Supporting Information

Copper-Catalyzed y-Selective Allyl–Alkyl Coupling between Allylic Phosphates and Alkylboranes

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Instrumentation and Chemical

NMR spectra were recorded on a Varian Gemini 2000 spectrometer, operating at 300 MHz for ¹H NMR and 75.4 MHz for ¹³C NMR, and a JEOL ECA-600, operating at 600 MHz for ¹H NMR, 150.9 MHz for ¹³C NMR and 192.6 MHz for ¹¹B NMR. Chemical shift values for ¹H and ¹³C are referenced to Me₄Si and the residual solvent resonances, respectively. Chemical shift values for ¹¹B is referenced to BF₃·OEt (δ 0 ppm). Chemical shifts are reported in δ ppm. IR spectra were recorded on a Perkin-Elmer Spectrum One. Mass spectra were obtained with Thermo Fisher Scientific Exactive, JEOL JMS-T100 GC or JEOL JMS-T100LP at the Center for Instrumental Analysis, Hokkaido University. Elemental analysis was performed at the Center for Instrument Analysis of Hokkaido University. TLC analyses were performed on commercial glass plates bearing 0.25-mm layer of Merck Silica gel 60F₂₅₄. Silica gel (Kanto Chemical Co., Silica gel 60 N, spherical, neutral) and aluminum oxide (Nacalai Tesuque, Alumina Activated 200) were used for column chromatography. HPLC analyses were conducted on a HITACHI ELITE LaChrom system with a HITACHI L-2400 UV detector or a Shimadzu LC-6A system with a Shimadzu SPD-10A UV detector. Gas chromatographic (GLC) analyses were conducted on a Shimadzu GC-14B equipped with a flame ionization detector. Gel permeation chromatography (GPC) was performed by LC-908 (Japan Analytical Industry Ltd., two in-line JAIGEL-2H, CHCl₃, 3.5 mL/min, UV and RI **S**1

detectors).

All reactions were carried out under nitrogen or argon atmosphere. Materials were obtained from commercial suppliers or prepared according to standard procedures unless otherwise noted. *t*-BuOK (1.0 M THF solution) and CuOAc were purchased from Aldrich Chemical Co., stored under nitrogen, and used as it is. THF was purchased from Kanto Chemical Co., stored under argon. Alkenes **1a–g** was well known compounds. 20(21)-Methylene steroid **1n** was prepared from pregrenone acetate according to the reported procedure.¹

Preparation of Allylic Phosphates

Preparation of Allylic Phosphates 3a, a' and 3c–e. Allylic substrates were prepared by the phosphorylation of the corresponding allylic alcohols. The allylic alcohols were prepared by the reduction of the corresponding propargylic alcohols by Cp_2TiCl_2/i -BuMgBr according to the reported procedure.² The obtained crude allylic alcohols were purified by silica gel chromatography and GPC (CHCl₃).

The preparation of **3a** is representative. To a solution of (*Z*)-1-phenyl-4-nonen-3-ol (437.7 mg, 2 mmol) in pyridine (2.2 mL, 5 mL per 1 gram of alcohol), $(EtO)_2P(O)Cl$ (405 μ L, 2.8 mmol) and DMAP (61.1 mg, 0.5 mmol) were sequentially added at 0 °C. After being stirred at rt for 2 h, the reaction mixture was diluted with EtOAc (66 mL) and quenched with H₂O (5 mL). The resulting mixture was washed with sat. CuSO₄ (10 mL × 3) and brine, and was dried over anhydrous MgSO₄, filtered, and evaporated under reduced pressure. The residue was purified through a short plug of aluminum oxide (ether) to provide **3a** in 86% yield (612.8 mg, 1.7 mmol).

Preparation of Allylic Phosphates 3b, 3f and 3g. Allylic phosphate **3b** was synthesized as follows: 2,3-*O*-isopropylidene-glyceraldehyde was subjected to *Z*-selective Horner-Wadsworth-Emmons-type reaction with pentyltriphenylphosphonium bromide in the presence of NaHMDS to give the corresponding *Z*-alkene. Next, deprotection of the acetal, silylation and phosphorylation produced **3b**. Allylic substrates **3f** and **3g** were obtained by the phosphorylation of the corresponding commercial available alcohols.

Preparation of (*S*, *Z*)-**Diethyl 3-Octen-2-yl Phosphate** (**3h**). (*S*)-3-Octyn-2-ol was prepared by the asymmetric reduction of 3-octyn-2-one according to the reported procedure.³ (*S*)-3-Octyn-2-ol was reduced with Cp₂TiCl₂/*i*-BuMgBr according to the reported procedure,² producing (*S*, *Z*)-3-octen-2-ol, which then was converted to the corresponding allylic phosphate (*S*)-(*Z*)-**3h**. The ee value of (*S*)-(*Z*)-**3h** (95% ee) was determined by HPLC analysis of the *p*-nitrobenzoate derivative of (*S*, *Z*)-3-octen-2-ol (CHIRALCEL[®] OD-3 column, 4.6 mm × 250 mm, Daicel Chemical Industries, hexane, 0.5 mL/min, 40 °C, 254 nm UV detector, retention time = 32.8 min for the *R* isomer and 34.3 min for the *S* isomer). The absolute configuration of (*S*)-(*Z*)-**3h** was determined by optical rotations of the precursor compounds, 3-octyn-2-ol.³ **Preparation of Cyclic Allylic Phosphates.** Allylic substrates **3i** and **3k–m** were prepared by the phosphorylation of the corresponding known allylic alcohols. Allylic phosphate (1R,4S)-**3j** was synthesized from commercial available (1R,3S)-*cis*-4-cyclopenetene-1,3-diol 1-acetate by silylation, deacetylation and phosphorylation.

Characterization Data for Allylic Phosphates

(Z)-Diethyl 1-Phenyl-4-nonen-3-yl Phosphate (3a)

Bu OP(O)(OEt)₂ Ph **3a**

Oil. ¹H NMR (300 MHz, CDCl₃) δ 0.88 (t, J = 7.2 Hz, 3H), 1.28–1.35 (m, 10H), 1.80–1.89 (m, 1H), 2.06–2.16 (m, 3H), 2.65–2.71 (m, 2H), 4.02–4.15 (m, 4H), 5.15 (dq, J = 9.0, 6.9 Hz, 1H), 5.46 (dd, J = 11.1, 9.0 Hz, 1H), 5.59 (dt, J = 11.1, 7.5 Hz, 1H), 7.18–7.21 (m, 3H), 7.26–7.31 (m, 2H). ¹³C NMR (75.4 MHz, CDCl₃) δ 13.77, 15.90 (d, J = 4.0 Hz), 16.00 (d, J = 3.4 Hz), 22.18, 27.38, 31.03, 31.48, 38.06 (d, J = 6.9 Hz), 63.40 (d, J = 5.7 Hz), 63.42 (d, J = 5.7 Hz), 74.28 (d, J = 6.3 Hz), 125.99, 128.30 (d, J = 3.4 Hz), 128.40, 128.45, 134.18, 141.44. HRMS–ESI (m/z): [M+Na]⁺ calcd for C₁₉H₃₁O₄PNa, 377.18522; found, 377.18552.

(Z)-Diethyl 1-Phenyl-3-nonen-5-yl Phosphate (3a')



Oil. ¹H NMR (300 MHz, CDCl₃) δ 0.87 (t, *J* = 6.9 Hz, 3H), 1.17–1.43 (m, 11H), 1.64 (m, 1H), 2.40–2.52 (m, 2H), 2.56–2.81 (m, 2H), 3.98–4.14 (m, 4H), 5.03 (dq, *J* = 9.3, 6.6 Hz, 1H), 5.41 (dd, *J* = 11.1, 9.3 Hz, 1H), 5.59 (dt, *J* = 11.1, 7.5 Hz, 1H), 7.16–7.21 (m, 3H), 7.22–7.31 (m, 2H). ¹³C NMR (75.4 MHz, CDCl₃) δ 13.85, 15.93 (d, *J* = 2.9 Hz), 16.30 (d, *J* = 2.9 Hz), 22.31, 26.80, 29.52, 35.50, 35.81 (d, *J* = 6.3 Hz), 63.34 (d, *J* = 5.7 Hz), 63.36 (d, *J* = 5.7 Hz), 74.73 (d, *J* = 6.3 Hz), 126.02, 128.41, 128.59, 129.50 (d, *J* = 2.9 Hz), 132.42, 141.47. HRMS–ESI (*m*/*z*): [M+Na]⁺ calcd for C₁₉H₃₁O₄PNa, 377.1858; found, 377.1862.

(Z)-1-(*tert*-Butyldimethylsiloxy)-3-octen-2-yl Diethyl Phosphate (3b)



Oil. ¹H NMR (300 MHz, CDCl₃) δ 0.06 (s, 6H), 0.88–0.92 (m, 12H), 1.28–1.40 (m, 10H), 2.12–2.16 (m, 2H), 3.61 (ddd, *J* = 10.8, 5.1, 1.8 Hz, 1H), 3.73 (dd, *J* = 10.8, 6.3 Hz, 1H), 4.02–4.17 (m, 4H), 5.12 (m, 1H), 5.40 (ddt, *J* = 10.8, 9.3, 1.2 Hz, 1H), 5.64 (dtd, *J* = 10.8, 7.5, 1.2 Hz, 1H). ¹³C NMR (75.4 MHz, CDCl₃) δ –5.56, 13.81, 15.91, 16.01, 18.25, 22.23, 25.75, 27.58, 31.57, 63.38

 $(d, J = 5.6 \text{ Hz}), 63.42 (d, J = 5.6 \text{ Hz}), 65.85 (d, J = 7.4 \text{ Hz}), 74.96 (d, J = 5.6 \text{ Hz}), 125.67 (d, J = 2.8 \text{ Hz}), 135.60. \text{HRMS-ESI} (m/z): [M+Na]^+ calcd for C_{18}H_{39}O_5PSiNa, 417.22021; found, 417.22031.$

(Z)-Diethyl 6-Phenyl-2-hexen-4-yl Phosphate (3c)

Oil. ¹H NMR (300 MHz, CDCl₃) δ 1.28 (td, J = 6.6, 0.9 Hz, 3H), 1.30 (td, J = 6.6, 0.9 Hz, 3H), 1.70 (dd, J = 6.9, 1.5 Hz, 3H), 1.82 (m, 1H), 2.10 (m, 1H), 2.63–2.72 (m, 2H), 4.02–4.15 (m, 4H), 5.17 (m, 1H), 5.50 (ddq, J = 13.2, 9.0, 1.5 Hz, 1H), 5.70 (dq, J = 13.2, 6.9 Hz 1H), 7.17–7.21 (m, 3H), 7.27–7.31 (m, 2H). ¹³C NMR (75.4 MHz, CDCl₃) δ 13.30, 15.92 (d, J = 3.4 Hz), 16.01 (d, J = 3.4 Hz), 31.00, 37.81 (d, J = 6.9 Hz), 63.43 (d, J = 6.3 Hz), 63.45 (d, J = 6.3 Hz), 73.89 (d, J = 6.3 Hz), 126.01, 128.42, 128.45, 128.48, 129.40 (d, J = 3.4 Hz), 141.41. HRMS–ESI (m/z): [M+Na]⁺ calcd for C₁₆H₂₅O₄PNa, 335.13881; found, 335.13924.

(Z)-Diethyl 7-Methyl-1-phenyl-4-octen-3-yl Phosphate (3d)



Oil. ¹H NMR (300 MHz, CDCl₃) δ 0.87 (d, *J* = 6.9 Hz, 3H), 0.90 (d, *J* = 6.9 Hz, 3H), 1.30 (dt, *J* = 6.9, 1.2 Hz, 3H), 1.33 (dt, *J* = 6.9, 1.2 Hz, 3H), 1.63 (septet, *J* = 6.9 Hz, 1H), 1.86 (m, 1H), 1.99 (t, *J* = 7.2 Hz, 2H), 2.10 (m, 1H), 2.61–2.77 (m, 2H), 4.02–4.17 (m, 4H), 5.16 (dt, *J* = 14.7, 7.2, 1H), 5.57 (m, 2H), 7.17–7.19 (m, 3H), 7.21–7.31 (m, 2H). ¹³C NMR (75.4 MHz, CDCl₃) δ 15.95 (d, *J* = 4.0 Hz), 16.05 (d, *J* = 4.0 Hz), 22.18, 22.21, 28.39, 31.07, 36.62, 38.12 (d, *J* = 6.3 Hz), 63.43 (d, *J* = 6.3 Hz), 63.45 (d, *J* = 6.3 Hz), 74.40 (d, *J* = 6.3 Hz), 126.02, 128.45, 128.48, 129.09 (d, *J* = 3.4 Hz), 132.90, 141.52. HRMS–ESI (*m*/*z*): [M+Na]⁺ calcd for C₁₉H₃₁O₄PNa, 377.18576; found, 377.18750.

(Z)-Diethyl 2-Methyl-5-decen-4-yl Phosphate (3e)



Oil. ¹H NMR (300 MHz, CDCl₃) δ 0.88–0.97 (m, 9H), 1.28–1.39 (m, 11H), 1.62–1.74 (m, 2H), 2.12–2.19 (m, 2H), 4.01–4.15 (m, 4H), 5.17 (m, 1H), 5.39 (tt, *J* = 10.8, 1.2, 1H), 5.54 (dt, *J* = 10.8, 7.5, 1H). ¹³C NMR (75.4 MHz, CDCl₃) δ 13.81, 15.91 (d, *J* = 3.4 Hz), 16.01 (d, *J* = 3.4 Hz), 22.24 (d, *J* = 3.4 Hz), 22.70, 24.08, 27.35, 31.54, 45.40 (d, *J* = 6.3 Hz), 63.26 (d, *J* = 5.7 Hz), 63.30 (d, *J* = 5.7 Hz), 73.35 (d, *J* = 5.7 Hz), 128.99 (d, *J* = 2.9 Hz), 133.62. HRMS–ESI (*m*/*z*): [M+Na]⁺ calcd for C₁₅H₃₁O₄PNa, 329.18522; found, 329.18579.



Oil. ¹H NMR (300 MHz, CDCl₃) δ 0.90 (t, J = 7.5 Hz, 3H), 1.34 (td, J = 7.2, 1.2 Hz, 6H), 1.36–1.46 (m, 2H), 2.07 (q, J = 6.9 Hz, 2H), 4.11 (quint., J = 7.2 Hz, 4H), 4.60 (dd, J = 8.1, 6.0 Hz, 2H), 5.55–5.70 (m, 2H). ¹³C NMR (75.4 MHz, CDCl₃) δ 13.34, 15.80, 15.89, 22.25, 29.22, 62.84, 62.91, 63.51 (d, J = 5.7 Hz), 124.09 (d, J = 6.8 Hz), 135.11. HRMS–ESI (m/z): [M+Na]⁺ calcd for C₁₀H₂₁O₄PNa, 259.10697; found, 259.10698.

(Z)-Diethyl 3,7-Dimethylocta-2,6-dienyl Phosphate (3g)



Oil. ¹H NMR (300 MHz, CDCl₃) δ 1.33 (t, J = 6.9 Hz, 6H), 1.60 (s, 3H), 1.68 (s, 3H), 1.77 (s, 3H), 2.05–2.15 (m, 4H), 4.10 (quint., J = 6.9 Hz, 4H), 4.54 (t, J = 7.5 Hz, 2H), 5.08 (m, 1H), 5.41 (t, J = 7.5 Hz, 1H). ¹³C NMR (75.4 MHz, CDCl₃) δ 15.95, 16.04, 17.53, 23.37, 25.56, 26.48, 32.01, 63.53 (d, J = 5.7 Hz), 63.71 (d, J = 5.7 Hz), 119.96 (d, J = 6.8 Hz), 123.51, 132.36, 142.74. HRMS–ESI (m/z): [M+Na]⁺ calcd for C₁₄H₂₇O₄PNa, 313.15392; found, 313.15407.

(S, Z)-Diethyl 3-Octen-2-yl Phosphate (3h).



Oil. ¹H NMR (300 MHz, CDCl₃) δ 0.90 (t, *J* = 6.9 Hz, 3H), 1.29–1.36 (m, 10H), 1.38 (d, *J* = 6.3 Hz, 3H), 2.09–2.16 (m, 2H), 4.02–4.14 (m, 4H), 5.31 (dq, *J* = 12.6, 6.3 Hz, 1H), 5.41–5.55 (m, 2H). ¹³C NMR (75.4 MHz, CDCl₃) δ 13.79, 15.90 (d, *J* = 1.1 Hz), 15.99 (d, *J* = 1.1 Hz), 22.17, 22.67 (d, *J* = 5.1 Hz), 27.22, 31.53, 63.55 (d, *J* = 5.7 Hz), 63.40 (d, *J* = 5.7 Hz), 71.28 (d, *J* = 5.7 Hz), 129.92 (d, *J* = 5.1 Hz), 132.76. HRMS–ESI (*m*/*z*): [M+Na]⁺ calcd for C₁₂H₂₅O₄PNa, 287.13881; found, 287.13795. [α]_D²⁴ +46.4 (*c* 1.54, CHCl₃).

cis-4-Cyclopentene-1,3-diyl Tetraethyl Diphosphate (3i)



Oil. ¹H NMR (300 MHz, CDCl₃) δ 1.35 (t, *J* = 7.2 Hz, 12H), 2.05 (dt, *J* = 14.4, 4.5 Hz, 1H), 2.93 (dt, *J* = 14.4, 6.9 Hz, 1H), 4.12 (quint., *J* = 7.2 Hz, 8H), 5.23 (dd, *J* = 6.9, 4.5 Hz, 2H), 6.15 (s, 2H). ¹³C NMR (75.4 MHz, CDCl₃) δ 16.00 (d, *J* = 5.7 Hz), 39.51, 63.81 (d, *J* = 5.7 Hz), 79.14 (d, *J* = 5.7 Hz), 135.15 (d, *J* = 5.1 Hz). HRMS–ESI (*m*/*z*): [M+Na]⁺ calcd for C₁₃H₂₆O₈P₂Na, 395.09951; found,

Diethyl (1R, 4S)-4-(Triisopropylsiloxy)-2-cyclopentenyl Phosphate (3j)



Oil. ¹H NMR (300 MHz, CDCl₃) δ 1.05–1.08 (m, 21H), 1.34 (td, J = 7.2, 0.9 Hz, 6H), 1.80 (dt, J = 13.5, 5.4 Hz, 1H), 2.86 (dt, J = 13.5, 7.2 Hz, 1H), 4.11 (quint., J = 7.2 Hz, 4H), 4.75 (t, J = 5.1 Hz, 1H), 5.18 (q, J = 6.0 Hz, 1H), 5.96 (dm, J = 5.4 Hz, 1H), 6.02 (dm, J = 5.4 Hz, 1H). ¹³C NMR (75.4 MHz, CDCl₃) δ 11.91, 15.95, 16.04, 17.80, 17.81, 42.61 (d, J = 4.5 Hz), 63.59 (d, J = 1.2 Hz), 63.67 (d, J = 1.2 Hz), 74.54, 79.84 (d, J = 5.8 Hz), 131.84 (d, J = 5.8 Hz), 139.21. HRMS–ESI (m/z): [M+Na]⁺ calcd for C₁₈H₃₇O₅PSiNa, 415.20401; found, 415.20462. [α]_D²⁵–16.4 (c 1.1, C₆H₆).

2-Cyclohexenyl Diethyl Phosphate (3k)



Oil. ¹H NMR (300 MHz, CDCl₃) δ 1.34 (t, J = 7.2 Hz, 6H), 1.55–2.18 (m, 6H), 4.11 (quint., J = 7.2 Hz, 4H), 4.88 (brs, 1H), 5.79 (d, J = 10.2 Hz, 1H), 5.95 (dt, J = 10.2, 3.6 Hz, 1H), ¹³C NMR (75.4 MHz, CDCl₃) δ 15.89, 15.98, 18.27, 24.56, 29.69 (d, J = 4.6 Hz), 63.38 (d, J = 5.7 Hz), 63.45 (d, J = 5.7 Hz), 72.03 (d, J = 5.7 Hz), 126.29 (d, J = 4.6 Hz), 132.73. HRMS–ESI (m/z): [M+Na]⁺ calcd for C₁₀H₁₉O₄PNa, 257.09186; found, 257.09175.

Diethyl 4,4-Dimethyl-2-cyclohexenyl Phosphate (3l)



Oil. ¹H NMR (300 MHz, CDCl₃) δ 0.97 (s, 3H), 1.03 (s, 3H), 1.34 (td, *J* = 6.6, 1.2 Hz, 6H), 1.42 (m, 1H), 1.63 (m, 1H), 1.82–2.01 (m, 2H), 4.11 (quintet d, *J* = 7.2, 1.2 Hz, 4H), 4.82 (q, *J* = 5.4 Hz, 1H), 5.63 (s, 2H). ¹³C NMR (75.4 MHz, CDCl₃) δ 15.91, 16.00, 26.90 (d, *J* = 4.0 Hz), 28.44, 28.82, 31.56, 32.88, 63.40, 63.47 (d, *J* = 6.3 Hz), 72.32 (d, *J* = 5.7 Hz), 123.75 (d, *J* = 5.1 Hz), 142.56. HRMS–ESI (*m*/*z*): [M+Na]⁺ calcd for C₁₂H₂₃O₄PNa, 285.12262; found, 285.12304.

2-Cycloheptenyl Diethyl Phosphate (3m)



Oil. ¹H NMR (300 MHz, CDCl₃) δ 1.34 (t, J = 7.2 Hz, 6H), 1.42 (m, 1H), 1.56–1.84 (m, 3H), 1.92–2.09 (m, 3H), 2.19 (m, 1H), 4.11 (quint., J = 7.2 Hz, 4H), 5.01 (t, J = 9.0 Hz, 1H), 5.74–5.87 (m, 2H). ¹³C NMR (75.4 MHz, CDCl₃) δ 16.00, 16.06, 26.09, 26.34, 28.29, 34.37 (d, J = 4.6 Hz), 63.53 (d, J = 6.3 Hz), 63.54 (d, J = 6.3 Hz), 78.34 (d, J = 5.7 Hz), 131.71, 134.15 (d, J = 5.1 Hz). HRMS–ESI (m/z): [M+Na]⁺ calcd for C₁₁H₂₁O₄PNa, 271.10751; found, 271.10732.

Procedures for Copper-Catalyzed Allyl–Alkyl Coupling

Typical Procedure for Hydroboration/Allyl–Alkyl Coupling Sequence (Scheme 1). In a glove box, $(9\text{-}BBN\text{-}H)_2$ (91.5 mg, 0.375 mmol) was placed in a vial containing a magnetic stirring bar. Then, the vial was sealed with a cap equipped with a Teflon-coated silicon rubber septum. The vial was removed from the glove box. THF (0.3 mL) and styrene (105 μ L, 0.9 mmol) were sequentially added, and the mixture was stirred at 60 °C for 1 hour to prepare alkylborane 2a. On the other hand, in a glove box, CuOAc (6.1 mg, 0.05 mmol) was placed in another vial. *t*-BuOK (1 M in THF, 0.5 ml, 0.5 mmol) was added to alkylborane 2a prepared in advance at 25 °C, and the mixture was stirred at 25 °C for 5 min to produce the corresponding alkylborate. Next, the alkylborate was then transferred to another vial containing Cu salt. Finally, allylic phosphate 3a (177.2 mg, 175 μ L, 0.5 mmol) was added. After 8 h stirring at 60 °C, CH₂Cl₂ was added to the mixture. Then, the mixture was filtered through a short plug of silica gel, which was washed with diethyl ether. After the solvent was removed under reduced pressure, flash chromatography on silica gel (hexane) provided 4a (122 mg, 0.4 mmol) in 80% yield.

Procedure for Hydroboration/Allyl–Alkyl Coupling Sequence (Scheme 2). In a glove box, $(9\text{-BBN-H})_2$ (33.6 mg, 0.1375 mmol) and steroid **1n** (102.8 mg, 0.286 mmol) were placed in a vial containing a magnetic stirring bar. Then the vial was sealed with a cap equipped with a Teflon-coated silicon rubber septum. The vial was removed from the glove box. The mixture was dissolved in THF (0.37 mL) at 0 °C, and the mixture was stirred at 25 °C for 24 hour to prepare alkylborane **2n**. After the vial was brought into a glove box, *t*-BuOK (1 M in THF, 0.25 ml, 0.25 mmol) was added at 25 °C, and the mixture was stirred at 25 °C for 5 min to produce the corresponding alkylborate. Next, CuOAc (3.1 mg, 0.025 mmol) was added, and then the vial was removed from the glove box. Finally, allylic phosphate (1*R*,4*S*)-**3j** (98.1 mg, 0.25 mmol) was added. After 8 h stirring at 60 °C, CH₂Cl₂ was added to the mixture. Then, the mixture was filtered through a short plug of silica gel, which was washed with diethyl ether. After the solvent was removed under reduced pressure, flash chromatography on silica gel (hexane) and GPC (CHCl₃) provided **5** (74.1 mg, 0.125 mmol) in 50% yield.

Characterization Data for Coupling Products

Coupling products $4k^4$ and $4m^5$ were reported in the literature.

(*E*)-5-(2-Phenylethyl)-1-phenyl-3-nonene (4aa)

Oil. ¹H NMR (300 MHz, CDCl₃) δ 0.86 (t, J = 6.9 Hz, 3H), 1.08–1.37 (m, 6H), 1.45 (m, 1H), 1.62 (m, 1H), 1.88 (m, 1H), 2.36 (q, J = 7.2 Hz, 2H), 2.44 (ddd, J = 13.8, 10.2, 6.3 Hz, 1H), 2.57 (ddd, J = 13.8, 10.2, 5.2 Hz, 1H), 2.70 (t, J = 7.2 Hz, 2H), 5.16 (dd, J = 15.3, 8.9 Hz, 1H), 5.40 (dt, J = 15.3, 7.2 Hz, 1H), 7.12–7.20 (m, 6H), 7.24–7.30 (m, 4H). ¹³C NMR (75.4 MHz, CDCl₃) δ 14.00, 22.70, 29.29, 33.53, 34.30, 35.16, 36.13, 37.22, 42.45, 125.55, 125.77, 128.27, 128.31, 128.49, 128.59, 129.78, 135.51, 142.15, 143.19. Anal. Calcd for C₂₃H₃₀: C, 90.13; H, 9.87%. Found: C, 89.95; H, 10.07%. The regioselectivity of **4aa** was assigned on the basis of ¹H and ¹³C NMR of 3-phenylpropanoic acid obtained by ozonolytic cleavage followed by Jones oxidation.

(E)-3-(2-Phenylethyl)-1-phenyl-4-nonene (4a'a)

Oil. ¹H NMR (300 MHz, CDCl₃) δ 0.92 (t, J = 7.2 Hz, 3H), 1.25–1.44 (m, 4H), 1.47–1.60 (m, 2H), 1.63–1.75 (m, 2H), 1.93–2.10 (m, 3H), 2.50 (ddd, J = 13.8, 10.2, 6.3 Hz, 2H), 2.64 (ddd, J = 13.8, 10.2, 5.4 Hz, 2H), 5.20 (dd, J = 15.3, 8.7 Hz, 1H), 5.43 (dt, J = 15.3, 6.9 Hz, 1H), 7.11–7.18 (m, 6H), 7.21–7.29 (m, 4H). ¹³C NMR (75.4 MHz, CDCl₃) δ 13.86, 22.10, 31.83, 32.27, 33.54, 37.37, 42.24, 125.61, 128.32, 128.49, 131.79, 134.08, 143.07. Anal. Calcd for C₂₃H₃₀: C, 90.13; H, 9.87%. Found: C, 90.10; H, 9.88%. The regioselectivity of **4a'a** was assigned on the basis of ¹H and ¹³C NMR of pentanoic acid obtained by ozonolytic cleavage followed by Jones oxidation.

(E)-1-(*tert*-Butyldimethylsiloxy)-4-(2-phenylethyl)-2-octene (4b)

Oil. ¹H NMR (300 MHz, CDCl₃) δ 0.88 (s, 6H), 0.84–0.89 (m, 3H), 0.92 (s, 9H), 1.12–1.40 (m, 6H), 1.54 (m, 1H), 1.66 (m, 1H), 1.99 (m, 1H), 2.50 (ddd, *J* = 13.8, 10.2, 6.6 Hz, 1H), 2.63 (ddd, *J* = 13.8, 10.2, 5.4 Hz, 1H), 4.17 (d, *J* = 5.1 Hz, 2H), 5.41 (dd, *J* = 15.3, 8.4 Hz, 1H), 5.53 (dt, *J* = 15.3, 5.1 Hz, 1H), 7.15–7.18 (m, 3H), 7.24–7.30 (m, 2H). ¹³C NMR (75.4 MHz, CDCl₃) δ –5.19, 13.98, 18.33, 22.70, 25.86, 29.26, 33.51, 34.90, 36.99, 41.96, 63.99, 125.61, 128.32, 128.48, 129.74, 135.31, 143.04. Anal. Calcd for C₂₂H₃₈OSi: C, 76.23; H, 11.05%. Found: C, 76.15; H, 10.94%. The regioselectivity of **4b** was assigned on the basis of ¹H and ¹³C NMR of 2-(2-phenylethyl)hexanoic acid⁶ obtained by ozonolytic cleavage followed by Jones oxidation.

(E)-5-Butyl-10-triisopropylsiloxy-1-phenyl-3-decene (4ab)

Oil. ¹H NMR (300 MHz, CDCl₃) δ 0.87 (t, *J* = 7.2 Hz, 3H), 1.00–1.25 (s, 33H), 1.49–1.54 (m, 2H), 1.84 (m, 1H), 2.31 (td, *J* = 7.8, 6.6 Hz, 2H), 2.67 (t, *J* = 7.8 Hz, 2H), 3.66 (t, *J* = 6.6 Hz, 2H), 5.10 (dd, *J* = 15.3, 8.7 Hz, 1H), 5.34 (dt, *J* = 15.3, 6.6 Hz, 1H), 7.17–7.19 (m, 3H), 7.25–7.30 (m, 2H). ¹³C NMR (75.4 MHz, CDCl₃) δ 11.88, 14.00, 17.92, 22.70, 25.82, 26.95, 29.36, 32.95, 34.33, 35.11, 35.39, 36.18, 42.64, 63.49, 125.70, 128.26, 128.59, 129.00, 135.96, 142.27. Anal. Calcd for C₂₉H₅₂OSi: C, 78.31; H, 11.78%. Found: C, 78.10; H, 11.80%.

(E)-5-Butyl-10,10-trimethylenedioxy-1-phenyl-3-decene (4ac)



Oil. ¹H NMR (300 MHz, CDCl₃) δ 0.86 (t, J = 6.9 Hz, 3H), 1.11–1.36 (m, 13H), 1.53–1.60 (m, 2H), 1.82 (m, 1H), 2.07 (m, 1H), 2.31 (td, J = 7.5, 6.6 Hz, 2H), 2.66 (t, J = 7.5 Hz, 2H), 3.76 (td, J = 12.0, 2.4 Hz, 2H), 4.10 (ddd, J = 12.0, 5.1, 1.2 Hz, 2H), 4.49 (t, J = 5.1 Hz, 1H), 5.09 (dd, J = 15.0, 8.7 Hz, 1H), 5.34 (dt, J = 15.0, 6.6 Hz, 1H), 7.14–7.22 (m, 3H), 7.24–7.30 (m, 2H). ¹³C NMR (75.4 MHz, CDCl₃) δ 14.01, 22.71, 23.97, 25.76, 26.93, 29.34, 34.34, 35.10, 35.16, 35.23, 36.18, 42.55, 66.88, 102.50, 125.72, 128.27, 128.59, 129.07, 135.88, 142.26. HRMS–EI (m/z): [M–H]⁺ calcd for C₂₃H₃₅O₂, 343.26370; found, 343.26306.

(E)-Methyl 6-Butyl-3,3-dimethyl-10-phenyl-7-decenoate (4ad)



Oil. ¹H NMR (300 MHz, CDCl₃) δ 0.87 (t, *J* = 6.9 Hz, 3H), 0.95 (s, 6H), 1.06–1.29 (m, 10H), 1.75 (m, 1H), 2.17 (s, 2H), 2.32 (q, *J* = 7.2 Hz, 2H), 2.68 (t, *J* = 7.2 Hz, 2H), 3.64 (s, 3H), 5.09 (dd, *J* = 15.3, 8.7 Hz, 1H), 5.34 (dt, *J* = 15.3, 7.2 Hz, 1H), 7.17–7.20 (m, 3H), 7.26–7.30 (m, 2H). ¹³C NMR (75.4 MHz, CDCl₃) δ 14.00, 22.69, 27.14, 27.19, 29.33, 29.58, 33.05, 34.25, 35.12, 36.14, 39.82, 43.25, 45.74, 51.00, 125.74, 128.28, 128.58, 129.27, 135.77, 142.20, 173.09. HRMS–EI (*m*/*z*): [M]⁺ calcd for C₂₃H₃₆O₂, 344.27153; found, 344.24148.

(E)-1-(3,4-Dimethoxyphenyl)-7-(2-phenylethyl)-5-decene (4a'e)



Oil. ¹H NMR (300 MHz, CDCl₃) δ 0.90 (t, J = 6.9 Hz, 3H), 1.18–1.69 (m, 10H), 1.92–2.06 (m, 3H), 2.43–2.53 (m, 3H), 2.64 (ddd, J = 14.1, 9.9, 5.4 Hz, 1H), 3.86 (s, 3H), 3.87 (s, 3H), 5.13 (dd, J = 15.3, 6.9 Hz, 1H), 5.39 (dd, J = 15.3, 6.9 Hz, 1H), 6.67–6.71 (m, 2H), 6.76–6.80 (m, 1H), 7.14–7.19 (m, 3H), 7.24–7.29 (m, 2H). ¹³C NMR (75.4 MHz, CDCl₃) δ 13.83, 22.07, 29.22, 31.83, 32.22, 33.55, 35.07, 35.48, 37.30, 42.32, 55.69, 55.84, 111.12, 111.72, 120.16, 125.58, 128.29, 128.49, 131.25, 134.31, 135.60, 143.12, 147.06, 148.80. Anal. Calcd for C₂₆H₃₆O₂: C, 82.06; H, 9.53%. Found: C, 82.01; H, 9.55%.

(E)-7-(2-Phenylethyl)-12-phthalimide-5-dodecene (4a'f)



Oil. ¹H NMR (300 MHz, CDCl₃) δ 0.89 (t, *J* = 6.9 Hz, 3H), 1.18–1.36 (m, 10H), 1.45 (m, 1H), 1.60–1.69 (m, 3H), 1.89 (m, 1H), 2.01 (q, *J* = 6.3 Hz, 2H), 2.48 (ddd, *J* = 13.5, 10.2, 6.6 Hz, 1H), 2.62 (ddd, *J* = 13.5, 10.2, 5.1 Hz, 1H), 3.66 (t, *J* = 7.2 Hz, 2H), 5.11 (dd, *J* = 15.0, 8.7 Hz, 1H), 5.35 (dt, *J* = 15.0, 6.6 Hz, 1H), 7.14–7.17 (m, 3H), 7.24–7.34 (m, 2H), 7.70 (dd, *J* = 6.0, 3.0 Hz, 2H), 7.84 (dd, *J* = 6.0, 3.0 Hz, 2H). ¹³C NMR (75.4 MHz, CDCl₃) δ 13.83, 22.06, 26.68, 26.87, 28.50, 31.79, 32.20, 33.55, 35.27, 37.29, 38.00, 42.41, 123.21, 125.55, 128.29, 128.48, 131.14, 132.26, 133.91, 134.34, 143.18, 168.62. Anal. Calcd for C₂₈H₃₅NO₂: C, 80.53; H, 8.45; N, 3.35%. Found: C, 80.40; H, 8.45; N, 3.31%.

(*E*)-1-Phenyl-5-(2-phenylpropyl)-3-nonene (4ag)

Oil. ¹H NMR (300 MHz, CDCl₃) δ 0.80 (t, J = 7.2 Hz, 0.5 × 3H), 0.88 (t, J = 7.2 Hz, 0.5 × 3H), 1.05–1.36 (m, 0.5 × 9H, 0.5 × 9H), 1.40 (m, 0.5 × 1H), 1.46–1.48 (m, 0.5 × 2H), 1.58–1.65 (m, 0.5 × 2H), 1.97 (m, 0.5 × 1H), 2.31–2.38 (m, 0.5 × 2H, 0.5 × 2H), 2.62–2.73 (m, 0.5 × 3H, 0.5 × 3H), 5.08 (dd, J = 15.5, 9.0 Hz, 0.5 × 1H), 5.08 (dd, J = 15.5, 9.0 Hz, 0.5 × 1H), 5.23 (dt, J = 15.5, 6.3 Hz, 0.5 × 1H), 5.36 (dt, J = 15.5, 6.3 Hz, 0.5 × 1H), 7.09–7.31 (m, 0.5 × 10H, 0.5 × 10H). ¹³C NMR (75.4 MHz, CDCl₃) δ 13.94, 13.99, 20.93, 22.67 (× 2C), 23.61, 29.13, 29.22, 34.21, 34.29, 35.30, 35.62, 36.09 (× 2C), 36.75, 37.24, 40.43, 40.52, 43.91, 44.28, 125.71, 125.74, 125.76 (× 2C), 126.98, 127.33, 128.30 (× 4C), 128.60 (× 2C), 129.41, 129.66, 135.49, 135.68, 142.21 (× 2C), 147.60, 148.78. HRMS–EI (m/z): [M]⁺ calcd for C₂₄H₃₂, 320.25040; found, 320.25048. The regioselectivity of **4ag** was assigned on the basis of ¹H and ¹³C NMR of 3-phenylpropanoic acid obtained by ozonolytic cleavage followed by Jones oxidation.

(*E*)-5-Methyl-1,7-diphenyl-3-heptene (4c)



Oil. ¹H NMR (300 MHz, CDCl₃) δ 0.97 (d, J = 6.9 Hz, 3H), 1.50–1.62 (m, 2H), 2.10 (septet, J = 6.9 Hz, 1H), 2.33 (q, J = 6.9 Hz, 2H), 2.46–2.61 (m, 2H), 2.69 (t, J = 7.2 Hz, 2H), 5.33 (dd, J = 15.9, 6.9 Hz, 1H), 5.43 (dt, J = 15.9, 7.2 Hz, 1H), 7.14–7.21 (m, 6H), 7.24–7.30 (m, 4H). ¹³C NMR (75.4 MHz, CDCl₃) δ 20.86, 33.55, 34.33, 36.09, 36.33, 38.78, 125.60, 125.78, 128.24, 128.31 (× 2C), 128.49, 128.60, 136.78, 142.20, 143.04. Anal. Calcd for C₂₀H₂₄: C, 90.85; H, 9.15%. Found: C, 90.81; H, 9.19%.

(E)-7-Methyl-1-phenyl-5-(2-phenylethyl)-3-octene(4d)



Oil. ¹H NMR (300 MHz, CDCl₃) δ 0.79 (d, *J* = 6.6 Hz, 3H), 0.82 (d, *J* = 6.6 Hz, 3H), 1.10 (dt, *J* = 8.7, 5.7 Hz, 2H), 1.35–1.65 (m, 3H), 2.00 (m, 1H), 2.38 (m, 2H), 2.45 (ddd, *J* = 13.8, 10.2, 6.3 Hz, 1H), 2.56 (ddd, *J* = 13.8, 10.2, 5.4 Hz, 1H), 2.70 (t, *J* = 7.5 Hz, 2H), 5.12 (dd, *J* = 15.3, 9.0 Hz, 1H), 5.40 (dt, *J* = 15.3, 6.6 Hz, 1H), 7.12–7.20 (m, 6H), 7.24–7.30 (m, 4H). ¹³C NMR (75.4 MHz, CDCl₃) δ 21.68, 23.47, 25.08, 33.52, 34.31, 36.12, 37.60, 40.34, 44.85, 125.55, 125.77, 128.27, 128.30, 128.48, 128.61, 129.68, 135.51, 142.13, 143.19. Anal. Calcd for C₂₃H₃₀: C, 90.13; H, 9.87%. Found: C, 90.04; H, 9.81%. The regioselectivity of **4d** was assigned on the basis of ¹H and ¹³C NMR of 3-phenylpropanoic acid obtained by ozonolytic cleavage followed by Jones oxidation.

(E)-2-Methyl-6-(2-phenylethyl)-4-decene (4e)



Oil. ¹H NMR (300 MHz, CDCl₃) δ 0.86 (t, *J* = 6.6 Hz, 3H), 0.89 (d, *J* = 6.6 Hz, 3H), 0.90 (d, *J* = 6.6 Hz, 3H), 1.13–1.39 (m, 6H), 1.41–1.72 (m, 3H), 1.89–1.94 (m, 3H), 2.51 (ddd, *J* = 13.5, 10.2, 6.6 Hz, 1H), 2.64 (ddd, *J* = 13.5, 10.2, 5.2 Hz, 1H), 5.13 (dd, *J* = 15.0, 9.0 Hz, 1H), 5.36 (dt, *J* = 15.0, 7.2 Hz, 1H), 7.13–7.18 (m, 3H), 7.24–7.30 (m, 2H). ¹³C NMR (75.4 MHz, CDCl₃) δ 14.00, 22.19, 22.22, 22.66, 28.45, 29.39, 33.66, 35.22, 37.40, 42.03, 42.59, 125.55, 128.29, 128.48, 129.59, 135.84, 143.28. Anal. Calcd for C₁₉H₃₀: C, 88.30; H, 11.70%. Found: C, 88.01; H, 11.85%.

3-(2-Phenylethyl)-1-hexene (4f)



Oil. ¹H NMR (300 MHz, CDCl₃) δ 0.86 (t, *J* = 6.3 Hz, 3H), 1.24–1.37 (m, 4H), 1.57 (m, 1H), 1.68 (m, 1H), 2.00 (m, 1H), 2.51 (ddd, *J* = 13.8, 10.2, 6.3 Hz, 1H), 2.65 (ddd, *J* = 13.8, 10.2, 5.4 Hz,

1H), 4.98 (dd, J = 16.8, 2.1 Hz, 1H), 5.02 (dd, J = 10.2, 2.1 Hz, 1H), 5.57 (ddd, J = 16.8, 10.2, 9.0 Hz, 1H), 7.16–7.18 (m, 3H), 7.25–7.30 (m, 2H). ¹³C NMR (75.4 MHz, CDCl₃) δ 14.03, 20.08, 33.45, 36.76, 37.20, 43.47, 114.66, 125.63, 128.32, 128.48, 143.02, 143.20. Anal. Calcd for C₁₄H₂₀: C, 89.29; H, 10.71%. Found: C, 89.10; H, 10.83%.

3,7-Dimethyl-3-(2-phenyletnyl)-1,6-octadiene (4g)



Oil. ¹H NMR (300 MHz, CDCl₃) δ 1.05 (s, 3H), 1.25–1.38 (m, 2H), 1.54–1.62 (m, 2H), 1.59 (s, 3H), 1.68 (s, 3H), 1.87–1.95 (m, 2H), 2.48–2.54 (m, 2H), 4.96 (dd, J = 17.7, 1.5 Hz, 1H), 5.06 (dd, J = 11.1, 1.5 Hz, 1H), 5.10 (m, 1H), 5.76 (dd, J = 17.7, 1.1 Hz, 1H), 7.15–7.19 (m, 3H), 7.25–7.30 (m, 2H). ¹³C NMR (75.4 MHz, CDCl₃) δ 17.49, 22.41, 22.69, 25.61, 30.63, 39.56, 40.65, 42.88, 112.06, 124.99, 125.63, 128.40 (× 2C), 131.27, 143.46, 146.97. HRMS–EI (*m*/*z*): [M]⁺ calcd for C₁₈H₂₆, 242.20345; found, 242.20325.

(*S*, *E*)-4-(2-Phenylethyl)-2-octene (4h)



Oil. ¹H NMR (300 MHz, CDCl₃) δ 0.87 (d, J = 6.9 Hz, 3H), 1.18–1.35 (m, 6H), 1.53 (m, 1H), 1.69 (dd, J = 6.3, 1.5 Hz, 3H), 1.70 (m, 1H), 1.90 (m, 1H), 2.48 (ddd, J = 13.8, 10.2, 6.6 Hz, 1H), 2.63 (ddd, J = 13.8, 10.2, 5.4 Hz, 1H), 5.18 (ddq, J = 15.0, 8.7, 1.5 Hz, 1H), 5.38 (dq, J = 15.0, 6.3 Hz, 1H), 7.14–7.18 (m, 3H), 7.23–7.29 (m, 2H). ¹³C NMR (75.4 MHz, CDCl₃) δ 14.01, 17.89, 22.74, 29.36, 33.59, 35.19, 37.24, 42.49, 125.01, 125.55, 128.29, 128.49, 135.91, 143.25. Anal. Calcd for C₁₆H₂₄: C, 88.82; H, 11.18%. Found: C, 88.54; H, 11.17%. [α]_D²⁴ +1.0 (c 0.8, CHCl₃). The ee value of (S)-(E)-**4h** was determined by chiral HPLC (CHIRALCEL® OD-3 column, 4.6 mm × 250 mm, Daisel Chemical Industries, hexane, 0.5 mL/min, 40 °C, 220 nm UV detector, retention time = 8.57 min for the *S* isomer and 8.98 min for the *R* isomer). The (S) absolute configuration of **4h** was determined by optical rotation of 2-(2-phenylethyl)-1-hexanol obtained in two steps from (S)-(E)-**4h**. (S)-(E)-**4h** was converted to 2-(2-phenylethyl)-1-hexanol (46% ee): [α]_D²⁵ +0.4 (c 1.1, CHCl₃) [Lit⁶, (R) isomer, [α]_D =-0.5 (c 3.2, CHCl₃)].

Diethyl trans-2-(2-Phenylethyl)-3-cyclopentenyl Phosphate (4i)



Oil. ¹H NMR (300 MHz, CDCl₃) δ 1.32 (td, J = 7.2, 0.9 Hz, 3H), 1.33 (td, J = 7.2, 0.9 Hz, 3H), 1.62–1.80 (m, 2H), 2.52 (m, 1H), 2.69 (t, J = 7.8 Hz, 2H), 2.76–2.83 (m, 2H), 4.09 (quint., J = 7.2, 2.4 Hz, 4H), 4.75 (septet, J = 3.3 Hz, 1H), 5.67–5.74 (m, 2H), 7.18–7.30 (m, 5H). ¹³C NMR (75.4 MHz, CDCl₃) δ 15.88, 15.98, 33.50, 34.36, 39.63 (d, J = 3.9 Hz), 52.41 (d, J = 6.3 Hz), 63.51, 63.60, 82.92 (d, J = 6.3 Hz), 125.84, 127.58, 128.36, 128.38, 132.31, 141.99. HRMS–ESI (m/z): [M+Na]⁺ calcd for C₁₇H₂₅O₄PNa, 347.13827; found, 347.13850. The *anti* stereochemistry of **4i** was assigned on the basis of ¹H NMR of *trans*-2-(2-phenylethyl)-3-cyclopentenol⁷ obtained by dephosphorylation with LiAlH₄. The *cis* isomer was not detected by ¹H NMR.

(1*S*,5*S*)-1-Triisopropylsiloxy-5-[(4-methoxycarbonyl-3,3-dimethyl)butyl]-3-cyclopentene (4j)



Oil. ¹H NMR (300 MHz, CDCl₃) δ 0.97 (s, 6H), 1.05–1.08 (m, 21H), 1.15–1.57 (m, 4H), 2.19 (s, 2H), 2.27 (m, 1H), 2.49 (m, 1H), 2.62 (m, 1H), 3.64 (s, 3H), 4.15 (dt, *J* = 6.3, 3.3 Hz, 1H), 5.62–5.69 (m, 2H). ¹³C NMR (75.4 MHz, CDCl₃) δ 12.10, 17.93, 27.07, 27.12, 27.64, 33.04, 39.99, 42.30, 45.64, 51.05, 55.45, 78.30, 127.87, 132.93, 172.94. Anal. Calcd for C₂₂H₄₂O₃Si: C, 69.05; H, 11.06%. Found: C, 68.99; H, 10.93%. [α]_D²⁴ +82.1 (*c* 1.00, CHCl₃). The de value of (1*S*,5*S*)-**4j** was determined by chiral HPLC analysis of the *p*-nitrobenzoate derivative obtained by desilylation followed by benzoylation from (1*S*,5*S*)-**4j**. HPLC analysis (CHIRALCEL[®] OD-3 column, 4.6 mm × 250 mm, Daisel Chemical Industries, hexane/2-propanol = 99:1, 0.5 mL/min, 40 °C, 254 nm UV detector, retention time = 28.4 min for the *cis* isomer and 36.7 min for the *trans* isomer) revealed that the diastereomeric excess of the *p*-nitrobenzoate derivative was 94%. The *anti* stereochemistry of **4j** was determined by comparison of the couping constants in the ¹H NMR for the corresponding alcohol (obtained by desilylation) with that for *trans*-2-(2-phenylethyl)-3-cyclopentenol.⁷ The alcohol from **4j**: δ 4.11 (CHOH, dt, *J* = 6.0, 3.0 Hz).

4,4-Dimethyl-3-(2-phenylethyl)-1-cyclohexene (4l)



Oil. ¹H NMR (300 MHz, CDCl₃) δ 0.76 (s, 3H), 0.88 (s, 3H), 1.23–1.41 (m, 3H), 1.74 (m, 1H), 1.83 (m, 1H), 1.98–2.03 (m, 2H), 2.51 (ddd, *J* = 13.5, 10.5, 6.3 Hz, 1H), 2.84 (ddd, *J* = 13.5, 10.5, 4.8 Hz, 1H), 5.58–5.78 (m, 2H), 7.15–7.21 (m, 3H), 7.25–7.31 (m, 2H). ¹³C NMR (75.4 MHz, CDCl₃) δ 21.48, 22.96, 28.76, 31.55, 32.21, 34.19, 35.87, 44.57, 125.72, 126.01, 128.37, 128.51, 129.50, 143.02. HRMS–EI (*m*/*z*): [M]⁺ calcd for C₁₆H₂₂, 214.17215; found, 214.17219. The

regioselectivity of **4** was determined on the basis of ¹³C NMR and DEPT experiments. The chemical shift of the quaternary homoallylic carbon is diagnostic of the regiochemitry. The corresponding ¹³C resonance appeared at higher field (δ 31.55) than that expected for the quaternary allylic carbon.

Steroid 5



White Solid. ¹H NMR (300 MHz, CDCl₃) δ 0.68 (s, 3H), 0.87–1.23 (m, 36H), 1.43–1.60 (m, 8H), 1.79–2.05 (m, 4H), 2.03 (s, 3H), 2.22–2.32 (m, 3H), 2.59–2.64 (m, 2H), 4.05 (dt, J = 6.3, 3.3 Hz, 1H), 4.58 (m, 1H), 5.38 (m, 1H), 5.63 (dq, J = 6.0, 1.5 Hz, 1H), 5.73 (dq, J = 6.0, 1.8 Hz, 1H). ¹³C NMR (75.4 MHz, CDCl₃) δ 11.76, 12.10, 17.93, 18.31, 19.19, 20.90, 21.33, 24.11, 27.64, 28.30, 31.71, 31.77, 34.35, 36.48, 36.88, 38.01, 39.65, 40.38, 42.04, 42.33, 49.92, 52.25, 56.66, 56.89, 73.94, 79.36, 122.67, 127.74, 132.98, 139.74, 170.71. HRMS–ESI (*m*/*z*): [M+Na]⁺ calcd for C₃₈H₆₄O₃SiNa, 619.45169; found, 619.45145. [α]_D²⁴ +33.6 (*c* 2.9, CHCl₃). The *anti* stereochemistry of **5** was determined by comparing the couping constants in the ¹H NMR of the corresponding alcohol (obtained by desilylation) with that of *trans*-2-(2-phenylethyl)-3-cyclopentenol.⁷ The alcohol derived from **5**: δ 4.01 (CHOH, dt, J = 6.0, 3.0 Hz). The *cis* isomer was not detected by ¹H NMR.

NMR Studies (Figures S1a–f, Table S1)

The mixture of $(9\text{-BBN-H})_2$ (122.4 mg, 0.5 mmol) and styrene (109.4 μ 1, 1.05 mmol) in THF- d_8 (0.4 mL) was stirred at 60 °C for 1 hour to prepare alkylborane **2a** [δ 65.7 ppm (¹¹B), Figure S1a. See also Figure S1d for the ¹H NMR spectrum].⁸ Next, *t*-BuOK (1 M in THF, 1.0 ml, 1.0 mmol) was added at 25 °C, and the mixture was stirred at 25 °C for 5 min. The ¹¹B NMR spectrum of the mixture showed a peak corresponding to a tetravalent borate (δ –1.4 ppm) (Figure S1b. See also Figure S1e for the ¹H NMR spectrum).⁹ Subsequently, CuOAc (121.6 mg, 1.0 mmol, B/Cu 1:1) was added to the borate solution, and the solution was heated at 60 °C for 1 h: the peak of the borate disappered completely and a signal that corresponds to 9-BBN-O'Bu¹⁰ appeared at δ 55.1 ppm as a major peak in the ¹¹B NMR spectrum (Figure S1c). Meanwhile, the formation of styrene and ethylbenzene were observed by ¹H NMR spectroscopy (4% and 13% NMR yields, respectively. Dibenzyl was used as an internal standard) (Figure S1f). These compounds seem to be produced by β -hydride elimination and protonation of an alkylcopper species. The signals for styrene and ethylbenzene did not increase furthermore with prolonged heating (60 °C, 20 h).

Figure S1a. [¹¹B NMR spectrum (192.6 MHz, THF- d_8)]



Figure S1b. [¹¹B NMR spectrum (192.6 MHz, THF- d_8)]



Figure S1c. [¹¹B NMR spectrum (192.6 MHz, THF- d_8)]



Figure S1d. [¹H NMR spectrum (300 MHz, THF- d_8)]



Figure S1e. [¹H NMR spectrum (300 MHz, THF-*d*₈)]



Figure S1f. [¹H NMR spectrum (300 MHz, THF- d_8)]



Table S1. Summary of the ¹¹B NMR Data for the Organoboron Compounds

this work	literature	
βB_{Ph} $\delta 65.7 \text{ ppm (THF-}d_8)$	Soderquist, J. A.; Kock, I.; Estrella, M. E. Org. Process Res. Dev. 2006, 10, 1076–1079.	$\beta 72.2 \text{ ppm} (C_6 D_6)$
$\overset{K^{+}}{\overbrace{B}^{-O^{*}Bu}}_{Ph}$ $\delta - 1.4 \text{ ppm (THF-}d_8)$	Köster, R.; Seidel, G.; Wagner, K.; Wrackmeyer, B. Chem. Ber. 1993 , <i>126</i> , 305–317.	$\delta - 1.5 \text{ ppm (THF-}d_8)$
$\frac{1}{\delta 55.1 \text{ ppm (THF-}d_8)}$	Brown, H. C.; Cha, J. S.; Nazer, B. J. Org. Chem. 1985 , 50, 549–553.	δ 55.5 ppm (neat)

References

- (1) Bruke, E, -J. Tetrahedron 1979, 35, 781–788.
- (2) Sato, F.; Ishikawa, H.; Sato, M. Tetarahedron Lett. 1981, 22, 85–88.
- (3) Matsumura, K.; Hashiguchi, S.; Ikariya, T.; Noyori, R. J. Am. Chem. Soc. 1997, 119, 8738–8739.
- (4) Lysenko, I. L.; Kim, K.; Lee, H. G.; Cha, J. K. J. Am. Chem. Soc. 2008, 130, 15997–16002.
- (5) Langlois, J.-B.; Alexakis, A. Chem. Comm. 2009, 3868–3870.
- (6) Spino, C.; Gund, V. G.; Nadeau, C. J. Comb. Chem. 2005, 7, 345–352.
- (7) (a) Partridge, J. J.; Chadha, N. K.; Uskoković, M. R. J. Am. Chem. Soc. 1973, 95, 532–540. (b) Danheiser, R. L.; Martinez-Davila, C.; Auchus, R. J.; Kadonaga, J. T. J. Am. Chem. Soc. 1981, 103, 2443–2446.
- (8) Soderquist, J. A.; Kock, I.; Estrella, M. E. Org. Process Res. Dev. 2006, 10, 1076-1079.
- (9) (a) Köster, R.; Seidel, G.; Wagner, K.; Wrackmeyer, B. Chem. Ber. 1993, 126, 305–317. (b)
 Fry, A.; Vishwakarma, L. C. J. Org. Chem. 1980, 45, 5306–5308.
- (10) Brown, H. C.; Cha, J. S.; Nazer, B. J. Org. Chem. 1985, 50, 549–553.











Original File: Date Nov 5 09 Comment Nov 5 09 Comment Statkard Observe Stick-come Tune-6.4 Match=0.4 Obsfue 192 Obsfue 192 Obsfue 996.3672 Hz Point 32768 Frequecy(Span) 18761.73 Hz Scan 192 AcqTime 1.4992 s Pf 1.501 s Pulsel 6.0 μ s Temperature 29.0 C Solvent CDC13 Reference 77.0 pm Broad.Factor 0.2863 Hz RGain 90.0 Dec/21 20:35:19 Operator		Bu OP(O)(OEt) ₂ OTBDMS 3b
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Scan	512
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PD	1.501 s
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Temperature	29.0 °C
Solvent	CDC 13
Reference	77.0 ppm
Broad.Factor	0.25 Hz
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