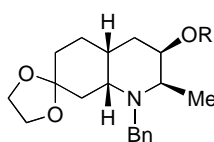
^a Values for compounds **13**, **17**, **18**, **19** and **21** were assigned on the basis of gHSQC spectra ^b Values taken from an NMR spectrum of a mixture of **9b** and **16**. ^c Values taken from an NMR spectrum of a mixture of **11c** and **19**.



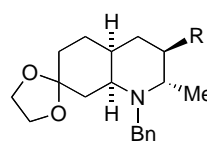
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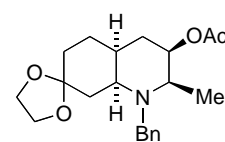
14



15 R = H
19 R = Ac



16 R = Cl
17 R = OAc
21 R = OH



18

Experimental Section

General: All reactions were carried out under an argon atmosphere with dry, freshly distilled solvents under anhydrous conditions. Analytical thin-layer chromatography was performed on SiO₂ (silica gel 60 F₂₅₄) or Al₂O₃ (ALOX N/UV₂₅₄), and the products were located with iodoplatinate spray. Chromatography refers to flash chromatography and was carried out on SiO₂ (silica gel 60, 230-240 mesh ASTM) or Al₂O₃ (aluminium oxide 90). Drying of organic extracts was performed over anhydrous Na₂SO₄. Evaporation of solvent was accomplished with a rotatory evaporator. Chemical shifts of ¹H and ¹³C NMR spectra are reported in ppm downfield (δ) from Me₄Si. Only noteworthy IR absorptions (cm⁻¹) are listed.

Methyl (2*S*,3*aS*,7*aS*)-1-Benzyl-6-oxooctahydroindole-2-carboxylate ethylene acetal (3). To a solution of ketone **1** (3.28 g, 11 mmol) in toluene (350 mL) were added a catalytic amount of TsOH and ethyleneglycol (1.84 mL, 33 mmol), and the reaction mixture was heated at reflux temperature for 4 h in a flask incorporating a Dean-Stark apparatus. The cooled solution was diluted with CH₂Cl₂ and washed with aqueous saturated NaHCO₃ (100 mL). The organic phase was dried and concentrated to give 3.64 g of **3** as a yellowish oil, which was used in the next step without further purification. An analytical sample was obtained by chromatography (SiO₂, 1% MeOH in CH₂Cl₂). *R*_f = 0.37 (SiO₂, hexane/EtOAc 3:2); [α]_D²⁰ -41 (*c* = 0.9, CHCl₃); ¹H NMR (400 MHz, CDCl₃, gCOSY) 1.46-1.58 (m, 1H), 1.64-1.85 (m, 5H), 1.87-1.95 (m, 1H), 2.02-2.11 (m, 1H), 2.13 -2.26 (m, 1H), 2.99 (q, *J* = 7.5 Hz, 1H, H-7a), 3.41 (dd, *J* = 9.1, 7.7 Hz, 1H, H-2), 3.54 (s, 3H, OCH₃), 3.74-3.92 (m, 6H), 7.18-7.40 (m, 5H, ArH); ¹³C NMR (100 MHz, CDCl₃, DEPT, gHSQC), see Table 1. Anal. Calcd for C₁₉H₂₅NO₄: C 68.86, H 7.60, N 4.22. Found: C 68.48, H 7.55, N 4.20.

Methyl (2*S*,3*aR*,7*aR*)-1-Benzyl-6-oxooctahydroindole-2-carboxylate ethylene acetal (4). Operating as above, from ketone **2** (1.03 g, 3.6 mmol), acetal **4** was obtained (1.12 g) as yellowish crystals and used in the next step without further purification. An analytical sample was obtained by chromatography (SiO₂, 1% MeOH in CH₂Cl₂): *R*_f = 0.24 (SiO₂, hexane/EtOAc 3:2); mp 64-66 °C; [α]_D²⁰ -53 (*c* 0.4, CHCl₃); ¹H NMR (300 MHz, CDCl₃) 1.44-1.72 (m, 4H), 1.79-1.94 (m, 3H), 2.12

(dt, $J = 12.9, 10.7$ Hz, 1H), 2.46 -2.58 (m, 1H), 3.37 (dt, $J = 10.5, 5.3$ Hz, 1H, H-7a), 3.50 (dd, $J = 10.2, 3.6$ Hz, 1H, H-2), 3.55 (s, 3H, OCH₃), 3.74 (d, $J = 13.2$ Hz, 1H), 3.81 (d, $J = 13.2$ Hz, 1H), 3.83 -3.96 (m, 4H), 7.18-7.40 (m, 5H, ArH). ¹³C NMR (75 MHz, CDCl₃, DEPT), see Table 1. Anal. calcd for C₁₉H₂₅NO₄: C 68.86, H 7.60, N 4.22. Found: C 68.56, H 7.85, N 4.24.

(2*S*,3*aS*,7*aS*)-1-Benzyl-2-hydroxymethyl-6-oxooctahydroindole ethylene acetal (5). Ester **3** (603 mg, 1.82 mmol) was dissolved in THF (9 mL) and then cooled to 0 °C. LiBH₄ (2 M in THF, 2.8 mL, 5.46 mmol, 3 equiv) was slowly added, and the reaction mixture was stirred at rt for 24 h. The reaction was quenched by adding H₂O (5 mL) and the organic layer was dried and concentrated to give a residue, which was purified by chromatography (SiO₂, 1% MeOH in CH₂Cl₂) to afford 441 mg (80% from **1**) of **5** as a colourless oil: $R_f = 0.31$ (SiO₂, CH₂Cl₂/MeOH 95:5); $[\alpha]_D^{20} -15$ (c 0.2, CHCl₃); ¹H NMR (400 MHz, CDCl₃, gCOSY) 1.49 (dddd, $J = 12.0, 5.0, 5.0, 1.2$ Hz, 1H, H-5eq), 1.65 (m, 3H, H-5 and H-7), 1.75 (m, 3H, H-3 and H-4), 1.85 (dd, $J = 12.4, 7.2$ Hz, 1H, H-3), 2.20 (m, 1H, H-3a), 2.40 (brs, 1H, OH), 2.98 (dddd, $J = 8.0, 7.5, 5.4, 1.8$ Hz, 1H, H-2), 3.02 (q, $J = 7.2$ Hz, 1H, H-7a), 3.33 (dd, $J = 11.0, 1.2$ Hz, 1H, CH₂OH), 3.42 (dd, $J = 11.0, 3.6$ Hz, 1H, CH₂OH), 3.69-3.88 (m, 6H, OCH₂ and NCH₂), 7.26-7.32 (m, 5H, ArH); ¹³C NMR (100 MHz, CDCl₃, gHSQC), see Table 1. HRFABMS calcd for C₁₈H₂₆NO₃ 304.1906 (MH⁺), found 304.1913.

(2*S*,3*aR*,7*aR*)-1-Benzyl-2-hydroxymethyl-6-oxooctahydroindole ethylene acetal (6). Operating as above from ester **4** (627 mg, 1.89 mmol), alcohol **6** (464 mg, 81% from **2**) was obtained after chromatography (SiO₂, 1% MeOH in CH₂Cl₂), as white crystals: $R_f = 0.40$ (SiO₂, CH₂Cl₂/MeOH 95:5); mp 72-74 °C; $[\alpha]_D^{20} -72$ (c 0.25, CHCl₃); ¹H NMR (400 MHz, COSY, CDCl₃) 1.46 (t, $J = 12.0$ Hz, 1H, H-7ax), 1.52 (dq, $J = 12.5, 2.5$ Hz, 1H, H-4eq), 1.63 (dm, $J = 12.0$ Hz, 1H, H-5eq), 1.64 (td, $J = 10.0, 3.2$ Hz, 1H, H-5ax), 1.78 (m, 1H, H-4ax), 1.80 (m, 1H, H-3β), 1.83 (dd, $J = 12.0, 5.5$ Hz, 1H, H-7eq), 2.05 (q, $J = 12.0$ Hz, 1H, H-3α), 2.30 (m, 1H, H-3a), 2.99 (dt, $J = 10.0, 3.2$ Hz, 1H, H-2), 3.24 (ddd, $J = 12.0, 5.5, 5.5$ Hz, 1H, H-7a), 3.38 (dm $J = 10.8$ Hz, 1H, CH₂OH), 3.55 (dd, $J = 10.8, 3.2$ Hz, 1H, CH₂OH), 3.64 and 3.71 (2d, $J = 13.6$ Hz, 1H each,

NCH₂), 3.77-3.93 (m, 4H, OCH₂), 7.26-7.32 (m, 5H, ArH); ¹³C NMR (100 MHz, CDCl₃, DEPT), see Table 1. Anal. Calcd for C₁₈H₂₅NO₃: C 71.26, H 8.31, N 4.62. Found: C 71.05, H 8.20, N 4.57.

(2*S*,3*aS*,7*aS*)-2-Acetyl-1-benzyl-octahydroindol-6-one ethylene acetal (7). To a solution of ester **3** (2.45 g, 7.4 mmol) in THF (140 mL) cooled to -20 °C was added Me(MeO)NH.HCl (1.82 g, 18.5 mmol, 2.5 eq) and then over 30 min was added a solution of *i*-PrMgCl in THF (18.5 mL, 2.0 M, 5 equiv) maintaining the temperature at -10 °C. The mixture was stirred for 40 min and quenched with saturated aqueous NH₄Cl solution (50 mL). The organic layer was dried and concentrated to afford 2.65 g of the corresponding Weinreb amide as a yellow oil, which was used without purification: *R*_f = 0.21 (SiO₂, CH₂Cl₂/MeOH 96:4). To a solution of the aforementioned Weinreb amide (2.65 g, 7.4 mmol) in THF (95 mL) cooled to 0 °C was added dropwise MeMgBr in Et₂O (6.4 mL, 3 M, 19.24 mmol, 2.6 eq). The reaction mixture was stirred at 0 °C for 1 h and quenched with saturated aqueous NH₄Cl solution (50 mL). The organic layer was dried and concentrated to give ketone **7** (2.32 g) as an oil, which was used without further purification. *R*_f = 0.52 (SiO₂, CH₂Cl₂/MeOH 95:5); ¹H NMR (300 MHz, CDCl₃) 1.53 (m, 1H), 1.61-1.85 (m, 6H), 2.02 (dt, *J* = 12.3, 7.2 Hz, 1H), 2.04 (s, 3H, CH₃), 2.25 (m, 1H, H-3a), 3.02 (q, *J* = 6.9 Hz, 1H, H-7a), 3.29 (dd, *J* = 9.3, 7.8 Hz, 1H, H-2), 3.63 and 3.83 (2d, *J* = 13.5 Hz, 1H each, NCH₂Ar), 3.75-3.95 (m, 4H, OCH₂), 7.20- 7.35 (m, 5H, Ar); ¹³C NMR (50 MHz, CDCl₃, DEPT), see Table 1.

(2*S*,3*aR*,7*aR*)-2-Acetyl-1-benzyl-octahydroindol-6-one ethylene acetal (8). The above procedure was applied to ester **4** (1.12 g) to afford the corresponding Weinreb amide (1.30 g): *R*_f = 0.16 (SiO₂, CH₂Cl₂/MeOH 95:5). The Weinreb amide was treated with MeMgBr in Et₂O (3.12 mL, 3 M, 9.36 mmol, 2.6 eq) and operating as in the formation of ketone **7**, 1.21 g of ketone **8** was isolated, which was used without further purification. *R*_f = 0.49 (SiO₂, CH₂Cl₂/MeOH 95:5); ¹H NMR (300 MHz, CDCl₃) 1.40 (t, *J* = 12.0 Hz, 1H, H-5ax), 1.50-1.70 (m, 3H), 1.75-1.90 (m, 3H), 2.07 (s, 3H, CH₃), 2.16 (dt, *J* = 13.5, 11.4 Hz, 1H), 2.50 (m, 1H, H-3a), 3.36 (ddd, *J* = 11.4, 5.7, 5.7 Hz, 1H, H-7a), 3.35 (dd, *J* = 11.0, 1.5 Hz, 1H, H-2), 3.57 and 3.71 (2d, *J* = 13.2 Hz, 1H each, CH₂Ar), 3.75-3.94 (m, 4H, OCH₂), 7.20- 7.35 (m, 5H, Ar); ¹³C NMR (50 MHz, CDCl₃, DEPT), see Table 1.

Reduction of ketone 7. To a solution of amino ketone **7** (2.29 g, 7.25 mmol) in MeOH (85 mL) at -20 °C was added NaBH₄ (571 mg, 14.5 mmol) in small portions. The resulting mixture was maintained at this temperature for 6 h. Then, water (25 mL) was added and the mixture was extracted with Et₂O (3x50 mL). The organic extracts were washed with brine, dried, and concentrated. Purification of the residue by chromatography (SiO₂, hexane to hexane/EtOAc 1:1) provided 1.05 g (46%) of alcohol **9a** as a colourless oil and 862 mg (37%) of alcohol **10a** as a colorless oil, after two successive purifications. Overall yield for three steps (**3** → **9a** + **10a**): 83%; 1.2:1 ratio of alcohols **9a**:**10a**.

(2*S*,3*aS*,7*aS*)-1-Benzyl-2-[(1'*R*)-(1-hydroxyethyl)]octahydroindol-6-one ethylene acetal (9a**):** R_f = 0.30 (SiO₂, CH₂Cl₂/MeOH 95:5); $[\alpha]_D^{20}$ - 41 (c 1.3, CHCl₃); ¹H NMR (400 MHz, CDCl₃, gCOSY) 1.09 (d, J = 6.6 Hz, 3H, CH₃), 1.43-1.81 (m, 8H, H-3, H-4, H-5, and H-7), 2.20 (m, 1H, H-3a), 2.80 (ddd, J = 9.3, 6.9, 3.3 Hz, 1H, H-2), 3.02 (q, J = 7.2 Hz, 1H, H-7a), 3.66-3.85 (m, 7H, H-1', NCH₂Ar, and OCH₂), 7.26-7.32 (m, 5H, ArH); ¹³C NMR (100 MHz, CDCl₃, gHSQC), see Table 1. HRFABMS: calcd for C₁₉H₂₈NO₃ 318.2069 (MH⁺), found 318.2070.

(2*S*,3*aS*,7*aS*)-1-Benzyl-2-[(1'*S*)-(1-hydroxyethyl)]octahydroindol-6-one ethylene acetal (10a**):** R_f = 0.30 (SiO₂, CH₂Cl₂/MeOH 95:5); $[\alpha]_D^{20}$ - 32 (c 0.8, CHCl₃); ¹H NMR (400 MHz, CDCl₃, gCOSY) 1.16 (d, J = 6.4 Hz, 3H, CH₃), 1.50 (m, 2H, H-5 and H-7), 1.60 (m, 2H, H-3 and H-5), 1.63 (t, J = 12.0 Hz, 1H, H-7ax), 1.77 (m, 2H, H-4), 1.88 (ddd, J = 12.0, 8.0, 7.0 Hz, 1H, H-3), 2.34 (m, 1H, H-3a), 2.82 (q, J = 7.5 Hz, 1H, H-2), 2.97 (dt, J = 12.0, 6.0 Hz, 1H, H-7a), 3.54 (quint, J = 6.2 Hz, 1H, H-1'), 3.74-3.86 (m, 6H, OCH₂, NCH₂Ar), 7.20-7.40 (m, 5H, ArH); ¹³C NMR (100 MHz, CDCl₃, gHSQC), see Table 1. HRFABMS: calcd for C₁₉H₂₈NO₃ 318.2069 (MH⁺), found 318.2074.

Reduction of ketone 8. The above procedure was followed using ketone **8** (1.06 g, 3.36 mmol). Purification by chromatography (SiO₂, hexane to hexane-EtOAc 1:1) afforded 622 mg (54%) of alcohol **11a** as a white solid and then 365 mg (32%) of alcohol **12a** as a white solid. Overall yield for three steps (**4** → **11a** + **12a**): 86%; 1.7:1 ratio of alcohols **11a** and **12a**.

(2*S*,3*aR*,7*aR*)-1-Benzyl-2-[(1'*S*)-(1-hydroxyethyl)]octahydroindol-6-one ethylene acetal

(11a): R_f = 0.22 (SiO₂, CH₂Cl₂/MeOH 98:2); mp 73-75 °C; $[\alpha]_D^{20}$ -100 (c 0.7, CHCl₃); ¹H NMR (400 MHz, CDCl₃, gCOSY) 1.13 (d, J = 6.4 Hz, 3H, CH₃), 1.41 (t, J = 12.0 Hz, 1H, H-7_{ax}), 1.50 (dm, J = 12.0 Hz, 1H), 1.60-1.68 (m, 2H), 1.73-1.90 (m, 4H), 2.21 (m, 1H, H-3_a), 2.76 (dq, J = 10.0, 2.4 Hz, 1H, H-2), 3.21 (ddd, J = 12.0, 5.5, 5.5 Hz, 1H, H-7_a), 3.61 and 3.77 (2d, J = 14.0 Hz, 1H each, NCH₂Ar), 3.78-3.93 (m, 5H, OCH₂ and H-1'), 7.20-7.40 (m, 5H, ArH); ¹³C NMR (100 MHz, CDCl₃, gHSQC), see Table 1. HRFABMS: calcd for C₁₉H₂₈NO₃ 318.2069 (MH⁺), found 318.2074.

(2*S*,3*aR*,7*aR*)-1-Benzyl-2-[(1'*R*)-(1-hydroxyethyl)]octahydroindol-6-one ethylene acetal

(12a): R_f = 0.11 (SiO₂, CH₂Cl₂/MeOH 98:2); mp 91-93 °C; $[\alpha]_D^{20}$ - 36 (c 1.0, CHCl₃); ¹H NMR (400 MHz, CDCl₃, gCOSY) 1.22 (d, J = 6.6 Hz, 3H, CH₃), 1.43 (t, J = 12.0 Hz, 1H, H-7_{ax}), 1.50-1.75 (m, 4H, H-3, H-4, H-5, H-7), 1.80 (m, 2H, H-4 and H-5), 2.15 (m, 1H, H-3), 2.32 (m, 1H, H-3_a), 2.88 (ddd, J = 9.8, 4.8, 2.2 Hz, 1H, H-2), 3.22 (ddd, J = 11.0, 5.4, 5.4 Hz, 1H, H-7_a), 3.69 (m, 1H, H-1'), 3.73-3.92 (m, 6H, NCH₂ and OCH₂), 3.96 (br s, 1H, OH), 7.20-7.40 (m, 5H, ArH); ¹³C NMR (100 MHz, CDCl₃, gHSQC), see Table 1. HRFABMS: calcd for C₁₉H₂₈NO₃ 318.2069 (MH⁺), found 318.2063.

Conversion of alcohols 9a-12a to their corresponding chlorides 9b-12b.

Compounds **9b-12b** were prepared according to the following procedure: to a solution of alcohol (**9a-12a**, 50 mg, 0.16 mmol) in THF (1 mL) at 0 °C was added MsCl (0.014 mL, 0.18 mmol, 1.1 equiv), followed by Et₃N (0.09 mL, 0.64 mmol, 4.0 equiv). After 4 h at reflux, the reaction mixture was poured into an aqueous 2.5 M NaOH solution (1 mL). After extraction with CH₂Cl₂ (3 x 2 mL), the organic phase was dried and concentrated to afford compounds **9b-12b**, which were used without further purification.

(2*S*,3*aS*,7*aS*)-1-Benzyl-2-[(1'*R*)-(1-chloroethyl)]octahydroindol-6-one ethylene acetal (9b).

This compound was obtained together with the expanded chloride **16**; ¹H NMR (300 MHz, CDCl₃) 1.53 (d, J = 6.6 Hz, 3H, CH₃), 1.40-2.10 (m, 8H), 2.25 (m, 1H), 2.85-3.00 (m, 2H), 3.80-4.00 (m,

7H), 7.20-7.40 (m, 5H); ^{13}C NMR (75 MHz, CDCl_3), see Table 1. HRFABMS calcd for $\text{C}_{19}\text{H}_{27}^{35}\text{ClNO}_3$ 336.1730 (MH^+), found 336.1731.

(2*S*,3*aS*,7*aS*)-1-Benzyl-2-[(1'*S*)-(1-chloroethyl)]octahydroindol-6-one ethylene acetal (10b): yellow oil; ^1H NMR (400 MHz, CDCl_3 , gCOSY) 1.49 (d, $J = 6.8$ Hz, 3H, CH_3), 1.55-1.65 (m, 4H, H-5 and H-7), 1.70-1.80 (m, 3H, H-3, H-4), 1.88 (ddd, $J = 12.0, 8.0, 7.0$ Hz, 1H, H-3), 2.25 (m, 1H, H-3*a*), 2.94 (dt, $J = 10.5, 6.8$ Hz, 1H, H-7*a*), 3.15 (dt, $J = 9.2, 6.4$ Hz, 1H, H-2), 3.73 and 3.87 (2d, $J = 13.5$ Hz, 1H each, NCH_2Ar), 3.70-3.84 (m, 4H, OCH_2), 3.95 (dq, $J = 11.5, 6.4$ Hz, 1H, H-1'), 7.20-7.38 (m, 5H, ArH); ^{13}C NMR (100 MHz, CDCl_3 , gHSQC), see Table 1). HRFABMS calcd for $\text{C}_{19}\text{H}_{27}^{35}\text{ClNO}_3$ 336.1730 (MH^+), found 336.1727.

(2*S*,3*aR*,7*aR*)-1-Benzyl-2-[(1'*S*)-(1-chloroethyl)]octahydroindol-6-one ethylene acetal (11b): white solid; ^1H NMR (400 MHz, CDCl_3 , gCOSY) 1.39 (t, $J = 12.0$ Hz, 1H, H-7*ax*), 1.44 (d, $J = 6.8$ Hz, 3H, CH_3) 1.50-1.65 (m, 4H, H-3 and H-4), 1.75-1.95 (m, 3H, H-7*eq* and H-5), 2.42 (m, 1H, H-3*a*), 2.96 (dt, $J = 10.0, 3.0$ Hz, 1H, H-2), 3.23 (ddd, $J = 11.6, 5.6, 5.6$ Hz, 1H, H-7*a*), 3.72 and 3.84 (2d, $J = 14$ Hz, 1H each, NCH_2), 3.78-3.90 (m, 4H, OCH_2), 4.05 (qd, $J = 6.8, 3.2$ Hz, 1H, H-1'), 7.20-7.40 (m, 5H, ArH); ^{13}C NMR (100 MHz, CDCl_3 , gHSQC), see Table 1. HRFABMS calcd for $\text{C}_{19}\text{H}_{27}^{35}\text{ClNO}_3$ 336.1730 (MH^+), found 336.1741.

(2*S*,3*aR*,7*aR*)-1-Benzyl-2-[(1'*R*)-(1-chloroethyl)]octahydroindol-6-one ethylene acetal (12b): yellow oil; ^1H NMR (300 MHz, CDCl_3) 1.48 (d, $J = 6.6$ Hz, CH_3), 1.40-2.05 (m, 8H), 2.25 (m, 1H), 3.20 (m, 2H), 3.6-4.0 (m, 7H), 7.20-7.35 (m, 5H); ^{13}C NMR (75 MHz, CDCl_3), see Table 1.

Ring expansion of chlorides **9b-12b**

Method A. A solution of the appropriate chloride derivative **9b-12b** (0.16 mmol) in THF (1 mL) was treated with AgOAc (0.48 mmol, 3 equiv) at reflux for 4 h. The reaction mixture was filtered through a bed of Celite and diluted with CH_2Cl_2 . The organic layer was washed with saturated NaHCO_3 (10 mL), dried and concentrated to afford the corresponding mixture of acetates **9c-12c** and **17-20**. (See Table 1 in the main paper for results in each series and below for the NMR data of formed acetates).

- From **9b**, a mixture of acetates **9c** and **17** was obtained (80% overall yield) in a 1:1.3 ratio according to the NMR spectrum. Purification and separation of the compounds was performed by chromatography (SiO₂, CH₂Cl₂/EtOAc; 9:1).
- From **10b**, a mixture of acetates **10c** and **18** was obtained (70 % overall yield) in a 2.9:1 ratio according to the NMR spectrum. Purification and separation of the compounds was performed by chromatography (SiO₂, CH₂Cl₂/EtOAc; 9:1).
- From **11b**, a non-separable mixture of acetates **11c** and **19** was obtained (80 % overall yield) in a 5.7:1 ratio according to the NMR spectrum and GC-MS analysis. Purification of the mixture of compounds was performed by chromatography (SiO₂, CH₂Cl₂/EtOAc 9:1).
- From **12b**, a non-separable mixture of acetates **12c** and **20** was formed (60 % overall yield) in a 13:1 ratio according to the NMR spectrum. Purification of the mixture of compounds was performed by chromatography (SiO₂, CH₂Cl₂/EtOAc 9:1).

Ring expansion of alcohols **9a-12a**

Method B. A solution of the appropriate alcohol derivative **9a – 12a** (0.16 mmol) in THF (1 mL) was treated with MsCl (0.19 mmol, 1.2 equiv) and Et₃N (0.64 mmol, 4 equiv) under an argon atmosphere at – 20 °C for 1 h. AgOAc (0.48 mmol, 3 equiv) was added and the resulting mixture was warmed to rt over a period of 1 h. The reaction mixture was filtered through a bed of Celite and diluted with CH₂Cl₂. The organic layer was washed with a saturated aqueous NaHCO₃ solution (10 mL), dried and concentrated to afford the corresponding mixture of acetates.

From 9a. A mixture of acetates **9c** and **17** was obtained in a 1:2.2 ratio according the NMR spectrum in 78 % yield. Purification and separation of the compounds was performed by chromatography (SiO₂, CH₂Cl₂/EtOAc 9:1). For analytical data of **9c** and **17**, see the main text and Tables 1 and 2.

From 10a. A mixture of acetates **10c** and **18** was obtained in a 1.1:1 ratio according the NMR spectrum in 66 % yield. Purification and separation of the compounds was performed by chromatography (SiO₂, CH₂Cl₂/EtOAc 9:1).

(2*S*,3*aS*,7*aS*)-1-Benzyl-2-[(1'*S*)-(1-acetoxyethyl)]octahydroindol-6-one ethylene acetal (10c):

Colourless oil. $R_f = 0.26$ (SiO₂, CH₂Cl₂/EtOAc 9:1). $[\alpha]_D^{20} - 57$ (c 0.2, CHCl₃); IR 1732 cm⁻¹; ¹H NMR (400 MHz, CDCl₃, gCOSY) 1.21 (d, $J = 6.0$ Hz, 3H, CH₃), 1.50 (t, $J = 11.0$ Hz, 1H, H-5), 1.50-1.80 (m, 7H), 1.90 (s, 3H, OAc), 2.25 (m, 1H, H-3a), 2.85 (dt, $J = 11.0, 6.4$ Hz, 1H, H-7a), 3.00 (dt, $J = 10.0, 7.0$ Hz, 1H, H-2), 3.70 and 3.94 (2d, $J = 14.0$ Hz, 1H each, NCH₂Ar), 3.74-3.86 (m, 4H, OCH₂), 4.91 (quint, $J = 6.2$ Hz, 1H, H-1'), 7.20-7.35 (m, 5H, ArH); ¹³C NMR (100 MHz, CDCl₃), see Table 1. HRFABMS: calcd for C₂₁H₃₀NO₄ 360.2175 (MH⁺), found 360.2163.

(2*R*,3*R*,4*aS*,8*aS*)-3-Acetoxy-1-benzyl-2-methyl-7-oxodecahydroquinoline ethylene acetal (18):

Colourless oil. $R_f = 0.58$ (SiO₂, CH₂Cl₂/EtOAc 9:1). $[\alpha]_D^{20} - 24$ (c 1.0, CHCl₃); IR 1733 cm⁻¹; ¹H NMR (400 MHz, CDCl₃, gCOSY) 1.10 (d, $J = 7.0$ Hz, 3H, CH₃), 1.48 (m, 1H, H-5), 1.52 (m, 1H, H-4), 1.59 (m, 2H, H-6), 1.72 (t, $J = 12.4$ Hz, 1H, H-8ax), 1.80 (m, 1H, H-5), 1.87 (brt, $J = 13.0$ Hz, 1H, H-4ax), 1.94 (dm, $J = 12.4$ Hz, 1H, H-8eq), 2.00 (s, 3H, OAc), 2.08 (m, 1H, H-4a), 3.00 (dt, $J = 12.8, 4.4$ Hz, 1H, H-8a), 3.20 (q, $J = 6.5$ Hz, 1H, H-2); 3.78 and 3.84 (2d, $J = 14.4$ Hz, 1H each, NCH₂Ar), 3.86-3.95 (m, 4H, OCH₂), 4.99 (ddd, $J = 12.0, 5.6, 4.4$ Hz, 1H, H-3), 7.20-7.35 (m, 5H, ArH); ¹³C NMR (100 MHz, CDCl₃, gHSQC), see Table 2 HRFABMS: calcd for C₂₁H₃₀NO₄ 360.2175 (MH⁺), found 360.2171.

From 11a. **11c** and **19** were obtained in a 2.2:1 ratio according the NMR spectrum in 76 % yield as a unseparable mixture. Purification of the mixture of compounds was performed by chromatography (SiO₂, CH₂Cl₂/EtOAc 9:1).

(2*S*,3*aR*,7*aR*)-1-Benzyl-2-[(1'*S*)-(1-acetoxyethyl)]octahydroindol-6-one ethylene acetal (11c):

Colourless oil. $R_f = 0.31$ (SiO₂, CH₂Cl₂); ¹H NMR (400 MHz, CDCl₃, gCOSY) 1.23 (d, $J = 6.4$ Hz, 3H, CH₃), 1.35 (t, $J = 12.4$ Hz, 1H, H-7ax), 1.45-2.05 (m, 7H), 2.07 (s, 3H, OAc), 2.25 (m, 1H, H-3a), 2.89 (dt, $J = 10.0, 2.8$ Hz, 1H, H-2), 3.08 (dt, $J = 12.0, 6.0$ Hz, 1H, H-7a), 3.52 and 3.88 (2d, $J = 13.6$ Hz, 1H each, NCH₂), 3.80-4.00 (m, 4H, OCH₂), 5.14 (qd, $J = 6.4, 2.4$ Hz, 1H, H-1'), 7.15-7.35 (m, 5H, ArH); ¹³C NMR (100 MHz, CDCl₃, gHSQC), see Table 1. HRFABMS: calcd for C₂₁H₃₀NO₄ 360.2175 (MH⁺), found 360.2158.

(2*R*,3*R*,4*aR*,8*aR*)-3-Acetoxy-1-benzyl-2-methyl-7-oxodecahydroquinoline ethylene acetal

(19): Colourless oil. R_f = 0.31 (SiO₂, CH₂Cl₂); ¹H NMR (400 MHz, CDCl₃) 1.04 (d, J = 7.2 Hz, 3H, CH₃), 1.45-2.05 (m, 8H), 2.07 (s, 3H, OAc), 2.34 (m, 1H, H-4a), 2.89 (qd, J = 7.2, 1.2 Hz, 1H, H-2ax), 3.09 (dt, J = 11.5, 4.4 Hz, 1H, H-8a), 3.68-3.80 (m, 6H, NCH₂Ar and OCH₂), 4.82 (q, J = 2.8 Hz, 1H, H-3eq), 7.15-7.30 (m, 5H, ArH); ¹³C NMR (100 MHz, CDCl₃), see Table 2. HRFABMS: calcd for C₂₁H₃₀NO₄ 360.2175 (MH⁺), found 360.2158.

From 12a. Acetate **12c** and traces of **20** were formed in a 13:1 ratio, according the NMR spectrum and GC-MS analysis, in 40% yield as a unseparable mixture. Purification of the mixture of compounds was performed by chromatography (SiO₂, CH₂Cl₂/EtOAc 9:1).

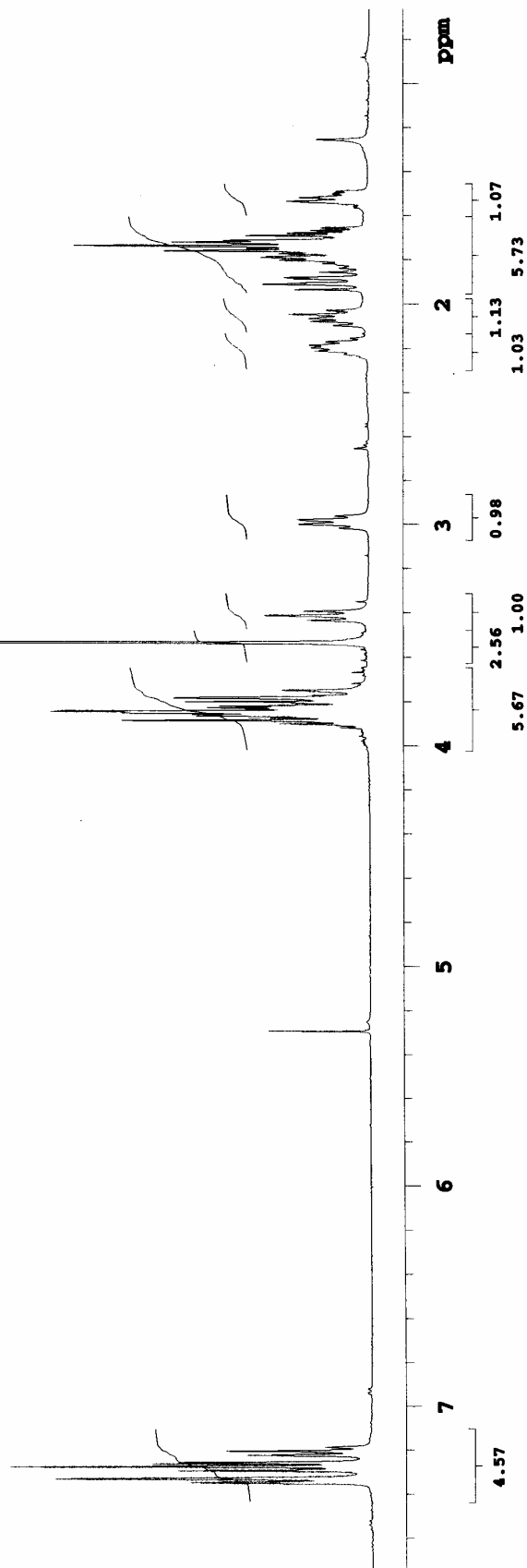
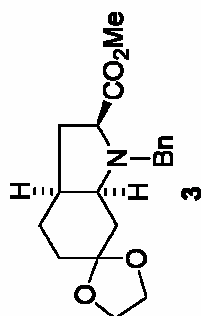
(2*S*,3*aR*,7*aR*)-1-Benzyl-2-[(1'*R*)-(1-acetoxyethyl)]octahydroindol-6-one ethylene acetal (12c):

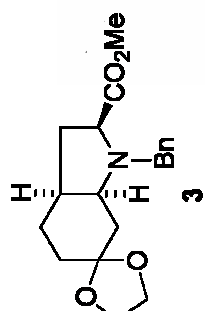
White Solid. R_f = 0.31 (SiO₂, CH₂Cl₂); ¹H NMR (400 MHz, CDCl₃, gCOSY) 1.24 (d, J = 6.4 Hz, 3H, CH₃), 1.37 (t, J = 12.0 Hz, 1H, H-7ax), 1.49 (dm, J = 12.0 Hz, 1H, H-5eq), 1.60-1.70 (m, 3H, H-5 and H-4), 1.75 (m, 1H, H-3), 1.80-1.90 (m, 2H, H-3 and H-7eq), 1.96 (s, 3H, OAc), 2.28 (m, 1H, H-3a), 3.05 (ddd, J = 8.8, 6.4, 2.8 Hz, 1H, H-2), 3.15 (ddd, J = 11.2, 5.6, 5.6 Hz, 1H, H-7a), 3.69 and 3.88 (2d, J = 14.4 Hz, 1H each, NCH₂Ar), 3.73-3.90 (m, 4H, OCH₂), 4.95 (quint, J = 6.4 Hz, 1H, H-1'), 7.20-7.35 (m, 5H, ArH); ¹³C NMR (100 MHz, CDCl₃), see Table 1. HRFABMS: calcd for C₂₁H₃₀NO₄ 360.2175 (MH⁺), found 360.2163.

Ring expansion of octahydroindole 9a using silver trifluoroacetate. A solution of alcohol **9a** (62 mg, 0.2 mmol) in THF (1.4 mL) was treated with MsCl (0.019 mL, 0.24 mmol, 1.2 equiv) and Et₃N (0.11 mL, 0.8 mmol, 4 equiv) under argon atmosphere at – 20 °C for 1 h. CF₃CO₂Ag was added (221 mg, 1 mmol, 5 equiv) and the resulting mixture was warmed to room temperature over a period of 1 h. The mixture was treated with 2.5 N NaOH (1 mL) and stirred for 3 h. The reaction mixture was filtered through a bed of Celite and diluted with CH₂Cl₂. The organic layer was dried and concentrated to afford a mixture of **9a** and **21**, which was purified by chromatography (Al₂O₃, hexane/EtOAc 9:1) to give 16 mg (26%) of **9a** and 36 mg (58%) of (2*S*,3*R*,4*aS*,8*aS*)-1-Benzyl-3-hydroxy-2-methyl-7-oxodecahydroquinoline ethylene acetal (**21**): Colourless oil. R_f = 0.10 (Al₂O₃, Hexane/EtOAc 8:2). $[\alpha]_D^{20}$ +1.5 (c 0.6, CHCl₃); ¹H NMR (300 MHz, CDCl₃, COSY) 1.17 (d, J =

6.0 Hz, 3H, CH₃), 1.43-1.49 (m, 2H, H-5 and H-6), 1.55-1.74 (m, 5H, H-4, H-5, H-6, and H-8eq), 1.88 (t, $J = 12.6$ Hz, 1H, H-8ax), 2.04 (m, 1H, H-4a), 2.57 (dq, $J = 9.0, 6.0$ Hz, 1H, H-2ax), 2.93 (dt, $J = 12.6, 4.5$ Hz, 1H, H-8a), 3.36 (td, $J = 9.0, 7.2$ Hz, 1H, H-3ax), 3.57 and 3.90 (2d, $J = 14.4$ Hz each, NCH₂Ar), 3.79-3.94 (m, 4H, OCH₂), 7.18-7.35 (m, 5H, ArH); ¹³C NMR (75 MHz, CDCl₃, gHSQC), see Table 2. HRFABMS: calcd for C₁₉H₂₈NO₃ 318.2069 (MH⁺), found 318.2064.

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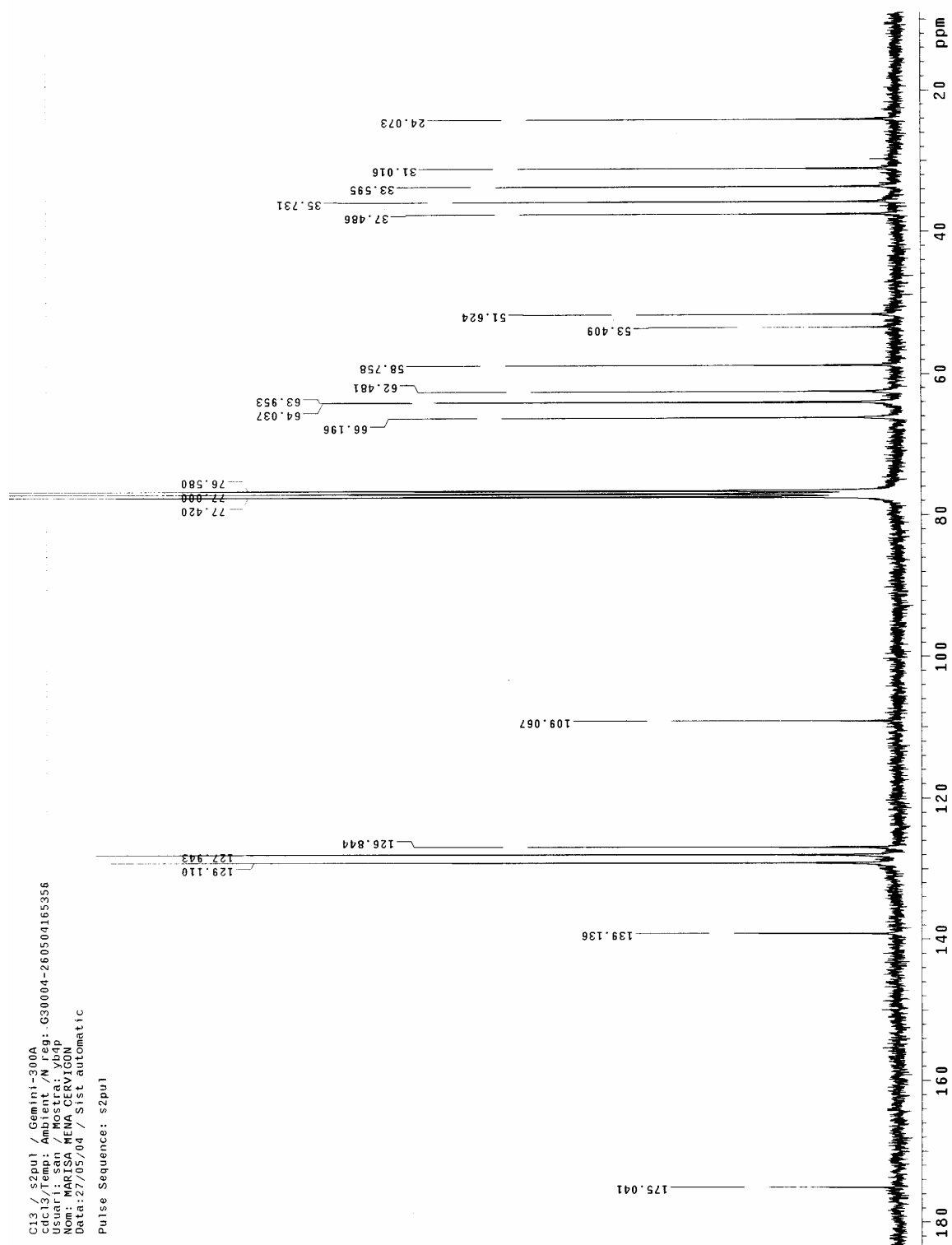


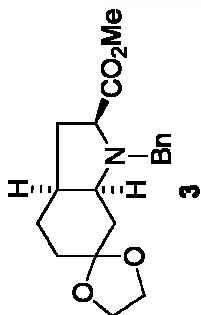


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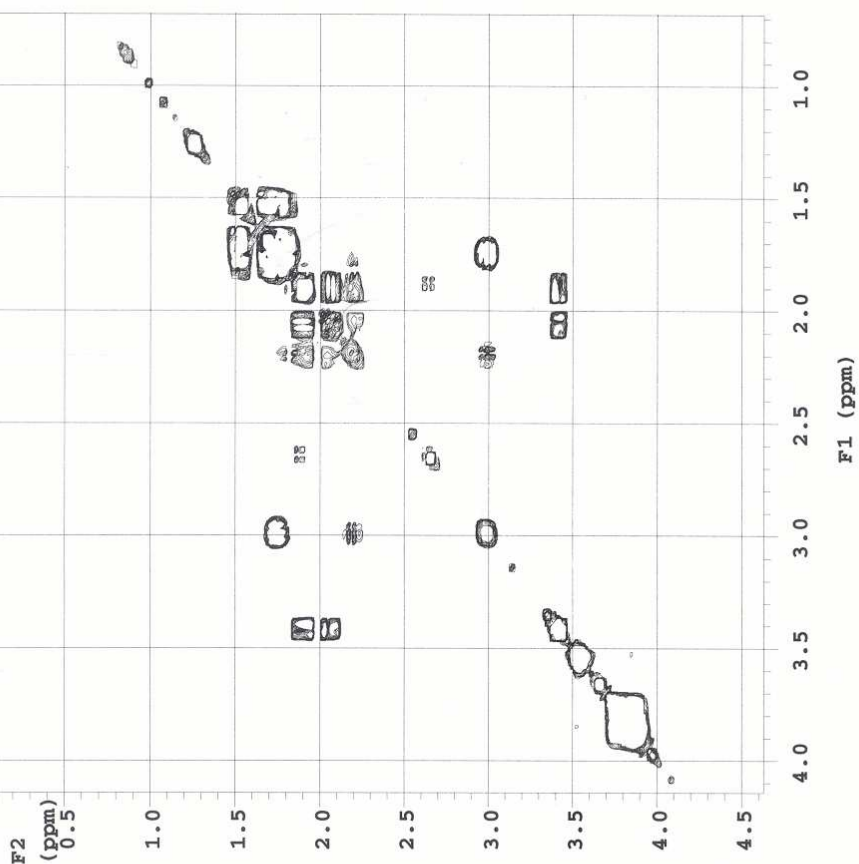


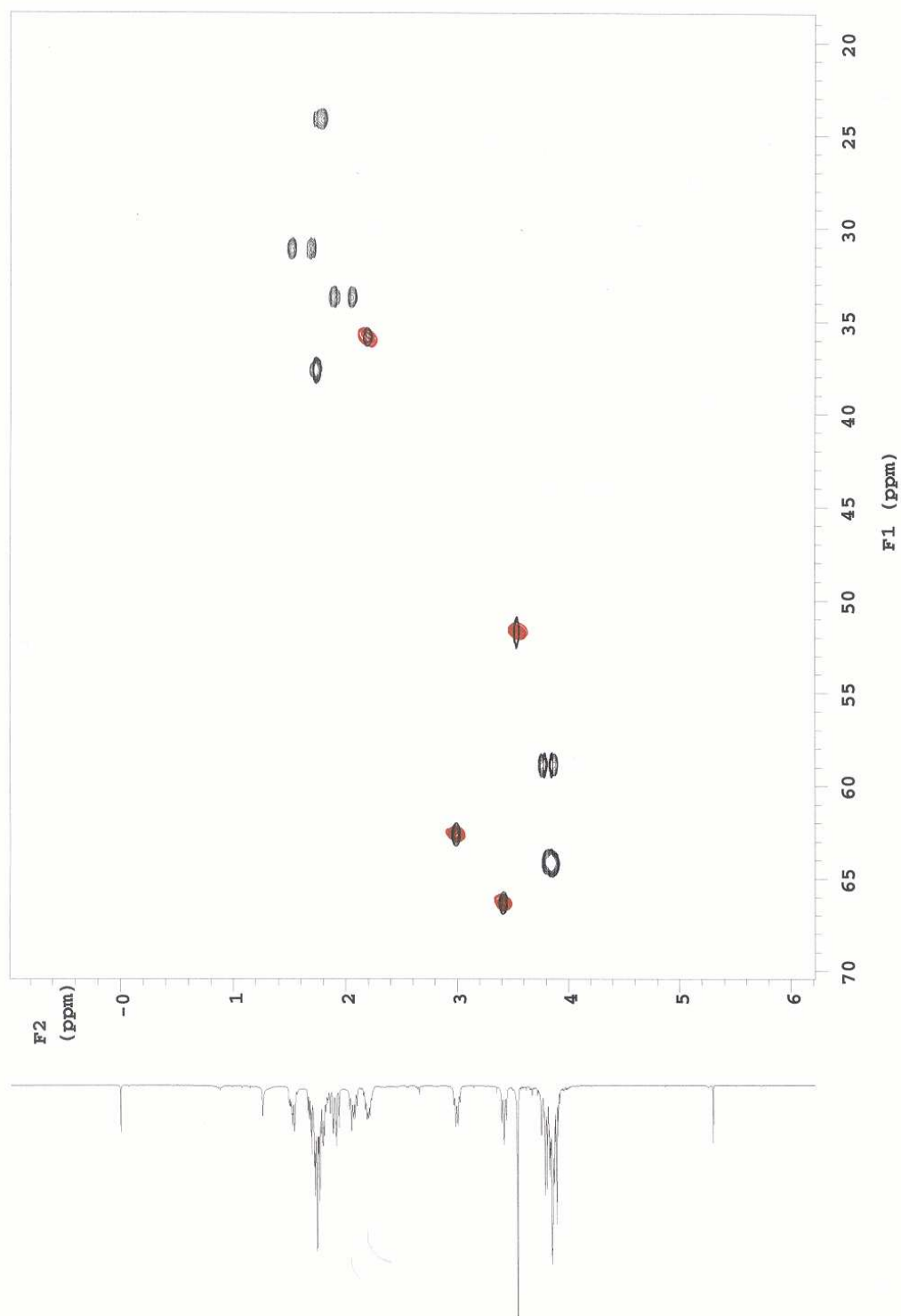
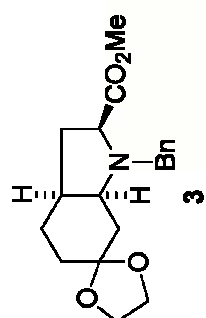
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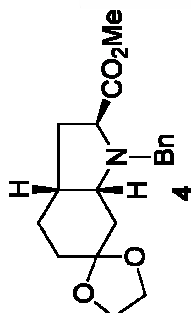
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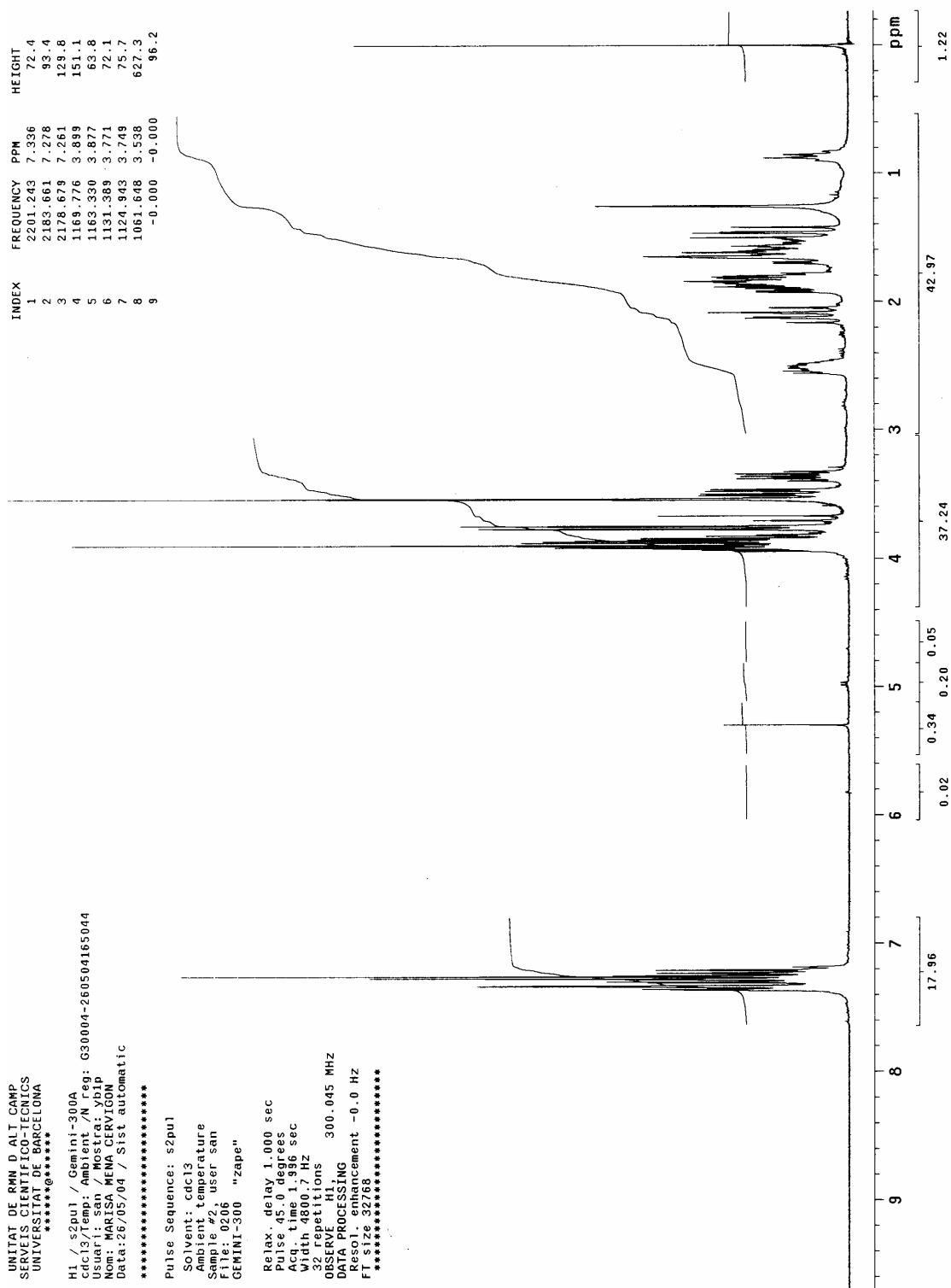
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UNIVERSITAT DE BARCELONA
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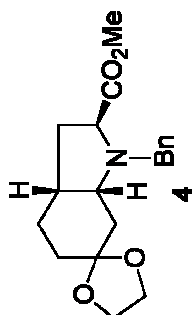
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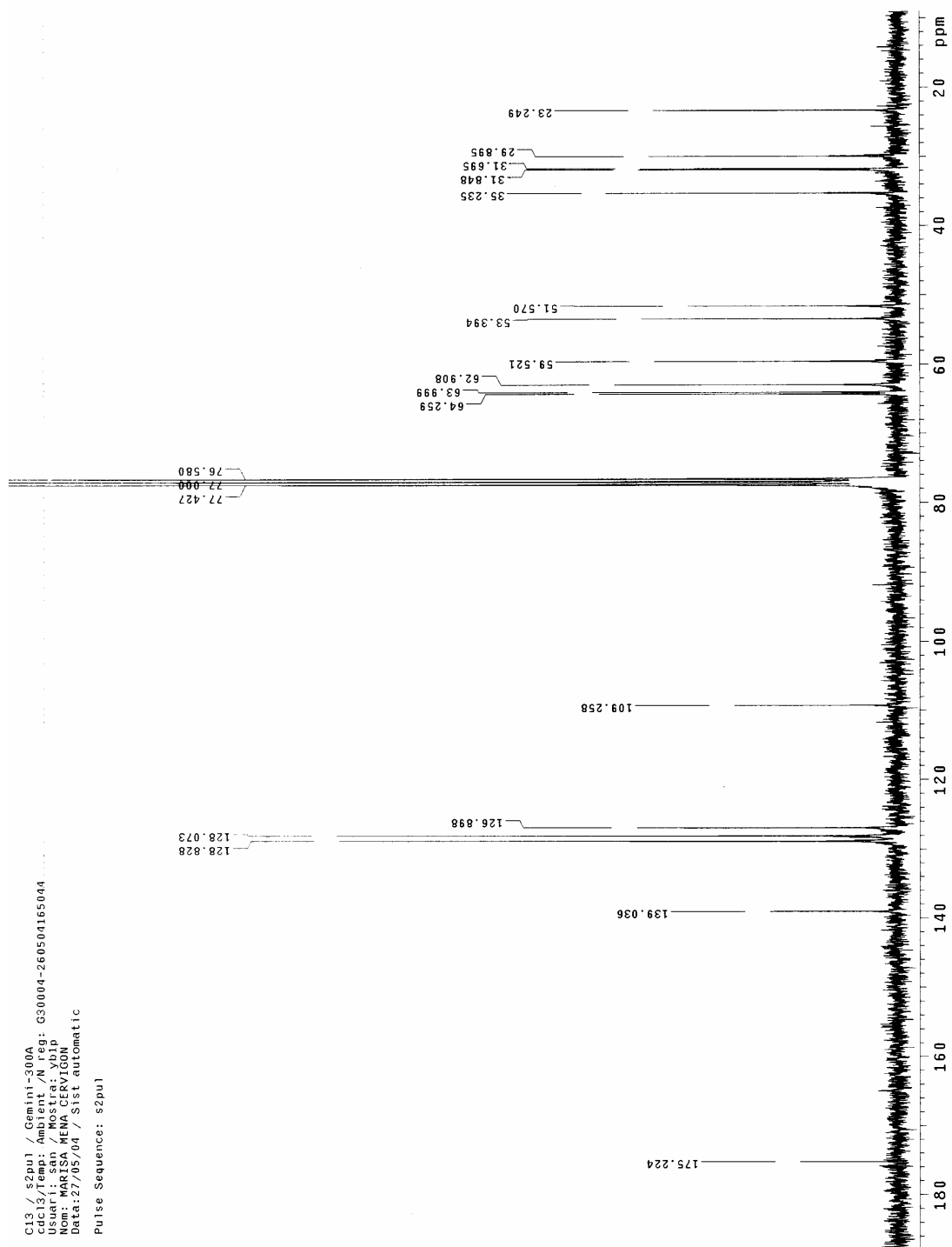
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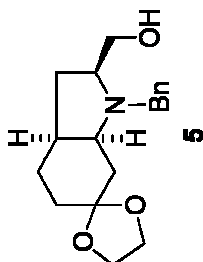
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Data: 02/08/04 / Sist automatic
Data: 02/08/04 / Sist automatic

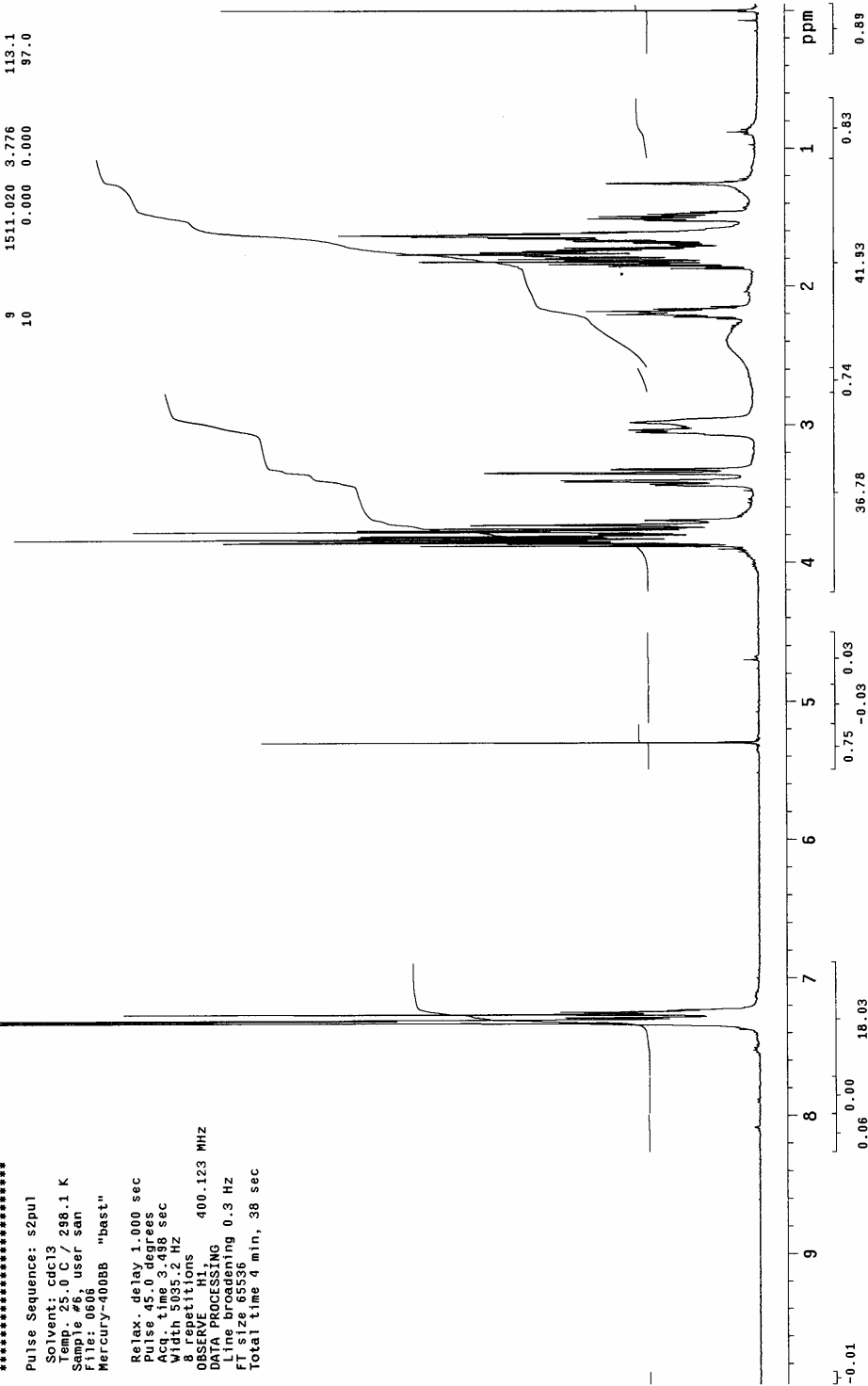
*****@*****

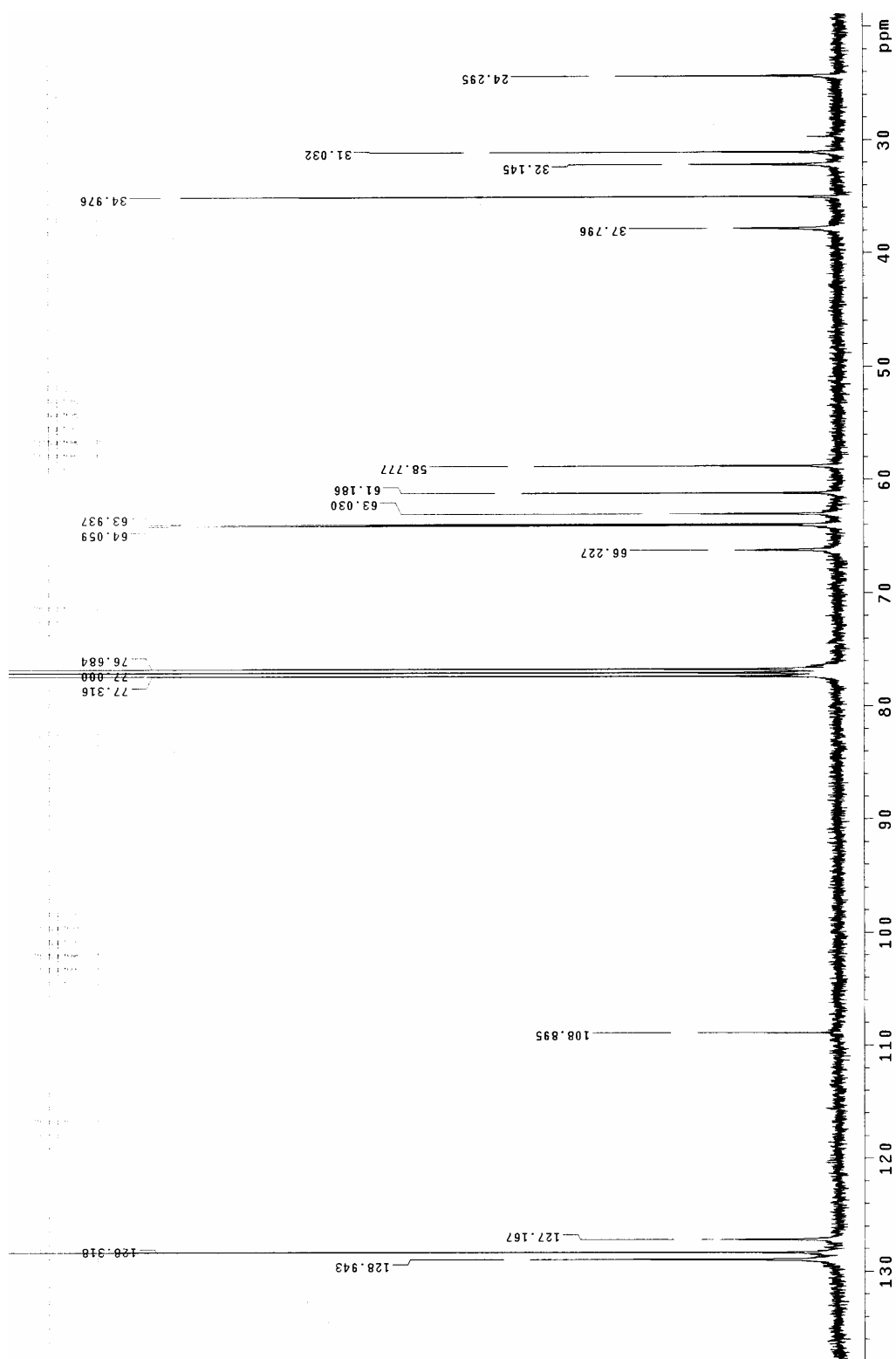
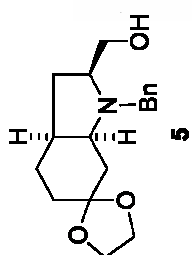
Pulse Sequence: s2pul

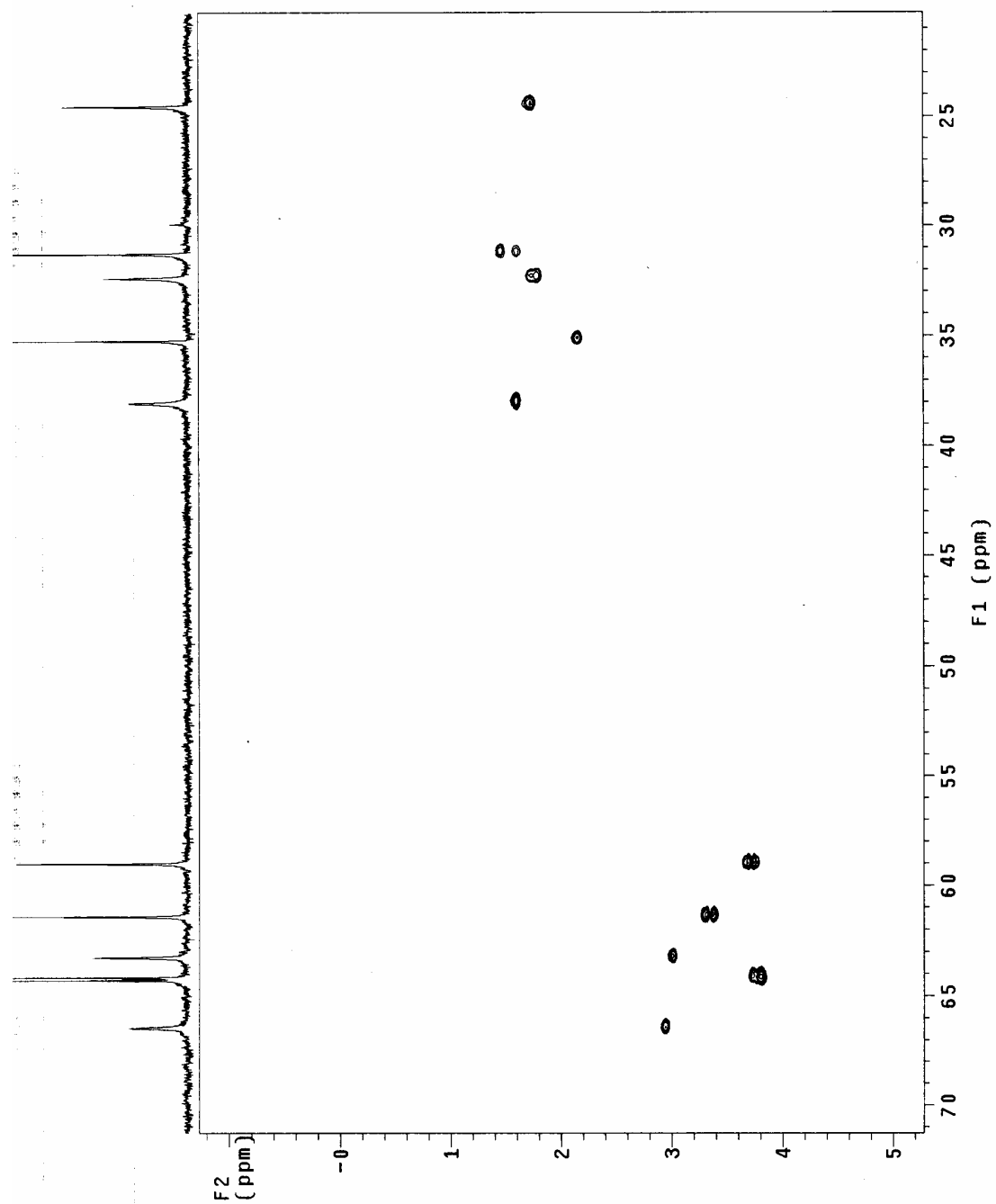
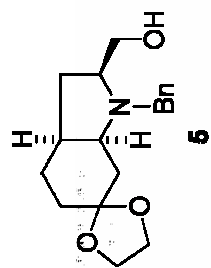
Solvent: cdc13
Temp: 25.0 C / 298.1 K
Sample #6, user san
File: 0606
Mercury-400088 "bast"

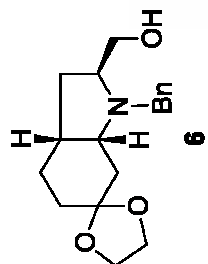
Relax. delay 1.000 sec
Pulse 45.0 degrees
Acq. time 3.498 sec
Width 3035.2 Hz
Sensitivity 8.00000000
OBSERVE H1 400.123 MHZ
DATA PROCESSING
Line broadening 0.3 Hz
FT size 65536
Total time 4 min, 38 sec

INDEX	FREQUENCY	PPM	HEIGHT
1	2929.392	7.323	253.2
2	2927.394	7.318	127.2
3	2923.692	7.307	149.4
4	2906.173	7.263	115.0
5	2119.238	5.296	90.0
6	1542.829	3.856	96.8
7	1541.292	3.852	90.2
8	1534.070	3.834	134.5
9	1511.020	3.776	113.1
10	0.000	0.000	97.0

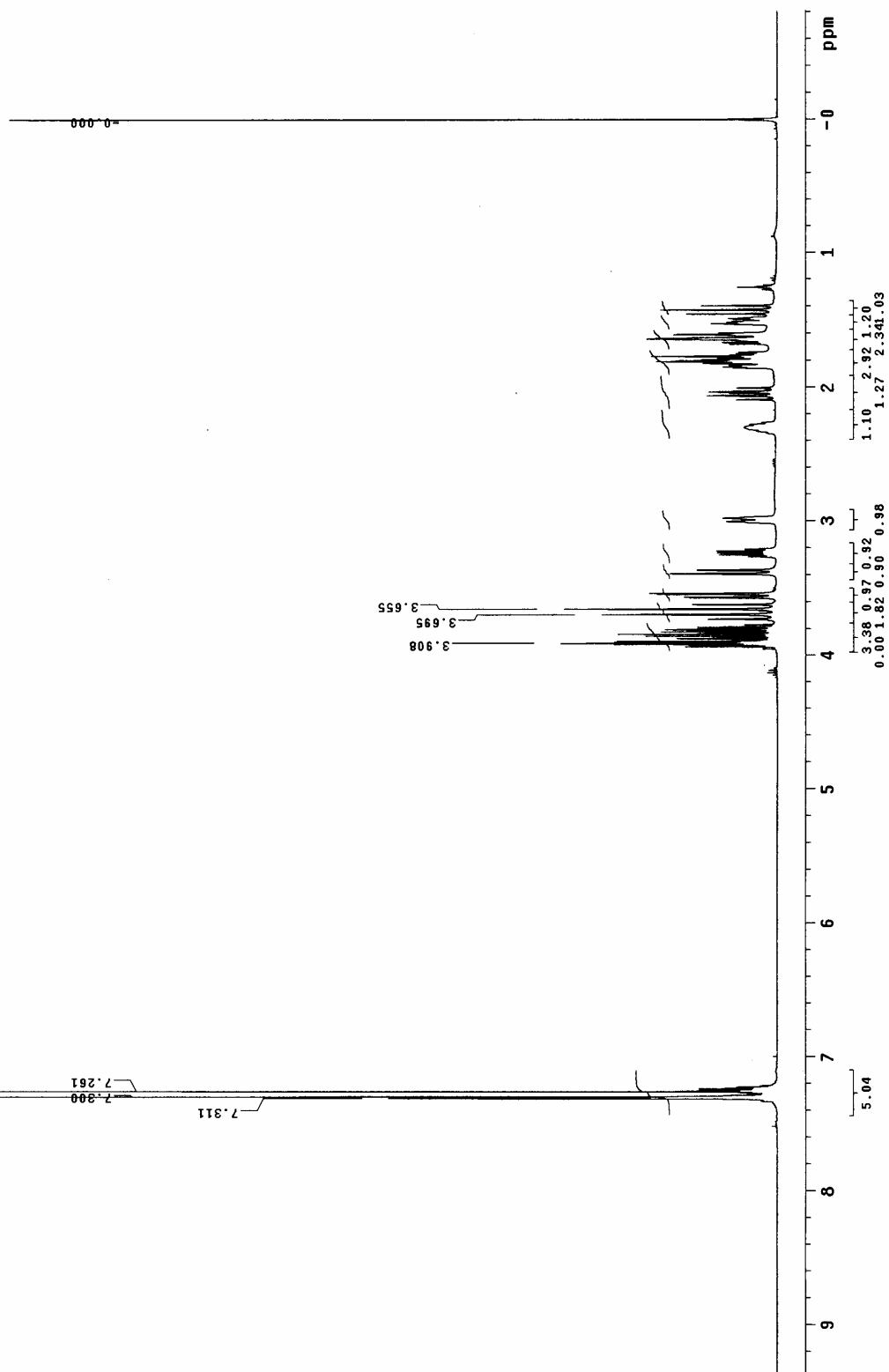


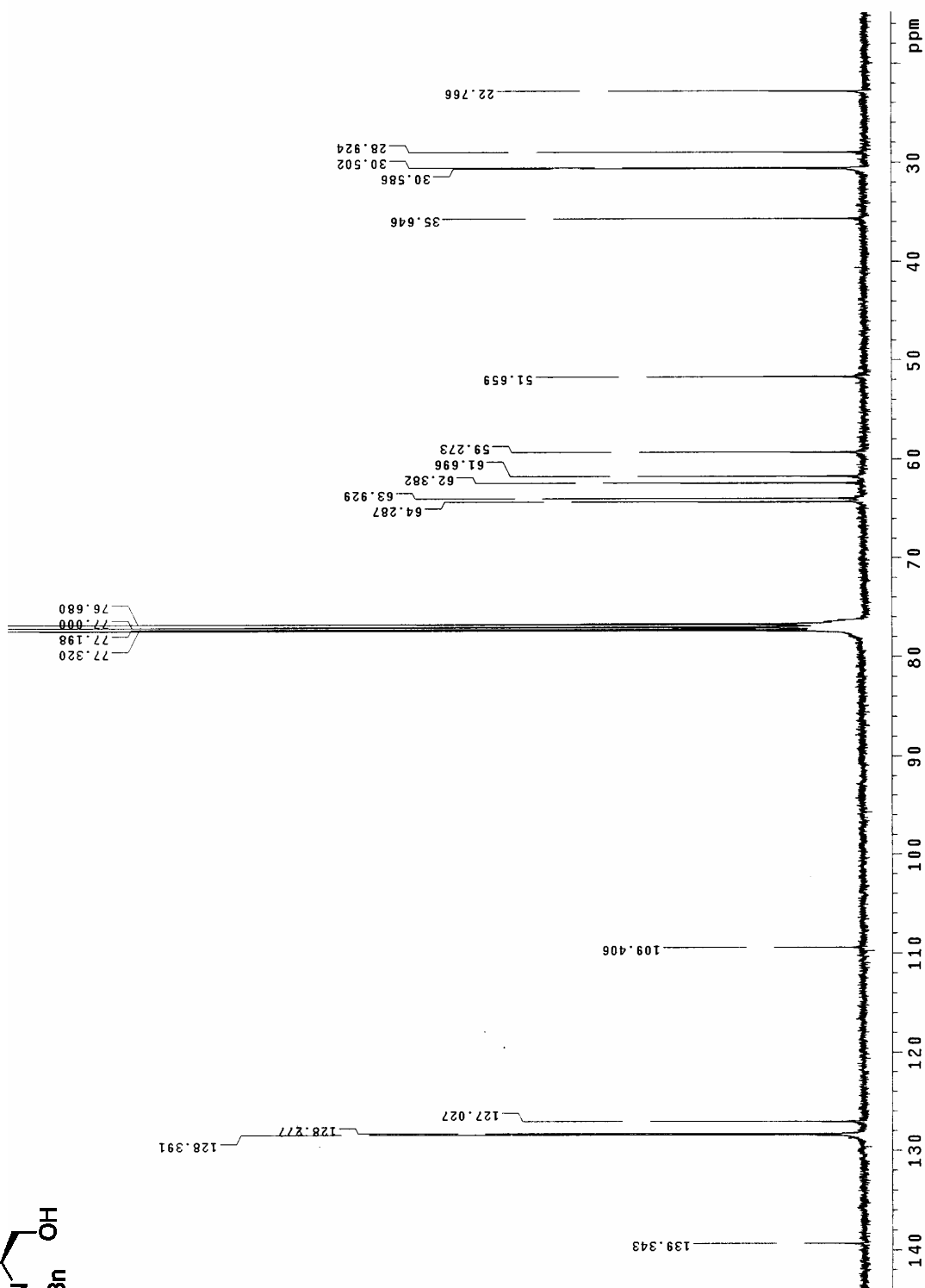
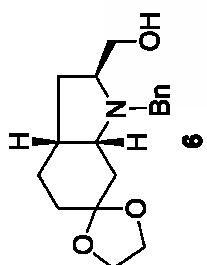


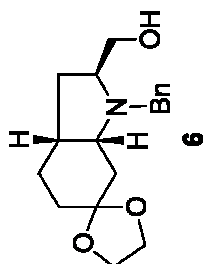




H1 / s2pul / Mercury-400
 Cdc13/epa: 25C / N Te95 M40004-010404155212
 Usuar1: san / Mostra: y22f
 Nom: MARISA MENA CERVIGNO
 Data: 01/04/04 / Sist automatic
 Pulse Sequence: s2pul



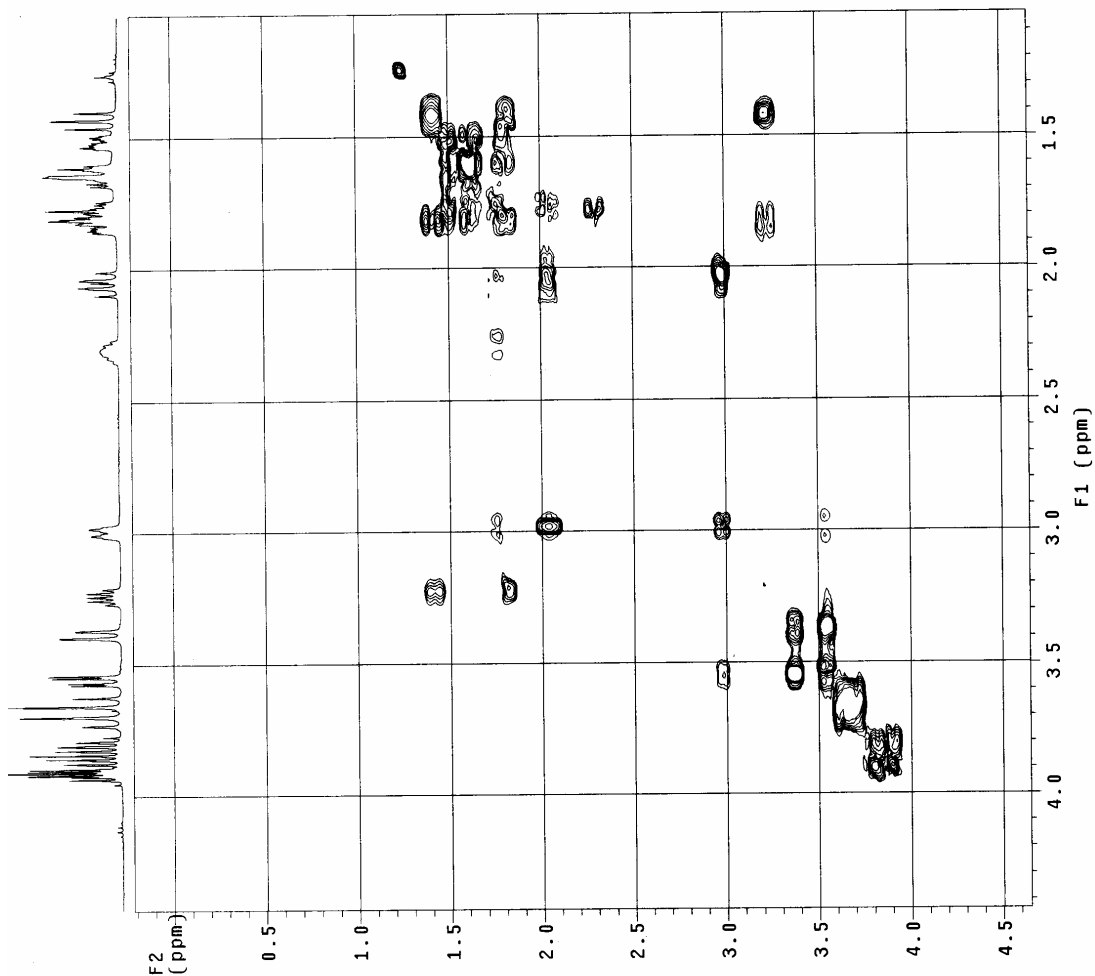


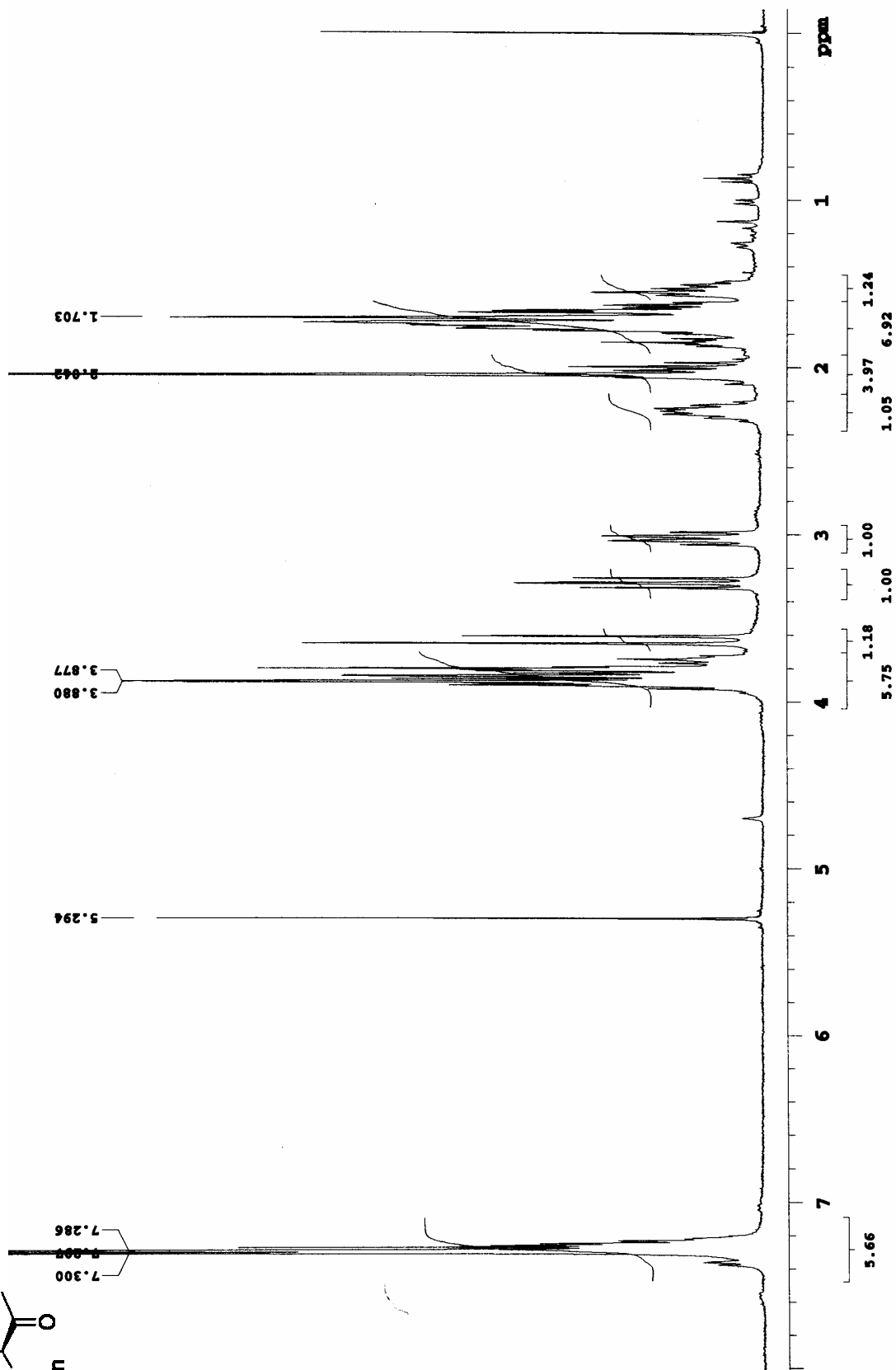
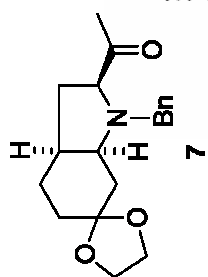


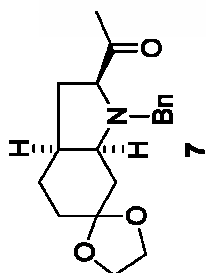
H1 / gCOSY / Mercury-400
 212 / temp: 23C / N Reg: M40004-010404165
 Usuari: san / Mostra: Vb2f
 Nom: MARISA MENA CERVIGON
 Data: 02/04/04 / Sist automatic

exp3 gCOSY

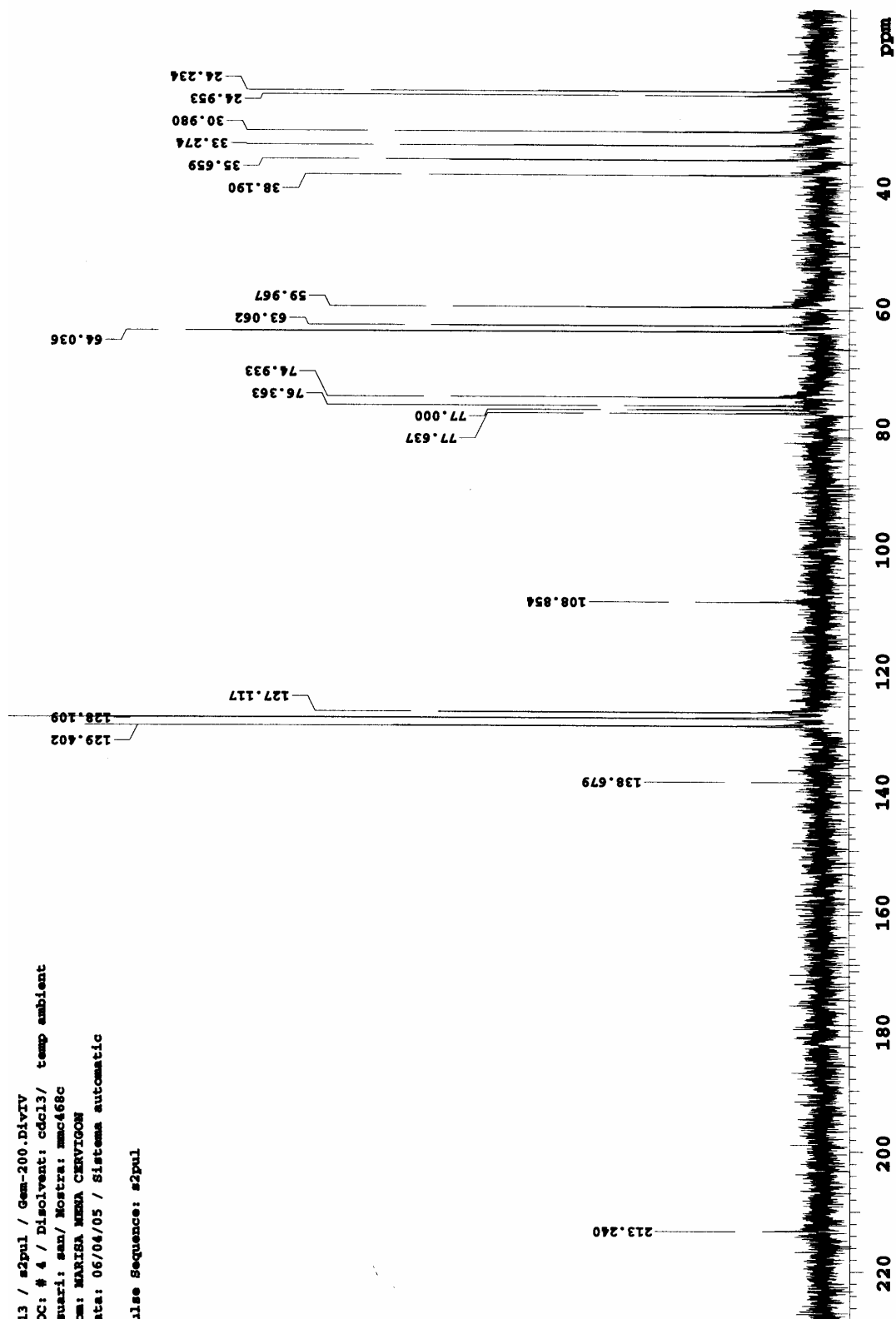
SAMPLE		FLAGS	
date	2-2004	hs	nn
solvent	cdc13	sspu1	
sample	auto_01Apr2~ hsg1v1		1005
004-21:33:29	SPECIAL		25.0
ACQUISITION			
sw	3876.0	gain	28
at	0.122	sb	0
fp	1022	f2	PROCESSING
ss	not used	sb	-0.066
d1	16	sbs	not used
nt	1.000	fn	2048
2D ACQUISITION			
sw1	3876.0	sb1	-0.066
ni	256	sbs1	not used
tn	TRANSMITTER	h1	DISPLAY
tn	400.124	sp	-86.7
sfreq	-528.6	wp	1947.5
tpwr	58	sp1	413.4
pw	12.400	wp1	1360.2
g21v11	1005	rfp	465.6
g21v11	0.000500	rfp1	465.6
gstab	0.000500	rfp1	
DECOUPLER			
dn	C13	wc	153.0
dm	nnn	sc	6.2
		wc2	153.0
		sc2	0
		vs	282
		vn	3
		at	cdc av

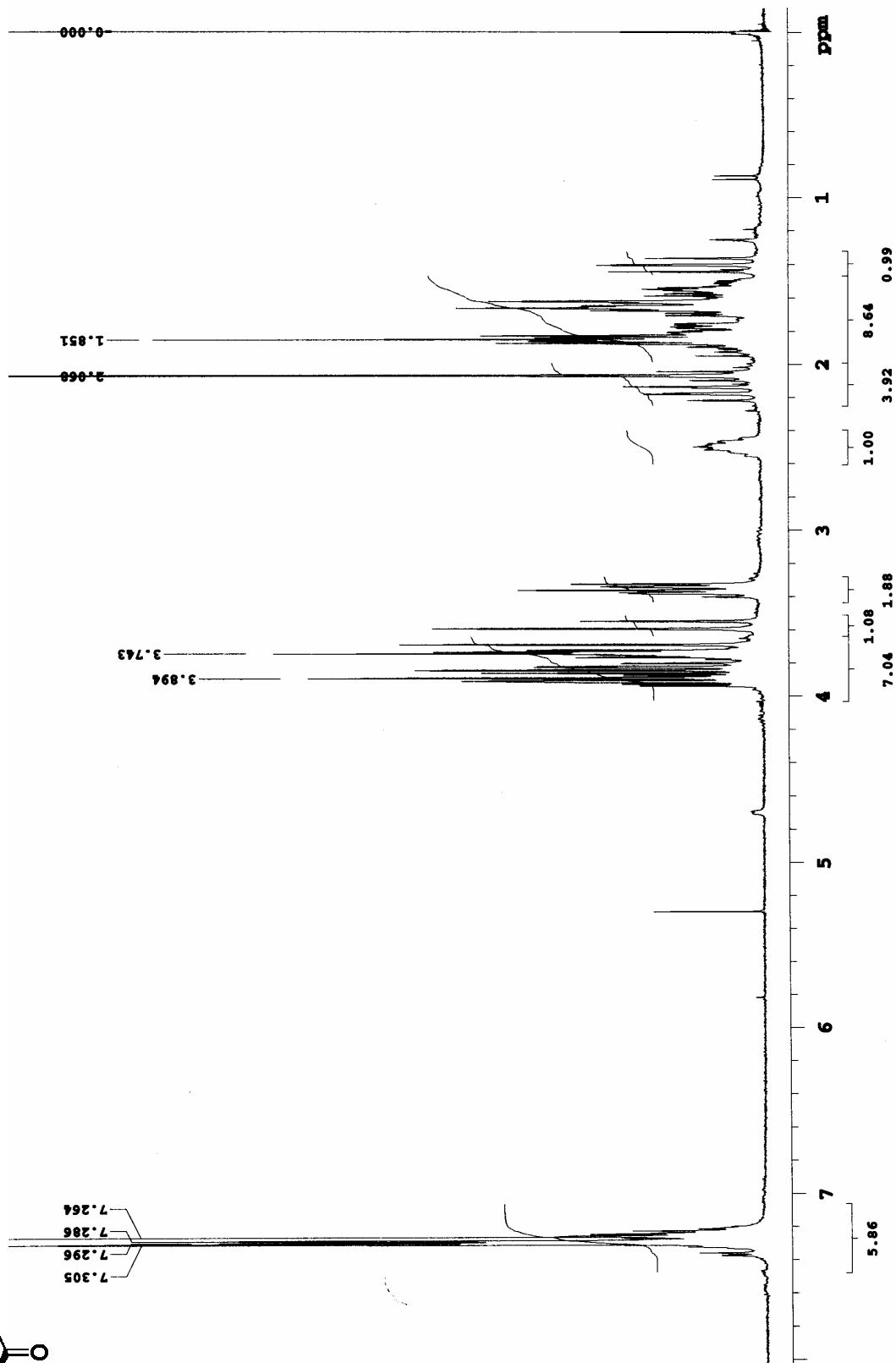
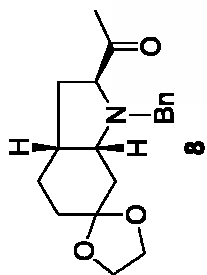


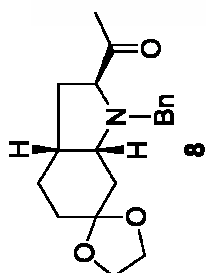




C13 / s2pul / Gem-200.D1VIV
 LOC: # 4 / Disolvent: cdcl3 / temp ambient
 Usuari: san/ Mostra: mmc468c
 Nom: MARIJA MENA CERVIGON
 Data: 06/04/05 / Sistema automatic
 Pulse Sequence: s2pul

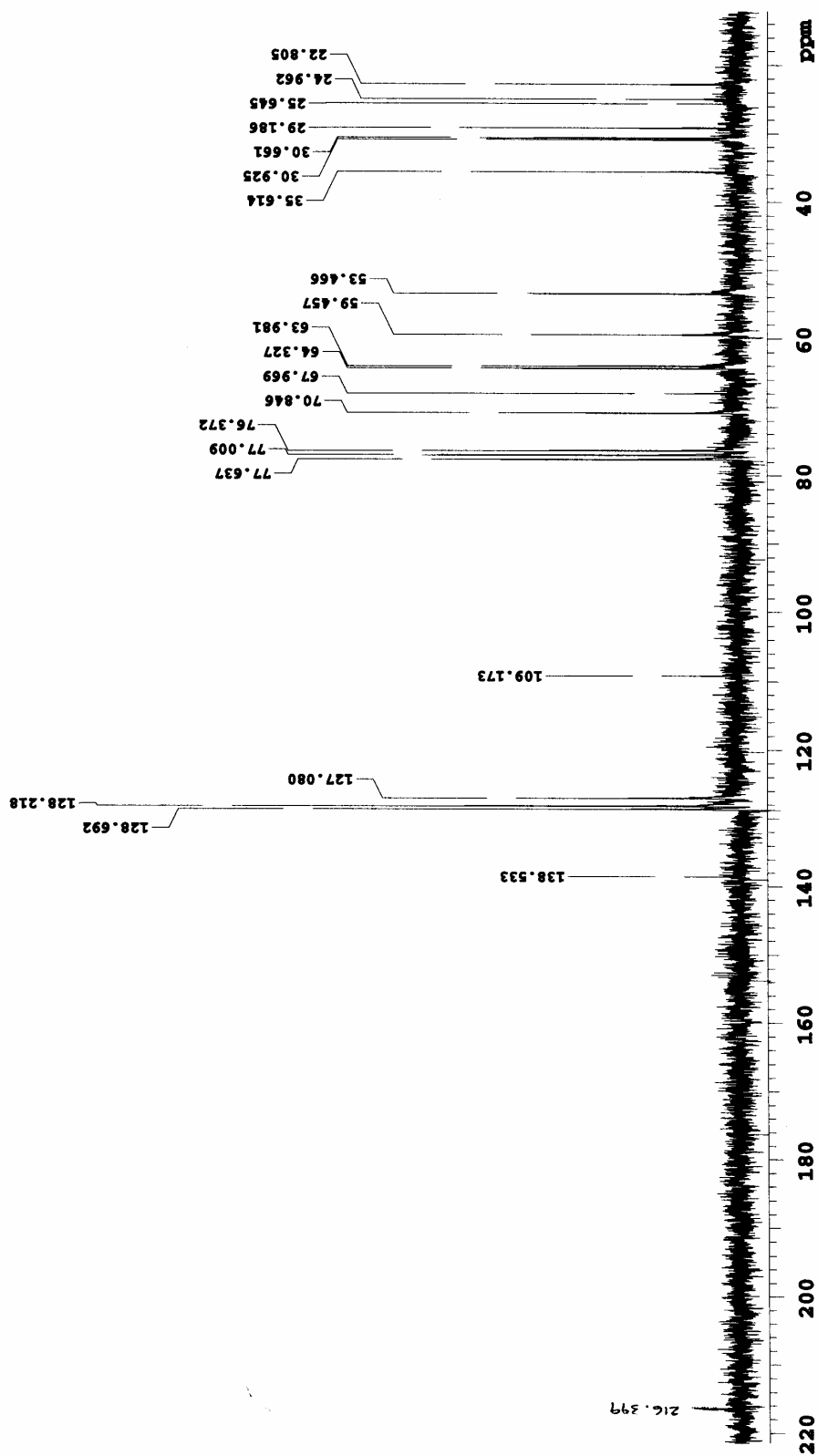


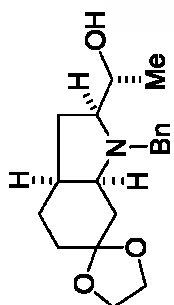




C13 / s2pul / Gem-200.DivIV
 LOC: # 5 / Disolvent: cdcl3/ temp ambient
 Usuari: san/ Mostra: mmc469c
 Nom: MARISSA NENA CERVIGON
 Data: 06/04/05 / Sistema automatic

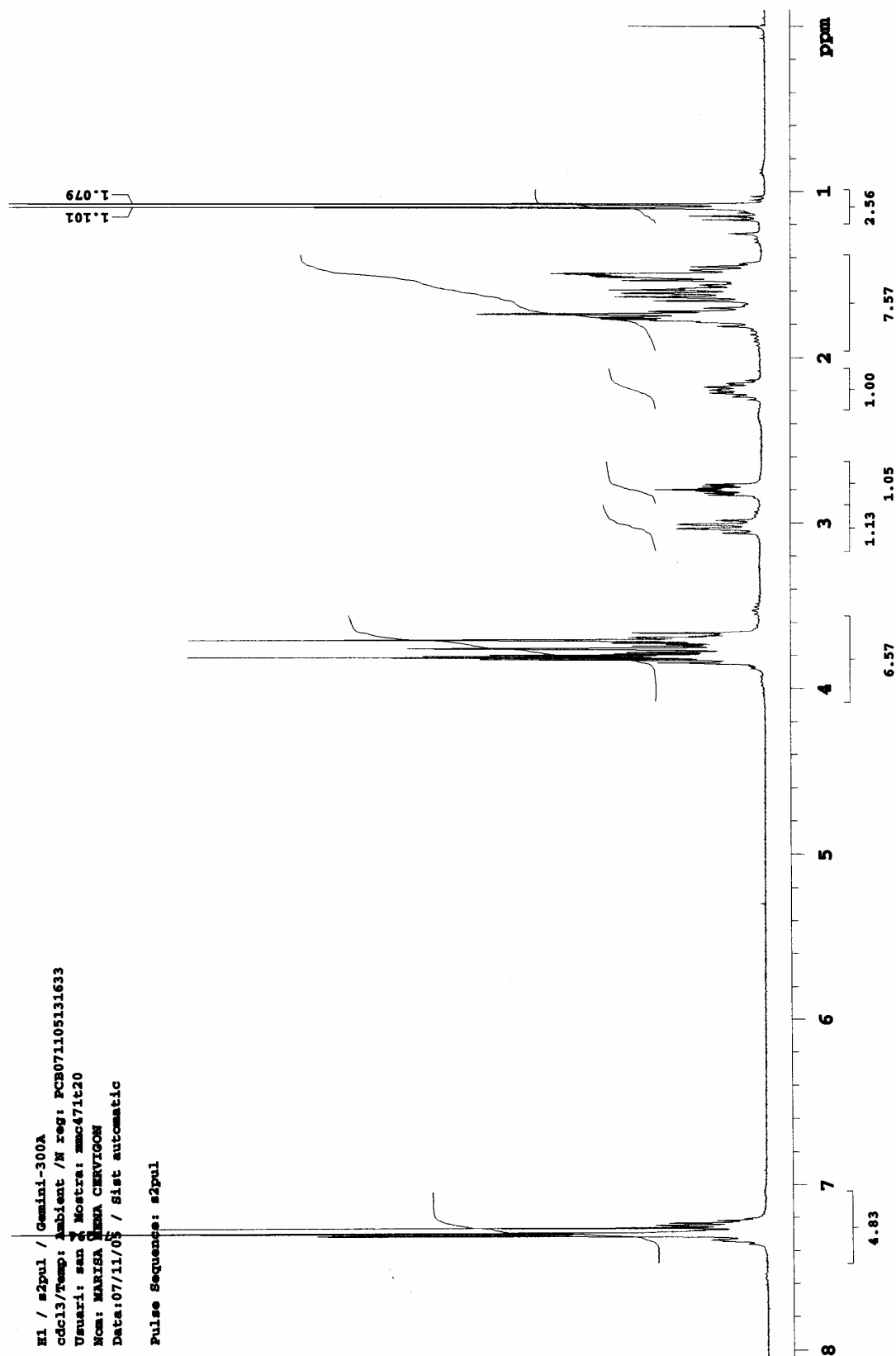
Pulse Sequence: s2pul

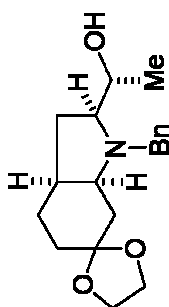




9a

H1 / s2pul / Gemini-300A
 cdcl3/Temp: Ambient /W reg: PCB071105131633
 Usuari: san q Mostra: mmc471t20
 Nom: MARISA XENA CERVIGON
 Data:07/11/05 / Sist automatic
 Pulse Sequence: s2pul

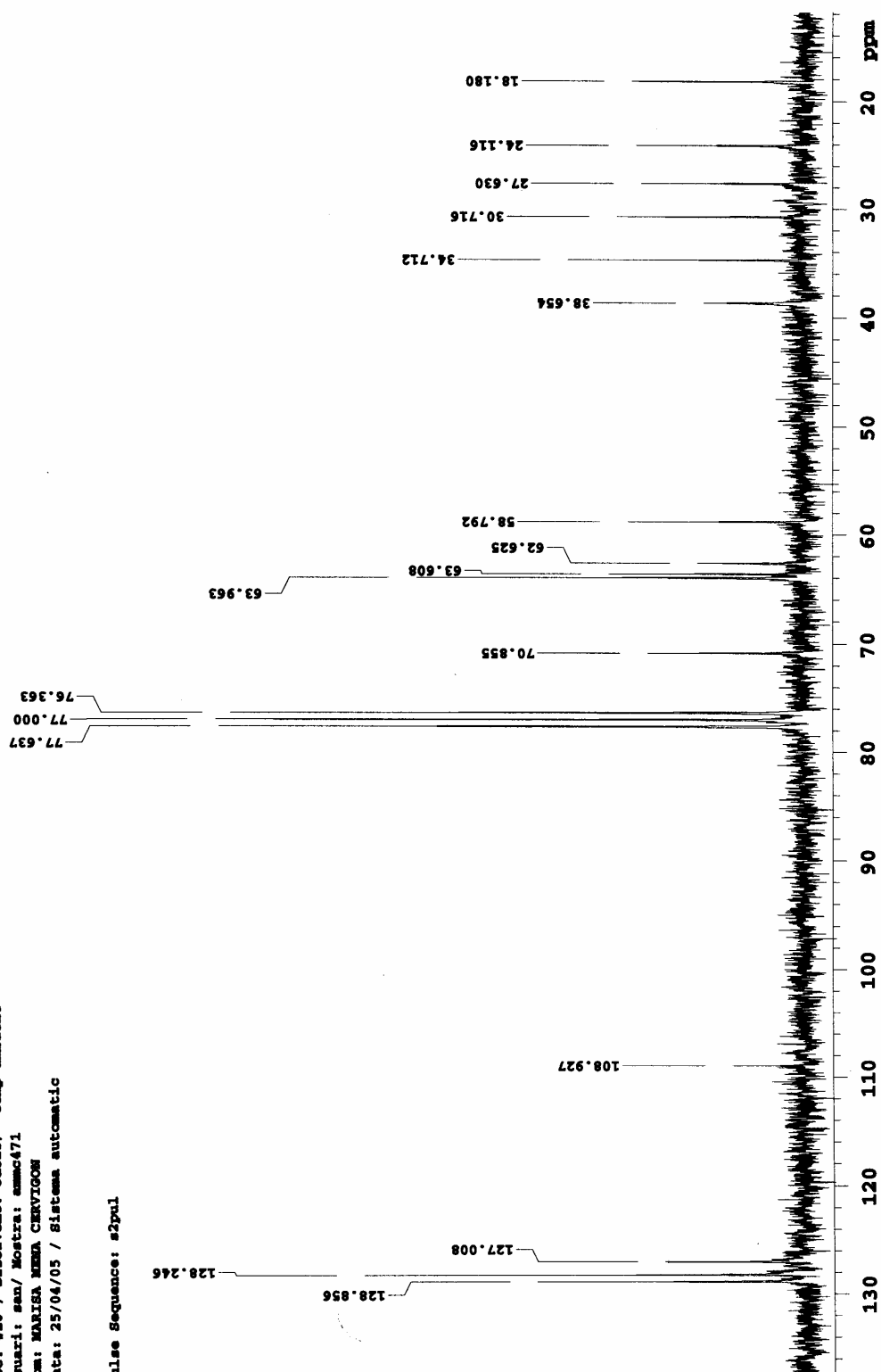


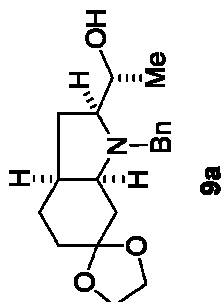


9a

H1 / s2pul / Gem-200.D1vTV
 LOC: #10 / Dissolvent: cdcl3 / temp ambient
 Usuari: san / Mostra: smac471
 Nom: MARIA NEHA CERVIGON
 Data: 25/04/05 / Sistema automatic

Pulse Sequence: s2pul

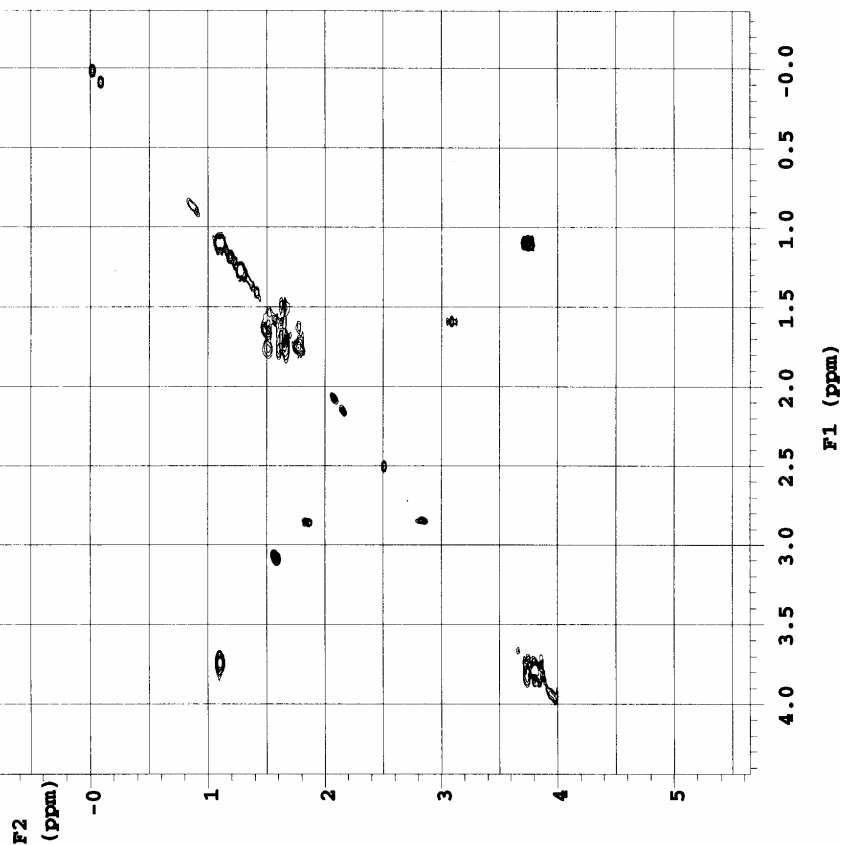


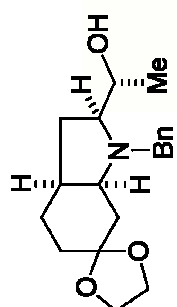


H1 / gCOSY / Mercury-600_qui
 cdc13/Temp: 25C /N reg: M40005-3560
 Usuari: san / Mostra: mmc47219a60
 Nom: MARISA MENA CERVIGON
 Data:23/09/05 / Sist automatic

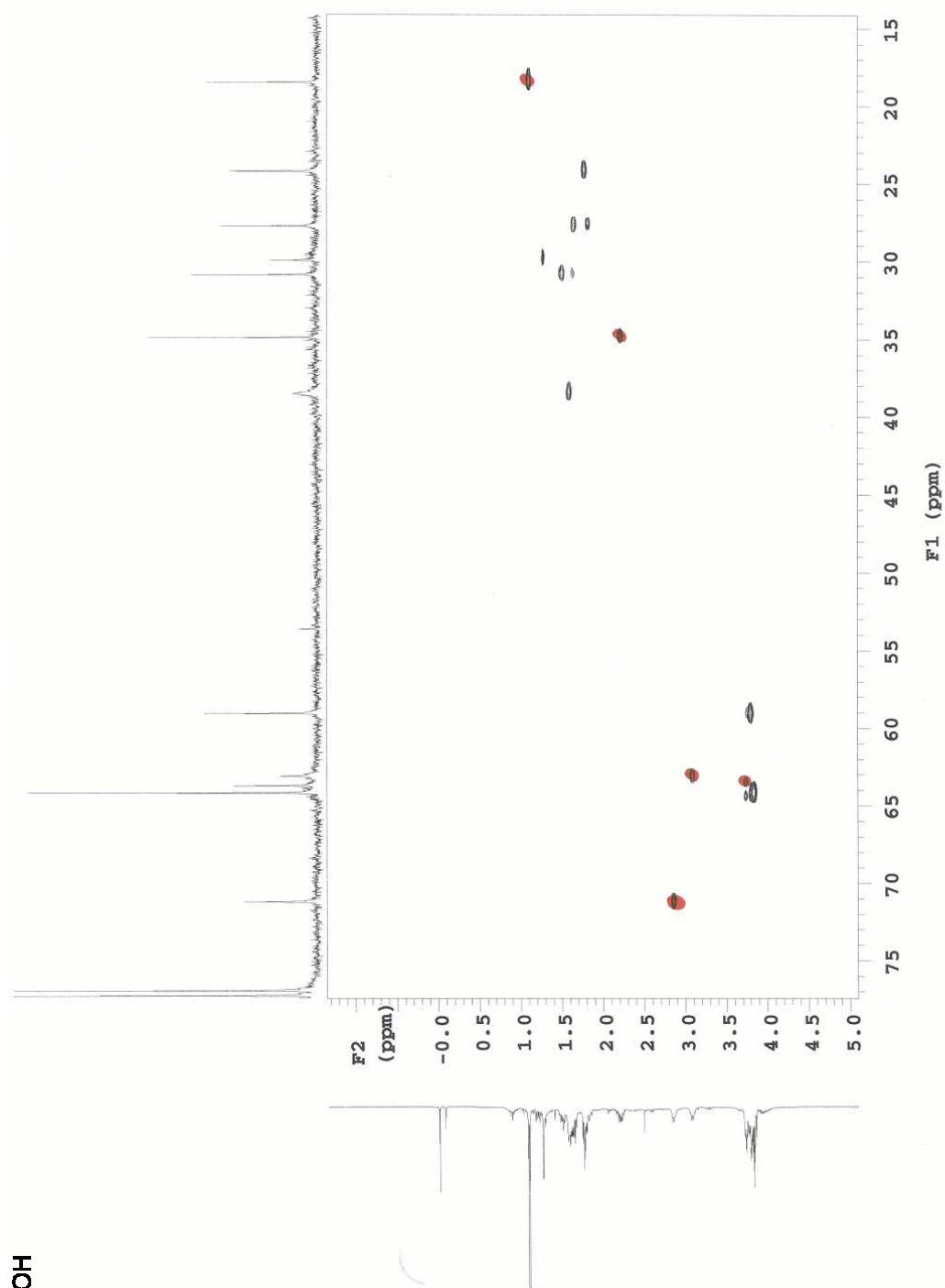
exp5 gCOSY

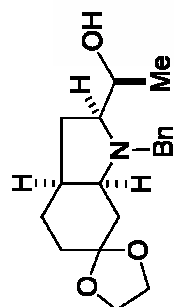
SAMPLE		FLAGS	
date	Sep 23 2005	hs	nm
solvent	cdcl3	aspu1	n
sample	auto_Custom-hsglv1	1002	
Q_23Sep2005 SPECIAL			
ACQUISITION		temp	25.0
sw	5211.0	gain	26
at	0.150	spin	0
np	1564	F2 PROCESSING	
fb	not used	ab	-0.075
ss	16	abs	not used
dl	1.000	fn	2048
nt	2	F1 PROCESSING	
2D ACQUISITION		ab1	-0.049
sw1	5211.0	abs1	not used
ni	256	procl	lp
PRESATURATION		fn1	2048
satmode		n	DISPLAY
satfrq	0	sp	-348.3
satdly	0	wp	2608.1
satpwr	0	sp1	-144.5
TRANSMITTER		wp1	1920.4
tn	H1	rf1	592.8
sfrq	400.114	rfp	0
tof	-17.4	rf11	592.8
tpwr	58	rfp1	0
pw	12.200	PLOT	
GRADIENTS		wc	131.8
gzlwl1	1002	sc	6.2
gtl1	0.001000	wc2	131.8
gstab	0.000500	sc2	0
DECOUPLER		vs	243
dn	C13	th	14
dm	nm	ai	cdc
		av	





9a

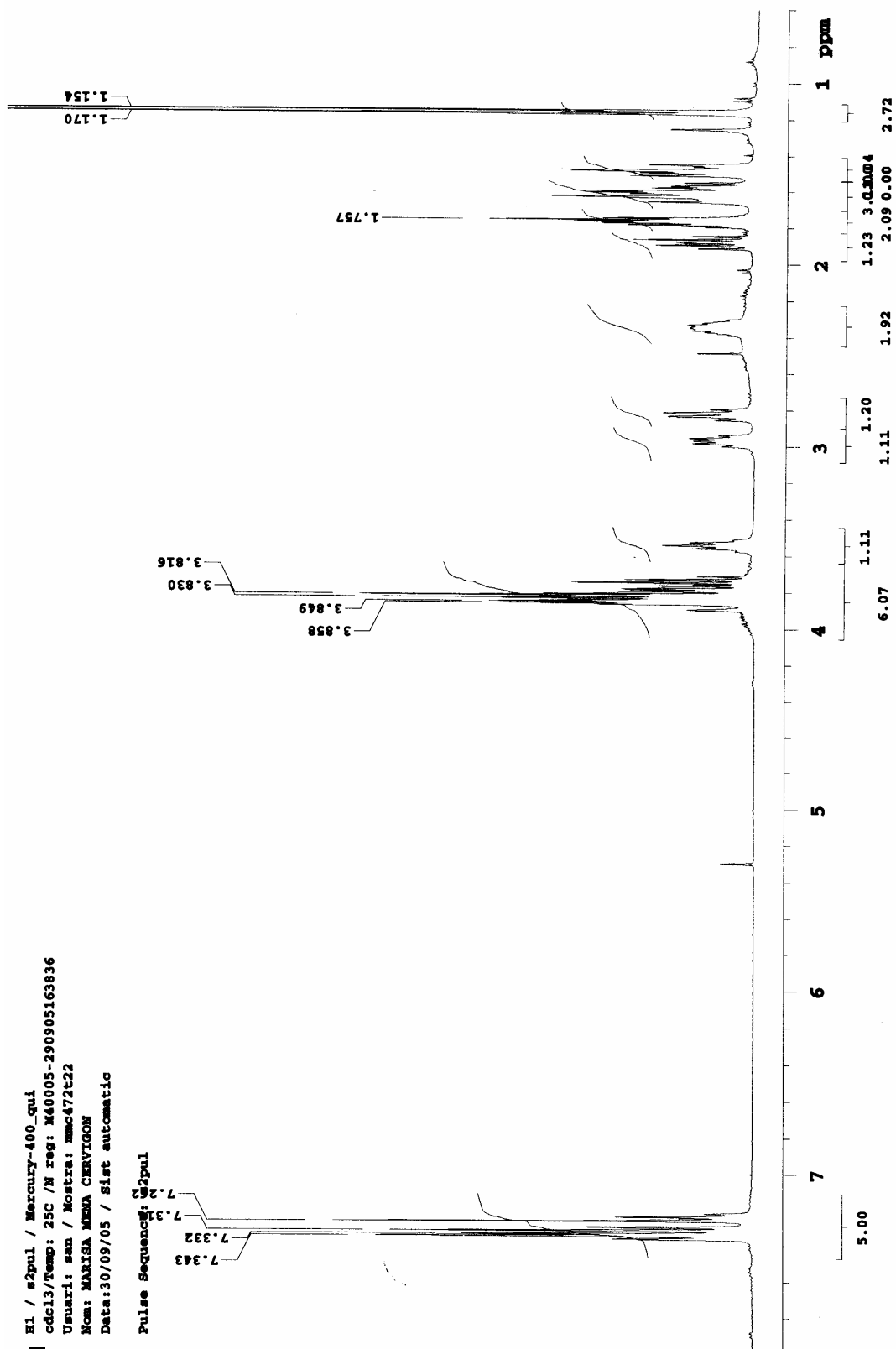


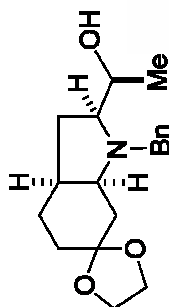


10a

H1 / s2pul / Mercury-400_qul
cdcl3/Temp: 25C / N reg: M40005-290905163836
Usuari: san / Mostra: mmc472t22
Nom: MARISA MERA CERVIGON
Data:30/09/05 / Sist automatic

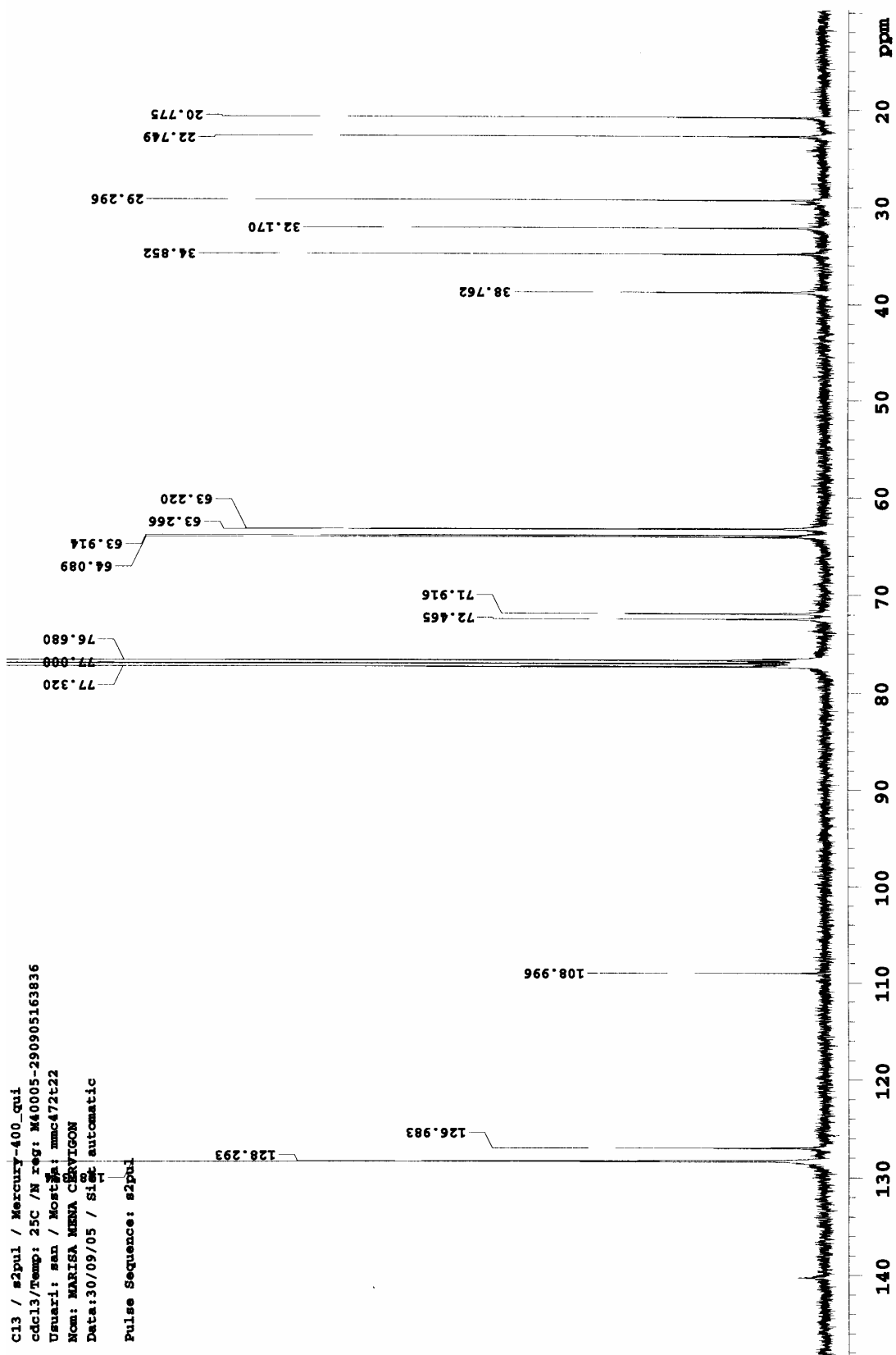
Pulse Sequence: zgpg30

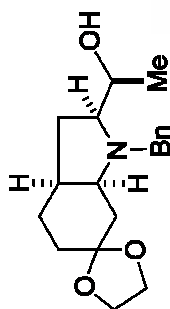




10a

C13 / s2pul / Mercury-400 qui
 cdcl3/Temp: 25C / N reg: M40005-290905163836
 Usuari: san / Most: mmc472t22
 Nom: MARISA NENA CARRIQUON
 Data:30/09/05 / Size automatic
 Pulse Sequence: s2pul





10a

H1 / gCOSY / Mercury-400_gui
cdcl3/Tmp: 25C / N reg: M40005-290905163

836

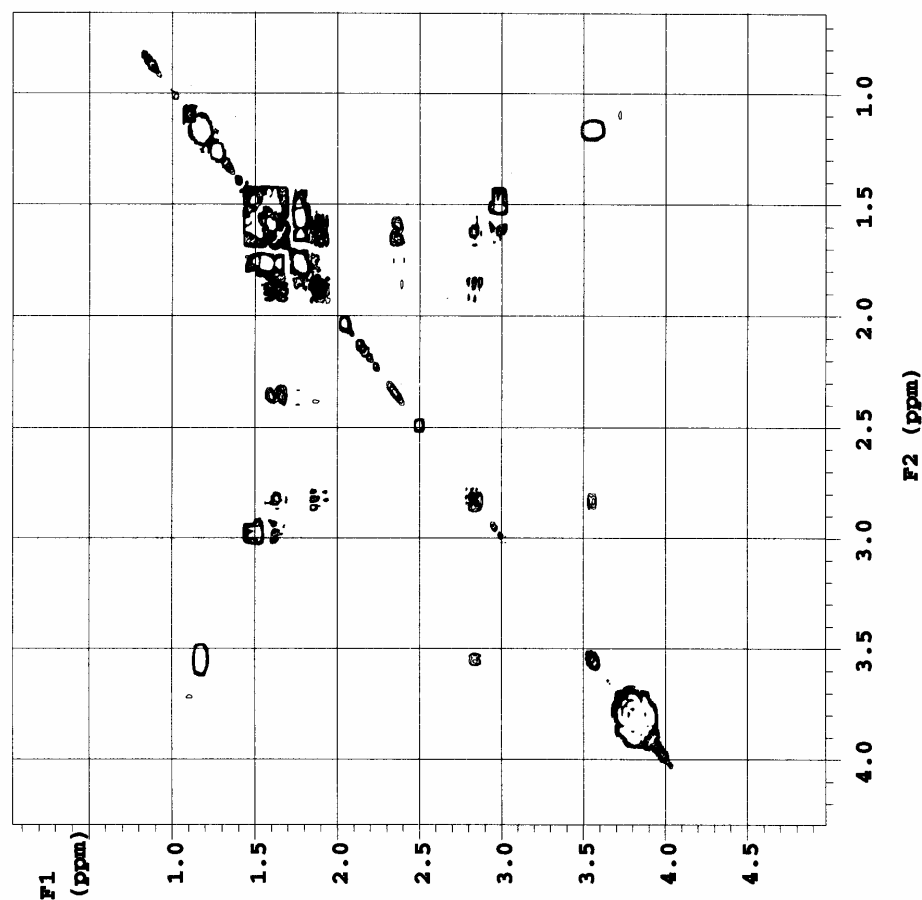
Usuari: san / Mostra: mmc472t22

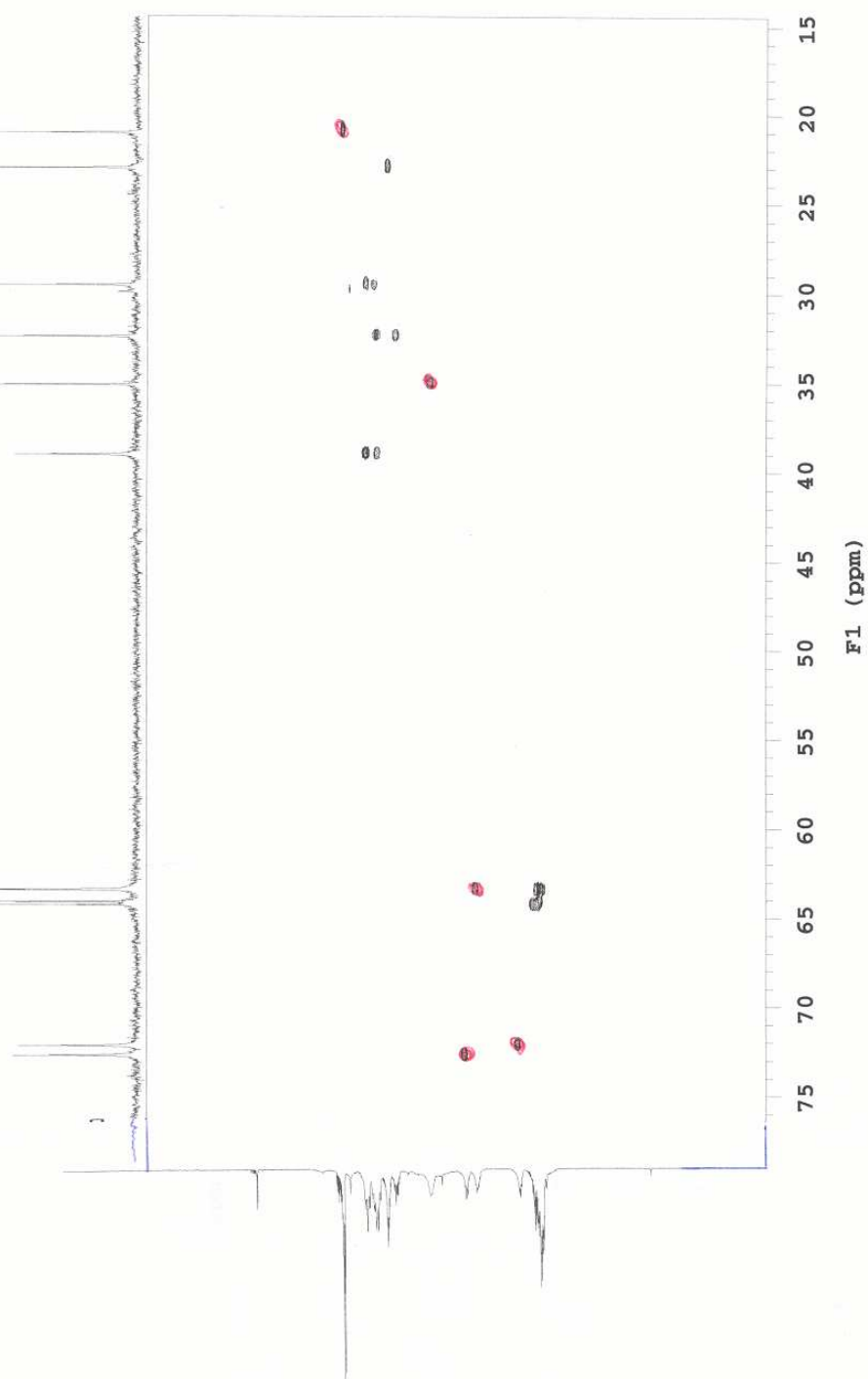
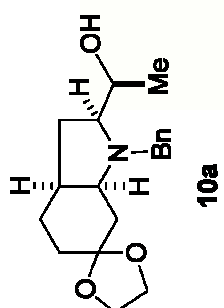
Nom: MARISA MENA CERVIGON

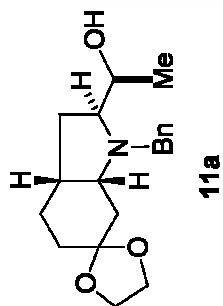
Data:30/09/05 / Sist automatic

exp22 gCOSY

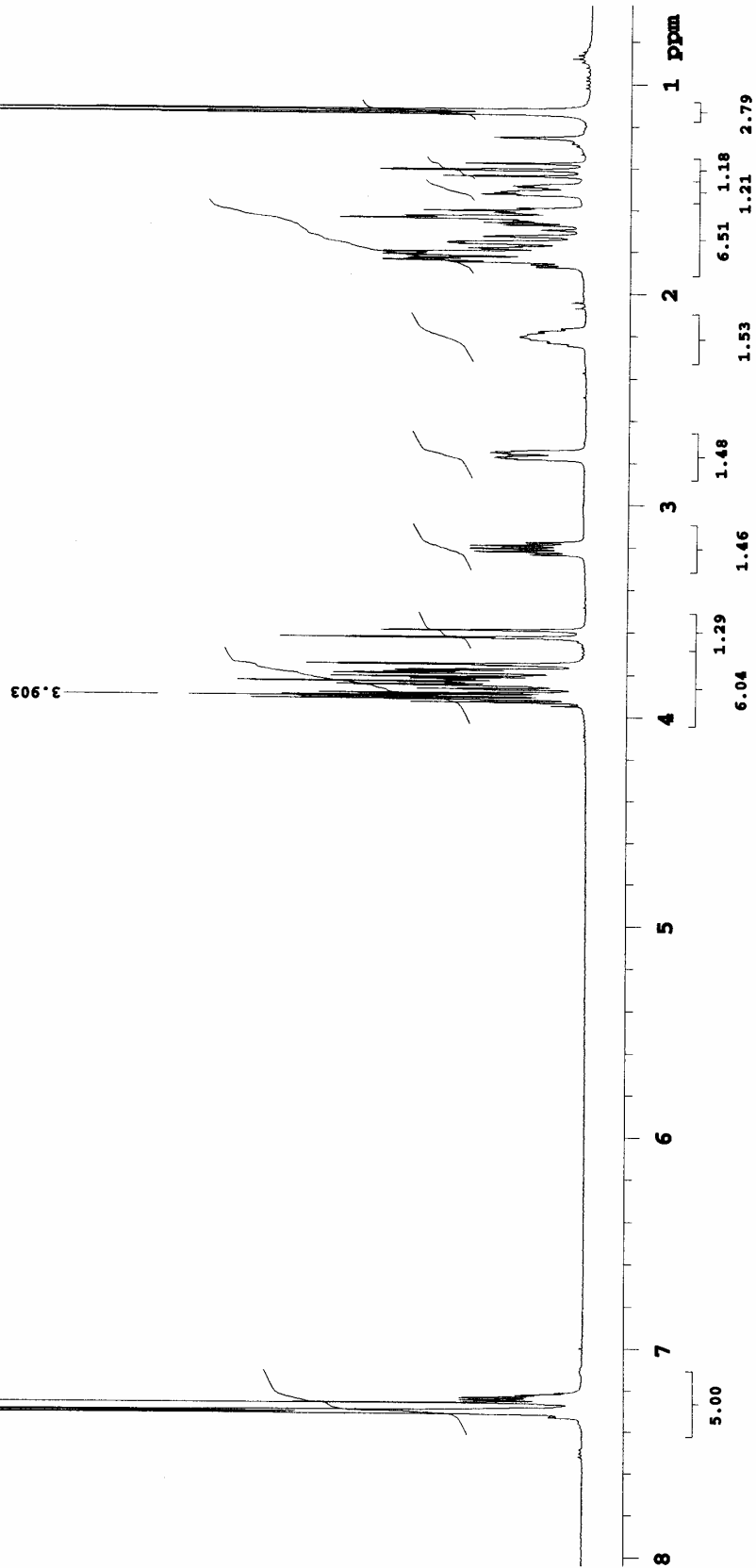
SAMPLE		FLAGS	
date	Sep 30 2005	hs	mn
solvent	cdcl3	sspul	n
sample	auto_Custom- hsglv1	1002	
Q_30Sep2005			
ACQUISITION			
sw	5211.0	gain	25.0
at	0.150	spin	0
np	1564	F2 PROCESSING	
fb	not used	sb	-0.075
ss	16	abs	not used
d1	1.000	fn	2048
nt	4	F1 PROCESSING	
2D ACQUISITION			
sw1	5211.0	abs1	-0.049
ni	256	proc1	lp
PRESATURATION			
satmode	n	DISPLAY	
satfrq	0	sp	254.8
satdly	0	vp	1461.9
satpwr	0	sp1	18.1
TRANSMITTER			
tn	H1	xf1	1971.3
sfreq	400.114	rfp	595.9
tof	-17.1	xf11	593.2
tpwr	58	xfp1	0
PLOT			
pw	12.200	vc	131.8
GRADIENTS			
gslw11	1002	sc	6.2
gt1	0.001000	sc2	131.8
gtab	0.000500	sc2	0
DECOUPLER			
dn	C13	th	493
dm	mn	ai	cdc
		av	8

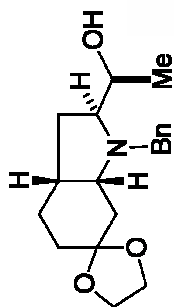






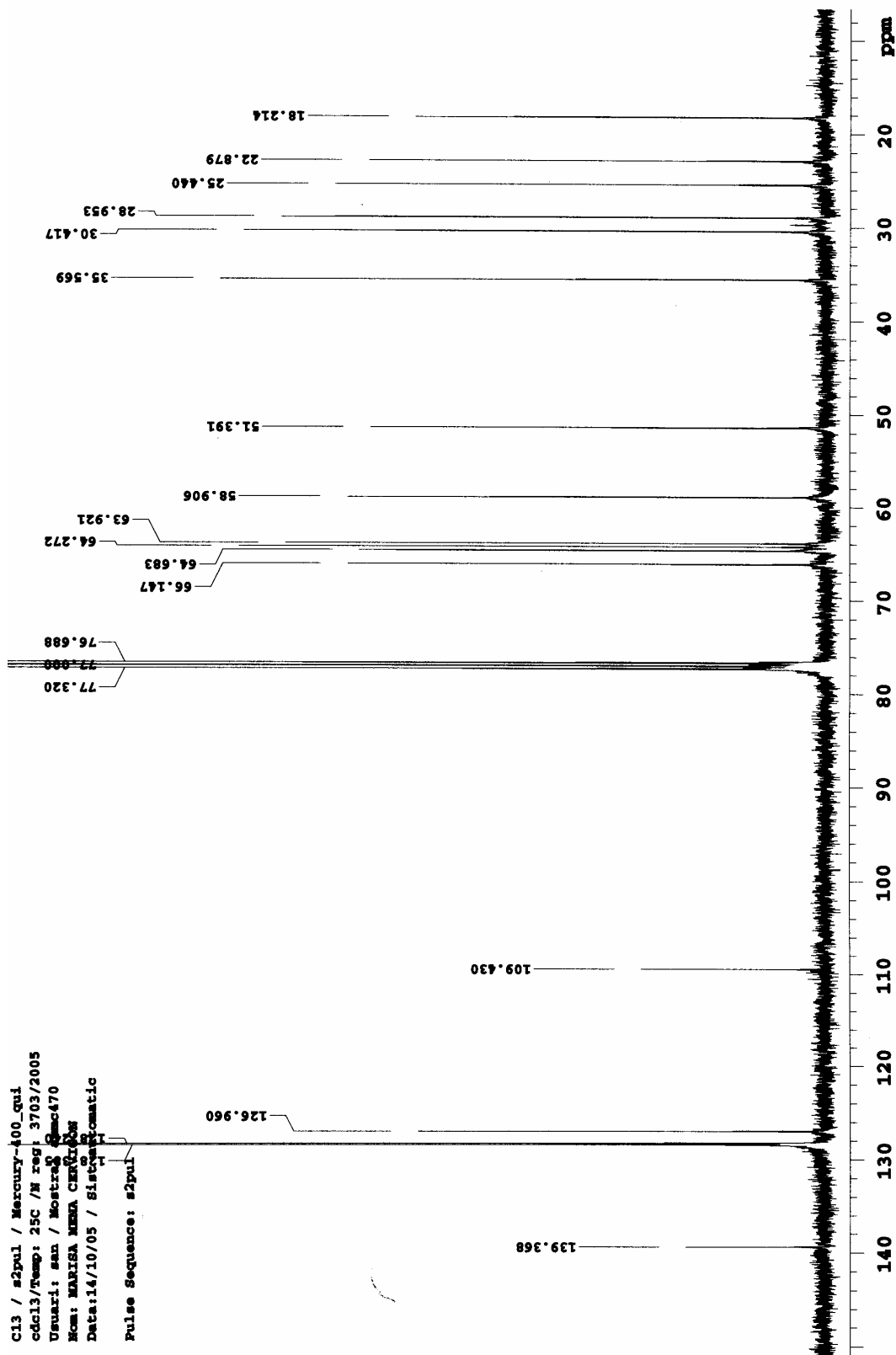
H1 / s2pul / Mercury-400_qui
 cdcl3/Temp: 25C / N reg: M40005-141005165205
 Usuari: san / Spectra: amac470
 Nom: MARISA / M40005-141005165205
 Data: 14/10/05 / Sist automatic
 Pulse Sequence: s2pul

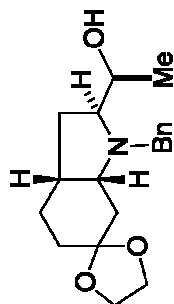




11a

C13 / s2pul / Mercury-400_qui
 cdcl3/Tmp: 25C / N reg: 3703/2005
 Usuari: san / Mostra: 200470
 Nom: MARIA NINA CERRON
 Data:14/10/05 / Sist:Automatic
 Pulse Sequence: s2pul





11a

H1 / gCOSY / Mercury-400.qui
cdcl3/Temp: 25C /N reg: M40005-141005165

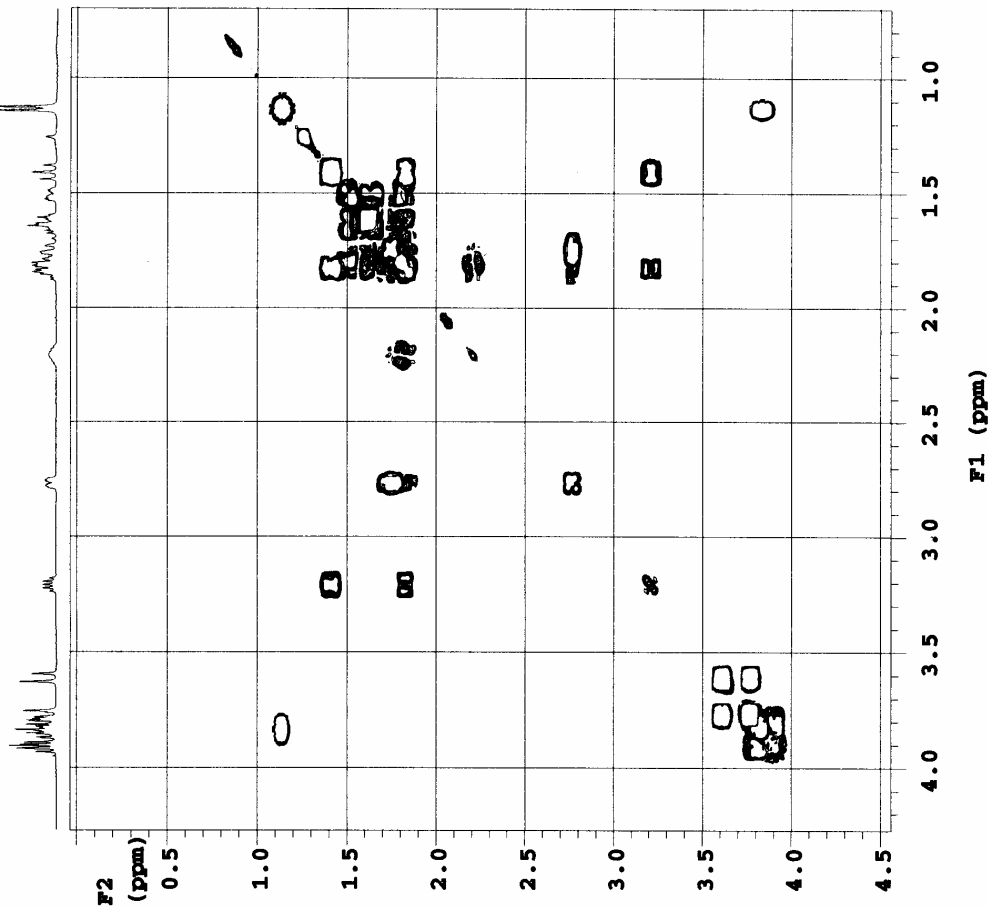
205

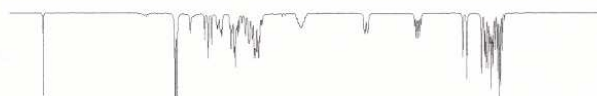
Usuari: san / Mostra: amac470
Nom: MARISA MENA CERVIGON

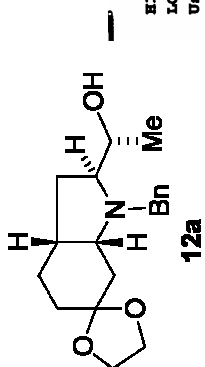
Data:14/10/05 / Sist automatic

exp5 gCOSY

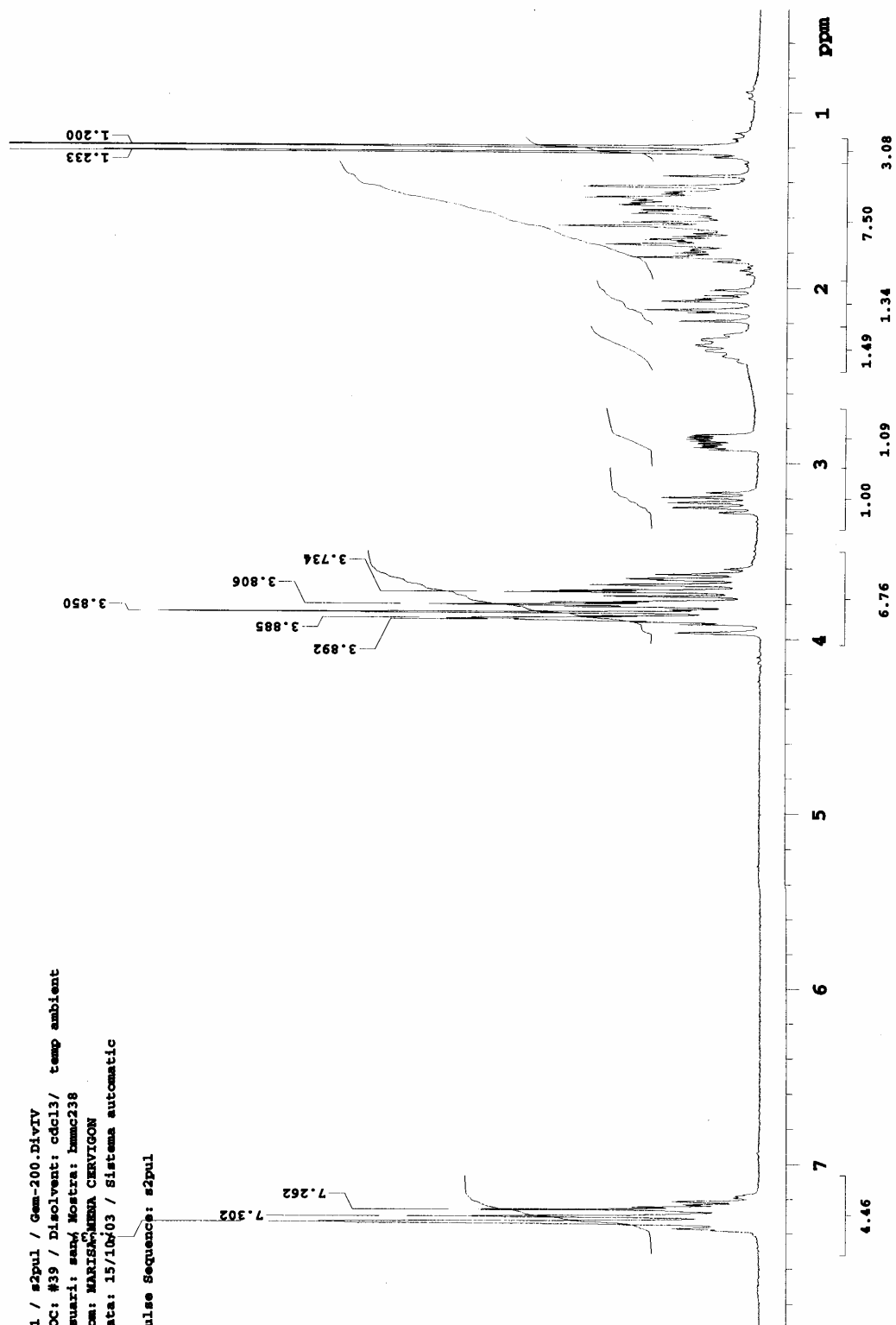
SAMPLE		FLAGS	
date	Oct 14 2005	hs	nm
solvent	cdcl3	sspul	n
sample	auto_Custom-haglv1	1002	
Q_14Oct2005-17:48:- SPECIAL			
15 temp 25.0			
ACQUISITION			
sw	3906.2	gain	28
at	0.150	F2 PROCESSING	0
tp	1172	sb	-0.075
fb	not used	sbs	not used
ss	16	fn	2048
d1	1.000	F1 PROCESSING	
nt	4	sb1	-0.066
2D ACQUISITION			
sw1	3906.2	proc1	lp
ni	256	fn1	2048
PRESATURATION DISPLAY			
satmode	n	sp	-13.6
satfrq	0	wp	1836.7
satlty	0	sp1	280.5
satpwr	0	wp1	1428.1
TRANSMITTER			
tn	H1	rfl	483.2
sfreq	400.113	rfl1	483.2
tof	-553.6	rflp1	0
tpwr	58	PLOT	
pw	12.200	wc	131.8
GRADIENTS			
gzlv11	1002	sc	6.2
gt1	0.001000	sc2	131.8
gstab	0.000500	vs	361
DECOUPLER			
dn	C13	ai	cdc
dm	nm	av	

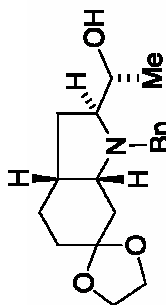






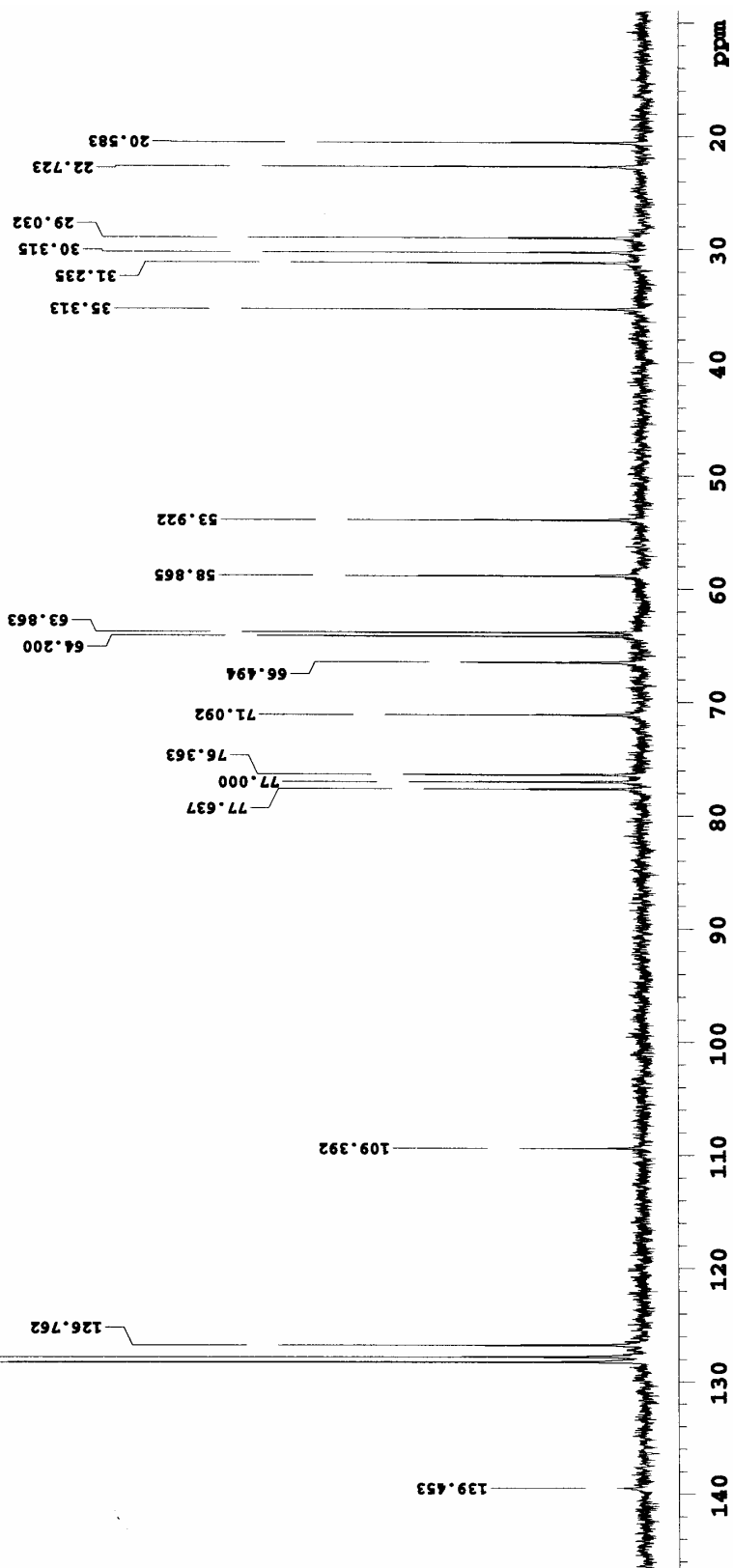
H1 / s2pul / Gem-200.DIVIV
 LOC: #39 / Disolvent: cdcl3/ temp ambient
 Usuari: sanja Mostra: hmc238
 Nom: MARIANA CERVIGON
 Data: 15/10/03 / Sistema automatic
 Pulse Sequence: s2pul

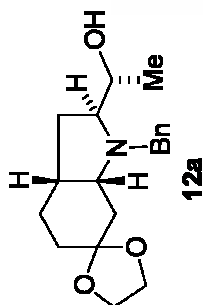




12a

C13 / s2pul / vsm-400.000.MAV14
 LOC: #20 / Disolvent: cdcl3/ temp ambient
 Usuari: san/ Mostres: 18_mmc238
 Nom: SANDRA DIAZ FINE
 Data: 16/10/03 / Sig: ca automatic
 Pulse Sequence: s2pul

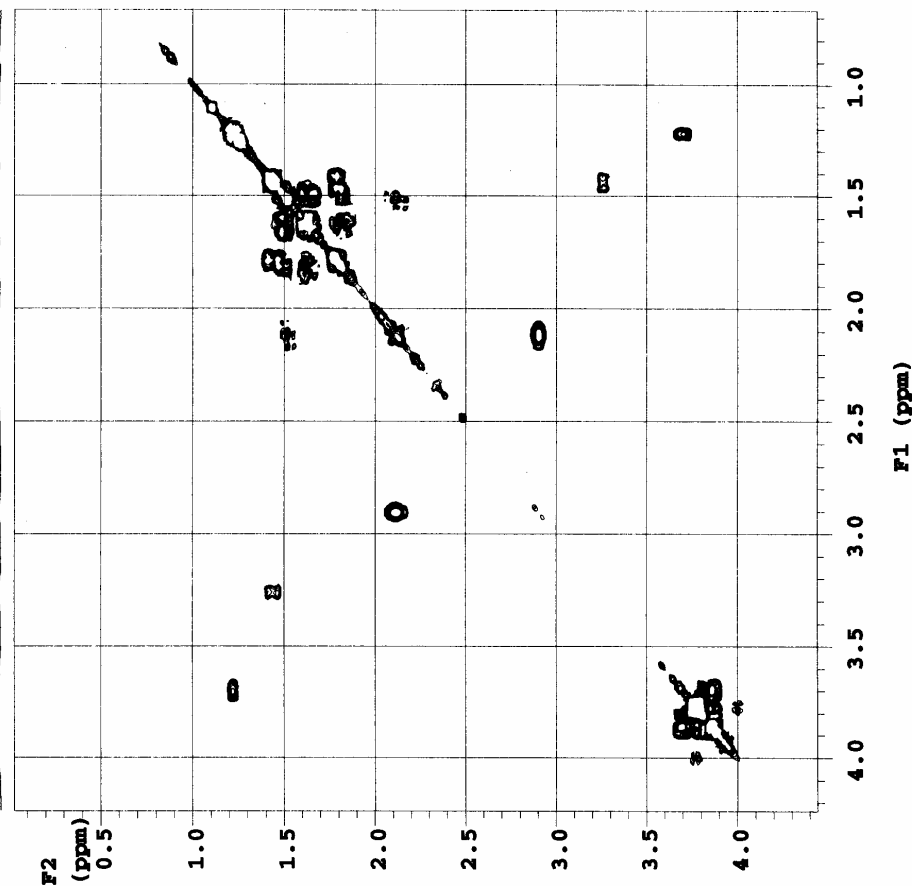


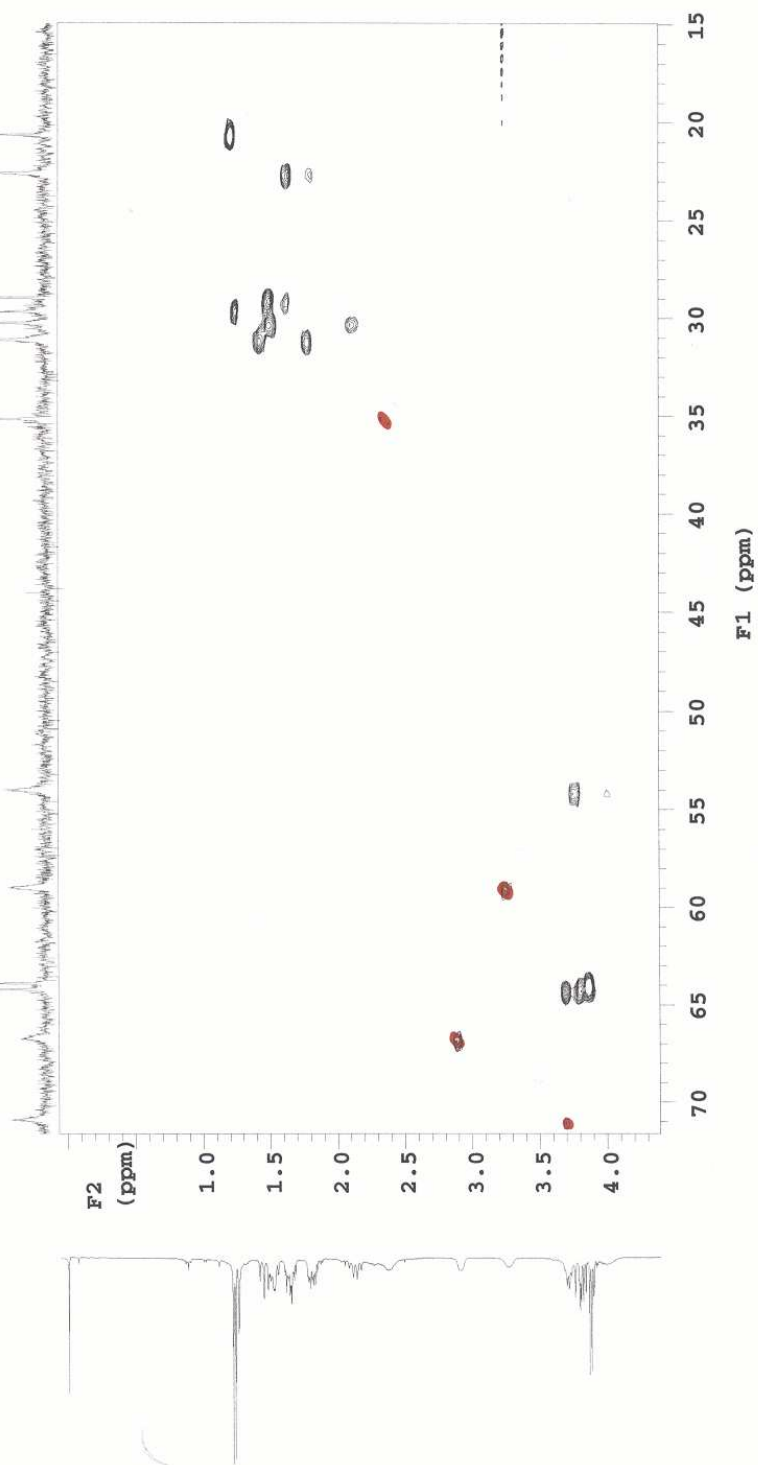
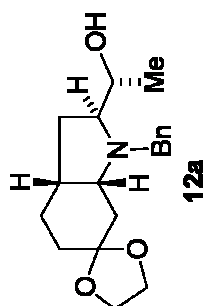


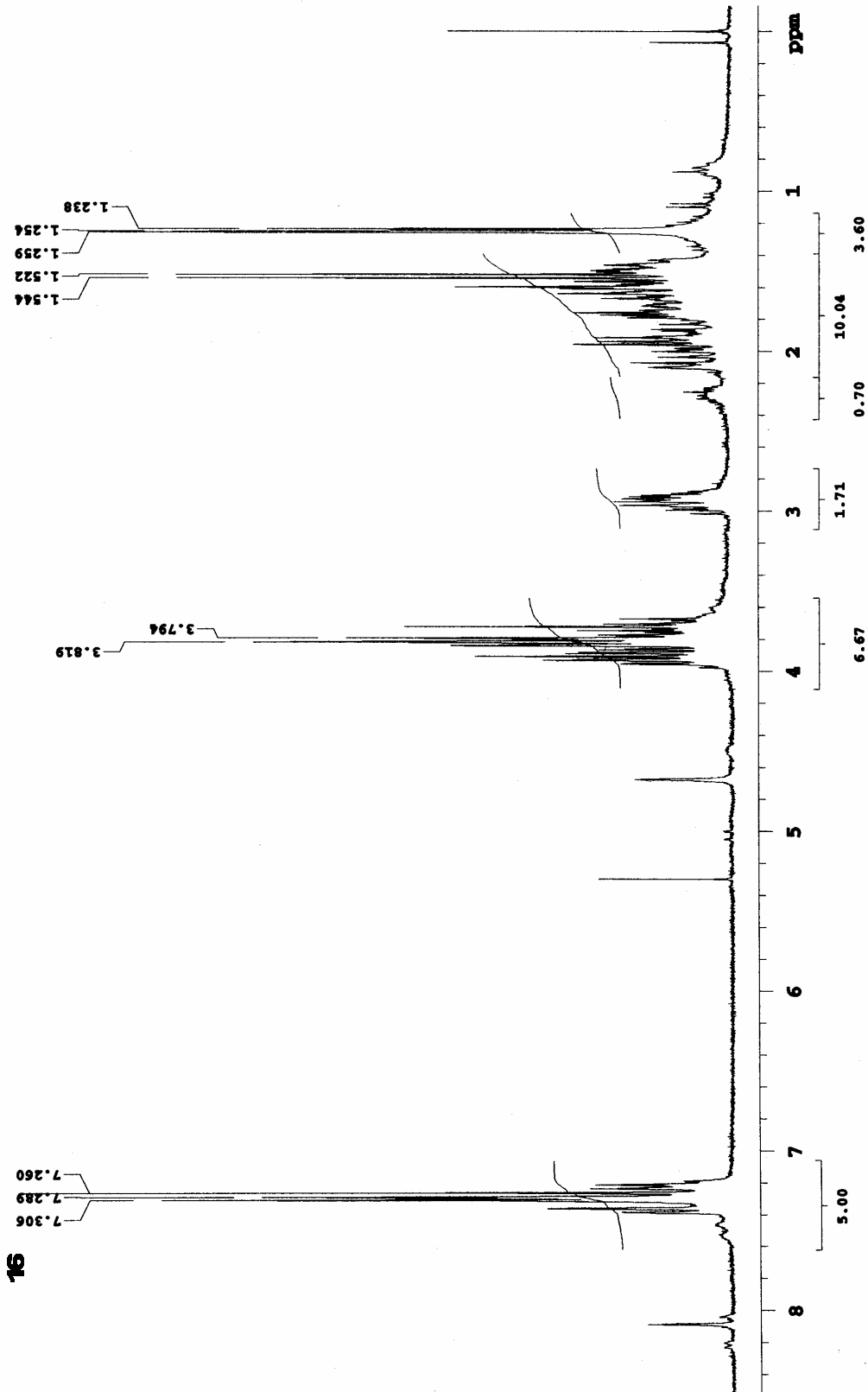
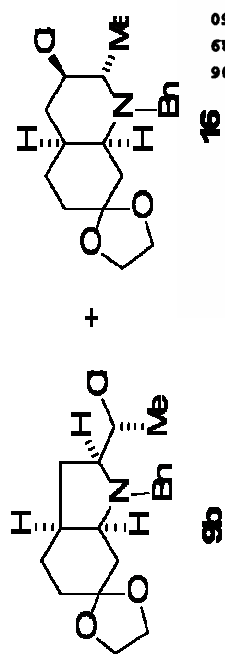
H1 / gCOSY / Mercury-400.qui
 cdc13/Temp: 25C / N reg: M40005-211005163
 841
 Usuari: san / Mostra: benzoh
 Nom: MARIA MENA CERVIGION
 Data:21/10/05 / sist automatic

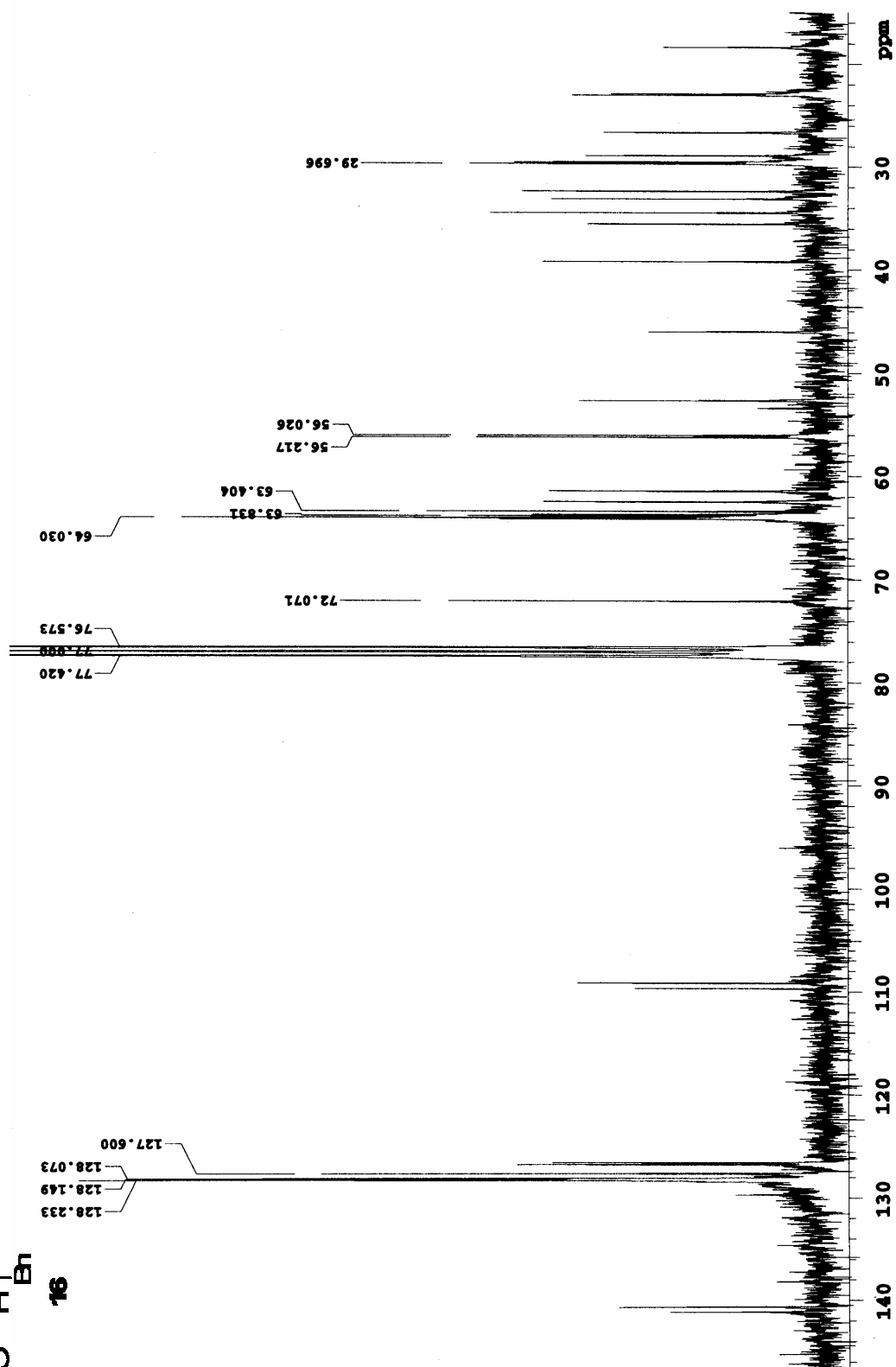
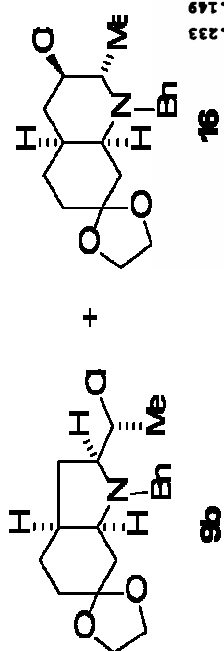
exp6 gCOSY

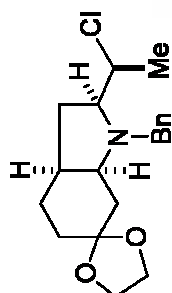
SAMPLE		FLAGS	
date	Oct 21 2005	hs	nn
solvent	cdc13	aspl	n
sample auto	Custom-hs	gvl	1002
Q_21Oct2005-18:41:-	SPECIAL		
55	temp	25.0	
ACQUISITION			
sw	3937.0	gain	32
at	0.150	F2 PROCESSING	0
np	1182	sb	-0.075
fb	not used	sbs	not used
ss	16	fn	2048
dl	1.000	F1 PROCESSING	
nt	8	sb1	-0.065
2D ACQUISITION			
sw1	3937.0	procl	lp
nl	256	fn1	2048
PRESATURATION			
satmode	n	sp	13.2
satfrq	0	wp	1762.6
satdly	0	sp1	267.2
satpr	0	wp1	1427.8
TRANSMITTER			
tn	H1	rfd	0
sfrq	400.113	rfl1	487.1
tof	-538.3	rfd1	0
tpwz	58	PLOT	
pw	12.200	wc	131.8
GRADIENTS			
gvlv11	1002	sc	6.2
gt1	0.001000	sc2	0
gstab	0.000500	vs	369
DECOUPLER			
dn	C13	al	cdc
dm	nn	av	



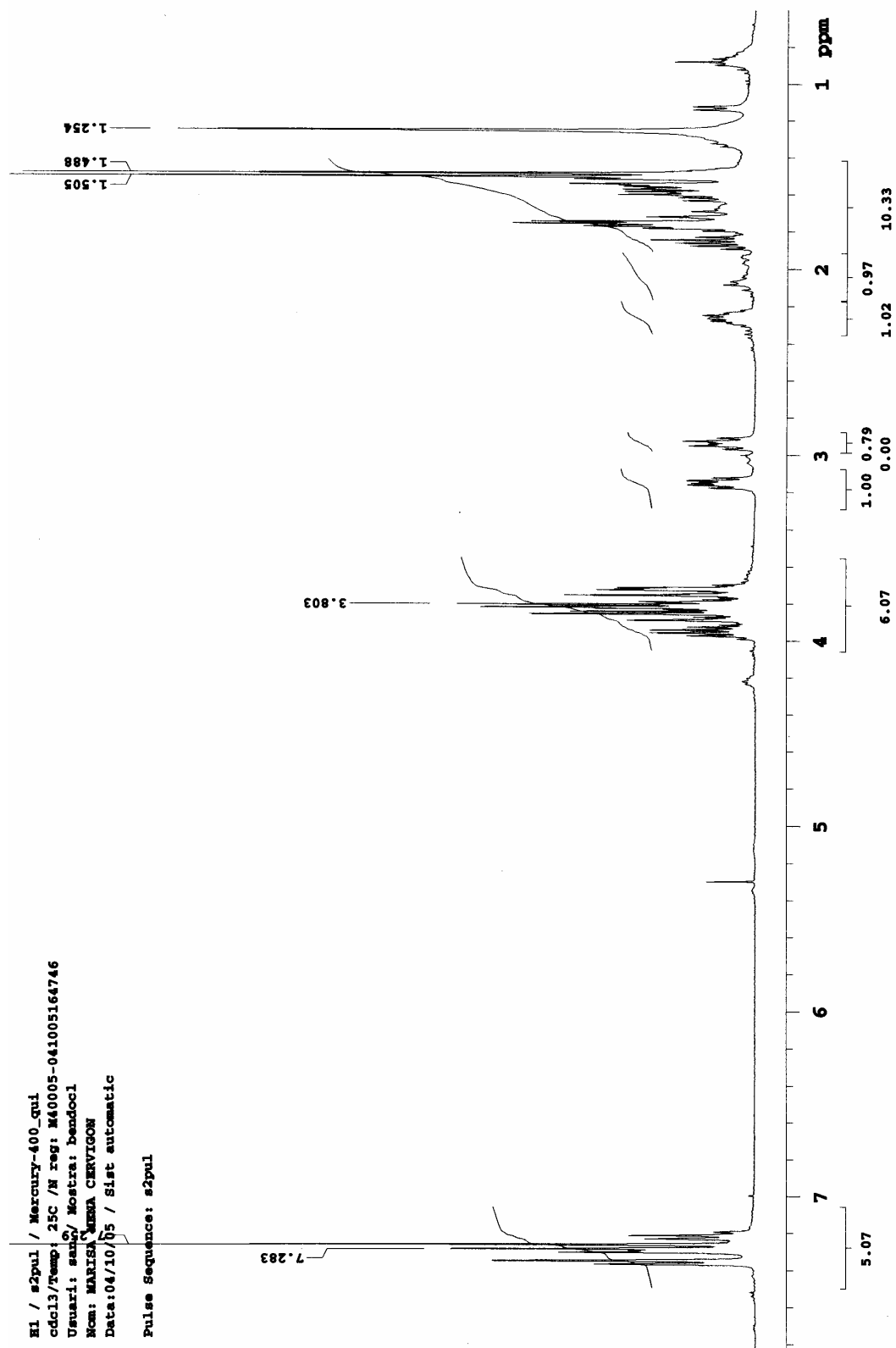


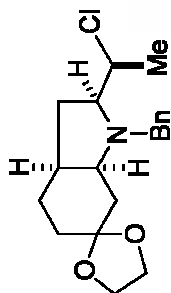




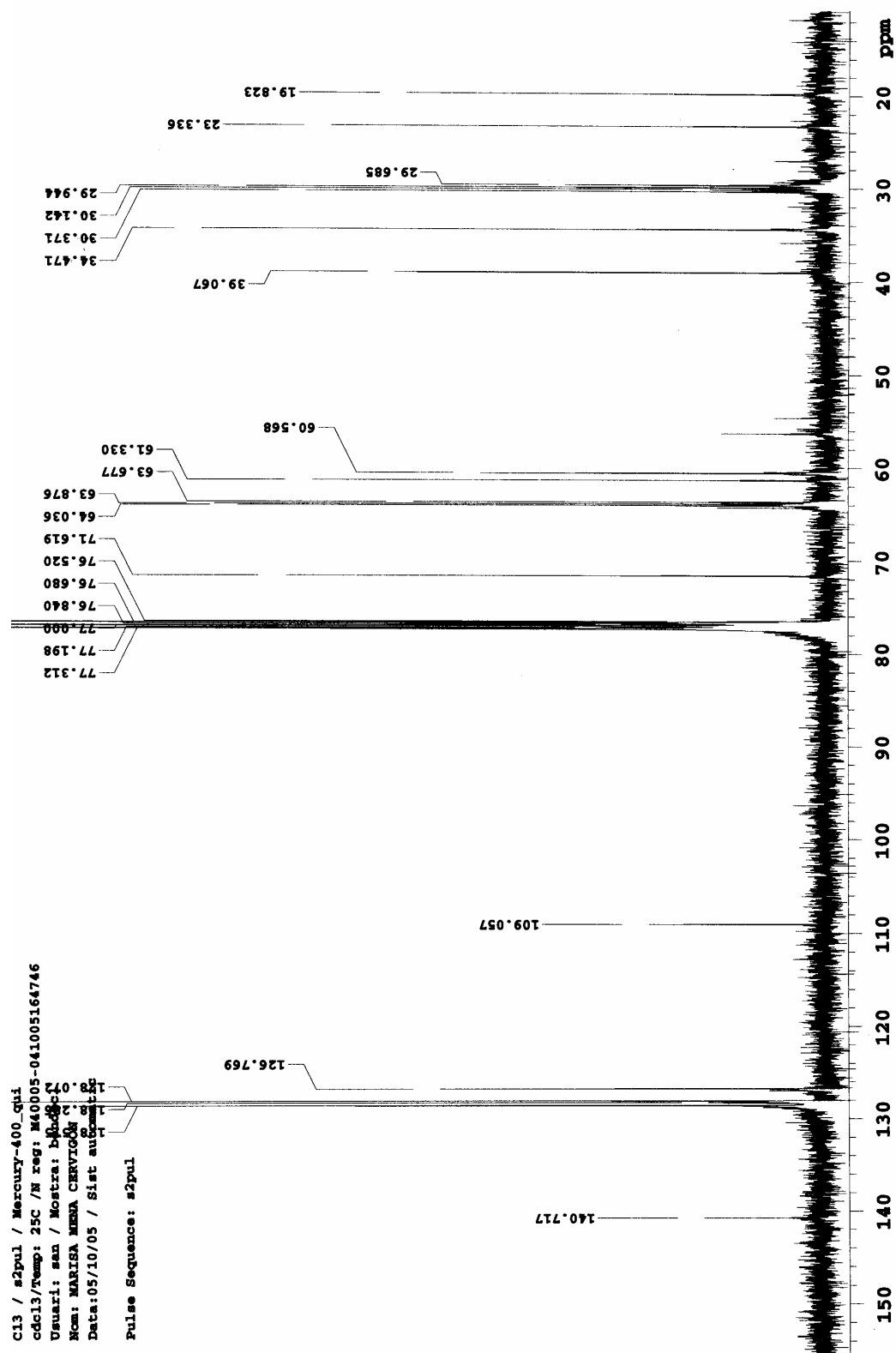


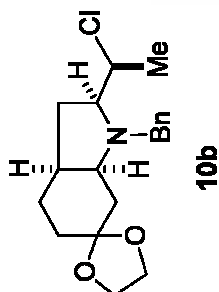
H1 / s2pul / Mercury-400_qui
 cdcl3/Temp: 25C /N reg: M40005-041005164746
 Usuari: sala / Mostra: bendocl
 Nom: MARISA / JENIA CERVIGON
 Data:04/10/05 / Sist automatic
 Pulse Sequence: s2pul





C13 / s2pul / Mercury-400.qhi
 cdcl3/Tmp: 25C / N reg: M40005-041005164746
 Usuari: san / Mostra: b00001
 Nom: MARISA MENA CERVIGÓN
 Data: 05/10/05 / Sist aut: b00001
 Pulse Sequence: s2pul





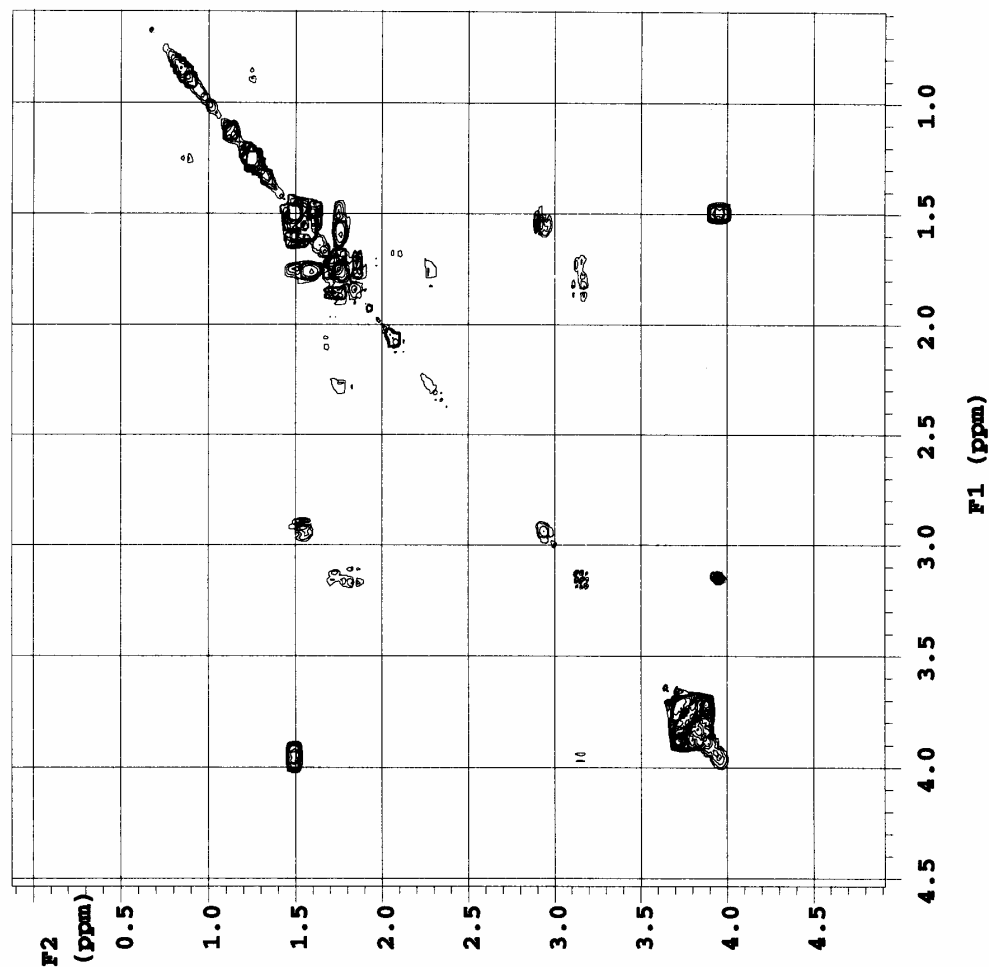
cdcl3/Temp: 25C /M reg: M40005-041005164

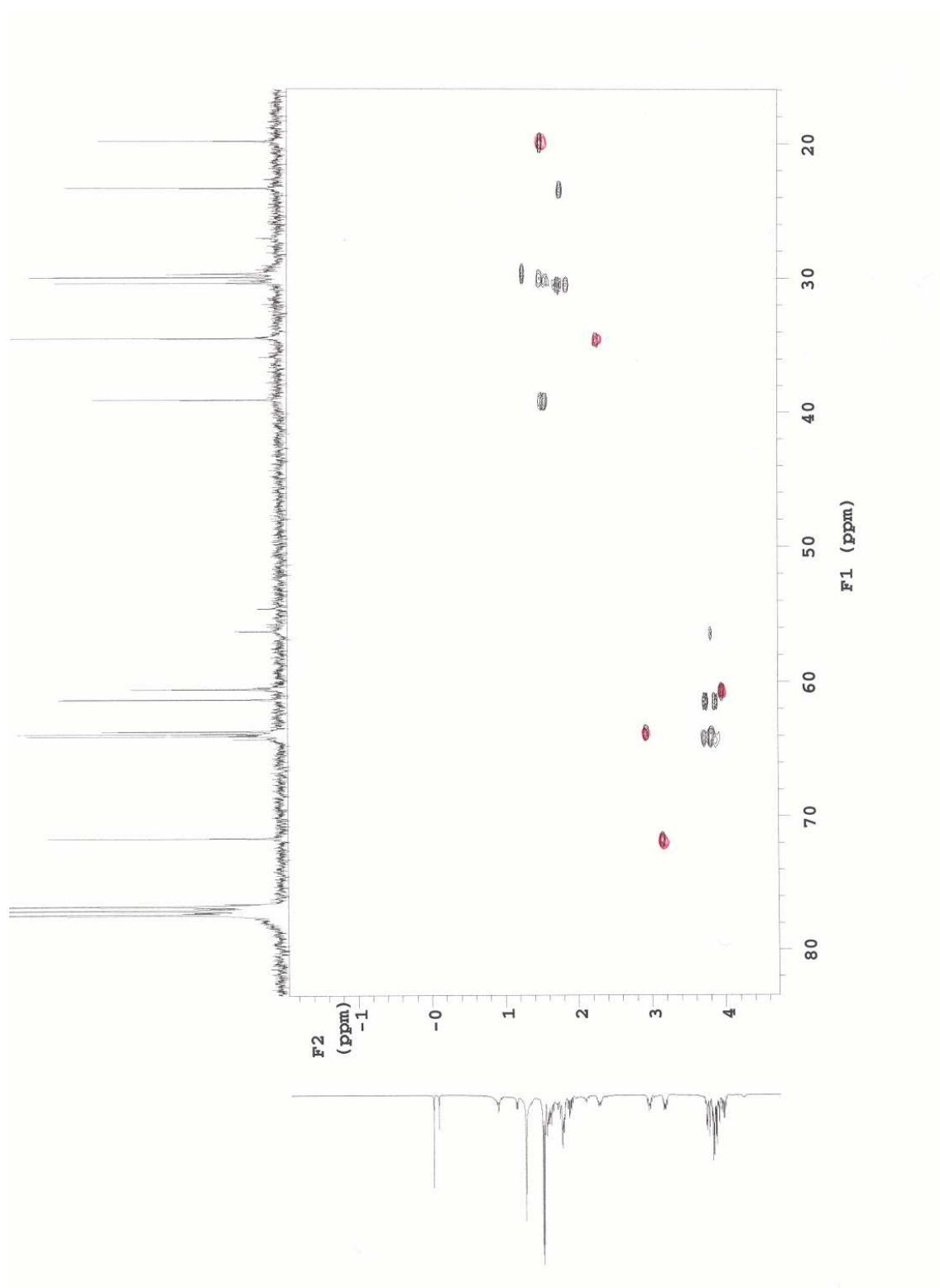
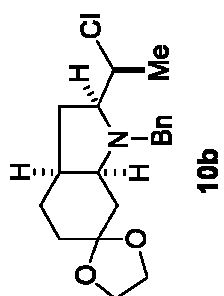
746

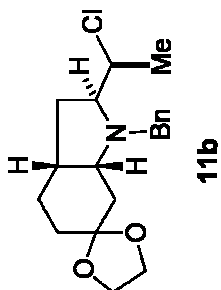
Usuari: san / Mostra: bendocl
Nom: MARISA NENA CERVIGNON
Data: 05/10/05 / Sist automatic

exp22 gCOSY

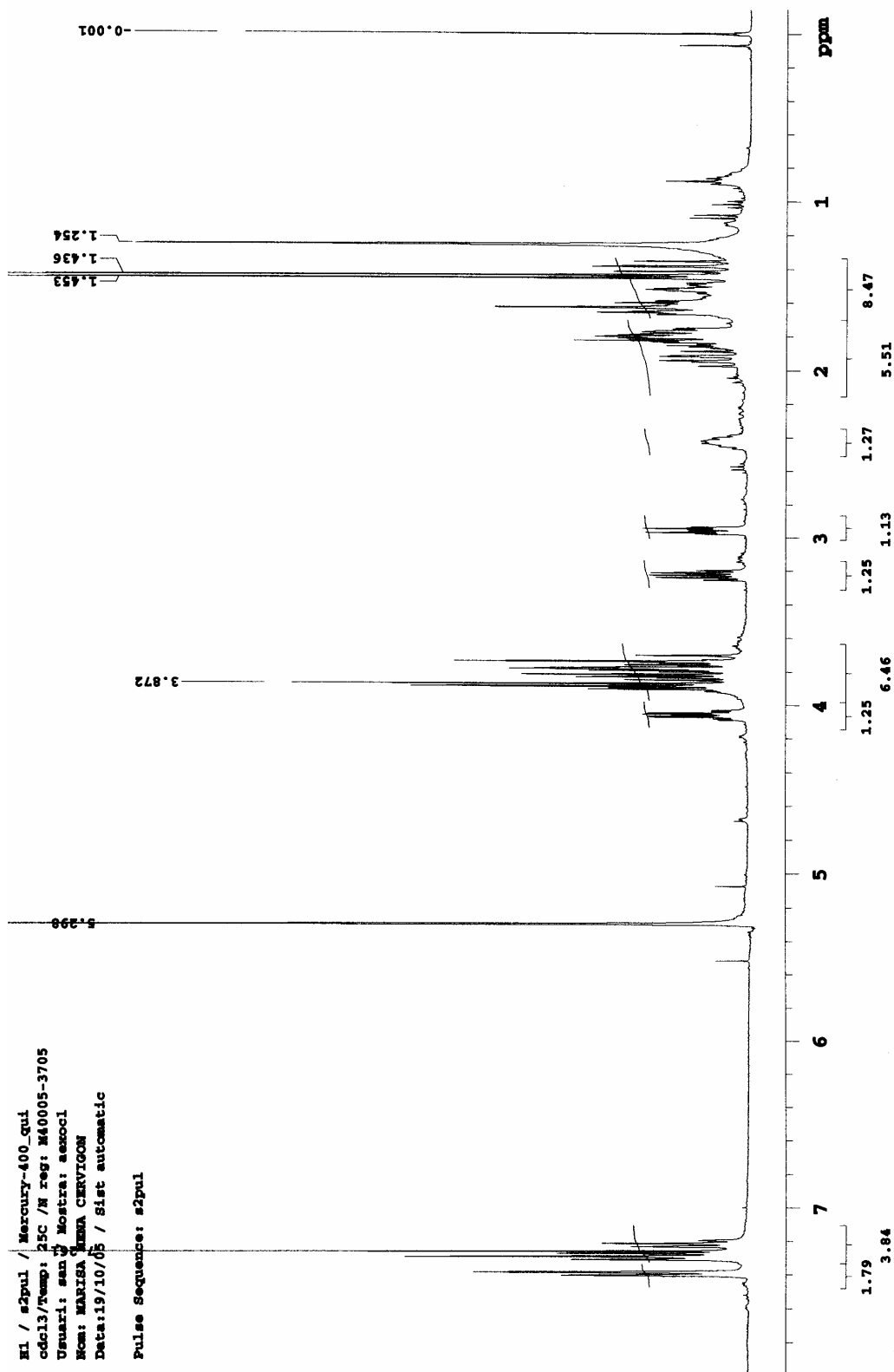
SAMPLE		FLAGS	
date	Oct 5 2005	hs	nn
solvent	cdcl3	sspul	n
sample	auto_Custom- hsglv1		1002
Q_04Oct2005 SPECIAL			
ACQUISITION	temp	25.0	
sw	6410.3	gain	32
at	0.150	spin	0
np	1924	F2 PROCESSING	
fb	not used	sb	-0.075
ss	16	sbs	not used
dl	1.000	fn	2048
nt	4	F1 PROCESSING	
2D ACQUISITION	sb1	-0.040	
sw1	6410.3	sbs1	not used
nl	256	procl	lp
PRESATURATION	fn1	2048	
satmode	n	DISPLAY	
satfrq	0	sp	-47.3
satdly	0	wp	2011.4
satpwr	0	sp1	234.6
TRANSMITTER	wp1		1579.1
tn	H1	rf1	799.3
sfrq	400.114	rfp	0
tof	381.9	rf11	799.3
tpwr	58	rfp1	0
PW	12.200	PLOT	
GRADIENTS	wc		131.8
gvlv11	1002	sc	6.2
gt1	0.001000	wc2	131.8
gstab	0.000500	sc2	0
DECOUPLER	vs		252
dn	C13	th	7
dm	nmn	ai	cdc
		av	

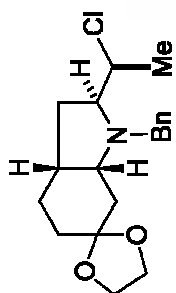






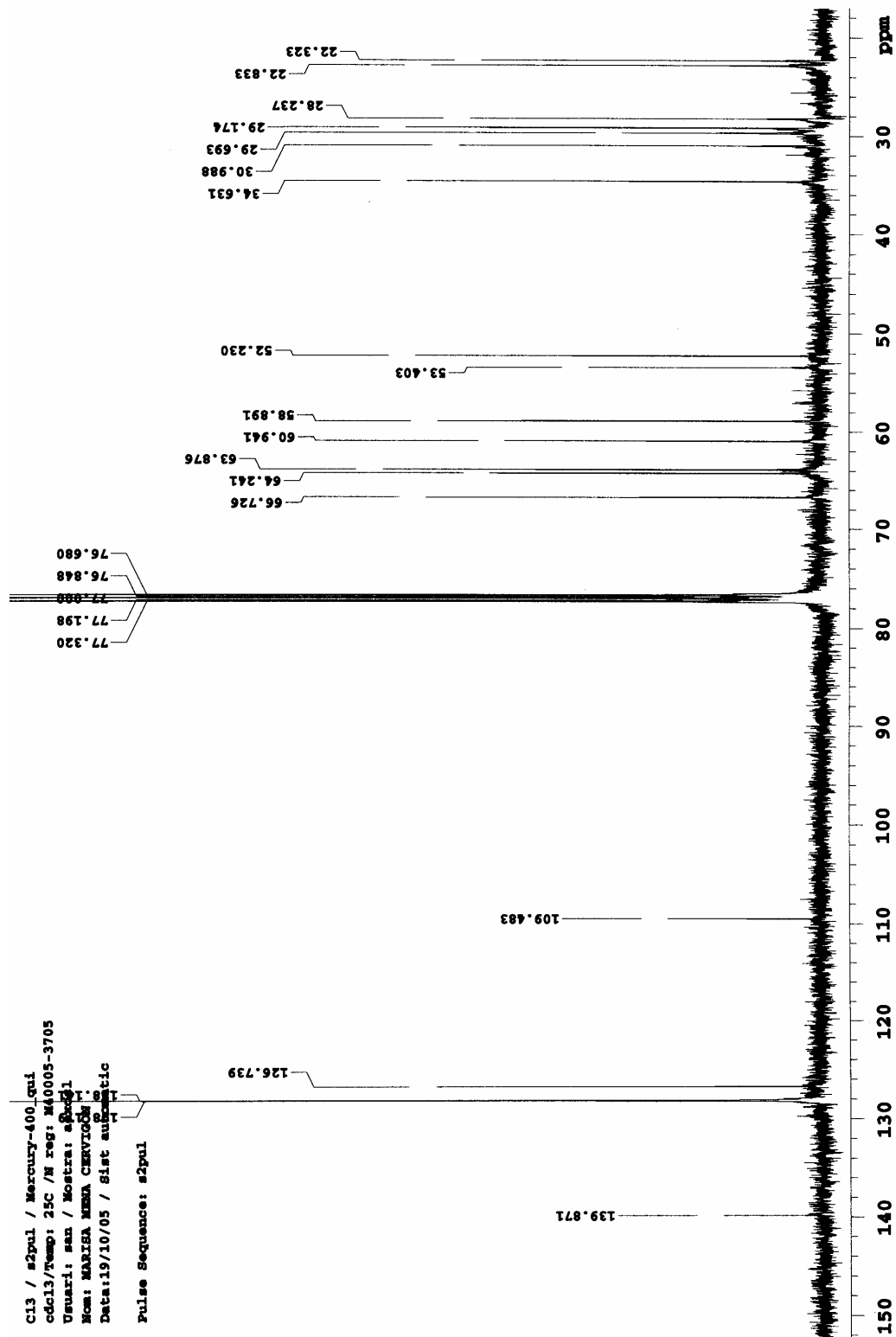
H1 / s2pul / Mercury-400_qui
 cdcl3/Temp: 25C /N reg: M40005-3705
 Usuari: san J Mostra: aescocl
 Nom: MARISA BENA CERVIGON
 Data:19/10/05 / Sist automatic
 Pulse Sequence: s2pul

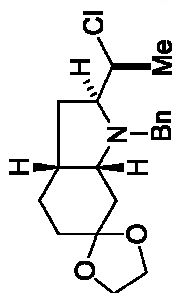




11b

C13 / s2pul / Mercury-400.qui
 cdcl3/Temp: 25C /M reg: M40005-3705
 Usuari: san / Mostra: s2pul
 Nom: MARISA MENA CERVIGÓN
 Data:19/10/05 / Sist aut:estic
 Pulse Sequence: s2pul



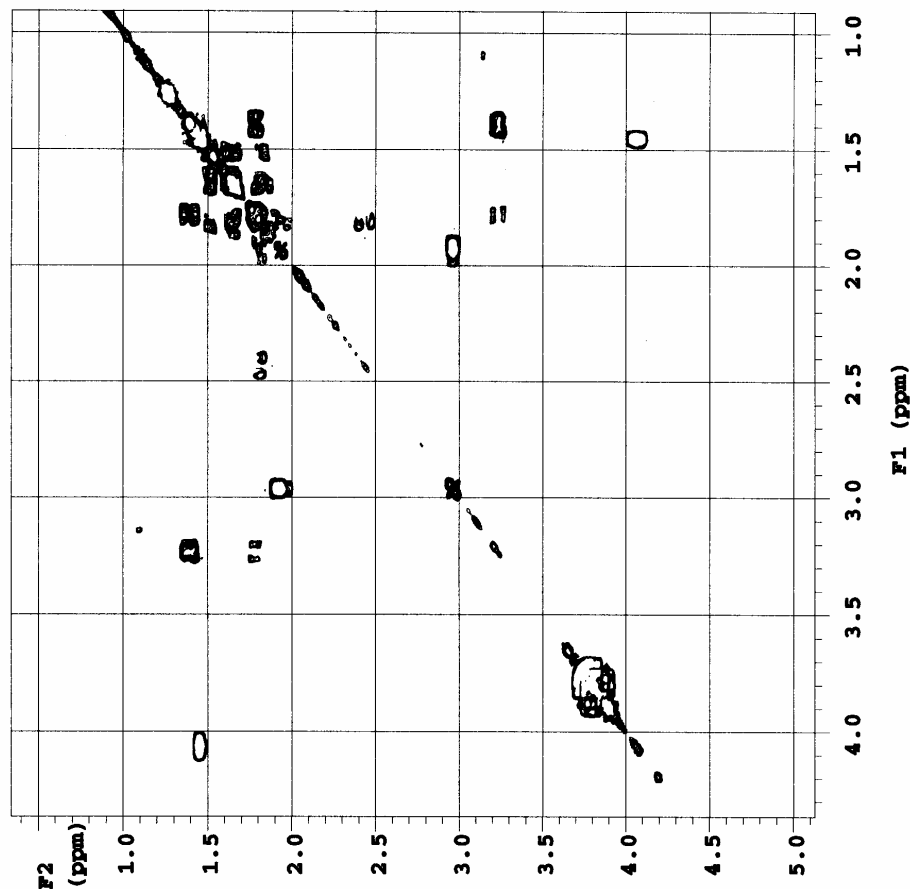


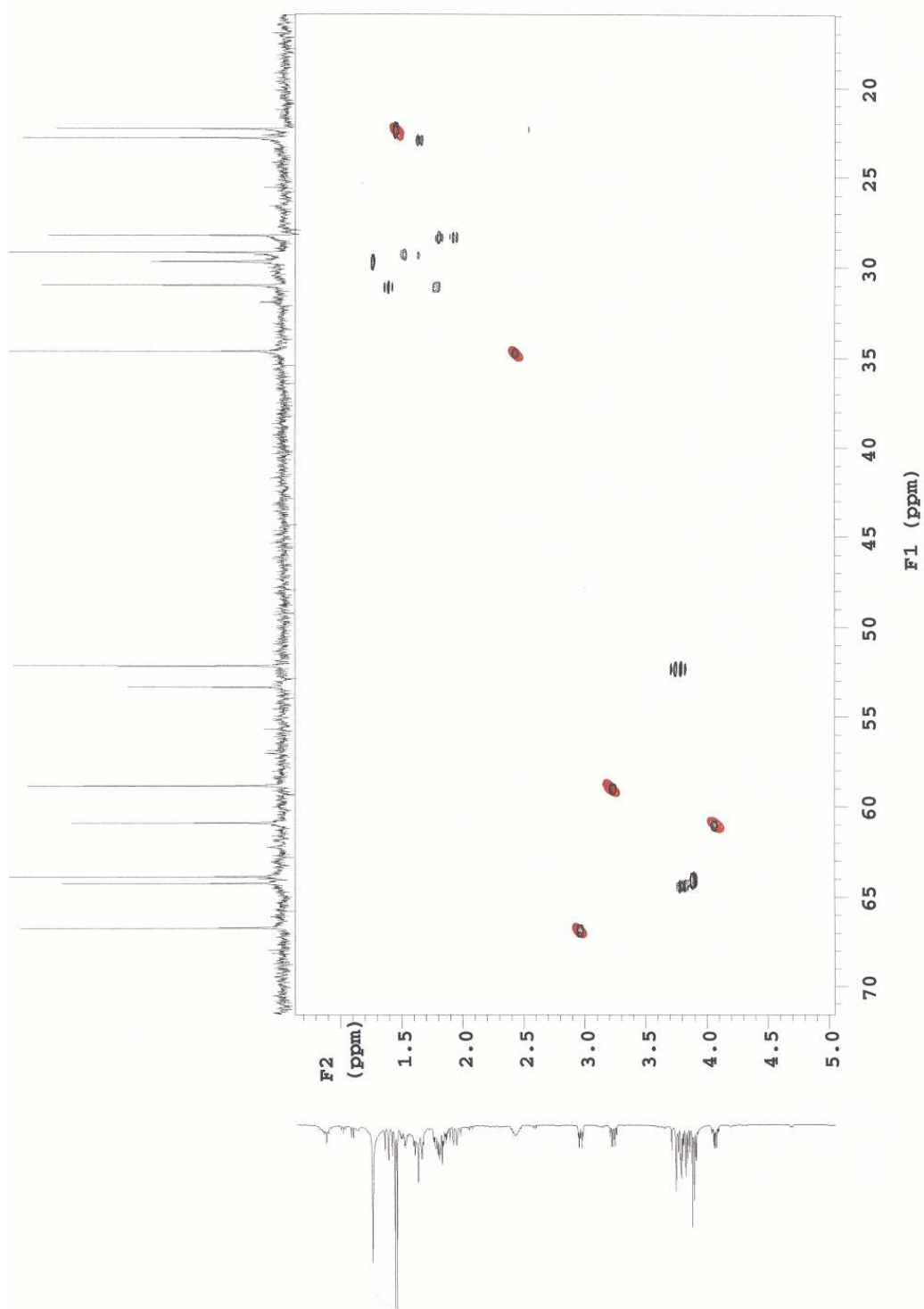
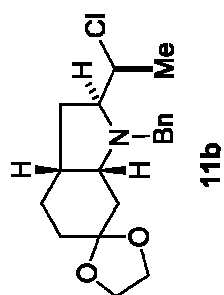
11b

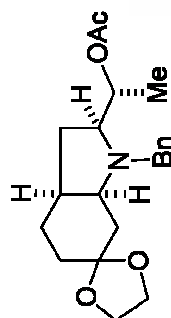
H1 / gCOSY / Mercury-400_qui
 cdcl3/Temp: 25C /M reg: M40005-3705
 Usuari: san / Mostra: aexocl
 Nom: MARISA MENA CERVIGON
 Data:19/10/05 / Sist automatic

exp4 gCOSY

SAMPLE		FLAGS	
date	Oct 19 2005	hs	nn
solvent	cdcl3	sepl	n
sample	auto Custom- hsglvi	1002	
Q_19oct2005	SPECIAL		
ACQUISITION temp 25.0			
sw	4273.5	gain	26
at	0.150	spin	0
np	1282	F2 PROCESSING	
fb	not used	ab	-0.075
ss	16	sbs	not used
d1	1.000	fn	2048
nt	4	F1 PROCESSING	
2D ACQUISITION sb1 -0.060			
sw1	4273.5	abs1	not used
ni	256	proc1	lp
PREPARATION fn1 2048			
satmode n DISPLAY			
satfrq	0	sp	137.7
satdly	0	vp	1913.3
satpr	0	sp1	363.3
TRANSMITTER wd1 1382.7			
tn	H1	rf1	514.0
sfrq	400.113	rfp	0
tof	-400.6	rf11	514.0
tpwr	58	rfp1	0
pw 12.200 PLOT			
GRADIENTS wc 131.8			
gvlv1	1002	sc	6.2
gt1	0.001000	vc2	131.8
gstab	0.000500	sc2	0
DECOUPLER vs 590			
dn	C13	th	7
dm	nm	ai	cdc
			av



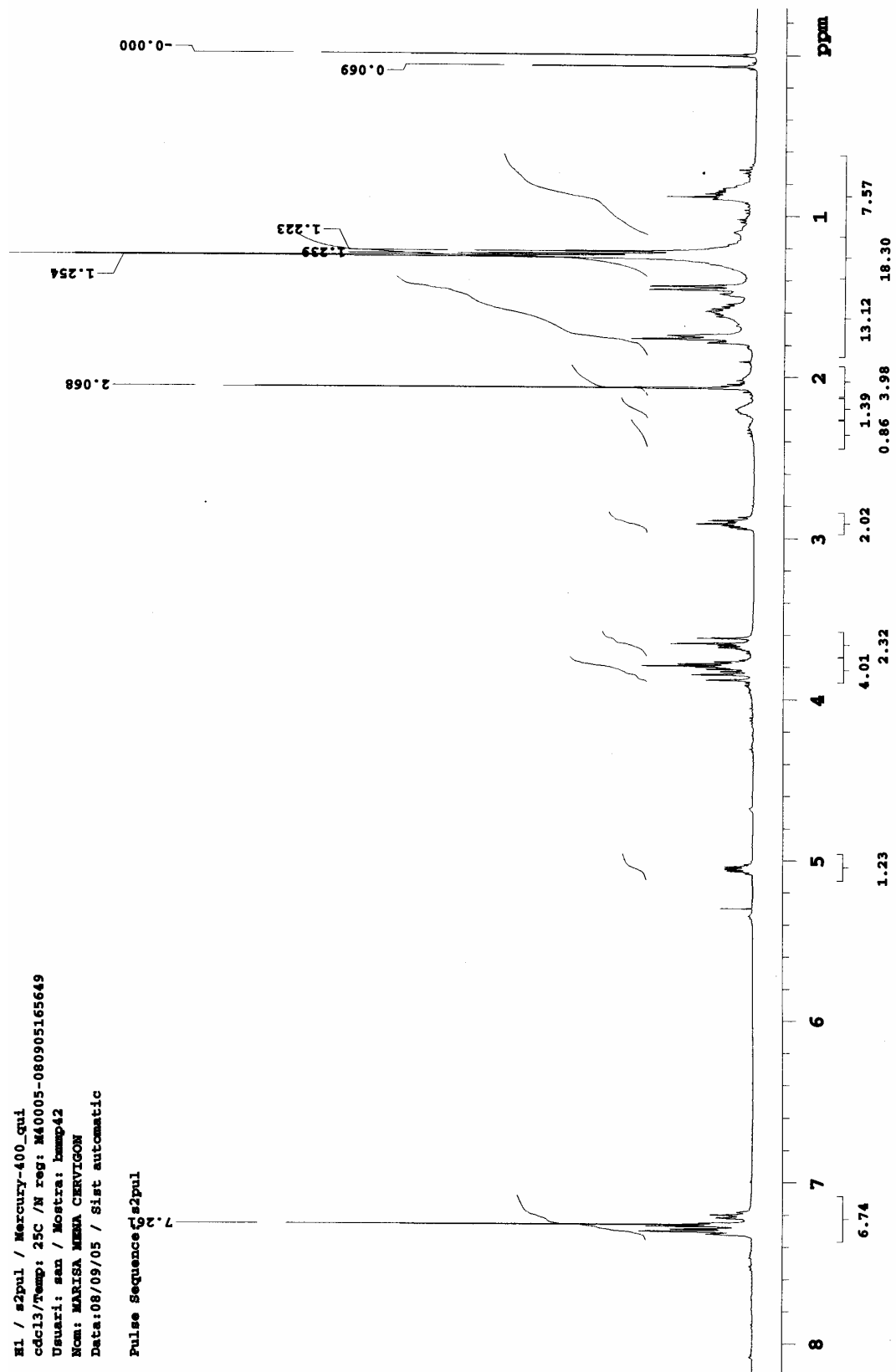


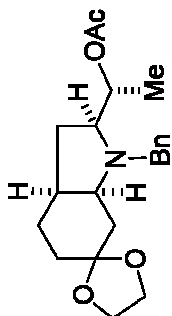


9c

H1 / s2pul / Mercury-400_qui
 cdc13/Temp: 25C / N reg: M40005-080905165649
 Usuari: san / Nostra: hump42
 Nom: MARISA MEMA CERVIGON
 Data:08/09/05 / Sist automatic

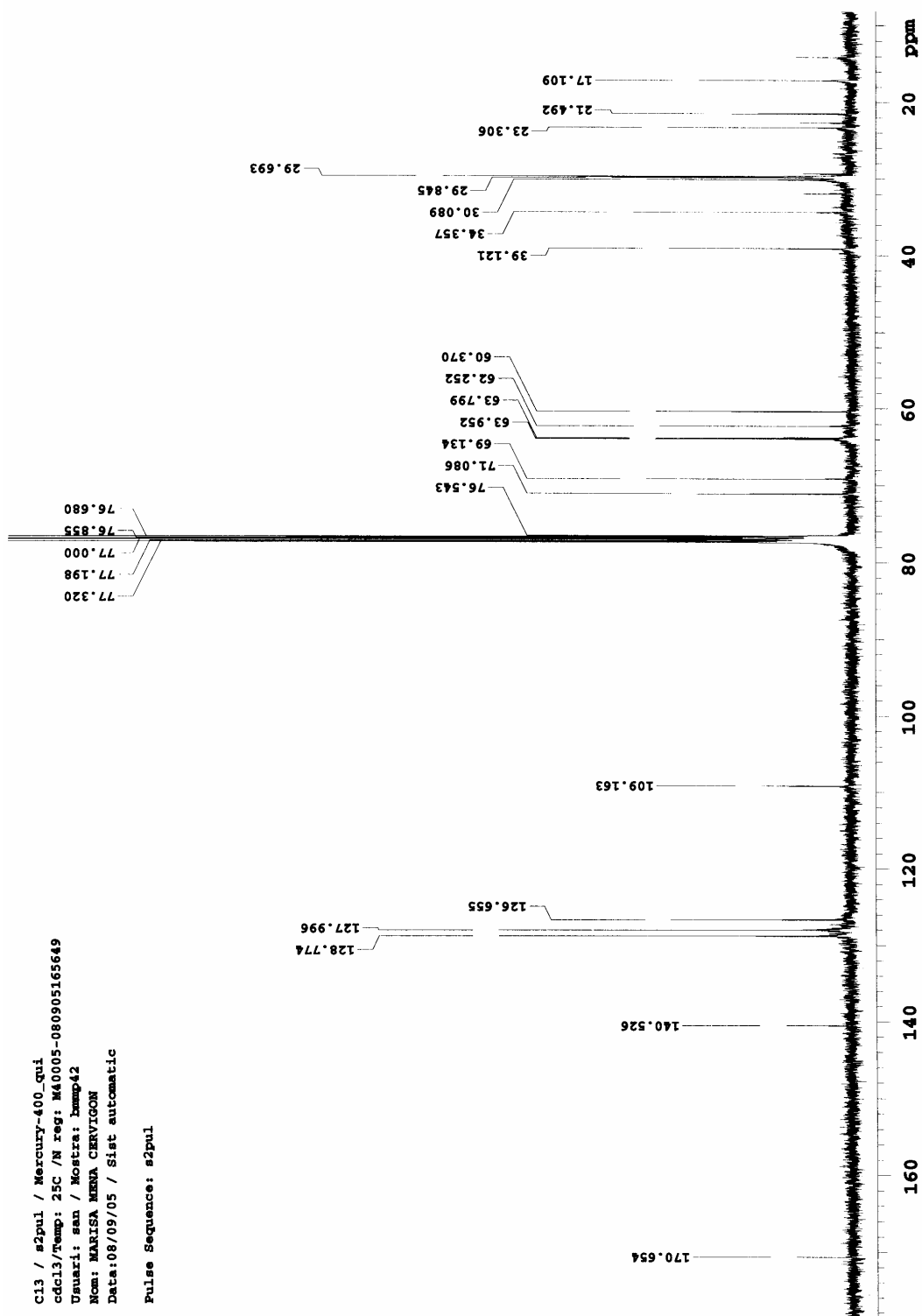
Pulse Sequence: c13s2pul

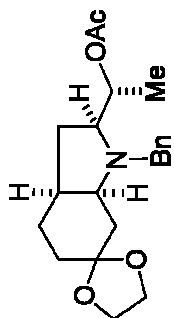




9c

C13 / s2pul / Mercury-400_qui
 cdcl3/Temp: 25C / N reg: M40005-080905165649
 Usuari: san / Mostra: bump42
 Nom: MARISA MENA CERVIGON
 Data:08/09/05 / Siat automatic
 Pulse Sequence: s2pul





9c

H1 / gCOSY / Mercury-400.qui
cdcl3/Tmp: 25C / N reg: M40005-290905163

622

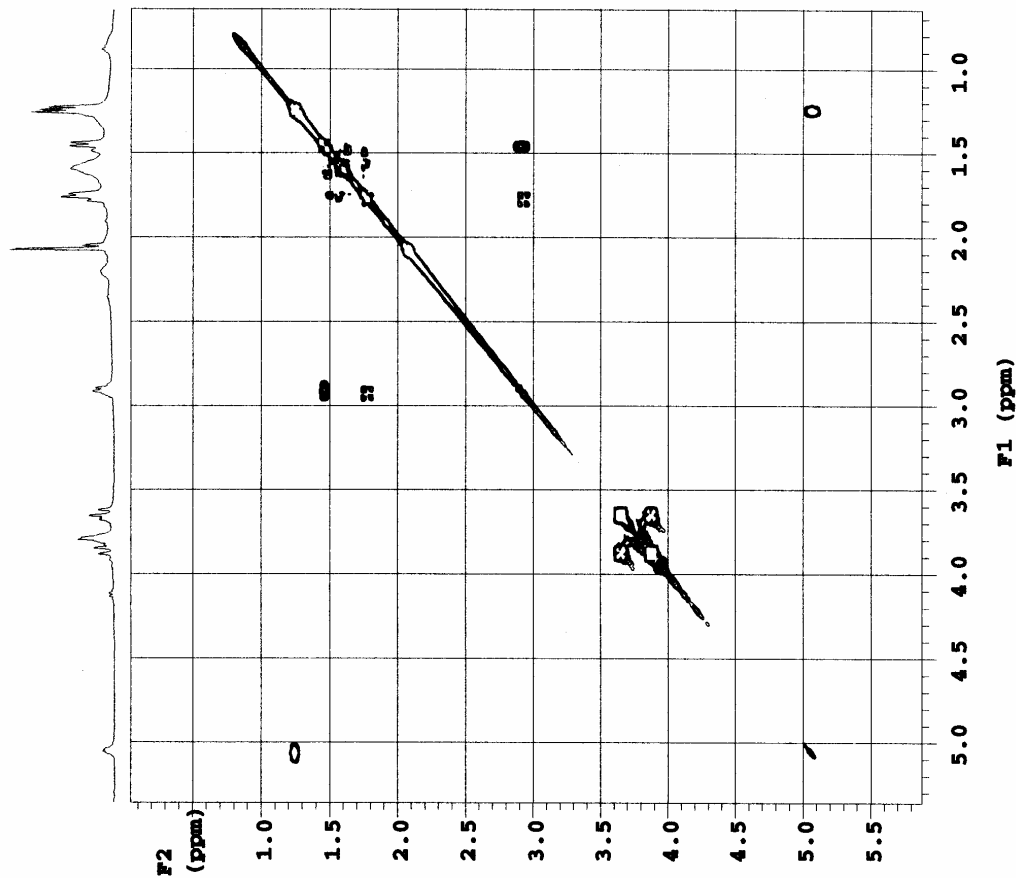
Usuari: sen / Mostra: hmp42

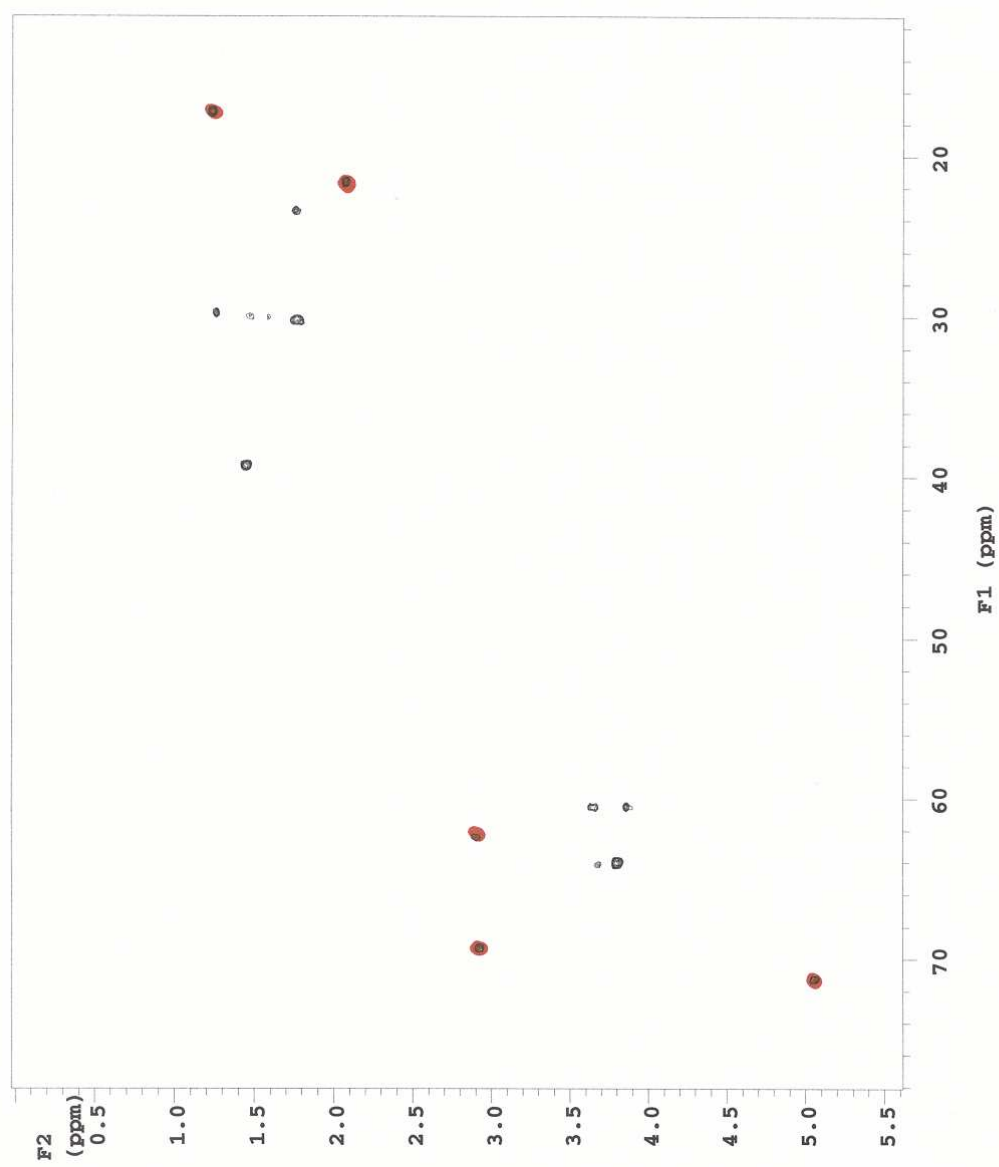
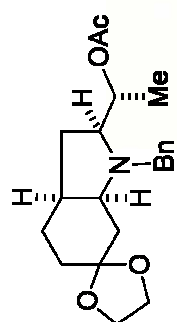
MCm: MARISA MEMA CERVIQOM

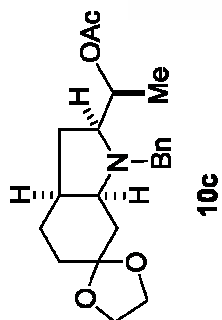
Data:29/09/05 / Sist automatic

exp6 gCOSY

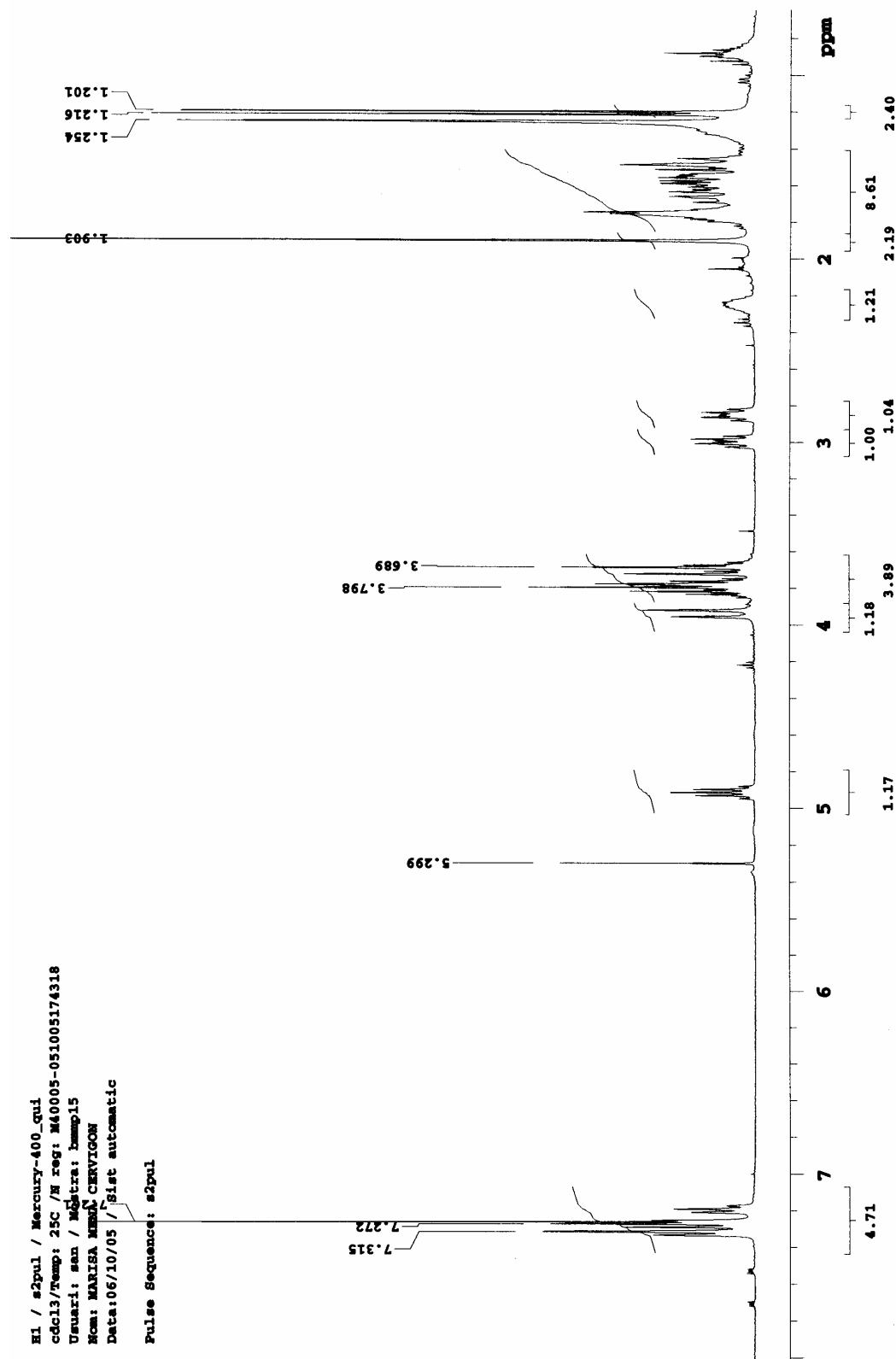
SAMPLE	FLAGS	nm
date Sep 29 2005	hs	nm
solvent cdcl3	sepul	n
sample auto_Custom-hsgly	1002	
Q_29Sep2005	SPECIAL	
ACQUISITION	temp	25.0
sw	3906.2 gain	36
at	0.150 spin	0
up	1172 F2 PROCESSING	
fb	not used ab	-0.075
ss	16 sbs	not used
dl	1.000 fn	2048
nt	4 F1 PROCESSING	
2D ACQUISITION	abl	-0.066
sw1	3906.2 sbs1	not used
ni	256 proc1	lp
PRESATURATION	fn1	2048
satmode	n	DISPLAY
satfrq	0 sp	21.1
satdly	0 wp	2329.2
satpwr	0 sp1	257.8
TRANSMITTER	wp1	1886.3
tn	H1 rfl	483.0
stfrq	400.113 rfp	0
tof	-556.0 rfl1	483.0
tpwr	58 rfp1	0
pw	12.200	PLOT
GRADIENTS	wc	131.8
gslv11	1002 sc	6.2
gt1	0.001000 wc2	131.8
gstab	0.000500 sc2	0
DECOUPLER	vs	1677
dn	C13 th	8
dm	nmn al	cdc av

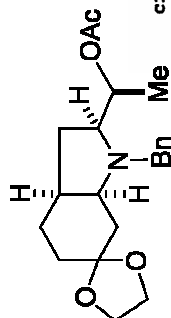






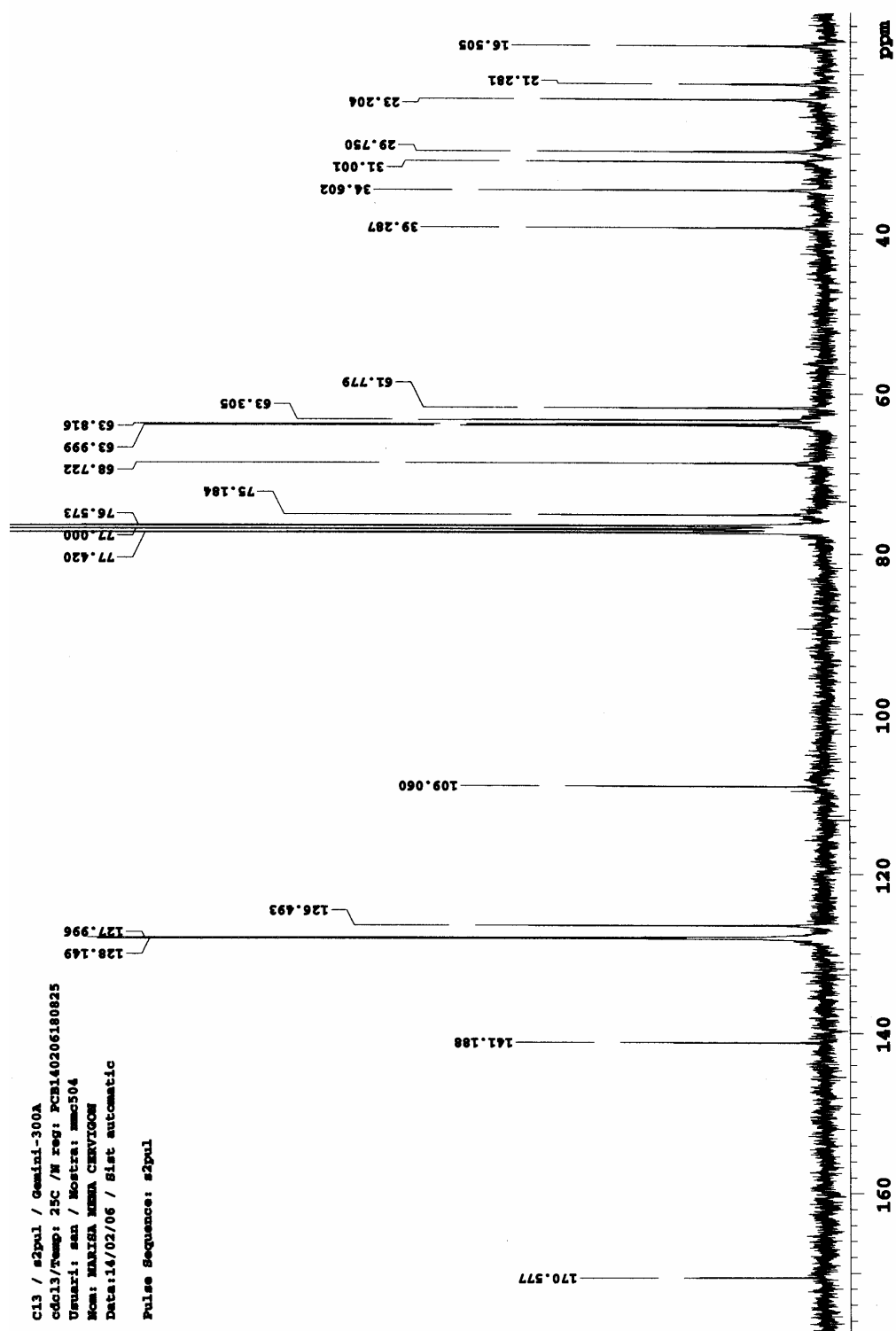
H1 / s2pul / Mercury-400_qul
 cdcl3/Temp: 25C / N reg: M40005-051005174318
 Usuari: san / Maestra: bump15
 Nom: MARISA MENA CERVIGON
 Data:06/10/05 / Sist automatic
 Pulse Sequence: s2pul

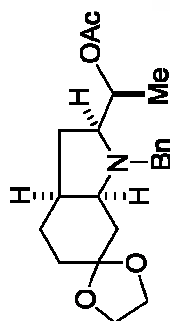




10c

C13 / s2pul / Gemini-300A
 cdcl3/Temp: 25C / W reg: PCHE40206180825
 Uruari: san / Mostra: mmc504
 Nom: MARISA MENA CERVIJON
 Data:14/02/06 / Sist automatic
 Pulse Sequence: s2pul





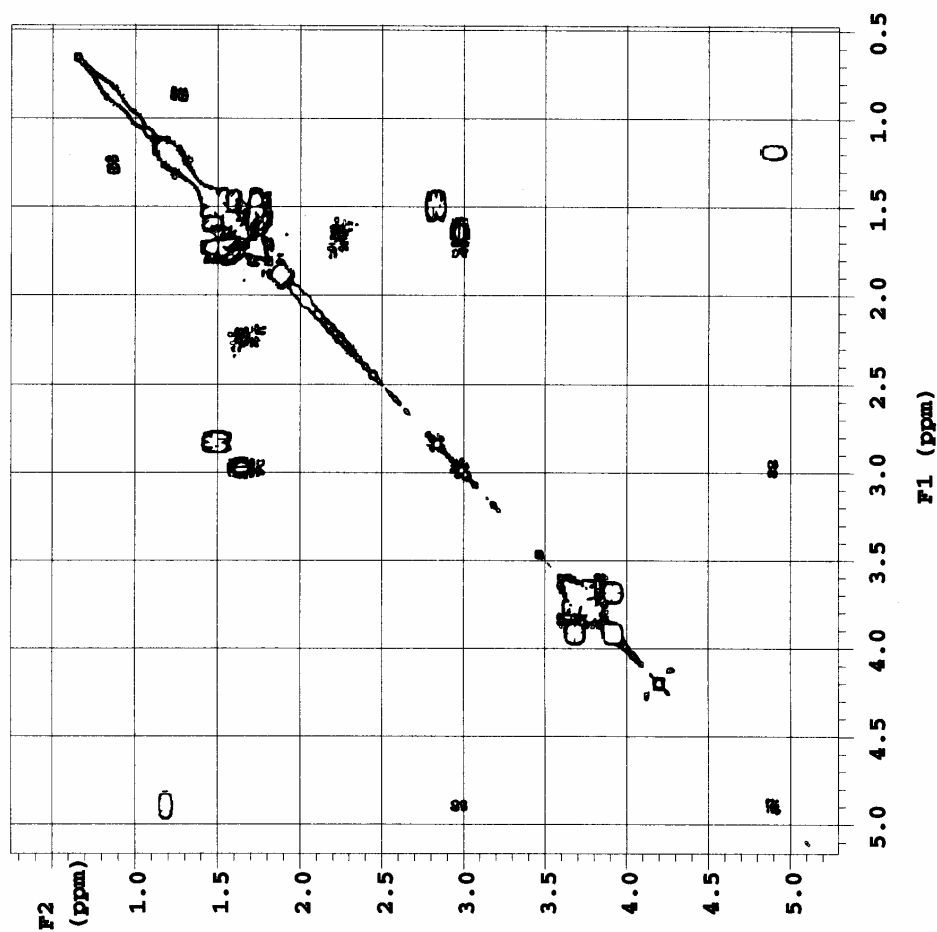
10c

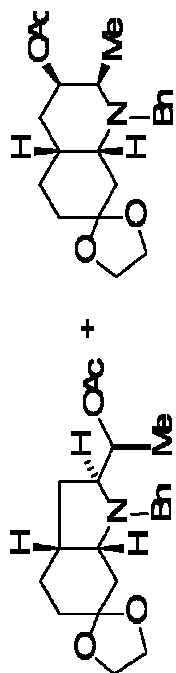
H1 / gCOSY / Mercury-400_qui
cdcl3/Tmp: 25C /W reg: M40005-051005174
318

Usuari: san / Mostra: hamp15
Nom: MARISA MENA CERVIQON
Data:06/10/05 / Sist automatic

exp22 gCOSY

SAMPLE		FLAGS	
date	Oct 6 2005	hs	nn
solvent	cdcl3	sepul	n
sample	auto_Custom- hsgl1	1002	
Q_06Oct2005 SPECIAL			
ACQUISITION		temp	25.0
sw	4065.0	gain	26
at	0.150	spin	0
np	1220	F2 PROCESSING	
fb	not used	sb	-0.075
ss	16	sbs	not used
dl	1.000	fn	2048
nt	8	F1 PROCESSING	
2D ACQUISITION		sb1	-0.063
sw1	4065.0	sbs1	not used
ni	256	procl	lp
PRESATURATION		fn1	2048
satmode n DISPLAY			
satfrq	0	sp	102.2
satdly	0	wp	2014.6
satpwr	0	sp1	189.6
TRANSMITTER		wp1	1875.6
tn	H1	xf1	497.8
sfrq	400.113	rfp	0
tof	-479.0	rf11	497.8
tpwr	58	rfp1	0
PLOT			
GRADIENTS		wc	131.8
gzlv11	1002	sc	6.2
gt1	0.001000	wc2	131.8
gstab	0.000500	sc2	0
DECOUPLER		vs	365
dn	C13	th	5
dm	nm	ai	cdc
		av	



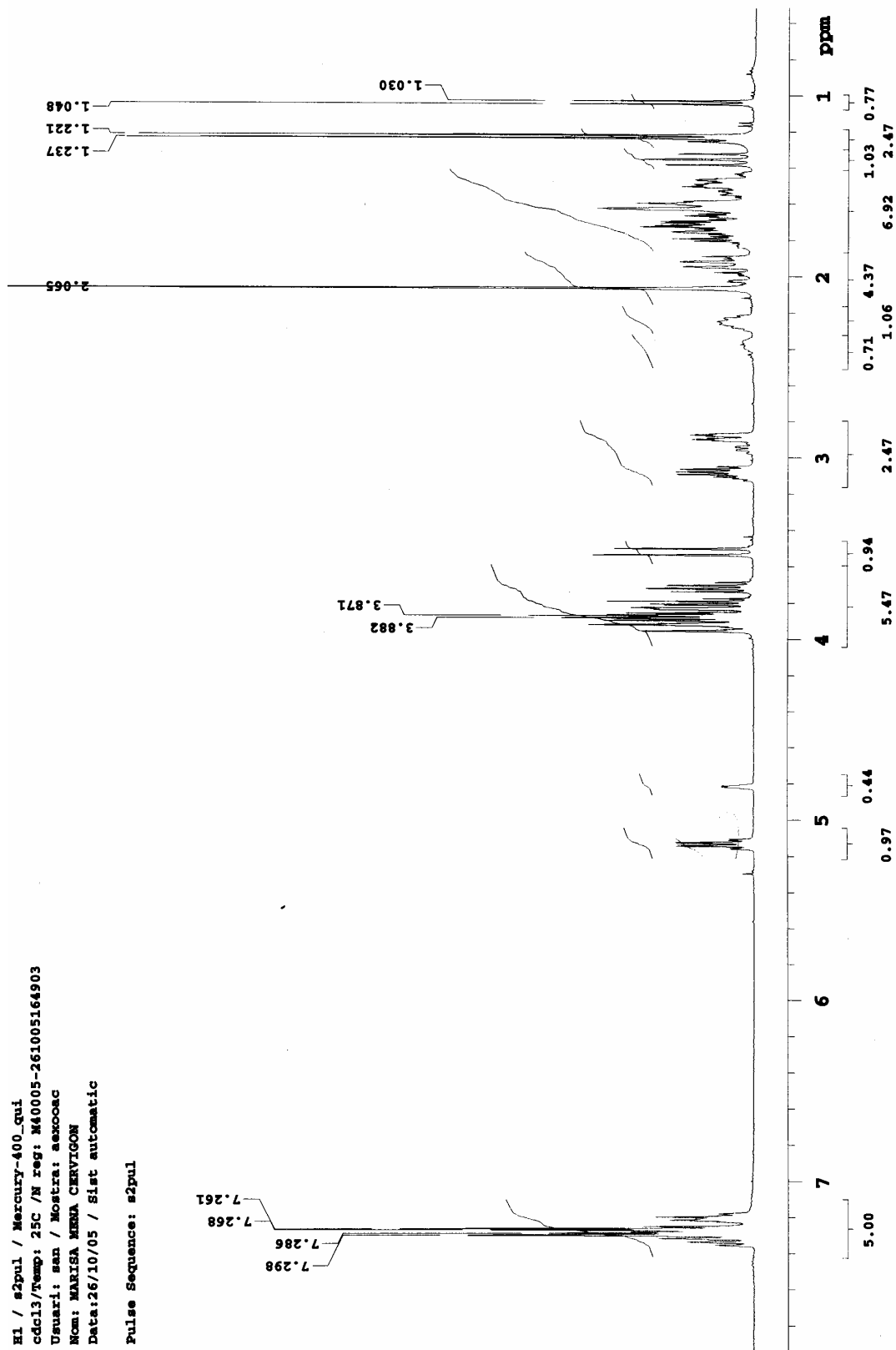


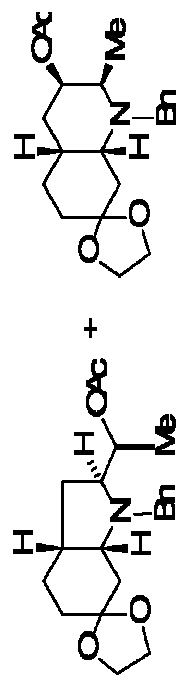
11c

19

H1 / s2pul / Mercury-400.qui
 cdcl3/Temp: 25C /N reg: M40005-261005164903
 Usuari: san / Mostra: sexooac
 Nom: MARISSA MESA CERVIGON
 Data:26/10/05 / Sist automatic

Pulse Sequence: s2pul

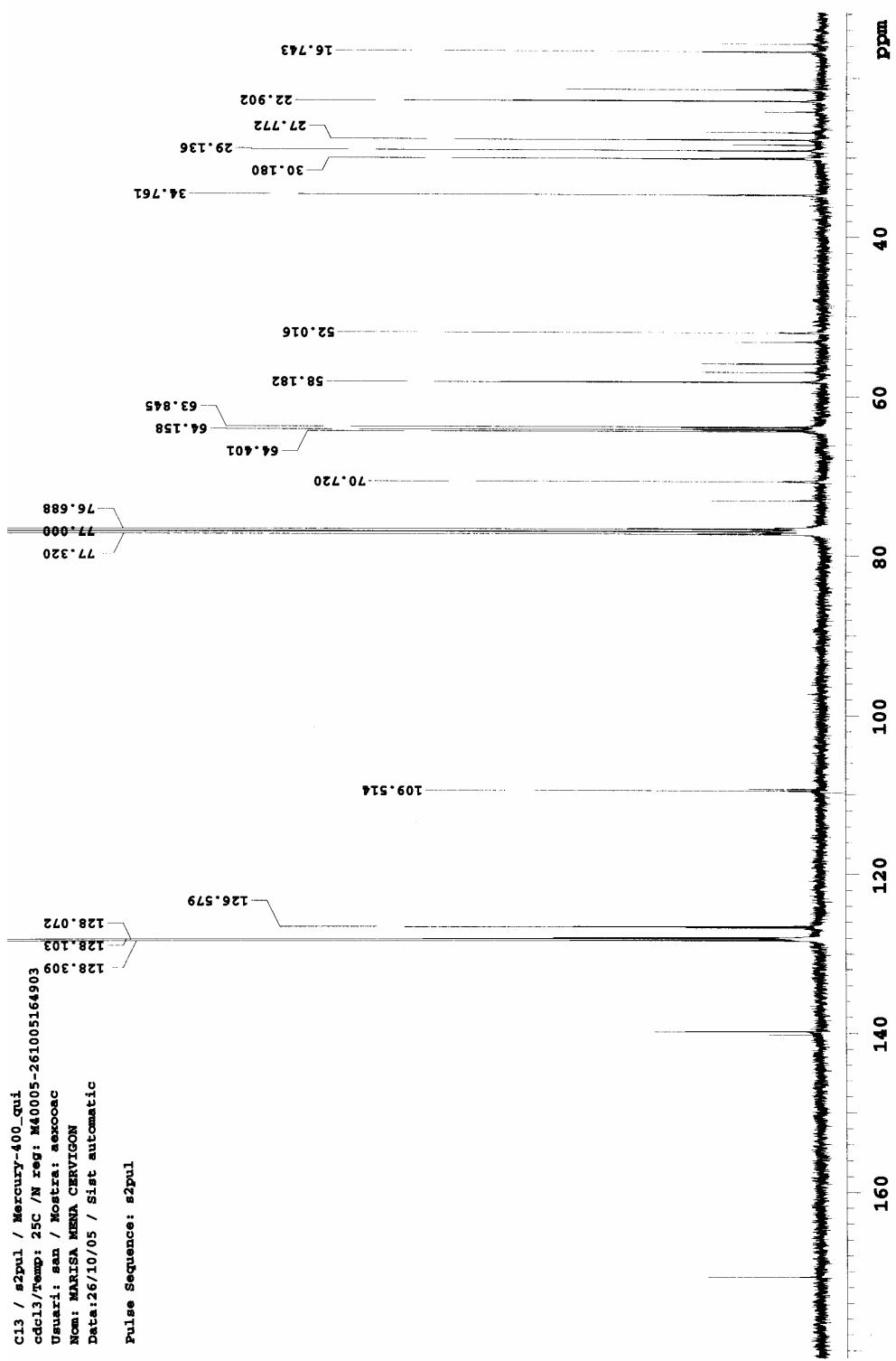




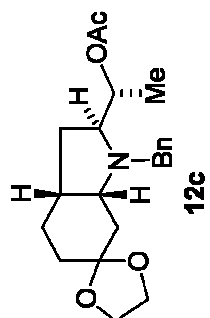
11c

19

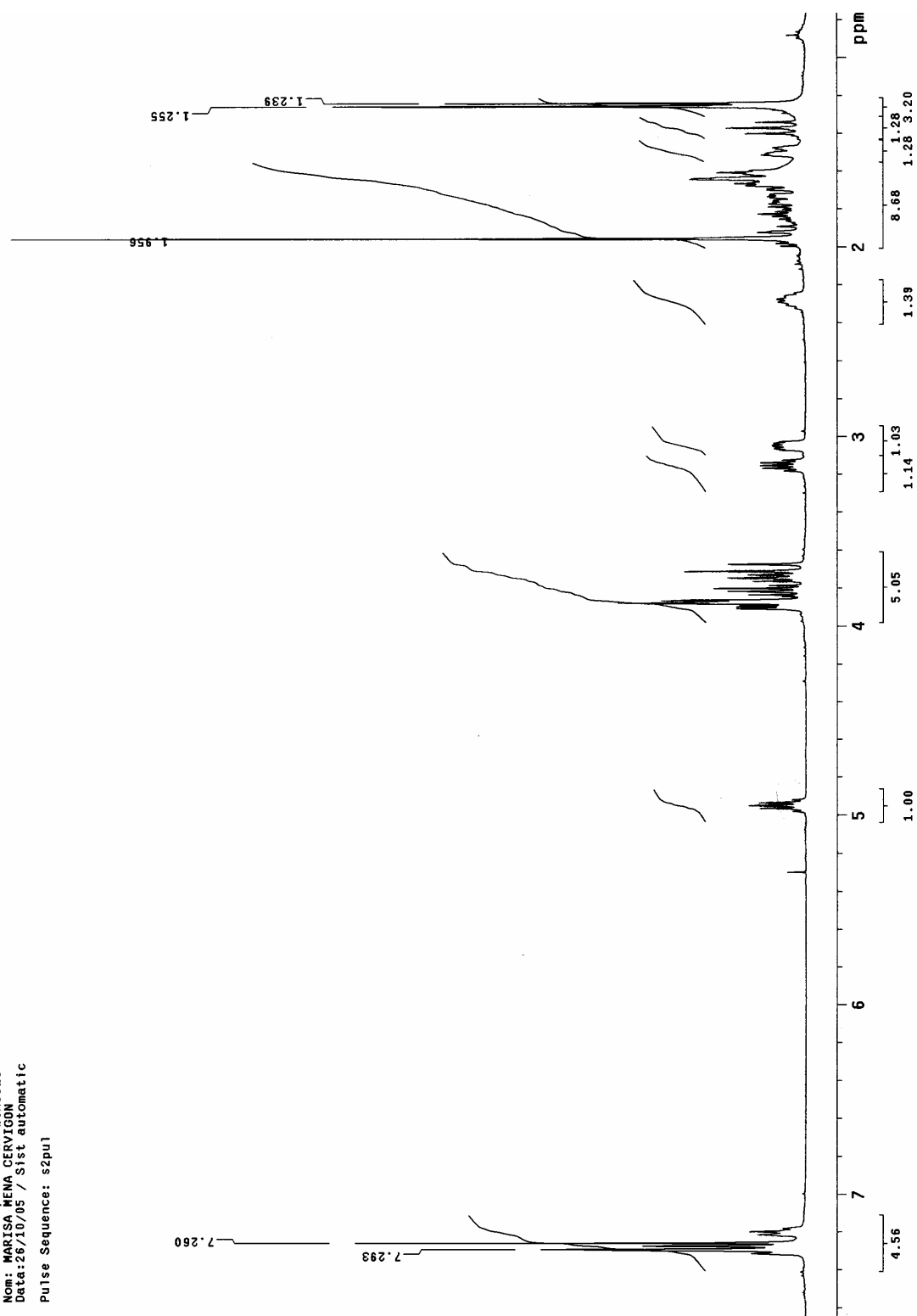
C13 / s2pul / Mercury-400_qui
 cdc13/Temp: 25C /N reg: M4005-261005164903
 Usuari: san / Mostra: sexocac
 Nom: MARISA MENA CERVIGON
 Data:26/10/05 / Sist automatic
 Pulse Sequence: s2pul

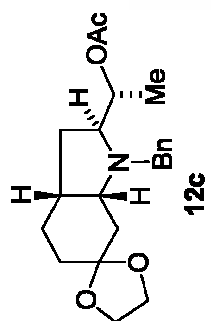




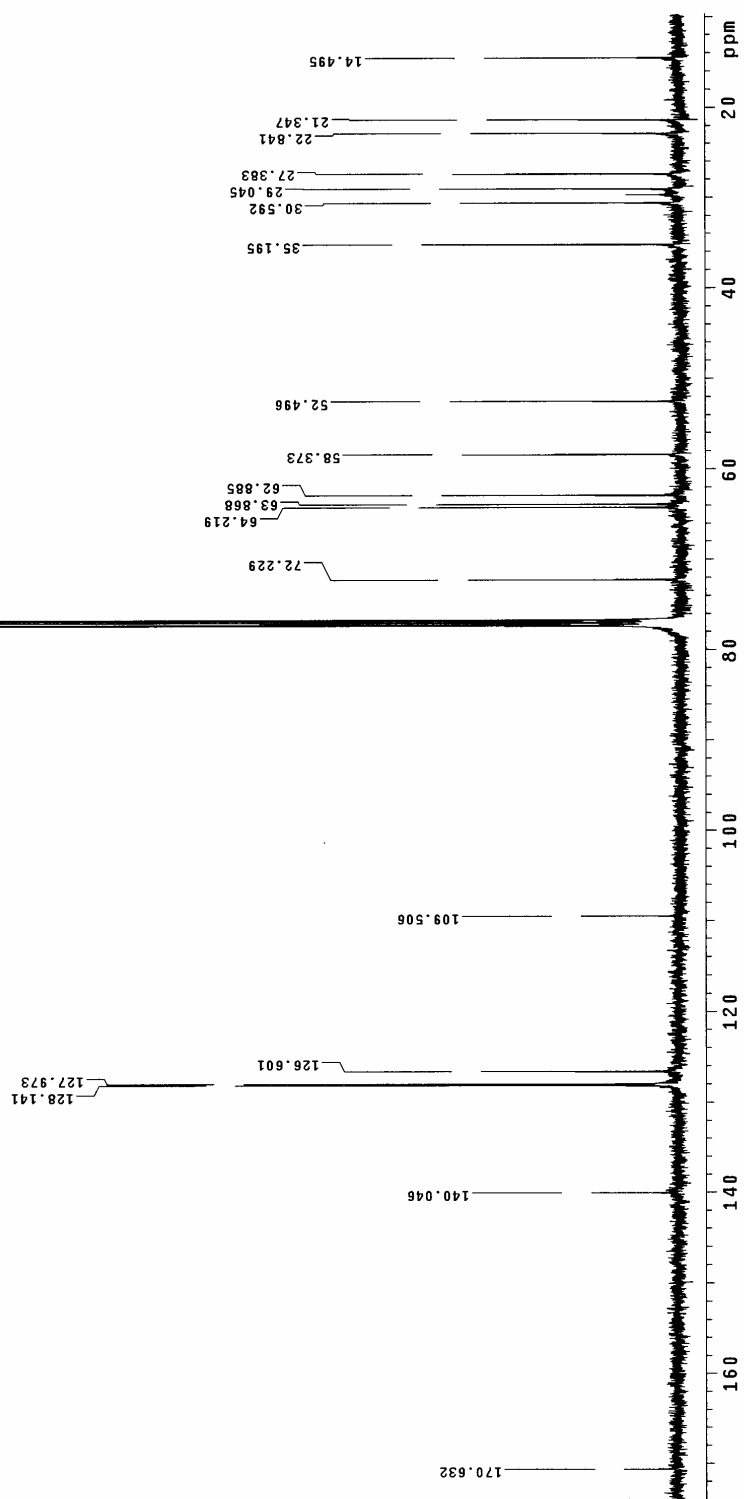


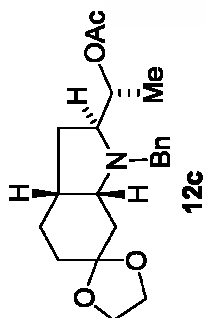
cdc13/Temp: 25C /N reg: M40005-261005165718
 Usuari: san / Nostra: bexooac
 Nom: MARISA MENA CERVIGON
 Data:26/10/05 / Sist automatic
 Pulse Sequence: s2pul



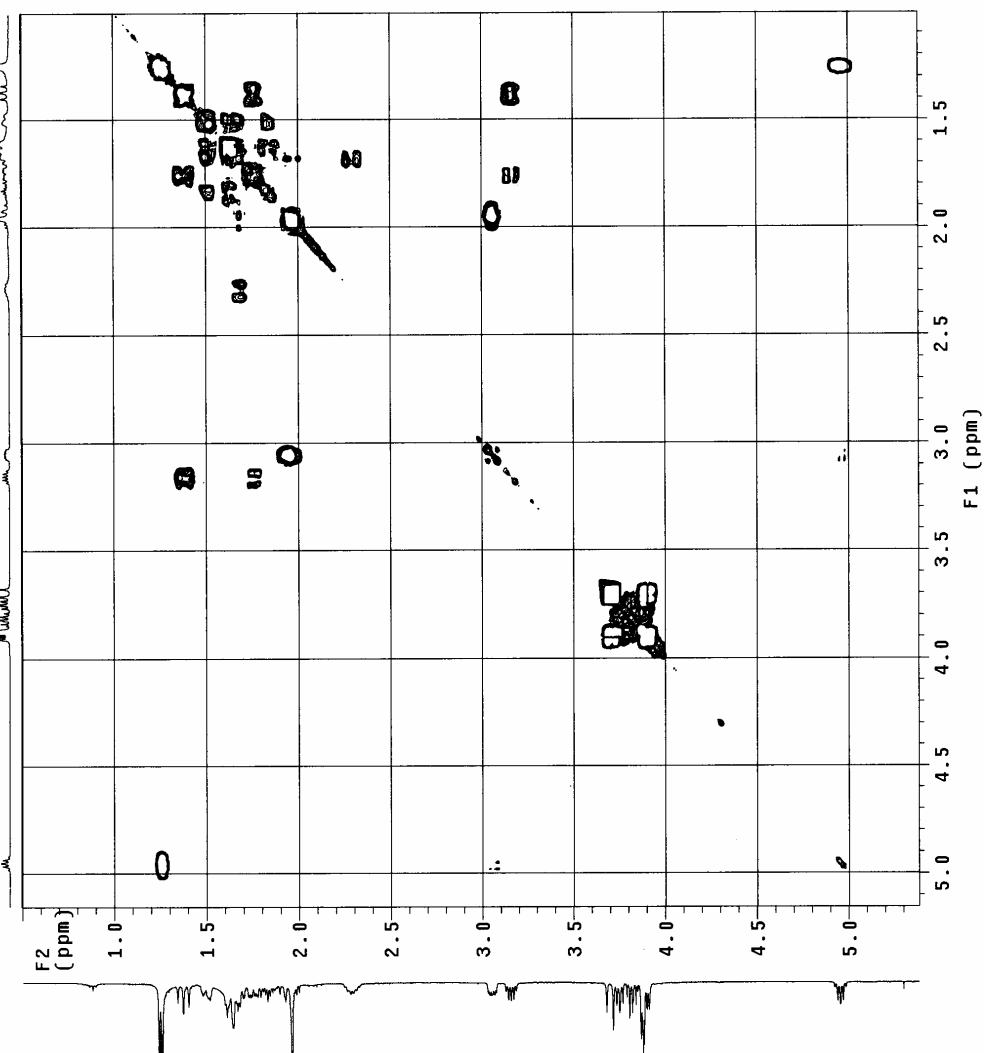


cdc13/Temp: 25C /N req: M40005-261005165718
 Usuari: san / Mostra: bexooac
 Nom: MARISA MENA CERVIGON
 Data: 27/10/05 / Sist automatic
 Pulse Sequence: s2pul1

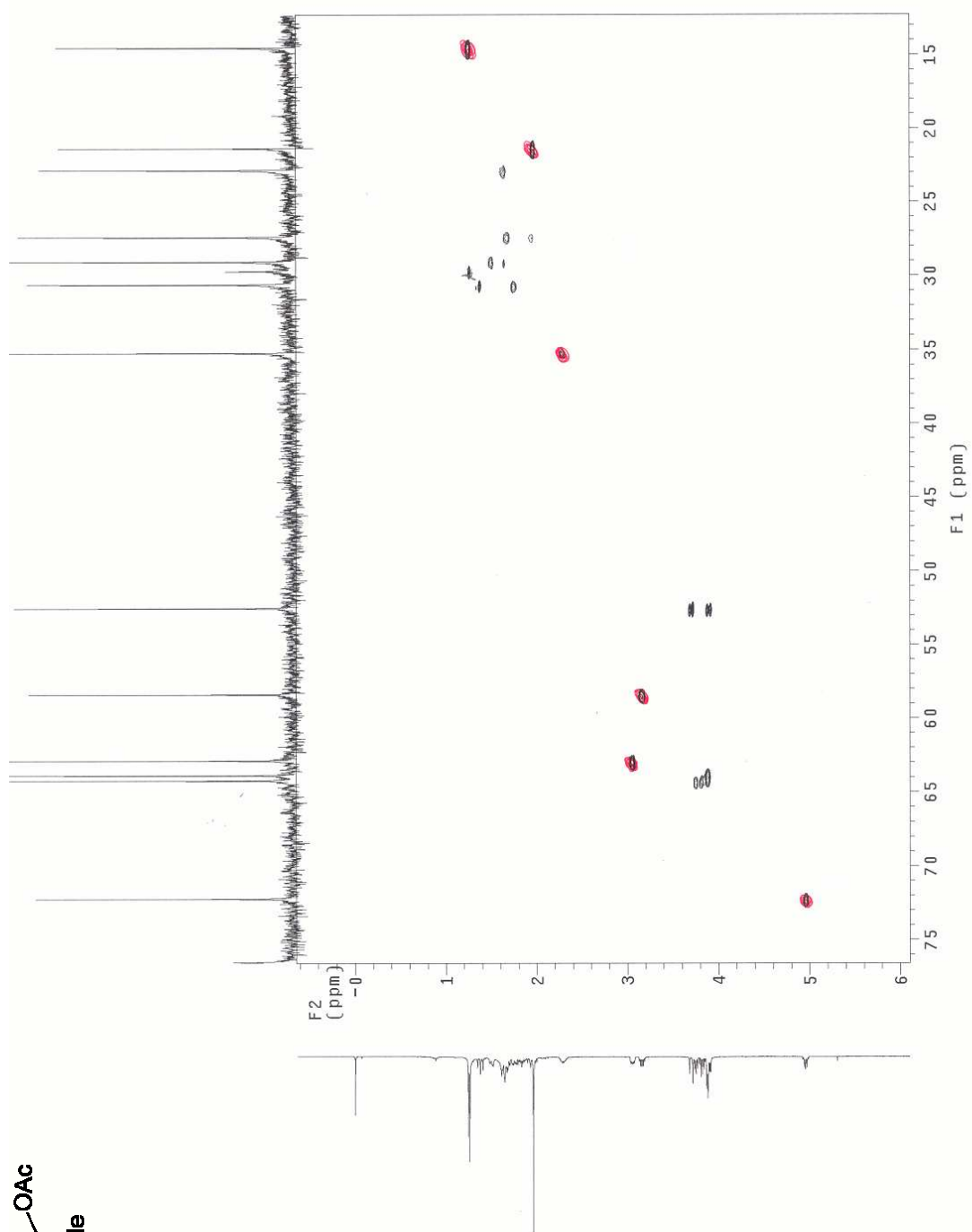
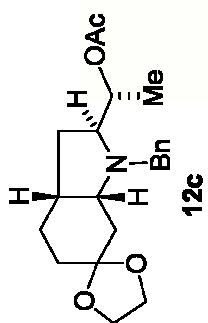


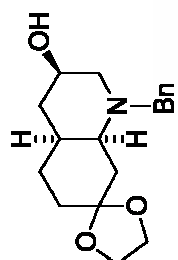


H1 / gCOSY / Mercury-400_rui
 cdc13/Temp: 25C /N reg: M40005-261005165
 718
 Usuari: san / Mostra: bexooac
 Nom: MARISA MENA CERVIGON
 Data: 27/10/05 / Sist automatic
 exp4 gCOSY

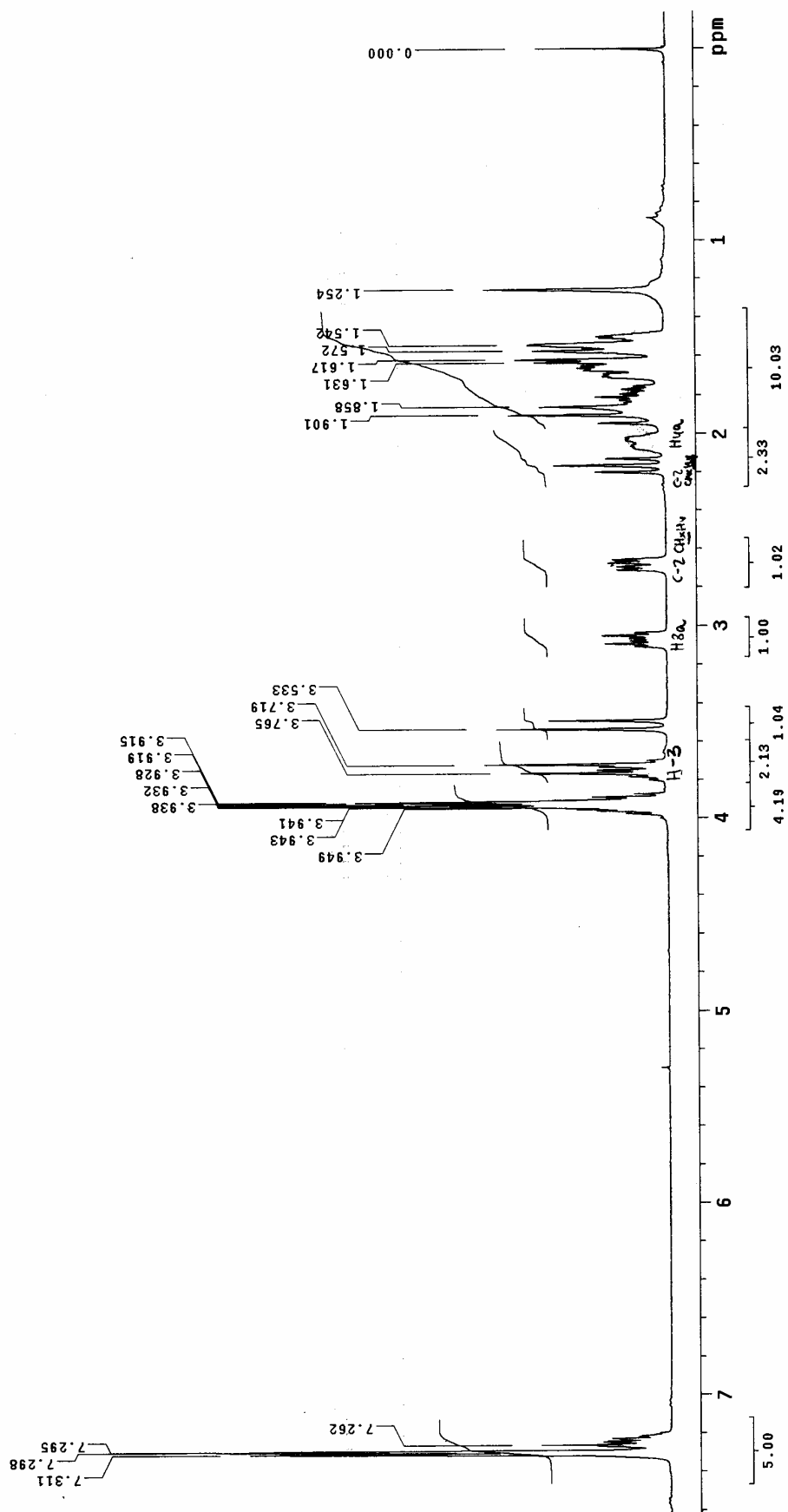


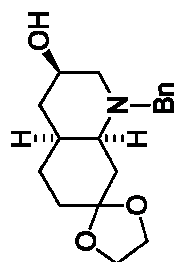
SAMPLE		FLAGS	
date	Oct 27 2005	hs	nn
solvent	cdc13	sspl	n
sample	autoCustom~	hsgl	1000
Q_26Oct2005		SPECIAL	25.0
ACQUISITION			
sw	333.7	0	gdn
at	0.150	sp1	28
nb	1182	sp1	0
fb	not used	sb	-0.075
ss	16	sbs	not used
di	1.000	fn	2048
nt	4	f1	PROCESSING
2D ACQUISITION	0	sb1	-0.065
sw1	333.7	0	sdsl
nt	256	sp1	not used
PRESATURATION	0	fn1	2048
satmode	n	DISPLAY	
satfrq	0	sp	133.0
satdly	0	wp	1355.0
satpwr	0	sp1	402.9
TRANSMITTER			
trnq	400.113	rf1	1658.7
trnf	400.113	rf1	486.1
tof	-533.5	rf1	486.1
tpwr	58	rfp1	0
GRADIENTS			
pw	12.300	wc	153.0
g2lv11	1000	sc	6.2
g1	0.001000	wc2	153.0
gstab	0.000500	sc2	0
dn	DECOUPLER	c13	333
dm	nnn	a1	cdc
		av	7





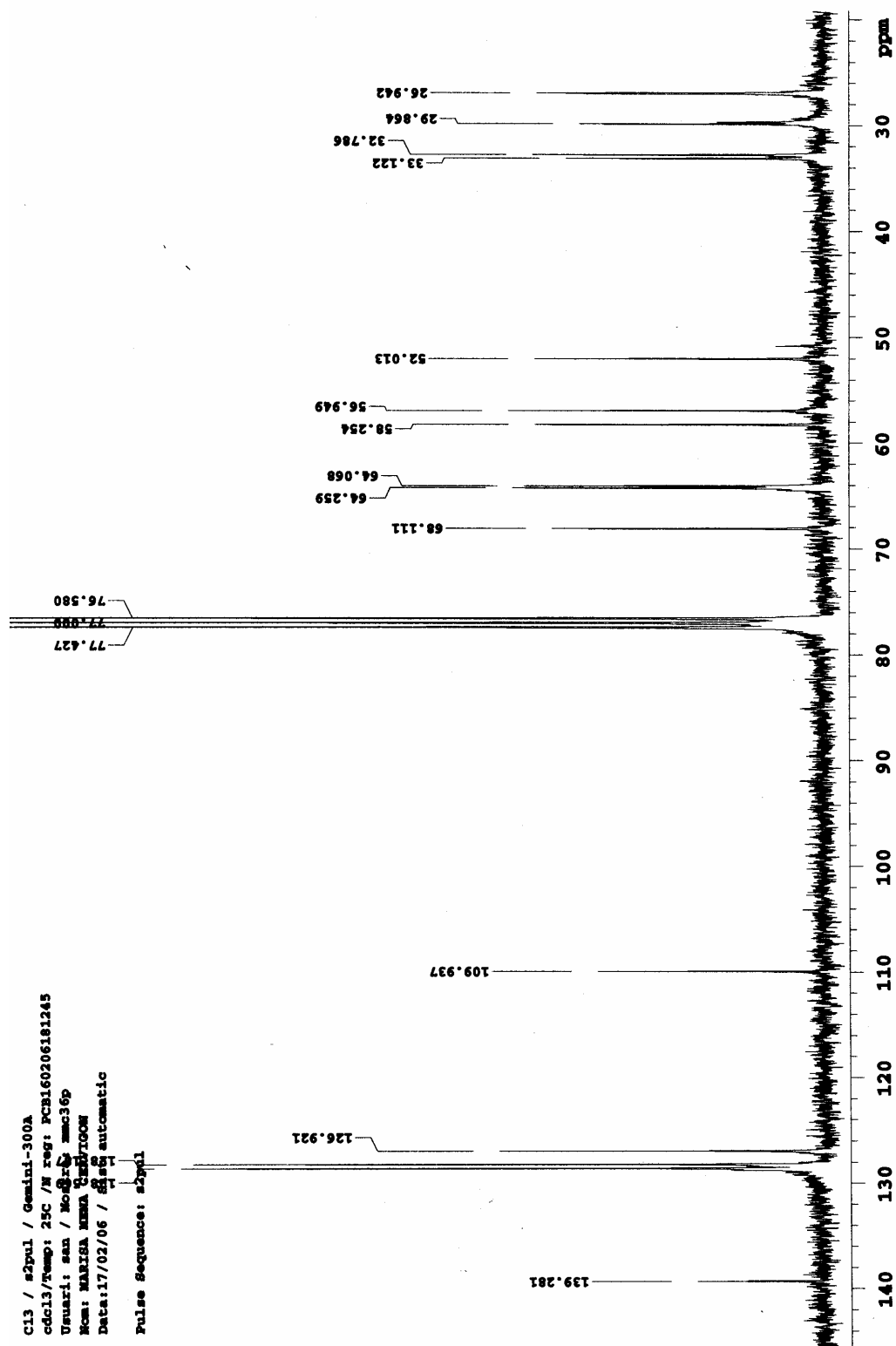
13

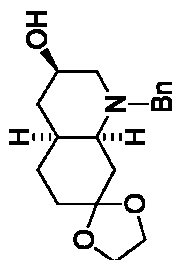




13

C13 / s2pul / Gemini-300A
 cdcl3/Temp: 25C / W reg: PCB160206181245
 Usuari: san / Mod: mmo36p
 Nom: MARISA MARIA CAS/100M
 Data: 17/02/06 / 25C automatic
 Pulse Sequence: s2pul





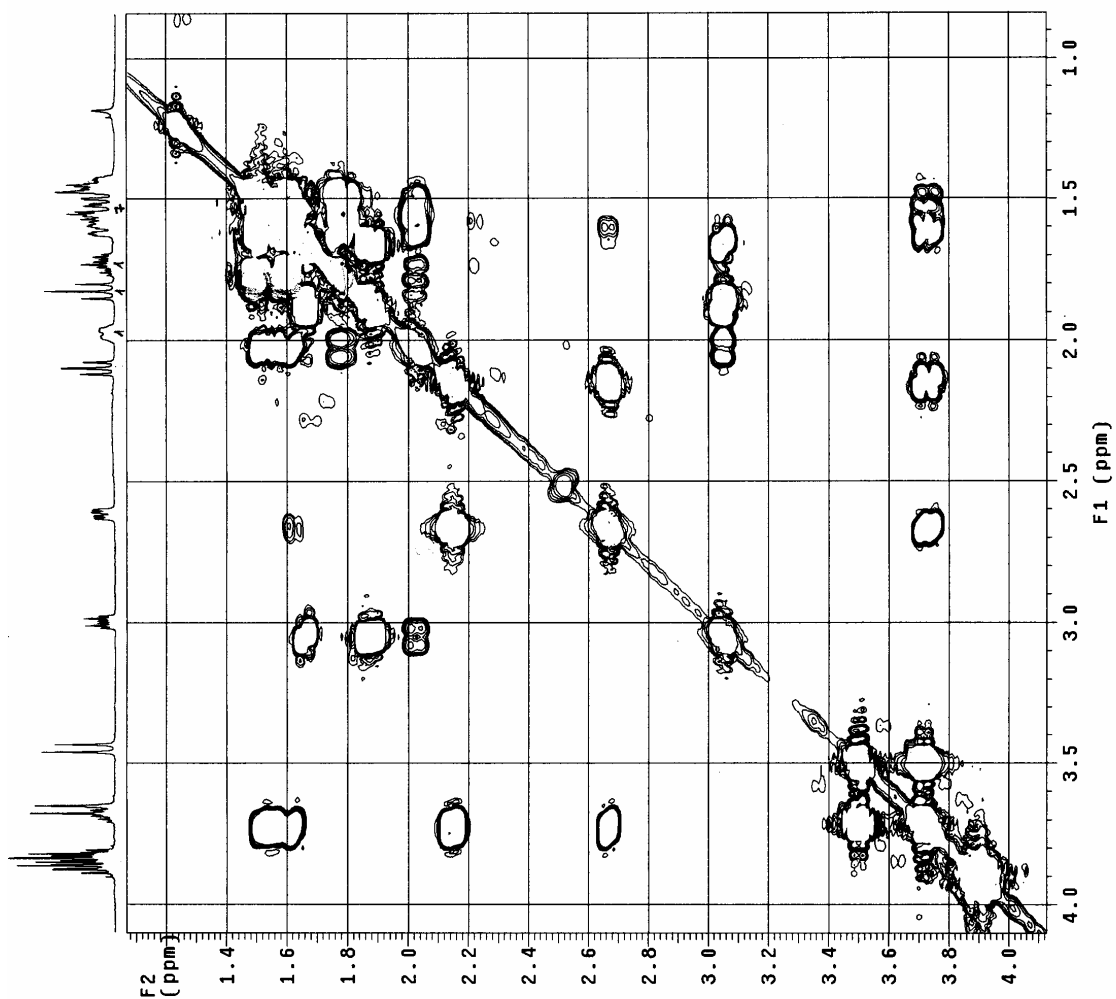
13

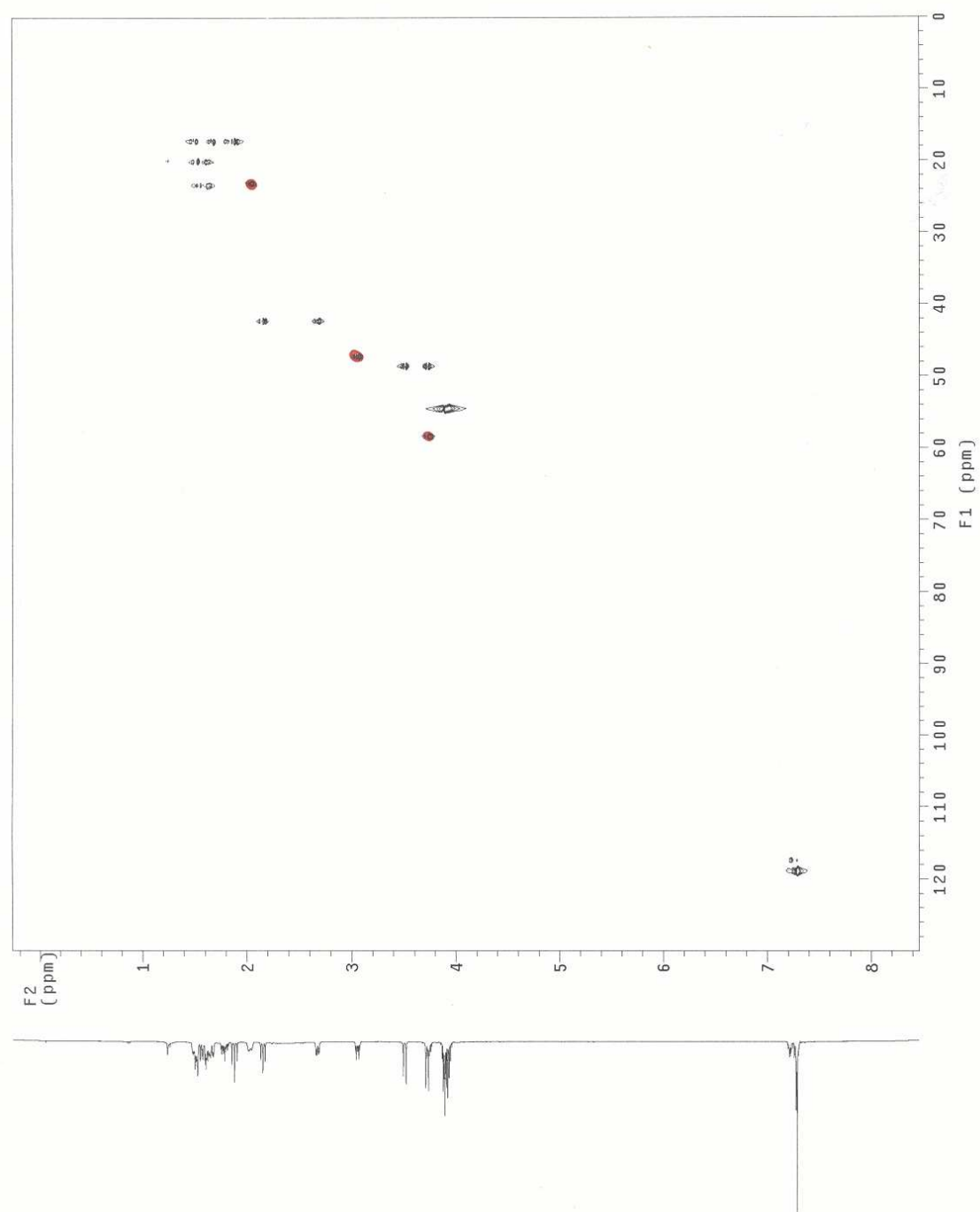
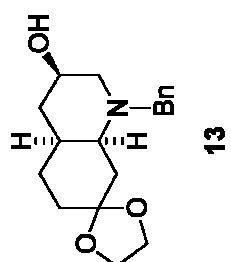
H1 / gcosy / Bruker DMX500
cdcl3 / 25C / N Reg.: 00425
exp1 / m005 / MWet.: 00425
Data: 22/05/02 / Op.: MA Molins

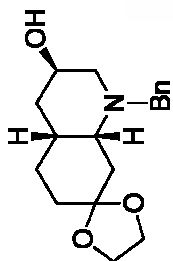
los datos en formato original estan en:
/data3/8500/san/May2002/san-00425

exp1 cosy9s

SAMPLE DEC. & VT
date 05-22-02 dfrq 500.621
solvent cdc13 dof 3257.4
file 3.cv dhp 20
ACQUISITION
sfrq 500.620 temp 25.0
at 0.469 sb 0.059
np 2048 ss not used
pw 4370.6 hzid 70
ss 16 wtfille
pw 3.8 proc
rd 1.000 fn 2048
tof 2069.7 math
nt 4 2D PROCESSING
ct 0 sb1 0.029
dp 2D ACQUISITION y wtfille1
sw1 4370.6 procl
n1 256 f1
2D DISPLAY
sp 535.6
sp1 420.3 wp 1529.5
wp1 1632.0 vs 500
sc2 0 sc 6
wc2 153 wc 153
rf11 4370.6 hzmm 10.00
rfp1 4235.5 ls 500.00
th 4235.5 rfp
th 100.00
ins
nm cdc av

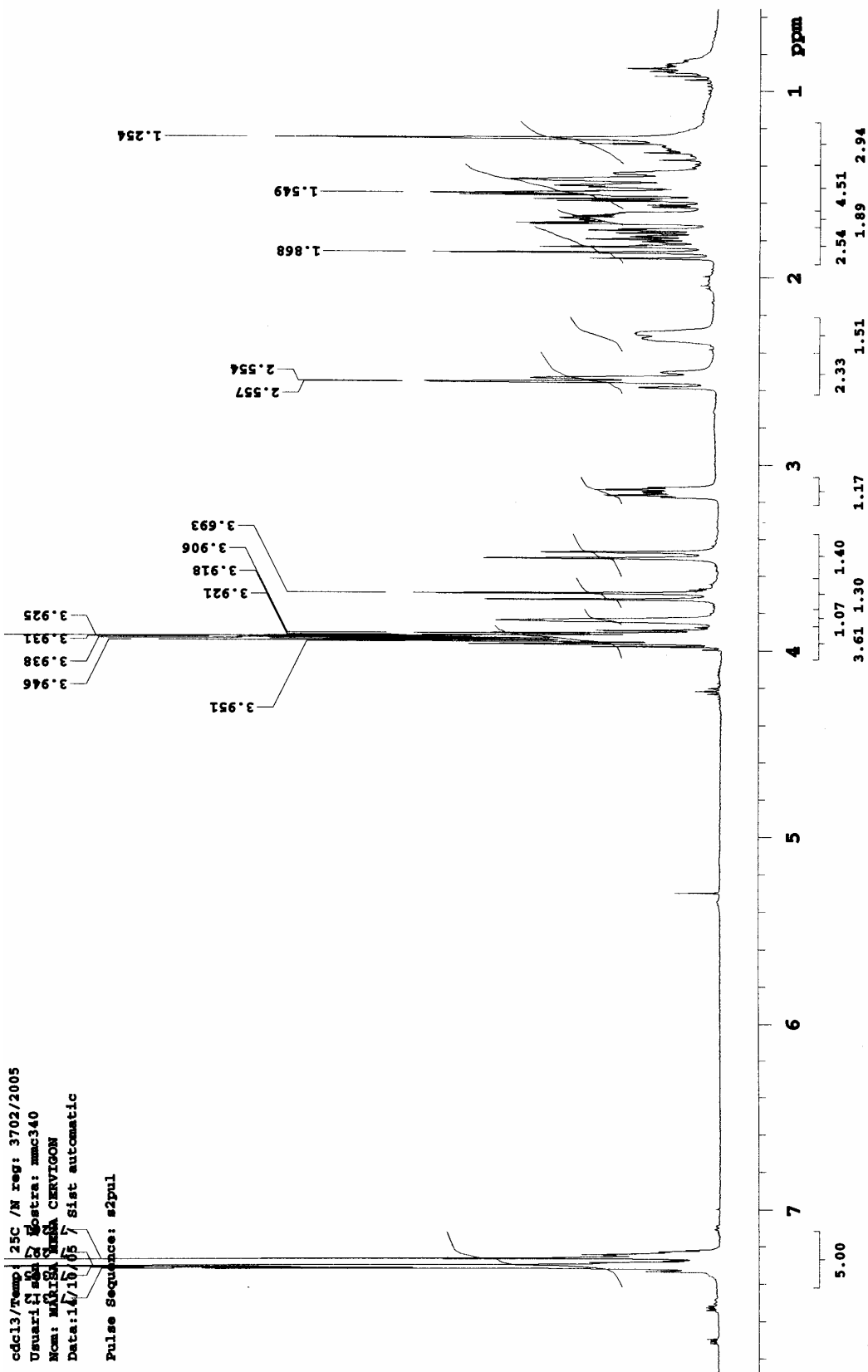


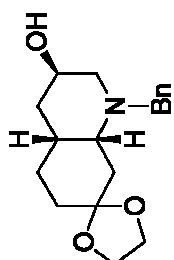




14

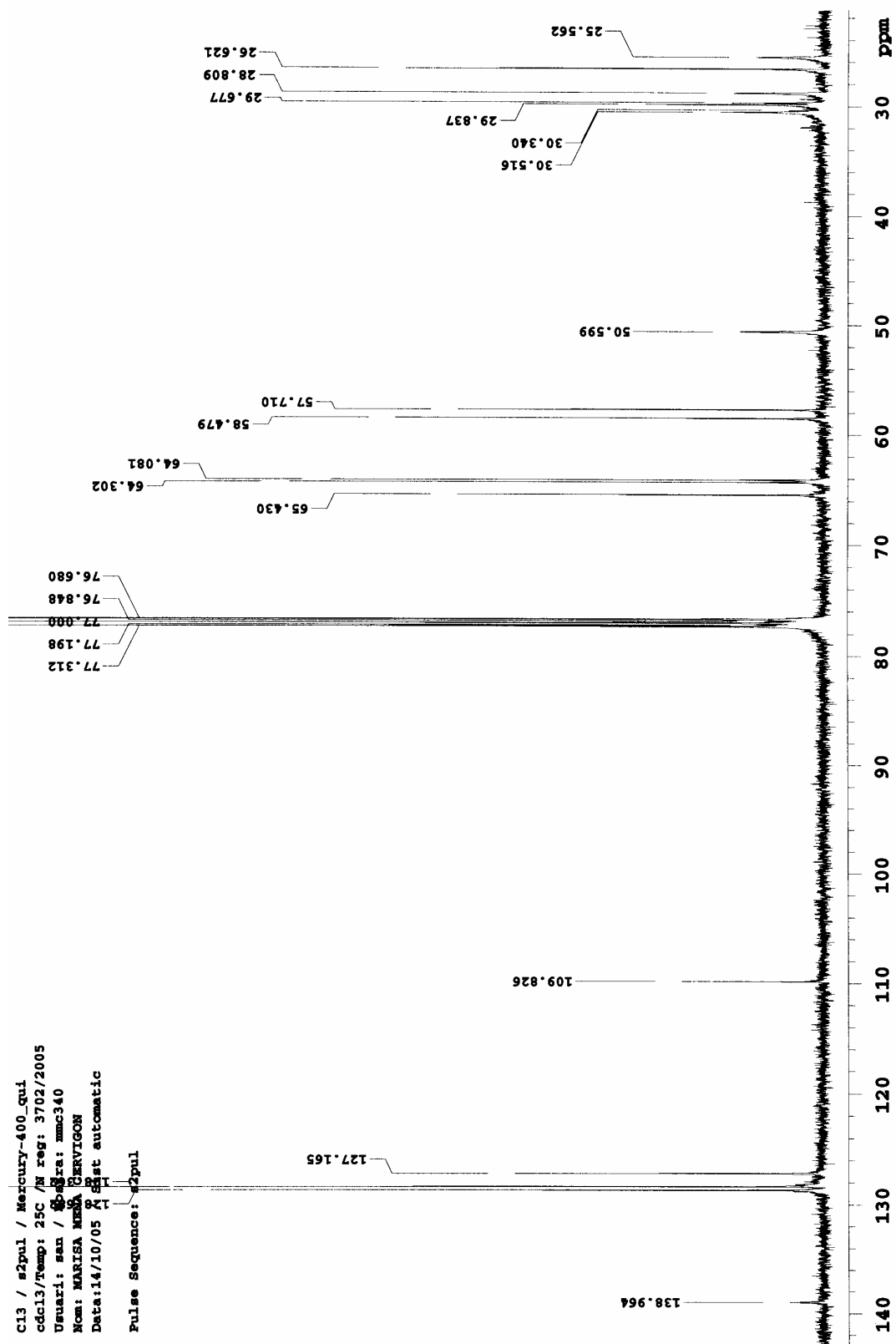
cdci3/Temp: 25C /N reg: 3702/2005
 Usuari: jlab /Q: 2 /Mostra: mmc340
 Nom: MARIA CRISTINA CERVIGON
 Data: 14/10/05 / Sist automatic
 Pulse Sequence: s2pul



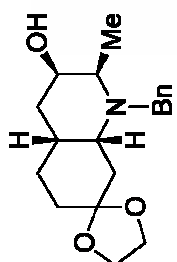


14

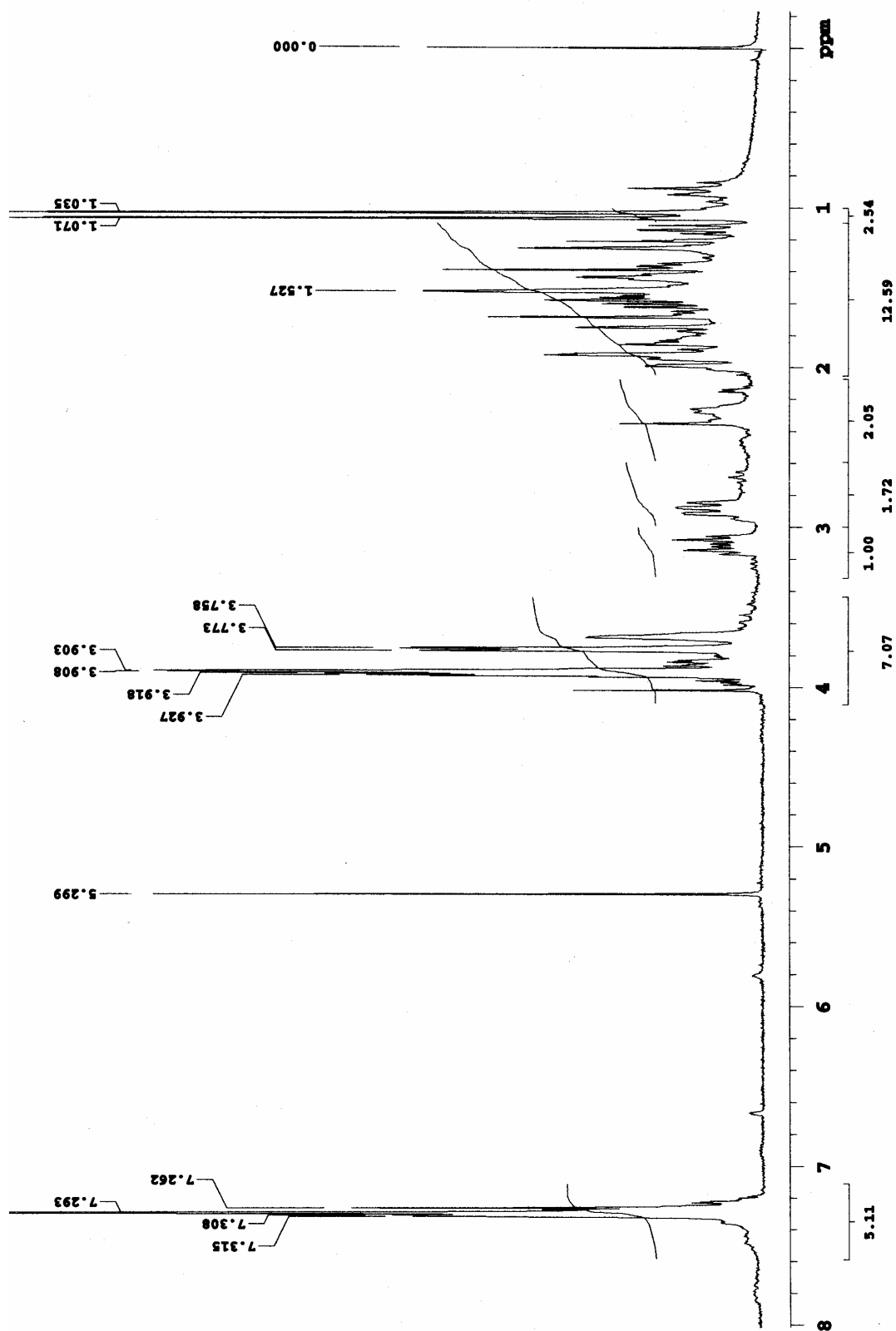
C13 / s2pul / Mercury-400_qui
 cdcl3/Temp: 25C / N reg: 3702/2005
 Usuari: san / 40-33ra: mmc340
 Nom: MARISA MORA / CERVIGON
 Data:14/10/05 14:58
 Pulse Sequence: s2pul

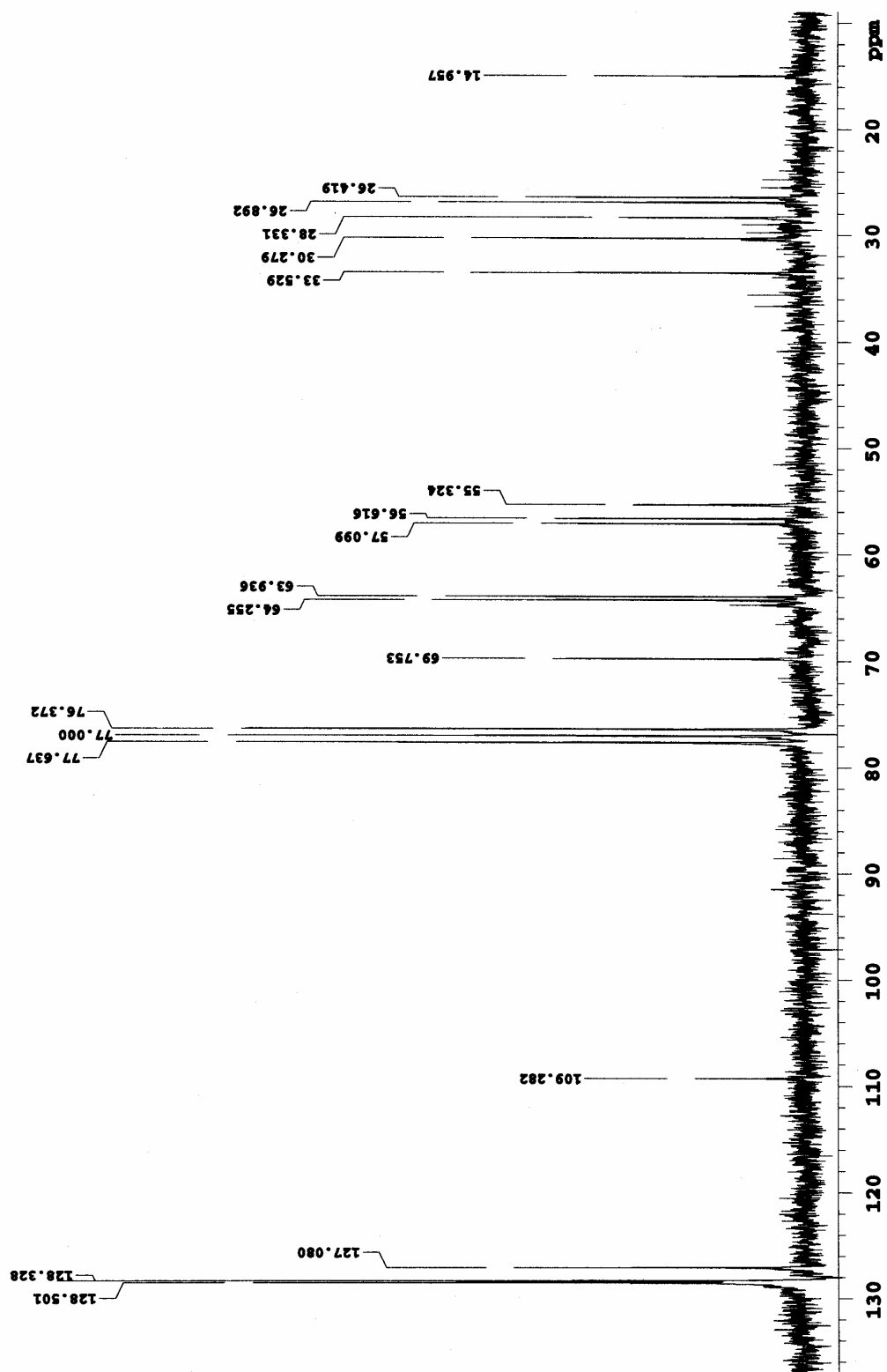
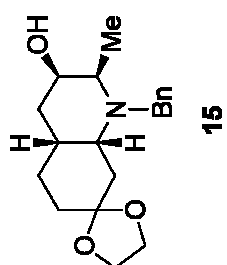


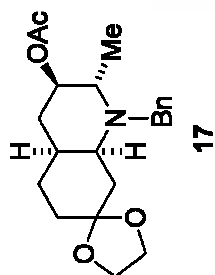




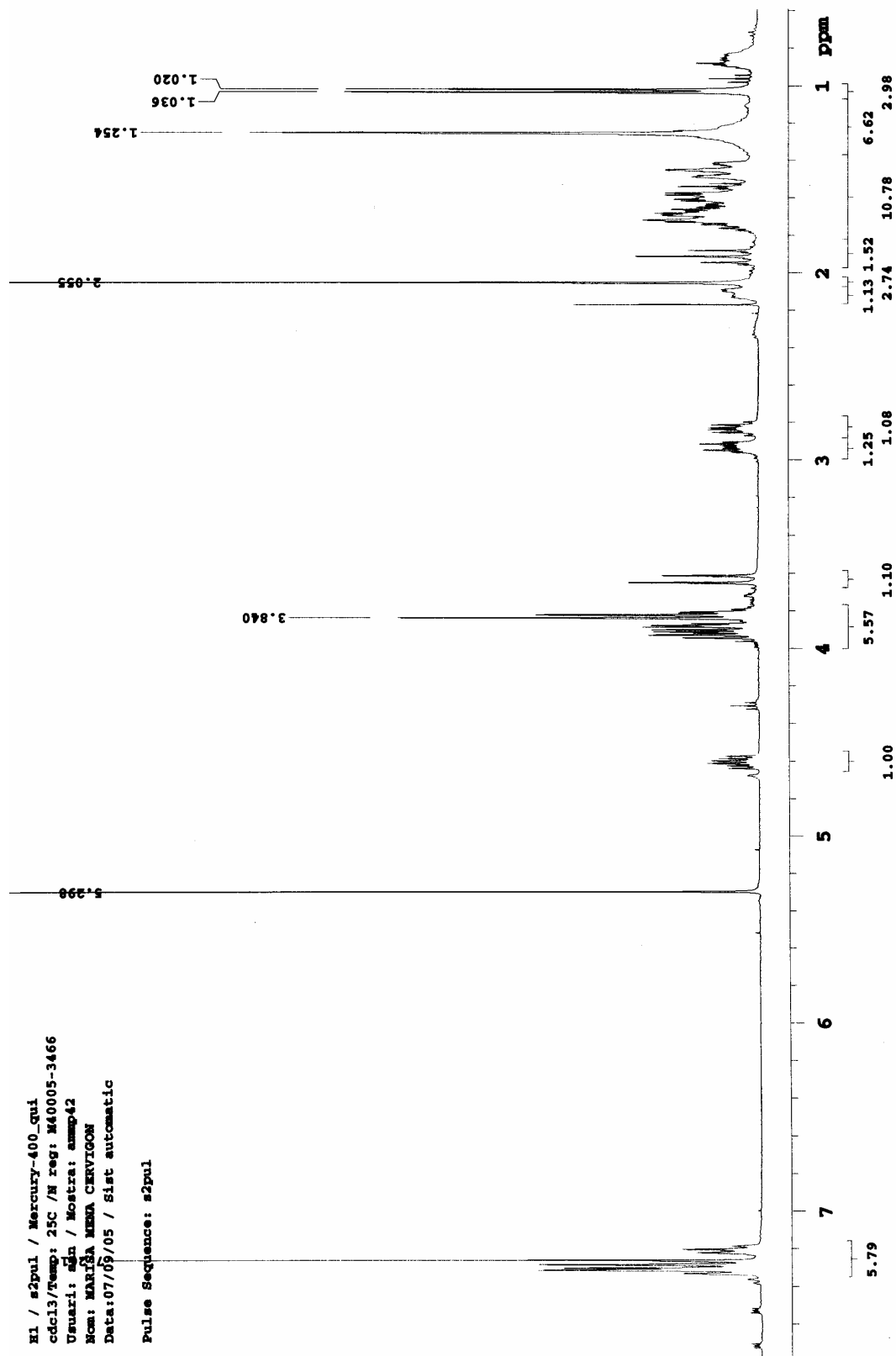
15

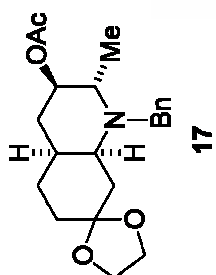




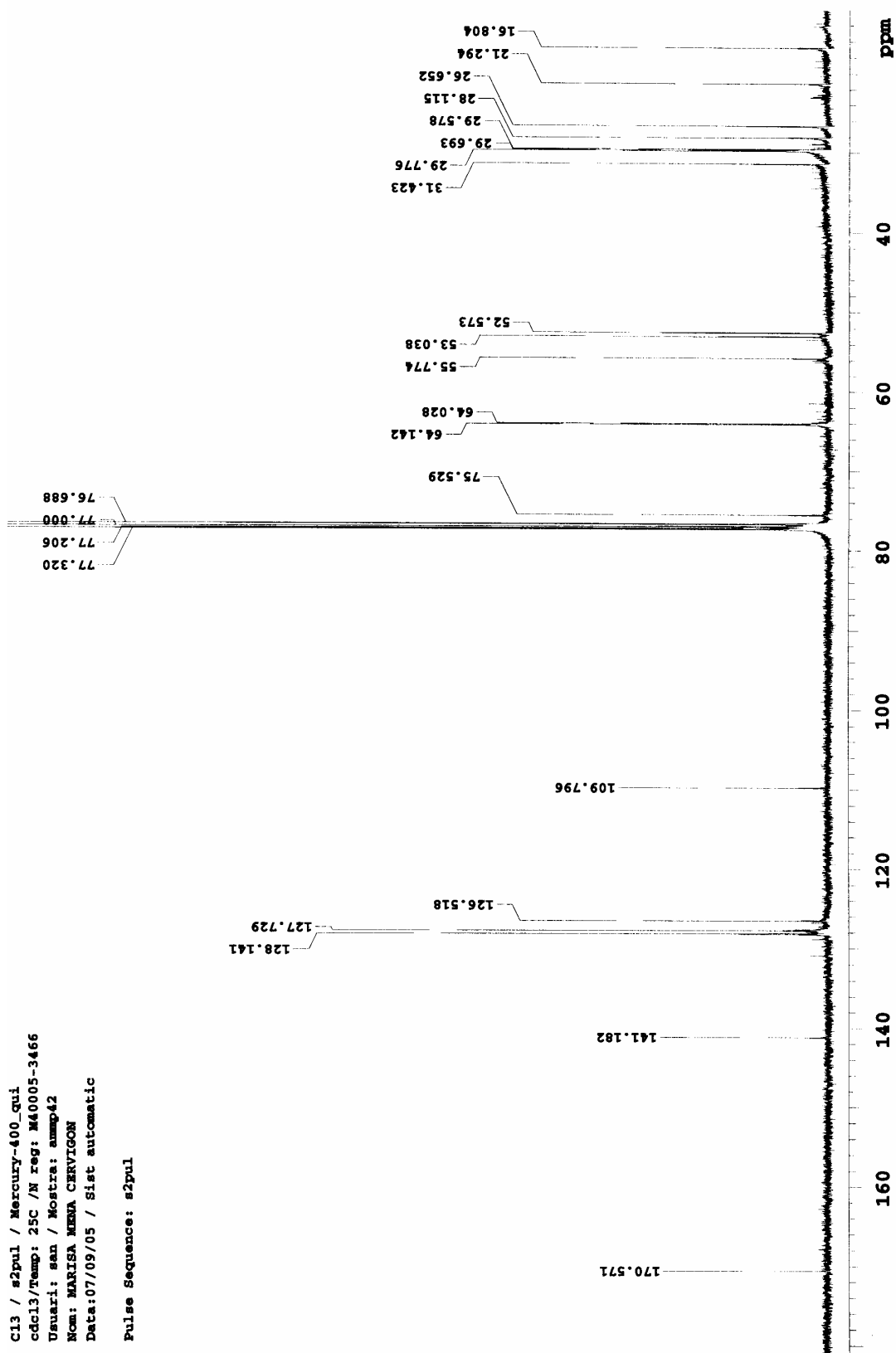


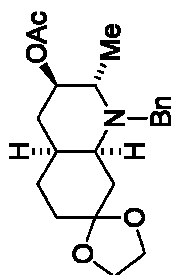
E1 / s2pul / Mercury-400_qul
 cdc13/Temp: 25C / N reg: M40005-3466
 Usuari: sdn / Mostra: ammp42
 Nom: MARIA NEMA CERVIGON
 Data: 07/09/05 / Sist automatic
 Pulse Sequence: s2pul





Cl3 / s2pul / Mercury-400_qui
 cdcl3/Temp: 25C / N reg: M40005-3466
 Usuari: san / Mostra: ammp42
 Nom: MARISA MENA CERVIGON
 Data:07/09/05 / Sist automatic
 Pulse Sequence: s2pul



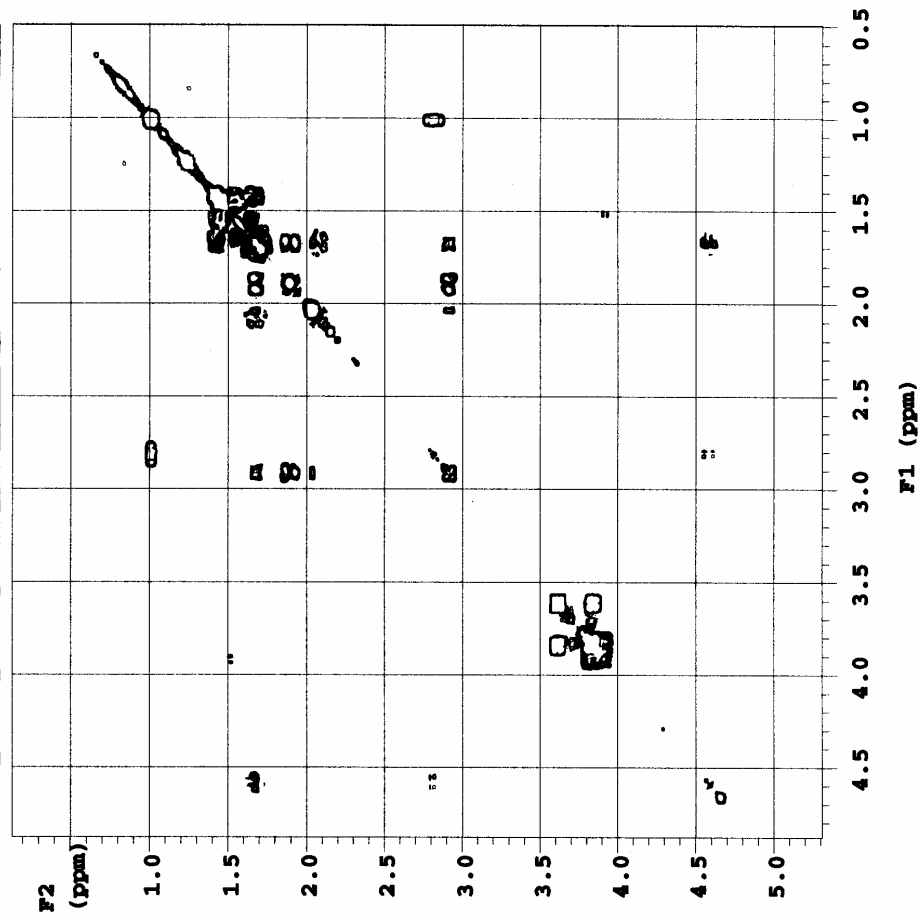


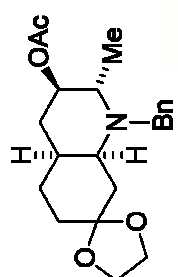
17

H1 / gCOSY / Mercury-400_qui
cdcl3/temp: 25C /M reg: M40005-3466
Usuari: san / Mostra: ammp42
Nom: MARISA MEMA CERVIGON
Data: 07/09/05 / Sist automatic

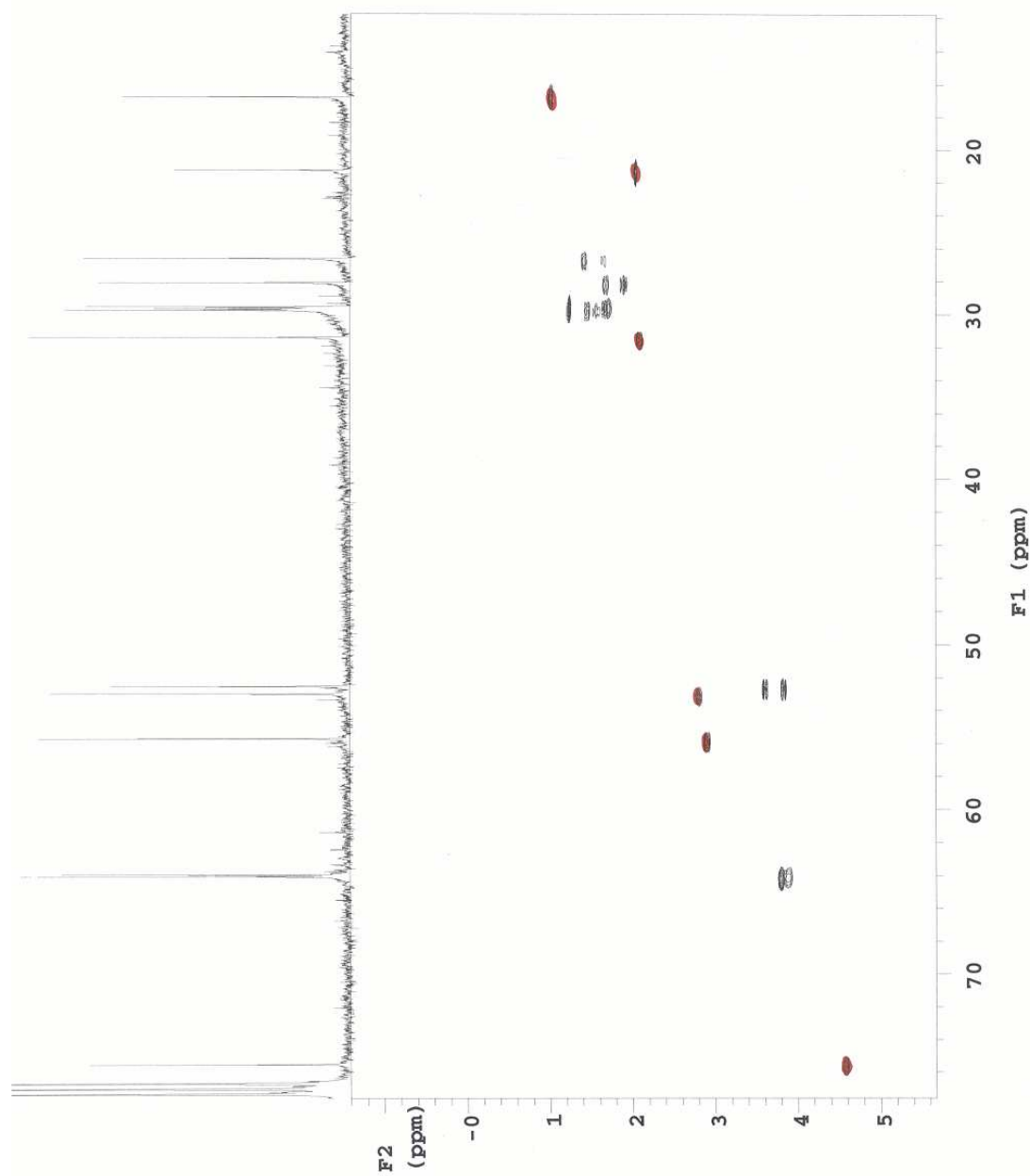
exp3 gCOSY

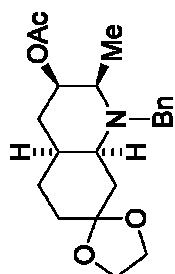
SAMPLE	date	hs	flags
Sep 7 2005	hs	mn	
solvent	cdcl3	sepu	n
sample	auto_Custom-hs1v1	1002	
Q_07Sep2005	SPECIAL		
ACQUISITION	temp	25.0	
sw	4310.3	gain	28
at	0.150	spin	0
np	1294	F2 PROCESSING	
fb	not used	sb	-0.075
ss	16	abs	not used
d1	1.000	fn	2048
nt	4	F1 PROCESSING	
2D ACQUISITION	abi	-0.059	
sv1	4310.3	sbs1	not used
ni	256	procl	lp
PRESATURATION	fn1	2048	
satmode	n	DISPLAY	
satfrq	0	sp	54.4
satdly	0	wp	2068.8
satpwr	0	sp1	197.7
TRANSMITTER	wp1	1752.8	
tn	H1	xf1	573.4
sfrq	400.113	rfp	0
tof	-430.2	rf11	573.4
tpwr	58	xfp1	0
PW	12.200	PLOT	
GRADIENTS	wc	131.8	
gvlv11	1002	sc	6.2
gt1	0.001000	wc2	131.8
gstab	0.000500	sc2	0
DECOUPLER	vs	171	
dn	Cl3	th	7
dm	nm	ai	cdc
		av	





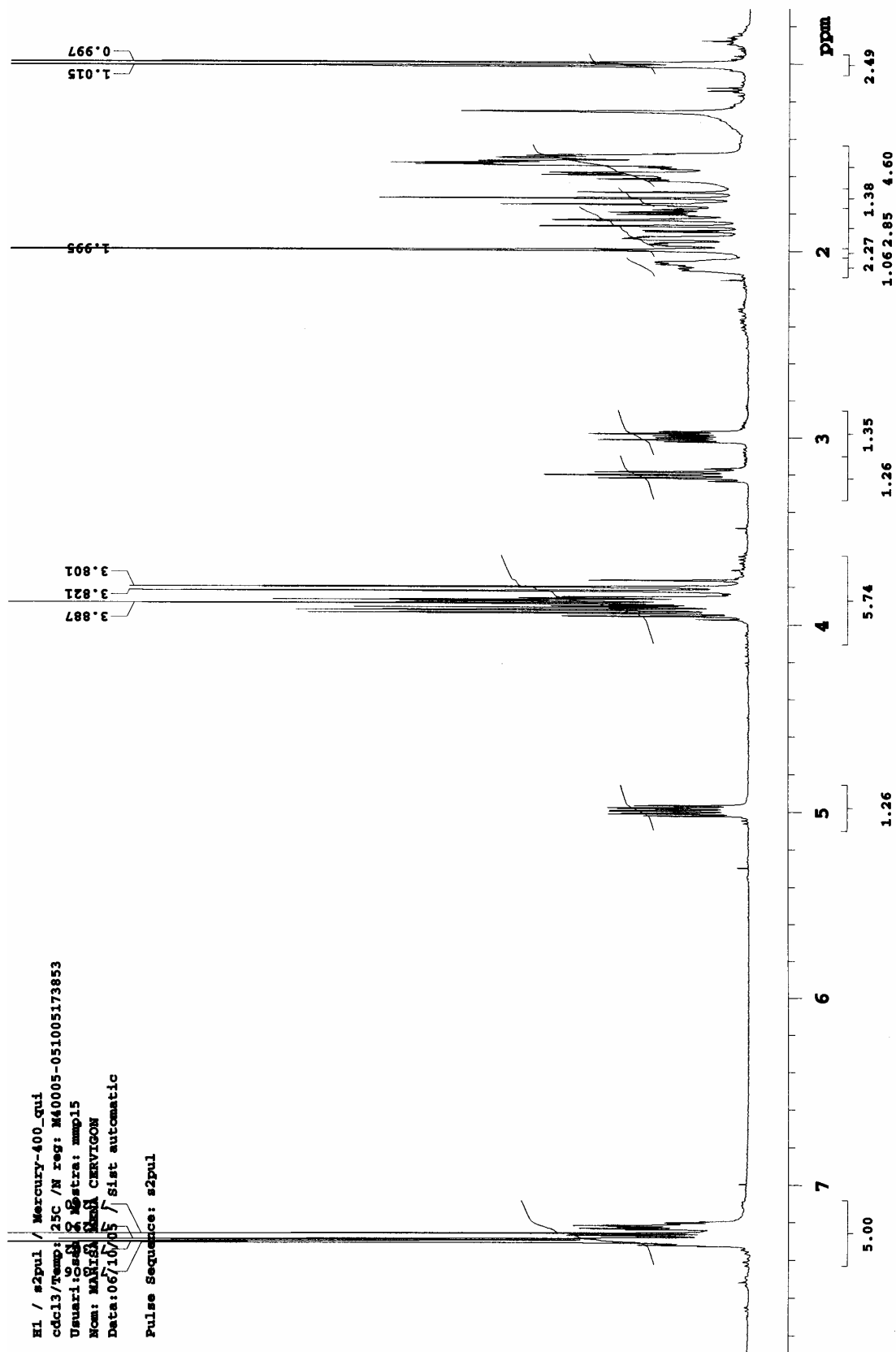
17

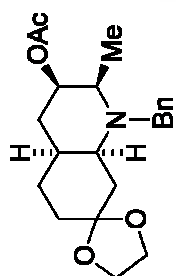




18

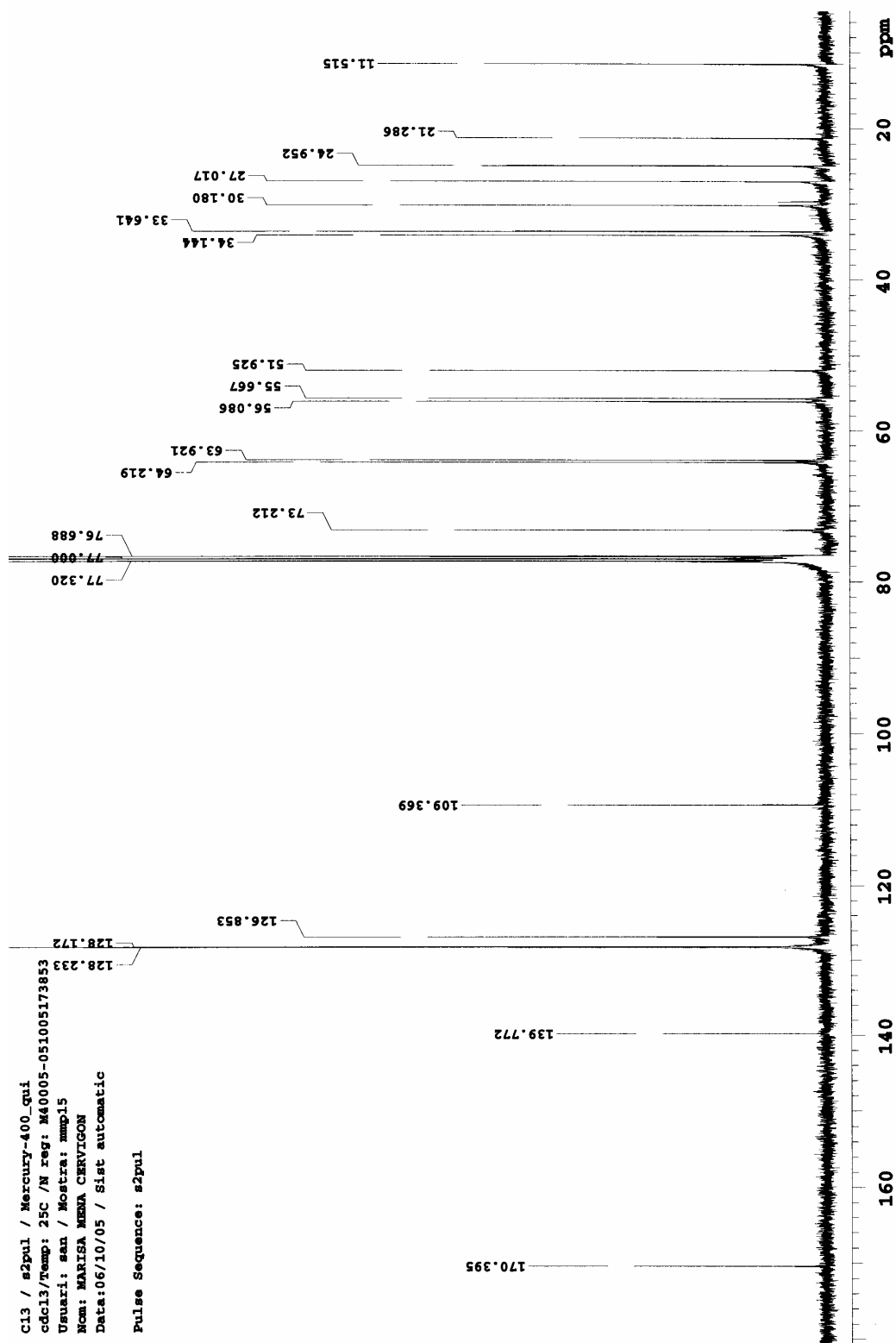
H1 / s2pul / Mercury-400_qui
 cdcl3/Temp: 25C /N reg: M40005-051005173853
 Usuari:Cs33 / 2 12estra: mmp15
 Nom: MARI6 / 2001 CERVIGON
 Data:06/16/05 / sist automatic
 Pulse Sequence: s2pul

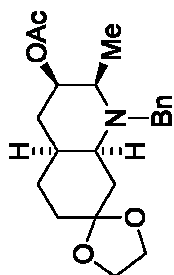




18

C13 / s2pul / Mercury-400.qui
 cdc13/Temp: 25C / N reg: M40005-051005173853
 Usuari: san / Mostra: mmp15
 Nom: MAPISA MEMA CERVIGON
 Data:06/10/05 / Sist automatic
 Pulse Sequence: s2pul



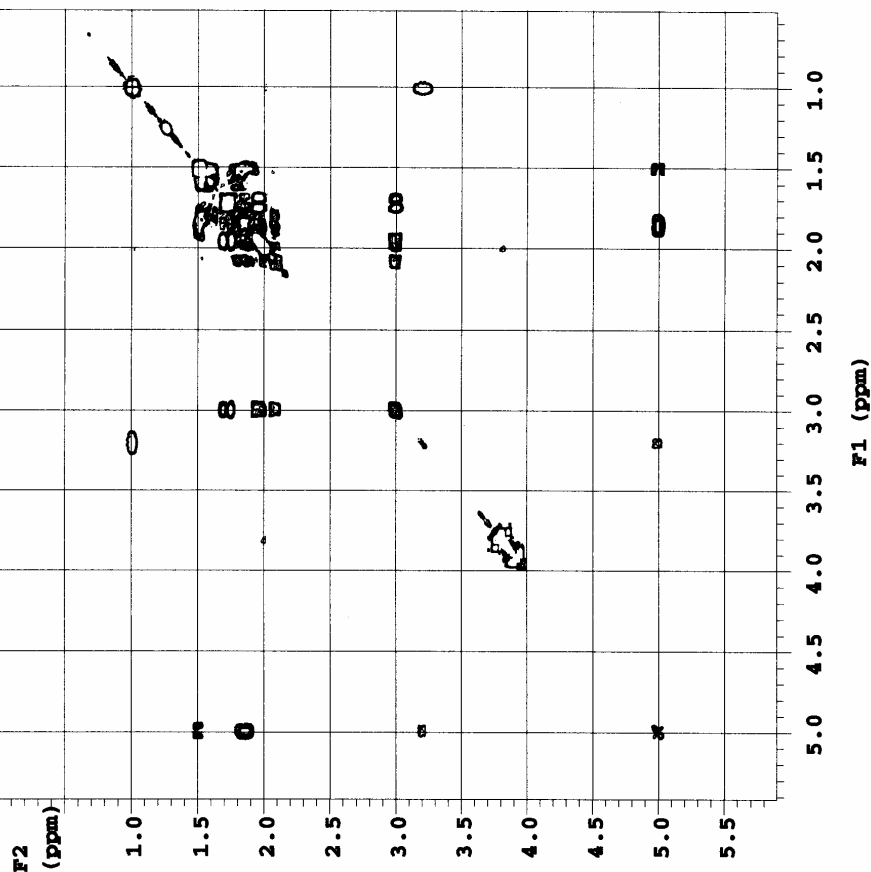


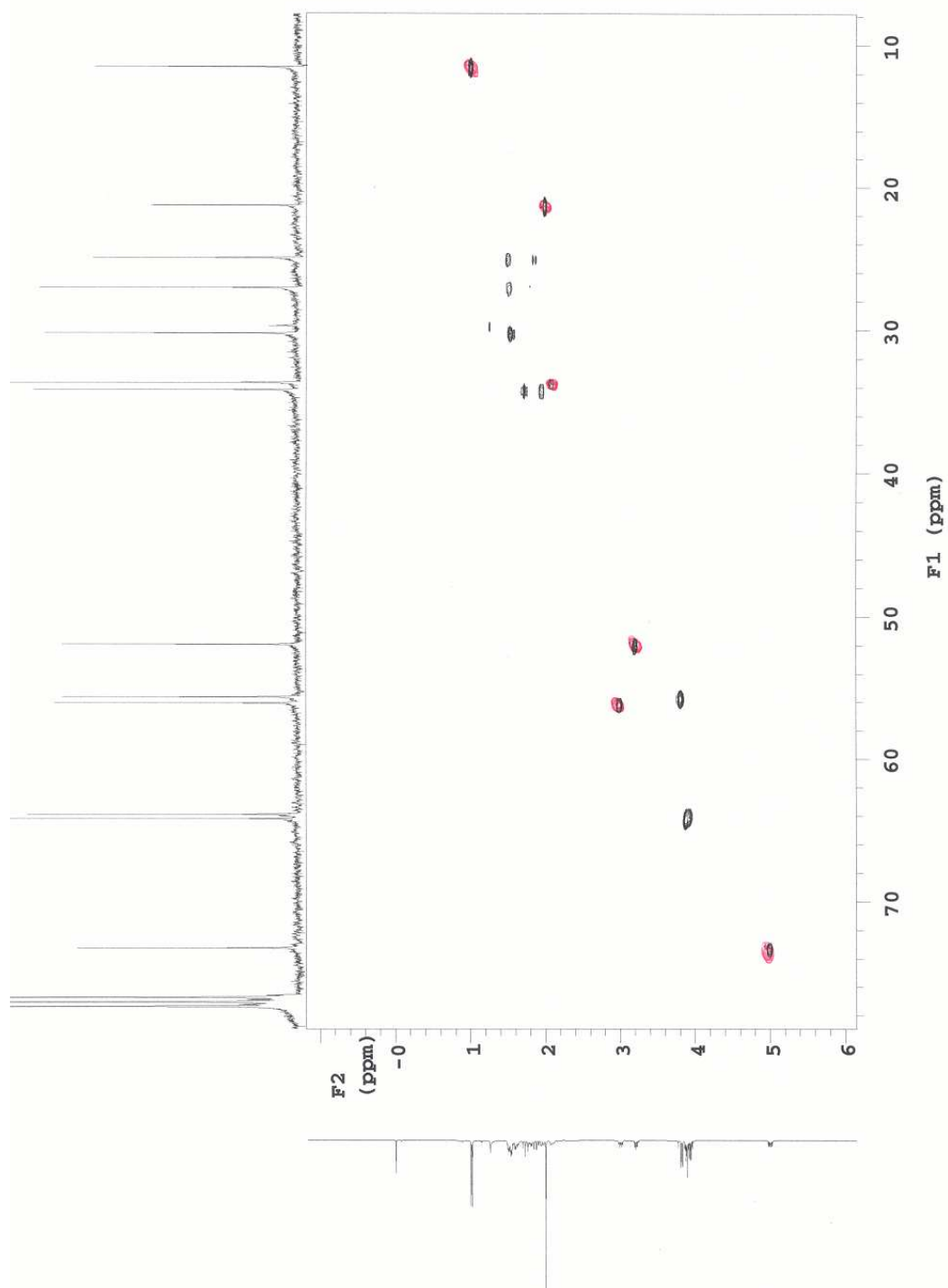
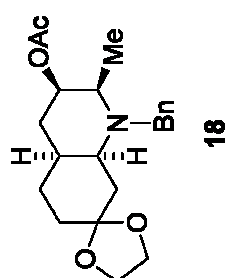
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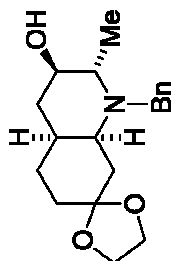
H1 / gCOSY / Mercury-400_qui
cdcl3/Temp: 25C / N reg: M40005-051005173
853
Usuari: san / Mostra: mmp15
Nom: MARISA NEMA CERVIGNON
Data:06/10/05 / Sist automatic

exp22 gCOSY

SAMPLE		FLAGS	
date	Oct 6 2005	hs	mn
solvent	cdcl3	sspul	n
sample	auto_Custom-hsglv1		1002
Q_06Oct2005	SPECIAL		
ACQUISITION			
sw	3937.0	gain	25.0
at	0.150	spin	0
np	1182	F2 PROCESSING	
fb	not used	sb	-0.075
ss	16	abs	not used
d1	1.000	fn	2048
nt	4	F1 PROCESSING	
2D ACQUISITION			
sw1	3937.0	sb1	-0.065
nd	256	proc1	lp
PRESATURATION			
satmode	n	DISPLAY	
satfrq	0	sp	-31.7
satdly	0	wp	2389.9
satpwr	0	sp1	210.7
TRANSMITTER			
tn	H1	rf1	1955.0
sfrq	400.113	rfp	485.8
tof	-540.4	xf11	485.8
tpwr	58	rfp1	0
PLOT			
pw	12.200	wc	131.8
GRADIENTS			
gslv11	1002	sc	6.2
gt1	0.001000	vc2	131.8
gstab	0.000500	sc2	0
DECOUPLER			
dn	C13	th	139
dm	nmn	ai	cdc
		av	6

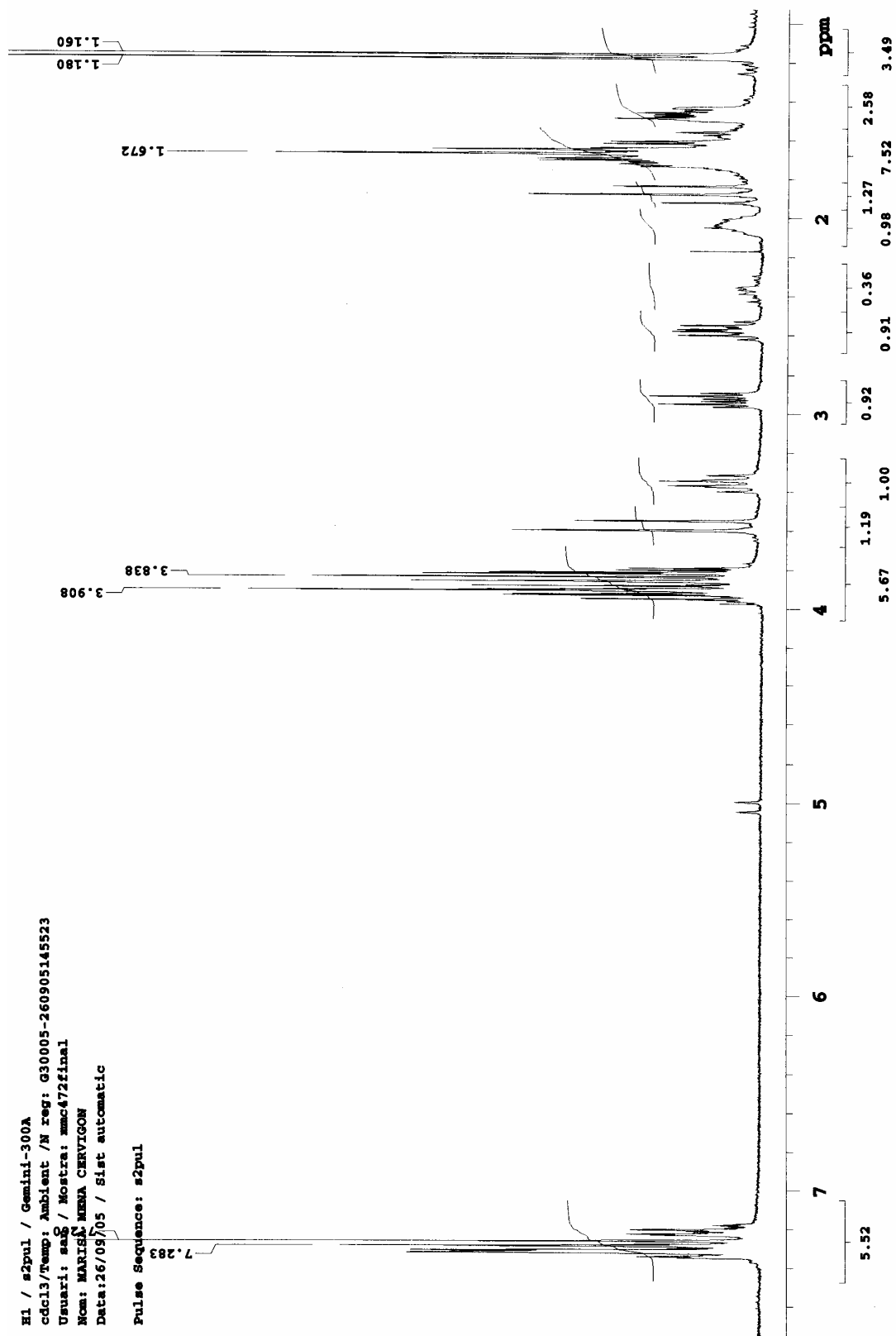


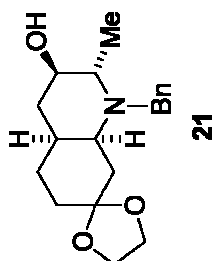




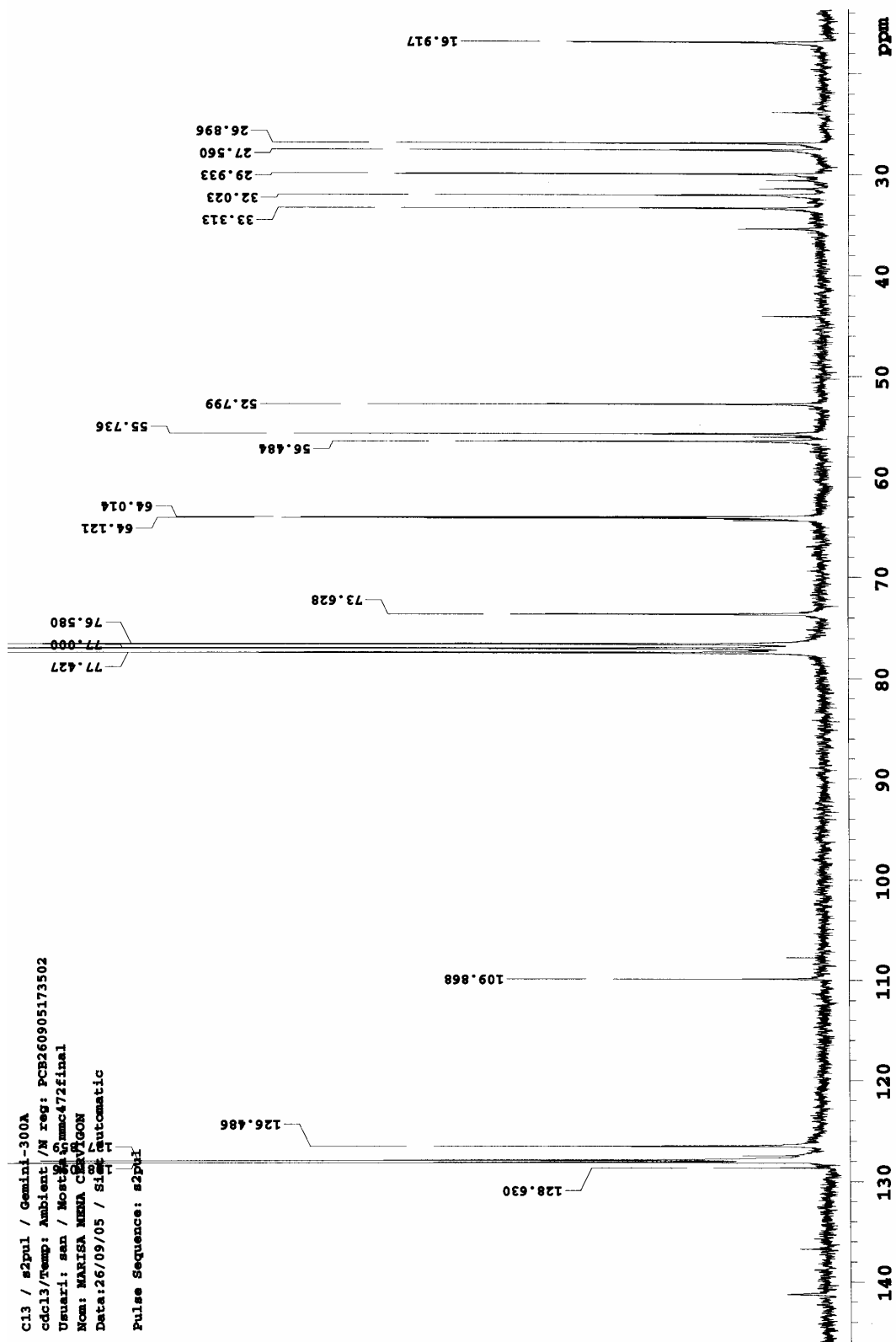
21

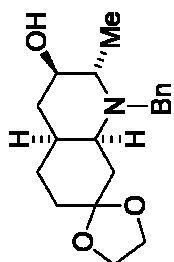
H1 / s2pul / Gemini-300A
 cdcl3/Temp: Ambient /N reg: G30005-260905145523
 Usuari: sas / Mostra: msc472final
 Nom: MARIS MESA CERVIGON
 Data:26/09/05 / Sist automatic
 Pulse Sequence: s2pul





C13 / s2pul / Gemini-300A
 cdcl3/Temp: Ambient / N reg: PCB260905173502
 Usuari: san / Mostre: 5 mmc472final
 Nom: WARISA MENA CAYUON
 Data: 26/09/05 / Shift automatic
 Pulse Sequence: s2pul





21

h1 / COSY / uemini-juva
cdcl3/Temp: Ambient /N reg: 030005-26090

5145523

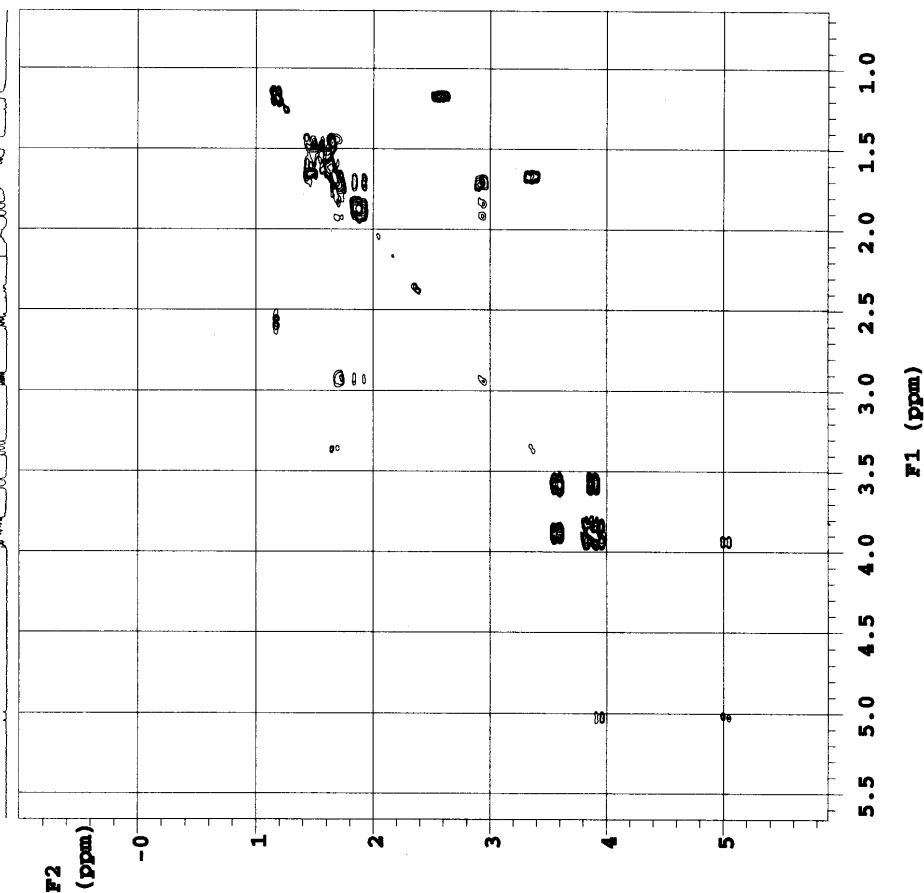
Usuari: san / Mostra: mmc472final

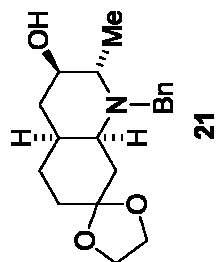
Nom: MARISA MENA CERVIGON

Data:26/09/05 / Sist automatic

exp7 COSY

SAMPLE		SPECIAL	
date	Sep 26 2005	temp	not used
solvent	cdcl3	gain	22
sample	auto_26Sep2- spin	0	
005 GRADIENTS			
ACQUISITION			
sv	2837.4	hsgl1	2000
at	0.180	hsgt	0.005000
np	1024	F2 PROCESSING	
fb	2250	ab	-0.090
ss	2	abs	not used
dl	1.000	fn	2048
nt	4	F1 PROCESSING	
2D ACQUISITION	abl	-0.090	
sw1	2837.4	abs1	not used
nl	256	procl	lp
TRANSMITTER	fn1	2048	
tn	h1	DISPLAY	
sfrq	300.046	sp	-299.4
tof	-397.5	wp	2066.3
tpwr	not used	sp1	191.5
pw	14.800	wp1	1506.0
PRESATURATION	rf1	316.1	
satmode	n	rfp	0
satpwr	0	xf11	316.1
satdly	0	rfp1	0
satfrq	0	PLOT	
DECOUPLER	wc	131.8	
dn	h1	sc	6.2
dm	nmn	wc2	131.8
FLAGS	sc2	0	
hs	nm	vs	4771
aspul	n	th	11
	ai	cdc	av





H1 / gHSQC / Mercury-400.qui
 cdc13/Temp: 25C /N reg: M40005-3561
 Usuari: san / Mostra: mmc472final
 Nom: MARISA MENA CERVIGON
 Data:24/09/05 / Sist automatic
 Pulse Sequence: gHSQC

