

Supporting Information

Iridium/Diene Complex Catalyzes Allylation Reactions of Ketones and Imines

Timothy J. Barker and Elizabeth R. Jarvo

Department of Chemistry, 1102 Natural Sciences II, University of California, Irvine, CA 92697

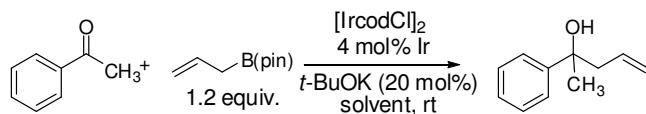
I. General procedures	(S1)
II. Solvent screen for allylation of ketones	(S2)
III. B(OH) ₃ and KOtBu additives screen for allylation of ketones	(S2)
IV. Base screen for allylation of ketones	(S3)
V. Procedure for allylations of ketones and imines	(S3)
VI. Characterization data for products	(S4)
VII. References	(S7)
VIII. Spectral Data	(S8)

I. General Procedures.

All reactions were carried out in a glovebox under an atmosphere of N₂. All glassware was either oven or flame-dried prior to use. Tetrahydrofuran (THF), diethyl ether (Et₂O), and dichloromethane (CH₂Cl₂) were degassed with argon and then passed through two 4 x 36 inch columns of anhydrous neutral A-2 alumina (8 x 14 mesh; LaRoche Chemicals; activated under a flow of argon at 350 °C for 12 h) to remove H₂O. All other solvents utilized were purchased “anhydrous” commercially, or purified as described (*vide infra*). Molarities of organolithium and organomagnesium reagents were determined by titration with menthol/bipyridine.¹ ¹H NMR spectra were recorded on Bruker GN-500 (500 MHz ¹H, 125.7 MHz ¹³C), CRYO-500 (500 MHz ¹H, 125.7 MHz ¹³C) or DRX-400 (400 MHz ¹H, 100MHz ¹³C) spectrometers. Proton chemical shifts are reported in ppm (δ) relative to the solvent resonance (CDCl₃, δ 7.27). Data are reported as follows: chemical shift [multiplicity [singlet (s), broad singlet (br s), doublet (d), doublet of doublets (dd), triplet (t), doublet of triplets (dt), doublet of doublet of triplets (ddt), triplet of triplets (tt), quartet (q), multiplet (m), apparent doublet (ad), broad doublet (br d) and broad multiplet (br m)], coupling constants [Hz], integration]. Carbon chemical shifts are reported in ppm (δ) relative to TMS with the respective solvent resonance as the internal standard (CDCl₃, δ 77.23 ppm). Unless otherwise indicated, NMR data were collected at 25 °C. Infrared spectra were obtained on a Mattson Instruments *Galaxy 5000* spectrometer. Analytical thin-layer chromatography (TLC) was performed using Silica Gel 60 F254 precoated plates (0.25 mm thickness). Visualization was accomplished by irradiation with a UV lamp and/or staining with KMnO₄, ceric ammonium molybdate (CAM), or p-anisaldehyde (PAA) solutions. Flash chromatography was performed using Silica Gel 60A (170-400 mesh) from Fisher Scientific. Allylboronic acid pinacol ester was purchased from Frontier Scientific, Inc. and was distilled through a 15 cm Vigreux fractionating column connected to a short-path distillation head (95 °C, 17 Torr) to remove B(OH)₃. [Ir(cod)Cl]₂ and [Ir(cod)OMe]₂ was purchased from Strem, stored in the glovebox, and used as received. [Ir(coe)₂Cl]₂ was synthesized according to a known procedure.² Potassium *tert*-butoxide was purchased from Alfa Aesar, stored in a

glovebox and used as received. Deutero-allylboronic acid pinacol ester **9** was prepared according to the procedure reported by Morken et. al.³ Deuterochloroiodomethane used for the synthesis of **9** was synthesized according to a known procedure.⁴ All ketones were purchased commercially and liquids were distilled prior to use. High resolution mass spectrometry was performed by the University of California, Irvine Mass Spectrometry Center.

II. Solvent Screen for Allylation of Ketones.

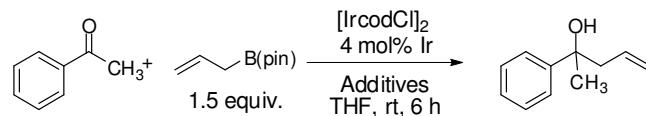


entry	solvent	time (h)	conversion (%) ^a
1	DCM	24	25
2	DMF	24	6
3	THF	24	>95
4	Toluene	24	27

^a 1,4-bis(trifluoromethyl)benzene was used as an internal standard.

III. B(OH)₃ and KOtBu Additives Screen for Allylation of Ketones.

Commercially available allylboronic acid pinacol ester was used in entry 1 and was found to be contaminated with boric acid by ¹³B NMR. In entries 2-9, the allylboronic acid pinacol ester was distilled and was analytically pure by ¹³B NMR. Adding 20 mol% of boric acid and 40 mol% KOtBu to reactions with analytically pure allylboronic acid pinacol ester were the optimized conditions that provided reproducible results.



entry	KOtBu (mol%)	B(OH) ₃ (mol%)	conversion (%) ^a
1	20	none	100
2	20	none	60
3	50	none	82
4	100	none	65
5	50	5	82
6	50	20	94
7	40	20	95
8	30	20	80
9	20	20	15

^a 1,4-bis(trifluoromethyl)benzene was used as an internal standard.

IV. Base Screen for Allylation of Ketones.

entry	base	time (h)	conversion (%) ^a
1	KOtBu	3	>95
2	NaOtBu	24	<5
3	LiOtBu	24	<5
4	LaO <i>i</i> Pr	24	38
5	KOPh	24	0 ^b

^a 1,4-bis(trifluoromethyl)benzene was used as an internal standard.

^b Under these reaction condition the allylboronic acid pinacol ester was isomerized to the 1-propenylboronic acid pinacol ester.

V. General Procedure for the Allylation of Ketones and Imines.

*Representative Procedure for Allylation of Acetophenone **1** with ligands.*

To a flame-dried 5 mL round-bottom flask in a glovebox, $[\text{Ir}(\text{cod})\text{Cl}]_2$ (7 mg, 0.010 mmol) and 1,2-bis(dicyclohexylphosphino)ethane (11 mg, 0.025 mmol) were stirred in 1 mL of THF for 3 h. After 3 h, potassium *tert*-butoxide (11 mg, 0.10 mmol), acetophenone (58 μ L, 0.50 mmol), allylboronic acid pinacol ester (140 μ L, 0.75 mmol), and 1,4-bis(trifluoromethyl)benzene (79 μ L, 0.50 mmol) (as an internal standard) were added. The reaction mixture was capped with a septa and stirred in the glovebox for 24 h at room temperature. After 24 h, an aliquot was removed and an $^1\text{H-NMR}$ was acquired. Conversion and yield of allylated product **3** was then calculated by comparison to the 1,4-bis(trifluoromethyl)benzene standard.

*Representative Procedure for Allylation of Acetophenone **5a**.*

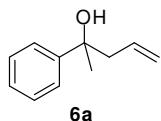
To a flame-dried 5 mL round-bottom flask in a glovebox, was added $[\text{Ir}(\text{cod})\text{Cl}]_2$ (7 mg, 0.010 mmol), potassium *tert*-butoxide (22 mg, 0.20 mmol), boric acid (6.2 mg, 0.10 mmol), THF (1 mL), acetophenone (58 μ L, 0.50 mmol) and allylboronic acid pinacol ester (140 μ L, 0.75 mmol). The reaction mixture was capped with a septa and stirred in the glovebox for 3 h at room temperature. After 3 h, the reaction was removed from the glovebox and quenched with 1 mL of saturated NH_4Cl solution and stirred for 10 min. The aqueous layer was extracted (3×5 mL Et_2O) and the combined organic layers were dried over MgSO_4 , filtered, and concentrated in vacuo. Purification was performed using silica gel chromatography (90:10 pentane/ Et_2O affording **6a** as a colorless oil (63 mg, 78%). Compounds **6e**, **6f**, and **6j** were purified using silver nitrate impregnated silica gel.⁵

*Representative Procedure for Allylation of N-benzylidene-4-methoxyaniline **5m**.*

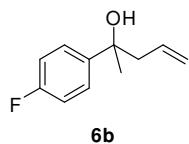
To a flame-dried 5 mL round-bottom flask in a glovebox, was added $[\text{Ir}(\text{cod})\text{Cl}]_2$ (6.7 mg, 0.010 mmol), potassium *tert*-butoxide (11 mg, 0.10 mmol), THF (1 mL), *N*-benzylidene-4-methoxyaniline (110 μ L, 0.50 mmol) and allylboronic acid pinacol ester (140 μ L, 0.75 mmol). The reaction mixture was capped with a septa and stirred in the

glovebox for 12 h at room temperature. After 12 h, the reaction was removed from the glovebox and quenched with 2 mL of H₂O and stirred for 10 min. The aqueous layer was extracted (3 x 8 mL CH₂Cl₂) and the combined organic layers were dried over Na₂SO₄, filtered, and concentrated in vacuo. Purification was performed using silica gel chromatography (3–5% Et₂O in pentane) affording **6m** as a pale yellow oil (86 mg, 68%).

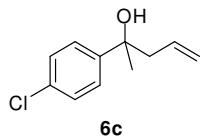
VI. Characterization Data for Products.



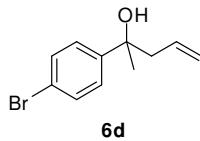
2-Phenyl-4-penten-2-ol (6a): ¹H NMR (CDCl₃, 400 MHz) δ 7.44 (d, *J* = 7.8 Hz, 2H), 7.34 (t, *J* = 7.4 Hz, 2H), 7.23 (d, *J* = 7.2 Hz, 1H), 5.68–5.56 (m, 1H), 5.14 (d, *J* = 18.8 Hz, 1H), 5.12 (d, *J* = 8.5 Hz, 1H), 2.69 (dd, *J* = 13.4, 6.7 Hz, 1H) 2.50 (dd, *J* = 13.8, 8.2 Hz, 1H), 2.03 (s, 1H), 1.54 (d, *J* = 1.54 Hz, 3H). The spectral values obtained were consistent with those reported in the literature.⁶



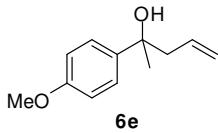
2-(4-Fluorophenyl)-4-penten-2-ol (6b): ¹H NMR (CDCl₃, 400 MHz) δ 7.43–7.39 (m, 2H), 7.05–7.00 (m, 2H), 5.66–5.58 (m, 1H), 5.17–5.12 (m, 2H), 2.66 (dd, *J* = 13.8, 6.6 Hz, 1H) 2.50 (dd, *J* = 13.7, 8.2 Hz, 1H), 2.02 (s, 1H), 1.55 (s, 3H). The spectral values obtained were consistent with those reported in the literature.⁷



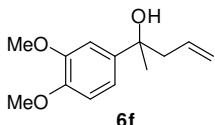
2-(4-Chlorophenyl)-4-penten-2-ol (6c): ¹H NMR (CDCl₃, 400 MHz) δ 7.38 (dd, *J* = 8.8 Hz, 2H), 7.31 (dd, *J* = 8.8 Hz, 2H), 5.64–5.56 (m, 1H), 5.17–5.12 (m, 2H), 2.66 (dd, *J* = 13.7, 6.5 Hz, 1H) 2.49 (dd, *J* = 13.7, 8.3 Hz, 1H), 2.02 (s, 1H), 1.54 (s, 3H). The spectral values obtained were consistent with those reported in the literature.⁵



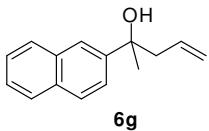
2-(4-Bromophenyl)-4-penten-2-ol (6d): ¹H NMR (CDCl₃, 400 MHz) δ 7.47 (d, *J* = 8.6 Hz, 2H), 7.32 (d, *J* = 8.6 Hz, 2H), 5.66–5.55 (m, 1H), 5.17–5.13 (m, 2H), 2.66 (dd, *J* = 13.7, 6.5 Hz, 1H) 2.49 (dd, *J* = 13.7, 8.3 Hz, 1H), 2.03 (s, 1H), 1.54 (s, 3H). The spectral values obtained were consistent with those reported in the literature.⁶



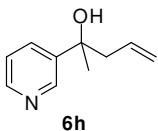
2-(4-Methoxyphenyl)-4-penten-2-ol (6e): **¹H NMR** (CDCl_3 , 400 MHz) δ 7.37 (d, J = 8.9 Hz, 2H), 6.89 (d, J = 8.9 Hz, 1H), 5.70–5.59 (m, 1H), 5.16–5.11 (m, 2H), 3.82 (s, 3H), 2.67 (dd, J = 13.7, 6.5 Hz, 1H), 2.50 (dd, J = 13.7, 8.2 Hz, 1H), 2.00 (s, 1H), 1.54 (s, 3H). The spectral values obtained were consistent with those reported in the literature.⁶



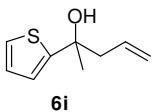
2-(3,4-Dimethoxyphenyl)-4-penten-2-ol (6f): **¹H NMR** (CDCl_3 , 400 MHz) δ 7.04 (d, J = 2.1 Hz, 1H), 6.94 (dd, J = 8.4, 2.1 Hz, 1H), 6.84 (d, J = 8.4 Hz, 1H), 5.71–5.60 (m, 1H), 5.15 (d, J = 17.5 Hz, 1H), 5.14 (d, J = 9.9 Hz, 1H), 3.91 (s, 3H), 3.89 (s, 3H), 2.70 (dd, J = 13.7, 6.5 Hz, 1H), 2.50 (dd, J = 13.7, 8.2 Hz, 1H), 2.01 (s, 1H), 1.55 (s, 3H); **¹³C NMR** (CDCl_3 , 125 MHz) δ 148.8, 147.8, 140.7, 134.0, 119.6, 117.0, 110.9, 108.7, 73.7, 56.1, 48.7, 30.2, 24.8; **IR** (thin film, cm^{-1}) 3482, 3076, 2976, 2934, 1261, 1143, 1028; **HRMS** (TOF MS ES+) m/z calculated for $\text{C}_{13}\text{H}_{18}\text{O}_3$ 245.1154 [$\text{M}+\text{Na}^+$] found 245.1146; **TLC R_f** = 0.3 (65:35 hexanes/EtOAc).



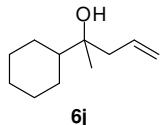
2-(Naphthalen-2-yl)-4-penten-2-ol (6g): **¹H NMR** (CDCl_3 , 500 MHz) δ 7.93 (s, 1H), 7.86–7.83 (m, 3H), 7.55 (dd, J = 8.6, 1.7 Hz, 1H), 7.50–7.46 (m, 2H), 5.67–5.59 (m, 1H), 5.18 (d, J = 17.1 Hz, 1H), 5.13 (d, J = 10.1 Hz, 1H), 2.82 (dd, J = 13.8, 6.3 Hz, 1H), 2.60 (dd, J = 13.8, 8.4 Hz, 1H), 2.18 (s, 1H), 1.65 (s, 3H). The spectral values obtained were consistent with those reported in the literature.⁵



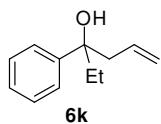
2-(Pyridin-3-yl)-4-penten-2-ol (6h): **¹H NMR** (CDCl_3 , 400 MHz) δ 8.70 (d, J = 1.8 Hz, 1H), 8.50 (d, J = 4.6 Hz, 1H), 8.00 (d, J = 8.0 Hz, 1H), 7.27 (dd, J = 7.7, 4.8 Hz, 1H), 5.68–5.60 (m, 1H), 5.17 (d, J = 10.3 Hz, 1H), 5.16 (d, J = 17.8 Hz, 1H), 2.68 (dd, J = 13.8, 6.7 Hz, 1H), 2.54 (dd, J = 13.8, 8.1 Hz, 1H), 2.30 (s, 1H), 1.59 (s, 3H); **¹³C NMR** (CDCl_3 , 125 MHz) δ 148.2, 147.1, 143.0, 133.0, 132.9, 123.2, 120.5, 72.7, 48.6, 30.0; **IR** (thin film, cm^{-1}) 3267, 2976, 1639, 1579, 1419; **HRMS** (TOF MS ES+) m/z calculated for $\text{C}_{10}\text{H}_{13}\text{O}_1$ 164.1075 [$\text{M}+\text{H}^+$] found 164.1074; **TLC R_f** = 0.2 (6:4 hexanes/EtOAc).



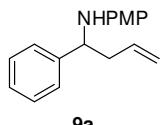
2-(Thiophen-2-yl)-4-penten-2-ol (6i): **¹H NMR** (CDCl_3 , 400 MHz) δ 7.21 (dd, $J = 5.0, 1.2$ Hz, 1H), 6.96 (dd, $J = 5.0, 3.6$ Hz, 1H), 5.79–5.70 (m, 1H), 5.20–5.16 (m, 2H), 2.72 (dd, $J = 13.7, 6.6$ Hz, 1H), 2.58 (dd, $J = 13.6, 8.1$ Hz, 1H), 2.27 (s, 1H), 1.63 (s, 3H). The spectral values obtained were consistent with those reported in the literature.⁵



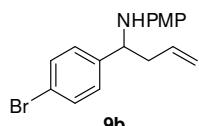
α -Methyl- α -(2-propenyl)cyclohexanemethanol (6j): **¹H NMR** (CDCl_3 , 500 MHz) δ 5.94–5.86 (m, 1H), 5.16 (d, $J = 17.1$ Hz, 1H), 5.12 (d, $J = 10.4$ Hz, 1H), 2.29–2.19 (m, 2H), 1.88–1.58 (m, 5H), 1.42 (s, 1H) 1.33–0.98 (m, 5H); **¹³C NMR** (CDCl_3 , 125 MHz) δ 134.4, 118.8, 74.1, 47.7, 44.4, 27.8, 27.1, 27.0, 26.9, 26.8, 23.9; **IR** (thin film, cm^{-1}) 3431, 3076, 2927, 2854, 1639, 1450; **HRMS** (TOF MS ES+) m/z calculated for $\text{C}_{11}\text{H}_{20}\text{O}$ 191.1412 [M+Na]⁺ found 191.1415; **TLC R_f** = 0.2 (9:1 pentane/Et₂O).



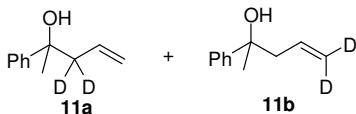
2-Phenyl-4-hexen-2-ol (6k): **¹H NMR** (CDCl_3 , 500 MHz) δ 7.40 (d, $J = 7.2$ Hz, 2H), 7.37 (t, $J = 7.7$ Hz, 2H), 7.25 (t, $J = 7.2$ Hz, 1H), 5.63–5.55 (m, 1H), 5.15 (d, $J = 19.3$ Hz, 1H), 5.12 (d, $J = 10.6$ Hz, 1H), 2.74 (dd, $J = 13.7, 6.0$ Hz, 1H) 2.51 (dd, $J = 13.7, 8.7$ Hz, 1H), 2.02 (s, 1H), 1.91–1.81 (m, 2H), 0.78 (t, $J = 7.4$ Hz, 3H). The spectral values obtained were consistent with those reported in the literature.⁸



4-methoxy-N-(1-phenylbut-3-enyl)aniline (9a): **¹H NMR** (CDCl_3 , 500 MHz) δ 7.39–7.17 (m, 5H), 6.67 (ad, $J = 8.9$ Hz, 2H), 6.45 (ad, $J = 8.9$ Hz, 2H), 5.81–5.70 (m, 1H), 5.21–5.09 (m, 2H), 4.30 (dd, $J = 8.3, 5.1$ Hz, 1H), 3.87 (br s, 1H), 3.67 (s, 3H), 2.63–2.53 (m, 1H), 2.51–2.41 (m, 1H). Analytical data is consistent with reported literature values.⁹



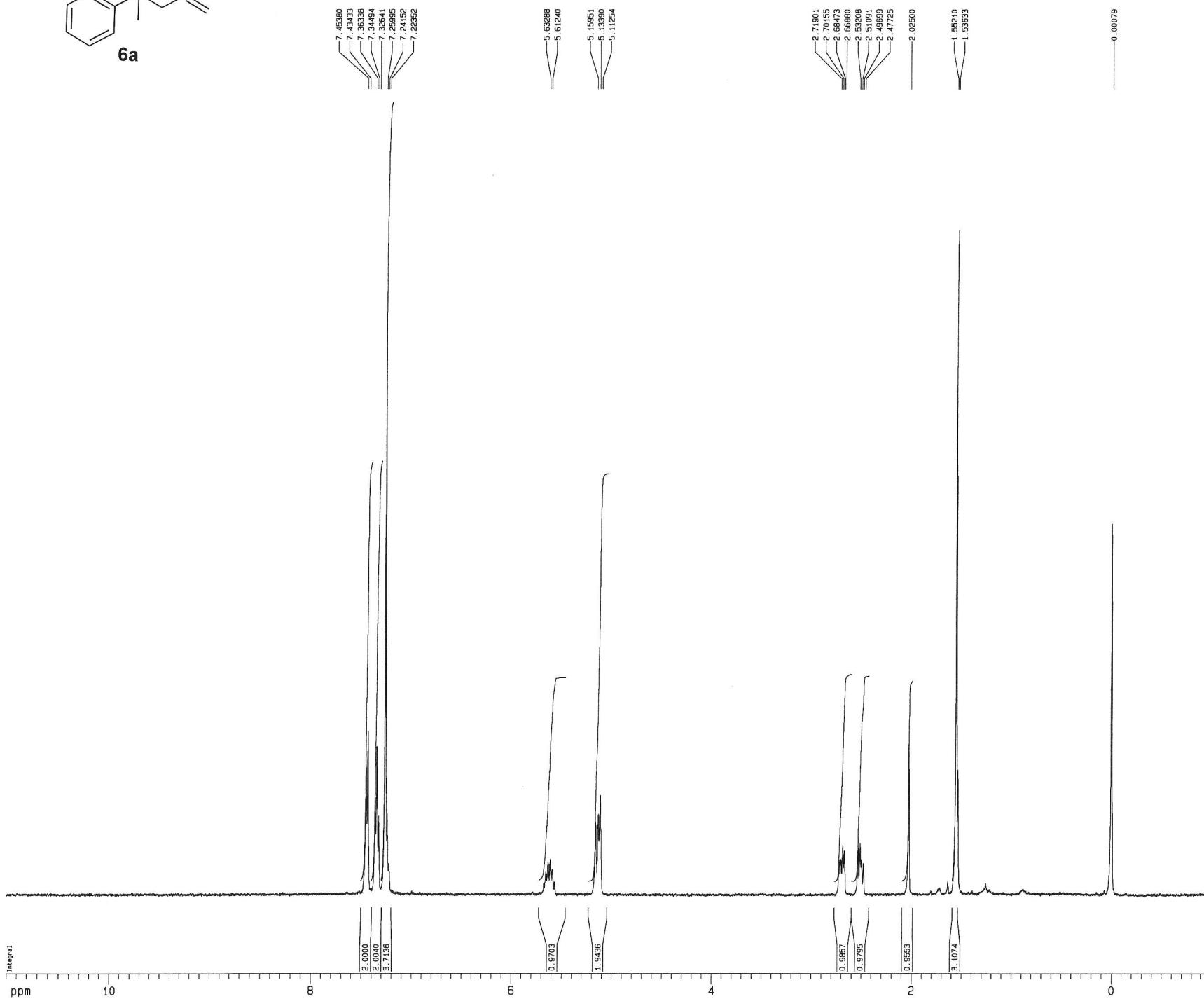
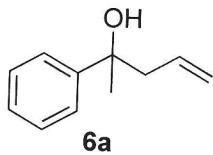
N-(1-(4-bromophenyl)but-3-enyl)-4-methoxyaniline (9b): **¹H NMR** (CDCl_3 , 500 MHz) δ 7.45 (d, $J = 8.4$ Hz, 2H), 7.25 (d, $J = 8.4$ Hz, 2H), 6.68 (d, $J = 8.9$ Hz, 2H), 6.42 (d, $J = 8.9$ Hz, 2H), 5.80–5.69 (m, 1H), 5.18 (d, $J = 17.1$ Hz, 1H), 5.16 (d, $J = 9.6$ Hz, 1H), 4.26 (m, 1H), 3.90 (s, 1H), 3.70 (s, 1H), 2.60–2.53 (m, 1H), 2.48–2.40 (m, 1H). **¹³C NMR** (CDCl_3 , 125 MHz) δ 152.5, 143.0, 141.2, 134.5, 131.9, 128.4, 120.8, 118.9, 115.0, 114.9, 57.8, 55.9, 43.4; **IR** (film, cm^{-1}) 3402, 3076, 2999, 2933, 2908, 2833, 1512, 1240; **HRMS** (TOF MS ES+) (m/z): [M+H]⁺ calculated for $\text{C}_{17}\text{H}_{18}\text{NOBr}$ 332.0650, found 332.0647; **TLC R_f** = 0.5 (5% Et₂O in pentane).



3,3-dideutero-2-phenyl-4-penten-2-ol (11a) and 5,5-dideutero-2-phenyl-4-penten-2-ol (11b): **¹H NMR** (CDCl_3 , 400 MHz) δ 7.44 (d, $J = 7.8$ Hz, 2H), 7.34 (t, $J = 7.4$ Hz, 2H), 7.23 (d, $J = 7.2$ Hz, 1H), 5.68–5.56 (m, 1H), 5.14–5.12 (m, 1H), 2.69–2.50 (m, 1H), 2.03 (s, 1H), 1.54 (d, $J = 1.54$ Hz, 3H); **¹³C NMR** (CDCl_3 , 125 MHz) δ 147.8, 133.8, 133.6, 128.4, 126.8, 125.0, 119.8, 73.8, 48.6, 30.13, 30.08; **IR** (thin film, cm^{-1}) 3425, 3061, 2976, 1601, 1446, 1373; **HRMS** (TOF MS APCI+) m/z calculated for $\text{C}_{11}\text{H}_{12}\text{D}_2\text{O}$ 187.1068 ($\text{M}+\text{Na}^+$) found 187.1067; **TLC** $R_f = 0.2$ (9:1 Pentane/Et₂O).

VII. References.

-
- ¹ Black, H.T. Titrating Alkyllithium Reagents.
<http://www.ux1.eiu.edu/~cfthb/research/handbook/titrating.htm> (accessed July 2008).
- ² van der Ent, A.; Onderdelinden, A. L. *Inorg. Synth.* **1990**, *28*, 90-92.
- ³ Sieber, J.D.; Morken, J.P. *J. Am. Chem. Soc.* **2008**, *130*, 4978-4983.
- ⁴ Miyano, S.; Hashimoto, H. *Bull. Chem. Soc. Japan* **1971**, *44*, 2864-2865
- ⁵ For a review on the use of silver nitrate in chromatography see: Williams, C.M.; Mander, L.N. *Tetrahedron* **2001**, *57*, 425-447.
- ⁶ Schneider, U.; Kobayashi, S. *Angew. Chem. Int. Ed.* **2007**, *46*, 5909-5912.
- ⁷ Zhang, X.; Chen, D.; Liu, X.; Feng, X. *J. Org. Chem.* **2007**, *72*, 5227-5233.
- ⁸ Canales, E.; Prasad, K. G.; Soderquist, J. A. *J. Am. Chem. Soc.* **2005**, *127*, 11572-11573.
- ⁹ Shimizu, M.; Kimura, M.; Watanabe, T.; Tamaru, Y. *Org. Lett.* **2005**, *7*, 637-640.



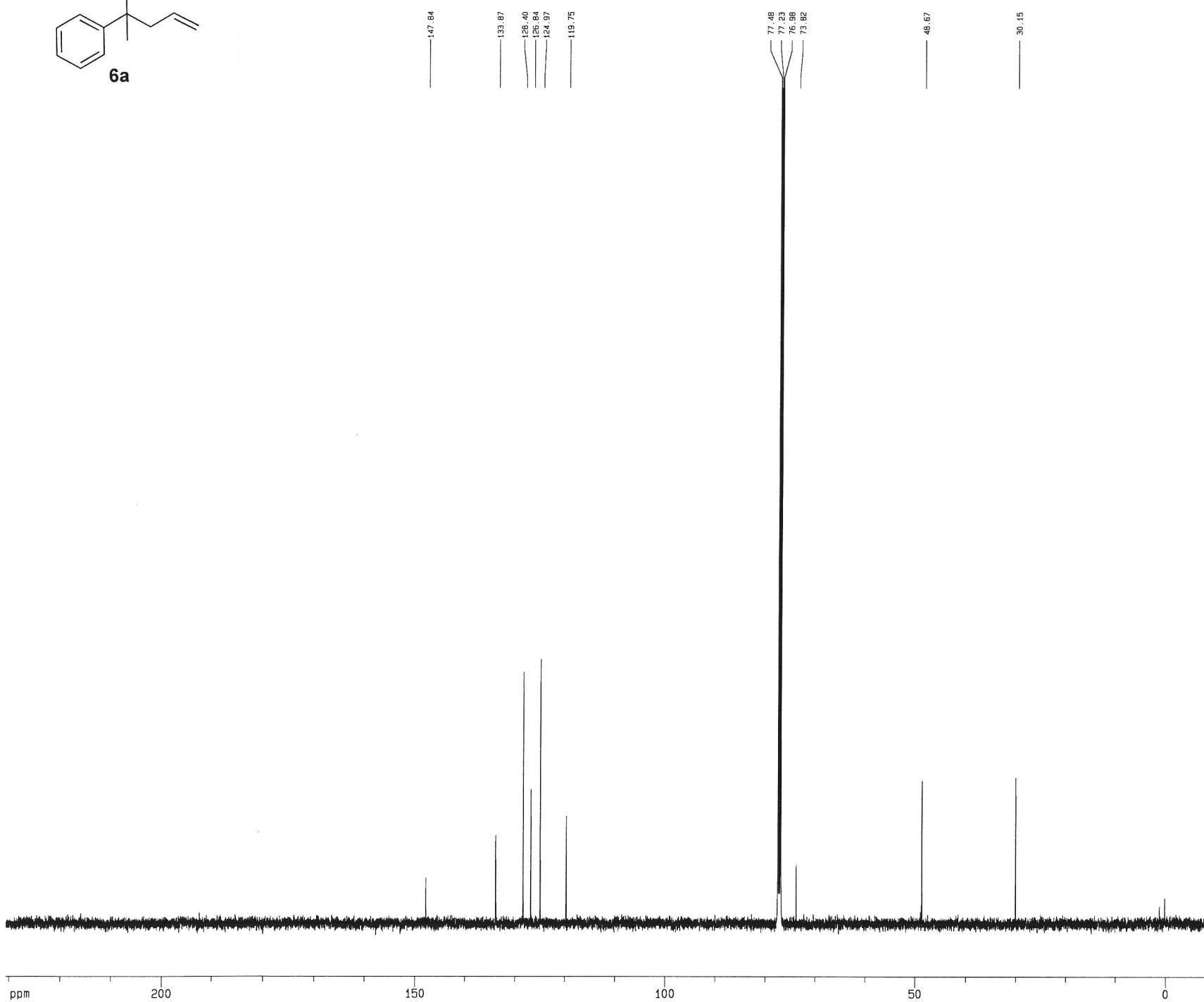
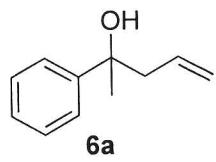
Current Data Parameters
 USER *tjbark*
 NAME *1-JB-171*
 EXPNO 3
 PROCNO 1

F2 - Acquisition Parameters
 Date 20080216
 Time 17.32
 INSTRUM *drx400*
 PROBHD 5 mm QNP H/F/P
 PULPROG *zg30*
 TD 65536
 SOLVENT *CDC13*
 NS 8
 DS 2
 SWH 6410.256 Hz
 FIDRES 0.097813 Hz
 AQ 5.1118579 sec
 RG 1149.4
 DW 76.000 usec
 DE 4.50 usec
 TE 298.0 K
 D1 0.1000000 sec
 MCREST 0.0000000 sec
 MCWRK 0.0150000 sec

===== CHANNEL f1 =====
 NUC1 1H
 P1 12.00 usec
 PL1 0.00 dB
 SF01 400.1328009 MHz

F2 - Processing parameters
 SI 65536
 SF 400.1300221 MHz
 WDM EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 2.00

1D NMR plot parameters
 CX 22.80 cm
 CY 10.00 cm
 F1P 11.000 ppm
 F1 4401.43 Hz
 F2P -1.000 ppm
 F2 -400.13 Hz
 PPMCM 0.52632 ppm/cm
 HZCM 210.59476 Hz/cm



Current Data Parameters
 USER *tjbarb*
 NAME *i-TB-278*
 EXPNO 5
 PROCNO 1

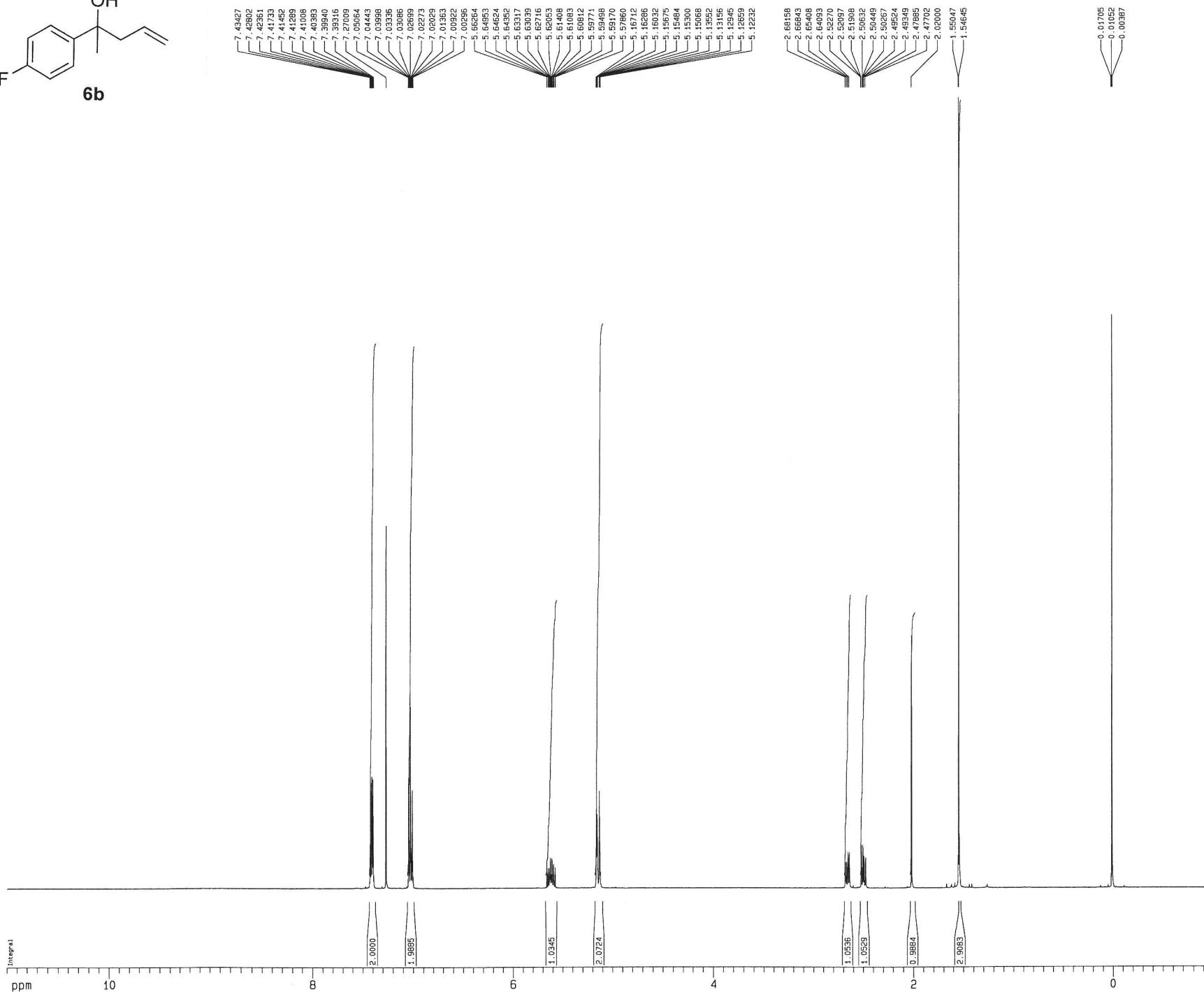
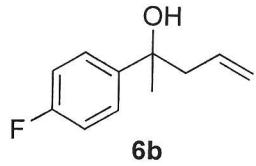
F2 - Acquisition Parameters
 Date_ 20080503
 Time 10.08
 INSTRUM cryo500
 PROBHD 5 mm CP/T1 1H-
 PULPROG zg3d30
 TD 65418
 SOLVENT CDC13T
 NS 513
 DS 4
 SWH 30303.031 Hz
 FIDRES 0.453222 Hz
 AQ 1.0794470 sec
 RG 14595.5
 DW 16.500 usec
 DE 6.00 usec
 TE 299.0 K
 D1 0.2500000 sec
 d11 0.0300000 sec
 MCREST 0.0000000 sec
 MCWRK 0.01500000 sec

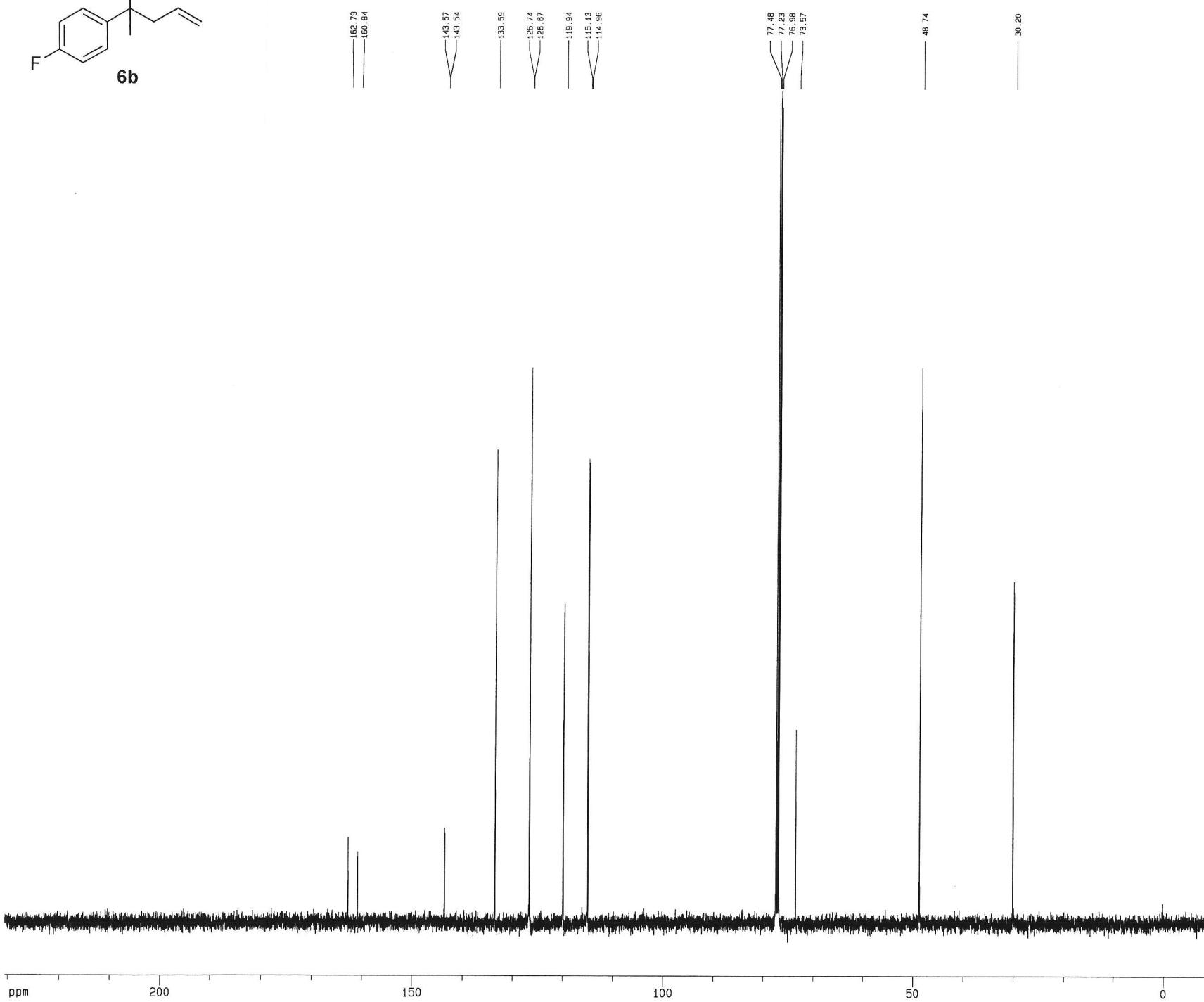
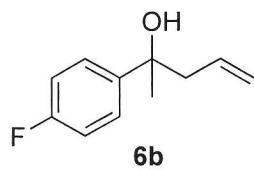
===== CHANNEL f1 =====
 NUC1 ¹³C
 P1 14.75 usec
 PL1 -1.00 dB
 SF01 125.7942548 MHz

===== CHANNEL f2 =====
 CPDPRG2 waltz16
 NUC2 ¹H
 PCPD2 100.00 usec
 PL2 1.60 dB
 PL12 24.80 dB
 SF02 500.2225011 MHz

F2 - Processing parameters
 SI 65536
 SF 125.7803983 MHz
 WDM EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 2.00

1D NMR plot parameters
 CX 22.80 cm
 CY 50.00 cm
 F1P 230.625 ppm
 F1 29008.05 Hz
 F2P -10.296 ppm
 F2 -1294.98 Hz
 PPMCM 10.55667 ppm/cm
 HZCM 1329.08032 Hz/cm





Current Data Parameters
 USER tjbark
 NAME 2-TJB-15
 EXPNO 5
 PROCNO 1

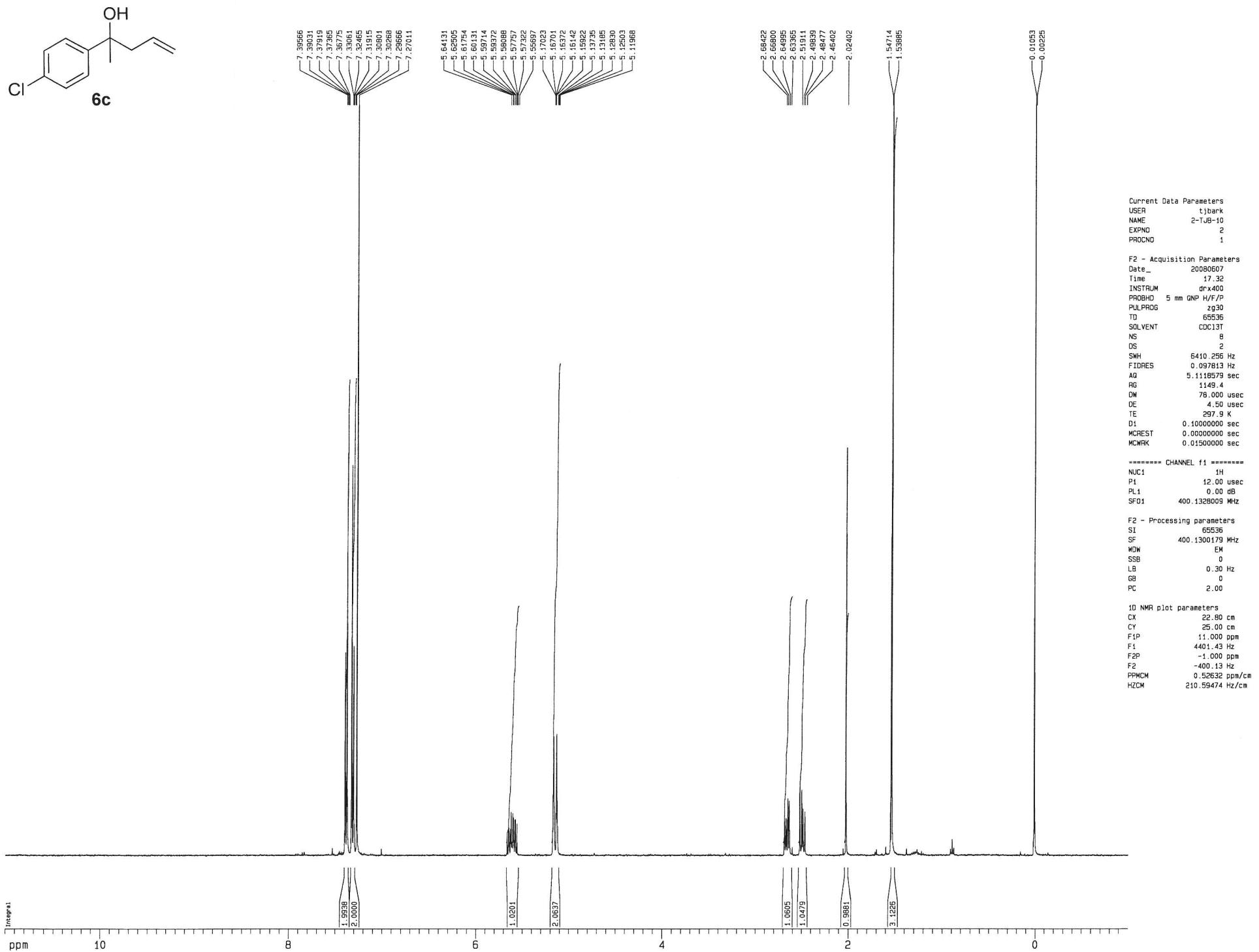
F2 - Acquisition Parameters
 Date_ 20080617
 Time 8.34
 INSTRUM cryo500
 PROBHD 5 mm CPTCI 1H-
 PULPROG zgdc30
 TD 65418
 SOLVENT CDCl3T
 NS 129
 DS 4
 SWH 30303.031 Hz
 FIDRES 0.463222 Hz
 FOAMES 1.0734470 sec
 A0 13004
 RG 16.500 usec
 DE 6.00 usec
 TE 298.0 K
 D1 0.2500000 sec
 d11 0.0300000 sec
 MCREST 0.0000000 sec
 MCWAK 0.0150000 sec

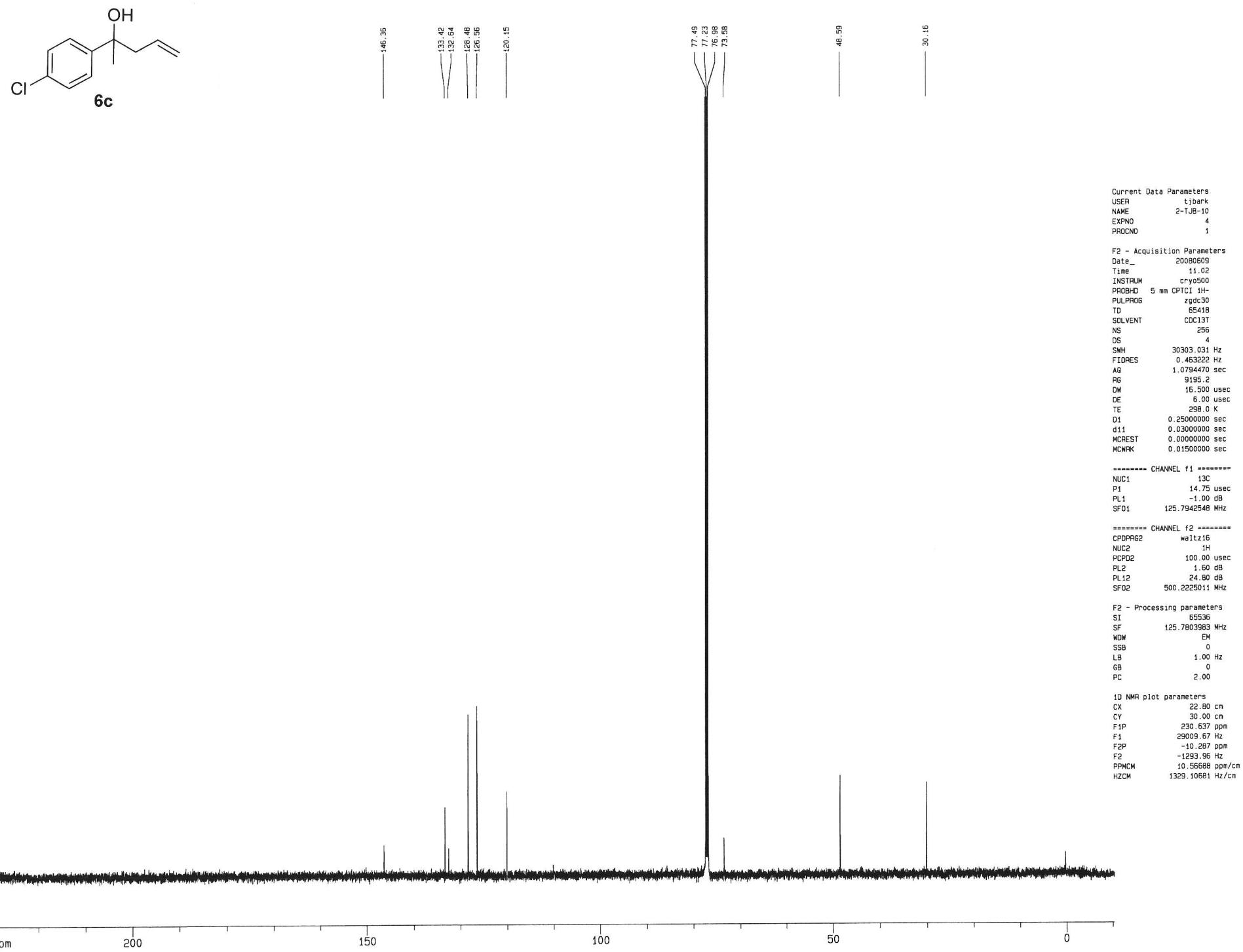
===== CHANNEL f1 =====
 NUC1 ^{13}C
 P1 14.75 usec
 PL1 -1.00 dB
 SF01 125.7942548 MHz

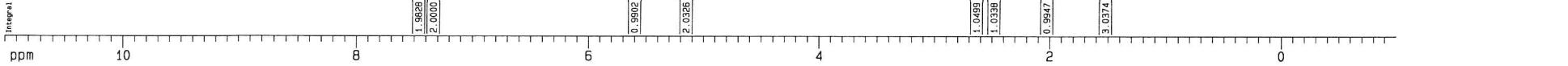
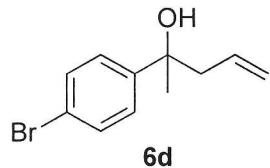
===== CHANNEL f2 =====
 CPDPRG2 waltz16
 NUC2 ^1H
 PCP02 100.00 usec
 PL2 1.60 dB
 PL12 24.80 dB
 SF02 500.2225011 MHz

F2 - Processing parameters
 SI 65536
 SF 125.7804006 MHz
 WDM EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 2.00

1D NMR plot parameters
 CX 22.80 cm
 CY 15.65 cm
 F1P 230.637 ppm
 F1 29009.68 Hz
 F2P -10.287 ppm
 F2 -1293.96 Hz
 PPMCM 10.56668 ppm/cm
 HZCM 1329.10693 Hz/cm







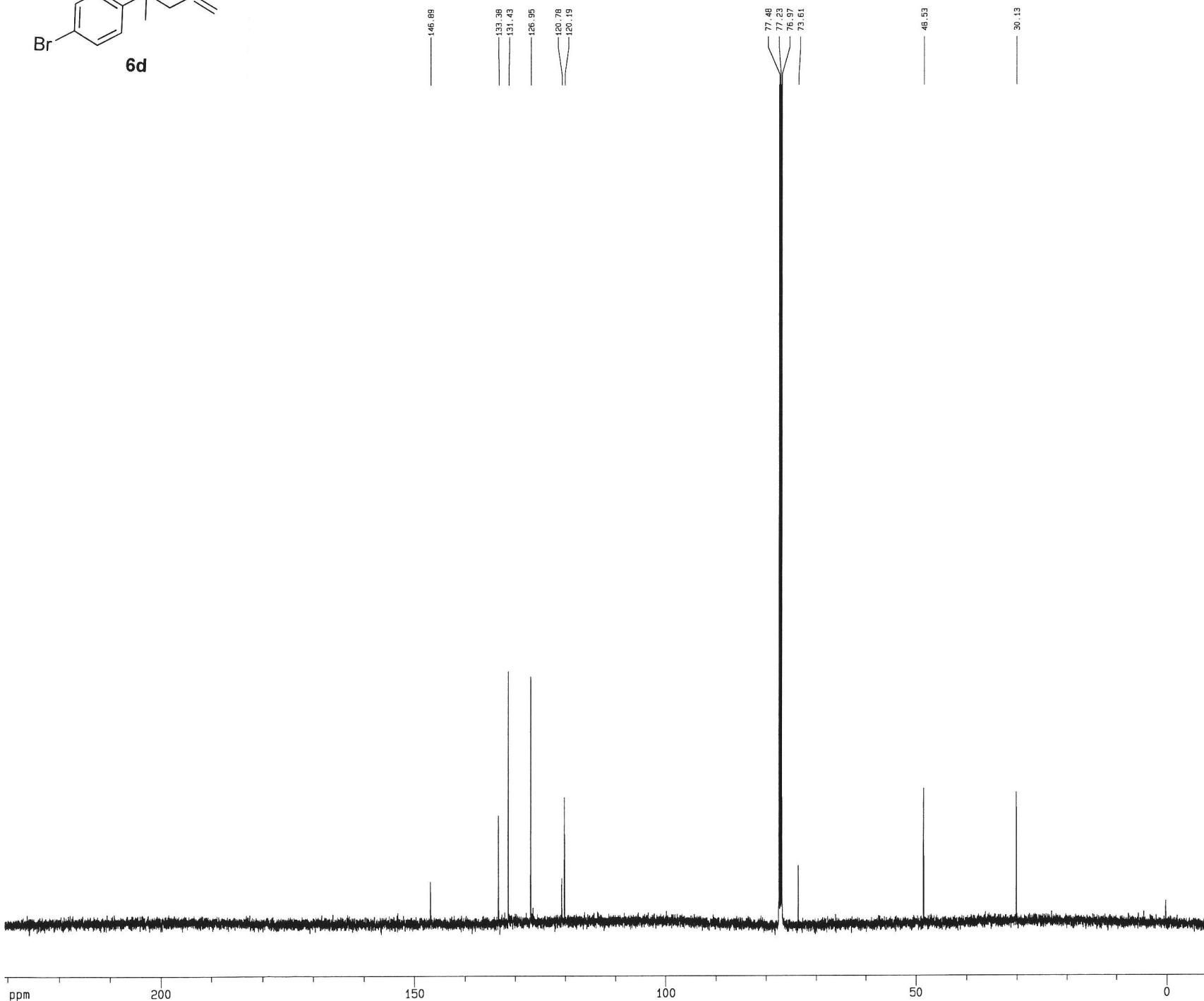
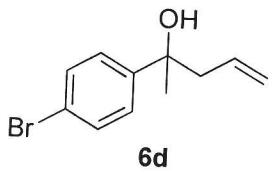
Current Data Parameters
 USER tjbark
 NAME 2-TB-81
 EXPNO 4
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20080801
 Time 16.10
 INSTRUM drx400
 PROBHD 5 mm QNP H/F/P
 PULPROG zg30
 TD 65536
 SOLVENT CDC13T
 NS 8
 DS 2
 SWH 6410.256 Hz
 FIDRES 0.097813 Hz
 AQ 5.1118579 sec
 RG 406.4
 DW 78.000 usec
 DE 4.50 usec
 TE 299.0 K
 D1 0.10000000 sec
 MCREST 0.00000000 sec
 MCWRK 0.01500000 sec

===== CHANNEL f1 =====
 NUC1 1H
 P1 12.00 usec
 PL1 0.00 dB
 SF01 400.1328009 MHz

F2 - Processing parameters
 SI 65536
 SF 400.1300180 MHz
 WDM EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 2.00

1D NMR plot parameters
 CX 22.80 cm
 CY 30.00 cm
 F1P 11.000 ppm
 F1 4401.43 Hz
 F2P -1.000 ppm
 F2 -402.13 Hz
 PPMCM 0.52632 ppm/cm
 HZCM 210.59474 Hz/cm



Current Data Parameters
 USER tjbark
 NAME 2-TJB-17
 EXPNO 2
 PROCN0 1

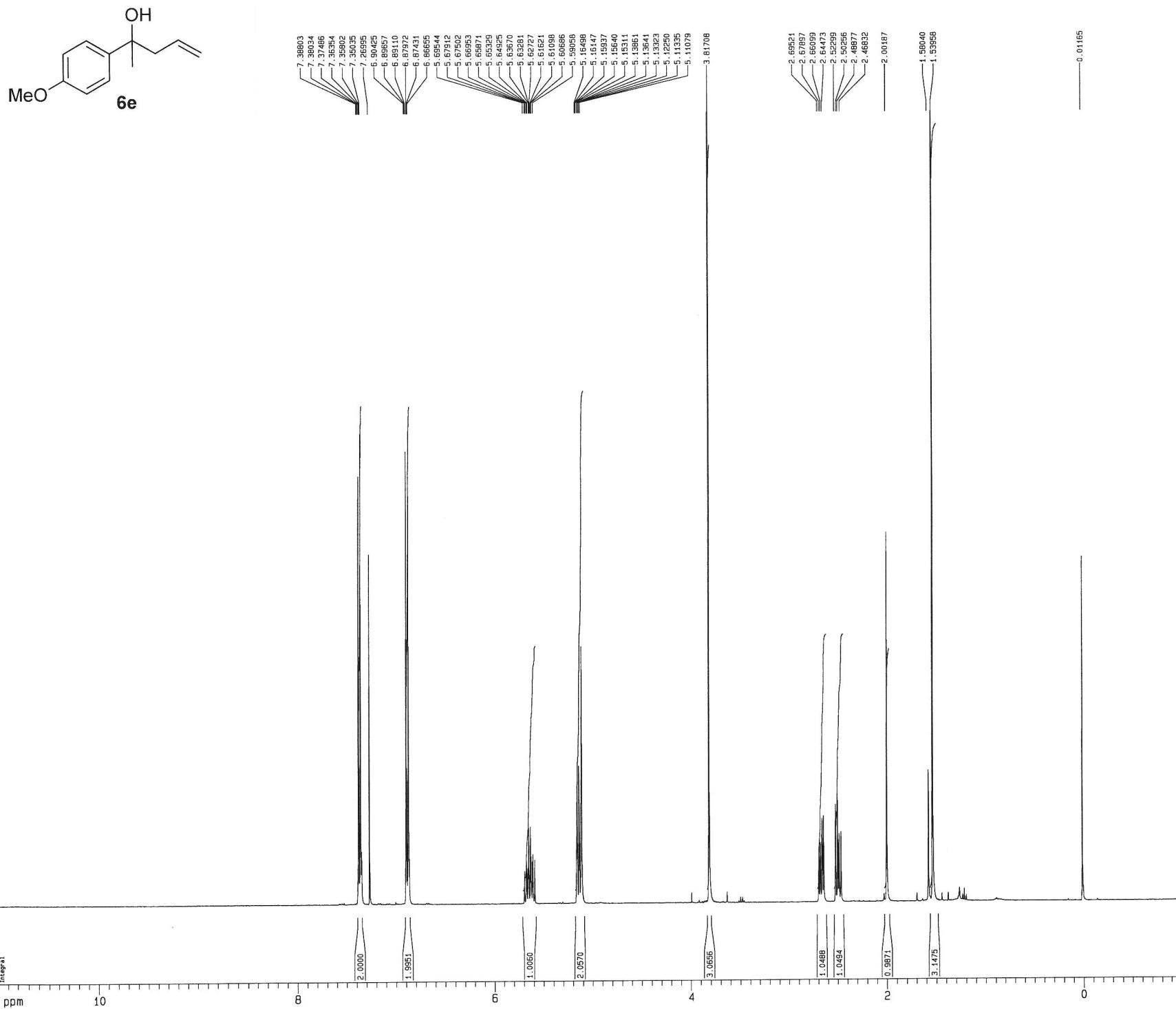
F2 - Acquisition Parameters
 Date_ 20080514
 Time 14.28
 INSTRUM cryo500
 PROBHD 5 mm CPTCI 1H-
 PULPROG zg30
 TD 65536
 SOLVENT CDCl3T
 NS 145
 DS 4
 SWH 30303.031 Hz
 FIORES 0.462388 Hz
 AQ 1.0813840 sec
 RG 13004
 DW 16.500 usec
 DE 5.00 usec
 TE 299.0 K
 D1 0.2500000 sec
 d11 0.0300000 sec
 MCREST 0.0000000 sec
 MCWRK 0.0150000 sec

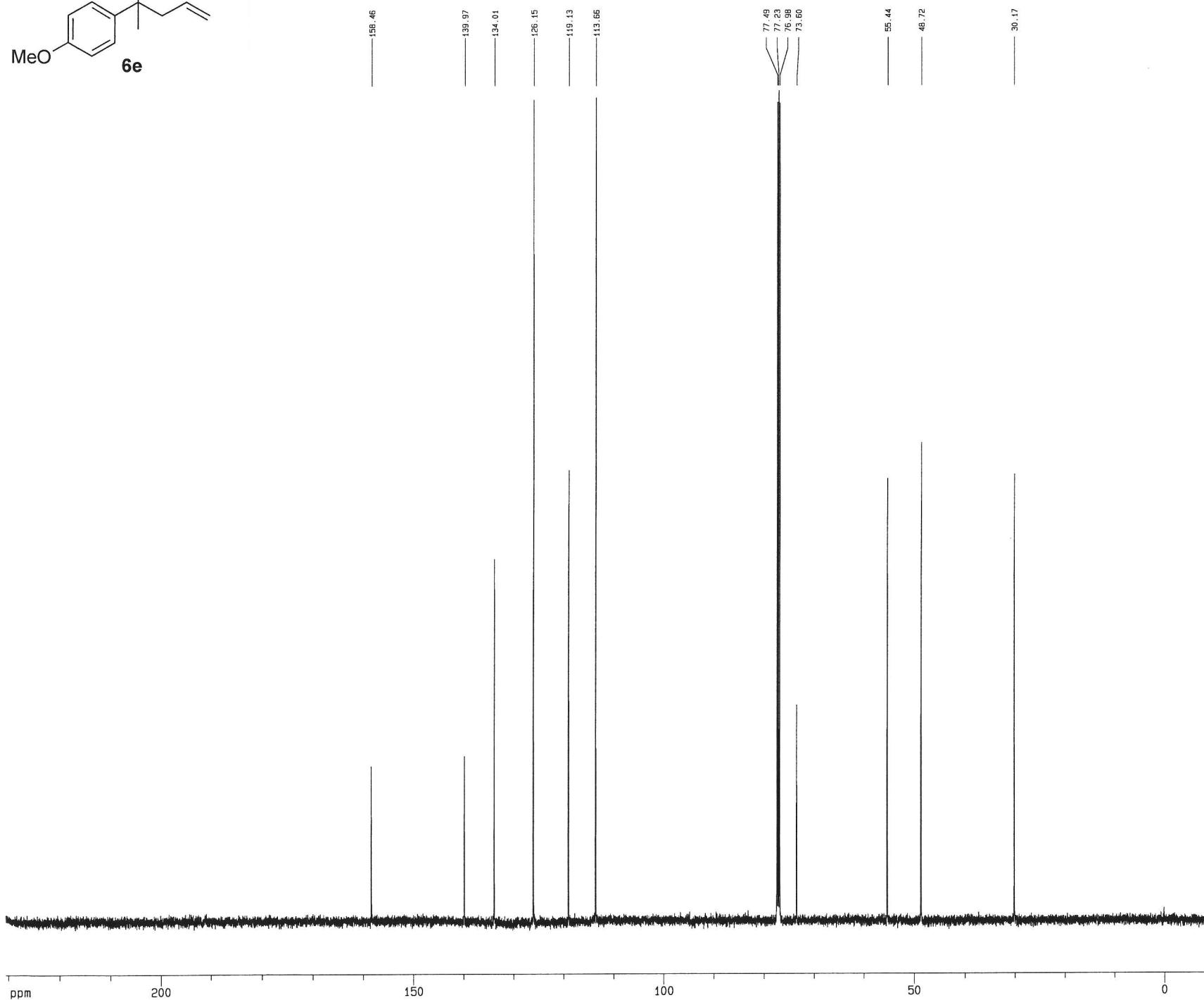
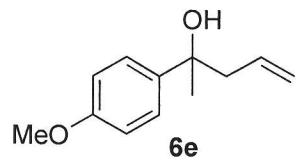
===== CHANNEL f1 =====
 NUC1 13C
 P1 14.75 usec
 PL1 -1.00 dB
 SF01 125.7942548 MHz

===== CHANNEL f2 =====
 CPDPRG2 waltz16
 NUC2 1H
 PCPD2 100.00 usec
 PL2 1.60 dB
 PL12 24.80 dB
 SF02 500.2225011 MHz

F2 - Processing parameters
 SI 65536
 SF 125.7803988 MHz
 WDN EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 2.00

1D NMR plot parameters
 CX 22.80 cm
 CY 30.00 cm
 F1P 230.637 ppm
 F1 29009.67 Hz
 F2P -10.287 ppm
 F2 -1293.96 Hz
 PPMCM 10.56688 ppm/cm
 HZCM 1329.10681 Hz/cm





Current Data Parameters
 USER tjbark
 NAME 2-TJB-76
 EXPNO 6
 PROCNO 1

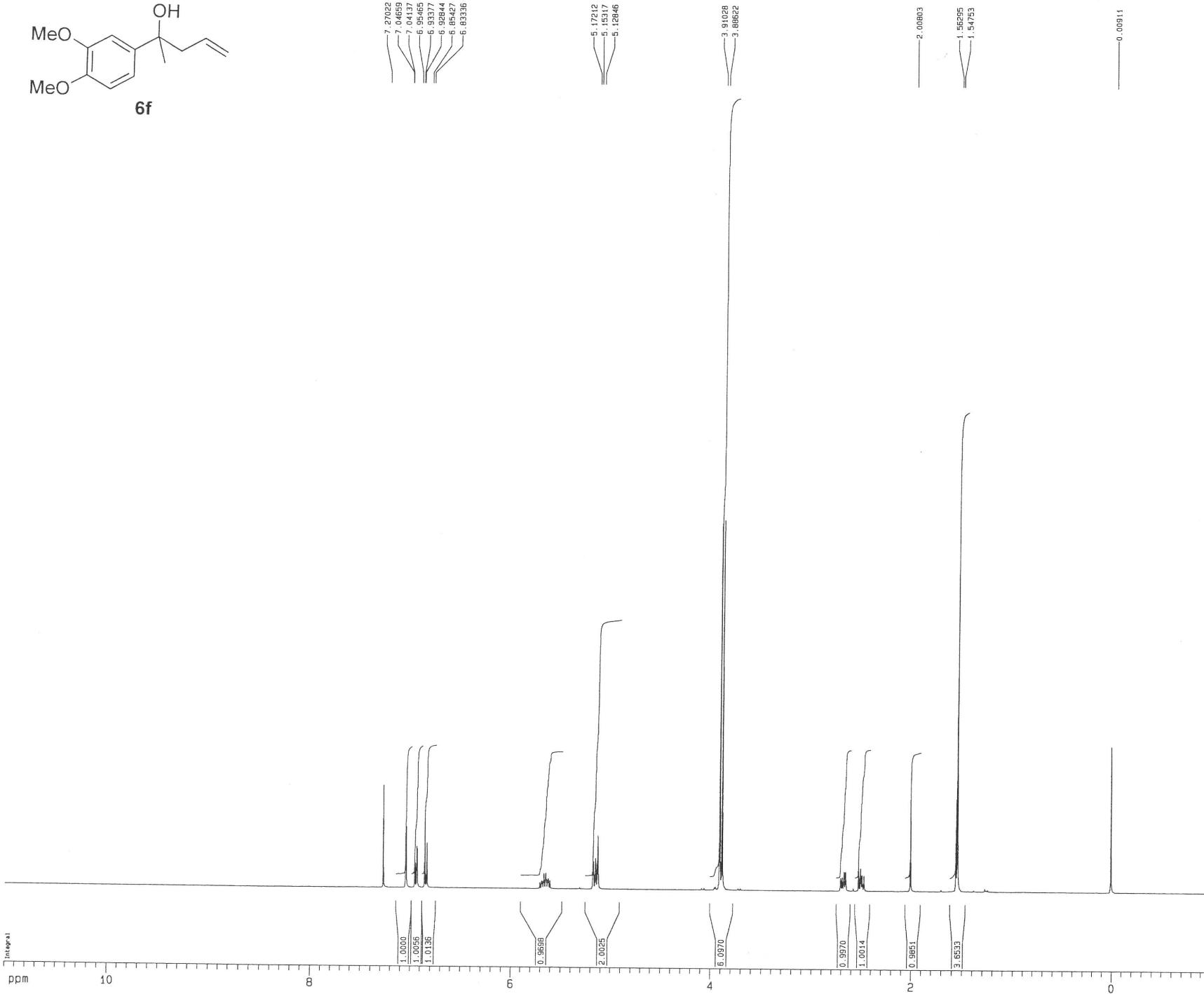
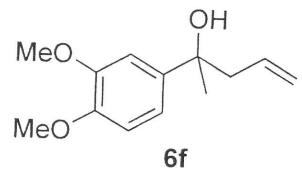
F2 - Acquisition Parameters
 Date_ 20080726
 Time 14.47
 INSTRUM cryo500
 PROBHD 5 mm CPTCI 1H-
 PULPROG zgdc30
 TD 65418
 SOLVENT CDCl3T
 NS 128
 DS 4
 SWH 30303.031 Hz
 FIDRES 0.463222 Hz
 AQ 1.0794470 sec
 RG 13004
 DW 16.500 usec
 DE 6.00 usec
 TE 298.0 K
 D1 0.2500000 sec
 d11 0.0300000 sec
 MCREST 0.0000000 sec
 MCWRK 0.0150000 sec

***** CHANNEL f1 *****
 NUC1 13C
 P1 14.75 usec
 PL1 -1.00 dB
 SF01 125.7942548 MHz

***** CHANNEL f2 *****
 CPDPRG2 waltz16
 NUC2 1H
 PCPD2 100.00 usec
 PL2 1.60 dB
 PL12 24.80 dB
 SF02 500.2225011 MHz

F2 - Processing parameters
 SI 65536
 SF 125.7804006 MHz
 WDW EM
 SS8 0
 LB 1.00 Hz
 GB 0
 PC 2.00

1D NMR plot parameters
 CX 22.80 cm
 CY 15.65 cm
 F1P 230.637 ppm
 F1 2909.68 Hz
 F2P -10.287 ppm
 F2 -1293.96 Hz
 PPWCM 10.55688 ppm/cm
 HZCM 1329.10693 Hz/cm



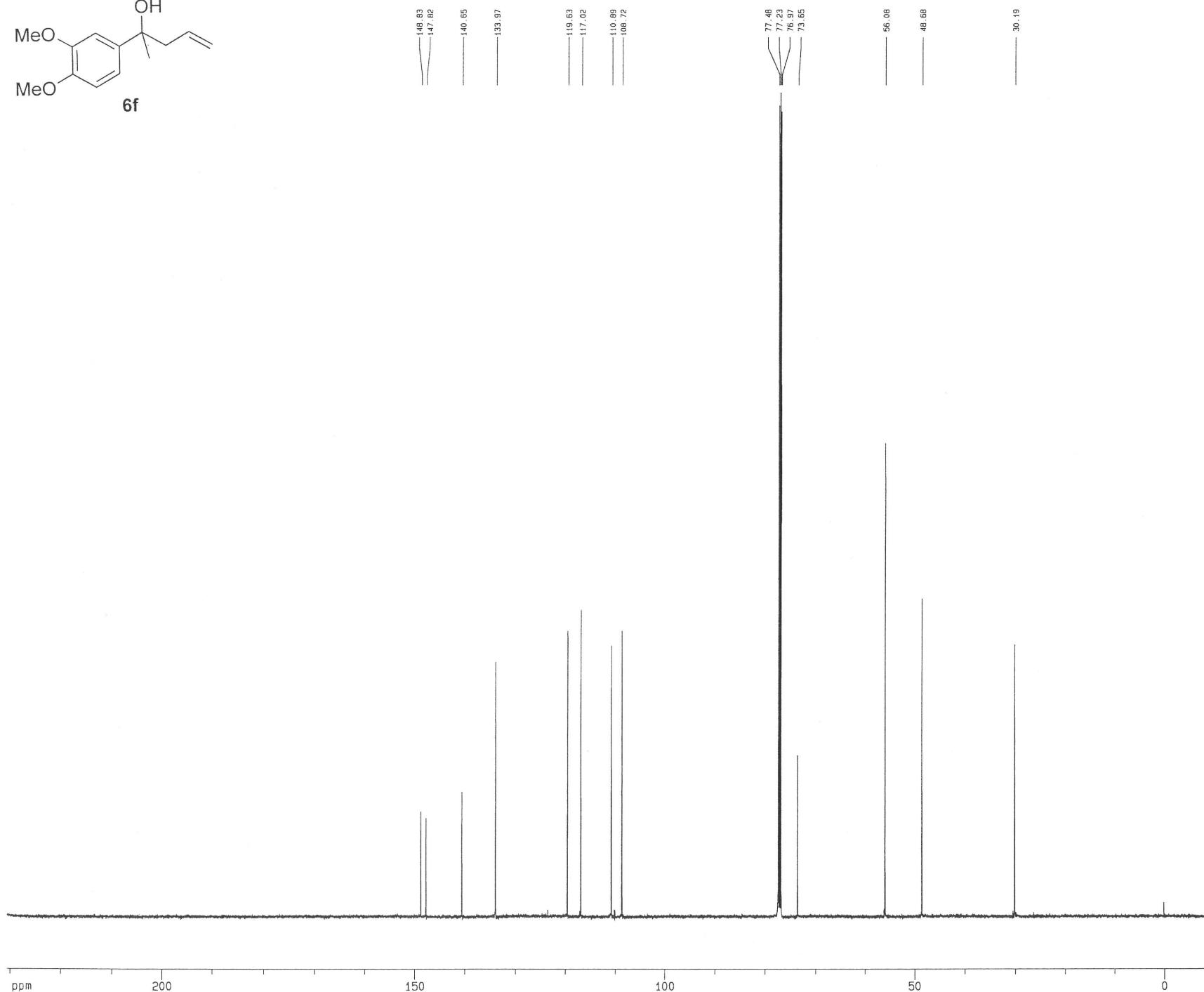
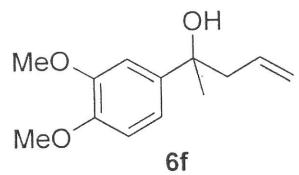
Current Data Parameters
 USER tbark
 NAME 2-TJB-46
 EXPNO 5
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20080705
 Time 13.20
 INSTRUM drx400
 PROBHD 5 mm DNP H/F/P
 PULPROG zg30
 TD 65536
 SOLVENT CDCl3T
 NS 8
 DS 2
 SWH 6410.256 Hz
 FIDRES 0.097813 Hz
 AQ 5.1118579 sec
 RG 645.1
 DM 78.000 usec
 DE 4.50 usec
 TE 298.0 K
 D1 0.1000000 sec
 MCREST 0.0000000 sec
 MCWRK 0.01500000 sec

===== CHANNEL f1 =====
 NUC1 1H
 P1 12.00 usec
 PL1 0.00 dB
 SF01 400.1326009 MHz

F2 - Processing parameters
 SI 65536
 SF 400.1300179 MHz
 WDW EM
 SS8 0
 LB 0.30 Hz
 GB 0
 PC 2.00

1D NMR plot parameters
 CX 22.80 cm
 CY 7.00 cm
 F1P 11.000 ppm
 F1 4401.43 Hz
 F2P -1.000 ppm
 F2 -400.13 Hz
 PPMCM 0.52632 ppm/cm
 HZCM 210.59474 Hz/cm



Current Data Parameters
 USER tibank
 NAME 2-TJB-20
 EXPNO 5
 PROCNO 1

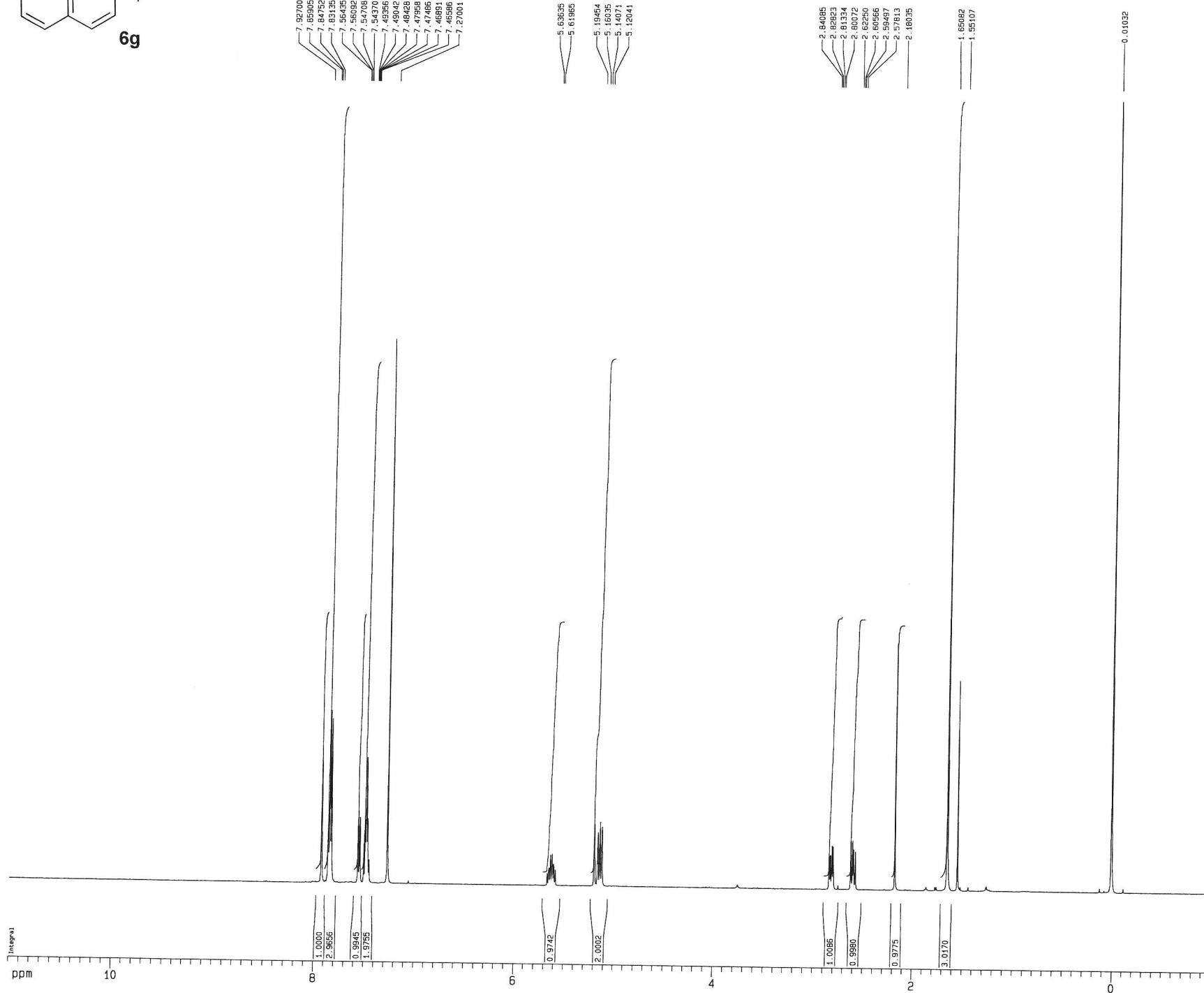
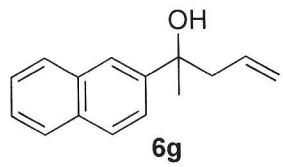
F2 - Acquisition Parameters
 Date_ 20080621
 Time 11.11
 INSTRUM cryo500
 PROBHD 5 mm CPTCI 1H-
 PULPROG zgdc30
 TD 65416
 SOLVENT CDCl3T
 NS 636
 DS 4
 SWH 30393.031 Hz
 FIDRES 0.463222 Hz
 AQ 1.0794470 sec
 RG 9195.2
 DW 16.500 usec
 DE 5.00 usec
 TE 298.0 K
 D1 0.2500000 sec
 d11 0.0300000 sec
 MCREST 0.0000000 sec
 MCWRK 0.0150000 sec

===== CHANNEL f1 =====
 NUC1 13C
 P1 14.75 usec
 PL1 -1.00 dB
 SF01 125.7942548 MHz

===== CHANNEL f2 =====
 CPDPRG2 waltz16
 NUC2 1H
 PCPQ2 100.00 usec
 PL2 1.60 dB
 PL12 24.80 dB
 SF02 500.2225011 MHz

F2 - Processing parameters
 SI 65536
 SF 125.7904011 MHz
 WDM EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 2.00

1D NMR plot parameters
 CX 22.80 cm
 CY 15.65 cm
 F1P 230.603 ppm
 F1 29005.29 Hz
 F2P -10.318 ppm
 F2 -1297.74 Hz
 PPWCM 10.56667 ppm/cm
 HZCM 1329.08044 Hz/cm



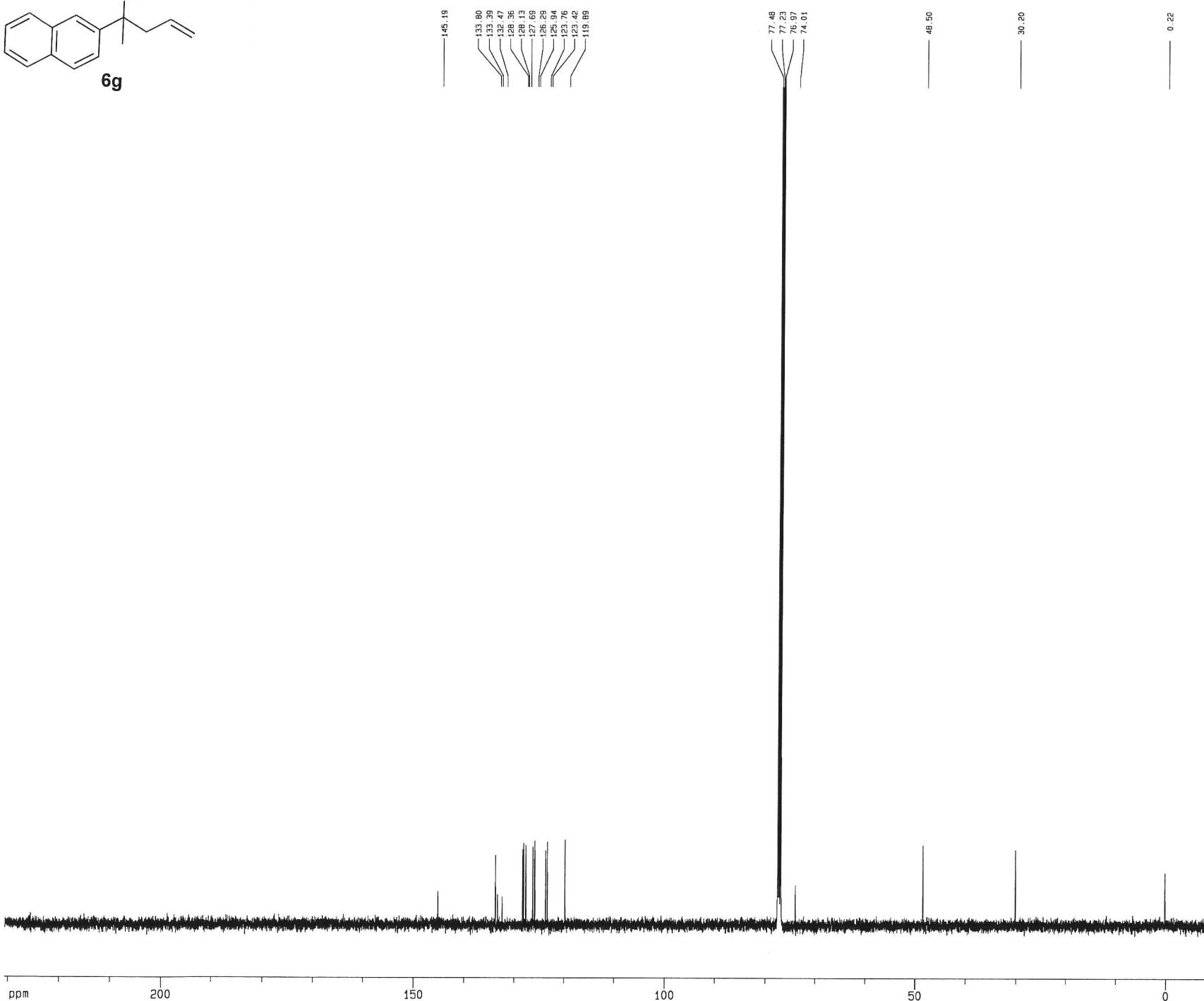
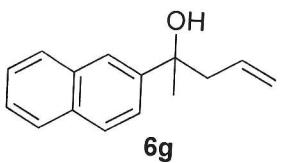
Current Data Parameters
 USER tjbark
 NAME 1-TB-295
 EXPNO 6
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20080614
 Time 14.08
 INSTRUM cryo500
 PROBH0D 5 mm CPTCI 1H-
 PULPROG zg30
 TD 81728
 SOLVENT CDCl3
 NS 8
 DS 2
 SWH 8012.820 Hz
 FIDRES 0.098043 Hz
 AQ 5.0998774 sec
 RG 18
 DM 62.400 usec
 DE 6.00 usec
 TE 299.0 K
 D1 0.1000000 sec
 MCWRT 0.0000000 sec
 MCWRK 0.0150000 sec

===== CHANNEL f1 =====
 NUC1 1H
 P1 7.38 usec
 PL1 1.60 dB
 SF01 500.2235015 MHz

F2 - Processing parameters
 SI 65536
 SF 500.2200262 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 4.00

1D NMR plot parameters
 CX 22.80 cm
 CY 15.00 cm
 F1P 11.000 ppm
 F1 5502.42 Hz
 F2P -1.000 ppm
 F2 -500.22 Hz
 PPMM 0.52632 ppm/cm
 HZCM 263.27371 Hz/cm



Current Data Parameters
 USER *tibark*
 NAME *1-TB-295*
 EXPNO 5
 PROCNO 1

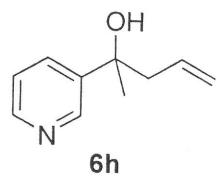
F2 - Acquisition Parameters
 Date_ 20080514
 Time 13.53
 INSTRUM cryo500
 PROBHD 5 mm CP/TCI 1H-
 PULPROG zgdc30
 TD 65418
 SOLVENT CDCl3T
 NS 602
 DS 4
 SWH 30303.031 Hz
 FIDRES 0.453222 Hz
 AQ 1.0734470 sec
 RG 13004
 DM 16.500 usec
 DE 6.00 usec
 TE 299.0 K
 D1 0.2500000 sec
 d11 0.0300000 sec
 MCDEST 0.0000000 sec
 MCWRK 0.01500000 sec

===== CHANNEL f1 ======
 NUC1 13C
 P1 14.75 usec
 PL1 -1.00 dB
 SF01 125.7942548 MHz

===== CHANNEL f2 ======
 CPDPRG2 waltz16
 NUC2 1H
 PCPD2 100.00 usec
 PL2 1.60 dB
 PL12 24.80 dB
 SFQ2 500.2225011 MHz

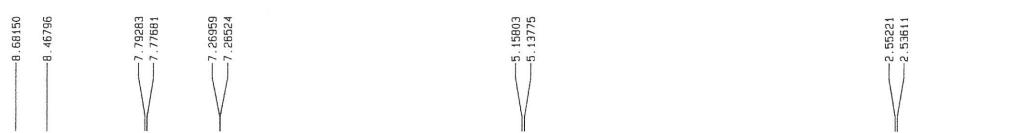
F2 - Processing parameters
 SI 65536
 SF 125.7803983 MHz
 WDM EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 2.00

1D NMR plot parameters
 CX 22.80 cm
 CY 45.00 cm
 F1P 230.625 ppm
 F1 29008.05 Hz
 F2P -10.296 ppm
 F2 -1294.98 Hz
 PPWCM 10.56667 ppm/cm
 HZCM 1329.08032 Hz/cm



Integral

10 8 6 4 2 0 1



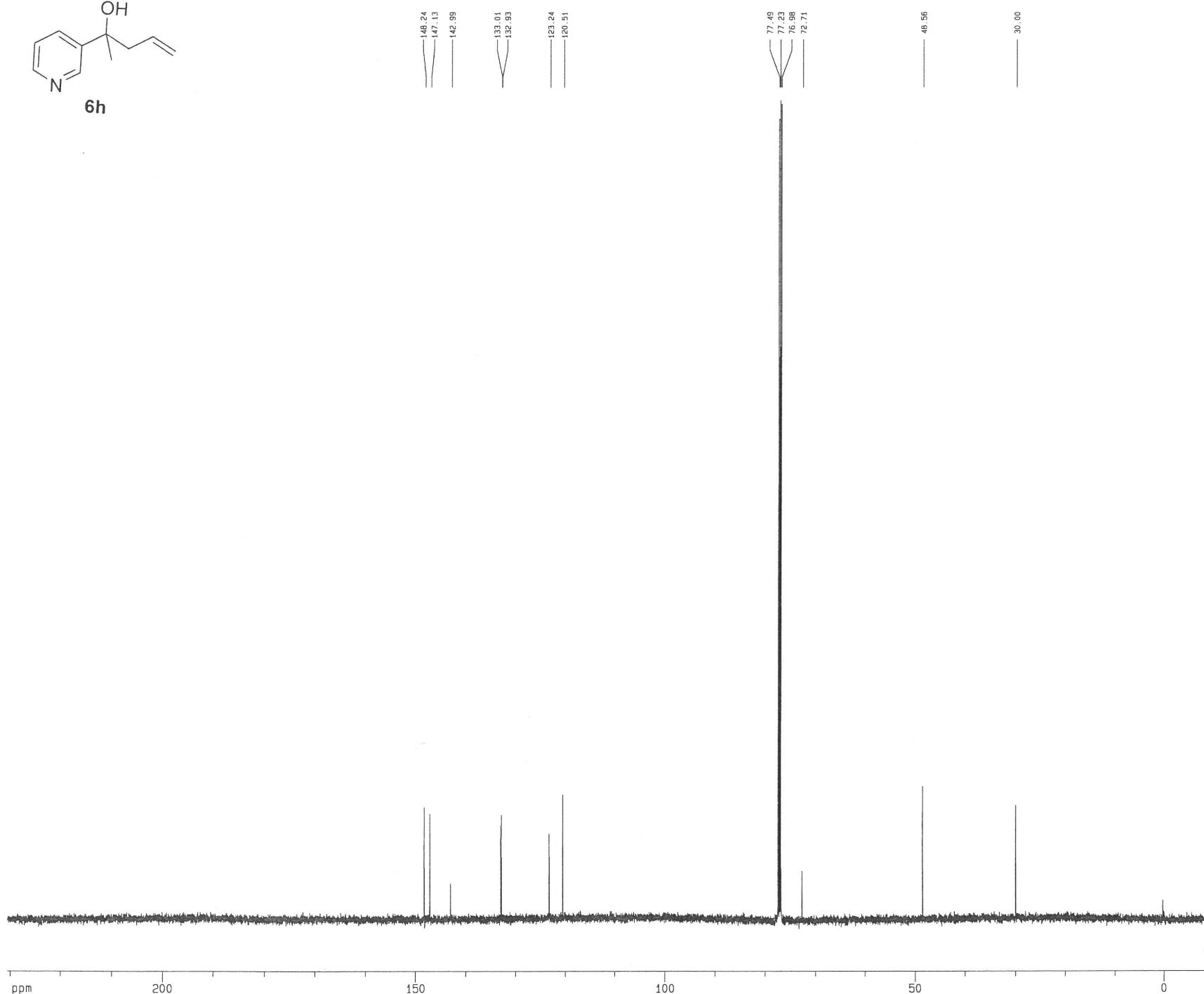
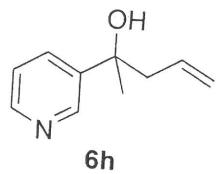
Current Data Parameters
 USER tbark
 NAME 1-TJB-298
 EXPNO 7
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20080827
 Time 8.21
 INSTRUM cryo500
 PROBHD 5 mm CPTCI 1H-
 PULPROG zg30
 TD 81728
 SOLVENT CDCl3T
 NS 8
 DS 2
 SWH 8012.820 Hz
 FIDRES 0.098043 Hz
 AQ 5.0998774 sec
 RG 6.3
 DW 62.400 usec
 DE 6.00 usec
 TE 298.0 K
 D1 0.1000000 sec
 MCREST 0.0000000 sec
 MCWRK 0.01500000 sec

===== CHANNEL f1 =====
 NUC1 1H
 PI 7.38 usec
 PL1 1.60 dB
 SF01 500.223501 MHz

F2 - Processing parameters
 SI 65536
 SF 500.2200289 MHz
 MW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 4.00

1D NMR plot parameters
 CX 22.80 cm
 CY 5.00 cm
 F1P 11.000 ppm
 F1 5502.42 Hz
 F2P -1.000 ppm
 F2 -500.22 Hz
 PPMCM 0.52632 ppm/cm
 HZCM 263.27371 Hz/cm



Current Data Parameters
 USER tjbark
 NAME 2-TJB-24
 EXPNO 6
 PROCNO 1

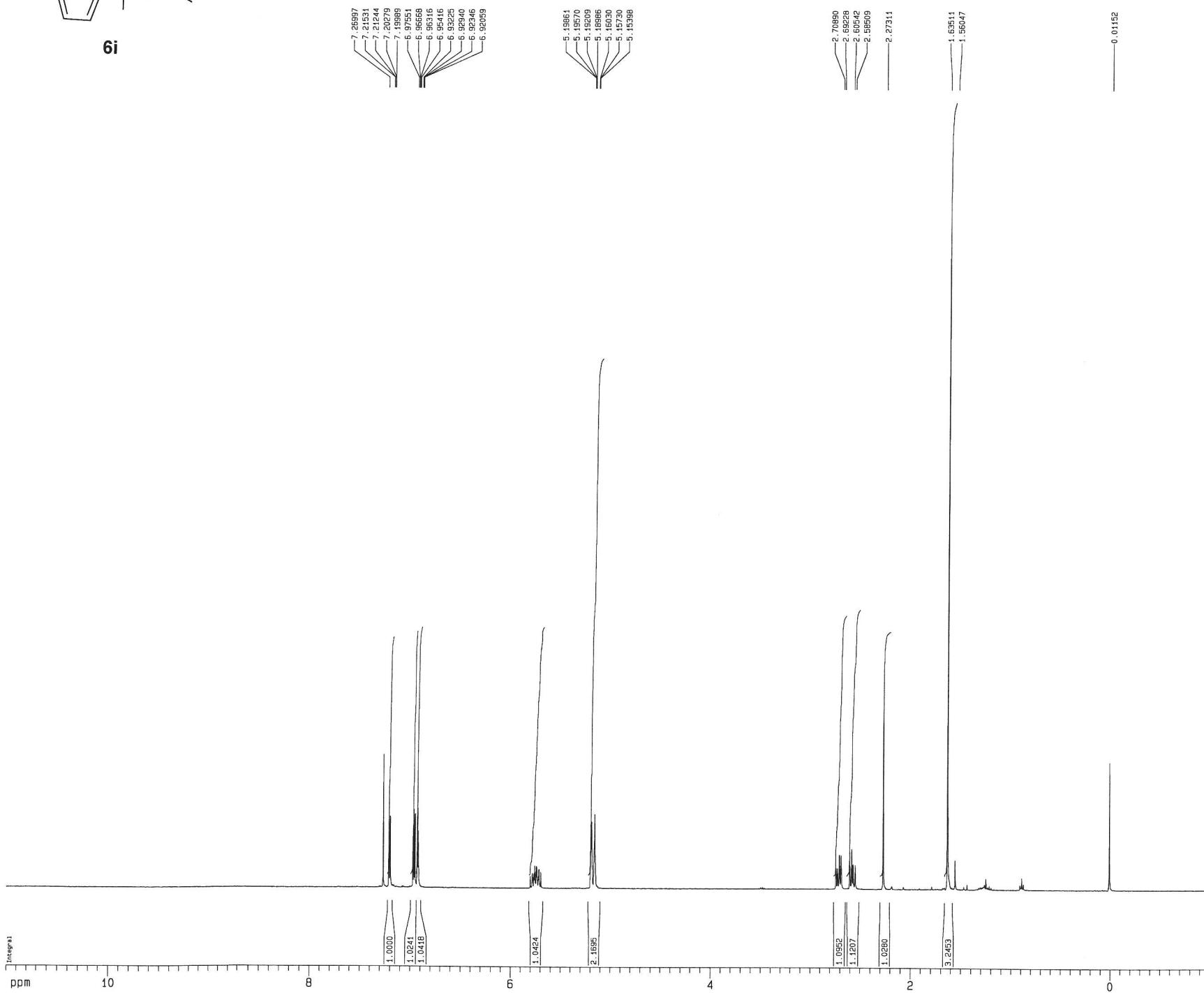
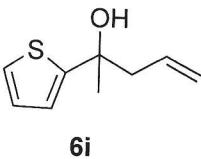
F2 - Acquisition Parameters
 Date_ 20080521
 Time 11.01
 INSTRUM cryo500
 PROBHD 5 mm CPTCI 1H-
 PULPROG zgdc30
 TD 65536
 SOLVENT CDCl3T
 NS 108
 DS 4
 SWH 30303.031 Hz
 FIDRES 0.452388 Hz
 AQ 1.0813940 sec
 RG 14595.5
 DW 16.500 usec
 DE 6.00 usec
 TE 298.0 K
 D1 0.2500000 sec
 d11 0.0300000 sec
 MCREST 0.0000000 sec
 MCWRFK 0.0150000 sec

***** CHANNEL f1 *****
 NUC1 13C
 P1 14.75 usec
 PL1 -1.00 dB
 SF01 125.7942548 MHz

***** CHANNEL f2 *****
 CPDPFG2 waltz16
 NUC2 1H
 PCPQ2 100.00 usec
 PL2 1.60 dB
 PL12 24.80 dB
 SF02 500.2225011 MHz

F2 - Processing parameters
 SI 65536
 SF 125.7803988 MHz
 MW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 2.00

1D NMR plot parameters
 CX 22.80 cm
 CY 15.65 cm
 F1P 230.637 ppm
 F1 29009.67 Hz
 F2P -10.287 ppm
 F2 -1293.96 Hz
 PPMCM 10.56688 ppm/cm
 HZCM 1329.10681 Hz/cm



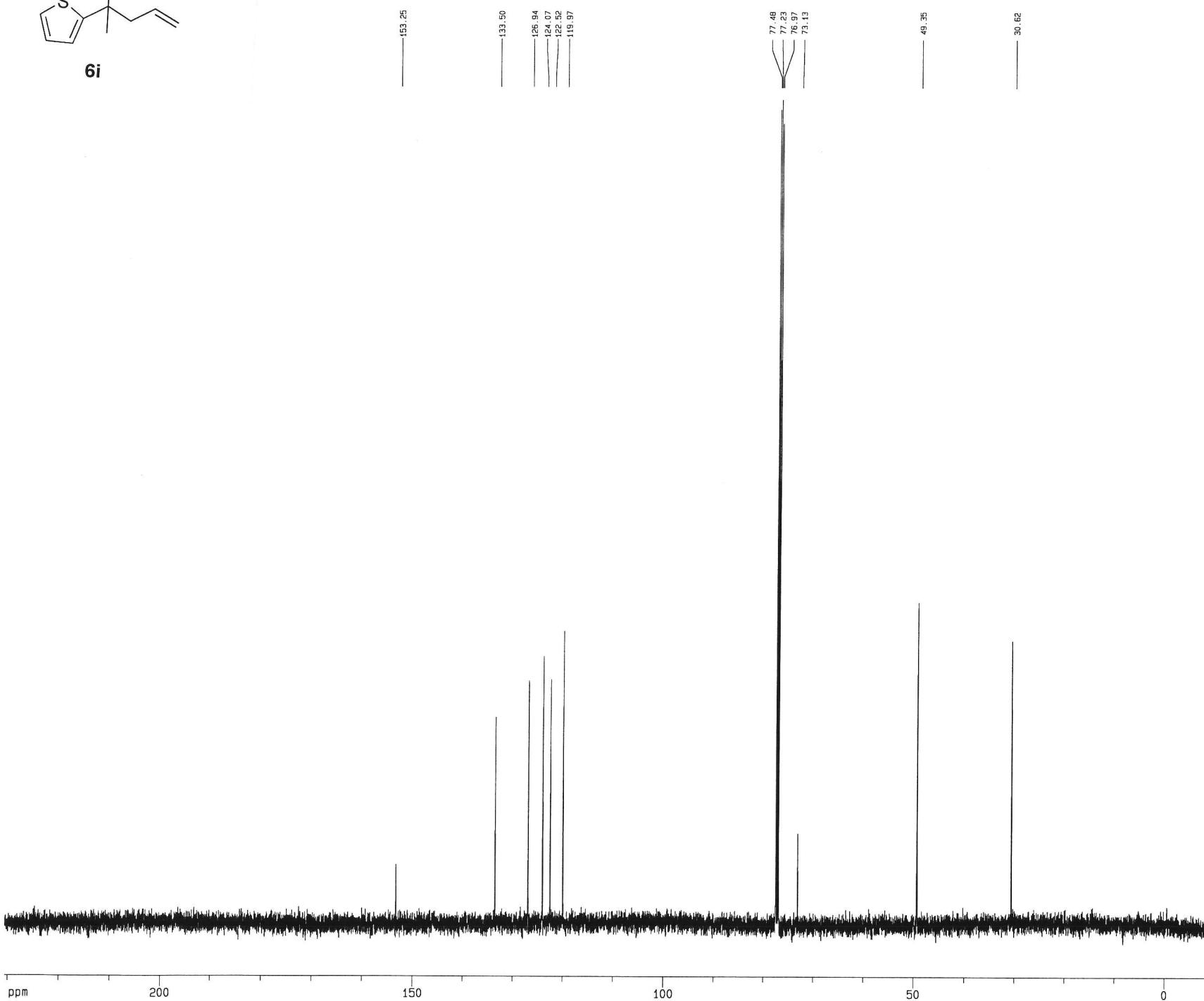
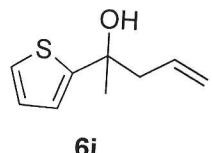
Current Data Parameters
 USER t1dark
 NAME 2-TJB-32
 EXPNO 8
 PROCNO 1

F2 - Acquisition Parameters
 Date- 2008120
 Time 8.52
 INSTRUM dtx400
 PROBHD 5 mm QNP H/F/P
 PULPROG zg30
 TD 65536
 SOLVENT CDCl3T
 NS 8
 DS 2
 SWH 6410.256 Hz
 FIDRES 0.097813 Hz
 AQ 5.1118579 sec
 RG 512
 DW 78.000 usec
 DE 4.50 usec
 TE 298.1 K
 D1 0.1000000 sec
 MCREST 0.0000000 sec
 MCW琪 0.0150000 sec

===== CHANNEL f1 =====
 NUC1 1H
 P1 12.00 usec
 PL1 0.00 dB
 SF01 400.1326009 MHz

F2 - Processing parameters
 SI 65536
 SF 400.1300179 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 2.00

1D NMR plot parameters
 CX 22.80 cm
 CY 12.00 cm
 F1P 11.000 ppm
 F1 4401.43 Hz
 F2P -1.000 ppm
 F2 -400.13 Hz
 PPMCM 0.52632 ppm/cm
 HZCM 210.59474 Hz/cm



Current Data Parameters
 USER tjbark
 NAME 2-TJB-32
 EXPNO 4
 PROCN0 1

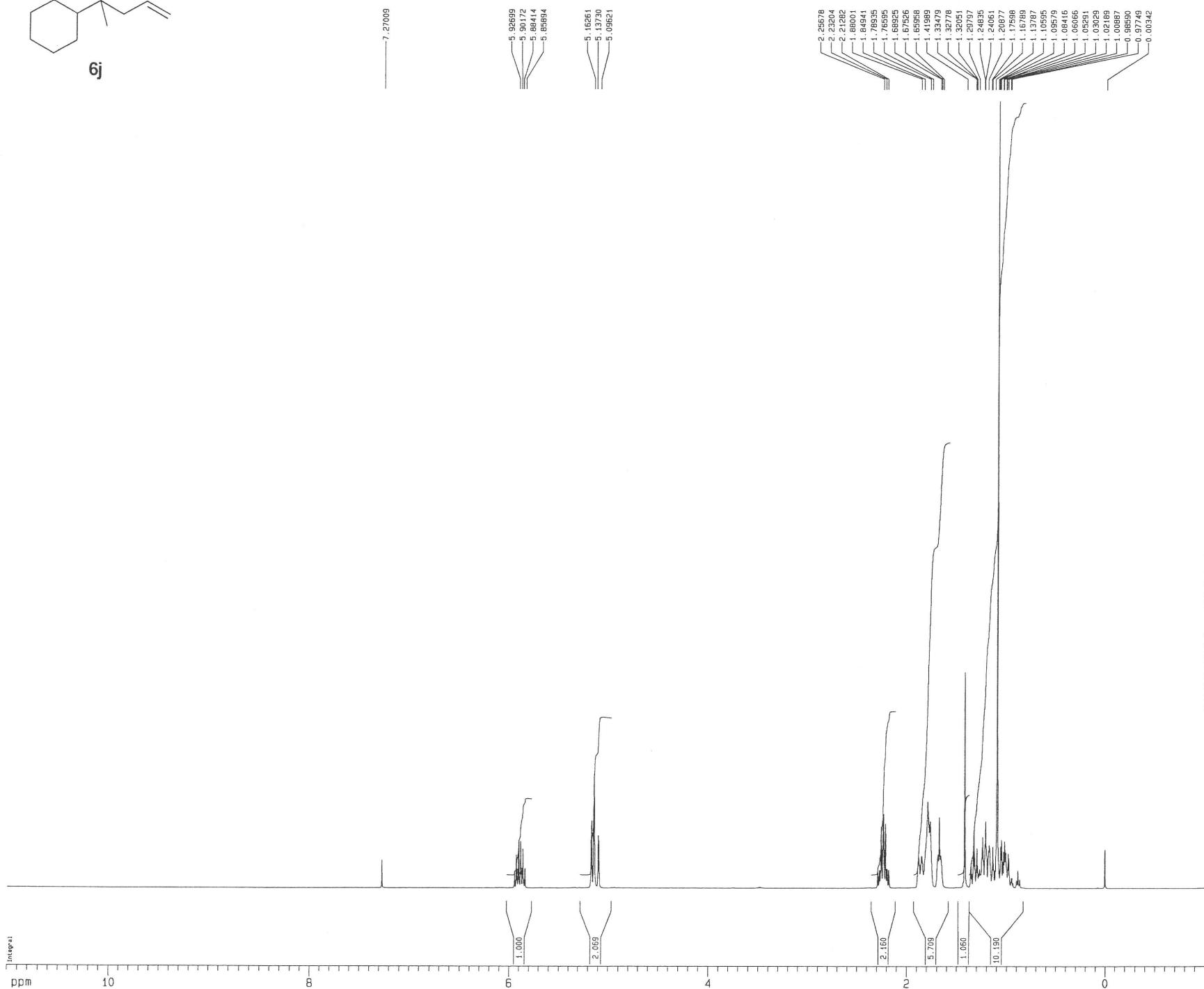
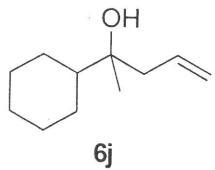
F2 - Acquisition Parameters
 Date_ 20080621
 Time 10.54
 INSTRUM cryo500
 PROBHD 5 mm OPTCI 1H-
 PULPROG zg30
 TD 65536
 SOLVENT CDC13T
 NS 33
 DS 4
 SWH 30303.031 Hz
 FIDRES 0.462388 Hz
 AQ 1.0813940 sec
 RG 13004
 DW 16.500 usec
 DE 6.00 usec
 TE 299.0 K
 D1 0.2500000 sec
 d11 0.0300000 sec
 MCRES 0.0000000 sec
 MCWKR 0.01500000 sec

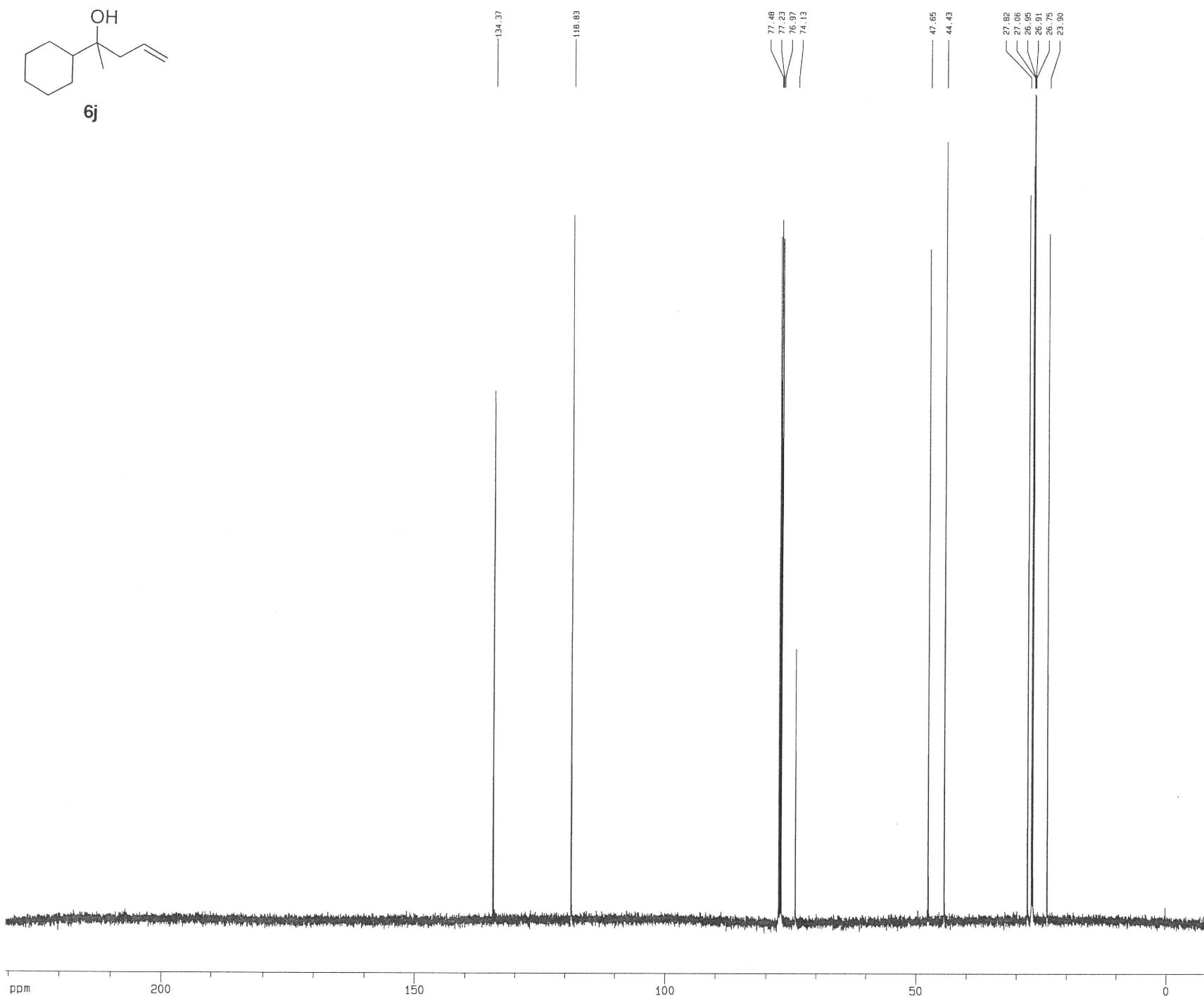
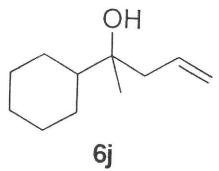
===== CHANNEL f1 =====
 NUC1 13C
 P1 14.75 usec
 PL1 -1.00 dB
 SF01 125.7942548 MHz

===== CHANNEL f2 =====
 CPDPRG2 waltz16
 NUC2 1H
 PCPD2 100.00 usec
 PL2 1.60 dB
 PL12 24.80 dB
 SF02 500.2225011 MHz

F2 - Processing parameters
 SI 65536
 SF 125.7803997 MHz
 WDM EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 2.00

1D NMR plot parameters
 CX 22.80 cm
 CY 15.65 cm
 F1P 230.637 ppm
 F1 29009.68 Hz
 F2P -10.287 ppm
 F2 -1293.96 Hz
 PPWCM 10.56688 ppm/cm
 HZCM 1329.10693 Hz/cm





Current Data Parameters
 USER tbark
 NAME 2-TJB-82
 EXPNO 3
 PROCN 1

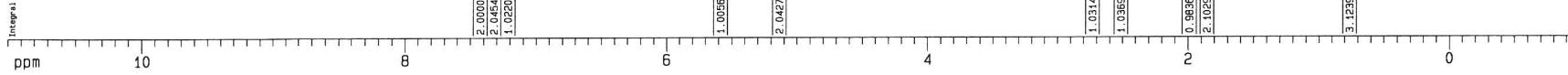
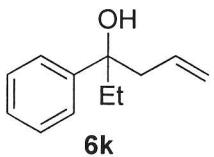
F2 - Acquisition Parameters
 Date_ 20080826
 Time 19.26
 INSTRUM cryo500
 PROBHD 5 mm CPTCI 1H
 PULPROG zg30
 TD 65536
 SOLVENT CDCl3T
 NS 128
 DS 4
 SWH 30303.031 Hz
 FIDRES 0.462388 Hz
 AQ 1.0813940 sec
 RG 13004
 DW 16.500 usec
 DE 6.00 usec
 TE 298.0 K
 D1 0.2500000 sec
 d11 0.0300000 sec
 MCREST 0.0000000 sec
 MCRAK 0.0150000 sec

***** CHANNEL f1 *****
 NUC1 13C
 P1 14.75 usec
 PL1 -1.00 dB
 SF01 125.7942548 MHz

***** CHANNEL f2 *****
 CPDPRG2 waltz16
 NUC2 1H
 PCP02 100.00 usec
 PL2 1.60 dB
 PL12 24.80 dB
 SF02 500.2225011 MHz

F2 - Processing parameters
 SI 65536
 SF 125.7804002 MHz
 WDM EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 2.00

1D NMR plot parameters
 CX 22.80 cm
 CY 15.65 cm
 F1P 290.637 ppm
 F1 29009.68 Hz
 F2P -10.287 ppm
 F2 -1293.96 Hz
 PPMCM 10.56688 ppm/cm
 HZCM 1329.10693 Hz/cm



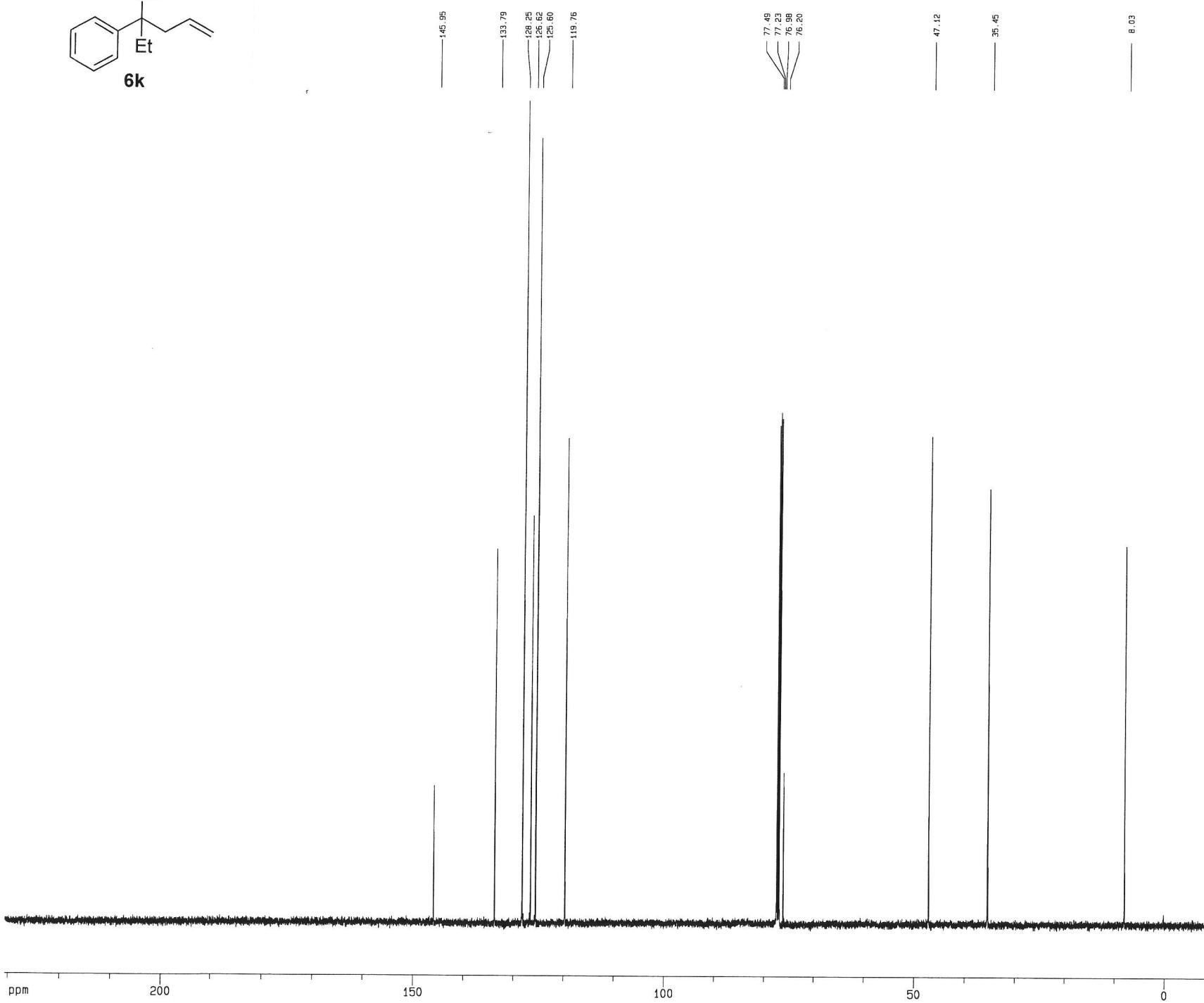
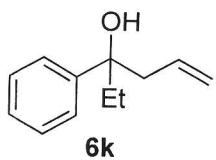
Current Data Parameters
 USER tjbark
 NAME 2-TJ6-37
 EXPNO 2
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20080625
 Time 9.26
 INSTRUM drx400
 PROBHD 5 mm QNP H/F/P
 PULPROG zg30
 TD 65536
 SOLVENT CDCl3T
 NS 8
 DS 2
 SWH 6410.256 Hz
 FIDRES 0.097813 Hz
 AQ 5.1118579 sec
 RG 912.3
 DW 78.000 usec
 DE 4.50 usec
 TE 298.0 K
 D1 0.1000000 sec
 MCREST 0.0000000 sec
 MCWRK 0.0150000 sec

===== CHANNEL f1 =====
 NUC1 1H
 P1 12.00 usec
 PL1 0.00 dB
 SF01 400.1328009 MHz

F2 - Processing parameters
 SI 65536
 SF 400.1300179 MHz
 WM EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 2.00

1D NMR plot parameters
 CX 22.80 cm
 CY 10.00 cm
 F1P 11.000 ppm
 F1 4401.43 Hz
 F2P -1.000 ppm
 F2 -400.13 Hz
 PPMCM 0.52632 ppm/cm
 HZCM 210.59474 Hz/cm



Current Data Parameters
 USER tjbark
 NAME 2-TJB-37
 EXPNO 4
 PROCN0 1

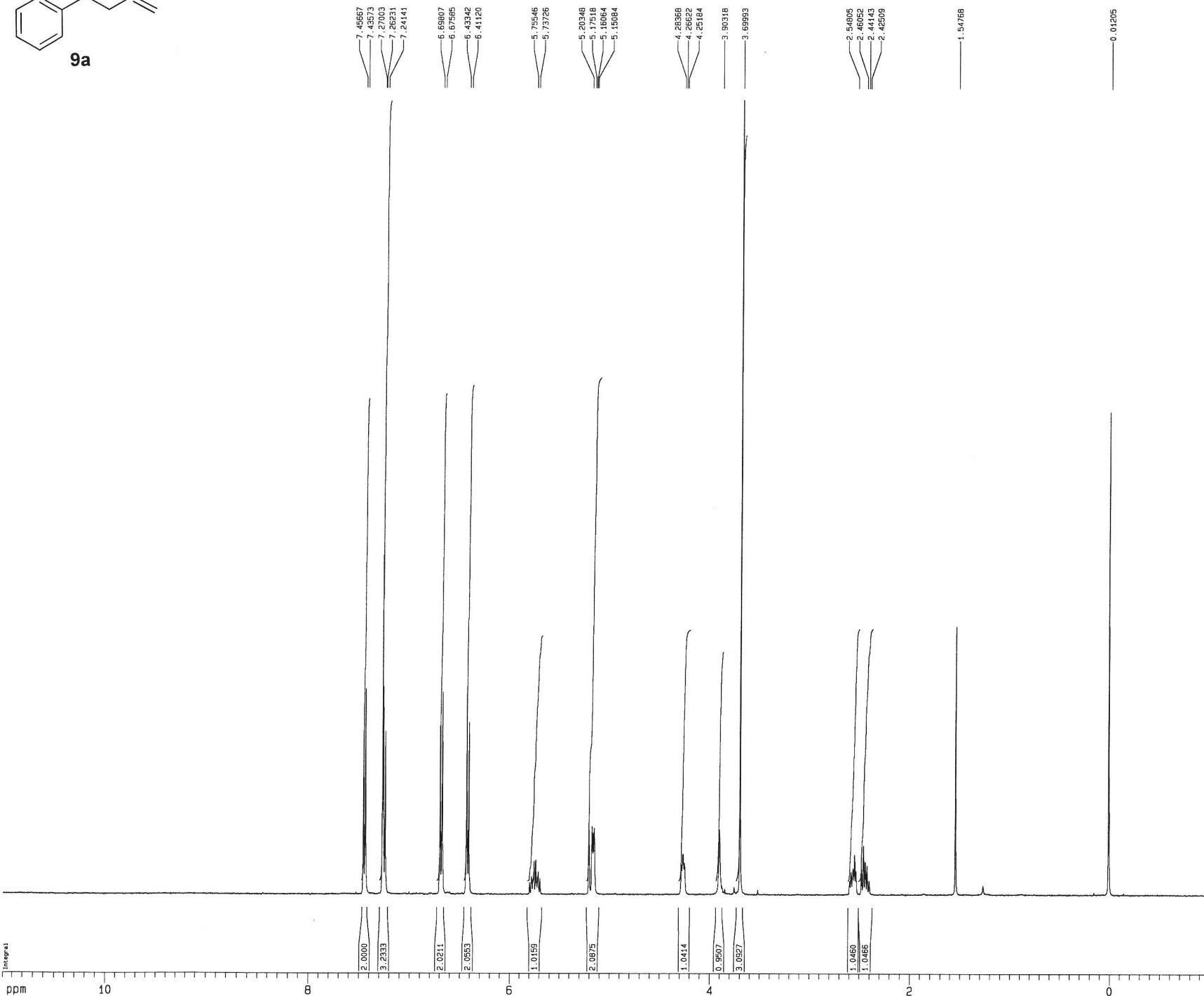
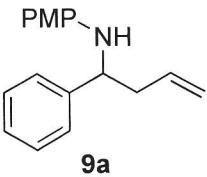
F2 - Acquisition Parameters
 Date_ 20080628
 Time 11.27
 INSTRUM cryo500
 PROBHD 5 mm CPTCI 1H-
 PULPROG zgdc30
 TD 65418
 SOLVENT CDCl3T
 NS 128
 DS 4
 SWH 30303.031 Hz
 FIDRES 0.463222 Hz
 AQ 1.0794470 sec
 RG 14596.5
 DW 16.300 usec
 DE 6.00 usec
 TE 299.0 K
 D1 0.2500000 sec
 d11 0.03000000 sec
 MCREST 0.00000000 sec
 MCWRFK 0.01500000 sec

===== CHANNEL f1 ======
 NUC1 13C
 P1 14.75 usec
 PL1 -1.00 dB
 SF01 125.7942548 MHz

===== CHANNEL f2 ======
 CPDPRG2 waltz16
 NUC2 1H
 PCP02 100.00 usec
 PL2 1.60 dB
 PL12 24.80 dB
 SF02 500.2225011 MHz

F2 - Processing parameters
 SI 65536
 SF 125.7804006 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 2.00

1D NMR plot parameters
 CX 22.80 cm
 CY 15.66 cm
 F1P 230.606 ppm
 F1 29005.75 Hz
 F2P -10.314 ppm
 F2 -1297.28 Hz
 PPMCM 10.56667 ppm/cm
 HZCM 1329.08044 Hz/cm



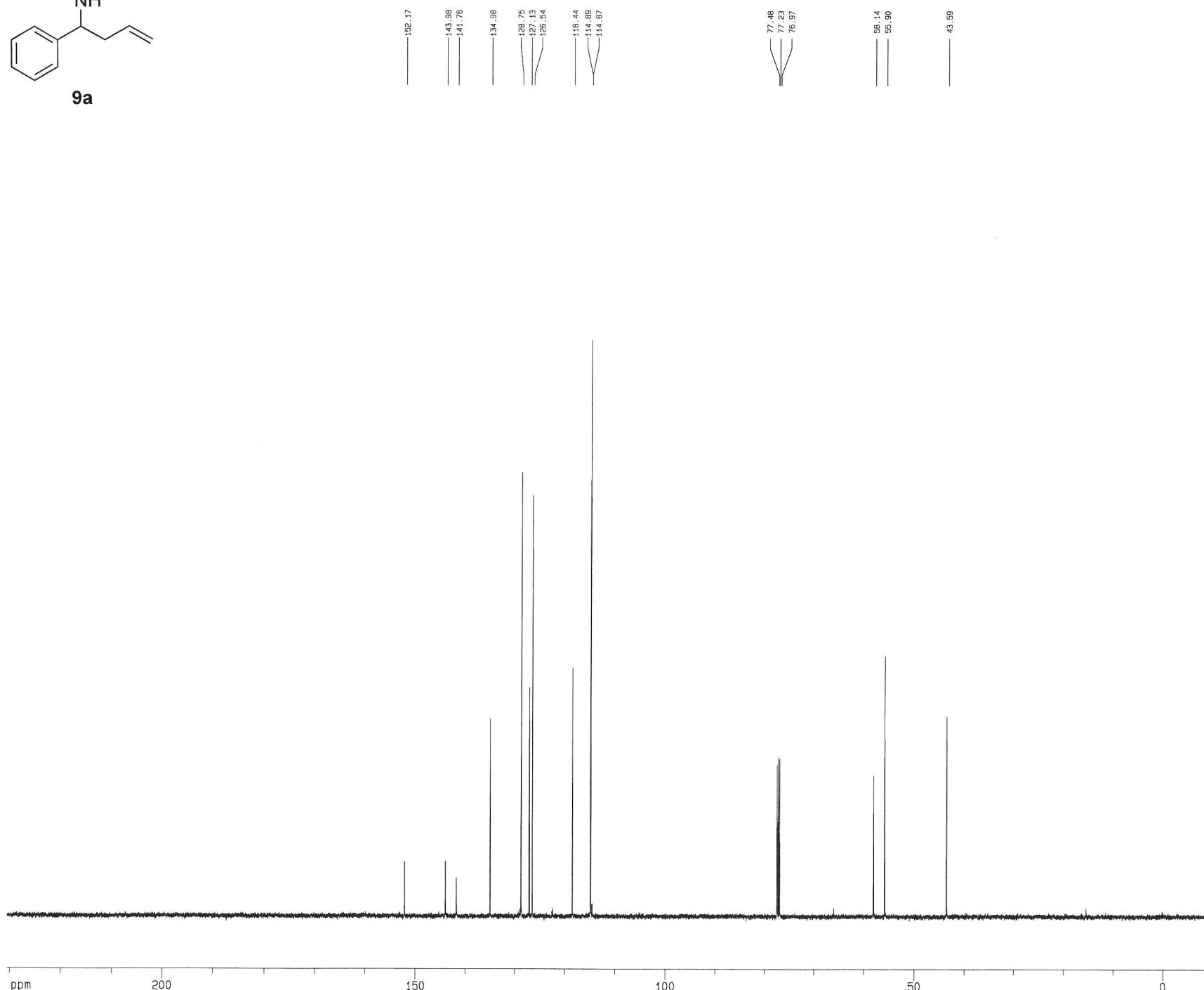
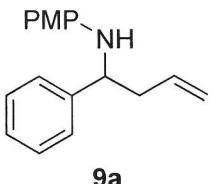
Current Data Parameters
 USER tjbark
 NAME 2-TJB-110
 EXPNO 2
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20080823
 Time 11:26
 INSTRUM drx400
 PROBHD 5 mm QNP H/F/P
 PULPROG zg30
 TD 65536
 SOLVENT CDC13T
 NS 8
 DS 2
 SWH 6410.256 Hz
 FIDRES 0.097813 Hz
 AQ 5.1116579 sec
 RG 724.1
 DW 78.000 usec
 DE 4.50 usec
 TE 298.0 K
 D1 0.1000000 sec
 MCREST 0.0000000 sec
 MCWAK 0.0150000 sec

===== CHANNEL f1 =====
 NUC1 1H
 P1 12.00 usec
 PL1 0.00 dB
 SF01 400.1328009 MHz

F2 - Processing parameters
 SI 65536
 SF 400.1300181 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 2.00

1D NMR plot parameters
 CX 22.80 cm
 CY 15.00 cm
 F1P 11.000 ppm
 F1 4401.43 Hz
 F2P -1.000 ppm
 F2 -400.13 Hz
 PPMCM 0.52632 ppm/cm
 HZCM 210.59474 Hz/cm



Current Data Parameters
 USER tjbark
 NAME 2-TJB-110
 EXPNO 4
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20081119
 Time 17.58
 INSTRUM cryo500
 PROBHD 5 mm CPT1 1H
 PULPROG SpinEchoes30gp.prd
 TD 65536
 SOLVENT CDCl3T
 NS 71
 DS 16
 SWH 30303.031 Hz
 FIDRES 0.462388 Hz
 AQ 1.0814105 sec
 RG 3251
 DW 16.500 usec
 DE 5.00 usec
 TE 298.0 K
 D1 0.2500000 sec
 d11 0.0002000 sec
 D16 0.0000000 sec
 d17 0.0001950 sec
 MCREFST 0.0000000 sec
 MCWRK 0.0150000 sec
 P2 29.70 usec

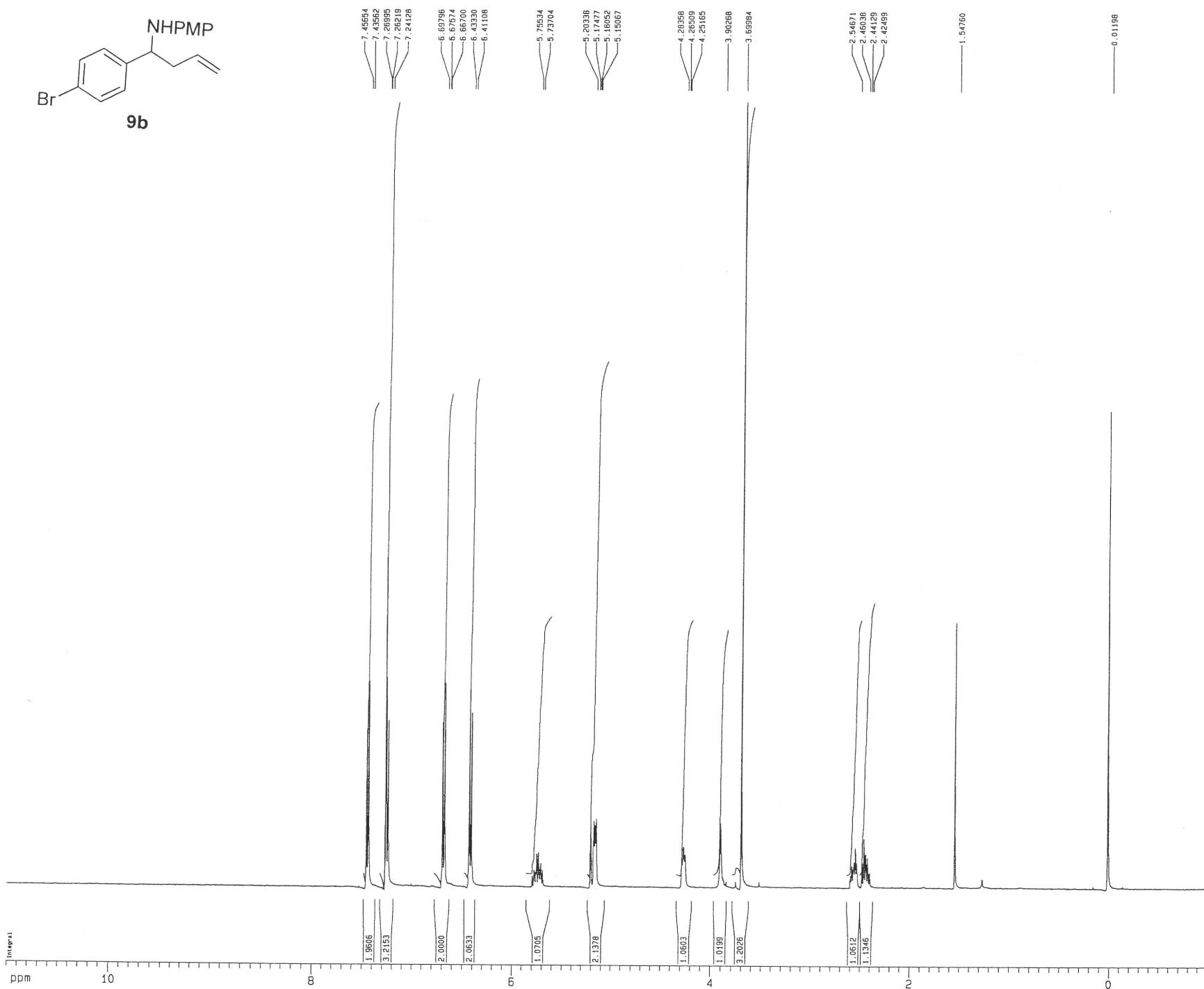
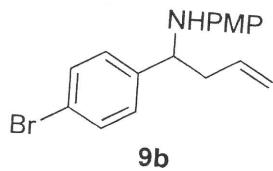
===== CHANNEL f1 =====
 NUC1 13C
 P1 14.85 usec
 P11 500.00 usec
 P12 2000.00 usec
 PL0 120.00 dB
 PL1 -1.00 dB
 SF01 125.7942548 MHz
 SP1 3.60 dB
 SP2 3.60 dB
 SPNAME1 Crp60.0_5.20_1
 SPNAME2 Crp60comp.4
 SP0FF1 0.00 Hz
 SP0FF2 0.00 Hz

===== CHANNEL f2 =====
 CPDPHG2 waltz16
 NUC2 1H
 PCP02 100.00 usec
 PL2 1.60 dB
 PL12 24.60 dB
 SF02 500.2225011 MHz

===== GRADIENT CHANNEL =====
 GPNAME1 SINE_100
 GPNAME2 SINE_100
 GPX1 0.00 %
 GPY2 0.00 %
 GPY1 0.00 %
 GPY2 0.00 %
 GPZ1 30.00 %
 GPZ2 50.00 %
 p15 500.00 usec
 p16 1000.00 usec

F2 - Processing parameters
 SI 65536
 SF 125.7804048 MHz
 MDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 2.00

1D NMR plot parameters
 CX 22.80 cm
 CY 11.00 cm
 F1P 230.63 ppm
 F1 29000.68 Hz
 F2P -19.287 ppm
 F2 -1293.95 Hz
 PPMCM 10.56688 ppm/cm
 HZCM 1329.10593 Hz/cm



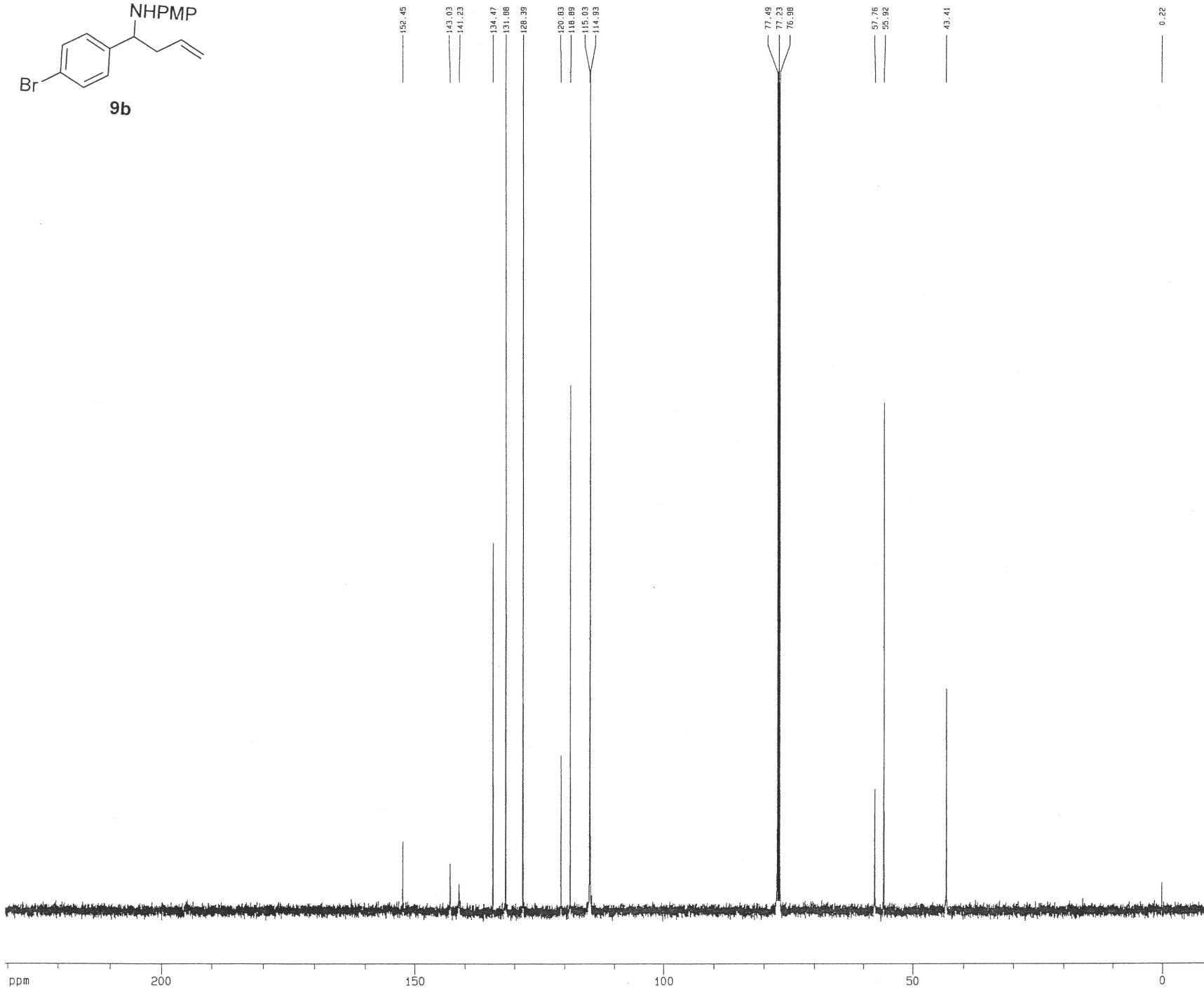
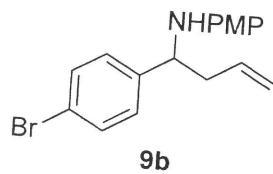
Current Data Parameters
 USER tjbark
 NAME 2-TB-110
 EXPNO 2
 PROCN0 1

F2 - Acquisition Parameters
 Date 20080823
 Time 11.26
 INSTRUM drx400
 PROBHD 5 mm DNP H/F/P
 PULPROG zg30
 TD 65536
 SOLVENT CDCl3T
 NS 8
 DS 2
 SWH 6410.256 Hz
 FIDRES 0.097813 Hz
 AQ 5.1118579 sec
 RG 724.1
 DW 78.000 usec
 DE 4.50 usec
 TE 298.0 K
 D1 0.1000000 sec
 MCREST 0.0000000 sec
 MCWPK 0.0150000 sec

***** CHANNEL f1 *****
 NUC1 1H
 PI 12.00 usec
 PL1 0.00 dB
 SF01 400.1328009 MHz

F2 - Processing parameters
 SI 65536
 SF 400.1300181 MHz
 DW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 2.00

1D NMR plot parameters
 CX 22.80 cm
 CY 15.00 cm
 F1P 11.000 ppm
 F1 4401.43 Hz
 F2P -1.000 ppm
 F2 -400.13 Hz
 PPVCM 0.52632 ppm/cm
 HZCM 210.59474 Hz/cm



Current Data Parameters
 USER tbark
 NAME 2-TJB-112
 EXPNO 12
 PROCNO 1

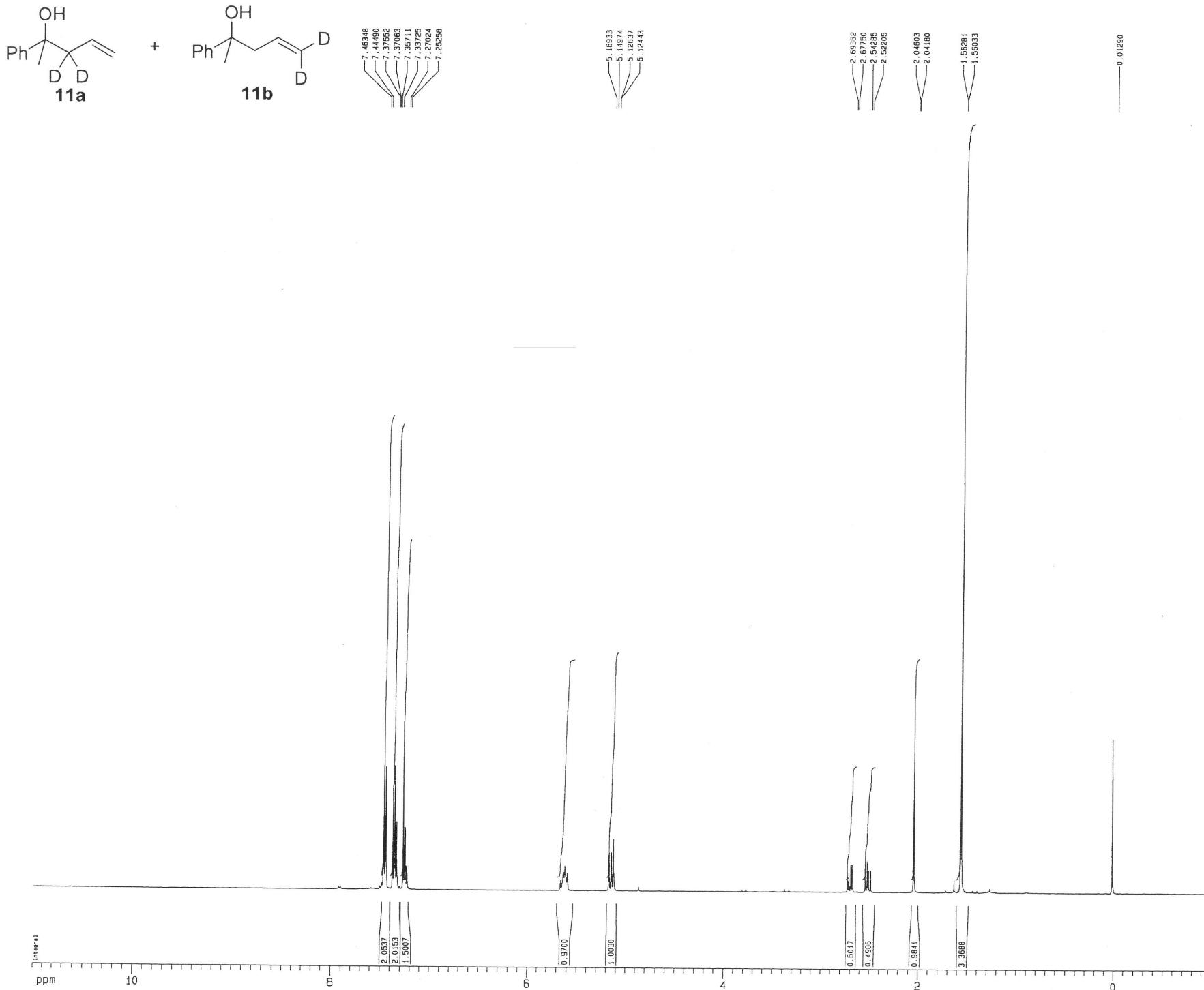
F2 - Acquisition Parameters
 Date_ 20080830
 Time 10.13
 INSTRUM cryo500
 PROBHD 5 mm CPTCI 1H-
 PULPROG zgdc30
 TD 65418
 SOLVENT CDCl3T
 NS 300
 DS 4
 SWH 30303.031 Hz
 FIDRES 0.463222 Hz
 AQ 1.0794470 sec
 RG 14596.5
 DW 16.500 usec
 DE 6.00 usec
 TE 298.0 K
 D1 0.2500000 sec
 d11 0.0300000 sec
 MCREST 0.0000000 sec
 MCWAK 0.0150000 sec

===== CHANNEL f1 =====
 NUC1 13C
 P1 14.75 usec
 PL1 -1.00 dB
 SF01 125.7942548 MHz

===== CHANNEL f2 =====
 CPDPG2 waltz16
 NUC2 1H
 PCPD2 100.00 usec
 PL2 1.60 dB
 PL12 24.80 dB
 SF02 500.2225011 MHz

F2 - Processing parameters
 SI 65536
 SF 125.7804006 MHz
 WDM EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 2.00

1D NMR plot parameters
 CX 22.80 cm
 CY 22.00 cm
 F1P 230.637 ppm
 F1 29009.68 Hz
 F2P -10.287 ppm
 F2 -1293.96 Hz
 PPMCM 10.56688 ppm/cm
 HZCM 1329.10693 Hz/cm



Current Data Parameters
 USER tjbank
 NAME 2-TJB-72
 EXPNO 4
 PROCNO 1

F2 - Acquisition Parameters
 Date 20080721
 Time 10.53
 INSTRUM drx400
 PROBHD 5 mm QNP H/F/P
 PULPROG zg30
 TD 65536
 SOLVENT CDCl₃
 NS 1
 DS 0
 SWH 6410.256 Hz
 FIDRES 0.097813 Hz
 AQ 5.118579 sec
 RG 456.1
 DW 78.000 usec
 DE 4.50 usec
 TE 298.0 K
 D1 0.1000000 sec
 MCREST 0.0000000 sec
 MCWRK 0.0150000 sec

***** CHANNEL f1 *****
 NUC1 1H
 P1 12.00 usec
 PL1 0.00 dB
 SF01 400.1328009 MHz

F2 - Processing parameters
 S1 65536
 SF 400.1300179 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 2.00

1D NMR plot parameters
 CX 22.80 cm
 CY 8.00 cm
 F1P 11.000 ppm
 F1 4401.43 Hz
 F2P -1.000 ppm
 F2 -400.13 Hz
 PPDM 0.52632 ppm/cm
 HZCM 210.59474 Hz/cm

