Supplemental Information

Discovery of inducible Nitric Oxide Synthase (iNOS) inhibitor Development Candidate KD7332 (Part 1): Identification of a Novel, Potent and Selective series of Quinolinone iNOS Dimerization Inhibitors that are Orally Active in Rodent Pain Models

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TABLE OF CONTENTS

- S-1 Title Page.
- S-2 to S-15 Experimental Section (Chemistry)
- S-16 to S-19 Experimental Section (Methods)
- S-20 to S-75 ¹H NMR, ¹³C NMR (when available), and LC trace (for purity).
- S-76 to S-77 High Resolution Mass Spectrum (table format).
- S-78 to S-82 Crystal data, structure refinement and Atomic coordinates for Cpd 12

4-((2-Chlorophenylamino)methyl)-8-fluoroquinolin-2(1H)-one (**55).** Compound **55** was synthesized as described for compound **53** using 4-(bromomethyl)-8-fluoroquinolin-2(1H)-one (**52)** and 2-chloroaniline as starting materials. ¹H NMR (400 MHz, DMSO-d₆) δ 11.63 (s, 1H), 7.67 (d, 1H, *J* = 8.0 Hz), 7.39 (dd, 1H, *J* = 11.2, 8.4 Hz), 7.23 (dd, 1H, *J* = 8.0, 1.6 Hz), 7.18-7.13 (m, 1H), 6.99 (td, 1H, *J* = 7.8, 1.6 Hz), 6.54 (td, 1H, *J* = 7.8, 1.0 Hz), 6.45 (dd, 1H, *J* = 8.4, 1.2 Hz), 6.23 (s, 1H), 6.14 (t, 1H, *J* = 5.6 Hz), 4.59 (d, 2H, *J* = 5.2 Hz). LRMS (ESI+) *m/z*: calcd for C₁₆H₁₂ClFN₂O 303.1, found 303.2 [M + H]⁺.

4-((3-Chlorophenylamino)methyl)-8-fluoroquinolin-2(1H)-one (56). Compound **56** was synthesized as described for compound **53** using 4-(bromomethyl)-8-fluoroquinolin-2(1H)-one (**52**) and 3-chloroaniline as starting materials. ¹H NMR (400 MHz, DMSO-d₆) δ 11.67 (s, 1H), 7.65 (d, 1H, *J* = 8.4 Hz), 7.42 (ddd, 1H, *J* = 10.8, 8.0, 0.8 Hz), 7.19 (m, 1H), 7.06 (t, 1H, *J* = 8.4 Hz), 6.62 (m, 2H), 6.55 (d, 2H, *J* = 7.8 Hz), 6.42 (s, 1H), 4.53 (d, 2H, *J* = 5.2 Hz). LRMS (ESI+) *m/z*: calcd for C₁₆H₁₂ClFN₂O 303.1, found 303.2 [M + H]⁺.

4-((4-Chlorophenylamino)methyl)-8-fluoroquinolin-2(1H)-one (**57).** Compound **57** was synthesized as described for compound **53** using 4-(bromomethyl)-8-fluoroquinolin-2(1H)-one (**52)** and 4-chloroaniline as starting materials. ¹H NMR (400 MHz, DMSO-d₆) δ 11.63 (s, 1H), 7.60 (d, 1H, *J* = 8.4 Hz), 7.38 (dd, 1H, *J* = 10.8, 7.2 Hz), 7.13 (m, 1H), 7.02 (m, 2H), 6.53 (d, 2H, *J* = 6.8 Hz), 6.48 (m, 1H), 6.36 (s, 1H), 4.46 (bs, 2H). LRMS (ESI+) *m/z*: calcd for C₁₆H₁₂ClFN₂O 303.1, found 303.2 [M + H]⁺.

8-Fluoro-4-((3-fluorophenylamino)methyl)quinolin-2(1H)-one (58). Compound 58 was synthesized as described for compound 53 using 4-(bromomethyl)-8-fluoroquinolin-2(1H)-one (52) and 3-fluoroaniline as starting materials. ¹H NMR (400 MHz, DMSO-d₆) δ 11.68 (s, 1H), 7.65 (d, 1H, J = 8.4 Hz), 7.42 (ddd, 1H, J = 11.2, 8.4, 1.2 Hz), 7.18 (m, 1H), 7.07 (m, 1H), 6.63

(t, 1H, J = 5.6 Hz), 6.43-6.29 (m, 4H), 4.53 (d, 2H, J = 6.0 Hz). LRMS (ESI+) m/z: calcd for C₁₆H₁₂F₂N₂O 287.1, found 287.2 [M + H]⁺.

8-Fluoro-4-((3-cyanophenylamino)methyl)quinolin-2(1H)-one (59). Compound 59 was synthesized as described for compound 53 using 4-(bromomethyl)-8-fluoroquinolin-2(1H)-one (52) and 3-aminobenzonitrile as starting materials. ¹H NMR (400 MHz, DMSO-d₆) δ 11.64 (s, 1H), 7.60 (d, 1H, *J* = 8.4 Hz), 7.44-7.33 (m, 1H), 7.22-7.12 (m, 2H), 6.92-6.78 (m, 3H), 6.87 (m, 1H), 6.36 (s, 1H), 4.54 (d, 2H, *J* = 2.8 Hz). LRMS (ESI+) *m/z*: calcd for C₁₇H₁₂FN₃O 294.1, found 294.2 [M + H]⁺.

8-Fluoro-4-((*m*-tolylamino)methyl)quinolin-2(1H)-one (60). Compound 60 was synthesized as described for compound 53 using 4-(bromomethyl)-8-fluoroquinolin-2(1H)-one (52) and *m*-toluidine as starting materials. ¹H NMR (400 MHz, DMSO-d₆) δ 11.60 (s, 1H), 7.62 (d, 1H, *J* = 8.4 Hz), 7.38 (ddd, 1H, *J* = 10.8, 7.6, 0.8 Hz), 7.13 (m, 1H), 6.89 (m, 1H), 6.39-6.31 (m, 4H), 6.17 (m, 1H), 4.48 (bs, 2H), 2.09 (s, 3H). LRMS (ESI+) *m/z*: calcd for C₁₇H₁₅FN₂O 283.1, found 283.2 [M + H]⁺.

8-Fluoro-4-((3-methoxyphenylamino)methyl)quinolin-2(1H)-one (61). Compound 61 was synthesized as described for compound 53 using 4-(bromomethyl)-8-fluoroquinolin-2(1H)-one (52) and 3-methoxyaniline as starting materials. ¹H NMR (400 MHz, DMSO-d₆) δ 11.61 (s, 1H), 7.62 (d, 1H, *J* = 8.4 Hz), 7.38 (ddd, 1H, *J* = 10.8, 8.0, 0.8 Hz), 7.13 (m, 1H), 6.90 (t, 1H, *J* = 7.6 Hz), 6.39 (s, 1H), 6.28 (m, 1H), 6.14-6.08 (m, 3H), 4.45 (bs, 2H), 3.58 (s, 3H). LRMS (ESI+) *m/z*: calcd for C₁₇H₁₅FN₂O₂ 299.1, found 299.2 [M + H]⁺.

2-(*tert***-Butyldimethylsilyloxy)-4-(chloromethyl)-8-fluoroquinoline (62).** To a stirred solution of 4-(bromomethyl)-8-fluoroquinolin-2(1H)-one (**52**, 1.28 g, 5.0 mmol) in DMF (50 mL) at 25°C was added *tert*-butyldimethylsilyl chloride (1.51 g, 10.0 mmol) followed by Et_3N (2.4 mL, 17.5 mmol). After 4 h, the reaction mixture was poured into ice water (500 mL), and the resulting Page **3** of **82**

precipitate was collected by vacuum filtration. The filter cake was washed with water (100 mL), then dried for 18 h to afford 2-(*tert*-butyldimethylsilyloxy)-4-(chloromethyl)-8-fluoroquinoline (**62**, 1.42 g, 88%) as a tan solid. ¹H NMR (400 MHz, DMSO-d₆) δ 7.86 (d, 1H, *J* = 8.0 Hz), 7.52-7.45 (m, 2H), 7.23 (s, 1H), 5.18 (s, 2H), 0.98 (s, 9H), 0.38 (s, 6H). LRMS (ESI+) *m/z*: calcd for C₁₆H₂₁ClFNOSi 326.1, found 211.9 [M - TBDMS + H]⁺.

2-(*tert*-**Butyldimethylsilyloxy**)-**8-fluoro-4-**(iodomethyl)quinoline (63). Sodium iodide (157 mg, 1.05 mmol) was added to a stirred solution of 2-(tert-butyldimethylsilyloxy)-4- (chloromethyl)-8-fluoroquinoline (62, 325 mg, 1.0 mmol) in dry acetone at 25°C. After 2 h, the heterogeneous mixture was diluted with DCM (200 mL), dried over MgSO₄, filtered, and concentrated under reduced pressure to afford 2-(*tert*-butyldimethylsilyloxy)-8-fluoro-4- (iodomethyl)quinoline (63, 390 mg, 94%) as an orange solid. ¹H NMR (400 MHz, CDCl₃) δ 7.72 (d, 1H, *J* = 8.4 Hz), 7.37 (m, 1H), 7.30 (m, 1H), 7.00 (s, 1H), 4.67 (s, 2H), 1.02 (s, 9H), 0.44 (s, 6H). LRMS (ESI+) *m/z*: calcd for C₁₆H₂₁FINOSi 418.0, found 303.8 [M - TBDMS + H]⁺.

N-(3-Chlorophenyl)-1-methyl-1*H*-imidazole-5-carboxamide (66). O-(7-Azabenzotriazol-1yl)-*N*,*N*,*N*',*N*'-tetramethyluronium hexafluorophosphate (HATU, 912 mg, 2.4 mmol) was added to a stirred mixture of 1-methyl-1*H*-imidazole-5-carboxylic acid (65, 252 mg, 2.0 mmol), 3chloroaniline (64, 273 μ L, 2.6 mmol), and Et₃N (1.1 mL, 8.0 mmol) in DMF (10 mL). After 18 h at 25°C, the reaction mixture was diluted with 1:1 hexanes:EtOAc (200 mL), washed with 5% brine (3 x 50 mL), dried over MgSO₄, filtered, and concentrated under reduced pressure. The resulting crude residue was purified by silica gel column chromatography (9:1 DCM/ACN) to afford N-(3-chlorophenyl)-1-methyl-1H-imidazole-5-carboxamide (66, 358 mg, 76%) as a tan solid. ¹H NMR (400 MHz, DMSO-d₆) δ 10.03 (s, 1H), 8.04 (s, 1H), 7.82 (s, 1H), 7.75 (m, 2H), 7.30 (t, 1H, *J* = 8.2 Hz), 7.07 (d, 1H, *J* = 8.0 Hz), 3.71 (s, 3H). **3-Chloro-N-((2-chloropyridin-4-yl)methyl)aniline (68).** A mixture of 4-(bromomethyl)-2chloropyridine (**67**,¹¹ 3.6 g, 15.8 mmol), 3-chloroaniline (2.0 g, 15.47 mmol), and K₂CO₃ (2.2 g, 15.78 mmol) in DMF (50 mL) was stirred for 2 h at 60°C. The resulting solution was diluted with water (150 mL) and extracted with EtOAc (2 x 100 mL). The organics were combined, dried over Na₂SO₄, filtered, and concentrated to dryness under reduced pressure. The residue was purified by silica gel column chromatography (1:5 EtOAc/Hexanes) to afford 2.8 g (67%) of 3-chloro-N-((2-chloropyridin-4-yl)methyl)aniline (**68**) as a yellow solid. LRMS (ESI+) *m/z*: calcd for C₁₂H₁₀Cl₂N₂ 253.0, found 253.0 [M + H]⁺.

N-(3-Chlorophenyl)-N-((2-chloropyridin-4-yl)methyl)-4-methylthiazole-5-carboxamide

(69). 4-Methylthiazole-5-carbonyl chloride (synthesis of acid chloride described for compound 7, 1.43 g, 7.99 mmol) was added in several batches to 3-chloro-N-((2-chloropyridin-4-yl)methyl)aniline (68, 1.12 g, 3.98 mmol) in DMF (20 mL) cooled to 0°C. The resulting solution was stirred at 25°C for 2 h. The solvent was removed by evaporation under reduced pressure and the residue was purified by silica gel column chromatography (1:2 EtOAc/Hexanes) to afford 0.8 g (46%) of N-(3-chlorophenyl)-N-((2-chloropyridin-4-yl)methyl)-4-methylthiazole-5-carboxamide (69) as a yellow oil. ¹H NMR (300 MHz, DMSO-d₆) δ 8.93 (s, 1H), 8.36 (d, 1H), 7.50 (m, 2H), 7.43 (d, 1H), 7.34 (m, 2H), 7.18 (d, 1H), 5.12 (s, 2H), 2.42 (s, 3H). LRMS (ESI+) *m/z*: calcd for C₁₇H₁₃Cl₂N₃OS 378.0, found 378.0 [M + H]⁺.

(2-Chloro-5,6,7,8-tetrahydroquinolin-4-yl)methanol (71). LiAlH₄ (190 mg, 5.00 mmol) was added to methyl 2-chloro-5,6,7,8-tetrahydroquinoline-4-carboxylate (70,ⁱ 200 mg, 0.88 mmol) in THF (10 mL) at 0°C. The resulting slurry was stirred at 0°C for 30 min. The reaction mixture was then quenched by adding water and 15% NaOH solution. The precipitate was filtered and dried to afford 250 mg (crude) of (2-chloro-5,6,7,8-tetrahydroquinolin-4-yl)methanol (71) as a white solid. LRMS (ESI+) m/z: calcd for C₁₀H₁₂ClNO 198.1, found 198.0 [M + H]⁺.

4-(Bromomethyl)-2-chloro-5,6,7,8-tetrahydroquinoline (72). PBr₃ (1.71 g, 6.31 mmol) in DCM (10 mL) was added dropwise to (2-chloro-5,6,7,8-tetrahydroquinolin-4-yl)methanol (71, 250 mg, 1.26 mmol) in DCM (5 mL) at 0°C. The resulting solution was stirred at 25°C for 30 min. The reaction mixture was then quenched with water (30 mL), neutralized to pH 7 using aqueous sodium bicarbonate, and extracted with DCM (4 x 120 mL). The organics were combined, dried over Na₂SO₄, and concentrated under reduced pressure to afford 160 mg (crude) of 4-(bromomethyl)-2-chloro-5,6,7,8-tetrahydroquinoline (72) as a white solid.

3-Chloro-*N***-**((**2-chloro-5,6,7,8-tetrahydroquinolin-4-yl)methyl)aniline** (**73**). A mixture of 4-(bromomethyl)-2-chloro-5,6,7,8-tetrahydroquinoline (**72**, 860 mg, 3.30 mmol), 3-chloroaniline (420 mg, 3.28 mmol), and Na₂CO₃ (350 mg) in DMF (20 mL) was heated to 60°C for 2 h. The resulting solution was diluted with water (60 mL) and extracted with EtOAc (5 x 300 mL). The organics were combined, dried over Na₂SO₄, and concentrated under reduced pressure. The residue was purified by silica gel column chromatography (1:20 EtOAc/Hexanes) to afford 600 mg of 3-chloro-*N*-((2-chloro-5,6,7,8-tetrahydroquinolin-4-yl)methyl)aniline (**73**) as a light yellow solid. LRMS (ESI+) *m/z*: calcd for C₁₆H₁₆Cl₂N₂ 307.1, found 307.0 [M + H]⁺

N-((2-Chloro-5,6,7,8-tetrahydroquinolin-4-yl)methyl)-*N*-(3-chlorophenyl)-4-methylthiazole-5-carboxamide (74). 4-Methylthiazole-5-carbonyl chloride (synthesis of acid chloride described in compound 7, 700 mg, 4.32 mmol) in DMF (15 mL) was added dropwise to 3-chloro-*N*-((2chloro-5,6,7,8-tetrahydroquinolin-4-yl)methyl)aniline (73, 510 mg, 1.66 mmol) in DMF (5 mL). The resulting solution was stirred at 25°C for 7 h. The reaction mixture was then quenched with water (60 mL) and extracted with EtOAc (4 x 240 mL). The organics were combined, dried over Na₂SO₄, and concentrated under reduced pressure. The residue was purified by silica gel column chromatography (1:5 EtOAc/Hexanes) to afford 420 mg of *N*-((2-chloro-5,6,7,8tetrahydroquinolin-4-yl)methyl)-*N*-(3-chlorophenyl)-4-methylthiazole-5-carboxamide (74) as a light yellow solid. LRMS (ESI+) *m/z*: calcd for C₂₁H₂₁Cl₂N₃OS 432.0, found 432.0 [M + H]⁺. Page 6 of 82 8-Fluoro-4-methyl-2*H*-chromen-2-one (77). A mixture of 1-(3-fluoro-2-hydroxyphenyl)ethanone (75, 13 g, 75.91 mmol) and (triphenylphosphoranylidene) acetic acid ethyl ester (76,ⁱⁱ 30 g, 86.2 mmol) in toluene (200 mL) was stirred at 120°C for 18 h. The mixture was concentrated under reduced pressure and the residue was partitioned between EtOAc (100 mL) and water (200 mL). The organic layer was isolated, washed with water (200 mL), dried over Na₂SO₄, filtered, and concentrated to dryness under reduced pressure. The residue was purified by silica gel column chromatography (1:25 EtOAc/Hexanes) to afford 5 g (35%) of 8-fluoro-4-methyl-2*H*-chromen-2-one (77) as a yellow solid. LRMS (ESI+) *m/z*: calcd for C₁₀H₇FO₂ 179.0, found 179.2 [M + H]⁺.

4-(Bromomethyl)-8-fluoro-2*H***-chromen-2-one (78).** A mixture of 8-fluoro-4-methyl-2*H*-chromen-2-one (77, 5g, 25.3 mmol), NBS (6g, 33.71 mmol), AIBN (cat., 0.1 g) in CCl₄ (300 mL) was refluxed for 18 h. The reaction mixture was concentrated to dryness and the residue was purified by silica gel chromatography (1:20 EtOAc/Hexanes) to afford 1.17 g (17%) of 4-(bromomethyl)-8-fluoro-2*H*-chromen-2-one (**78**) as a yellow solid. LRMS (ESI+) m/z: calcd for C₁₀H₆BrFO₂ 256.9, found 257.0 [M + H]⁺.

4-((**3**-Chlorophenylamino)methyl)-8-fluoro-2*H*-chromen-2-one (**79**). A mixture of 4-(bromomethyl)-8-fluoro-2*H*-chromen-2-one (**78**, 500 mg, 1.94 mmol) and 3-chloroaniline (0.8 mL, 7.78 mmol) in DMSO (5 mL) was heated to 70°C for 1 h. The cooled reaction mixture was then poured into ice water, and extracted with DCM (3 x 50 mL). The organics were combined, dried over Na₂SO₄, filtered, and concentrated to dryness under reduced pressure. The residue was purified by silica gel chromatography (20-50% EtOAc/Hexanes) to afford 500 mg of 4-((3-chlorophenylamino)methyl)-8-fluoro-2*H*-chromen-2-one (**79**) as a yellow solid. LRMS (ESI+) m/z: calcd for C₁₆H₁₁ClFNO₂ 304.0, found 303.9 [M + H]⁺. **4-(Bromomethyl)-8-fluoroquinoline (81).** A mixture of 8-fluoro-4-methylquinoline (**80**,ⁱⁱⁱ 500 mg, 3.11 mmol), NBS (553 mg, 3.11 mmol), and AIBN (cat.) in CCl₄ (30 mL) was refluxed for 2 h. The reaction mixture was filtered to remove insoluble materials, and the filtrate was concentrated to dryness under reduced pressure. The residue was purified by silica gel chromatography (1:20 EtOAc/Hexanes) to afford 380 mg (51%) of 4-(bromomethyl)-8-fluoroquinoline (**81**) as a white solid. LRMS (ESI+) *m/z*: calcd for C₁₀H₇BrFN 239.9, found 240.0 [M + H]⁺.

3-Chloro-*N***-((8-fluoroquinolin-4-yl)methyl)aniline (82).** A mixture of 4-(bromomethyl)-8fluoroquinoline (81, 380 mg, 1.59 mmol), 3-chloroaniline (400 mg, 3.15 mmol), and K₂CO₃ (440 mg, 3.19 mmol) in DMF (20 mL) was heated to 60°C for 2 h. The reaction mixture was filtered to remove insoluble materials, and then concentrated to dryness under reduced pressure. The residue was purified by silica gel chromatography (1:20 EtOAc/Hexanes) to afford 320 mg (70%) of 3-chloro-*N*-((8-fluoroquinolin-4-yl)methyl)benzenamine (82) as a white solid. LRMS (ESI+) m/z: calcd for C₁₆H₁₂CIFN₂ 287.1, found 287.0 [M + H]⁺.

N-Allyl-*N*-benzyl-2-fluoro-6-iodobenzamide (85). A mixture of 2-fluoro-6-iodobenzoic acid (84, 30 g, 113 mmol) in SOCl₂ (60 mL) was heated to 80°C for 1.5 h. Excess SOCl₂ was removed under reduced pressure and the residue was taken up with a solution of Et₃N (13.7 g, 136 mmol) in THF (50 mL). This solution was then added dropwise to a separate solution of N-benzylprop-2-en-1-amine (83, 16.6 g, 113 mmol) in THF (150 mL) at 5°C. The resulting reaction mixture was stirred at 25°C for 2 h. The mixture was concentrated under reduced pressure and the residue was dissolved in EtOAc (200 mL). The organic layer was washed with aqueous NaHSO₄ (2 x 50 mL), aqueous sodium bicarbonate (50 mL), and brine (50 mL), dried over Na₂SO₄, and concentrated to dryness under reduced pressure to afford 30 g (67%) of N-allyl-N-benzyl-2-fluoro-6-iodobenzamide (85) as a white solid. LRMS (ESI+) m/z: calcd for C₁₇H₁₅FINO 396.0, found 396.0 [M + H]⁺. Page 8 of 82

2-Benzyl-8-fluoro-4-methylisoquinolin-1(*2H*)-one (**86**). A mixture of *N*-allyl-*N*-benzyl-2-fluoro-6-iodobenzamide (**85**, 198 mg, 0.50 mmol), dicyclohexylamine (362 mg, 2.00 mmol), PPh₃ (13.1 mg, 0.05 mmol), and Pd(OAc)₂ (12 mg, 0.03 mmol) in DMA (7 mL) was heated to 100°C for 18 h. The mixture was concentrated under reduced pressure and the residue was purified by silica gel chromatography (1:30 EtOAc/Hexanes) to afford 83 mg (62%) of 2-benzyl-8-fluoro-4-methylisoquinolin-1(2*H*)-one (**86**) as a white solid. LRMS (ESI+) *m/z*: calcd for $C_{17}H_{14}FNO$ 268.1, found 268.0 [M + H]⁺.

8-Fluoro-4-methylisoquinolin-1(*2H*)-one (**87**). A solution of 2-benzyl-8-fluoro-4methylisoquinolin-1(2*H*)-one (**86**, 30 g, 112.36 mmol) in H₂SO₄ (450 g) was heated to 150°C for 2.5 h. Adjustment of the pH to 9 was accomplished by the addition of NaOH (4M). The resulting aqueous solution was extracted with EtOAc (3 x 100 mL). The organics were combined, washed with brine (2 x 500 mL), filtered, dried over Na₂SO₄, and concentrated under reduced pressure to afford 15.8 g (79%) of 8-fluoro-4-methylisoquinolin-1(2*H*)-one (**87**) as a white solid. LRMS (ESI+) *m/z*: calcd for C₁₀H₈FNO 178.1, found 178.0 [M + H]⁺.

1-Chloro-8-fluoro-4-methylisoquinoline (**88**). A mixture of 8-fluoro-4-methylisoquinolin-1(2*H*)-one (**87**, 3 g, 16.95 mmol) in POCl₃ (40 mL) was heated to 80°C for 2 h. The mixture was concentrated to dryness and the residue was partitioned between EtOAc and H₂O (1:1, 100 mL). Adjustment of the pH to 8 was accomplished by the addition of NH₃.H₂O (25%). The two layers were separated and the aqueous layer was extracted with EtOAc (50 mL). The organics were combined, dried over Na₂SO₄, filtered, and concentrated under reduced pressure to afford 3.3 g (99%) of 1-chloro-8-fluoro-4-methylisoquinoline (**88**) as a white solid. LRMS (ESI+) *m/z*: calcd for C₁₀H₇CIFN 196.0, found 196.0 [M + H]⁺. **4-(Bromomethyl)-1-chloro-8-fluoroisoquinoline** (**89**). Compound **89** was synthesized as described for compound **81** using 1-chloro-8-fluoro-4-methylisoquinoline (**88**) as the starting material. LRMS (ESI+) m/z: calcd for C₁₀H₆BrClFN 273.9, found 274.0 [M + H]⁺.

3-Chloro-*N***-((1-chloro-8-fluoroisoquinolin-4-yl)methyl)benzenamine (90)**. Compound **90** was synthesized as described for compound **82** using 1-chloro-8-fluoro-4-methylisoquinoline (**89**) and 3-chloroaniline as starting materials. LRMS (ESI+) m/z: calcd for C₁₆H₁₁Cl₂FN₂ 321.0, found 321.0 [M + H]⁺.

3-Chloro-*N***-((8-fluoroisoquinolin-4-yl)methyl)benzenamine (91)**. A solution of 3-chloro-*N*-((1-chloro-8-fluoroisoquinolin-4-yl)methyl)benzenamine (**90**, 500 mg, 1.56 mmol) and Pd/C (cat.) in MeOH (20 mL) was hydrogenated (with a balloon of hydrogen) for 4 h at 25°C. A filtration through celite was performed to remove the Pd/C and the filtrate was evaporated to dryness under reduced pressure. The residue was dissolved in EtOAc (30 mL) and the organic layer was washed with aqueous sodium bicarbonate (2 x 20 mL) and brine (2 x 20 mL). The organic layer was dried over Na₂SO₄, filtered, and concentrated under reduced pressure to afford 380 mg (85%) of 3-chloro-*N*-((8-fluoroisoquinolin-4-yl)methyl)benzenamine (**91**) as a beige solid. ¹HNMR (400MHz, DMSO-d₆) δ 9.41 (s, 1H), 8.59 (s, 1H), 7.99 (d, 1H, *J* = 8.4 Hz), 7.84 (m, 1H), 7.52 (dd, 1H, *J* = 10.8, 8.0 Hz), 7.06 (t, 1H, *J* = 8.4 Hz), 6.61 (m, 1H), 6.62-6.53 (m, 3H), 4.69 (d, 2H, *J* = 5.2 Hz). LRMS (ESI+) *m/z*: calcd for C₁₆H₁₂CIFN₂ 287.1, found 287.0 [M + H]⁺.

N-(2-Chlorophenyl)-3-oxobutanamide (98). Compound 98 was synthesized as described for compound 49 using 3-oxobutanoate and 2-chloroaniline (92) as starting materials. ¹H NMR (400 MHz, CDCl₃) δ 9.62 (s, 1H), 8.32 (dd, 1H, *J* = 8.2, 0.8 Hz), 7.37 (d, 1H, *J* = 8.0 Hz), 7.25 (t, 1H, *J* = 8.0 Hz), 7.05 (t, 1H, *J* = 8.0 Hz), 3.64 (s, 2H), 2.34 (s, 3H).

N-(**3-Fluorophenyl**)-**3-oxobutanamide** (**99**). Compound **99** was synthesized as described for compound **49** using 3-oxobutanoate and 3-fluoroaniline (**93**) as starting materials. LRMS (ESI+) m/z: calcd for C₁₀H₁₀FNO₂ 196.0, found 195.9 [M + H]⁺.

N-(**4-Fluorophenyl**)-**3-oxobutanamide** (**100**). Compound **100** was synthesized as described for compound **49** using 3-oxobutanoate and 4-fluoroaniline (**94**) as starting materials. LRMS (ESI+) m/z: calcd for C₁₀H₁₀FNO₂ 196.0, found 196.0 [M + H]⁺.

N-(2-Bromo-5-fluorophenyl)-3-oxobutanamide (101). Compound 101 was synthesized as described for compound 49 using 3-oxobutanoate and 2-bromo-5-fluoroaniline (95) as starting materials. ¹H NMR (400 MHz, CDCl₃) δ 9.73 (s, 1H), 8.20 (dd, 1H, *J* = 11.0, 3.0 Hz), 7.48 (dd, 1H, *J* = 8.8, 5.6 Hz), 6.72 (m, 1H), 3.65 (s, 2H), 2.33 (s, 3H). LRMS (ESI+) *m/z*: calcd for C₁₀H₉BrFNO₂ 273.9, found 277.9 [M + H]⁺.

N-(2,3-Difluorophenyl)-3-oxobutanamide (102). Compound 102 was synthesized as described for compound 49 using 3-oxobutanoate and 2,3-difluoroaniline (96) as starting materials. ¹H NMR (400 MHz, CDCl₃) δ 9.52 (s, 1H), 7.99 (m, 1H), 7.04 (m, 1H), 6.89 (s, 1H), 3.64 (s, 2H), 2.33 (s, 3H). LRMS (ESI+) *m/z*: calcd for C₁₀H₉F₂NO₂ 214.0, found 214.1 [M + H]⁺.

4-Bromo-*N***-(2-chlorophenyl)-3-oxobutanamide** (103). Compound 103 was synthesized as described for compound 50 using *N*-(2-chlorophenyl)-3-oxobutanamide (98) as the starting material. ¹H NMR (400 MHz, CDCl₃) δ 9.10 (s, 1H), 8.30 (dd, 1H, J = 8.4, 1.2 Hz), 7.39 (dd, 1H, J = 8.4, 1.4 Hz), 7.27 (m, 1H), 7.07 (td, 1H, J = 8.2, 1.6 Hz), 4.07 (s, 2H), 3.87 (s, 2H).

4-Bromo-*N***-(3-fluorophenyl)-3-oxobutanamide (104).** Compound **104** was synthesized as described for compound **50** using *N*-(3-fluorophenyl)-3-oxobutanamide (**99**) as the starting material. LRMS (ESI+) m/z: calcd for C₁₀H₉BrFNO₂ 273.9, found 273.8 [M + H]⁺.

4-Bromo-*N***-(4-fluorophenyl)-3-oxobutanamide (105).** Compound **105** was synthesized as described for compound **50** using *N*-(4-fluorophenyl)-3-oxobutanamide (**100**) as the starting material. LRMS (ESI+) m/z: calcd for C₁₀H₉BrFNO₂ 273.9, found 274.0 [M + H]⁺.

4-Bromo-*N***-(2-bromo-5-fluorophenyl)-3-oxobutanamide** (106). Compound 106 was synthesized as described for compound 50 using *N*-(2-bromo-5-fluorophenyl)-3-oxobutanamide (101) as the starting material. ¹H NMR (400 MHz, DMSO-d₆) δ 9.79 (s, 1H), 7.72 (m, 2H), 7.03 (m, 1H), 4.49 (s, 2H), 3.87 (s, 2H).

4-Bromo-*N***-(2,3-difluorophenyl)-3-oxobutanamide (107).** Compound **107** was synthesized as described for compound **50** using *N*-(2,3-difluorophenyl)-3-oxobutanamide (**102**) as the starting material. ¹H NMR (400 MHz, CDCl₃) δ 8.91 (s, 1H), 8.05 (m, 1H), 7.09 (m, 1H), 6.95 (m, 1H), 4.02 (s, 2H), 3.82 (s, 2H). LRMS (ESI+) *m/z*: calcd for C₁₀H₈BrF₂NO₂ 291.9, found 292.0 [M + H]⁺.

4-Bromo-2-fluoro-*N*-(**2-fluorophenyl**)-**3-oxobutanamide** (**108**). A mixture of 4-bromo-*N*-(2-fluorophenyl)-3-oxobutanamide (**50**, 1 g, 3.65 mmol) and Selectfluor® (0.7 g, 4.74 mmol) in ACN (30 mL) was heated to 60°C for 2 h. The solvent was removed under reduced pressure and the residue was dissolved in DCM (30 mL). The organic layer was washed with water (2 x 10 mL), dried over MgSO₄, filtered, and concentrated under reduced pressure. The crude residue was purified by silica gel column chromatography (0- 50% EtOAc/Hexanes) to afford 0.5 g of 4-bromo-2-fluoro-N-(2-fluorophenyl)-3-oxobutanamide (**108**) as a yellow solid. ¹H NMR (400 MHz, CDCl₃) δ 8.21 (s, 1H), 8.19 (m, 1H), 7.17-7.11 (m, 3H), 5.84 (d, 1H, *J* = 48.4 Hz), 4.34 (dd, 2H, *J* = 69.6, 12.8 Hz).

4-(Bromomethyl)-8-chloroquinolin-2(1H)-one (109). Compound **109** was synthesized as described for compound **52** using 4-bromo-*N*-(2-chlorophenyl)-3-oxobutanamide (**103**) as the starting material. LRMS (ESI+) m/z: calcd for C₁₀H₇BrClNO 271.9, found 272.8 [M + H]⁺. Page **12** of **82**

4-(Bromomethyl)-7-fluoroquinolin-2(1H)-one (110). Compound **110** was synthesized as described for compound **52** using 4-bromo-*N*-(3-fluorophenyl)-3-oxobutanamide (**104**) as the starting material. ¹H NMR (400 MHz, DMSO-d₆) δ 11.91 (s, 1H), 7.90 (dd, 1H, *J* = 9.0, 5.8 Hz), 7.11 (m, 2H), 6.70 (s, 1H), 4.89 (s, 2H). LRMS (ESI+) *m/z*: calcd for C₁₀H₇BrFNO 255.9, found 255.8 [M + H]⁺.

4-(Bromomethyl)-6-fluoroquinolin-2(1H)-one (111). Compound **111** was synthesized as described for compound **52** using 4-bromo-*N*-(4-fluorophenyl)-3-oxobutanamide (**105**) as the starting material. LRMS (ESI+) m/z: calcd for C₁₀H₇BrFNO 255.9, found 256.0 [M + H]⁺.

8-Bromo-4-(bromomethyl)-5-fluoroquinolin-2(1H)-one (112). Compound 112 was synthesized as described for compound 52 using 4-bromo-*N*-(2-bromo-5-fluorophenyl)-3-oxobutanamide (106) as the starting material. ¹H NMR (400 MHz, DMSO-d₆) δ 10.64 (s, 1H), 7.91 (dd, 1H, *J* = 9.0, 5.0), 7.14 (m, 1H), 6.90 (s, 1H), 4.88 (s, 2H).

4-(Bromomethyl)-7,8-difluoroquinolin-2(1H)-one (113). Compound **113** was synthesized as described for compound **52** using 4-bromo-*N*-(2,3-difluorophenyl)-3-oxobutanamide (**107**) as the starting material. ¹H NMR (400 MHz, DMSO-d₆) δ 7.70 (m, 1H), 7.34 (m, 1H), 6.78 (s, 1H), 4.90 (s, 2H).

4-(Bromomethyl)-3,8-difluoroquinolin-2(1H)-one (114). Compound **114** was synthesized as described for compound **52** using 4-bromo-2-fluoro-*N*-(2-fluorophenyl)-3-oxobutanamide (**108**) as the starting material. ¹H NMR (400 MHz, DMSO-d₆) δ 12.50 (s, 1H), 7.72 (d, 1H, *J* = 8.0 Hz), 7.47 (dd, 1H, *J* = 10.8, 7.6 Hz), 7.31 (m, 1H), 4.91 (s, 2H). LRMS (ESI+) *m/z*: calcd for C₁₀H₆BrF₂NO 273.9, found 273.7 [M + H]⁺.

4-((3-Chlorophenylamino)methyl)quinolin-2(1H)-one (115). Compound 115 was synthesized as described for compound 53 using 4-(bromomethyl) quinolin-2(1H)-one (51, commercially

available) and 3-chloroaniline as starting materials. ¹H NMR (400 MHz, DMSO-d₆) δ 11.68 (s, 1H), 7.82 (dd, 1H, J = 8.2, 1.2 Hz), 7.51 (td, 1H, J = 8.4, 1.2 Hz), 7.33 (dd, 1H, J = 8.4, 1.2 Hz), 7.20 (td, 1H, J = 8.0, 1.2 Hz), 7.06 (t, 1H, J = 8.0 Hz), 6.61 (s, 1H), 6.54 (m, 2H), 6.37 (s, 1H), 4.54 (s, 2H). LRMS (ESI+) m/z: calcd for C₁₆H₁₃ClN₂O 285.1, found 285.2 [M + H]⁺.

8-Chloro-4-((**3-chlorophenylamino)methyl)quinolin-2(1H)-one** (**116**). Compound **116** was synthesized as described for compound **53** using 4-(bromomethyl)-8-chloroquinolin-2(1H)-one (**109**) and 3-chloroaniline as starting materials. ¹H NMR (400 MHz, DMSO-d₆) δ 10.92 (s, 1H), 7.84 (d, 1H, *J* = 8.4 Hz), 7.69 (d, 1H, *J* = 8.0 Hz), 7.23 (t, 1H, *J* = 7.8 Hz), 7.08 (t, 1H, *J* = 8.0 Hz), 6.64-6.56 (m, 3H), 6.46 (s, 1H), 4.58 (m, 2H). LRMS (ESI+) *m/z*: calcd for C₁₆H₁₂Cl₂N₂O 319.0, found 318.9 [M + H]⁺.

4-((3-Chlorophenylamino)methyl)-7-fluoroquinolin-2(1H)-one (117). Compound **117** was synthesized as described for compound **53** using 4-(bromomethyl)-7-fluoroquinolin-2(1H)-one (**110**) and 3-chloroaniline as starting materials. LRMS (ESI+) m/z: calcd for C₁₆H₁₂ClFN₂O 303.1, found 302.9 [M + H]⁺.

4-((3-Chlorophenylamino)methyl)-6-fluoroquinolin-2(1H)-one (118). Compound **118** was synthesized as described for compound **53** using 4-(bromomethyl)-6-fluoroquinolin-2(1H)-one (**111**) and 3-chloroaniline as starting materials.

8-Bromo-4-((3-chlorophenylamino)methyl)-5-fluoroquinolin-2(1H)-one (119). Compound 119 was synthesized as described for compound 53 using 8-bromo-4-(bromomethyl)-5-fluoroquinolin-2(1H)-one (112) and 3-chloroaniline as starting materials. LRMS (ESI+) m/z: calcd for C₁₆H₁₁BrClFN₂O 380.9, found 380.6 [M + H]⁺.

4-((3-Chlorophenylamino)methyl)-7,8-difluoroquinolin-2(1H)-one (120). Compound **120** was synthesized as described for compound **53** using 4-(bromomethyl)-7,8-difluoroquinolin-2(1H)-

one (**113**) and 3-chloroaniline as starting materials. ¹H NMR (400 MHz, DMSO-d₆) δ 11.96 (s, 1H), 7.70 (m, 1H), 7.30 (m, 1H), 7.08 (t, 1H, *J* = 8.0 Hz), 6.63-6.55 (m, 3H), 6.40 (s, 1H), 4.54 (m, 2H). LRMS (ESI+) *m/z*: calcd for C₁₆H₁₁ClF₂N₂O 321.0, found 320.9 [M + H]⁺.

4-((3-Chlorophenylamino)methyl)-3,8-difluoroquinolin-2(1H)-one (121). Compound **121** was synthesized as described for compound **53** using 4-(bromomethyl)-3,8-difluoroquinolin-2(1H)-one (**114**) and 3-chloroaniline as starting materials. LRMS (ESI+) m/z: calcd for C₁₆H₁₁ClF₂N₂O 321.0, found 320.8 [M + H]⁺.

N-((8-Bromo-5-fluoro-2-oxo-1,2-dihydroquinolin-4-yl)methyl)-N-(3-chlorophenyl)-4-

methylthiazole-5-carboxamide (122). Compound 122 was synthesized as described for compound 7 using 8-bromo-4-((3-chlorophenylamino)methyl)-5-fluoroquinolin-2(1H)-one (119) and 4-methylthiazole-5-carboxylic acid as starting materials. ¹H NMR (400 MHz, DMSO-d₆) δ 10.59 (s, 1H), 8.97 (s, 1H), 7.90 (dd, 1H, J = 8.4, 4.8 Hz), 7.56 (s, 1H), 7.34-7.31 (m, 2H), 7.27 (m, 1H), 7.10 (dd, 1H, J = 12.0, 8.4 Hz), 6.54 (s, 1H), 5.36 (s, 2H), 2.44 (s, 3H). LRMS (ESI+) m/z: calcd for C₂₁H₁₄BrClFN₃O₂S 505.9, found 505.7 [M + H]⁺.

Low Temperature SDS-PAGE: RAW 264.7 cells were seeded into 6-well dishes at a density of 1.5×10^6 cells/well in 2 mL of growth media. Cells were incubated for 2-3 h at 37°C and 10% CO₂. Next, 1 mL of media was removed from each well and replaced with 1 mL of a 2X cocktail containing IFNy (100 U/mL final), LPS (2 µg/mL final) and compound or vehicle (0.1% DMSO final) diluted in DMEM without serum. Cells were incubated overnight (~15 hr) at 37°C and 10% CO₂. Cells were washed once with ice cold PBS and then 200 µL of ice-cold lysis buffer containing protease inhibitors was added to each well. Cells were scraped from the dish and transferred to micro-centrifuge tubes on ice. Samples were sonicated for 5 seconds at setting 4 and centrifuged for 10 minutes at 4°C. Supernatants were transferred to clean tubes and stored on ice. The concentration of protein in the cell extracts was determined using a Bradford assay with a BSA standard curve as described by the manufacturer (Advanced). Equivalent amounts of protein extracts were added to micro-centrifuge tubes and the volume was adjusted to 10 µL with lysis buffer. An equal volume of ice-cold 2X loading buffer was added to each sample and gently pipetted to mix. Samples were stored on ice until ready to load. Twenty microliters of each sample was loaded onto a 15-well 4-20% Tris-glycine polyacrylamide gel. The gel was run in pre-chilled 1X SDS running buffer in a cold room for 2.5 h at 125 V. Proteins were transferred to nitrocellulose for 2.5 h at 70 V in 1X transfer buffer. The membrane was blocked overnight in Blotto diluted in western wash buffer at 4°C. The membrane was incubated in a 1:2500 dilution of mouse anti-iNOS antibody (BD Biosciences) in Blotto for 1 h at room temperature followed by 3 x 5 min washes in western wash buffer. The blot was then incubated in a 1:2000 dilution of goat anti-mouse HRP-conjugated secondary antibody (Santa Cruz Biotech) in Blotto for 1 hour at room temperature. Following 4 x 5 min washes, the proteins were visualized by chemiluminescence using West Dura detection reagent as described by the manufacturer (Pierce). The data were captured on a CCD-based imaging device (Alpha Inotech).

X-Ray Diffraction Data: *Crystal data*: C₂₁H₁₅CIFN₃O₂S, f.w. = 427.87, colorless needle, triclinic, *P*-1, *a* = 8.8921(13), *b* = 17.982(3), *c* = 18.935(3) Å, *a* = 72.680(2), β = 82.543(2), γ = 77.465(2)°, *V* = 2814.5 Å³, *Z* = 6, *Z*' = 3, *d*(calcd) = 1.515 g cm⁻³. 16620 reflections were collected at 100 K using a Bruker D8 platform diffractometer with an APEX CCD detector and MoKa radiation. The structure was refined using 9455 independent reflections with all nonhydrogen atoms anisotropic and hydrogen atoms treated as idealized contributions. The symmetry independent structure consists of three chemically similar molecules. All software is contained in the APEX, SAINT and SHELXTL libraries of programs maintained by Bruker-AXS (Madison, WI).

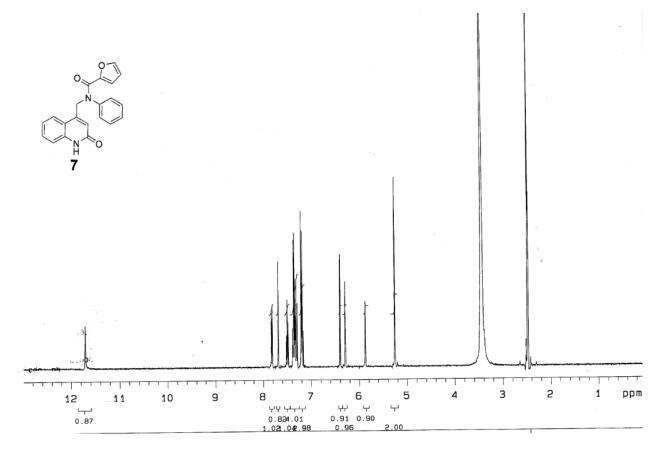
Mouse Pharmacokinetics: Male (Balb/c) mice (6 to 8 weeks of age) were obtained from Charles-River Laboratories (Hollister, CA). Animals were acclimated for a minimum of 3 days prior to performing any experimental procedures. A 12-hr light: 12-hr dark cycle was maintained throughout the course of all experiments in a temperature and humidity controlled vivarium facility. Animals were fed standard laboratory rodent chow, and water ad libitum. For intravenous (IV) administration, 12 overnight fasted animals received a single dose of 3 or 1 mg/kg body weight (3 or 1 mL/kg) via penile vein injection under light isoflurane anesthesia in 10% DMSO; 30% PEG-400 and 60% HPBCD (30% w/v in water). For oral (PO) administration, 9 overnight fasted animals received a single dose of 10 mg/kg body weight (10 mL/kg) via oral gavage in a suspension of 0.5% Tween-80, 0.5% PVP-K30, 9% PEG-400 and 90% CMC in water (0.5% w/v). PO dose suspensions were dispersed with an electric homogenizer, followed by treatment with a sonicator probe on wet ice. Blood samples (~ 125μ L) were collected by puncture of the submandibular vein (i.e. the cheek pouch method) into lithium heparin pretreated tubes at 0 (pre-dose), 5, 10, 20 and 40 minutes, 1, 2, 4, 8, 12 and 24 hours following IV administration and at 0 (pre-dose), 0.25, 0.5, 1, 2, 4, 8, 12 and 24 hours post-dose following PO

administration, such that no animal was sampled more than 3 times. Plasma was isolated by centrifugation and stored at -80°C until analysis by LC-MS/MS.

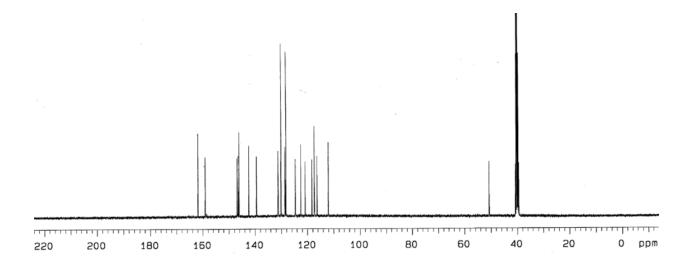
Sample analyses were performed by LC-MS using a PE Sciex API 4000 Q-Trap mass spectrometer tandem with Agilent 1100 chromatography system. Polar-RP Synergi column from Phenomenex (2.0 x 3.0 mm, 4.0 micron). Chromatography was performed using a gradient system using two mobile phases A and B. The mobile phase A consisted of 0.1% formic acid in water and mobile phase B of 0.1% formic acid in acetonitrile. Mobile phase B was ramped from 0% to 100% from 0.0 to 1.0 min and was held at 100% B from 1.0 to 1.9 min. From 2.0 to 2.7 min, column was equilibrated to the initial condition at 100% A. Flow rate for the gradient was set at 1.0 mL/min. Mass spectrometry detection was carried out on a PE Sciex API4000 Q-Trap equipped with a Turbo IonSpray source. ESI+ mass spectra were acquired with multiple reaction monitoring (MRM). The plasma samples were prepared by protein precipitation monitored with an internal standard. Standard curve was prepared by spiking control plasma with the compound and serial dilutions from this spiked solution generated different levels of the standard curve. A weighted linear least squares regression was then performed to generate a correlation between concentration and the relative amounts of analyte and internal standard for compound. Twelve standard curve were used, highest level at 10,000 ng/mL. Lowest level of detection was at 4.0 ng/mL.

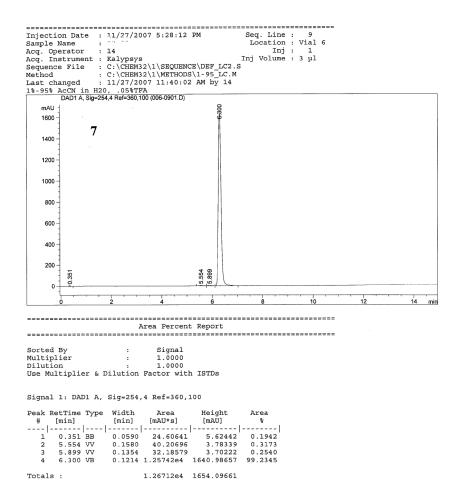
Mouse Liver Microsome Assay: Test article in DMSO was added to incubation buffer such that the final DMSO concentration was 0.4% (v/v). Microsomes were stored at -70°C until use, thawed on ice and then diluted to 5 mg/mL in Phosphate buffer immediately before use. Microsome incubations were performed in 100 mM Phosphate buffer (pH 7.4), containing 0.1% BSA (fatty acid free), 1 mM EDTA, 3 mM MgCl2, alamethicin (50 µg/mg microsomal protein) and 1.25 mM D-Saccharic acid 1,4-lactone. D-saccharic acid 1,4-lactone is an inhibitor of βglucuronidase, an enzyme which can rapidly hydrolyze glucuronide conjugates. Microsomes Page **18** of **82** and test article in incubation buffer were pre-warmed in triplicate at 37°C for 5 minutes. Reactions were initiated by addition of 25 μ L of NADPH-regenerating system (consisting of 20 mM NADP, 100 mM glucose-6-phosphate, 20 U/mL glucose-6-phosphate dehydrogenase and 100 mM UDPGA). Final incubation volume was 0.5 mL with a test article concentration of 5 μ M. At specified time points, 100 μ L aliquots were removed and combined with 300 μ L of chilled quenching reagent (1:2 water: acetonitrile) containing internal standard (IS). Sample plates were then centrifuged at 1,500 g for 15 minutes and supernatants submitted for LC-MS/MS analysis. Relative test article concentrations were defined by the analyte/IS peak ratios in each sample.

Cytochrome P450 Inhibition Assay: Compounds were evaluated for biochemical inhibition of cytochrome P450 using the Vivid CYP450 Blue assay (Invitrogen) adapted to 1536-well plate format. Briefly, individual isozymes and their blue substrates were diluted to a 2X working concentration in 1X assay buffer. Isozyme concentrations were: 3A4, 10 nM; 2D6, 40 nM; 1A2, 12 nM; 2C9, 20 nM; and 2C19, 10 nM. 2X Regeneration System was included in the isozyme mix. Substrate concentrations were: 40 nM for 3A4 and 2D6; 20 μ M for 1A2, 2C9, and 2C19. NADP+ was added to the substrate mix at a concentration of 400 μ M for the assays. A volume of 2.5 μ L of the isozyme mix was dispensed into black Greiner 1536-well plates. Then, 30 nL of compound was added to the wells. Subsequently, 2.5 μ L of the blue substrate was added and plates were incubated for 1 hour at room temperature. Four replicate data points are obtained per compound at each concentration. Inhibition of substrate turnover was measured using a fluorescent plate reader with excitation at 405 nm and emission at 450 nm (Molecular Devices).

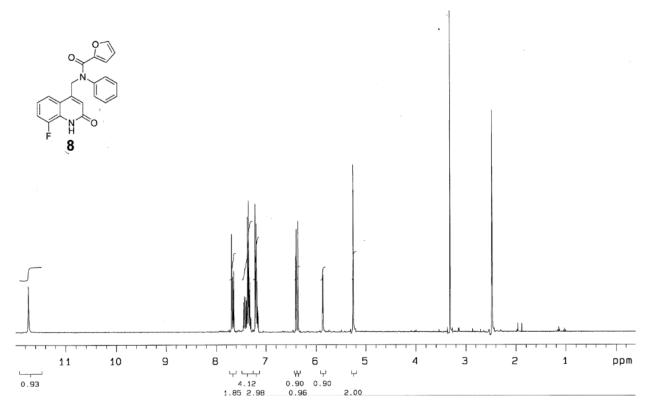


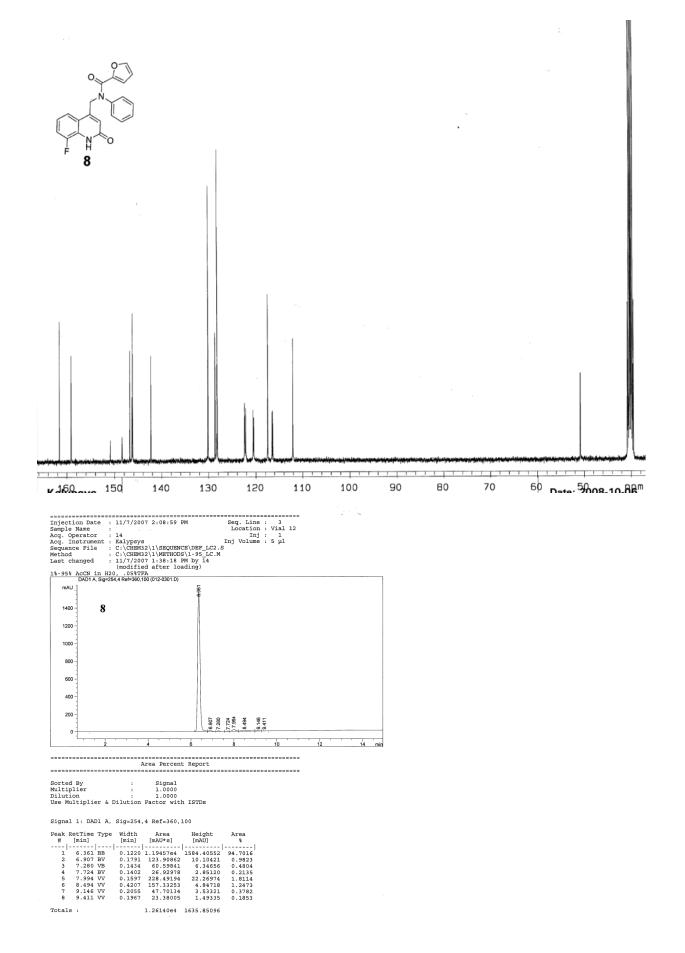
Compound 7: N-((2-Oxo-1,2-dihydroquinolin-4-yl)methyl)-N-phenylfuran-2-carboxamide



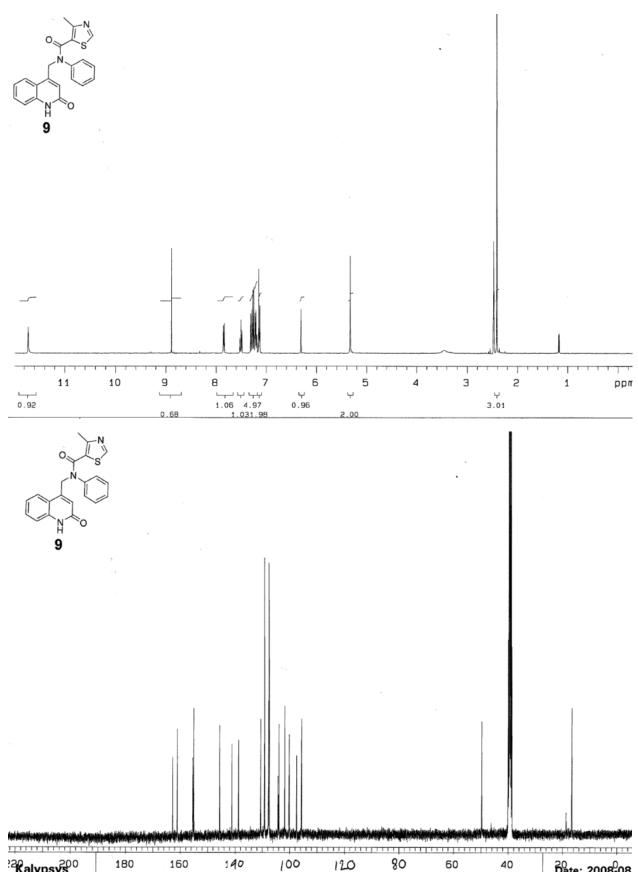


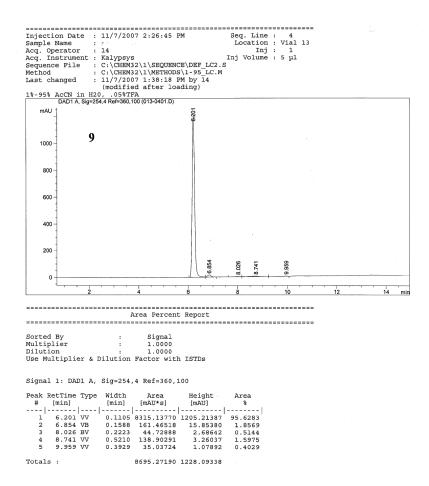
Compound 8: *N*-((8-Fluoro-2-oxo-1,2-dihydroquinolin-4-yl)methyl)-*N*-phenylfuran-2-carboxamide

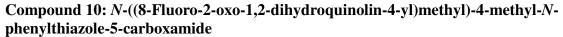


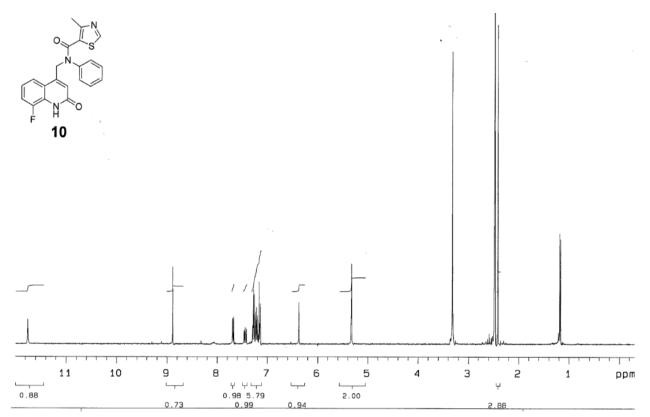


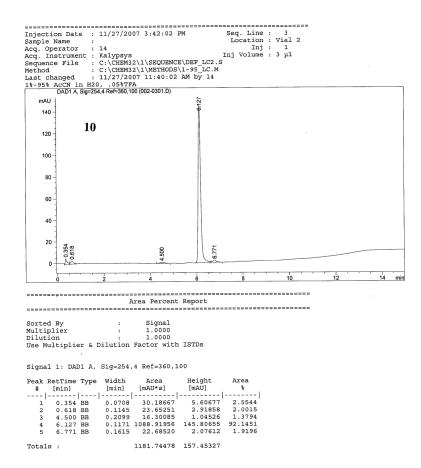
Compound 9: 4-Methyl-*N*-((2-oxo-1,2-dihydroquinolin-4-yl)methyl)-*N*-phenylthiazole-5-carboxamide

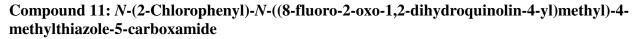


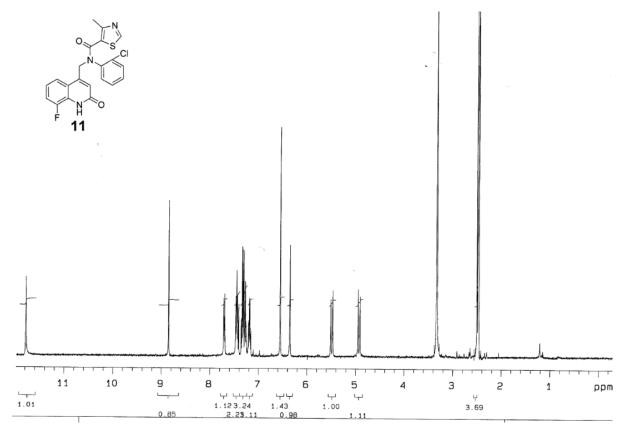


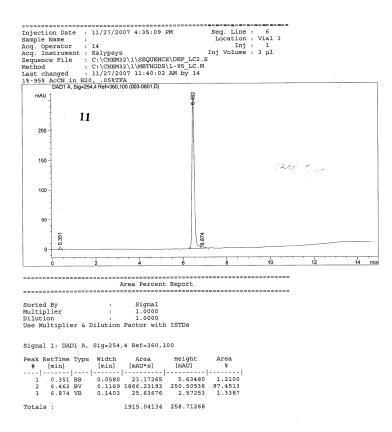


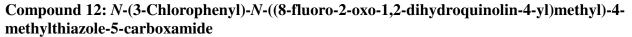


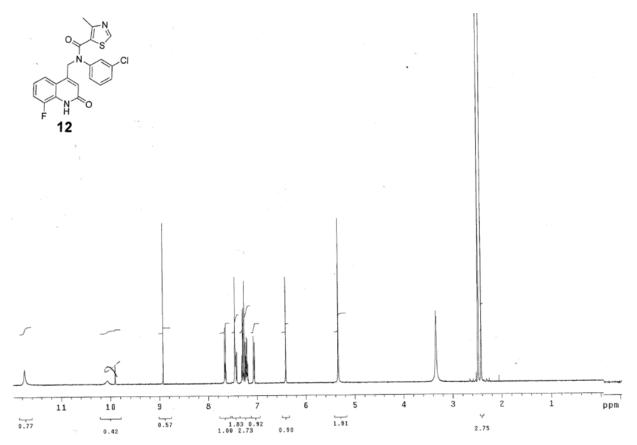


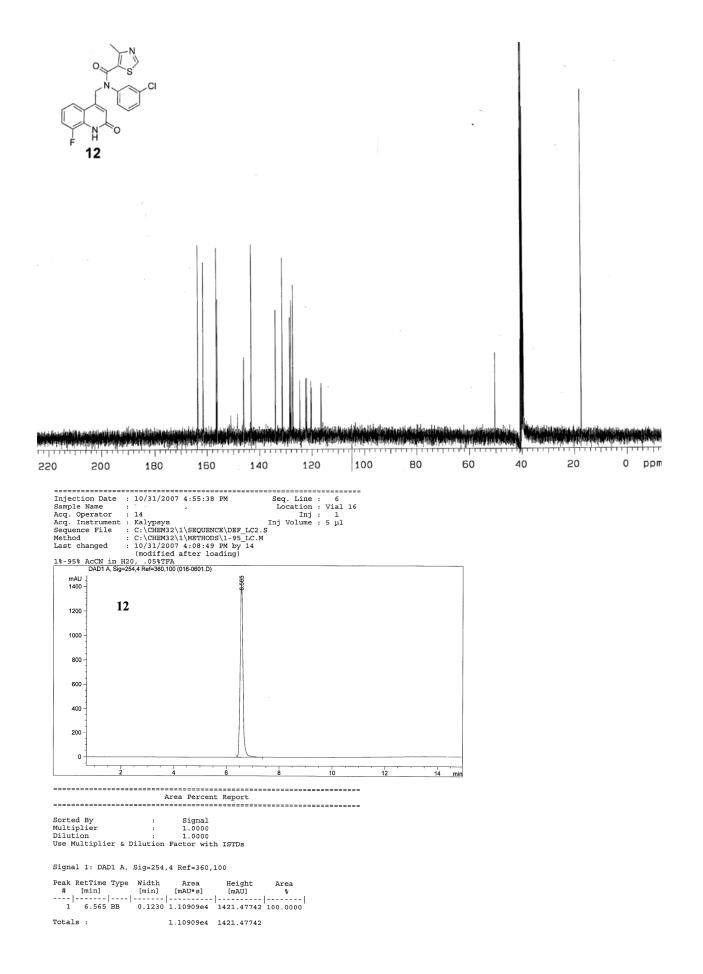




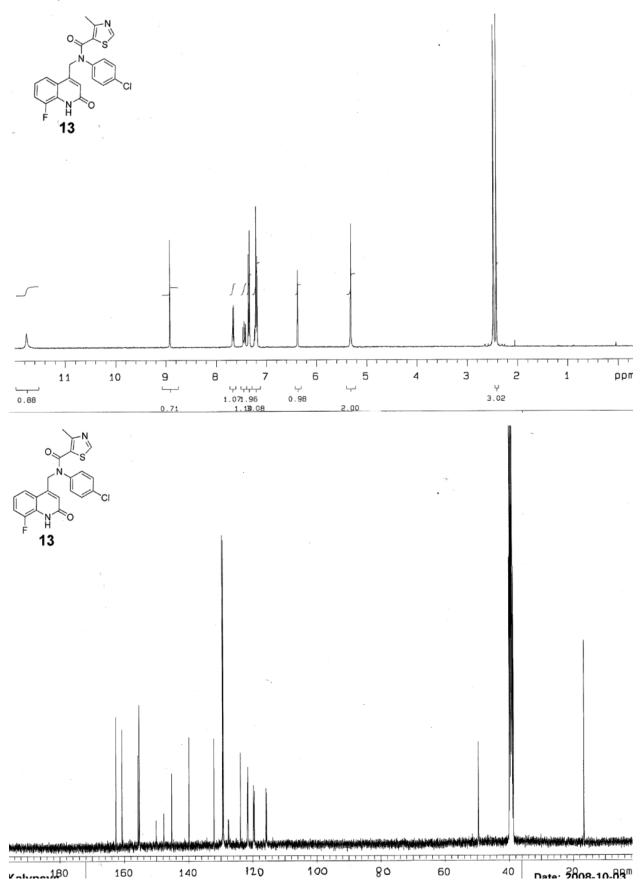




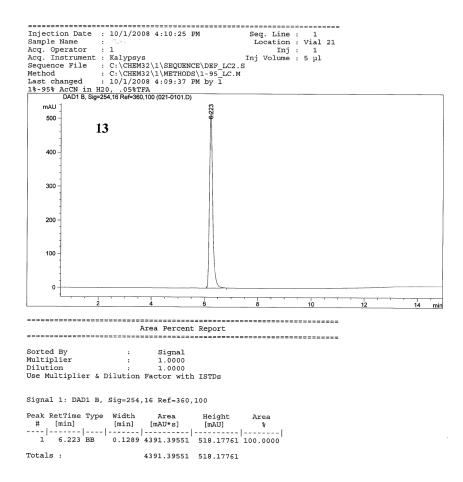




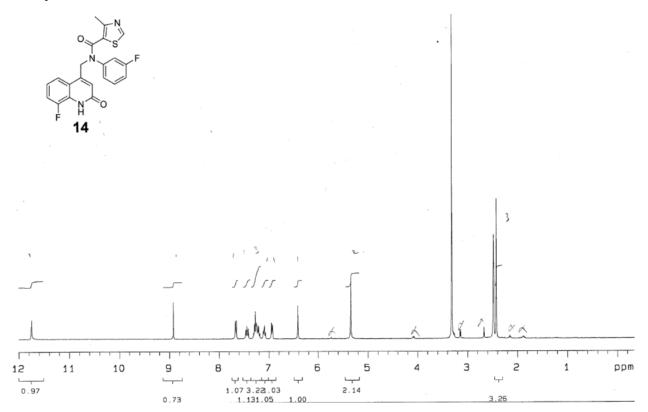
Compound 13: N-(4-Chlorophenyl)-N-((8-fluoro-2-oxo-1,2-dihydroquinolin-4-yl)methyl)-4-methylthiazole-5-carboxamide

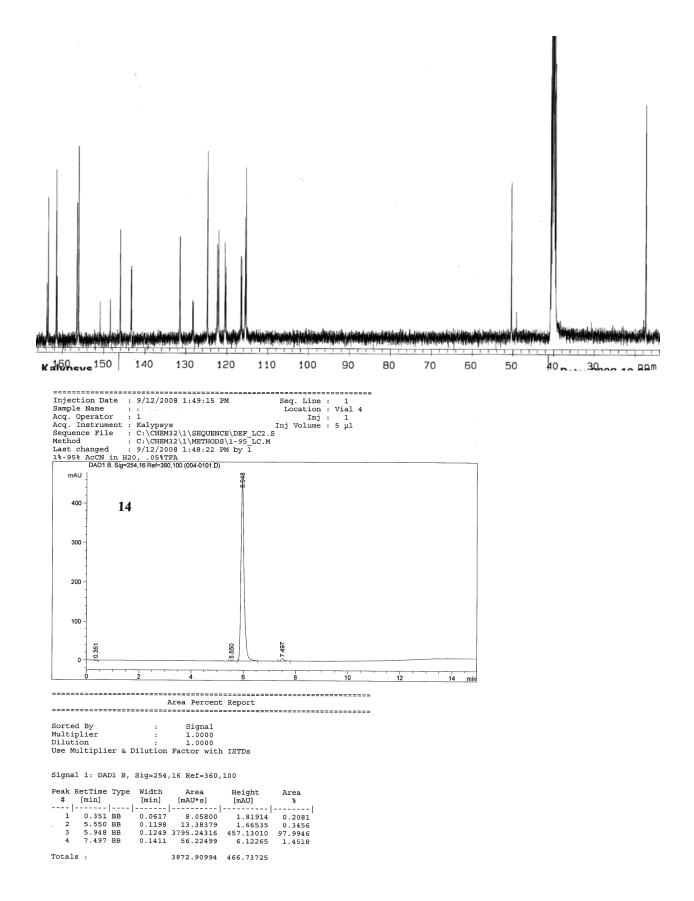


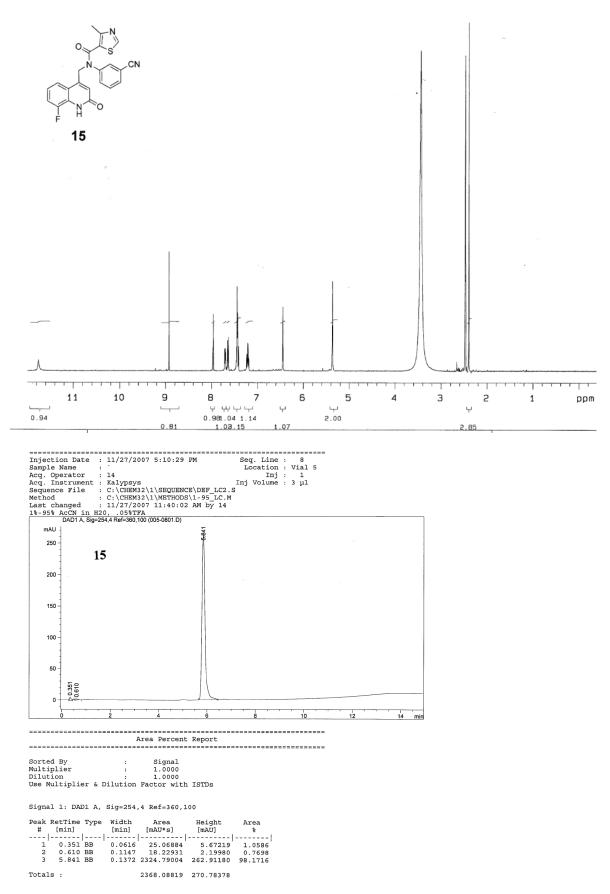
Page **28** of **82**



Compound 14: *N*-((8-Fluoro-2-oxo-1,2-dihydroquinolin-4-yl)methyl)-*N*-(3-fluorophenyl)-4-methylthiazole-5-carboxamide

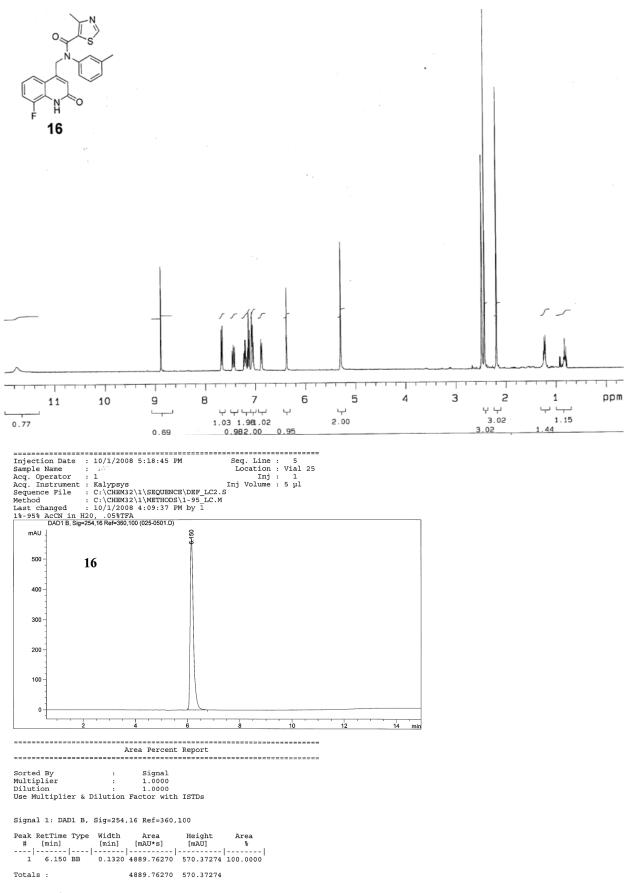






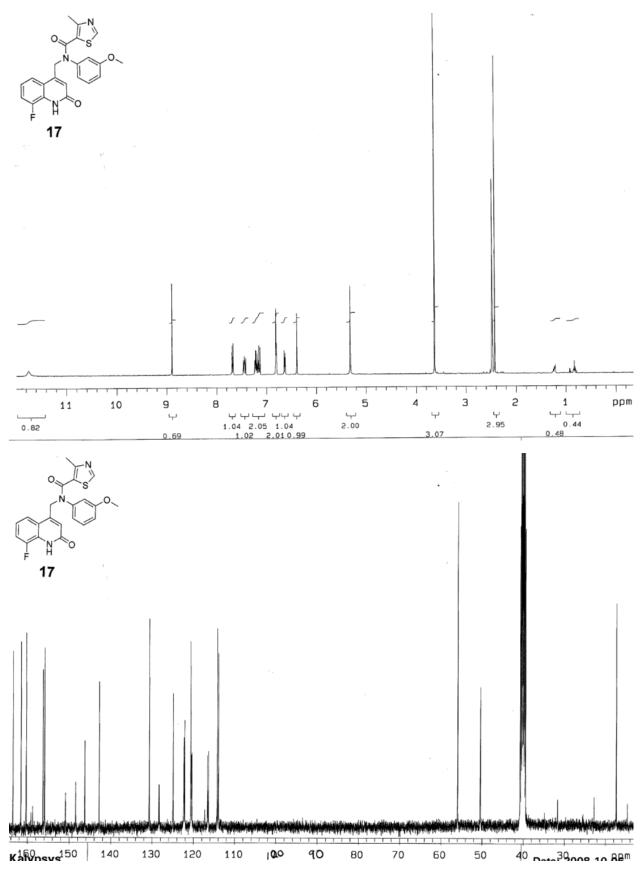
Page **31** of **82**

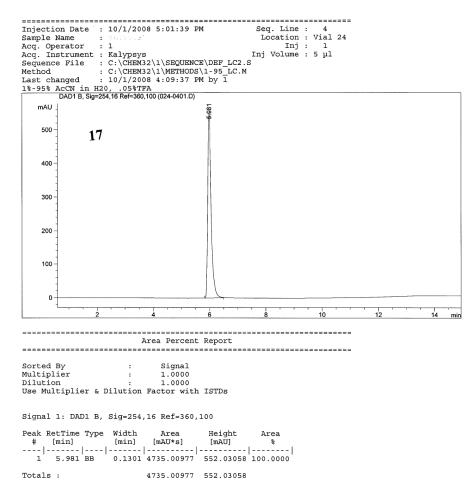
Compound 16: *N*-(3-Methylphenyl)-*N*-((8-fluoro-2-oxo-1,2-dihydroquinolin-4-yl)methyl)-4-methylthiazole-5-carboxamide



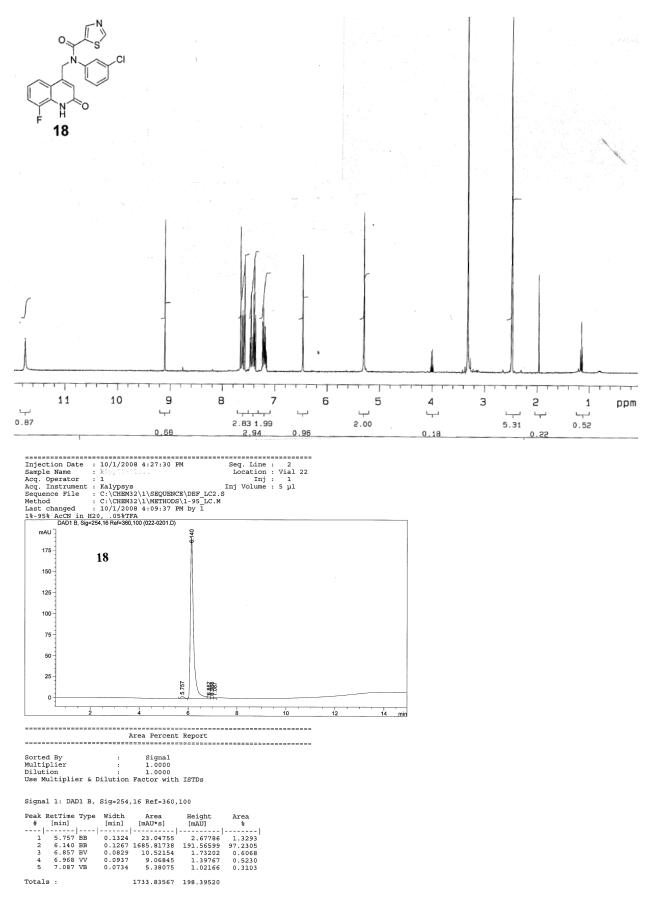


Compound 17: N-(3-Methoxyphenyl)-N-((8-fluoro-2-oxo-1,2-dihydroquinolin-4-yl)methyl)-4-methylthiazole-5-carboxamide



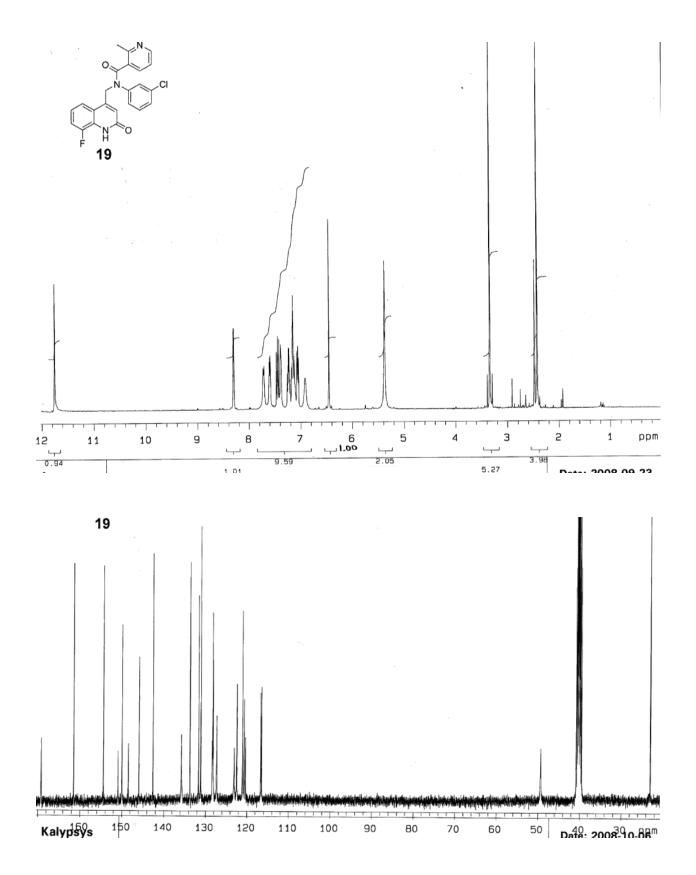


Compound 18: *N*-(3-Chlorophenyl)-*N*-((8-fluoro-2-oxo-1,2-dihydroquinolin-4-yl)methyl)thiazole-5-carboxamide



Compound 19: *N*-(3-Chlorophenyl)-*N*-((8-fluoro-2-oxo-1,2-dihydroquinolin-4-yl)methyl)-2-methylnicotinamide

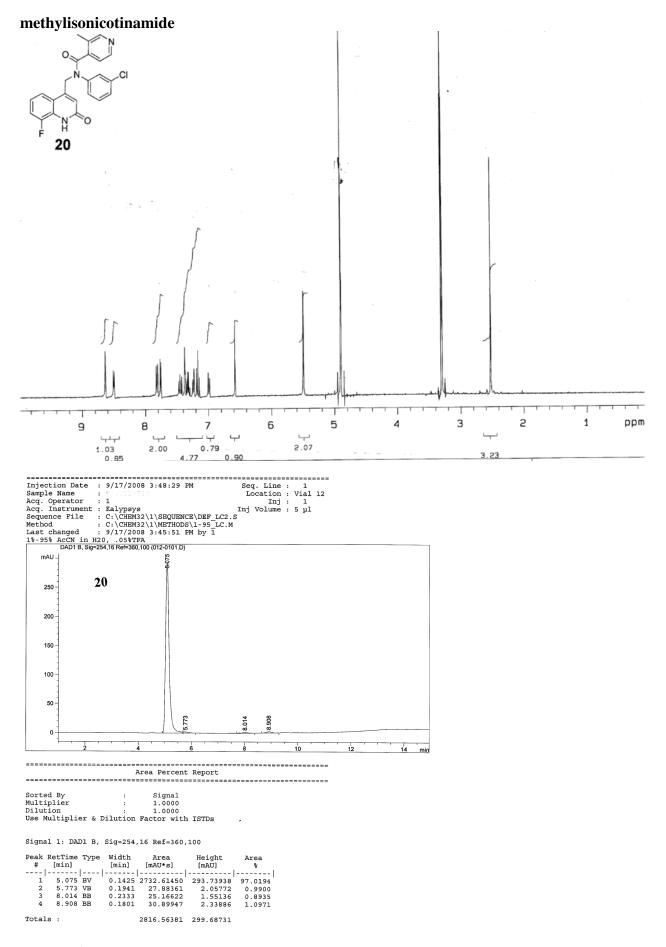
Page **35** of **82**



	08 9:06:13 AM	Seq. Line		
Sample Name :			: Vial 3	
Acq. Operator : 1		Inj		
Acq. Instrument : Kalypsy Sequence File : C:\CHEN	'S 132\1\SEQUENCE\D	Inj Volume	: 5 µl	
	132\1\METHODS\1-			
	08 9:05:30 AM b			
1%-95% AcCN in H20, .05%		1 -		
DAD1 A, Sig=254,4 Ref=360			CONTRACTOR AND	
mAU	6-088			
400 19	Ĩ			
350				
300 -				
250 -				
200				
150	-			
100				
50 - Se		.855		
0 0 0		8.9		
1				
0 2	4	6 8	10	12 14 mi

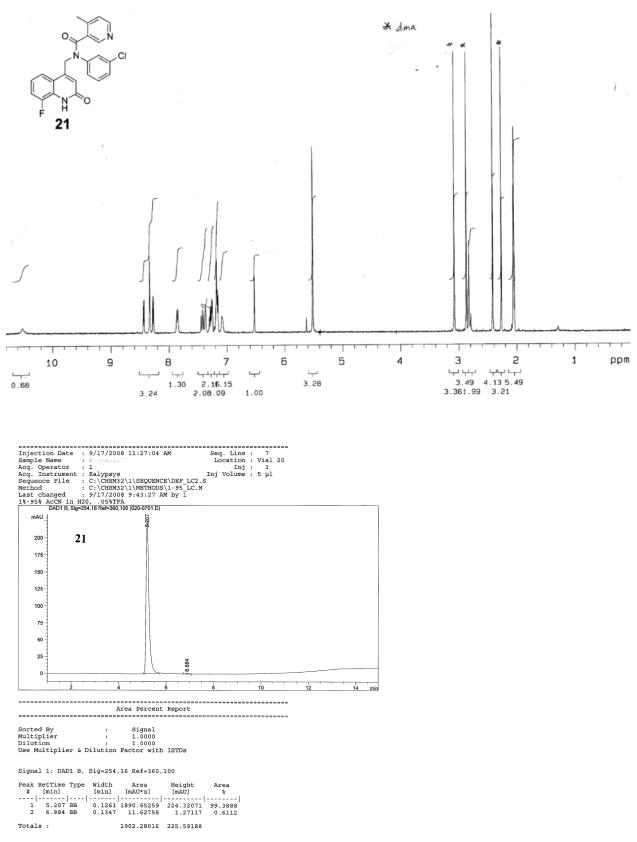
	Area Percent Rep	port		
Sorted By :	Signal			
Multiplier :	1.0000			
Dilution :	1.0000			
Use Multiplier & Dilution	Factor with IST	IDs		
Signal 1: DAD1 A, Sig=254	,4 Ref=360,100			
Peak RetTime Type Width	Area He	eight Area		
# [min] [min]		nAU] %		
1 0.350 BB 0.0633				
	9.55625 2 4193.13330 426	2.01589 0.2243 5.41476 98.3991		
3 6.855 VB 0.1300		5.22083 1.3767		
Totals :	4261.35497 434			
	aco1.3342/ 434			

Compound 20: *N*-(3-Chlorophenyl)-*N*-((8-fluoro-2-oxo-1,2-dihydroquinolin-4-yl)methyl)-3-



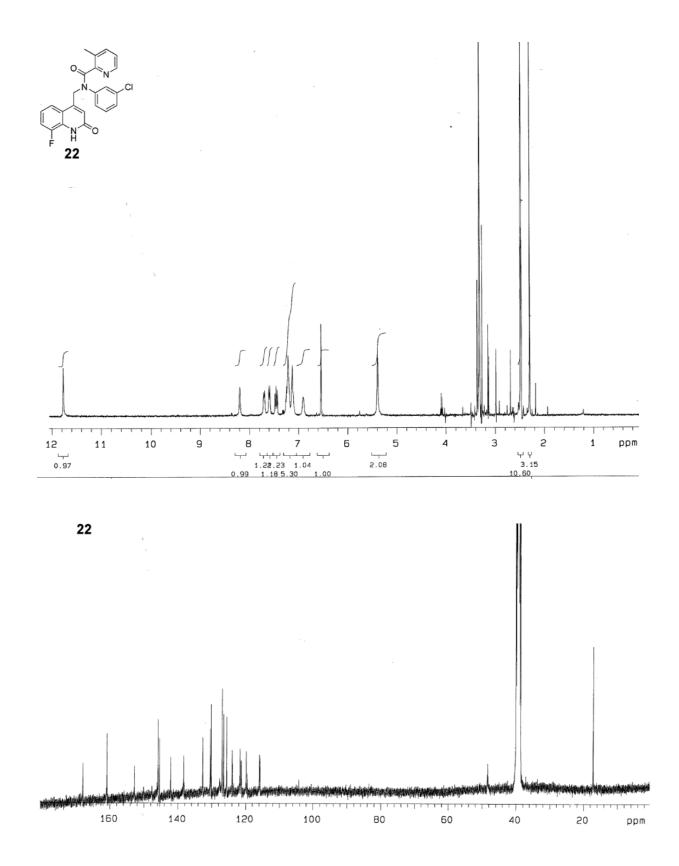
Page 38 of 82

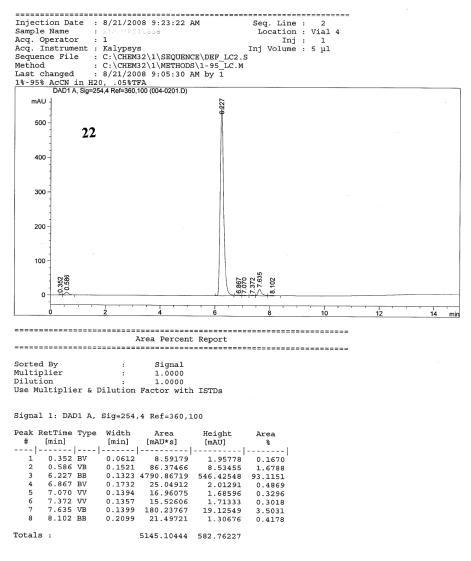
Compound 21: *N*-(3-Chlorophenyl)-*N*-((8-fluoro-2-oxo-1,2-dihydroquinolin-4-yl)methyl)-4-methylnicotinamide



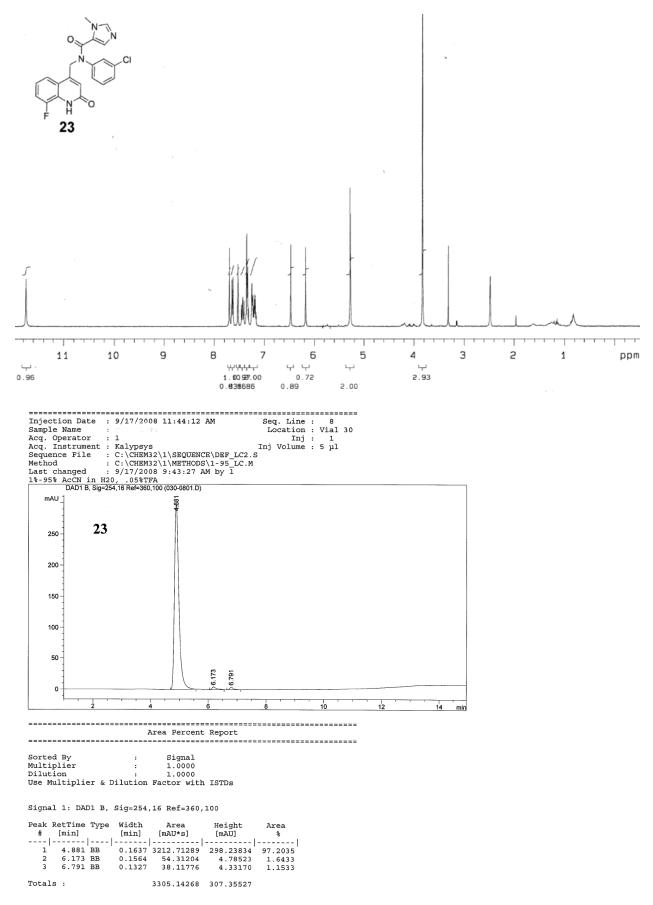
Compound 22: *N*-(3-Chlorophenyl)-*N*-((8-fluoro-2-oxo-1,2-dihydroquinolin-4-yl)methyl)-3-methylpicolinamide

Page **39** of **82**



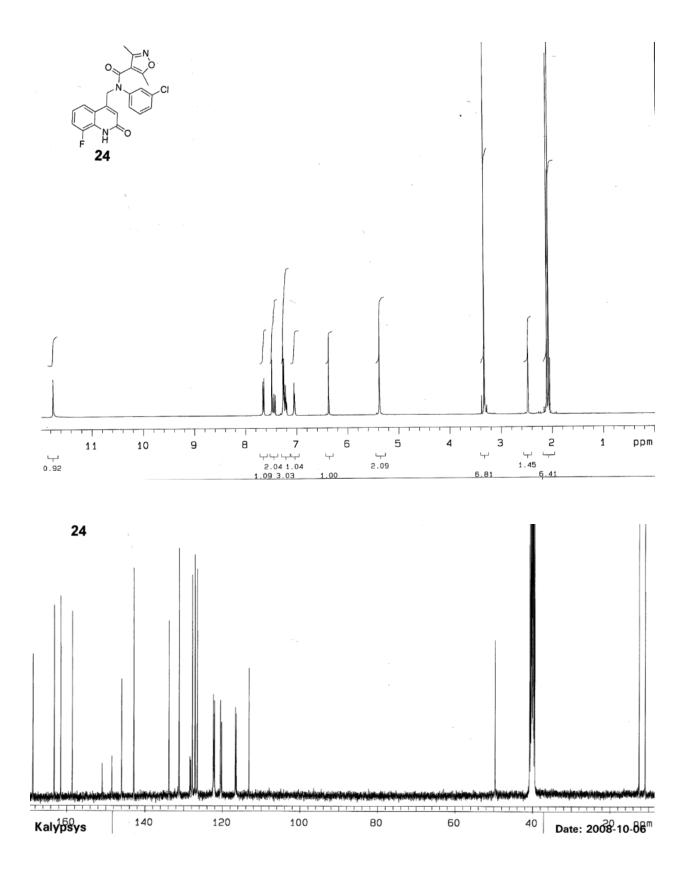


Compound 23: *N*-(3-Chlorophenyl)-*N*-((8-fluoro-2-oxo-1,2-dihydroquinolin-4-yl)methyl)-1-methyl-1*H*-imidazole-5-carboxamide

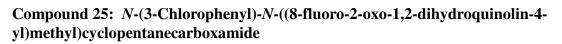


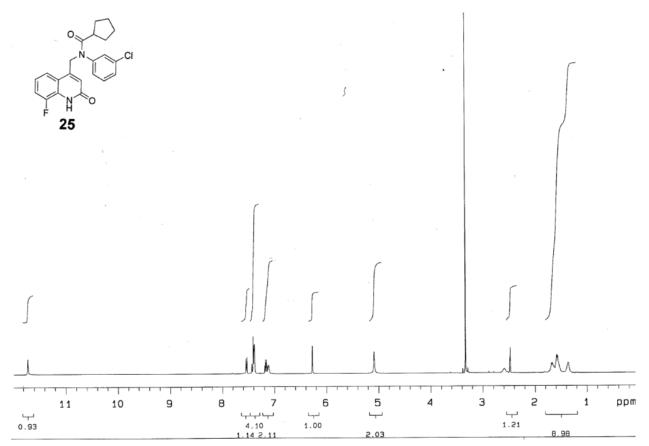
Compound 24: *N*-(3-Chlorophenyl)-*N*-((8-fluoro-2-oxo-1,2-dihydroquinolin-4-yl)methyl)-3,5-dimethylisoxazole-4-carboxamide

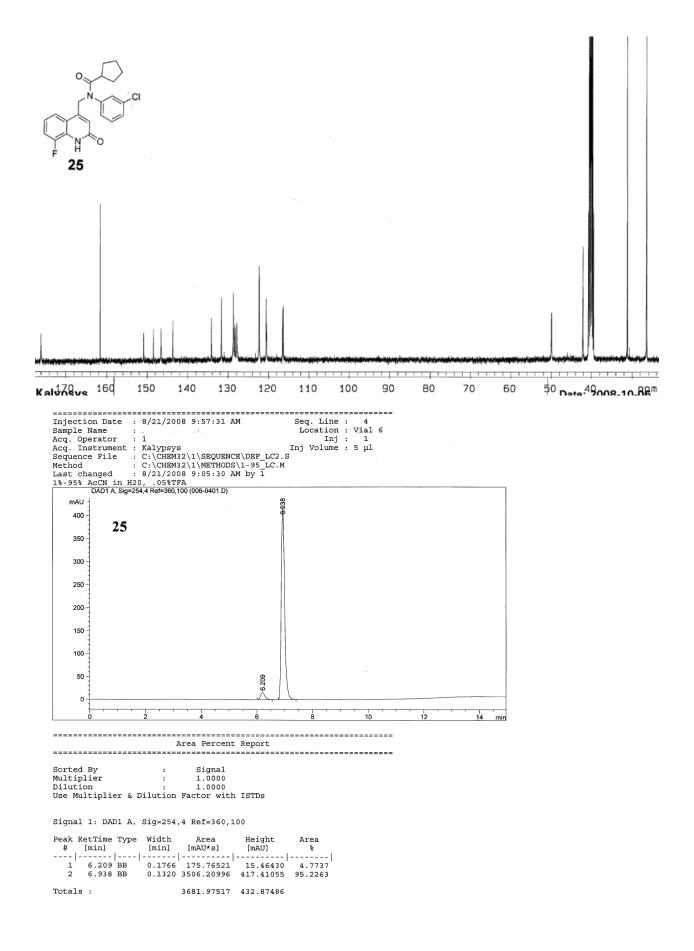
Page **42** of **82**



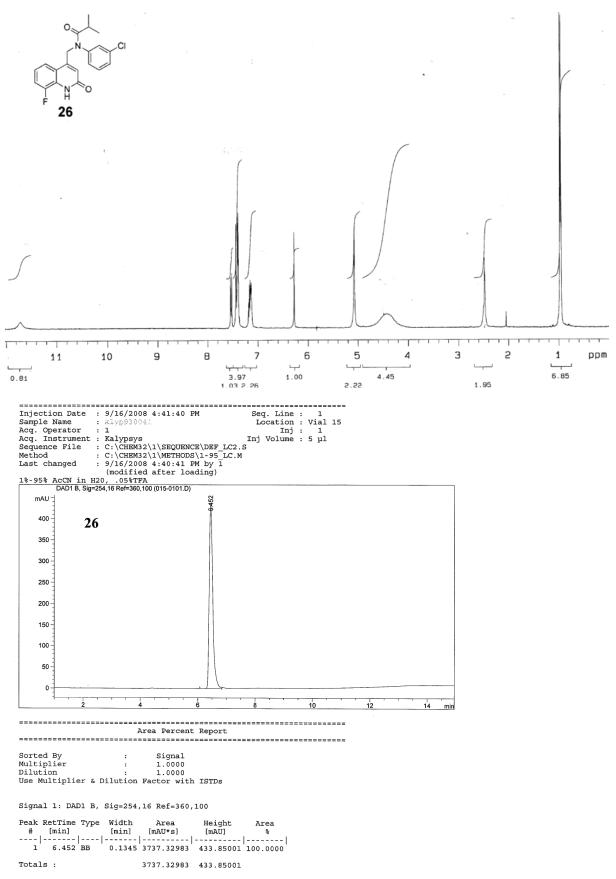
Injection Date : 8/21/2008 9:40:27 AM Seq. Line : 3 Sample Name : RUMINICATION Location : Vial 5 Acq. Instrument : Kalypsys Inj : 1 Acq. Instrument : Kalypsys Inj Volume : 5 µl Sequence File : C:\CHEM32\1\SEQUENCE\DEF_LC2.S Method : C:\CHEM32\1\METHODS\1-95_LC.M Last changed : 8/21/2008 9:05:30 AM by 1 l%-95% ACCN in H20, 05%TFA DAD4.Sig=244 AFeF360.000 (005-0301D)							
mAU 1 8							
400-24							
300 -							
200 -							
100 -							
0							
0 2 4 6 8 10 12 14 min							
Area Percent Report							
Sorted By : Signal Multiplier : 1.0000 Dilution : 1.0000 Use Multiplier & Dilution Factor with ISTDs							
Signal 1: DAD1 A, Sig=254,4 Ref=360,100							
Peak RetTime Type Width Area Height Area # [min] [mAU] % 1 6.300 BB 0.1308 3914.82495 462.19650 100.0000							
Totals : 3914.82495 462.19650							



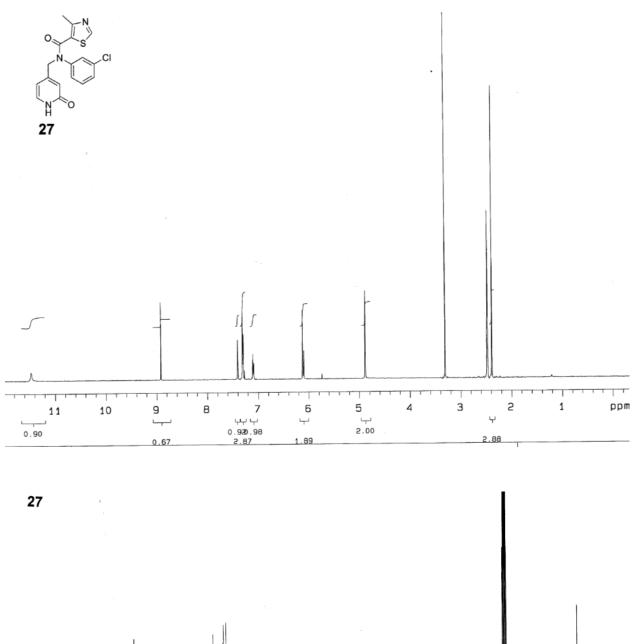




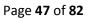
Compound 26: *N*-(3-Chlorophenyl)-*N*-((8-fluoro-2-oxo-1,2-dihydroquinolin-4-yl)methyl)isobutyramide

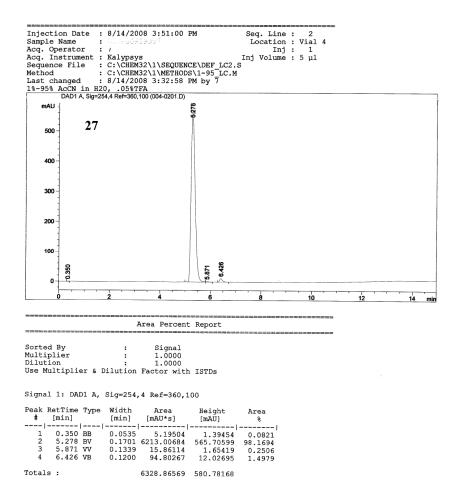


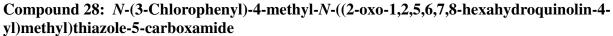
Compound 27: *N*-(3-Chlorophenyl)-4-methyl-*N*-((2-oxo-1,2-dihydropyridin-4-yl)methyl)thiazole-5-carboxamide

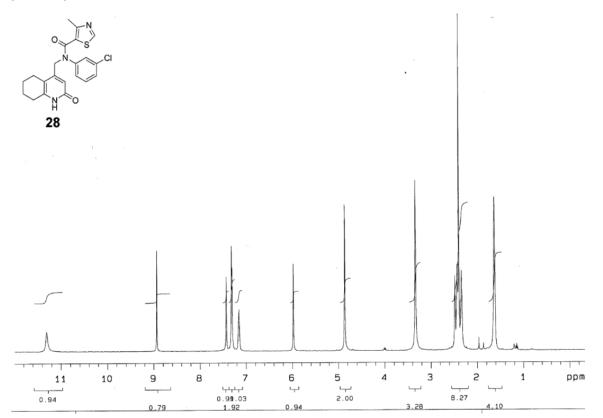


Date: 2008-08-27m

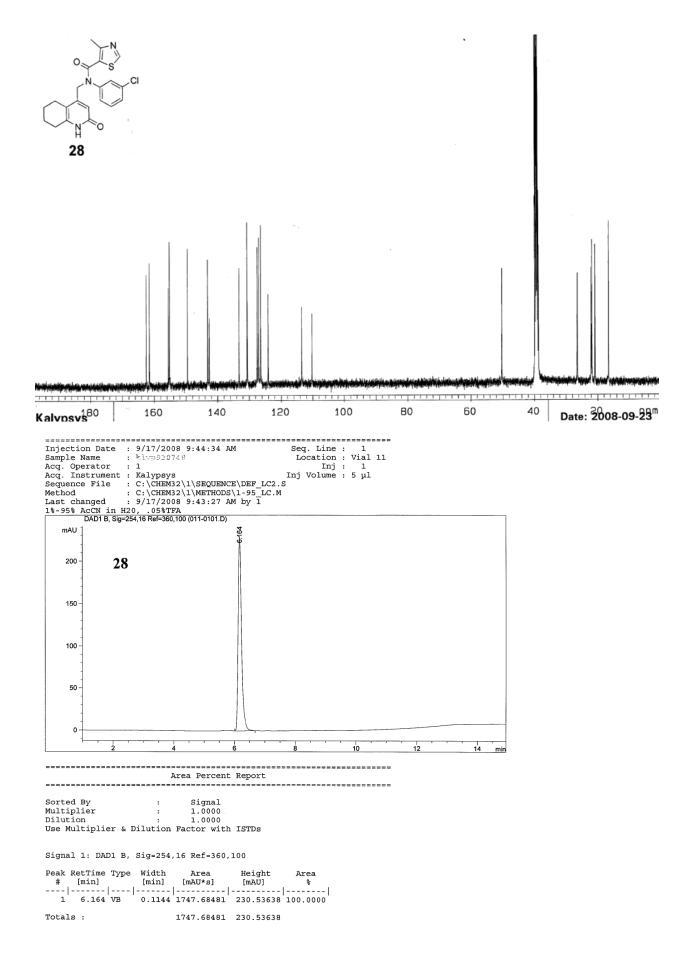




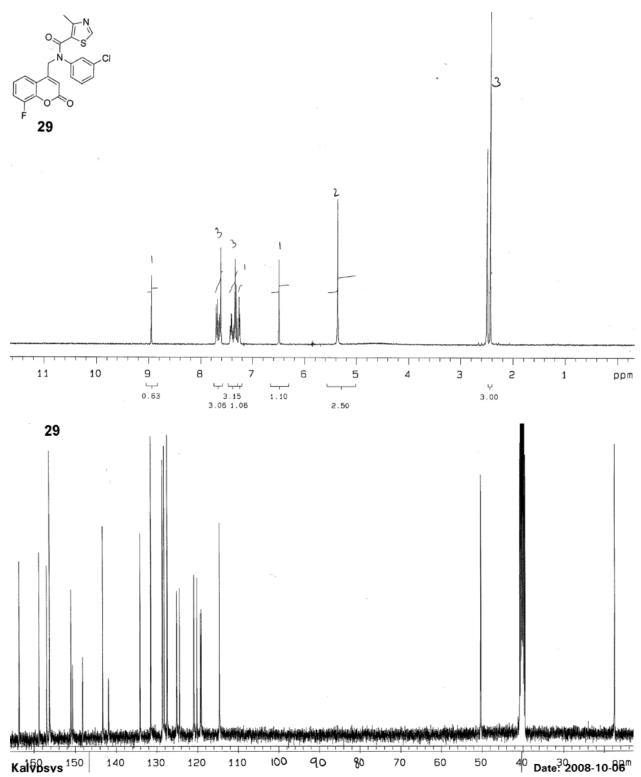


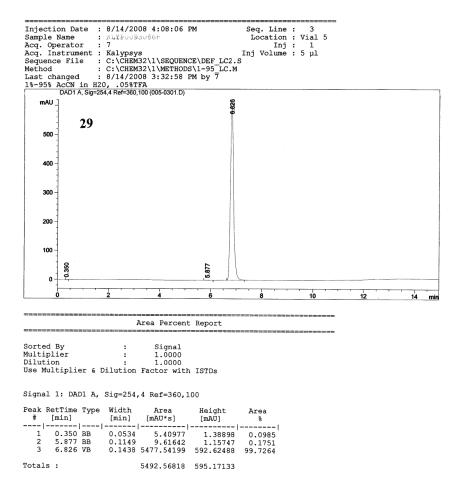


Page **48** of **82**

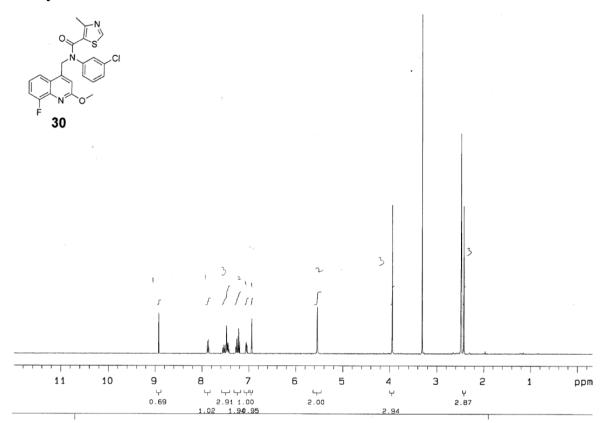


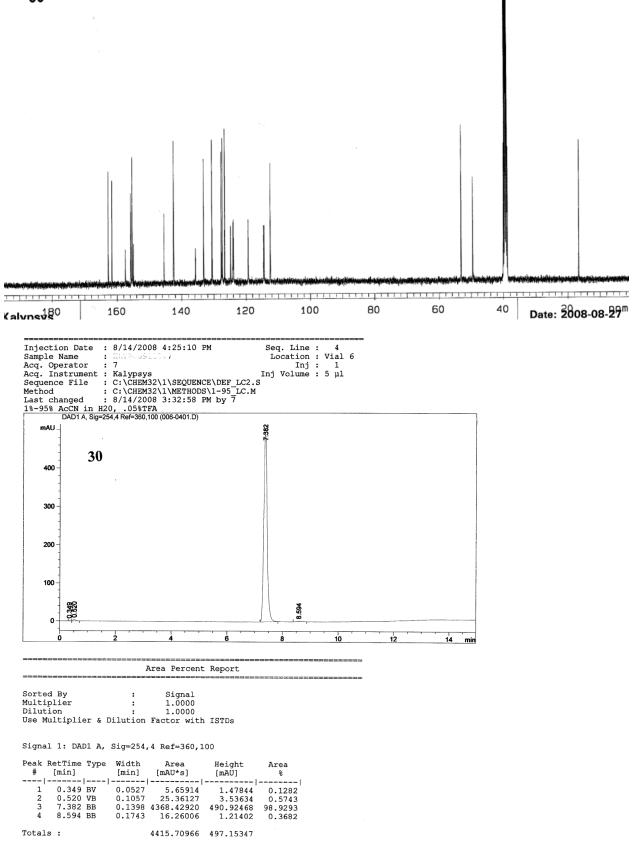
Compound 29: *N*-(3-Chlorophenyl)-*N*-((8-fluoro-2-oxo-2*H*-chromen-4-yl)methyl)-5-methylthiazole-4-carboxamide



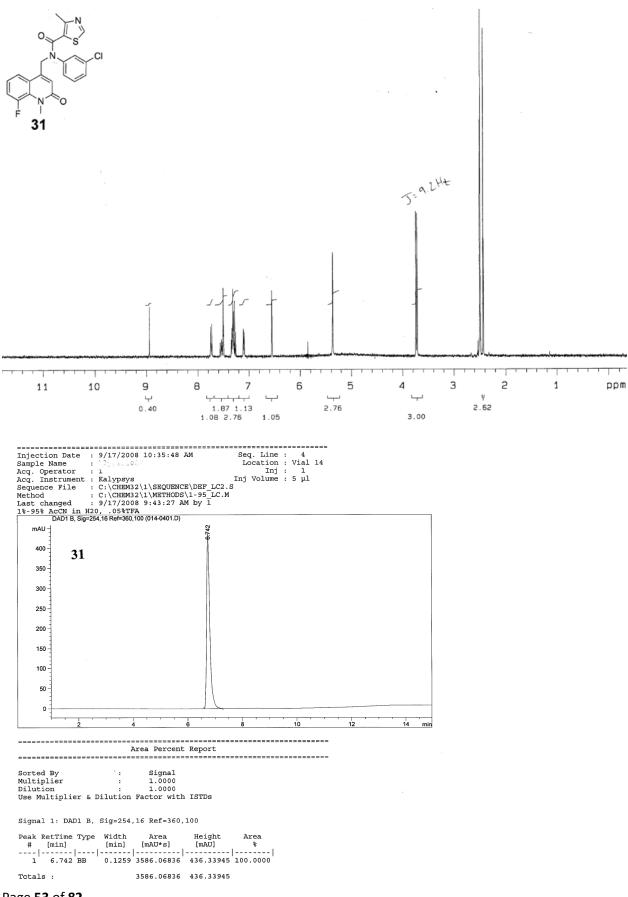


Compound 30: *N*-(3-Chlorophenyl)-*N*-((8-fluoro-2-methoxyquinolin-4-yl)methyl)-4-methylthiazole-5-carboxamide



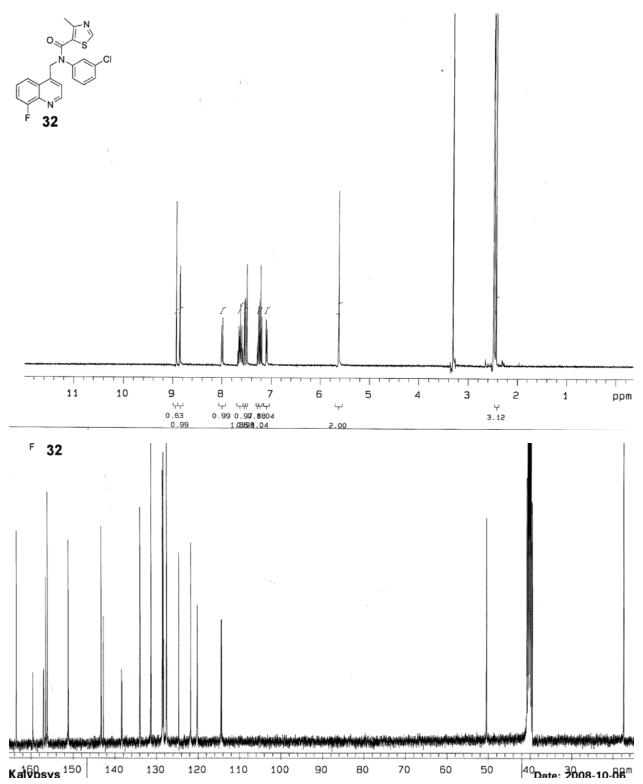


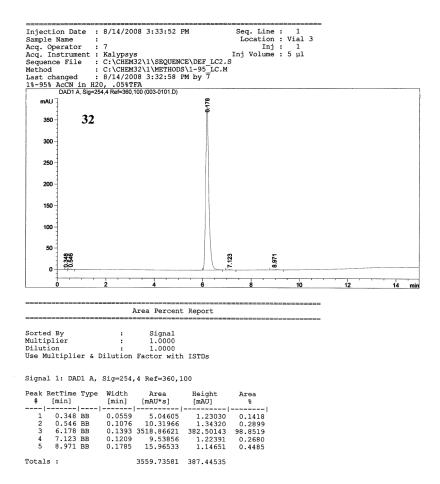
$Compound \ 31: \ N-(3-chlorophenyl)-N-((8-fluoro-1-methyl-2-oxo-1,2-dihydroquinolin-4-yl)methyl)-4-methylthiazole-5-carboxamide$

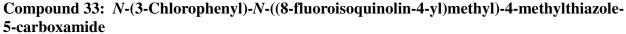


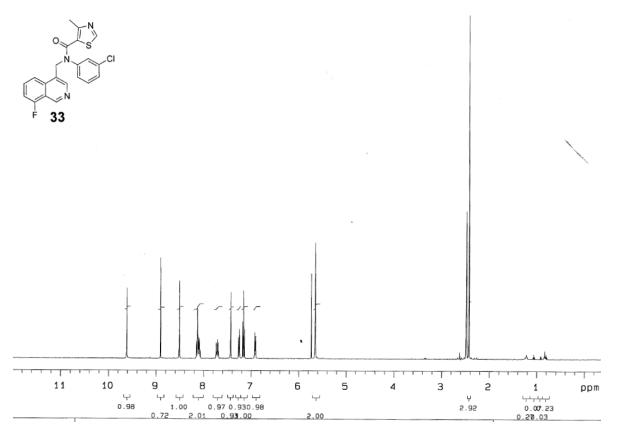
Page 53 of 82

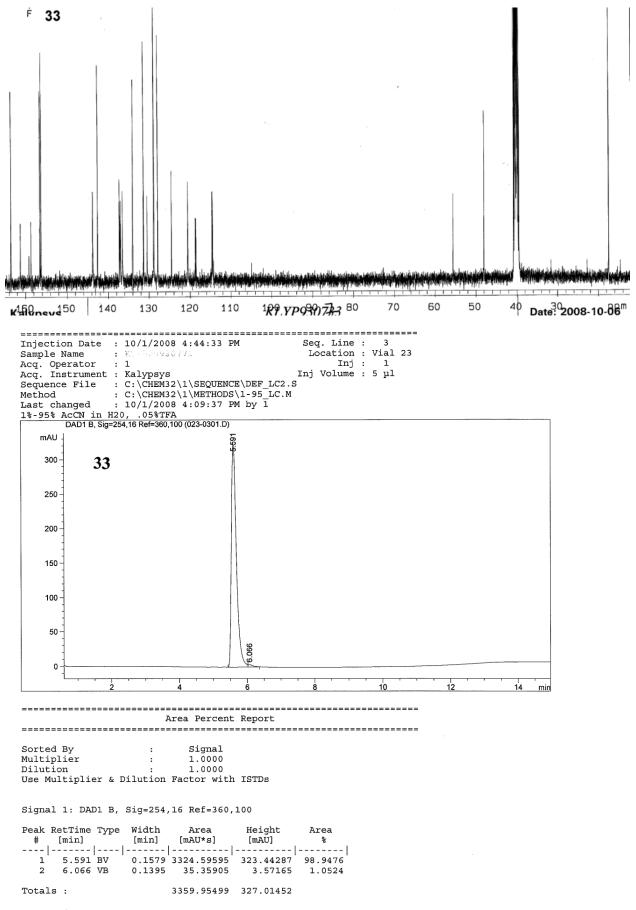
$\label{eq:compound 32: N-(3-Chlorophenyl)-N-((8-fluoroquinolin-4-yl)methyl)-4-methylthiazole-5-carboxamide$





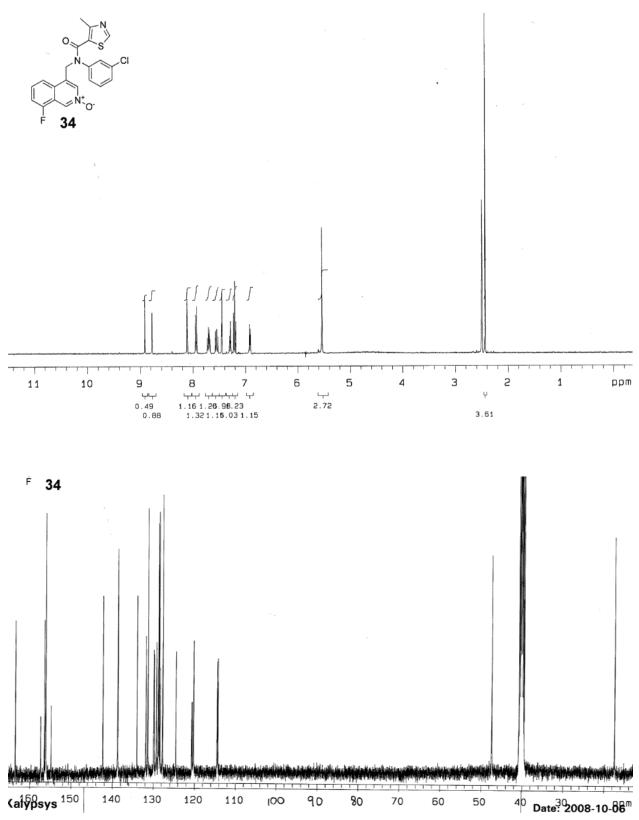


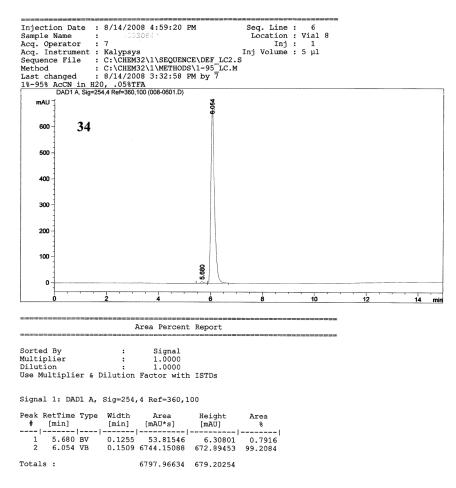


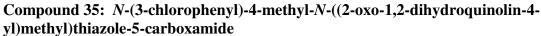


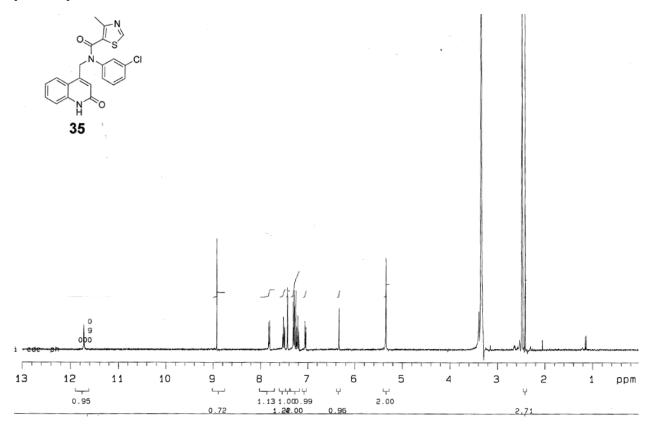
Page 56 of 82

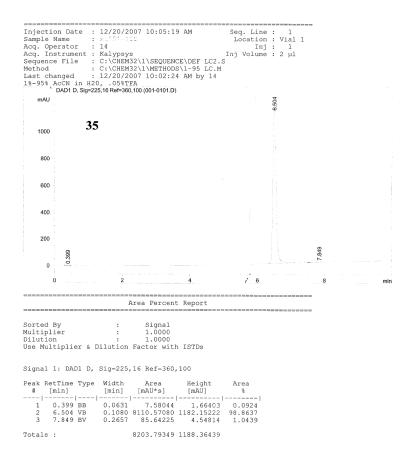
Compound 34: 4-((*N*-(3-Chlorophenyl)-4-methylthiazole-5-carboxamido)methyl)-8-fluoroisoquinoline 2-oxide

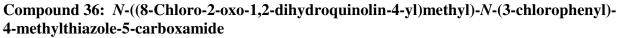


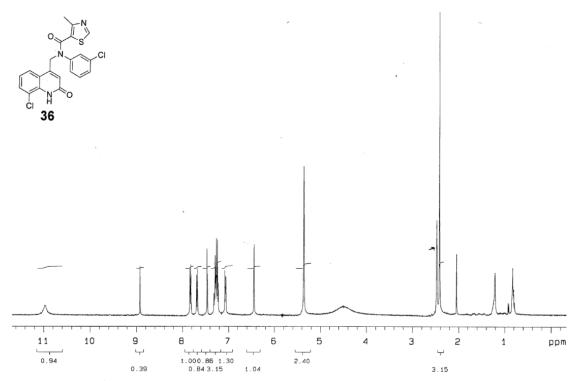


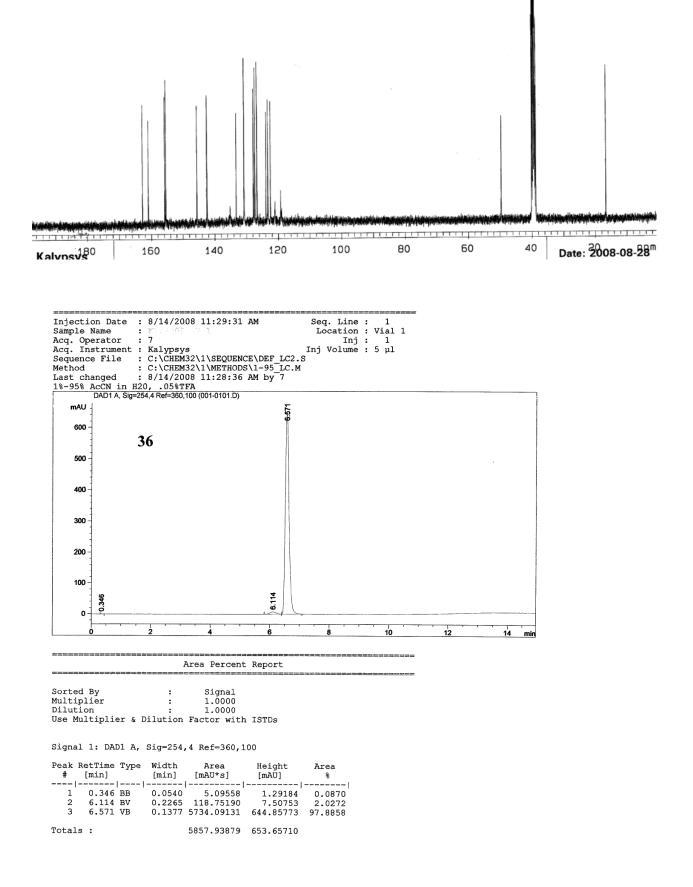




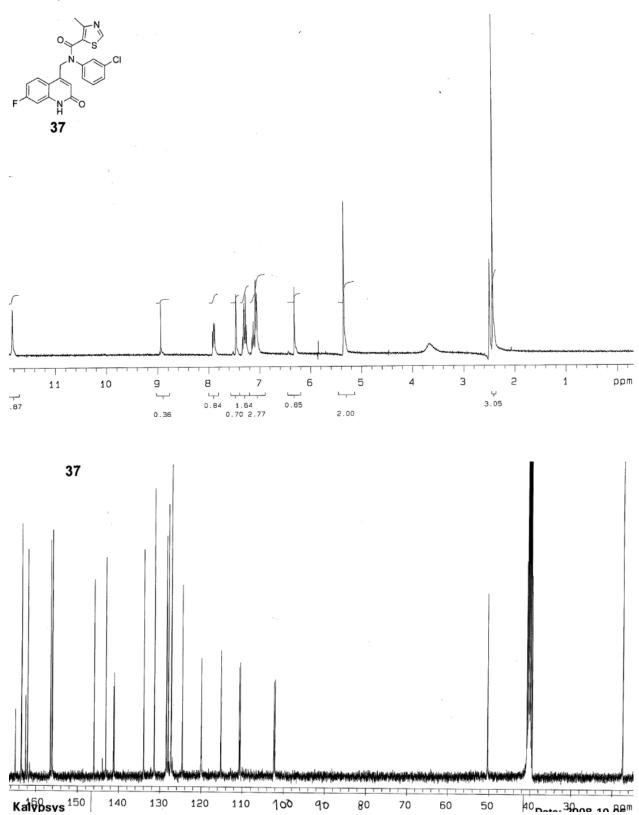






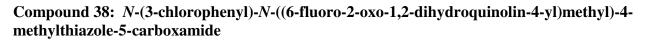


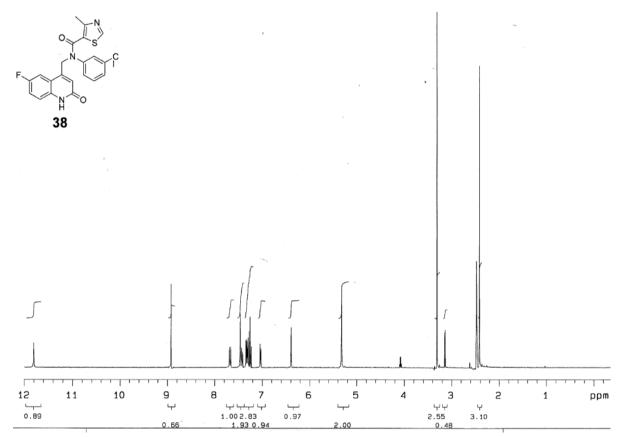
Compound 37: *N*-(3-Chlorophenyl)-*N*-((7-fluoro-2-oxo-1,2-dihydroquinolin-4-yl)methyl)-4-methylthiazole-5-carboxamide

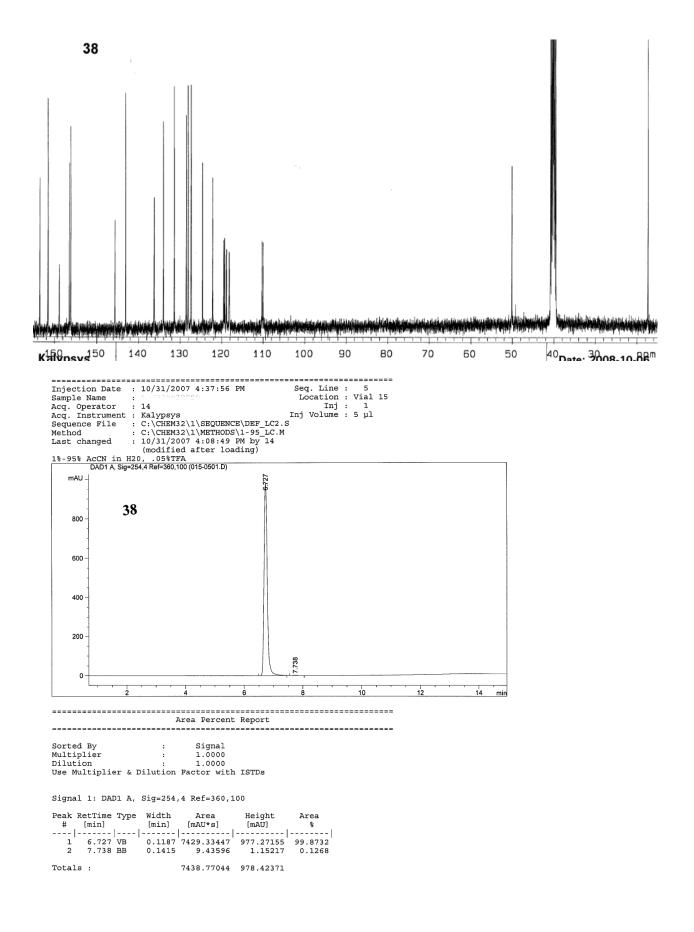


Sample 1 Acq. Ope Acq. In: Sequence Method Last cha	Name : erator : strument : e File : : anged : AcCN in H2	C:\CHEM32\1\SEQUE C:\CHEM32\1\METHO 11/1/2007 8:47:36 (modified after 1- 0, .05%TFA	NCE\DEF_LC2. DS\1-95_LC.M AM by 14 pading)		: Vial 2 : 1		
mAU	DAD1 A, Sig=28	54,4 Ref=360,100 (002-0201.D)	6.7 94				
800 -	37						
600 -							
400 -							
200 -							
0-		· · · · · · · · · · · · · · · · · · ·	Π.				
	2	4	6	8	10	12	14 min

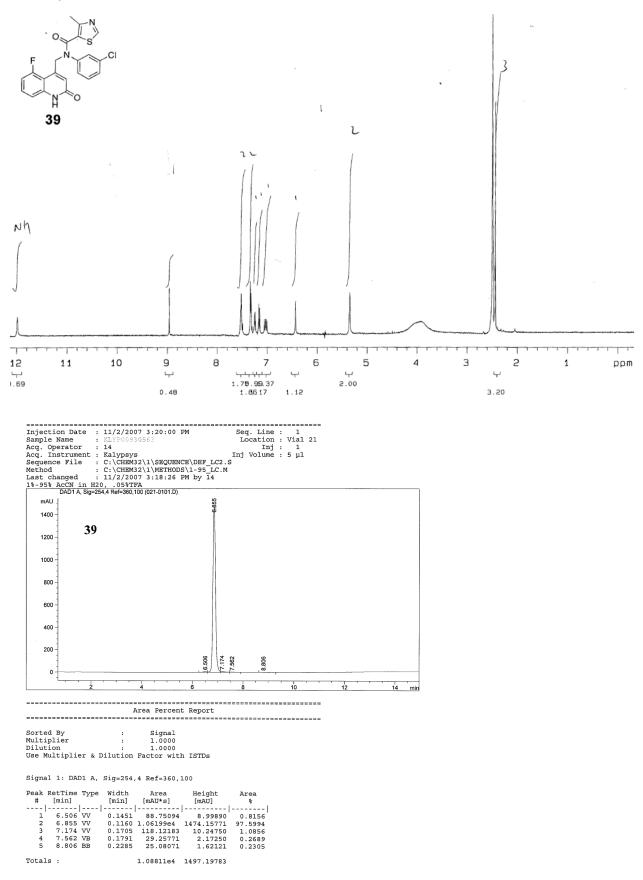
		Area Perce					
Sorted I Multipl: Dilution Use Mult	ier n	: Signal : 1.0000 : 1.0000 Dilution Factor wit	th ISTDs				
Signal :	1: DAD1 A,	Sig=254,4 Ref=360	,100				
# [1		Width Area [min] [mAU*s] 0.1070 6161.8481					



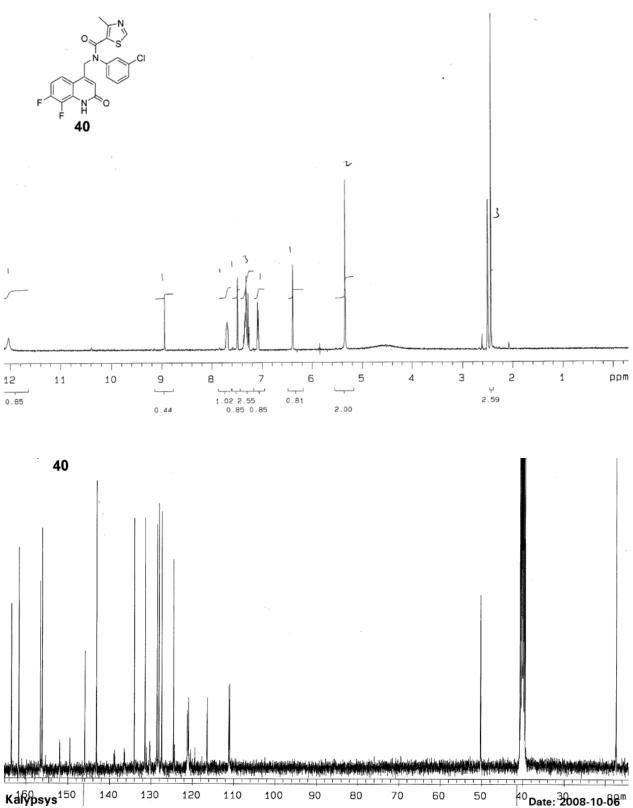


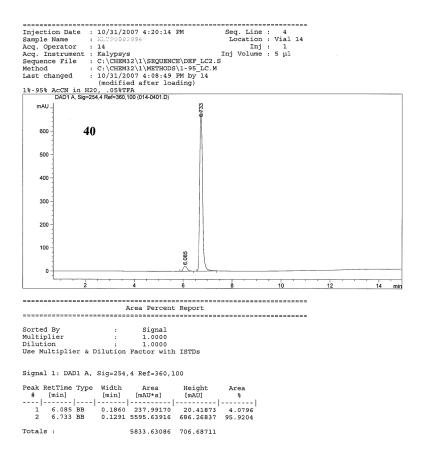


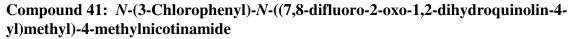
Compound 39: *N*-(3-Chlorophenyl)-*N*-((5-fluoro-2-oxo-1,2-dihydroquinolin-4-yl)methyl)-4-methylthiazole-5-carboxamide

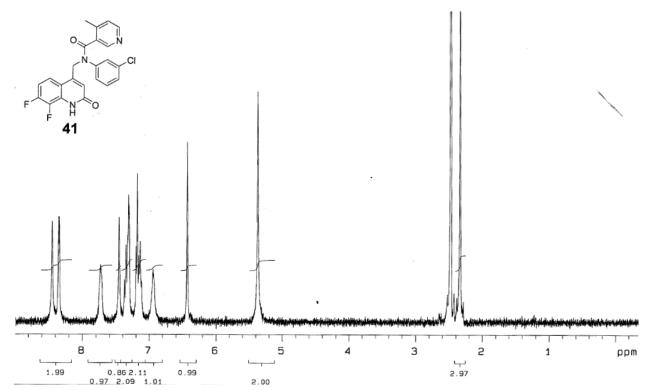


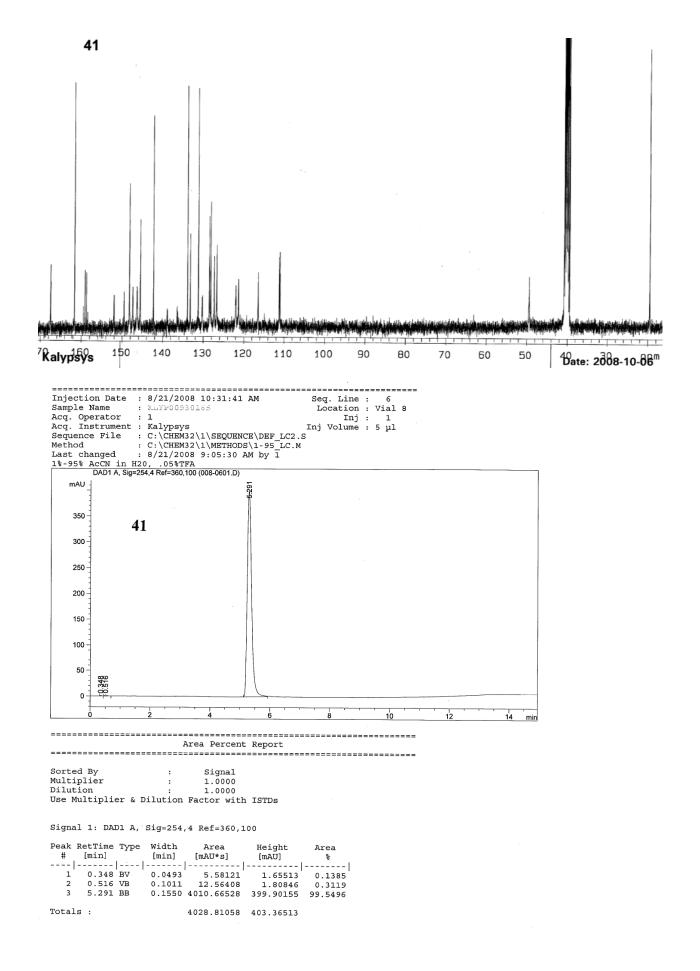
Compound 40: *N*-(3-Chlorophenyl)-*N*-((7,8-difluoro-2-oxo-1,2-dihydroquinolin-4-yl)methyl)-4-methylthiazole-5-carboxamide

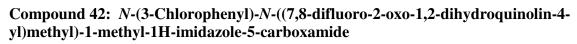


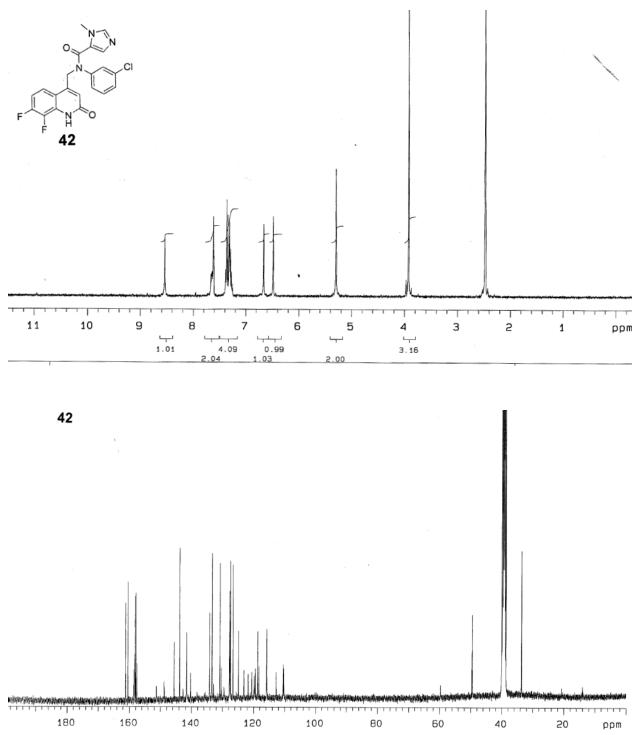


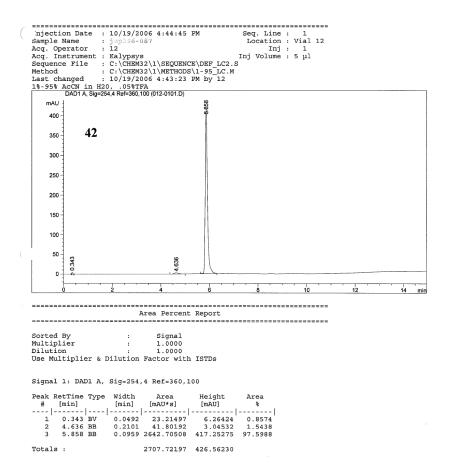


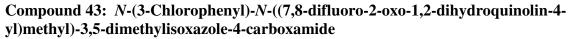


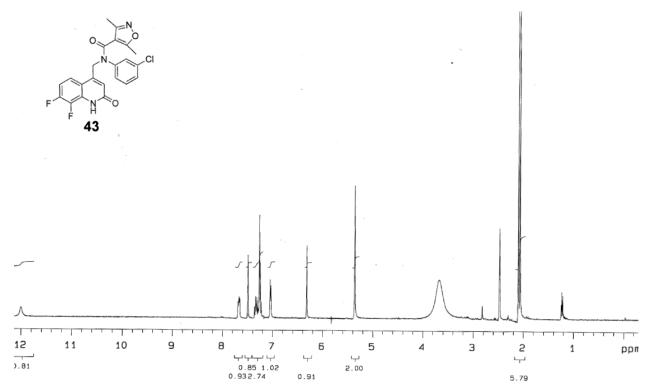


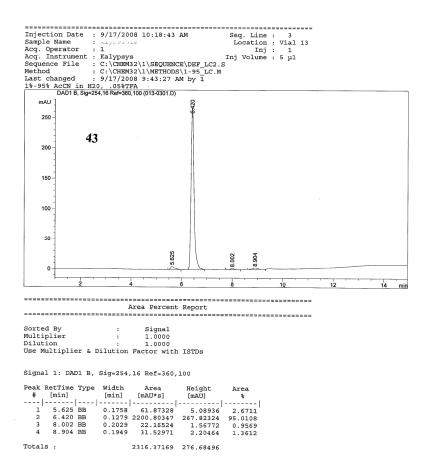


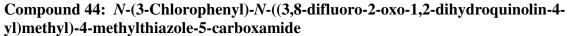


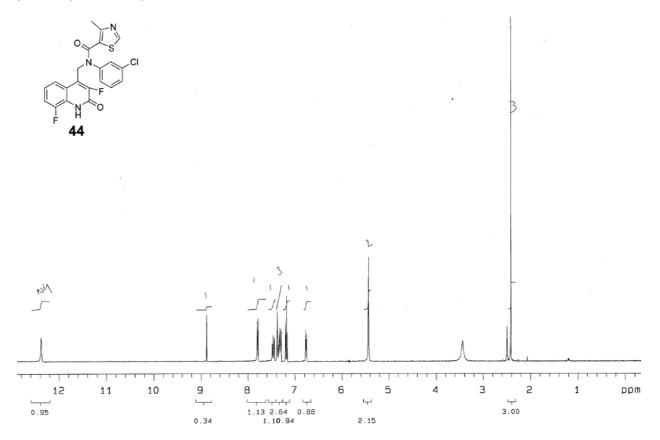


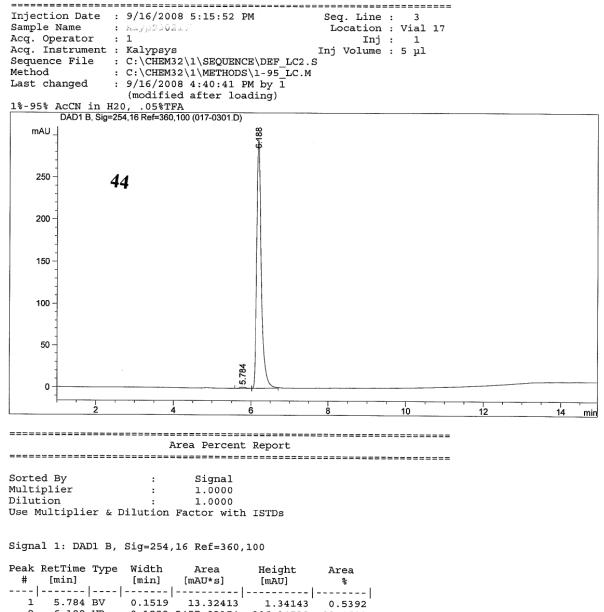








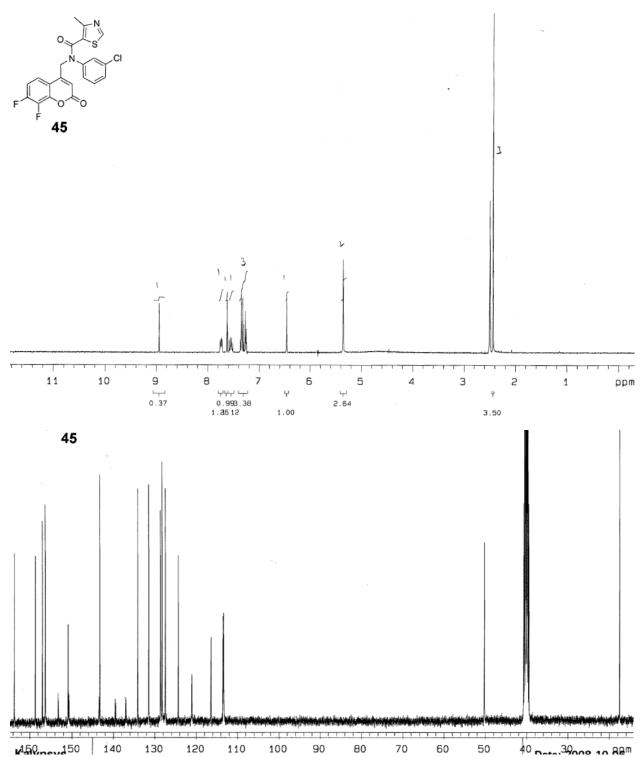




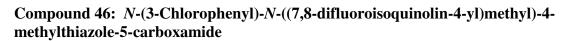
	_			0.2025		T' 24740	0.3352
•	2	6.188	VB	0.1228	2457.83154	296.34518	99.4608

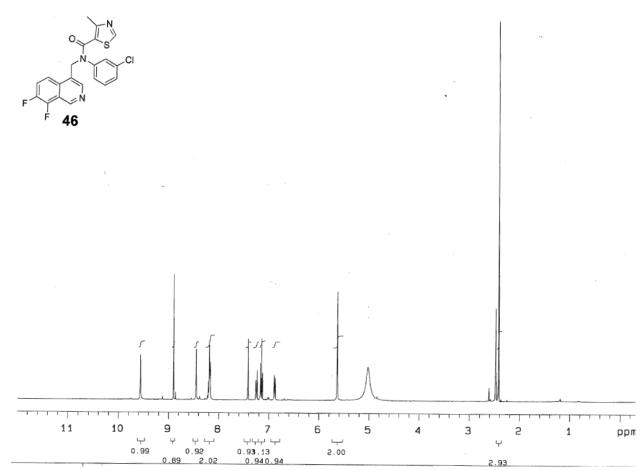
	Totals :	2471.15567 297.68662
--	----------	----------------------

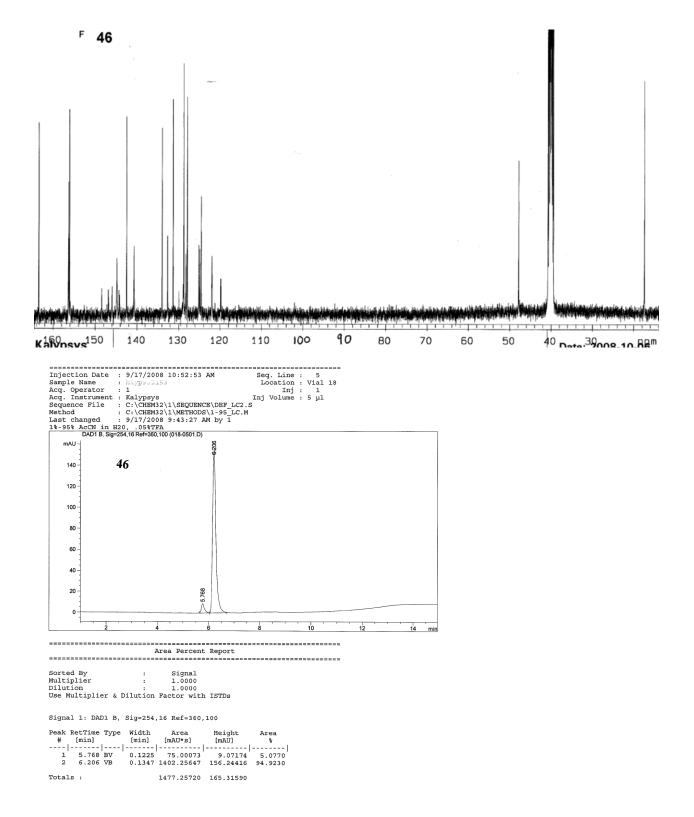
Compound 45: *N*-(3-Chlorophenyl)-*N*-((7,8-difluoro-2-oxo-2H-chromen-4-yl)methyl)-4-methylthiazole-5-carboxamide



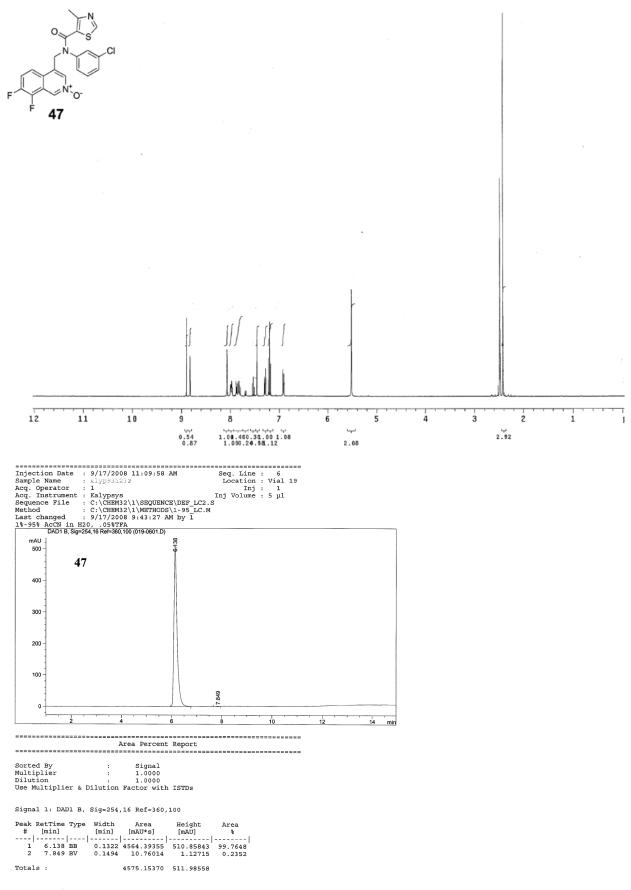
Injection Date : 8/25/2008 2:51:48 PM Seq. Line : 1 Sample Name : Kalrevoussker Location : Vial 13 Acq. Operator : 1 Inj : 1 Acq. Instrument : Kalypeys Sequence File : C:\CHEM32\\\SEQUENCE\DEF_LC.S Method : C:\CHEM32\\\METHODS\1-95_LC.M
Last changed : 8/25/2008 2:50:32 PM by 1
1%-95% AcCN in H20, .05%TFA DAD1 B, Sig=254,16 Ref=360,100 (013-0101.D)
mAU
175 - 45
150
125
100
76
50
25
<u>0 2 4 6 8 10 12 14 min</u>
Area Percent Report
Sorted By : Signal Multiplier : 1.0000 Dilution : 1.0000 Use Multiplier & Dilution Factor with ISTDs
Signal 1: DAD1 B, Sig=254,16 Ref=360,100
Peak RetTime Type Width Area Height Area # [min] [min] [mAU] % 1 1 7.016 BB 0.1384 1735.29309 197.66359 100.0000
Totals : 1735.29309 197.66359







Compound 47: 4-((*N*-(3-Chlorophenyl)-4-methylthiazole-5-carboxamido)methyl)-7,8-difluoroisoquinoline 2-oxide



Page 75 of 82

Compound #	MF	MW	ESI+ Exact m/z	ESI+ Measured m/z	ESI+ Mass Defect (ppm)	ESI- Exact m/z	ESI- Measured m/z	ESI- Mass Defect (ppm)
7	C21H16N2O3	344.1161	345.1239	345.1239	0.00	343.1083	343.1085	0.58
8	C21H15N2O3F	362.1067	363.1145	363.1141	-1.10	361.0989	361.0985	-1.11
9	C21H17N3O2S	375.1041	376.1119	376.1121	0.53	374.0963	374.0959	-1.07
10	C21H16N3O2FS	393.0947	394.1025	394.1026	0.25	392.0869	392.0870	0.26
11	C21H15N3O2FSCl	427.0558	428.0636	428.0641	1.17	426.0480		
12	C21H15N3O2FSCl	427.0558	428.0636	428.0641	1.17	426.0480	426.0486	1.41
13	C21H15N3O2FSCl	427.0558	428.0636			426.0480	426.0478	-0.47
14	C21H15N3O2F2S	411.0853	412.0931	412.0923	-1.94	410.0775	410.0771	-0.98
15	C22H15N4O2FS	418.09	419.0978	419.0971	-1.67	417.0822	417.0821	-0.24
16	C22H18N3O2FS	407.1104	408.1182	408.1176	-1.47	406.1026	406.1027	0.25
17	C22H18N3O3FS	423.1053	424.1131	424.1125	-1.41	422.0975	422.0974	-0.24
18	C20H13N3O2FSCl	413.0401	414.0479	414.0472	-1.69	412.0323	412.0323	0.00
19	C23H17N3O2FCl	421.0993	422.1071			420.0915	420.0919	0.95
20	C23H17N3O2FCl	421.0993	422.1071			420.0915	420.0915	0.00
21	C23H17N3O2FCl	421.0993	422.1071			420.0915	420.0919	0.95
22	C23H17N3O2FCl	421.0993	422.1071			420.0915	420.0919	0.95
23	C21H16N4O2FCl	410.0946	411.1024	411.1019	-1.22	409.0868	409.0863	-1.22
24	C22H17N3O3FC1	425.0942	426.1020	426.1028	1.88	424.0864	424.0865	0.24
25	C22H20N2O2FCI	398.1197	399.1275	399.1274	-0.25	397.1119	397.1122	0.76
26	C20H18N2O2FCl	372.1041	373.1119	373.1118	-0.27	371.0963	371.0957	-1.62

27	C17H14N3O2SCI	359.0495	360.0573	360.0576	0.83	358.0417	358.0417	0.00
28	C21H20N3O2SC1	413.0965	414.1043	414.1046	0.72	412.0887	412.0889	0.49
29	C21H14N2O3FSCl	428.0398	429.0476	429.0470	-1.40	427.0320	427.0321	0.23
30	C22H17N3O2FSCI	441.0714	442.0792	442.0785	-1.58	440.0636	440.0640	0.91
31	C22H17N3O2FSCl	441.0714	442.0792	442.0789	-0.68	440.0636		
32	C21H15N3OFSCl	411.0608	412.0686	412.0681	-1.21	410.0530	410.0533	0.73
33	C21H15N3OFSCI	411.0608	412.0686	412.0682	-0.97	410.0530		
34	C21H15N3O2FSCI	427.0558	428.0636	428.0644	1.87	426.0480	426.0486	1.41
35	C21H16N3O2SCl	409.0652	410.0730	410.0733	0.73	408.0574	408.0568	-1.47
36	C21H15N3O2SCl2	443.0262	444.0340			442.0184	442.0186	0.45
37	C21H15N3O2FSCl	427.0558	428.0636	428.0641	1.17	426.0480	426.0482	0.47
38	C21H15N3O2FSCI	427.0558	428.0636	428.0628	-1.87	426.0480	426.0472	-1.88
39	C21H15N3O2FSCl	427.0558	428.0636			426.0480	426.0487	1.64
40	C21H14N3O2F2SCl	445.0463	446.0541	446.0537	-0.90	444.0385	444.0381	-0.90
41	C23H16N3O2F2Cl	439.0899	440.0977	440.0974	-0.68	438.0821	438.0820	-0.23
42	C21H15N4O2F2Cl	428.0852	429.0930	429.0922	-1.86	427.0774	427.0773	-0.23
43	C22H16N3O3F2Cl	443.0848	444.0926			442.0770	442.0767	-0.68
44	C21H14N3O2F2SCl	445.0463	446.0541			444.0385	444.0380	-1.13
45	C21H13N2O3F2SCl	446.0303	447.0381	447.0375	-1.34	445.0225	445.0220	-1.12
46	C21H14N3OF2SCl	429.0514	430.0592	430.0585	-1.63	428.0436		
47	C21H14N3O2F2SCl	445.0463	446.0541	446.0542	0.22	444.0385	444.0382	-0.68

Identification code	Cpd 12
Empirical formula	C21 H15 CI F N3 O2 S
Formula weight	427.87
Temperature	100(2) K
Wavelength	0.71073 Å
Crystal system	Triclinic
Space group	P-1
Unit cell dimensions	$a = 8.8921(13) \text{ Å}$ $\alpha = 72.680(2)^{\circ}$
	$b = 17.982(3) \text{ Å}$ $\beta = 82.543(2)^{\circ}$
	$c = 18.935(3) \text{ Å}$ $\gamma = 77.465(2)^{\circ}$
Volume	2814.5(7) Å ³
Z	6
Density (calculated)	1.515 g/cm ³
Absorption coefficient	0.349 mm ⁻¹
F(000)	1320
Crystal size	0.22 x 0.08 x 0.04 mm ³
Theta range for data collection	1.87 to 25.00°
Index ranges	-10<=h<=9, -21<=k<=21, -22<=l<=22
Reflections collected	16620
Independent reflections	9455 [R(int) = 0.0512]
Completeness to theta = 25.00°	98.2 %
Absorption correction	None
Max. and min. transmission	0.9862 and 0.9272
Refinement method	Full-matrix least-squares on F ²
Data / restraints / parameters	9455 / 0 / 794
Goodness-of-fit on F ²	1.011
Final R indices [I>2sigma(I)]	R1 = 0.0584, $wR2 = 0.1363$
R indices (all data)	R1 = 0.1099, wR2 = 0.1593
Largest diff. peak and hole	0.837 and -0.406 e Å ⁻³

	Х	У	Z	U(eq)
Cl(1")	15733(1)	1875(1)	-4548(1)	40(1)
Cl(1')	7275(2)	129(1)	4646(1)	34(1)
Cl(2')	7634(6)	2718(3)	2227(3)	38(2)
Cl(1)	-591(1)	6491(1)	2212(1)	40(1)
F(1")	9436(3)	-1324(1)	724(1)	32(1)
F(1')	3760(3)	2140(1)	-858(1)	34(1)
F(1)	7590(3)	4120(1)	-774(1)	34(1)
S(1')	3114(1)	2472(1)	4075(1)	30(1)
S(1")	9455(2)	1661(1)	-5145(1)	39(1)
S (1)	3886(1)	5846(1)	4048(1)	33(1)
O(1')	6577(3)	-148(2)	825(2)	31(1)
O(1")	6706(3)	967(2)	-1006(1)	29(1)
O (1)	9523(3)	5310(2)	791(2)	29(1)
D(2')	424(4)	2050(2)	2883(2)	36(1)
D(2")	9226(4)	459(2)	-3675(2)	45(1)
D(2)	6936(4)	4254(2)	3388(2)	35(1)
N(1")	8461(4)	-126(2)	-492(2)	24(1)
N(1')	4837(4)	966(2)	345(2)	26(1)
N(1)	8103(4)	4660(2)	349(2)	25(1)
N(2')	1854(4)	3907(2)	3441(2)	30(1)
N(2")	9220(4)	3036(2)	-4959(2)	39(1)
N(2)	6244(5)	5937(2)	4643(2)	34(1)
N(3')	2974(4)	1470(2)	2834(2)	26(1)
N(3")	10935(4)	833(2)	-3131(2)	24(1)
N(3)	4964(4)	5036(2)	2717(2)	25(1)
C(1")	10276(5)	-1303(2)	66(2)	25(1)
C(1')	2995(5)	2140(3)	-188(2)	29(1)
C(1)	6562(5)	4034(2)	-174(2)	26(1)
C(2")	11541(5)	-1881(2)	35(2)	28(1)
C(2')	1719(5)	2719(3)	-144(2)	29(1)
C(2)	5325(5)	3698(2)	-159(2)	26(1)
C(3")	12379(5)	-1847(3)	-650(2)	29(1)
2(3')	961(5)	2698(3)	559(2)	33(1)

Table 2. Atomic coordinates (x 10⁴) and equivalent isotropic displacement parameters ($Å^2x$ 10³)for Cpd 12. U(eq) is defined as one third of the trace of the orthogonalized U^{ij} tensor.

C(3)	4282(5)	3622(2)	474(2)	30(1)
C(4')	1498(5)	2123(3)	1178(2)	30(1)
C(4")	11930(5)	-1245(2)	-1266(2)	26(1)
C(4)	4521(5)	3888(2)	1049(2)	28(1)
C(5")	10625(5)	-650(2)	-1226(2)	22(1)
C(5')	2810(5)	1534(2)	1129(2)	27(1)
C(5)	5795(5)	4238(2)	1031(2)	22(1)
C(6")	10120(5)	16(2)	-1855(2)	23(1)
C(6')	3437(5)	904(2)	1762(2)	25(1)
C(6)	6080(5)	4559(2)	1608(2)	21(1)
C(7")	8844(5)	548(2)	-1781(2)	25(1)
C(7')	4654(5)	350(2)	1648(2)	27(1)
C(7)	7293(5)	4926(2)	1517(2)	26(1)
C(8")	7916(5)	491(2)	-1087(2)	24(1)
C(8')	5446(5)	358(3)	924(2)	26(1)
C(8)	8385(5)	4987(2)	873(2)	25(1)
C(9')	3564(5)	1548(2)	425(2)	26(1)
C(9")	9776(5)	-686(2)	-549(2)	22(1)
C(9)	6840(5)	4314(2)	401(2)	22(1)
C(10')	2676(5)	846(3)	2537(2)	30(1)
C(10")	11106(5)	52(2)	-2575(2)	28(1)
C (10)	5035(5)	4446(2)	2316(2)	26(1)
C(11')	4521(5)	1425(3)	3012(2)	26(1)
C(11")	12176(5)	1247(2)	-3168(2)	27(1)
C(11)	3925(5)	5780(2)	2462(2)	25(1)
C(12")	13188(5)	1383(2)	-3789(2)	27(1)
C(12')	5101(5)	839(3)	3637(2)	30(1)
C(12)	2356(5)	5774(3)	2486(2)	25(1)
C(13")	14414(5)	1737(3)	-3775(2)	28(1)
C(13')	6532(5)	845(2)	3848(2)	29(1)
C(13)	1365(5)	6485(3)	2209(2)	28(1)
C(14')	7372(5)	1418(3)	3448(2)	34(1)
C(14")	14675(5)	1947(3)	-3162(2)	34(1)
C(14)	1908(6)	7192(3)	1938(2)	33(1)
C(15")	13661(6)	1790(3)	-2539(2)	36(1)
C(15')	6806(5)	1988(3)	2819(2)	32(1)
C(15)	3452(5)	7193(3)	1922(2)	32(1)
C(16")	12420(5)	1442(3)	-2535(2)	31(1)

C(16')	5371(5)	1990(3)	2599(2)	30(1)
C(16)	4486(5)	6476(3)	2175(2)	29(1)
C(17")	9987(5)	962(3)	-3686(2)	31(1)
C(17')	1766(5)	2028(3)	3004(2)	29(1)
C(17)	5924(5)	4855(3)	3277(2)	27(1)
C(18")	9828(5)	1717(3)	-4287(2)	30(1)
C(18')	2118(5)	2643(2)	3306(2)	24(1)
C(18)	5692(5)	5387(2)	3770(2)	27(1)
C(19')	2660(5)	3473(3)	3998(2)	30(1)
C(19")	9106(5)	2669(3)	-5440(2)	39(1)
C(19)	4747(6)	6133(3)	4647(2)	37(1)
C(20')	1530(5)	3436(3)	3043(2)	27(1)
C(20")	9648(5)	2491(3)	-4288(2)	35(1)
C(20)	6805(5)	5482(2)	4153(2)	31(1)
C(21')	557(5)	3831(3)	2401(2)	36(1)
C(21")	9882(6)	2804(3)	-3679(2)	42(1)
C(21)	8487(5)	5150(3)	4110(3)	40(1)

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