

Supporting Information for

3,3'-Anisyl-Substituted BINOL, H₄BINOL and H₈BINOL Ligands:

**Asymmetric Synthesis of Diverse Propargylic Alcohols and Their
Ring-Closing Metathesis to Chiral Cycloalkenes**

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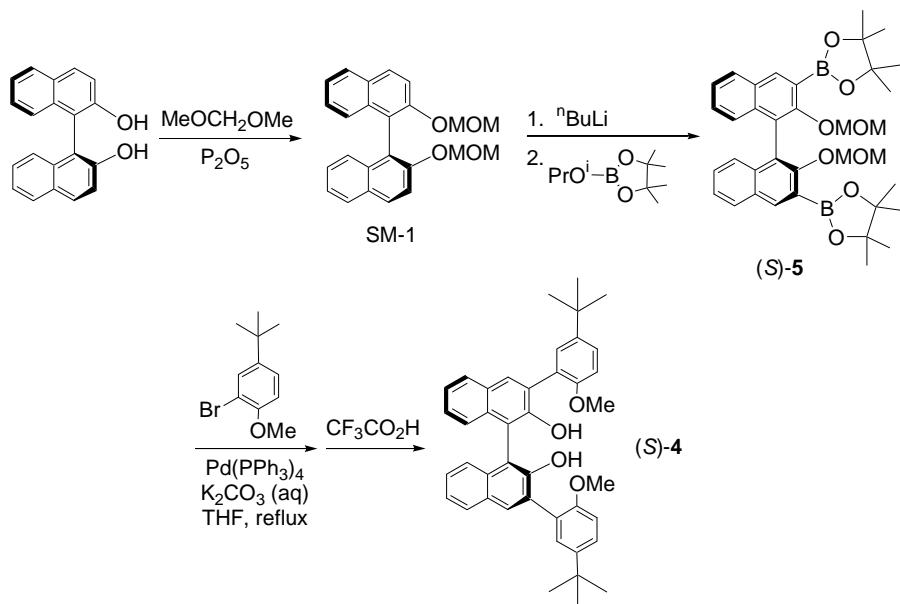
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General Data:

Toluene, THF, and 1,4-dioxane were distilled over sodium and benzophenone under nitrogen atmosphere. Methylene chloride and diethyl ether were dried by passing through activated alumina columns under nitrogen. Solvents were stored over 4 Å molecular sieves.

Methyl propiolate was distilled under reduced pressure from 4 Å molecular sieves prior to use. All aldehydes were passed through a plug of alumina and distilled from 4 Å molecular sieves prior to use.

Preparation of the Chiral Ligands

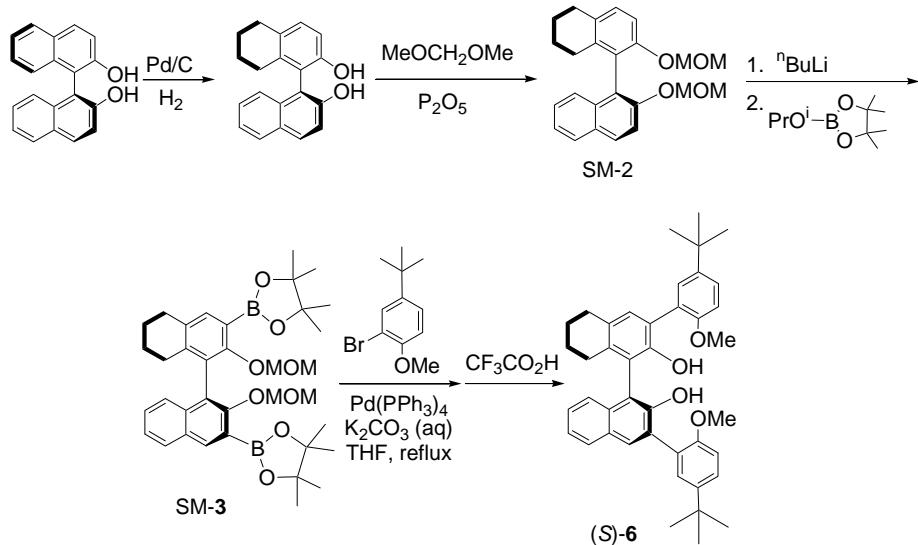


Preparation of (S)-2,2'-bis(methoxymethoxy)-1,1'-binaphthyl, SM-1. To a 250 mL flask, (S)-BINOL (6.0 g, 21 mmol), $\text{CH}_3\text{OCH}_2\text{OCH}_3$ (21.0 g, 271 mmol) and 150 mL CH_2Cl_2 were added. P_2O_5 (25 g) was then added to the reaction mixture with 5 portions. After the mixture was stirred for 1 h, the reaction was quenched with Na_2CO_3 (saturated, aqueous 100 mL). The organic layer was separated and the aqueous layer was extracted with CH_2Cl_2 (2 x 50 mL). The resulting organic layers were dried and concentrated. The residue was purified by flash chromatography on silica gel eluted with 10% $\text{EtOAc}/\text{hexanes}$ to yield the product as a white solid in 84% yield. $^1\text{H NMR}$ (300 MHz, CDCl_3) δ 3.15 (s, 6H), 4.98 (d, 2H, $J = 6.9$ Hz), 5.09 (d, 2H, $J = 6.6$ Hz), 7.15-7.25 (m, 4H), 7.32-7.37 (m, 2H), 7.58 (d, 2H, $J = 9.0$ Hz), 7.88 (d, 2H, $J = 8.1$ Hz), 7.96 (d, 2H, $J = 9.0$ Hz).

Preparation of 2-((1'S)-2,2'-bis(methoxymethoxy)-3'-(4,4,5-trimethyl-1,3,2-dioxa-borolan-2-yl)-1,1'-binaphthyl-3-yl)-4,4,5,5-tetramethyl-1,3,2-dioxaborolane, (S)-5. (S)-2,2'-Bis(methoxymethoxy)-1,1'-binaphthyl (5.0 g, 13 mmol) was dissolved in hexanes and cooled to 0 °C under nitrogen. $n\text{BuLi}$ (2.4 M, 22 mL, 52 mmol) was added and the

reaction mixture was allowed to warm to room temperature over 3 h, during which a white precipitate formed. The reaction mixture was cooled to -78 °C and 2-isopropoxy-4,4,5,5-tetramethyl-1,3,2-dioxaborolane (15 mL, 73.6 mmol) in THF (30 mL) was added via cannula. The mixture was allowed to warm to room temperature and stirred for 16 h. The generated salts were removed by filtration through a Buchner funnel and rinsed with CH₂Cl₂. The combined organic layer was concentrated and purified by flash chromatography over a short silica gel column (10% EtOAc/hexanes), yielding (*S*)-**5** as a white solid in 80% yield. (Extended time on the column would result in partial hydrolysis to the boronic acid.) ¹H NMR (300 MHz, CDCl₃) δ 1.42 (s, 24H), 2.30 (s, 6H), 4.91 (s, 4H), 7.22 (d, 2H, J = 8.4 Hz), 7.26-7.32 (m, 2H), 7.39-7.43 (m, 2H), 7.93 (d, 2H, J = 8.1 Hz), 8.49 (s, 2H). ¹³C NMR (75 MHz, CDCl₃) δ 25.2, 55.8, 84.1, 100.3, 124.9, 126.1, 127.0, 127.4, 128.5, 130.4, 136.4, 139.4, 157.3.

Preparation of (*S*)-3,3'-bis(5-tert-butyl-2-methoxyphenyl)-1,1'-binaphthyl-2,2'-diol, (*S*)-4**.** Under nitrogen, (*S*)-**5** (3.15 g, 5 mmol), 2-bromo-4-tert-butyl-1-methoxybenzene (4.86 g, 20 mmol, 4 equiv), and Pd(PPh₃)₄ (410 mg, 0.5 mmol, 10 mol%) were placed in a 2-neck flask equipped with a reflux condenser that was connected with a vacuum adaptor. Degassed THF (50 mL) and 2M K₂CO₃ (40 mL) were transferred into the flask via cannula. To ensure the removal of oxygen, the reaction vessel was freeze-pumped at -78 °C and refilled with nitrogen three times. The reaction mixture was then heated at 95 °C for 24 h. After cooled to room temperature, the mixture was extracted with CH₂Cl₂ (3 x 40 mL). The combined organic layer was washed with 2M HCl (3 x 40 mL), dried, and concentrated. The crude product was dissolved in CH₂Cl₂ (25 mL) and trifluoroacetic acid (5 mL) was added. After the reaction mixture was stirred at room temperature for 10 h, it was concentrated and purified by column chromatography on silica gel eluted with 10% EtOAc/hexanes to afforded (*S*)-**4** as a white solid in 84% yield. ¹H NMR (300 MHz, CDCl₃) δ 1.37 (s, 18H), 3.81 (s, 6H), 5.95 (s, 2H), 6.97 (d, 2H, J = 8.7 Hz), 7.26-7.44 (m, 8H), 7.53 (d, 2H, J = 2.4 Hz), 7.90-7.94 (m, 4H). These data are consistent with those reported.¹



¹ Moore, D.; Huang, W. -S.; Pu, L. *Tetrahedron Lett.* **2002**, *43*, 8831-8834.

Preparation of (S)-H₄BINOL and (S)-H₈BINOL. The reaction was carried out by slightly modifying a literature procedure.² (S)-BINOL (5.0 g, 17.5 mmol), 5% Pd/C (0.75 g) and ethanol (95%, 50 mL) were placed in a parr reactor and stirred under 700 psi hydrogen pressure at 70 °C. The reaction was monitored by TLC. Usually (S)-BINOL disappeared in 6 h. A longer reaction time and higher temperature will generate more (S)-H₈BINOL. The mixture was then cooled down to room temperature. After the catalyst was removed by filtration and rinsed with ethanol (3 x 50 mL), the combined filtrates were concentrated in vacuum. The residue was purified by flash chromatography on silica gel eluted with 10% EtOAc/hexanes to give (S)-H₄BINOL³ [~60% yield, more polar than (S)-H₈BINOL and less polar than (S)-BINOL] and (S)-H₈BINOL (~40% yield).

Preparation of (S)-2,2'-bis(methoxymethoxy)-5,6,7,8-tetrahydro-1,1'-binaphthyl, SM-2. To a 250 mL flask, (S)-H₄BINOL (6 g, 21 mmol), CH₃OCH₂OCH₃ (21 g, 271 mmol) and 150 mL CH₂Cl₂ were added. P₂O₅ (25 g) was then added to the reaction mixture with 5 portions. After the reaction mixture was stirred for 1 h, it was quenched with Na₂CO₃ (saturated, aqueous, 100 mL). The organic layer was separated and the aqueous layer was extracted with CH₂Cl₂ (2 x 50 mL). The resulting organic layers were dried and concentrated. The residue was purified by flash chromatography on silica gel eluted with 10% EtOAc/hexanes to yield the product as a white solid in 84% yield. ¹H NMR (300 MHz, CDCl₃) δ 1.56-1.63 (m, 2H), 1.72-1.75 (m, 2H), 2.02-2.12 (m, 1H), 2.26-2.36 (m, 1H), 2.83 (t, 2H, J = 6.0 Hz), 3.08 (s, 3H), 3.35 (s, 3H), 4.84 (d, 1H, J = 6.9 Hz), 4.93 (d, 1H, J = 6.6 Hz), 5.09 (d, 1H, J = 6.9 Hz), 5.17 (d, 1H, J = 6.9 Hz), 7.05 (d, 1H, J = 8.4 Hz), 7.14 (d, 1H, J = 8.4 Hz), 7.29-7.38 (m, 3H), 7.51 (d, 1H, J = 9.0 Hz), 7.81-7.87 (m, 2H). ¹³C NMR (75 MHz, CDCl₃) δ 23.4, 27.4, 29.7, 55.8, 56.2, 95.0, 95.6, 113.2, 117.5, 124.2, 125.3, 126.5, 128.1, 129.1, 129.6, 130.1, 131.3, 133.6, 138.2, 152.0, 153.2.

Preparation of 2-((1'S)-2,2'-bis(methoxymethoxy)-3'-(4,4,5-trimethyl-1,3,2-dioxaborolan-2-yl)-5',6',7',8'-tetrahydro-1,1'-binaphthyl-3-yl)-4,4,5,5-tetramethyl-1,3,2-dioxaborolane, SM-3. Under nitrogen, (S)-2,2'-bis(methoxymethoxy)-5,6,7,8-tetrahydro-1,1'-binaphthyl (5.0 g, 13 mmol) was dissolved in hexanes and cooled to 0 °C. ⁿBuLi (2.4 M, 22 mL, 52 mmol) was added and the reaction mixture was allowed to warm to room temperature over 3 h, during which a white precipitate formed. It was then cooled to -78 °C and 2-isopropoxy-4,4,5,5-tetramethyl-1,3,2-dioxaborolane (15 mL, 73.6 mmol) in THF (30 mL) was added via cannula. The mixture was allowed to warm to room temperature and stirred for 16 h. The generated salts were removed by filtration through a Buchner funnel and rinsed with CH₂Cl₂. The combined organic layer was concentrated and purified by flash chromatography over a short silica gel column (10% EtOAc/hexanes), yielding SM-3 as a white solid in 80% yield. (Extended time on the column would result in partial hydrolysis to the boronic acid.) ¹H NMR (300 MHz, CDCl₃) ¹H NMR (300 MHz, CDCl₃) δ 1.32 (s, 12H), 1.37 (s, 12H), 1.53-1.59 (m, 2H), 1.66-1.76 (m, 2H), 2.07-2.15 (m, 1H), 2.17 (s, 3H), 2.36-2.46 (m, 1H), 2.76 (s, 3H), 2.79-2.85 (m, 2H), 4.72 (d, 1H, J = 6.0 Hz), 4.79 (d, 1H, J = 5.7 Hz), 4.95 (d, 1H, J = 6.3

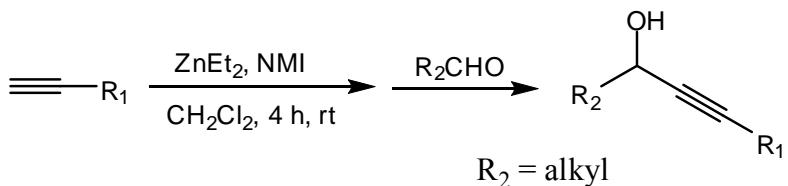
² Korostylev, A.; Tararov, V. I.; Fischer, C.; Monsees, A.; Brner, A. *J. Org. Chem.* **2004**, 69, 3220-3221.

³ Heumann, L. V.; Keck, G. E. *J. Org. Chem.* **2008**, 73, 4725-4727.

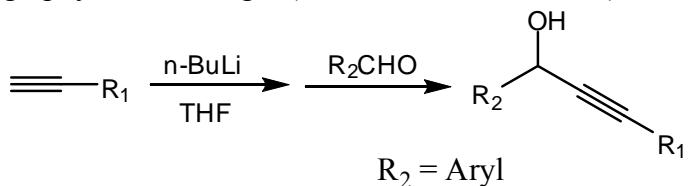
Hz), 5.05 (d, 1H, J = 6.3 Hz), 7.35-7.36 (m, 3H), 7.57 (s, 1H), 7.82-7.85 (m, 1H), 8.36 (s, 1H). ^{13}C NMR (75 MHz, CDCl_3) δ 23.3, 25.1, 28.2, 29.8, 55.6, 56.3, 83.7, 84.0, 100.4, 100.6, 124.8, 126.4, 127.4, 128.6, 130.4, 132.7, 135.6, 137.4, 139.0, 142.7, 156.5, 159.2.

Partial racemization of (*S*)-4, (*S*)-6, and (*S*)-8 for determination of the enantiomeric purity. Under nitrogen, (*S*)-4, (*S*)-6, and (*S*)-8 (10 mg) were stirred in NMP (2 mL) at 190 °C for 40 h. After cooled, the mixture was diluted with diethyl ether and washed three times with water. The aqueous layer was extracted once with diethyl ether. The combined organic layers were dried, concentrated, and purified by flash chromatography over a short silica gel column (10% EtOAc/hexanes). The purity of the racemized ligands was confirmed by ^1H NMR spectroscopy, and the partially racemized ligands were separated by HPLC (OD column, 99:1 hexanes: $^i\text{PrOH}$, flow rate = 0.3 mL/min).

Preparation of Racemic Propargylic Alcohols



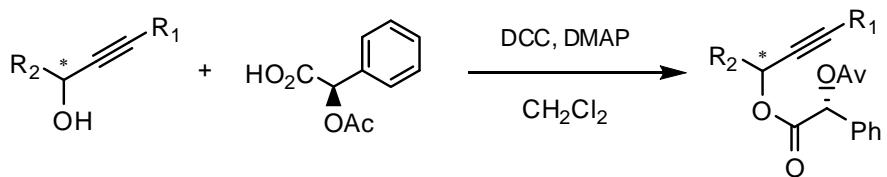
Alkyne (0.5 mmol) was dissolved in CH_2Cl_2 . ZnEt_2 (51.5 μL , 0.5 mmol) and N-methylimidazole (2 μL , 0.025 mmol) were added and the mixture was stirred for 4 h. An aldehyde (0.25 mmol) was added, and the reaction mixture was stirred overnight. The reaction was quenched with saturated aqueous ammonium chloride (5 mL), extracted with CH_2Cl_2 (3 x 10 mL), dried, and concentrated. The racemic alcohols were purified by flash chromatography over silica gel (10-20% EtOAc/hexanes).⁴



Under nitrogen, a solution of alkyne (0.75 mmol) in THF (3 mL) in a 10 mL round bottom flask was cooled to -78 °C by a dry ice/acetone bath, and $^n\text{BuLi}$ (0.12 mL, 2.5 M in hexane) was added. The mixture was stirred for 15 min. An aldehyde (0.25 mmol) was then added and the reaction mixture was continuously stirred for another 15 min. Ice was added to quench the reaction and methylene chloride was used for extraction. The extract was dried over magnesium sulfate. After the volatile solvent was removed by roto-evaporation, the residue was passed through a short silica gel eluted with petroleum ether/ethyl acetate (9/1) to afford the product.

⁴ Yue, Y.; Yu, X.-Q.; Pu, L. *Chem. Eur. J.* **2009**, 15, 5104-5107.

Preparation of the Mandelic Acetate Derivatives of the Propargylic Alcohols for ^1H NMR Analysis



A propargylic alcohol (20 mg), DCC (2 equiv), DMAP (2 equiv), and (*R*)-O-acetylmandelic acid (2 equiv) were dissolved in CH_2Cl_2 (5 mL). The reaction was monitored by TLC. After consumption of the starting material (about 20 min), the crude mixture was passed through a short silica gel column eluted with 30% EtOAc/hexanes. The ee determination was based on the proton signal at $\delta \sim 5.9$ ppm in the ^1H NMR spectrum.

Characterization of the New Optically Active Propargylic Alcohol Products:

1-Phenylnon-3-yn-5-ol, P-1. 78% yield. 84% ee determined by HPLC analysis: AD column, 99:1 hexanes: $^i\text{PrOH}$, flow rate = 0.3 mL/min, λ = 221 nm, retention time: $t_{\text{major}} = 51.43$ min $t_{\text{minor}} = 54.968$ min. $[\alpha]_D = -6.20$ ($c = 0.34$, THF). ^1H NMR (300 MHz, CDCl_3) δ 0.93-0.98 (t, 3H, $J = 7.2$ Hz), 1.37-1.48 (m, 4H), 1.65-1.74 (m, 2H), 2.02 (b, 1H), 2.55 (td, 2H, $J_t = 8.4$ Hz, $J_d = 2.1$ Hz), 2.87 (t, 2H, $J = 7.5$ Hz), 4.37 (tt, 1H, $J_1 = 6.6$ Hz, $J_2 = 2.1$ Hz), 7.25-7.37 (m, 5H). ^{13}C NMR (75 MHz, CDCl_3) δ 14.3, 21.2, 22.7, 27.6, 35.3, 38.1, 63.0, 82.5, 84.9, 126.6, 128.6, 128.7, 140.9. HRMS (M-H^+) for $\text{C}_{15}\text{H}_{19}\text{O}$ Calcd: 215.1430. Found: 215.1434.

9-Phenylnon-1-en-6-yn-5-ol, P-2. 67% yield. 80% ee determined by HPLC analysis: AD column, 99:1 hexanes: $^i\text{PrOH}$, flow rate = 0.3 mL/min, λ = 254 nm, retention time: $t_{\text{major}} = 58.59$ min $t_{\text{minor}} = 62.91$ min. $[\alpha]_D = -2.58$ ($c = 1.09$, THF). ^1H NMR (300 MHz, CDCl_3) δ 1.76-1.84 (m, 2H), 2.10 (b, 1H), 2.19-2.27 (m, 2H), 2.57 (t, 2H, $J = 7.5$ Hz), 2.88 (t, 2H, $J = 7.5$ Hz), 4.41 (b, 1H), 5.03-5.14 (m, 2H), 5.81-5.95 (m, 1H), 7.28-7.38 (m, 5H). ^{13}C NMR (75 MHz, CDCl_3) δ 21.2, 29.7, 35.3, 37.3, 62.4, 82.2, 85.2, 115.4, 126.6, 128.7, 128.8, 138.1, 140.8. HRMS (MH^+) for $\text{C}_{15}\text{H}_{19}\text{O}$ Calcd: 215.1430. Found: 215.1429.

Ethyl 4-hydroxynon-8-en-2-ynoate, P-3. 63% yield. 94% ee by HPLC analysis: OD column, 98:2 hexanes: $^i\text{PrOH}$, flow rate = 1.0 mL/min, λ = 221 nm, retention time: $t_{\text{major}} = 20.36$ min $t_{\text{minor}} = 16.53$ min. $[\alpha]_D = -7.35$ ($c = 1.10$, THF). ^1H NMR (300 MHz, CDCl_3) δ 1.30 (t, 3H, $J = 7.0$ Hz), 1.57-1.62 (m, 2H), 1.74-1.81 (m, 2H), 2.09 (q, 2H, $J = 7.1$ Hz), 2.26 (b, 1H), 4.23 (q, 1H, $J = 7.1$ Hz), 4.49 (t, 2H, $J = 6.6$ Hz), 4.96-5.05 (m, 2H), 5.72-5.85 (m, 1H). ^{13}C NMR (75 MHz, CDCl_3) δ 14.2, 24.3, 29.9, 33.4, 36.4, 62.2, 62.4, 87.9, 115.3, 138.2, 153.6. HRMS (MH^+) for $\text{C}_{11}\text{H}_{17}\text{O}_3$ Calcd: 197.1172. Found: 197.1171.

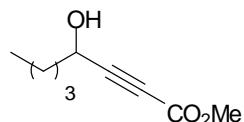
1-Phenyloct-7-en-1-yn-3-ol, P-4. 84% yield. 87% ee determined by analyzing the ^1H

NMR spectrum of the (*R*)-mandalate ester derivative. $[\alpha]_D = -2.11$ ($c = 1.09$, THF). ^1H NMR (300 MHz, CDCl_3) δ 1.64-1.72 (m, 2H), 1.83-1.90 (m, 2H), 2.13-2.20 (m, 2H), 2.45 (bs, 1H), 4.66 (q, 2H, $J = 5.1$ Hz), 5.00-5.12 (m, 2H), 5.75-5.88 (m, 1H). ^{13}C NMR (75 MHz, CDCl_3) δ 24.76, 33.61, 37.54, 63.05, 85.17, 90.44, 115.13, 122.92, 128.56, 128.65, 131.96, 138.71. HRMS (MH^+) for $\text{C}_{14}\text{H}_{17}\text{O}$ Calcd: 201.1279. Found: 201.1284.

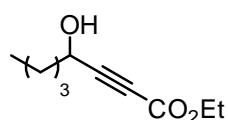
1-Phenyldec-9-en-3-yn-5-ol, P-5. 74% yield. 81% ee determined by analyzing the ^1H NMR spectrum of the (*R*)-mandalate ester derivative. $[\alpha]_D = -4.07$ ($c = 1.01$, THF). ^1H NMR (300 MHz, CDCl_3) δ 1.53 (q, 2H, $J = 6.0$ Hz), 1.63-1.72 (m, 2H), 2.00 (d, 1H, $J = 5.1$ Hz), 2.09 (q, 2H, $J = 6.6$ Hz), 2.52 (t, 2H, $J = 7.5$ Hz), 2.83 (t, 2H, $J = 7.5$ Hz), 4.34 (d, 1H, $J = 5.7$ Hz), 4.99-5.08 (m, 2H), 5.75-5.89 (m, 1H), 7.21-7.31 (m, 5H). ^{13}C NMR (75 MHz, CDCl_3) δ 21.1, 24.6, 33.6, 35.3, 37.7, 62.7, 82.3, 84.9, 115.0, 126.6, 128.6, 128.7, 138.8, 140.8. HRMS (MH^+) for $\text{C}_{16}\text{H}_{21}\text{O}$ Calcd: 229.1587. Found: 229.1592.

1-(4-Chlorophenyl)-5-phenylpent-2-yn-1-ol, P-6. 72% yield. 80% ee determined by analyzing the ^1H NMR spectrum of the (*R*)-mandalate ester derivative. $[\alpha]_D = -19.24$ ($c = 1.01$, THF). ^1H NMR (300 MHz, CDCl_3) δ 2.23 (d, 2H, $J = 6.1$ Hz), 2.58 (td, 2H, $J_t = 7.5$ Hz, $J_d = 1.8$ Hz), 2.86 (t, 2H, $J = 7.5$ Hz), 5.38 (dt, 2H, $J_d = 6.0$ Hz, $J_t = 1.8$ Hz), 7.20-7.39 (m, 9H). ^{13}C NMR (75 MHz, CDCl_3) δ 21.1, 35.0, 64.3, 80.7, 87.4, 126.6, 128.3, 128.7, 128.8, 128.8, 134.2, 139.7, 140.6. HRMS ($\text{M}-\text{H}^-$) for $\text{C}_{17}\text{H}_{14}\text{OCl}$ Calcd: 269.0728. Found: 269.0725.

Characterization of the Known Propargylic Alcohols



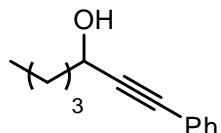
Methyl 4-hydroxyoct-2-yneate, SM-aa. Reaction time: 10 h. 84% yield. 94% ee determined by HPLC analysis: Chiracel OD column, 98:2 hexanes: $^i\text{PrOH}$, flow rate = 0.5 mL/min, $\lambda = 221$ nm, retention time: $t_{\text{major}} = 16.44$ min $t_{\text{minor}} = 14.97$ min. $[\alpha]_D = -4.30$ ($c = 1.01$, CHCl_3). ^1H NMR (300 MHz, CDCl_3) δ 0.92 (t, 3H, $J = 7.2$ Hz), 1.32-1.39 (m, 4H), 1.73-1.79 (m, 2H), 1.94-1.96 (m, 1H), 3.79 (s, 3H), 4.5 (t, 1H, $J = 6.2$ Hz). These data are consistent with those reported.⁵



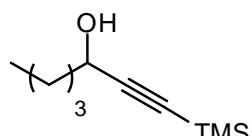
Ethyl 4-hydroxyoct-2-yneate, SM-ab. 60% yield. 94% ee determined by HPLC analysis: Chiracel OD column, 98:2 hexanes: $^i\text{PrOH}$, flow rate = 1.0 mL/min, $\lambda = 221$ nm, retention time: $t_{\text{major}} = 17.24$ min $t_{\text{minor}} = 14.93$ min. ^1H NMR (300 MHz, CDCl_3) δ 0.93 (t, 3H, $J = 7.2$ Hz), 1.33 (t, 3H, $J = 7.2$ Hz), 1.42-1.52 (m, 4H), 1.75-1.82 (m, 2H), 2.66 (d, 1H, $J = 5.4$ Hz), 4.22-4.29 (m, 2H), 4.47-4.53 (m, 1H). These data are

⁵ Rajaram, A.R.; Pu, L. *Org. Lett.* **2006**, 8, 2019-2021.

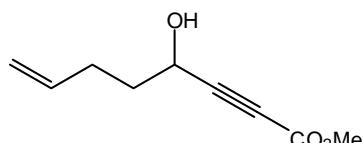
consistent with those reported.⁶



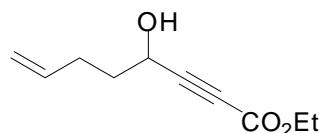
1-Phenylhept-1-yn-3-ol, SM-ac. 88% yield. 81% ee determined by HPLC analysis: Chiracel OD column, 90:10 hexanes:ⁱPrOH, flow rate = 1.0 mL/min, λ = 254 nm, retention time: $t_{\text{major}} = 18.55$ min $t_{\text{minor}} = 7.05$ min. ¹H NMR (300 MHz, CDCl₃) δ 0.93 (t, 3H, J = 7.2 Hz), 1.38-1.51 (m, 4H), 1.79-1.82 (m, 2H), 1.94 (d, 1H, J = 5.4 Hz), 4.59 (q, 1H, J = 6.2 Hz), 7.29-7.31 (m, 3H), 7.41-7.44 (m, 2H). These data are consistent with those reported.⁷



1-(Trimethylsilyl)hept-1-yn-3-ol, SM-ae. 65% yield. 97% ee determined by analyzing the ¹H NMR spectrum of the (R)-mandalate ester derivative. ¹H NMR (300 MHz, CDCl₃) δ 0.17 (s, 9H), 0.91 (t, 3H, J = 7.2 Hz), 1.29-1.47 (m, 4H), 1.66-1.71 (m, 2H), 1.77 (d, 1H, J = 5.4 Hz), 4.34 (q, 1H, J = 6.3 Hz). These data are consistent with those reported.⁸



Methyl 4-hydroxyoct-7-en-2-yoate, SM-ba. 63% yield. 95% ee determined by HPLC analysis: Chiracel OD column, 98:2 hexanes:ⁱPrOH, flow rate = 1.0 mL/min, λ = 221 nm, retention time: $t_{\text{major}} = 14.88$ min $t_{\text{minor}} = 12.84$ min. $[\alpha]_D = 9.99$ (c = 1.06, CHCl₃). ¹H NMR (300 MHz, CDCl₃) δ 1.84-1.91 (m, 2H), 1.97 (bs, 1H), 2.26 (q, 2H, J = 7.8 Hz), 3.79 (s, 3H), 4.53 (q, 1H, J = 5.7 Hz), 5.01-5.12 (m, 2H), 5.75-5.88 (m, 1H). ¹³C NMR (75 MHz, CDCl₃) δ 29.04, 35.71, 52.84, 61.38, 76.32, 87.97, 115.81, 136.92, 153.82. HRMS (MH⁺) for C₉H₁₃O₃ Calcd: 169.0859. Found: 169.0863.



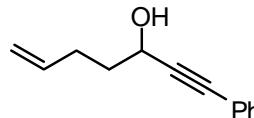
Ethyl 4-hydroxyoct-7-en-2-yoate, SM-bb. 60% yield. 95% ee determined by HPLC analysis: Chiracel OD column, 98:2 hexanes:ⁱPrOH, flow rate = 1.0 mL/min, λ = 221 nm, retention time: $t_{\text{major}} = 18.47$ min $t_{\text{minor}} = 16.27$ min. $[\alpha]_D = 13.57$ (c = 1.04, CHCl₃). ¹H NMR (300 MHz, CDCl₃) δ 1.32 (t, 3H, J = 7.1 Hz), 1.84-1.95 (m, 3H), 2.26 (q, 2H, J = 7.5 Hz), 4.25 (q, 2H, J = 7.2 Hz), 4.52 (q, 1H, J = 6.3 Hz), 5.03 (d, 1H, J = 5.4 Hz).

⁶ Herrmann, J. L.; Berger, M. H. ; Schlessinger, R. H. *J. Am. Chem. Soc.* **1979**, *101*, 1544-1549.

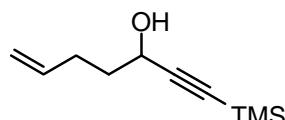
⁷ Gao, G.; Moore, D. ; Xie, R.-G. ; Pu, L. *Org. Lett.* **2002**, *4*, 4143-4146.

⁸ Ivanov, I. V.; Romanov, S. G.; Groza, N. V.; Nigam, S.; Kuhn, H.; Myagkova, G. I. *Bioorg. Med. Chem.* **2002**, *10*, 2335-2343.

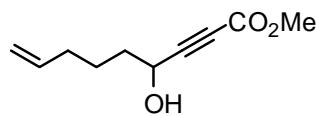
= 10.2 Hz), 5.09 (d, 1H, J= 17.1 Hz), 5.75-5.86 (m, 1H). These data are consistent with those reported.⁹



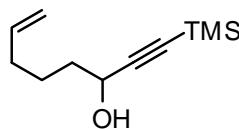
1-Phenylhept-6-en-1-yn-3-ol, SM-bc. 97% yield. 81% ee determined by HPLC analysis: Chiracel OD column, 90:10 hexanes:ⁱPrOH, flow rate = 1.0 mL/min, λ = 254 nm, retention time: $t_{\text{major}} = 21.33$ min $t_{\text{minor}} = 7.57$ min. ¹H NMR (300 MHz, CDCl₃) δ 1.89-1.95 (m, 2H), 1.99 (bs, 1H), 2.27-2.35 (m, 2H), 4.63 (t, 1H, J = 6.3 Hz), 5.00-5.14 (m, 2H), 5.81-5.94 (m, 1H), 7.30-7.32 (m, 3H), 7.42-7.45 (m, 2H). These data are consistent with those reported.¹⁰



1-(Trimethylsilyl)hept-6-en-1-yn-3-ol, SM-be. 69% yield. 94.5% ee determined by analyzing the ¹H NMR spectrum of the (*R*)-mandalate ester derivative. ¹H NMR (300 MHz, CDCl₃) δ 0.21 (s, 9H), 1.69-1.87 (m, 2H), 1.94 (d, 1H, J = 5.4 Hz), 2.23-2.30 (m, 2H), 4.41 (q, 1H, J = 6.0 Hz), 5.01-5.14 (m, 2H), 5.80-5.94 (m, 1H). These data are consistent with those reported.¹¹



Methyl 4-hydroxynon-8-en-2-yoate, SM-ca. 67% yield. 95% ee determined by HPLC analysis: Chiracel OD column, 98:2 hexanes:ⁱPrOH, flow rate = 1.0 mL/min, λ = 221 nm, retention time: $t_{\text{major}} = 25.96$ min $t_{\text{minor}} = 20.31$ min. ¹H NMR (300 MHz, CDCl₃) δ 1.57-1.65 (m, 2H), 1.77-1.84 (m, 2H), 2.09-2.16 (m, 2H), 2.47 (bs, 1H), 3.81 (s, 3H), 4.53 (d, 1H, J = 4.5 Hz), 4.99-5.08 (m, 2H), 5.75-5.89 (m, 1H). These data are consistent with those reported.¹²



1-(Trimethylsilyloct-7-en-1-yn-3-ol, SM-ce. 68% yield. 94% ee determined by analyzing the ¹H NMR spectrum of the (*R*)-mandalate ester derivative. ¹H NMR (300 MHz, CDCl₃) δ 0.19 (s, 9H), 1.51-1.60 (m, 2H), 1.66-1.73 (m, 2H), 2.04-2.12 (m, 3H), 4.35 (q, 1H, J = 5.4 Hz), 4.94-5.05 (m, 2H), 5.73-5.86 (m, 1H). These data are consistent with those reported.¹³

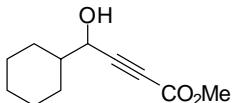
⁹ Johansson, M.; Kopcke, B.; Anke, H.; Sterner, O. *Tetrahedron* **2002**, 58, 2523-2528.

¹⁰ Weber, B.; Seebach, D. *Tetrahedron* **1994**, 50, 7473-7484.

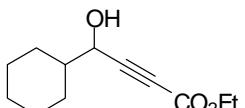
¹¹ Frstner, A.; Nevado, C.; Waser, M.; Tremblay, M.; Chevrier, C.; Tepl, F.; Assa, C.; Moulin, E.; Mller, O. *J. Am. Chem. Soc.* **2007**, 129, 9150-9161.

¹² Albert, B. J.; Koide, K. *J. Org. Chem.* **2008**, 73, 1093-1098.

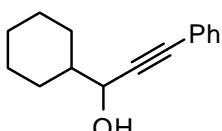
¹³ Satcharoen, V.; Mclean, N. J.; Kemp, S. C.; Camp, N. P.; Brown, R. C.D. *Org. Lett.* **2007**, 9, 1867-1869.



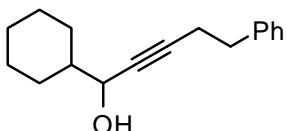
Methyl 4-cyclohexyl-4-hydroxybut-2-yneoate, SM-da. 84% yield. 95% ee determined by HPLC analysis: Chiralpak AD-H column, 99:1 hexanes:ⁱPrOH, flow rate = 0.3 mL/min, λ = 221 nm, retention time: $t_{\text{major}} = 73.66$ min $t_{\text{minor}} = 78.68$ min. $[\alpha]_D = 5.04$ ($c = 1.01$, CHCl₃). ¹H NMR (300 MHz, CDCl₃) δ 1.06-1.33 (m, 5H), 1.57-1.70 (m, 2H), 1.76-1.88 (m, 5H), 3.78 (s, 3H), 4.28 (t, 1H, J = 6.2 Hz). These data are consistent with those reported.¹



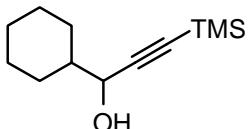
Ethyl 4-cyclohexyl-4-hydroxybut-2-yneoate, SM-db. 81% yield. 99.4% ee determined by HPLC analysis: Chiralpak AD-H column, 99:1 hexanes:ⁱPrOH, flow rate = 0.3 mL/min, λ = 221 nm, retention time: $t_{\text{major}} = 48.83$ min $t_{\text{minor}} = 52.78$ min. ¹H NMR (300 MHz, CDCl₃) δ 1.04-1.23 (m, 5H), 1.30 (t, 3H, J = 7.2 Hz), 1.62-1.68 (m, 2H), 1.75-1.86 (m, 4H), 2.31 (d, 1H, J = 6.0 Hz), 4.19-4.28 (m, 3H). These data are consistent with those reported.¹⁴



1-Cyclohexyl-3-phenylprop-2-yn-1-ol, SM-dc. 94% yield. 82% ee determined by HPLC analysis: Chiracel OD column, 90:10 hexanes:ⁱPrOH, flow rate = 1.0 mL/min, λ = 221 nm, retention time: $t_{\text{major}} = 15.98$ min $t_{\text{minor}} = 7.09$ min. ¹H NMR (300 MHz, CDCl₃) δ 1.11-1.35 (m, 5H), 1.67-1.95 (m, 6H), 2.33 (d, 1H, J = 5.7 Hz), 4.38 (t, 1H, J = 6.0 Hz), 7.28-7.31 (m, 3H), 7.43-7.46 (m, 2H). These data are consistent with those reported.⁶



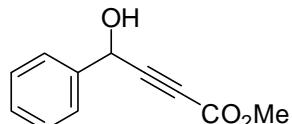
1-Cyclohexyl-5-phenylpent-2-yn-1-ol, SM-dd. 83% yield. 77% ee determined by analyzing the ¹H NMR spectrum of the (R)-mandelate ester derivative. ¹H NMR (300 MHz, CDCl₃) δ 0.97-1.32 (m, 5H), 1.59-1.80 (m, 7H), 2.52 (td, 2H, J_t = 7.5 Hz, J_d = 1.5 Hz), 2.83 (t, 2H, J = 7.5 Hz), 4.10 (d, 1H, J = 6.0 Hz), 7.21-7.32 (m, 5H). These data are consistent with those reported.¹⁵



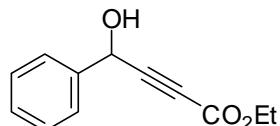
¹⁴ Villeneuve, K.; Tam, W. *Organometallics*. **2007**, 26, 6082-6090.

¹⁵ Takita, R.; Yakura, K.; Ohshima, T.; Shibasaki, M. *J. Am. Chem. Soc.* **2005**, 127, 13760-13761.

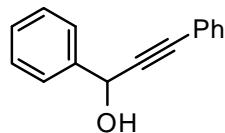
1-Cyclohexyl-3-(trimethylsilyl)prop-2-yn-1-ol, SM-de. 81% yield. 92% ee determined by analyzing the ^1H NMR spectrum of the (R)-mandalate ester derivative. ^1H NMR (300 MHz, CDCl_3) δ 0.17 (s, 9H), 1.02-1.32 (m, 5H), 1.66-1.87 (m, 7H), 4.13 (t, 1H, $J = 5.7$ Hz). These data are consistent with those reported.⁶



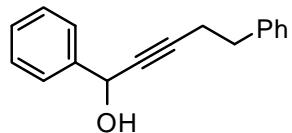
Methyl 4-phenyl-4-hydroxybut-2-yneoate, SM-ea. 64% yield. 90% ee determined by HPLC analysis: Chiracel OD column, 95:5 hexanes: $i\text{PrOH}$, flow rate = 1.0 mL/min, $\lambda = 254$ nm, retention time: $t_{\text{major}} = 24.30$ min $t_{\text{minor}} = 29.45$ min. ^1H NMR (300 MHz, CDCl_3) δ 2.29 (bs, 1H), 3.80 (s, 3H), 5.58 (d, 1H, $J = 6.3$ Hz), 7.39-7.41 (m, 3H), 7.51-7.54 (m, 2H). These data are consistent with those reported.¹⁶



Ethyl 4-phenyl-4-hydroxybut-2-yneoate, SM-eb. 52% yield. 88% ee determined by HPLC analysis: Chiracel OD column, 90:10 hexanes: $i\text{PrOH}$, flow rate = 1.0 mL/min, $\lambda = 254$ nm, retention time: $t_{\text{major}} = 10.99$ min $t_{\text{minor}} = 13.65$ min. ^1H NMR (300 MHz, CDCl_3) δ 1.31 (t, 3H, $J = 7.2$ Hz), 2.61 (d, 1H, $J = 6.3$ Hz), 4.24 (q, 2H, $J = 7.2$ Hz), 5.57 (d, 1H, $J = 6.3$ Hz), 7.34-7.43 (m, 3H), 7.50-7.53 (m, 2H). These data are consistent with those reported.¹³

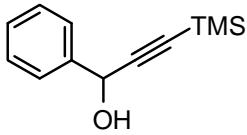


1,3-Diphenylprop-2-yn-1-ol, SM-ec. 91% yield. 83% ee determined by HPLC analysis: Chiracel OD column, 90:10 hexanes: $i\text{PrOH}$, flow rate = 1.0 mL/min, $\lambda = 254$ nm, retention time: $t_{\text{major}} = 25.28$ min $t_{\text{minor}} = 13.43$ min. ^1H NMR (300 MHz, CDCl_3) δ 2.48 (bs, 1H), 5.74 (bs, 1H), 7.37-7.53 (m, 8H), 7.65-7.67 (m, 2H). These data are consistent with those reported.⁶

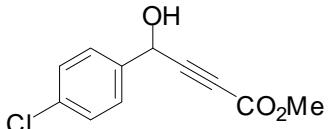


1,5-Diphenylpent-2-yn-1-ol, SM-ed. 61% yield. 92% ee determined by HPLC analysis: Chiracel OD column, 90:10 hexanes: $i\text{PrOH}$, flow rate = 1.0 mL/min, $\lambda = 254$ nm, retention time: $t_{\text{major}} = 34.07$ min $t_{\text{minor}} = 16.76$ min. ^1H NMR (300 MHz, CDCl_3) δ 2.43 (d, 1H, $J = 5.7$ Hz), 2.63-2.68 (m, 2H), 2.90-2.96 (m, 2H), 5.49 (bs, 1H), 7.30-7.54 (m, 10H). These data are consistent with those reported.¹⁴

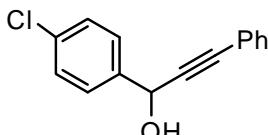
¹⁶ Gao, G.; Wang Q.; Yu, X.-Q.; Xie, R.-G.; Pu, L. *Angew. Chem. Int. Ed.* **2006**, 45, 122-125.



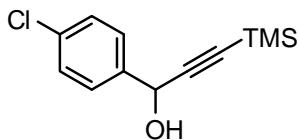
1-Phenyl-3-(trimethylsilyl)prop-2-yn-1-ol, SM-ee. 50% yield. 97% ee determined by HPLC analysis: Chiracel OD column, 95:5 hexanes:ⁱPrOH, flow rate = 1.0 mL/min, λ = 254 nm, retention time: $t_{\text{major}} = 7.41$ min $t_{\text{minor}} = 10.41$ min. ¹H NMR (300 MHz, CDCl₃) δ 0.25 (s, 9H), 2.29 (d, 1H, J = 6.3 Hz), 5.49 (d, 1H, J = 6.3 Hz), 7.37-7.46 (m, 3H), 7.58-7.60 (m, 2H). These data are consistent with those reported.¹⁷



Methyl 4-(4-chlorophenyl)-4-hydroxybut-2-yoate, SM-fa. 83% yield. 92% ee determined by HPLC analysis: Chiracel OD column, 95:5 hexanes:ⁱPrOH, flow rate = 1 mL/min, λ = 221 nm, retention time: $t_{\text{major}} = 19.97$ min $t_{\text{minor}} = 18.47$ min. ¹H NMR (300 MHz, CDCl₃) δ 2.80 (bs, 1H), 3.80 (s, 3H), 5.55 (bs, 1H), 7.34-7.38 (m, 2H), 7.43-7.46 (m, 2H). These data are consistent with those reported.¹⁵



1-(4-Chlorophenyl)-3-phenylprop-2-yn-1-ol, SM-fc. 87% yield. 84% ee determined by HPLC analysis: Chiracel OD column, 90:10 hexanes:ⁱPrOH, flow rate = 1 mL/min, λ = 254 nm, retention time: $t_{\text{major}} = 40.32$ min $t_{\text{minor}} = 11.01$ min. ¹H NMR (300 MHz, CDCl₃) δ 2.82 (d, 1H, J = 6.0 Hz), 5.66 (d, 1H, J = 5.7 Hz), 7.32-7.37 (m, 5H), 7.45-7.48 (m, 2H), 7.52-7.55 (m, 2H). These data are consistent with those reported.⁶



1-(4-Chlorophenyl)-3-(trimethylsilyl)prop-2-yn-1-ol, SM-fe: 61% yield. 94% ee determined by HPLC analysis: Chiracel OD column, 90:10 hexanes:ⁱPrOH, flow rate = 1 mL/min, λ = 254 nm, retention time: $t_{\text{major}} = 14.28$ min $t_{\text{minor}} = 17.18$ min. ¹H NMR (300 MHz, CDCl₃) δ 0.20 (s, 9H), 2.24 (d, 1H, J = 6.0 Hz), 5.42 (d, 1H, J = 5.7 Hz), 7.33-7.36 (m, 2H), 7.45-7.48 (m, 2H). These data are consistent with those reported.¹⁸

Characterization of the Ring-Closing Metathesis Products:

2-(1-Phenylvinyl)cyclopent-2-enyl acetate, P-7. 94% yield. $[\alpha]_D = -19.67$ (c = 1.73, CH₂Cl₂). ¹H NMR (300 MHz, CDCl₃) δ 1.92-1.98 (m, 1H), 2.05 (s, 3H), 2.36-2.45 (m,

¹⁷ Asano, Y.; Hara, K.; Ito, H.; Sawamura, M. *Org. Lett.* **2007**, 9, 3901-3904.

¹⁸ Qiu, L.; Wang, Q.; Lin, L.; Liu, X.; Jiang, X.; Zhao, Q.; Hu, G.; Wang, R. *Chirality*. **2009**, 21, 316-323.

2H), 2.57-2.61 (m, 1H), 5.15 (s, 1H), 5.22 (s, 1H), 5.92 (t, 1H, J = 2.7 Hz), 6.07-6.10 (m, 1H), 7.30-7.33 (m, 5H). ^{13}C NMR (75 MHz, CDCl_3) δ 21.6, 31.2, 31.8, 79.3, 114.9, 127.6, 128.3, 128.5, 137.4, 141.8, 141.9, 144.2, 171.4. HRMS (M-H) for $\text{C}_{15}\text{H}_{15}\text{O}_2$ Calcd: 227.1067 Found: 227.1070.

2-(4-Phenylbutan-2-yl)cyclopent-2-enyl acetate, P-8. 85% yield. $[\alpha]_D$ = -3.95 (c = 1.66, CH_2Cl_2). ^1H NMR (300 MHz, CDCl_3) δ 1.85-1.93 (m, 1H), 2.05 (s, 3H), 2.29-2.47 (m, 2H), 2.56-2.61 (m, 3H), 2.80-2.85 (m, 2H), 4.97 (s, 1H), 4.99 (s, 1H), 6.01-6.03 (m, 1H), 6.15 (t, 1H, J = 2.4 Hz), 7.19-7.22 (m, 3H), 7.27-7.32 (m, 2H). ^{13}C NMR (75 MHz, CDCl_3) δ 21.6, 31.2, 31.7, 35.1, 36.5, 79.2, 113.0, 126.1, 128.5, 128.7, 133.6, 141.3, 141.7, 142.3, 171.3. EI-HRMS (M-AcOH) for $\text{C}_{15}\text{H}_{16}$ Calcd: 196.1252. Found: 196.1260.

2-(1-Phenylvinyl)cyclohex-2-enyl acetate, P-9. 97% yield. $[\alpha]_D$ = -140.17 (c = 1.95, CH_2Cl_2). ^1H NMR (300 MHz, CDCl_3) δ 1.65-1.83 (m, 3H), 1.92-1.95 (m, 1H), 1.99 (s, 3H), 2.14-2.27 (m, 2H), 5.08 (s, 1H), 5.19 (s, 1H), 5.65 (t, 1H, J = 3.6 Hz), 5.94 (t, 1H, J = 3.9 Hz), 7.26-7.34 (m, 5H). ^{13}C NMR (75 MHz, CDCl_3) δ 17.7, 21.5, 25.9, 29.1, 67.2, 113.0, 127.6, 128.3, 128.6, 134.1, 136.4, 141.7, 149.3, 170.8. HRMS (MH $^+$) for $\text{C}_{16}\text{H}_{19}\text{O}_2$ Calcd: 243.1380. Found: 243.1389

2-(4-Phenylbutan-2-yl)cyclohex-2-enyl acetate, P-10. 87% yield. $[\alpha]_D$ = -77.02 (c = 1.00, CH_2Cl_2). ^1H NMR (300 MHz, CDCl_3) δ 1.66-1.69 (m, 3H), 1.98-2.00 (m, 1H), 2.05 (s, 3H), 2.09-2.17 (m, 1H), 2.26-2.33 (m, 1H), 2.49-2.55 (m, 2H), 2.74-2.80 (m, 2H), 4.88 (s, 1H), 4.97 (s, 1H), 5.70 (b, 1H), 6.17-6.20 (m, 1H), 7.18-7.29 (m, 5H). ^{13}C NMR (75 MHz, CDCl_3) δ 17.2, 21.7, 26.0, 29.1, 35.3, 36.1, 66.6, 111.0, 125.9, 126.1, 128.7, 128.7, 130.5, 134.7, 142.4, 145.6, 171.0. EI-HRMS (M-AcOH) for $\text{C}_{16}\text{H}_{18}$ Calcd: 210.1408. Found: 210.1412.

2-(2-Oxo-2,5-dihydrofuran-3-yl)cyclopent-2-enyl acetate, P-11. 61% yield. ^1H NMR (300 MHz, C_6D_6) δ 1.58-1.63 (m, 1H), 1.68 (s, 3H), 1.92-2.04 (m, 2H), 2.26-2.33 (m, 1H), 3.79 (s, 2H), 5.93-5.96 (m, 1H), 6.42 (b, 1H), 7.32 (b, 1H). ^{13}C NMR (75 MHz, C_6D_6) δ 20.8, 30.8, 31.5, 69.1, 79.4, 126.0, 139.2, 142.8, 170.1, 171.6. HRMS (MNH $_4^+$) for $\text{C}_{11}\text{H}_{16}\text{NO}_4$ Calcd: 226.1074. Found: 226.1075

Characterization of the Tandem Ring-Closing Metathesis and Hydrogenation Products.

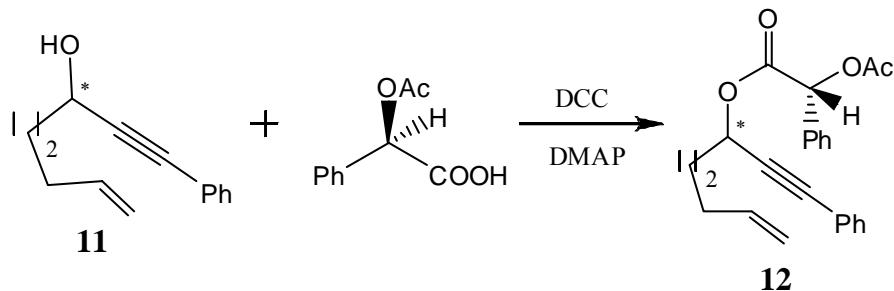
Methyl 2-(5-acetoxycyclopent-1-enyl)propanoate, P-12. 88% yield. The ratio of the two diastereomers is 2:1 determined by analyzing the ^1H NMR spectrum. $[\alpha]_D$ = -1.77 (c = 1.10, CH_2Cl_2). ^1H NMR (300 MHz, CDCl_3) δ 1.34 (d, 3H, J = 7.2 Hz), 1.72-1.90 (m, 2H), 2.02 (s, 3H), 2.31-2.51 (m, 2H), 3.20-3.30 (m, 1H), 3.66 (s, 3H), 5.70-5.73 (m, 1H), 5.90 (bs, 1H). Following are the resolved signals of another diastereomer: δ 1.29 (d, 3H, J = 7.2 Hz), 3.68 (s, 3H), 5.86 (b, 1H). ^{13}C NMR (75 MHz, CDCl_3) δ 16.0, 21.5, 30.4, 31.2, 38.6, 52.2, 80.5, 132.9, 141.0, 171.2, 174.6. Following are the resolved signals of another diastereomer: δ 16.8, 39.6, 80.6, 132.7, 174.9. HRMS (MNa $^+$) for

$C_{11}H_{16}O_4Na$ Calcd: 235.0946. Found: 235.0949.

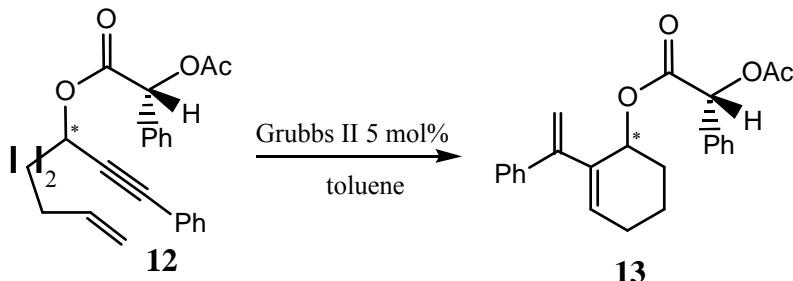
Ethyl 2-(5-acetoxycyclopent-1-enyl)propanoate, P-13. 89% yield. The ratio of the two diastereomers is 2:1 determined by analyzing the 1H NMR spectrum. $[\alpha]_D = -0.44$ ($c = 1.70, CH_2Cl_2$). 1H NMR (300 MHz, $CDCl_3$) δ 1.22 (t, 3H, $J = 7.2$ Hz), 1.33 (d, 3H, $J = 7.2$ Hz), 1.72-1.90 (m, 1H), 2.02 (s, 3H), 2.26-2.51 (m, 3H), 3.17-3.27 (m, 1H), 4.07-4.15 (m, 2H), 5.70-5.73 (m, 1H), 5.89 (b, 1H). Following are the resolved signals of another diastereomer: δ 1.28 (d, 3H, $J = 7.2$ Hz), 2.01 (s, 3H), 5.86 (bs, 1H). ^{13}C NMR (75 MHz, $CDCl_3$) δ 14.4, 15.9, 21.4, 30.4, 31.3, 38.7, 60.9, 80.6, 132.6, 141.1, 171.2, 174.1. Following are the resolved signals of another diastereomer: δ 16.9, 21.5, 30.4, 39.8, 60.8, 132.6, 171.2, 174.5. HRMS (MNa $^+$) for $C_{12}H_{18}O_4Na$ Calcd: 249.1103 Found: 249.1094.

Methyl 2-(6-acetoxycyclohex-1-enyl)propanoate, P-14. 90% yield. The ratio of the two diastereomers is 2:1 determined by analyzing the 1H NMR spectrum. $[\alpha]_D = -42.64$ ($c = 0.61, CH_2Cl_2$). 1H NMR (300 MHz, $CDCl_3$) δ 1.23 (d, 3H, $J = 7.2$ Hz), 1.57-1.70 (m, 2H), 1.76-1.88 (m, 3H), 2.01 (s, 3H), 2.09-2.20 (m, 1H), 3.02-3.14 (m, 1H), 3.64 (s, 3H), 5.33 (t, 1H, $J = 4.5$ Hz), 5.89 (t, 1H, $J = 3.9$ Hz). Following are the resolved signals of another diastereomer: δ 2.02 (s, 3H), 3.65 (s, 3H), 5.85 (t, 1H, $J = 3.9$ Hz). ^{13}C NMR (75 MHz, $CDCl_3$) δ 15.8, 18.3, 21.5, 25.4, 29.1, 43.2, 52.1, 68.8, 130.1, 134.9, 170.9, 175.2. Following are the resolved signals of another diastereomer: δ 17.1, 18.0, 44.0, 69.2, 171.0, 175.3. HRMS (MNH $_4^+$) for $C_{12}H_{22}NO_4$ Calcd: 244.1543 Found: 244.1546.

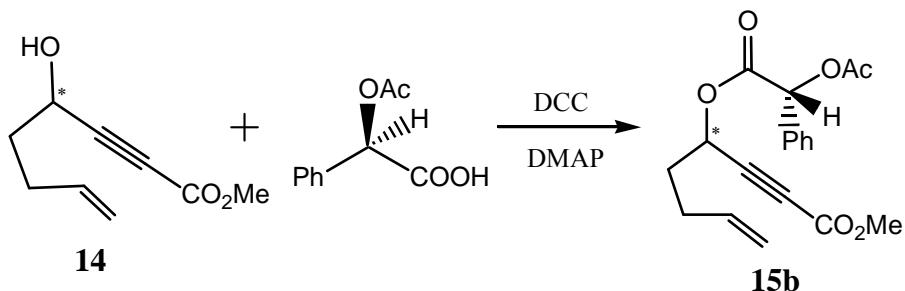
Ethyl 2-(6-acetoxycyclohex-1-enyl)propanoate, P-15. 91% yield. The ratio of the two diastereomers is 3:1 determined by analyzing the ^{13}C NMR spectrum. $[\alpha]_D = -59.92$ ($c = 1.05, CH_2Cl_2$). 1H NMR (300 MHz, $CDCl_3$) δ 1.20-1.26 (m, 6H), 1.60-1.66 (m, 2H), 1.76-1.80 (m, 2H), 2.03 (s, 3H), 2.12-2.26 (m, 2H), 3.01-3.14 (m, 1H), 4.10 (q, 2H, $J = 7.2$ Hz), 5.34 (t, 1H, $J = 4.5$ Hz), 5.86-5.91 (m, 1H). ^{13}C NMR (75 MHz, $CDCl_3$) δ 14.4, 15.8, 18.3, 21.5, 25.4, 29.1, 43.1, 60.8, 69.1, 129.8, 135.0, 170.9, 174.8. The following are the resolved signals of another diastereomer: δ 17.2, 18.0, 21.6, 44.2, 69.2, 171.0, 174.9. HRMS (MNa $^+$) for $C_{13}H_{20}O_4Na$ Calcd: 263.1259. Found: 263.1263.



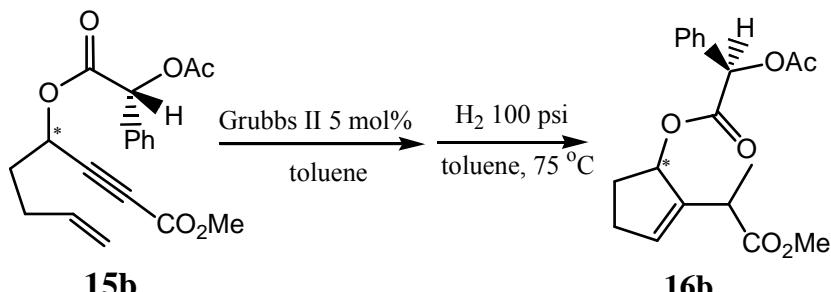
1-Phenyl-7-en-1-yn-3-ol (**11**) (1.0 equiv) was dissolved in CH_2Cl_2 and cooled to 0 °C. (*R*)-(−)- α -Acetylmandelic acid (1.5 equiv), DCC (1.5 equiv) and DMAP (0.1 equiv) were added in one portion. After 20 min the reaction mixture was passed through a short silica gel column eluted with 20% EtOAc/hexanes to give **12**.



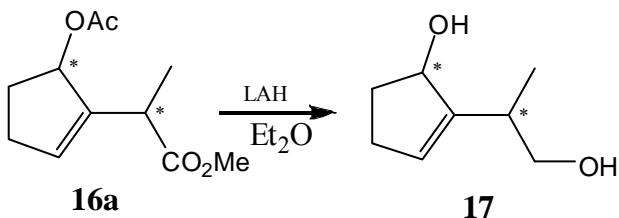
(R)-((S)-1-phenyloct-7-en-1-yn-3-yl) 2-acetoxy-2-phenylacetate (**12**) (1 equiv) was dissolved in toluene (0.14 M) and combined with Grubbs II (5 mol%) under nitrogen. After the mixture was stirred at room temperature for 1 h, it was passed through a short silica gel column eluted with 20% EtOAc/hexanes to give **13**. The ee was determined by analyzing the ¹H NMR spectrum. It shows that the enantiomeric purity of the products generated from the asymmetric methyl propiolate addition to the aldehydes is maintained in the subsequent RCM reaction.



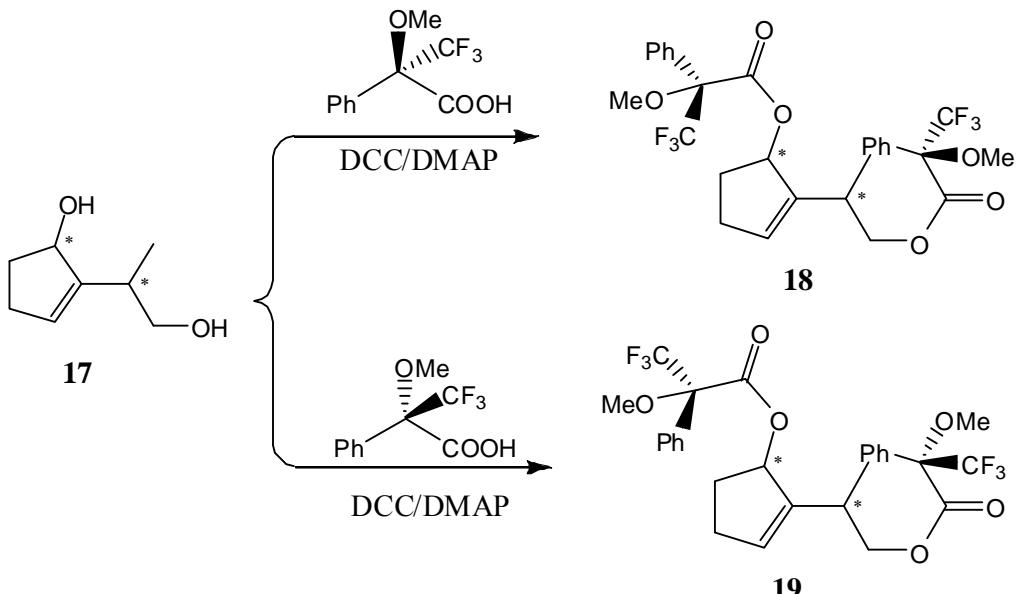
Methyl 4-hydroxyoct-7-en-2-ynoate (**14**) (1.0 equiv) was dissolved in CH₂Cl₂ and cooled to 0 °C. (R)-(-)-α-Acetylmandelic acid (1.5 equiv), DCC (1.5 equiv) and DMAP (0.1 equiv) were added in one portion. After 20 min the reaction mixture was passed through a short silica gel column eluted with 20% EtOAc/hexanes to give **15b**.



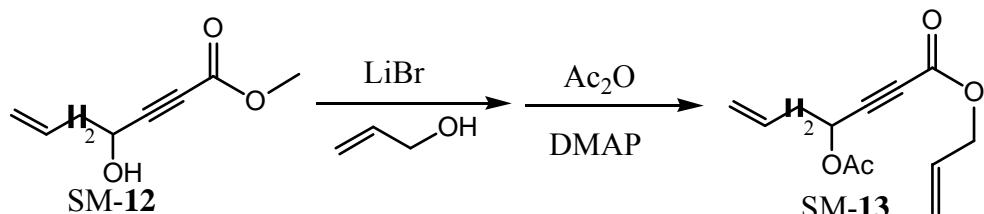
To a 50 mL round bottom flask, methyl 4-((R)-2-acetoxy-2-phenylacetoxymethyl)oct-7-en-2-ynoate (**15b**) (1 equiv) was dissolved in toluene (0.14 M) and Grubbs II (5 mol%) was added under nitrogen. After stirring 1 h at room temperature, the mixture was transferred to a parr reactor. Hydrogen (100 psi) was introduced and the reactor was heated at 75 °C with stirring for 10 h. The crude mixture was passed through a short silica gel column eluted with 20% EtOAc/hexanes. The ee was determined by analyzing its ¹H NMR spectrum. The enantiomeric purity of the product generated from the asymmetric methyl propiolate addition to the aldehydes is maintained in **16b**.



To a 10 mL flask, **16a** (1.0 equiv) and LAH (3 equiv) were dissolved in Et₂O under nitrogen. After the mixture was stirred for 30 min, it was passed through a short silica gel column eluted with 50% EtOAc/hexanes to give **17** in 92% yield. ¹H NMR (300 MHz, CDCl₃) δ 1.08 (d, 3H, J = 7.2 Hz), 1.73-1.82 (m, 1H), 2.18-2.25 (m, 2H), 2.43-2.53 (m, 1H), 2.66-2.77 (m, 1H), 3.21 (bs, 2H), 3.59 (d, 2H, J = 5.4 Hz), 4.77-4.79 (m, 1H), 5.68-5.70 (m, 1H). The following are the resolved signals of another diastereomer: δ 1.07 (d, 3H, J = 7.2 Hz), 3.43 (dd, 1H, J₁ = 8.7 Hz, J₂ = 10.5 Hz), 3.69 (dd, 1H, J₁ = 10.5 Hz, J₂ = 4.5 Hz), 4.63-4.65 (m, 1H).



To a 10 mL flask, **17** (1.0 equiv) was dissolved in CH₂Cl₂ and cooled to 0 °C. (R)-(+)-α-Methoxy-α-trifluoromethyl-phenylacetic acid or (S)-(-)-α-methoxy-α-(trifluoromethyl)phenylacetic acid (3.0 equiv), DCC (3.0 equiv) and DMAP (0.1 equiv) were added in one portion. After 1 h the reaction mixture was passed through a short silica gel column eluted with 10% EtOAc/hexanes to give the product **18** or **19**.

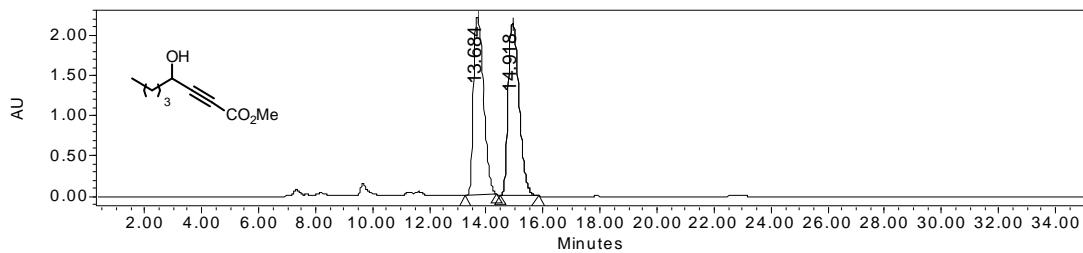


Preparation of allyl 4-acetoxyoct-7-en-2-ynoate, SM-13. To a 25 mL flask, methyl 4-hydroxyoct-7-en-2-yneoate (SM-12) (84 mg, 0.5 mmol), LiBr (86 mg, 1 mmol) and allyl alcohol (15 mL) were added. After the mixture was stirred at 50 °C for 2 d, the solvent

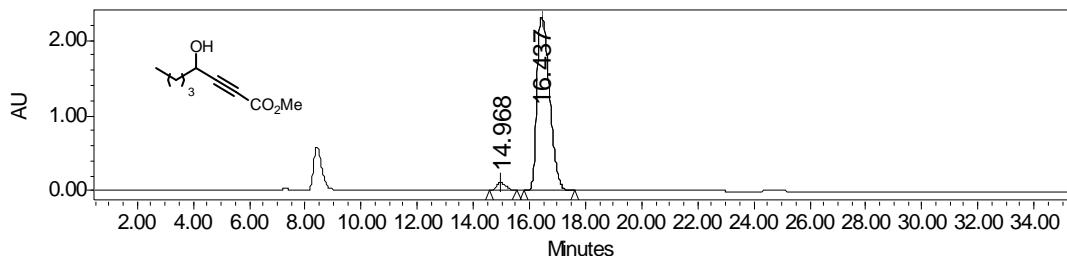
was removed under vacuum. The crude product was dissolved in CH₂Cl₂ (25 mL), and DMAP (12.2 mg, 0.1 mmol) and Ac₂O (0.5 mL) were added. The reaction mixture was stirred at room temperature for 10 min and then concentrated. The residue was purified by column chromatography on silica gel eluted with 10% EtOAc/hexanes to give SM-13 as a white solid in 83% yield. ¹H NMR (300 MHz, CDCl₃) δ 1.93-2.00 (m, 2H), 2.13 (s, 1H), 2.22-2.29 (m, 2H), 4.70 (d, 2H, J = 6.0 Hz), 5.04-5.13 (m, 2H), 5.31-5.43 (m, 2H), 5.49 (t, 1H, J = 6.6 Hz), 5.75-6.02 (m, 2H). ¹³C NMR (75 MHz, CDCl₃) δ 21.0, 29.2, 33.3, 62.8, 66.9, 77.7, 84.6, 116.3, 119.8, 131.2, 136.5, 152.9, 169.9. HRMS (MH⁺) for C₁₁H₁₅O₃ Calcd: 195.1016. Found: 195.1020.

HPLC Plots

SM-aa:

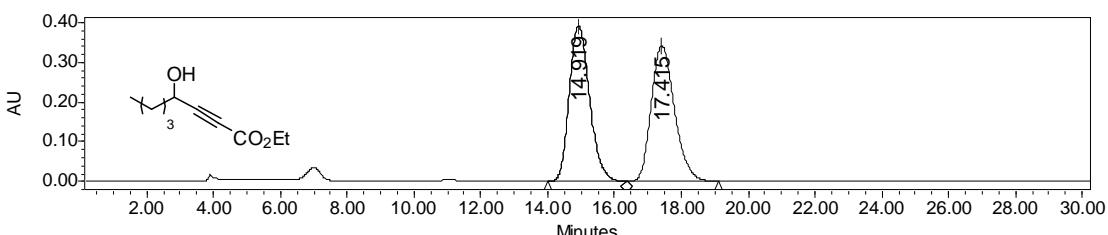


Name	Retention Time	Area	% Area
1	13.684	54300971	49.02
2	14.918	56481027	50.98

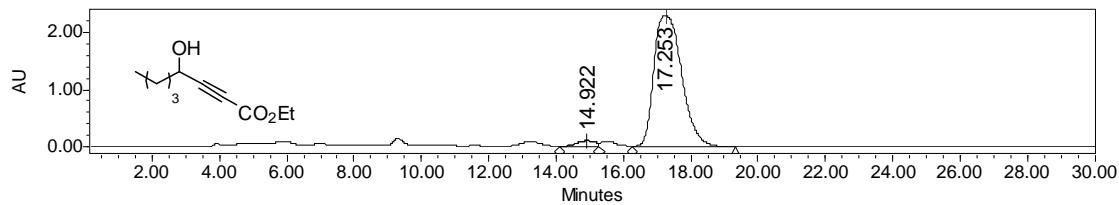


Name	Retention Time	Area	% Area
1	14.968	2402438	3.17
2	16.437	73302699	96.83

SM-ab:

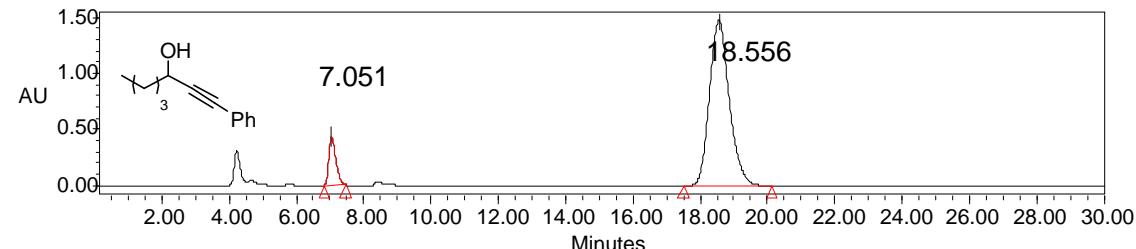
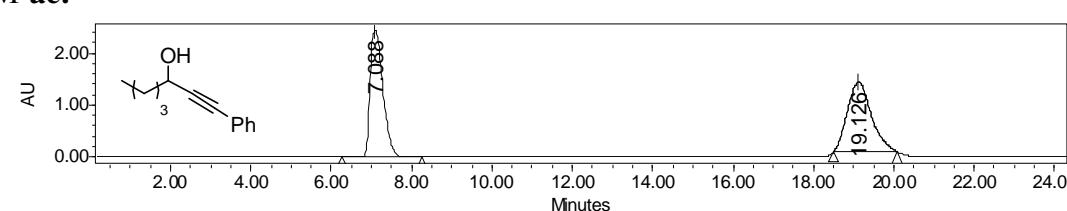


Name	Retention Time	Area	% Area
1	14.919	16476403	50.02
2	17.415	16462301	49.98

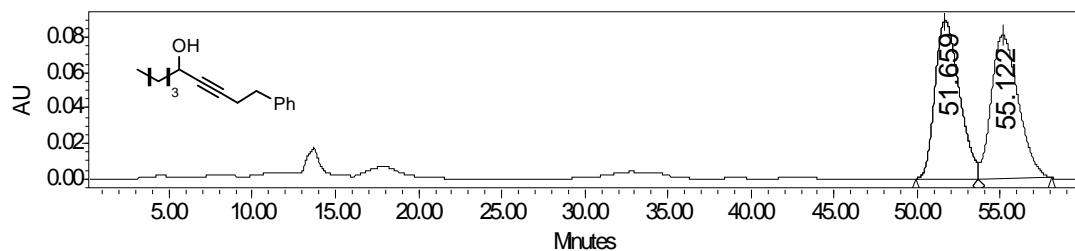


SM-ac:

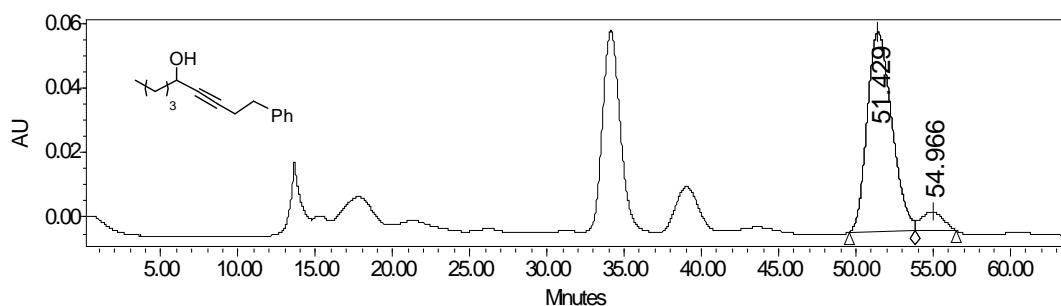
Name	Retention Time	Area	% Area
1	14.922	3899368	2.86
2	17.253	132522065	97.14



SM-ad (= P-1):

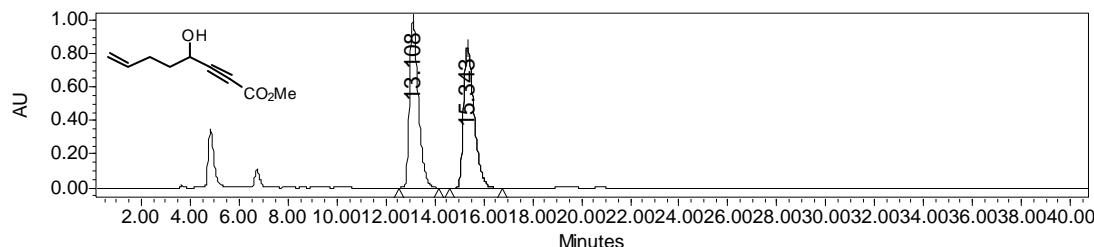


Name	Time	Area	% Area
1	51.659	9131629	49.87
2	55.122	9179394	50.13

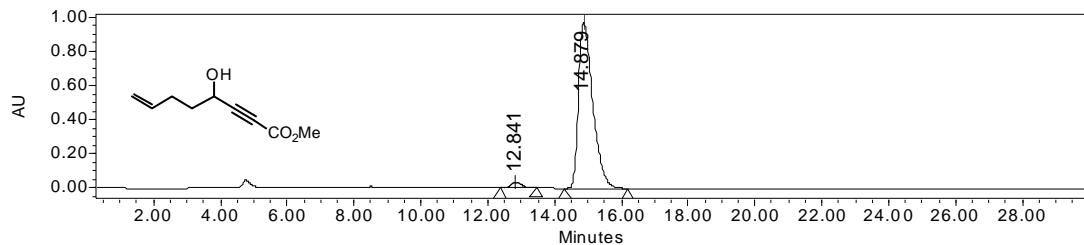


Name	Time	Area	% Area
1	51.429	6929610	91.92
2	54.966	6087848.08	

SM-ba:

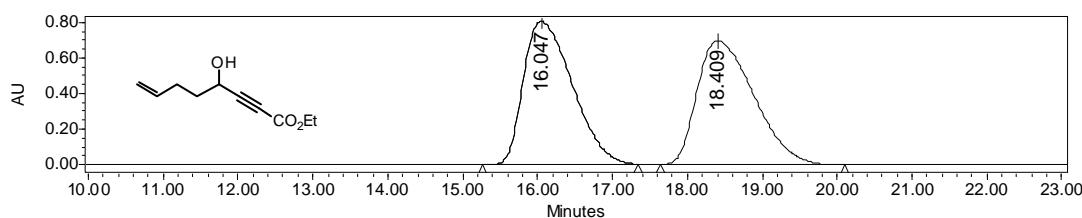


Name	Retention Time	Area	% Area
1	13.108	24899891	49.82
2	15.343	25075437	50.18

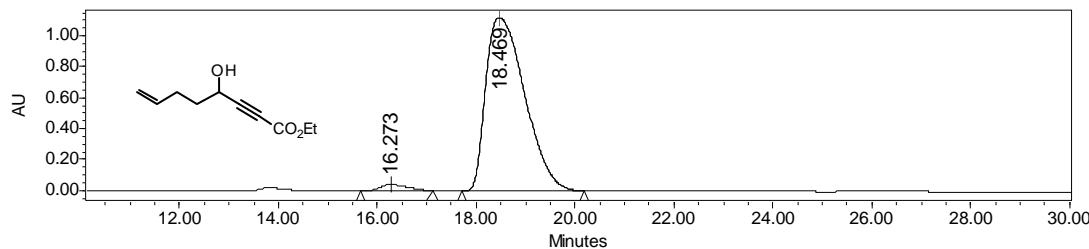


Name	Retention Time	Area	% Area
1	12.841	665996	2.26
2	14.879	28782019	97.74

SM-bb:

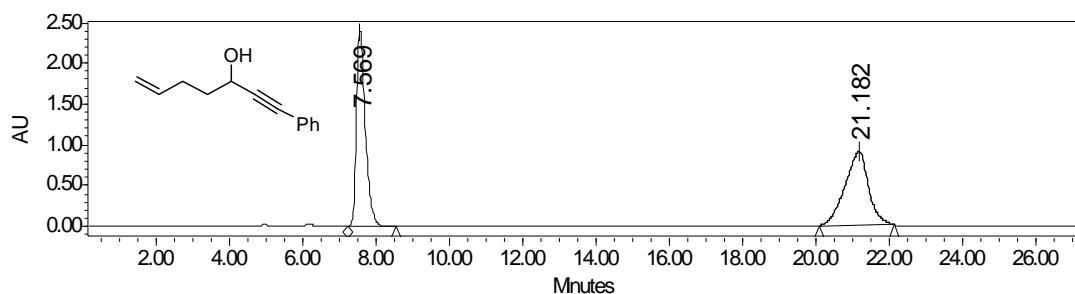


Name	Retention Time	Area	% Area
1	16.047	36550833	49.86
2	18.409	36751392	50.14

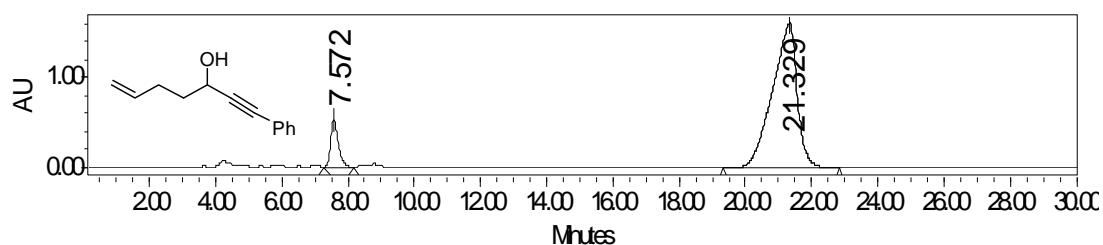


Name	Retention Time	Area	% Area
1	16.273	1634508	2.59
2	18.469	61570530	97.41

SM-bc:

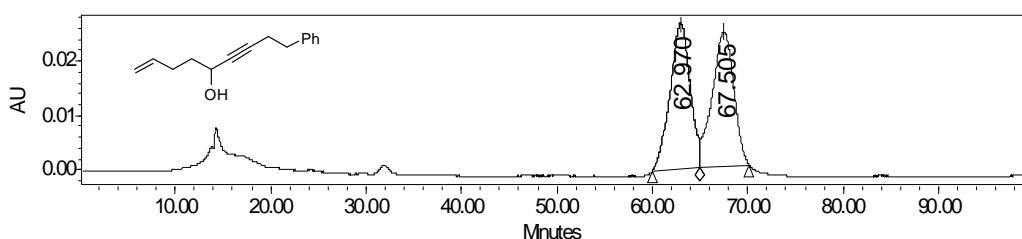


Name	Retention Time	Area % Area
1	7.569	40538257 49.99
2	21.182	40561237 50.01

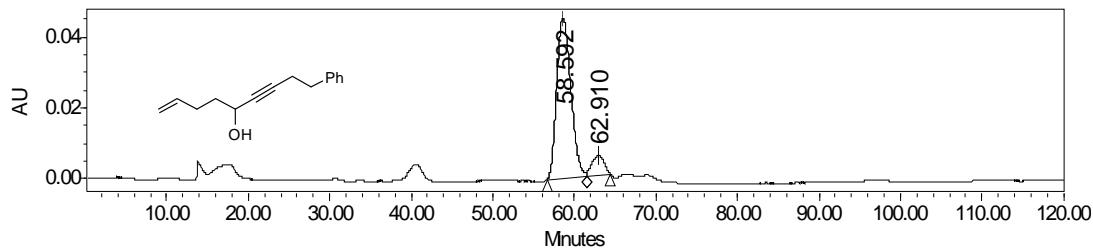


Name	Retention Time	Area % Area
1	7.572	8574269 9.29
2	21.329	83748327 90.71

SM-bd (= P-2):

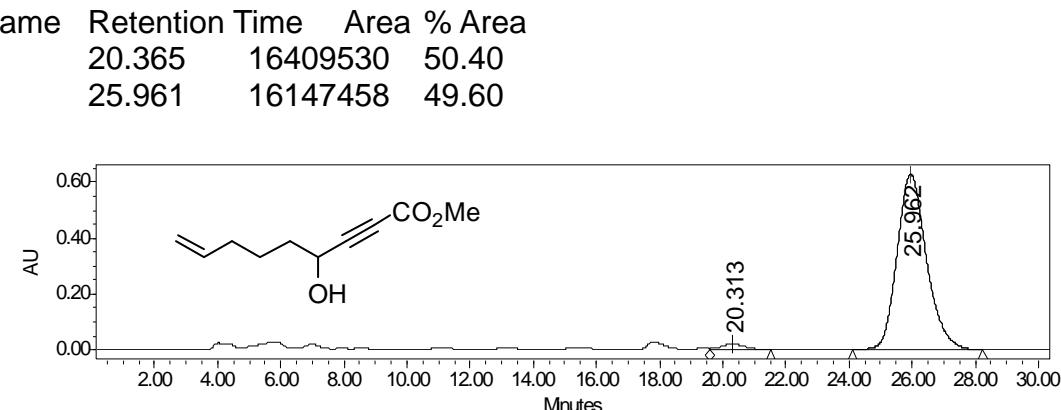
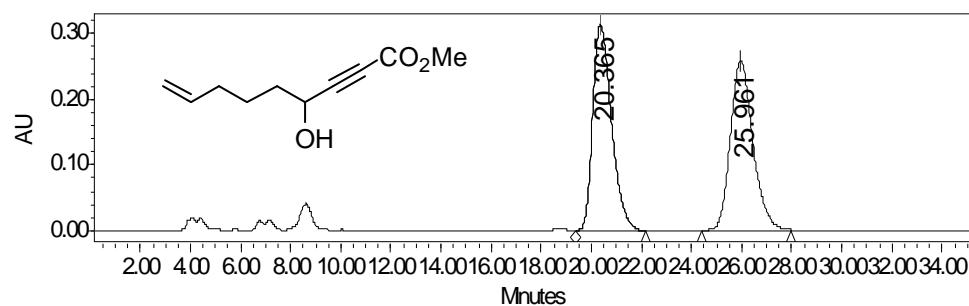


Name	Retention Time	Area % Area
1	62.970	3940203 50.11
2	67.505	3922836 49.89



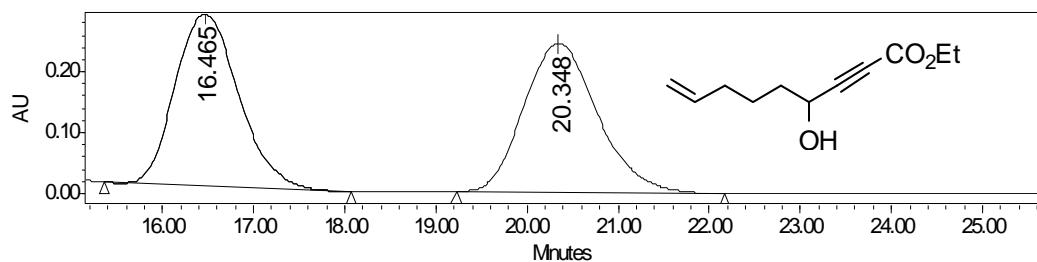
Name	Retention Time	Area %	Area
1	58.592	5447451	89.90
2	62.910	611677	10.10

SM-ca:

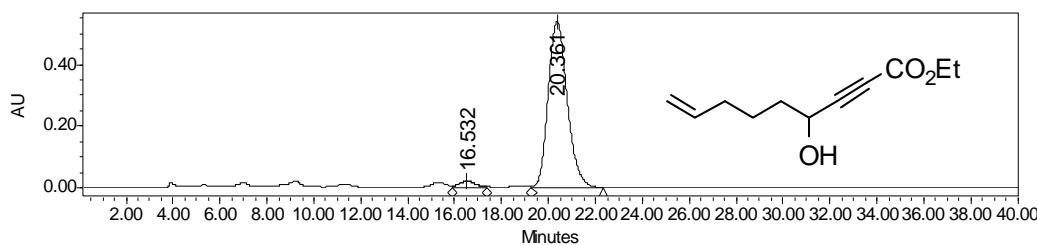


Name	Retention Time	Area %	Area
1	20.313	1021514	2.45
2	25.962	40742263	97.55

SM-cb (= P-3):

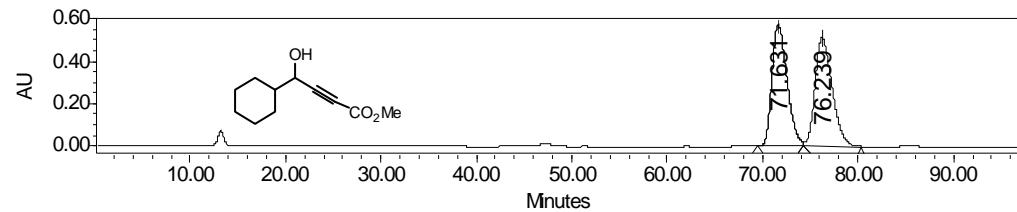


Name	Retention Time	Area	% Area
1	16.465	13654426	49.84
2	20.348	13744620	50.16

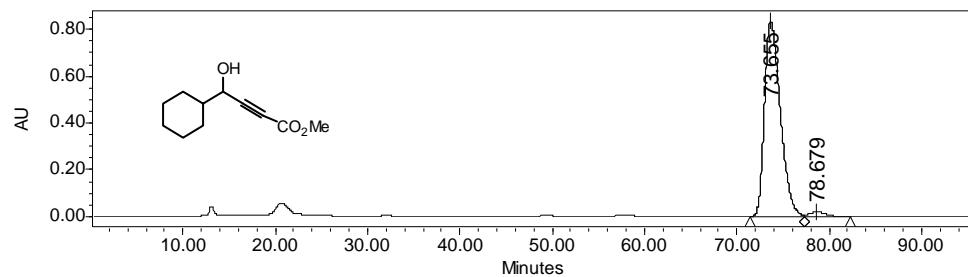


Name	Retention Time	Area	% Area
1	16.532	980870	3.12
2	20.361	30418112	96.88

SM-da:

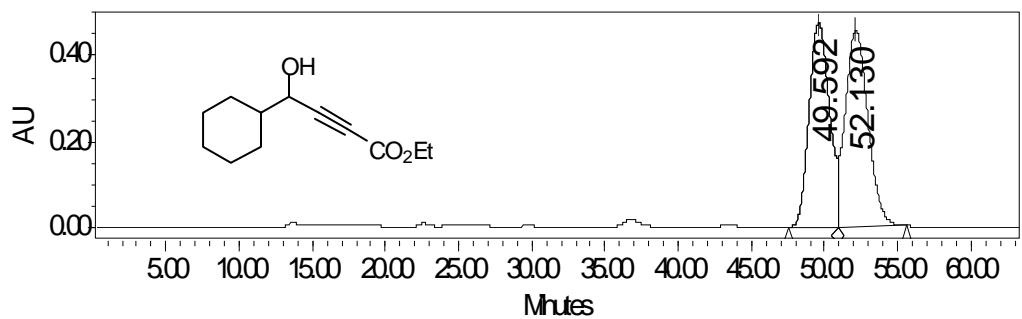


Name	Retention Time	Area	% Area
1	71.631	63675367	49.92
2	76.239	63880927	50.08

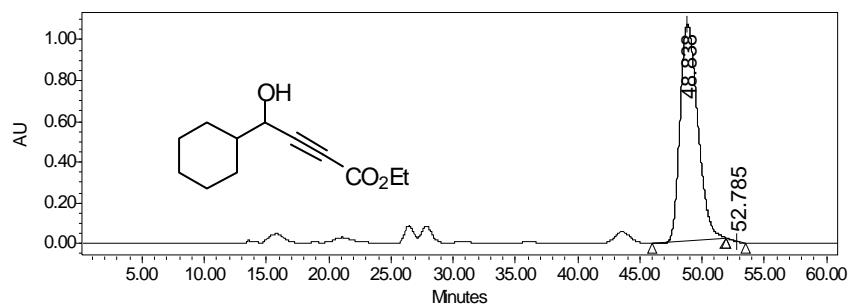


Name	Retention Time	Area	% Area
1	73.655	99728514	97.36
2	78.679	2705025	2.64

SM-db:

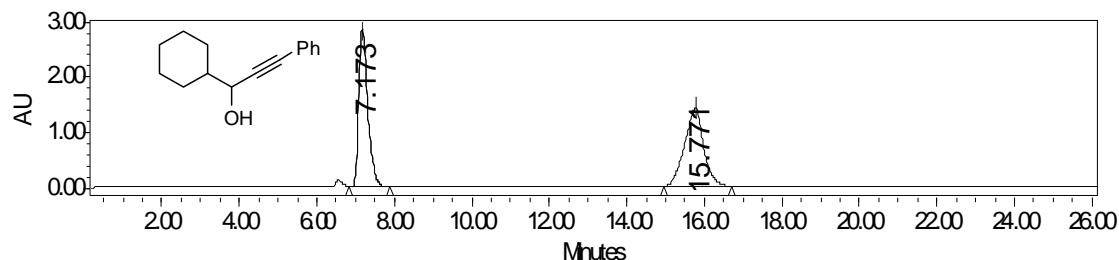


Name	Retention Time	Area	% Area
1	49.592	47746772	49.71
2	52.130	48308387	50.29

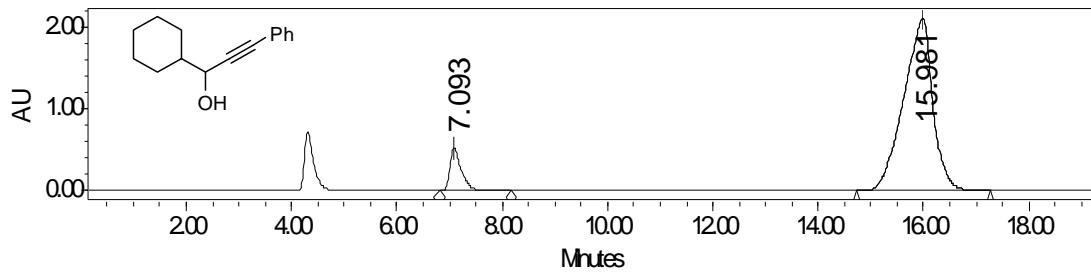


Name	Retention Time	Area	% Area
1	48.838	105122240	99.70
2	52.785	320249	0.30

SM-dc:



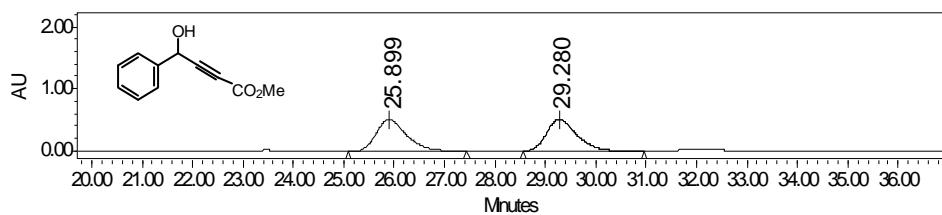
Name	Retention Time	Area	% Area
1	7.173	46556220	48.80
2	15.771	48843347	51.20



Name Retention Time Area % Area

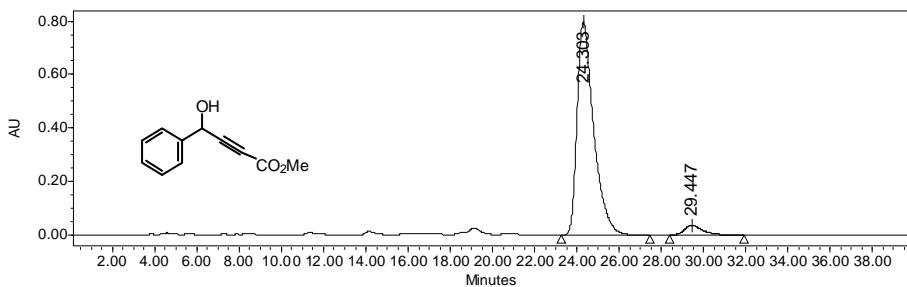
1	7.093	7916130	8.84
2	15.981	81618006	91.16

SM-ea:



Retention Time Area % Area

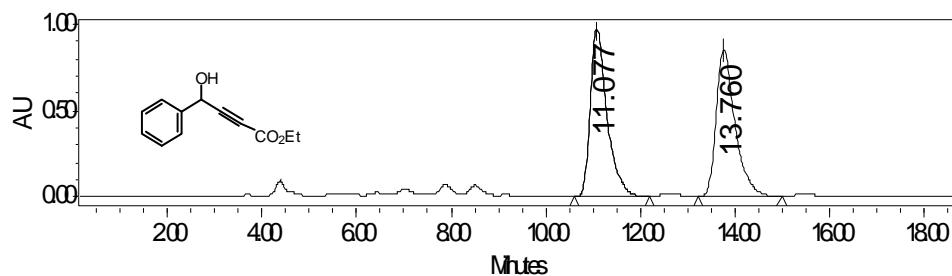
25.899	21903551	50.20
29.280	21731884	49.80



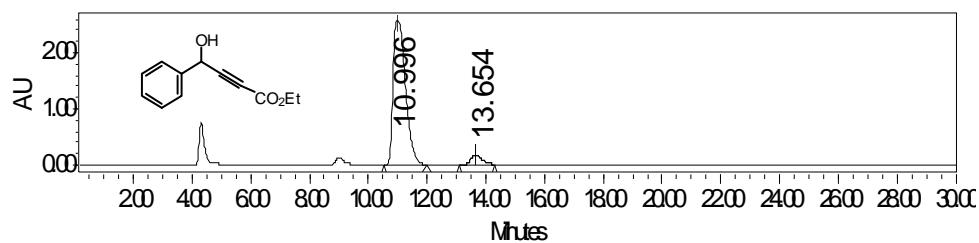
Retention Time Area % Area

24.303	43379467	95.13
29.447	2221559	4.87

SM-eb:

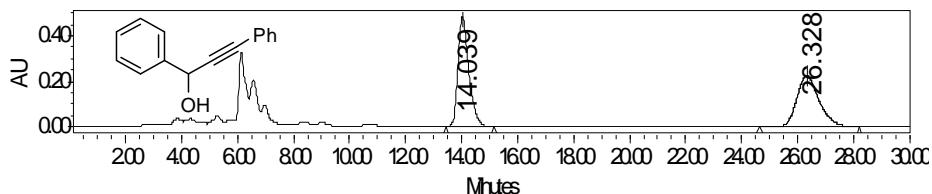


Name	Retention Time	Area % Area
1	11.077	24191565 50.25
2	13.760	23948441 49.75

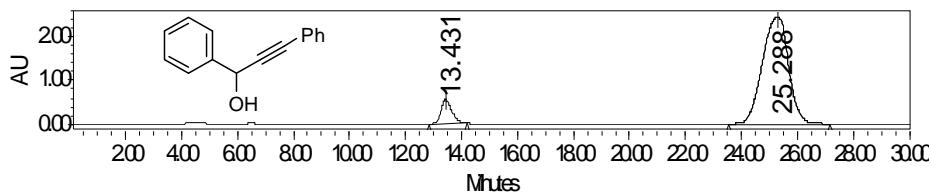


Name	Retention Time	Area % Area
1	10.996	75247293 94.32
2	13.654	4535437 5.68

SM-ec:

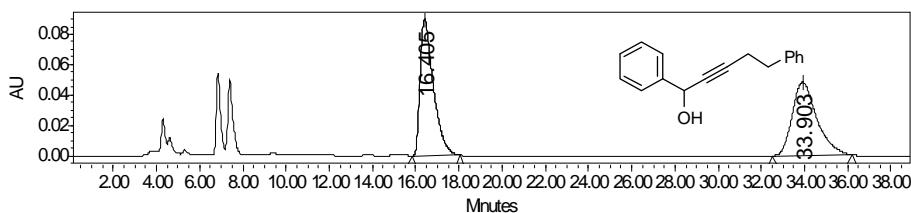


Name	Retention Time	Area % Area
1	14.039	12881844 50.48
2	26.328	12637699 49.52

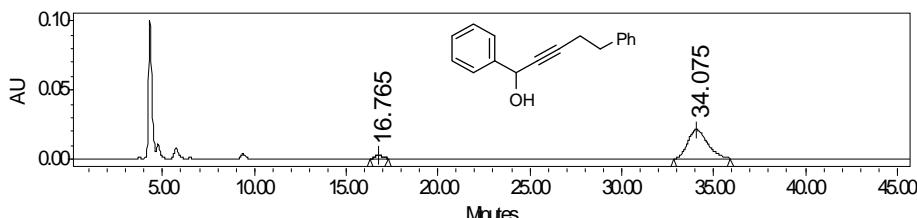


Name	Retention Time	Area % Area
1	13.431	15453516 8.78
2	25.288	160539328 91.22

SM-ed:

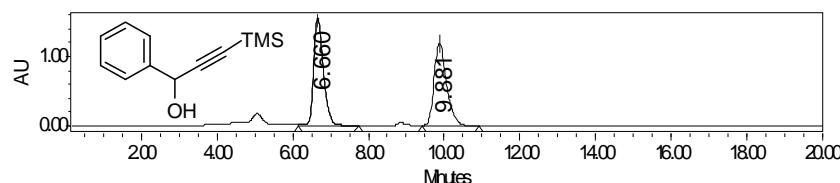


Name	Retention Time	Area	% Area
1	16.405	3835258	50.10
2	33.903	3820121	49.90

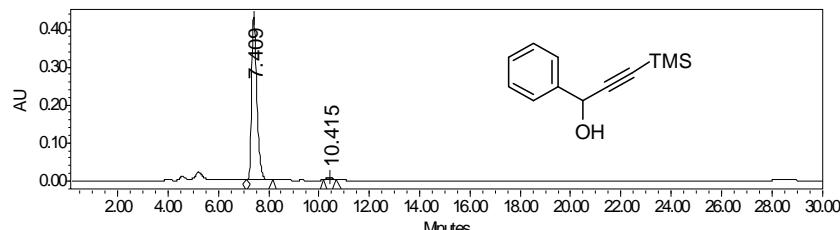


Name	Retention Time	Area	% Area
1	16.765	66450	4.04
2	34.075	1576547	95.96

SM-ee:

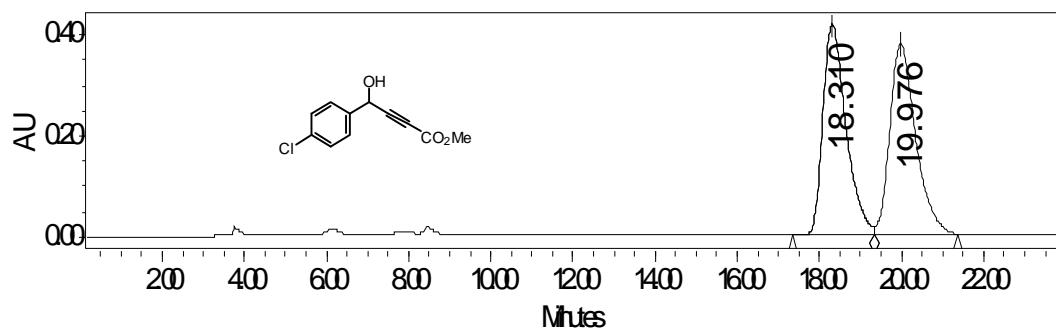


Name	Retention Time	Area	% Area
1	6.660	28021123	50.14
2	9.881	27865086	49.86

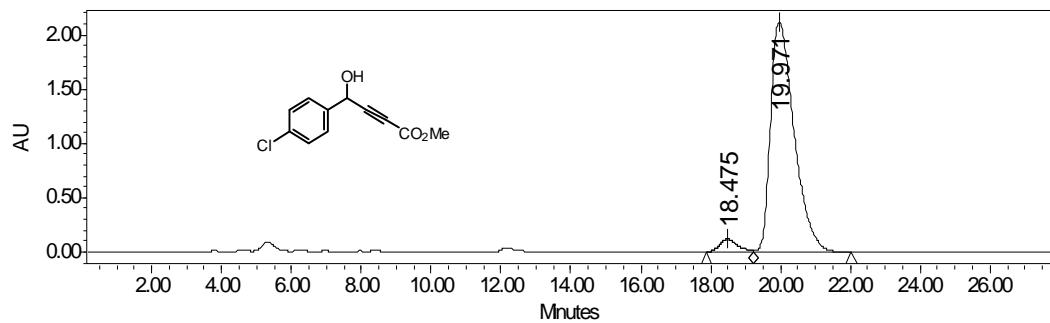


Name	Retention Time	Area	% Area
1	7.409	6157853	98.43
2	10.415	98374	1.57

SM-fa:

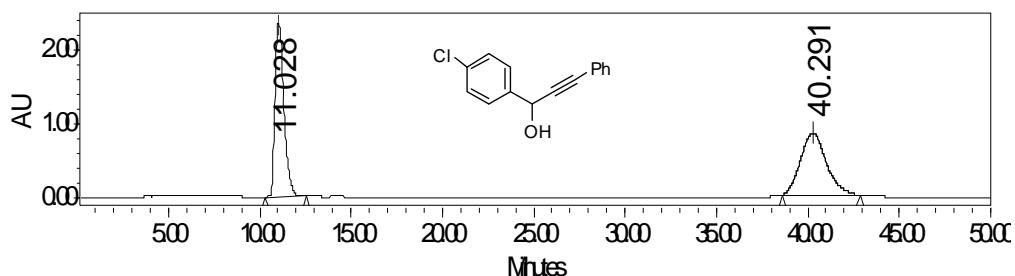


Name	Retention Time	Area	% Area
1	18.310	16463402	49.84
2	19.976	16569596	50.16

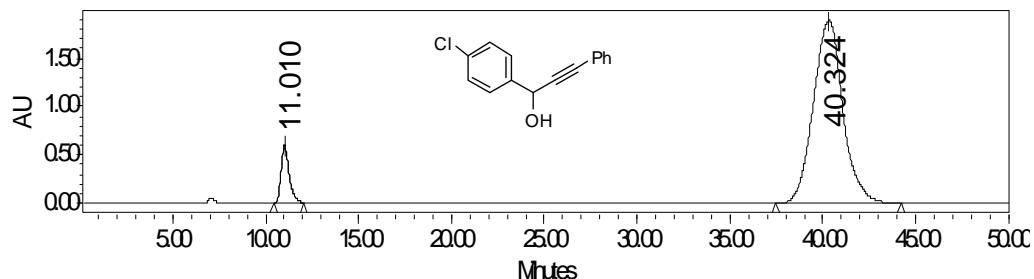


Name	Retention Time	Area	% Area
1	18.475	4246562	4.10
2	19.971	99225708	95.90

SM-fc:

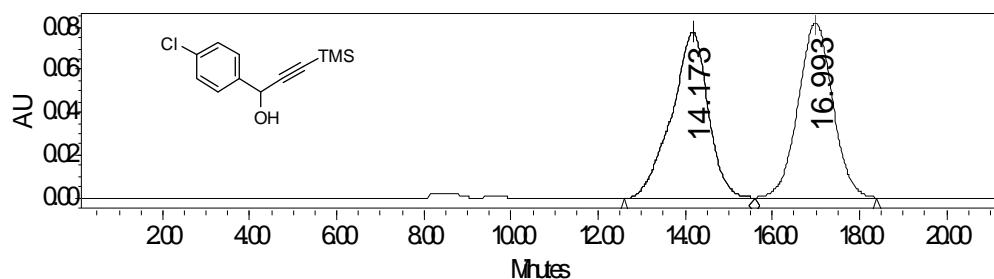


Name	Retention Time	Area	% Area
1	11.028	81629826	49.22
2	40.291	84205753	50.78

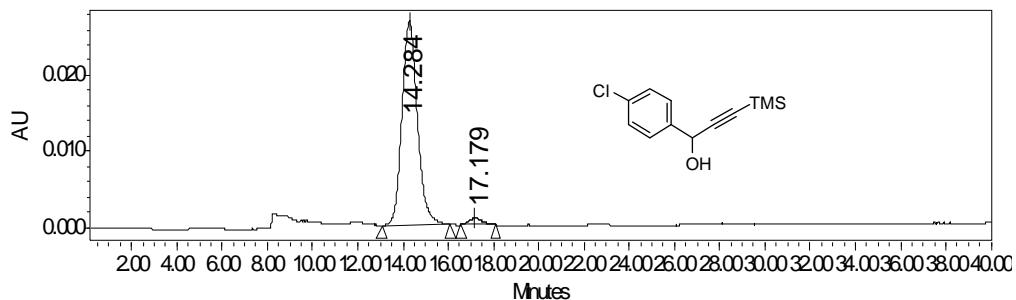


Name	Retention Time	Area	% Area
1	11.010	17816482	7.88
2	40.324	208423689	92.12

SM-fe:

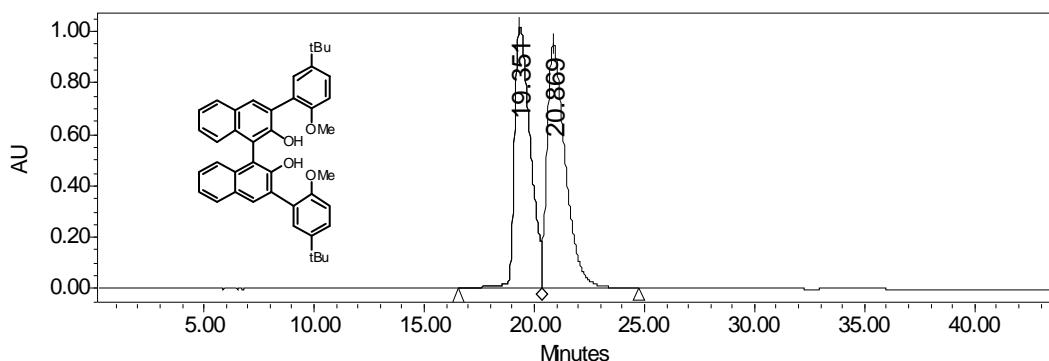


Name	Retention Time	Area	% Area
1	14.173	4595271	50.18
2	16.993	4562491	49.82

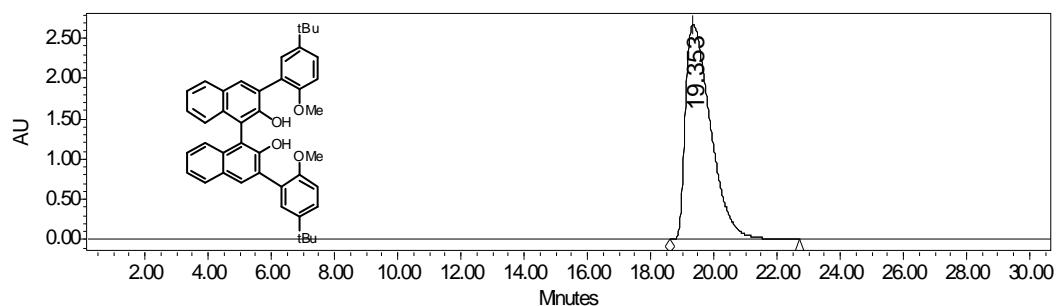


Name	Retention Time	Area	% Area
1	14.284	1270145	97.07
2	17.179	38350	2.93

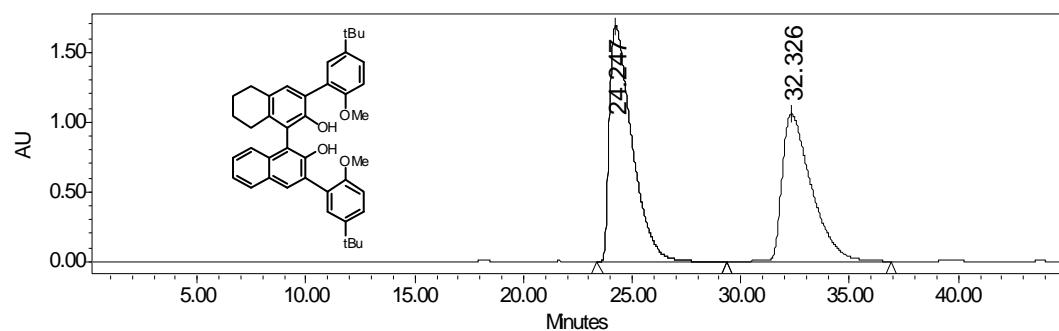
Partially racemized (*S*)-4



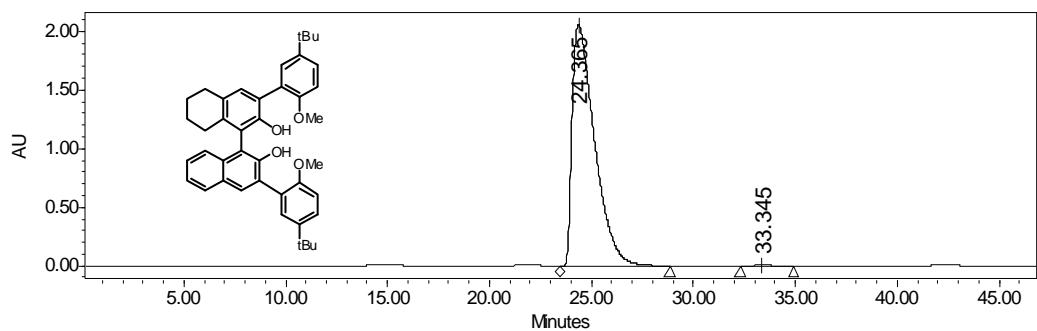
(*S*)-4



Partially racemized (*S*)-6

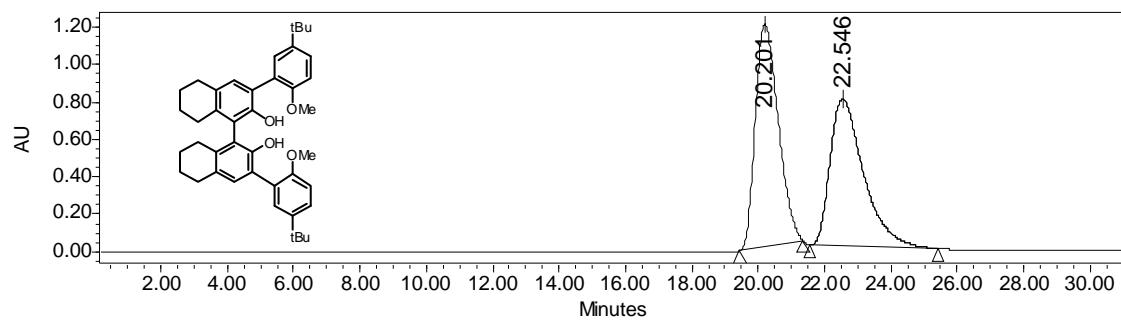


(S)-6



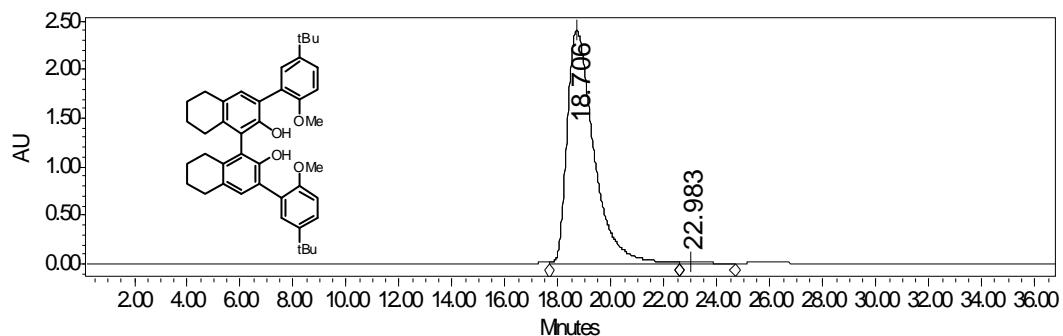
Name	Retention Time	Area	% Area
1	24.365	157558152	99.62
2	33.345	600295	0.38

Racemic 7



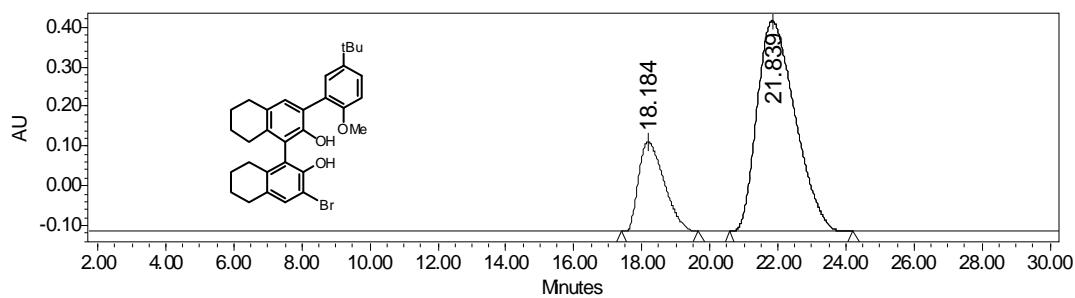
Name	Retention Time	Area	% Area
1	20.201	57158610	50.79
2	22.546	55378818	49.21

(S)-7



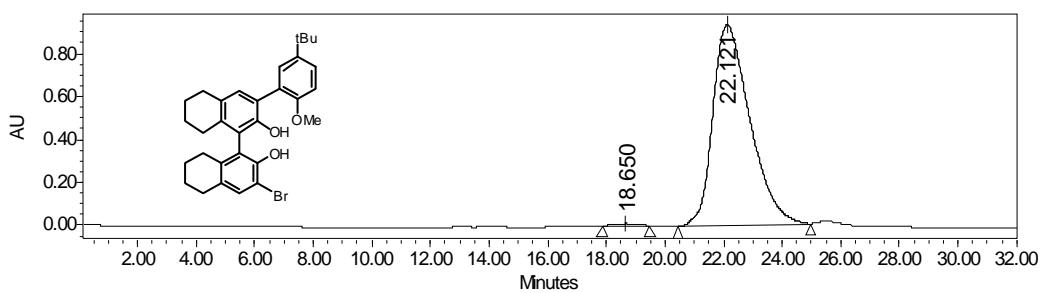
Name	Retention Time	Area	% Area
1	18.706	169810183	99.10
2	22.983	1544576	0.90

Partially racemized (*S*)-8



Name	Retention Time	Area	% Area
1	18.184	11658394	22.11
2	21.839	41076770	77.89

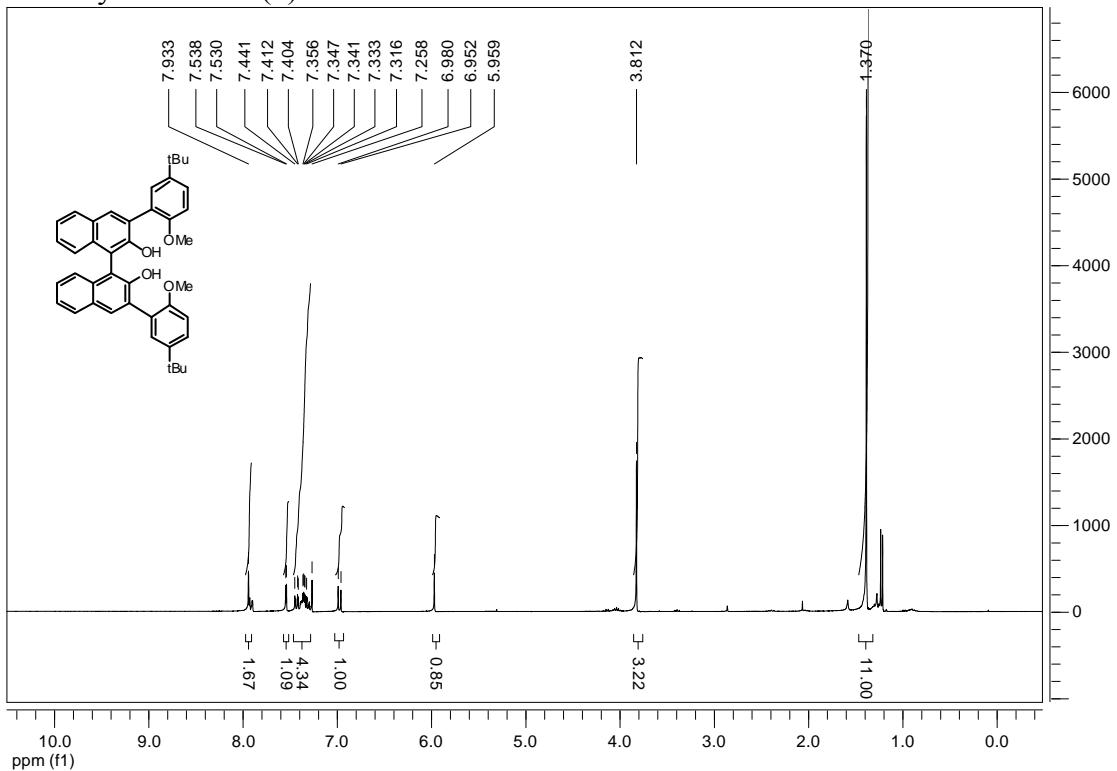
(*S*)-8



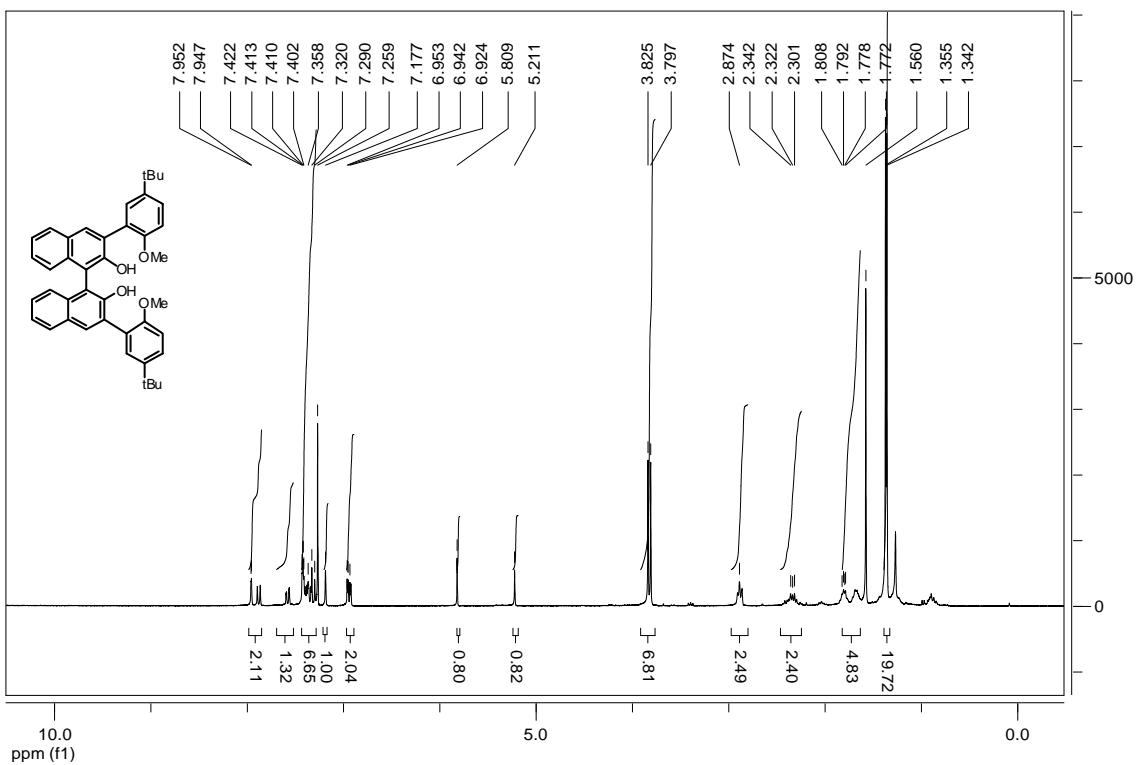
Name	Retention Time	Area	% Area
1	18.650	541518	0.65
2	22.121	82555238	99.35

¹H NMR Spectra of Partially Racemized Ligands to Determine Purity

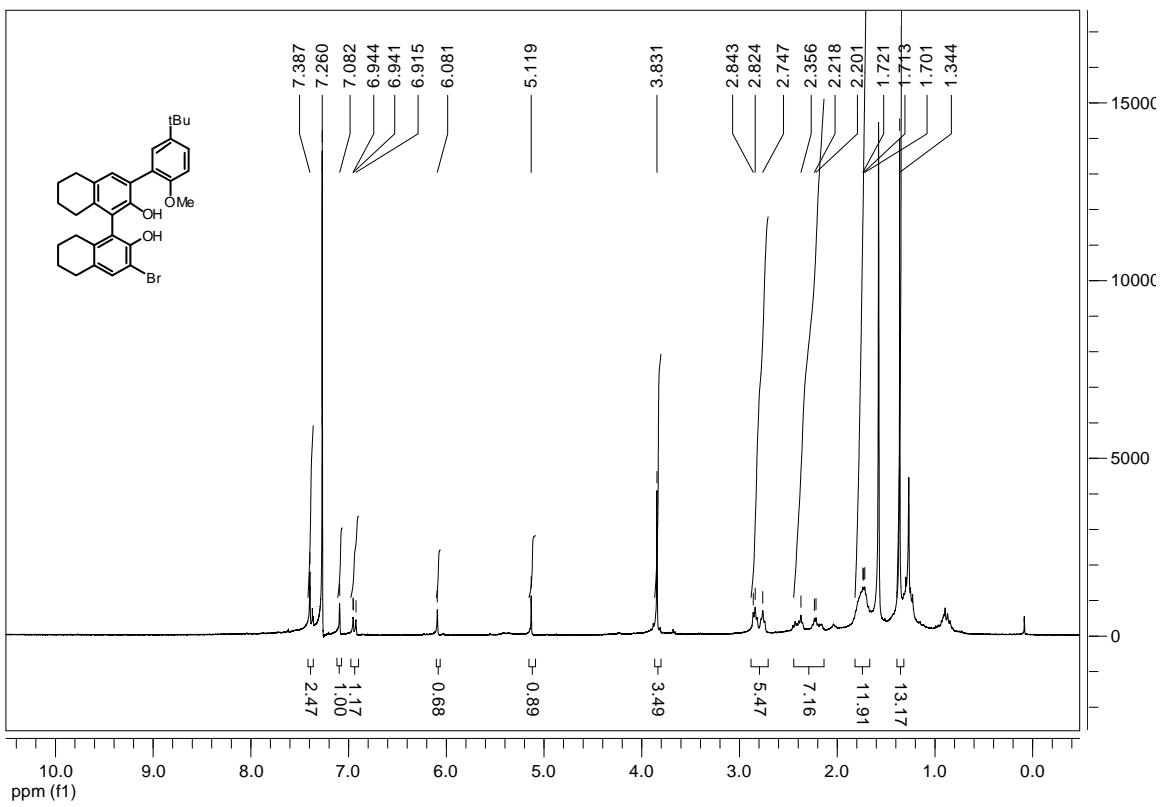
Partially racemized (*S*)-4



Partially racemized (*S*)-6

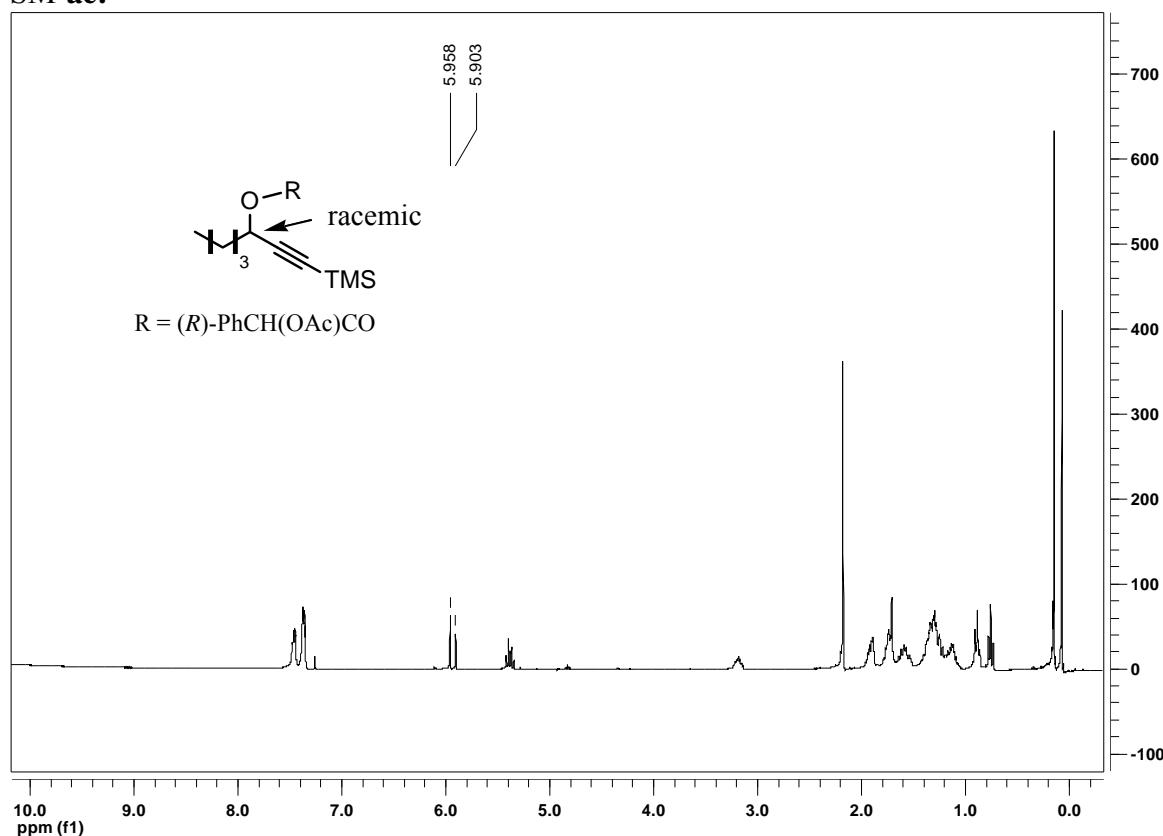


Partially racemized (*S*)-8

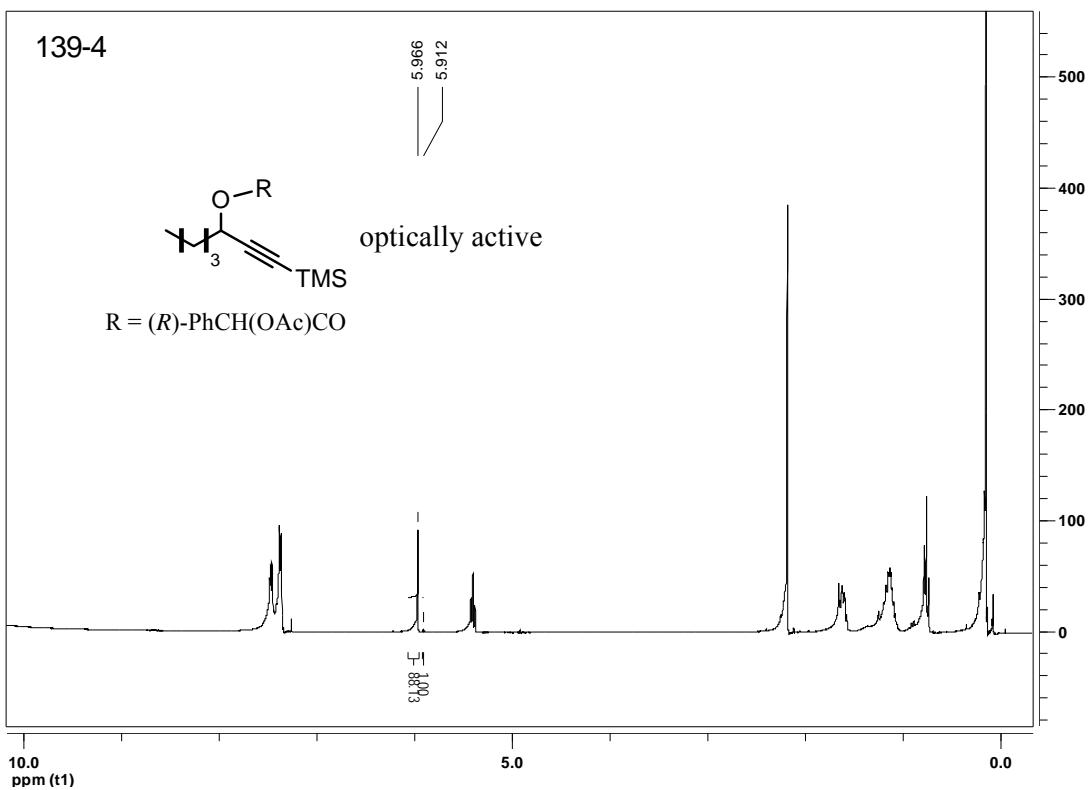
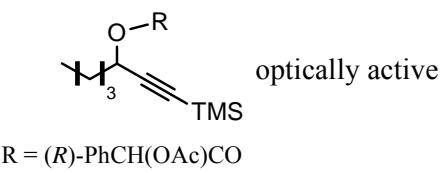


^1H NMR Spectra of the Esters Prepared from the Propargylic Alcohols and $(R)\text{-PhCH(OAc)CO}_2\text{H}$ for ee Determination

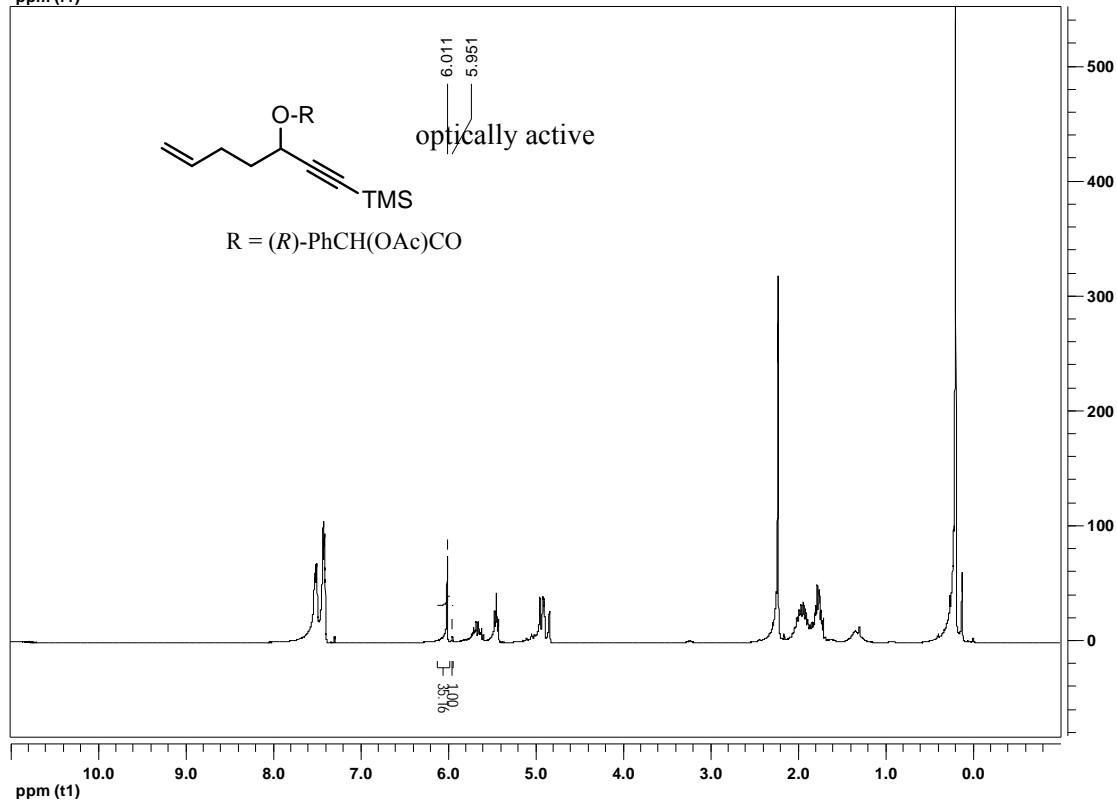
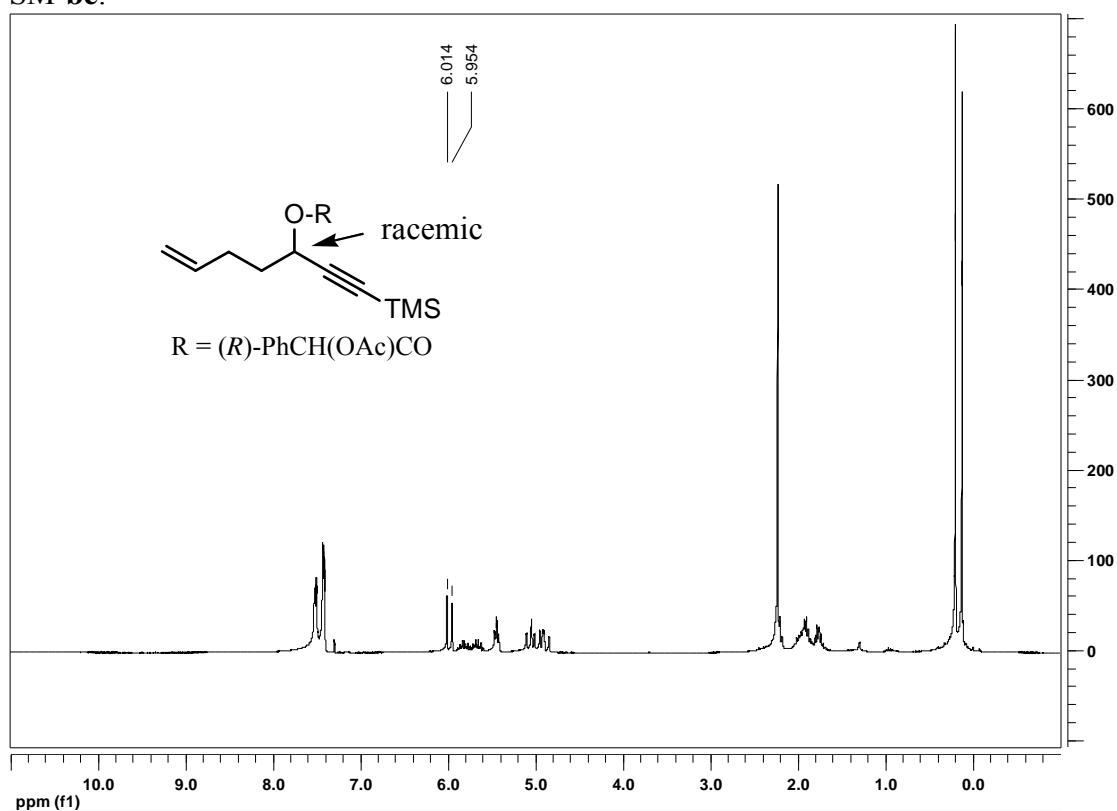
SM-ae:



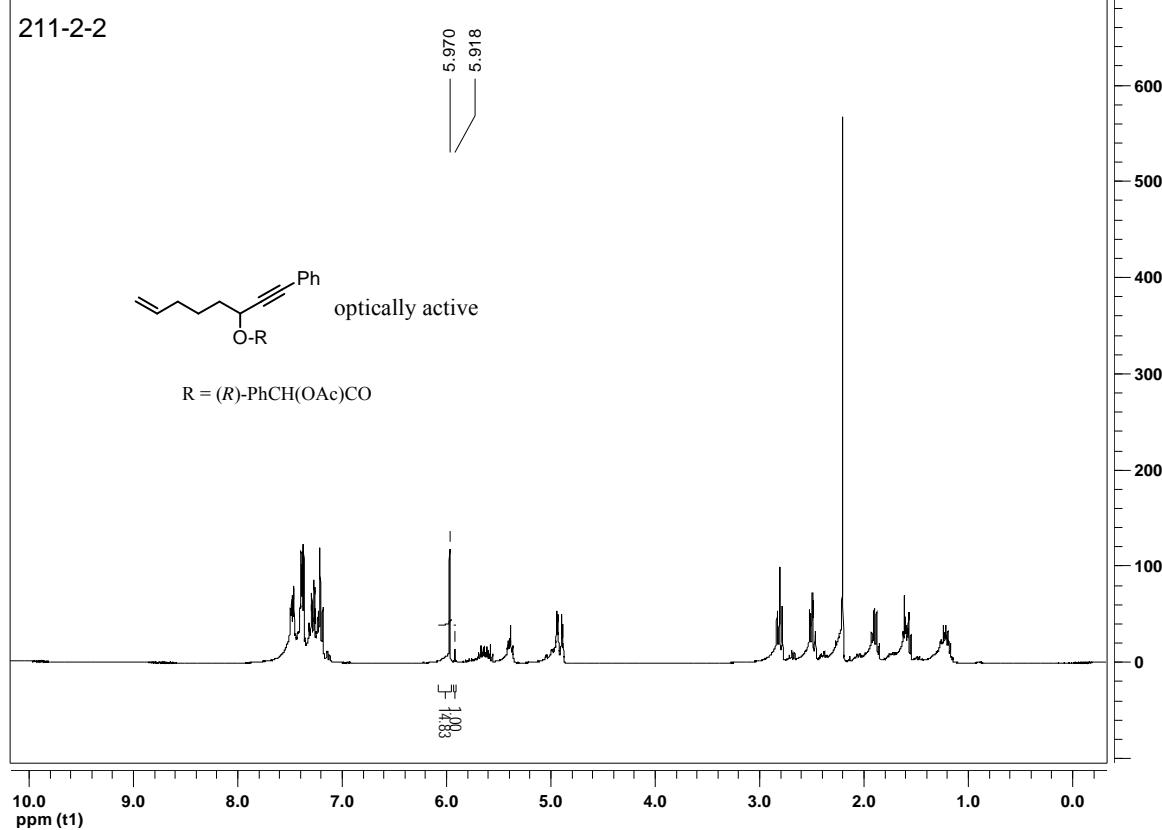
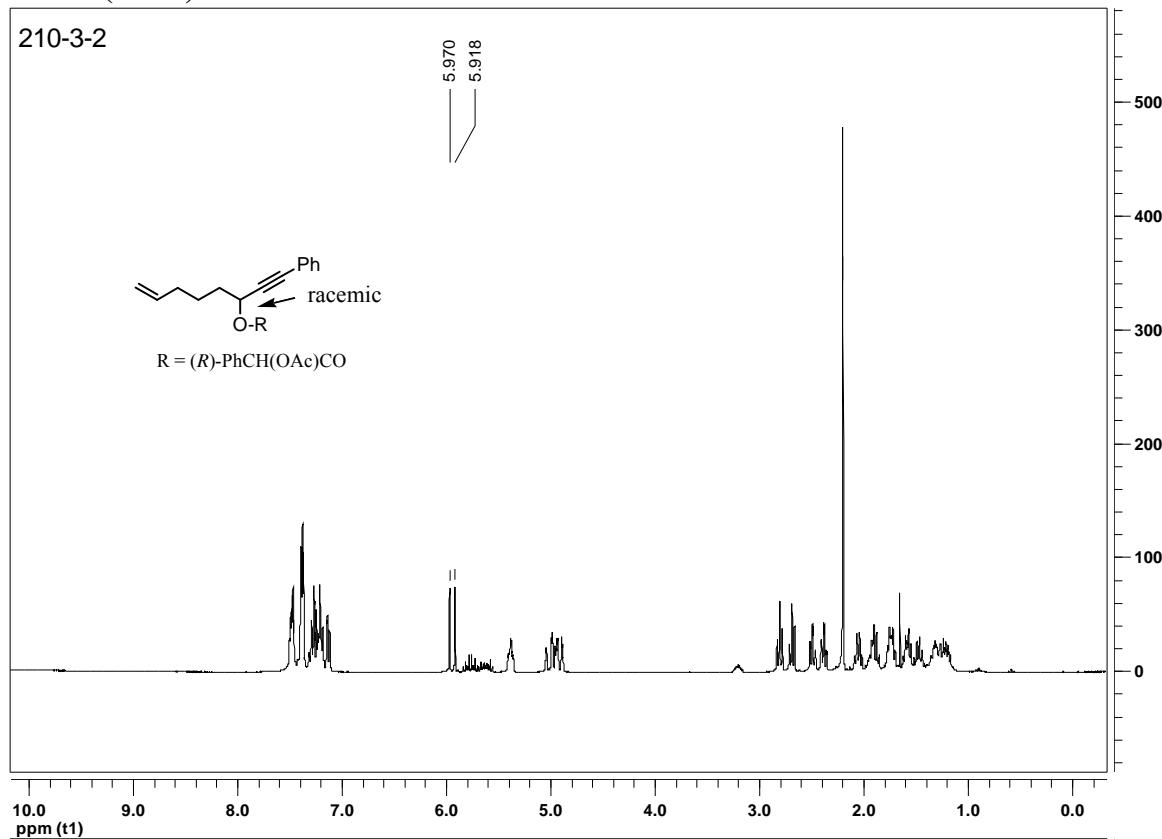
139-4



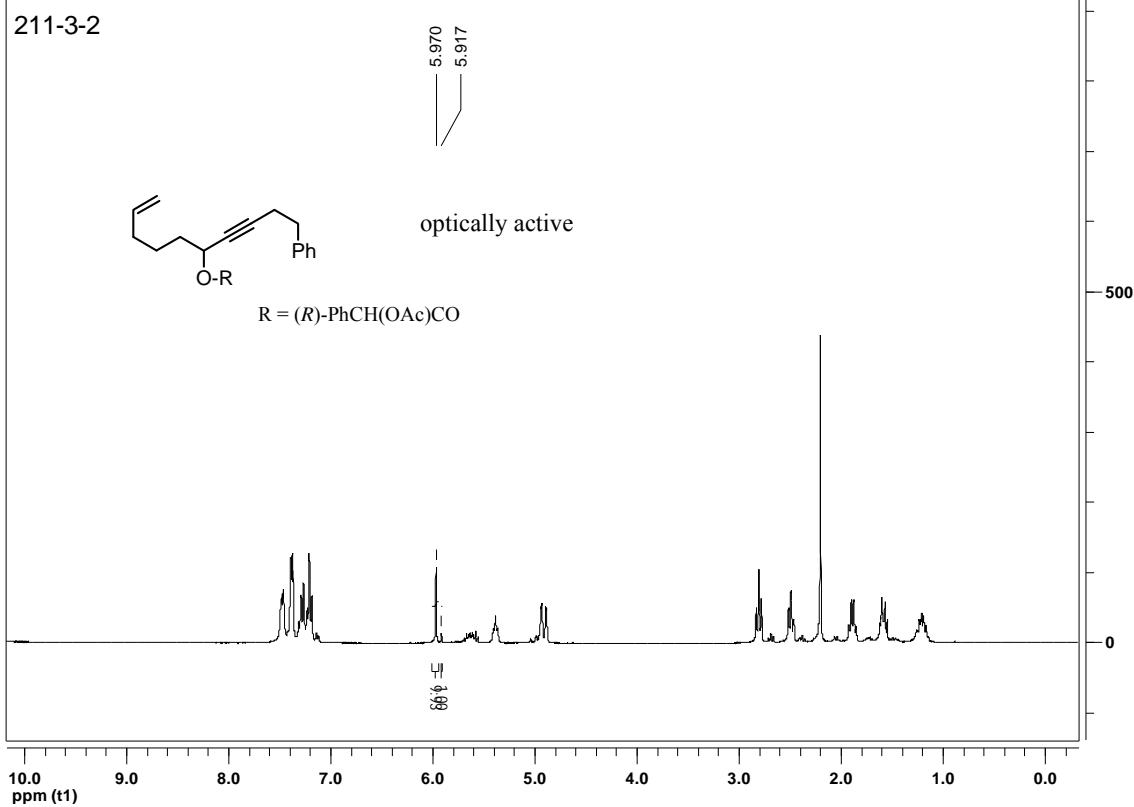
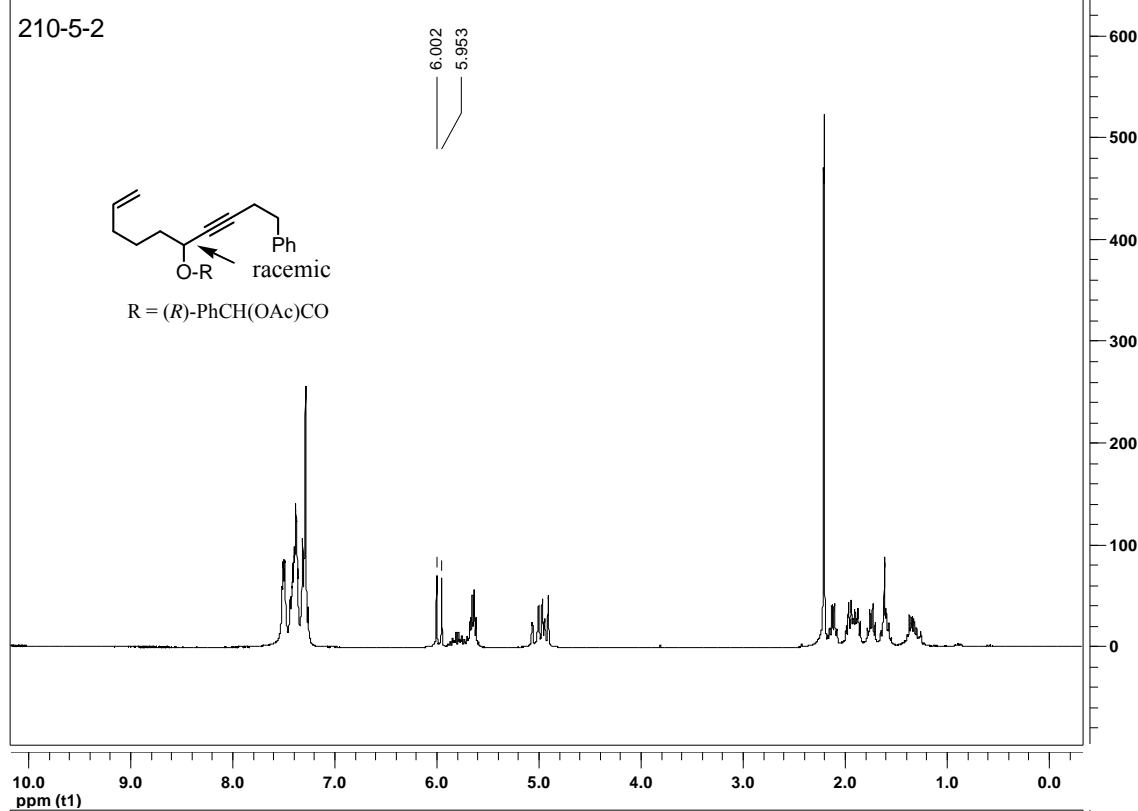
SM-be:



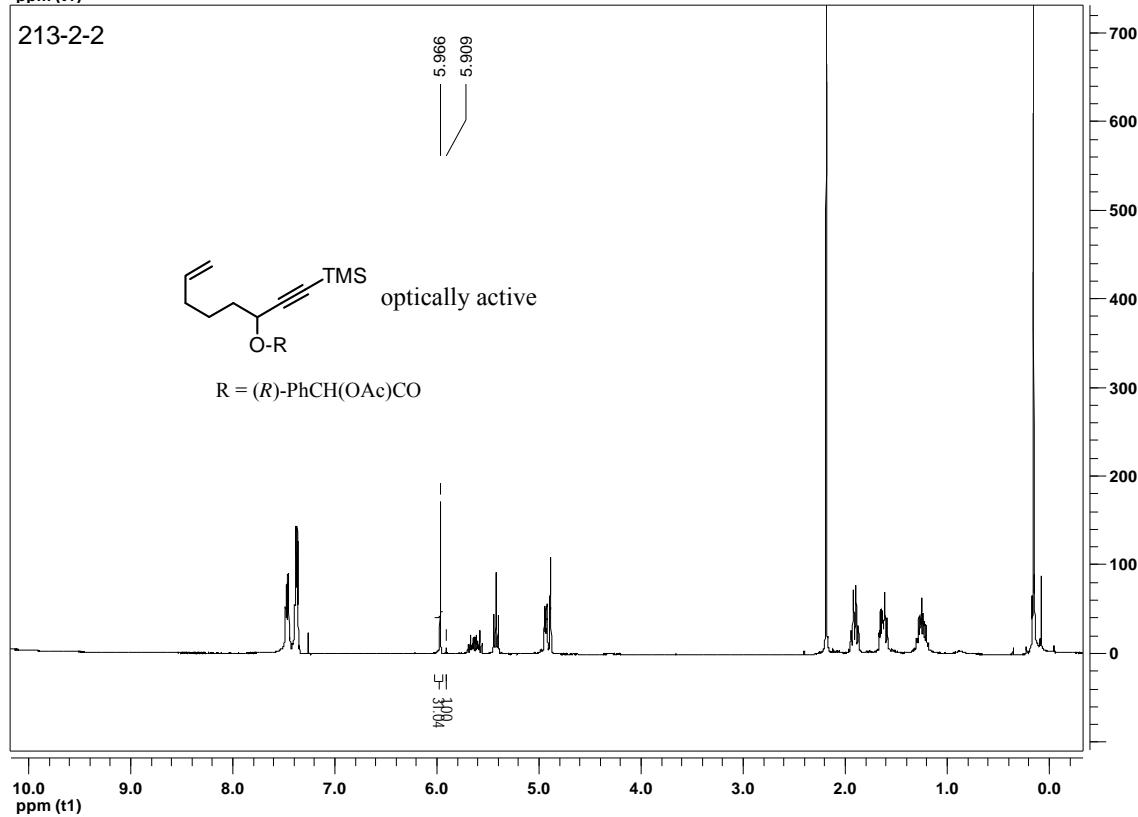
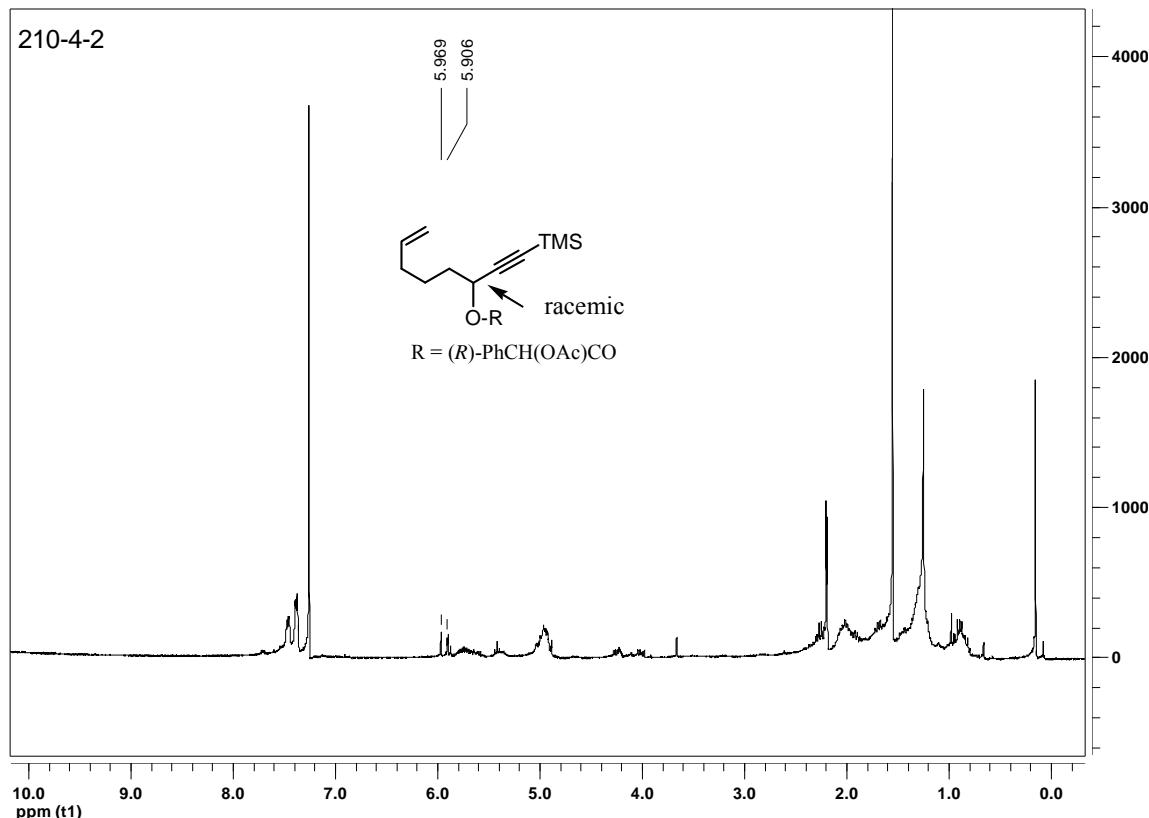
SM-cc (= P-4):



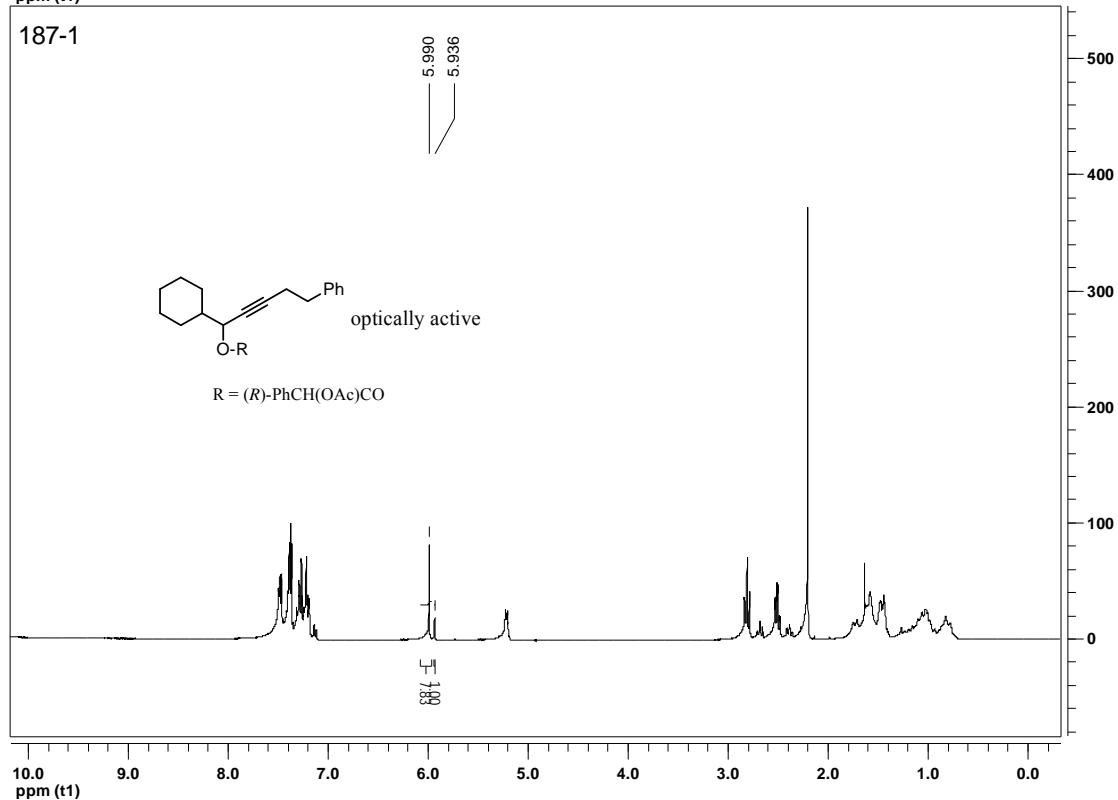
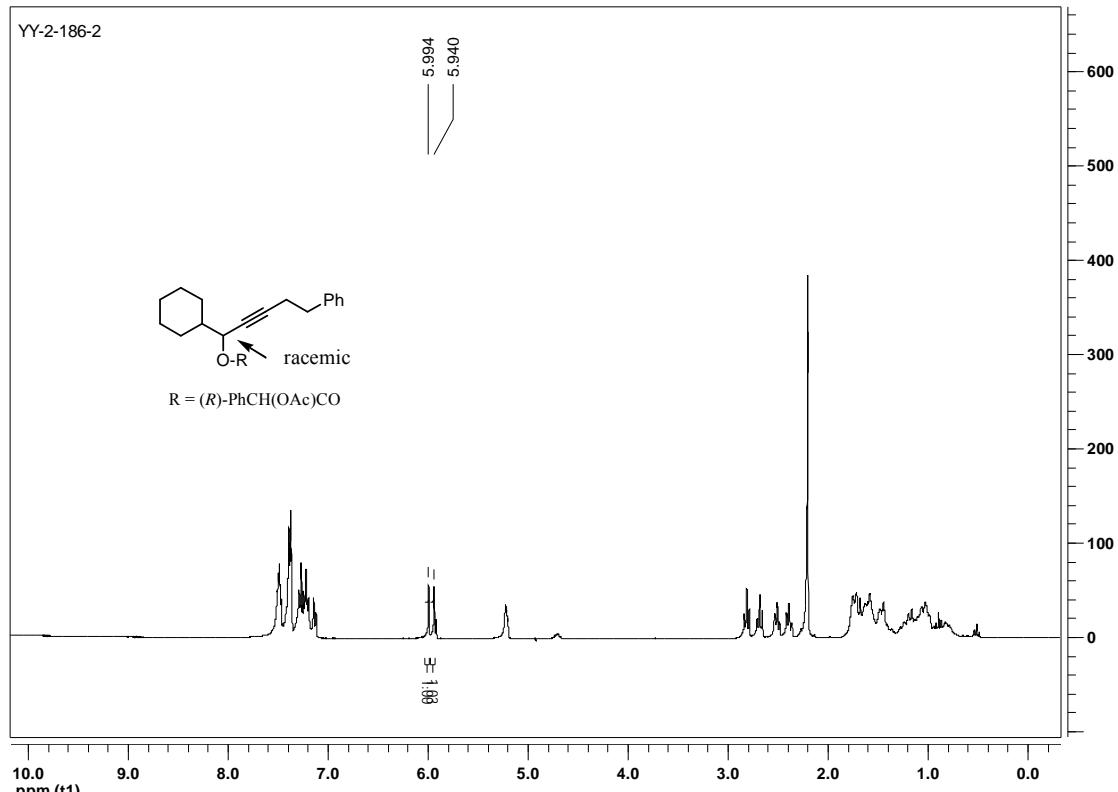
SM-cd (= P-5)



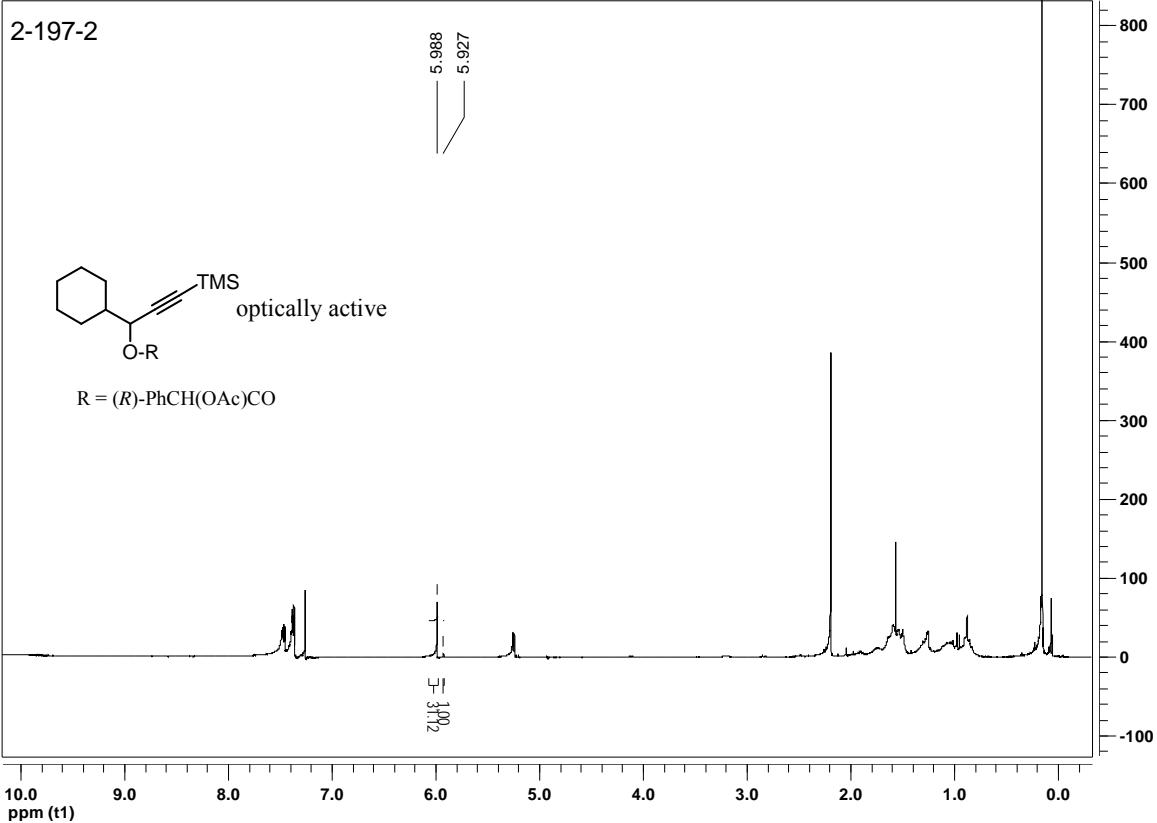
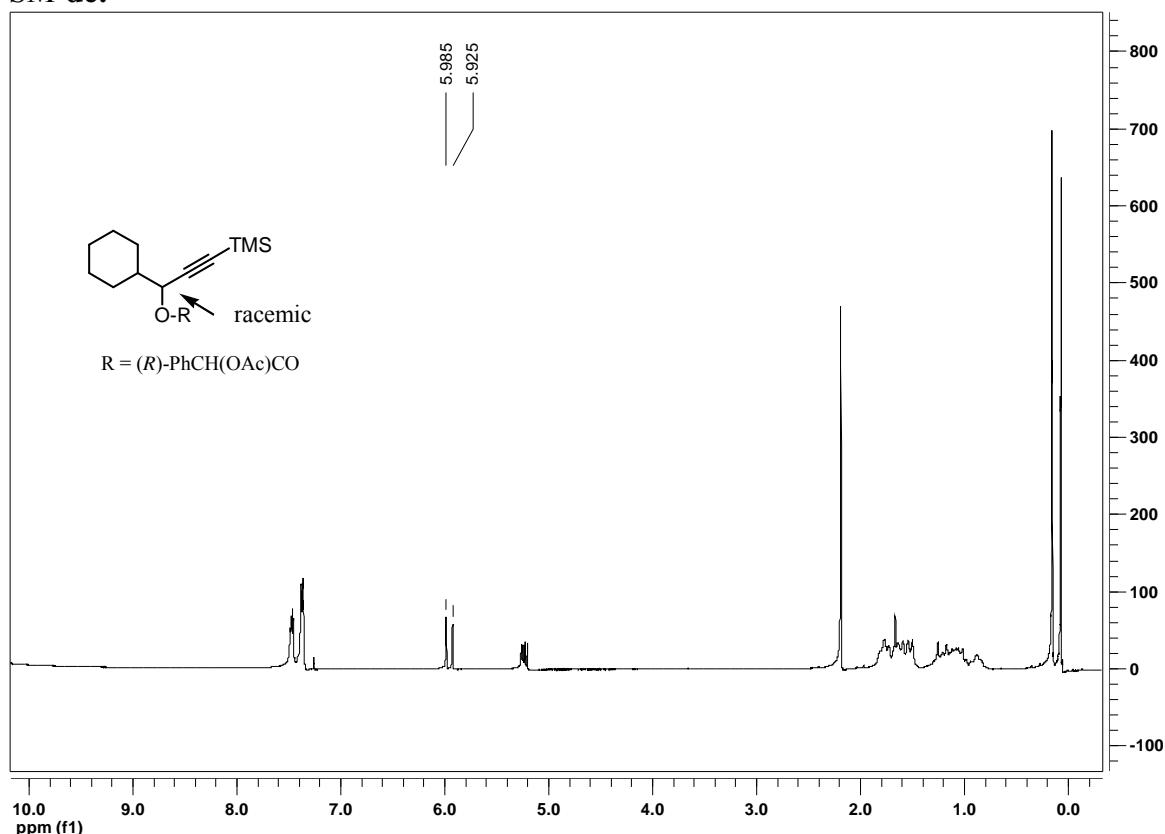
SM-ce:



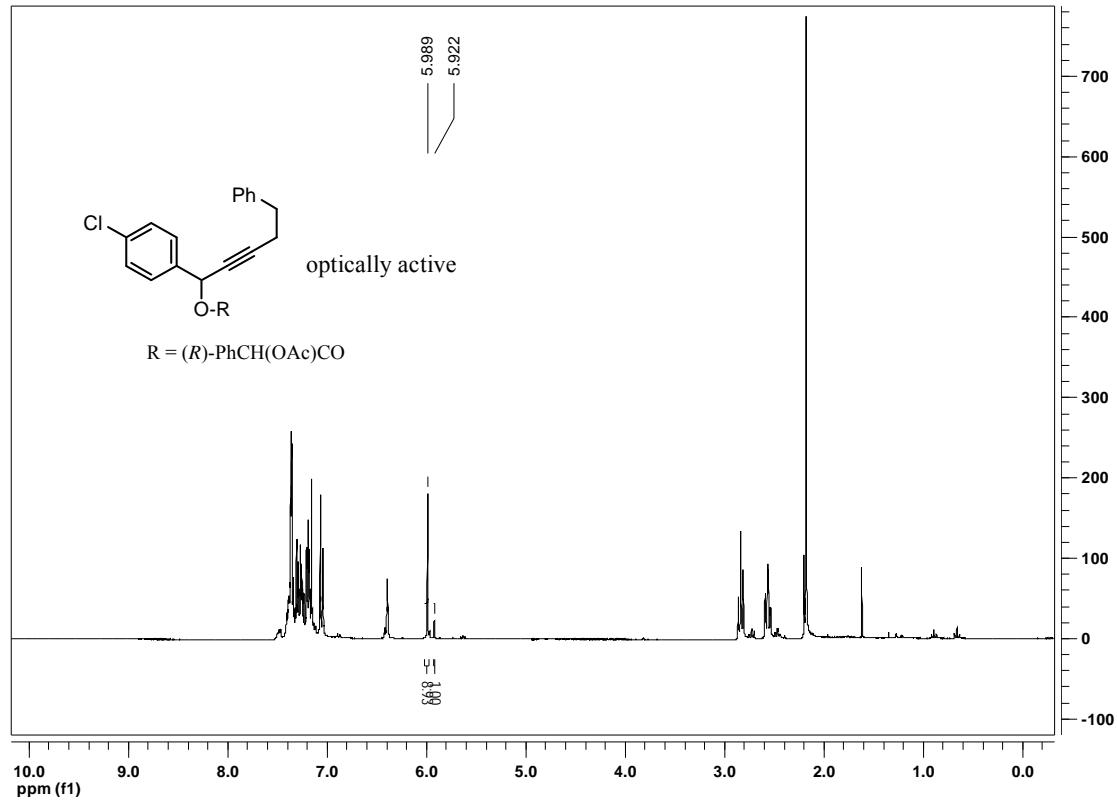
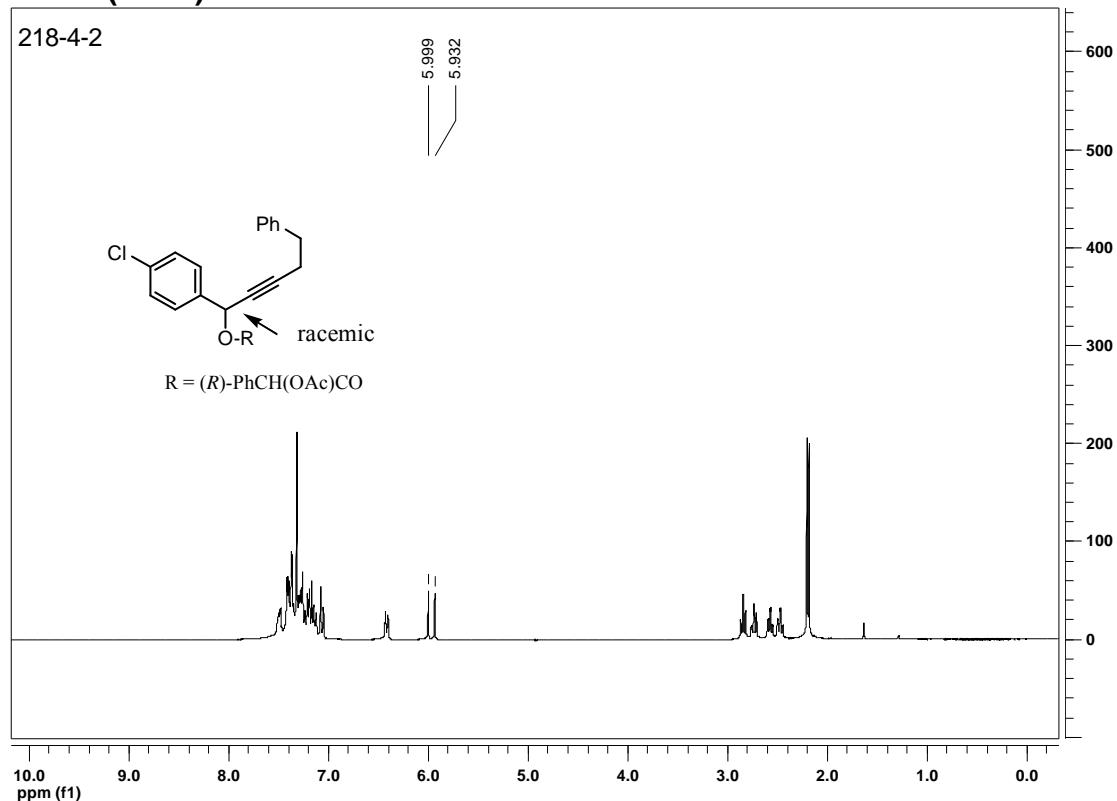
SM-dd:



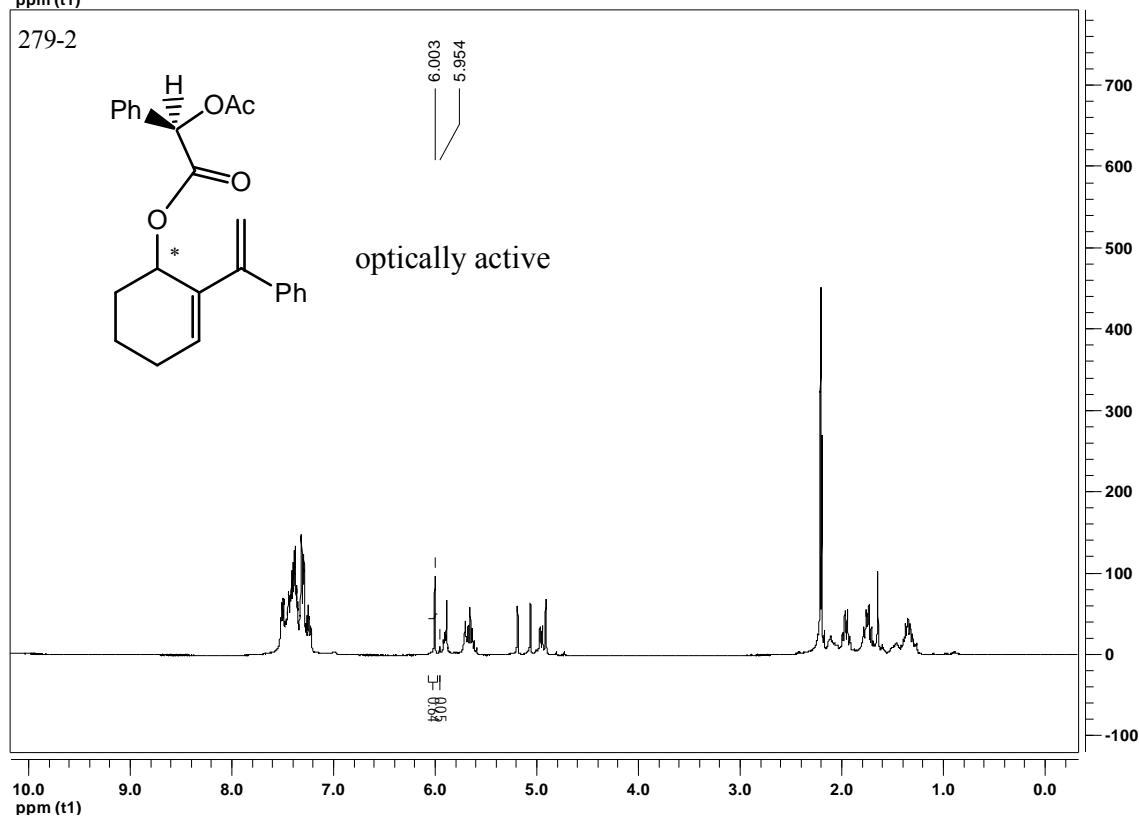
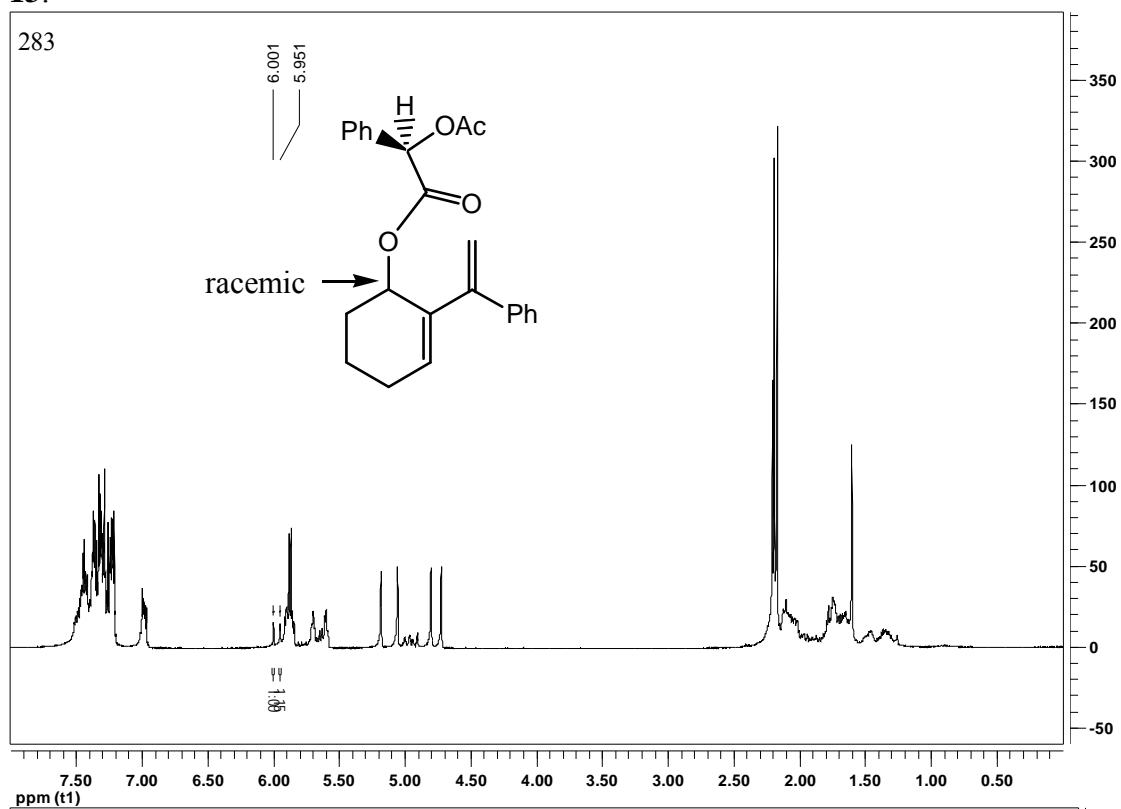
SM-de:



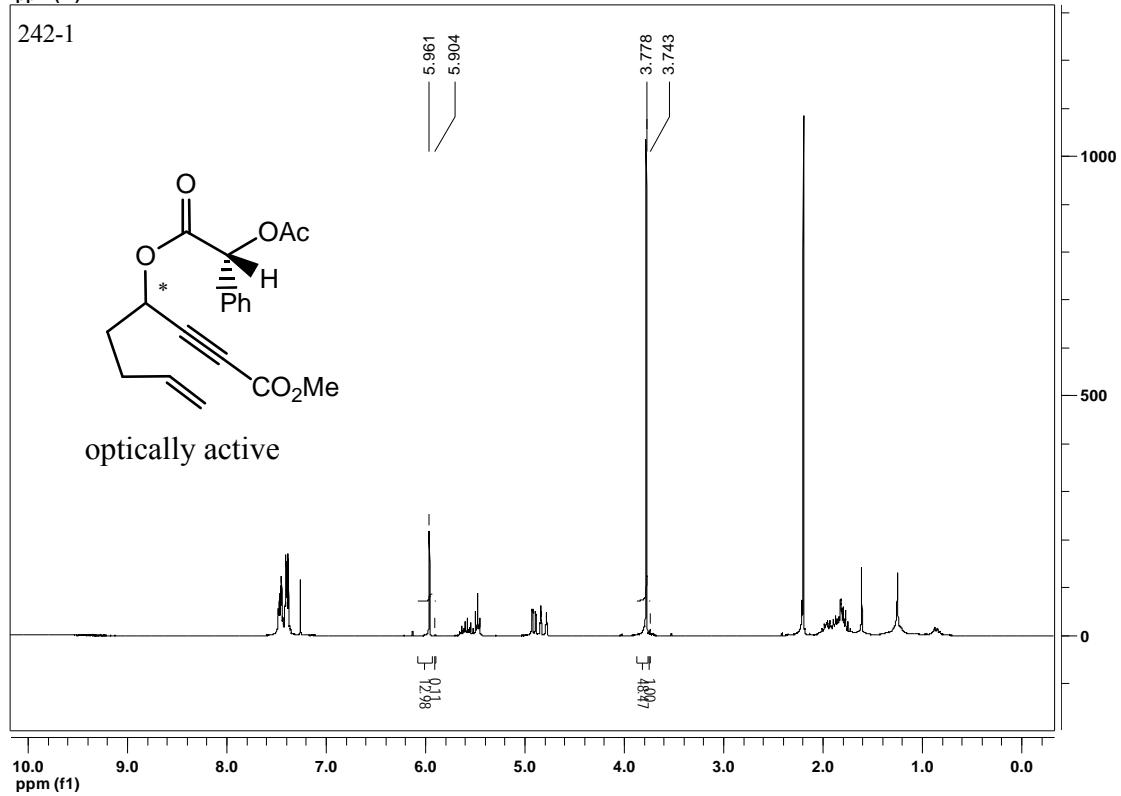
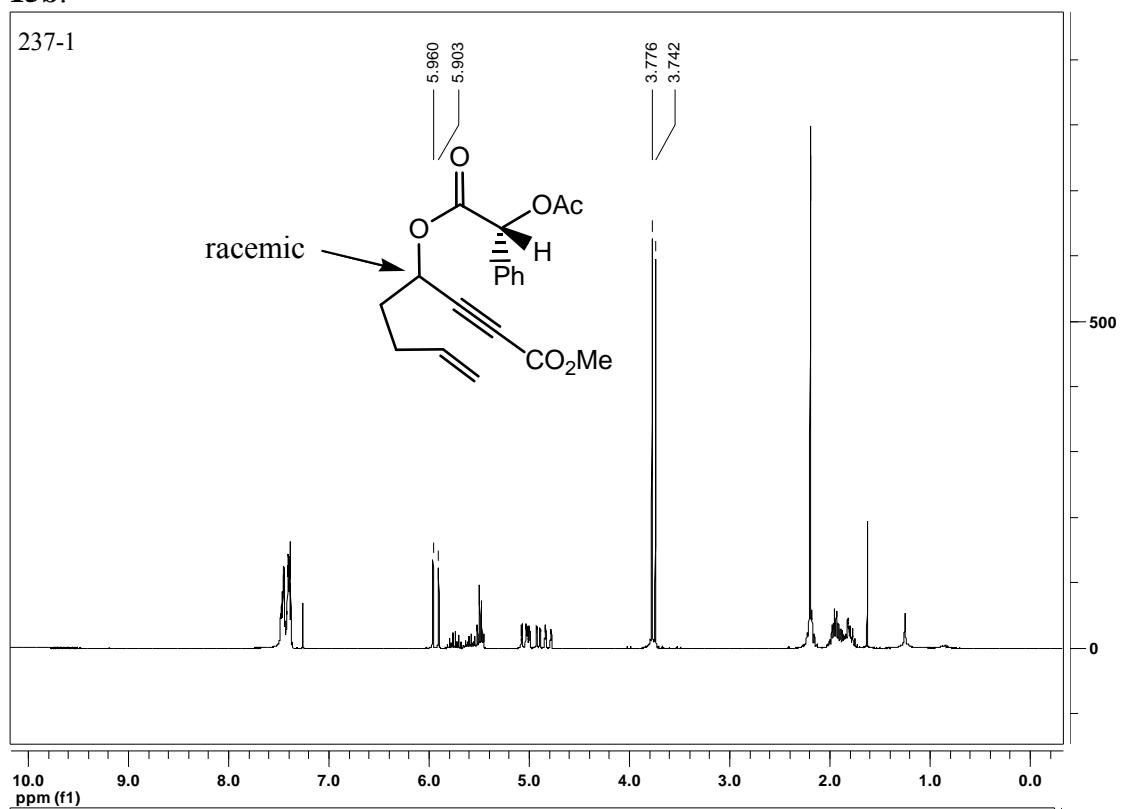
SM-fd (= P-6):



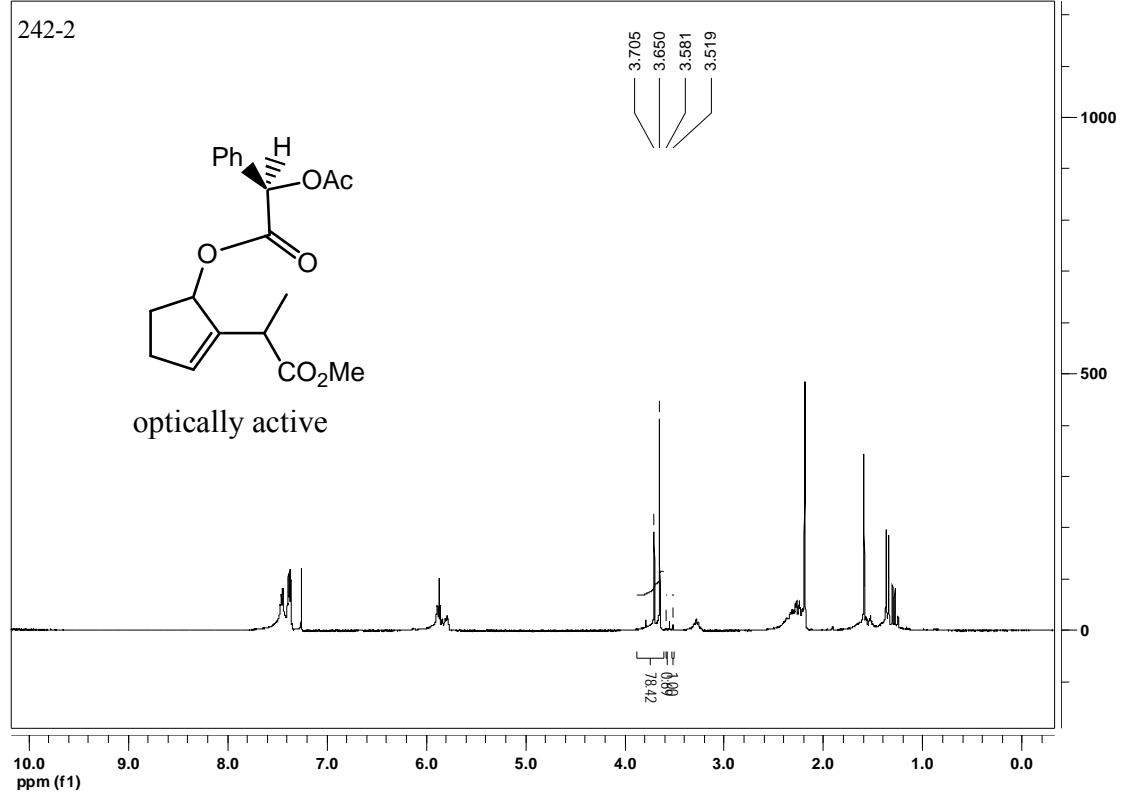
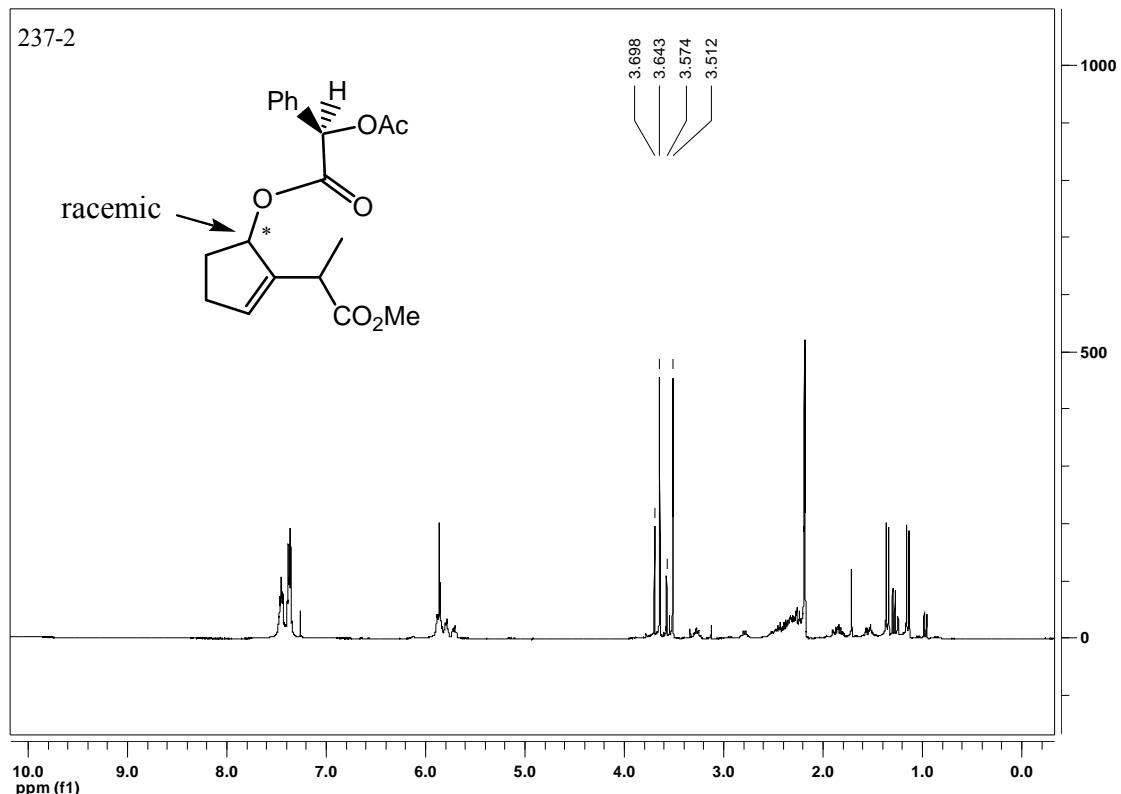
13:



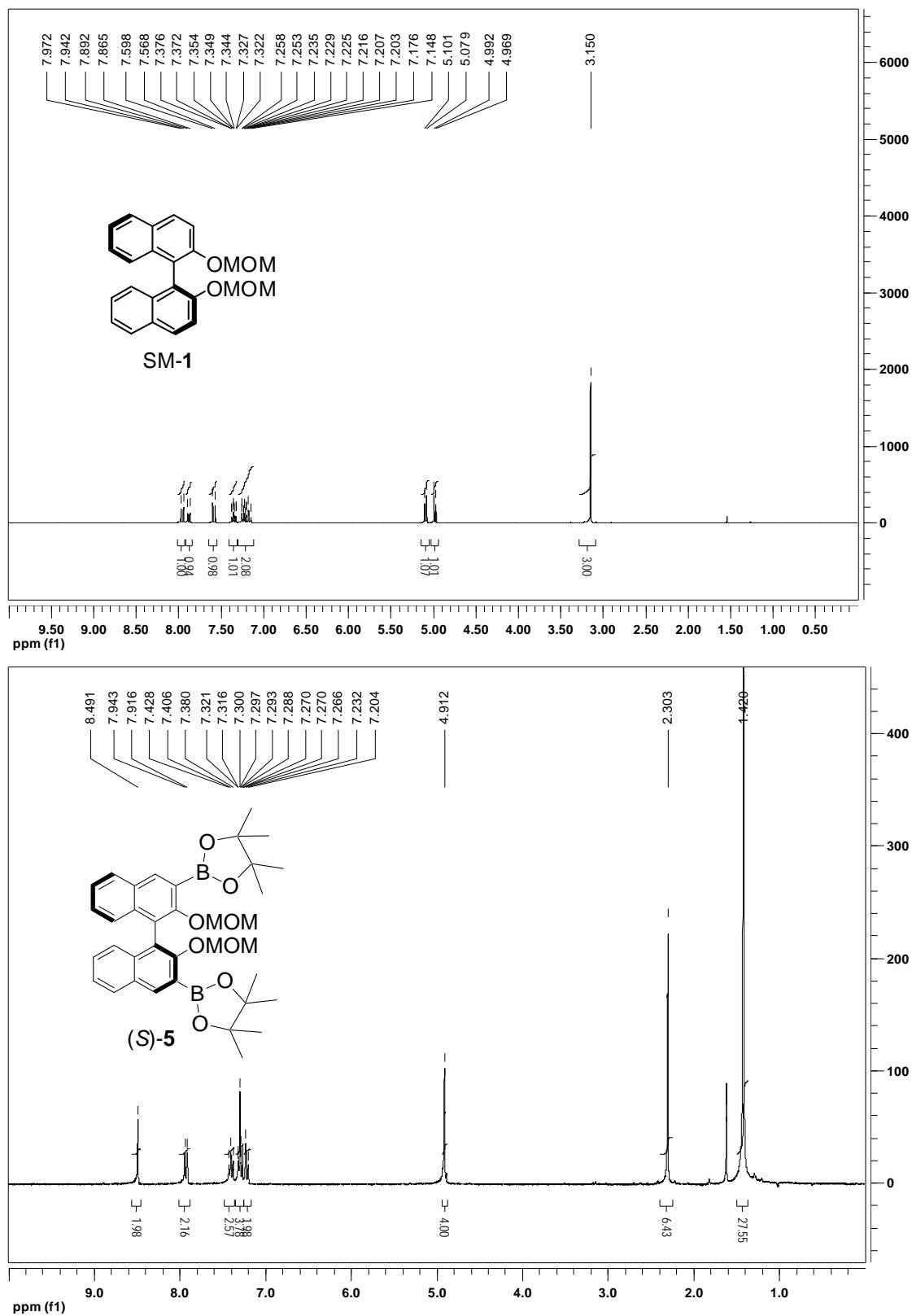
15b:

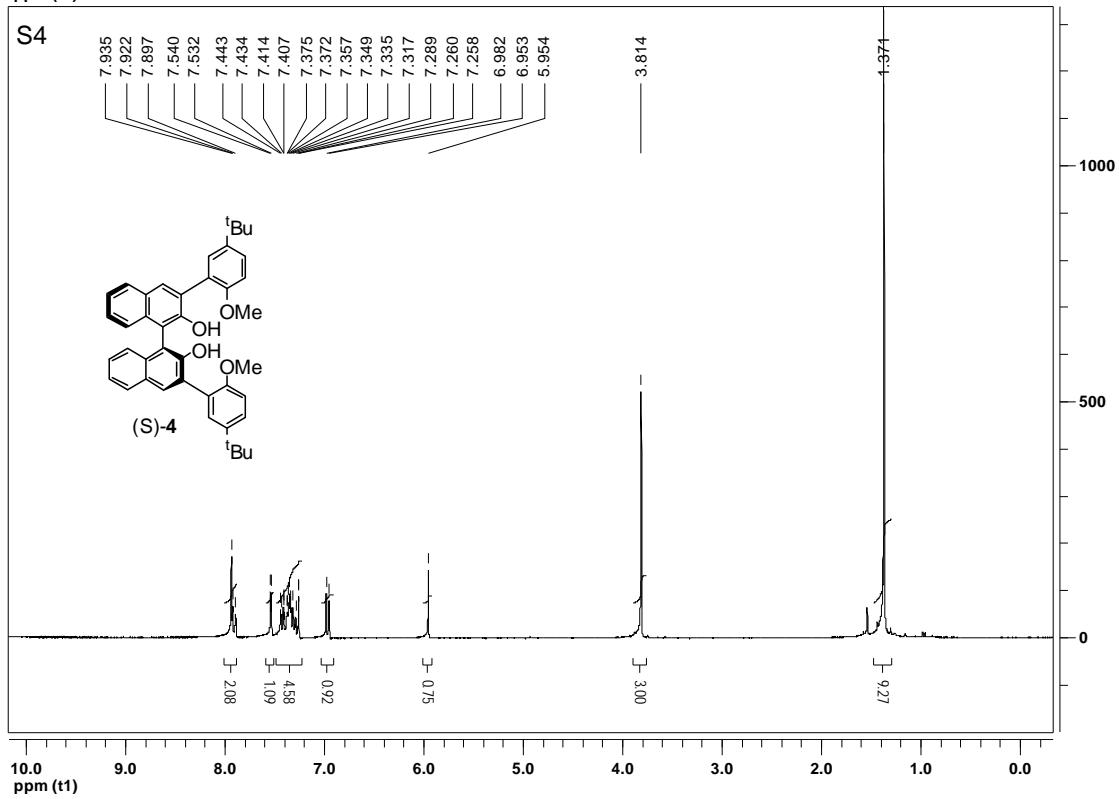
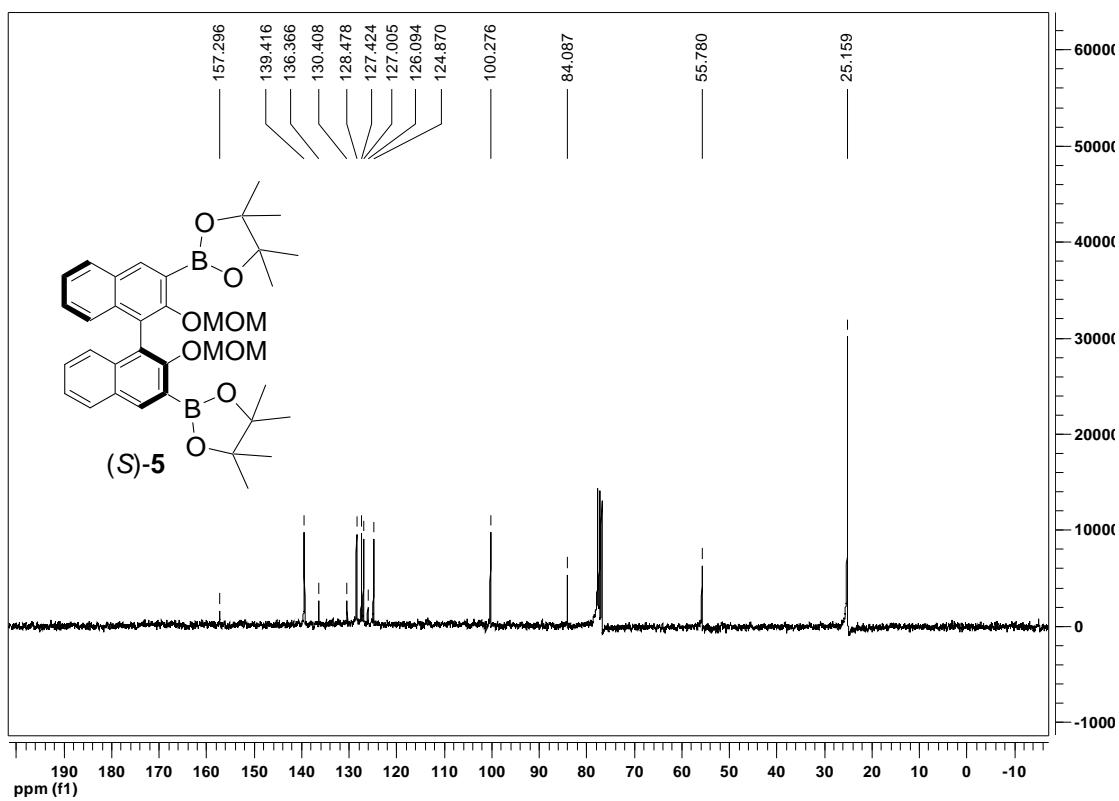


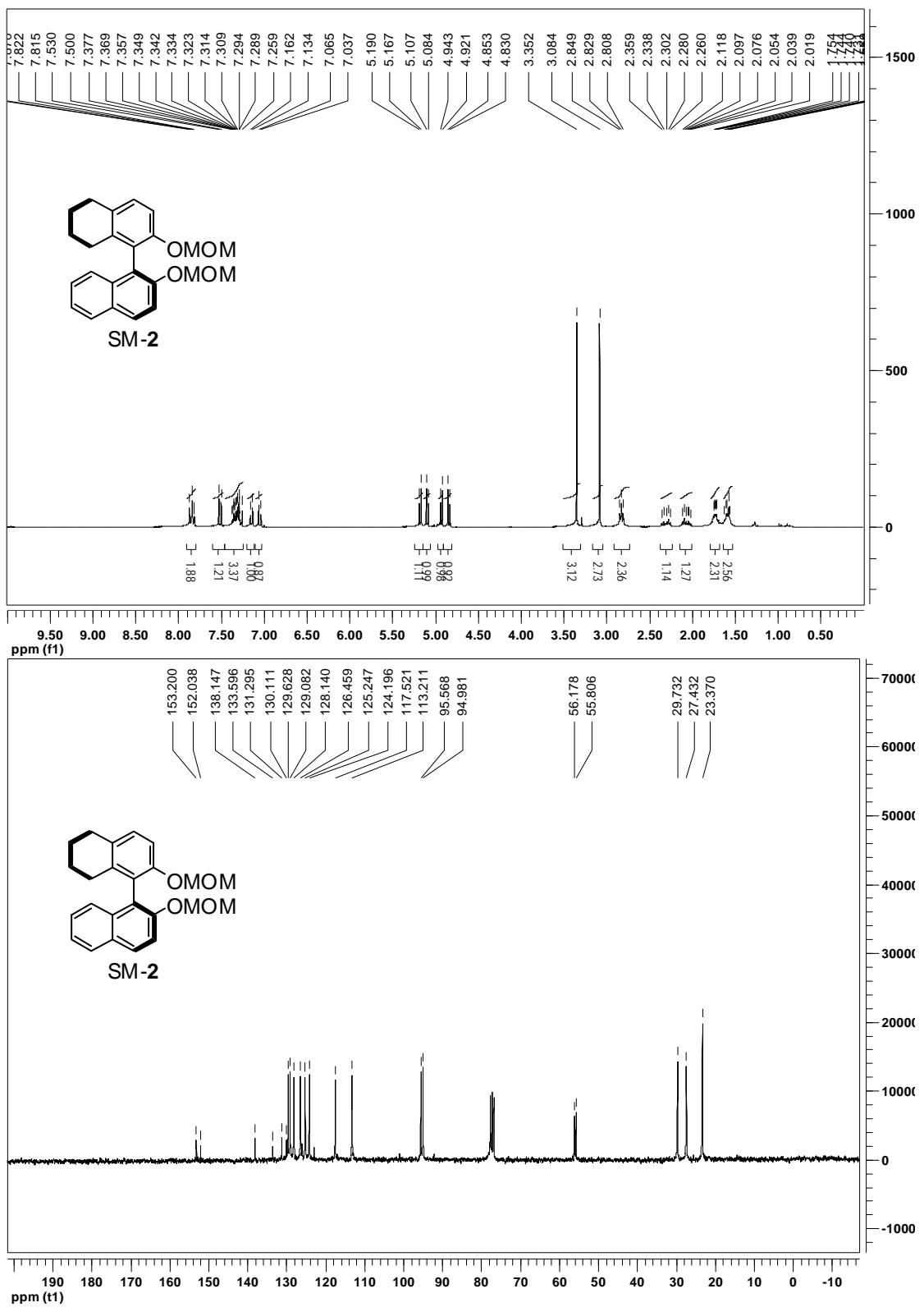
16b:

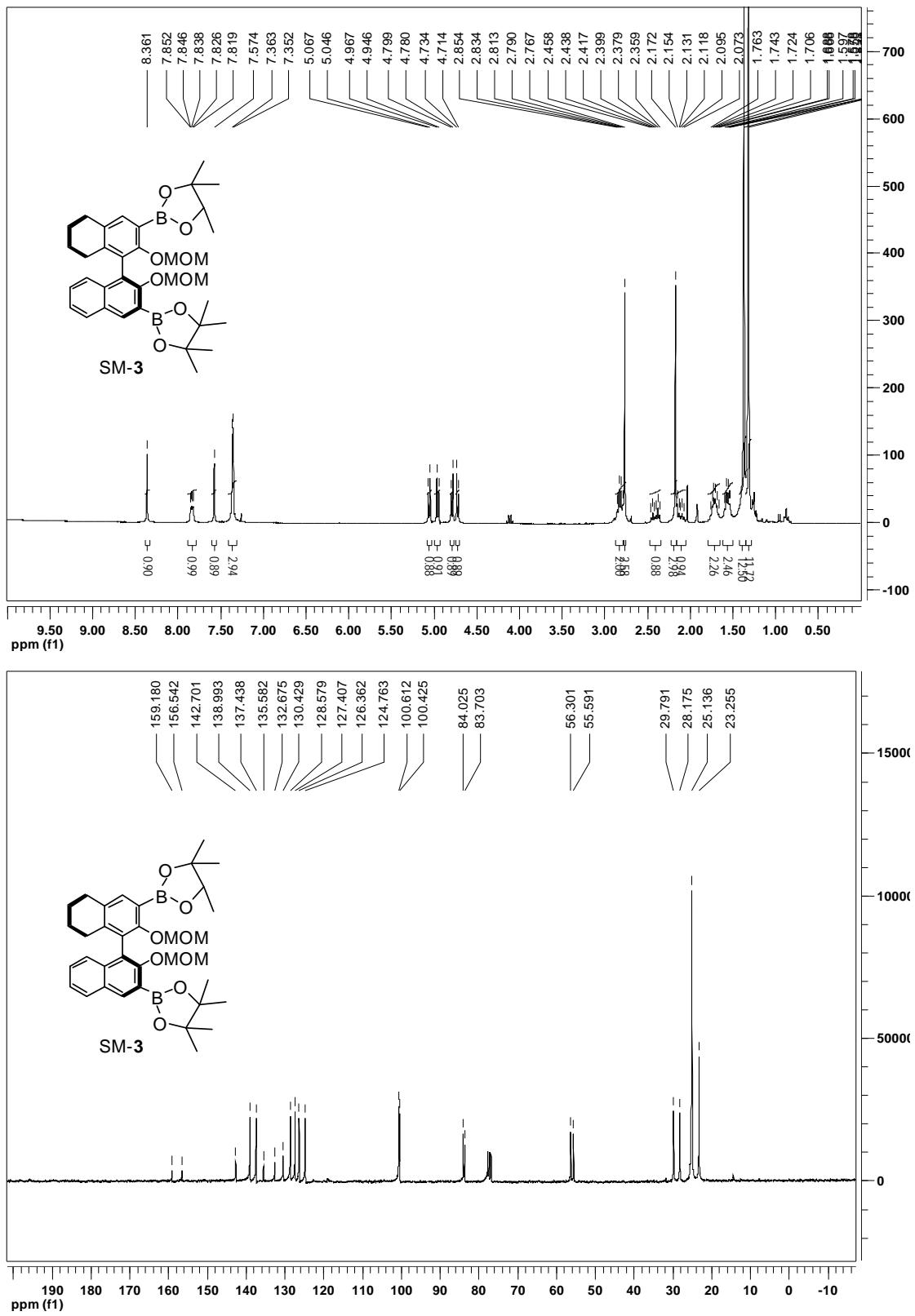


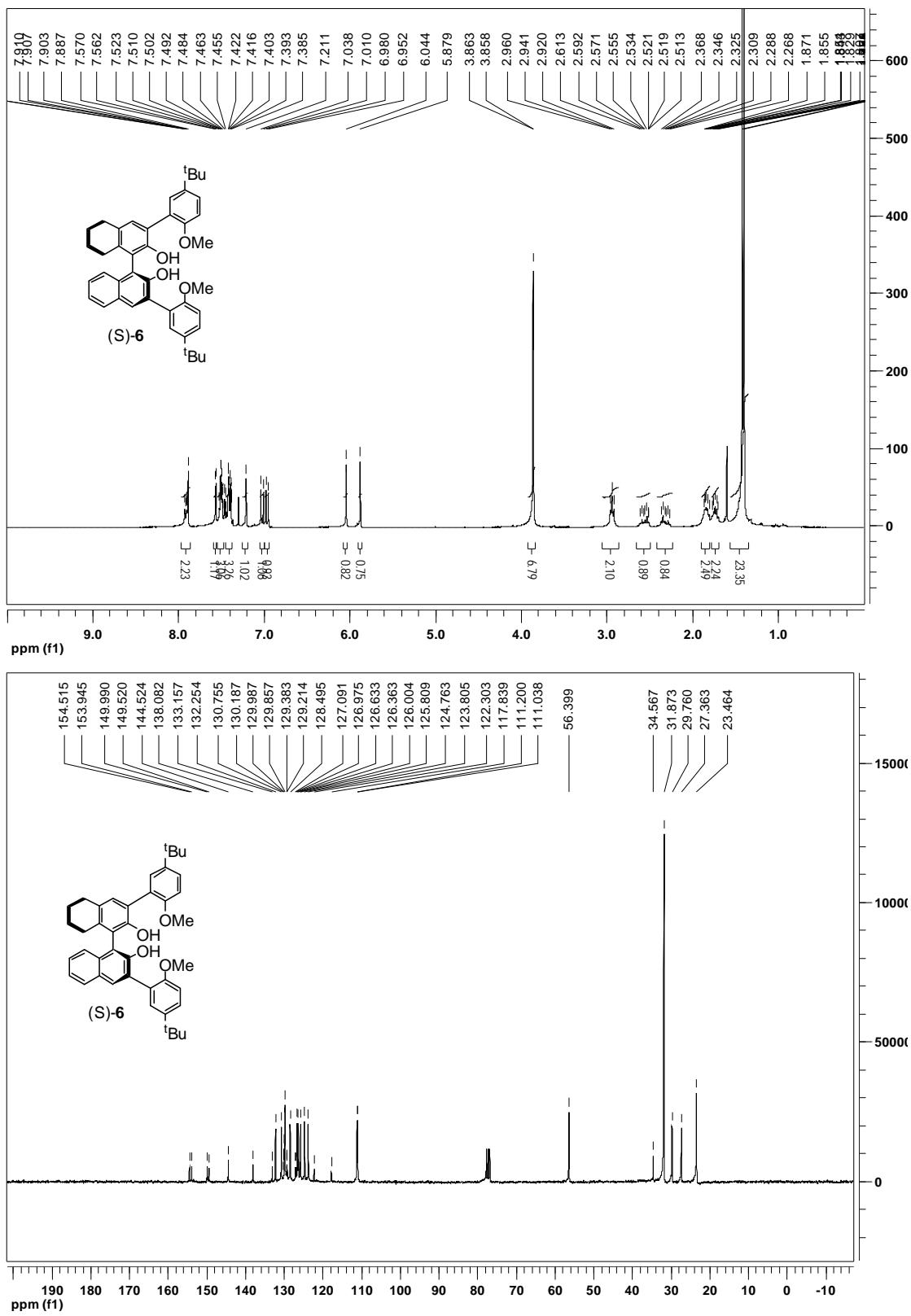
¹H and ¹³C NMR Spectra

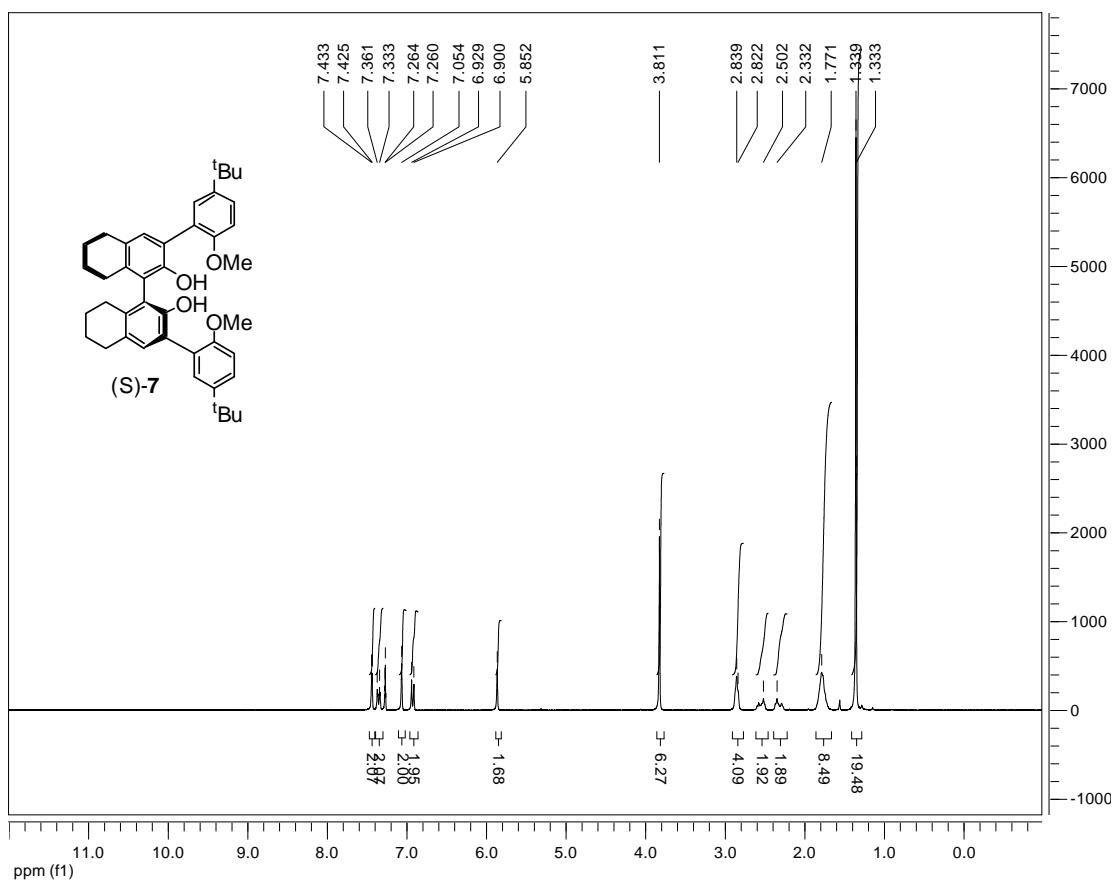


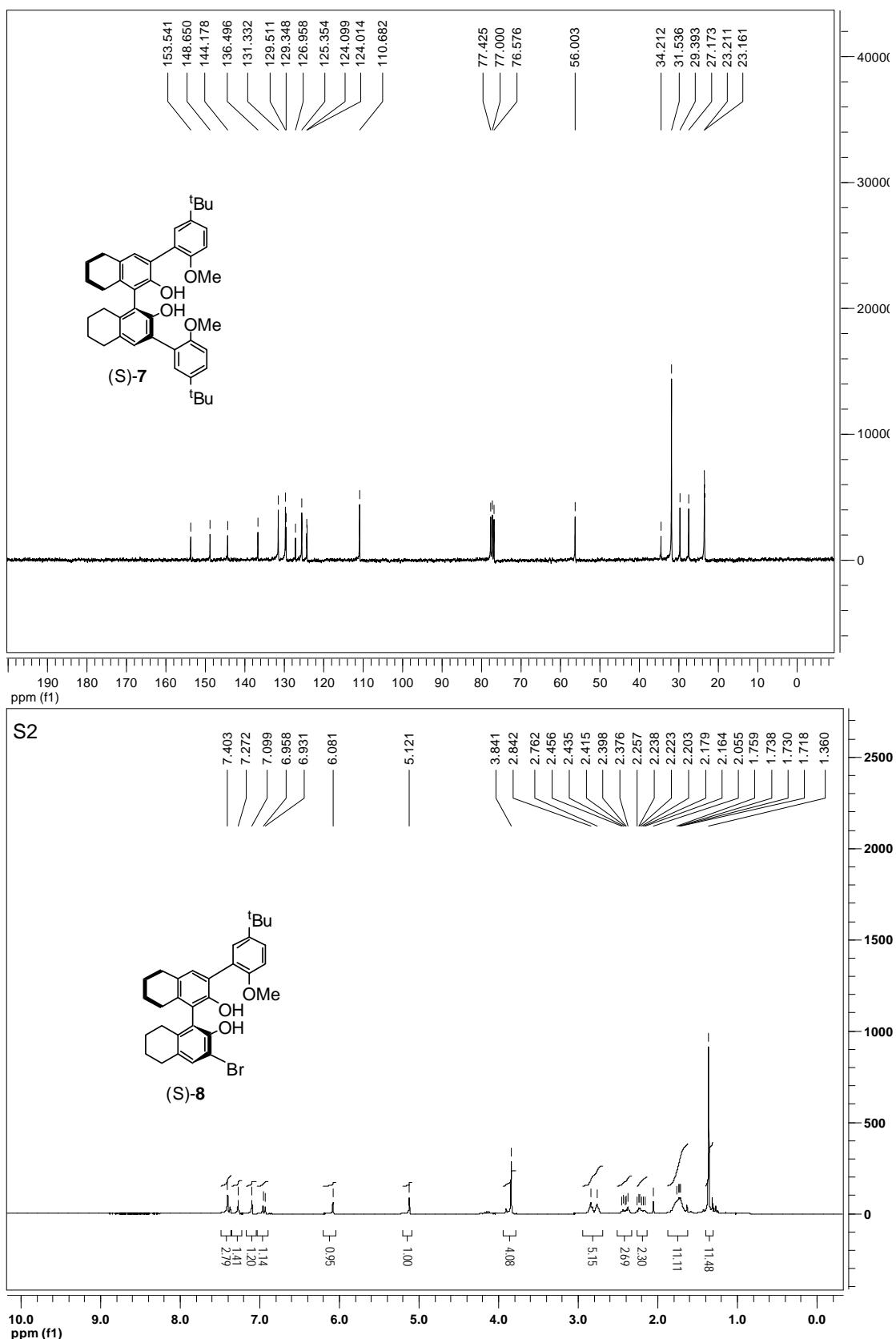




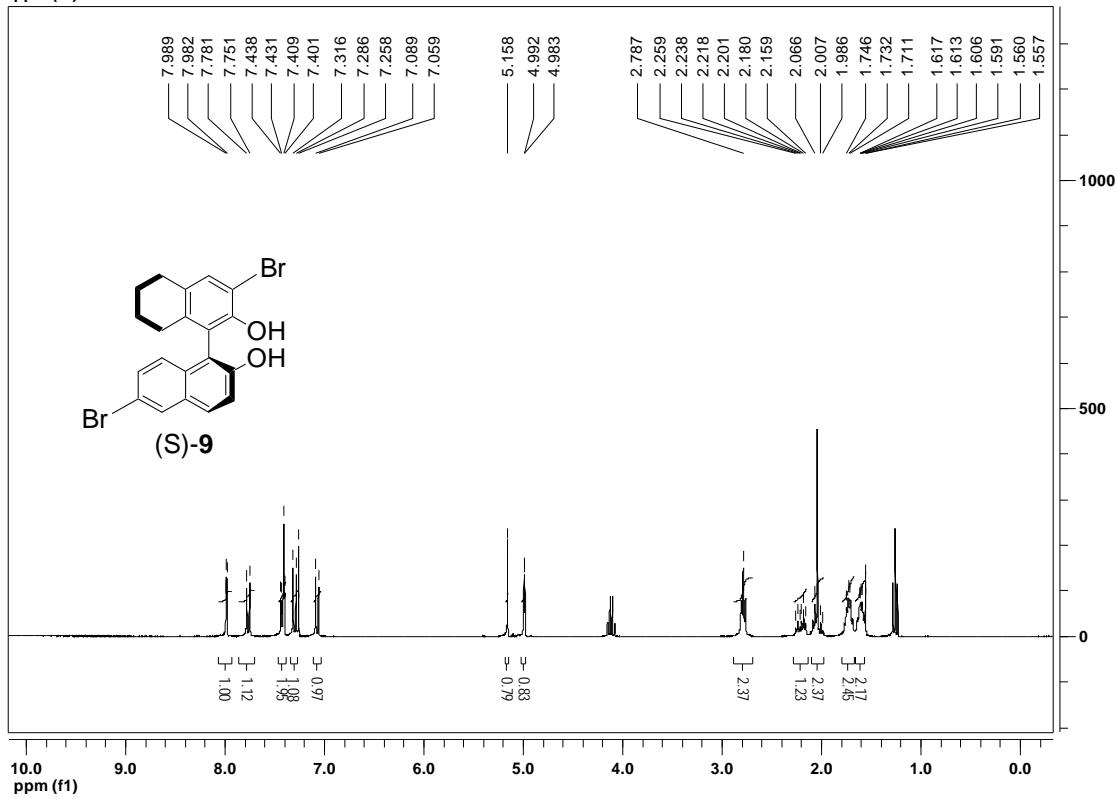
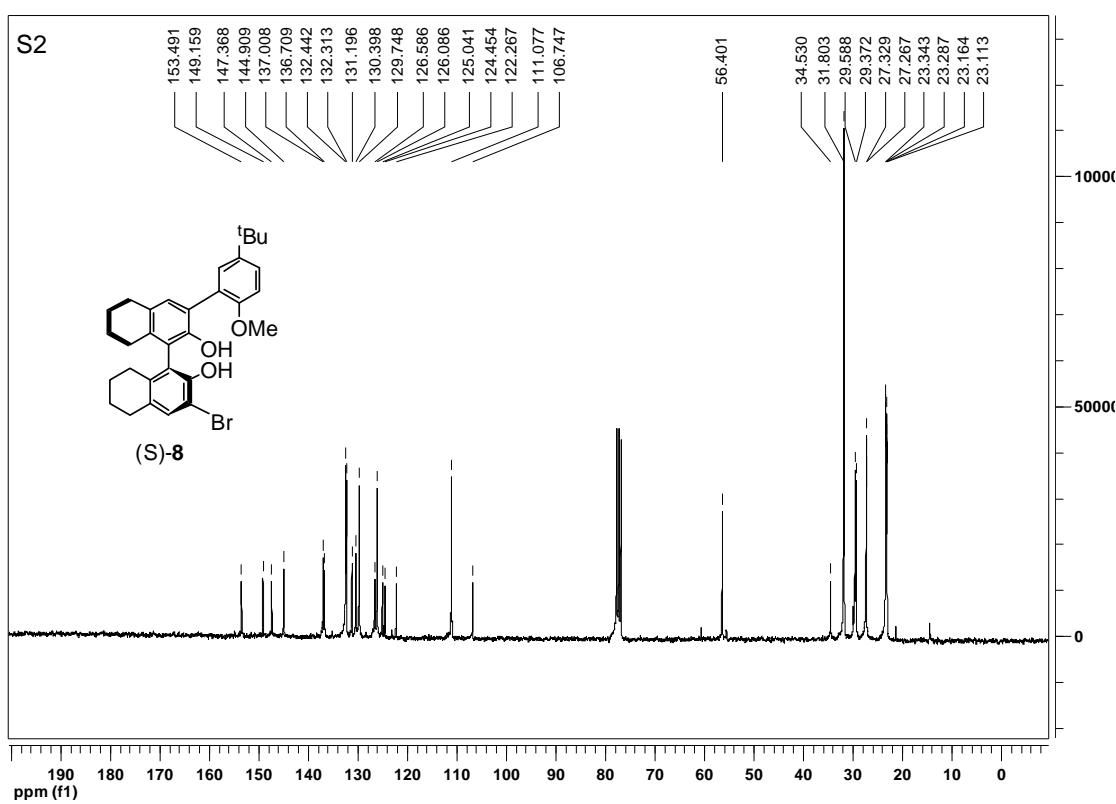


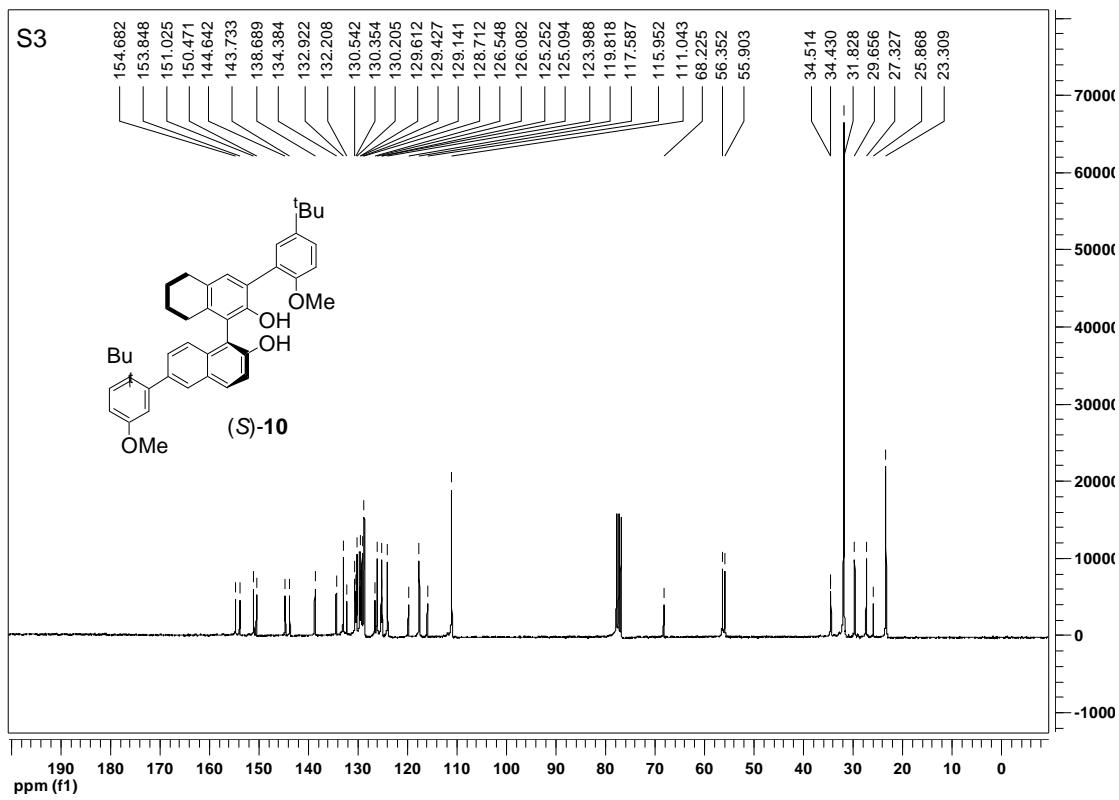
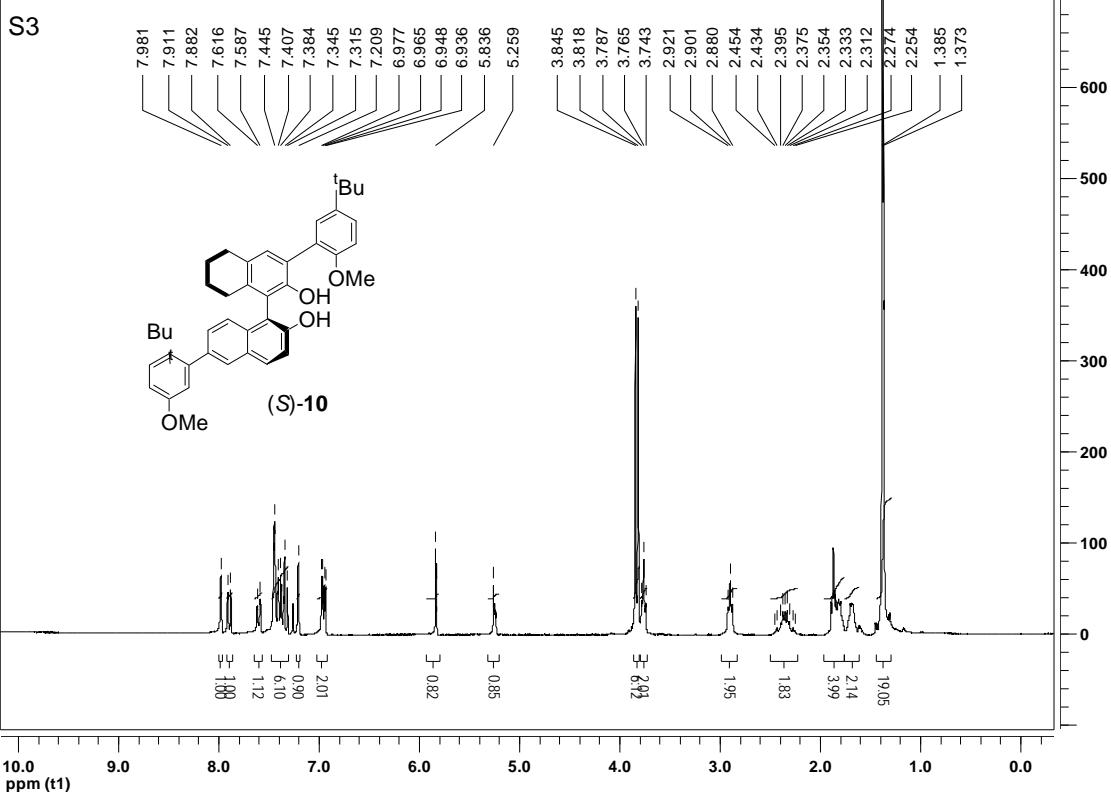


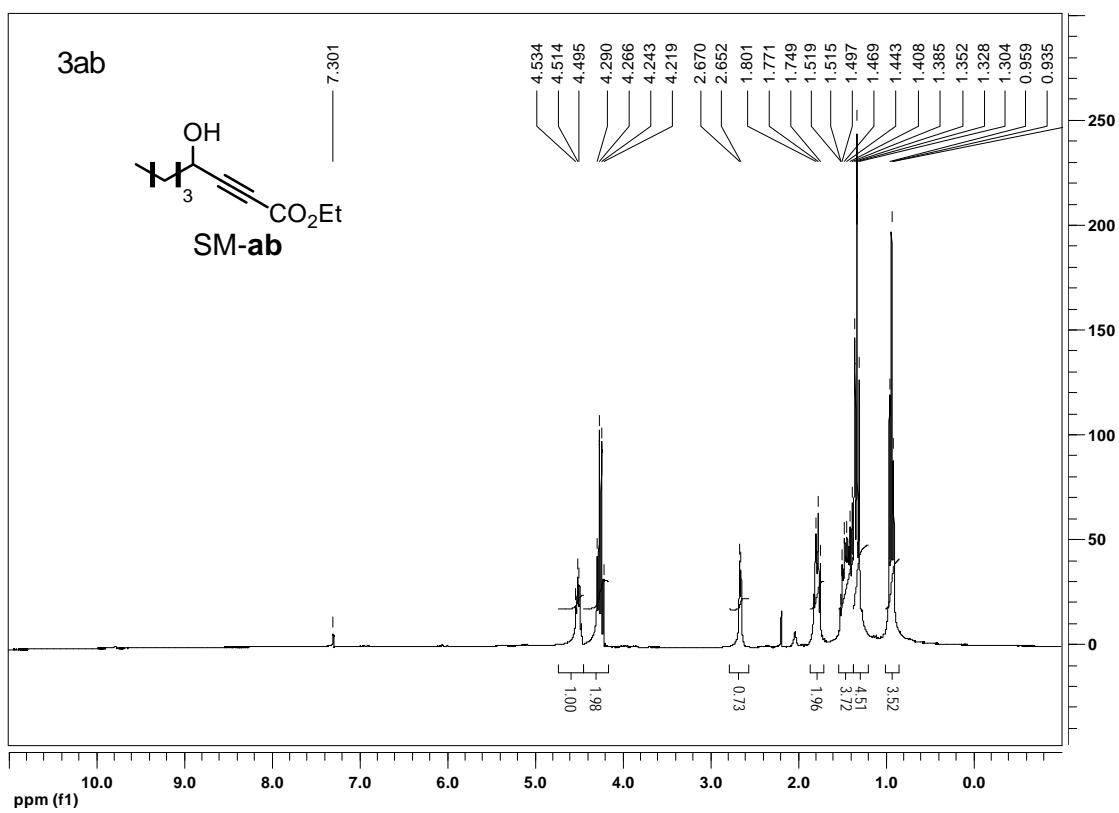
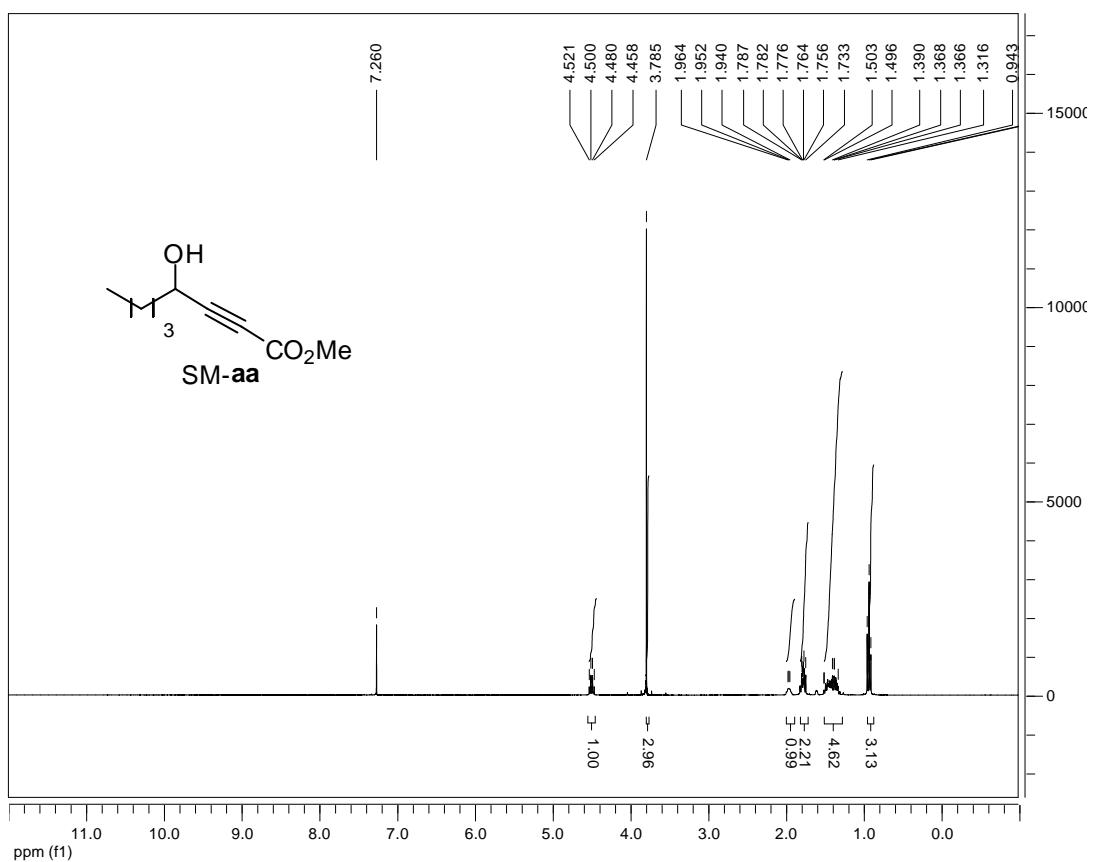


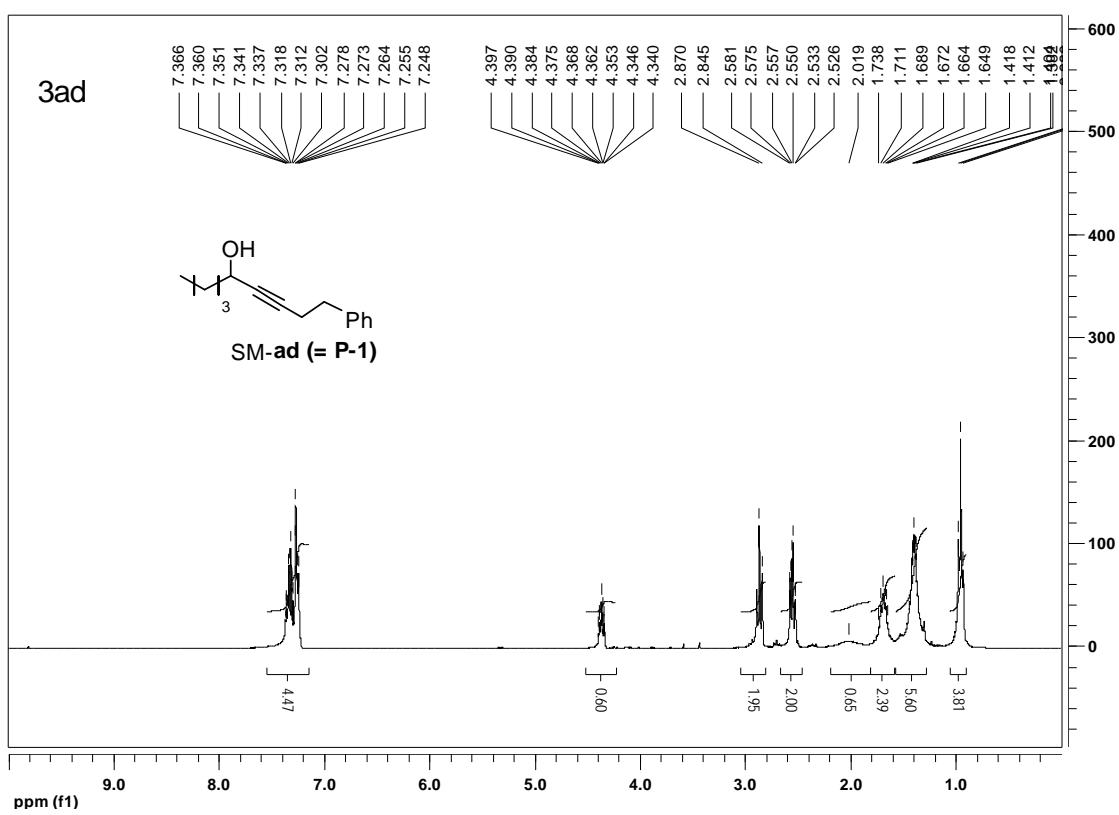
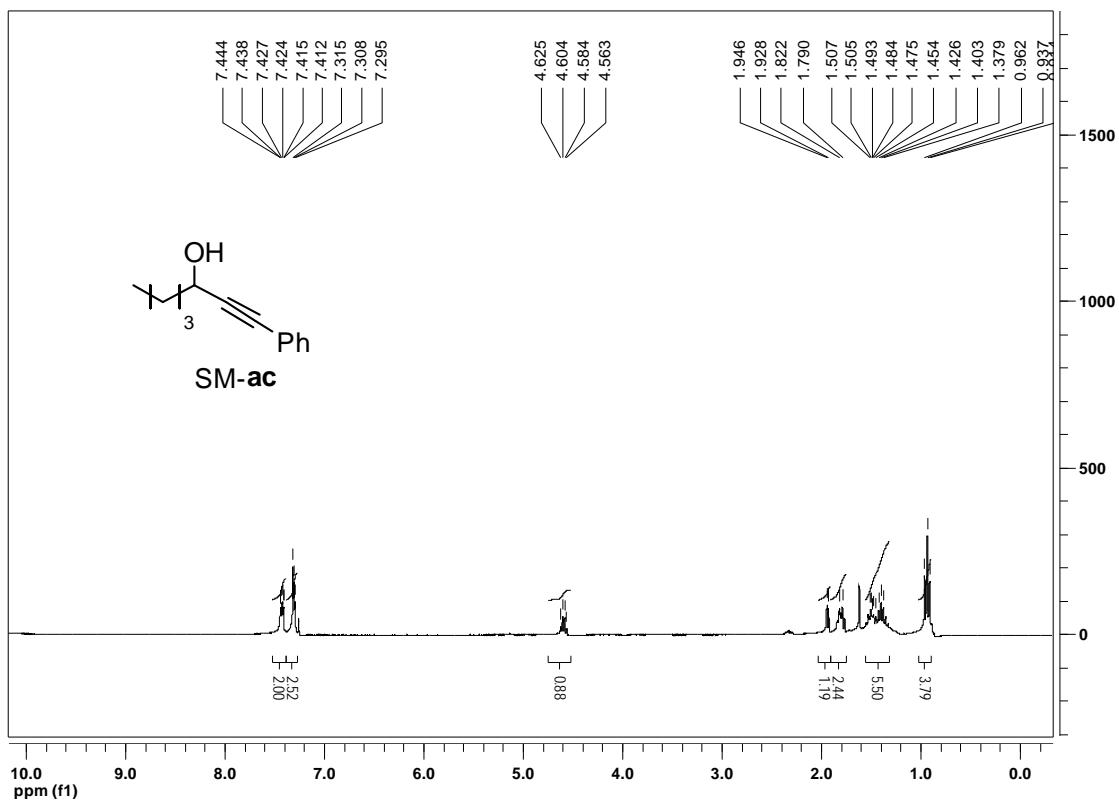


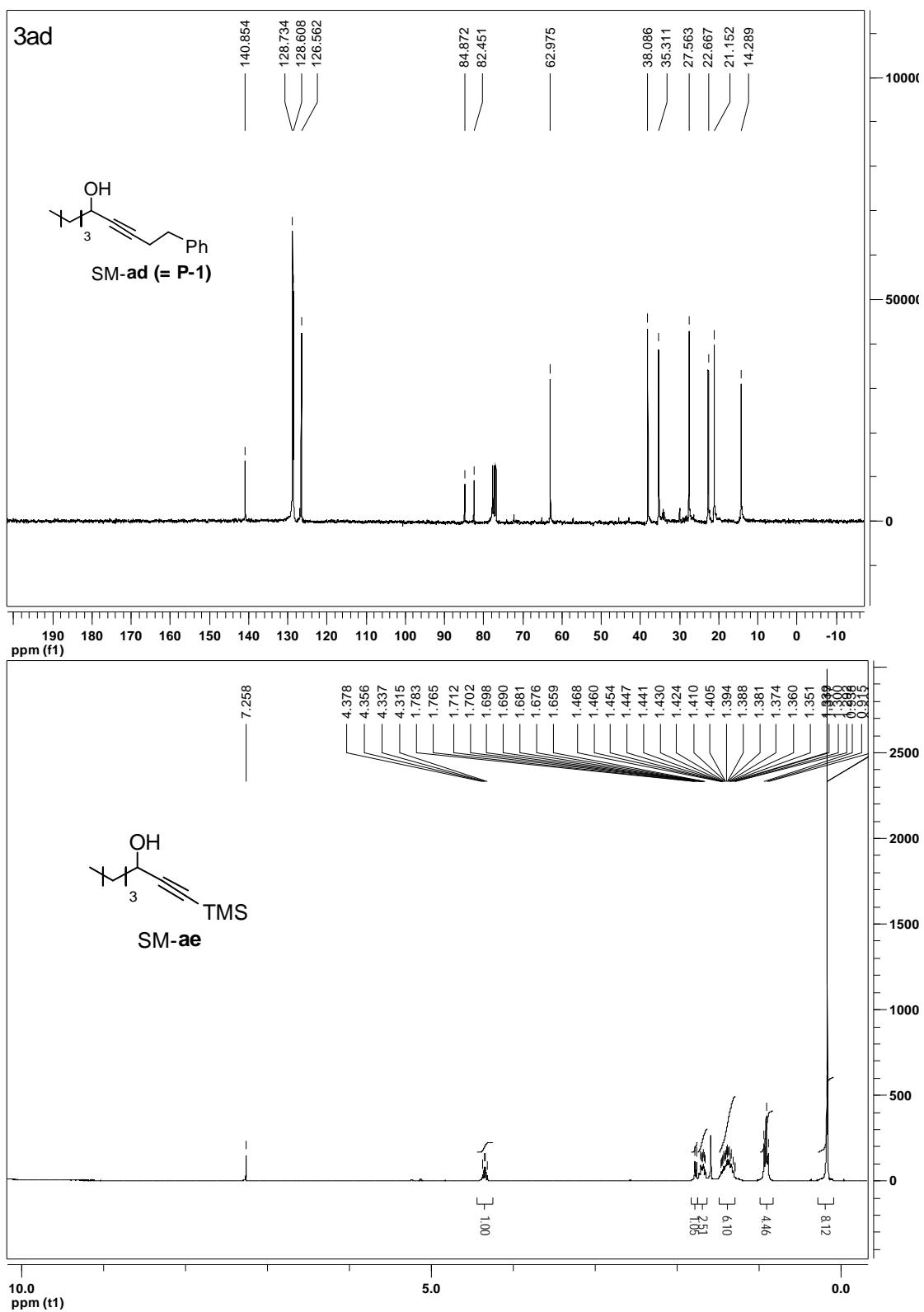
S2

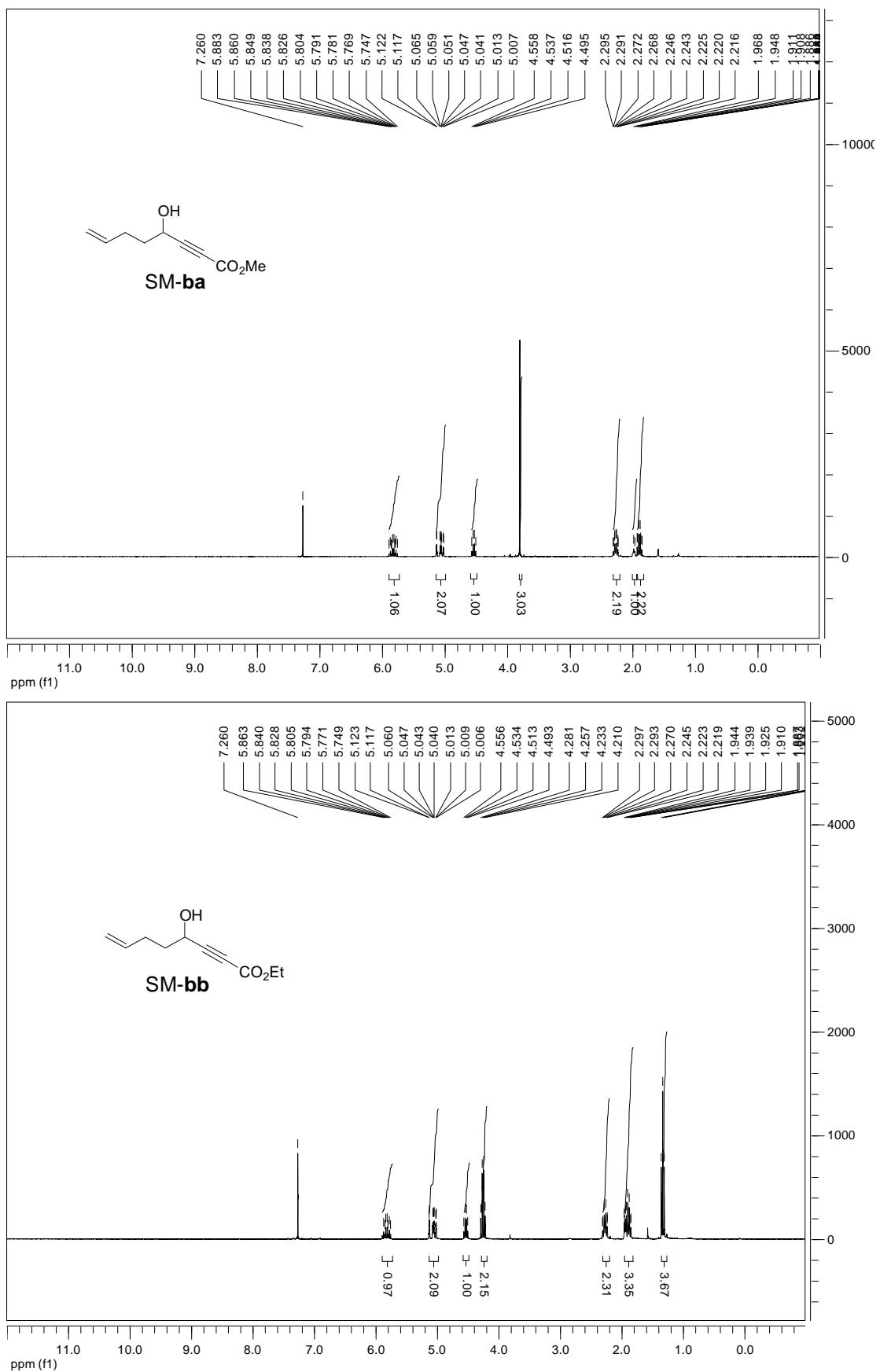


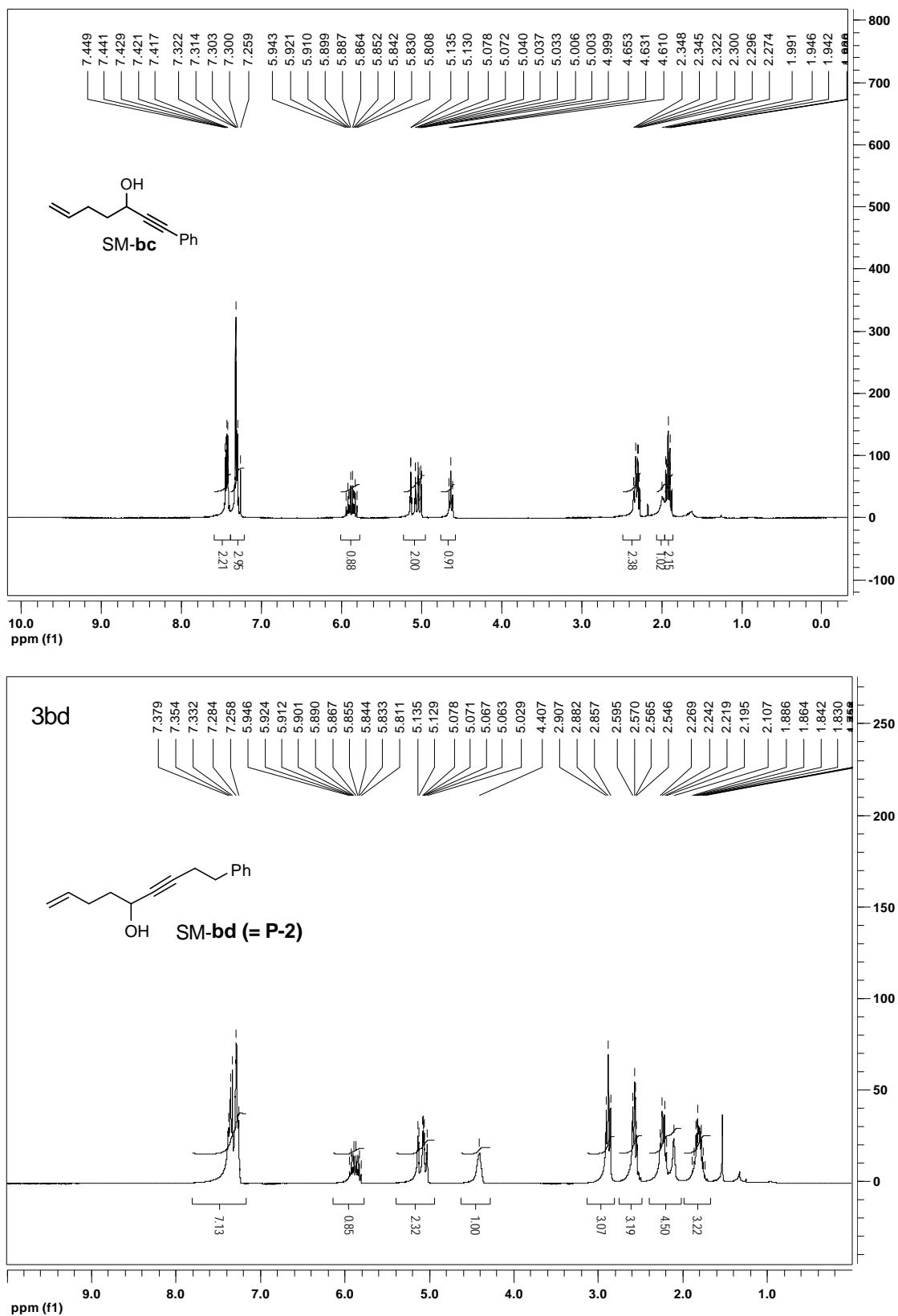


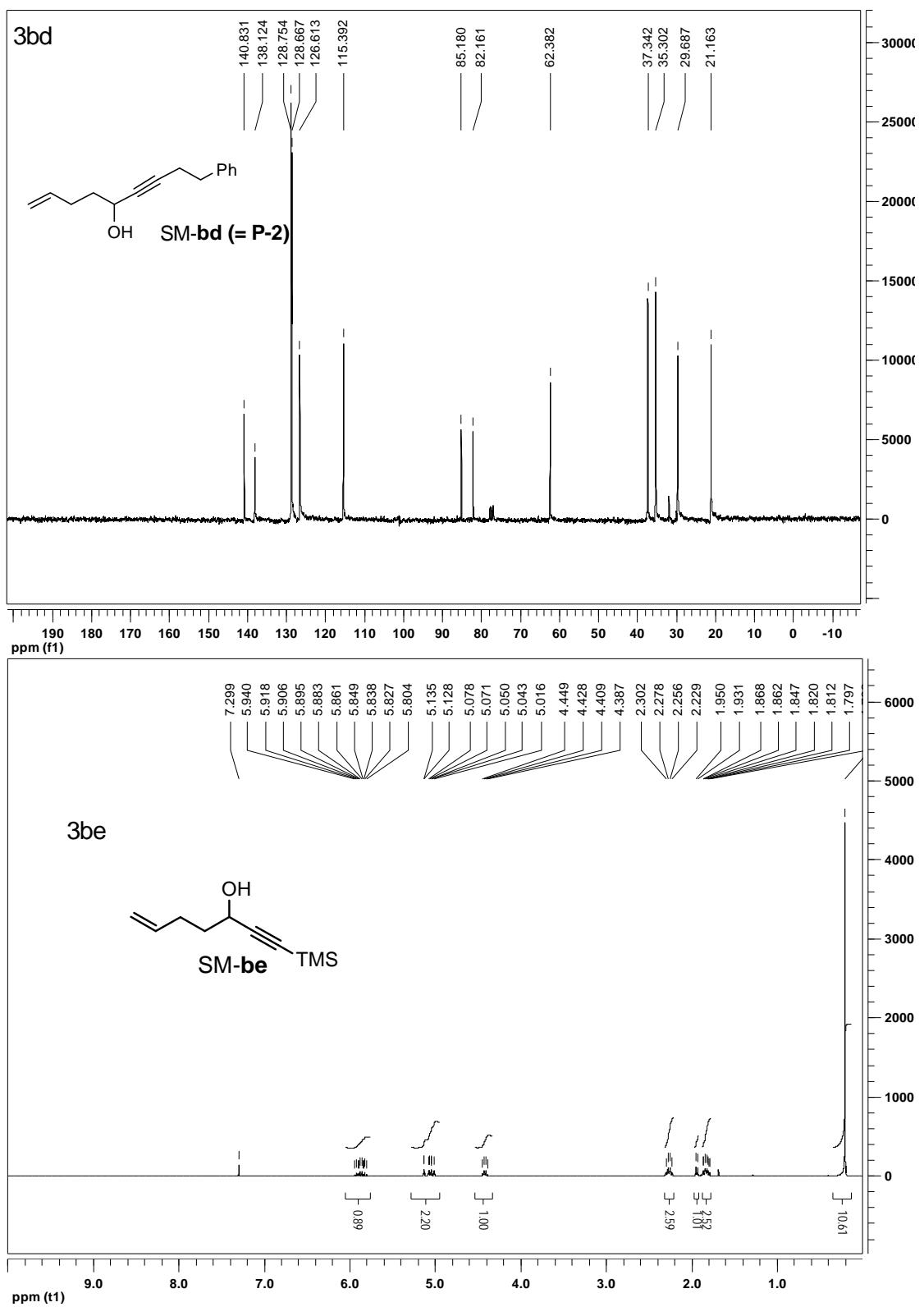


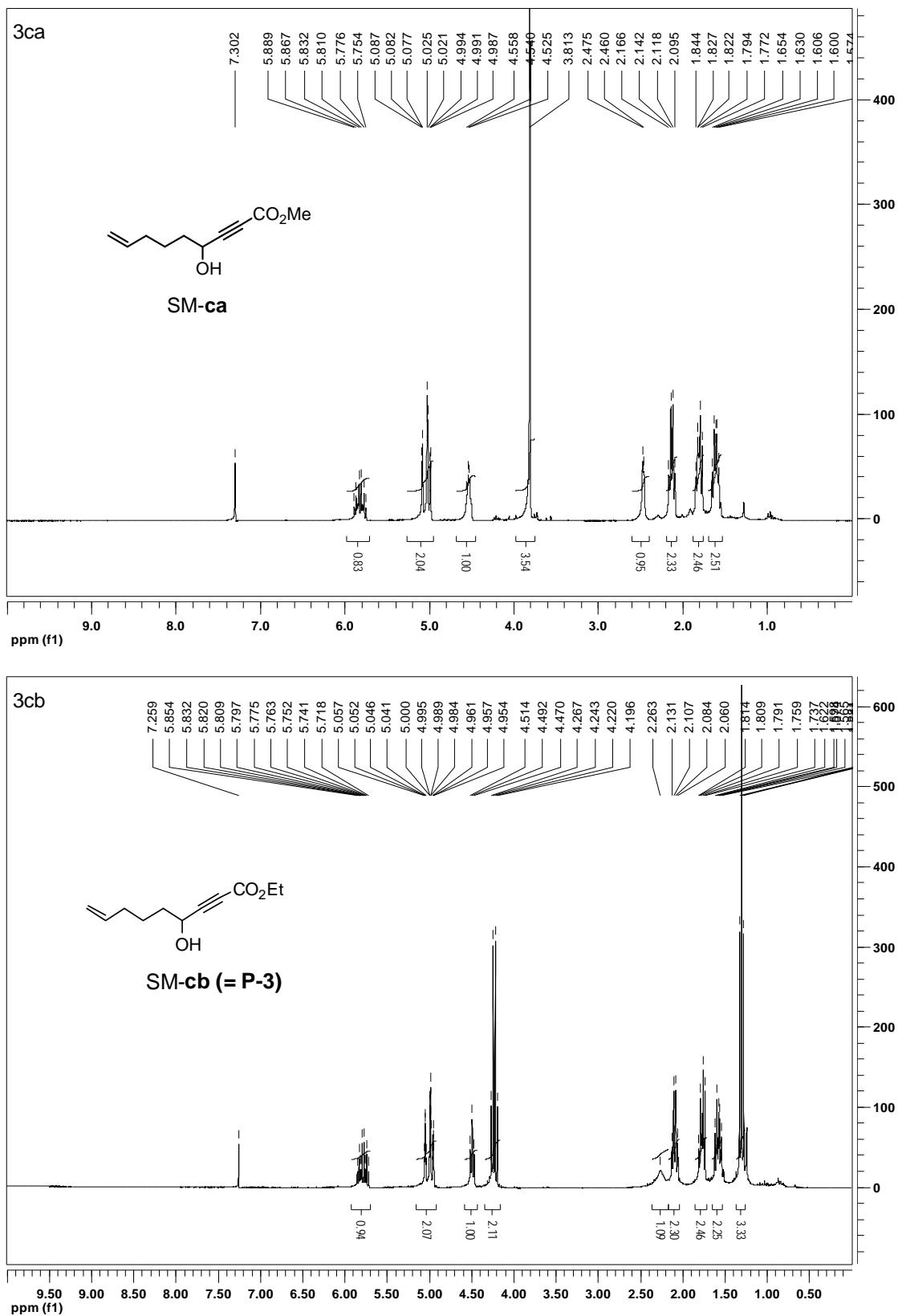


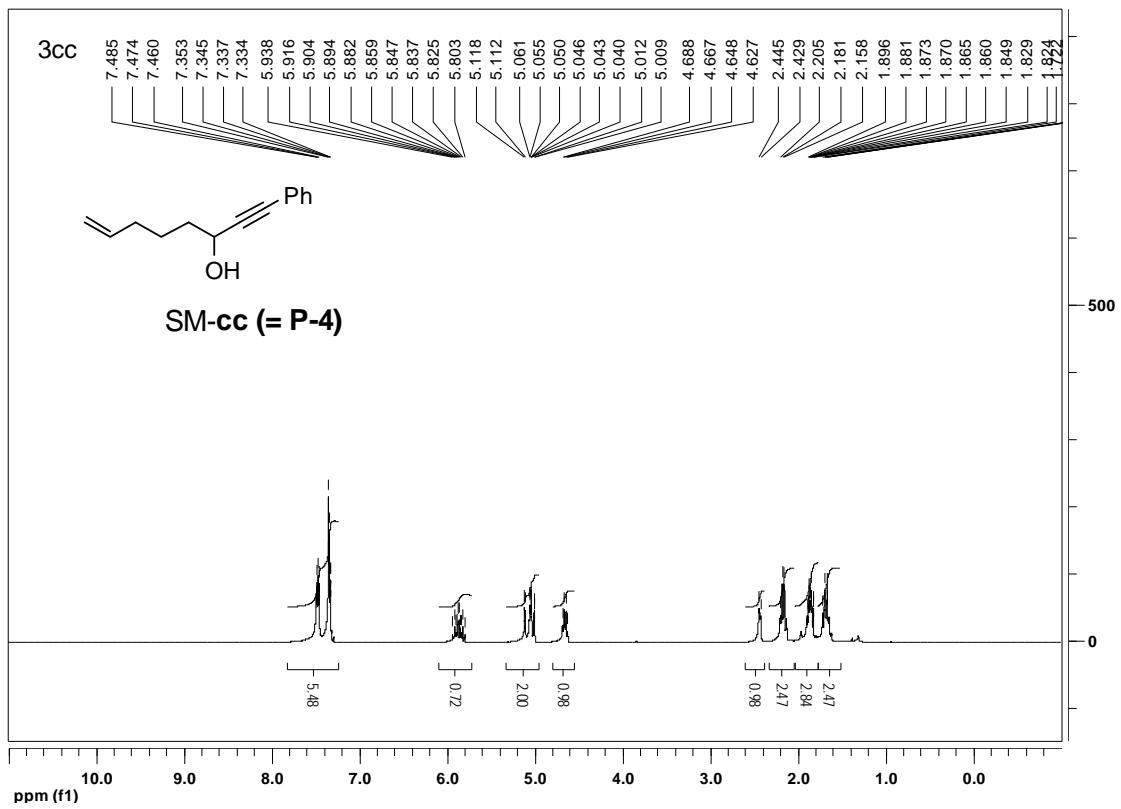
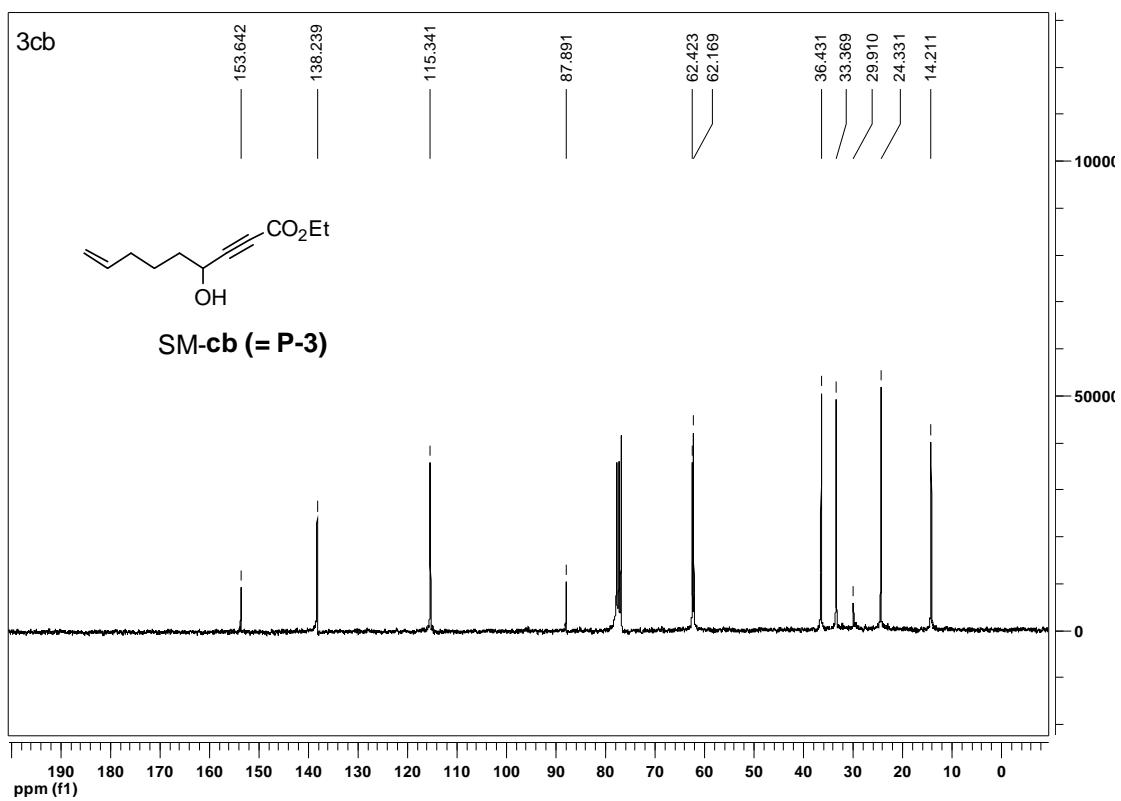


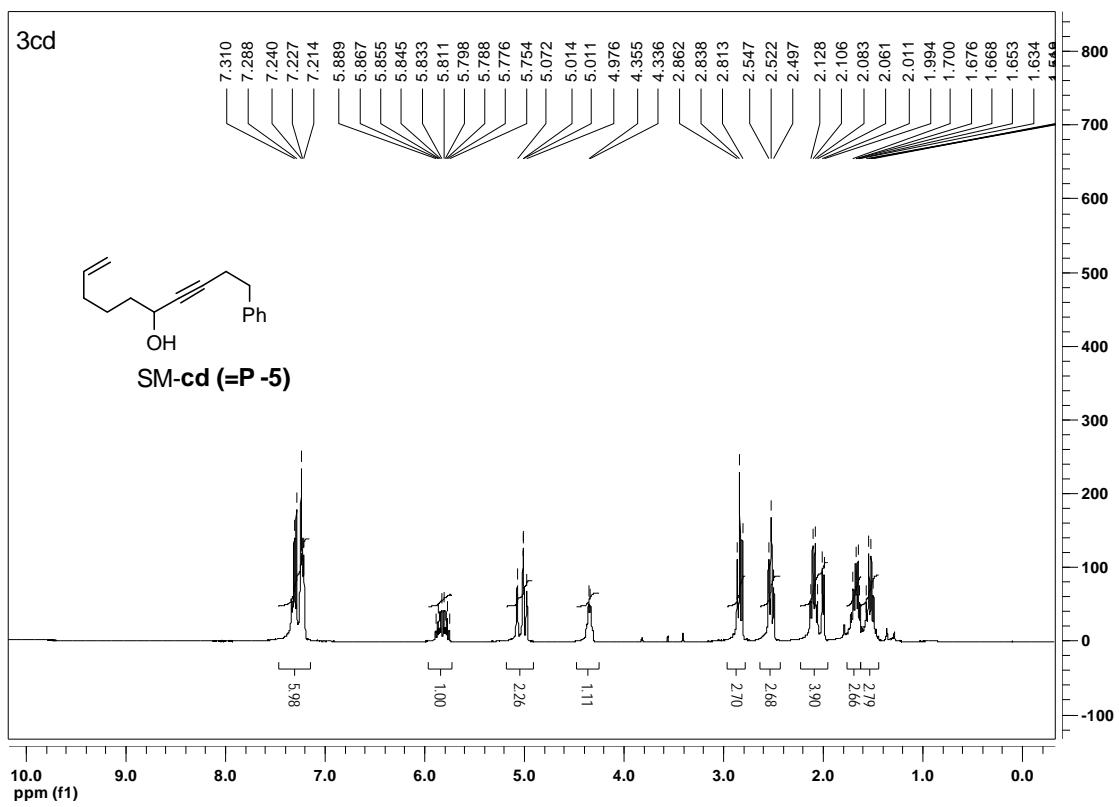
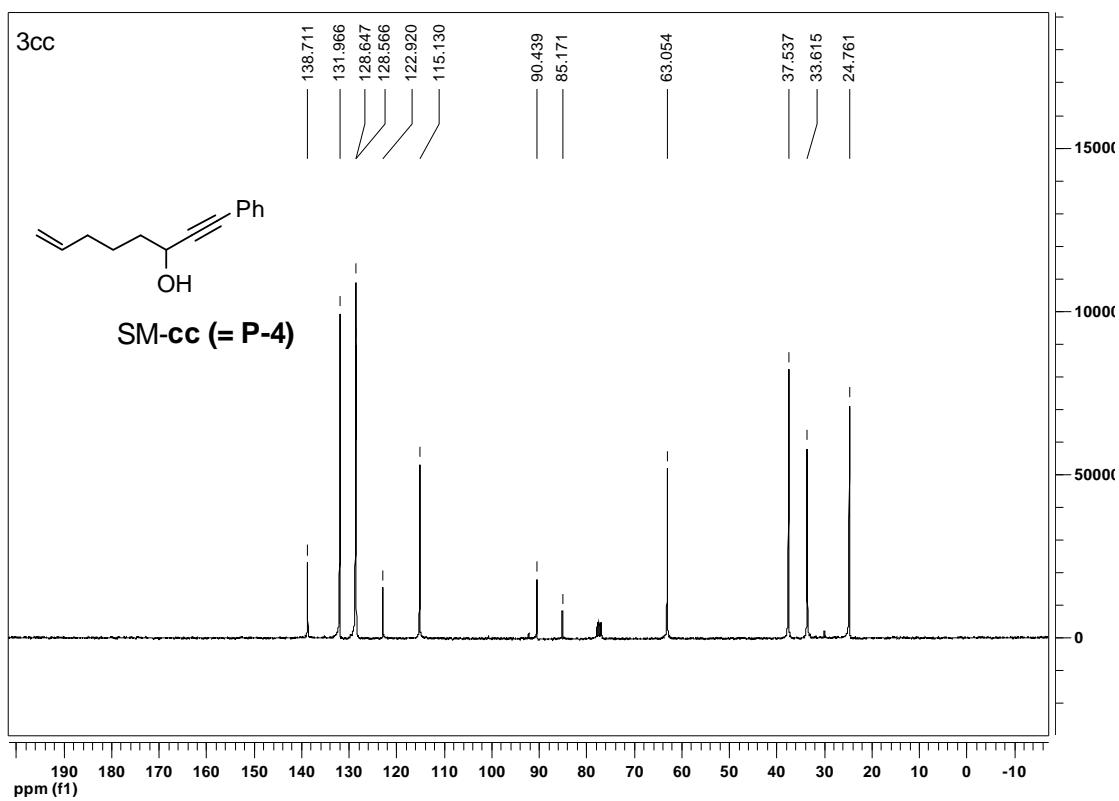


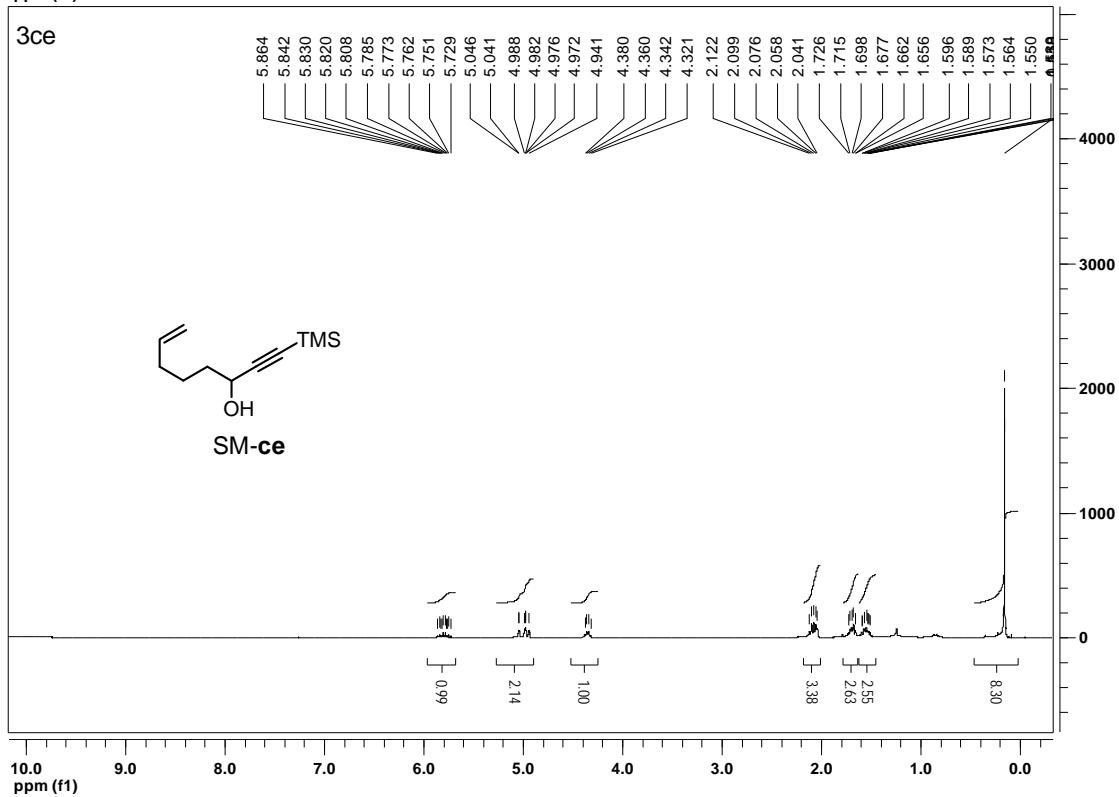
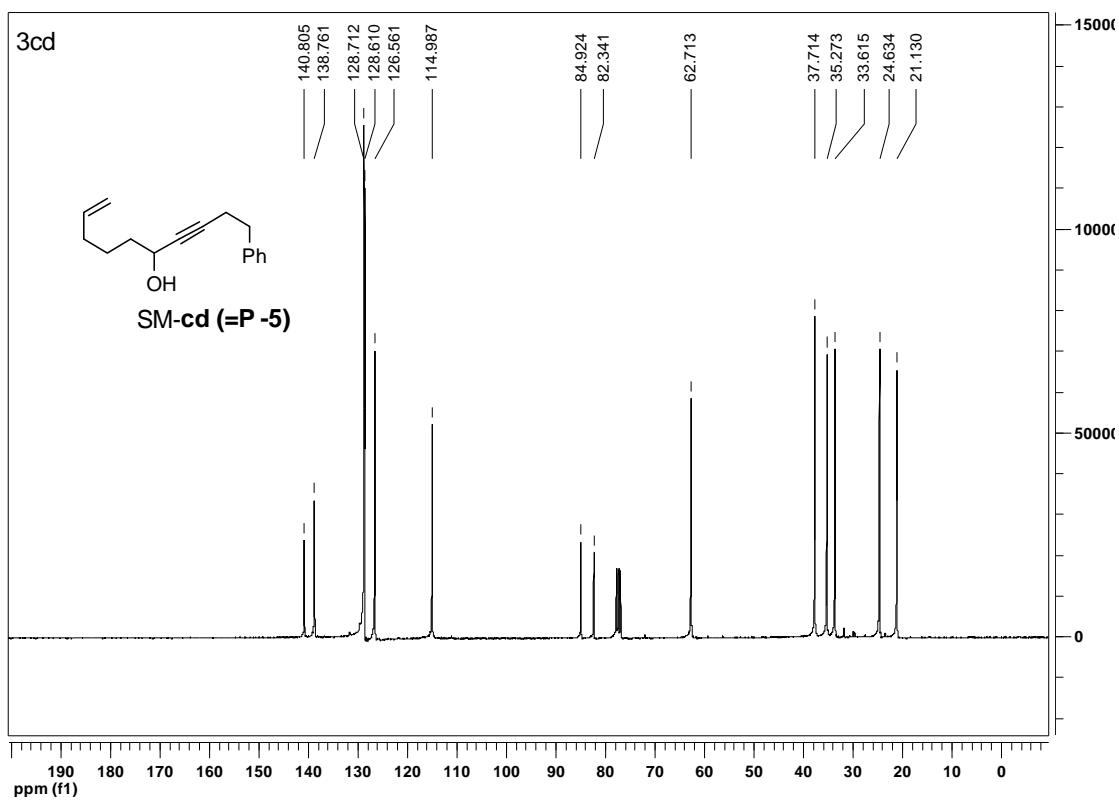


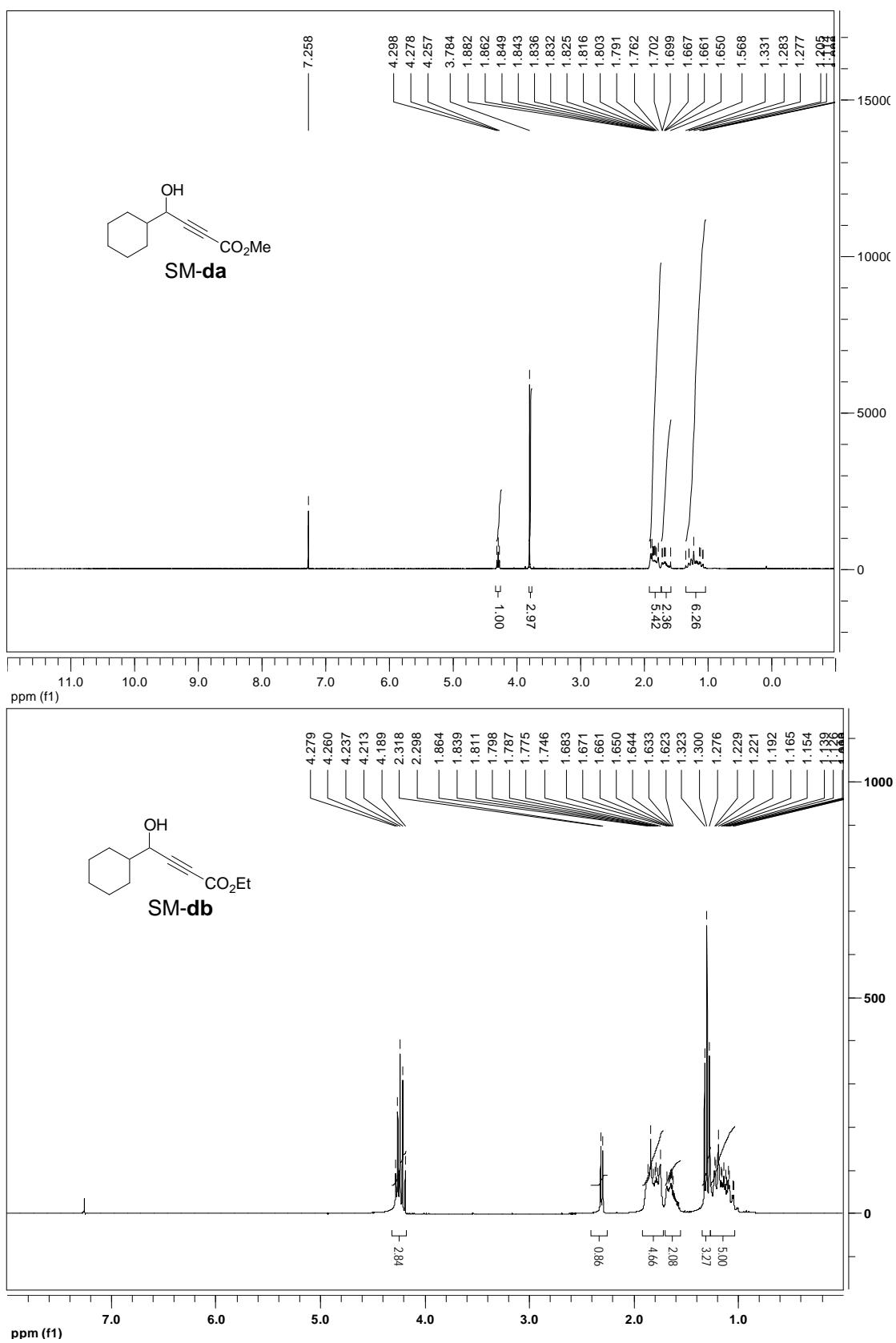


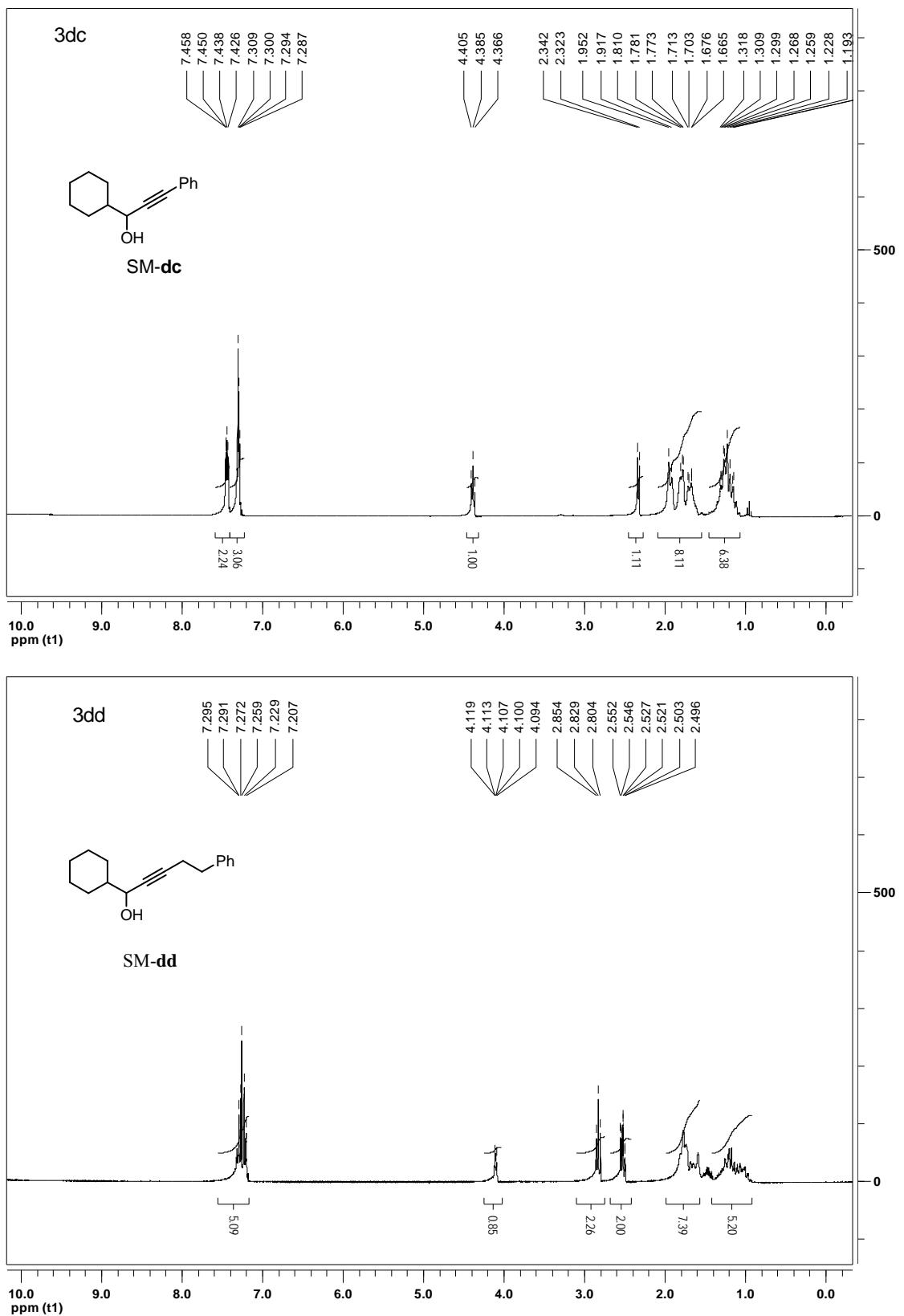


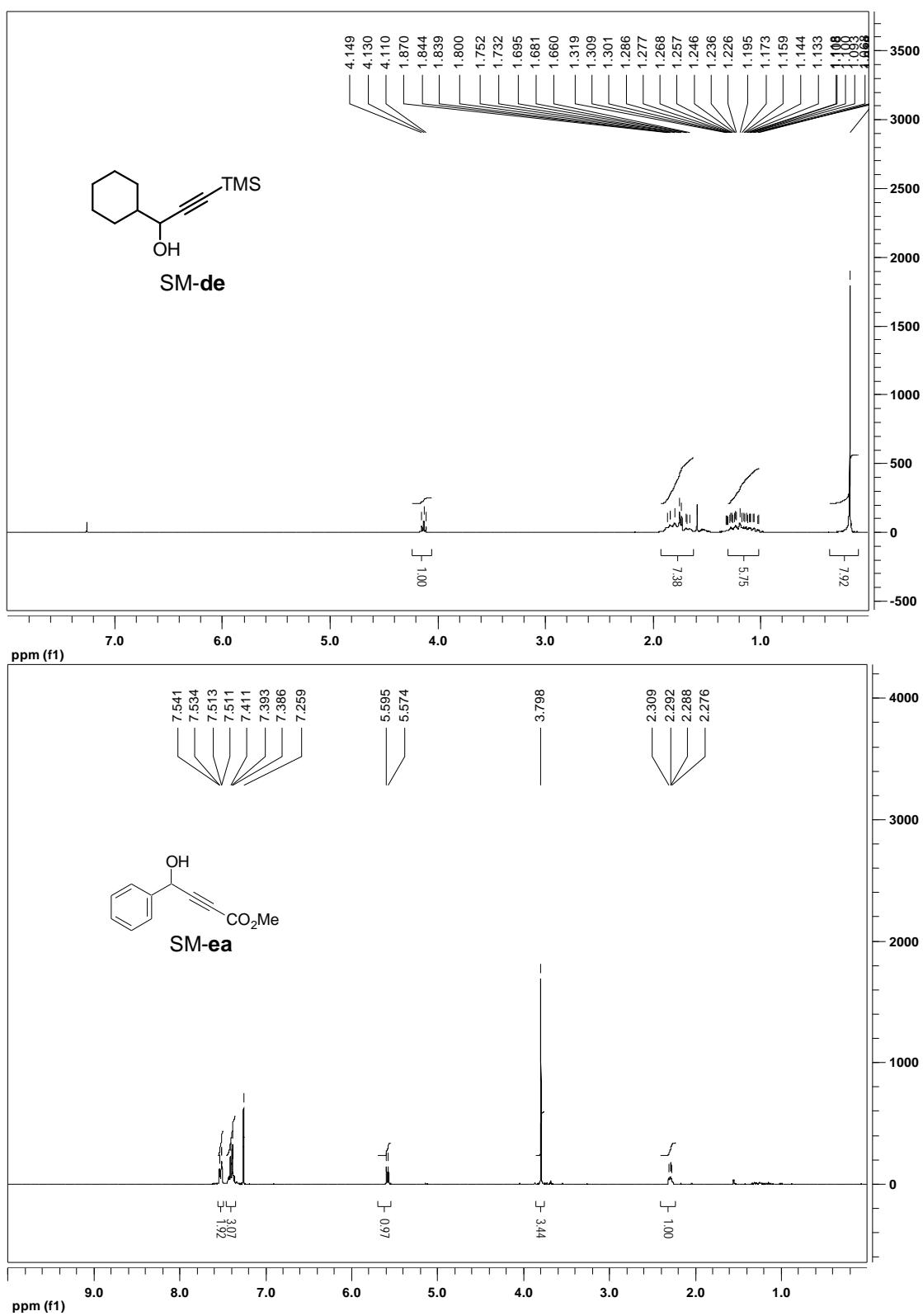


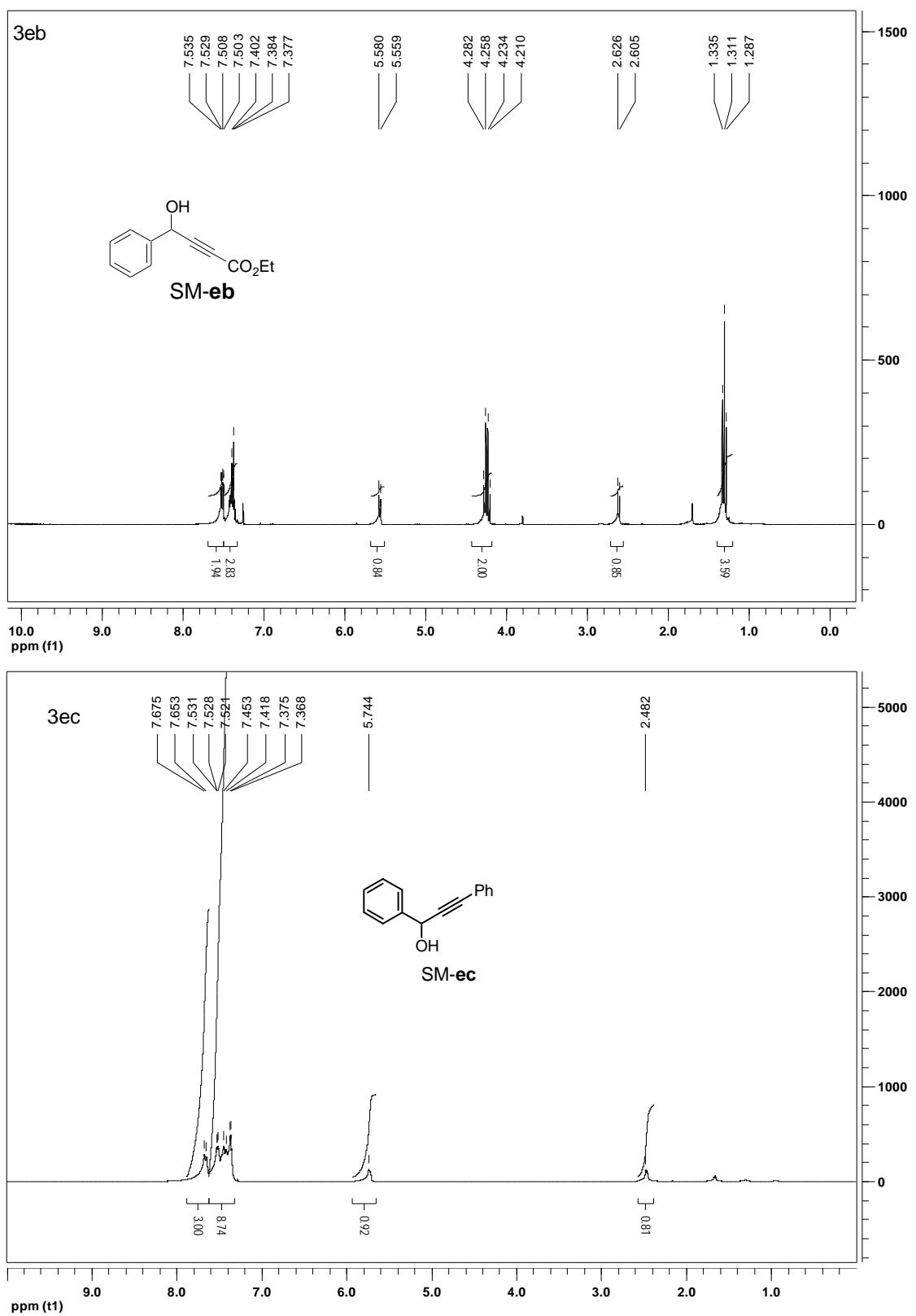


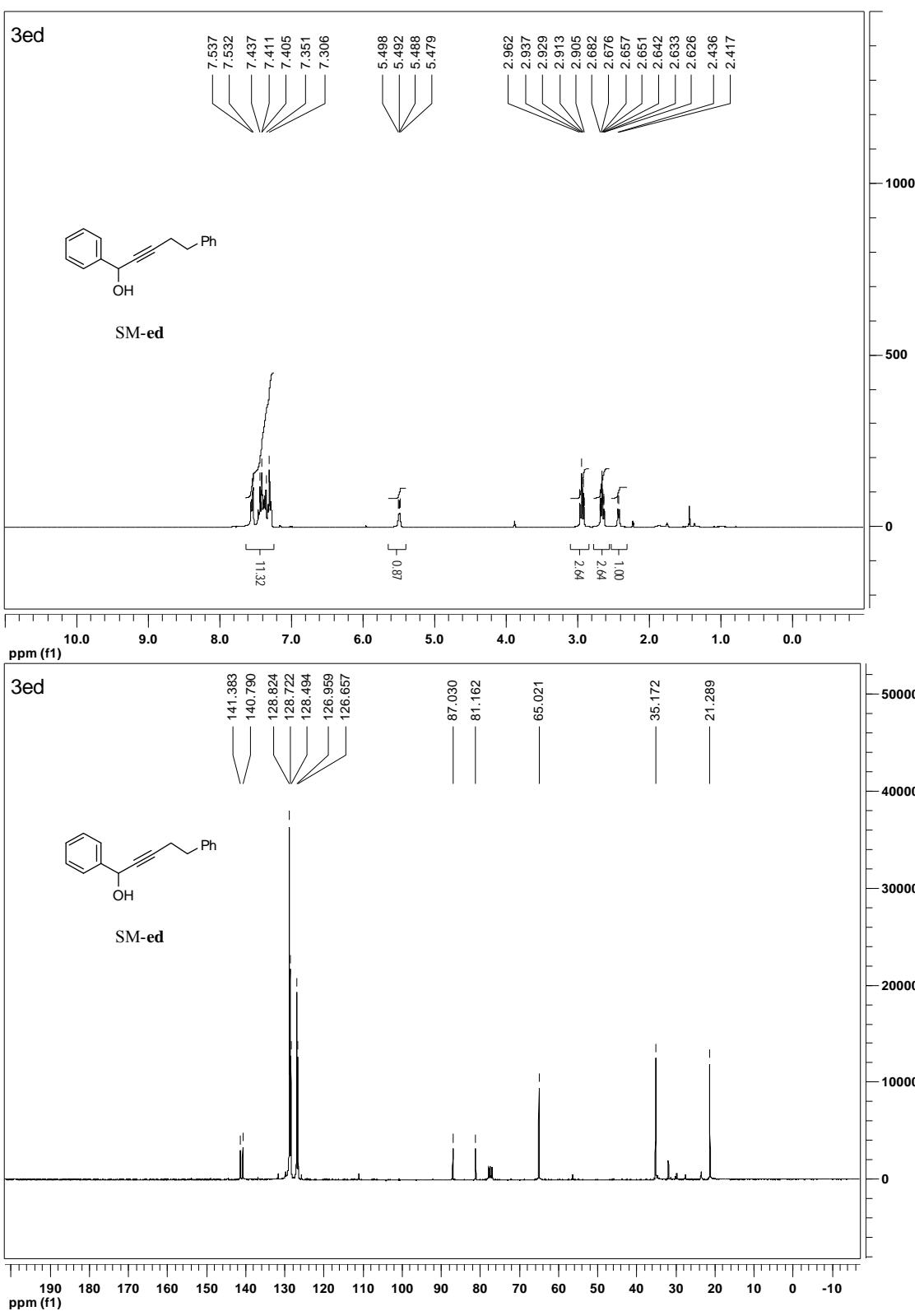


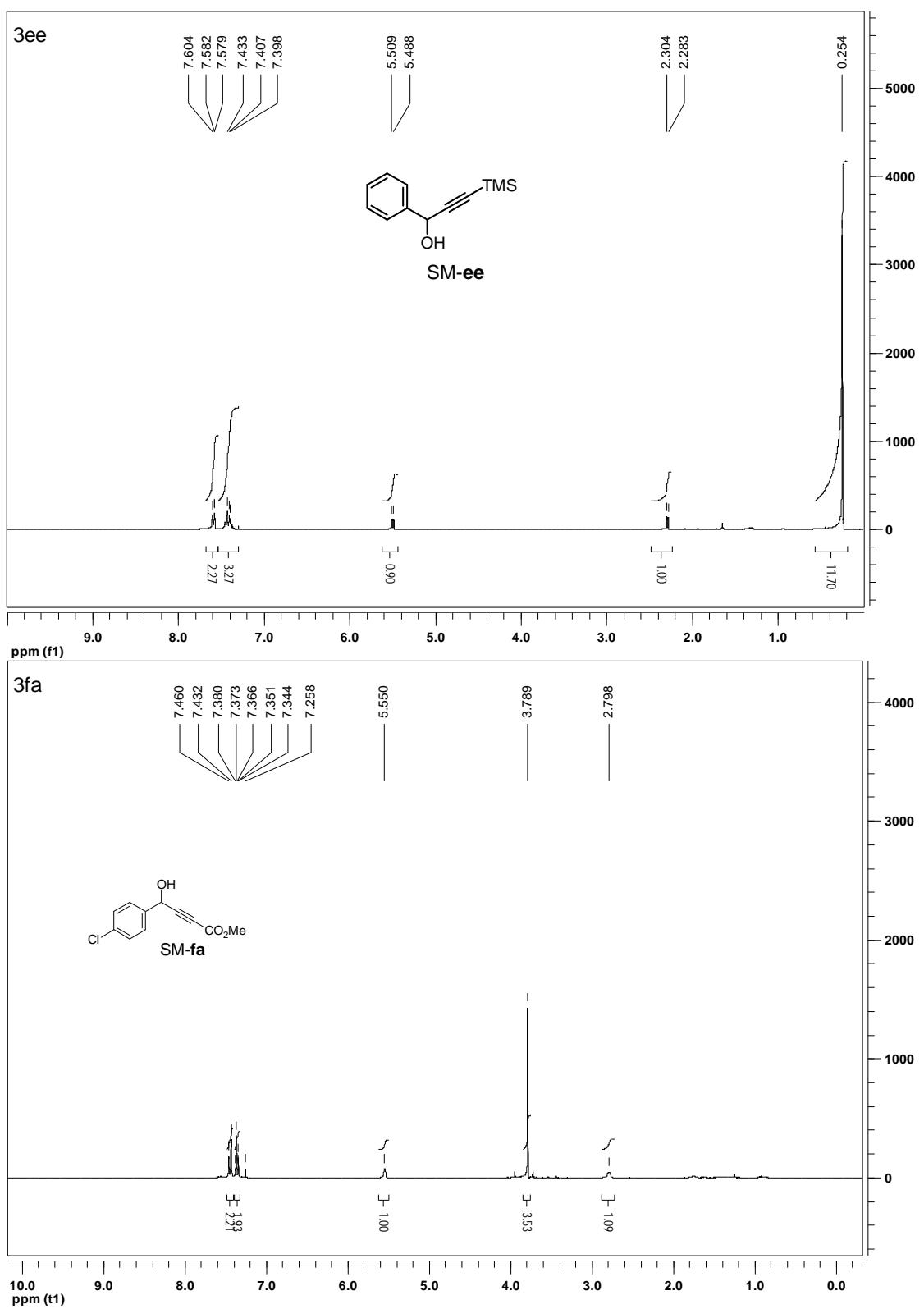


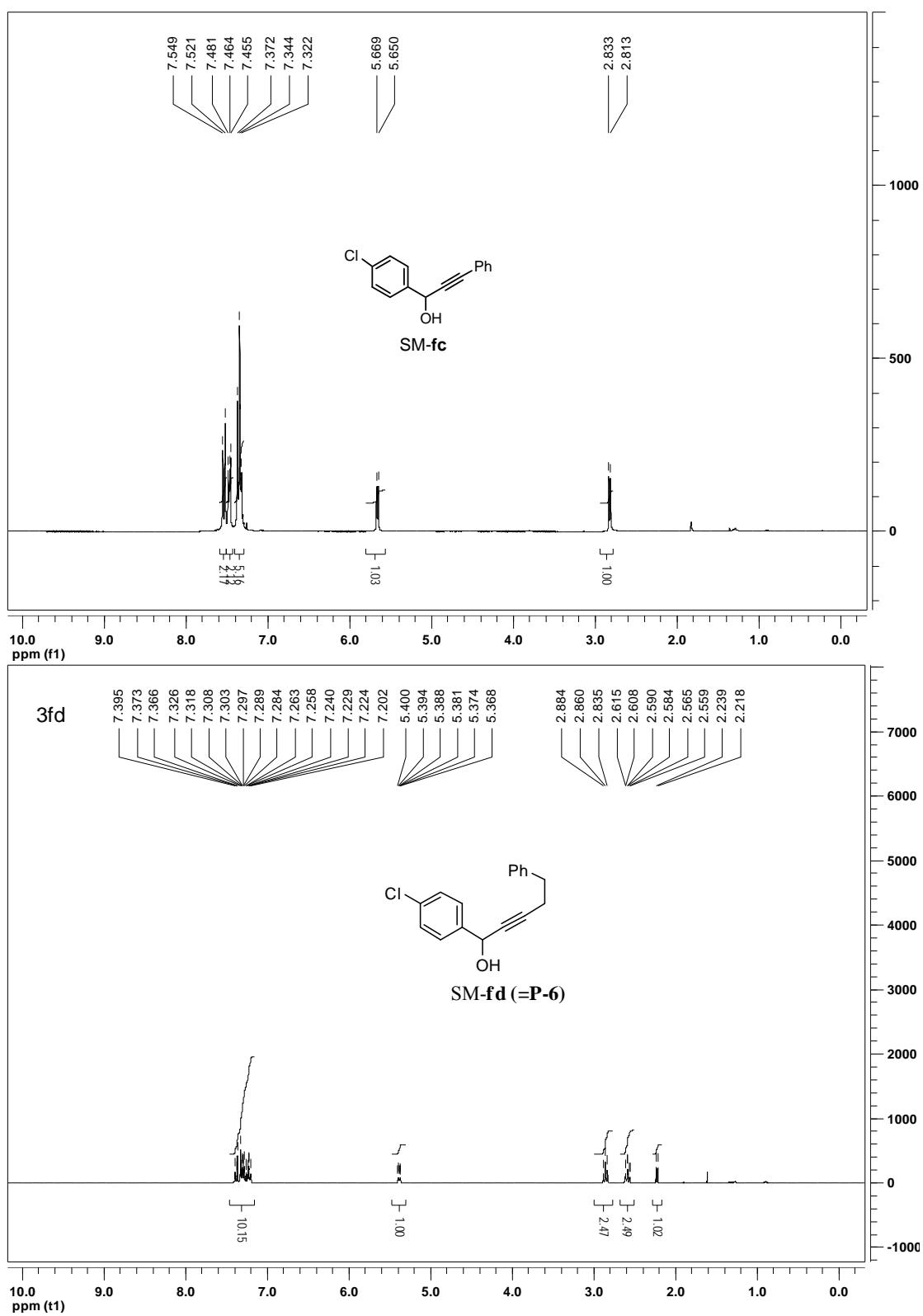


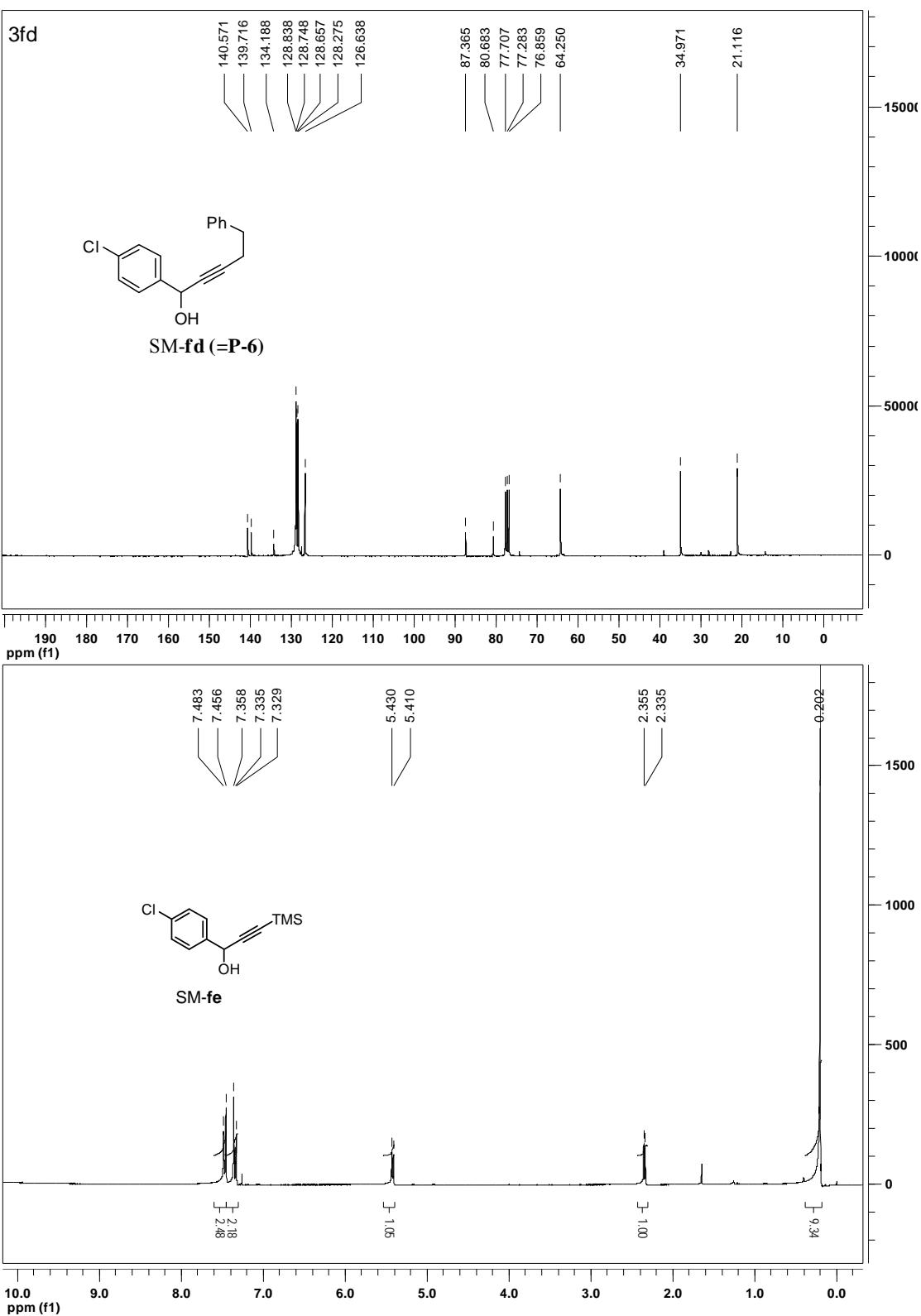


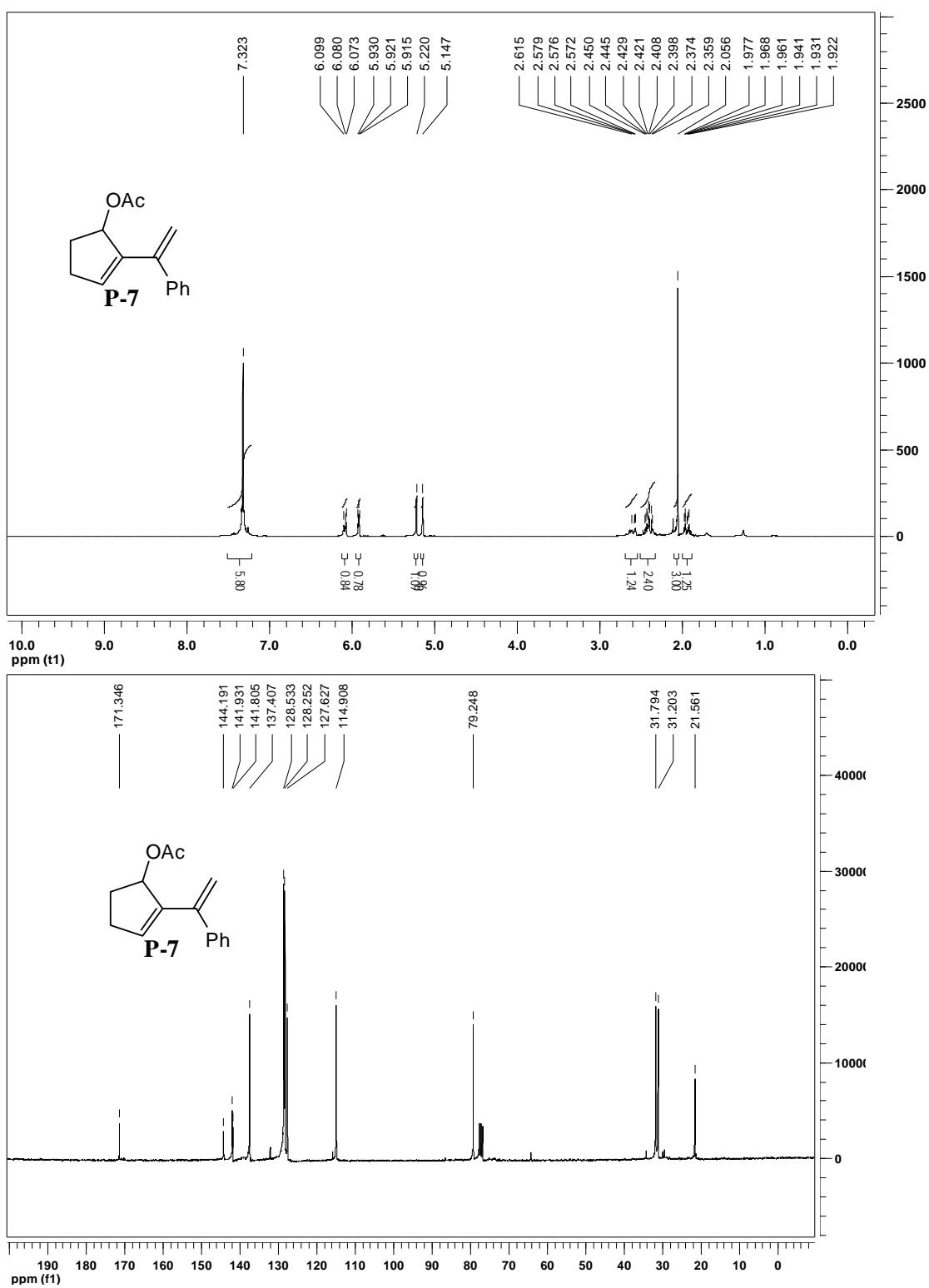


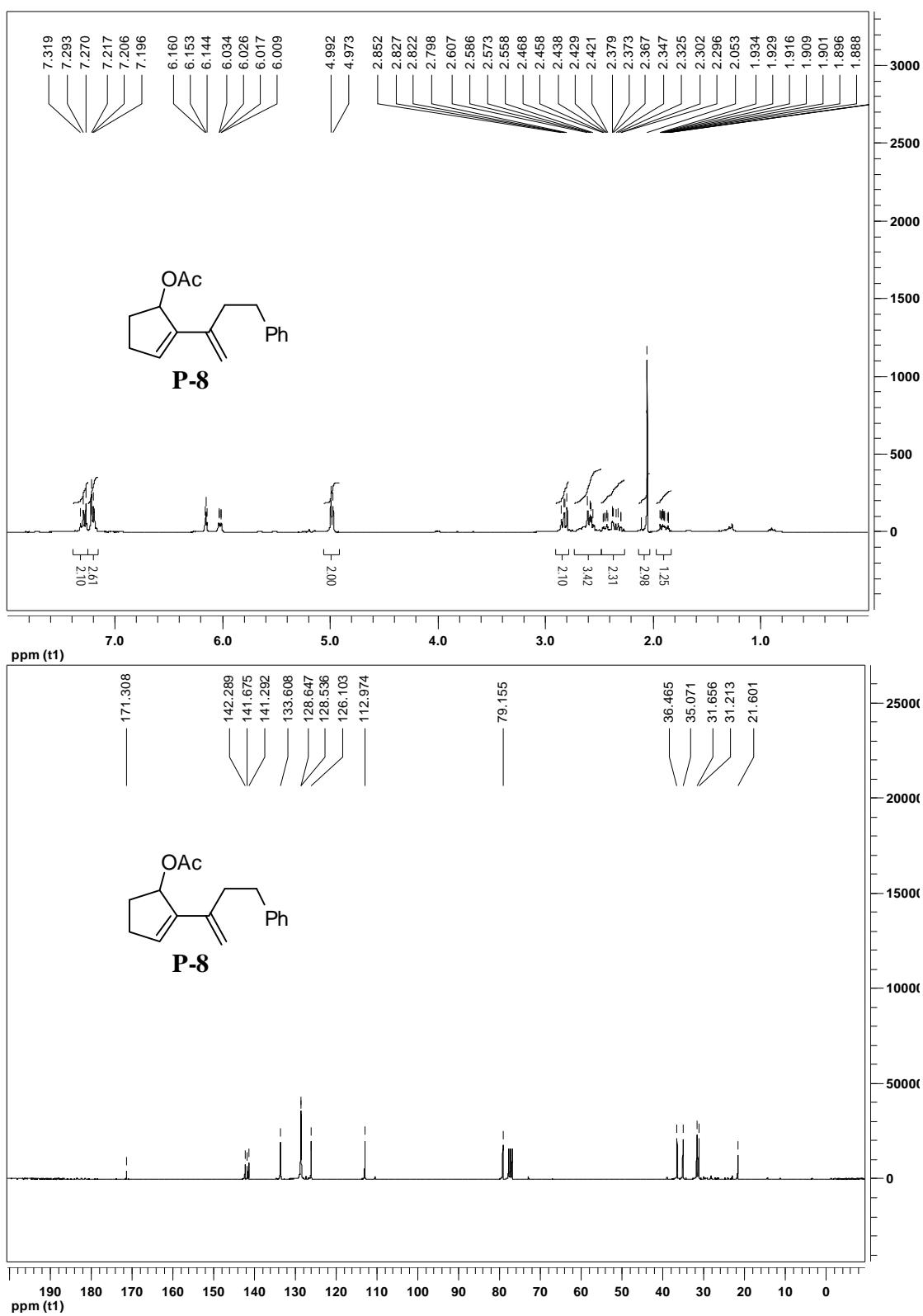












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