

Supporting Information

Asymmetric aldol reaction of acetaldehyde and isatin derivatives for the synthesis of convolutamydine E, CPC-1, and a 3a-hydroxyfuroindoline part of madindoline A and B

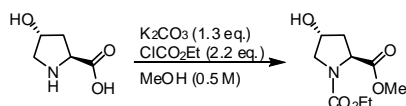
Takahiko Itoh, Hayato Ishikawa, Yujiro Hayashi*

Department of Industrial Chemistry, Faculty of Engineering, Tokyo University of Science,
Kagurazaka, Shinjuku-ku, Tokyo 162-8601, Japan

General Remarks

All reactions were carried out under argon atmosphere and monitored by thin-layer chromatography using Merck 60 F254 precoated silica gel plates (0.25 mm thickness). FT-IR spectra were recorded on a JASCO FT/IR-410 spectrometer. ^1H and ^{13}C NMR spectra were recorded on a Bruker AM400 (400 MHz for ^1H NMR, 100 MHz for ^{13}C NMR) instrument. Data for ^1H NMR are reported as chemical shift (δ ppm), coupling constant (Hz), integration, and assignment. Data for ^{13}C NMR are reported as chemical shift. High-resolution mass spectral analyses (HRMS) were carried out using Bruker ESI-TOF MS. Preparative thin layer chromatography was performed using Wakogel B-5F purchased from Wako Pure Chemical Industries, Tokyo, Japan. Flash chromatography was performed using silica gel 60N of Kanto Chemical Co. Int., Tokyo, Japan. GC-MS was performed on Shimadzu GC-MS QP2010, equipped with a split-mode capillary injection system and electron ionization detectors using Bodman ChiralDEX Γ -TA (30 m \times 0.25 mm). HPLC analysis was performed on a HITACHI Elite LaChrom Series HPLC, UV detection monitored at appropriate wavelength respectively, using Chiralcel OJ-H (0.46 cm \times 25 cm).

(2*S*,4*R*)-1-Ethyl 2-methyl 4-hydroxypyrrolidine-1,2-dicarboxylate



To a solution of (2*S*,4*R*)-4-hydroxypyrrolidine-2-carboxylic acid (15.1 g, 115.7 mmol) and potassium carbonate (20.7 g, 150.4 mmol) in MeOH (230 mL) was added ethyl chloroformate (24.4 mL, 254.5 mmol) at 0 °C. The reaction mixture was stirred for 12 h at room temperature. The resulting mixture was quenched with H₂O (115 mL) and organic materials were extracted with ethyl acetate three times, and then combined organic extracts were dried over anhydrous Na₂SO₄, and concentrated in vacuo after filtration. Purification by column chromatography (ethyl acetate : hexane = 1 : 1) gave (2*S*,4*R*)-1-Ethyl 2-methyl 4-hydroxypyrrolidine-1,2-dicarboxylate (17.5 g, 80.7 mmol) in 70% yield.

NMR spectra data was observed as a mixture of rotamer.

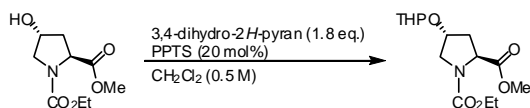
^1H NMR (CDCl₃, 400 MHz): δ 1.15 (1.5H, t, J = 7.2 Hz), 1.21 (1.5H, t, J = 7.2 Hz), 1.96–2.06 (1H, m), 2.17–2.31 (1H, m), 2.69 (1H, br-s), 3.41–3.62 (2H, m), 3.68 (1.5H, m), 3.70 (1.5H, m), 3.95–4.15 (2H, m), 4.33–4.45 (2H, m);

^{13}C NMR (CDCl₃, 100 MHz): δ 14.5, 14.6, 38.5, 39.2, 52.2, 52.3, 54.6, 55.1, 57.7, 57.8, 61.5, 61.6, 69.4, 70.1, 154.9, 155.3, 173.2, 173.3;

IR (KBr): ν 3439, 2984, 2954, 1749, 1682, 1434, 1383, 1350, 1205, 1174 cm⁻¹;

HRMS (ESI): $[M+Na]^+$ calculated for $[C_9H_{15}NO_5Na]$: 240.0842, found: 240.0849;
 $[\alpha]_D^{23} = -75.5$ (c 1.36, MeOH).

(2*S*,4*R*)-1-Ethyl 2-methyl 4-(tetrahydro-2*H*-pyran-2-yloxy)pyrrolidine-1,2-dicarboxylate



To a solution of (2*S*,4*R*)-1-ethyl 2-methyl 4-hydroxypyrrolidine-1,2-dicarboxylate (17.5 g, 80.7 mmol) and pyridinium *p*-toluenesulfonate (4.0 g, 15.9 mmol) in methylene chloride (167 mL) was added 3,4-dihydro-2*H*-pyran (11.2 mL, 142.6 mmol) at 0 °C. The reaction mixture was stirred for 3 h at room temperature. The resulting mixture was quenched with pH 7.0 phosphate buffer solution and organic materials were extracted with ethyl acetate three times, and then combined organic extracts were dried over anhydrous Na_2SO_4 , and concentrated in vacuo after filtration. Purification by column chromatography (ethyl acetate : hexane = 1 : 2) gave (2*S*,4*R*)-1-ethyl 2-methyl 4-(tetrahydro-2*H*-pyran-2-yloxy)-pyrrolidine-1,2-dicarboxylate (20.2 g, 67.0 mmol) in 83% yield.

NMR spectra data was observed as a mixture of diastereomer and rotamer.

1H NMR ($CDCl_3$, 400 MHz): δ 1.06 (1.5H, t, $J = 7.2$ Hz), 1.13 (1.5 H, t, $J = 7.2$ Hz), 1.32–1.50 (4H, m), 1.52–1.76 (2H, m), 1.88–2.07 (1H, m), 2.11–2.38 (1H, m), 3.33–3.64 (6H, m), 3.66–3.76 (1H, m), 3.86–4.09 (2H, m), 4.20–4.39 (2H, m), 4.48–4.58 (1H, m);

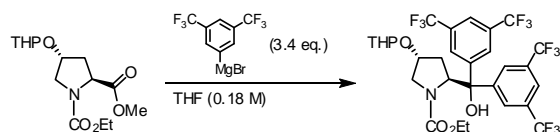
^{13}C NMR ($CDCl_3$, 100 MHz): δ 14.2, 14.3, 18.9, 19.28, 19.34, 25.0, 25.1, 30.33, 30.37, 30.44, 35.1, 35.9, 36.5, 37.4, 38.1, 38.9, 51.2, 51.5, 51.76, 51.82, 51.9, 52.4, 52.7, 54.2, 54.6, 57.3, 57.4, 57.5, 57.6, 57.8, 60.99, 61.06, 61.14, 62.1, 62.2, 62.4, 62.5, 62.6, 68.7, 69.4, 73.1, 74.0, 74.1, 94.2, 97.2, 97.55, 97.63, 97.7, 154.2, 154.4, 154.89, 154.93, 172.8, 172.9, 173.0, 176.2;

IR (KBr): ν 3461, 2952, 2870, 1756, 1681, 1469, 1442, 1270, 1203, 1122, 1022 cm^{-1} ;

HRMS (ESI): $[M+Na]^+$ calculated for $[C_{14}H_{23}NO_6Na]$: 324.1418, found: 324.1418;

$[\alpha]_D^{23} = -60.2$ (c 1.13, MeOH).

(2*S*,4*R*)-Ethyl 2-(bis(3,5-bis(trifluoromethyl)phenyl)(hydroxy)methyl)-4-(tetrahydro-2*H*-pyran-2-yloxy)pyrrolidine-1-carboxylate



To a solution of (2*S*,4*R*)-1-ethyl 2-methyl 4-(tetrahydro-2*H*-pyran-2-yloxy)-pyrrolidine-1,2-dicarboxylate (7.6 g, 25.2 mmol) in tetrahydrofuran (30 mL) was added (3,5-bis(trifluoromethyl)phenyl) magnesium bromide tetrahydrofuran solution (0.75 M solution, 114 mL) at 0 °C. The reaction mixture was stirred for 4 h at room temperature. The resulting mixture was quenched with aqueous NH_4Cl solution and organic materials were extracted with ethyl acetate three times, and then combined organic extracts were dried over anhydrous Na_2SO_4 , and concentrated in vacuo after filtration. Purification by column chromatography (ethyl acetate : hexane = 1 : 2) gave (2*S*,4*R*)-ethyl 2-(bis(3,5-bis(trifluoromethyl)phenyl)(hydroxy)methyl)-4-(tetrahydro-2*H*-pyran-2-yloxy)pyrrolidine-1-carboxylate (16.2 g, 23.0 mmol) in 92% yield.

NMR spectra data was observed as a mixture of diastereomer and rotamer.

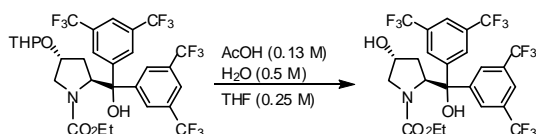
^1H NMR (CDCl_3 , 400 MHz): δ 1.10–1.38 (3H, m), 1.14–1.58 (3H, m), 1.60–1.80 (4H, m), 1.84–1.95 (1H, m), 1.96–2.20 (1H, m), 3.36–3.46 (1H, m), 3.68–3.97 (3H, m), 3.98–4.17 (2H, m), 4.42–4.51 (1H, m), 4.92–5.06 (1H, m), 7.81–7.85 (3H, m), 7.87–7.93 (3H, m);

IR (KBr): ν 2954, 2875, 1735, 1513, 1451, 1247, 1144, 1109, 1005, 736 cm^{-1} ;

HRMS (ESI): $[\text{M}+\text{Na}]^+$ calculated for $[\text{C}_{29}\text{H}_{27}\text{NO}_5\text{F}_{12}\text{Na}]$: 720.1590, found: 720.1566;

$[\alpha]_{\text{D}}^{23} = +48.3$ (c 1.72, MeOH).

(2*S*,4*R*)-Ethyl 2-(bis(3,5-bis(trifluoromethyl)phenyl)(hydroxy)methyl)-4-hydroxypyrrolidine-1-carboxylate



To a solution of (2*S*,4*R*)-ethyl 2-(bis(3,5-bis(trifluoromethyl)phenyl)(hydroxy)methyl)-4-(tetrahydro-2*H*-pyran-2-yloxy)pyrrolidine-1-carboxylate (17.7 g, 25.2 mmol) in tetrahydrofuran (95 mL) and H_2O (47 mL) was added acetic acid (188 mL, 197.4 mmol) at room temperature. The reaction mixture was stirred for 3 h at 60 $^{\circ}\text{C}$. The resulting mixture was quenched with pH 7.0 phosphate buffer solution and organic materials were extracted with ethyl acetate three times, and then combined organic extracts were dried over anhydrous Na_2SO_4 , and concentrated in vacuo after filtration. Purification by column chromatography (ethyl acetate : hexane = 1 : 1) gave (2*S*,4*R*)-ethyl 2-(bis(3,5-bis(trifluoromethyl)phenyl)(hydroxy)methyl)-4-hydroxypyrrolidine-1-carboxylate (14.5 g, 23.4 mmol) in 93% yield.

^1H NMR (CDCl_3 , 400 MHz): δ 1.13 (3H, t, J = 6.8 Hz), 1.77–1.88 (1H, m), 1.90–1.99 (2H, m), 3.04 (1H, dd, J = 4.0, 12.8 Hz), 3.70–3.77 (1H, m), 3.91–4.14 (3H, m), 4.15–4.20 (1H, m), 5.04 (1H, t, J = 8.4 Hz), 7.81–7.87 (3H, m), 7.88–7.94 (3H, m);

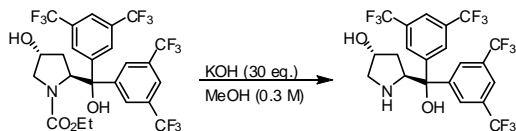
^{13}C NMR (CDCl_3 , 100 MHz): δ 14.6, 39.6, 56.7, 62.8, 67.4, 69.0, 80.3, 107.9 (4C), 121.7 (4C), 131.4 (4C), 144.9 (2C), 146.5 (2C), 158.4;

IR (KBr): ν 3393, 2987, 1675, 1429, 1372, 1348, 1279, 1173, 1134, 682 cm^{-1} ;

HRMS (ESI): $[\text{M}+\text{Na}]^+$ calculated for $[\text{C}_{24}\text{H}_{19}\text{NO}_4\text{F}_{12}\text{Na}]$: 636.1015, found: 636.1020;

$[\alpha]_{\text{D}}^{22} = +54.5$ (c 1.49, MeOH).

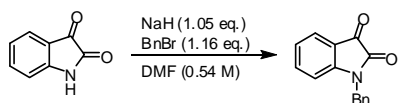
(3*R*,5*S*)-5-(Bis(3,5-bis(trifluoromethyl)phenyl)(hydroxy)methyl)pyrrolidin-3-ol (6)



To a solution of (2*S*,4*R*)-ethyl 2-(bis(3,5-bis(trifluoromethyl)phenyl)(hydroxy)methyl)-4-hydroxypyrrolidine-1-carboxylate (10.7 g, 17.3 mmol) in MeOH (57 mL) was added potassium hydroxide (29.0 g, 519 mmol) at room temperature. The reaction mixture was stirred for 3 h at 90 $^{\circ}\text{C}$. The resulting mixture was quenched with pH 7.0 phosphate buffer solution and organic materials were extracted with ethyl acetate three times, and then combined organic extracts were dried over anhydrous Na_2SO_4 , and concentrated in vacuo after filtration. Purification by column chromatography (ethyl acetate : hexane = 1 : 1) gave (3*R*,5*S*)-5-(bis(3,5-bis(trifluoromethyl)phenyl)(hydroxy)methyl)pyrrolidin-3-ol (7.1 g, 13.1 mmol) in 88% yield.

^1H NMR (CDCl_3 , 400 MHz): δ 1.42–1.51 (1H, m), 1.68–1.78 (1H, m), 3.08 (1H, dd, $J = 1.6, 10.0$ Hz), 3.17 (1H, dd, $J = 4.0, 12.0$ Hz), 4.41–4.46 (1H, m), 4.72 (1H, dd, $J = 6.0, 10.0$ Hz), 7.77 (2H, d, $J = 10.8$ Hz), 7.94 (2H, s), 8.11 (2H, s);
 ^{13}C NMR (CD_3OD , 100 MHz): δ 37.1, 56.3, 64.6, 73.0, 78.7, 122.2, 123.4, 126.1, 127.5 (2C), 128.2 (2C), 128.9, 132.8 (2C, d, $J = 4.0$ Hz), 133.1 (2C, d, $J = 4.0$ Hz), 149.3 (2C), 150.2 (2C);
 IR (KBr): ν 3365, 1372, 1278, 1174, 1131, 902, 844, 711, 682 cm^{-1} ;
 HRMS (ESI): $[\text{M}+\text{H}]^+$ calculated for $[\text{C}_{21}\text{H}_{16}\text{NO}_2\text{F}_{12}]$: 542.0984, found: 542.0984;
 $[\alpha]_{\text{D}}^{23} = +27.7$ (c 1.64, MeOH).

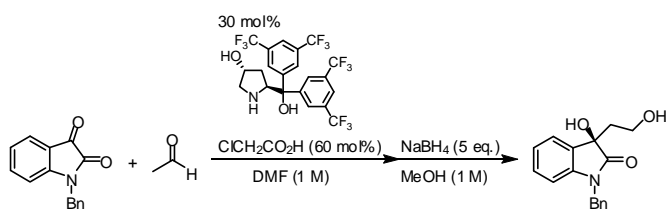
1-Benzylindoline-2,3-dione (7a)



A solution of isatin (5.0 g, 34 mmol) in DMF (62 mL) was cooled to 0 °C (ice bath). NaH (60% dispersion in mineral oil, 1.4 g, 36 mmol) was added portionwise to the orange solution. The color of solution changed to deep purple. When the gas evolution stopped, benzyl bromide (6.7 g, 39 mmol) was added slowly, whereupon the mixture turned red-brown. After the reaction mixture was stirred for 15 min at room temperature, H_2O (300 mL) was introduced to precipitate the product. After filtration, the product was washed with hexane to afford 1-benzylindoline-2,3-dione (7.6 g, 95%) after drying.

^1H NMR (CDCl_3 , 400 MHz): δ 4.85 (2H, s), 6.70 (1H, d, $J = 8.0$ Hz), 7.01 (1H, dt, $J = 0.4, 7.2$ Hz), 7.19–7.31 (5H, m), 7.40 (1H, dt, $J = 1.2, 8.0$ Hz), 7.53 (1H, dd, $J = 0.4, 7.2$ Hz);
 ^{13}C NMR (CDCl_3 , 100 MHz): δ 44.1, 111.0, 117.7, 123.9, 125.4, 127.4 (2C), 128.2, 129.1 (2C), 134.5, 138.3, 150.8, 158.3, 183.2;
 IR (KBr): ν 1731, 1613, 1471, 1349, 1177, 1078, 1004, 766, 754, 694 cm^{-1} ;
 HRMS (ESI): $[\text{M}+\text{Na}]^+$ calculated for $[\text{C}_{15}\text{H}_{11}\text{NO}_2\text{Na}]$: 260.0682, found: 260.0681;

(R)-1-Benzyl-3-hydroxy-3-(2-hydroxyethyl)indolin-2-one (8a) (Table 1, entry 13)



To a solution of (2*S*,4*R*)-4-hydroxy-2-(bis-[3,5-bis(trifluoromethyl)phenyl]hydroxymethyl)pyrrolidine (48 mg, 0.090 mmol), chloroacetic acid (17 mg) and 1-benzylindoline-2,3-dione (71 mg, 0.30 mmol) in DMF (0.30 mL) was added acetaldehyde (84 μL , 1.50 mmol) in the sealed tube (ACE GLASS, product number 5027-05) at 4 °C. After the reaction mixture was stirred for 48 h at 4 °C, MeOH (0.5 mL) and NaBH_4 (56 mg, 1.5 mmol) were added, and the mixture was stirred for 1 h at –20 °C. The resulting mixture was quenched with pH 7.0 phosphate buffer solution and the organic materials were extracted with ethyl acetate three times, and then combined organic extracts were dried over anhydrous Na_2SO_4 , and concentrated in vacuo after filtration. Purification by preparative thin layer chromatography (ethyl acetate : hexane = 1:1) gave (*R*)-1-benzyl-3-hydroxy-3-(2-hydroxyethyl)indolin-2-one (47 mg, 0.17 mmol) in 55% yield with 86% ee.

^1H NMR (CDCl_3 , 400 MHz): δ 2.01–2.10 (1H, m), 2.23–2.32 (1H, m), 3.90–4.01 (2H, m), 4.75 (1H, d, J = 15.6 Hz), 4.94 (1H, d, J = 15.6 Hz), 6.68 (1H, d, J = 8.0 Hz), 7.04 (1H, t, J = 7.6 Hz), 7.17 (1H, dt, J = 1.2, 7.6 Hz), 7.20–7.31 (5H, m), 7.38 (1H, dd, J = 0.8, 8.0 Hz);

^{13}C NMR (CDCl_3 , 100 MHz): δ 39.4, 43.9, 58.6, 76.1, 109.6, 123.3, 123.9, 127.2 (2C), 127.8, 128.9 (2C), 129.7, 130.7, 135.4, 142.0, 178.5;

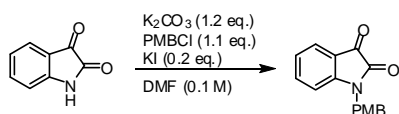
IR (KBr): ν 3393, 2946, 1705, 1614, 1489, 1468, 1368, 1174, 1080, 753 cm^{-1} ;

HRMS (ESI): $[\text{M}+\text{Na}]^+$ calculated for $[\text{C}_{17}\text{H}_{17}\text{NO}_3\text{Na}]$: 306.1101, found: 306.1106;

$[\alpha]_{\text{D}}^{22} = +23.4$ (c 0.95, MeOH).

Enantiometric excess was determined by HPLC with a Chiralcel OJ-H column (hexane : 2-propanol = 10 : 1, λ = 254 nm), 1.0 mL / min; major enantiomer t_{R} = 24.3 min, minor enantiomer t_{R} = 19.3 min.

1-(4-Methoxybenzyl)indoline-2,3-dione (7c)



A solution of isatin (10.2 g, 69 mmol) in DMF (690 mL) was cooled to 0 °C (ice bath). Potassium carbonate (11.5 g, 83 mmol) and potassium iodide (2.3 g, 13.9 mmol) were added to the orange solution. The color of solution changed to deep purple. When the gas evolution stopped, *p*-methoxybenzyl chloride (11.2 mL, 76 mmol) was added slowly. The reaction mixture was stirred for 3 h at 110 °C. The reaction was quenched with aqueous 1N-HCl and organic materials were extracted with ethyl acetate three times, and then combined organic extracts were dried over anhydrous Na_2SO_4 , and concentrated in vacuo after filtration. The product was washed with hexane to afford 1-(4-methoxybenzyl)indoline-2,3-dione (17.5 g, 96%) after drying.

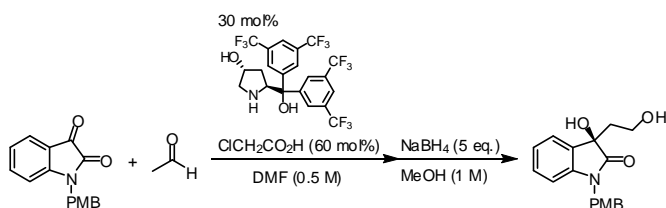
^1H NMR (CDCl_3 , 400 MHz): δ 3.77 (3H, s), 4.85 (2H, s), 6.79 (1H, d, J = 8.0 Hz), 6.82–6.88 (2H, m), 7.06 (1H, dt, J = 0.4, 7.6 Hz), 7.22–7.29 (2H, m), 7.47 (1H, dt, J = 1.2, 7.6 Hz), 7.57 (1H, dd, J = 0.8, 6.8 Hz);

^{13}C NMR (CDCl_3 , 100 MHz): δ 43.6, 55.3, 111.0, 114.4 (2C), 117.7, 123.8, 125.4, 126.5, 128.9 (2C), 138.3, 150.8, 158.3, 159.5, 183.4;

IR (KBr): ν 1735, 1610, 1513, 1467, 1353, 1248, 1182, 1021, 856, 762 cm^{-1} ;

HRMS (ESI): $[\text{M}+\text{Na}]^+$ calculated for $[\text{C}_{16}\text{H}_{13}\text{NO}_3\text{Na}]$: 290.0788, found: 290.0782;

(R)-3-Hydroxy-3-(2-hydroxyethyl)-1-(4-methoxybenzyl)indolin-2-one (8c)



To a solution of (2*S*,4*R*)-4-hydroxy-2-(bis-[3,5-bis(trifluoromethyl)phenyl]hydroxymethyl)pyrrolidine (48 mg, 0.090 mmol), chloroacetic acid (17 mg) and 1-(4-methoxybenzyl)indoline-2,3-dione (80 mg, 0.30 mmol) in DMF (0.60 mL) was added acetaldehyde (84 μL , 1.50 mmol) in the sealed tube (ACE GLASS, product number 5027-05) at 4 °C. After the reaction mixture was stirred for 48 h at 4 °C, MeOH (0.5 mL) and NaBH_4 (56 mg, 1.5 mmol) were added, and the reaction mixture was stirred for 1 h at –20 °C. The

resulting mixture was quenched with pH 7.0 phosphate buffer solution and organic materials were extracted with ethyl acetate three times, and then combined organic extracts were dried over anhydrous Na₂SO₄, and concentrated in vacuo after filtration. Purification by preparative thin layer chromatography (ethyl acetate : hexane = 1:1) gave (*R*)-3-hydroxy-3-(2-hydroxyethyl)-1-(4-methoxybenzyl)indolin-2-one (68 mg, 0.22 mmol) in 73% yield with 86% ee.

¹H NMR (CDCl₃, 400 MHz): δ 2.03–2.12 (1H, m), 2.23–2.32 (1H, m), 3.75 (3H, s), 3.92–3.99 (2H, m), 4.71 (1H, d, *J* = 15.2 Hz), 4.88 (1H, d, *J* = 15.2 Hz), 6.73 (1H, d, *J* = 4.8 Hz), 6.79–6.86 (2H, m), 7.05 (1H, dt, *J* = 0.8, 7.6 Hz), 7.20 (3H, dt, *J* = 0.6, 7.6 Hz), 7.39 (1H, dd, *J* = 0.8, 7.2 Hz);

¹³C NMR (CDCl₃, 100 MHz): δ 39.5, 43.3, 55.3, 58.6, 76.1, 109.7, 114.2 (2C), 123.7, 123.9, 127.5, 128.6 (2C), 129.6, 130.7, 142.0, 159.2, 178.5;

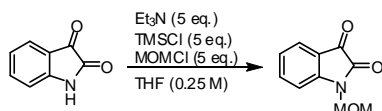
IR (KBr): ν 3395, 1704, 1614, 1513, 1468, 1367, 1249, 1176, 1033, 751 cm⁻¹;

HRMS (ESI): [M+Na]⁺ calculated for [C₁₈H₁₉NO₄Na]: 336.1206, found: 336.1206;

[α]_D²² = +21.5 (*c* 1.53, MeOH).

Enantiometric excess was determined by HPLC with a Chiralcel OJ-H column (hexane : 2-propanol = 10 : 1, λ = 254 nm), 1.0 mL / min; major enantiomer *t*_R = 29.7 min, minor enantiomer *t*_R = 27.9 min.

1-(Methoxymethyl)indoline-2,3-dione (7d)



A solution of isatin (1.0 g, 6.8 mmol) in THF (27 mL) was cooled to 0 °C (ice bath). Triethylamine (4.7 mL, 34 mmol) and trimethylsilyl chloride (4.3 mL, 34 mmol) were added to the orange solution. The reaction mixture was stirred for 2 h at 80 °C. After cooled the reaction mixture to 0 °C, chloro(methoxy)methane (3.0 mL, 34 mmol) was added slowly. The reaction mixture was stirred for 6 h at 80 °C, then the resulting mixture was quenched with pH 7.0 phosphate buffer solution. Organic materials were extracted with ethyl acetate three times, and then combined organic extracts were dried over anhydrous Na₂SO₄, and concentrated in vacuo after filtration. Purification by column chromatography (ethyl acetate : hexane = 1 : 9) gave 1-(methoxymethyl)indoline-2,3-dione (694 mg, 3.6 mmol) in 53% yield.

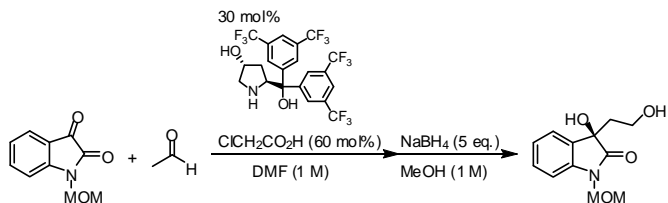
¹H NMR (CDCl₃, 400 MHz): δ 3.37 (3H, s), 5.15 (2H, s), 7.11 (1H, d, *J* = 8.0 Hz), 7.18 (1H, dt, *J* = 0.8, 8.0 Hz), 7.58–7.67 (2H, m);

¹³C NMR (CDCl₃, 100 MHz): δ 56.6, 71.7, 111.6, 117.5, 124.3, 125.4, 138.6, 150.1, 158.4, 182.8;

IR (KBr): ν 2944, 1726, 1605, 1467, 1346, 1287, 1183, 1076, 910, 755 cm⁻¹;

HRMS (ESI): [M+Na]⁺ calculated for [C₁₀H₉NO₃Na]: 214.0475, found: 214.0480;

(R)-3-Hydroxy-3-(2-hydroxyethyl)-1-(methoxymethyl)indolin-2-one (8d)



To a solution of (2*S*,4*R*)-4-hydroxy-2-(bis-[3,5-bis(trifluoromethyl)phenyl]hydroxymethyl)pyrrolidine (48 mg, 0.090 mmol), chloroacetic acid (17 mg) and 1-(methoxymethyl)indoline-2,3-dione (58 mg, 0.30 mmol) in DMF (60 mL) was added acetaldehyde (84 μ L, 1.50 mmol) in the sealed tube (ACE GLASS, product number 5027-05) at 4 °C. After the reaction mixture was stirred for 48 h at 4 °C, MeOH (0.5 mL) and NaBH₄ (56 mg, 1.5 mmol) were added, and the reaction mixture was stirred for 1 h at –20 °C. The resulting mixture was quenched with pH 7.0 phosphate buffer solution and organic materials were extracted with ethyl acetate three times, and then combined organic extracts were dried over anhydrous Na₂SO₄, and concentrated in vacuo after filtration. Purification by preparative thin layer chromatography (ethyl acetate : hexane = 1:1) gave (*R*)-3-Hydroxy-3-(2-hydroxyethyl)-1-(methoxymethyl)indolin-2-one (52 mg, 0.22 mmol) in 72% yield with 86% ee.

¹H NMR (CDCl₃, 400 MHz): δ 2.01–2.09 (1H, m), 2.25–2.35 (1H, m), 3.34 (3H, s), 3.90–4.03 (2H, m), 5.05 (1H, d, *J* = 14.0 Hz), 5.13 (1H, d, *J* = 14.0 Hz), 7.03 (1H, d, *J* = 6.8 Hz), 7.14 (1H, dt, *J* = 0.8, 8.4 Hz), 7.32 (1H, dt, *J* = 0.8, 8.4 Hz), 7.41 (1H, dd, *J* = 0.8, 6.8 Hz);

¹³C NMR (CDCl₃, 100 MHz): δ 39.4, 56.4, 58.6, 71.6, 76.3, 110.0, 123.8, 124.0, 129.9, 130.0, 141.2, 178.8;

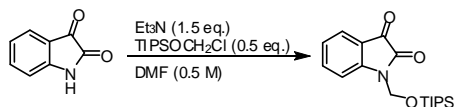
IR (KBr): ν 3399, 1718, 1614, 1487, 1468, 1350, 1182, 1094, 913, 755 cm^{–1};

HRMS (ESI): [M+Na]⁺ calculated for [C₁₂H₁₅NO₄Na]: 260.0893, found: 260.0891;

[α]_D²² = +12.5 (*c* 0.44, MeOH).

Enantiometric excess was determined by HPLC with a Chiralcel OJ-H column (hexane : 2-propanol = 30 : 1, λ = 215 nm), 1.0 mL / min; major enantiomer *t*_R = 49.1 min, minor enantiomer *t*_R = 53.2 min.

1-[(Triisopropylsilyloxy)methyl]indoline-2,3-dione (7e)



A solution of isatin (7.3 g, 49.8 mmol) in DMF (100 mL) was cooled to 0 °C (ice bath). Triethylamine (10.4 mL, 75 mmol) and (chloromethoxy)triisopropylsilane (5.6 g, 24.9 mmol) was added to the orange solution. The reaction mixture was stirred for 24 h at room temperature, then the resulting mixture was quenched with pH 7.0 phosphate buffer solution. Organic materials were extracted with ethyl acetate three times, and then combined organic extracts were dried over anhydrous Na₂SO₄, and concentrated in vacuo after filtration. Purification by column chromatography (ethyl acetate : hexane = 1 : 10) gave 1-[(triisopropylsilyloxy)methyl]indoline-2,3-dione (2.1 g, 6.3 mmol) in 25% yield.

¹H NMR (CDCl₃, 400 MHz): δ 1.06 (18H, d, *J* = 6.8 Hz), 1.12–1.22 (3H, m), 5.41 (2H, s), 7.12–7.19 (2H, m), 7.62 (2H, dt, *J* = 1.6, 7.6 Hz);

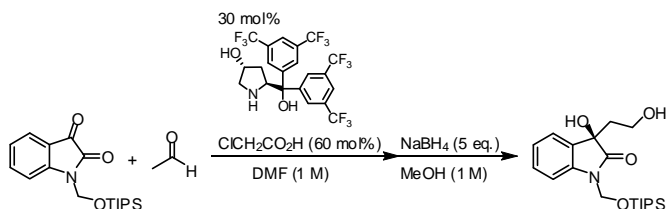
¹³C NMR (CDCl₃, 100 MHz): δ 11.9 (3C), 17.7 (6C), 64.8, 111.8, 117.5, 124.0, 125.3, 138.4, 150.5,

157.2, 183.3;

IR (KBr): ν 2941, 2865, 1746, 1600, 1452, 1311, 1274, 1226, 1159, 1092, cm^{-1} ;

HRMS (ESI): $[\text{M}+\text{Na}]^+$ calculated for $[\text{C}_{18}\text{H}_{27}\text{NO}_3\text{SiNa}]$: 356.1652, found: 356.1660;

(R)-3-Hydroxy-3-(2-hydroxyethyl)-1-((triisopropylsilyloxy)methyl)indolin-2-one (8e)



To a solution of (2*S*,4*R*)-4-hydroxy-2-(bis-[3,5-bis(trifluoromethyl)phenyl]hydroxymethyl)pyrrolidine (71 mg, 0.13 mmol), chloroacetic acid (25 mg, 0.26) and 1-[(triisopropylsilyloxy)methyl]indoline-2,3-dione (146 mg, 0.44 mmol) in DMF (0.44 mL) was added acetaldehyde (123 μL , 2.2 mmol) in the sealed tube (ACE GLASS, product number 5027-05) at 4 $^{\circ}\text{C}$. After the reaction mixture was stirred for 72 h at 4 $^{\circ}\text{C}$, MeOH (0.5 mL) and NaBH_4 (82 mg, 2.2 mmol) were added, and the reaction mixture was stirred for 1 h at -20°C . The resulting mixture was quenched with pH 7.0 phosphate buffer solution and organic materials were extracted with ethyl acetate three times, and then combined organic extracts were dried over anhydrous Na_2SO_4 , and concentrated in vacuo after filtration. Purification by preparative thin layer chromatography (ethyl acetate : hexane = 1:1) gave (*R*)-3-hydroxy-3-(2-hydroxyethyl)-1-((triisopropylsilyloxy)methyl)indolin-2-one (121 mg, 0.31 mmol) in 73% yield with 85% ee.

^1H NMR (CDCl_3 , 400 MHz): δ 1.06 (18H, d, J = 8.8 Hz), 1.11–1.22 (3H, m), 1.94–2.04 (1H, m), 2.22–2.32 (1H, m), 2.60–2.75 (1H, m), 3.90–4.08 (3H, m), 5.30 (1H, d, J = 7.6 Hz), 5.42 (1H, d, J = 7.6 Hz), 7.09 (1H, d, J = 11.6 Hz), 7.13 (1H, dt, J = 0.8, 7.6 Hz), 7.33 (1H, dt, J = 0.8, 7.6 Hz), 7.41 (1H, dd, J = 11.6 Hz);

^{13}C NMR (CDCl_3 , 100 MHz): δ 11.9 (3C), 17.8 (6C), 39.2, 58.6, 64.9, 76.4, 110.3, 123.4, 123.7, 129.7, 130.2, 141.4, 177.3;

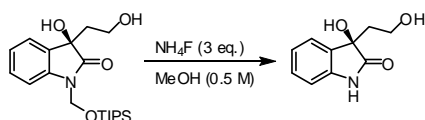
IR (KBr): ν 3392, 2944, 2866, 1718, 1615, 1469, 1364, 1279, 1092, 751 cm^{-1} ;

HRMS (ESI): $[\text{M}+\text{Na}]^+$ calculated for $[\text{C}_{20}\text{H}_{33}\text{NO}_4\text{SiNa}]$: 402.2071, found: 402.2078;

$[\alpha]_{\text{D}}^{20} = +12.8$ (c 0.78, MeOH).

Enantiometric excess was determined by HPLC with a Chiralcel OJ-H column (hexane : 2-propanol = 10 : 1, λ = 254 nm), 1.0 mL / min; major enantiomer t_{R} = 13.1 min, minor enantiomer t_{R} = 13.7 min.

(R)-3-Hydroxy-3-(2-hydroxyethyl)indolin-2-one (14)



To a solution of (*R*)-3-hydroxy-3-(2-hydroxyethyl)-1-((triisopropylsilyloxy)methyl)indolin-2-one (498 mg, 1.3 mmol) in MeOH (2.6 mL) was added ammonium fluoride (147 mg, 3.9 mmol) at room temperature. The reaction mixture was stirred for 12 h at 70 $^{\circ}\text{C}$. After that reaction mixture was concentrated in vacuo, purification by preparative thin layer chromatography (ethyl acetate) gave (*R*)-3-hydroxy-3-(2-hydroxyethyl)indolin-2-one (225 mg, 1.2 mmol) in 88% yield.

^1H NMR (CDCl_3 , 400 MHz): δ 2.00–2.09 (1H, m), 2.22–2.31 (1H, m), 3.90–4.03 (2H, m), 6.86 (1H, d, J = 7.6 Hz), 7.08 (1H, t, J = 7.6 Hz), 7.25 (1H, dt, J = 0.8, 7.6 Hz), 7.38 (1H, d, J = 7.6 Hz), 7.93 (1H, br-s);

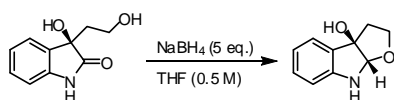
^{13}C NMR (CDCl_3 , 100 MHz): δ 39.2, 58.6, 76.3, 110.3, 123.3, 124.3, 129.8, 131.0, 139.9, 180.2;

IR (KBr): ν 3284, 2923, 1358, 1715, 1621, 1471, 1279, 1178, 753, 584 cm^{-1} ;

HRMS (ESI): $[\text{M}+\text{Na}]^+$ calculated for $[\text{C}_{10}\text{H}_{11}\text{NO}_3\text{Na}]$: 216.0631, found: 216.0631;

$[\alpha]_{\text{D}}^{20}$ = +21.6 (c 0.23, MeOH).

(3a*R*,8a*S*)-3,3a,8,8a-Tetrahydro-2*H*-furo[2,3-*b*]indol-3a-ol (15)



To a solution of (*R*)-3-hydroxy-3-(2-hydroxyethyl)indolin-2-one (118 mg, 0.61 mmol) in tetrahydrofuran (1.2 mL) was added NaBH_4 (116 mg, 3.1 mmol) at 0 °C. The reaction mixture was stirred for 48 h at room temperature. The resulting mixture was quenched with pH 7.0 phosphate buffer solution and organic materials were extracted with ethyl acetate three times, and then combined organic extracts were dried over anhydrous Na_2SO_4 , and concentrated in vacuo after filtration. Purification by preparative thin layer chromatography (ethyl acetate : hexane = 1:1) gave (3a*R*,8a*S*)-3,3a,8,8a-tetrahydro-2*H*-furo[2,3-*b*]indol-3a-ol (51 mg, 0.28 mmol) in 47% yield.

^1H NMR (CDCl_3 , 400 MHz): δ 2.36 (1H, ddd, J = 2.2, 5.4, 12.0 Hz), 2.48 (1H, ddd, J = 7.6, 11.0, 12.0 Hz), 3.69 (1H, ddd, J = 5.4, 9.0, 11.0 Hz), 4.08 (1H, ddd, J = 2.2, 7.6, 9.0 Hz), 4.57 (1H, br-s), 5.41 (1H, s), 6.62 (1H, d, J = 7.6 Hz), 6.82 (1H, t, J = 7.6 Hz), 7.18 (1H, dt, J = 1.2, 7.6 Hz), 7.33 (1H, dt, J = 1.5, 7.6 Hz);

^{13}C NMR (CDCl_3 , 100 MHz): δ 40.9, 67.4, 89.5, 99.4, 109.5, 119.5, 124.1, 130.1, 130.3, 149.6;

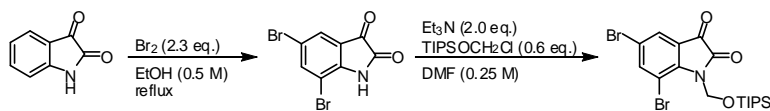
IR (KBr): ν 3382, 2952, 2873, 1668, 1613, 1471, 1313, 1111, 1021, 949, 747 cm^{-1} ;

HRMS (ESI): $[\text{M}+\text{Na}]^+$ calculated for $[\text{C}_{10}\text{H}_{11}\text{NO}_2\text{Na}]$: 200.0682, found: 200.0681;

$[\alpha]_{\text{D}}^{22}$ = -114.8 (c 0.77, CHCl_3);

lit.^{S1)} $[\alpha]_{\text{D}}$ = -144 (c 0.84, CHCl_3).

5,7-Dibromo-1-((triisopropylsilyloxy)methyl)indoline-2,3-dione (9)



5,7-Dibromoindoline-2,3-dione was prepared by known method^{S2)}. A solution of 5,7-dibromoindoline-2,3-dione (9.9 g, 32.5 mmol) in DMF (130 mL) was cooled to 0 °C (ice bath). Triethylamine (9.0 mL, 65 mmol) and (chloromethoxy)triisopropylsilane (4.8 g, 21.6 mmol) were added to the orange solution. The reaction mixture was stirred for 24 h at room temperature, then the resulting mixture was quenched with pH 7.0 phosphate buffer solution. Organic materials were extracted with ethyl acetate three times, and then combined organic extracts were dried over anhydrous Na_2SO_4 , and concentrated in vacuo after filtration. Purification by column chromatography (ethyl acetate : hexane = 1 : 10) gave 5,7-dibromo-1-((triisopropylsilyloxy)methyl)indoline-2,3-dione (4.5 g, 9.1 mmol) in 42% yield.

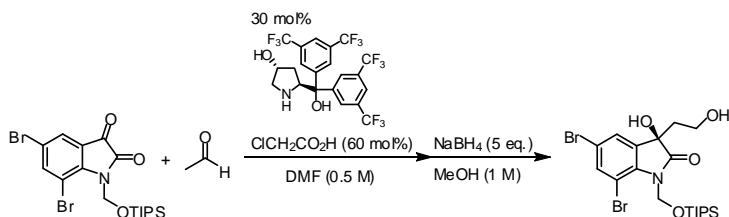
^1H NMR (CDCl_3 , 400 MHz): δ 1.02–1.08 (18H, m), 1.10–1.22 (3H, m), 5.73 (2H, s), 7.72 (1H, d, $J = 2.0$ Hz), 7.91 (1H, d, $J = 2.0$ Hz);

^{13}C NMR (CDCl_3 , 100 MHz): δ 11.2 (3C), 17.1 (6C), 63.7, 105.3, 116.4, 120.6, 126.5, 144.4, 145.9, 156.8, 180.6;

IR (KBr): ν 2941, 2468, 1746, 1600, 1452, 1313, 1274, 1227, 1160, 1092 cm^{-1} ;

HRMS (ESI): $[\text{M}+\text{Na}]^+$ calculated for $[\text{C}_{18}\text{H}_{25}\text{Br}_2\text{NO}_3\text{SiNa}]$: 513.9843, found: 513.9864;

(R)-5,7-Dibromo-3-hydroxy-3-(2-hydroxyethyl)-1-((triisopropylsilyloxy)methyl)indolin-2-one (11)



To a solution of (2*S*,4*R*)-4-hydroxy-2-(bis-[3,5-bis(trifluoromethyl)phenyl]hydroxymethyl)pyrrolidine (71 mg, 0.13 mmol), chloroacetic acid (25 mg, 0.26) and 5,7-dibromo-1-((triisopropylsilyloxy)methyl)indoline-2,3-dione (220 mg, 0.44 mmol) in DMF (0.89 mL) was added acetaldehyde (123 μL , 2.2 mmol) in the sealed tube (ACE GLASS, product number 5027-05) at 4 $^\circ\text{C}$. After the reaction mixture was stirred for 48 h at 4 $^\circ\text{C}$, MeOH (0.5 mL) and NaBH_4 (82 mg, 2.2 mmol) were added, and the reaction mixture was stirred for 1 h at -20 $^\circ\text{C}$. The resulting mixture was quenched with pH 7.0 phosphate buffer solution and organic materials were extracted with ethyl acetate three times, and then combined organic extracts were dried over anhydrous Na_2SO_4 , and concentrated in vacuo after filtration. Purification by preparative thin layer chromatography (ethyl acetate : hexane = 1:1) gave (R)-5,7-Dibromo-3-hydroxy-3-(2-hydroxyethyl)-1-((triisopropylsilyloxy)methyl)indolin-2-one (200 mg, 0.83 mmol) in 83% yield with 81% ee.

^1H NMR (CDCl_3 , 400 MHz): δ 1.06 (18H, dd, $J = 2.0, 9.2$ Hz), 1.10–1.20 (3H, m), 1.89 (1H, ddd, $J = 4.0, 4.8, 15.2$ Hz), 2.27 (1H, ddd, $J = 4.8, 9.2, 15.2$ Hz), 3.85–3.96 (1H, m), 4.06–4.18 (1H, m), 5.66 (2H, s), 7.46 (1H, d, $J = 2.0$ Hz), 7.63 (1H, d, $J = 2.0$ Hz);

^{13}C NMR (CDCl_3 , 100 MHz): δ 11.8 (3C), 17.8 (6C), 36.4, 58.3, 65.1, 77.5, 113.1, 119.8, 123.8, 126.9, 129.7, 144.4, 176.3;

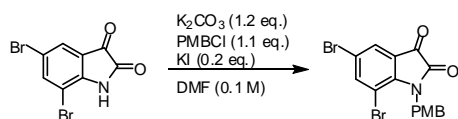
IR (KBr): ν 3356, 2941, 2864, 1725, 1598, 1464, 1278, 1174, 1093, 881 cm^{-1} ;

HRMS (ESI): $[\text{M}+\text{Na}]^+$ calculated for $[\text{C}_{20}\text{H}_{31}\text{Br}_2\text{NO}_4\text{SiNa}]$: 535.0389, found: 535.0394;

$[\alpha]_D^{22} = +14.2$ (c 0.85, MeOH).

Enantiometric excess was determined by HPLC with a Chiralcel OJ-H column (hexane : 2-propanol = 30 : 1, $\lambda = 298$ nm), 1.0 mL / min; major enantiomer $t_R = 6.2$ min, minor enantiomer $t_R = 7.2$ min.

5,7-Dibromo-1-(4-methoxybenzyl)indoline-2,3-dione



A solution of 5,7-dibromoindoline-2,3-dione (2.3 g, 7.5 mmol) in DMF (75 mL) was cooled to 0 $^\circ\text{C}$ (ice bath). Potassium carbonate (1.3 g, 9.1 mmol) and potassium iodide (0.25 g, 1.5 mmol) were added to the orange solution. The color of solution changed to deep purple. When the gas evolution stopped, *p*-methoxybenzyl chloride (1.2 mL, 8.3 mmol) was added slowly. The reaction mixture was stirred for 4 h

at 110 °C. The resulting mixture was quenched with aqueous 1N-HCl and organic materials were extracted with ethyl acetate three times, and then combined organic extracts were dried over anhydrous Na₂SO₄, and concentrated in vacuo after filtration. The product was washed with hexane to afford 5,7-dibromo-1-(4-methoxybenzyl)indoline-2,3-dione (3.0 g, 95%) after drying.

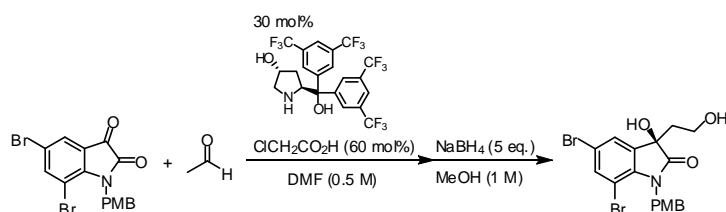
¹H NMR (CDCl₃, 400 MHz): δ 3.77 (3H, s), 5.35 (2H, s), 6.85 (2H, d, *J* = 8.4 Hz), 7.20 (2H, d, *J* = 8.4 Hz), 7.70 (1H, d, *J* = 2.0 Hz), 7.82 (1H, d, *J* = 2.0 Hz);

¹³C NMR (CDCl₃, 100 MHz): δ 44.2, 55.3, 105.2, 114.1, 114.3, 117.1, 121.5, 127.5, 127.6, 128.0, 129.2, 145.3, 146.8, 158.4, 159.3, 181.4;

IR (KBr): ν 1744, 1600, 1513, 1447, 1315, 1247, 1178, 1144, 1031, 724 cm⁻¹;

HRMS (ESI): [M+Na]⁺ calculated for [C₁₆H₁₁Br₂NO₃Na]: 445.8998, found: 445.8985;

(R)-5,7-Dibromo-3-hydroxy-3-(2-hydroxyethyl)-1-(4-methoxybenzyl)indolin-2-one



To a solution of (2*S*,4*R*)-4-hydroxy-2-(bis-[3,5-bis(trifluoromethyl)phenyl]hydroxymethyl)pyrrolidine (63 mg, 0.12 mmol), chloroacetic acid (22 mg, 0.23 mmol) and 5,7-dibromo-1-(4-methoxybenzyl)indoline-2,3-dione (163 mg, 0.38 mmol) in DMF (0.77 mL) was added acetaldehyde (108 μL, 1.93 mmol) in the sealed tube (ACE GLASS, product number 5027-05) at 4 °C. After the reaction mixture was stirred for 48 h at 4 °C, MeOH (0.5 mL) and NaBH₄ (56 mg, 1.5 mmol) were added, and the mixture stirred for 1 h at -20 °C. The resulting mixture was quenched with pH 7.0 phosphate buffer solution and organic materials were extracted with ethyl acetate three times, and then combined organic extracts were dried over anhydrous Na₂SO₄, and concentrated in vacuo after filtration. Purification by preparative thin layer chromatography (ethyl acetate : hexane = 1:1) gave (R)-5,7-dibromo-3-hydroxy-3-(2-hydroxyethyl)-1-(4-methoxybenzyl)indolin-2-one (153 mg, 0.32 mmol) in 85% yield.

¹H NMR (CDCl₃, 400 MHz): δ 1.92–2.01 (1H, m), 2.24–2.35 (1H, m), 3.76 (3H, s), 3.88–3.96 (1H, m), 4.01–4.16 (1H, m), 5.20–5.33 (2H, m), 6.80 (2H, d, *J* = 12.0 Hz), 7.13 (2H, d, *J* = 8.4 Hz), 7.48 (1H, d, 1.6 Hz), 7.52–7.56 (1H, m);

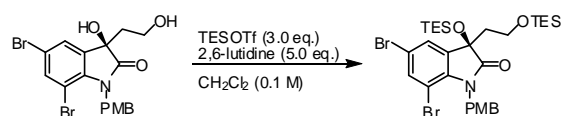
¹³C NMR (CDCl₃, 100 MHz): δ 39.2, 44.0, 55.2, 58.5, 75.5, 103.4, 114.1 (2C), 116.4, 126.5, 127.7 (2C), 128.6, 135.6, 137.3, 138.8, 158.9, 178.7;

IR (KBr): ν 3393, 2362, 2330, 1716, 1513, 1452, 1247, 1177, 1144, 1033 cm⁻¹;

HRMS (ESI): [M+Na]⁺ calculated for [C₁₈H₁₇Br₂NO₄Na]: 491.9417, found: 491.9423;

[α]_D²² = +15.5 (*c* 1.53, MeOH).

(R)-5,7-Dibromo-1-(4-methoxybenzyl)-3-(triethylsilyloxy)-3-(2-(triethylsilyloxy)ethyl)indolin-2-one



To a solution of (*R*)-5,7-dibromo-3-hydroxy-3-(2-hydroxyethyl)-1-(4-methoxybenzyl)indolin-2-one (238 mg, 0.51 mmol), 2,6-lutidine (380 μ L, 2.55 mmol) in methylene chloride (5.1 mL) were added triethylsilyl trifluoromethanesulfonate (354 μ L, 1.53 mmol). The reaction mixture was stirred for 1 h at 0 $^{\circ}$ C. The resulting mixture was quenched with pH 7.0 phosphate buffer solution. Organic materials were extracted with ethyl acetate three times, and then combined organic extracts were dried over anhydrous Na_2SO_4 and concentrated in vacuo after filtration. Purification by column chromatography (ethyl acetate : hexane = 1 : 7) gave (*R*)-5,7-dibromo-1-(4-methoxybenzyl)-3-(triethylsilyloxy)-3-(2-(triethylsilyloxy)ethyl)indolin-2-one (320 mg, 0.46 mmol) in 90% yield.

^1H NMR (CDCl_3 , 400 MHz): δ 0.35–0.54 (12H, m), 0.83 (9H, t, J = 8.0 Hz), 0.88 (9H, t, J = 8.0 Hz), 2.04–2.13 (1H, m), 2.18–2.27 (1H, m), 3.54–3.67 (2H, m), 3.77 (3H, s), 5.21 (1H, d, J = 16.0 Hz), 5.24 (1H, d, J = 16.0 Hz), 6.80–6.85 (2H, m), 7.19–7.25 (1H, m), 7.36 (1H, d, J = 2.0 Hz), 7.36 (1H, d, J = 2.0 Hz), 7.54 (1H, d, J = 2.0 Hz);

^{13}C NMR (CDCl_3 , 100 MHz): δ 4.3 (3C), 5.7 (3C), 6.7 (6C), 43.1, 43.9, 55.2, 57.8, 75.9, 77.3, 102.7, 113.9 (2C), 115.6, 126.7, 128.4 (2C), 129.1, 136.8, 138.8, 158.9, 177.3;

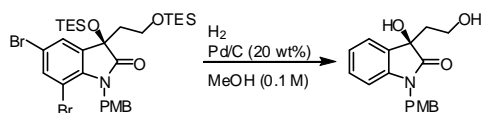
IR (KBr): ν 2954, 2875, 1735, 1513, 1450, 1247, 1144, 1110, 1005, 736 cm^{-1} ;

HRMS (ESI): $[\text{M}+\text{Na}]^+$ calculated for $[\text{C}_{30}\text{H}_{45}\text{Br}_2\text{NO}_4\text{Si}_2\text{Na}]$: 720.1146, found: 720.1120;

$[\alpha]_{\text{D}}^{21} = -15.3$ (c 1.36, MeOH).

The absolute configuration of (*R*)-5,7-Dibromo-3-hydroxy-3-(2-hydroxyethyl)-1-((triisopropylsilyloxy)methyl)indolin-2-one (**11**) was determined after debromination of (*R*)-5,7-dibromo-3-hydroxy-3-(2-hydroxyethyl)-1-(4-methoxybenzyl)indolin-2-one;

(*R*)-3-Hydroxy-3-(2-hydroxyethyl)-1-(4-methoxybenzyl)indolin-2-one

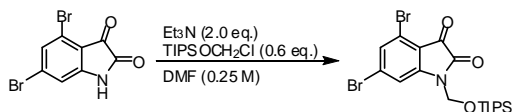


To a solution of (*R*)-5,7-dibromo-1-(4-methoxybenzyl)-3-(triethylsilyloxy)-3-(2-(triethylsilyloxy)ethyl)indolin-2-one (27.0 mg, 0.039 mmol) in MeOH (0.39 mL) was added 20% Pd/C (5.4 mg, 0.0078 mmol) at room temperature. The reaction mixture was stirred for 1h under H_2 atmosphere. Warmed MeOH (5 mL) was added, and the resulting mixture was filtered through a pad of celite, and concentrated in vacuo. Purification by preparative thin layer chromatography (ethyl acetate : hexane = 1:1) gave (*R*)-3-Hydroxy-3-(2-hydroxyethyl)-1-(4-methoxybenzyl)indolin-2-one (10.7 mg, 0.019 mmol) in 88% yield. In this reaction, the deprotection of TES group proceeded in addition to debromination.

Enantiometric excess was determined by HPLC with a Chiralcel OJ-H column (hexane : 2-propanol = 10 : 1, λ = 254 nm), 1.0 mL / min; major enantiomer t_{R} = 28.2 min, minor enantiomer t_{R} = 26.8 min.

The absolute configuration was determined by the comparison with retention time of previous synthetic compound **7c**; see page S5.

4,6-Dibromo-1-((triisopropylsilyloxy)methyl)indoline-2,3-dione (10)



4,6-Dibromoindoline-2,3-dione was prepared from *p*-nitroaniline by known method^{S3}. A solution of 4,6-dibromoindoline-2,3-dione (6.1 g, 20.1 mmol) in DMF (80 mL) was cooled to 0 °C (ice bath). Triethylamine (5.6 mL, 40 mmol) and (chloromethoxy)triisopropylsilane (3.0 g, 13.4 mmol) were added to the orange solution. The reaction mixture was stirred for 24 h at room temperature, then the resulting mixture was quenched with pH 7.0 phosphate buffer solution. Organic materials were extracted with ethyl acetate three times, and then combined organic extracts were dried over anhydrous Na₂SO₄, and concentrated in vacuo after filtration. Purification by column chromatography (ethyl acetate : hexane = 1 : 10) gave 4,6-dibromo-1-((triisopropylsilyloxy)methyl)indoline-2,3-dione (1.6 g, 3.2 mmol) in 24% yield.

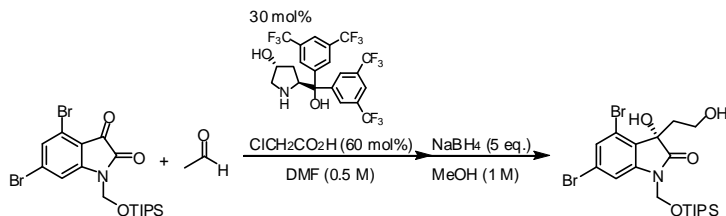
¹H NMR (CDCl₃, 400 MHz): δ 1.04–1.09 (18H, m), 1.01–1.23 (3H, m), 5.40 (2H, s), 7.31 (1H, d, *J* = 1.6 Hz), 7.49 (1H, d, *J* = 1.6 Hz);

¹³C NMR (CDCl₃, 100 MHz): δ 11.2 (3C), 17.1 (6C), 63.7, 105.3, 116.4, 120.6, 126.5, 144.4, 145.9, 156.8, 180.6;

IR (KBr): ν 2942, 2865, 1745, 1591, 1393, 1321, 1239, 1096, 883, 689 cm⁻¹;

HRMS (ESI): [M+Na]⁺ calculated for [C₁₈H₂₅Br₂NO₃SiNa]: 513.9843, found: 513.9861;

(S)-4,6-Dibromo-3-hydroxy-3-(2-hydroxyethyl)-1-((triisopropylsilyloxy)methyl)indolin-2-one (12)



To a solution of (2*S*,4*R*)-4-hydroxy-2-(bis-[3,5-bis(trifluoromethyl)phenyl]hydroxymethyl)pyrrolidine (71 mg, 0.13 mmol), chloroacetic acid (25 mg, 0.26) and 4,6-dibromo-1-((triisopropylsilyloxy)methyl)indoline-2,3-dione (220 mg, 0.44 mmol) in DMF (0.89 mL) was added acetaldehyde (123 μL, 2.2 mmol) in the sealed tube (ACE GLASS, product number 5027-05) at 4 °C. After the reaction mixture was stirred for 48 h at 4 °C, MeOH (0.5 mL) and NaBH₄ (82 mg, 2.2 mmol) were added, and the reaction mixture was stirred for 1 h at –20 °C. The resulting mixture was quenched with pH 7.0 phosphate buffer solution and organic materials were extracted with ethyl acetate three times, and then combined organic extracts were dried over anhydrous Na₂SO₄, and concentrated in vacuo after filtration. Purification by preparative thin layer chromatography (ethyl acetate : hexane = 1:1) gave (*S*)-4,6-dibromo-3-hydroxy-3-(2-hydroxyethyl)-1-((triisopropylsilyloxy)methyl)indolin-2-one (207 mg, 0.86 mmol) in 86% yield with 82% ee.

¹H NMR (CDCl₃, 400 MHz): δ 1.03–1.09 (18H, m), 1.10–1.20 (3H, m), 2.01–2.14 (1H, m), 2.75–2.86 (1H, m), 3.79–4.00 (3H, m), 5.30 (1H, d, *J* = 9.6 Hz), 5.35 (1H, d, *J* = 9.6 Hz), 7.20 (1H, d, *J* = 0.8 Hz), 7.40 (1H, d, *J* = 0.8 Hz);

¹³C NMR (CDCl₃, 100 MHz): δ 11.8 (3C), 17.8 (6C), 36.4, 58.3, 65.1, 77.5, 113.1, 119.8, 123.8, 126.9, 129.7, 144.4, 176.3;

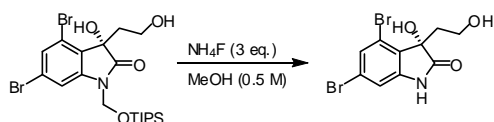
IR (KBr): ν 3356, 2941, 2864, 1725, 1598, 1464, 1278, 1174, 1093, 881 cm⁻¹;

HRMS (ESI): $[M+Na]^+$ calculated for $[C_{20}H_{31}Br_2NO_4SiNa]$: 535.0389, found: 535.0391;

$[\alpha]_D^{17} = -11.2$ (c 0.76, MeOH).

Enantiometric excess was determined by HPLC with a Chiralcel OJ-H column (hexane : 2-propanol = 50 : 1, λ = 222 nm), 0.5 mL / min; major enantiomer t_R = 22.9 min, minor enantiomer t_R = 21.1 min.

ent-Convolutamydine E (13)



To a solution of (*S*)-4,6-dibromo-3-hydroxy-3-(2-hydroxyethyl)-1-((triisopropylsilyloxy)methyl)indolin-2-one (40 mg, 0.082 mmol) in MeOH (0.16 mL) was added ammonium fluoride (9.1 mg, 0.25 mmol) at room temperature. The reaction mixture was stirred for 12 h at 70 °C. After that reaction was concentrated in vacuo, purification by preparative thin layer chromatography (ethyl acetate) gave *ent*-Convolutamydine E (19.0 mg, 0.062 mmol) in 76% yield.

1H NMR (pyridine- d_5 , 400 MHz): δ 3.20 (2H, t, J = 6.8 Hz), 3.62 (1H, s), 4.00–4.04 (2H, m), 7.12 (1H, s), 7.49 (1H, s);

^{13}C NMR (pyridine- d_5 , 100 MHz): δ 39.2, 58.0, 77.4, 112.5, 120.8, 128.1, 130.1, 130.1, 146.7, 180.6;

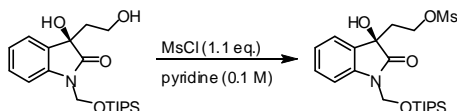
IR (KBr): ν 3275, 2923, 2380, 2348, 1729, 1607, 1574, 1424, 1168, 1082 cm^{-1} ;

HRMS (ESI): $[M+Na]^+$ calculated for $[C_{10}H_9Br_2NO_3Na]$: 371.8841, found: 371.8832;

$[\alpha]_D^{20} = -7.8$ (c 0.67, MeOH, *ent*-Convolutamydine E).

lit.^{S4)} $[\alpha]_D = +12.6$ (c 1.00, MeOH, Convolutamydine E).

(R)-2-(3-Hydroxy-2-oxo-1-((triisopropylsilyloxy)methyl)indolin-3-yl)ethyl methanesulfonate (16)



To a solution of (*R*)-3-hydroxy-3-(2-hydroxyethyl)-1-((triisopropylsilyloxy)methyl)indolin-2-one (492 mg, 1.30 mmol) in pyridine (13 mL) was added methanesulfonyl chloride (111 μ L, 1.43 mmol) at 0 °C. The reaction mixture was stirred for 3 h at room temperature. The resulting mixture was quenched with aqueous 1N-HCl and organic materials were extracted with ethyl acetate three times, and then combined organic extracts were dried over anhydrous Na_2SO_4 , and concentrated in vacuo after filtration. Purification by column chromatography (ethyl acetate : hexane = 1 : 3) gave (*R*)-2-(3-hydroxy-2-oxo-1-((triisopropylsilyloxy)methyl)indolin-3-yl)ethyl methanesulfonate (445 mg, 0.97 mmol) in 75% yield.

1H NMR ($CDCl_3$, 400 MHz): δ 1.03–1.09 (18H, m), 1.11–1.24 (3H, m), 2.33 (1H, dt, J = 6.4, 14.4 Hz), 2.41–2.51 (1H, m), 2.90 (3H, s), 4.32–4.38 (2H, m), 5.36 (2H, d, J = 1.2 Hz), 7.09–7.18 (2H, m), 7.33–7.43 (2H, m);

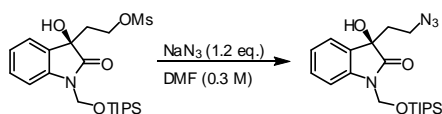
^{13}C NMR ($CDCl_3$, 100 MHz): δ 11.9 (3C), 17.8 (6C), 37.2, 37.3, 65.0, 74.8, 77.3, 110.6, 123.6, 124.0, 128.7, 130.3, 141.8, 176.1;

IR (KBr): ν 3437, 2944, 2866, 1730, 1615, 1469, 1359, 1175, 1094, 752 cm^{-1} ;

HRMS (ESI): $[M+Na]^+$ calculated for $[C_{21}H_{35}NSO_6SiNa]$: 480.1847, found: 480.1835;

$[\alpha]_D^{21} = +7.8$ (c 0.80, MeOH).

(R)-3-(2-Azidoethyl)-3-hydroxy-1-((triisopropylsilyloxy)methyl)indolin-2-one (17)



To a solution of (*R*)-2-(3-hydroxy-2-oxo-1-((triisopropylsilyloxy)methyl)indolin-3-yl)ethyl methane sulfonate (572 mg, 1.25 mmol) in DMF (4.2 mL) was added sodium azide (98 mg, 1.50 mmol) at room temperature. The reaction mixture was stirred for 6 h at 60 °C. The resulting mixture was quenched with pH 7.0 phosphate buffer solution and organic materials were extracted with ethyl acetate three times, and then combined organic extracts were dried over anhydrous Na₂SO₄, and concentrated in vacuo after filtration. Purification by column chromatography (ethyl acetate : hexane = 1 : 3) gave (*R*)-2-(3-hydroxy-2-oxo-1-((triisopropylsilyloxy)methyl)indolin-3-yl)ethyl methanesulfonate (419 mg, 1.04 mmol) in 83% yield.

¹H NMR (CDCl₃, 400 MHz): δ 1.06 (18H, dd, *J* = 2.4, 7.2 Hz), 1.10–1.23 (3H, m), 2.09–2.20 (1H, m), 2.28 (1H, dt, *J* = 8.0, 13.6 Hz), 3.34–3.45 (2H, m), 5.31 (1H, d, *J* = 9.6 Hz), 5.41 (1H, d, *J* = 9.6 Hz), 7.08–7.16 (2H, m), 7.32–7.41 (2H, m);

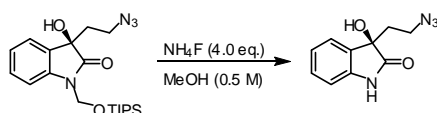
¹³C NMR (CDCl₃, 100 MHz): δ 11.9 (3C), 17.8 (6C), 37.0, 46.2, 64.9, 75.2, 110.5, 123.4, 123.8, 128.9, 130.1, 141.7, 176.3;

IR (KBr): ν 3402, 2943, 2866, 2097, 1715, 1616, 1469, 1366, 1092, 751 cm⁻¹;

HRMS (ESI): [M+Na]⁺ calculated for [C₂₀H₃₂N₄O₃SiNa]: 427.2136, found: 427.2133;

[α]_D²⁰ = +12.5 (*c* 1.20, MeOH).

(R)-3-(2-Azidoethyl)-3-hydroxyindolin-2-one (18)



To a solution of (*R*)-3-(2-azidoethyl)-3-hydroxy-1-((triisopropylsilyloxy)methyl)indolin-2-one (390 mg, 0.97 mmol) in MeOH (2.0 mL) was added ammonium fluoride (144 mg, 3.8 mmol) at room temperature. The reaction mixture was stirred for 3 h at 70 °C. The resulting mixture was concentrated in vacuo. Purification by preparative thin layer chromatography (ethyl acetate : hexane = 1:1) gave (*R*)-3-(2-azidoethyl)-3-hydroxyindolin-2-one (207 mg, 0.97 mmol) in 98% yield.

¹H NMR (CDCl₃, 400 MHz): δ 2.14–2.23 (1H, m), 2.28 (1H, dt, *J* = 7.6, 14.0 Hz), 3.41 (2H, dt, *J* = 2.4, 7.6 Hz), 6.89 (1H, d, *J* = 8.0 Hz), 7.18 (1H, dt, *J* = 0.8, 7.6 Hz), 7.29 (1H, dt, *J* = 1.6, 8.0 Hz), 7.36 (1H, d, *J* = 7.6 Hz);

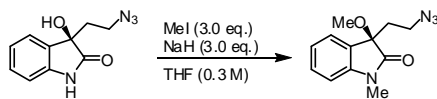
¹³C NMR (CDCl₃, 100 MHz): δ 36.9, 46.3, 75.3, 110.4, 123.4, 124.4, 129.6, 130.1, 140.1, 178.9;

IR (KBr): ν 3313, 2358, 2098, 1706, 1471, 1333, 1264, 1212, 1183, 753 cm⁻¹;

HRMS (ESI): [M+Na]⁺ calculated for [C₁₀H₁₀N₄O₂Na]: 241.0696, found: 241.0692;

[α]_D²⁴ = +2.8 (*c* 0.31, MeOH).

(R)-3-(2-Azidoethyl)-3-methoxy-1-methylindolin-2-one (19)



To a solution of (*R*)-3-(2-azidoethyl)-3-hydroxyindolin-2-one (218 mg, 0.99 mmol) in THF (3.3 mL) was cooled to 0 °C (ice bath). Then NaH (60% dispersion in mineral oil, 129 mg, 2.97 mmol) was added portionwise. When the gas evolution stopped, methyl iodide (0.18 mL, 2.97 mmol) was added slowly. The reaction mixture was stirred for 4 h at room temperature. The resulting mixture was quenched with pH 7.0 phosphate buffer solution and organic materials were extracted with ethyl acetate three times, and then combined organic extracts were dried over anhydrous Na₂SO₄, and concentrated in vacuo after filtration. Purification by column chromatography (ethyl acetate : hexane = 1 : 3) gave (*R*)-3-(2-azidoethyl)-3-methoxy-1-methylindolin-2-one (207 mg, 0.84 mmol) in 85% yield.

¹H NMR (CDCl₃, 400 MHz): δ 2.06–2.16 (1H, m), 2.20–2.31 (1H, m), 2.99 (3H, s), 3.22 (3H, s), 3.27–3.45 (2H, m), 6.87 (1H, d, *J* = 8.0 Hz), 7.11–7.16 (1H, m), 7.29 (1H, d, *J* = 7.2 Hz), 7.34–7.41 (1H, m);

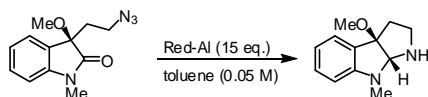
¹³C NMR (CDCl₃, 100 MHz): δ 26.1, 36.6, 45.9, 52.8, 80.9, 108.5, 123.1, 124.1, 126.3, 130.2, 143.9, 175.3;

IR (KBr): ν 2934, 2098, 1725, 1613, 1492, 1469, 1372, 1348, 1105, 754 cm⁻¹;

HRMS (ESI): [M+Na]⁺ calculated for [C₁₂H₁₄N₄O₂Na]: 269.1009, found: 269.1005;

[α]_D²³ = −18.8 (*c* 1.20, MeOH).

(3a*R*,8a*R*)-3a-Methoxy-8-methyl-1,2,3,3a,8,8a-hexahydropyrrolo[2,3-*b*]indole (20)



To a solution of (*R*)-3-(2-azidoethyl)-3-methoxy-1-methylindolin-2-one (14 mg, 0.058 mmol) in toluene (1.2 mL) was added sodium bis(2-methoxyethoxy)aluminum hydride (65 wt% in toluene; 0.26 mL, 0.87 mmol) at 0 °C. The reaction mixture was stirred for 1.5 h at room temperature. After that the solution was heated to 80 °C and maintained at 80 °C for 8 h. After cooling to room temperature, the resulting mixture was quenched with saturated aqueous sodium potassium tartrate (7 mL), diluted with ethyl acetate (5 mL), and stirred vigorously for 45 min. Organic materials were extracted with ethyl acetate three times, and then combined organic extracts were dried over anhydrous Na₂SO₄, and concentrated in vacuo after filtration. Purification by preparative thin layer chromatography (methanol : chloroform = 1 : 10) gave (3a*R*,8a*R*)-3a-methoxy-8-methyl-1,2,3,3a,8,8a-hexahydropyrrolo[2,3-*b*]indole (7.2 mg, 0.035 mmol) in 61% yield.

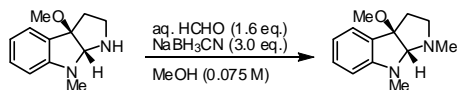
¹H NMR (CDCl₃, 400 MHz): δ 2.17–2.23 (2H, m), 2.75 (1H, dt, *J* = 8.4, 11.2 Hz), 2.86 (3H, s), 3.09 (3H, s), 3.11–3.17 (1H, m), 4.78 (1H, s), 6.41 (1H, d, *J* = 8.0 Hz), 6.69 (1H, dt, *J* = 0.8, 7.6 Hz), 7.14–7.21 (2H, m);

¹³C NMR (CDCl₃, 100 MHz): δ 31.8, 41.9, 45.4, 52.9, 86.4, 94.7, 106.0, 117.1, 124.4, 126.7, 129.9, 152.3;

IR (KBr): ν 2937, 2360, 2341, 1609, 1490, 1294, 1099, 938, 743 cm⁻¹;

HRMS (ESI): $[M+Na]^+$ calculated for $[C_{12}H_{16}N_2ONa]$: 227.1155, found: 227.1164;
 $[\alpha]_D^{19} = -79.1$ (c 0.64, MeOH).

CPC-1 (21)



To a solution of (3a*R*,8a*R*)-3a-methoxy-8-methyl-1,2,3,3a,8,8a-hexahydropyrrolo[2,3-*b*]indole (37 mg, 0.18 mmol) in MeOH (2.4 mL) was added formaldehyde (37% water solution, 23 μ L, 0.29 mmol) at 0 °C. The reaction mixture was stirred for 1 h at room temperature. Then, NaBH₃CN (34 mg, 0.54 mmol) was added and the reaction mixture was stirred for 1 h at room temperature. The resulting mixture was quenched with pH 7.0 phosphate buffer solution and organic materials were extracted with ethyl acetate three times, and then combined organic extracts were dried over anhydrous Na₂SO₄, and concentrated in vacuo after filtration. Purification by preparative thin layer chromatography (methanol : chloroform = 1 : 10) gave CPC-1 (27 mg, 0.12 mmol) in 67% yield.

¹H NMR (CDCl₃, 400 MHz): δ 2.13 (1H, ddd, J = 4.5, 6.0, 12.3 Hz), 2.35 (1H, ddd, J = 6.8, 8.3, 12.3 Hz), 2.58 (3H, s), 2.62 (1H, dt, J = 6.0, 8.8 Hz), 2.80 (1H, ddd, J = 4.4, 6.8, 9.2 Hz), 2.97 (3H, s), 3.04 (3H, s), 4.36 (1H, s), 6.51 (1H, d, J = 7.6 Hz), 6.75 (1H, dt, J = 1.2, 7.6 Hz), 7.16 (1H, dd, J = 1.2, 7.6 Hz), 7.20 (1H, dt, J = 1.2, 7.6 Hz);

¹³C NMR (CDCl₃, 100 MHz): 36.2, 38.6, 39.3, 52.4, 52.5, 91.7, 94.1, 107.8, 117.9, 124.1, 128.1, 129.7, 153.1

IR (KBr): ν 2938, 2791, 1608, 1489, 1162, 1098, 1041, 933, 742 cm⁻¹;

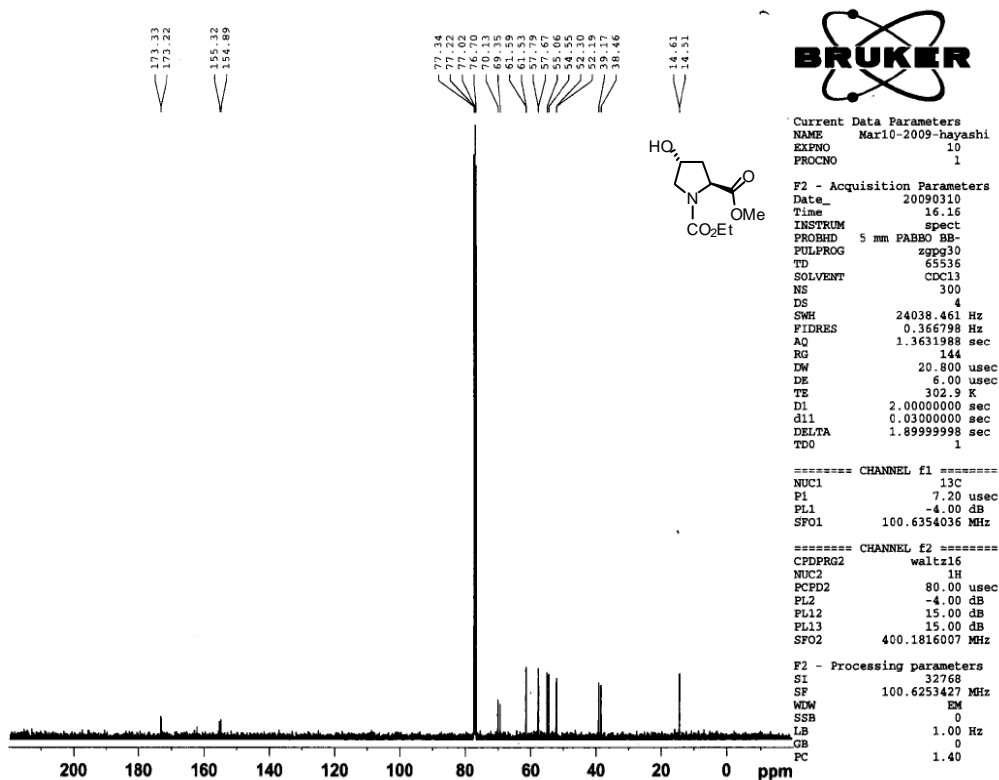
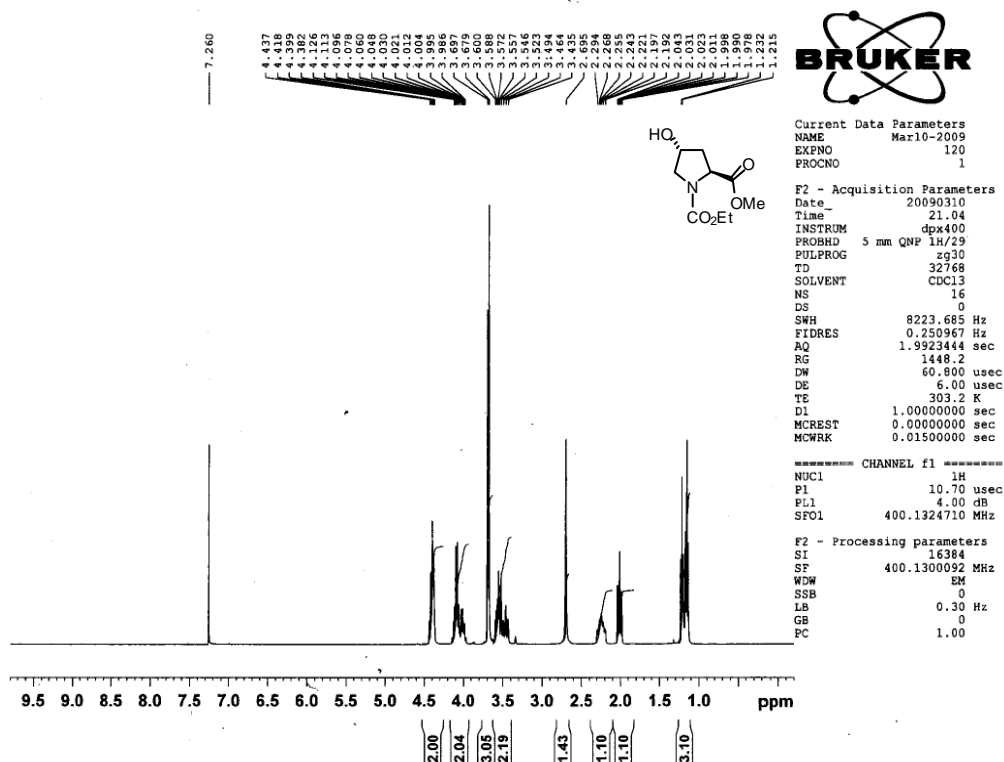
HRMS (ESI): $[M+Na]^+$ calculated for $[C_{13}H_{18}N_2ONa]$: 241.1311, found: 241.1302;

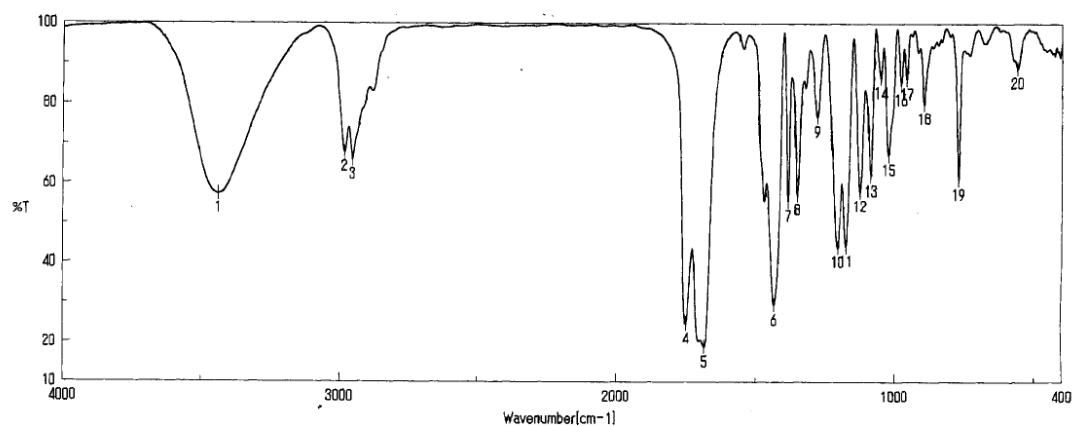
$[\alpha]_D^{19} = -83.0$ (c 0.73, MeOH).

lit.^{S5)} $[\alpha]_D^{26} = -88$ (c 0.1, MeOH).

References;

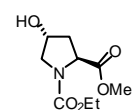
- S1) T. Sunazuka, T. Hirose, T. Shirahata, Y. Harigaya, M. Hayashi, K. Komiyama, S. Omura, A. B. Smith III, *J. Am. Chem. Soc.* **2000**, *122*, 2122.
S2) H. G. Lindwall, J. Bandes, I. Weinberg, *J. Am. Chem. Soc.* **1931**, *53*, 317.
S3) S. J. Garden, J. C. Torres, A. A. Ferreira, R. B. Silva, A. C. Pinto, *Tetrahedron Lett.* **1997**, *38*, 1501.
S4) T. Nakamura, S. Shirokawa, S. Hosokawa, A. Nakazaki, S. Kobayashi, *Org. Lett.* **2006**, *8*, 677.
S5) M. Kitajima, I. Mori, K. Arai, N. Kogure, H. Takayama, *Tetrahedron Lett.* **2006**, *47*, 3199.





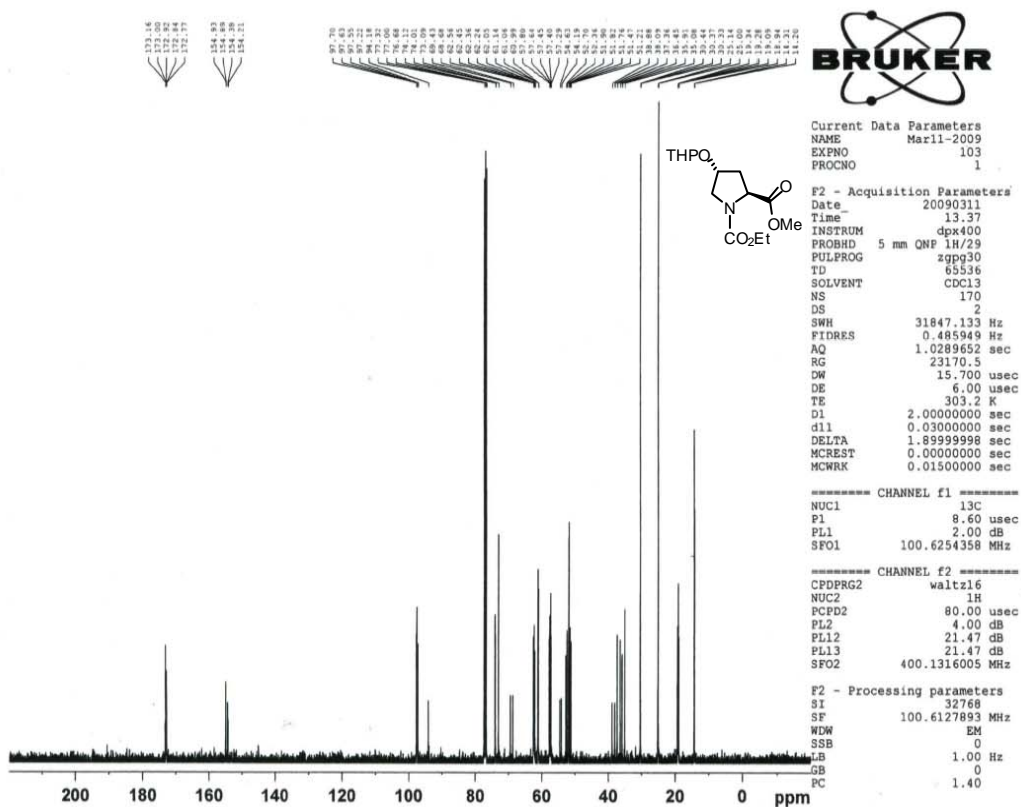
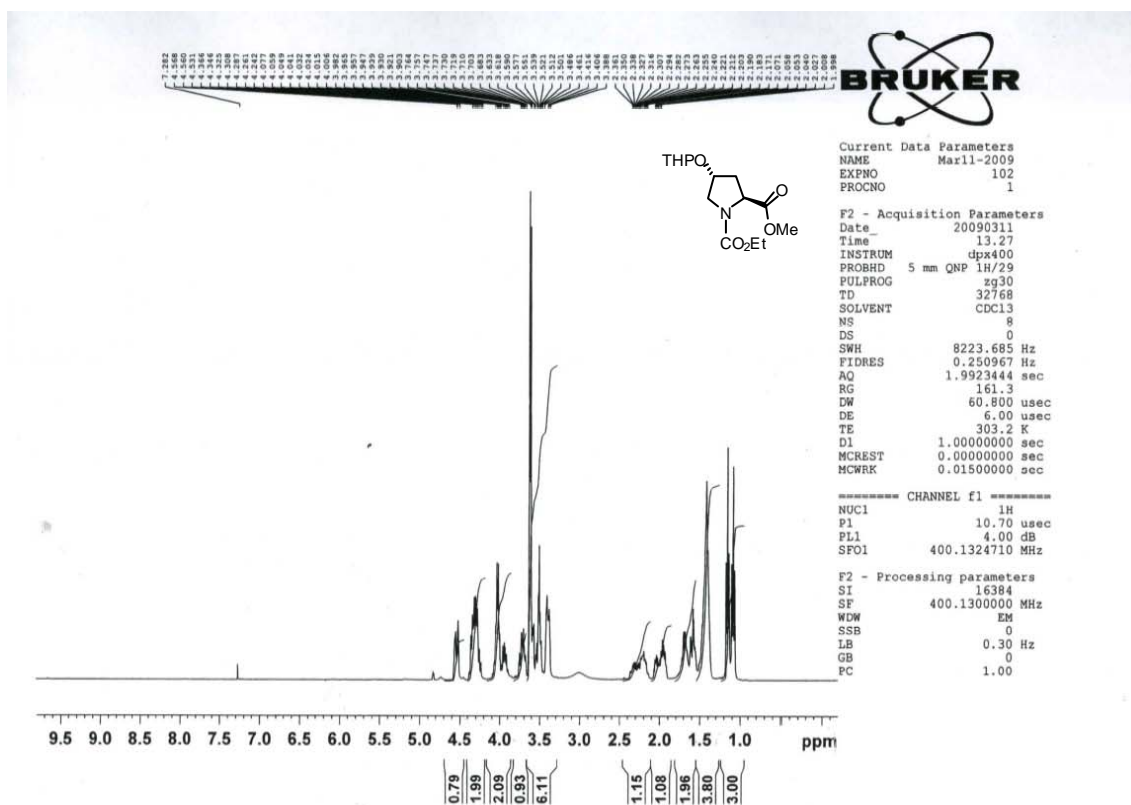
積算回数 16
 ゼロフィリング ON
 ゲイン 2
 日時 10/03/10 18:40
 測定者
 ファイル名 Memory#3
 サンプル名 background
 コメント

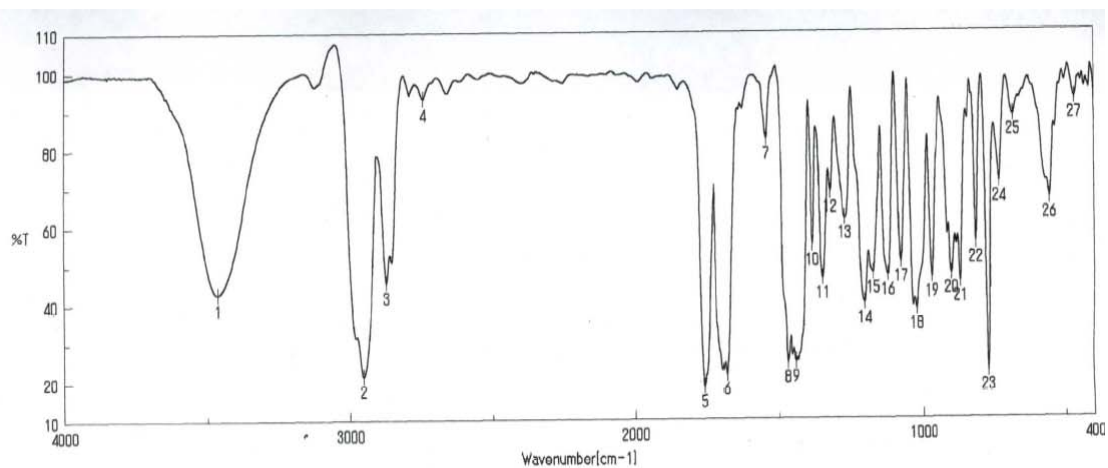
分解 4 cm⁻¹
 アポダイゼーション Cosine
 スキャンスピード 2 mm/sec



1: 3439.42, 57.3560	2: 2984.30, 67.8789	3: 2954.41, 65.9904	4: 1749.12, 24.7730
5: 1682.59, 18.8285	6: 1434.78, 29.3856	7: 1383.68, 55.5817	8: 1350.89, 57.1559
9: 1277.61, 76.6739	10: 1205.29, 43.9232	11: 1174.44, 44.1738	12: 1126.22, 58.0948
13: 1086.69, 61.8590	14: 1050.05, 86.5472	15: 1024.02, 67.1091	16: 978.70, 84.7850
17: 957.48, 86.1910	18: 696.74, 90.0592	19: 773.32, 61.0680	20: 562.15, 88.9576

積算回数=the number of accumulation	分解=resolution
ゼロフィリング=zero filling	アポダイゼーション=apodization
ゲイン=gain	スキャンスピード=scan speed
日時=date	
測定者=user name	
ファイル名=file name	
サンプル名=sample name	
コメント=comment	



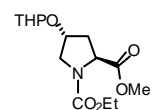


積算回数
 ゼロファイリング
 ゲイン
 日時
 測定者
 ファイル名
 サンプル名
 コメント

16
 ON
 2
 109/03/11 19:48
 Memory#3
 buckground

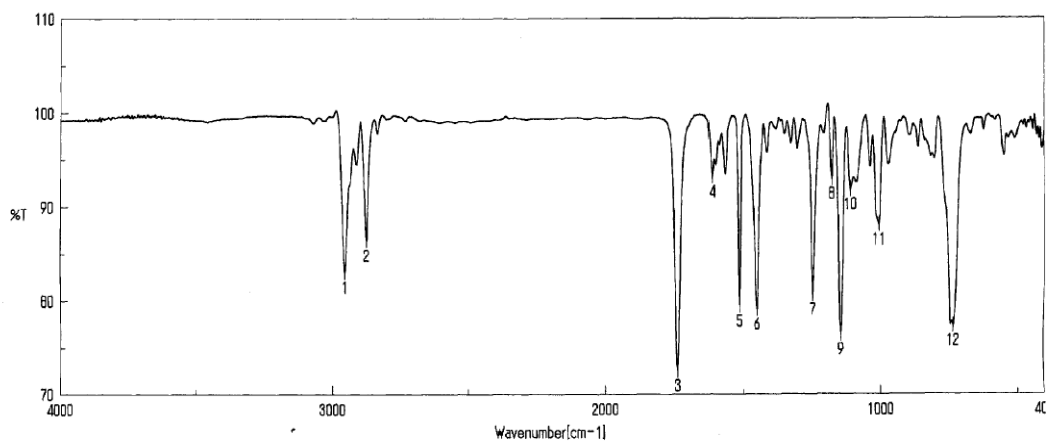
分解
 アボダイゼーション
 スキャンスピード

4 cm-1
 Cosine
 2 mm/sec



1: 3461.60, 42.8242	2: 2952.48, 21.6119	3: 2870.52, 45.6707	4: 2741.32, 93.0654
5: 1756.83, 18.5669	6: 1681.62, 21.7069	7: 1543.74, 82.5993	8: 1469.49, 24.7528
9: 1442.49, 24.9478	10: 1383.68, 55.1146	11: 1347.03, 46.4233	12: 1321.00, 68.6932
13: 1270.86, 62.0144	14: 1203.36, 40.0492	15: 1173.47, 47.6789	16: 1122.37, 47.2491
17: 1077.05, 50.6431	18: 1022.09, 38.4555	19: 969.05, 46.7079	20: 900.59, 47.5202

21: 869.74, 45.5185	22: 813.81, 55.4627	23: 772.35, 22.4313	24: 733.78, 71.2444
25: 684.61, 88.4793	26: 557.33, 67.0409	27: 469.68, 92.8297	

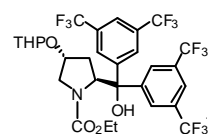


積算回数
ゼロファイリング
ゲイン
日時
測定者
ファイル名
サンプル名
コメント

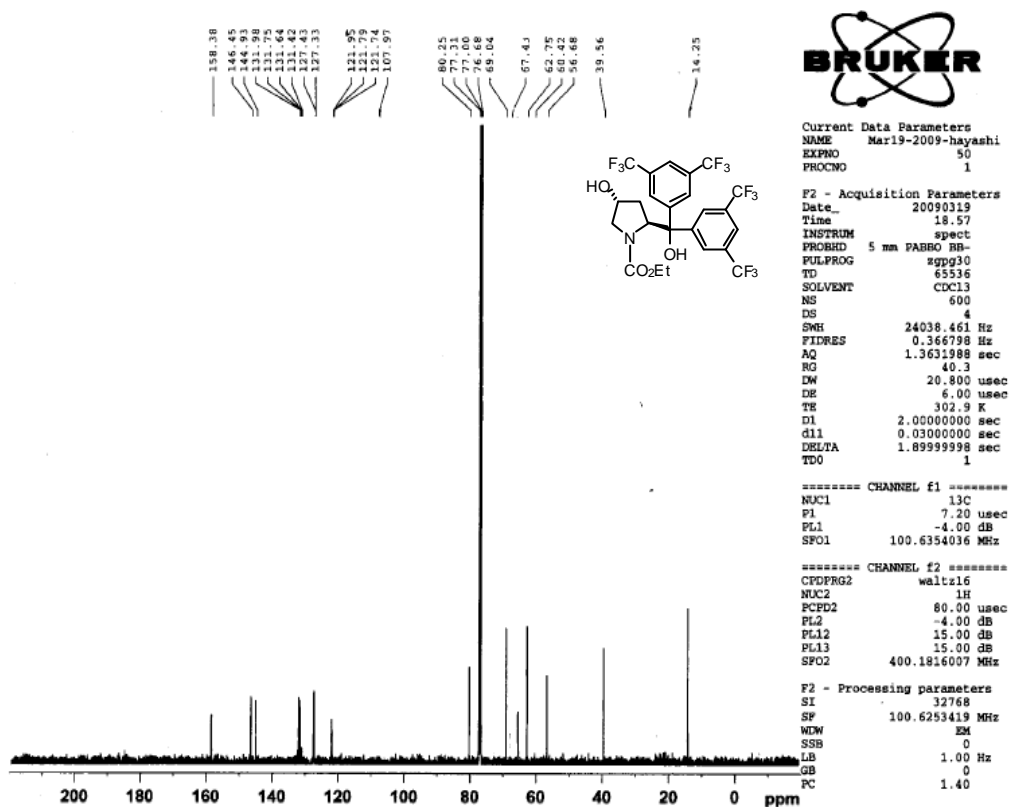
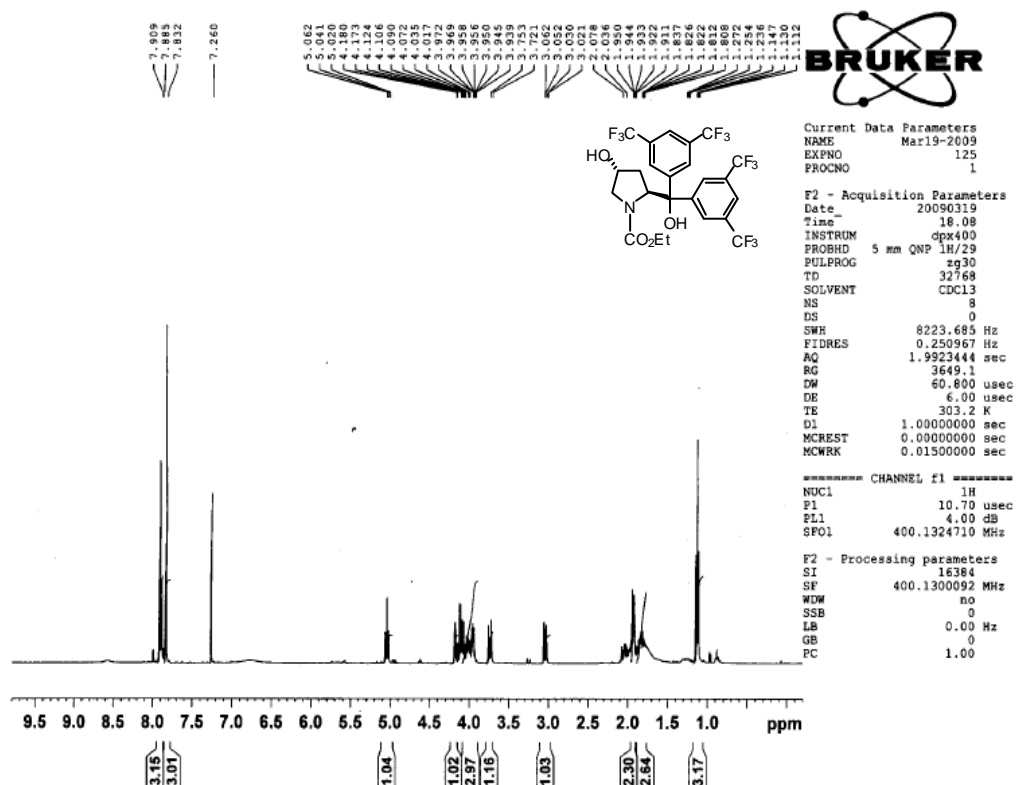
16
ON
2
106/03/11 20:19
Memory#5
background

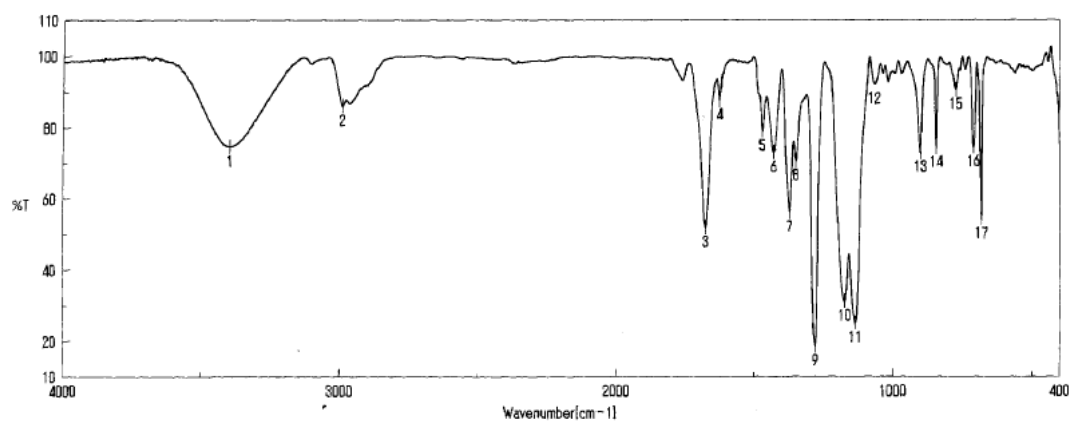
分解
アポダイゼーション
スキャンスピード

4 cm-1
Cosine
2 mm/sec



1: 2954.41, 83.0789	2: 2875.34, 86.4379	3: 1735.62, 72.5595	4: 1613.16, 93.2559
5: 1513.85, 79.4440	6: 1451.17, 79.1066	7: 1247.72, 80.7302	8: 1177.33, 93.0283
9: 1144.55, 78.4633	10: 1109.83, 91.9178	11: 1005.70, 88.1501	12: 736.67, 77.3761



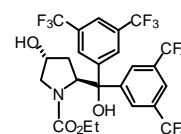


積算回数
ゼロフィリング
ゲイン
日時
測定者
ファイル名
サンプル名
コメント

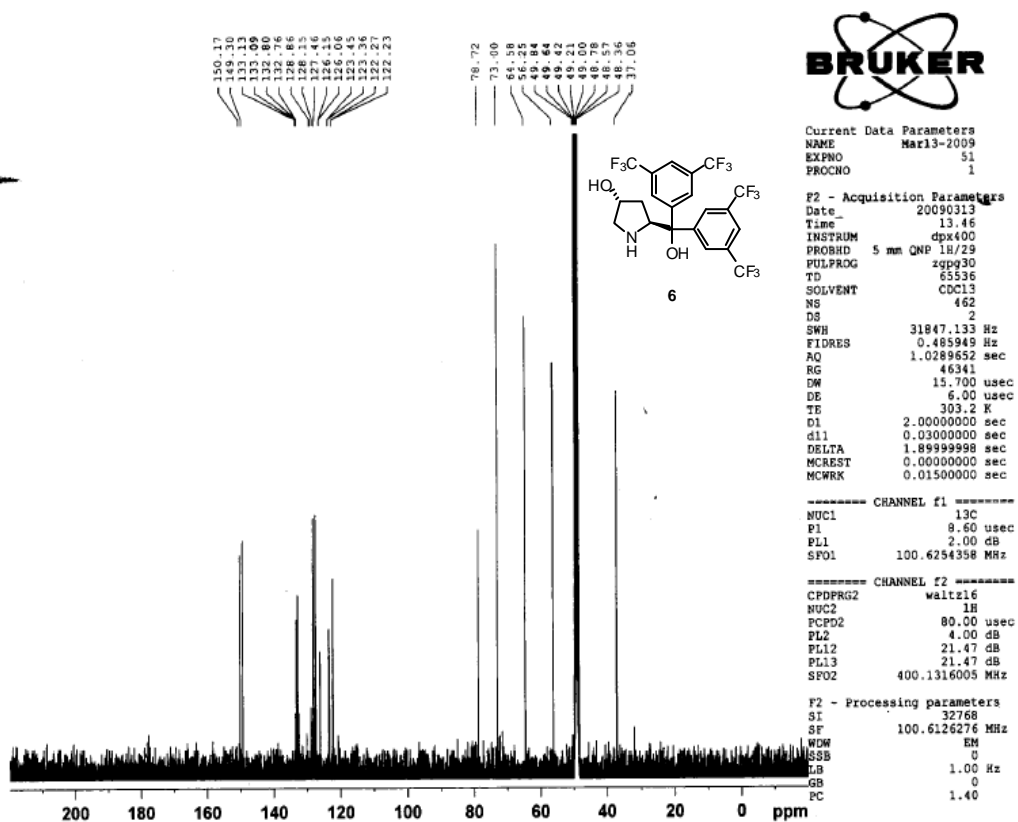
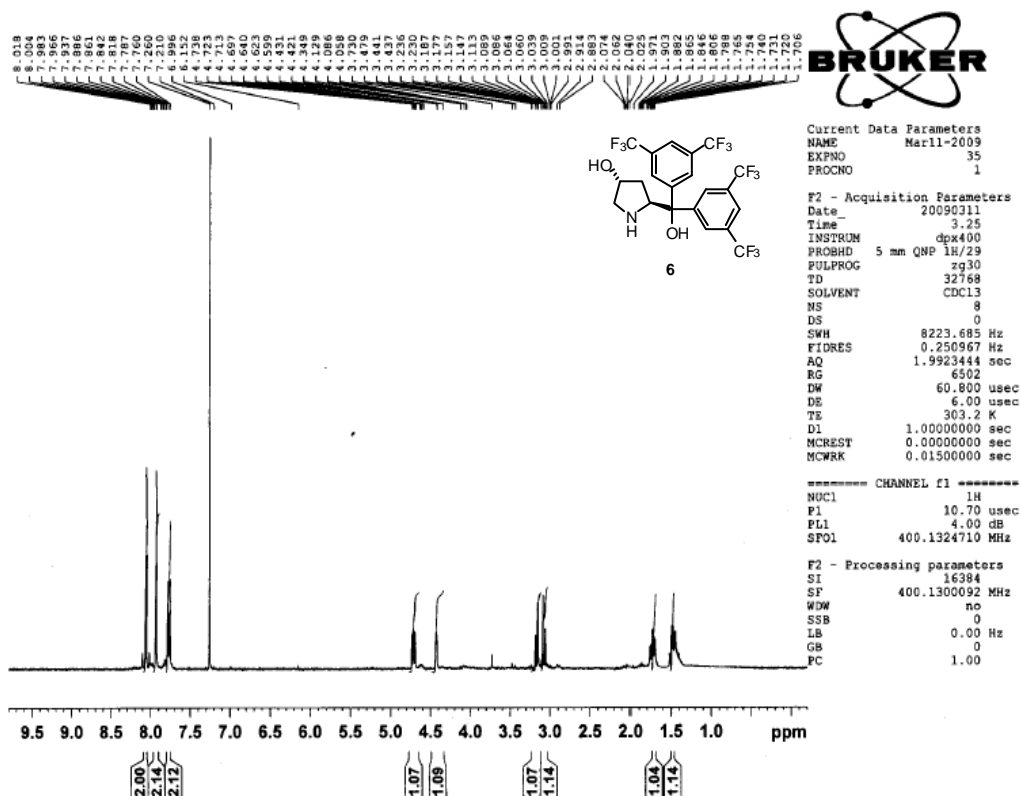
32
ON
2
109/04/24 11:39
Memory#3
background

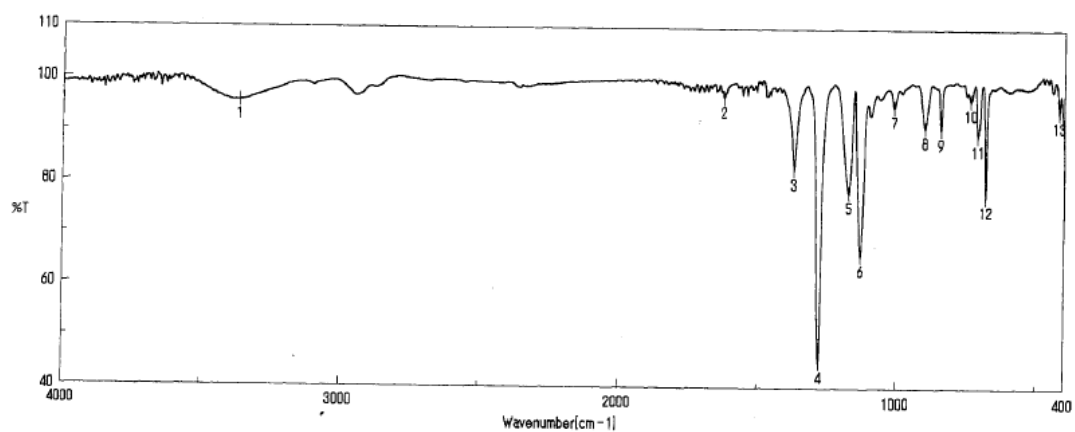
分解
アポダイゼーション
スキャンスピード

4 cm⁻¹
Cosine
2 cm/sec



1: 3393.14, 74.6334	2: 2987.20, 66.1250	3: 1675.84, 51.9780	4: 1624.73, 67.7940
5: 1470.46, 79.0937	6: 1429.96, 73.1706	7: 1372.10, 56.2901	8: 1348.96, 70.7330
9: 1279.54, 18.7468	10: 1173.47, 31.4150	11: 1134.90, 25.2293	12: 1066.44, 92.3618
13: 901.56, 73.1558	14: 844.67, 74.4791	15: 773.32, 90.7548	16: 710.64, 74.6391
17: 682.88, 54.4788			



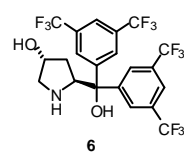


積算回数
ゼロファイリング
ゲイン
日時
測定者
ファイル名
サンプル名
コメント

16
ON
2
109/03/11 20:09
Memory#2
background

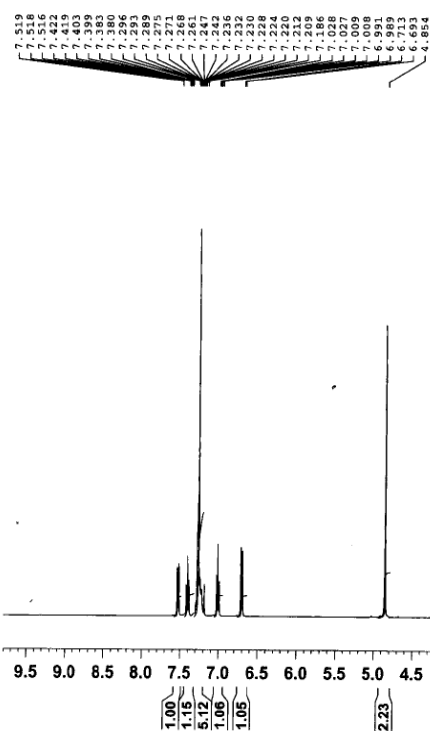
分解
アポダイゼーション
スキャンスピード

4 cm-1
Cosine
2 mm/sec



1: 3385.17, 95.7224	2: 1624.73, 96.6898	3: 1372.10, 82.6798	4: 1278.57, 45.1105
5: 1174.44, 78.2102	6: 1131.05, 65.8814	7: 1013.41, 95.0227	8: 902.52, 90.9982
9: 844.67, 90.6281	10: 737.64, 96.4286	11: 711.60, 89.3870	12: 682.66, 77.4696
13: 418.48, 93.9506			

Bn-isatin



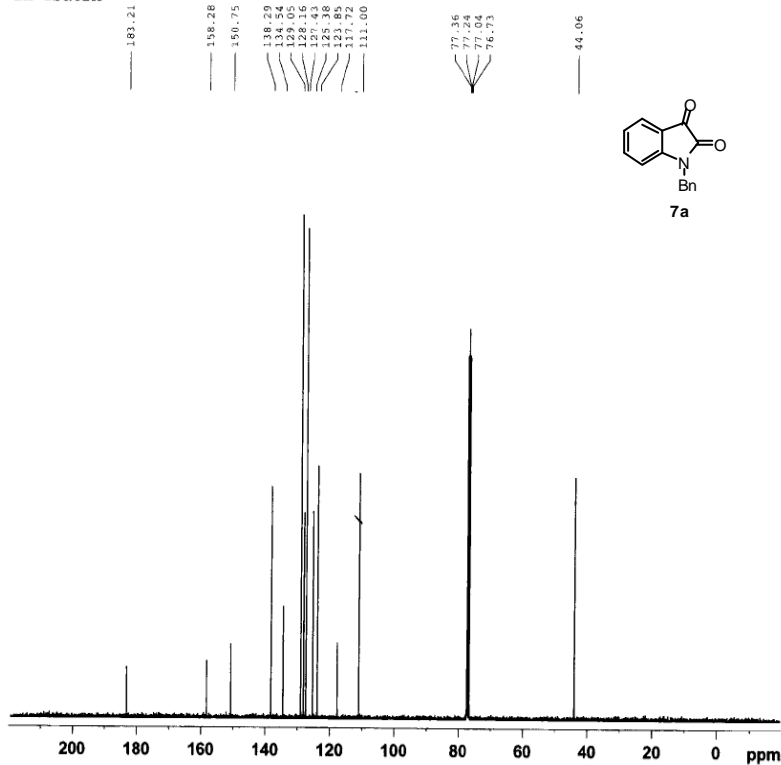
Current Data Parameters
 NAME Dec06-2008-hayashi
 EXPNO 80
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20081206
 Time 11.44
 INSTRUM spect
 PROBRD 5 mm PABBO BB-
 PULPROG zg30
 TD 65536
 SOLVENT CDCl3
 NS 16
 DS 2
 SWH 8223.685 Hz
 FIDRES 0.125483 Hz
 AQ 3.9846387 sec
 RG 181
 DW 50.800 usec
 DE 6.00 usec
 TE 303.0 K
 D1 1.00000000 sec
 TDO 1

===== CHANNEL f1 =====
 NUC1 1H
 P1 12.00 usec
 PL1 -4.00 dB
 SFO1 400.1824713 MHz

F2 - Processing parameters
 SI 32768
 SF 400.1800369 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00

Bn-isatin



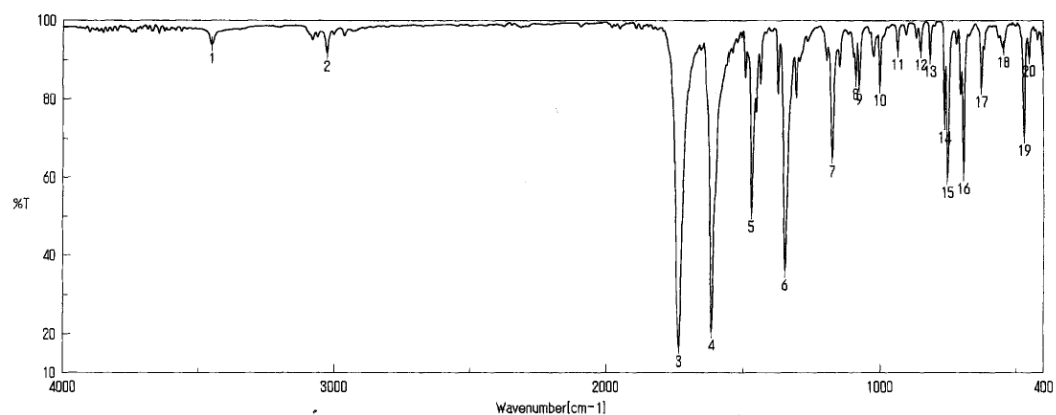
Current Data Parameters
 NAME Dec06-2008-hayashi
 EXPNO 120
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20081206
 Time 16.40
 INSTRUM spect
 PROBRD 5 mm PABBO BB-
 PULPROG zgpg30
 TD 65536
 SOLVENT CDCl3
 NS 300
 DS 4
 SWH 24038.461 Hz
 FIDRES 0.366798 Hz
 AQ 1.3631988 sec
 RG 80.6
 DW 20.800 usec
 DE 6.00 usec
 TE 302.9 K
 D1 2.00000000 sec
 d11 0.03000000 sec
 DELTA 1.89999999 sec
 TDO 1

===== CHANNEL f1 =====
 NUC1 13C
 P1 7.20 usec
 PL1 -4.00 dB
 SFO1 100.6354036 MHz

===== CHANNEL f2 =====
 CPDPRG2 waltz16
 NUC2 1H
 PCPD2 80.00 usec
 PL2 -4.00 dB
 PL12 15.00 dB
 PL13 15.00 dB
 SFO2 400.1816007 MHz

F2 - Processing parameters
 SI 32768
 SF 100.6253448 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40

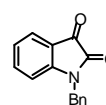


種算回数
ゼロファイリング
ゲイン
日時
測定者
ファイル名
サンプル名
コメント

64
ON
2
109/01/03 15:25
Brisatin. JWS
background

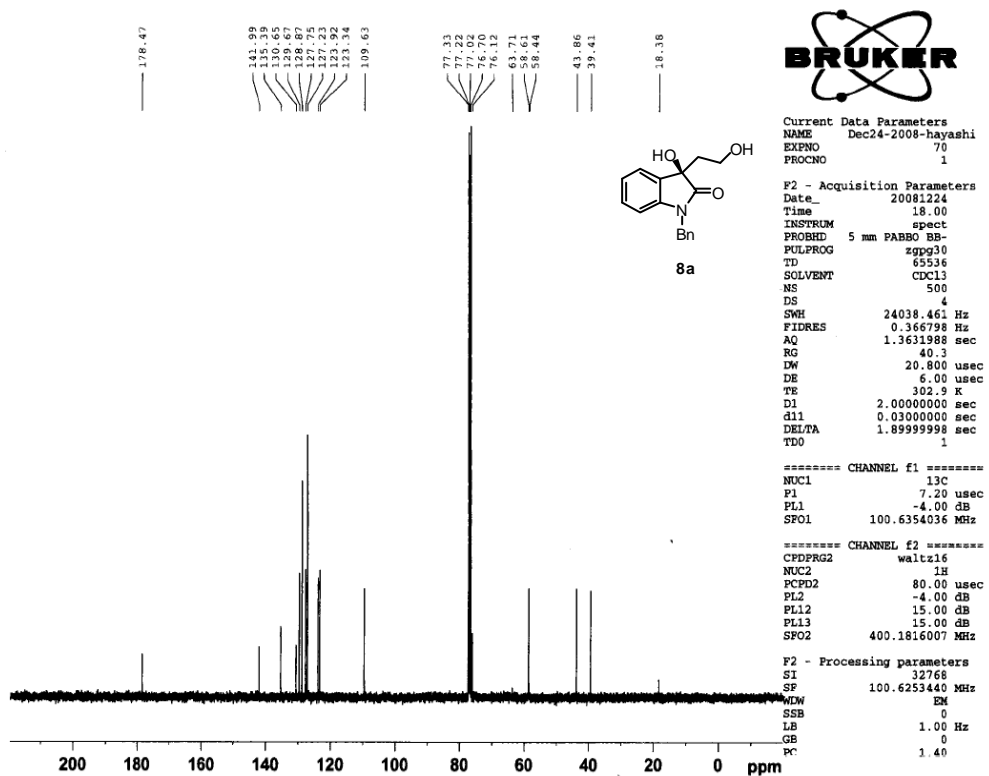
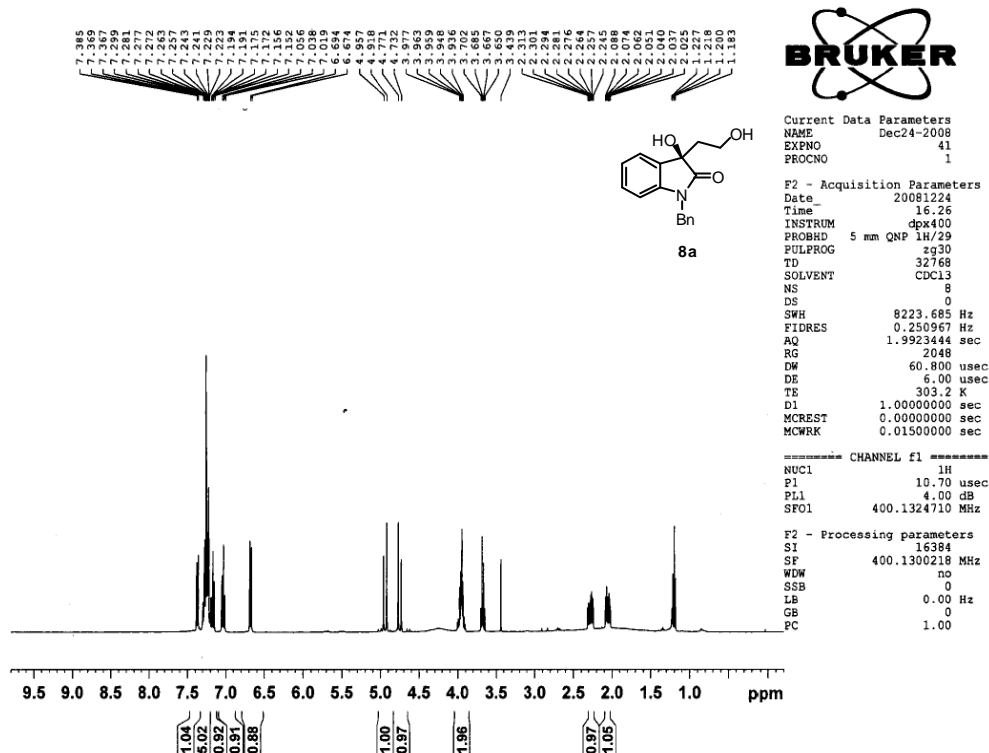
分解
アポダイゼーション
スキャンスピード

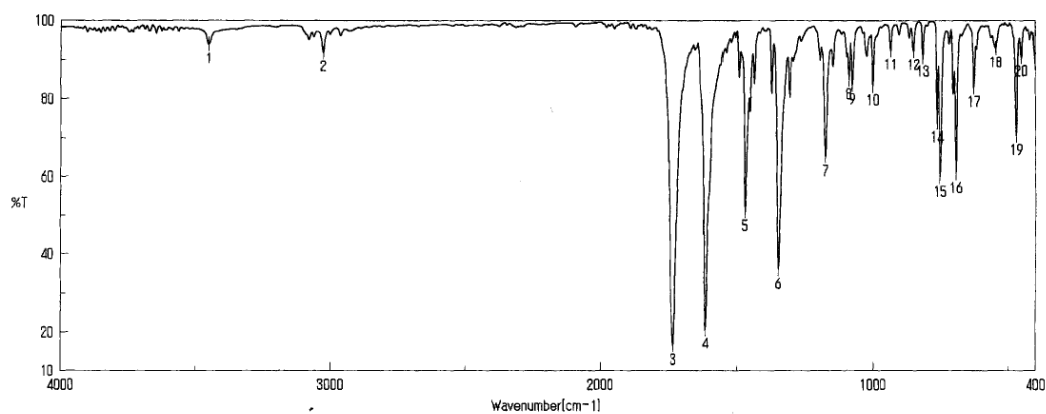
4 cm-1
Cosine
2 mm/sec



7a

1: 3453.88, 94.0413	2: 3029.62, 91.9169	3: 1731.76, 16.4638	4: 1613.16, 20.6778
5: 1471.42, 50.9012	6: 1348.96, 36.1033	7: 1177.33, 65.0943	8: 1091.51, 64.8911
9: 1078.01, 83.6301	10: 1003.77, 83.4161	11: 938.20, 92.4271	12: 853.35, 92.3515
13: 818.63, 90.5934	14: 765.60, 73.8559	15: 754.03, 59.8688	16: 694.25, 60.7783
17: 627.72, 82.9286	18: 547.68, 93.0124	19: 471.51, 70.6046	20: 453.19, 90.6829



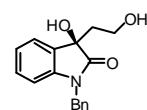


積算回数
ゼロフィリング
ゲイン
日時
測定者
ファイル名
サンプル名
コメント

64
ON
2
109/01/03 15:25
Bnisatin, JWS
background

分解
アボダイゼーション
スキャンスピード

4 cm⁻¹
Cosine
2 mm/sec

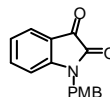
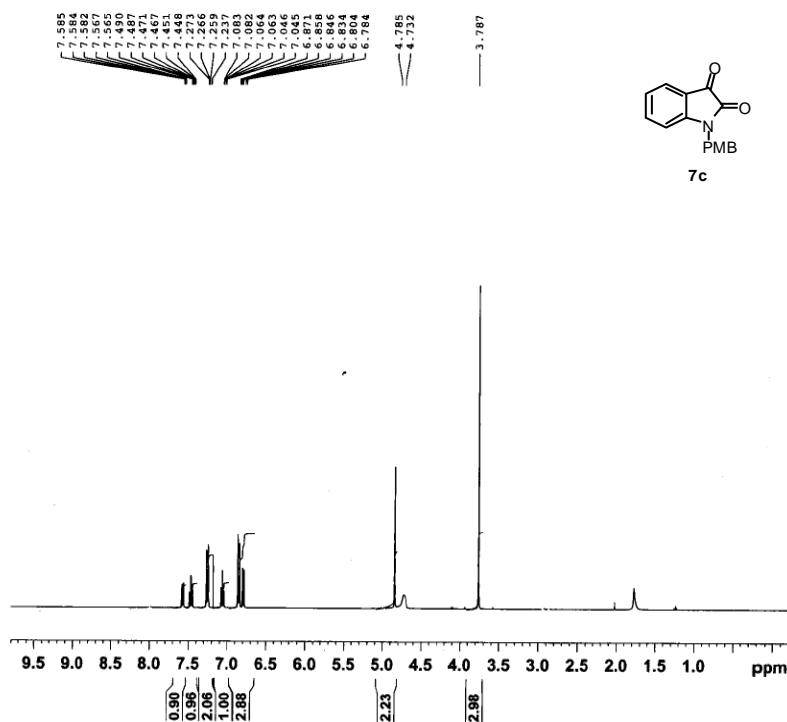


8a

1: 3453.88, 94.0413	2: 3029.62, 91.9189	3: 1731.76, 18.4638	4: 1613.16, 20.6778
5: 1471.42, 50.9012	6: 1348.96, 36.1033	7: 1177.33, 65.0943	8: 1091.51, 84.8911
9: 1078.01, 83.8301	10: 1003.77, 83.4161	11: 938.20, 92.4271	12: 853.35, 92.3515
13: 818.83, 90.5934	14: 765.60, 73.8559	15: 754.03, 59.8688	16: 694.25, 60.7783
17: 627.72, 82.9286	18: 547.68, 93.0124	19: 471.51, 70.6046	20: 453.19, 90.6829

7.585 7.586 7.587 7.588 7.589 7.590 7.591 7.592 7.593 7.594 7.595 7.596

7.585 7.586 7.587 7.588 7.589 7.590 7.591 7.592 7.593 7.594 7.595 7.596



```
Current Data Parameters
NAME      Dec06-2008-hayashi
EXPNO          90
PROCNO        1
```

```

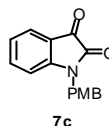
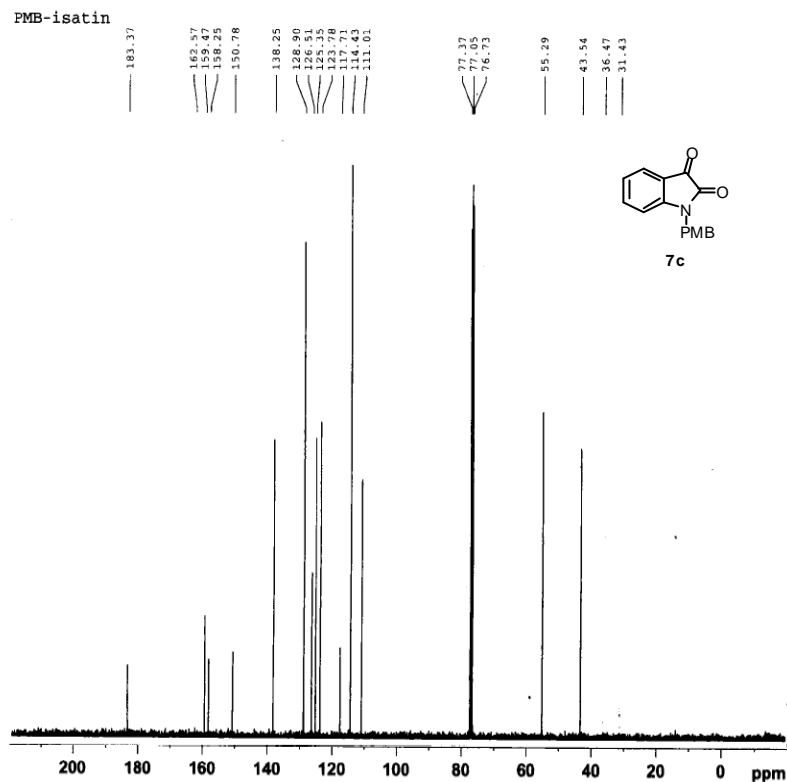
F2 - Acquisition Parameters
Date_      200812206
Time       11.52
INSTRUM    spect
PROBHD     5 mm PABBO BB-
PULPROG    zg30
TD          65536
SOLVENT     CDCl3
NS          16
DS          2
SWH         8223.685      Hz
FIDRES     0.125483      Hz
AQ          3.9846387    sec
RG          80.6
DE         60.800        usec
TE         5.00          usec
TW         303.0        K
D1         1.00000000    sec
TD0        1

```

```
===== CHANNEL f1 =====  
NUC1                1H  
P1                  12.00 usec  
PL1                 -4.00 dB  
SFO1               400.1824713 MHz
```

```
F2 - Processing parameters
SI              32768
SF             400.1800078 MHz
WDW             EM
SSB              0
LB              0.30 Hz
GB              0
PC              1.00
```

PMB-isatin



```
Current Data Parameters
NAME      Dec06-2008-hayashi
EXPNO      130
PROCNO     1
```

```

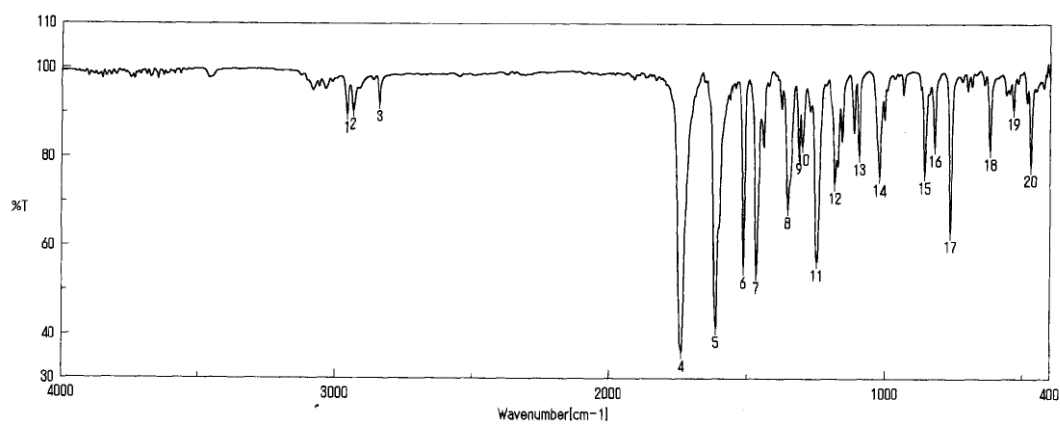
P2 - Acquisition Parameters
Date_          20081206
Time_          17.05
INSTRUM        5 mm PABBO BB-
PROBHD         spect
PULPROG        zgpg30
TD             65536
AQ             CDC13
SOLVENT        300
NS             4
DS             24038.461 Hz
SWH            0.366798 Hz
FIDRES        1.363198 sec
AQ           RG 40.3
DW            20.800 usec
DE            6.00 usec
TE           302.9 K
D1            2.0000000 sec
t1            0.0300000 sec
DELTA         1.6999999 sec
TD0           1

```

```
===== CHANNEL f1 =====
NUC1                13C
P1                  7.20 usec
PL1                 -4.00 dB
SFO1               100.6354036 MHz
```

```
===== CHANNEL f2 =====
CPDPRG2          waltz16
NUC2              1H
PCPD2             80.00 usec
PL2              -4.00 dB
PL12             15.00 dB
PL13             15.00 dB
SFO2             400.1816007 MHz
```

```
F2 - Processing parameters
SI              32768
SF              100.6253410 MHz
WDW             EM
SSB             0
LB              1.00 Hz
GB              0
PC              1.40
```

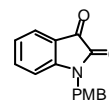



積算回数
ゼロファイリング
ゲイン
日時
測定者
ファイル名
サンプル名
コメント

64
ON
2
109/01/03 15:22
PMB isatin, JWS
background

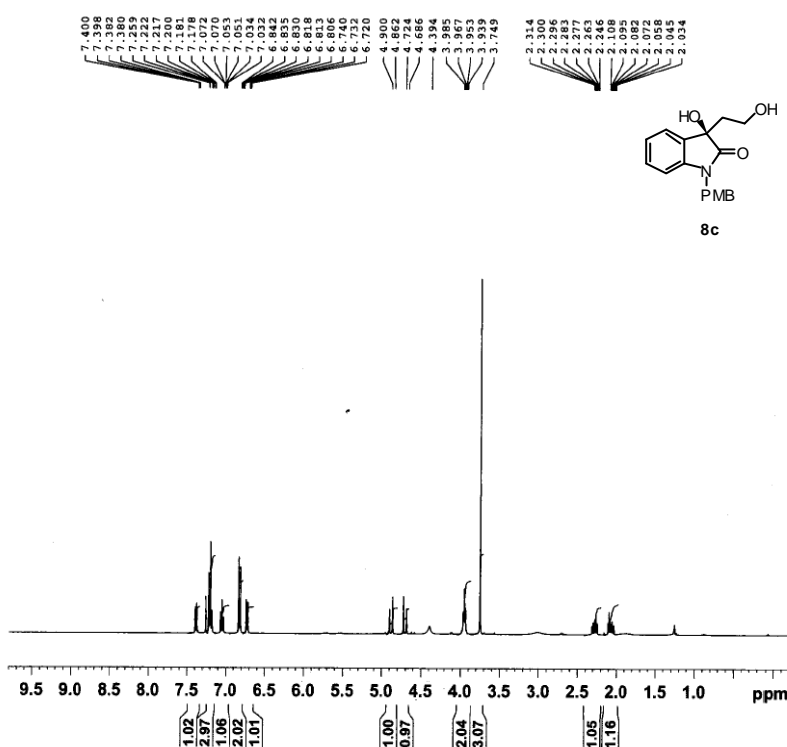
分解
アポダイゼーション
スキャンスピード

4 cm-1
Cosine
2 mm/sec



7c

1: 2980.20, 99.8313	2: 2936.09, 90.5909	3: 2840.63, 92.4719	4: 1735.62, 36.1058
5: 1610.27, 41.5858	6: 1513.85, 55.0553	7: 1487.56, 53.8085	8: 1353.78, 59.4944
9: 1314.25, 80.8667	10: 1299.79, 82.8880	11: 1248.66, 56.8697	12: 1182.15, 74.1755
13: 1094.41, 80.6432	14: 1021.12, 75.9802	15: 856.24, 76.5139	16: 819.60, 82.5785
17: 762.71, 63.2523	18: 618.07, 81.7643	19: 533.22, 90.9339	20: 470.55, 78.0972

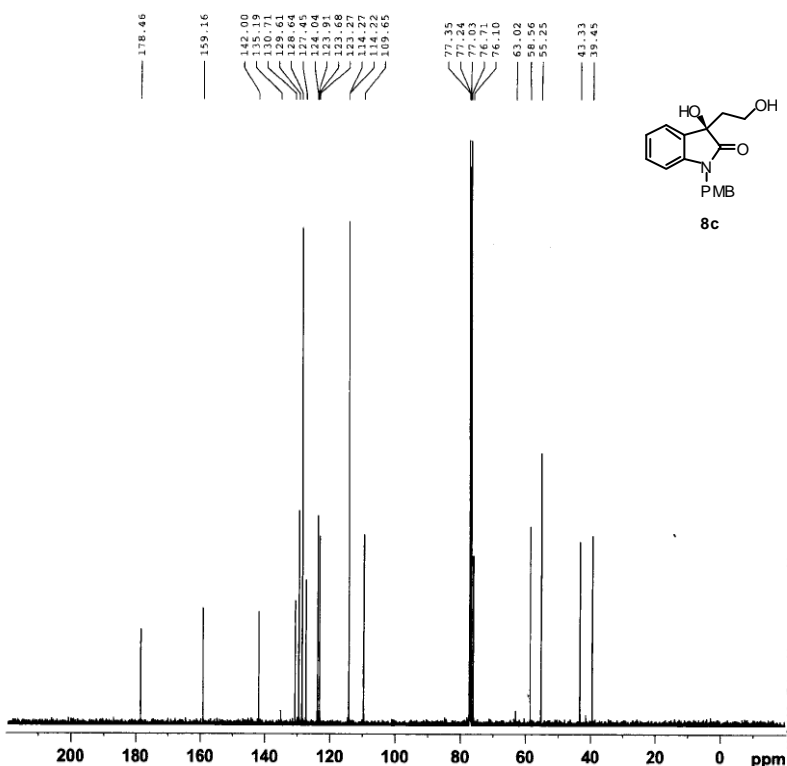


Current Data Parameters
 NAME Dec18-2008-hayashi
 EXPNO 50
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20081218
 Time 16.01
 INSTRUM spect
 PROBHD 5 mm PABBO BB-
 PULPROG zg30
 TD 65536
 SOLVENT CDCl3
 NS 16
 DS 2
 SWH 8223.685 Hz
 FIDRES 0.125483 Hz
 AQ 3.9846387 sec
 RG 144
 DW 60.800 usec
 DE 6.00 usec
 TE 303.0 K
 D1 1.00000000 sec
 TDO 1

===== CHANNEL f1 =====
 NUC1 1H
 P1 12.00 usec
 PL1 -4.00 dB
 SFO1 400.1824713 MHz

F2 - Processing parameters
 SI 32768
 SF 400.1800079 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



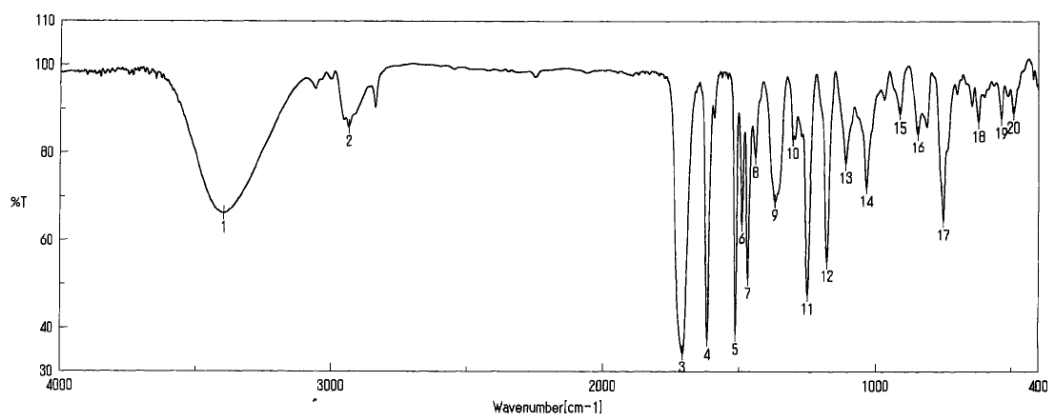
Current Data Parameters
 NAME Dec18-2008-hayashi
 EXPNO 51
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20081218
 Time 16.32
 INSTRUM spect
 PROBHD 5 mm PABBO BB-
 PULPROG zgpg30
 TD 65536
 SOLVENT CDCl3
 NS 500
 DS 4
 SWH 24038.461 Hz
 FIDRES 0.366798 Hz
 AQ 1.3631986 sec
 RG 40.3
 DW 20.800 usec
 DE 6.00 usec
 TE 302.9 K
 D1 2.00000000 sec
 d11 0.03000000 sec
 DELTA 1.89999998 sec
 TDO 1

===== CHANNEL f1 =====
 NUC1 13C
 P1 7.20 usec
 PL1 -4.00 dB
 SFO1 100.6354036 MHz

===== CHANNEL f2 =====
 CPDPRG2 waltz16
 NUC2 1H
 PCPD2 80.00 usec
 PL2 -4.00 dB
 PL12 15.00 dB
 PL13 15.00 dB
 SFO2 400.1816007 MHz

F2 - Processing parameters
 SI 32768
 SF 100.6253437 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40

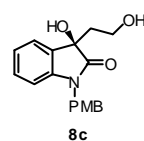


積算回数
ゼロフィリング
ゲイン
日時
測定者
ファイル名
サンプル名
コメント

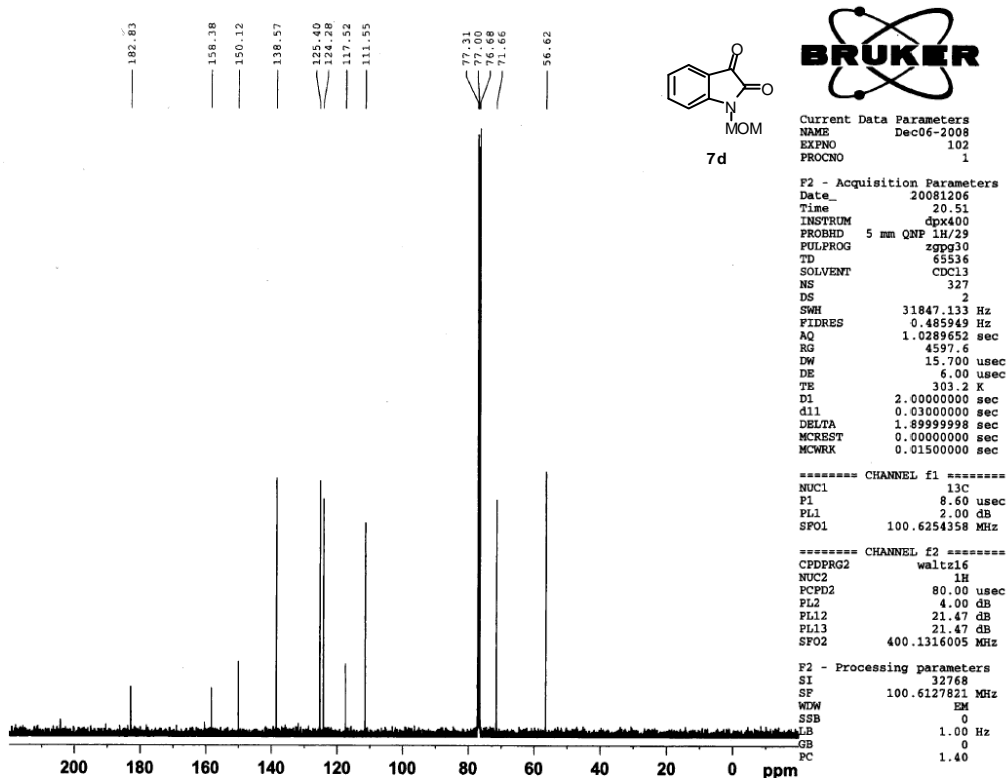
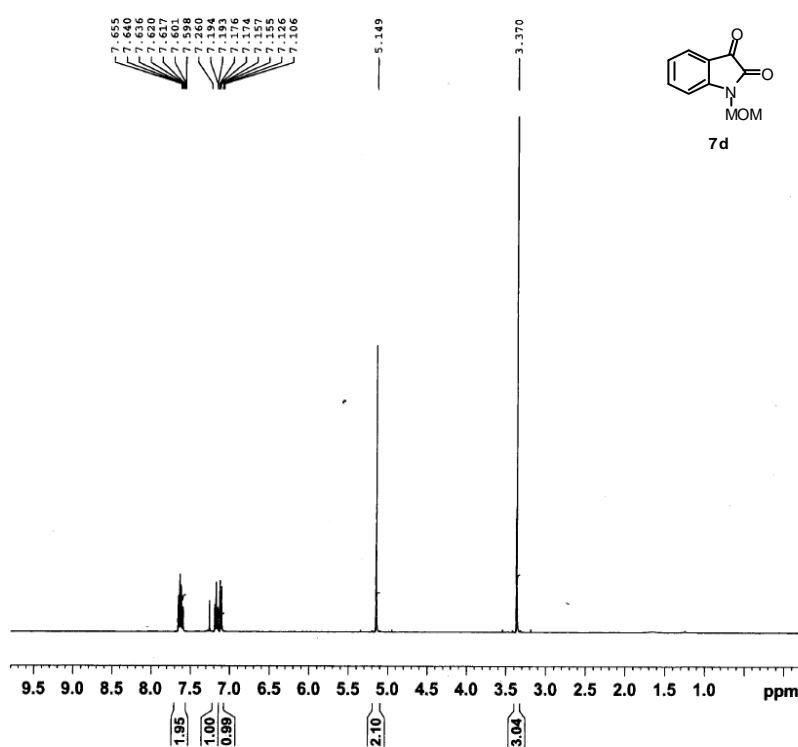
23
ON
2
108/12/24 18:53
Memory#7
background

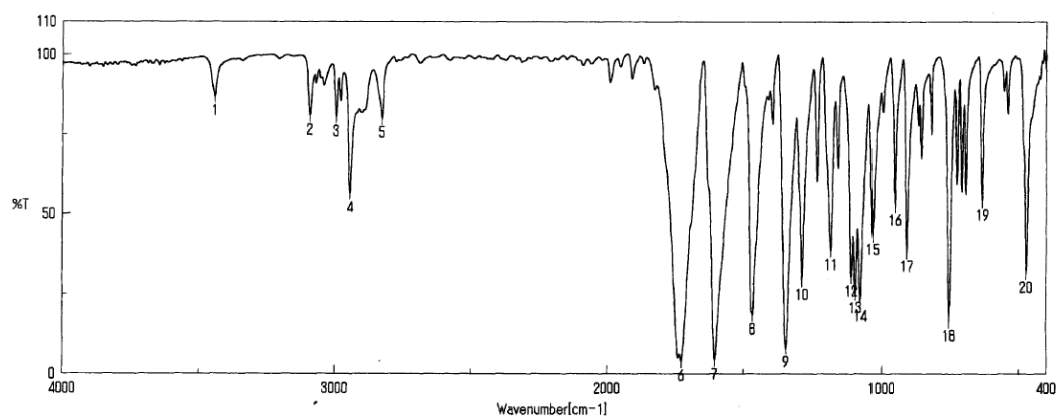
分解
アポダイゼーション
スキャンスピード

4 cm⁻¹
Cosine
2 mm/sec



1: 3395.07, 66.2394	2: 2933.20, 86.0409	3: 1704.76, 34.3685	4: 1614.13, 37.2696
5: 1513.85, 38.4710	6: 1487.81, 63.7787	7: 1467.56, 51.2527	8: 1439.60, 78.9403
9: 1367.28, 69.1035	10: 1302.68, 63.2122	11: 1248.68, 47.7100	12: 1176.36, 55.1991
13: 1108.87, 77.7164	14: 1032.69, 72.3730	15: 910.24, 89.0698	16: 843.70, 84.6343
17: 751.14, 64.6205	18: 620.00, 87.1546	19: 537.06, 87.8828	20: 490.79, 99.1436



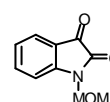


積算回数
ゼロフィリング
ゲイン
日時
測定者
ファイル名
サンプル名
コメント

64
ON
4
108/01/03 15:11
MOMisatin. JWS
background

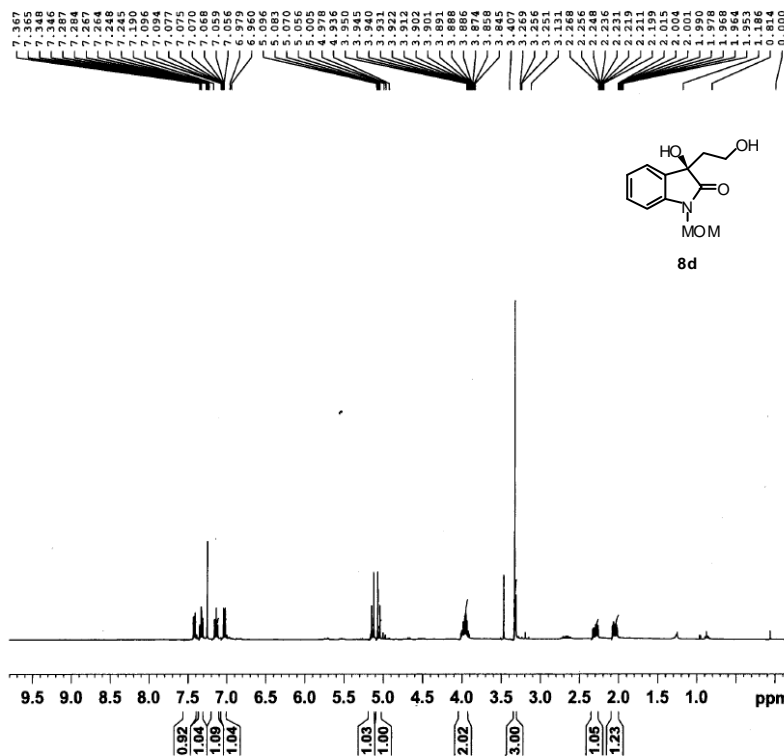
分解
アポダイゼーション
スキャンスピード

4 cm-1
Cosine
2 mm/sec



7d

1: 3443.28, 86.8512	2: 3094.23, 80.9017	3: 2996.84, 80.4734	4: 2944.77, 56.6455
5: 2828.10, 80.0723	6: 1726.94, 4.0359	7: 1605.45, 4.4528	8: 1487.58, 18.5314
9: 1346.07, 8.2906	10: 1287.25, 29.5032	11: 1183.11, 38.7052	12: 1108.87, 30.5332
13: 1094.41, 25.3041	14: 1076.08, 22.6380	15: 1031.73, 43.4538	16: 952.66, 52.6008
17: 910.24, 38.1401	18: 755.96, 16.5387	19: 634.47, 54.1929	20: 472.47, 31.7685

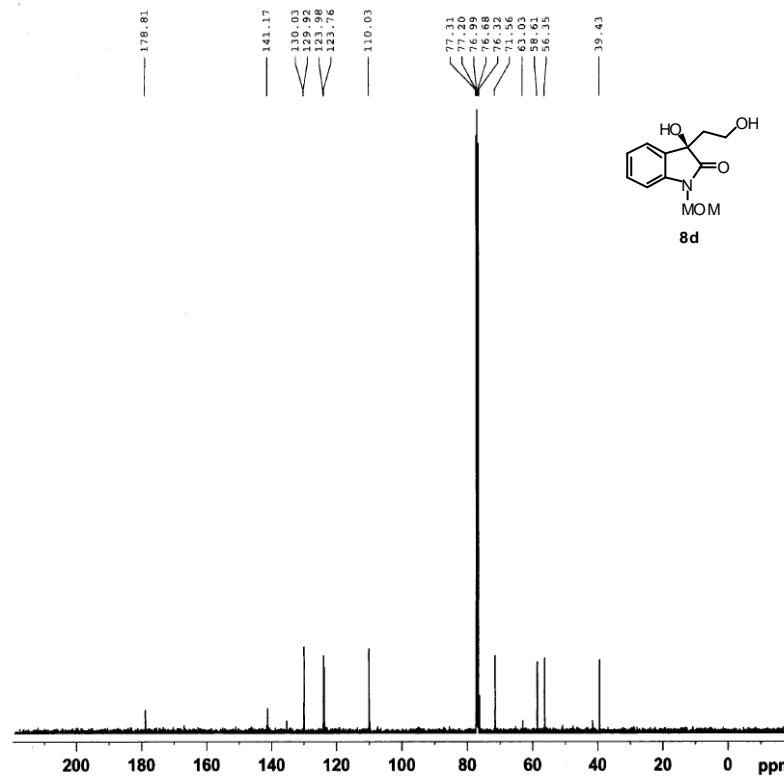


Current Data Parameters
 NAME Dec17-2008-hayashi
 EXPNO 53
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20081217
 Time 11.53
 INSTRUM spect
 PROBRD 5 mm PABBO BB-
 PULPROG zg30
 TD 65536
 SOLVENT CDCl3
 NS 32
 DS 2
 SWH 8223.685 Hz
 FIDRES 0.125483 Hz
 AQ 3.9846387 sec
 RG 256
 DW 60.800 usec
 DE 6.00 usec
 TE 303.0 K
 D1 1.00000000 sec
 TD0 1

===== CHANNEL f1 =====
 NUC1 1H
 P1 12.00 usec
 PL1 -4.00 dB
 SFO1 400.1824713 MHz

F2 - Processing parameters
 SI 32768
 SF 400.1800092 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



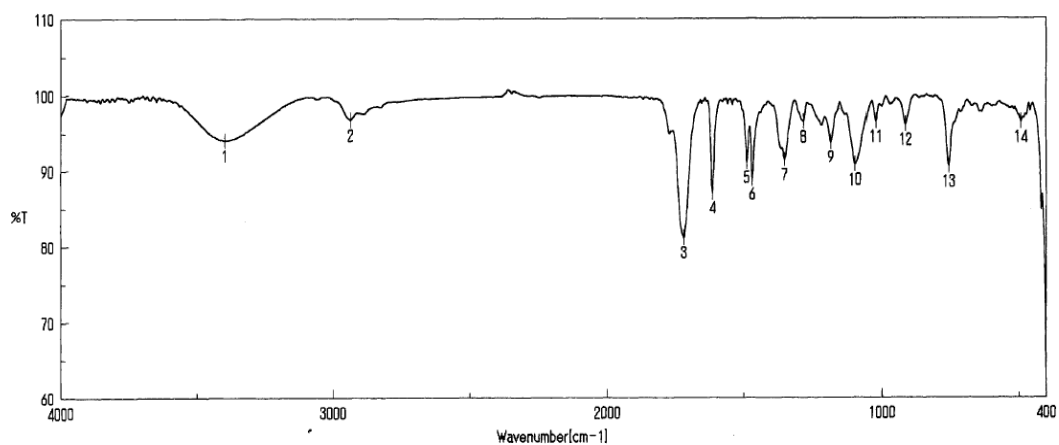
Current Data Parameters
 NAME Dec17-2008-hayashi
 EXPNO 54
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20081217
 Time 12.23
 INSTRUM spect
 PROBRD 5 mm PABBO BB-
 PULPROG zgpg30
 TD 65536
 SOLVENT CDCl3
 NS 500
 DS 4
 SWH 24038.461 Hz
 FIDRES 0.366798 Hz
 AQ 1.3631988 sec
 RG 114
 DW 20.800 usec
 DE 6.00 usec
 TE 302.9 K
 D1 2.00000000 sec
 d11 0.03000000 sec
 DELTA 1.89999998 sec
 TD0 1

===== CHANNEL f1 =====
 NUC1 13C
 P1 7.20 usec
 PL1 -4.00 dB
 SFO1 100.6354036 MHz

===== CHANNEL f2 =====
 CPDPRG2 waltz16
 NUC2 1H
 PCPD2 80.00 usec
 PL2 -4.00 dB
 PL12 15.00 dB
 PL13 15.00 dB
 SFO2 400.1816007 MHz

F2 - Processing parameters
 SI 32768
 SF 100.6253424 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40

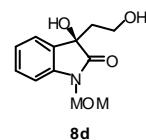


積算回数
ゼロファイリング
ゲイン
日時
測定者
ファイル名
サンプル名
コメント

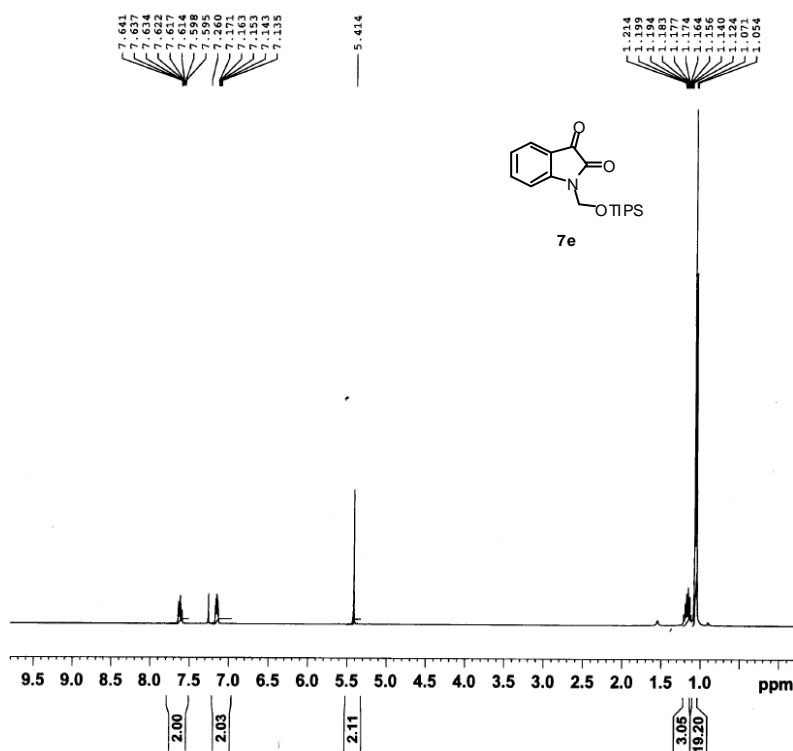
21
ON
2
108/12/24 18:34
Memory#3
background

分解
アボダイゼーション
スキャンスピード

4 cm⁻¹
Cosine
2 mm/sec



1: 3399.89, 94.0082	2: 2936.09, 96.7471	3: 1718.26, 81.1851	4: 1614.13, 87.1222
5: 1487.81, 91.1412	6: 1468.53, 88.9544	7: 1350.89, 91.4164	8: 1281.47, 96.4765
9: 1182.15, 93.7824	10: 1094.41, 90.8186	11: 1021.12, 96.5099	12: 913.13, 96.1381
13: 755.96, 90.5991	14: 492.72, 96.4407		

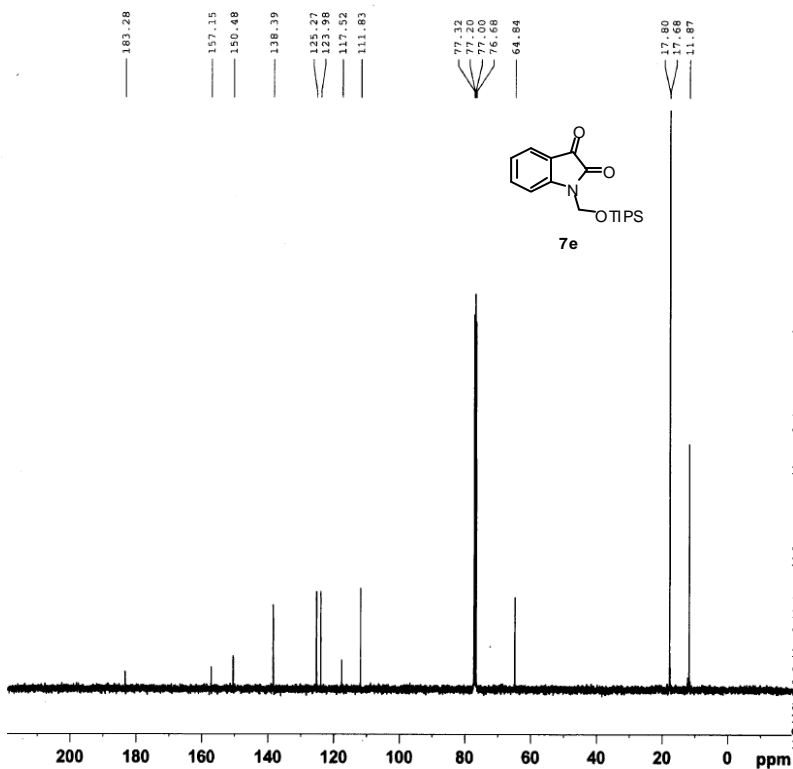


Current Data Parameters
NAME Dec17-2008-hayashi
EXPNO 30
PROCNO 1

F2 - Acquisition Parameters
Date_ 20081217
Time 10.32
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zg30
TD 65536
SOLVENT CDC13
NS 32
DS 2
SWH 8223.685 Hz
FIDRES 0.125483 Hz
AQ 3.9846387 sec
RG 161
DW 60.800 usec
DE 6.00 usec
TE 303.0 K
D1 1.0000000 sec
TDO 1

===== CHANNEL f1 =====
NUC1 1H
P1 12.00 usec
PL1 -4.00 dB
SFO1 400.1824713 MHz

F2 - Processing parameters
SI 32768
SF 400.1800075 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00



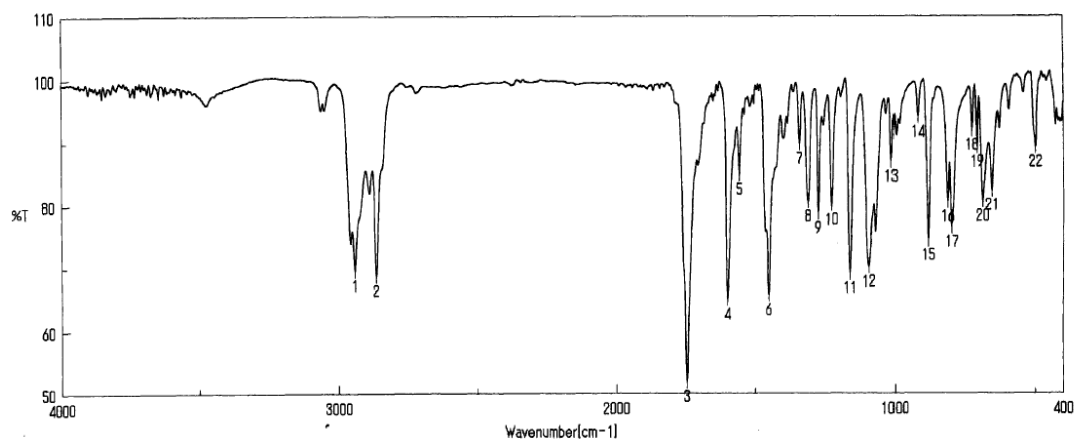
Current Data Parameters
NAME Dec17-2008-hayashi
EXPNO 31
PROCNO 1

F2 - Acquisition Parameters
Date_ 20081217
Time 10.51
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zgpg30
TD 65536
SOLVENT CDC13
NS 300
DS 4
SWH 24038.461 Hz
FIDRES 0.366798 Hz
AQ 1.3631988 sec
RG 203
DW 20.800 usec
DE 6.00 usec
TE 302.9 K
D1 2.0000000 sec
d11 0.0300000 sec
DELTA 1.89999998 sec
TDO 1

===== CHANNEL f1 =====
NUC1 13C
P1 7.20 usec
PL1 -4.00 dB
SFO1 100.6354036 MHz

===== CHANNEL f2 =====
CPDPRG2 waltz16
NUC2 1H
PCPD2 80.00 usec
PL2 -4.00 dB
PL12 15.00 dB
PL13 15.00 dB
SFO2 400.1816007 MHz

F2 - Processing parameters
SI 32768
SF 100.6253420 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

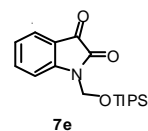


積算回数
ゼロフィリング
ゲイン
日時
測定者
ファイル名
サンプル名
コメント

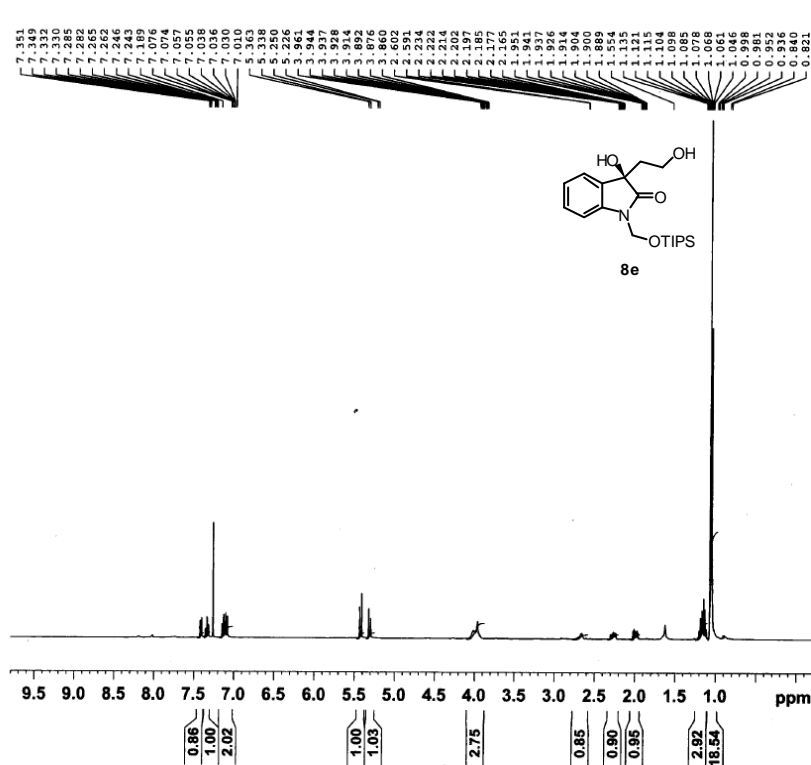
23
ON
2
108/12/24 19:06
Memory#11
background

分解
アポダイゼーション
スキャンスピード

4 cm-1
Cosine
2 mm/sec



1: 2941.68, 68.6614	2: 2885.70, 68.8611	3: 1746.23, 51.9457	4: 1600.63, 65.0892
5: 1559.17, 85.0539	6: 1452.14, 65.7717	7: 1343.18, 89.8150	8: 1311.36, 80.6876
9: 1274.72, 78.8715	10: 1226.51, 80.1445	11: 1159.97, 69.2797	12: 1082.48, 70.2863
13: 1013.41, 87.0269	14: 919.86, 94.1106	15: 853.24, 74.5372	16: 812.85, 80.6181
17: 799.35, 76.6129	18: 726.07, 92.0652	19: 707.75, 89.3292	20: 688.46, 80.7240

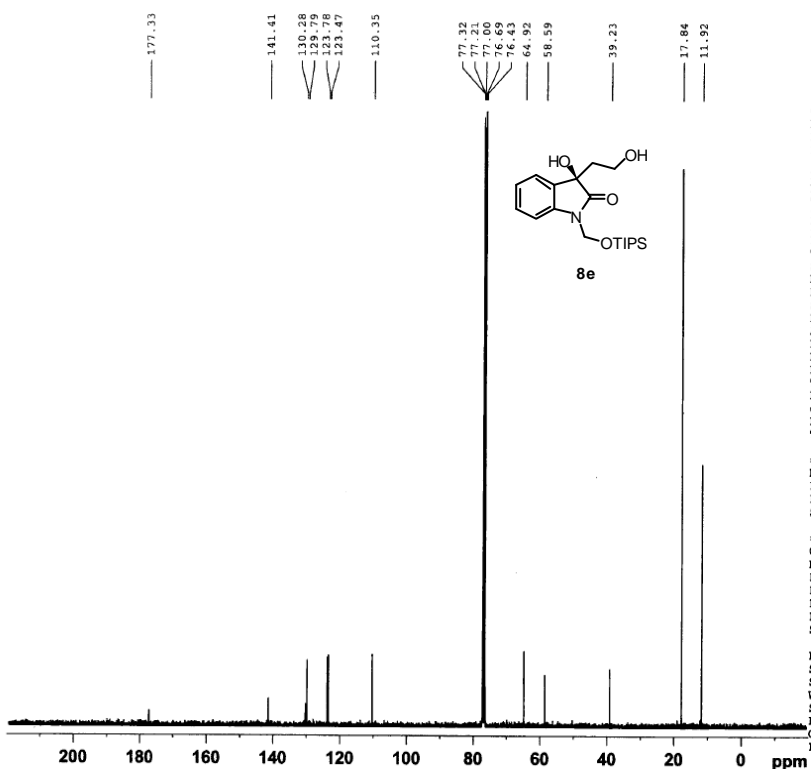


Current Data Parameters
 NAME Dec26-2008-hayashi
 EXPNO 10
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20081226
 Time 13.26
 INSTRUM spect
 PROBHD 5 mm PABBO BB-
 PULPROG zg30
 TD 65536
 SOLVENT CDCl3
 NS 16
 DS 2
 SWH 8223.685 Hz
 FIDRES 0.125483 Hz
 AQ 3.9846387 sec
 RG 181
 DW 60.800 usec
 DE 5.00 usec
 TE 303.0 K
 D1 1.00000000 sec
 TDO 1

===== CHANNEL f1 =====
 NUC1 1H
 P1 12.00 usec
 PL1 -4.00 dB
 SFO1 400.1824713 MHz

F2 - Processing parameters
 SI 32768
 SF 400.1800073 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



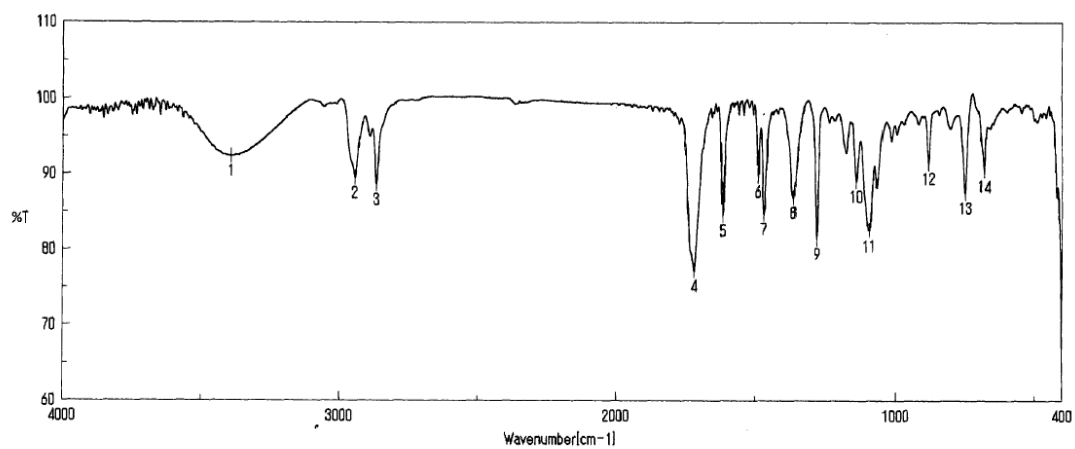
Current Data Parameters
 NAME Dec26-2008-hayashi
 EXPNO 11
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20081226
 Time 13.57
 INSTRUM spect
 PROBHD 5 mm PABBO BB-
 PULPROG zgpg30
 TD 65536
 SOLVENT CDCl3
 NS 500
 DS 4
 SWH 24038.461 Hz
 FIDRES 0.366798 Hz
 AQ 1.3631988 sec
 RG 228
 DW 20.800 usec
 DE 6.00 usec
 TE 302.9 K
 D1 2.00000000 sec
 d11 0.03000000 sec
 DELTA 1.89999998 sec
 TDO 1

===== CHANNEL f1 =====
 NUC1 13C
 P1 7.20 usec
 PL1 -4.00 dB
 SFO1 100.6354036 MHz

===== CHANNEL f2 =====
 CPDPRG2 waltz16
 NUC2 1H
 PCPD2 80.00 usec
 PL2 -4.00 dB
 PL12 15.00 dB
 PL13 15.00 dB
 SFO2 400.1816007 MHz

F2 - Processing parameters
 SI 32768
 SF 100.6253435 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40

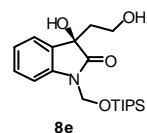


積算回数
ゼロファイリング
ゲイン
日時
測定者
ファイル名
サンプル名
コメント

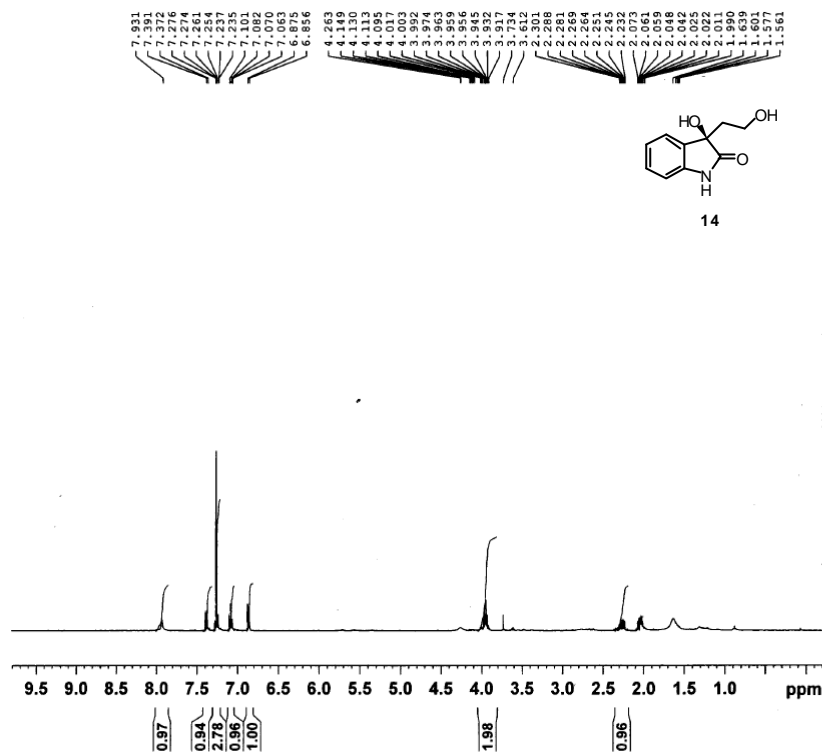
16
ON
1
109/03/09 20:44
Memory#5
background

分解
アボダイゼーション
スキャンスピード

4 cm⁻¹
Cosine
2 mm/sec



1: 3392.17,	92.4051	2: 2944.77,	89.5829	3: 2886.67,	88.6778	4: 1718.26,	77.2375
5: 1615.09,	84.5267	6: 1468.78,	89.7725	7: 1469.49,	84.7470	8: 1364.39,	86.9990
9: 1279.54,	81.5174	10: 1139.72,	89.1043	11: 1092.48,	82.5320	12: 882.27,	91.4709
13: 751.14,	87.6203	14: 682.68,	90.4104				

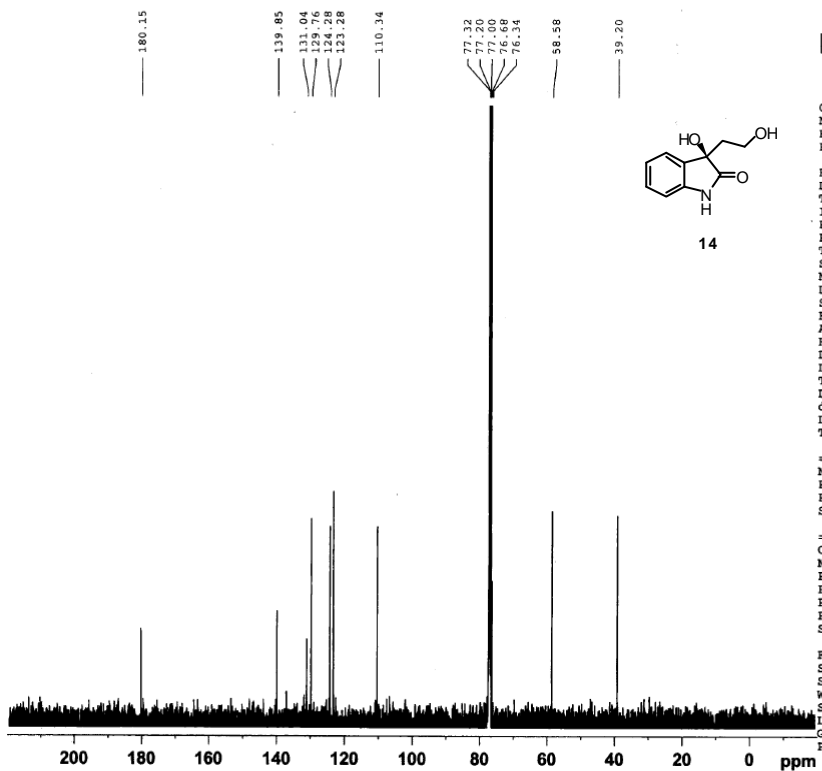


Current Data Parameters
NAME Mar21-2009
EXPNO 81
PROCNO 1

F2 - Acquisition Parameters
Date_ 20090321
Time 19.48
INSTRUM dpx400
PROBHD 5 mm QNP 1H/29
PULPROG zg30
TD 32768
SOLVENT CDCl3
NS 8
DS 0
SWH 8223.685 Hz
FIDRES 0.250967 Hz
AQ 1.9923444 sec
RG 5160.6
DW 60.800 usec
DE 6.00 usec
TE 303.2 K
D1 1.00000000 sec
MCREST 0.00000000 sec
MCWRK 0.01500000 sec

===== CHANNEL f1 =====
NUC1 1H
P1 10.70 usec
PL1 4.00 dB
SFO1 400.1324710 MHz

F2 - Processing parameters
SI 16384
SF 400.1300092 MHz
WDW no
SSB 0
LB 0.00 Hz
GB 0
PC 1.00



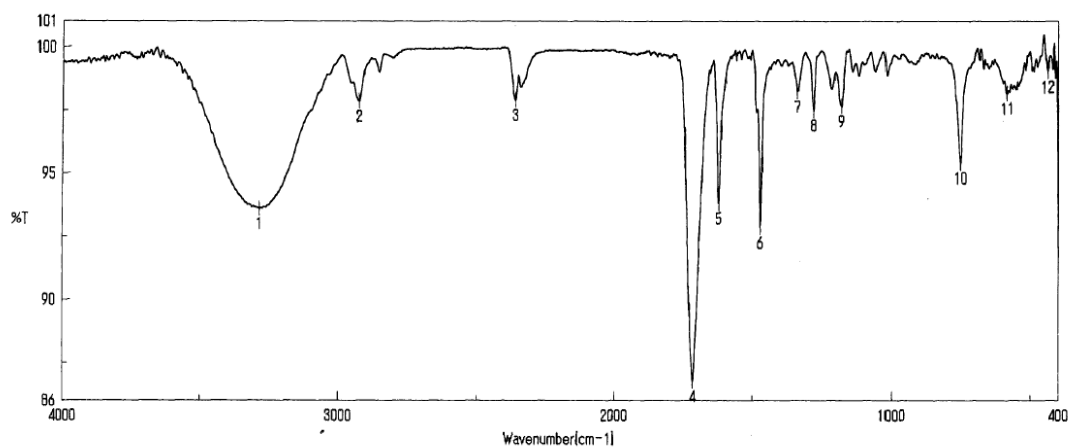
Current Data Parameters
NAME Mar21-2009-hayashi
EXPNO 110
PROCNO 1

F2 - Acquisition Parameters
Date_ 20090321
Time 21.52
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zgpg30
TD 65536
SOLVENT CDCl3
NS 2000
DS 4
SWH 24038.461 Hz
FIDRES 0.366798 Hz
AQ 1.3631988 sec
RG 114
DW 20.800 usec
DE 6.00 usec
TE 302.9 K
D1 2.00000000 sec
d11 0.03000000 sec
DELTA 1.89999998 sec
TD0 1

===== CHANNEL f1 =====
NUC1 13C
P1 7.20 usec
PL1 4.00 dB
SFO1 100.6354036 MHz

===== CHANNEL f2 =====
CPDPRG2 waltz16
NUC2 1H
PCPD2 80.00 usec
PL2 -4.00 dB
PL12 15.00 dB
PL13 15.00 dB
SFO2 400.1816007 MHz

F2 - Processing parameters
SI 32768
SF 100.6253405 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

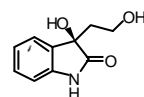


積算回数
ゼロフィリング
ゲイン
日時
測定者
ファイル名
サンプル名
コメント

16
ON
1
109/03/23 14:57
Memory#3
background

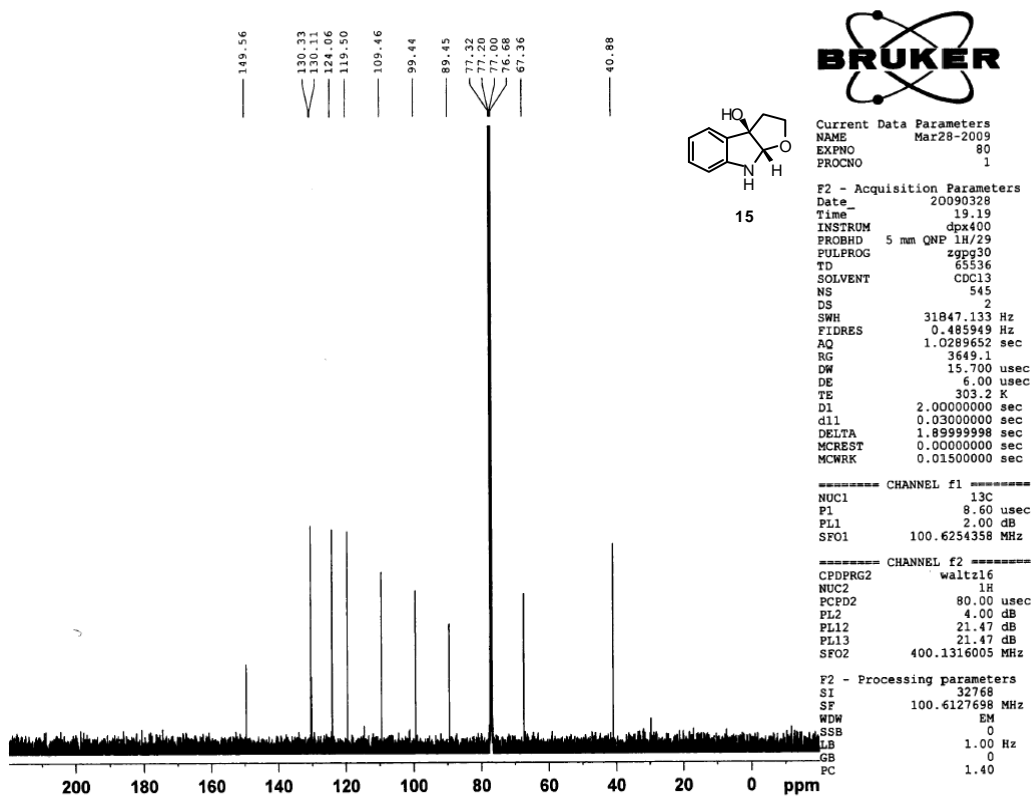
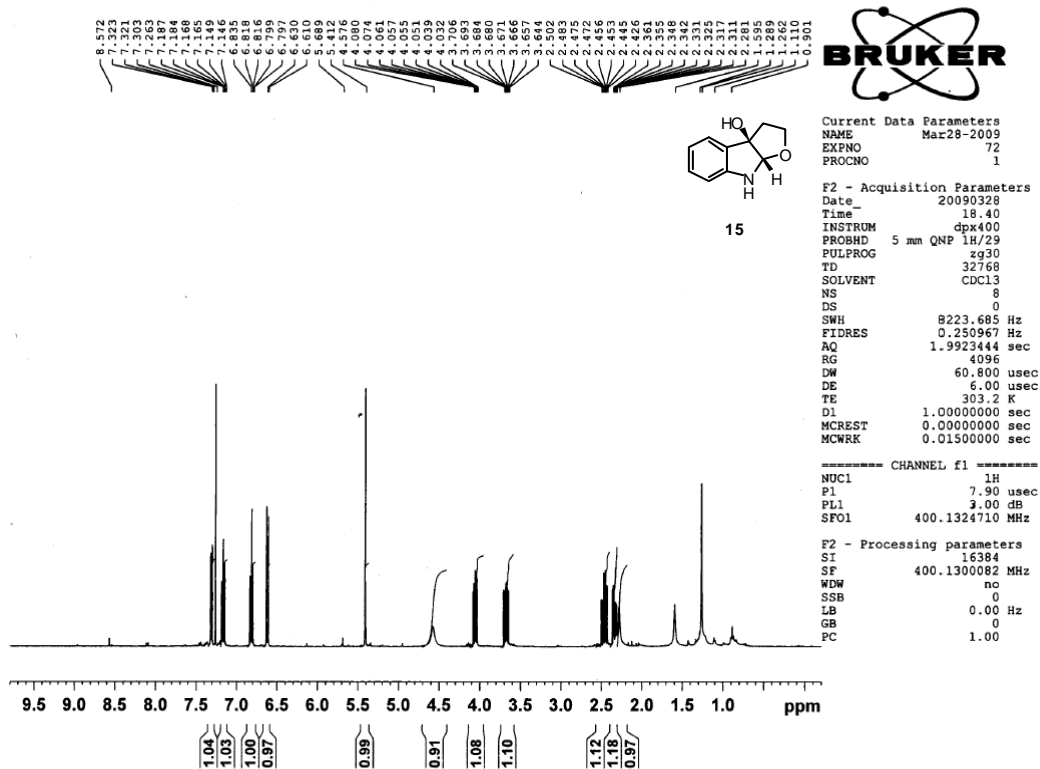
分解
アポダイゼーション
スキャンスピード

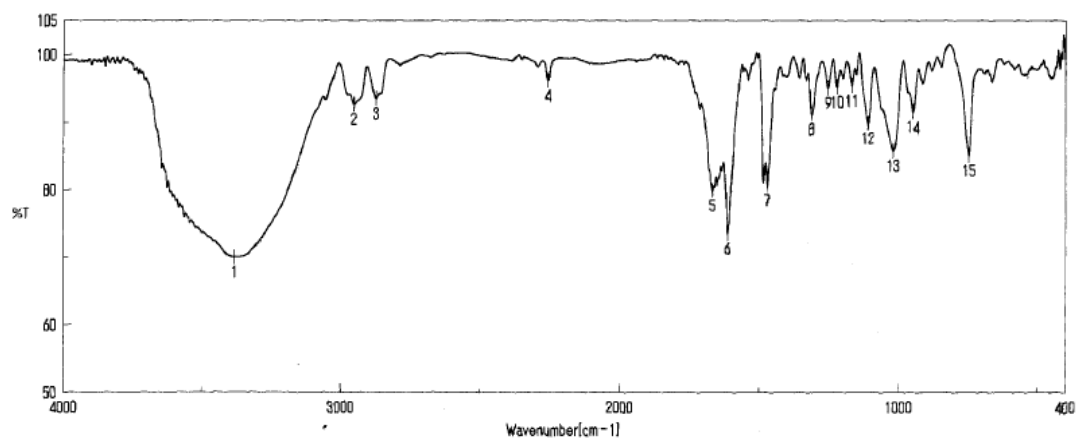
4 cm-1
Cosine
2 cm/sec



14

1: 3284.18, 93.6011	2: 2923.56, 97.8678	3: 2358.52, 97.9053	4: 1715.37, 86.7861
5: 1621.84, 93.8252	6: 1471.42, 92.8601	7: 1337.39, 98.2586	8: 1279.54, 97.5109
9: 1178.28, 97.6699	10: 753.07, 95.4080	11: 584.33, 98.1779	12: 437.76, 99.0196



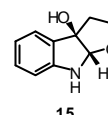


積算回数
ゼロファイリング
ゲイン
日時
測定者
ファイル名
サンプル名
コメント

32
ON
2
109/04/23 18:43
Memory#5
background

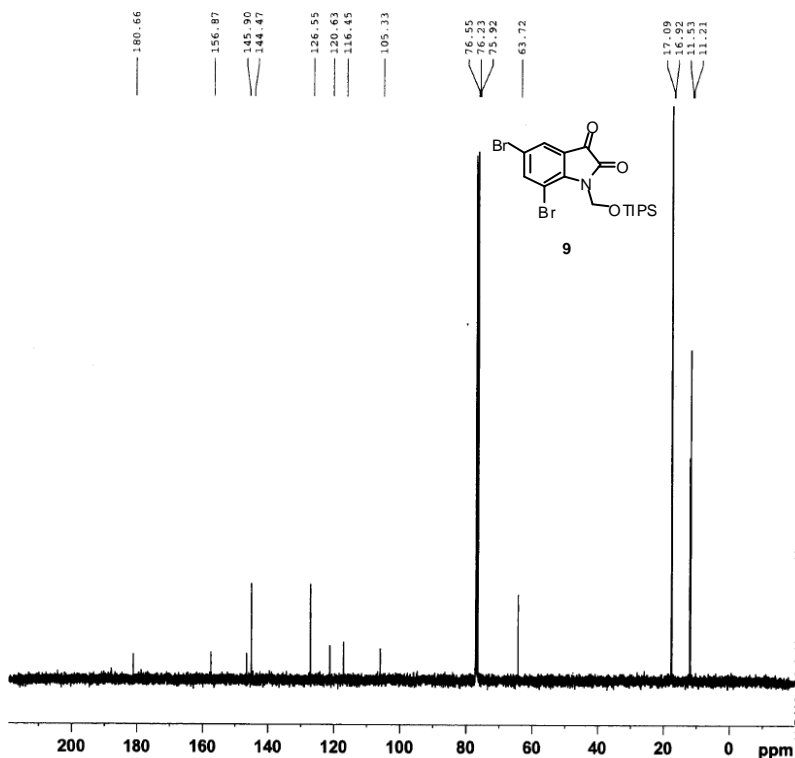
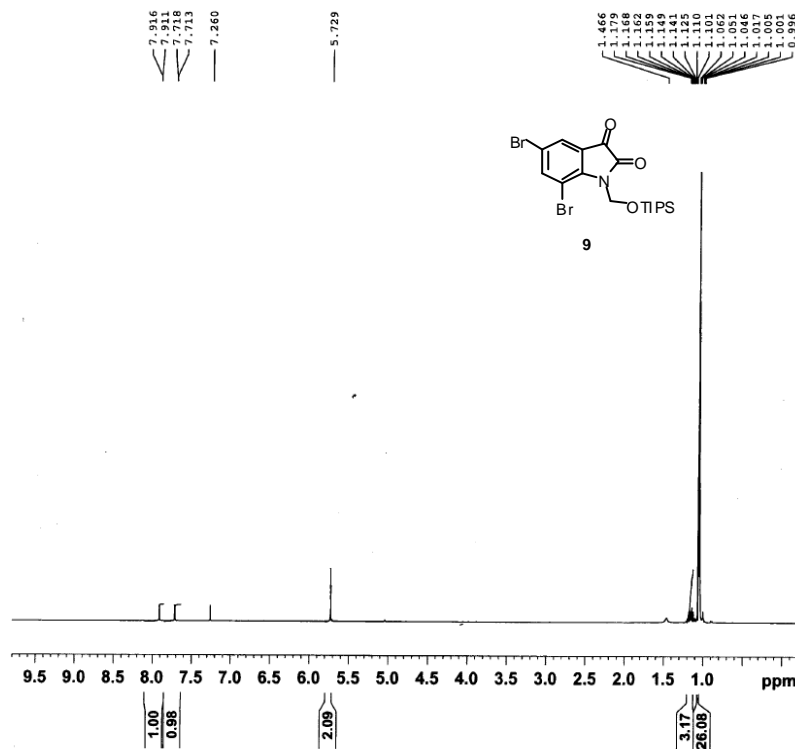
分解
アポダイゼーション
スキャンスピード

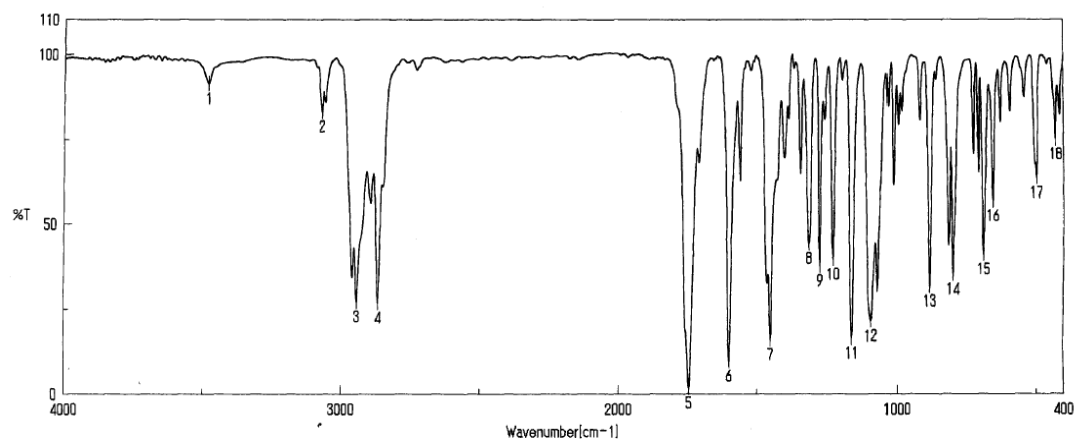
4 cm⁻¹
Coarse
2 mm/sec



15

1: 3382.53, 70.0549	2: 2952.48, 92.7123	3: 2873.42, 93.5309	4: 2255.34, 98.2442
5: 1898.12, 79.9558	6: 1613.16, 73.5297	7: 1471.42, 80.7052	8: 1313.29, 91.2658
9: 1255.43, 95.1091	10: 1223.61, 95.2541	11: 1199.62, 95.4512	12: 1111.76, 89.8875
13: 1021.12, 85.8415	14: 949.77, 91.5903	15: 747.28, 85.1480	



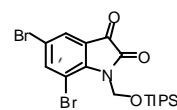


積算回数
ゼロフィリング
ゲイン
日時
測定者
ファイル名
サンプル名
コメント

64
ON
2
109/01/03 15:06
diBrisatin (OTIPS) not natural. JWS
background

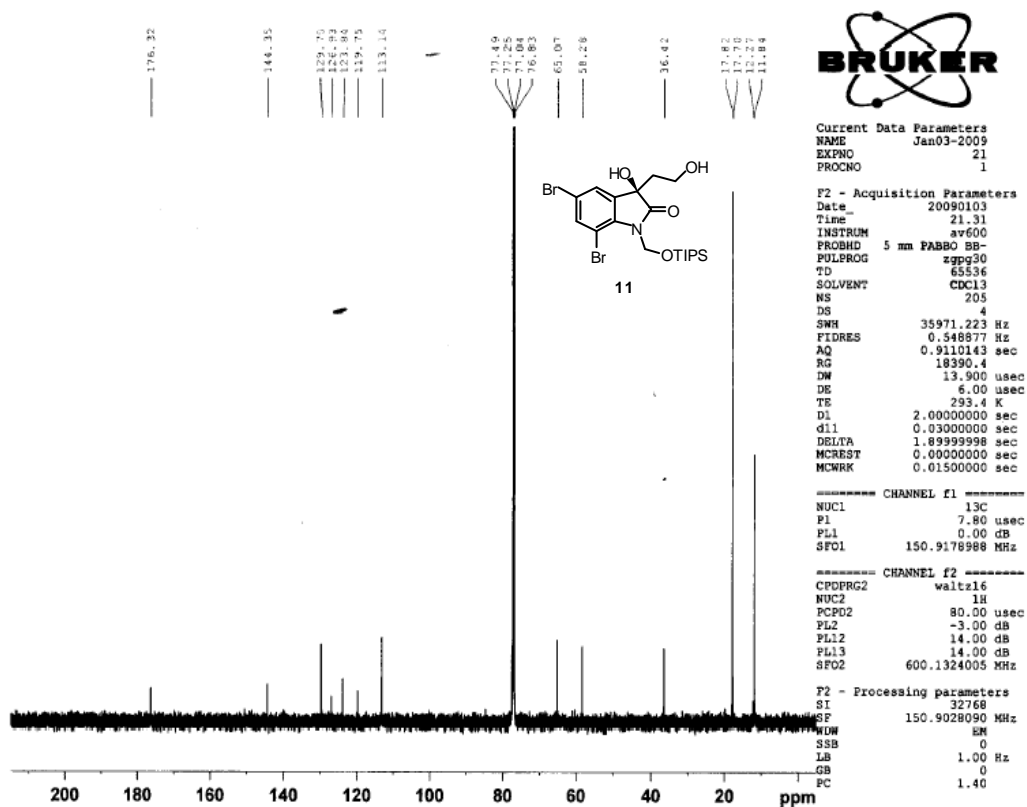
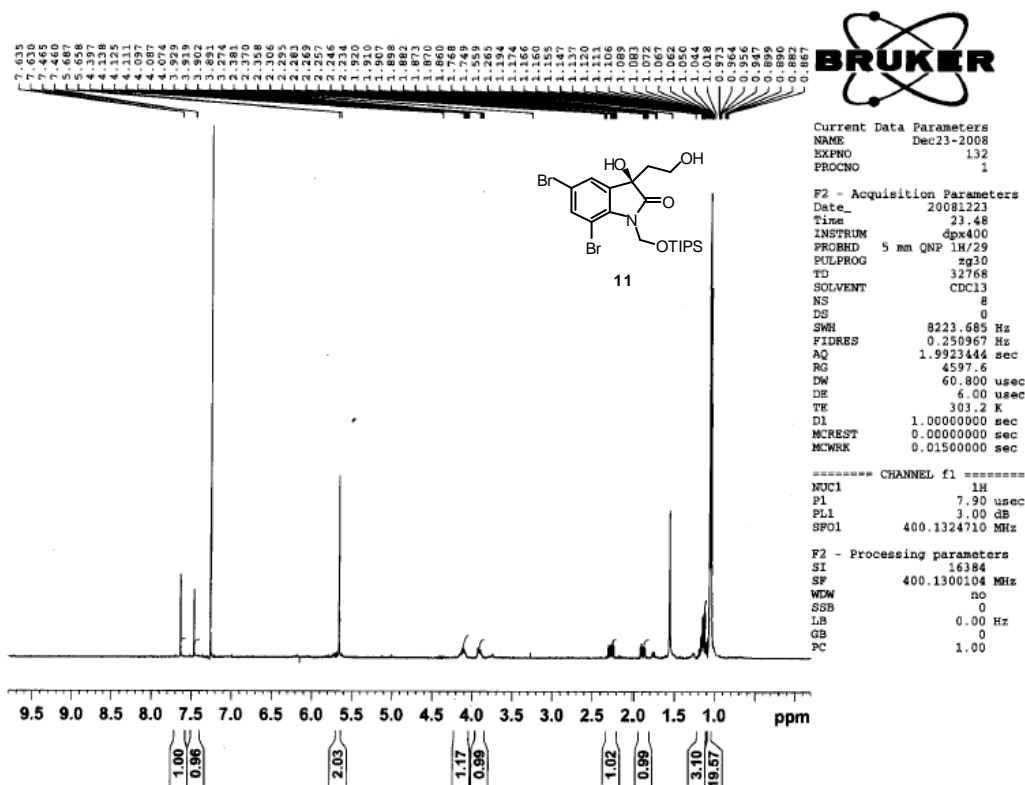
分解
アポダイゼーション
スキャンスピード

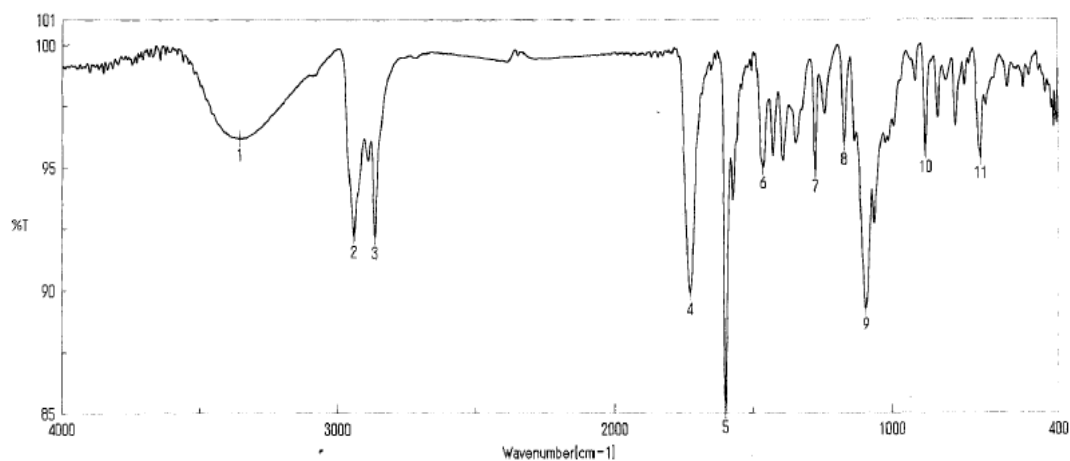
4 cm-1
Cosine
2 mm/sec



9

1: 3476.06, 91.4517	2: 3066.26, 83.2333	3: 2941.88, 27.3585	4: 2864.74, 27.0436
5: 1746.23, 1.8945	6: 1600.63, 9.7920	7: 1452.14, 16.2026	8: 1313.29, 44.5592
9: 1274.72, 37.7887	10: 1227.47, 39.7451	11: 1160.94, 16.6128	12: 1092.48, 21.6006
13: 884.20, 31.9686	14: 800.31, 35.3972	15: 689.43, 41.2033	16: 656.64, 56.9665
17: 499.47, 63.8390	18: 429.08, 75.1549		

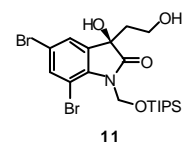




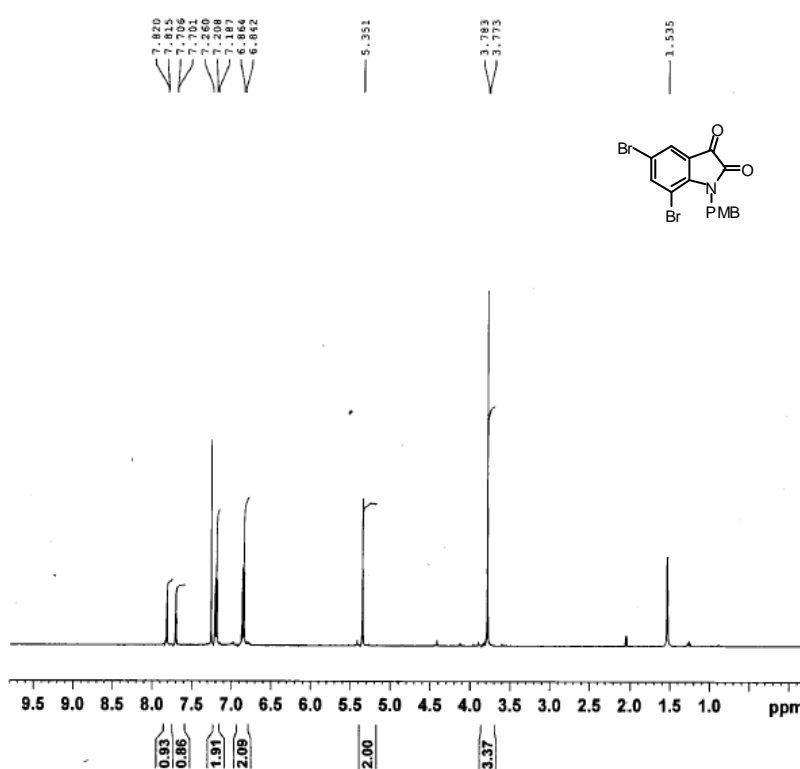
積算回数 64
 ゼロフィリング ON
 ゲイン 2
 日時 108/01/03 23:06
 測定者
 ファイル名 Memory#3
 サンプル名 background
 コメント

分解
 アポダイゼーション
 スキャンスピード

4 cm⁻¹
 Cosine
 2 mm/sec



1: 3356.50, 96.1773	2: 2941.88, 92.2225	3: 2864.74, 92.1813	4: 1725.98, 89.8775
5: 1598.70, 85.0725	6: 1464.67, 95.0076	7: 1278.57, 94.8629	8: 1174.44, 95.9840
9: 1093.44, 89.2615	10: 881.31, 95.6806	11: 683.64, 95.3996	

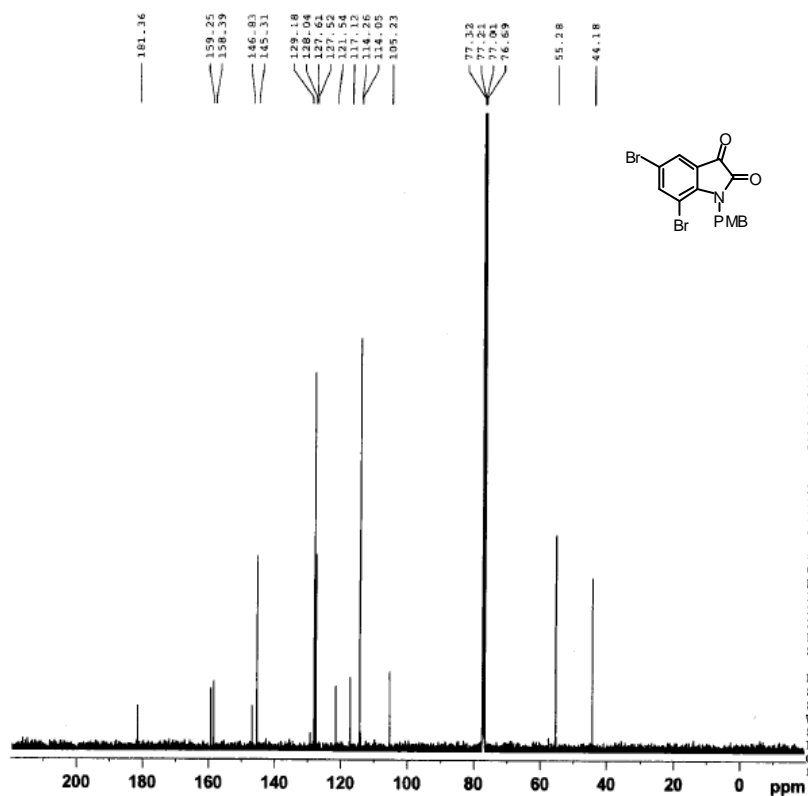


Current Data Parameters
 NAME Apr23-2009
 EXPNO 114
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20090423
 Time 19.02
 INSTRUM dpx400
 PROBHD 5 mm QNP 1H/29
 PULPROG zg30
 TD 32768
 SOLVENT CDCl3
 NS 8
 DS 0
 SWH 8223.685 Hz
 FIDRES 0.250967 Hz
 AQ 1.9923444 sec
 RG 10321.3
 DW 60.800 usec
 DE 5.00 usec
 TE 303.2 K
 D1 1.00000000 sec
 MCREST 0.00000000 sec
 MCWRC 0.01500000 sec

===== CHANNEL f1 =====
 NUC1 1H
 P1 10.70 usec
 PL1 4.00 dB
 SFO1 400.1324710 MHz

F2 - Processing parameters
 SI 16384
 SF 400.1300092 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



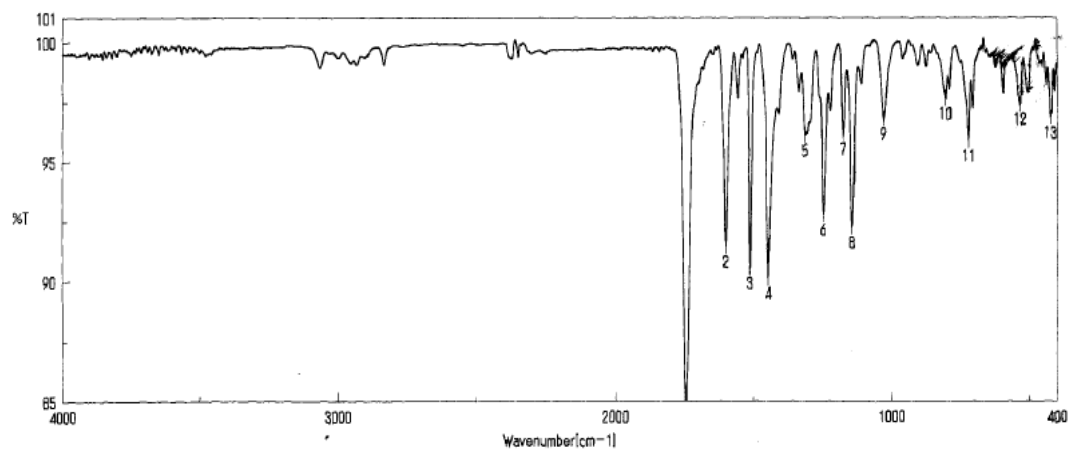
Current Data Parameters
 NAME Apr23-2009-hayaishi
 EXPNO 150
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20090423
 Time 19.20
 INSTRUM spect
 PROBHD 5 mm PABBO BB-
 PULPROG zgpg30
 TD 65536
 SOLVENT CDCl3
 NS 1024
 DS 4
 SWH 24038.461 Hz
 FIDRES 0.366798 Hz
 AQ 1.3631988 sec
 RG 228
 DW 20.800 usec
 DE 6.00 usec
 TE 302.9 K
 D1 2.00000000 sec
 d11 0.03000000 sec
 DELTA 1.89999999 sec
 TD0 1

===== CHANNEL f1 =====
 NUC1 13C
 P1 7.20 usec
 PL1 -3.50 dB
 SFO1 100.6354036 MHz

===== CHANNEL f2 =====
 CPDPRG2 waltz16
 NUC2 1H
 PCPD2 80.00 usec
 PL2 -4.00 dB
 PL12 14.50 dB
 PL13 14.50 dB
 SFO2 400.1816007 MHz

F2 - Processing parameters
 SI 32768
 SF 100.6253411 MHz
 WDW HM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40

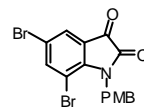


積算回数
ゼロファイリング
ゲイン
日時
測定者
ファイル名
サンプル名
コメント

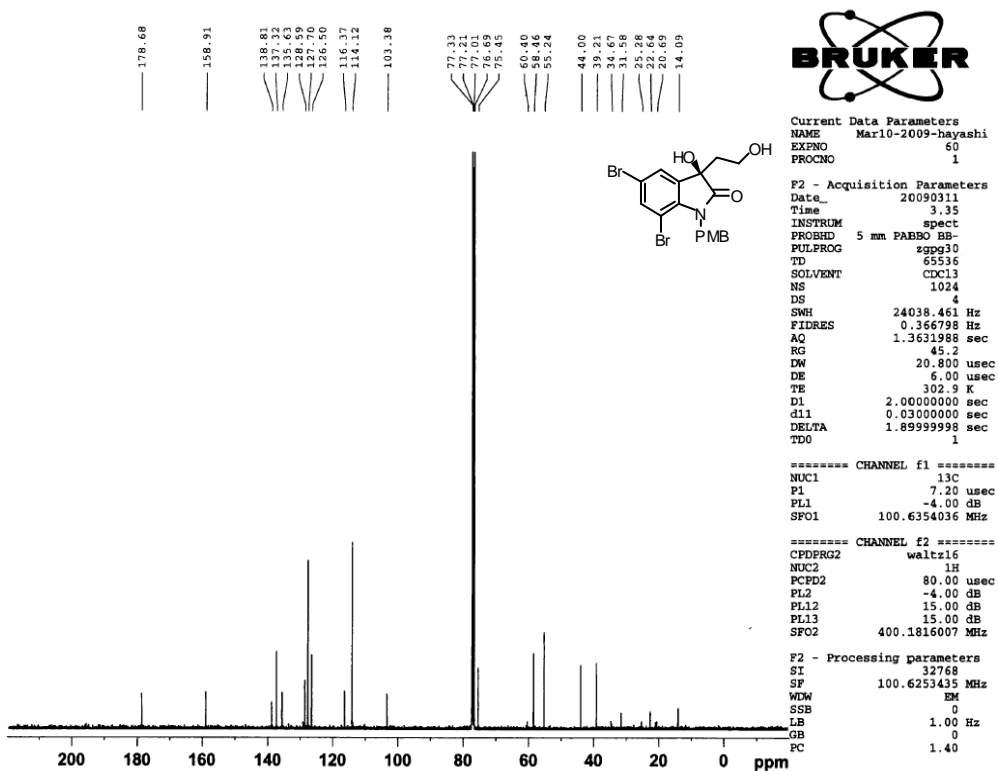
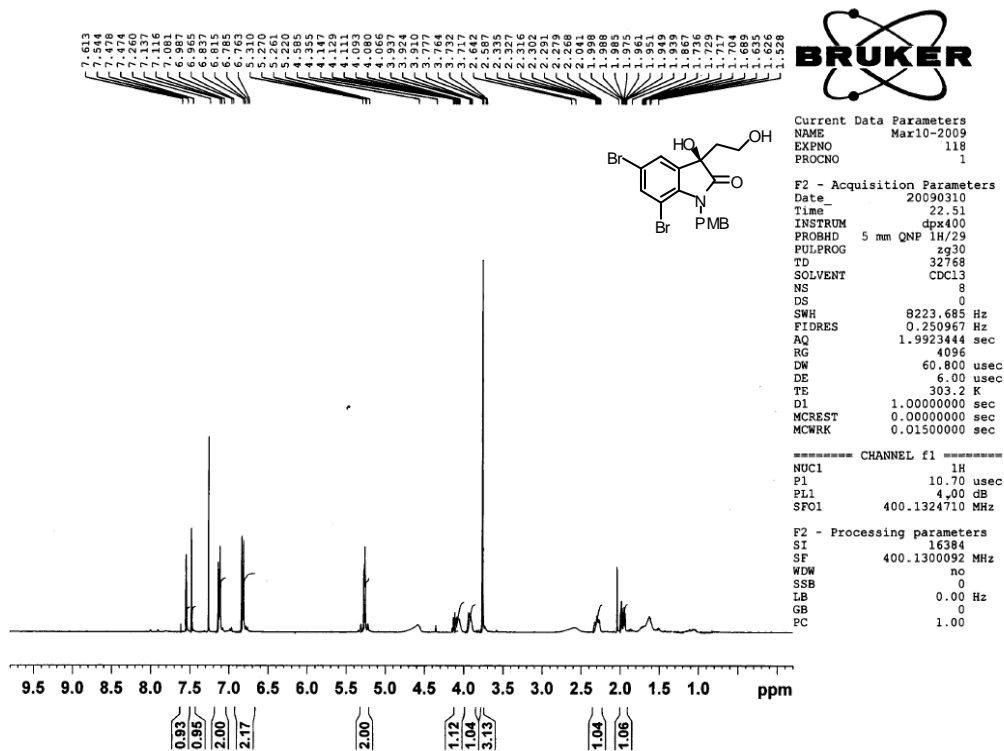
32
ON
2
109/04/23 18:38
Memory#3
background

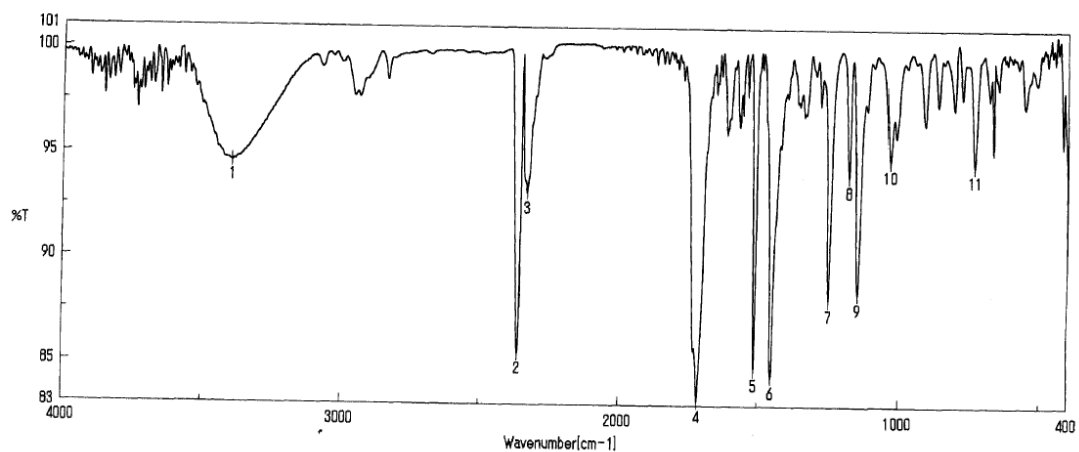
分解
アポダイゼーション
スキャンスピード

4 cm-1
Cosine
2 mm/sec



1: 1744.30, 85.0089	2: 1600.83, 91.4791	3: 1513.85, 90.5878	4: 1447.31, 90.1978
5: 1318.21, 96.1531	6: 1247.72, 92.8201	7: 1178.29, 98.2131	8: 1144.55, 92.2825
9: 1031.73, 96.8260	10: 808.03, 87.8855	11: 724.14, 95.9124	12: 537.06, 87.4485
13: 425.23, 96.8853			



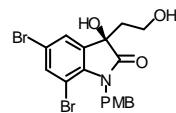


積算回数
ゼロファイリング
ゲイン
日時
測定者
ファイル名
サンプル名
コメント

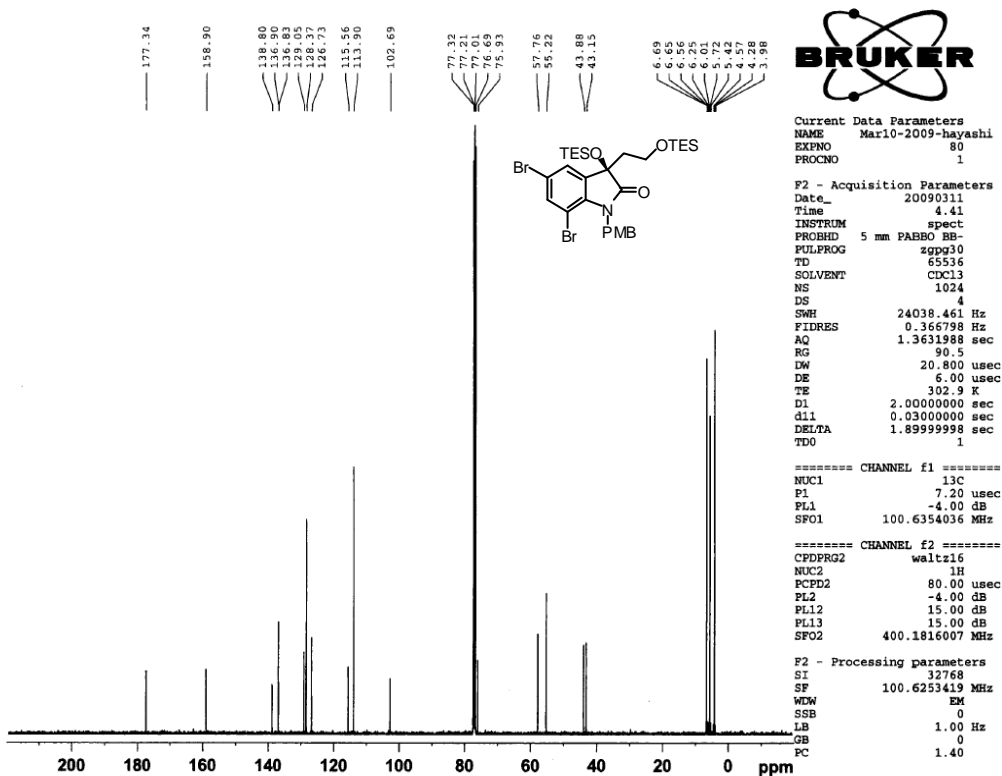
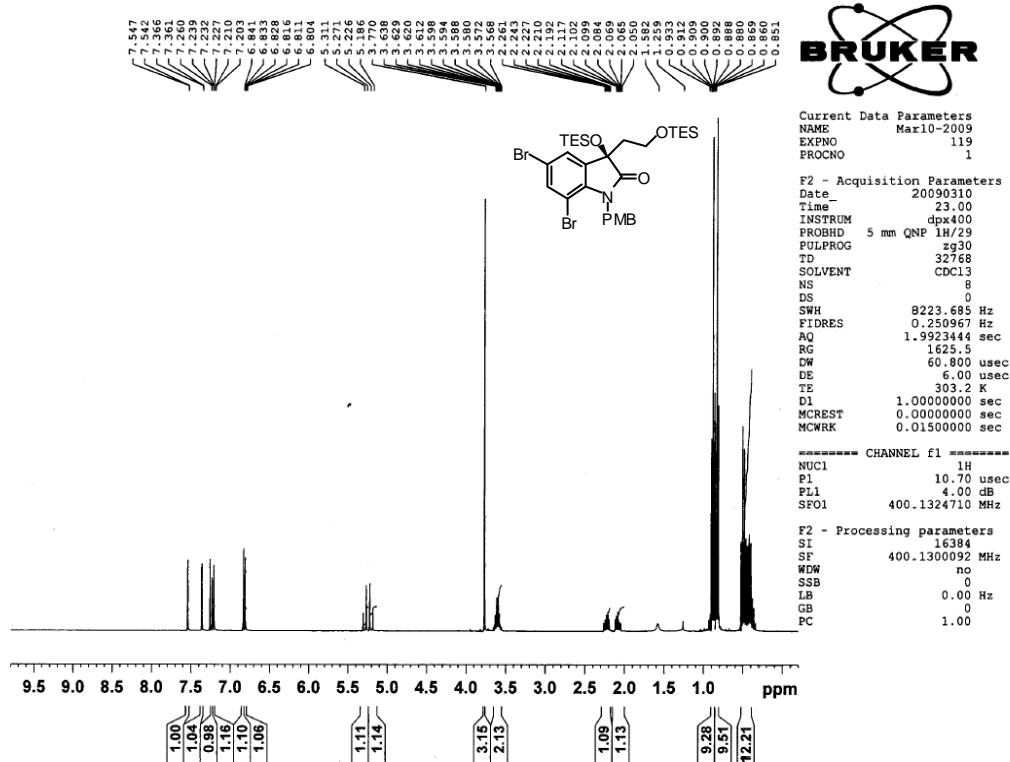
16
ON
2
108/03/11 20:03
Memory#3
background

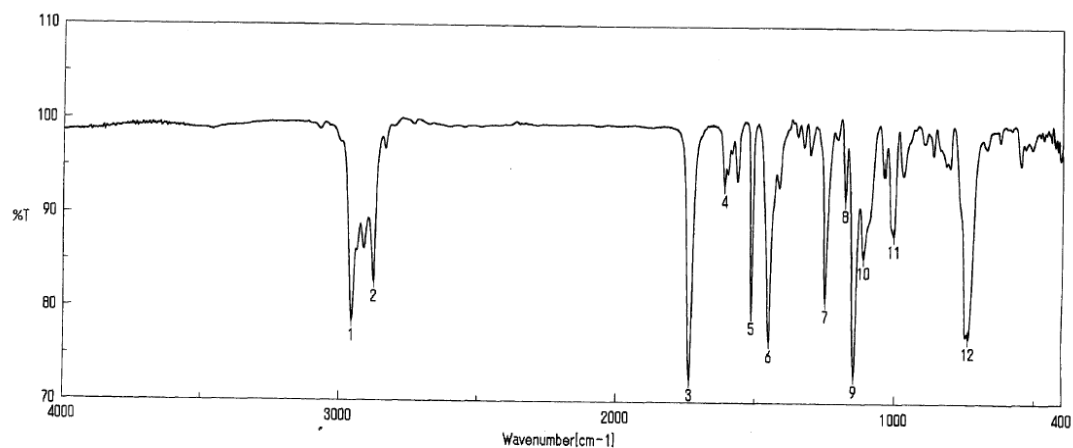
分解
アポダイゼーション
スキャンスピード

4 cm-1
Cosine
2 mm/sec



1: 3393.14, 94.5952	2: 2362.37, 85.4855	3: 2330.55, 93.1385	4: 1716.34, 83.2270
5: 1513.85, 84.7558	6: 1452.14, 84.3839	7: 1247.72, 88.0858	8: 1177.33, 93.9838
9: 1144.55, 88.3948	10: 1033.66, 94.7545	11: 735.71, 94.5325	



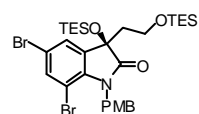


積算回数
ゼロファイリング
ゲイン
日時
測定者
ファイル名
サンプル名
コメント

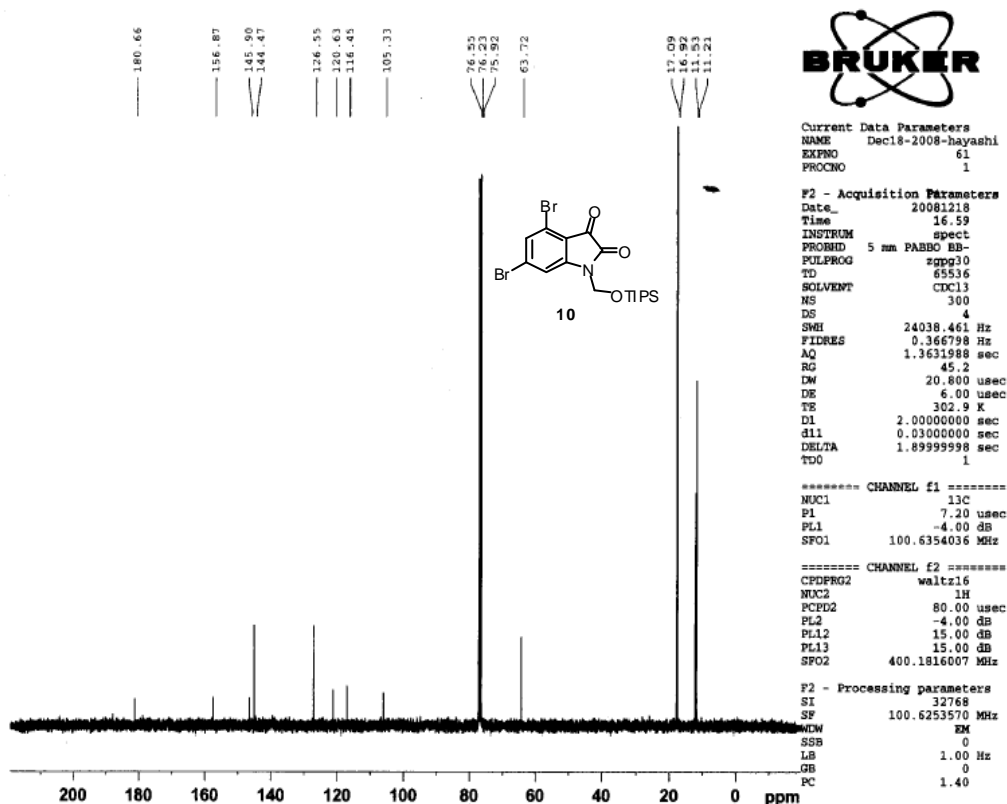
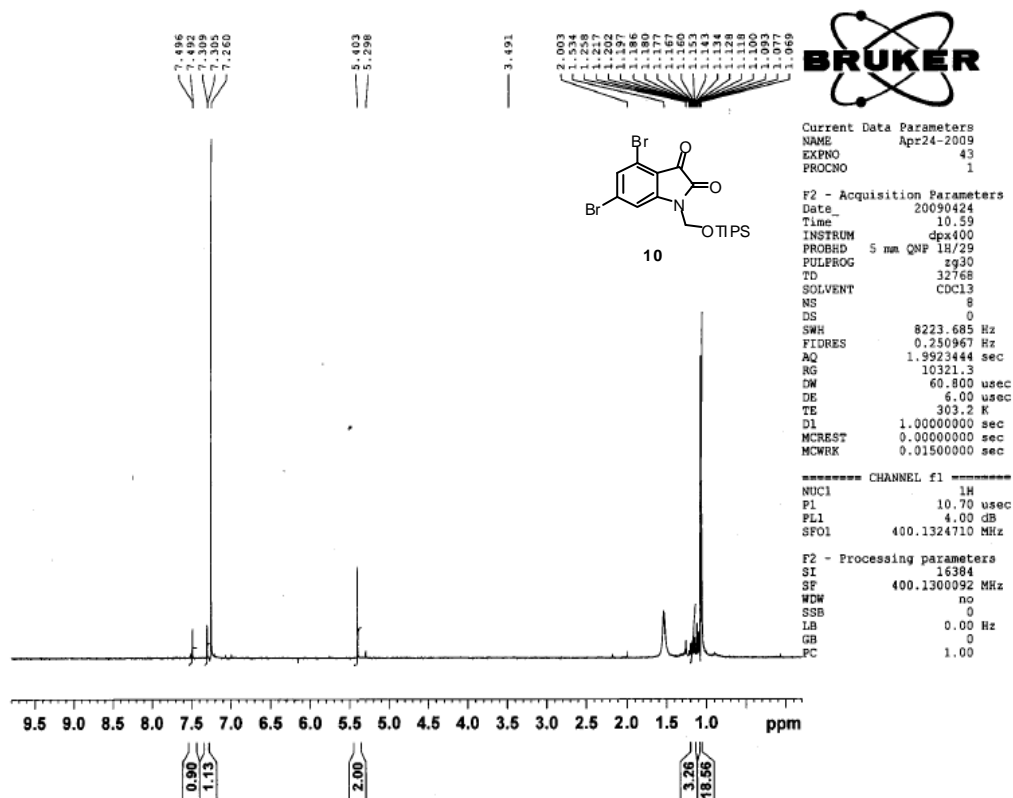
16
ON
2
109/03/11 20:19
Memory#4
background

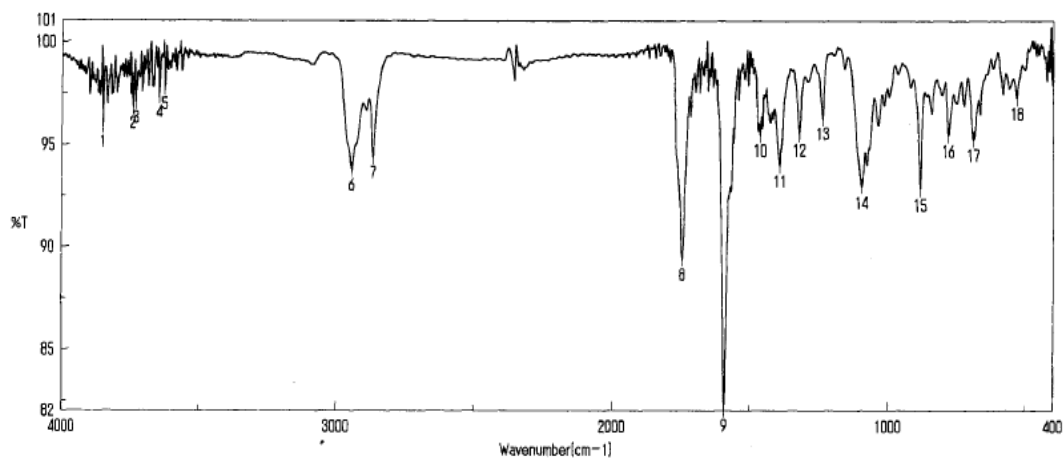
分解
アポダイゼーション
スキャンスピード

4 cm-1
Cosine
2 mm/sec



1: 2954.41, 78.5849	2: 2875.34, 82.7350	3: 1735.62, 72.4820	4: 1613.16, 93.1573
5: 1513.85, 79.8046	6: 1450.21, 78.8440	7: 1247.72, 80.9638	8: 1177.33, 91.6334
9: 1144.55, 72.9488	10: 1110.80, 85.6001	11: 1005.70, 87.8985	12: 736.67, 77.0784



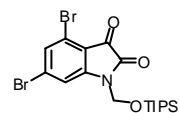


積算回数
ゼロフィリング
ゲイン
日時
測定者
ファイル名
サンプル名
コメント

64
ON
2
109/01/03 14:49
diBrisatin(OTIPS) natural. JMS
background

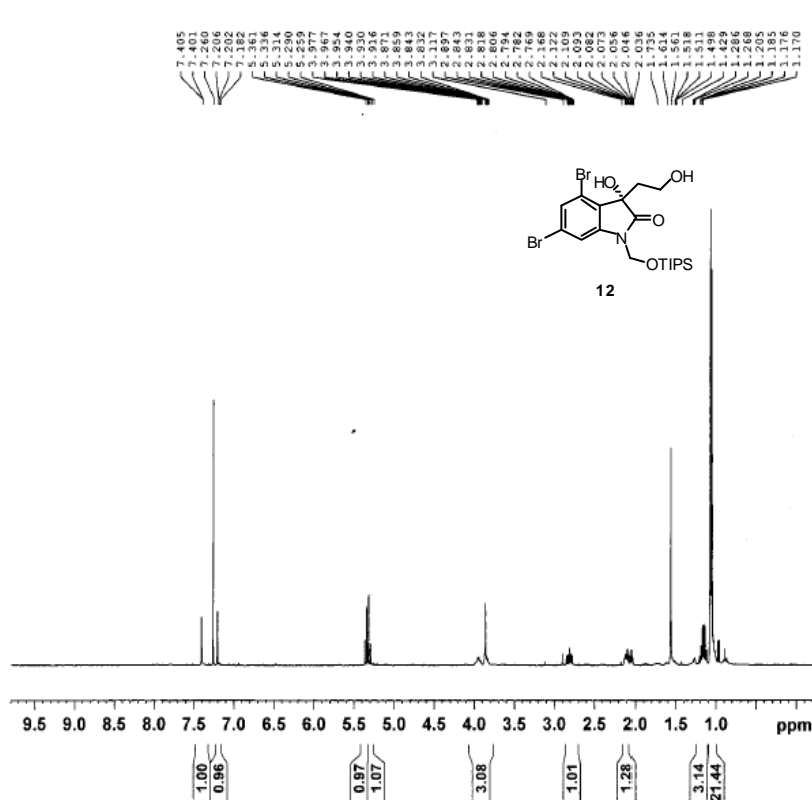
分解
アポダイゼーション
スキャンスピード

4 cm-1
Cosine
2 mm/sec



10

1: 3650.18, 95.8923	2: 3741.23, 96.8167	3: 3730.62, 97.0873	4: 3645.77, 97.3724
5: 3625.52, 97.7852	6: 2942.94, 93.8090	7: 2885.70, 94.3987	8: 1745.26, 89.4255
9: 1591.95, 82.0631	10: 1482.74, 95.4954	11: 1393.32, 93.9870	12: 1321.96, 95.5032
13: 1236.04, 98.3097	14: 1096.33, 92.9626	15: 883.24, 92.6413	16: 780.06, 95.4653
17: 689.43, 95.2644	18: 532.26, 97.2834		

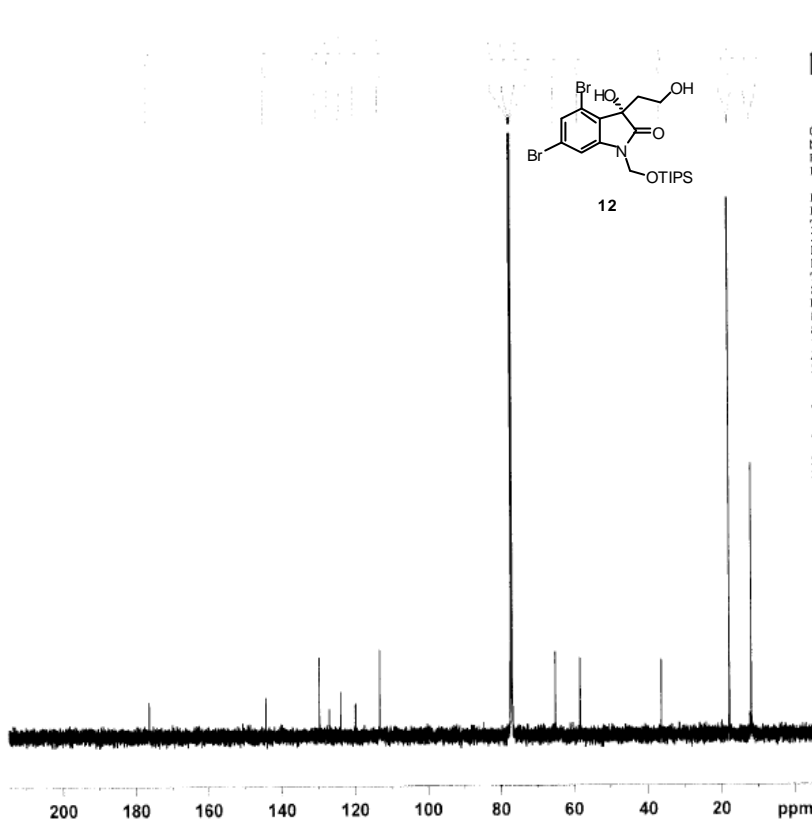


Current Data Parameters
NAME Jan03-2009
EXPNO 37
PROCNO 1

F2 - Acquisition Parameters
Date_ 20090103
Time 20.22
INSTRUM dpx400
PROBHD 5 mm QNP 1H/29
PULPROG zg30
TD 32768
SOLVENT CDCl3
NS 8
DS 0
SWH 8223.685 Hz
FIDRES 0.250967 Hz
AQ 1.9923444 sec
RG 4597.6
CW 60.800 usec
DE 6.00 usec
TE 303.2 K
D1 1.00000000 sec
MCREST 0.00000000 sec
MCWRK 0.01500000 sec

===== CHANNEL f1 =====
NUC1 1H
P1 10.70 usec
PL1 4.00 dB
SFO1 400.1324710 MHz

F2 - Processing parameters
SI 16384
SF 400.1300092 MHz
WDW no
SSB 0
LB 0.00 Hz
GB 0
PC 1.00



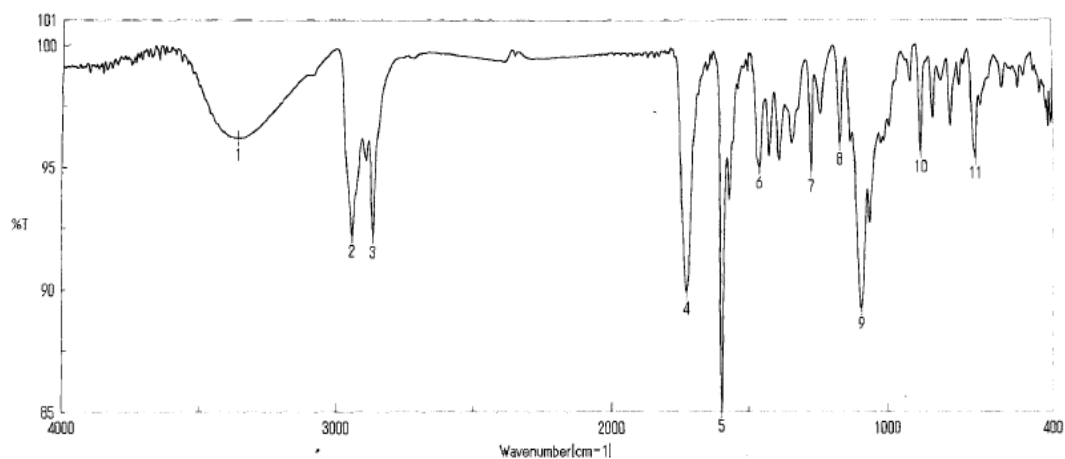
Current Data Parameters
NAME Jan03-2009
EXPNO 21
PROCNO 1

F2 - Acquisition Parameters
Date_ 20090103
Time 21.31
INSTRUM av600
PROBHD 5 mm PABBO BB-
PULPROG zgpg30
TD 65536
SOLVENT CDCl3
NS 205
DS 4
SWH 35971.223 Hz
FIDRES 0.548877 Hz
AQ 0.9110143 sec
RG 18390.4
CW 13.900 usec
DE 6.00 usec
TE 293.4 K
D1 2.00000000 sec
d11 0.03000000 sec
DELTA 1.89999998 sec
MCREST 0.00000000 sec
MCWRK 0.01500000 sec

===== CHANNEL f1 =====
NUC1 13C
P1 7.80 usec
PL1 0.00 dB
SFO1 150.9178989 MHz

===== CHANNEL f2 =====
CPDPRG2 waltz16
NUC2 1H
PCPD2 80.00 usec
PL2 -3.00 dB
PL12 14.00 dB
PL13 14.00 dB
SFO2 600.1324005 MHz

F2 - Processing parameters
SI 32768
SF 150.9028090 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

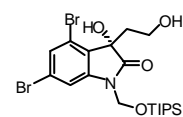


積算回数
ゼロフィリング
ゲイン
日時
測定者
ファイル名
サンプル名
コメント

64
ON
2
108/01/03 23:06
Memory#3
background

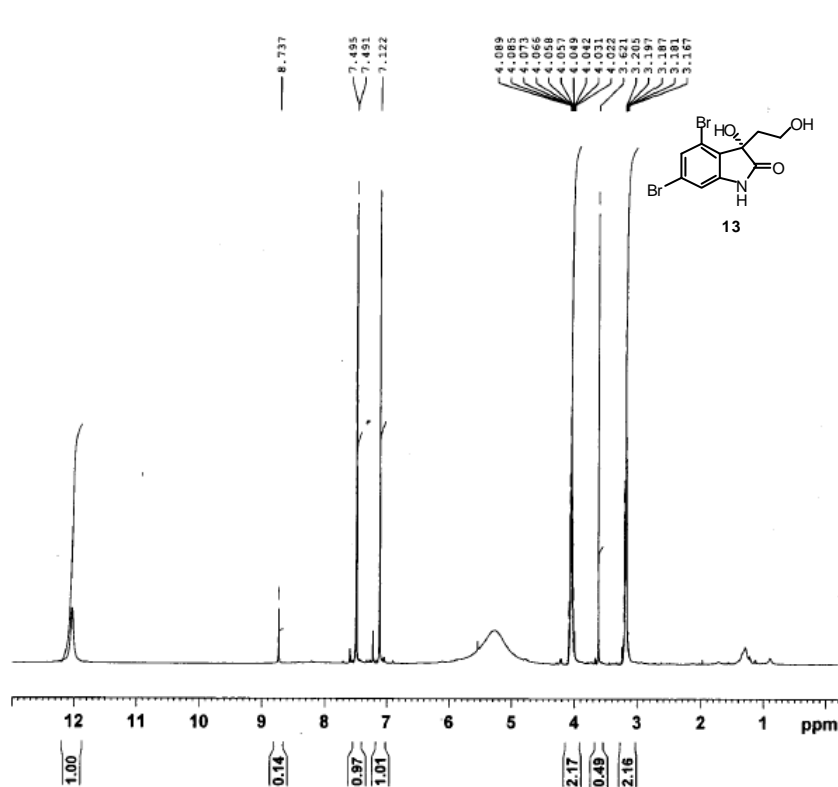
分解
アポダイゼーション
スキャンスピード

4 cm⁻¹
Cosine
2 mm/sec



12

1: 3356.50, 96.1773	2: 2941.88, 92.2225	3: 2864.74, 92.1813	4: 1725.98, 89.8775
5: 1598.70, 85.0725	6: 1484.57, 95.0076	7: 1278.57, 94.8629	8: 1174.44, 95.9840
9: 1093.44, 89.2615	10: 881.31, 95.6806	11: 683.64, 95.3998	

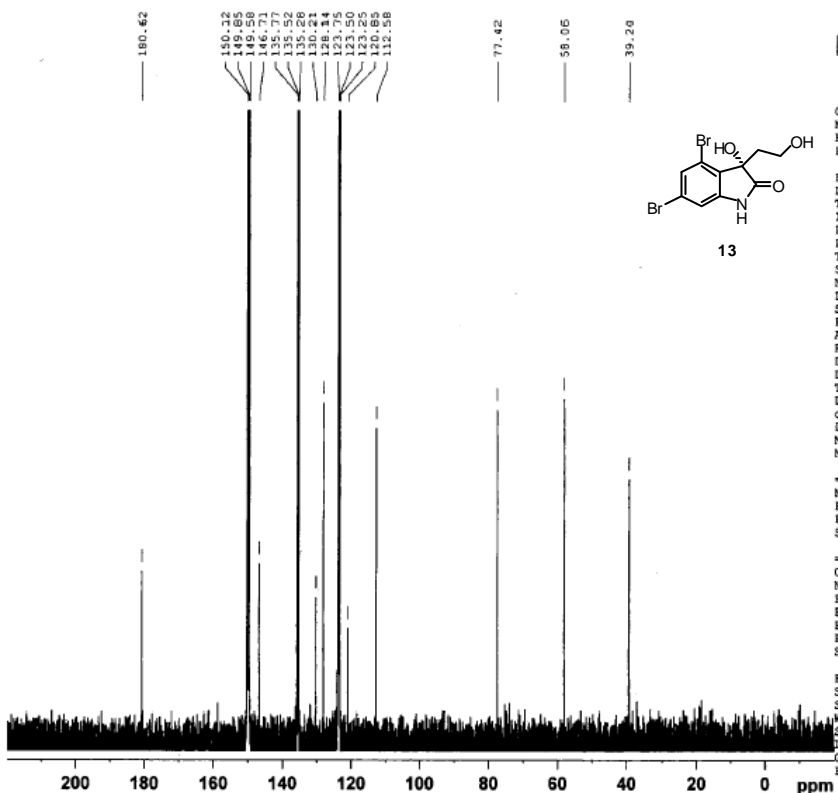


Current Data Parameters
NAME Mar31-2009
EXPNO 30
PROCNO 1

F2 - Acquisition Parameters
Date_ 20090331
Time 15.58
INSTRUM dpx400
PROBHD 5 mm QNP 1H/29
PULPROG zg30
TD 32768
SOLVENT Pyr
NS 32
DS 0
SWH 8223.685 Hz
FIDRES 0.250967 Hz
AQ 1.9923444 sec
RG 2580.3
DW 60.800 usec
DE 6.00 usec
TE 303.2 K
D1 1.00000000 sec
MCREST 0.00000000 sec
MCWRK 0.01500000 sec

===== CHANNEL f1 =====
NUC1 1H
P1 10.70 usec
PL1 4.00 dB
SFO1 400.1324710 MHz

F2 - Processing parameters
SI 16384
SF 400.1305874 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00



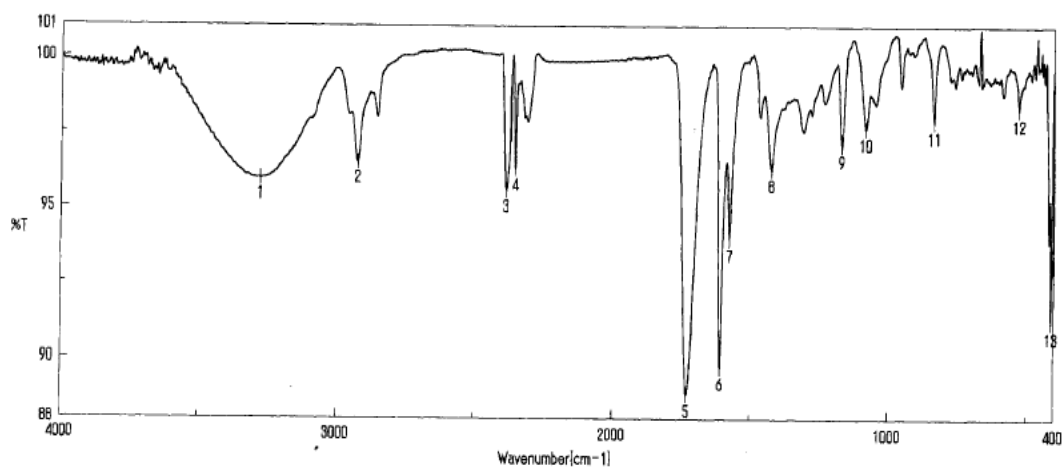
Current Data Parameters
NAME Mar31-2009
EXPNO 31
PROCNO 1

F2 - Acquisition Parameters
Date_ 20090331
Time 16.11
INSTRUM dpx400
PROBHD 5 mm QNP 1H/29
PULPROG zgpg30
TD 65536
SOLVENT Pyr
NS 180
DS 2
SWH 31847.133 Hz
FIDRES 0.405949 Hz
AQ 1.0289652 sec
RG 7298.2
DW 15.700 usec
DE 6.00 usec
TE 303.2 K
D1 2.00000000 sec
d11 0.03000000 sec
DELTA 1.89999998 sec
MCREST 0.00000000 sec
MCWRK 0.01500000 sec

===== CHANNEL f1 =====
NUC1 13C
P1 8.60 usec
PL1 2.00 dB
SFO1 100.6254358 MHz

===== CHANNEL f2 =====
CPDPRG2 waltz16
NUC2 1H
PCPD2 80.00 usec
PL2 4.00 dB
PL12 21.47 dB
PL13 21.47 dB
SFO2 400.1316005 MHz

F2 - Processing parameters
SI 32768
SF 100.6128973 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40



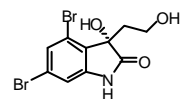
積算回数
ゼロフィリング
ゲイン
日時
測定者
ファイル名
サンプル名
コメント

16
ON
1
109/03/31 15:35

Memory#4
background

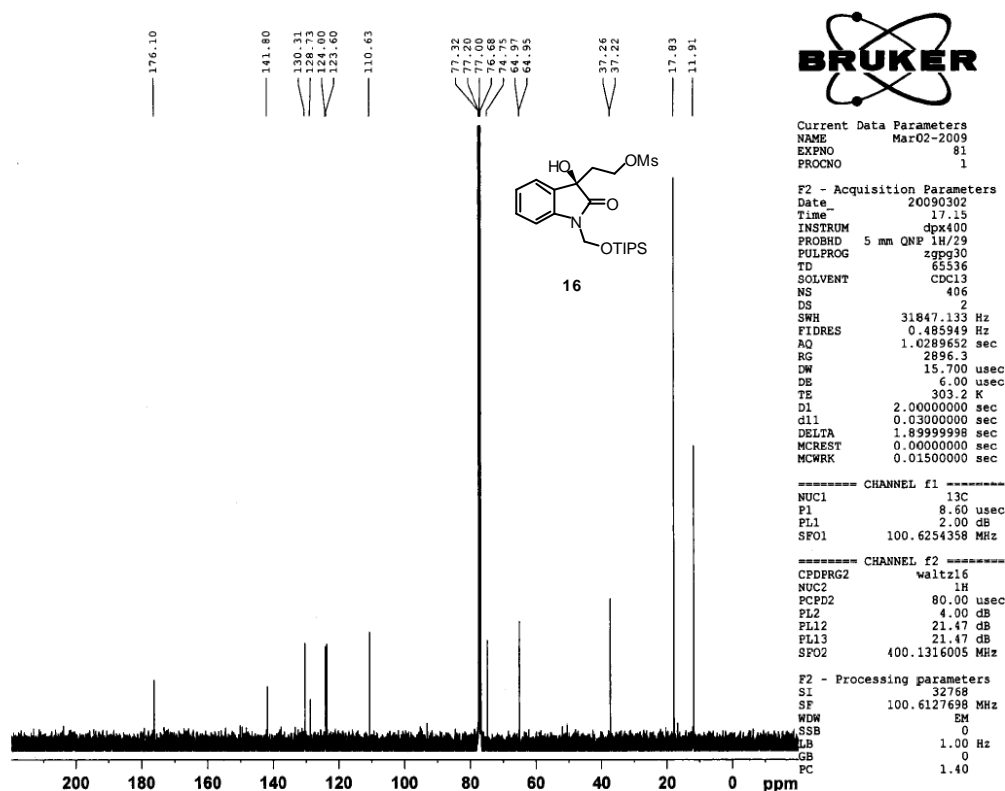
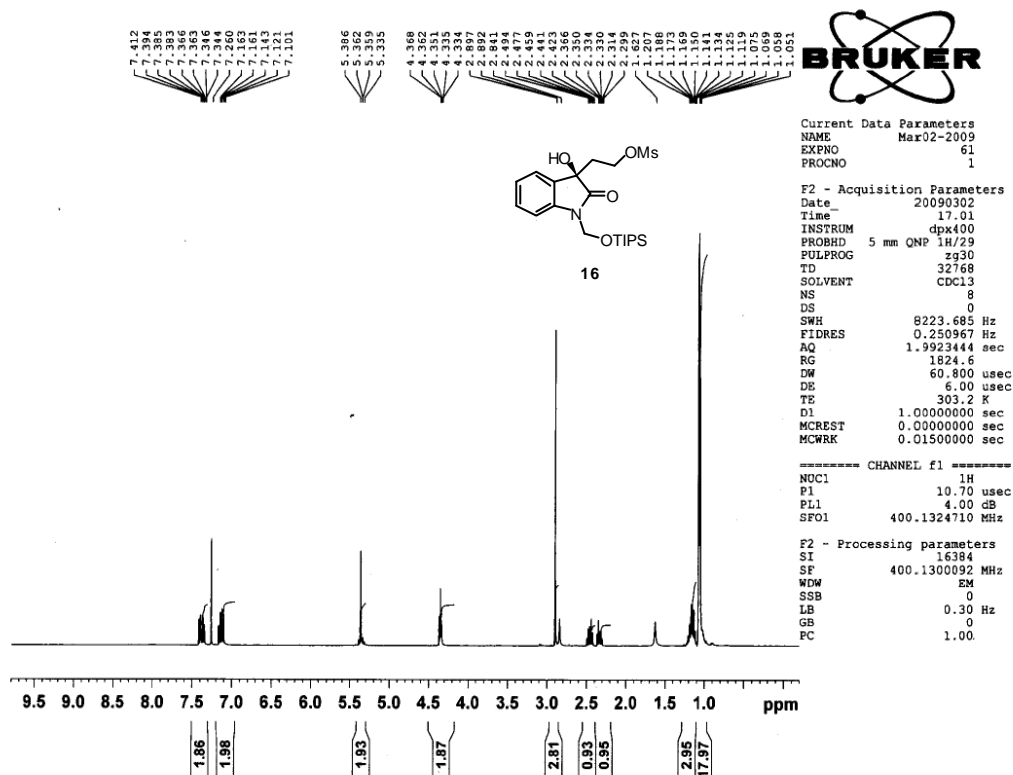
分解
アポダイゼーション
スキャンスピード

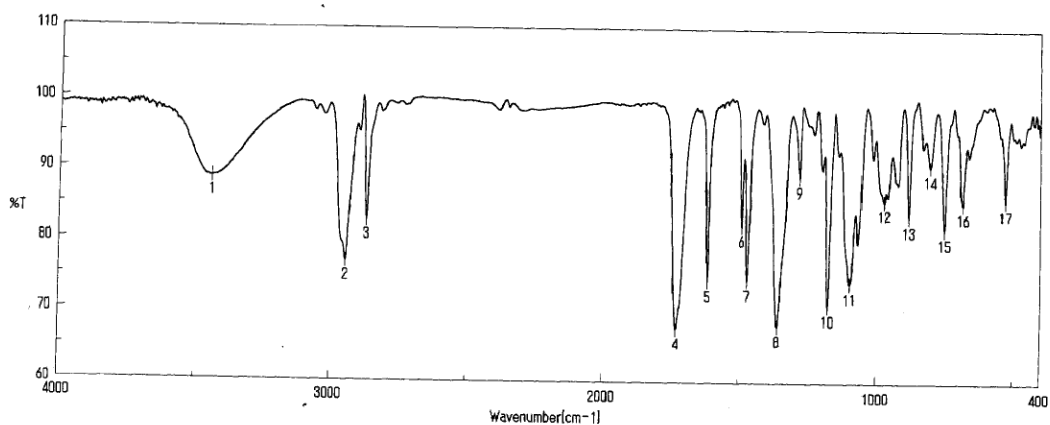
4 cm-1
Cosine
2 mm/sec



13

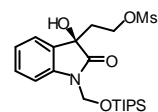
1: 3275.50, 95.9556	2: 2923.56, 96.5016	3: 2380.89, 95.5881	4: 2348.87, 96.2823
5: 1726.83, 88.8083	6: 1607.38, 89.6934	7: 1574.59, 93.9574	8: 1424.17, 96.2745
9: 1189.65, 97.0914	10: 1082.83, 97.6470	11: 836.95, 97.8771	12: 527.44, 98.2881
13: 498.83, 91.2279			





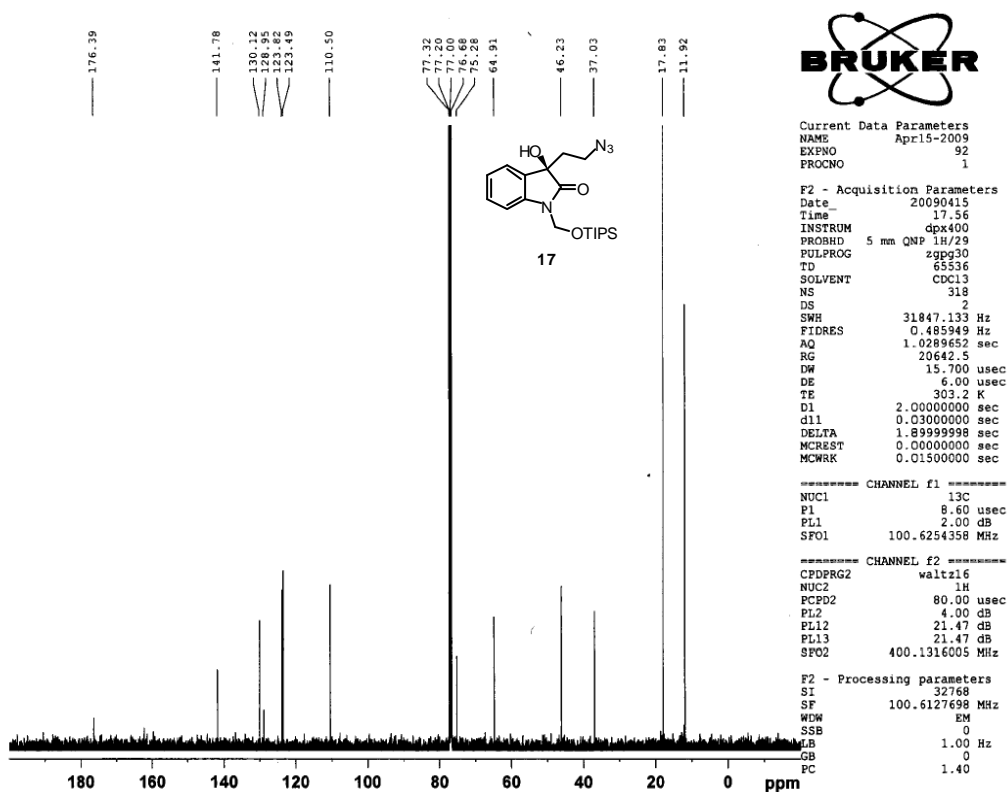
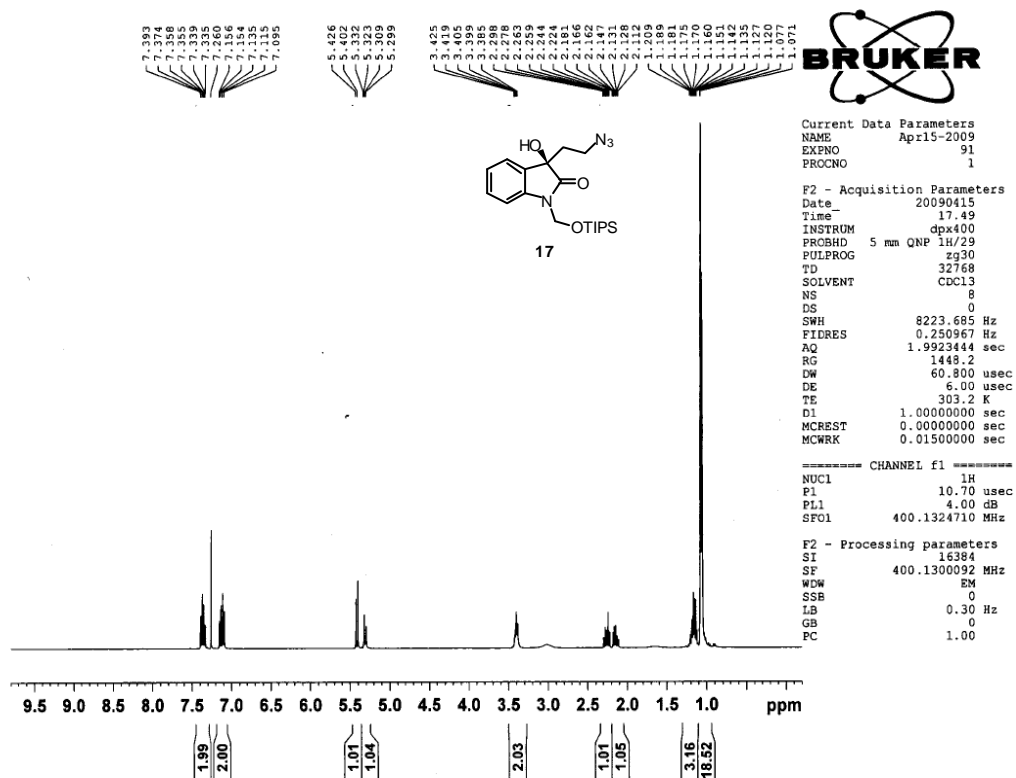
繰り返し回数 16
 ゼロファイリング ON
 ゲイン 1
 日時 109/03/09 20:36
 測定者
 ファイル名 Memory#3
 サンプル名 background
 コメント

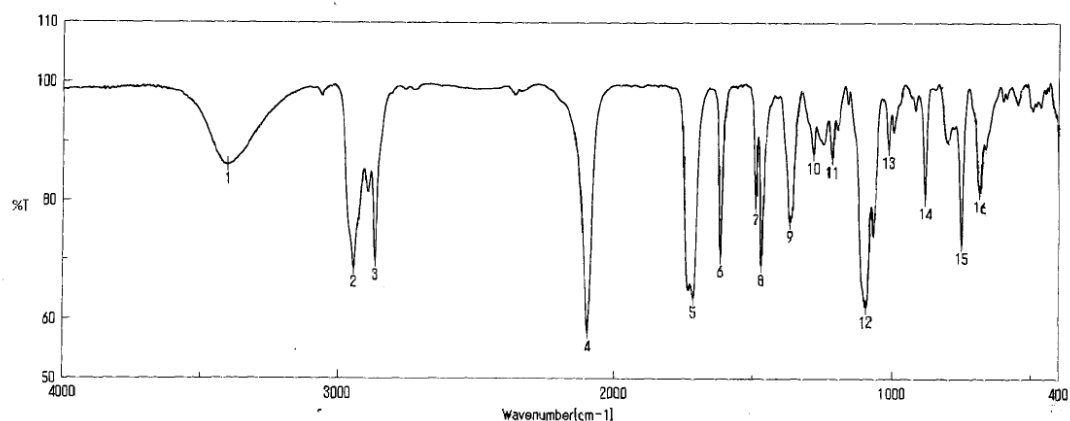
分解 4 cm⁻¹
 アポダイゼーション Cosine
 スキャンスピード 2 mm/sec



16

1: 3437.49, 88.7907	2: 2944.77, 76.9865	3: 2886.67, 82.8222	4: 1730.80, 67.6003
5: 1615.09, 74.3235	6: 1489.74, 82.1824	7: 1489.49, 74.4732	8: 1359.57, 68.0199
9: 1280.50, 89.1570	10: 1175.40, 70.8730	11: 1094.41, 74.1168	12: 973.88, 85.8708
13: 883.24, 83.5410	14: 806.10, 90.7340	15: 752.10, 81.8180	16: 684.61, 65.3872
17: 528.40, 85.6095			



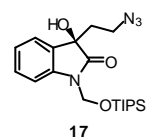


積算回数
ゼロフィリング
ゲイン
日時
測定者
ファイル名
サンプル名
コメント

16
ON
1
109/04/15 20:44
Memory#3
background

分解
アポダイゼーション
スキャンスピード

4 cm-1
Cosine
2 mm/sec



1: 3402.78, 86.0810	2: 2943.80, 68.7794	3: 2886.67, 69.9037	4: 2097.21, 57.9311
5: 1715.37, 63.7242	6: 1616.06, 70.5728	7: 1488.78, 79.2320	8: 1469.40, 69.1118
9: 1366.32, 76.5065	10: 1281.47, 87.9909	11: 1213.97, 87.2798	12: 1092.48, 62.0546
13: 1015.34, 88.9378	14: 882.27, 80.4073	15: 751.14, 72.8242	16: 685.57, 81.5139



```

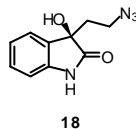
Current Data Parameters
NAME          May17-2009
EXPNO         1
PROCNO        1

F2 - Acquisition Parameters
Date          20090317
Time          13.00
INSTRUM       5 mm QNP
PROBHD        dpx400
PULPROG       zg30
TD            32768
SOLVENT       CDCl3
NS            8
DS            0
SWH           8223.685 Hz
FIDRES        0.25067 Hz
AQ            1.9923444 sec
RG            919.52
RO            60.800 usec
DE            6.000 usec
TE            303.2 K
D1            1.00000000 sec
d11           0.00000000 sec
MCRET        0.01500000 sec
MCWRK        0.01500000 sec

```

```
===== CHANNEL f1 =====
NUC1          1H
P1            7.90 usec
PL1           3.00 dB
SFO1          400.1324710 MHz
```

```
F2 - Processing parameters
SI                16384
SF                400.1300092 MHz
WDW               no
SSB               0
LB               0.00 Hz
GB               0
PC               1.00
```



```
Current Data Parameters
NAME      Mar17-2009-hayashi
EXPNO      20
PROCNO     1
```

```

P2 - Acquisition Parameters
Date_      20090317
Time       13.41
INSTRUM    spect
PROBHD     5 mm PABBO BB-
PULPROG    zgpg30
TD         65536
NS          CDX13
SOLVENT    DMSO
DS          600
SWH         4
FIDRES     0.366798 Hz
AQ         1.363198 sec
RG         128
DE         20.800 usec
WE         6.00 usec
TE         302.9 K
D1         2.00000000 sec
d11        0.03000000 sec
DELTA      1.89999999 sec
TD0        1

```

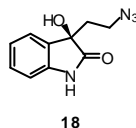
```
===== CHANNEL f1 =====
NUC1          13C
P1             7.20 usec
PL1           -4.00 dB
SFO1         100.6354036 MHz
```

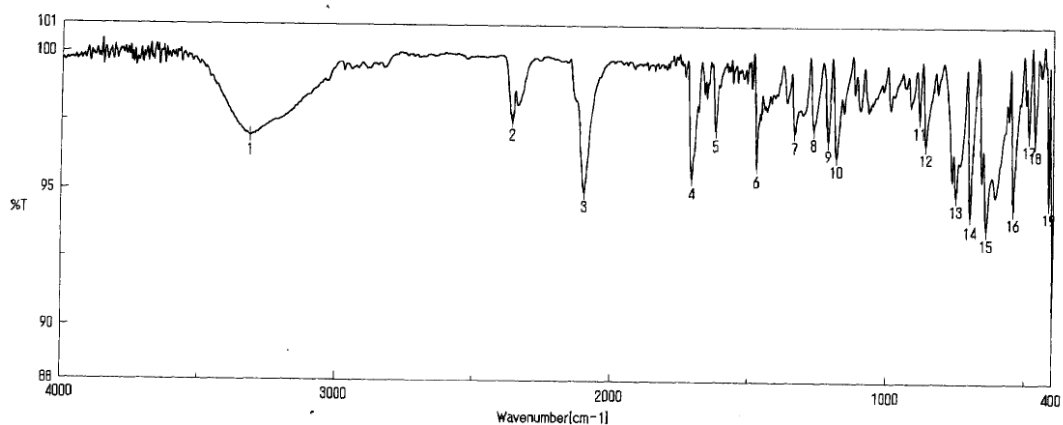
```

===== CHANNEL f2 =====
CPDFRG2          waltz16
NUC2              1H
PCPDF2           80.00 usec
PL2              -4.00 dB
PL12             15.00 dB
PL13             15.00 dB
SFO2            400.1816007 MHz

```

```
F2 - Processing parameters
SI                      32768
SF                      100.6253449 MHz
WDW                      EM
SSB                      0
LB                      1.00 Hz
GB                      0
PC                      1.40
```



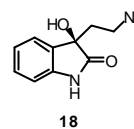


積算回数
ゼロフィリング
ゲイン
日時
測定者
ファイル名
サンプル名
コメント

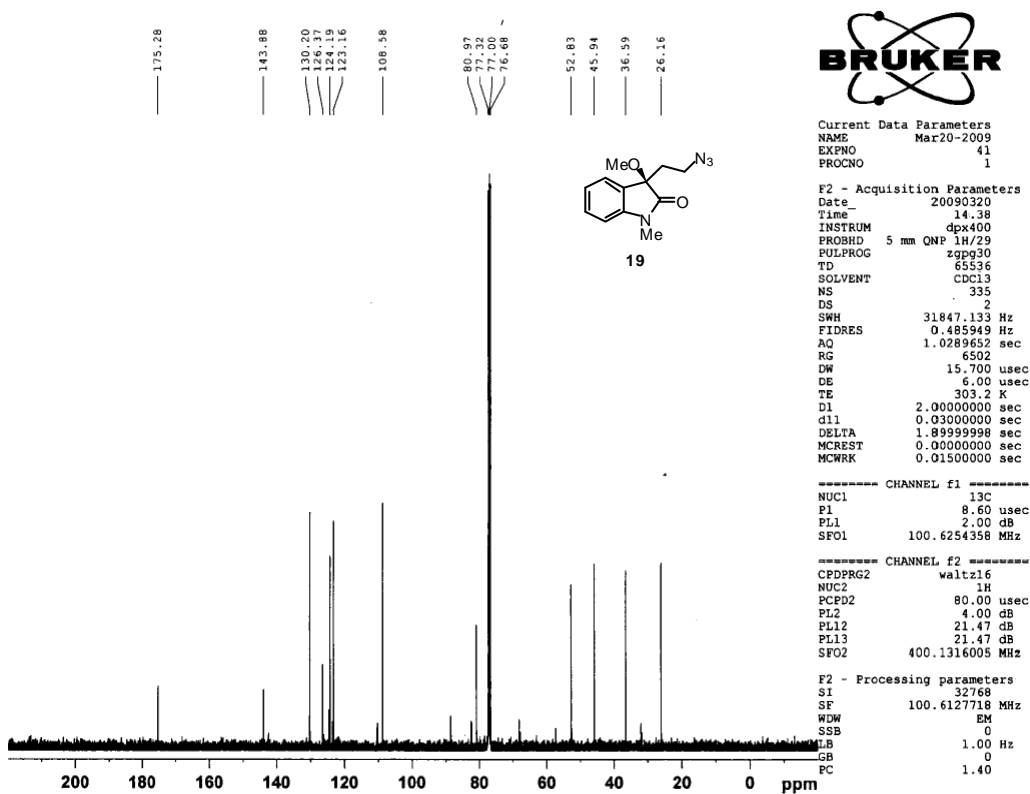
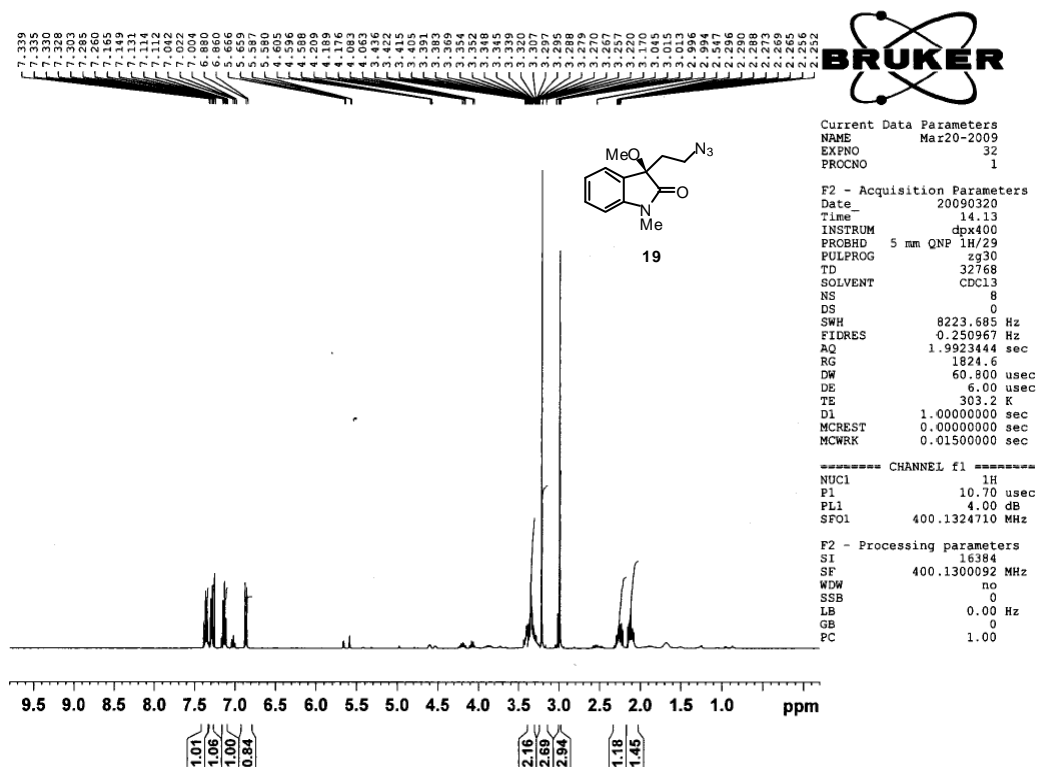
16
ON
2
109/03/17 14:08
Memory#3
background

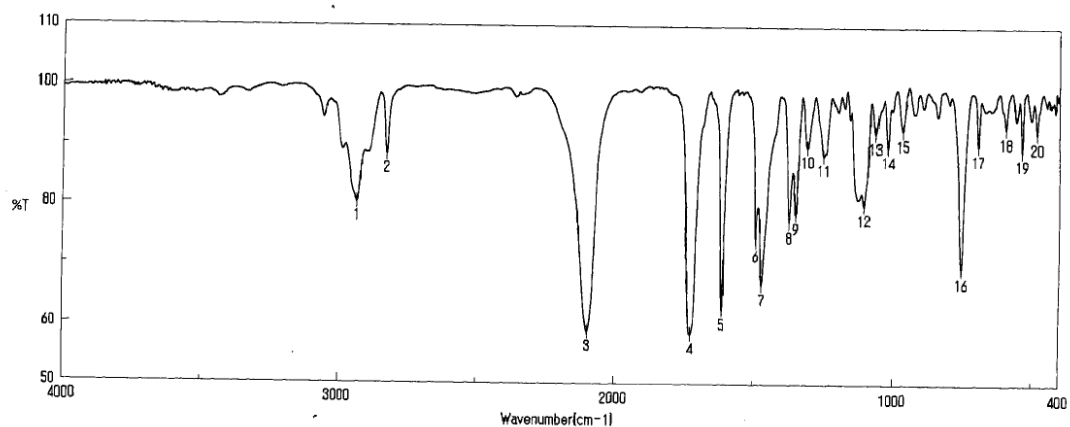
分解
アポダイゼーション
スキャンスピード

4 cm⁻¹
Cosine
2 mm/sec



1: 3313.11, 96.9334	2: 2358.52, 97.5331	3: 2098.17, 94.9395	4: 1706.69, 95.4665
5: 1620.88, 97.1981	6: 1471.42, 95.8913	7: 1333.53, 97.1408	8: 1264.11, 97.2578
9: 1212.04, 96.8497	10: 1183.11, 96.2697	11: 886.13, 97.6965	12: 862.99, 96.7435
13: 753.07, 94.8580	14: 702.93, 94.0977	15: 643.14, 93.0280	16: 545.76, 94.3675
17: 488.87, 97.0911	18: 466.99, 96.8830	19: 417.51, 94.5644	



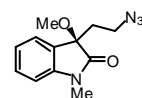


積算回数
ゼロファイリング
ゲイン
日時
測定者
ファイル名
サンプル名
コメント

16
ON
1
109/03/20 15:19
Memory#3
background

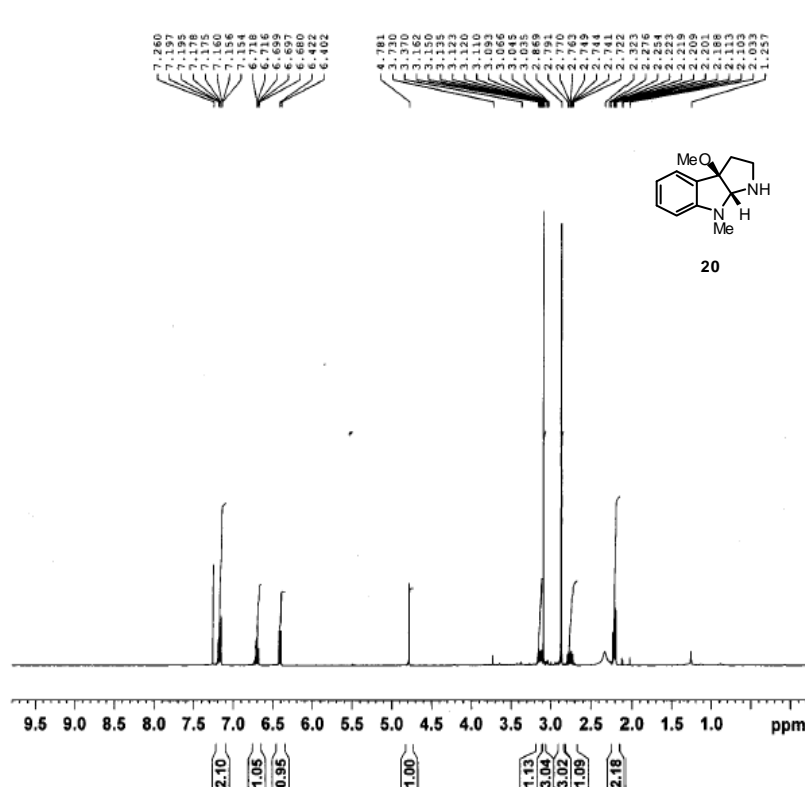
分解
アポダイゼーション
スキャンスピード

4 cm⁻¹
Cosine
2 mm/sec



19

1: 2934.16, 80.4971	2: 2827.13, 88.5275	3: 2098.17, 58.8342	4: 1725.01, 58.4829
5: 1613.16, 62.5633	6: 1492.83, 74.0805	7: 1469.49, 66.7502	8: 1372.10, 77.2481
9: 1348.96, 78.6395	10: 1309.43, 89.9629	11: 1250.81, 88.5780	12: 1105.01, 80.1304
13: 1085.48, 92.3101	14: 1023.05, 89.9927	15: 970.02, 92.7781	16: 754.99, 89.5357
17: 698.11, 90.2584	18: 597.82, 93.1563	19: 538.04, 89.4276	20: 484.05, 92.3390

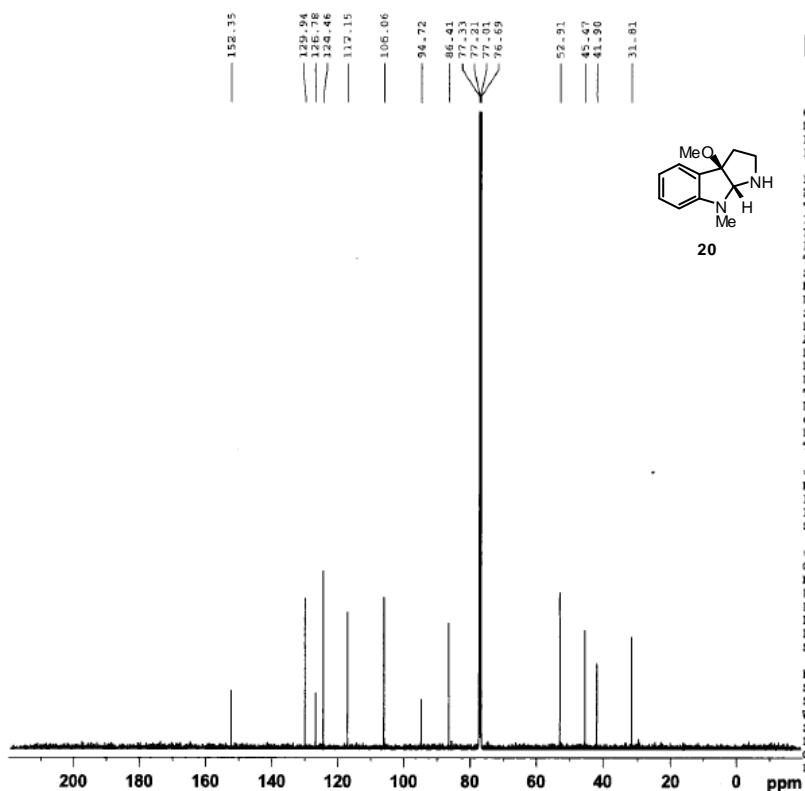


Current Data Parameters
 NAME Apr16-2009
 EXPNO 164
 PROCNO 1

F2 - Acquisition Parameters
 Date 20090416
 Time 21.11
 INSTRUM dpx400
 PROBD 5 mm QNP 1H/29
 PULPROG zg30
 TD 32768
 SOLVENT CDCl3
 NS 8
 DS 0
 SWH 8223.695 Hz
 FIDRES 0.250967 Hz
 AQ 1.9923444 sec
 RG 3649.1
 DW 60.800 usec
 DE 6.00 usec
 TE 303.2 K
 D1 1.00000000 sec
 MCREST 0.00000000 sec
 MCWKK 0.01500000 sec

===== CHANNEL f1 =====
 NUC1 1H
 P1 10.70 usec
 PL1 4.00 dB
 SFO1 400.1324710 MHz

F2 - Processing parameters
 SI 16384
 SF 400.1300092 MHz
 WDW no
 SSB 0
 LB 0.00 Hz
 GB 0
 PC 1.00



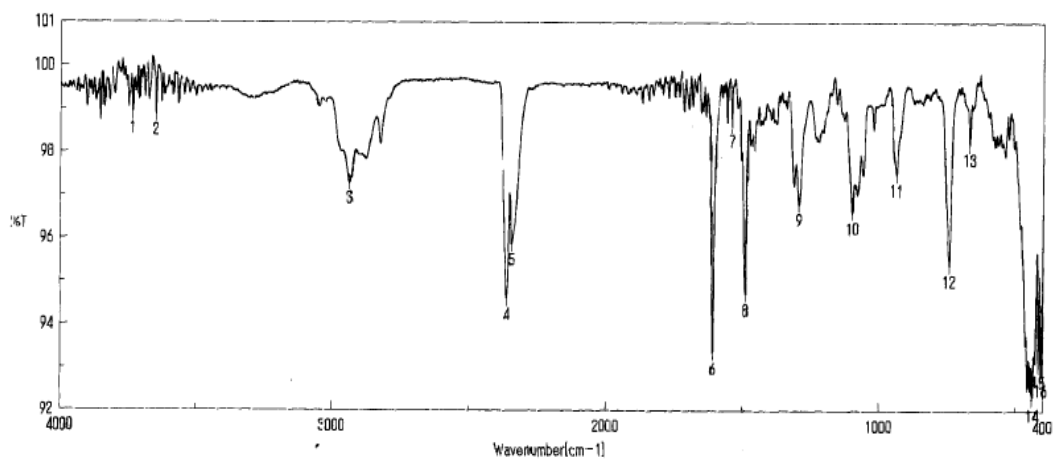
Current Data Parameters
 NAME Apr16-2009-hayashi
 EXPNO 70
 PROCNO 1

F2 - Acquisition Parameters
 Date 20090416
 Time 22.16
 INSTRUM spect
 PROBD 5 mm PABBO DB-
 PULPROG zgpg30
 TD 65536
 SOLVENT CDCl3
 NS 1024
 DS 4
 SWH 24038.461 Hz
 FIDRES 0.366798 Hz
 AQ 1.3631988 sec
 RG 203
 DW 20.800 usec
 DE 6.00 usec
 TE 296.9 K
 D1 2.00000000 sec
 d11 0.03000000 sec
 DELTA 1.89999998 sec
 TD0 1

===== CHANNEL f1 =====
 NUC1 13C
 P1 7.20 usec
 PL1 -3.50 dB
 SFO1 100.6354036 MHz

===== CHANNEL f2 =====
 CPDPRG2 waltz16
 NUC2 1H
 PCPD2 80.00 usec
 PL2 -4.00 dB
 PL12 14.50 dB
 PL13 14.50 dB
 SFO2 400.1816007 MHz

F2 - Processing parameters
 SI 32768
 SF 100.6253433 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40

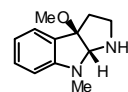


積算回数
ゼロファイリング
ゲイン
日時
測定者
ファイル名
サンプル名
コメント

16
ON
2
108/04/18 15:28
Memory#3
background

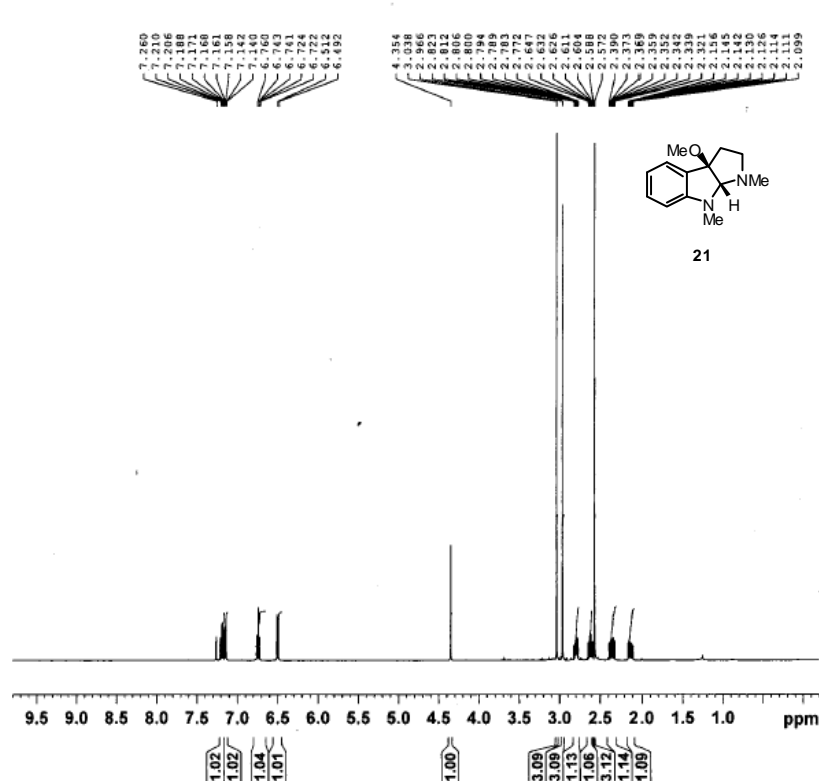
分解
アポダイゼーション
スキャンスピード

4 cm⁻¹
Cosine
2 mm/sec



20

1: 3734.48, 98.0191	2: 3548.66, 98.8743	3: 2937.08, 97.3080	4: 2360.44, 94.6147
5: 2341.15, 95.8702	6: 1809.31, 93.3449	7: 1540.85, 96.6429	8: 1490.70, 94.7051
9: 1294.00, 96.8044	10: 1089.23, 96.5995	11: 938.20, 97.4886	12: 743.42, 95.3726
13: 669.18, 98.2113	14: 437.76, 92.2808	15: 418.55, 93.0498	16: 408.91, 92.8490

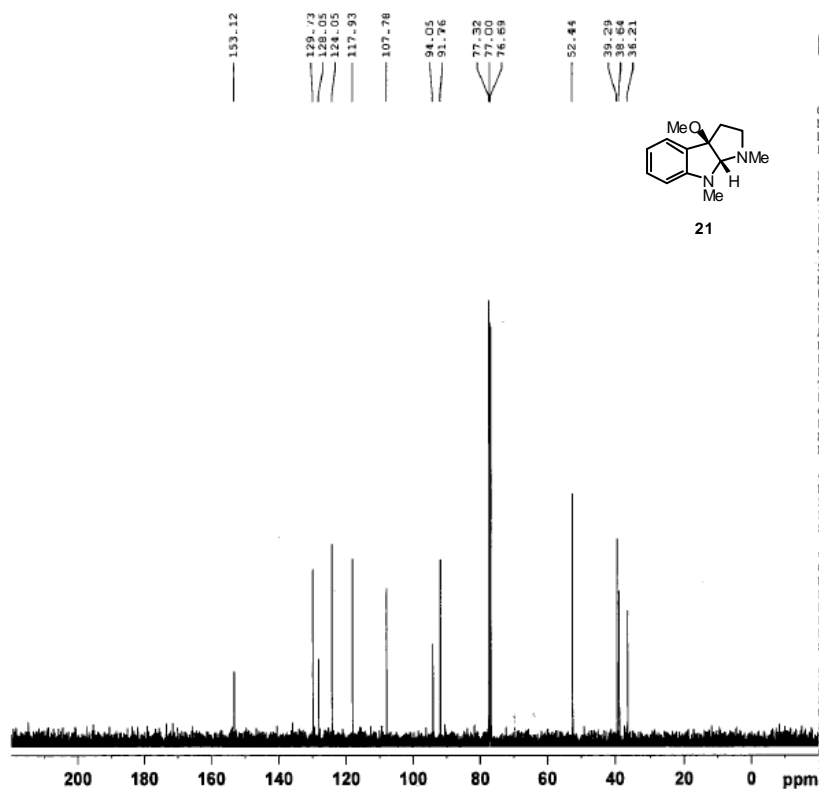


Current Data Parameters
 NAME Apr22-2009
 EXPNO 12
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20090422
 Time_ 10.05
 INSTRUM dpx400
 PROBHD 5 mm QNP 1H/29
 PULPROG zg30
 TD 32768
 SOLVENT CDCl3
 NS 8
 DS 0
 SWH 8223.685 Hz
 FIDRES 0.250967 Hz
 AQ 1.9923444 sec
 RG 1448.2
 DW 60.800 usec
 DE 6.00 usec
 TE 303.2 K
 D1 1.00000000 sec
 MCREST 0.00000000 sec
 MCWRR 0.01500000 sec

===== CHANNEL f1 =====
 NUC1 1H
 P1 10.70 usec
 PL1 4.00 dB
 SFO1 400.1324710 MHz

F2 - Processing parameters
 SI 16384
 SF 400.1300092 MHz
 WDW no
 SSB 0
 LB 0.00 Hz
 GB 0
 PC 1.00



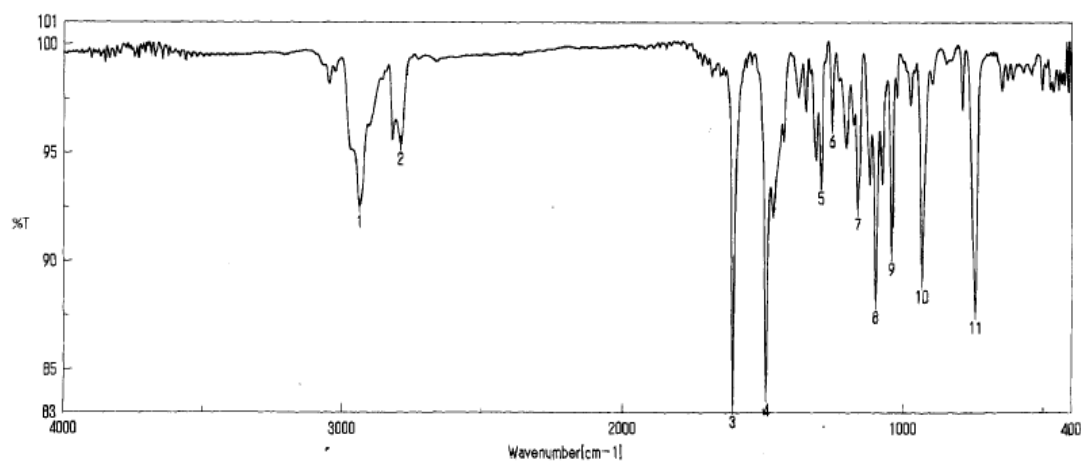
Current Data Parameters
 NAME Apr22-2009
 EXPNO 21
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20090422
 Time_ 10.19
 INSTRUM dpx400
 PROBHD 5 mm QNP 1H/29
 PULPROG zgpg30
 TD 65536
 SOLVENT CDCl3
 NS 64
 DS 2
 SWH 31847.133 Hz
 FIDRES 0.485949 Hz
 AQ 1.0289652 sec
 RG 7298.2
 DW 15.700 usec
 DE 6.00 usec
 TE 303.2 K
 D1 2.00000000 sec
 d11 0.03000000 sec
 DELTA 1.89999998 sec
 MCREST 0.00000000 sec
 MCWRR 0.01500000 sec

===== CHANNEL f1 =====
 NUC1 13C
 P1 8.60 usec
 PL1 2.00 dB
 SFO1 100.6254358 MHz

===== CHANNEL f2 =====
 CPDPRG2 waltz16
 NUC2 1H
 FCPD2 80.00 usec
 PL2 4.00 dB
 PL12 21.47 dB
 PL13 21.47 dB
 SFO2 400.1316005 MHz

F2 - Processing parameters
 SI 32768
 SF 100.6127718 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40

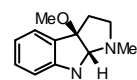


積算回数
ゼロフィリング
ゲイン
日時
測定者
ファイル名
サンプル名
コメント

32
ON
1
108/04/24 10:15
Memory#3
background

分解
アポダイゼーション
スキャンスピード

4 cm⁻¹
Oxline
2 mm/sec



21

1: 2938.02, 92.5473	2: 2791.46, 95.4216	3: 1608.34, 83.3025	4: 1489.74, 83.8498
5: 1294.00, 93.6511	6: 1254.47, 96.2809	7: 1162.87, 92.4345	8: 1088.26, 88.1443
9: 1041.37, 90.3837	10: 933.38, 89.0991	11: 742.46, 87.6738	