

SUPPORTING INFORMATION

A Practical Total Synthesis of Globo-H for Use in Anticancer Vaccines

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Ethyl 2,3,4-tri-*O*-benzyl- α -L-fucopyranosyl-(1 \rightarrow 2)-3,4-carbonyl-6-*O*-tri-isopropylsilyl- β -D-galactopyranosyl-(1 \rightarrow 3)-6-*O*-tri-isopropylsilyl-2-deoxy-2-phenylsulfonfylamino-1-thio- β -D-galactopyranoside (7).¹ A solution of thiofucosyl donor **15** (2.38 g, 4.41 mmol) and freshly activated 4 Å molecular sieves (2 g) in CH₂Cl₂/Et₂O (50 mL/25 mL) was stirred at rt for 1 h, then cooled to -78 °C. To the cooled solution was added AgOTf (3.4 g, 13.22 mmol) and 2,6-di-*tert*-butylpyridine (2.92 mL, 13.92 mmol). After 10 min, freshly distilled *p*-TolSCl (619.3 μ L, 4.41 mmol) was added. After 40 minutes, the characteristic yellow color of *p*-TolSCl in the reaction solution disappeared, indicating depletion of *p*-TolSCl. A solution of acceptor **4** (2.85 g, 4.41 mmol) in CH₂Cl₂/Et₂O (6 mL/3 mL) was added dropwise via a syringe. The reaction mixture was warmed to rt under stirring over 3 h, and stirred for an additional 1 h at rt. The mixture was filtered over Celite and further washed with CH₂Cl₂. After evaporation of solvent in vacuo, the mixture was diluted with ethyl acetate then washed with a saturated aqueous solution of NaHCO₃, water, and brine and dried over MgSO₄. After removal of the solvent, the crude was purified via silica gel flash chromatography using 10-15% EtOAc/hexane to give 3.66 g (78%) of α -trisaccharide **6**.¹ ¹H NMR (500 MHz, CDCl₃) δ 7.41-7.29 (m, 15H), 6.35 (d, *J* = 5.0 Hz, 1H), 5.01 (d, *J* = 3.7 Hz, 1H), 4.99 (d, *J* = 11.6 Hz, 1H), 4.92 (d, *J* = 4.7 Hz, 1H), 4.87 (d, *J* = 5.0 Hz, 1H), 4.84 (d, *J* = 11.6 Hz, 2H), 4.75 (d, *J* = 11.8 Hz, 1H), 4.71 (dd, *J* = 8.2, 3.6 Hz, 1H), 4.66 (d, *J* = 11.7 Hz, 2H), 4.54 (d, *J* = 6.3 Hz, 1H), 4.44 (m, 1H), 4.12-4.00 (m, 5H), 3.92-3.83 (m, 7H), 3.68 (m, 1H), 2.66 (d, *J* = 2.2 Hz, 1H), 1.15-1.07 (m, 45H); ESI-MS calcd for C₅₈H₈₆O₁₄Si₂Na⁺ 1085.55 (M + Na)⁺, found 1085.39. A mixture of trisaccharide glycal **6** (1 g, 0.94 mmol) and benzenesulfonamide (887 mg, 5.64 mmol) was azeotroped with C₆H₆ once and further dried on high vacuum for 1 h. It was dissolved in freshly distilled Et₂O (40 mL), and freshly activated 4 Å molecular sieves (1 g) was added. The resulting mixture was stirred at rt for 1 h, then cooled to -5 °C. To the solution was added I(*sym*-coll)₂ClO₄ (925 mg, 1.974 mmol) and the mixture was further stirred at -5 °C for 12 h. After filtration on Celite pad, the crude was washed with saturated Na₂S₂O₃ solution, saturated CuSO₄, brine and dried over MgSO₄. After concentration, the crude material, containing iodosulfonamide **17**, was immediately subjected to the next step without purification. To a solution of EtSH (695 μ L, 9.4 mmol) in DMF (5 mL) was added lithium bis-(trimethylsilyl)amide (LHMDS, 1.0 M in THF, 4.7 mL, 4.7 mmol) at -45 °C. After 15 min of stirring, the solution was transferred dropwise via a cannula to a flask containing iodosulfonamide **17** in DMF (25 mL) at -45 °C. The reaction mixture was allowed to warm to rt and stirred for a total of 3 h. After dilution with saturated NH₄Cl, the crude was extracted four times with ethyl acetate. The combined extracts were washed with water and brine and dried over MgSO₄. Concentration and purification by silica gel chromatography (15-25% EtOAc in hexane) afforded 903 mg (75% over two steps) of DEF donor **7**. ¹H NMR (500 MHz, CDCl₃) δ 7.90 (d, *J* = 7.8 Hz, 2H), 7.53 (t, *J* = 7.4 Hz, 1H), 7.47 (t, *J* = 7.4 Hz, 2H), 7.40-7.28 (m, 15H), 5.43 (s, 1H), 5.00 (d, *J* = 3.6 Hz, 1H), 4.98 (d, *J* = 11.6 Hz, 1H), 4.91 (d, *J* = 8.8 Hz, 1H), 4.84 (d, *J* = 7.9 Hz, 1H), 4.82 (d, *J* = 7.9 Hz, 1H), 4.74 (d, *J* = 11.8 Hz, 1H), 4.67-4.62 (m, 4H), 4.37 (d, *J* = 10.2 Hz, 1H), 4.12-4.08 (m, 4H), 4.00 (q, *J* = 6.5 Hz, 1H), 3.94-3.77 (m, 6H), 3.71 (s, 1H), 3.60-3.56 (m, 1H), 3.47 (m, 1H), 2.82 (m, 1H), 2.48-2.44 (m, 1H), 2.32-2.28 (m, 1H), 1.17 (d, *J* = 6.4 Hz, 3H), 1.15-1.05 (m, 45H); ¹³C NMR (150 MHz, CDCl₃) δ 155.5, 140.7, 138.9, 138.8, 132.9, 129.0, 128.8, 128.6, 128.5, 128.4, 128.2, 128.0, 127.8, 127.8, 127.6, 99.7, 98.1, 83.8, 81.5, 79.4, 77.8, 76.7, 75.1, 74.4, 73.2, 73.1, 71.7, 71.7, 69.8, 68.1, 68.0, 62.2, 62.0, 55.4, 23.8, 18.2, 18.2, 18.1, 17.0, 14.7, 12.3, 12.1, 11.9; ESI-MS calcd for C₆₆H₉₇O₁₆NS₂Si₂K⁺ 1318.54 (M + K)⁺, found 1318.58.

4-Pentenyl 2,4,6-tri-*O*-benzyl- α -D-galactopyranosyl-(1 \rightarrow 4)-2,3,6-tri-*O*-benzyl- β -D-galactopyranosyl-(1 \rightarrow 4)-2,3,6-tri-*O*-benzyl- β -D-glucopyranoside (8).² A solution of thiogalactosyl donor **16** (3.97 g, 5.86 mmol) and freshly activated 4 Å molecular sieves (4 g) in CH₂Cl₂/ Et₂O (100 mL/50 mL) was stirred at rt for 1 h, then cooled to -78 °C. To the cooled solution was added AgOTf (4.52 g, 17.6 mmol) and 2,6-di-*tert*-butylpyridine (3.9 mL, 17.6 mmol). After 10 min, freshly distilled *p*-TolSCl (824 μ L, 5.86 mmol) was added. After 10 minutes, the characteristic yellow color of *p*-TolSCl in the reaction solution disappeared, indicating depletion of *p*-TolSCl. A solution of acceptor **12** (4.65 g, 4.88 mmol) in CH₂Cl₂/ Et₂O (10 mL/5 mL) was added dropwise via a syringe. The reaction mixture was warmed to rt under stirring over 3 h, and then stirred for an additional 1 h at rt. The mixture was filtered over Celite and further washed with CH₂Cl₂. After evaporation of solvent in vacuo, the mixture was diluted with ethyl acetate then washed with a saturated aqueous solution of NaHCO₃, water, and brine and dried over MgSO₄. After removal of the solvent, the crude was purified via silica gel flash chromatography using 15% EtOAc/hexane to give 5.55 g (76 %) of α -trisaccharide **13**. This PMB protected **13** (5.25 g, 3.49 mmol) in CH₂Cl₂ (130 mL) at 0 °C was treated with phosphate buffer (20 mL, pH 7.2) and 2,3-dichloro-5,6-dicyano-1,4-benzoquinone (DDQ) (1.19 g, 5.24 mmol, 1.5 equiv.) and stirred at 0 °C for 5 h. The reaction mixture was diluted with saturated NaHCO₃ and further stirred at rt for 1 h. The mixture was extracted twice with CH₂Cl₂, then washed with brine, dried over MgSO₄, and concentrated to dryness. The crude material was purified by flash column chromatography using 16% EtOAc/hexane to give 4.06 g (84 %) of ABC acceptor **8**. ¹H NMR (500 MHz, CDCl₃) δ 7.42 - 7.16 (m, 45 H), 5.89-5.81 (m, 1H), 5.13 (d, *J* = 3.1 Hz, 1H), 5.10 (d, *J* = 11.4 Hz, 1H), 5.06-4.99 (m, 2H), 4.91-4.85 (m, 2H), 4.81-4.71 (m, 6H), 4.85-4.49 (m, 5H), 4.43-4.38 (m, 3H), 4.31 (s, 2H), 4.19-4.03 (m, 5H), 3.99-3.95 (m, 3H), 3.84-3.80 (m, 2H), 3.76 (d, *J* = 10.0 Hz, 1H), 3.64-3.32 (m, 9H), 3.22 (dd, *J* = 10.0, 5.0 Hz, 1H), 2.22-2.17 (m, 2H), 1.82-1.73 (m, 2H); ¹³C NMR (CDCl₃, 150 MHz) δ 139.5, 138.8, 138.7, 138.7, 138.6, 138.5, 138.4, 138.3, 138.2, 138.1, 128.5, 128.4, 128.31, 128.30, 128.28, 128.26, 128.24, 128.15, 128.14, 128.06, 128.05, 127.82, 127.77, 127.73, 127.65, 127.64, 127.61, 127.58, 127.57, 127.52, 127.45, 127.2, 114.9, 103.6, 102.8, 99.7, 83.0, 81.8, 81.6, 79.4, 77.7, 75.3, 75.2, 75.2, 75.1, 75.0, 74.9, 73.2, 73.14, 73.08, 73.06, 73.00, 72.2, 70.0, 69.29, 69.25, 68.4, 67.8, 67.7, 60.4, 30.3, 29.0; ESI-MS calcd for C₈₆H₉₄O₁₆Na⁺ 1405.64 (M + Na)⁺; found: 1406.02.

4-Pentenyl 2,3,4-tri-*O*-benzyl- α -L-fucopyranosyl-(1 \rightarrow 2)-3,4-carbonyl-6-*O*-tri-isopropylsilyl- β -D-galactopyranosyl-(1 \rightarrow 3)-6-*O*-tri-isopropylsilyl-2-deoxy-2-phenylsulfonylamino- β -D-galactopyranosyl-(1 \rightarrow 3)-2,4,6-tri-*O*-benzyl- α -D-galactopyranosyl-(1 \rightarrow 4)-2,3,6-tri-*O*-benzyl- β -D-galactopyranosyl-(1 \rightarrow 4)-2,3,6-tri-*O*-benzyl- β -D-glucopyranoside (9).² The DEF donor **7** (2.13 g, 1.66 mmol) and ABC acceptor **8** (1.15 g, 0.831 mmol) were combined, azeotroped twice with anhydrous benzene (2 \times 20 mL) and placed under high vacuum for 5 h. The mixture was then dissolved in CH₂Cl₂ (20 mL) and Et₂O (40 mL), treated with freshly activated 4 Å molecular sieves (2 g) and 2,6-di-*tert*-butylpyridine (734 μ L, 3.32 mmol), then cooled to -78 °C. Methyl triflate (4.0 equiv, 365 μ L) was added in one portion and the reaction was allowed to warm to rt slowly overnight. The reaction was quenched by the addition of Et₃N (5 mL), and filtered through Celite with Et₂O. The filtrate was washed with saturated NaHCO₃, and brine, and dried over MgSO₄. Concentration and purification by silica gel chromatography (15-20% EtOAc/hexane) afforded 1.551 g (72%) of hexasaccharide **9**. ¹H NMR (CDCl₃, 500 MHz): δ 7.66 (d, *J* = 7.3 Hz, 2H), 7.35-6.94 (m, 63H), 5.77-5.69 (m, 1H), 5.10 (s, 1H), 5.02-4.99 (m, 2H), 4.95-4.91 (m, 3H), 4.89-4.87 (m, 1H), 4.84-4.51 (m, 11H), 4.43-4.31 (m, 4H), 4.26-4.19 (m, 3H), 4.11-3.13 (m, 40H), 2.98-2.95 (m, 1H), 2.78-2.77 (m, 1H), 2.66-2.64 (m, 1H), 2.09-2.04 (m, 2H), 1.68-1.62 (m, 2H), 1.13 (d, *J* = 6.4 Hz, 3H), 1.03-0.97 (m, 42H); ¹³C NMR (CDCl₃, 150 MHz): δ 155.7, 141.0, 139.6, 139.2, 138.99, 138.96, 138.9, 138.8, 138.7, 138.6, 138.4, 138.3, 138.2, 132.3, 129.1, 128.9, 128.78, 128.75, 128.6, 128.54, 128.46, 128.43, 128.40, 128.37, 128.32, 128.30, 128.26, 128.16, 128.13, 127.96,

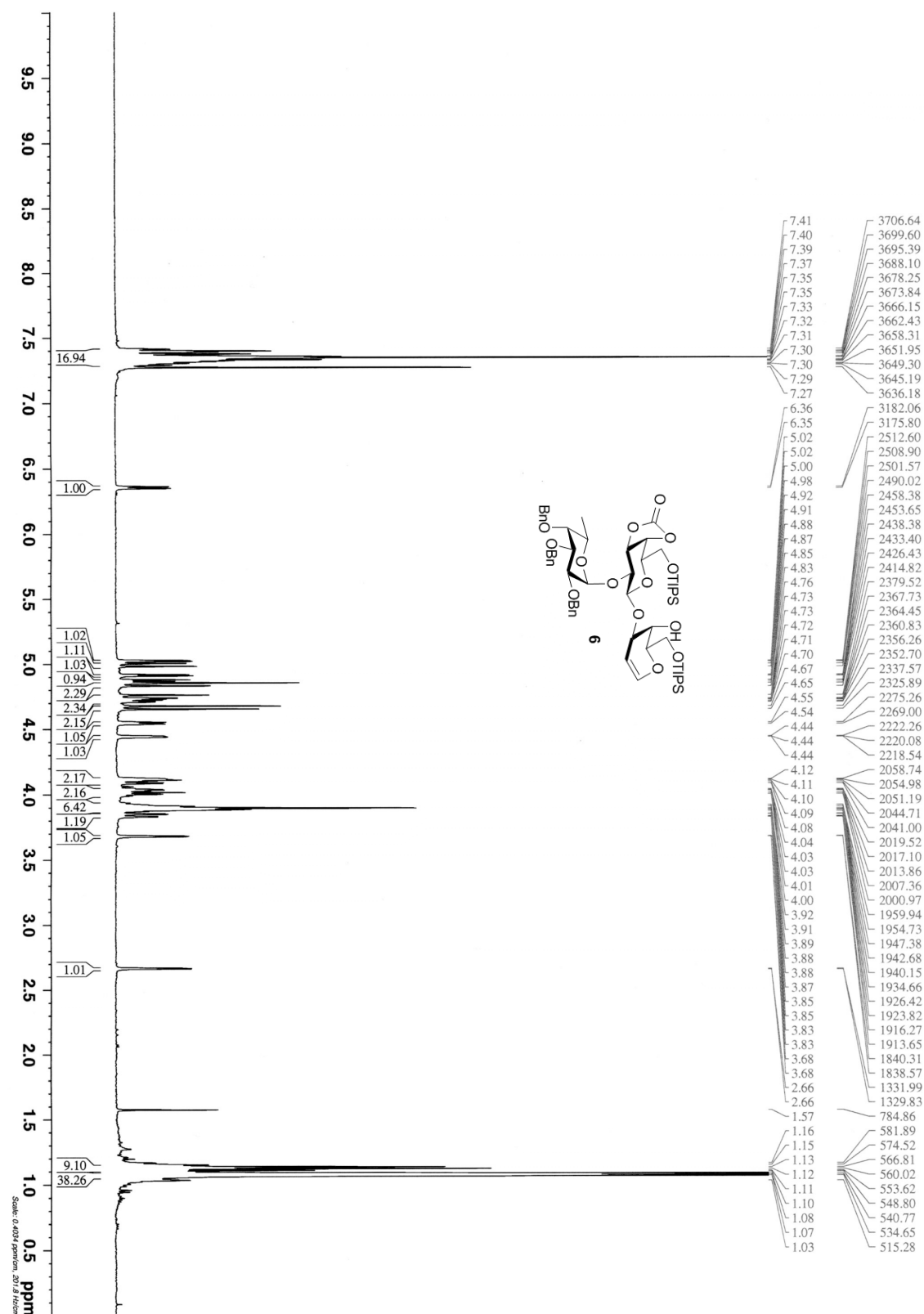
127.89, 127.86, 127.85, 127.8, 127.72, 127.66, 127.61, 127.59, 127.55, 127.5, 127.4, 115.1, 103.8, 103.4, 102.9, 99.9, 99.4, 98.1, 83.5, 81.9, 81.6, 80.7, 79.6, 79.2, 78.5, 78.1, 77.9, 77.7, 76.7, 75.6, 75.5, 75.4, 75.3, 75.2, 75.0, 74.2, 73.4, 73.3, 73.2, 72.8, 72.2, 72.1, 71.6, 71.5, 69.6, 69.53, 69.46, 68.5, 68.4, 68.3, 68.0, 67.8, 62.2, 62.0, 56.3, 30.4, 29.2, 18.27, 18.25, 18.21, 18.18, 18.1, 17.9, 17.1, 12.12, 12.09; ESI-MS calcd for $C_{150}H_{185}NO_{32}SSi_2Na^+$ 2623.20 (M + Na)⁺; found: 2624.41.

4-Pentenyl 2,3,4-tri-O-acetyl- α -L-fucopyranosyl-(1 \rightarrow 2)-3,4,6-tri-O-acetyl- β -D-galactopyranosyl-(1 \rightarrow 3)-4,6-O-tri-acetyl-2-acetylamino-2-deoxy- β -D-galactopyranosyl-(1 \rightarrow 3)-2,4,6-tri-O-acetyl- α -D-galactopyranosyl-(1 \rightarrow 4)-2,3,6-tri-O-acetyl- β -D-galactopyranosyl-(1 \rightarrow 4)-2,3,6-tri-O-acetyl- β -D-glucopyranoside (10).² TBAF (1.0 M in THF, 37.34 mL, 60 equiv.) was added to a solution of the hexasaccharide **9** (1.62 g, 0.622 mmol) and acetic acid (2.14 mL, 60 equiv.) in THF (35 mL). The reaction was stirred at rt for 3 days, poured into ice water and extracted with EtOAc. The organic extracts were washed with saturated NaHCO₃ and brine, dried over MgSO₄ and concentrated to an oil which was purified through a short plug of silica gel with EtOAc. The resulting triol was dissolved in anhydrous MeOH (20 mL) and sodium methoxide was added (0.75 mL of a 25% solution in MeOH). The reaction was stirred at rt for 18 h, neutralized with Dowex-H⁺, filtered with MeOH washings and concentrated. THF (5 mL) and condensed liquid NH₃ (~70 mL) were added at -78 °C to the resulting white solid. Sodium (~1.2 g) was added and the resulting blue solution stirred at -78 °C for 2 h. The reaction was quenched with anhydrous MeOH (~30 mL), brought to rt and concentrated under a stream of dry N₂ to a volume of ~15 mL. The reaction was neutralized with Dowex-H⁺, filtered with MeOH washing and concentrated to a white solid. The white solid was dissolved in pyridine (10 mL) and CH₂Cl₂ (10 mL) and cooled to 0 °C. A crystal of DMAP was added followed by acetic anhydride (10 mL). The ice bath was removed and the reaction stirred at rt overnight. The mixture was diluted with EtOAc and washed with water, saturated NaHCO₃ and brine, dried over MgSO₄. Concentration followed by purification by flash column chromatography (gradient elution 60% EtOAc/CH₂Cl₂) gave **10** as a white solid (470 mg, 42%). ¹H NMR (600 MHz, CDCl₃) δ 6.74 (d, *J* = 6.5 Hz, 1H), 5.80-5.73 (m, 1H), 5.58 (d, *J* = 2.8 Hz, 1H), 5.44 (d, *J* = 3.4 Hz, 1H), 5.39 (d, *J* = 3.2 Hz, 1H), 5.27 (dd, *J* = 10.9, 3.0 Hz, 1H), 5.24-5.08 (m, 6H), 5.04-4.92 (m, 6H), 4.87 (t, *J* = 8.0 Hz, 1H), 4.74 (dd, *J* = 10.9, 2.5 Hz, 1H), 4.52-4.40 (m, 6H), 4.36 (t, *J* = 6.1 Hz, 1H), 4.26 (dd, *J* = 10.6, 3.4 Hz, 1H), 4.18-4.04 (m, 8H), 3.99 (d, *J* = 1.9 Hz, 1H), 3.95 (t, *J* = 6.3 Hz, 1H), 3.88-3.82 (m, 3H), 3.76-3.74 (m, 1H), 3.61-3.59 (m, 1H), 3.49-3.46 (m, 1H), 3.06-3.01 (m, 1H), 2.15-2.14 (m, 12H), 2.10-2.03 (m, 29H), 2.00 (s, 3H), 1.96 (s, 2 \times 3H), 1.95 (s, 3H), 1.90 (s, 3H), 1.71-1.59 (m, 2H), 1.14 (d, *J* = 6.5 Hz, 3H); ¹³C NMR (CDCl₃, 150 MHz) δ 172.5, 171.7, 171.0, 170.81, 170.78, 170.75, 170.7, 170.60, 170.59, 170.5, 170.3, 170.0, 169.8, 169.7, 169.2, 138.0, 115.3, 102.2, 101.5, 100.7, 99.5, 99.0, 94.6, 76.3, 73.9, 73.6, 73.4, 72.8, 72.6, 72.1, 72.0, 71.8, 71.5, 71.1, 70.91, 70.86, 70.4, 70.3, 69.5, 69.4, 69.1, 68.3, 68.2, 67.8, 67.5, 64.7, 62.5, 62.0, 61.6, 61.4, 61.2, 56.4, 30.0, 28.8, 23.3, 21.10, 21.09, 21.03, 21.00, 20.98, 20.95, 20.91, 20.89, 20.87, 20.83, 20.82, 20.80, 20.74, 20.68, 16.1; ESI-MS calcd for $C_{77}H_{107}NO_{47}Na^+$ 1820.59 (M + Na)⁺; found: 1821.17.

References

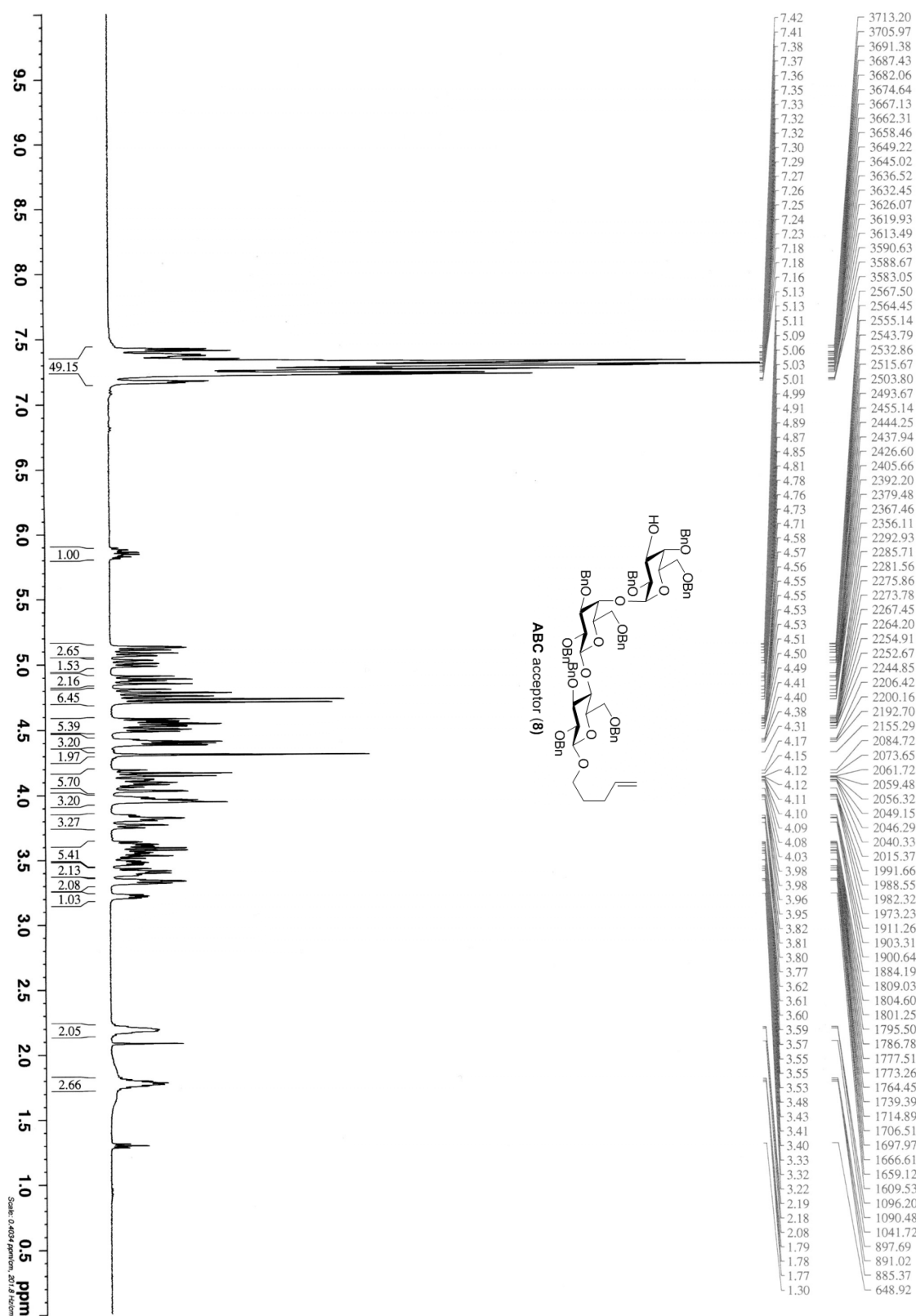
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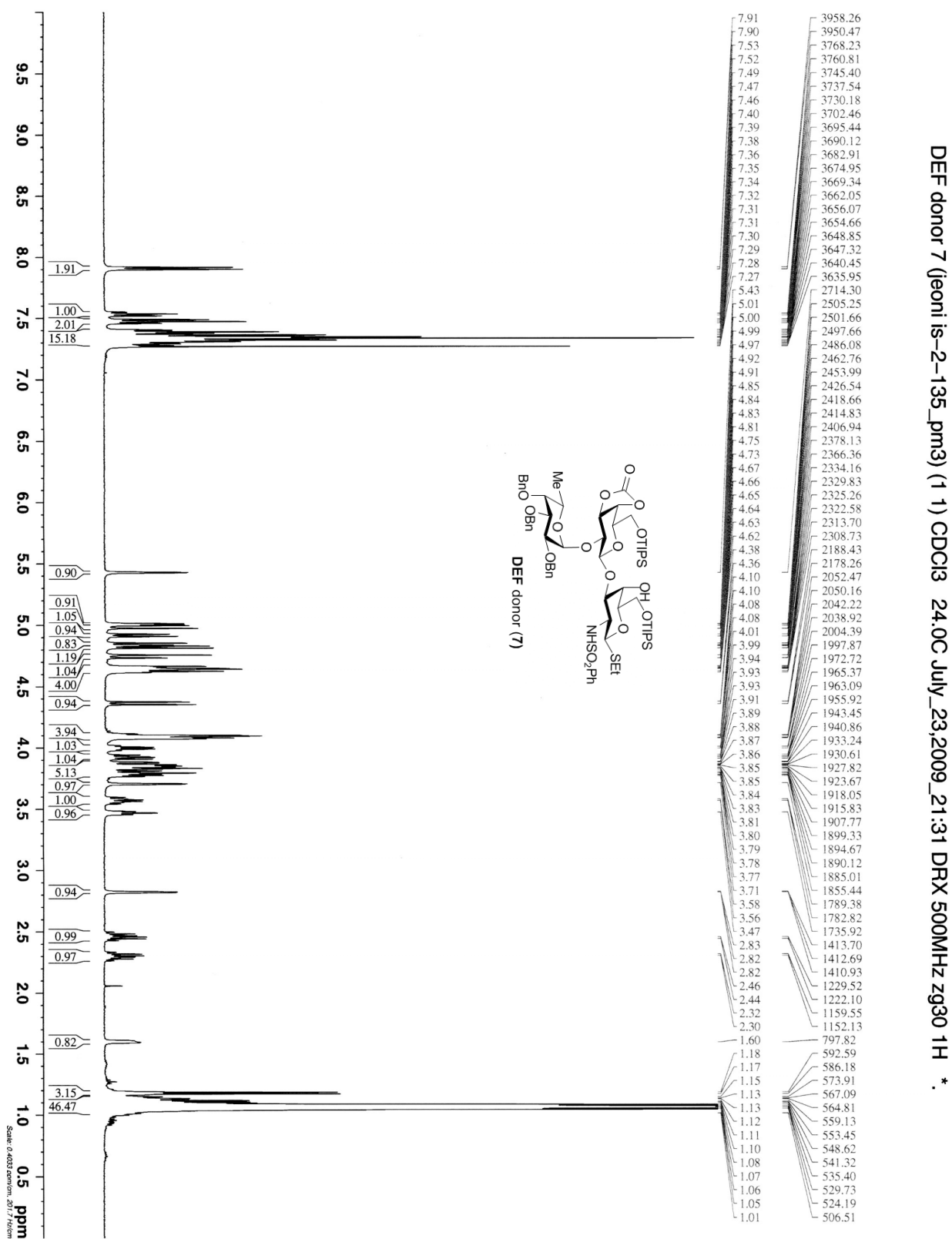


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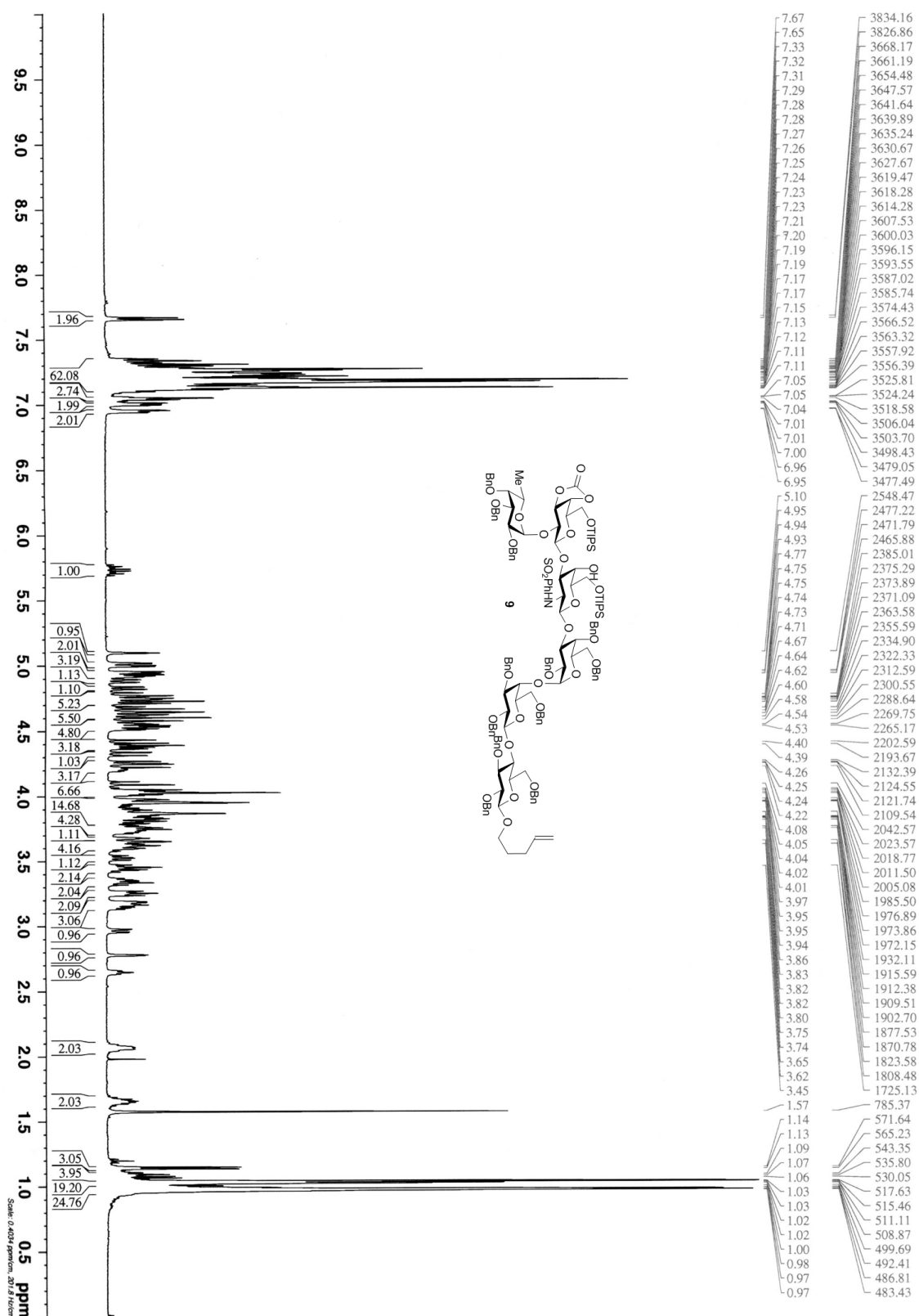
500 MHz ^1H NMR spectrum of ABC acceptor **8** in CDCl_3



500 MHz ^1H NMR spectrum of DEF donor 7 in CDCl_3

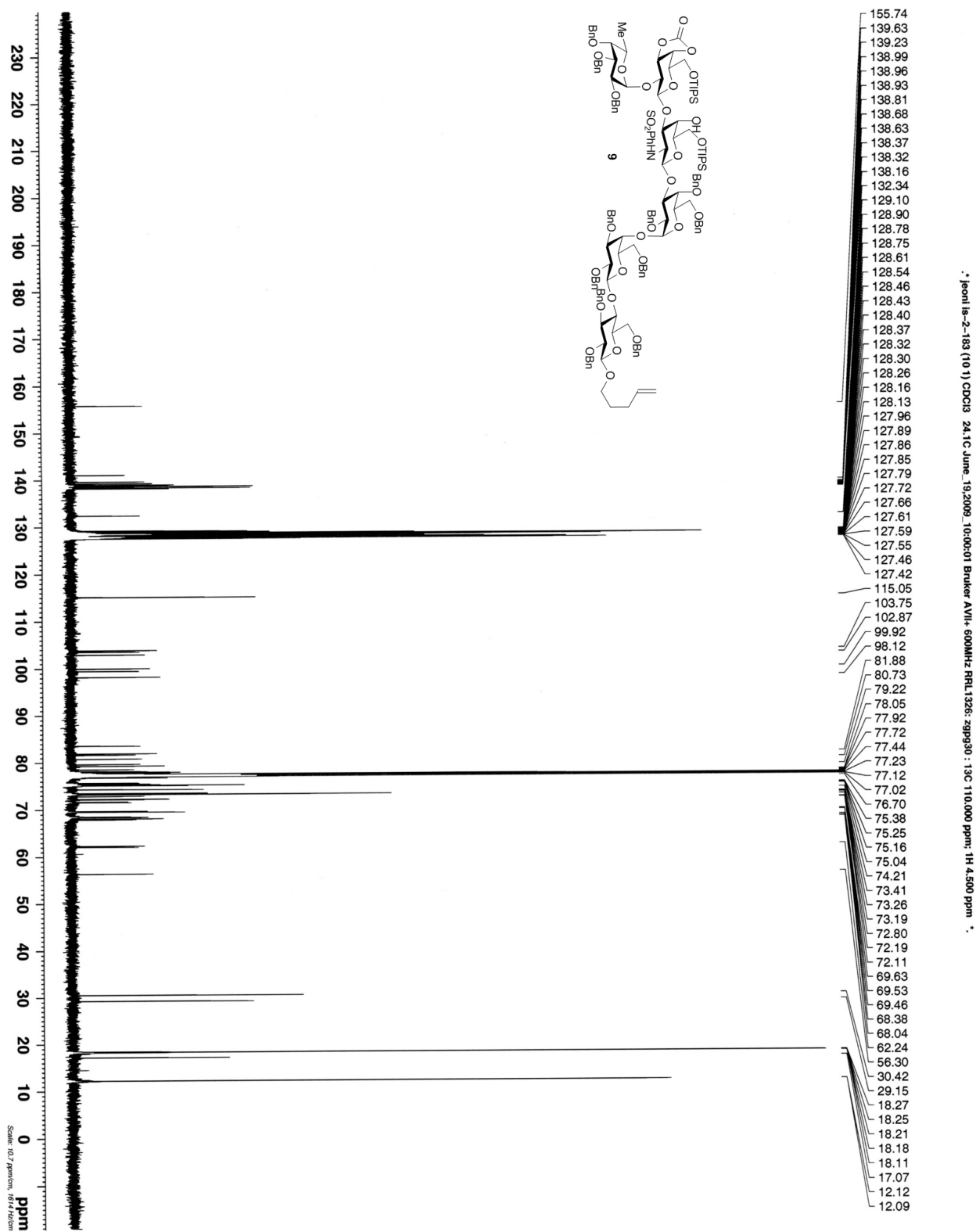


500 MHz ^1H NMR spectrum of hexasaccharide **9** in CDCl_3

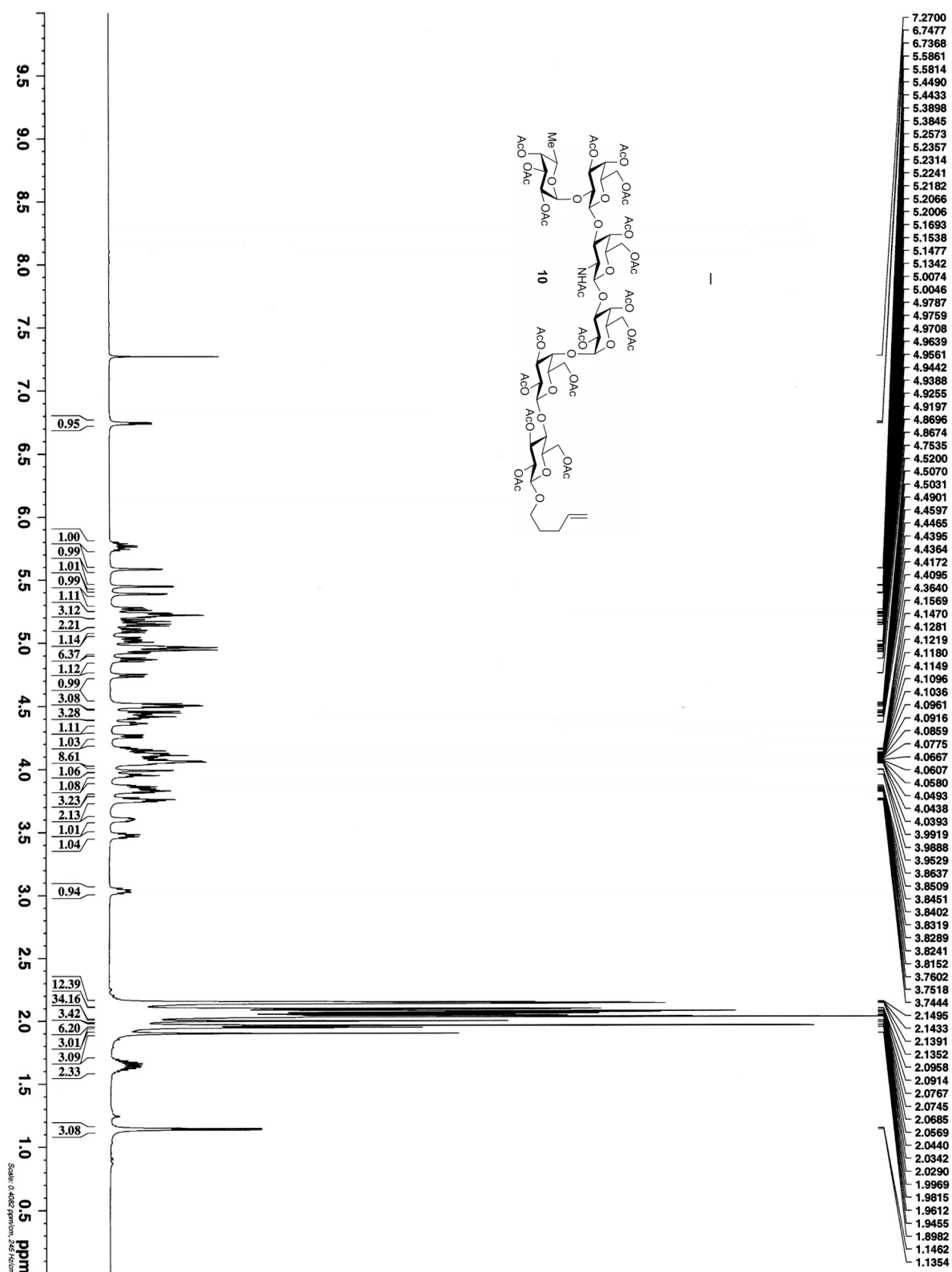


hexasaccharide **9** is-2-183 (1) CDCl_3 24.0C June_19,2009_09:10 DRX 500MHz zg30 1H *

150 MHz ^{13}C NMR spectrum of hexasaccharide **9** in CDCl_3



600 MHz ^1H NMR spectrum of hexasaccharide **10** in CDCl_3



150 MHz ^{13}C NMR spectrum of hexasaccharide **10** in CDCl_3

