Supporting Information for

Interaction of the neurotransmitter, neuropeptide Y (NPY), with phospholipid membranes: Film balance and fluorescence microscopy studies

Martina Dyck¹ and Mathias Lösche^{2,3,*}

University of Leipzig, Institute of Experimental Physics I, D–04103 Leipzig, Germany, Carnegie Mellon University, Department of Physics, Pittsburgh, PA 15213-3890, USA, and CNBT Consortium, NIST Center for Neutron Research, Gaithersburg, MD 20899-8562, USA

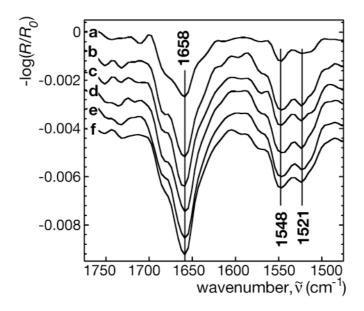


Figure S1: Amide region of external Fourier-transform infrared reflection-absorption spectra (FT-IRRA; p-polarization, incident angle, $\alpha_i = 40^\circ$, $T = 20^\circ$ C) of a hNPY film on a water subphase at various states of compression (a: $\pi = 0$ mN/m, b: 8 mN/m, c: 12 mN/m and d: 16 mN/m). The FT-IRRA spectra show a strong, narrow absorption band at 1658 cm⁻¹, the amide I mode, which is mainly associated with the C=O stretching mode of the peptide backbone. The position of this band is conformationally sensitive¹ and indicates that hNPY adopts an α-helical secondary structure at the air/water interface. In addition, the amide II band at 1548 cm⁻¹ also indicates an α-helical structure. The amide I band shows a high-frequency shoulder (~1685 cm⁻¹) that may arise from a turn or a loop in the peptide.¹ Since hNPY has 5 tyrosine residues, the band in the amide II region at ~ 1520 cm⁻¹ can be attributed to C–H in-plane bending and C–C stretching vibrations of tyrosine.² Since there is no amide I absorption detected near 1630 cm⁻¹, it is rather unlikely that the band at 1520 cm⁻¹ should be attributed to a β-sheet conformation.

The data have been collected in collaboration with Andreas Kerth and Alfred Blume at the Institute of Physical Chemistry at the Martin Luther University in Halle, Germany. A full account of this work will be presented elsewhere.³

(1) Goormaghtigh, E.; Cabiaux, V.; Ruysschaert, J.-M. Determination of Soluble and Membrane Protein Structure by Fourier Transform Infrared Spectroscopy. In *Physico-chemical Methods in the Study of Biomembranes*; Hilderson, H. J., Ralston, G. B., Eds.; Plenum Press, 1994; pp 405.

(2) Barth, A. Progress in Biophysics and Molecular Biology 2000, 74, 141.

(3) Dyck, M.; Kerth, A.; Blume, A.; Lösche, M. J. Phys. Chem. B 2006, submitted.