

Supporting information Available

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General Experimental Methods.

Melting points were determined using a hot-stage melting point apparatus and are uncorrected. Infrared spectra were recorded on a FT-IR spectrophotometer as potassium bromide disks of solids (KBr) or as thin films of liquids (neat) between sodium chloride plates. Nuclear magnetic resonance spectra (^1H , ^{31}P and ^{13}C NMR) were recorded on either 300 or 400 MHz spectrometers. ^1H and ^{13}C NMR assignments were confirmed using standard 2D COSY, NOESY, HSQC, and HMBC experiments. Low resolution electrospray ionisation (ESI) were recorded in the positive mode (ESI^+) on a QMS-quadrupole mass spectrometer. Accurate mass measurements were obtained at high resolution with a FTMS and a 4.7T superconducting magnet. The instrument was externally calibrated with FC5311. Analytical thin-layer chromatography (TLC) was performed on plastic slides coated with silica gel (Polygram SIL g/uv254). Flash chromatography was performed using Merck silica gel 60 (Merck no. 9385), 0.063-0.200 mm (230-400 mesh). Solvents were purified according to standard procedures. Chloroform used for optical rotations was of analytical purity. Degassed methanol and benzene were used in all hydrogenation reactions. Degassed dichloromethane was used in metathesis reactions. Deuterated chloroform was used as supplied. Deuterated dichloromethane was degassed by three freeze-pump-thaw cycles. Grubbs' catalyst refers to bis(tricyclohexylphosphine)benzylidene ruthenium dichloride.¹⁸ Second generation Grubbs' catalyst refers to tricyclohexylphosphine[1,3-bis(2,4,6-trimethylphenyl)-4,5-dihydroimidazol-2-ylidene][benzylidene]ruthenium dichloride.¹⁹ Wilkinson's catalyst refers to chlorotris(triphenylphosphine)rhodium (I).²³ Rh(I)-(S,S)-Et-DuPHOS refers to (+)-1,2-bis[(2S,5S)-2,5-diethylphospholano]benzene(1,5-cyclooctadiene)rhodium(I) tetrafluoroborate.²⁰ Rh(I)-(R,R)-Et-DuPHOS refers to (-)-1,2-bis[(2R,5R)-2,5-diethylphospholano]benzene(1,5-cyclooctadiene)rhodium(I) tetrafluoroborate.²⁰ All ruthenium-catalysed metathesis reactions were performed using standard Schlenk techniques under an atmosphere of nitrogen or in an argon filled drybox. Solvents were dried and degassed using standard procedures. In all rhodium-phosphine hydrogenations, high purity (<10 ppm of

oxygen) hydrogen and argon were used and purified by passage through a series of traps to remove water, oxygen and hydrocarbons. Ethylene and *cis*-2-butene were used as supplied. The enantiomeric excess of hydrogenation products was determined *via* analytical gas chromatography (GC) using a chiral column Model C-024 (column: 0.25 mm x 50 cm, 50CP2/XE60-SVALSAPEA) using helium as the carrier gas. Optical rotations were measured with a polarimeter (in a cell length of 1 dm) at a wavelength of 589 nm (sodium D line) at a temperature of 22°C.

(2R)-Methyl 2N-acetylaminopent-4-enoate (ent-4).¹⁶ (2Z)-Methyl 2N-acetylaminopenta-2,4-dienoate **3** (40.0 mg, 0.24 mmol), benzene (5 ml), [(COD)Rh(*R,R*)-Et-DuPHOS]OTf (3 mg), 30 psi H₂, 3 h, 88% yield, 96% ee (2*R*-**4**), *t*_R = 16.4 min (*R*) (GC chiral column Model C-024, 100°C for 1 min, 5°C min⁻¹ to 280°C for 9 min). [α]_D²² -43.0° (c = 0.47, CHCl₃). Spectral data were consistent with literature data.¹⁶

(2S)-Methyl 2N-benzoylaminopent-4-enoate (6).³² (2Z)-Methyl 2N-benzoylaminopenta-2,4-dienoate (100.0 mg, 0.43 mmol), benzene (8 ml), [(COD)Rh(*S,S*)-Et-DuPHOS]OTf (3 mg), 30 psi H₂, 3 h, 99% yield, 100% ee (2*S*-**6**), *t*_R = 27.0 min (GC chiral column Model C-024, 180°C for 1 min, 2°C min⁻¹ to 210°C for 20 min). [α]_D²² +49.3° (c = 1.12, CHCl₃). Spectral data were consistent with literature data.³²

(2R)-Methyl 2N-benzoylaminopent-4-enoate (ent-6).³² (2Z)-Methyl 2N-benzoylaminopenta-2,4-dienoate (100.0 mg, 0.43 mmol), benzene (8 ml), [(COD)Rh(*R,R*)-Et-DuPHOS]OTf (3 mg), 30 psi H₂, 3 h, 93% yield, 100% ee (2*R*-**6**), *t*_R = 26.4 min (GC chiral column Model C-024, 180°C for 1 min, 2°C min⁻¹ to 210°C for 20 min). [α]_D²² -49.7° (c = 0.64, CHCl₃). Spectral data were consistent with literature data.³²

(2S)-Methyl 2N-acetyl-amino-5-phenylpent-4-enoate (13).³⁰ (2Z)-Methyl 2N-acetyl-amino-5-phenylpenta-2,4-dienoate **12** (28.0 mg, 0.11 mmol), methanol (5 ml), [(COD)Rh(*S,S*)-Et-DuPHOS]OTf (1 mg), 75 psi H₂, 2 h, 97% yield, 99% ee (2*S*-**13**). [α]_D²² +90.0° (c = 0.64, CHCl₃). Spectral data were consistent with literature data.³⁰

(2R)-Methyl 2N-acetylamino-5-phenylpent-4-enoate (ent-13).³⁰ (2Z)-Methyl 2N-acetylamino-5-phenylpenta-2,4-dienoate **12** (27.4 mg, 0.11 mmol), methanol (5 ml), [(COD)Rh(*R,R*)-Et-DuPHOS]OTf (1 mg), 75 psi H₂, 2 h, 92% yield, 99% ee (**2R-13**). $[\alpha]_D^{22} -89.8^\circ$ (*c* = 1.03, CHCl₃). Spectral data were consistent with literature data.³⁰

(2S)-Methyl 2N-acetylamino-5-methylhex-4-enoate (15). (2Z)-Methyl 2N-acetylamino-5-methylhexa-2,4-dienoate **16** (74.0 mg, 0.38 mmol), methanol (5 ml), [(COD)Rh(*S,S*)-Et-DuPHOS]OTf (2 mg), 75 psi H₂, 2 h, 98% yield, 100% ee (**2S-15**), *t_R* = 24.2 min (GC chiral column Model C-024, 100°C for 1 min, 5°C min⁻¹ to 210°C for 7 min). $[\alpha]_D^{22} +58.2^\circ$ (*c* = 0.79, CHCl₃), m.p. 46-48°C. ν_{\max} (neat): 3288m, 2955w, 1746s, 1660s, 1538m, 1436m, 1377m, 1274w, 1210w, 1126w, 1030w, 736w cm⁻¹. ¹H NMR (300 MHz, CDCl₃): δ 1.59 (s, 3H, H6a), 1.69 (d, *J* 0.9 Hz, 3H, H6b), 2.00 (s, 3H, CH₃CO), 2.39-2.60 (m, 2H, H3), 3.72 (s, 3H, OCH₃), 4.63 (dt, *J* 7.9, 5.6 Hz, 1H, H2), 4.99 (t, *J* 7.5 Hz, 1H, H4), 6.02 (bs, 1H, NH). ¹³C NMR. (100 MHz, CDCl₃): δ 18.0, 26.0, ((CH₃)₂), 23.3 (CH₃CO), 30.8 (C3), 52.2, 52.4 (C2, OCH₃), 117.6 (C4), 136.6 (C5), 169.8, 172.8 (C1, CONH). HRMS (ESI⁺, MeOH): *m/z* calcd for C₁₀H₁₇NO₃Na [(M+Na)⁺] 222.1106, found 222.1105.

(2R)-Methyl 2N-acetylamino-5-methylhex-4-enoate (ent-15). (2Z)-Methyl 2N-acetylamino-5-methylhexa-2,4-dienoate **16** (24.5 mg, 0.12 mmol), methanol (5 ml), [(COD)Rh(*R,R*)-Et-DuPHOS]OTf (1 mg), 75 psi H₂, 2 h, 100% yield, 100% ee (**2R-15**), *t_R* = 23.9 min (GC chiral column Model C-024, 100°C for 1 min, 5°C min⁻¹ to 210°C for 7 min). $[\alpha]_D^{22} -58.3^\circ$ (*c* = 0.53, CHCl₃). Spectral data were consistent with that previously obtained.

(2S,7S)-Dimethyl 2,7-*N,N'*-dibenzoylamino-oct-4-enedioate (7). (2S)-Methyl 2N-benzoylaminopent-4-enoate **6** (49.0 mg, 0.21 mmol), dichloromethane (4 ml), 2nd generation Grubbs' **S5**

catalyst (18.0 mg, 0.02 mmol, 10 mol%), 12 h, 50°C, 100% conversion. Purification by flash chromatography (SiO₂, dichloromethane : light petroleum : ethyl acetate, 1:1:1) gave pure dimer **7** (32.8 mg, 36%) as a pale brown solid, m.p. 140-142°C. t_R (*E/Z*) = 13.5, 13.9 min (GC column 30QC5/BPX5), 150°C for 1 min, 10°C min⁻¹ to 280°C for 6 min). $[\alpha]_D^{22} +56.4^\circ$ (*c* = 0.27, CHCl₃). ν_{max} (neat): 3322bm, 2953m, 2358w, 1742s, 1644s, 1603w, 1580w, 1538m, 1488m, 1436m, 1267w, 1218m, 1027w, 973w, 802w, 736m cm⁻¹. ¹H NMR (300 MHz, CDCl₃): δ 2.57-2.69 (m, 4H, H3,6), 3.67 (s, 6H, OCH₃), 4.85-4.98 (m, 2H, H2,7), 5.49 (t, *J* 4.1 Hz, 2H, H4,5), 6.86 (bd, *J* 7.4 Hz, 2H, NH), 7.40-7.44 (m, 4H, H3',5'), 7.48-7.52 (m, 2H, H4'), 7.81-7.83 (m, 4H, H2',6'). ¹³C NMR (100 MHz, CDCl₃): δ 35.2 (C3,6), 52.5 (C2,7), 52.6 (OCH₃), 127.2 (C2',6'), 128.7 (C3',5'), 128.8 (C4,5), 131.9 (C4'), 133.9 (C1'), 167.1, 172.4 (C1,8, CONH). HRMS (ESI⁺, MeOH): *m/z* calcd for C₂₄H₂₆N₂O₆Na [(M+Na)⁺] 461.1689, found 461.1695.

Attempted dimerisation of (2*S*)-methyl 2*N*-acetylamino-5-phenylpent-4-enoate (13**).** *Trial 1:* (2*S*)-Methyl 2*N*-acetylamino-5-phenylpent-4-enoate **13** (59.3 mg, 0.24 mmol), dichloromethane (10 ml), Grubbs' catalyst (19.8 mg, 0.02 mmol, 10 mol%), 13 h, 50°C, 0% conversion of **13** into dimer **5**. *Trial 2:* (2*S*)-Methyl 2*N*-acetylamino-5-phenylpent-4-enoate **13** (59.3 mg, 0.24 mmol), dichloromethane (7 ml), 2nd generation Grubbs' catalyst (10.2 mg, 0.01 mmol, 5 mol%), 20 h, 50°C, 33% conversion of **13** into dimer **5**.

Dimerisation of (2*S*)-Methyl 2*N*-acetylaminohex-4-enoate (17**).** (2*S*)-Methyl 2*N*-acetylaminohex-4-enoate **17** (17.0 mg, 0.09 mmol), dichloromethane (4 ml), 2nd generation Grubbs' catalyst (4.2 mg, 0.005 mmol, 5 mol%), 17 h, 50°C, 100% conversion into dimer **5**. Spectroscopic data were in agreement with that previously obtained.

Cross metathesis of (2*S*)-methyl 2*N*-acetylaminopent-4-enoate (4**) in the presence of (2*S*)-methyl 2*N*-acetylaminopenta-2,4-dienoate (**3**).** (2*S*)-Methyl 2*N*-acetylaminopent-4-enoate **4** (34.0 mg, 0.20 mmol), (2*Z*)-methyl 2*N*-acetylaminopenta-2,4-dienoate **3** (33.6 mg, 0.20 mmol), dichloromethane (4 ml), Grubbs' catalyst (16.3 mg, 0.02 mmol, 10 mol%), 18 h, 50°C. The ¹H NMR spectrum displayed peaks characteristic of the starting allylglycine derivative **4** and dienamide **3** but no peaks characteristic of expected dimer **5**. The mass spectrum displayed peaks attributed to allylglycine derivative **4** and tricyclohexylphosphine-dienamide conjugate addition adduct. Mass Spectrum (ESI⁺, DCM/MeOH): *m/z* 194.1 C₈H₁₃NO₃Na [(M+Na)⁺], **4**. *m/z* 450.4 C₂₆H₄₅NO₃P⁺ [M⁺], tricyclohexylphosphine-dienamide adduct.

Cross metathesis of (2*S*)-methyl 2*N*-acetylaminopent-4-enoate (4**) in the presence of (2*S*)-methyl 2*N*-acetylaminopenta-2,4-dienoate (**12**).** (2*S*)-Methyl 2*N*-acetylaminopent-4-enoate **4** (18.1 mg, 0.11 mmol), (2*Z*)-methyl 2*N*-acetylaminopenta-2,4-dienoate **12** (26.1 mg, 0.11 mmol), dichloromethane (4 ml), Grubbs' catalyst (8.7 mg, 0.01 mmol, 10 mol%), 18 h, 50°C, 28% conversion of **4** into dimer **5**. Dienamide **12** did not react under these conditions.

Cross metathesis of (2*S*)-Methyl 2*N*-acetylaminopent-4-enoate (4**) in the presence of (2*S*)-Methyl 2*N*-acetylaminopenta-2,4-dienoate (**15**).** (2*S*)-Methyl 2*N*-acetylaminopent-4-enoate **4** (12.7 mg, 0.07 mmol), (2*S*)-methyl 2*N*-acetylaminopenta-2,4-dienoate **15** (12.0 mg, 0.07 mmol), dichloromethane (4 ml), Grubbs' catalyst (11.5 mg, 0.01 mmol, 20 mol%), 18 h, 50°C, 100% conversion of **4** into dimer **5**. **15** did not react under these conditions.

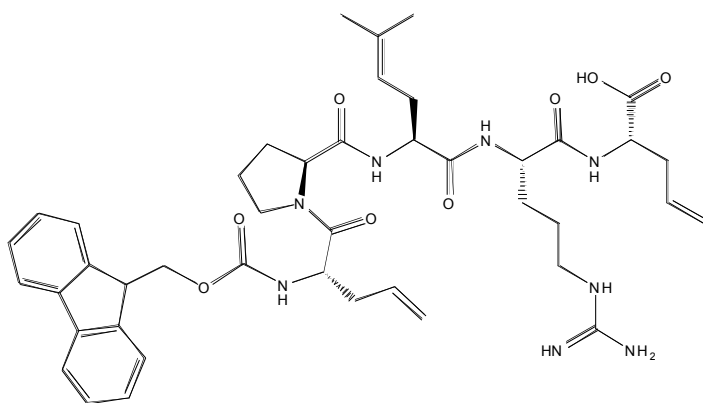
Wilkinson's hydrogenation of Dimer (7). (2*S*,7*S*)-Dimethyl 2,7-*N,N'*-dibenzoylamino-octanedioate (**18**). (2*S*,7*S*)-Dimethyl 2,7-*N,N'*-dibenzoylamino-octa-4-enedioate **7** (20.0 mg, 0.05

mmol), benzene (5 ml), Wilkinson's catalyst (2 mg), 50 psi H₂, 4 h, 100% of **18** as a brown oil. $t_R = 17.2$ min (GC: Column 30QC5/BPX5, 150°C for 1 min, 10°C min⁻¹ to 280°C for 6 min). ν_{\max} (neat): 3055m, 2986w, 2955w, 1741s, 1662s, 1603w, 1580w, 1518m, 1486m, 1438s, 1359w, 1286s, 1182m, 1120m, 1028w, 896m cm⁻¹. ¹H NMR (300 MHz, CDCl₃): δ 1.35-1.53 (m, 4H, H4,5), 1.80-2.02 (m, 4H, H3,6), 3.78 (s, 3H, OCH₃), 4.82 (dt, J 7.4, 5.3 Hz, 1H, H2,7), 6.73 (bd, J 7.7 Hz, 2H, NH), 7.40-7.49 (m, 6H, H3',4',5'), 7.78-7.82 (m, 4H, H3',5'). ¹³C NMR (100 MHz, CDCl₃): δ 24.9 (C4,5), 32.6 (C3,6), 52.5, 52.7 (C2, OCH₃), 127.2 (C2',6'), 128.6 (C3',5'), 131.9 (C4'), 134.1 (C1'), 167.2, 173.2 (C1,8, CONH). HRMS (ESI⁺, MeOH): m/z calcd for C₂₄H₂₈N₂O₆Na [(M+Na)⁺] 463.1845, found: 463.1842

Wilkinson's hydrogenation of (2S)-methyl 2N-acetylamino-5-phenylpenta-2,4-dienoate (12).

(2S)-Methyl 2N-acetylamino-5-phenylpenta-2,4-dienoate **12** (11.5 mg, 0.05 mmol), benzene (5 ml), Wilkinson's catalyst (1 mg), 50 psi H₂, 4 h, 99% yield of a 1:4 mixture of 13:14 as a brown oil. This mixture was subjected to the hydrogenation conditions previously described which led to 100% conversion into **14**.³⁴. $t_R = 10.8$ min (GC column 30QC5/BPX5, 150°C for 1 min, 10°C min⁻¹ to 280°C for 6 min). ν_{\max} (neat): 3054m, 2956m, 1736m, 1676m, 1509w, 1438m, 1372w, 1265s, 1174w, 1120m, 1028w, 738s cm⁻¹. ¹H NMR (300 MHz, CDCl₃): δ 1.53-1.65 (m, 4H, H3,4), 1.94 (s, 3H, CH₃CO), 2.52-2.59 (m, 2H, H5), 3.65 (s, 3H, OCH₃), 4.54-4.63 (m, 1H, H2), 5.90 (bd, J 7.2 Hz, 1H, NH), 7.07-7.29 (m, 5H, AromCH). ¹³C NMR (75 MHz, CDCl₃): δ 23.3 (CH₃CO), 27.2 (C4), 32.3 (C3), 35.5 (C5), 52.1, 52.5 (C2, OCH₃), 126.1, 128.5, 132.2 (Arom CH), 141.7, (Arom C), 169.9, 173.2 (C1, CONH). Mass Spectrum (ESI⁺, MeOH): m/z 272.2 C₁₄H₁₉NO₃Na [(M+Na)⁺].

Linear Pentapeptide: Fmoc-Hag-Pro-Pre-Arg-Hag-OH



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The procedure outlined in the General Section was used to attach the first amino acid, Fmoc-Hag-OH, to Wang resin. Quantities of the resin and coupling reagents are presented in Table 1. The first coupling reaction was shaken for 14 h.

Table 1 Quantities of Reagents used in the Synthesis of Fmoc-Hag-Pro-Pre-Arg-Hag-OH

<i>Reagent</i>	<i>Mass (mg) or Volume (μl)</i>	<i>Mole (mmol)</i>
Wang Resin	212 mg	0.19
Fmoc-L-Hag-OH	195 mg	0.58
DIC	90.6 μl	0.58
DMAP	7.1 mg	0.06

The procedure outlined in the General Section was used for subsequent coupling reactions in the synthesis of pentapeptide Fmoc-Hag-Pro-Pre-Arg-Hag-OH. Quantities of the coupling reagents HATU and NMM are tabulated below (Table 2) and remained constant throughout the synthesis. The quantities of successive amino acids and their reaction durations are detailed in Table 3.

Table 2 Quantities of Coupling Reagents used in the Synthesis of Fmoc-Hag-Pro-Pre-Arg-Hag-OH

<i>Coupling Reagent</i>	<i>Mass (mg) or Volume (ml)</i>	<i>Mole (mmol)</i>
HATU	147 mg	0.39
NMM	128 μ l	1.16

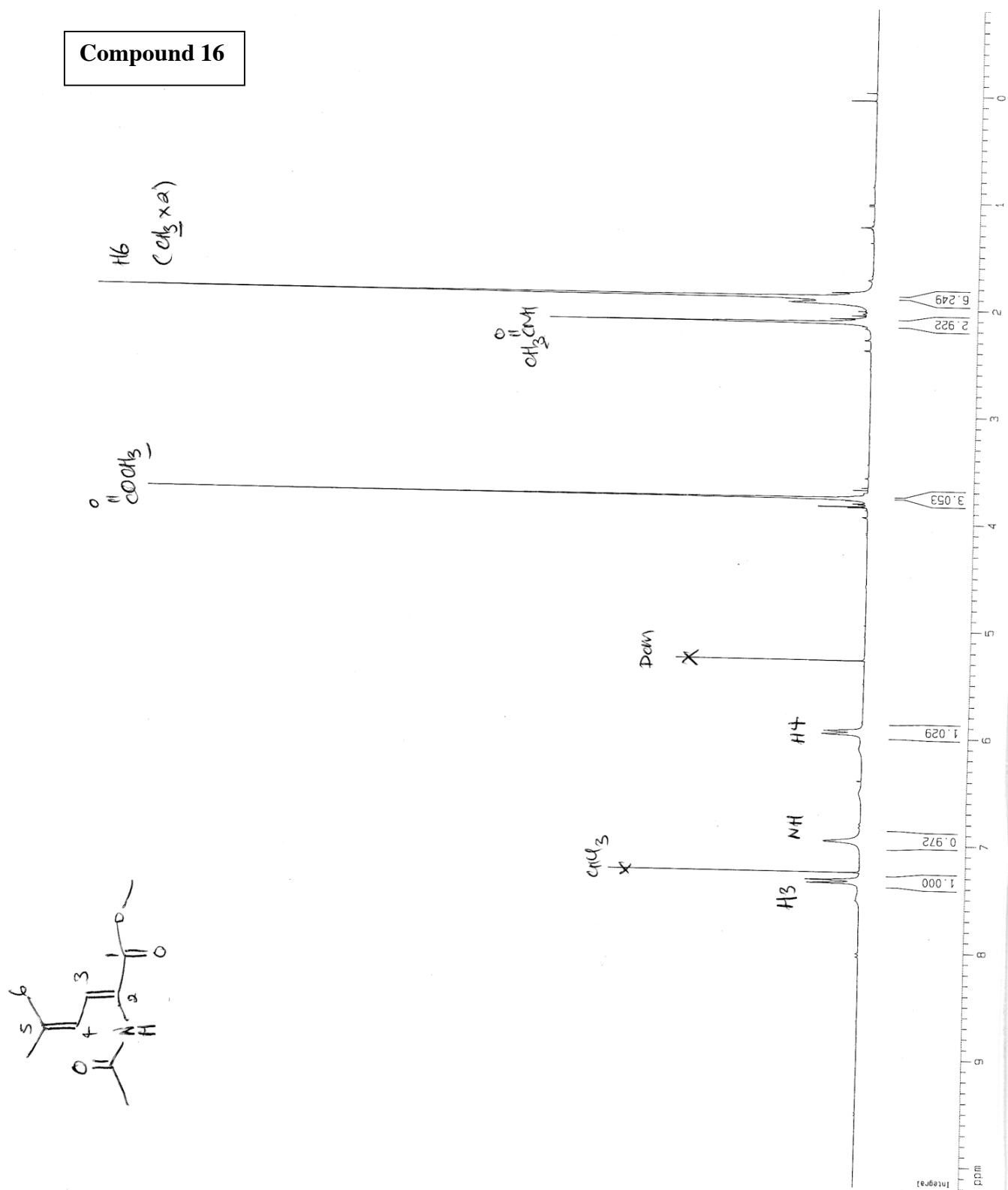
Table 3 Quantities of Amino Acids used in the Synthesis of Fmoc-Hag-Pro-Pre-Arg-Hag-OH

<i>Amino Acid</i>	<i>Mass (mg)</i>	<i>Mole (mmol)</i>	<i>Reaction Time (h)*</i>
Fmoc-L-Arg(Pbf)-OH	376	0.58	2
Fmoc-L-Pre-OH	211	0.58	3
Fmoc-L-Pro-OH	196	0.58	6
Fmoc-L-Hag-OH	195	0.58	2

* *Note:* Reaction times have not been optimised.

After the final amino acid coupling, a small aliquot of peptidyl-resin was subjected to the TFA-mediated cleavage procedure described in the General Section. Mass spectral analysis of the isolated residue confirmed formation of the linear pentapeptide (**19**). Mass spectrum (ESI⁺, MeCN/H₂O): m/z 813.6 (M+H)⁺, C₄₃H₅₇N₈O₈.requires 813.4; m/z 831.5 (M+H₂O+H)⁺, C₄₃H₅₉N₈O₉ requires 831.4; m/z 927.6 (M+TFA+H)⁺, C₄₅H₅₈ F₃N₈O₁₀ requires 927.4.

Compound 16



Compound 16

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300CS/BPX-5 1.0um

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IF

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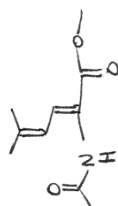
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min



JE-6-101-1300

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Compound 4

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16.605

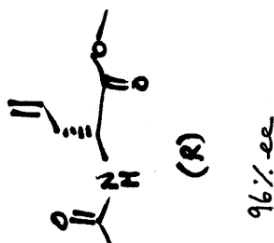
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Total area = 1.096E+007



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 1.6e6
 1.4e6
 1.2e6
 1.0e6
 8.0e5
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Area Percent Report

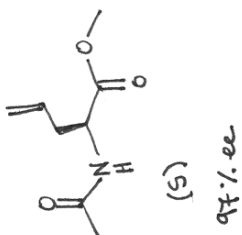
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Total area = 1646270

Compound 4



min

8.0e5

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5.0e5

4.0e5

3.0e5

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1.0e5

0

5

10

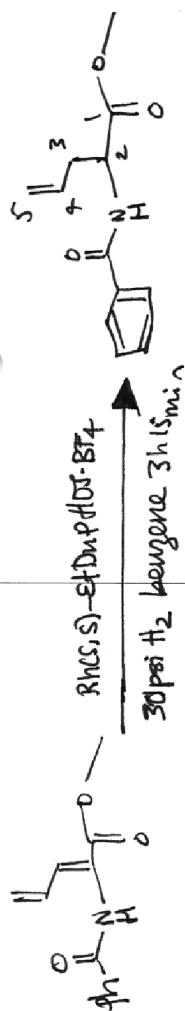
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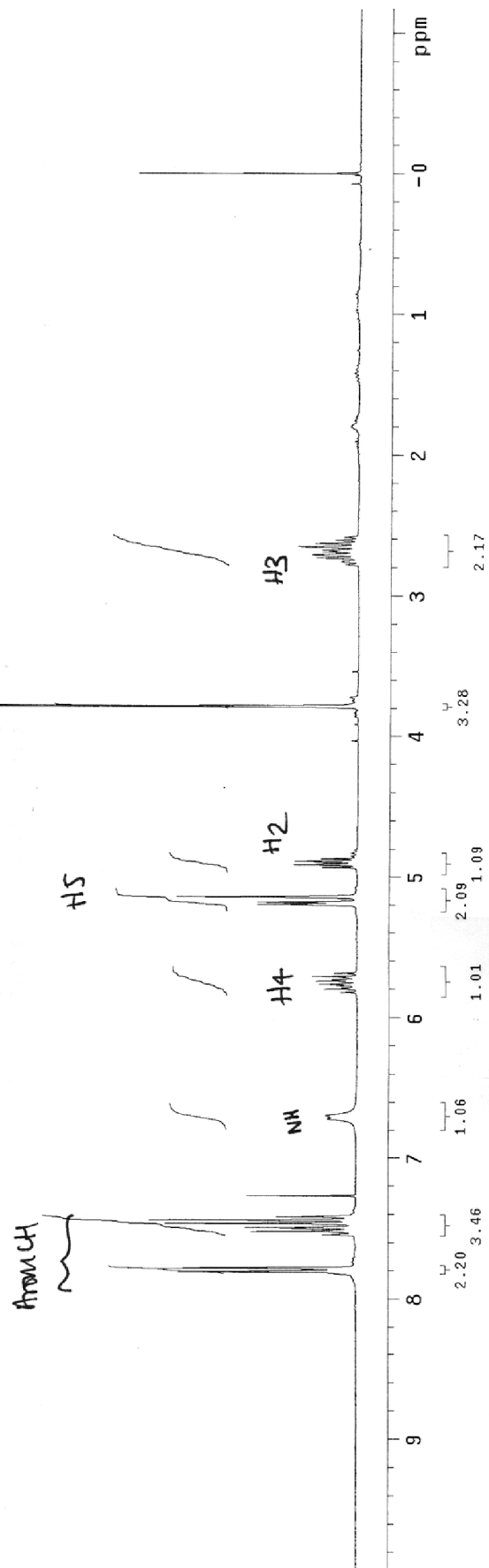
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Compound 6

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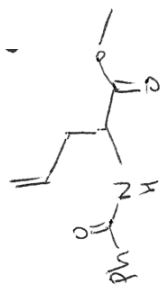


100% conversion



Compound 6

JE-6-146

From Rh(R,R)-DuPHOS- BF_4

210°C (20 min)

180°C

1 min

DCM (solvent)

JE-69b.

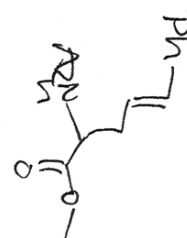
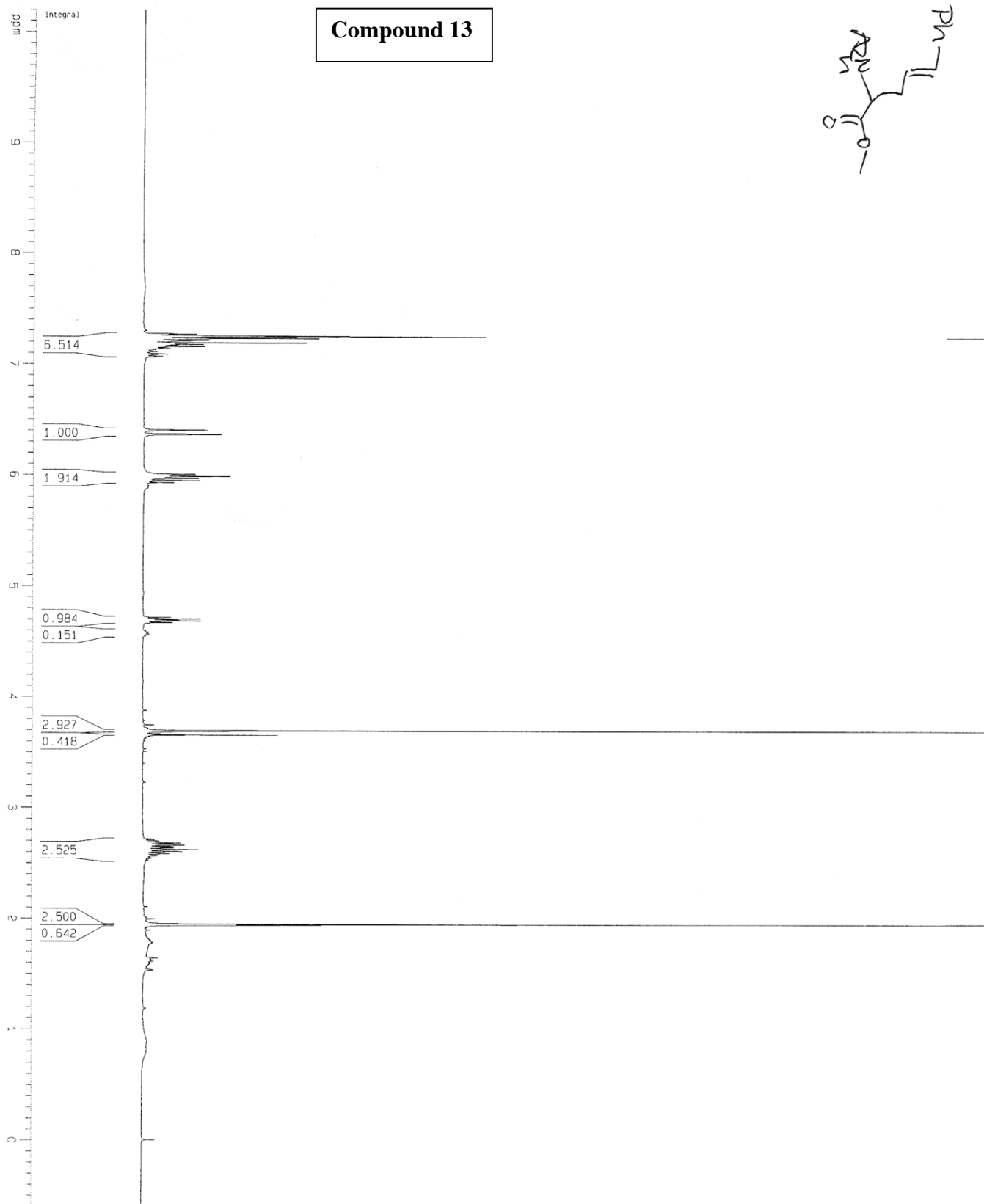


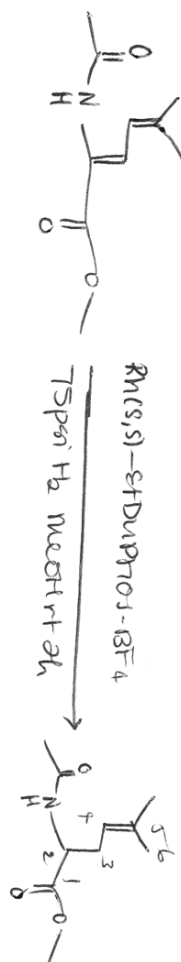
From (S, S) to (S, S)

$$\frac{1800}{200 \text{ min}^{-1}} = 9 \text{ min}$$

DCM (solvent)

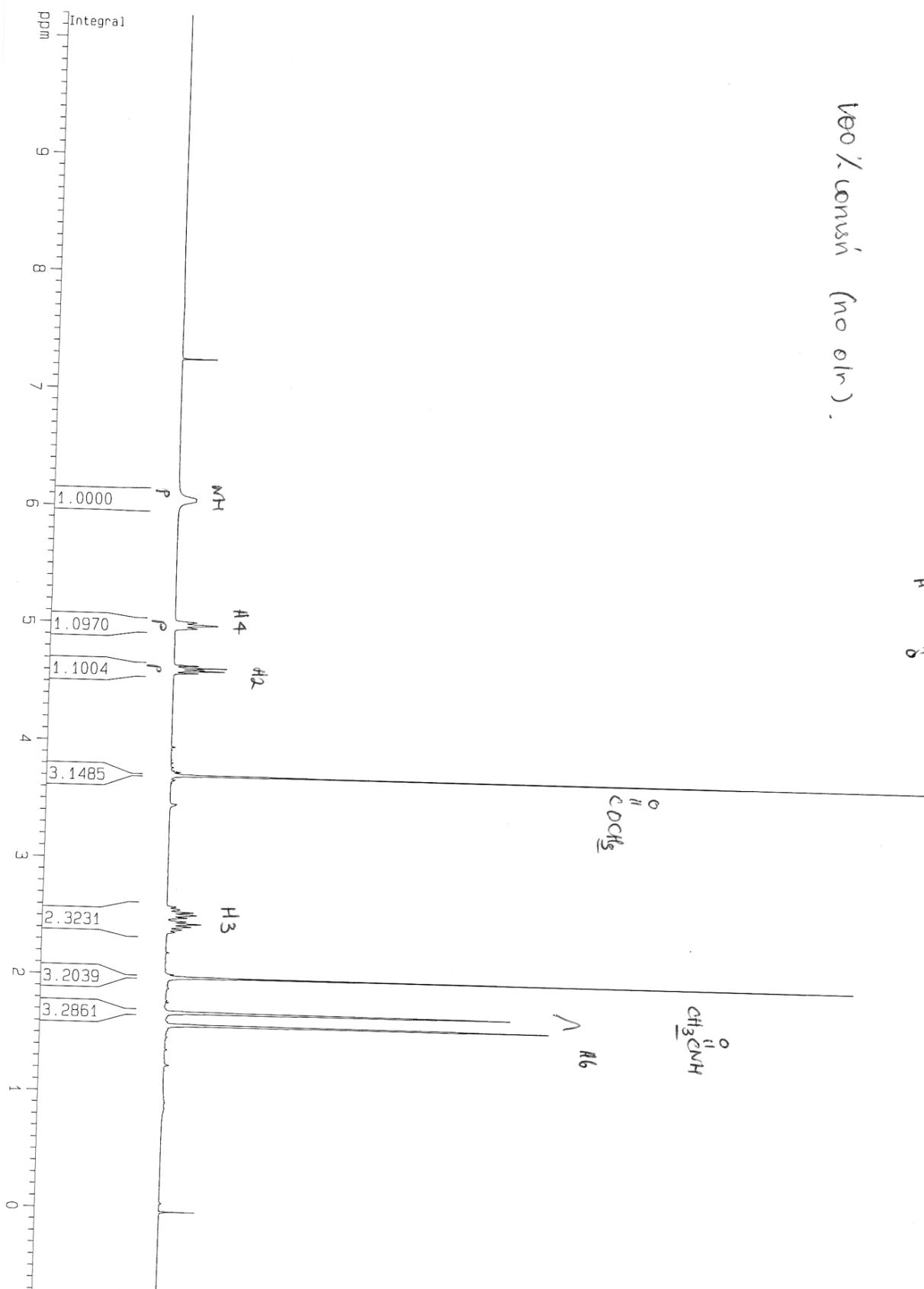
10

**Compound 13**

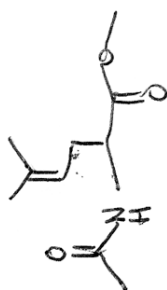


100% conversion (no dimer).

Compound 15



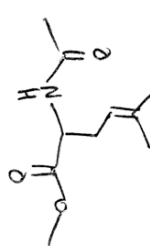
Compound 15

JE-6-133B Rh(S,S)-EtDUPHOS · BF₄ 24.2100°C
1 hr
210°C (7 min)
5°C/min

100% ee

100% ee

Compound 15



160°C 50 min
1 min
200°C (1 min)

JE-6-135B
(2R,3R)-2-((2S,3S)-3-methyl-2-penten-3-yl)-3-methylbutanamide

23.9

TIMEABLE STOP

C16=100 0.00001 0.010 H-003660004.END

RUN# 00 MAY 27, 1984 10:26:11

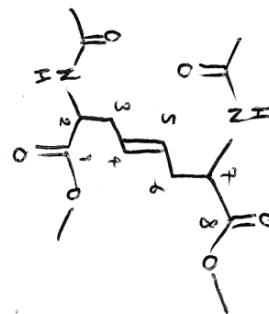
SIGNAL FILE: H-003660004.END

AREA

RT	AREA	TYPE	WIDTH	AREA
4.483	7120	FB	.000	.00000
4.000	145000	FB	.000	.00000
20.077	600000	FB	.044	.04400

TOTAL AREA=1.17600E+00

MUL FACTOR=1.00000E+00



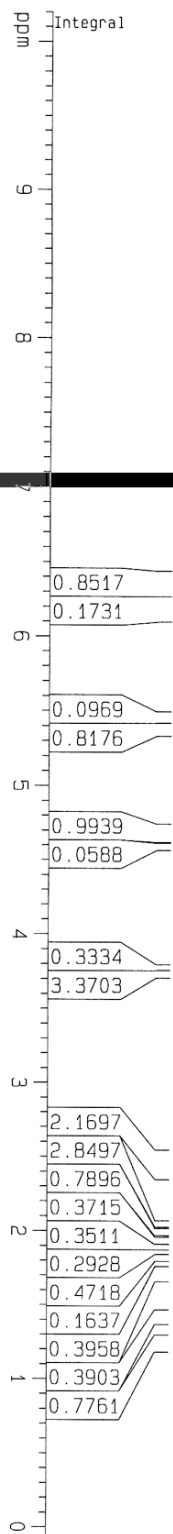
pure product!

Compound 5

JE-6-73-F7+8

Integral

ppm



COCH_3

$\text{CH}_3\text{C(=O)NH}$

H_{3,6}

H_{4,5}

H_{2,7}

NH

Compound 5
Area Percent Report

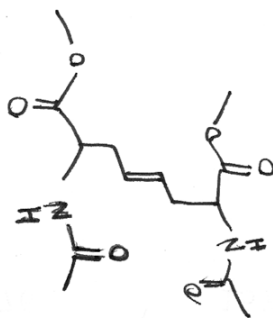
Data File Name : C:\HPCHEM\1\DATA\JE7.D
 Operator :
 Instrument : HP5890A
 Sample Name :
 Run Time Bar Code:
 Acquired on : NOV 14, 1904 23:02:42
 Report Created on: 15 Nov 04 00:36 AM

Page Number : 1
 Vial Number : 0
 Injection Number :
 Sequence Line :
 Instrument Method:
 Analysis Method : GC1.MTH

Sig. 1 in C:\HPCHEM\1\DATA\JE7.D

Pk#	Ret Time	Area	Height	Type	Width	Area %
1	12.483	322864	109482	BV	0.044	67.1459
2	12.676	157975	45524	VB	0.049	32.8541

Ac-Hag-divine



2.0e5

1.8e5

1.6e5

1.4e5

1.2e5

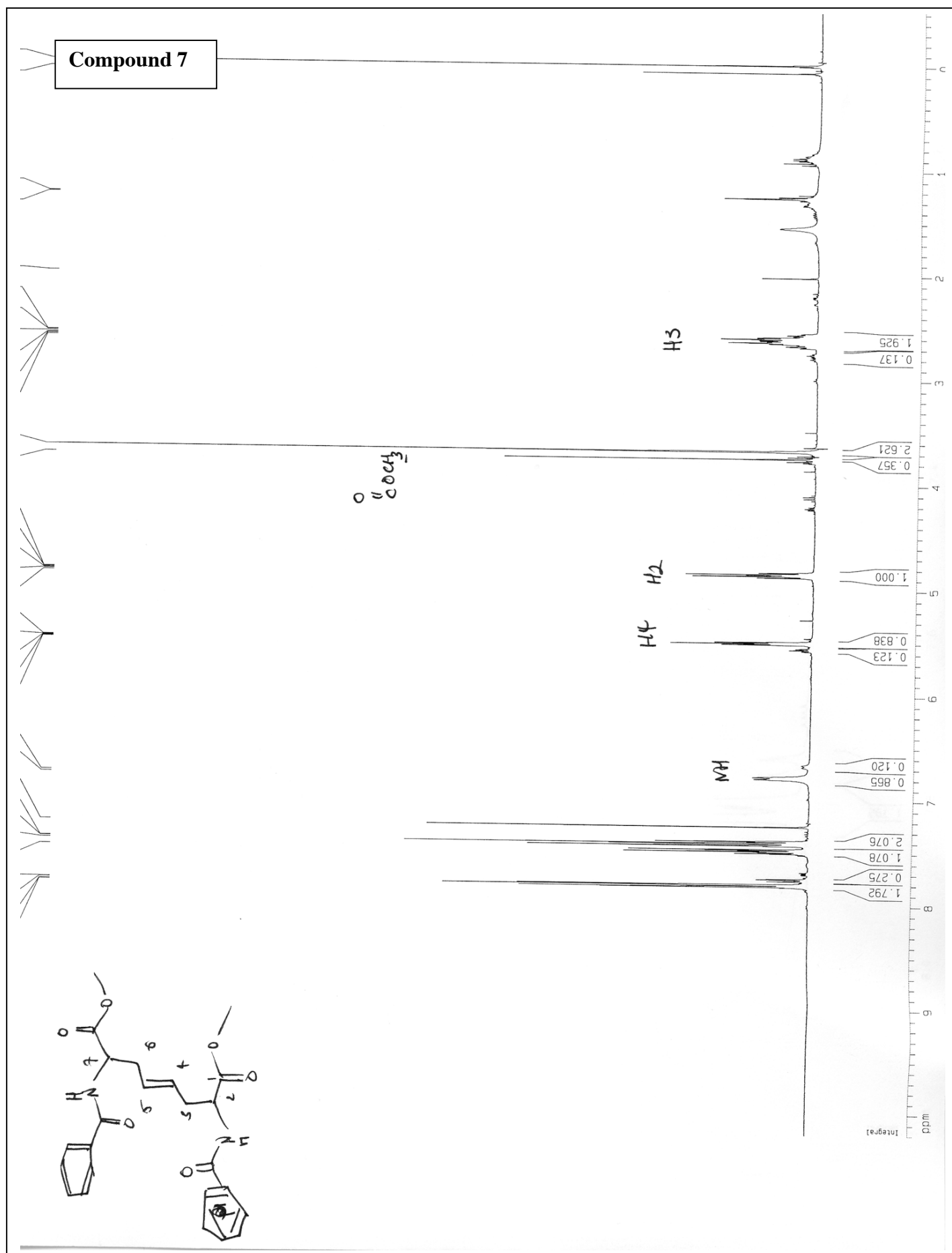
1.0e5

8.0e4

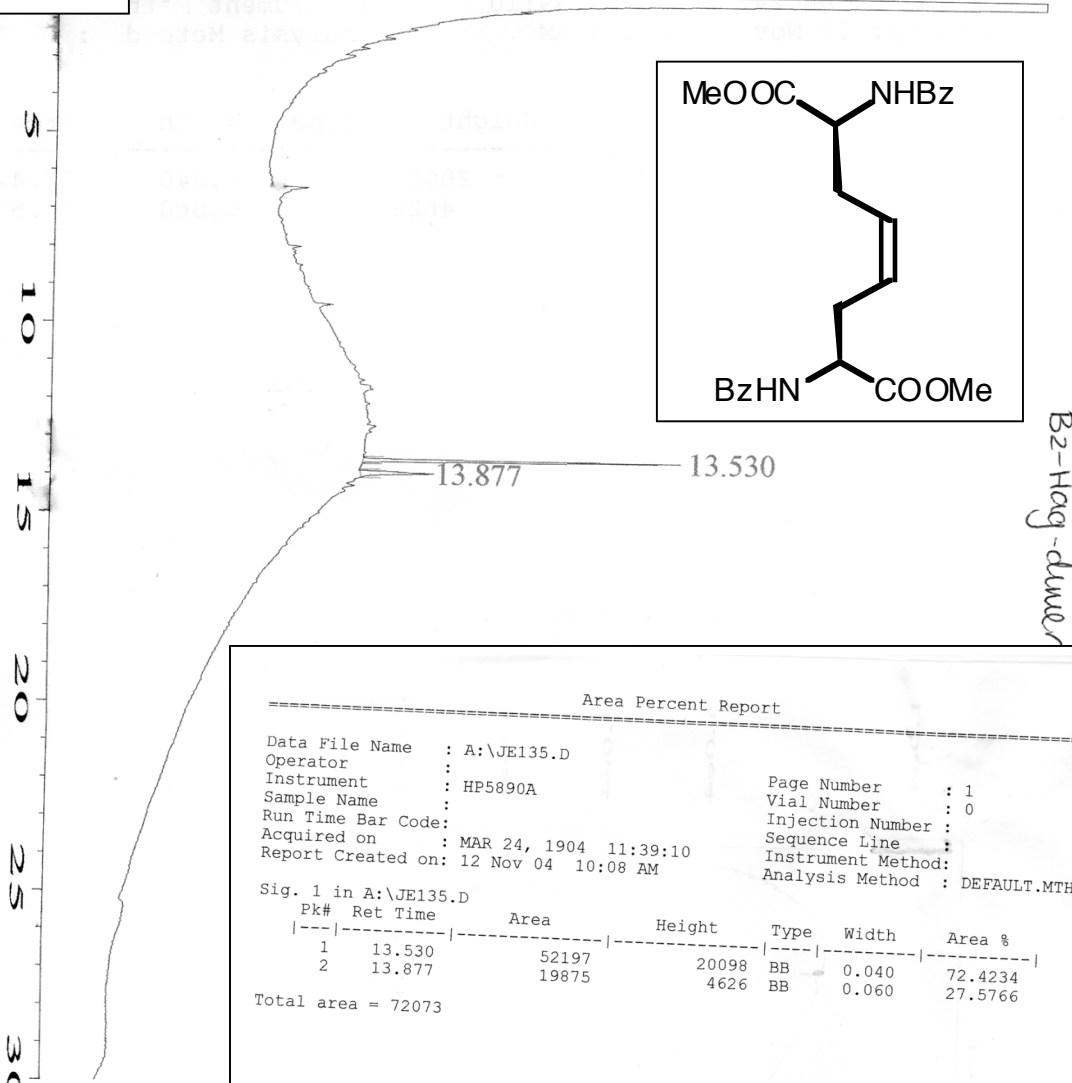
6.0e4

4.0e4

2.0e4



Compound 7



Data File Name : A:\JE135.D

Operator :

Instrument : HP5890A

Sample Name :

Run Time Bar Code:

Acquired on : MAR 24, 1904 11:39:10

Report Created on: 11 Nov 04 05:26 PM

Page Number : 1

Vial Number : 0

Injection Number :

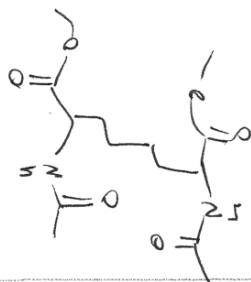
Sequence Line :

Instrument Method:

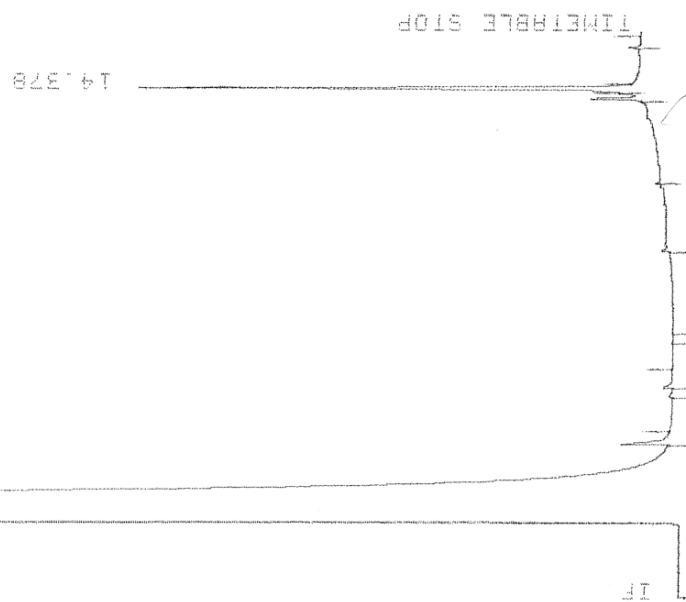
Analysis Method : DEFAULT.MTH

Compound 9

reduced Ar-Hag-dimers



150°C 1min
280°C (6min)
40°C min⁻¹

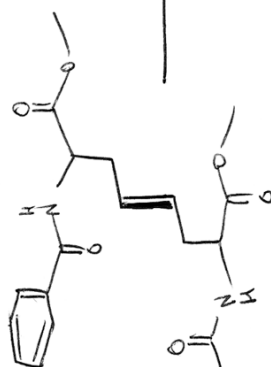
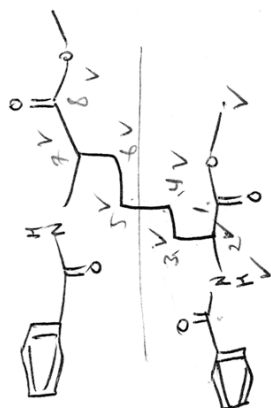


* TIME TO STOP
* RUN # 70
JUL 30, 1994 16:13:59
START

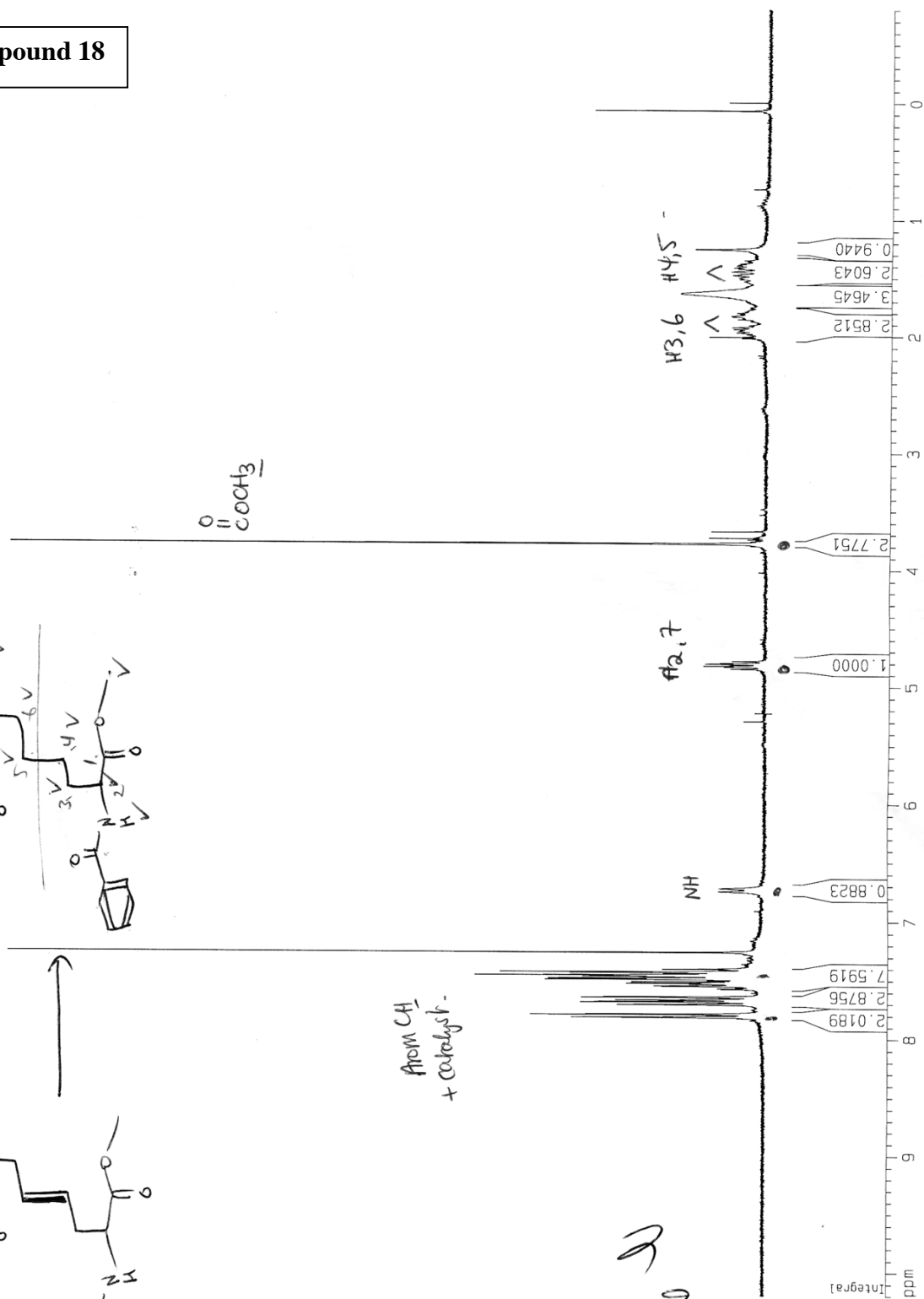
Closing signal file H:\08060808.BNC
RUN# 70 JUL 30, 1994 16:13:59
SIGNAL FILE: H:\08060808.BNC
30005/BPX-5 1.00W
AREA%
RT AREA TYPE WIDTH AREA%
14.378 480725 BB .067 100.00000
TOTAL AREA= 480725
MULTIPLIER=1.00000E+00

Compound 18

JE-6-155 (Bz-Hag-dimer H2'n)

from CH₂
+ catalyst

2.2



Compound 18

TOTAL AREA= 569301
MUL FACTOR=1.0000E+00

RT	AREA TYPE	WIDTH	AREA%
11.042	PB	1.052	2.66256
17.155	PB	1.136	97.33744

30005/BPX-5 1.0um

SIGNAL FILE: H:\08C5E938.BNC

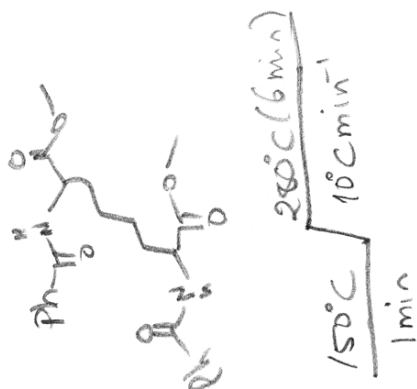
RUN# 67 JUL 30, 1994 13:33:43

Closing signal file H:\08C5E938.BNC

TIMEABLE STOP

11.042

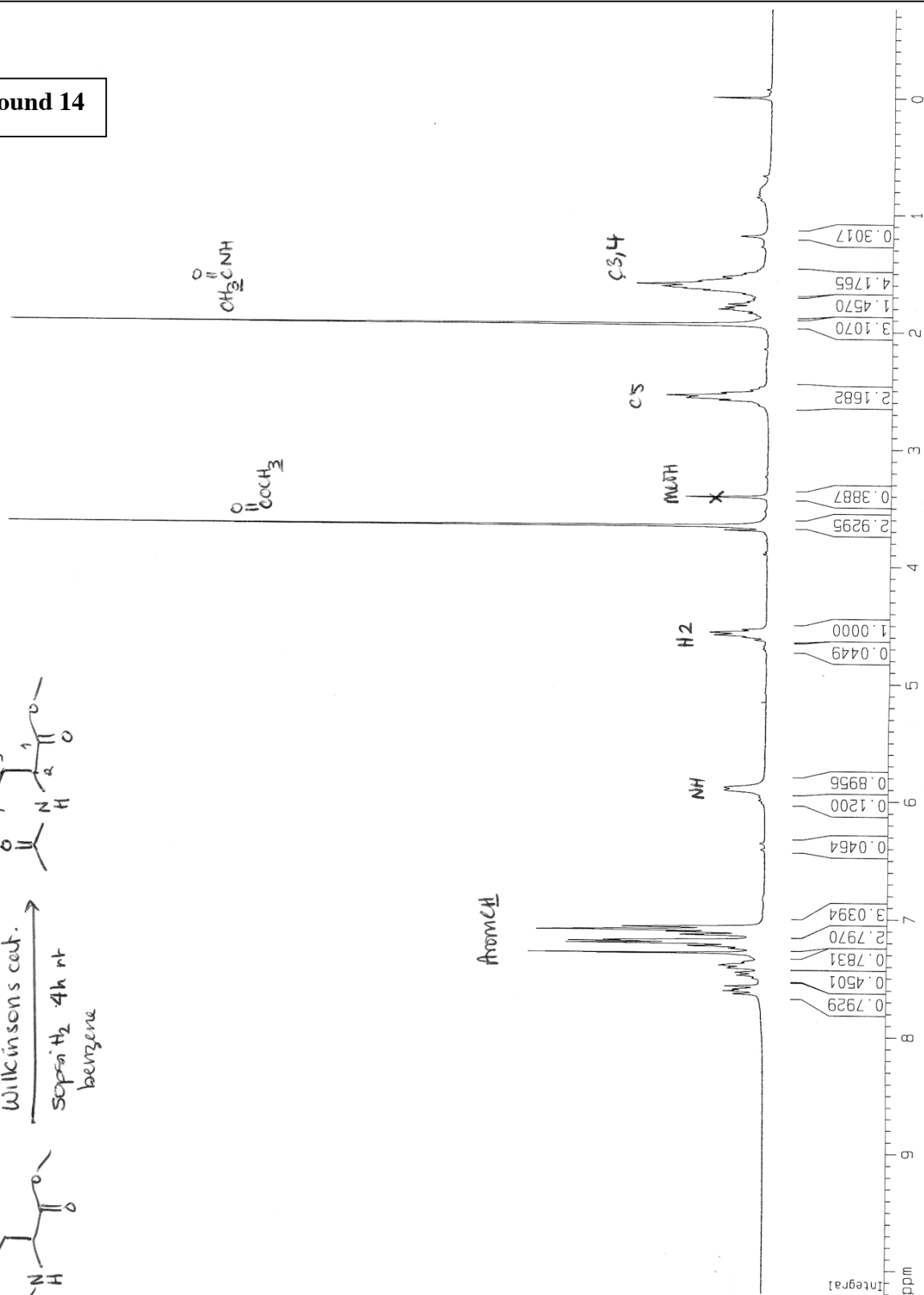
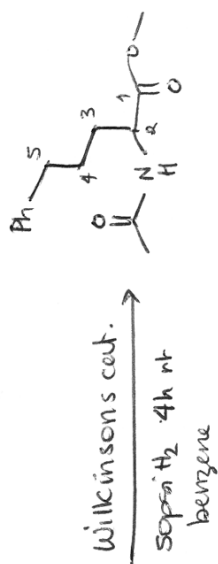
11.042



IT

IF

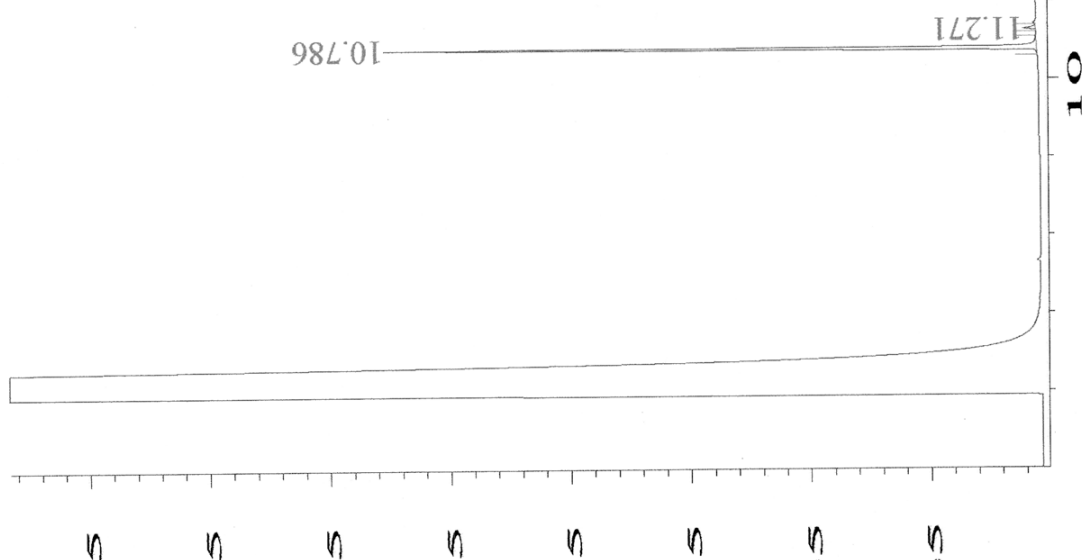
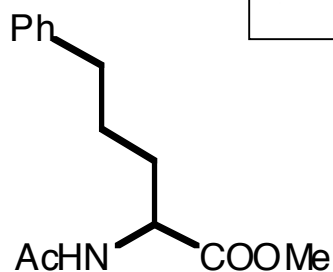
* LIST: RT 2 = 5
* RT 2 = 3 @
* RUN # 67 JUL 30, 1994 13:33:43
START

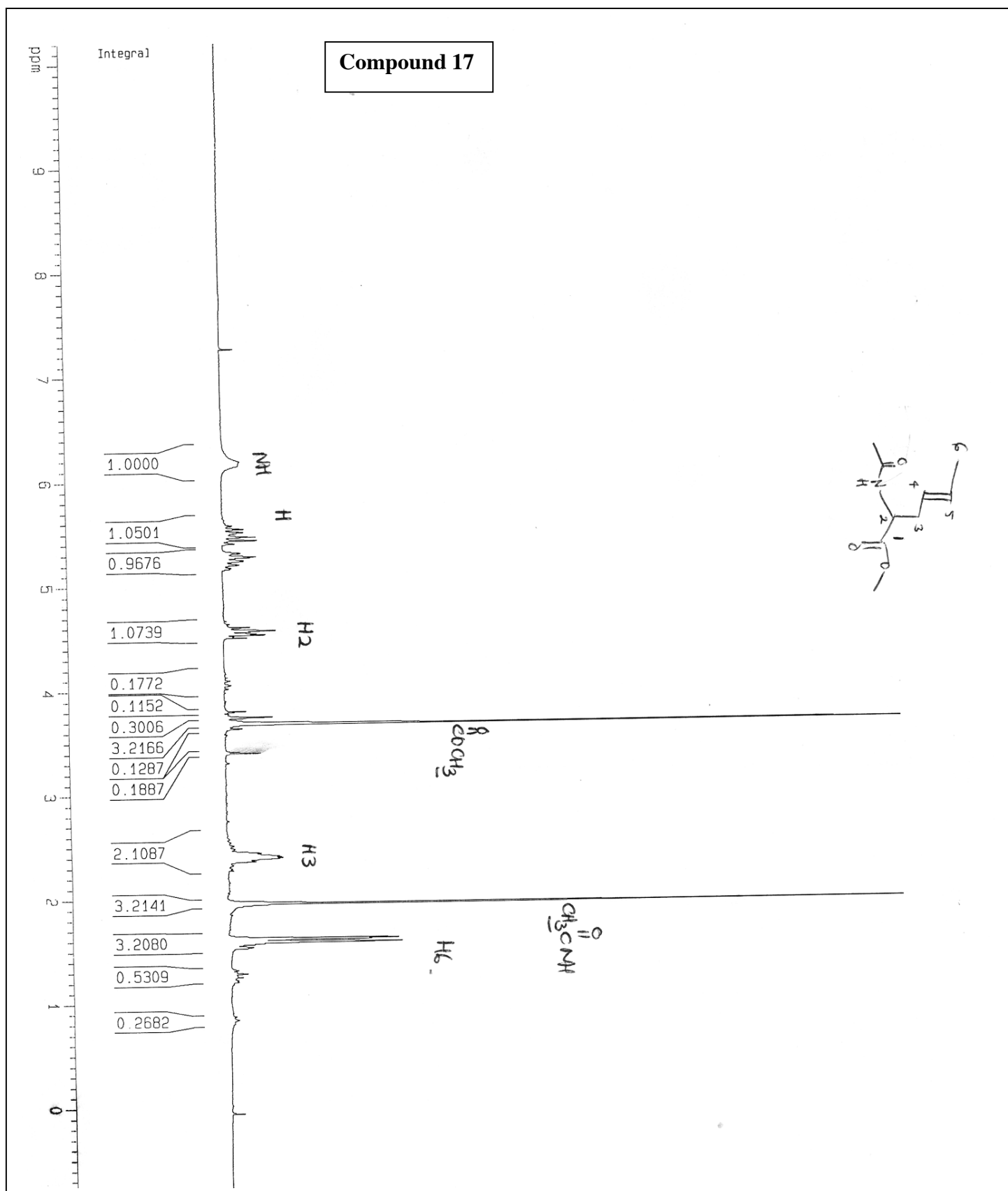
CC(=O)N[C@H](C/C=C/c1ccccc1)C(=O)OC

Data File Name : C:\HPCHEM\1\DATA\JE61.D
 Operator :
 Instrument : HP5890A
 Sample Name :
 Run Time Bar Code :
 Acquired on : JUL 28, 1904 11:38:51
 Report Created on: 12 Nov 04 10:48 AM
 Page Number : 1
 Vial Number : 0
 Injection Number :
 Sequence Line :
 Instrument Method: LC1.MTH

Compound 14

Area Percent Report						
=====						
Data File Name	:	C:\HPCHEM\1\DATA\JE61.D	Page Number	:	1	
Operator	:		Vial Number	:	0	
Instrument	:	HP5890A	Injection Number	:		
Sample Name	:		Sequence Line	:		
Run Time Bar Code	:		Instrument Method	:	LC1.MTH	
Acquired on	:	JUL 28, 1904 11:38:51	Analysis Method	:	LC1.MTH	
Report Created on:	:	12 Nov 04 10:48 AM				
Sig. 1 in C:\HPCHEM\1\DATA\JE61.D						
Pk#	Ret Time	Area	Height	Type	Width	Area %
1	10.786	1639190	545977	BB	0.044	98.2905
2	11.271	28509	10214	BV	0.042	1.7095



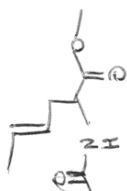
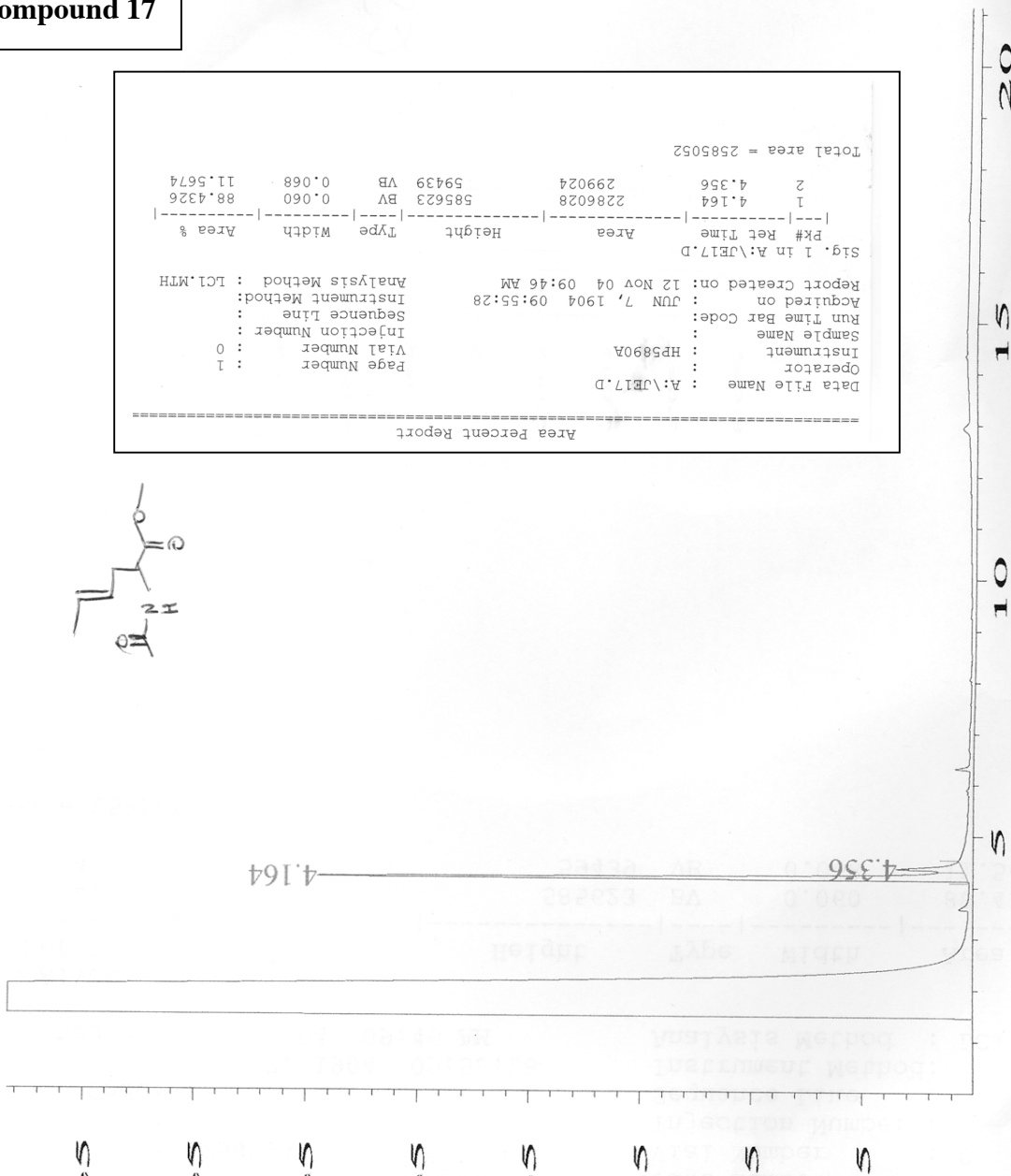


Data File Name : A:\JE17.D
 Operator :
 Instrument : HP5890A
 Sample Name :
 Run Time Bar Code :
 Acquired on : JUN 7, 1904 09:55:28
 Report Created on: 12 Nov 04 09:46 AM
 Page Number : 1
 Vial Number : 0
 Injection Number :
 Sequence Line :
 Instrument Method :
 Analysis Method : LCI.MTH

Compound 17

Area Percent Report

Data File Name : A:\JE17.D				
Operator :				
Instrument : HP5890A				
Sample Name :				
Run Time Bar Code :				
Acquired on : JUN 7, 1904 09:55:28				
Report Created on: 12 Nov 04 09:46 AM				
Analysis Method : LCI.MTH				
Page Number	: 1			
Vial Number	: 0			
Injection Number	:			
Sequence Line	:			
Instrument Method	:			
Total area = 2585052				
PK#	Ret Time	Area	Height	Type
1	4.164	2286028	585623	BV
2	4.356	299024	59439	VB
			0.068	
			0.060	
			88.4326	
			11.5674	



S23