Nagelamides Q and R, Novel Dimeric Bromopyrrole Alkaloids from Sponges *Agelas* sp.

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Experimental Section

General Experimental Procedures. Optical rotation was recorded on a JASCO P-1030 polarimeter. IR and UV spectra were recorded on a Shimadzu UV-1600PC and a JASCO FT/IR-5300 spectrophotometers, respectively. ¹H, ¹³C and 2D NMR spectra were measured on a JEOL ramuda-400 spectrometer and a Bruker AMX-600 spectrometer using 5 and 2.5 mm micro cells (Shigemi Co., Ltd.) for DMSO-*d*₆, respectively. The 2.49 and 39.5 ppm resonances of residual DMSO-*d*₆ and the 7.19 and 123.5 ppm resonances of residual C₅D₅N were used as internal references for ¹H and ¹³C NMR spectra, respectively. Positive mode ESIMS spectra were obtained on a JEOL JMS 700-TZ spectrometer at -80 V as a focus voltage using a sample dissolved in MeOH with flow rate of 200 mL/min.

Sponge Description. The sponge (SS-1134) Agelas sp. (order, Agelasida, family Agelasiidae) was collected off Seragaki, Okinawa, and kept frozen until used. The sponge is open textured sponge with convoluted surface with numerous superficial depressions and occasional oscules. The Sponge has a fine ridged pattern over the surface with large apical oscules approximately 5 mm wide. The surface patterning gives a 'brain-like' appearance. The sponge has a dense internal structure and is firm and compressible. Skeleton is reticulate with fibre development, with primary fibres cored by verticillate acanthostyles, 4 spicules across, fibres are 130 µm wide and mesh spaces are approximately 500 µm wide. Secondary fibres form a tight mesh and are either cored or unispicular. Spicules are verticillate spined acanthostyles, thick, slightly curved 250 x 20 µm, some thin forms occur. The sponge (SS-956) Agelas sp. (order, Agelasida, family Agelasiidae) was collected off Unten-Port, Okinawa, and kept frozen until used. The sponge is medium brown mounds with smooth, faintly patterned surfaces. Mesohyl is compact with numerous pores. The sponge is firm and compressible. Skeleton is reticulate with fibre development, with primary fibres cored and echinated by verticillate spined acanthostyles. Primary fibres are 90 µm wide with brushes of spicules apically. Meshes are small and compact. Spicules are verticillate spined acanthostyles, thick, slightly curved 230 x 20 µm, some thin forms occur. The voucher specimens were deposited at Graduate School of Pharmaceutical Sciences, Hokkaido University.

Extraction and Isolation. The sponge (SS-1134, 0.70 kg, wet weight) was extracted with MeOH (2L x 2), and the extract was partitioned between BuOH (500 mL x 3) and H₂O (500 mL). Parts (7.63 g) of BuOH soluble materials (10.63 g) were subjected to a

silica gel column (CHCl₃/MeOH/AcOH) and then a C₁₈ column (Cosmosil 140 C₁₈ PREP, Nakarai Tesque Inc.; eluent, MeOH/H₂O/CF₃CO₂H, 80:20:01) to give a crude fraction of alkaloids. This fraction was separated by C₁₈ MPLC (KUSANO C.I.G.; Kusano Science Corp.; eluent, CH₃OH/H₂O/CF₃CO₂H, 45:55:0.1 to 90:10:0.1 in 90 min; flow rate, 3.5 mL/min). A part of the fraction containing nagelamide Q (1) was purified by C₁₈ HPLC [YMC Hydrospere C18, YMC Co., Inc., 10 x 250 mm; eluent, CH₃CN/H₂O/CF₃CO₂H₂ 25:75:0.1; flow rate, 2.5 mL/min; UV detection at 255 nm] to afford nagelamide Q (1, 0.0012 %, wet weight). The sponge (SS-956, 0.50 kg, wet weight) was extracted with MeOH (1.5 L x 3), and the extract was partitioned between BuOH (50 mL x 3) and H₂O (50 mL). BuOH soluble materials (4.31 g) were subjected to a silica gel column (CHCl₃/MeOH/AcOH) and then a C₁₈ column (Cosmosil 140 C₁₈ PREP, Nakarai Tesque Inc.; eluent, MeOH/H₂O/CF₃CO₂H, 80:20:01) to give a crude fraction of alkaloids. This fraction was separated by C₁₈ MPLC (KUSANO C.I.G.; Kusano Science Corp.; eluent, CH₃OH/H₂O/CF₃CO₂H, 40:60:0.1 to 70:30:0.1 in 90 min; flow rate, 3.0 mL/min). A part of the fraction containing nagelamide R (2) was purified by C₁₈ HPLC [YMC Hydrospere C18, YMC Co., Inc., 10 x 250 mm; eluent, CH₃CN/H₂O/CF₃CO₂H, 20:80:0.1 to 40:60:0.1 in 40 min; flow rate, 2.5 mL/min; UV detection at 255 nm] to afford nagelamide R (2, 0.0013 %).

Nagelamide Q (1): colorless amorphous solid; $[\alpha]_D^{24}$ -5 (c 0.2, MeOH); UV (MeOH) λ_{max} 276 nm (ε 18000); IR (KBr) ν_{max} 3406, 1684, 1206, and 1044 cm⁻¹; ESIMS (pos.) m/z 896, 898, 900, 902, and 904 [1:4:6:4:1, M⁺]; HRESIMS (pos.) m/z 895.85868 [M⁺, calcd for $C_{24}H_{26}N_{11}O_5^{79}Br_4S$, 895.85726].

Nagelamide R (2): colorless amorphous solid; $[\alpha]_D^{22}$ -3 (c 0.2, MeOH); UV (MeOH) λ_{max} 278 nm (ε 27000), 400 nm (ε 1600); IR (KBr) ν_{max} 3407, 1685, 1202, and 1140 cm⁻¹; ESIMS (pos.) m/z 773, 775, 777, 779, and 781 [1:4:6:4:1, (M-H)⁺]; HRESIMS (pos.) m/z 772.85649 [(M-H)⁺, calcd for $C_{22}H_{21}N_{10}O_2^{79}Br_4$, 772.85825].

Conversion of Nagelamide R (2) to Nagelamide L: Nageamide R (2, 100 μ g) was dissolved in the mixture of DMSO and TFA (99:1) and stand at room temperature. After 24 h, DMSO and TFA in the reaction mixture were removed by lyophilization to afford nagelamide L (85 μ g), whose spectrum data were identical with those of natural nagelamide L.

Figure S1. ¹H NMR spectrum of nagelamide Q (1) in DMSO-*d*₆

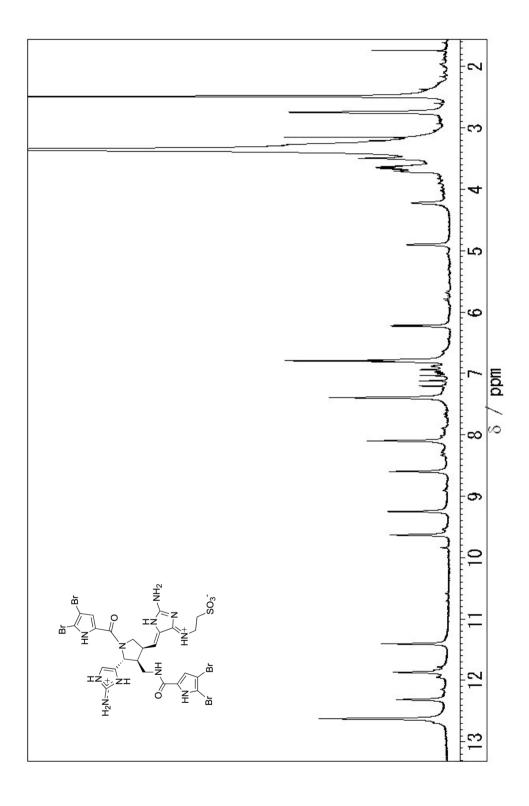
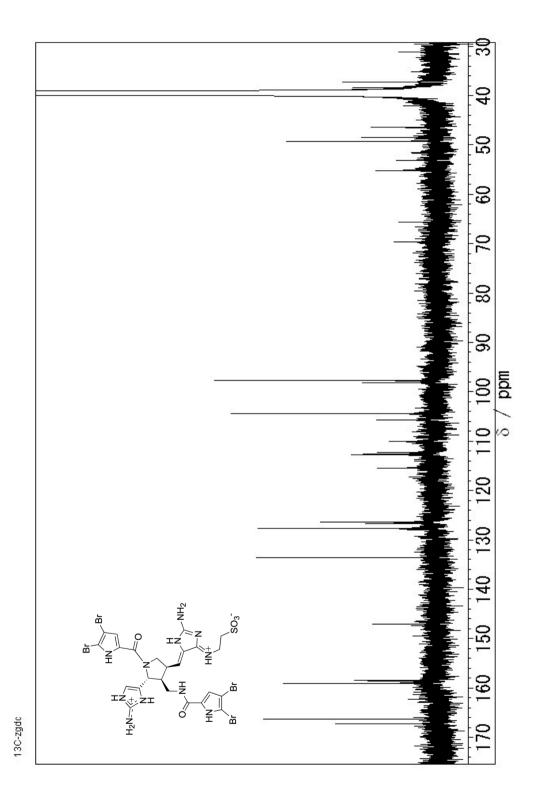


Figure S2. ¹³C NMR spectrum of nagelamide Q (1) in DMSO-d₆



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Figure S3. ¹H-¹H COSY spectrum of nagelamide Q (1) in DMSO-*d*₆

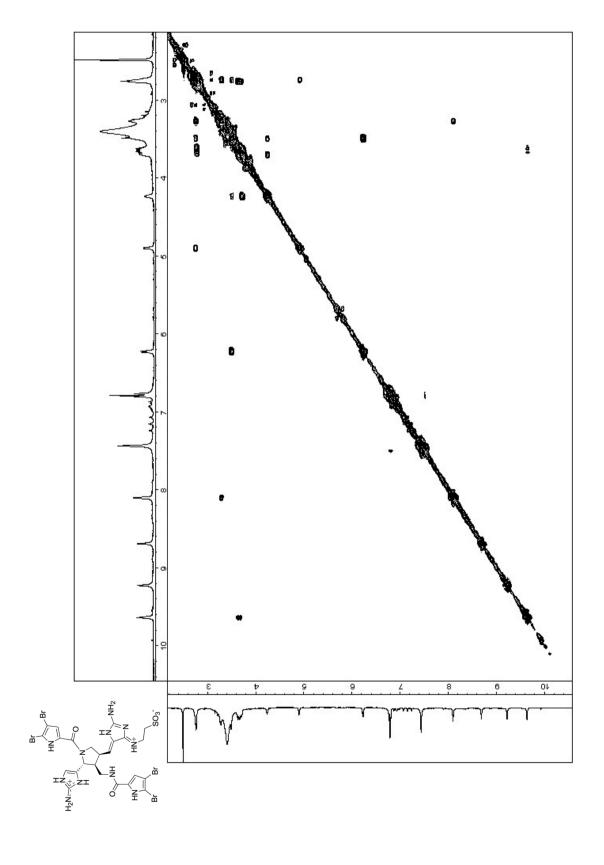


Figure S4. ROESY spectrum of nagelamide Q (1) in DMSO- d_6

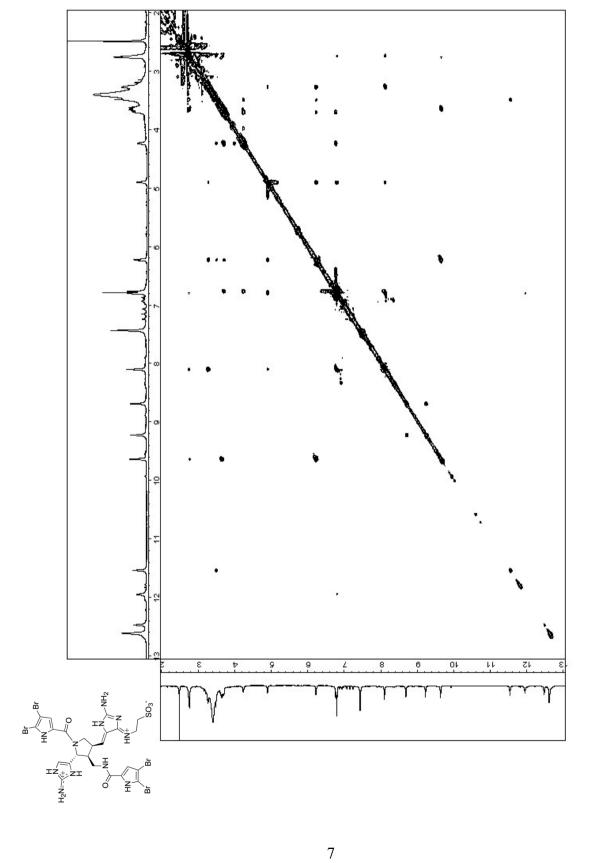


Figure S5. HMQC spectrum of nagelamide Q (1) in DMSO- d_6

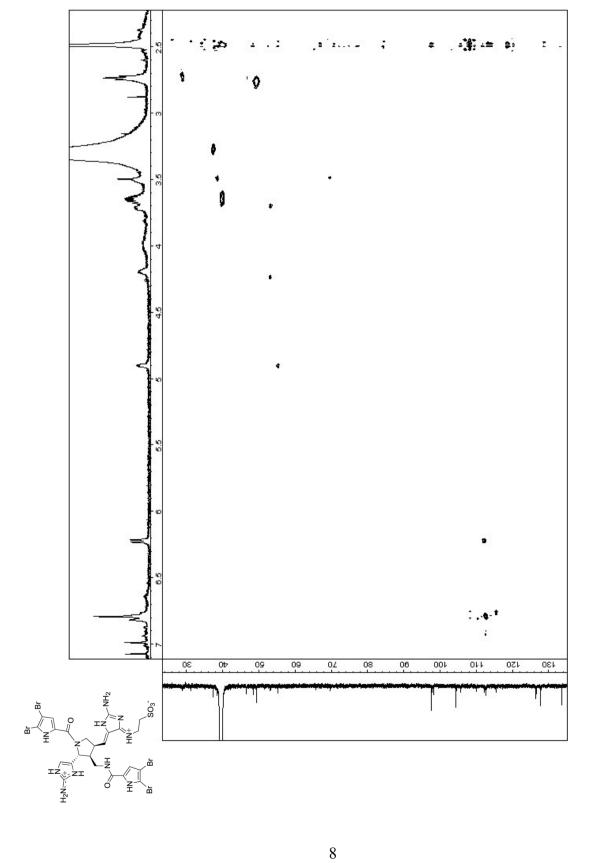


Figure S6. HMBC spectrum of nagelamide Q (1) in DMSO-d₆

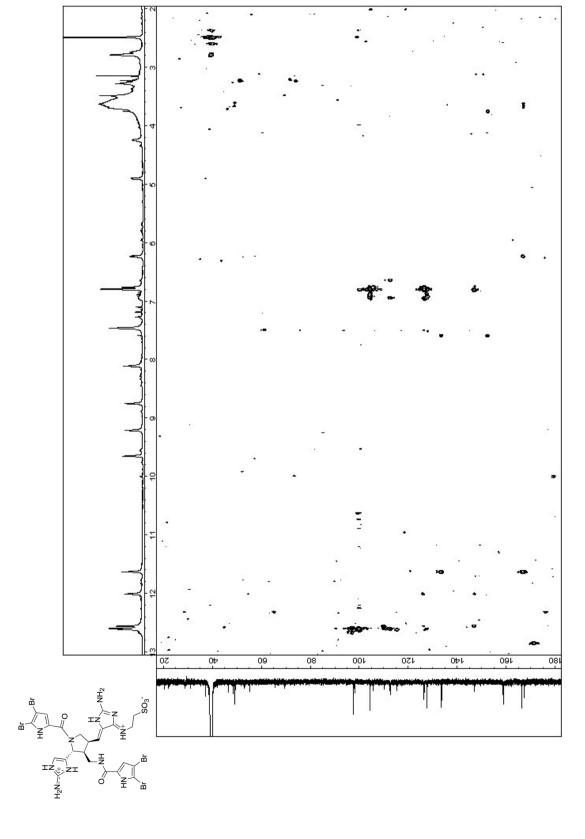


Figure S7. ¹H NMR spectrum of nagelamide R (2) in C₅D₅N

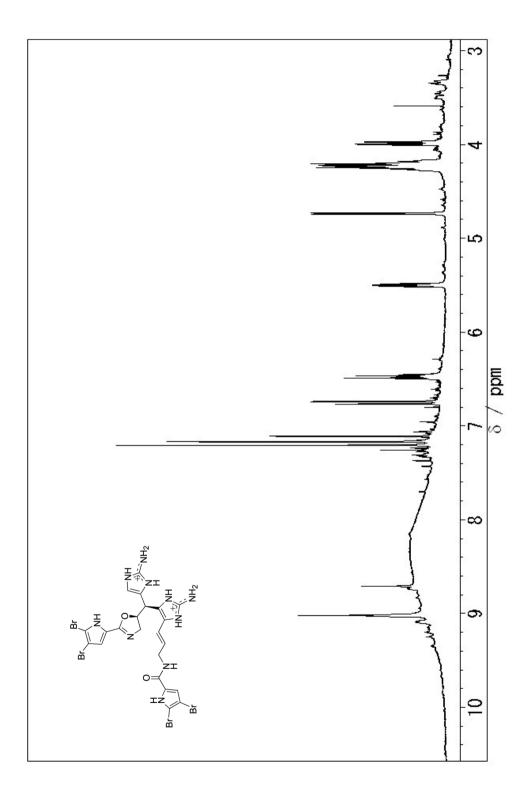


Figure S8. ¹³C NMR spectrum of nagelamide R (2) in C₅D₅N

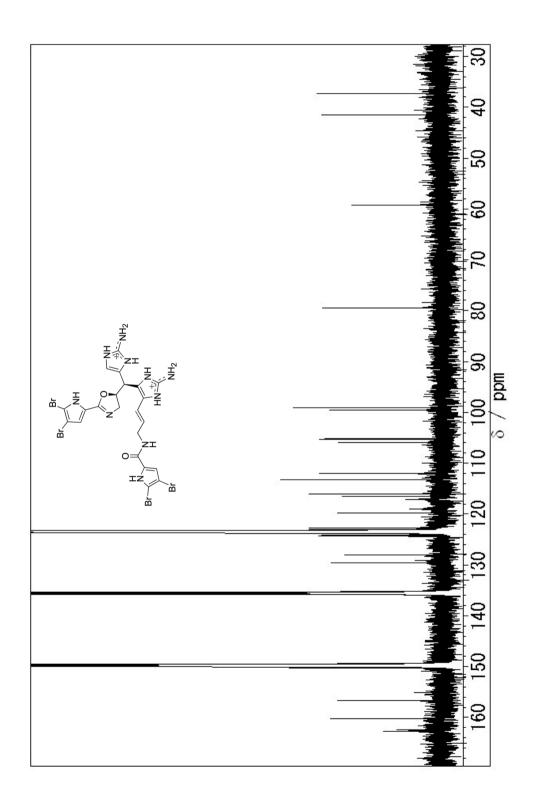


Figure S9. ¹H-¹H COSY spectrum of nagelamide R (2) in C₅D₅N

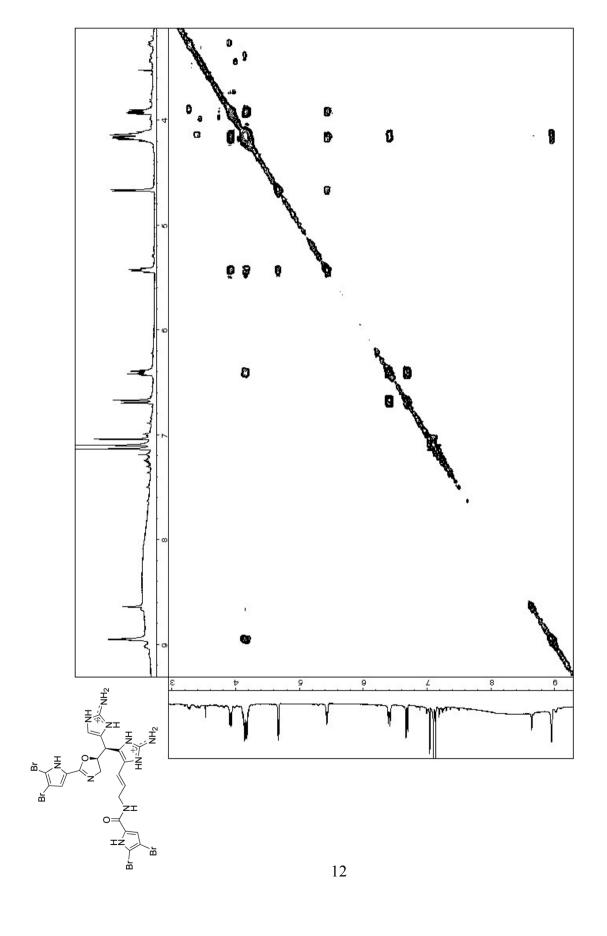


Figure S10. ROESY spectrum of nagelamide R (2) in C_5D_5N

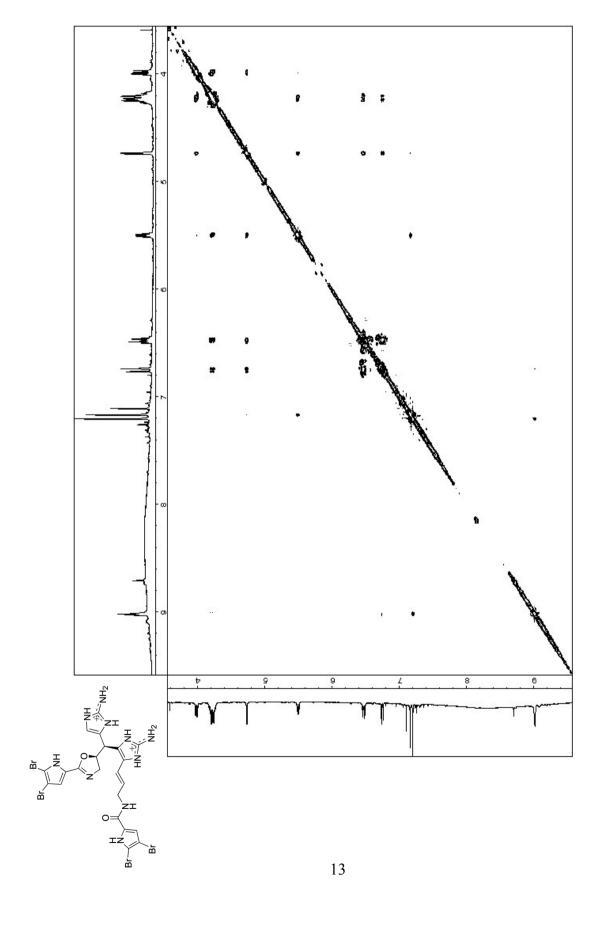


Figure S11. HMQC spectrum of nagelamide R (2) in C₅D₅N

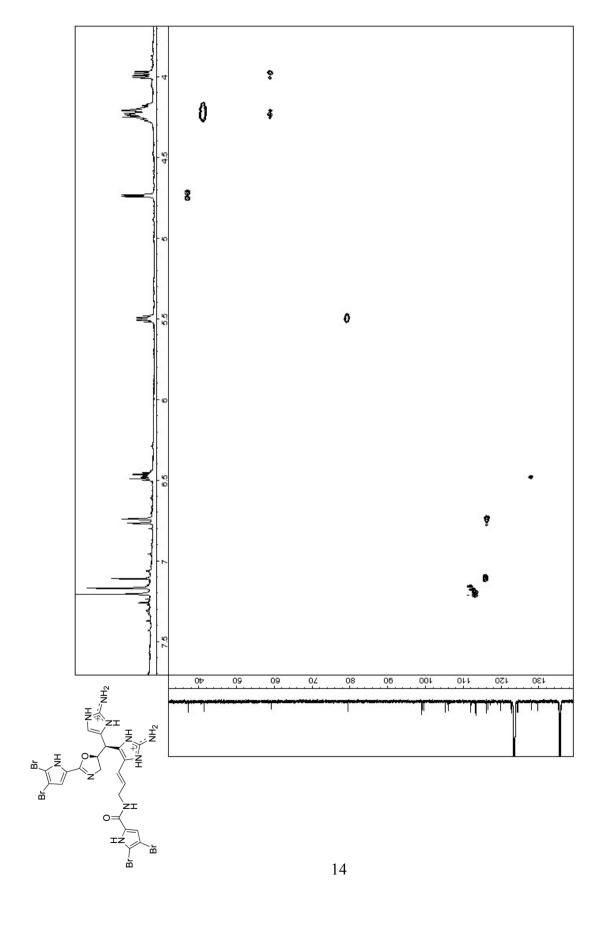


Figure S12. HMBC spectrum of nagelamide R (2) in C₅D₅N

