Lithiathion of N-Alkyl-(o-tolyl)aziridine: Stereoselective Synthesis of Isochromans.

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Experimental

General

All reactions involving air-sensitive reagents were performed in oven-dried glassware under an atmosphere of nitrogen using syringe-septum cap technique. Tetrahydrofuran (THF) was freshly distilled under a nitrogen atmosphere over sodium / benzophenone. Hexane was freshly distilled under a nitrogen atmosphere over finely powdered CaH₂.

Column chromatography was performed using the solvent systems indicated. Petroleum ether refers to the fraction boiling at 30-40 °C. The stationary phase used was silica gel 60 unless otherwise indicated.

¹H NMR and ¹³C NMR spectra were recorded at 400, 500, 600 MHz and 75, 100, 125, 150 MHz respectively with CDCl₃ as solvent. Data are expressed as chemical shifts in parts per million (ppm) relative to residual chloroform CDCl₃ (¹H δ 7.27), (¹³C δ 77.0). The multiplicity of each signal is designated by the following abbreviations: s, singlet; d, doublet; dd, doublet of doublets; dt, doublet of triplets; ddt, doublet of doublet of triplets; t, triplet; br, broad; m multiplet. Coupling constants *J* are given in Hz. Infra-red spectra of the compounds were recorded as a film or KBr disc as indicated. Low resolution mass spectra were obtained by GC-MS analysis using a gas chromatography with a BPX₅ column-HP 6890 plus (30 m, 0.25 mm i.d.) equipped with a 5973 mass selective detector operating at 70 eV (flow rate (He) = 1 mL/min). Specific rotations [*a*]_D were measured using a polarimeter with a cell of path length 1.0 dm, at 20 °C. Concentrations (c) are given in g/100 mL. Melting points were uncorrected. TLC was performed on Merck silica gel plates with F-254 indicator; detection was accomplished by UV light (254 nm), by exposing to I₂ vapours and spraying a solution of (5% W/V) ammonium molibdate and 0.2% W/V cerium(III)sulphate in 100 ml 17.6% aq. sulphuric acid and heating to 200 °C for some time until blue spots appear.

N,N,N',N'-tetramethylethylenediamine (TMEDA) was distilled over finely powdered CaH₂. Commercial solutions of *n*-BuLi (2.5 M hexane solution) and *sec*-BuLi (1.3 M cyclohexane solution) were titrated by using *N*-pivaloyl-*o*-toluidine prior use.¹

All other chemicals were of commercial grade and used without further purification.

¹ Suffert, J. J. Org. Chem. 1989, 54, 509-510.

Procedure for the preparation of aziridine (±)-1.



Aziridine (\pm)-1 was prepared using dimethyl-[2-bromo-1-(o-tolyl)ethyl]sulphonium bromide as intermediate, which was easily generated by reaction of bromine, dimethyl sulphide (*ca.* 1:5 molar ratio in CH₂Cl₂) and *o*-tolylstyrene.² A commercially available aqueous solution (40% p/v) of MeNH₂ (20 mmol) was added dropwise at 0 °C to the sulphonium bromide (1 mmol) and the resulting mixture was stirred overnight. The mixture was poured into 20 mL of saturated brine, extracted with diethyl ether (3 × 10 mL), dried (Na₂SO₄), and the solvent evaporated under reduced pressure. Kugelrohr distillation furnished the desired aziridine with 95% yield. Spectroscopic data for (\pm)-1 have been reported.³

 $Br \overset{\oplus}{S(CH_3)_2} [2-Bromo-1-(o-tolyl)ethyl]dimethylsulphonium bromide: white solid, mp 119–120 °C, 88%. ¹H-NMR (CDCl₃, 500 MHz) <math>\delta$ 4.87 (s, 3 H), 5.12 (s, 3 H), 5.45 (s, 3 H), 6.55 (t, *J* = 11.0 Hz, 1 H), 6.65 (dd, *J* = 5.5, 11.0 Hz, 1 H), 7.08 (q, *J* = 5.5 Hz, 1 H), 9.82–9.96 (m, 4 H). C¹³-NMR (CDCl₃, 500 MHz) δ 21.5, 25.5, 26.4, 31.4, 129.9, 131.0, 133.2, 134.2, 141.4. ESI-MS *m*/*z* (%): 197 (43), 118 (64). FT-IR (KBr, cm⁻¹): 3423, 2981, 2932, 2915, 1493, 1440, 1207, 1004, 737. Anal calcd for C₁₁H₁₆Br₂S: C, 38.84; H, 4.74%. Found: C, 37.79; H, 4.59%.

(S)-1 ($[\alpha]_D^{20} = +144$ (c = 1.0, CHCl₃), er > 98:2, 63% yield) was prepared by lithiation/MeItrapping sequence from S-(+)-1-methyl-2-phenylaziridine ($[\alpha]_D^{20} = +177$ (c = 0.5, CHCl₃), er > 98:2) according a reported procedure.³ S-(+)-1-methyl-2-phenylaziridine (40% yield) was synthetized starting from commercially available (*R*)-styrene oxide, by ring-opening reaction with MeNH₂ and a subsequent Mitsunobu ring-closure of the correspondent aminoalcohols.^{4,5,6} The

² Chow, Y. L.; Bakker, B. H.; Iwai, K. J. Chem. Soc. Chem. Comm. 1980, 521–522.

³ Capriati, V.; Florio, S.; Luisi, R.; Musio, B. Org. Lett. 2005, 7, 17, 3749-3752.

⁴ a) Sawamura, M; Hamashima, H; Ito, Y. *J. Org. Chem.* **1990**, 55, 5935.16. b) Huszthy, P; Oue, M; Bradshaw, J. S.; Zhu, C. Y.; Wang, T.; Dalley, N. K.; Curtis, J. C.; Izatt, R. M. *J. Org. Chem.* **1992**, 57, 5383-5394.

⁵ Anderson, W. K.; Milowsky, A. S. J. Med. Chem. 1986, 29, 2241 – 2249.

⁶ Fujita, S.; Imamura, K.; Nozaki, H. Bull. Chem. Soc. Jpn. 1971, 44, 1975-1977.

enantiomeric purity of the aziridine derivatives was ascertained by ¹H-NMR in presence of Mosher's acid (See page S28-S29).

Procedure for the lithiation/trapping sequence of aziridine 1:



To a stirred solution of aziridine (\pm)-1 (100 mg, 0.68 mmol) and TMEDA (204.0 µL, 1.36 mmol) in THF (4 mL) at -78 °C a solution of *sec*-BuLi (1.4 M in hexane, 972 µL, 1.36 mmol) was added dropwise . After 30 minutes at -78 °C the electrophile (1.36 mmol) was added neat if liquid and in 2.0 ml of solvent if solid. After 2 hours at -78 °C, the mixture was allowed to warm slowly to room temperature and the reaction mixture was poured in saturated aqueous NH₄Cl (10 mL) and extracted with Et₂O (3 x 10 mL). The combined organic layers were dried (Na₂SO₄), filtered and concentrated *in vacuo*. Flash chromatography (AcOEt/petroleum ether, 3/7) afforded the substituted aziridines **2a-j**. The same procedure was used for *S*-(+)-1-methyl-2-(*o*-tolyl)aziridine using hexane as the solvent.

2-(2-Ethylphenyl)-1-methylaziridine (2b): colourless oil, 95%. ¹H-NMR (CDCl₃, N Et 500 MHz) δ 1.26 (t, *J* = 7.6 Hz, 3 H), 1.61 (d, *J* = 6.5 Hz, 1 H), 1.85 (d, *J* = 3.5 Hz, 1 H), 2.39 (dd, *J* = 3.5, 6.5 Hz, 1 H), 2.51 (s, 3 H), 2.76 (dd, *J* = 2.0, 7.4 Hz, 1 H), 2.79 (dd, *J* = 2.0, 7.4 Hz, 1 H), 7.09-7.22 (m, 4 H). ¹³C-NMR (CDCl₃, 125 MHz) δ 25.7, 38.0, 40.0, 48.0, 58.6, 125.7, 125.9, 126.8, 127.8, 137.3, 142.4. GC-MS *m/z* (%): 161 [M⁺,

18], 160 (100), 146 (98), 131 (58), 117 (96), 91 (29). FT-IR (film, cm⁻¹): 3047, 2967, 2849, 1491, 1459, 1383, 1191, 1013, 800, 753, 732.

¹N Bu MHz) δ 0.98 (t, J = 7.4 Hz, 3 H), 1.41-1.48 (m, 2 H), 1.61-1.67 (m, 3 H), 1.87 (d, J = 3.4 Hz, 1 H), 2.41 (dd, J = 3.4, 6.5 Hz, 2 H), 2.52 (s, 3 H), 2.72-2.82 (m, 2 H), 7.13-7.26 (m, 4 H). ¹³C-NMR (CDCl₃, 100 MHz) δ : 14.0, 22.7, 32.5, 33.0, 38.0, 40.0, 47.9, 125.7, 125.9, 126.6, 128.7, 137.4, 141.1. GC-MS m/z (%): 189 [M⁺, 12], 188 (54), 174 (35), 160 (36), 145 (43), 144 (54), 131 (39), 117 (100), 91 (25). FT-IR (film, cm⁻¹): 2955, 2927, 2870, 1491, 1458, 1259, 1029, 759.



2-Methyl-1-[2-(1-methylaziridin-2-yl)phenyl]propan-2-ol (2d): waxy solid, 57%. ¹H-NMR (CDCl₃, 500 MHz) δ 1.13 (s, 3 H), 1.31 (s, 3 H), 1.60 (d, *J* = 6.7 Hz, 1 H), 1.90 (d, *J* = 3.6 Hz, 1 H), 2.42 (dd, *J* = 4.4, 7.4 Hz, 1 H), 2.44 (s, 3 H), 2.70 (d, *J* = 13.6 Hz, 1 H), 2.99 (d, *J* = 13.6 Hz, 1 H), 7.04-7.13 (m, 4 H). ¹³C-NMR (CDCl₃, 150 MHz) δ 28.1, 32.1, 36.4, 41.5, 45.5, 46.5, 69.3, 126.3, 126.7,

128.2, 131.5, 137.1, 138.2. ESI-MS *m*/*z* (%): 228 [M-Na⁺] (100). FT-IR (film, cm⁻¹): 3385, 2959, 1452, 1379, 1139, 744. Anal calcd for C₁₃H₁₉NO: C, 76.06; H, 9.33; N, 6.82%; Found: C, 75.96; H, 9.24; N, 6.71;



1-[2-(1-Methylaziridin-2-yl)benzyl]cyclohexanol (2e). waxy solid, 21%. ¹H-NMR (CDCl₃, 400 MHz) δ 1.23-1.71 (m, 10 H), 1.66 (d overlapping m at 1.23-1.71 ppm, J = 6.7 Hz, 1 H), 1.98 (d, J = 3.6 Hz, 1 H), 2.48 (dd overlapping s at 2.50 ppm, J = 3.0, 6.5 Hz, 1 H), 2.50 (s, 3 H), 2.80 (d, J =

13.6 Hz, 1 H), 2.93 (d, J = 13.6 Hz, 1 H), 5.28 (br s, 1 H), 7.09-7.18 (m, 4 H). ¹³C-NMR (CDCl₃, 100 MHz) δ 22.2, 22.6, 26.1, 36.3, 36.4, 40.7, 41.7, 44.7, 46.6, 69.9, 126.3, 126.7, 128.4, 131.5, 136.8, 138.3. GC-MS m/z (%): 245 [M⁺, 10], 227 (53), 202 (18), 147 (100), 115 (20). FT-IR (film, cm⁻¹): 3059, 2932, 2854, 1448, 1380, 1264, 986, 735. Anal calcd for C₁₆H₂₃NO: C, 78.32; H, 9.45; N, 5.71; Found: C, 78.12; H, 9.38; N, 5.65.



1-[2-(1-Methylaziridin-2-yl)phenyl]-2-phenylbutan-2-ol (2g). Mixture of diastereoisomers (dr = 70:30). Overall yield 74%.

Major diastereoisomer: yellow solid, mp 71-72 °C, 52%. ¹H-NMR (CDCl₃, 500 MHz) δ 0.66 (t, *J* = 7.3 Hz, 3 H), 1.68 (d, *J* = 6.7 Hz, 1 H), 1.84-1.99 (m, 2 H), 2.10 (d, *J* = 3.6 Hz, 1 H), 2.24 (s, 3 H), 2.37 (dd, *J* = 3.8, 6.5 Hz, 1

H), 3.20 (AB system, 2 H), 7.19-7.28 (m, 5 H), 7.39 (t, J = 7.7 Hz, 2 H), 7.59 (d, J = 7.4 Hz, 2 H). ¹³C-NMR (CDCl₃, 100 MHz) δ 8.0, 33.2, 35.5, 41.8, 46.0, 46.1, 74.8, 125.4, 125.9, 126.4, 127.0, 127.8, 129.0, 131.7, 137.2, 138.2, 149.2. GC-MS m/z (%): 281 [M⁺, 10], 280 (11), 252 (29), 237 (15), 219 (21), 179 (22), 146 (100), 132 (59), 105(17), 103 (28), 91 (35), 77 (41), 44 (41). FT-IR (KBr, cm⁻¹): 3140, 2961, 2953, 2931, 2850, 1442, 1378, 1199, 1020, 774, 753, 704. Anal calcd for C₁₉H₂₃NO: C, 81.10; H, 8,24; N, 4.98%. Found: C, 81.03; H, 8.33%; N, 4.76%.



3,3-Dimethyl-1-[2-(1-methylaziridin-2-yl)phenyl]butan-2-ol (2h). Separable mixture of diastereoisomers (dr = 80:20).

Major diastereoisomer (R^*, R^*)-2h: white solid, mp 38-40 °C, 55%. ¹H-NMR (CDCl₃, 500 MHz) δ 1.04 (s, 9 H), 1.68 (d, J = 6.7 Hz, 1 H), 1.98 (d, J = 3.6 Hz, 1 H), 2.51 (dd, J = 3.6, 6.6 Hz, 1 H), 2.53 (s, 3 H), 2.81 (dd, J = 10.5, 13.4

Hz, 1 H), 2.86 (dd, J = 2.9, 13.4 Hz, 1 H), 3.26 (dd, J = 2.9, 7.6 Hz, 1 H), 7.12-7.24 (m, 4 H). ¹³C-NMR (CDCl₃, 100 MHz) δ 25.9, 34.4, 35.6, 41.4, 46.7, 81.5, 125.8, 127.5, 128.5, 129.7, 137.4, 140.2. GC-MS m/z (%): 233 [M⁺, 3], 176 (100), 144 (24), 134 (39), 131 (37), 117 (25), 103 (27), 57 (22), 44 (51). FT-IR (KBr, cm⁻¹): 3212, 2951, 2866, 1479, 1452, 1380, 1361, 1077, 1015, 805, 758. Anal calcd for C₁₅H₂₃NO: C, 77.21; N, 6.00; H, 9.93%. Found: C, 77.58; N, 6.17; H, 9.92 %. Minor diastereoisomer (**R***,**S***)-2h: colourless oil, 27%. ¹H-NMR (CDCl₃, 500 MHz) δ 1.03 (s, 9 H), 1.59 (d, J = 6.6 Hz, 1 H), 1.93 (d, J = 3.5 Hz, 1 H), 2.40 (dd, J = 2.9, 6.5 Hz, 1 H), 2.51 (s, 3 H), 2.62 (dd, J = 10.7, 13.8 Hz, 1 H), 2.99 (dd, J = 2.0, 13.8 Hz, 1 H), 3.43 (dd, J = 2.0, 10.7 Hz, 1 H), 7.12-7.24 (m, 4 H). ¹³C-NMR (CDCl₃, 100 MHz) δ 25.8, 34.7, 35.1, 37.7, 40.9, 47.9, 80.0, 126.5, 126.9, 130.2, 138.0, 138.3. GC-MS m/z (%): 233 [M⁺ 4], 218 (15), 176 (100), 134 (45), 131 (42), 117 (28), 103 (32), 44 (57). FT-IR (film, cm⁻¹): 3383, 3061, 2952, 2867, 1463, 1454, 1385, 1361, 1069, 1013, 739. Anal calcd for C₁₅H₂₃NO: C, 77.21; N, 6.00; H, 9.93%. Found: C, 77.58; N, 6.17; H, 9.92 %.



1-(4-Chlorophenyl)-2-[2-(1-methylaziridin-2-yl)-phenyl]-ethanol (2i). Separable mixture of diastereoisomers (dr = 70:30).

Major diastereoisomer (R^*, R^*)-2i: white solid, mp 135-137 °C, 59%. ¹H-NMR (CDCl₃, 500 MHz) δ 1.72 (d, J = 6.7 Hz, 1 H), 2.03 (d, J = 3.6 Hz, 1

H), 2.48 (dd, J = 3.8, 6.5 Hz, 1 H), 2.53 (s, 3 H), 3.00 (dd, J = 3.9, 13.8 Hz, 1 H), 3.06 (dd, J = 10.6, 13.8 Hz, 1 H), 4.74 (dd, J = 3.8, 10.4 Hz, 1 H), 7.19-7.47 (m, 8 H). ¹³C-NMR (CDCl₃, 100 MHz) δ 36.1, 41.3, 43.3, 46.4, 74.5, 126.4, 126.7, 127.8, 127.9, 128.3, 128.7, 129.6, 132.2, 137.5, 138.4, 145.0. GC-MS m/z (%): 287 [M⁺, 3], 243 (8), 178 (15), 146 (20), 103 (19), 77 (14), 44 (100). ESI-MS m/z (%): 288 [M-H⁺] (100). FT-IR (KBr, cm⁻¹): 3357, 3060, 2950, 2855, 1489, 1089, 819, 737. Anal calcd for C₁₇H₁₈CINO: C, 70.95; H, 6,30; N, 4.87%. Found: C, 70.76; H, 6.35; N, 4.93%. The enantiomeric purity of (**1***S*,**2**'*S*)-**2i** ([α]²⁰_D = + 46.4, c 0.5, CHCl₃) was determined by HPLC analysis (OD-H chiral column; hexane:*i*PrOH 99:1; flow: 1.0 ml/min; for racemic (*R**,*R**)-**2i** t₁ = 9. min, t₂ = 48.2 min; for (**1***S*,**2**'*S*)-**2i** t = 48.2 min).

Minor diastereoisomer (R^*, S^*)-2i: yellow oil, 26%. ¹H-NMR (CDCl₃, 500 MHz) δ 1.64 (d, J = 6.7 Hz, 1 H), 1.96 (d, J = 3.4 Hz, 1 H), 2.33 (dd, J = 3.9, 6.4 Hz, 1 H), 2.48 (s, 3 H), 2.98 (dd, J = 5.4

Hz, 13.7 Hz, 1 H), 3.28 (dd, J = 5.5, 13.7 Hz, 1 H), 4.99 (t, J = 5.3 Hz, 1 H), 6.56 (d, J = 7.5 Hz, 1 H), 7.02-7.46 (m, 8 H). ¹³C-NMR (CDCl₃, 100 MHz) δ 36.5, 41.3, 41.8, 46.8, 72.3, 126.5, 126.7, 127.5, 127.8, 132.4, 135.5, 137.9, 142.8. GC-MS m/z (%): 287 [M⁺, 5], 243 (21), 146 (68), 131 (26), 103 (32), 77 (26), 44 (100). ESI-MS m/z (%): 288 [M-H⁺] (100). FT-IR (film, cm⁻¹): 3351, 3061, 2950, 2856, 1489, 1454, 1090, 1014, 738. The enantiomeric purity of (**1***R*,**2**'*S*)-**2i** ([α]²⁰_D = + 76.6, c 1.0, CHCl₃) was determined by HPLC analysis (OD-H chiral column; hexane:*i*PrOH 95:5; flow: 1.0 ml/min; for racemic (*R**,*S**)-**2i** t₁ = 9.3 min, t₂ = 10.5 min; for (**1***R*,**2**'*S*)-**2i** t = 10.5 min).

N OH

1-(Furan-2-yl)-2-[2-(1-methylaziridin-2-yl)phenyl]ethanol (2j).

Separable mixture of diastereoisomers (dr = 70:30).

Major diastereoisomer (R^*,R^*)-2j: yellow oil, 56%. ¹H-NMR (CDCl₃, 400 MHz) δ 1.67 (d, J = 6.6 Hz, 1 H), 2.00 (d, J = 3.6 Hz, 1 H), 2.46 (dd, J = 3.6, 6.5 Hz, 1 H), 2.51 (s, 3 H), 3.05 (dd, J = 3.7, 13.7 Hz, 1 H), 3.26 (dd, J

= 11.4, 13.6 Hz, 1 H), 4.66 (dd, J = 3.8, 11.3 Hz, 1 H), 6.24-6.29 (m, 2 H), 7.10-7.11 (m, 2 H), 7.18-7.25 (m, 2 H), 7.33-7.35 (m, 1 H). ¹³C-NMR (CDCl₃, 100 MHz) δ 35.8, 39.2, 41.5, 46.0, 68.6, 104.7, 110.0, 126.4, 127.8, 128.7, 129.4, 137.8, 138.1, 141.4, 158.5. GC-MS m/z (%): 243 [M⁺, 7], 228 (10), 182(17), 144 (26), 132 (22), 115 (26), 95 (23), 44 (100). FT-IR (film, cm⁻¹): 3353, 3115, 3061, 2950, 2854, 1451, 1381, 1144, 1064, 1012, 735. Anal calcd for C₁₅H₁₇NO₂: C, 74.05; H, 7.04; N, 5.76;%. Found: C, 74.28; H, 6.97; N, 5.65 %.

Minor diastereoisomer (\mathbb{R}^* , \mathbb{S}^*)-2j: yellow oil, 24%. ¹H-NMR (CDCl₃, 400 MHz) δ 1.55 (d, J = 6.6 Hz , 1 H), 1.88 (d, J = 3.6 Hz , 1 H), 2.31 (dd, J = 3.6, 6.6 Hz , 1 H), 2.39 (s, 3 H), 3.08 (dd, J = 5.6, 13.9 Hz, 2 H), 3.24 (dd, J = 5.3, 13.9 Hz, 1 H), 4.94 (t, J = 5.4 Hz, 1 H), 5.96-5.07 (m, 1 H), 6.17-6.18 (m, 1 H), 6.63 (d, J = 7.5 Hz, 1 H), 6.94-7.05 (m, 4 H). ¹³C-NMR (CDCl₃, 100 MHz) δ 36.6, 38.7, 41.3, 46.9, 67.8, 106.1, 110.2, 126.5, 126.9, 127.6, 130.5, 135.9, 137.9, 141.2, 156.4. GC-MS m/z (%): 243 [M⁺, 5], 182 (15), 144 (21), 132 (20), 115 (25), 95 (26), 44 (100). FT-IR (film, cm⁻¹): 3359, 3117, 2949, 1733, 1604, 1492, 1454, 1384, 1145, 1050, 1012, 807, 738.

General procedure for the preparation of aminomethylisochromans: A solution of the hydroxyalkylated phenylaziridine (1.0 mmol) in 3.0 mL of acetic acid was stirred at room temperature until the substrate disappeared (TLC monitoring, AcOEt/petroleum ether, 8/2). The resulting reaction mixture was poured into 20 mL of NaOH_{aq} (10%), extracted with CH₂Cl₂ (3×10 mL), dried (Na₂SO₄), and evaporated under reduced pressure. The crude reaction mixture was purified by flash-chromatography (silica gel; CH₂Cl₂/MeOH, 9/1) to give the corresponding isochromans showing the following data:

Spiro[cyclohexane-1,3'-(1'-methylamino)-3,4-dihydro-1H-isochromene] (3a): colourless oil,



80%. ¹H-NMR (CDCl₃, 400 MHz) δ 1.16-1.58 (m, 7 H), 1.69-1.81 (m, 3H), 2.54 (d, J = 15.6 Hz, 1 H), 2.60 (s, 3 H), 2.76 (d, J = 15.6 Hz, 1 H), 2.87 (dd, J = 9.0, 12.1 Hz, 1 H), 3.27 (dd, J = 2.4, 12.1 Hz, 1 H), 4.28 (br s, 1H), 4.96 (d, J = 7.8 Hz, 1 H), 7.02–7.07 (m, 2 H), 7.13-7.16 (m, 2 H). ¹³C-NMR

 $(CDCl_3, 75 \text{ MHz}) \delta 21.9, 22.0, 26.0, 35.3, 38.9, 39.5, 53.4, 56.1, 68.3, 72.2, 123.8, 126.1, 126.9, 129.5, 133.3, 134.5. ESI-MS$ *m*/*z*(%): 246 [M-H]⁺ (100). GC-MS*m*/*z*(%): 201 [M⁺ - 44], 183 (76), 141 (100), 117 (23). FT-IR (film, cm⁻¹): 2934, 2855, 1627, 1450, 1259, 1075, 742.

1-Methylaminomethyl-3*tert*-butyl-3,4-dihydro-1*H*-isochromene (R^*,S^*)-3c: yellow oil, 80%. ¹H-NMR (CDCl₃, 600 MHz) δ 1.0 (s, 9 H), 2.50 (s, 3 H), 2.59 (dd, J = 1.8, 15.7 Hz, 1 H), 2.73–2.85 (m, 2 H), 3.19 (dd, J = 2.9, 12.0 Hz, 1 H), 3.28 (dd, J = 2.6, 11.3 Hz, 1 H), 4.9 (d, J = 8.0 Hz, 1 H), 7.03–7.15 (m, 4 H). ¹³C-NMR (CDCl₃, 150 MHz) δ 25.8, 29.3, 34.0, 36.1, 55.2, 56.5, 75.8, 81.6, 123.6, 126.0, 126.4, 129.3, 135.0, 136.3. ESI-MS m/z (%): 234 [M-H]⁺ (100). FT-IR

(film, cm⁻¹): 2954, 1478, 1362, 1100, 744.

1-Methylaminomethyl-3*tert*-butyl-3,4-dihydro-1*H*-isochromene (R^*,R^*)-3c: yellow oil, 80%. ¹H-NMR (CDCl₃, 400 MHz) δ 1.0 (s, 9 H), 2.52 (s, 3 H), 2.58 (dd, J = 2.8, 16.0 Hz, 1 H), 2.69 (dd, J = 2.9, 12.8 Hz, 1 H), 2.99 (dd, J = 1.8, 12.6 Hz, 1 H), 3.42 (dd, J = 2.8, 11.3 Hz, 1 H), 4.97 (dd, J = 2.8, 10.7 Hz, 1 H), 7.03–7.29 (m, 4 H). ¹³C-NMR (CDCl₃, 150 MHz) δ 25.9, 28.9, 33.8, 36.1, 55.2, 74.2,

1-Methylaminomethyl-3-(4-chlorophenyl)-3,4-dihydro-1*H*-isochromene (*R**,*S**)-3d: colourless



oil, 43%. ¹H-NMR (CDCl₃, 600 MHz) δ 1.9 (br s, 1 H), 2.49 (s, 3 H), 2.93-3.07 (m, 3 H), 3.22 (dd, *J* = 2.9, 12.5 Hz, 1 H), 4.75 (dd, *J* = 2.9, 11.0 Hz, 1 H), 5.15 (d, *J* = 7.7 Hz, 1 H), 7.09–7.25 (m, 5 H), 7.34 (m, 3 H). ¹³C-MNR (CDCl₃, 100 MHz) δ 36.5, 36.9, 57.0, 75.3, 76.6, 124.1, 126.6, 126.6, 126.7, 127.3, 128.5, 128.9, 133.3, 134.0, 135.8, 140.7.

ESI-MS m/z (%): 288 [M-H]⁺ (100). GC-MS m/z (%): 245 [M⁺-H, 3], 243 (28), 178 (82), 131 (76), 103 (100).FT-IR (film, cm⁻¹): 3435, 2962, 1437, 1262, 1090, 802. The enantiomeric purity of (**1***R*,**3***S*)-**3d** ([α]²⁰_D = -19.3, c 0.2, CHCl₃) was determined by HPLC analysis (OD-H chiral column; hexane:*i*PrOH 95:5; flow: 1.5 ml/min; for racemic (*R**,*S**)-**3d** t₁ = 4.90 min, t₂ = 7.13 min; for (**1***R*,**3***S*)-**3d** resulted t = 4.90 min).

1-Methylaminomethyl-3-(4-chlorophenyl)-3,4-dihydro-1*H*-isochromene (*R**,*R**)-3d: yellow



oil, 32%. ¹H-NMR (CDCl₃, 400 MHz) δ 2.34 (br s, 1 H), 2.47 (s, 3 H), 2.84 (dd, J = 2.9, 13.2 Hz, 1 H), 2.94 (d, J = 6.6 Hz, 1 H), 3.12 (dd, J = 10.2, 12.9 Hz, 2 H), 4.89 (t, J = 6.6 Hz, 1 H), 5.11 (dd, J = 2.6, 10.1 Hz, 1 H), 7.06–7.19 (m, 4 H), 7.30-7.36 (m, 4 H). ¹³C-NMR (CDCl₃, 150 MHz) δ 36.0, 36.0₂, 55.4, 69.3, 74.2, 125.2, 126.4, 126.8, 127.5, 128.6, 128.7,

133.3, 133.3₂, 135.2, 140.4. ESI-MS m/z (%): 288 [M-H]⁺ (100). FT-IR (film, cm⁻¹): 2929, 1490, 1085, 749. The enantiomeric purity of (**1***R*,**3***R*)-**3d** ($[\alpha]^{20}_{D} = +6$, c 0.5, CHCl₃) was determined by HPLC analysis (OD-H chiral column; hexane:*i*PrOH 95:5; flow: 1.5 ml/min; for racemic (*R**,*R**)-**3d** t₁ = 6.3 min, t₂ = 10.0 min; for (**1***R*,**3***R*)-**3d** t = 6.3 min).







S12













S18











S23











S28



Enantiomeric purity of aziridine (S)-4.



Enantiomeric purity of aziridine (*S*)-1 and (*S*)-2a.



Stereochemical analysis on isochromans 3

COSY experiment (600 MHz) of (*R**,*S**)-3c.



Selective NOESY experiment (600 MHz) of (*R**,*S**)-3c.



COSY experiment (600 MHz) of (*R**,*R**)-3c.



Selective NOESY experiment (600 MHz) of (*R**,*R**)-3c.



Selective NOESY experiment (600 MHz) of (*R**,*S**)-3d.



Selective NOESY experiment (600 MHz) of (*R**,*R**)-3d.



X-ray crystal structure of (*R**,*R**)-2i (50% ellipsoid probability)