**Table I.** Antifungal peptides activity<sup>1,2</sup> against fungal strains determined by the broth microdilution method.

	Yeasts						
Antifungal agent	C.albicans ATCC 10231		C.albicans SA40				
"Bonn	$IC_{50}^{1}\mu g/ml$	MFC <sup>2</sup> µg/ml	$IC_{50} \mu g/ml$	MFC µg/ml			
Histatin 5	6.25 <sup>3</sup>	12.5	6.25	12.5			
ATCUN-C16	6.25	12.5	6.5	12.5			

<sup>1</sup> Growth inhibition obtained with fungal pathogens yeasts in the presence of synthetic proteins. Fungal cells  $(1 \times 10^4 \text{CFU/ml})$  were incubated with a dilution series of proteins in SDB (10%) supplemented with dialysed yeast extract, and growth inhibition was assessed spectrophotometrically at 620nm after 48 of incubation at 30 C (Helmerhorst E.J., Venuleo C., Beri A., Oppenheim F.G., Yeast 2005;22:705-14). The IC<sub>50</sub> values corresponded to the concentration of proteins that inhibited the maximum growth of that particular strain by 50%.

 $^{2}$  Cells were exposed to a dilution series of synthetic proteins in 10% SDB, pH7.0, after which cells were plated on SDA and incubated at 37 C for 48h. Cell viability was determined by colony counting. The minimum fungicidal concentration (MFC) was the lowest drug concentration resulting in the death of 99.9% or more of the initial inoculum.

<sup>3</sup> Average of three replicate experiments, each performed in duplicate.

Residue	$^{1}J_{C\alpha H\alpha}$ (Hz)				
D1	142.50				
<b>S2</b>	142.10				
Н3	144.20				
A4	144.20				
G5	nd <sup>b</sup>				
Y6	147.00				
K7	144.60				
<b>R8</b>	146.00				
К9	145.50				
F10	146.80				
H11	147.10				
E12	144.10				
K13	145.60				
H14	145.60				
H15	145.00				
S16	143.60				
H17	143.70				
R18	141.80				
G19	nd <sup>b</sup>				
Y20	142.90				
an accuracy of $\pm 0.5$ Hz					

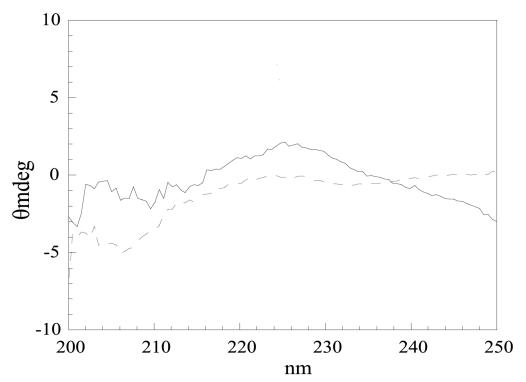
Table II.  ${}^{1}J_{C\alpha H\alpha}$  values in Hz for the ACTUN-C16 peptide measured at 25°C in a 1:1 TFE:H<sub>2</sub>O solution, pH 6.0.<sup>a</sup>

<sup>a</sup>: Coupling constants are reported with an accuracy of  $\pm 0.5$  Hz. <sup>b</sup>: nd: not determined.

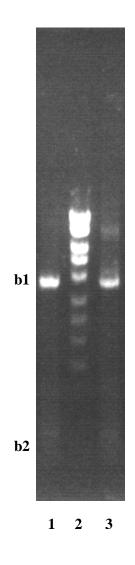
Residue	<sup>15</sup> N	NH	$^{13}C_{\alpha}$	${}^{1}H_{\alpha}$	$^{13}C_{\beta}$	${}^{1}H_{\beta}$	Others <sup>b</sup>
D1	-	-	51.9	3.97	49.5	2.69/2.69	
<b>S2</b>	116.7	7.92	57.1	4.37	61.42	3.82/3.82	
Н3	118.0	7.98	55.2	4.60	28.0	3.11/3.24	
A4	122.6	7.98	52.4	4.09	16.0	1.43	
G5	105.5	8.36	44.3	3.83/3.83	-	-	
Y6	120.5	7.78	58.2	4.28	35.9	3.07/3.07	
K7	119.0	7.98	57.1	4.00	30.1	1.86/1.86	γ 23.2 (1.38/1.55), δ 27.2
							(1.66/1.66), ε 39.7 (2.91/2.91)
R8	119.0	7.89	56.5	4.12	25.0	1.72/1.64	γ 27.7 (1.90/1.90), δ 41.0 (3.15/3.15)
К9	118.8	7.89	56.0	4.10	29.6	1.82/1.82	$\begin{array}{cccc} \gamma & 22.5 & (1.40/1.44), \ \delta & 26.4 \\ (1.63/1.63), & \epsilon & 39.7 \\ (2.89/2.89) \end{array}$
F10	119.4	8.28	58.5	4.20	36.7	3.12/2.96	
H11	118.5	8.21	57.1	4.34	28.1	3.23/3.23	
E12	119.2	8.31	56.4	4.03	27.7	2.11 /2.24	γ 34.1 (2.46/2.24)
K13	118.6	8.01	55.5	4.08	30.2	1.68/1.74	$\begin{array}{llllllllllllllllllllllllllllllllllll$
H14	118.8	7.89	55.7	4.36	28.0	2.94/2.73	
H15	119.0	7.98	54.9	4.49	28.0	2.94/2.94	
S16	114.6	7.95	57.1	4.35	61.7	3.86/3.83	
H17	118.3	7.97	54.4	4.55	27.8	3.13/3.03	
R18	120.2	7.95	54.1	4.24	28.5	1.73/1.64	γ 24.4 (1.42/1.42), δ 41.0 (3.01/3.01)
G19	108.0	8.14	42.9	3.96/3.74	-	-	
Y20	122.8	7.29	56.6	4.39	36.7	2.91/3.02	

Table III. <sup>1</sup>H and <sup>13</sup>C Chemical Shifts Assignments of the ACTUN-C16 peptide in a 1:1 TFE:H<sub>2</sub>O solution at 25°C and pH 6.0.<sup>a</sup>

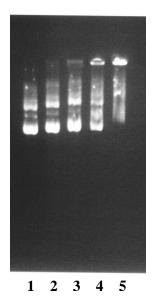
<sup>a</sup>: <sup>1</sup>H Chemical shifts are reported in ppm with an accuracy of ±0.02 ppm. <sup>13</sup>C Chemical shifts are reported in ppm with an accuracy of ±0.1 ppm. <sup>b</sup>: Carbon chemical shifts first, in brackets the proton chemical shift.



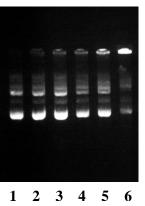
**Fig. 1** Circular dichroism spectra of the peptide with or without the plasmid DNA. CD spectra of ATCUN-C16 (50  $\mu$ M) in presence (—) and in absence (- -) of pDNA (Peptide/DNA 1:1 w/w) in 17mM Tris-HCl, pH 7.4, buffer. The ATCUN-C16-pDNA spectrum was subtracted of the contribution of the free pDNA.



**Fig. 2** Agarose gel electrophoresis (1% agarose) of the 0.25  $\mu$ g pQE30-rhdA after cleavage with SmaI (1U/ $\mu$ g) for 1h at 30°C, in the absence (lane 1) and in the presence (lane 3) of RTN32 peptide, in a ratio pDNA/peptide 1:10 w/w. **b**<sub>1</sub> and **b**<sub>2</sub> bands correspond to the SmaI digest fragments of about 3900 bp and 600 bp respectively. Molecular weight markers are shown in the lane 2.



**Fig. 3** Gel retardation shift assay at different Hst5/pDNA w/w ratios, using 0.9 µg pQE30-rhda and 1% agarose gel. Lane 1 (0), lane 2 (0.5), lane 3 (1), lane 4 (2) and lane 5 (5).



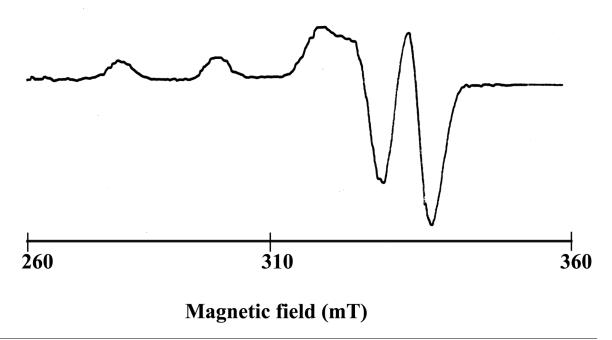
## 0' 15' 90' 3h 5h 20h

**Fig. 4** Agarose gel electrophoresis of pDNA treated with ATCUN-C16 peptide in 50 mM Tris-HCl buffer, pH 7.5, lanes 0-6 pDNA/ATCUN-C16 peptide mixture (ratio 1:1 w/w) in the presence of  $ZnCl_2$  (ATCUN-C16: $Zn^{2+}$  1:1 c/c) at different times (0, 15', 90', 3h, 5h and 20h).

## 0' 5' 15' 1h 3h



**Fig. 5** Agarose gel electrophoresis of pDNA treated with ATCUN-C16 peptide in 50 mM Tris-HCl buffer, pH 7.5, A) pDNA/ATCUN-C16 peptide mixture (ratio 1:1 w/w) in the presence of NiCl<sub>2</sub> and magnesium-monoperoxyphthalate (MMPP), at a molar ratio peptide/Ni<sup>2+/</sup> MMPP 1:1:10 c/c/c, incubated for 0 (lane 1), 5 (lane 2), 15 (lane 3), 60 (lane 4), 180 (lane 5) min at 37° C.



**Fig.6** Low-temperature EPR spectrum of Cu-ATCUN-C16 complex, 1:1 Cu/peptide molar ratio, in 50mM Hepes pH 7.2 buffer.