

Synthesis of Mono-ADP-Ribosylated Oligopeptides using Ribosylated Amino Acids Building Blocks

Gerbrand J. van der Heden van Noort, Maarten G. van der Horst, Herman S. Overkleef,

Gijsbert A. van der Marel, Dmitri V. Filippov*

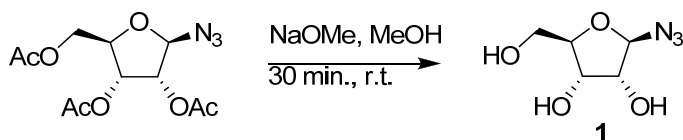
*Leiden Institute of Chemistry, Leiden University, PO Box 9502, 2300 RA
Leiden, The Netherlands*

filippov@chem.leidenuniv.nl

Experimental Section

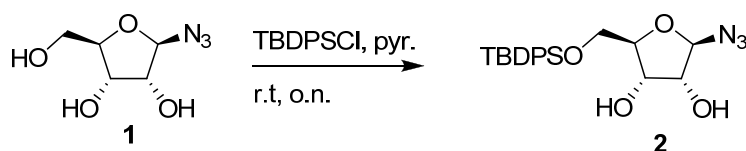
General procedures

Petroleum Ether (PE) with a boiling range of 40 - 60°C was used. THF and Et₂O were distilled over LiAlH₄ prior to use. DCM was distilled over CaH₂ prior to use. All other solvents used under anhydrous conditions were stored over 4Å molecular sieves except for methanol which was stored over 3Å molecular sieves. Solvents used for workup and column chromatography were of technical grade and distilled before use. Unless stated otherwise, solvents were removed by rotary evaporation under reduced pressure at 40°C. Reactions were monitored by TLC-analysis using Merck 25 DC plastikfolien 60 F254 with detection by spraying with 20% H₂SO₄ in EtOH, (NH₄)₆Mo₇O₂₄·4H₂O (25 g/L) and (NH₄)₄Ce(SO₄)₄·2H₂O (10 g/L) in 10% sulfuric acid or by spraying with a solution of ninhydrin (3 g/L) in EtOH / AcOH (20/1 v/v), followed by charring at approx. 150°C. Column chromatography was performed on Fluka silicagel (0.04 – 0.063 mm). For LC-MS analysis was used, a JASCO HPLC-system (detection simultaneously at 214 and 254 nm) equipped with an analytical C18 column (4.6 mmD × 50 mmL, 3μ particle size) in combination with buffers A: H₂O, B: MeCN and C: 0.5% aq. TFA and coupled to a PE/SCIEX API 165 single quadrupole mass spectrometer (Perkin-Elmer), unless stated otherwise. Alternatively a Thermo Finnigan LCQ Advantage MAX ion-trap mass spectrometer with an electrospray ion source coupled to Surveyor HPLC system (Thermo Finnigan) was used with the same analytical column. For reversed-phase HPLC purification of the final compounds, an automated HPLC system supplied with a C18 column (10.0 mmD × 250 mmL, 5μ particle size) was used. High resolution mass spectra spectra (except for the compounds **18**, **21** and **25**) were recorded by direct injection (2 μL of a 2 μM solution in water/acetonitrile; 50/50; v/v and 0.1% formic acid) on a mass spectrometer (Thermo Finnigan LTQ Orbitrap) equipped with an electrospray ion source in positive mode (source voltage 3.5 kV, sheath gas flow 10, capillary temperature 250 °C) with resolution R = 60000 at m/z 400 (mass range m/z = 150-2000) and dioctylphthalate (m/z = 391.2842) as a "lock mass". For the peptide derivatives **18**, **21** and **25** high resolution mass spectra were obtained from LC-MS analysis on Thermo Finnigan LTQ Orbitrap mass spectrometer equipped with an analytical C18 column (4.6 mmD × 50 mmL, 3μ particle size). The high resolution mass spectrometer was calibrated prior to measurements with a calibration mixture (Thermo Finnigan). ¹H- and ¹³C-NMR spectra were measured on a Brüker AV-400 (400 MHz), a Brüker AV-500 (500 MHz) or a Brüker DMX-600 (600 MHz). Chemical shifts are given in ppm (δ) relative to TMS (0 ppm) and coupling constants are given in Hz. Optical rotations are measured in CHCl₃ at a concentration of 10 mg/mL at 25 °C.



β -D-ribofuranosyl azide (1)

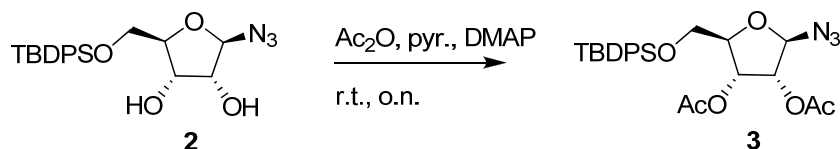
2,3,5-tri-*O*-acetyl- β -D-ribofuranosyl azide (2.60 g, 8.62 mmol), was dissolved in MeOH (40 mL) and NaOMe (3.3 eq., 1.54 g, 28.5 mmol) was added. After 30 min. TLC analysis (2 % MeOH/ DCM) indicated complete conversion. Amberlite- H^+ was added until pH = 6. Filtration and concentration *in vacuo* yielded the title compound as a colourless oil (1.27 g, 7.27 mmol, 85%). ^{13}C -NMR (100 MHz, MeOD-*d*4) δ : 96.2 (C1'), 85.2 (C4'), 76.2 (C2'), 71.8 (C3'), 63.8 (C5'). ^1H -NMR (400 MHz, MeOD-*d*4) δ : 5.17 (s, 1H, H1', $J_{1,2} = 1.3$ Hz), 4.85 (bs, 3H, OH), 4.03 (dd, 1H, H3', $J_{2,3} = 4.5$ Hz, $J_{3,4} = 6.6$ Hz), 3.93 (m, 1H, H4', $J_{4,5A} = 3.2$ Hz, $J_{4,5B} = 5.7$ Hz), 3.79 (dd, 1H, H2'), 3.71, 3.57 (AB, 2H, H5', $J_{5A,5B} = 12.1$ Hz). $[\alpha_D \text{CHCl}_3] = -195.9^\circ$. IR: 3330.3, 2113.8, 1706.1, 1234.7, 1019.0, 925.9.



5-*tert*-Butyldiphenylsilyl- β -D-ribofuranosyl azide (2)

β -D-ribofuranosyl azide **1** (0.95 g, 3.15 mmol) was used without further purification and after coevaporation with pyridine dissolved in pyridine (40 mL). TBDPSCI (1.1 eq., 3.46 mmol, 0.9 mL) was added. The reaction mixture was stirred overnight at room temperature. After concentration *in vacuo* the residue was taken up in DCM and washed with 5 % citric acid, dried (MgSO_4) and concentrated. Column chromatography yielded the title compound (1.46 g, 3.22 mmol, quantitative) as a colourless oil.

^{13}C -NMR (100 MHz, CDCl_3) δ : 135.6 (Arom.), 132.8 (Cq Arom.), 129.91, 129.87, 127.84, 127.80 (Arom.), 94.7 (C1'), 83.5 (C2'), 75.5 (C4'), 71.6 (C3'), 63.8 (C5'), 26.8 (CH_3 t-Bu), 19.2 (Cq t-Bu). ^1H -NMR (400 MHz, CDCl_3) δ : 7.73 – 7.67 (m, 4H, Arom.), 7.75 – 7.36 (m, 6H, Arom.), 5.31 (d, 1H, H1', $J_{1,2} = 1.9$ Hz), 4.37 (dd, 1H, H3', $J_{2,3} = 4.8$ Hz, $J_{3,4} = 6.3$ Hz), 4.05 (m, 1H, H4', $J_{4,5A} = 3.2$, $J_{4,5B} = 4.5$), 3.98 (dd, 1H, H2'), 3.88 – 3.81 (AB, 2H, H5', $J_{5A,5B} = 11.1$), 1.08 (s, 9H, t-Bu). $[\alpha_D \text{CHCl}_3] = -61.4^\circ$. IR: 3314.1, 2931.6, 2858.7, 2110.3, 1427.8, 1234.2, 1111.6, 988.8, 699.1, 501.6. HRMS $[\text{C}_{21}\text{H}_{27}\text{N}_3\text{O}_4\text{Si} + \text{Na}]^+$: 436.1661 found, 436.1663 calc.

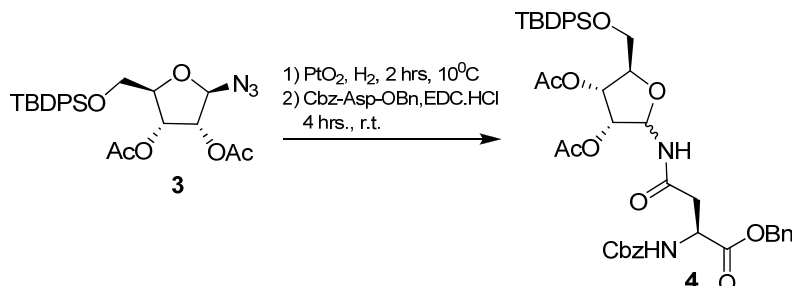


5-*tert*-Butyldiphenylsilyl-2,3-di-*O*-acetyl- β -D-ribofuranosyl azide (3)

Ribofuranosyl azide **2** (4.13, 10 mmol) was dissolved in pyridine (20 mL) and a catalytic amount of DMAP and acetic anhydride (5 eq., 50 mmol, 4.69 mL) were added. The reaction mixture was stirred overnight at room temperature. After concentration the residue was taken up in DCM and washed with 5 % citric acid, dried (MgSO_4) and purified using silica gel column chromatography (PE: EA, 100/0 – 50/50). The title compound (4.88 g, 9.8 mmol, 98%) was obtained as a colourless oil.

^{13}C -NMR (100 MHz, CDCl_3) δ : 169.4, 169.3 (CO OAc), 135.54, 134.50 (Arom.), 132.7 (Cq Arom.), 129.74, 129.67, 127.68, 127.66 (Arom.), 92.6 (C1'), 82.4 (C4'), 74.6 (C2'), 70.7 (C3'), 63.2 (C5'), 26.6 (t-Bu), 20.39, 20.37 (CH_3 OAc), 19.0 (Cq t-Bu). ^1H -NMR (400 MHz, CDCl_3) δ : 7.72 – 7.68 (m, 4H, arom.), 7.41 – 7.36 (m, 6H, arom.), 5.51 (t, 1H, H3', $J_{2,3} = 4.9$ Hz, $J_{3,4} = 5.3$), 5.41 (d, 1H, H1', $J_{1,2} = 3.0$ Hz), 5.21 (dd, 1H, H2'), 4.21 (m, 1H, H4', $J_{4,5A} = 3.5$ Hz, $J_{4,5B} = 3.6$ Hz), 3.88 – 3.71 (AB, 2H, H5', $J_{5A,5B} = 11.4$ Hz), 2.09, 2.03 (2x s, 6H, CH_3 OAc), 1.08 (s,

9H, t-Bu). $[\alpha_D \text{ CHCl}_3] = -53.8^\circ$. IR: 2933.1, 2858.7, 2112.2, 1751.8, 1214.1, 1230.2, 1109.6, 701.6, 502.0. HRMS $[\text{C}_{25}\text{H}_{31}\text{N}_3\text{O}_6\text{Si} + \text{Na}]^+$: 520.1866 found, 520.1874 calc.

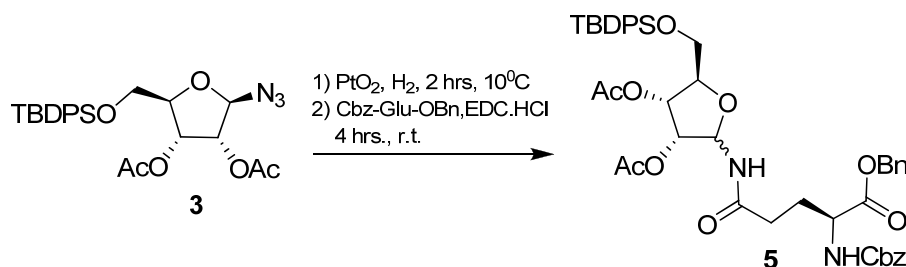


***N'*-(5-*tert*-Butyldiphenylsilyl-2,3-di-*O*-acetyl- α -D-ribosyl)-*N''*-benzyloxycarbonyl asparagine benzyl ester (4)**

Platinum(IV)oxide (110 mg, 0.48 mmol) was added to a solution of azide **3** (1.24 g, 2.5 mmol) in EtOAc (20 mL) and purged with argon before a hydrogen gas was bubbled through the reaction mixture for 2 hours at 10 °C, upon which TLC analysis (3:2, v/v, EA/PE) showed complete disappearance of the starting material. The suspension was filtered through a pad of celite and concentrated. The residue was dissolved in DCM (30 mL) and EDC.HCl (0.53 g, 2.8 mmol) and Z-Asp-OBn (0.89 g, 2.5 mmol) were added under an argon atmosphere. After stirring for 4 hours the reaction mixture was concentrated. The residue was taken up in DCM and washed with sat. aq. NaHCO_3 and sat. aq. NaCl. The organic layer was dried (MgSO_4), concentrated and the stereo-isomers were separated using silica gel column chromatography (PE: EA, 80/20 – 50/50). The title compound was obtained as a white foam (0.65 g, 0.80 mmol, 32 %) along with the β anomer (0.31 g, 0.38 mmol, 15%).

α -anomer: ^{13}C -NMR (100 MHz, CDCl_3) δ : 170.6, 169.5 (CO α -Asn, CO γ -Asn), 169.2, 169.0 (CO OAc), 158.2 (CO Cbz), 135.8 (Cq Arom.), 135.4 (Cq Arom.), 135.5 (Arom.), 132.6, 132.3 (Cq Arom.), 129.7, 128.4 – 127.7 (Arom.), 82.0 ($\text{C4}'$), 78.5 ($\text{C1}'$), 72.3 ($\text{C3}'$), 70.0 ($\text{C2}'$), 67.42, 66.9 (CH_2 Bn, CH_2 Cbz), 63.5 ($\text{C5}'$), 50.7 (CH α -Asn), 38.0 (CH_2 β -Asn), 26.6 (CH_3 , t-Bu), 20.6, 20.2 (CH_3 , OAc), 19.0 (Cq, t-Bu). ^1H -NMR (400 MHz, $\text{MeOD-}d_4$) δ : 7.89 (1H, NH), 7.72 – 7.68, 7.44 – 7.26 (3 x m, 20H, Arom.), 6.07 (d, 1H, $\text{H1}'$), 5.56 (dd, 1H, $\text{H3}'$), 5.46 (dd, 1H, $\text{H2}'$), 5.16, 5.08 (2 x m, 4H, CH_2 Bn, CH_2 Cbz), 4.68 (dd, 1H, $\text{H4}'$), 4.16 (m, 1H, CH α -Asn), 3.76 (ddd, 2H, $\text{H5}'$), 2.88 (d, 2H, CH_2 β -Asn), 2.09, 2.06 (2 x s, 6H, CH_3 OAc), 1.07 (s, 9H, t-Bu). α_D (CHCl_3) = + 35.8°. IR (cm^{-1}): 503.0, 698.4, 1112.8, 1211.9, 1505.7, 1733.7, 1747.7. LC-MS (50 - 90 % B in 15 min.) $\text{Rt} = 9.70$. HRMS $[\text{C}_{44}\text{H}_{50}\text{N}_2\text{O}_{11}\text{Si} + \text{H}]^+$: 811.3263 found, 811.3211 calc.

β -anomer: ^{13}C -NMR (100 MHz, CDCl_3) δ : 170.6, 170.1 (CO α -Asn, CO γ -Asn), 169.8, 169.7 (CO OAc), 156.0 (CO Cbz), 135.4 (Cq Arom.), 135.3 (Arom.), 133.1, 132.0 (Cq Arom.), 130.0, 128.4 – 127.7 (Arom.), 82.0, 81.9 ($\text{C4}'$, $\text{C1}'$), 74.1 ($\text{C2}'$), 71.2 ($\text{C3}'$), 67.2, 66.8 (CH_2 Bn, CH_2 Cbz), 63.3 ($\text{C5}'$), 50.2 (CH α -Asn), 37.1 (CH_2 β -Asn), 26.8 (CH_3 , t-Bu), 20.5, 20.4 (CH_3 OAc), 19.3 (Cq, t-Bu). ^1H -NMR (400 MHz, $\text{MeOD-}d_4$) δ : 7.88(s, 1H, NH Asn), 7.71 – 7.69, 7.43 – 7.40, 7.32 – 7.30 (3 x m, 20H, Arom.), 5.63 (d, 1H, $\text{H1}'$), 5.54 (m, 1H, $\text{H3}'$), 5.18 (t, 1H, $\text{H2}'$), 5.14, 5.07 (2 x s, 4H, CH_2 Bn, CH_2 Cbz), 4.64 (m, 1H, $\text{H4}'$), 4.08 (m, 1H, CH α -Asn), 3.80 (m, 2H, $\text{H5}'$), 2.77 – 2.71 (AB, 2H, CH_2 β -Asn), 2.07, 2.03 (2 x s, 6H, CH_3 OAc), 1.07 (s, 9H, t-Bu). α_D (CHCl_3) = +12.4°. IR: 503.3, 698.0, 1112.3, 1214.5, 1498.7, 1747.2. LC-MS (50 - 90 % B in 15 min.) $\text{Rt} = 9.63$. HRMS $[\text{C}_{44}\text{H}_{50}\text{N}_2\text{O}_{11}\text{Si} + \text{H}]^+$: 811.3262 found, 811.3211 calc.



***N*^δ-(5-*tert*-Butyldiphenylsilyl-2,3-di-*O*-acetyl- α -D-ribose)- *N* ^{α} -benzyloxycarbonyl glutamine benzyl ester (5)**

Platinum(IV)oxide (440 mg, 88 μmol) was added to a solution of azide **3** (5 g, 9.6 mmol) in EtOAc (100 mL) and purged with argon before hydrogen gas was bubbled through the reaction mixture for 2 hours at 10 $^\circ\text{C}$, upon which TLC analysis (3:2, v/v, EA/PE) showed complete disappearance of the starting material. The suspension was filtered through a pad of celite and concentrated. The residue was dissolved in DCM (100 mL) and EDC.HCl (85 mg, 9.4 mmol) and Z-Glu-OBn (3.0 g, 8.1 mmol) were added under an argon atmosphere. After stirring for 24 hours the reaction mixture was concentrated. The residue was taken up in DCM and washed with 1N HCl and sat. aq. NaCl. The organic layer was dried (MgSO_4), concentrated and the stereo-isomers were separated using silica gel column chromatography (PE: EA, 100/0 – 65/35). The title compound was obtained as a white foam (3.0 g, 3.7 mmol, 46 %) along with the β anomer (1.0 g, 1.2 mmol, 15%).

α -anomer: $^{13}\text{C-NMR}$ (CDCl_3) δ : 171.8, 171.5 (CO α -Gln, CO δ -Gln), 169.3, 169.0 (CO Ac), 156.0 (CO Cbz), 135.9, 135.4 (Cq Arom.), 135.5 (Arom.), 132.7, 132.4 (Cq Arom.), 129.6, 128.4 – 127.6 (Arom.), 81.6 (C4'), 78.6 (C1'), 72.2 (C3'), 70.1 (C2'), 67.1, 66.8 (CH_2 Bn, CH_2 Cbz), 63.5 (C5'), 53.4 (CH α -Gln), 32.4 (CH_2 β -Gln), 28.1 (CH_2 γ -Gln), 26.6 (t-Bu), 20.5, 20.3 (CH_3 OAc), 218.9 (Cq t-Bu). $^1\text{H-NMR}$ (CDCl_3) δ : 7.69 – 7.66, 7.42 – 7.37, 7.37 – 7.27 (3 x m, 20H, Arom.), 6.72 (d, 1H, NH), 6.10 (dd, 1H, H1'), 5.82 (d, 1H, NH), 5.58 (t, 1H, H3'), 5.49 (t, 1H, H2'), 5.14, 5.07 (2x s, 4H, CH_2 Bn, CH_2 Cbz), 4.42 (m, 1H, CH α -Gln), 4.10 (m, 1H, H4'), 3.71 (m, 2H, H5'), 2.33 (m, 2H, CH_2 β -Gln), 2.21 (m, 2H, CH_2 γ -Gln), 2.09, 2.07 (2x s, 6H, OAc), 1.07 (s, 9H, t-Bu). ESI-MS: m/z 825.20 [$\text{M}+\text{H}$]⁺. LC-MS (10 - 90 % B in 15 min), Rt = 9.94. IR (cm^{-1}): 3328, 2933, 2362, 1745, 1520, 1235, 1212, 1110, 1049, 741, 698, 604, 502. [$\alpha_{\text{D}}^{\text{CHCl}_3}$] = + 32.6 $^\circ$. HRMS [$\text{C}_{45}\text{H}_{52}\text{N}_2\text{O}_{11}\text{Si} + \text{H}$]⁺: 825.3419 found, 825.3413 calc.

β -anomer: $^{13}\text{C-NMR}$ (100 MHz, CDCl_3) δ : 172.0, 171.6 (CO Ac), 169.8, 169.7 (CO α -Gln, CO δ -Gln), 156.1 (CO Cbz), 136.0 (Cq Arom.), 135.5, 135.4 (Arom.), 135.1, 133.1, 132.2 (Cq Arom.), 130.0, 129.93, 128.5, 128.3, 128.2, 128.0, 127.9 (Arom.), 81.9 (C1', C4'), 74.0 (C2'), 71.4 (C3'), 67.2, 66.9 (CH_2 Bn, CH_2 Cbz), 63.5 (C5'), 53.3 (CH α -Gln), 31.8 (CH_2 β -Gln), 27.7 (CH_2 γ -Gln), 26.8 (t-Bu), 20.6, 20.4 (OAc), 19.3 (Cq t-Bu). $^1\text{H-NMR}$ (500 MHz, CDCl_3) δ : 7.68 – 7.65, 7.40 – 7.25 (2 x m, 20H, Arom.), 6.47, 6.46 (d, 1H, NH δ), 5.81 – 5.78 (dd, 1H, H1'), 5.65, 5.63 (d, 1H, NH α), 5.57 – 5.55 (dd, 1H, H3'), 5.32 – 5.30 (t, 1H, H2'), 5.12, 5.04 (2x s, 4H, CH_2 Bn), 4.31 – 4.27 (m, 1H, H α), 4.12 – 4.10 (m, 1H, H4'), 3.88 – 3.80 (AB, 2H, H5'), 2.08, 2.04 (2x s, 6H, OAc), 1.94 – 1.91 (m, 2H, H β), 2.13 – 2.10, 1.85 – 1.80 (2x m, 2H, H γ), 1.10 (s, 9H, t-Bu). $J_{1,2} = 5.7 \text{ Hz}$, $J_{2,3} = 5.3 \text{ Hz}$, $J_{3,4} = 3.4 \text{ Hz}$, $J_{1, \text{NH}} = 9.0$, $J_{4,5\text{A}} = 2.9 \text{ Hz}$, $J_{4,5\text{B}} = 1.9 \text{ Hz}$, $J_{5\text{A}, 5\text{B}} = 11.4 \text{ Hz}$. ESI-MS: m/z 825.20 [$\text{M}+\text{H}$]⁺. LC-MS (10 - 90 % B in 15 min), Rt = 9.88. IR (cm^{-1}): 3335.3, 2934.0, 1745.2, 1519.9, 1240.8, 1214.4, 1110.5, 1064.7, 738.9, 699.0, 502.2. [$\alpha_{\text{D}}^{\text{CHCl}_3}$] = - 0.8 $^\circ$. HRMS [$\text{C}_{45}\text{H}_{52}\text{N}_2\text{O}_{11}\text{Si} + \text{H}$]⁺: 825.3422 found, 825.3413 calc.

Both α -anomers of **4** and **5** showed the presence of a NOE cross-peak between H1' and H3' of ribose, whereas for the corresponding β -anomers a NOE cross-peak between H1' and H4' was observed (see Fig. 1.)

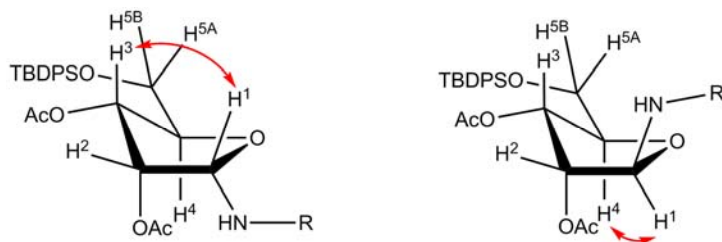
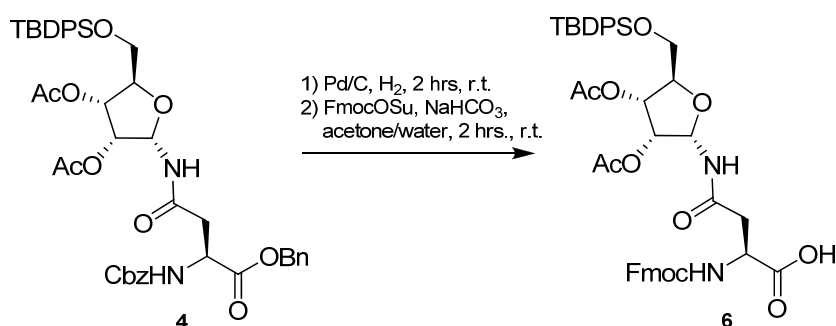
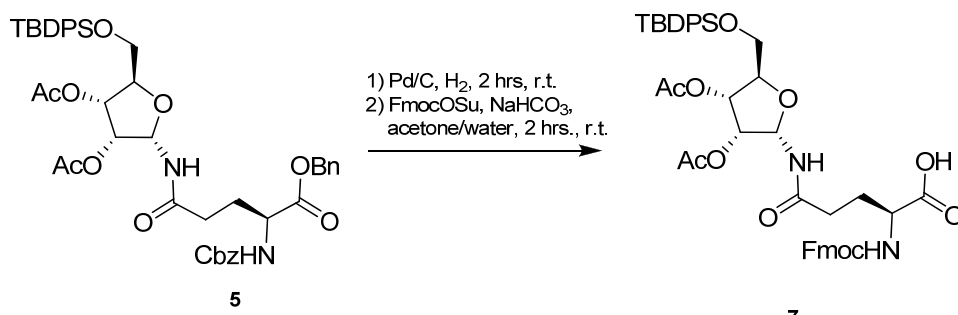


Fig. 1 NOESY observations



***N'*-(5- *tert*-Butyldiphenylsilyl-2, 3-di-*O*-acetyl- α -D-ribose)- *N'*-fluorenylmethoxycarbonyl asparagine (**6**)**

Amino acid **4a** (1.25 g, 1.54 mmol) was dissolved in MeOH (30 mL), Pd-C (10%, 190 mg) and a drop of conc. HCl were added after purging with argon. A flow of hydrogen gas was passed through the reaction mixture for 2 hours. LC-MS analysis (CN-Column, 10-90 % B in 26 min.) showed complete consumption of the starting material and formation of the unprotected amino acid. Next, NaHCO₃ (190 mg, 2.3 mmol) was added and the reaction mixture was filtered over a pad of celite. The solvent was removed *in vacuo* and the residue was dissolved in H₂O/acetone (40 mL, 1:1, v/v). Fmoc-OSu (0.51 g, 1.5 mmol) and NaHCO₃ (0.13 g, 1.5 mmol) were added after which the reaction mixture was stirred at room temperature for 2 hrs. The mixture was acidified to pH = 2 and acetone was removed under reduced pressure. The resulting solution was extracted with DCM, washed with citric acid 5% and H₂O, dried (MgSO₄) and concentrated. Purification using column chromatography (99.9 : 0.1 DCM/AcOH – 89.9/1/0.1 DCM/MeOH/AcOH) afforded the title compound as a white foam (1.1 g, 1.30 mmol, 84%). ¹³C-NMR (100 MHz, MeOD-*d*₄) δ : 173.2, 171.0 (CO γ -Asn, CO α -Asn), 170.2, 170.1 (CO OAc), 156.9 (CO Fmoc), 143.8, 143.8 (Cq Fmoc), 137.5 (Fmoc), 135.4, (Cq TBDPS), 129.7 – 119.6 (Arom. TBDPS, Fmoc), 81.2 (C4'), 79.2 (C1'), 71.5 (C3'), 70.7 (C2'), 66.8 (C5'), 63.5 (CH₂ Fmoc), 50.8 (CH α -Asn), 46.9 (CH Fmoc), 37.5 (CH₂ β -Asn), 26.1 (CH₃ *t*-Bu), 19.2, 19.1 (CH₃ OAc), 18.7 (Cq *t*-Bu). ¹H-NMR (400 MHz, MeOD-*d*₄) δ : 7.85 (s, 1H, NH), 7.71 – 7.58, 7.40 – 7.21 (m, 18H, Arom.), 6.06 (d, 1H, H1'), 5.51 (t, 1H, H3'), 5.43 (t, 1H, H2'), 4.42 (m, 1H, H α -Asn), 4.28 (m, 2H, CH₂ Fmoc), 4.16 (t, 1H, CH Fmoc), 4.07 (d, 1H, H4'), 3.65 (m, 2H, H5'), 2.87 – 2.74 (m, 2H, CH₂ β -Asn), 2.08, 2.05 (2 x s, 6H, CH₃ OAc), 1.01 (s, 9H, CH₃ *t*-Bu). α_D (CDCl₃) = +56.2°. LC-MS (10 – 90 % B in 15 min.), Rt = 11.34 min. IR: 503.5, 701.4, 1112.5, 1212.2, 1505.7, 1747.5. HRMS [C₄₄H₄₈N₂O₁₁Si + H]⁺: 809.3106 found, 809.3055 calc.



***N*^δ-(5-*tert*-Butyldiphenylsilyl-2,3-di-*O*-acetyl- α -D-ribose)-*N* ^{α} -fluorenylmethoxycarbonyl glutamine (7)**

Amino acid **5** (0.2 g, 0.24 mmol) was dissolved in MeOH (5 mL), Pd-C (10%) (30 mg) and a drop of conc. HCl were added after which hydrogen gas was passed through the reaction mixture for 5 hours. LC-MS analysis (CN-Column, 10-90 % B in 26 min.) showed complete consumption of the starting material and formation of the unprotected amino acid. After filtration over a pad of celite the solvent was concentrated and dissolved in H₂O/ acetone/ dioxane (3 mL, 1:1:2, v/v/v). Fmoc-OSu (80 mg, 0.24 mmol) and NaHCO₃ (20 mg, 0.24 mmol) were added after which the reaction mixture was stirred overnight. The mixture was washed with 5% citric acid and after drying (MgSO₄) the organic layer was concentrated. Purification using column chromatography (99.9 : 0.1 DCM/AcOH – 99.4 : 0.1 : 0.5 DCM/ AcOH/ MeOH) afforded the title compound as a white foam (0.19 g, 0.23 mmol, 97%).

¹³C-NMR (100 MHz, MeOD-*d*₄) δ : 173.9, 173.7 (CO δ -Gln, CO α -Gln), 170.2, 170.0 (CO OAc), 157.2 (CO Fmoc), 143.9, 143.8, 141.2 (Cq Arom.), 135.4, 135.3 (Arom.), 132.7, 132.6 (Cq Arom.), 129.7 - 119.6 (Arom. Fmoc/TBDPS), 81.1 (C4'), 79.2 (C1'), 71.5 (C3'), 70.7 (C2'), 66.7 (C5'), 63.4 (CH₂ Fmoc), 53.4 (CH α -Gln), 47.0 (CH Fmoc), 32.0 (CH₂ β -Gln), 27.3 (CH₂ γ -Gln), 26.0 (CH₃ *t*-Bu), 19.3, 19.1 (CH₃ OAc), 18.6 (Cq *t*-Bu). ¹H-NMR (400 MHz, MeOD-*d*₄) δ : 7.89 (d, 1H, NH), 7.79 – 7.65, 7.46 – 7.28 (2x m, 18H, Arom.), 6.10 (d, 1H, H1'), 5.58 (m, 1H, H3'), 5.49 (m, 1H, H3'), 4.38 – 4.16 (m, 5H, CH Fmoc, H5', H4', H α), 3.76 - 3.71 (m, 1H, CH₂ Fmoc), 2.45 (m, 2H, CH₂ β), 2.24, 2.02 (2x m, 2H, Hy), 2.11, 2.05 (CH₃ OAc), 1.06 (CH₃ *t*-Bu). ESI-MS: *m/z* 823.27 [M+H]⁺. LC-MS (50 - 90 % B in 15 min), Rt = 8.92. IR (cm⁻¹): 2933.1, 1742.5, 1519.6, 1212.6, 1108.1, 740.0, 702.8, 502.3. α_D (CHCl₃) = + 33.4°. HRMS [C₄₅H₅₀N₂O₁₁Si + H]⁺: 823.3263 found, 823.3256 calc.

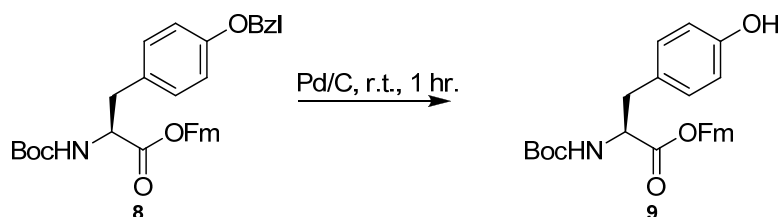


***N* ^{α} -*tert*-Butoxycarbonyl-4-*O*-benzyl-L-tyrosine fluorenylmethyl ester (8)**

Boc-Tyr(Bzl)-OH (2.0 mmol, 0.74 g) was coevaporated with MeCN and dissolved in DCM (40 mL) under an argon atmosphere. To this solution were added 9-fluorenylmethanol (2.2 mmol, 0.43 g), EDC·HCl (2.2 mmol, 0.42 g) and DMAP (0.2 mmol, 32 mg). The solution was stirred at room temperature for 2 hours, washed with 5% citric acid and H₂O. The combined organic layers were dried and concentrated *in vacuo*. The crude compound was purified by silica gel column chromatography (PE/EtOAc, 95/5) to give compound **8** as a white powder (0.84 g, 76%).

¹³C-NMR (100 MHz, CDCl₃) δ : 171.9 (CO α -Tyr), 157.8 (Cq Tyr.), 155.1 (CO Boc), 143.5, 143.4, 141.2 (Cq Fm), 136.9 (Cq Bn), 130.24 (Arom. Tyr.), 128.5 (Arom. Bn/Fm), 127.9 (Cq Tyr.), 127.3, 127.1 (Arom. Bn/Fm), 125.0, 120.0 (Arom. Fm), 114.9 (Arom. Tyr.), 79.8 (Cq Boc), 69.8 (CH₂ Bn), 67.0 (CH₂ Fm), 54.6 (CH α Tyr.), 46.6 (CH Fm), 37.4 (CH₂ β Tyr.), 28.3 (CH₃ Boc). ¹H-NMR (400 MHz, CDCl₃) δ : 7.72 (d, 2H, Arom. Bn/Fm), 7.48 (t, 2H, Arom. Bn/Fm), 7.35 – 7.26 (m, 8H, Arom. Bn/Fm), 6.98, 6.84 (2x d, 4H, Arom. Tyr.), 5.06 (d, 1H,

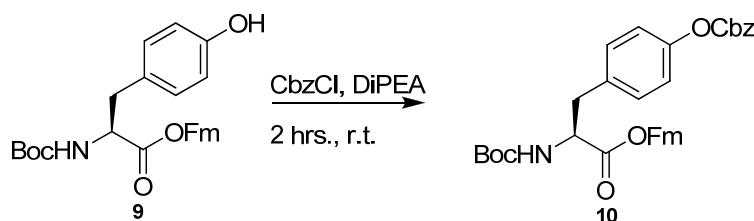
NH α -Tyr), 4.91 (s, 2H, CH₂ Bn), 4.60 (m, 1H, CH α -Tyr), 4.38 (m, 2H, CH₂ Fm), 4.10 (t, 1H, CH Fm), 2.94 (m, 2H, CH₂ β -Tyr), 1.42 (s, 9H, t-Bu Boc). IR (cm⁻¹): 3348, 2931, 1733, 1686, 1527, 1512, 1242, 1163, 1048, 736, 549. α_D (CHCl₃) = 0.0°. LC-MS (50 - 90 % B in 15 min), Rt = 9.62. HRMS [C₃₅H₃₅NO₅ + H]⁺: 550.2585 found, 550.2588 calc.



***N*^α-*tert*-Butoxycarbonyl-L-tyrosine fluorenylmethyl ester (9)**

Compound **8** (1.1 mmol, 0.58 g) was coevaporated with MeCN and dissolved in n-propanol/EtOAc (9:1, 30 mL). The solution was purged with argon prior to addition of Pd-C (10%, 100 mg) and HCl (89 μ L, 1.17 mmol). Hydrogen gas was bubbled through the solution for 1 hour after which the reaction mixture was filtered over celite, concentrated and purified by silica gel column chromatography (PE/EtOAc, 90/10 $\rightarrow$ 80/20) to give compound **9** as a white powder (0.36 g, 73%).

¹³C-NMR (100 MHz, CDCl₃): δ 172.2 (CO α -Tyr), 155.4, (CO Boc), 143.5, 143.3, 141.3, 141.3 (Cq Fm), 130.4 (Arom. Tyr), 127.9, 127.9 (Arom. Fm), 127.4 (Cq Tyr), 127.2, 125.1 (Arom. Fm), 120.1, 115.6 (Arom. Tyr), 80.3 (Cq Boc), 67.2 (CH₂ Fm), 54.7 (CH α Tyr), 46.7 (CH Fm), 37.5 (CH₂ β Tyr), 28.3 (CH₃ Boc). ¹H-NMR (400 MHz, CDCl₃): δ 7.78 (d, 2H, Arom. Fm), 7.48 (t, 2H, Arom. Fm), 7.34, 7.27 (2 x m, 4H, Arom. Fm), 6.89, 6.71 (2x d, 4H, Arom. Tyr), 5.12 (d, 1H, NH α -Tyr), 4.60 (m, 1H, CH α -Tyr), 4.39 (m, 2H, CH₂ Fm), 4.10 (t, 1H, CH Fm), 2.97 – 2.87 (m, 2H, CH₂ β -Tyr), 1.41 (s, 9H, Boc). IR (cm⁻¹): 3367, 2979, 1686, 1516, 1244, 1163, 530. α_D (CHCl₃) = -1.0°. LC-MS (50 - 90 % B in 15 min), Rt = 5.62. HRMS [C₂₈H₂₉NO₅ + H]⁺: 460.2116 found, 460.2118 calc.

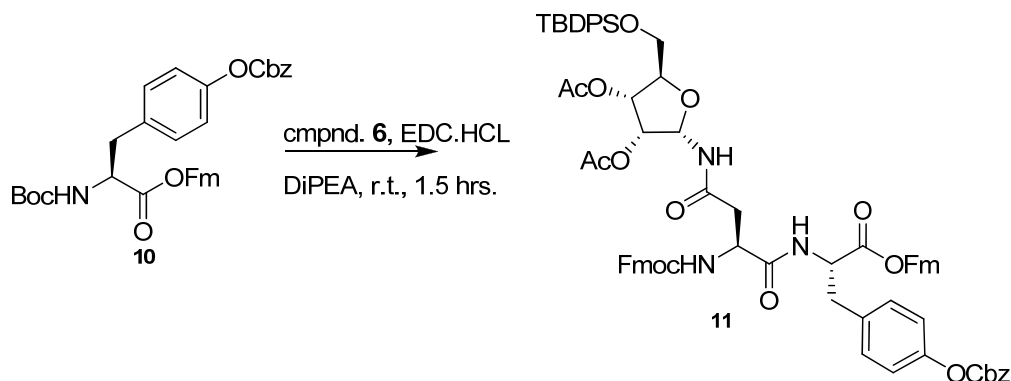


***N*^α-*tert*-Butoxycarbonyl-4-O-benzyloxycarbonyl-L-tyrosine fluorenylmethyl ester (10)**

Compound **9** (0.68 mmol, 0.31 g) was coevaporated with MeCN and dissolved in DCM (3.5 mL) under an argon atmosphere. To this solution were added benzylchloroformate (0.1 mL, 0.75 mmol) and DiPEA (0.12 mL, 0.75 mmol). The solution was stirred at room temperature for 2 hours, washed with 5% citric acid and H₂O. The combined organic layers were dried and concentrated *in vacuo*. The crude compound was purified by silica gel column chromatography (PE/EtOAc, 90/10 $\rightarrow$ 80/20) to give compound **10** as a white powder (0.34 g, 84%).

¹³C-NMR (100 MHz, CDCl₃): δ : 171.4 (CO α -Tyr), 154.9, 153.3 (CO Cbz, CO Boc), 149.9 (Cq Tyr), 143.2, 143.1, 141.0, 141.0 (Cq Fm), 134.5 (Cq Cbz), 133.7 (Cq Tyr), 130.2 (Arom. Tyr), 128.5, 128.4, 127.6, 126.9 (Arom. Fm/Cbz), 124.7, 120.8 (Arom. Fm), 119.8 (Arom. Tyr), 79.7 (Cq Boc), 70.0 (CH₂ Cbz), 66.8 (CH₂ Fm), 54.1 (CH α Tyr), 46.4 (CH Fm), 37.1 (CH₂ β Tyr), 28.0 (CH₃ Boc). ¹H-NMR (400 MHz, CDCl₃): δ 7.83 (d, 2H, Arom. Fm), 7.62 (t, 2H, Arom. Fm), 7.58 – 7.37 (m, 9H, Arom. Fm/Cbz), 7.12 (s, 4H, Arom. Tyr), 5.32 (s, 2H, CH₂ Cbz), 5.10 (d, 1H, NH α -Tyr), 4.71 (m, 1H, CH α -Tyr), 4.52 (m, 2H, CH₂ Fm), 4.23 (t, 1H, CH Fm), 3.10 – 3.00 (m, 2H, CH₂ β -Tyr), 1.50 (s, 9H, Boc). α_D (CHCl₃) = + 0.4°. IR (cm⁻¹): 3351,

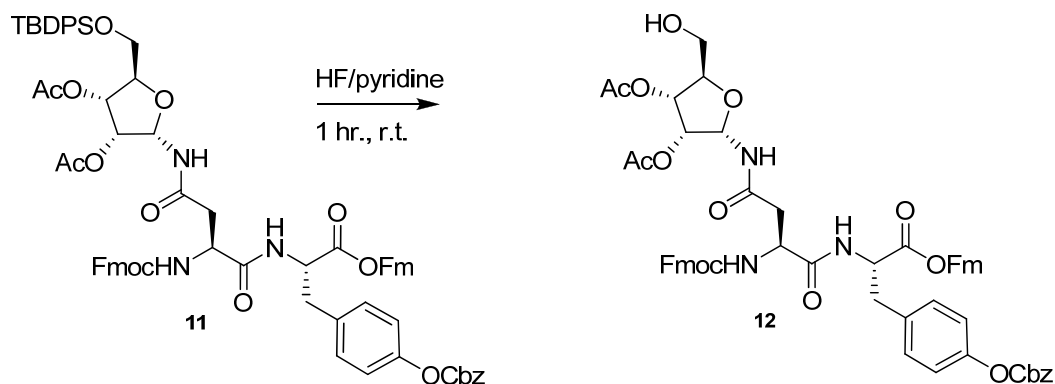
1748, 1683, 523, 1267, 1161, 738, 545. LC-MS (50 - 90 % B in 15 min), Rt = 9.34. HRMS [C₃₆H₃₅NO₇ + H]⁺: 616.2301 found, 616.2305 calc.



***N*'-(5-*tert*-Butyldiphenylsilyl-2,3-di-*O*-acetyl- α -D-ribose)-*N* ^{α} -fluorenylmethoxycarbonyl asparaginyl-(4-*O*-benzyloxycarbonyl)-L-tyrosine fluorenylmethyl ester (11)**

Compound **10** (0.55 mmol, 0.33 g) was dissolved in 3 M HCl/EtOAc and stirred at room temperature for 30 min. The reaction mixture was concentrated *in vacuo* and coevaporated with toluene. The residue was dissolved in DCM (10 mL) and a solution of **6** (0.5 mmol) in DCM (2.5 mL) was added, followed by EDC-HCl (0.6 mmol, 0.12 g) and DiPEA (0.6 mmol, 0.1 mL). After stirring at room temperature for 1.5 hr the reaction mixture was washed with 5% citric acid and water. The organic layer was dried (MgSO₄) and concentrated. The crude compound was purified by silica gel column chromatography (PE/EtOAc, 100/0 → 35/65) to give compound **11** as a white powder (0.59 g, 0.46 mmol, 84%).

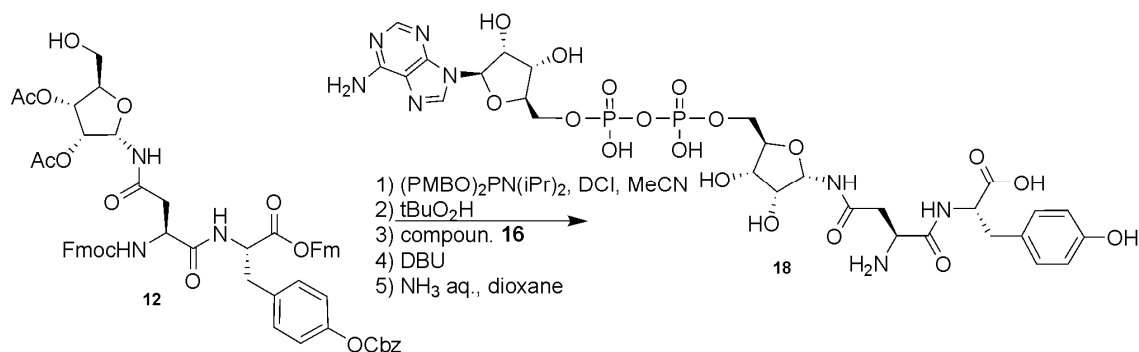
¹³C-NMR (100 MHz, CDCl₃): δ 171.3 (CO α -Tyr), 170.9, 170.6 (CO OAc), 169.6, 169.2 (CO α -Asn, CO γ -Asn), 156.2 (CO Fmoc), 153.5 (CO Cbz), 150.3 (Cq Tyr), 143.8, 143.7, 143.5, 143.4, 141.4, 141.3 (Cq Fm, Cq Fmoc), 135.7 (Arom. TBDPS), 134.8 (Cq Cbz), 133.6 (Cq Tyr), 132.9, 132.7 (Cq TBDPS), 130.4, 129.9, 128.8, 128.7, 128.5, 128.0, 127.9, 127.8, 127.3, 127.2, 125.2, 125.0, 121.2, 120.0 (Arom. Fmoc, Arom. Fm, Arom. TBDPS, Arom. Cbz), 82.2 (C4'), 78.7 (C1'), 72.4 (C3'), 70.3 (CH₂ Cbz), 70.2 (C2') 67.5, 67.2 (CH₂ Fmoc, CH₂ Fm), 63.7 (C5'), 53.7 (C α Tyr), 51.3 (C α Asn), 47.1, 46.7 (CH Fmoc, CH Fm), 38.0 (CH₂ β -Asn), 37.0 (CH₂ β -Tyr), 26.8 (CH₃ TBDPS), 20.8, 20.6 (CH₃ OAc), 19.2 (Cq t-Bu). ¹H-NMR (400 MHz, MeOD-d₄/CDCl₃): δ 7.57, 7.53, 7.51 – 7.48, 7.36, 7.26, 7.03 (6x m, Arom. Fmoc, Arom. Fm, Arom. TBDPS, Arom. Tyr, 35H), 6.71 (d, 1H, NH γ -Tyr), 6.25 (d, 1H, NH α -Asn), 6.06 (dd, 1H, H1'), 5.56 (s, 1H, H3'), 5.47 (m, 1H, H2'), 5.16 (CH₂ Cbz), 4.75 (m, 1H, CH α -Asn), 4.52 (m, 1H, CH α -Tyr), 4.45, 4.34 (2 x m, 4H, CH₂ Fmoc, CH₂ Fm), 4.18 (m, 2H, CH Fmoc, CH Fm), 4.11 (s, 1H, H4'), 3.71 (s, 2H, H5'), 3.02 – 2.62 (m, 4H, CH₂ β -Asn, β -Tyr), 2.07, 2.02 (2 x s, 6H, CH₃ OAc), 1.06 (s, 9H, t-Bu). α_D (CHCl₃) = + 29.6°. IR (cm⁻¹): 3278, 2933, 2360, 1747, 1696, 1648, 1534, 1211, 1104, 737, 699, 503. HRMS [C₇₅H₇₃N₃O₁₅Si + H]⁺: 1284.4903 found, 1284.4883 calc.



***N'*-(5-Hydroxy-2,3-di-*O*-acetyl- α -D-ribose)-*N'*-fluorenylmethoxycarbonyl asparaginyl-(4-*O*-benzyloxycarbonyl)-L-tyrosine fluorenylmethyl ester ; Fmoc-Asn-(*N'*-[2,3-di-*O*-acetyl- α -D-ribose])*-Tyr*(Cbz)-OFm (**12**)**

Silyl protected compound **11** (0.47 mmol, 0.61 g) was dissolved in pyridine (10 mL) and treated with a HF/pyridine solution (1.0 mL) for 1 hour at room temperature. The reaction was quenched with sat. aq. NaHCO₃ and extracted with DCM. After washing with water the organic layer was dried (MgSO₄) and concentrated *in vacuo*. The crude compound was purified by silica gel column chromatography (DCM/MeOH, 99/1) to give compound **12** as a white powder (0.42 g, 0.4 mmol, 85%).

¹³C-NMR (100 MHz, DMSO-*d*₆): δ 172.0, 171.6, 170.0, 169.9, 169.8 (CO α -Tyr, CO OAc, CO α -Asn, CO γ -Asn), 156.2 (CO Fmoc), 153.4 (CO Cbz), 149.9 (Cq Tyr.), 144.3, 144.1, 143.9, 141.3, 141.2, 141.1 (Cq Fm, Cq Fmoc), 135.48 (Cq Cbz), 135.45 (Cq Tyr.), 130.7, 129.0, 128.8, 128.2, 128.1, 127.8, 127.7, 127.6, 125.8, 125.6, 121.4, 120.6 (Arom. Fmoc, Arom. Fm, Arom. Cbz), 80.5 (C4'), 79.1 (C1'), 71.5 (C3'), 70.7 (C2'), 70.2 (CH₂ Cbz), 66.7, 66.3 (CH₂ Fmoc, CH₂ Fm), 61.4 (C5'), 54.2 (C α Tyr), 51.6 (C α Asn), 47.0, 46.7 (CH Fmoc, CH Fm), 38.0 (CH₂ β -Asn), 36.0 (CH₂ β -Tyr), 20.9 (CH₃ OAc). ¹H-NMR (400 MHz, CDCl₃): δ 7.58, 7.45, 7.41, 7.32, 7.09 (5 x m, 25H, Arom. Fmoc, Arom. Fm, Arom. Cbz), 6.69, 6.23 (2x d, 1H, NH's), 5.95 (m, 1H, H1'), 5.40, 5.31 (2x m, 4H, H2', H3'), 5.25 (s, 2H, CH₂ Cbz), 4.79, 4.56 (2x m, 2H, H α -Asn, H α -Tyr), 4.51, 4.38, (2x m, 6H, CH₂ Fmoc, CH₂ Fm), 4.24 (m, 2H, CH Fmoc, CH Fm), 4.12 (m, 1H, H4'), 3.87 – 3.64 (m, 2H, H5'), 3.03 – 2.89, 2.56 (m, 4H, CH₂ β Asn, CH₂ β Tyr) 2.14, 2.12 (2 x s, 6H, CH₃ OAc). α_D (CHCl₃) = + 26.8°. IR (cm⁻¹): 3288, 2924, 2360, 1746, 1652, 1538, 1269, 1212, 1046, 736. LC-MS (10 - 90 % B in 15 min), Rt = 11.30. HRMS [C₅₉H₅₅N₃O₁₅ + H]⁺: 1046.318 found, 1046.3705 calc.



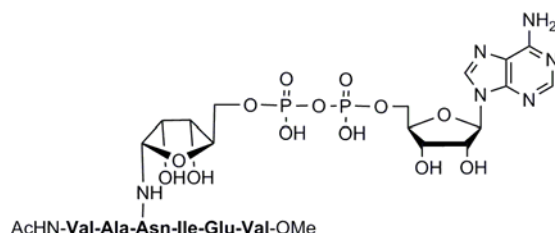
Compound 18: H-Asn(*N'* [5'-adenosine diphosphate- α -D-ribose])*-Tyr*-OH.

Ribosylated dipeptide **12** (0.1 mmol, 100 mg) was dissolved in dioxane/ MeCN (1:1, 4 mL) and cooled to 0 °C. Di(p-methoxybenzyl)-N,N-diisopropylphosphoramidite (0.11 mmol, 43 μ L) and a solution of DCl (0.11 mmol, 13 mg) in 200 μ L MeCN were added after which the reaction mixture was kept at 0 °C for 1 hour. 100 μ L 5.5 M solution of tBuO₂H in nonane was added and stirred for an additional hour at 0 °C. After trituration with hexane, the residue was

treated with 5% TFA in DCM to yield intermediate **14**. 6-N-Benzoyl-2', 3'-di-O-isobutyryladeniny-5-phosphorimidazolidate **16** (0.2 mmol) in 1 mL MeCN was added to the reaction mixture and stirred at room temperature for 16 hours. A silica column with isocratic elution (30% MeOH/DCM) and RP-HPLC purification 80 – 90 % MeOH in 20 mM NH₄OAc over 5 CV yielded **17** (26.2 mg, 15.4 μ mol, 15.4% based on **12**).

To a solution of **17** (14.5 mg, 8.5 μ mol) in dioxane/MeCN (1:1, 2 ml) 5 eq. DBU (43 μ mol, 6.5 μ l) was added and reacted for 30 min at room temperature. A solution of aqueous ammonia /dioxane (2:1, 6 ml) was added and reacted for 16 hours at room temperature. After concentration the title compound was obtained by means of gel filtration (4.1 mg, 4.9 μ mol, 58% based on **17**).

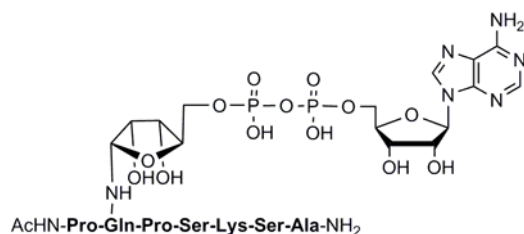
³¹P-NMR (162 MHz, D₂O): δ -10.45, -10.58, -10.75, -10.88. LC-MS (0 - 50 % MeCN in 10 mM NH₄OAc in 15 min), Rt = 4.0. HRMS [C₂₈H₃₈N₈O₁₈P₂ + H]⁺: 837.1861 found, 837.1852 calc.



Compound **21**; Ac-Val-Ala-Asn(N⁵ [5'-adenosine diphosphate- α -D-ribose])-Ile-Glu-Val-OMe

Intermediate **19** was prepared starting from Tentagel S HMB resin (100 mg, 23 μ mol) via standard Fmoc based Solid phase peptide synthesis, using HCTU as coupling reagent and a five fold excess of the appropriate amino acid, using Fmoc-Val-OH, Fmoc-Ala-OH, compound **6**, Fmoc-Ile-OH and Fmoc-Glu(Dmab)-OH. An analytical sample was deprotected and cleaved off the resin using the method described below, to determine quality of **19**. The resin was treated with 0.36 mL HF/pyridine diluted with pyridine (3 mL) (0 °C $\rightarrow$ r.t., 75 min). The resin was filtered and washed with pyridine and MeCN to give intermediate **20**.

The resin (80 mg, 18 μ mol) was coevaporated with MeCN and subsequently MeCN (3 mL), di(p-methoxybenzyl)-N,N-diisopropylphosphoramidite (0.23 mmol, 90 μ L) and a solution of DCI (0.23 mmol, 27 mg) in 1 mL MeCN were added after which the reaction mixture was kept at 0 °C for 1 hour. A 0.5 M solution of I₂ in pyridine was added until the reaction mixture kept the dark brown colour and stirred for an additional hour at 0 °C. 6-N-benzoyl-2', 3'-di-O-isobutyryladeniny-5'-phosphate monoester (0.23 mmol, 0.14 g) was added to the reaction mixture and stirred at room temperature for 24 hours. The resin was filtered and washed with MeCN and treated with 2% hydrazine hydrate in DMF (3 times 5 min.). The resin was filtered, washed with MeCN and treated with 7N NH₃/MeOH (10 mL) for 16 hours. Subsequent filtration into cold Et₂O and centrifuging (6.5 min at 4000 RPM) followed by decantation of the supernatant resulted in the crude title compound. RP-HPLC using a gradient of 0 – 60% MeCN in 1% TFA in H₂O followed by a gradient RP-HPLC using a gradient of 0 – 60% MeCN of 10 mM NH₄OAc, followed by lyophilisation of the appropriate fractions yielded the title compound **21** (0.78 mg, 0.6 μ mol, 3.4% based on initial loading of the resin). ³¹P-NMR (162 MHz, D₂O): δ -11.04, -11.17, -11.31, -11.44. (EDTA was added to sharpen signals). LC-MS (0 - 50 % MeCN [1% TFA] in 15 min), Rt = 8.07. HRMS [C₄₆H₇₄N₁₂O₂₄P₂ + H]⁺: 1241.4505 found, 1241.4486 calc.

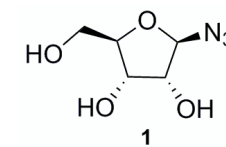


Compound 25: Ac-Pro-Gln-(N^δ [5'-adenosine diphosphate-α-D-ribose]) -Pro-Ser-Lys-Ser-Ala-NH₂

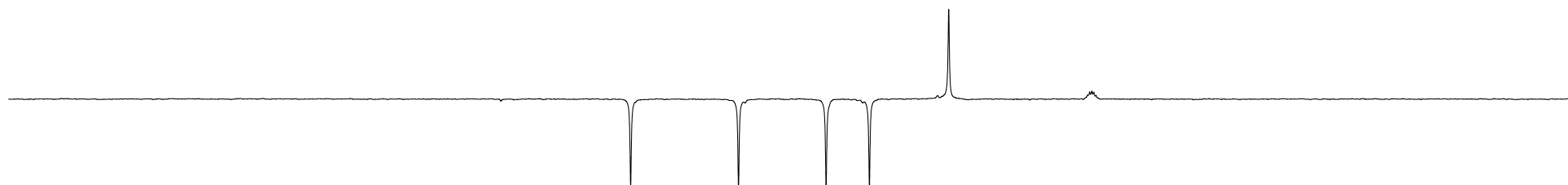
Starting from Tentagel S HMB resin (150 mg, 65 μmol) using standard Fmoc based Solid Phase Peptide Synthesis intermediate **22** was prepared using Fmoc-Pro-OH, compound **7**, Fmoc-Ser(Trt)-OH, Fmoc-Lys(TFA)-OH and Fmoc-Ala-OH. The resin was repeatedly treated with 5% TFA in DCM until no yellow colour appeared upon addition of the fresh TFA solution. After washing with DCM, acetic anhydride (0.5 M, 200 μL) and DMAP (0.05 M, 16 mg) in 2 mL NMP were added and the resin was shaken for 25 min. The resin was washed with NMP and DCM prior to coupling of the last two amino acids using the standard cycle to obtain **23**. An analytical sample was deprotected and cleaved off the resin using the method described below, to determine quality of **24**. The resin was coevaporated with MeCN and treated with a HF/pyridine (0.9 mL) diluted with pyridine (7.5 mL) (0 °C → r.t., 75 min). The resin was filtered and washed with pyridine and MeCN to obtain **24**. The resin was coevaporated with MeCN and subsequently MeCN (8 mL), di(*p*-methoxybenzyl)-*N,N*-diisopropylphosphoramidite (10 eq., 250 μL), dry 3 Å molsieves and a solution of DCI (10 eq., 75 mg) in 1 mL MeCN were added after which the reaction mixture was kept at 0 °C for 1 hour. Next the phosphate was oxidized using 5.5 M *t*BuO₂H (1.1 mL) in nonane for 1 hour at room temperature followed by addition of 3% TFA for 10 min. to deprotect the phosphor triester. 6-N-Benzoyl-2', 3'-di-O-isobutyryladenine-5-phosphorimidazolide **16** (0.4 mmol, 6 eq.) in 1 mL MeCN was added to the reaction mixture and stirred at room temperature for 16 hours. The resin was filtered, washed with MeCN and treated with 7N NH₃/MeOH (10 mL) for 16 hours. Subsequent filtration into cold Et₂O and centrifuging (6.5 min at 4000 RPM) followed by decantation of the supernatant resulted in the crude title compound. Preparative RP-HPLC using a gradient of 2.5 – 40% MeCN in 10 mM aq. NH₄OAc, followed by lyophilisation of the appropriate fractions yielded the title compound (1.3 mg, 1.0 μmol, 1.5% based on initial loading of the resin).

³¹P-NMR (162 MHz, D₂O): δ -10.46, -10.59, -10.74, -10.87. LC-MS (0 - 50 % MeCN [1% TFA] in 15 min), Rt = 4.49. HRMS [C₄₇H₇₅N₁₅O₂₄P₂ + H]⁺: 1296.4675 found, 1296.4657 calc.

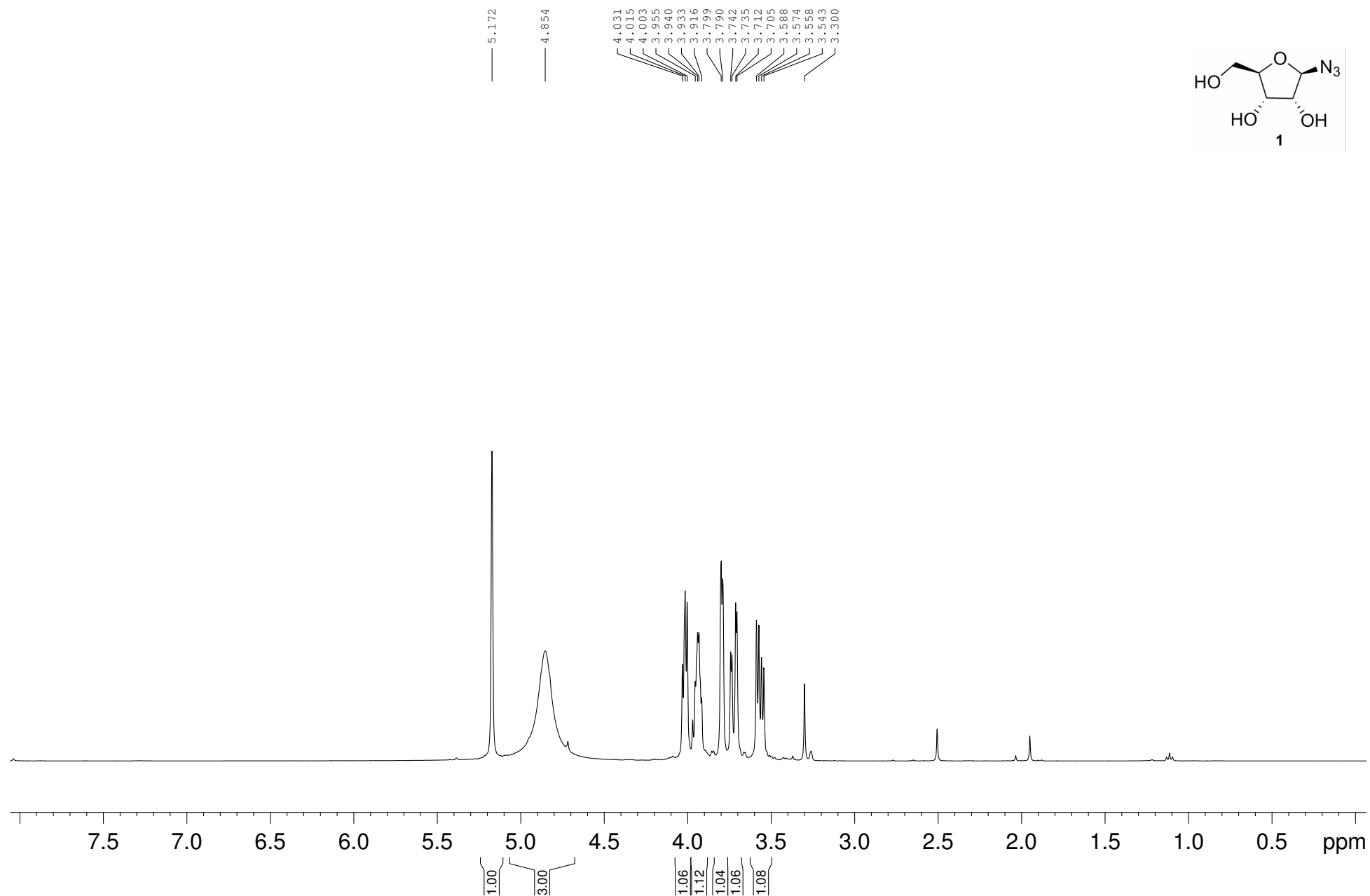
^{13}C -NMR, 100 MHz, MeOD- d_4 , cmpnd 1



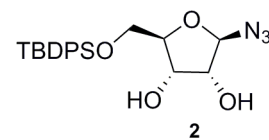
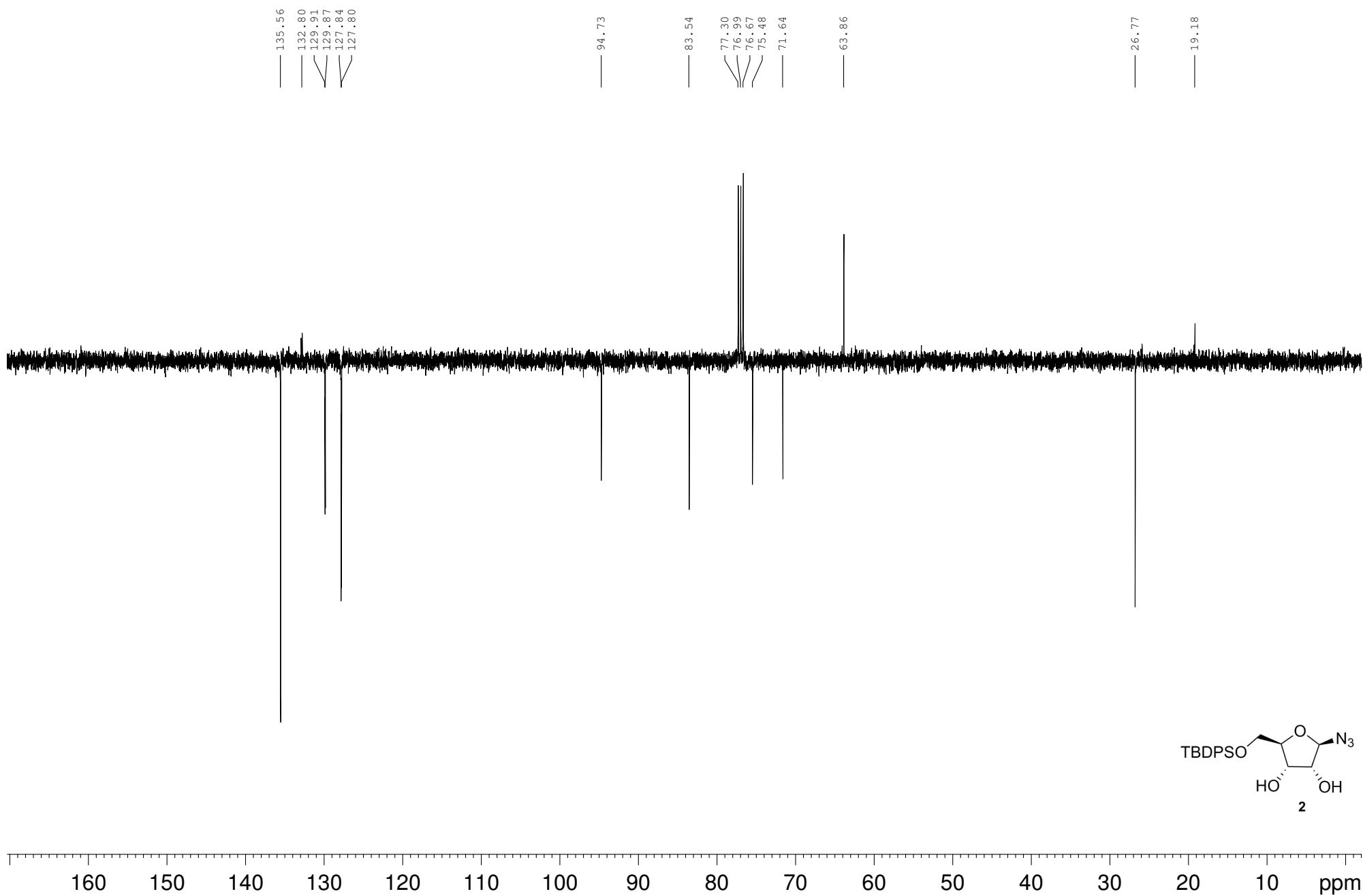
96.24
85.19
76.22
71.79
63.66



^1H NMR, 400 MHz, MeOD- d_4 , compnd 1



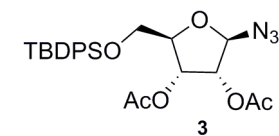
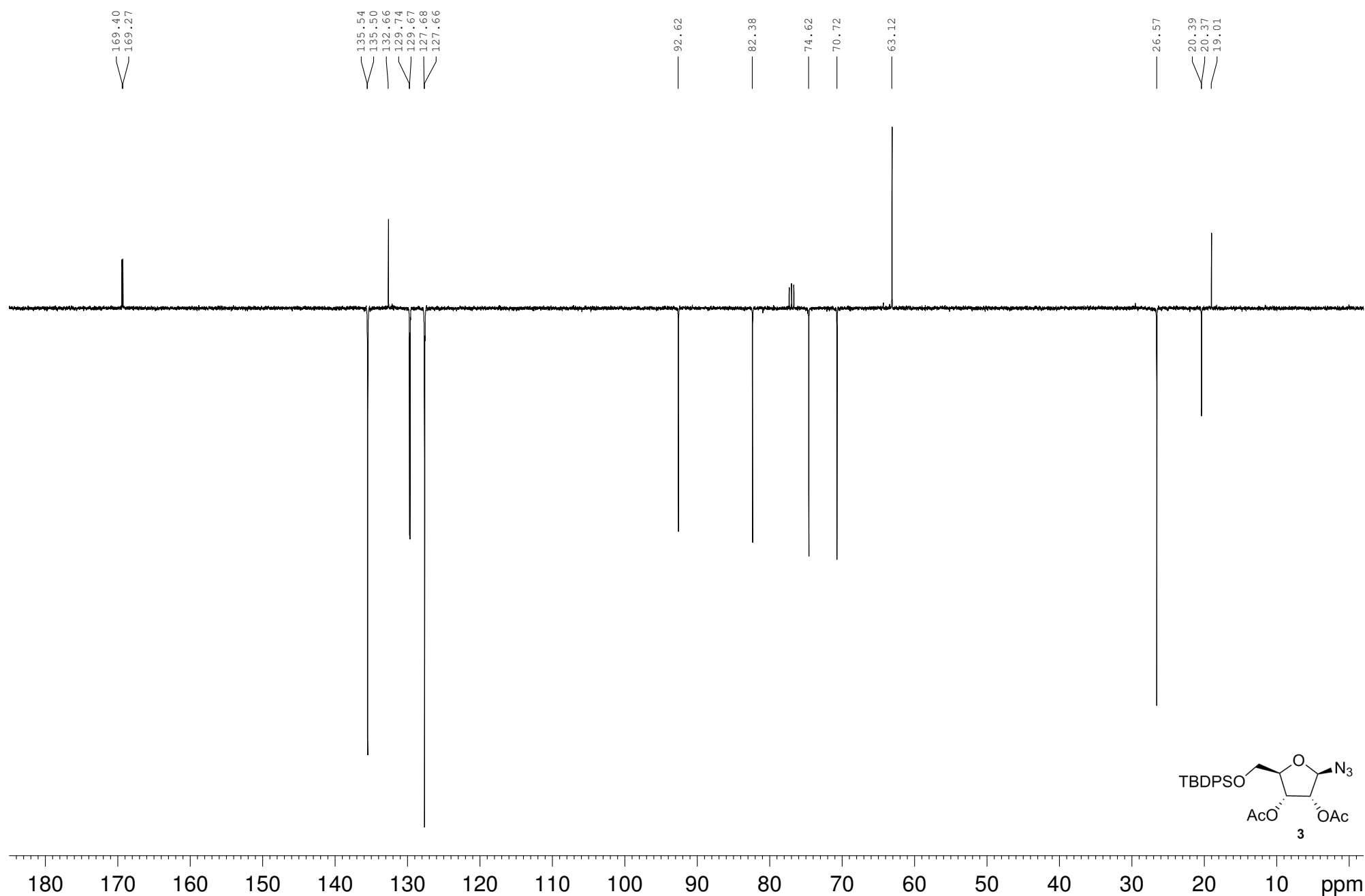
^{13}C -NMR, 100 MHz, CDCl_3 , compnd. 2



¹H NMR spectrum (CDCl₃) of compound 2.

Chemical structure of compound 2: C1=CC=C(C=C1)C2=CC=CC=C2C3=CC=CC=C3C4=CC=CC=C4C5=CC=CC=C5C6=CC=CC=C6C7=CC=CC=C7C8=CC=CC=C8C9=CC=CC=C9C10=CC=CC=C10C11=CC=CC=C11C12=CC=CC=C12C13=CC=CC=C13C14=CC=CC=C14C15=CC=CC=C15C16=CC=CC=C16C17=CC=CC=C17C18=CC=CC=C18C19=CC=CC=C19C20=CC=CC=C20C21=CC=CC=C21C22=CC=CC=C22C23=CC=CC=C23C24=CC=CC=C24C25=CC=CC=C25C26=CC=CC=C26C27=CC=CC=C27C28=CC=CC=C28C29=CC=CC=C29C30=CC=CC=C30C31=CC=CC=C31C32=CC=CC=C32C33=CC=CC=C33C34=CC=CC=C34C35=CC=CC=C35C36=CC=CC=C36C37=CC=CC=C37C38=CC=CC=C38C39=CC=CC=C39C40=CC=CC=C40C41=CC=CC=C41C42=CC=CC=C42C43=CC=CC=C43C44=CC=CC=C44C45=CC=CC=C45C46=CC=CC=C46C47=CC=CC=C47C48=CC=CC=C48C49=CC=CC=C49C50=CC=CC=C50C51=CC=CC=C51C52=CC=CC=C52C53=CC=CC=C53C54=CC=CC=C54C55=CC=CC=C55C56=CC=CC=C56C57=CC=CC=C57C58=CC=CC=C58C59=CC=CC=C59C60=CC=CC=C60C61=CC=CC=C61C62=CC=CC=C62C63=CC=CC=C63C64=CC=CC=C64C65=CC=CC=C65C66=CC=CC=C66C67=CC=CC=C67C68=CC=CC=C68C69=CC=CC=C69C70=CC=CC=C70C71=CC=CC=C71C72=CC=CC=C72C73=CC=CC=C73C74=CC=CC=C74C75=CC=CC=C75C76=CC=CC=C76C77=CC=CC=C77C78=CC=CC=C78C79=CC=CC=C79C80=CC=CC=C80C81=CC=CC=C81C82=CC=CC=C82C83=CC=CC=C83C84=CC=CC=C84C85=CC=CC=C85C86=CC=CC=C86C87=CC=CC=C87C88=CC=CC=C88C89=CC=CC=C89C90=CC=CC=C90C91=CC=CC=C91C92=CC=CC=C92C93=CC=CC=C93C94=CC=CC=C94C95=CC=CC=C95C96=CC=CC=C96C97=CC=CC=C97C98=CC=CC=C98C99=CC=CC=C99C100=CC=CC=C100C101=CC=CC=C101C102=CC=CC=C102C103=CC=CC=C103C104=CC=CC=C104C105=CC=CC=C105C106=CC=CC=C106C107=CC=CC=C107C108=CC=CC=C108C109=CC=CC=C109C110=CC=CC=C110C111=CC=CC=C111C112=CC=CC=C112C113=CC=CC=C113C114=CC=CC=C114C115=CC=CC=C115C116=CC=CC=C116C117=CC=CC=C117C118=CC=CC=C118C119=CC=CC=C119C120=CC=CC=C120C121=CC=CC=C121C122=CC=CC=C122C123=CC=CC=C123C124=CC=CC=C124C125=CC=CC=C125C126=CC=CC=C126C127=CC=CC=C127C128=CC=CC=C128C129=CC=CC=C129C130=CC=CC=C130C131=CC=CC=C131C132=CC=CC=C132C133=CC=CC=C133C134=CC=CC=C134C135=CC=CC=C135C136=CC=CC=C136C137=CC=CC=C137C138=CC=CC=C138C139=CC=CC=C139C140=CC=CC=C140C141=CC=CC=C141C142=CC=CC=C142C143=CC=CC=C143C144=CC=CC=C144C145=CC=CC=C145C146=CC=CC=C146C147=CC=CC=C147C148=CC=CC=C148C149=CC=CC=C149C150=CC=CC=C150C151=CC=CC=C151C152=CC=CC=C152C153=CC=CC=C153C154=CC=CC=C154C155=CC=CC=C155C156=CC=CC=C156C157=CC=CC=C157C158=CC=CC=C158C159=CC=CC=C159C160=CC=CC=C160C161=CC=CC=C161C162=CC=CC=C162C163=CC=CC=C163C164=CC=CC=C164C165=CC=CC=C165C166=CC=CC=C166C167=CC=CC=C167C168=CC=CC=C168C169=CC=CC=C169C170=CC=CC=C170C171=CC=CC=C171C172=CC=CC=C172C173=CC=CC=C173C174=CC=CC=C174C175=CC=CC=C175C176=CC=CC=C176C177=CC=CC=C177C178=CC=CC=C178C179=CC=CC=C179C180=CC=CC=C180C181=CC=CC=C181C182=CC=CC=C182C183=CC=CC=C183C184=CC=CC=C184C185=CC=CC=C185C186=CC=CC=C186C187=CC=CC=C187C188=CC=CC=C188C189=CC=CC=C189C190=CC=CC=C190C191=CC=CC=C191C192=CC=CC=C192C193=CC=CC=C193C194=CC=CC=C194C195=CC=CC=C195C196=CC=CC=C196C197=CC=CC=C197C198=CC=CC=C198C199=CC=CC=C199C200=CC=CC=C200C201=CC=CC=C201C202=CC=CC=C202C203=CC=CC=C203C204=CC=CC=C204C205=CC=CC=C205C206=CC=CC=C206C207=CC=CC=C207C208=CC=CC=C208C209=CC=CC=C209C210=CC=CC=C210C211=CC=CC=C211C212=CC=CC=C212C213=CC=CC=C213C214=CC=CC=C214C215=CC=CC=C215C216=CC=CC=C216C217=CC=CC=C217C218=CC=CC=C218C219=CC=CC=C219C220=CC=CC=C220C221=CC=CC=C221C222=CC=CC=C222C223=CC=CC=C223C224=CC=CC=C224C225=CC=CC=C225C226=CC=CC=C226C227=CC=CC=C227C228=CC=CC=C228C229=CC=CC=C229C230=CC=CC=C230C231=CC=CC=C231C232=CC=CC=C232C233=CC=CC=C233C234=CC=CC=C234C235=CC=CC=C235C236=CC=CC=C236C237=CC=CC=C237C238=CC=CC=C238C239=CC=CC=C239C240=CC=CC=C240C241=CC=CC=C241C242=CC=CC=C242C243=CC=CC=C243C244=CC=CC=C244C245=CC=CC=C245C246=CC=CC=C246C247=CC=CC=C247C248=CC=CC=C248C249=CC=CC=C249C250=CC=CC=C250C251=CC=CC=C251C252=CC=CC=C252C253=CC=CC=C253C254=CC=CC=C254C255=CC=CC=C255C256=CC=CC=C256C257=CC=CC=C257C258=CC=CC=C258C259=CC=CC=C259C260=CC=CC=C260C261=CC=CC=C261C262=CC=CC=C262C263=CC=CC=C263C264=CC=CC=C264C265=CC=CC=C265C266=CC=CC=C266C267=CC=CC=C267C268=CC=CC=C268C269=CC=CC=C269C270=CC=CC=C270C271=CC=CC=C271C272=CC=CC=C272C273=CC=CC=C273C274=CC=CC=C274C275=CC=CC=C275C276=CC=CC=C276C277=CC=CC=C277C278=CC=CC=C278C279=CC=CC=C279C280=CC=CC=C280C281=CC=CC=C281C282=CC=CC=C282C283=CC=CC=C283C284=CC=CC=C284C285=CC=CC=C285C286=CC=CC=C286C287=CC=CC=C287C288=CC=CC=C288C289=CC=CC=C289C290=CC=CC=C290C291=CC=CC=C291C292=CC=CC=C292C293=CC=CC=C293C294=CC=CC=C294C295=CC=CC=C295C296=CC=CC=C296C297=CC=CC=C297C298=CC=CC=C298C299=CC=CC=C299C300=CC=CC=C300C301=CC=CC=C301C302=CC=CC=C302C303=CC=CC=C303C304=CC=CC=C304C305=CC=CC=C305C306=CC=CC=C306C307=CC=CC=C307C308=CC=CC=C308C309=CC=CC=C309C310=CC=CC=C310C311=CC=CC=C311C312=CC=CC=C312C313=CC=CC=C313C314=CC=CC=C314C315=CC=CC=C315C316=CC=CC=C316C317=CC=CC=C317C318=CC=CC=C318C319=CC=CC=C319C320=CC=CC=C320C321=CC=CC=C321C322=CC=CC=C322C323=CC=CC=C323C324=CC=CC=C324C325=CC=CC=C325C326=CC=CC=C326C327=CC=CC=C327C328=CC=CC=C328C329=CC=CC=C329C330=CC=CC=C330C331=CC=CC=C331C332=CC=CC=C332C333=CC=CC=C333C334=CC=CC=C334C335=CC=CC=C335C336=CC=CC=C336C337=CC=CC=C337C338=CC=CC=C338C339=CC=CC=C339C340=CC=CC=C340C341=CC=CC=C341C342=CC=CC=C342C343=CC=CC=C343C344=CC=CC=C344C345=CC=CC=C345C346=CC=CC=C346C347=CC=CC=C347C348=CC=CC=C3

^{13}C -NMR, 100 MHz, CDCl_3 , compnd. 3

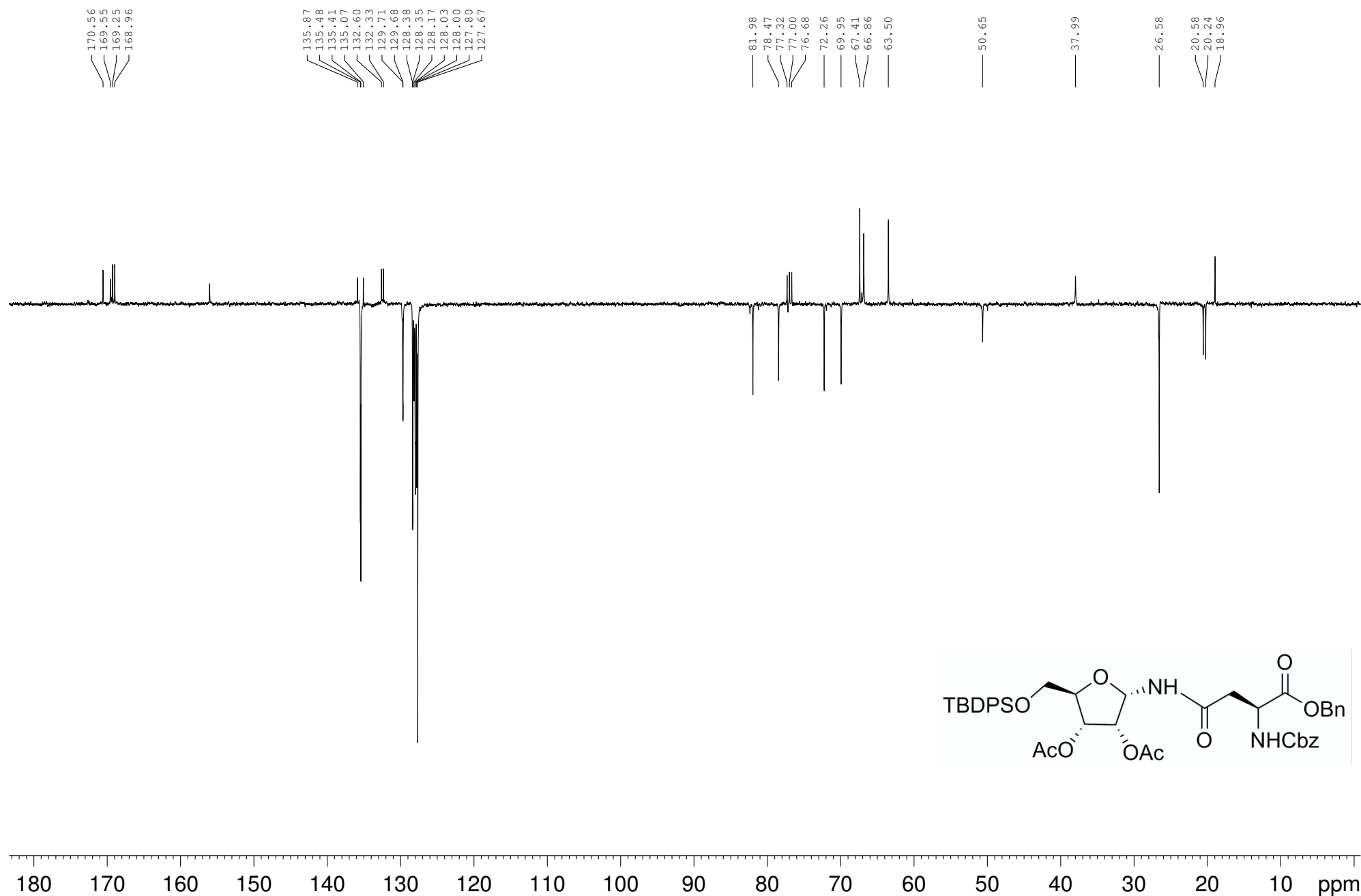


Chemical structure of compound 3: A furanose ring with a TBDPSO group at C2, an azide group at C3, and acetoxy groups at C4 and C5.

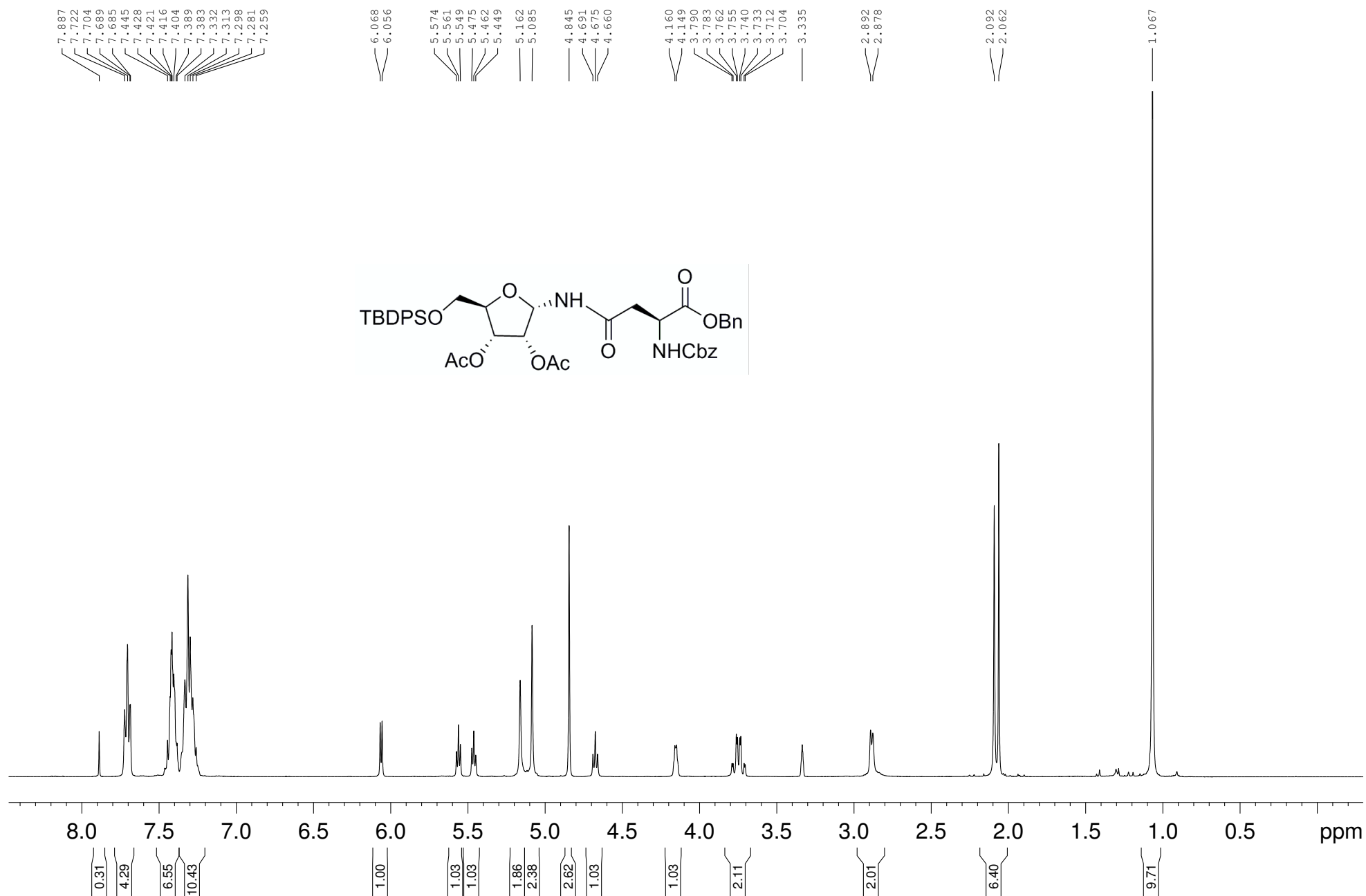
¹H NMR spectrum (CDCl₃):

Chemical Shift (ppm)	Integration
7.723, 7.704, 7.684, 7.681, 7.413, 7.397, 7.378, 7.362	4.08, 6.03
5.525, 5.511, 5.498, 5.418, 5.411, 5.222, 5.215, 5.211, 5.203	1.00, 0.87, 0.97
4.232, 4.223, 4.216, 4.210, 4.202, 3.876, 3.867, 3.847, 3.839, 3.751, 3.743, 3.723, 3.714	1.00, 0.99, 1.02
2.188, 2.093, 2.027	0.35, 3.00, 2.99
1.263, 1.088, 1.069	9.41

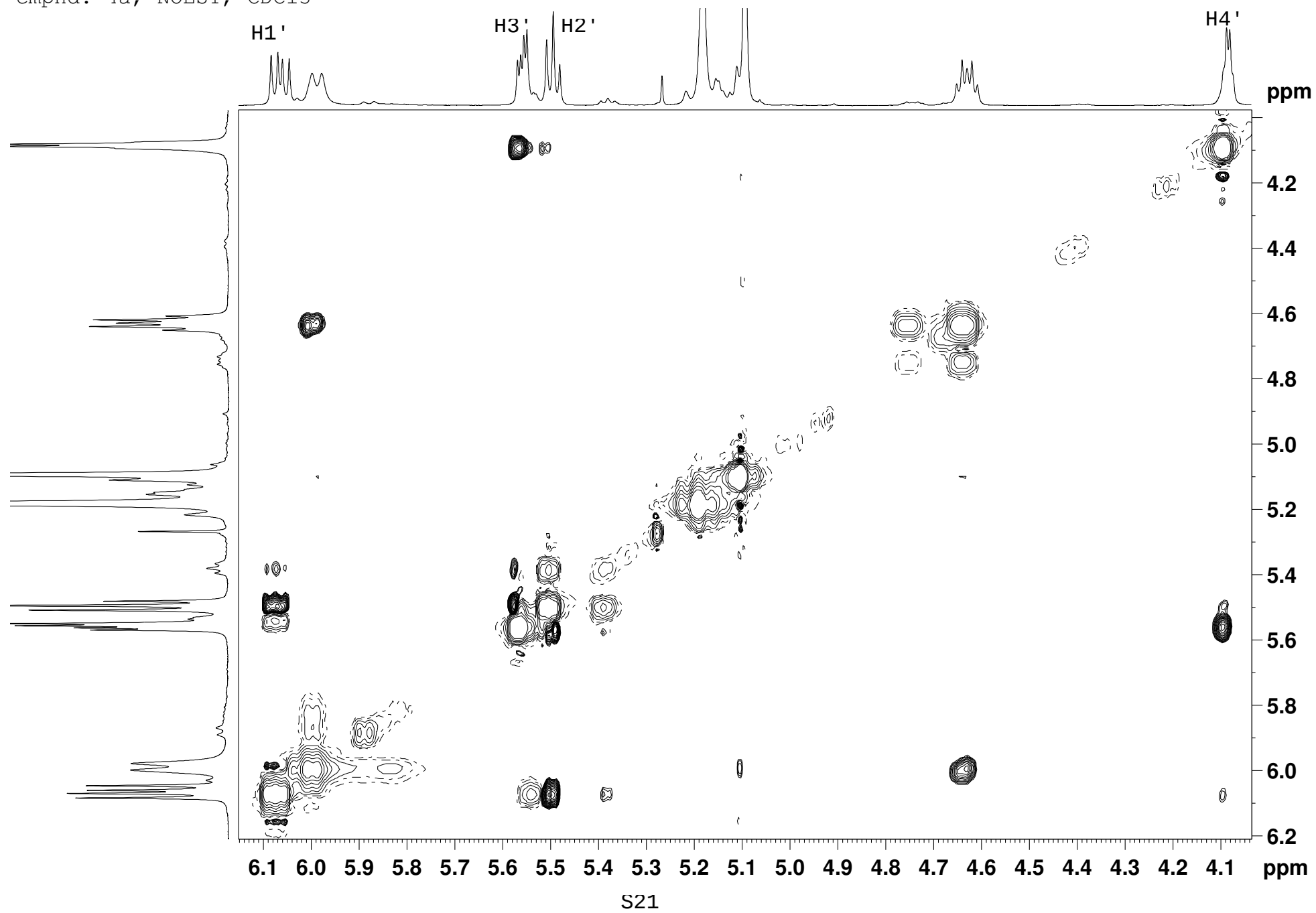
^{13}C -NMR, 100MHz, CDCl_3 , compnd 4A



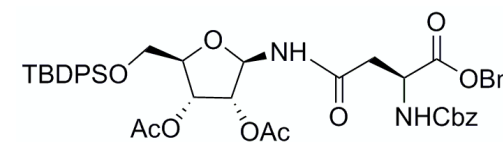
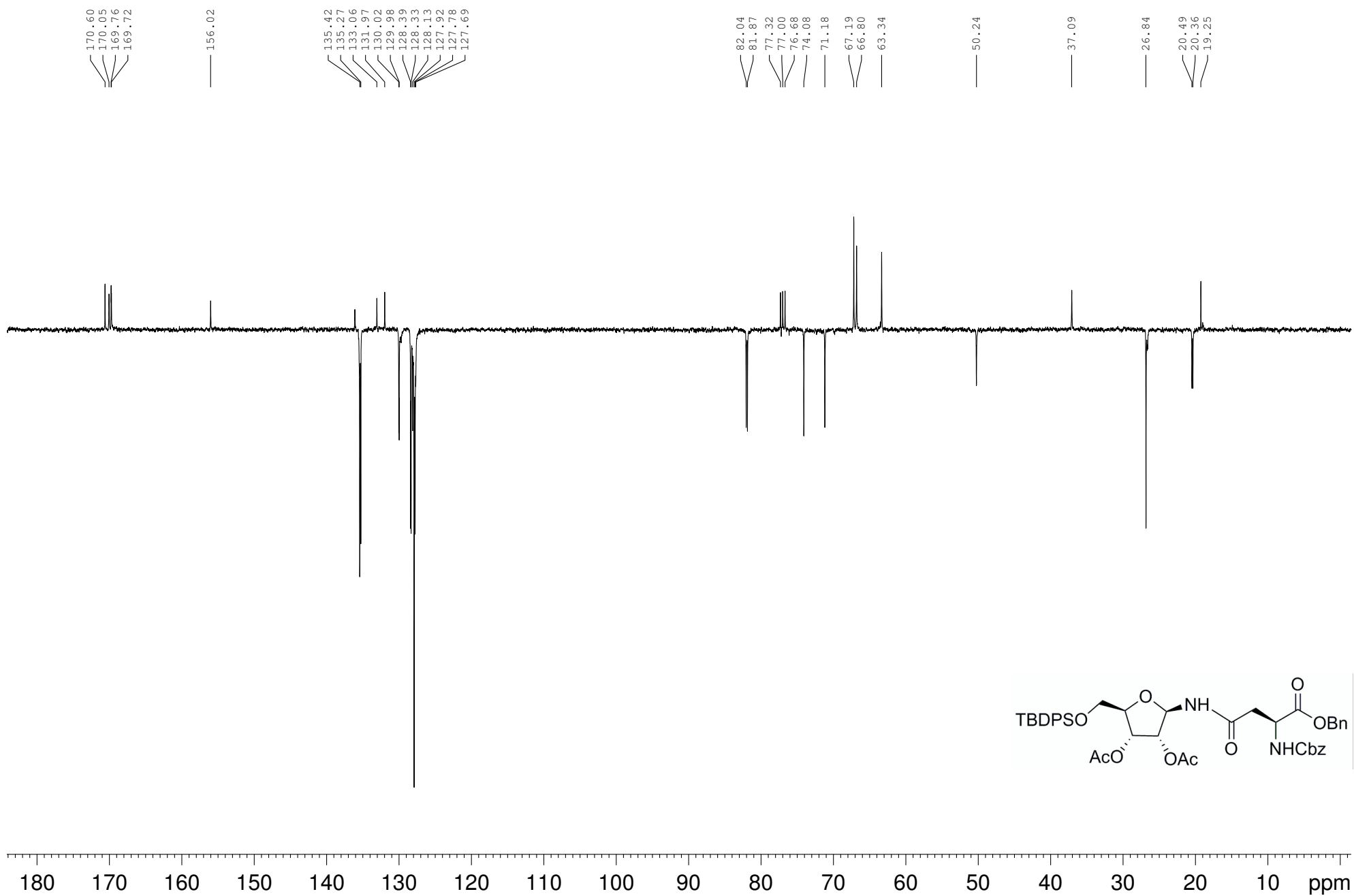
¹H-NMR, 400 MHz, MeOD-d₄, cmpnd 4A



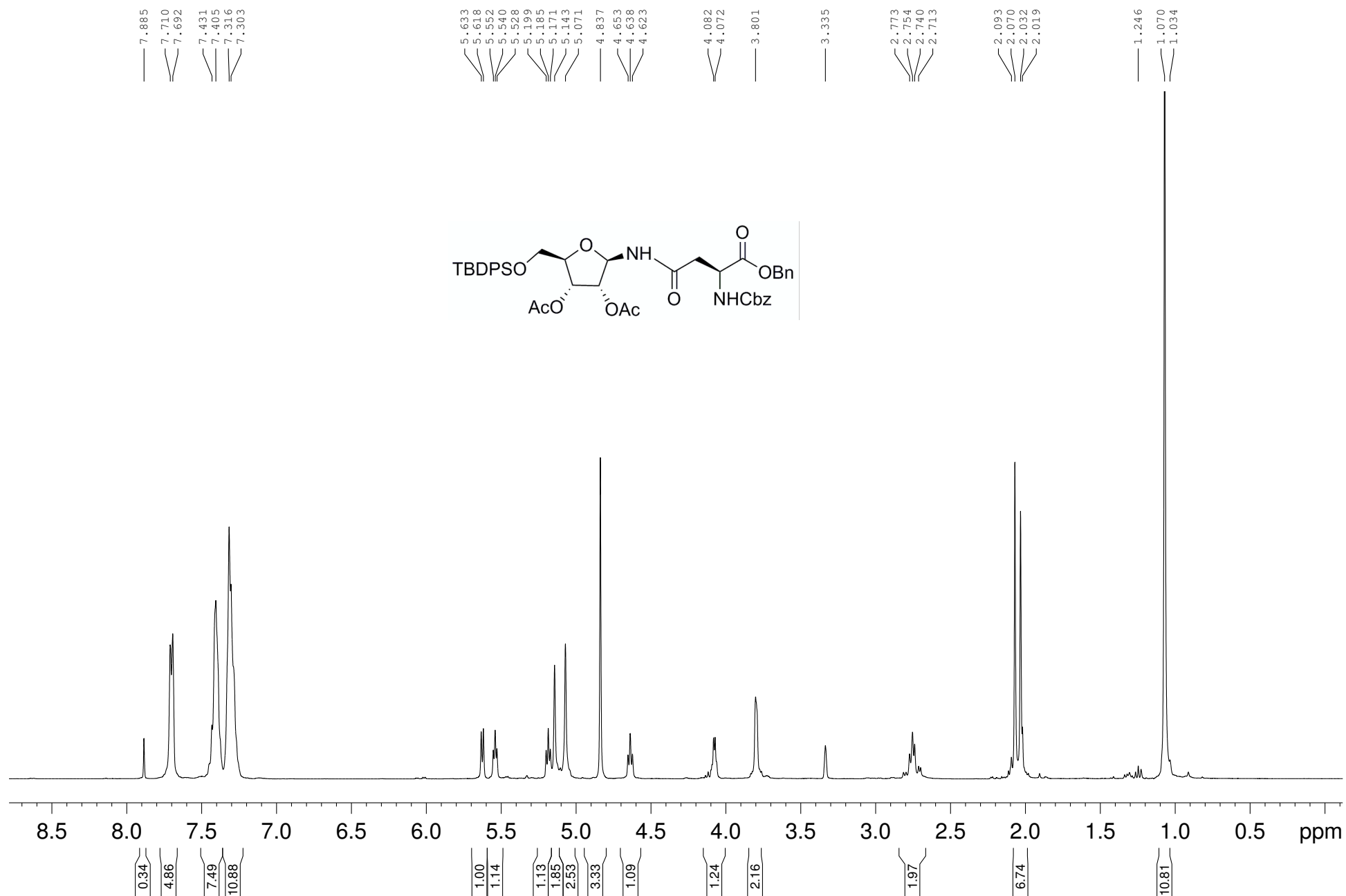
cmpnd. 4a, NOESY, CDCl₃



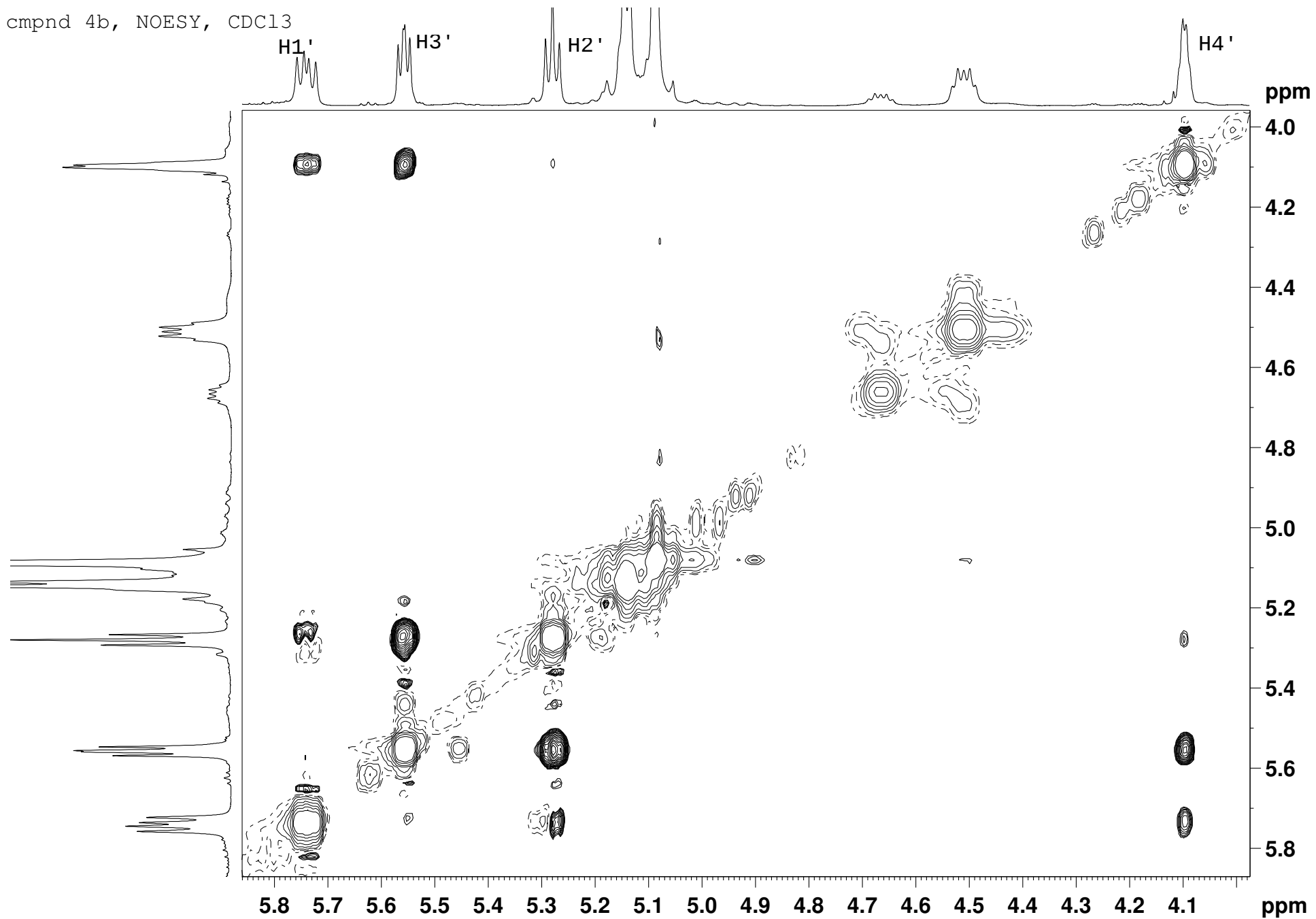
¹³C-NMR, 100 MHz, CDCl₃, compnd 4B



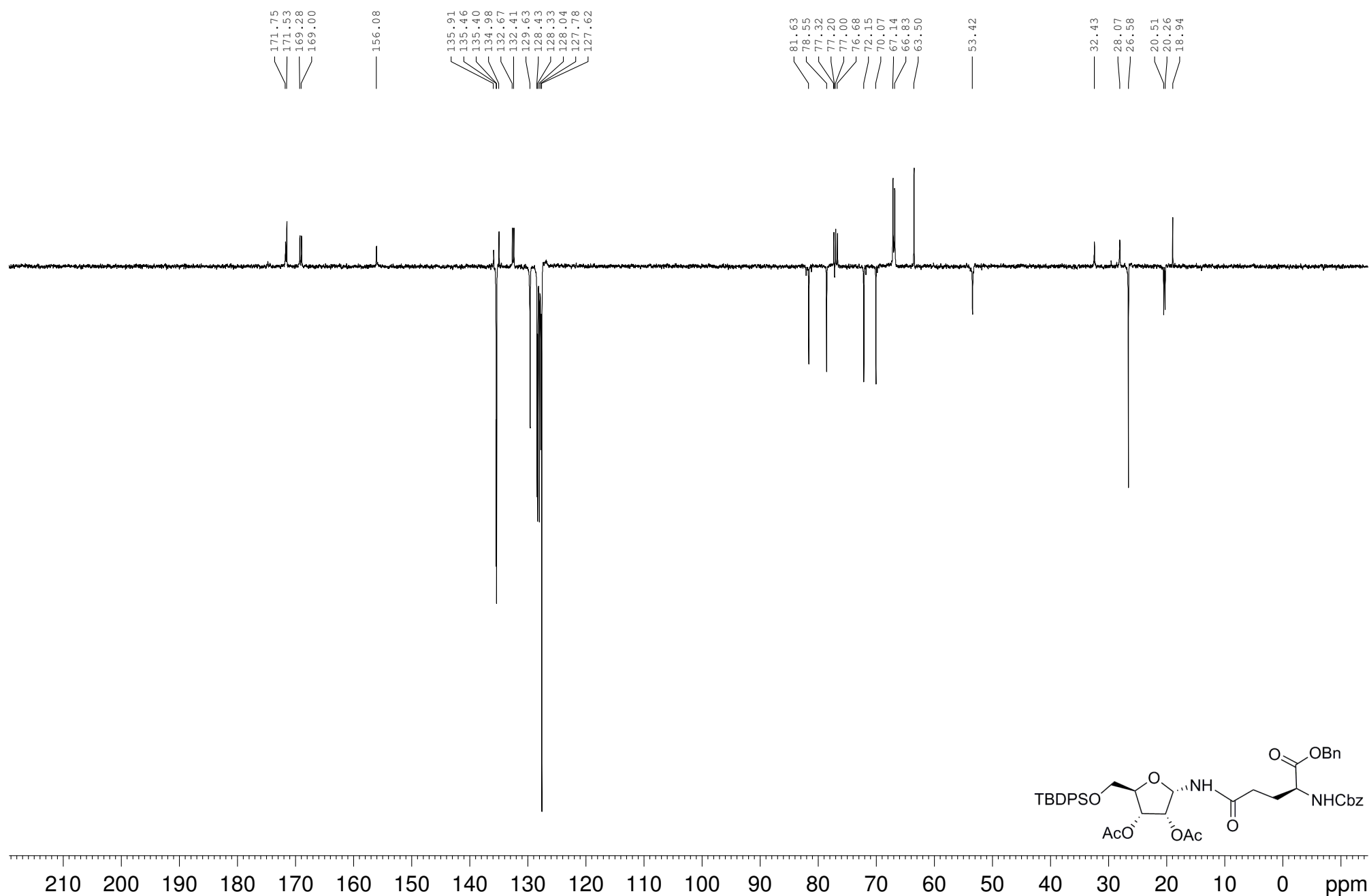
¹H-NMR, 400 MHz, MeOD-d₄, cmpnd 4B



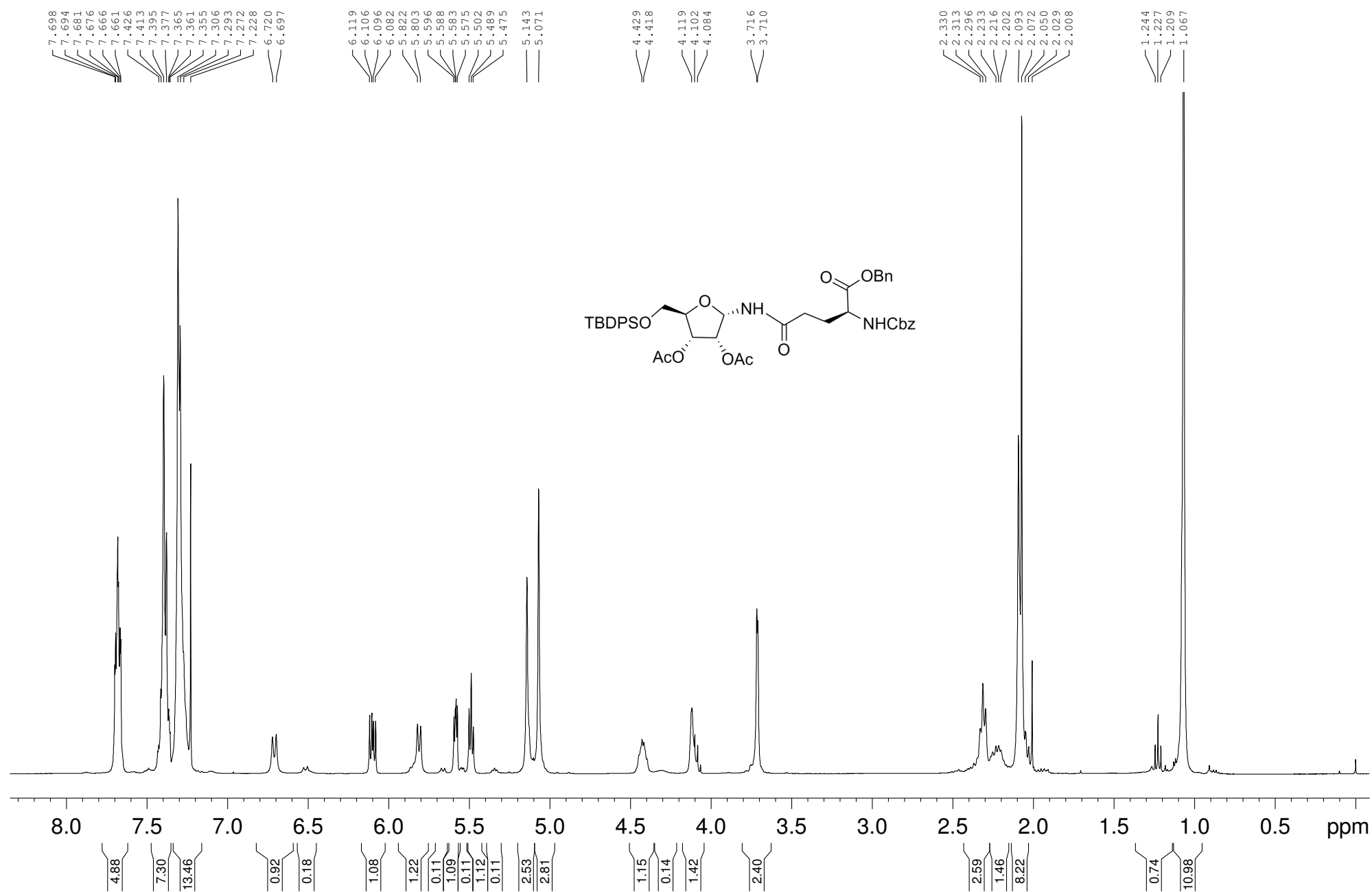
cmpnd 4b, NOESY, CDC13



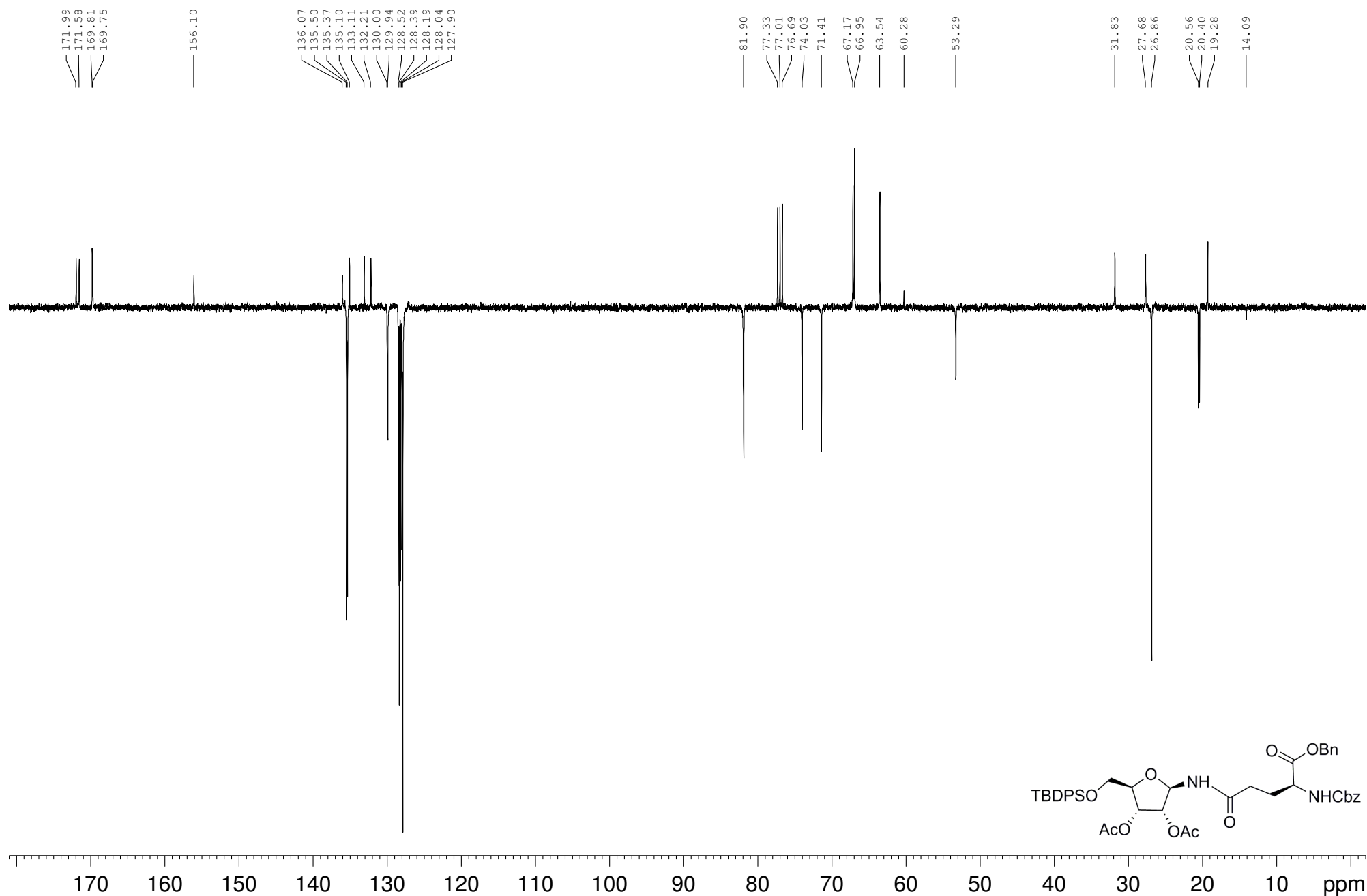
^{13}C -NMR, 100 MHz, CDCl_3 , compnd. 5a



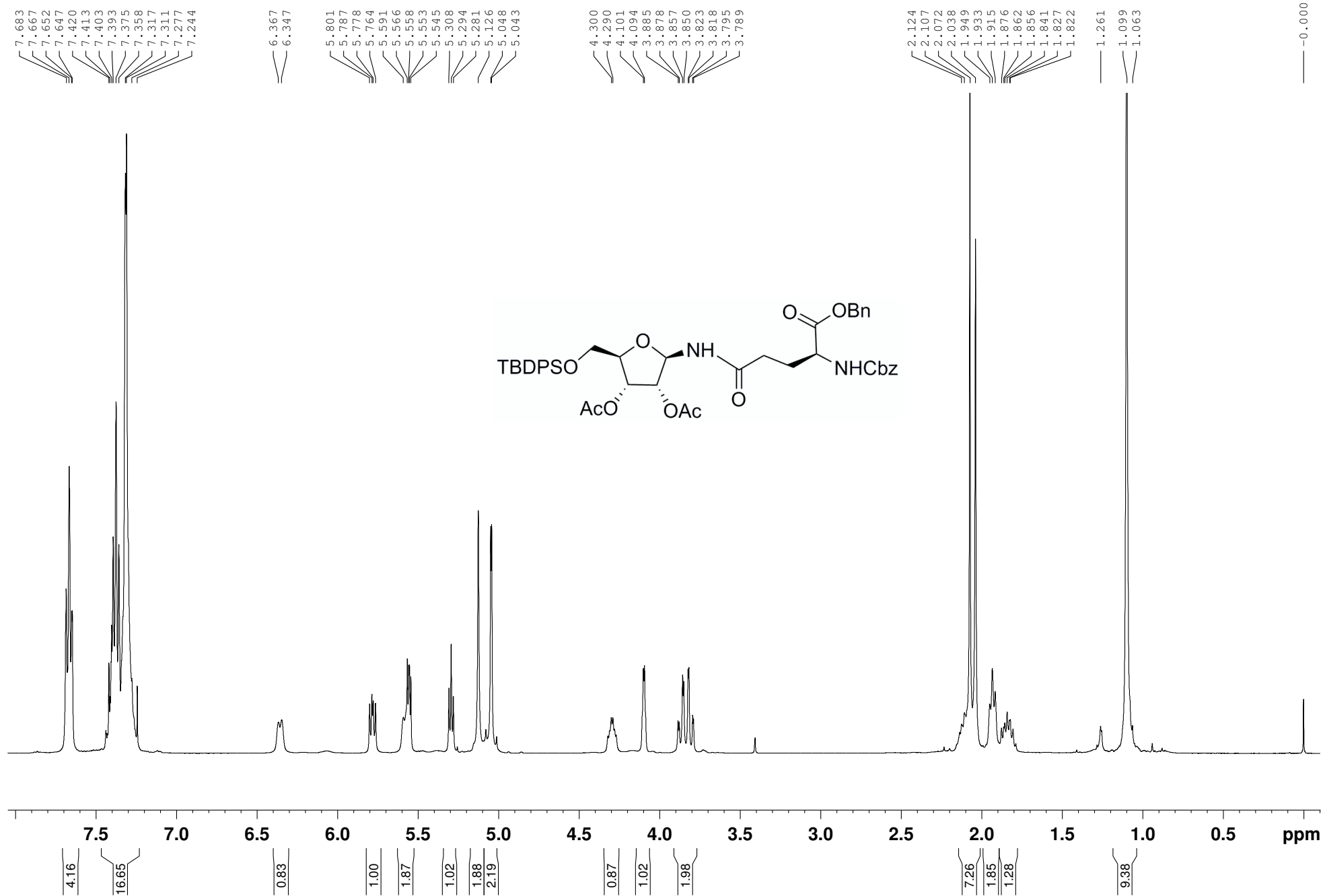
H-NMR, 400 MHz, CDCl₃, cmpnd 5a

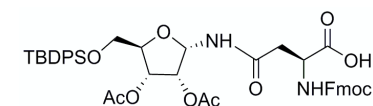


^{13}C -NMR, 100 MHz, CDCl_3 , compnd. 5b

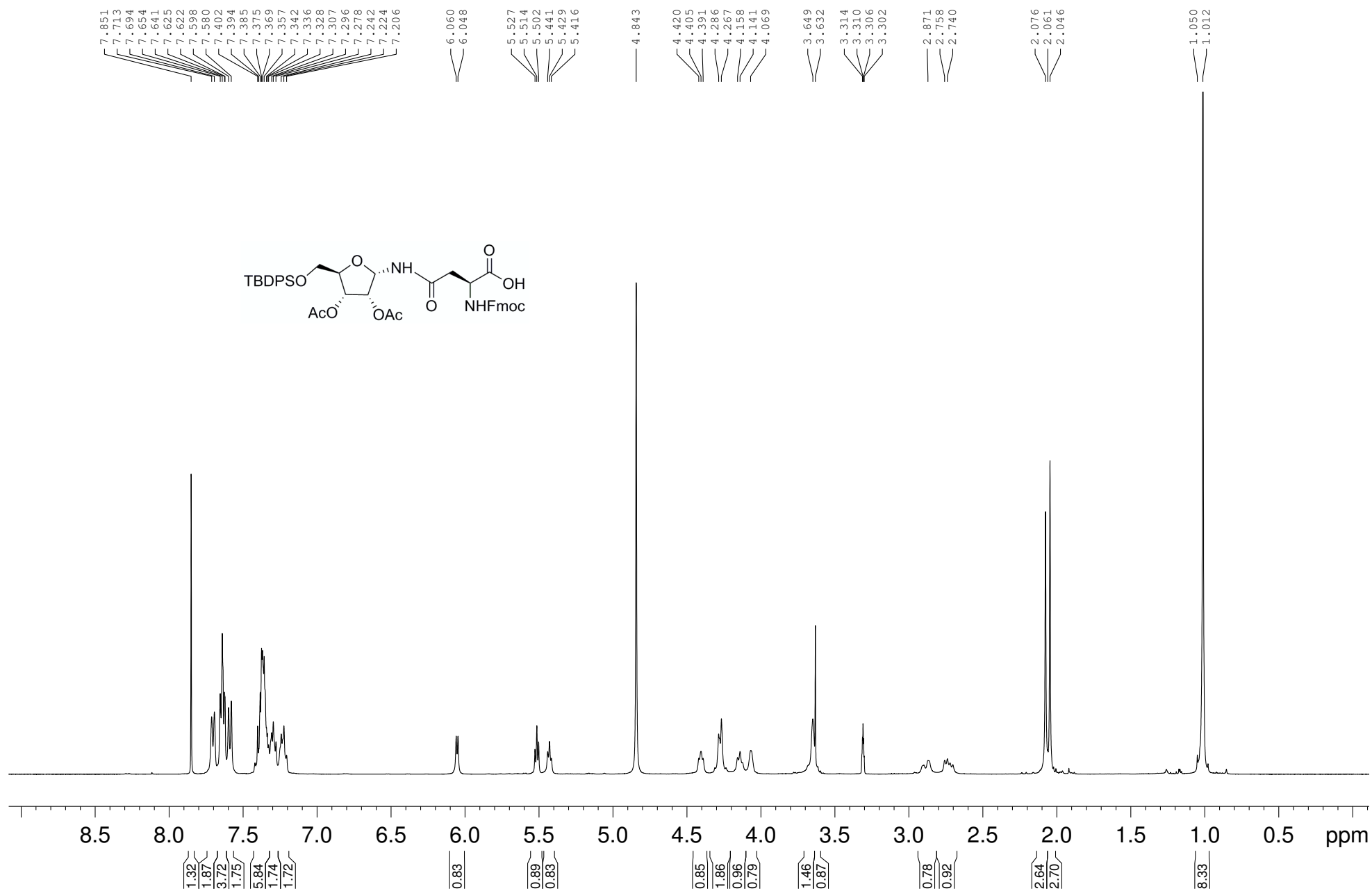


compnd. 5b, 400 MHz, ¹H-NMR, CDCl₃





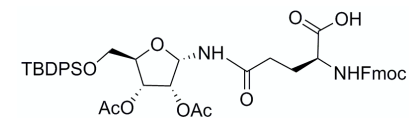
¹H NMR, 400 MHz, MeOD-d₄, cmpnd 6



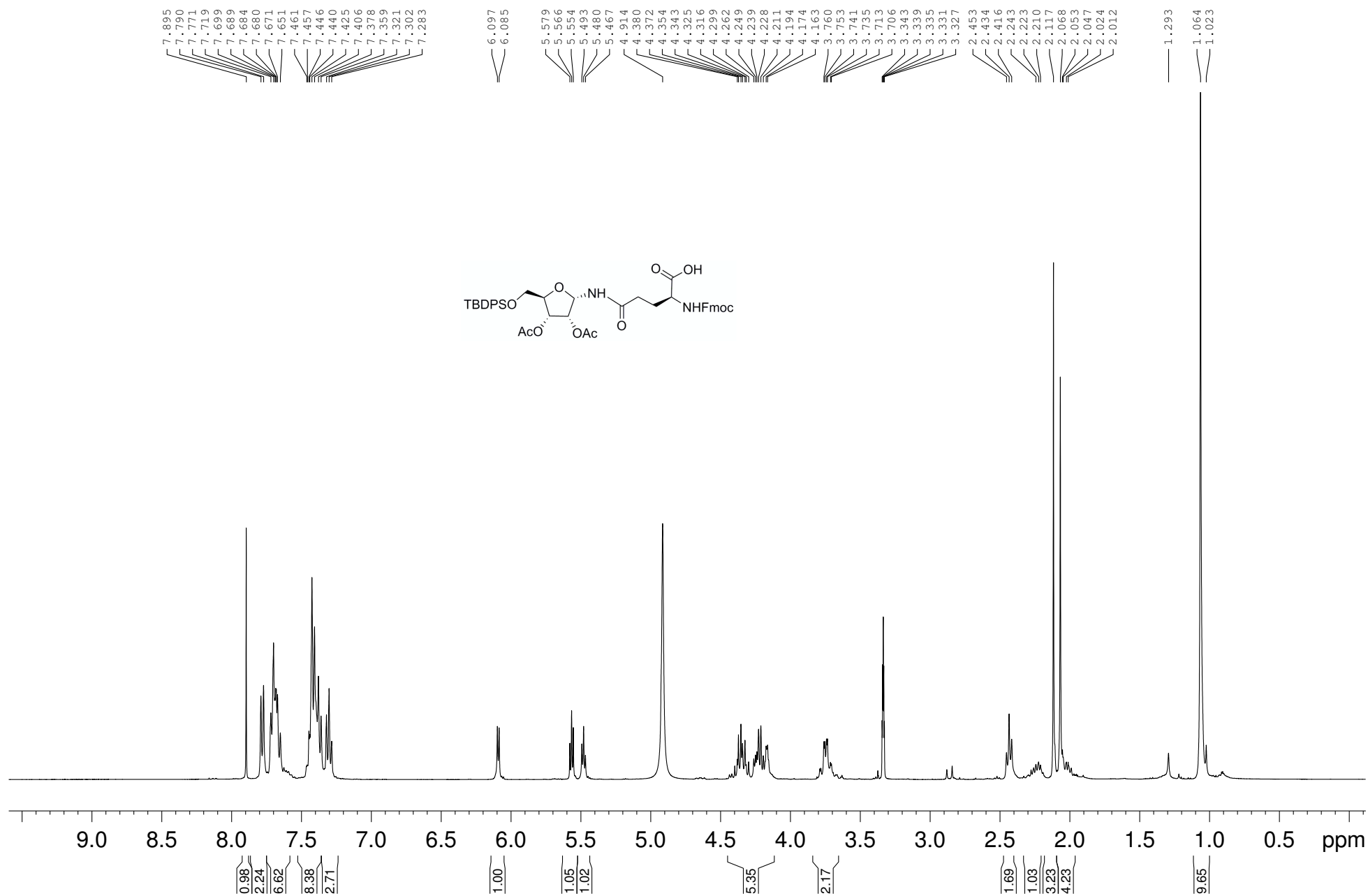
Chemical structure of compound 10 is shown in the bottom right corner. The structure is a complex molecule featuring a central five-membered ring (likely a furanose derivative) substituted with a TBDPSO group, an AcO group, and an NH group. The NH group is part of a side chain that includes a carboxylic acid group (COOH) and an NHFmoc group.

The ^{13}C NMR spectrum displays the following chemical shifts (ppm) for the peaks:

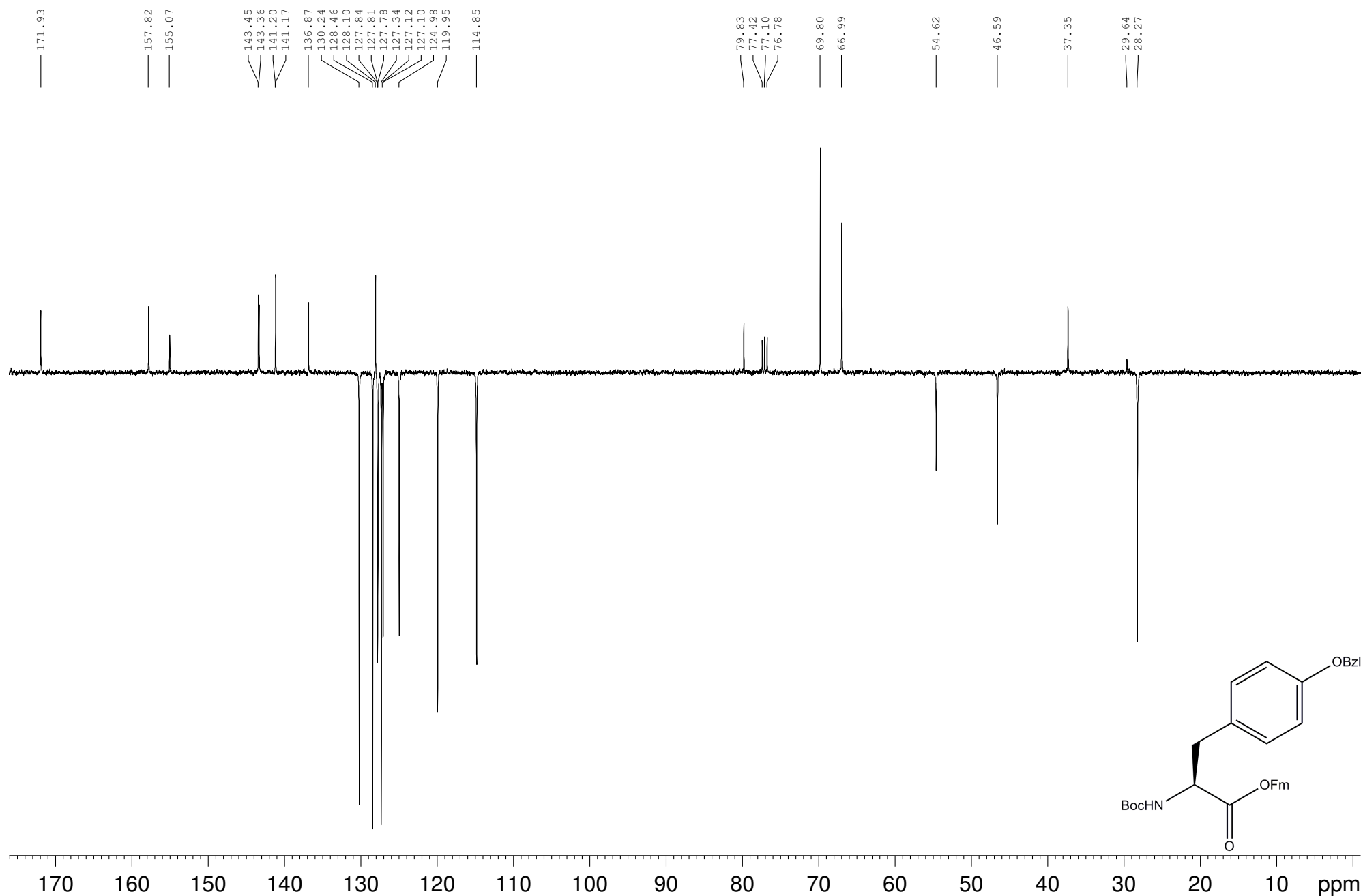
- 173.91, 173.67, 170.23, 169.97
- 157.24
- 143.92, 143.78, 141.17
- 135.40, 135.34, 132.73, 132.60, 129.66, 129.61, 127.55, 127.53, 127.42, 126.80, 124.94, 124.87, 119.56
- 81.11, 80.99, 79.22, 78.08
- 71.50, 70.67
- 67.53, 66.66
- 63.38
- 53.44, 48.30, 48.08, 47.87, 47.66, 47.45, 47.23, 47.02, 46.98
- 32.03
- 27.26, 25.96
- 19.29, 19.13, 18.64



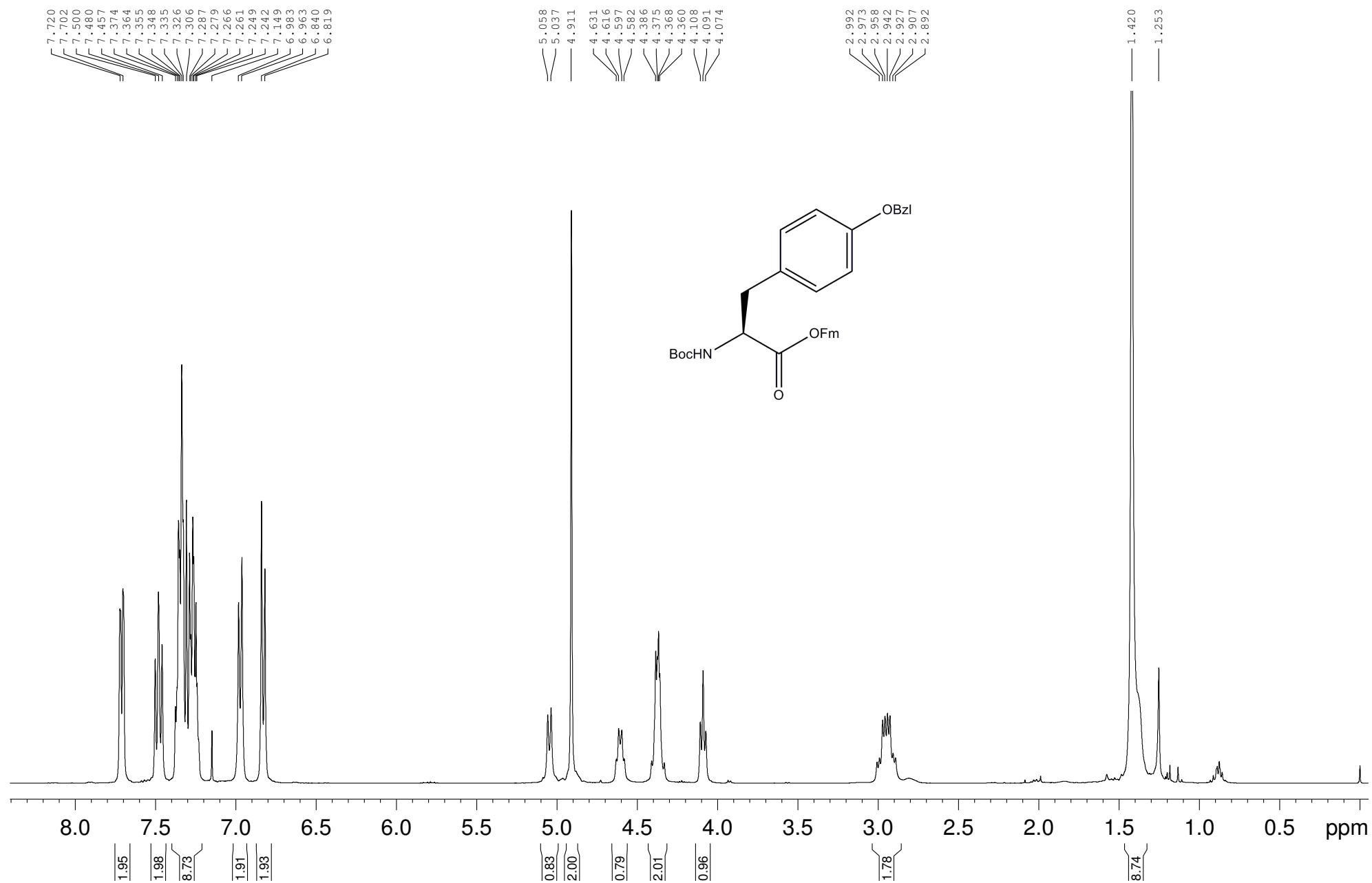
¹H-NMR, 400 MHz, MeOD-d₄, compnd 7



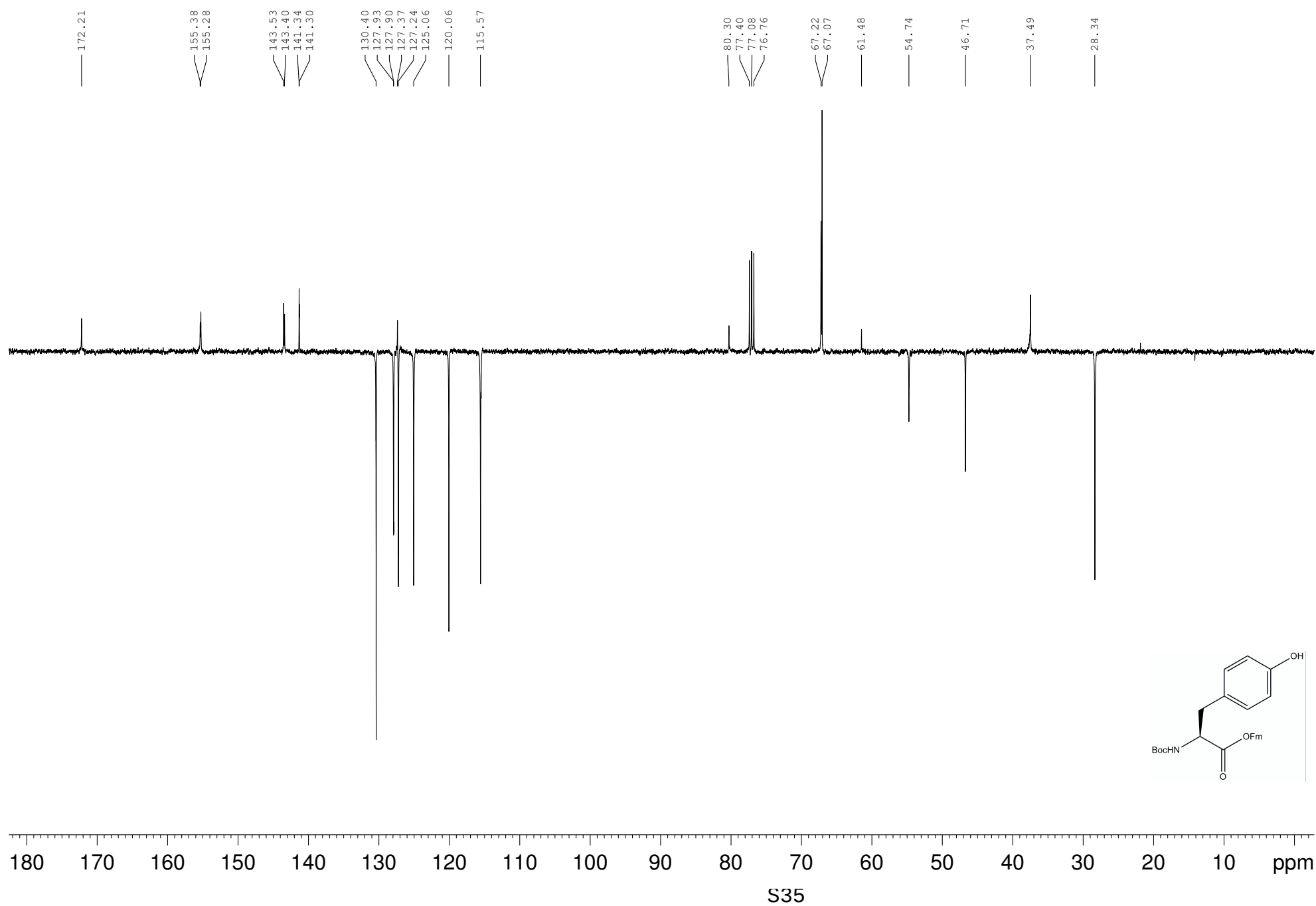
^{13}C -NMR, 100 MHz, CDCl_3 , compnd. 8



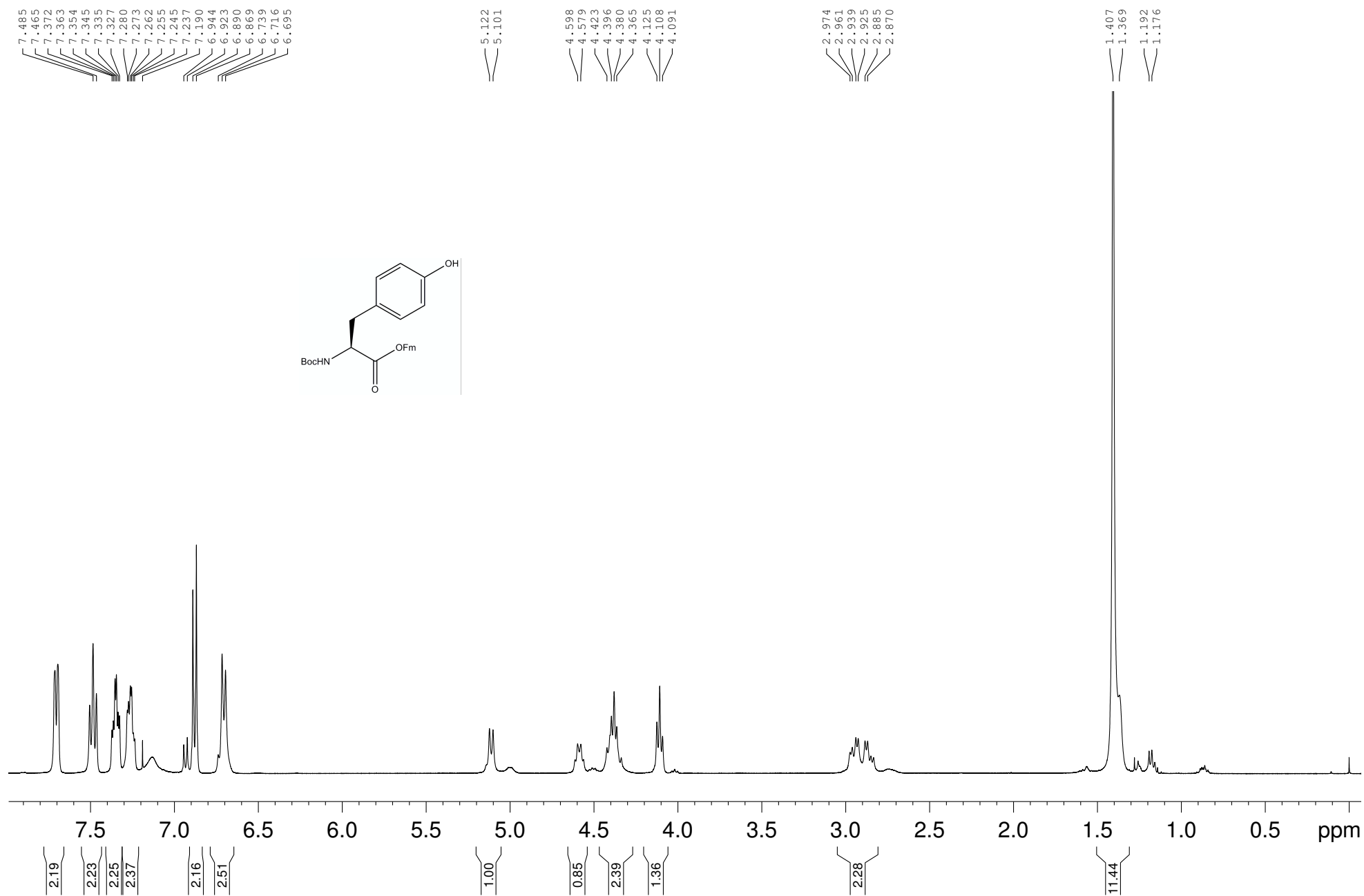
H-NMR, 400 MHz, CDCl₃, compnd. 8



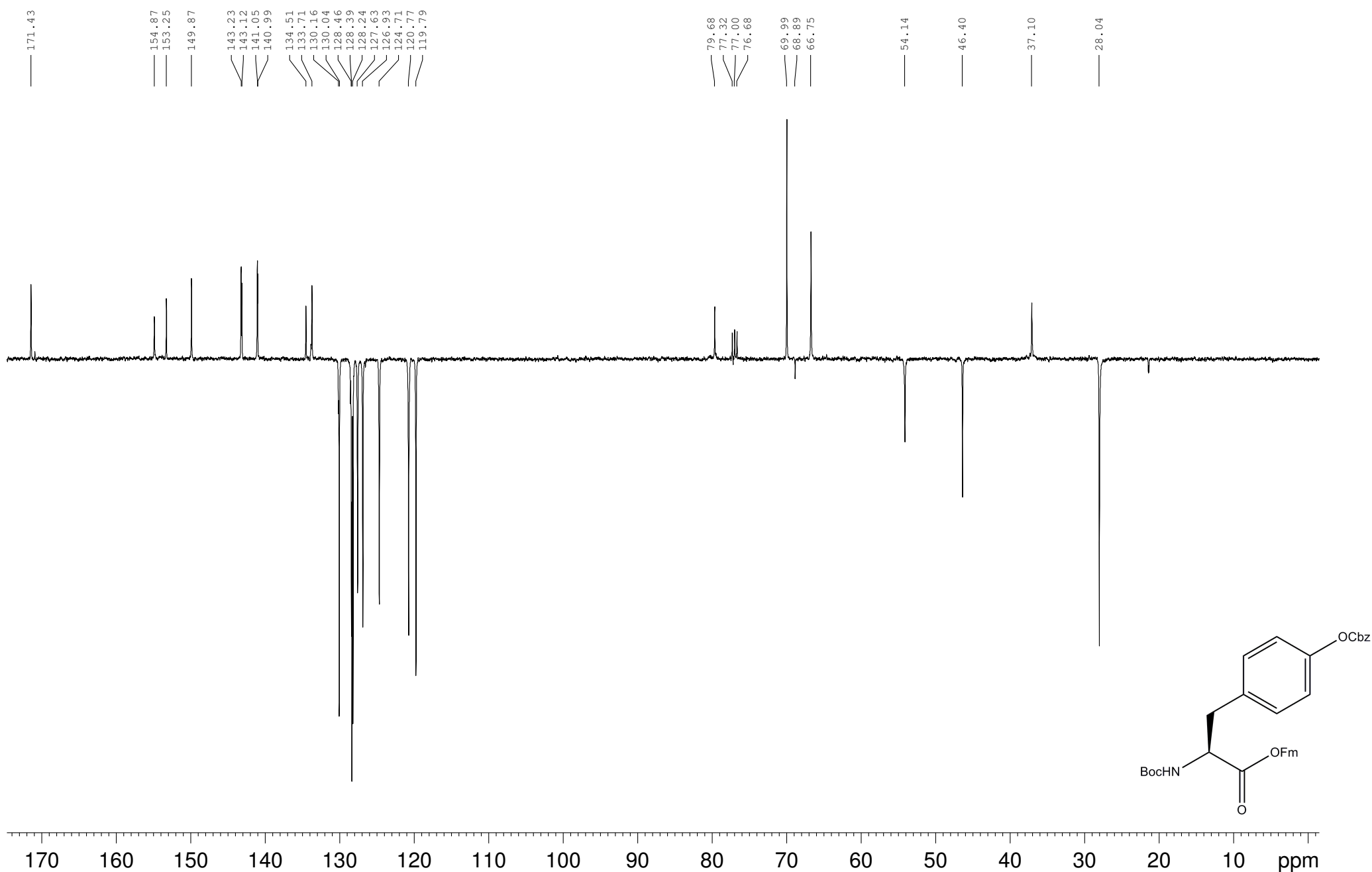
^{13}C -NMR, 100 MHz, CDCl_3 , compnd 9



^1H -nmr, 400 MHz, CDCl_3 , compnd 9



¹³C-NMR, 100 MHz, CDCl₃, compnd.10



¹H-NMR, 400 MHz, CDCl₃, compnd 10

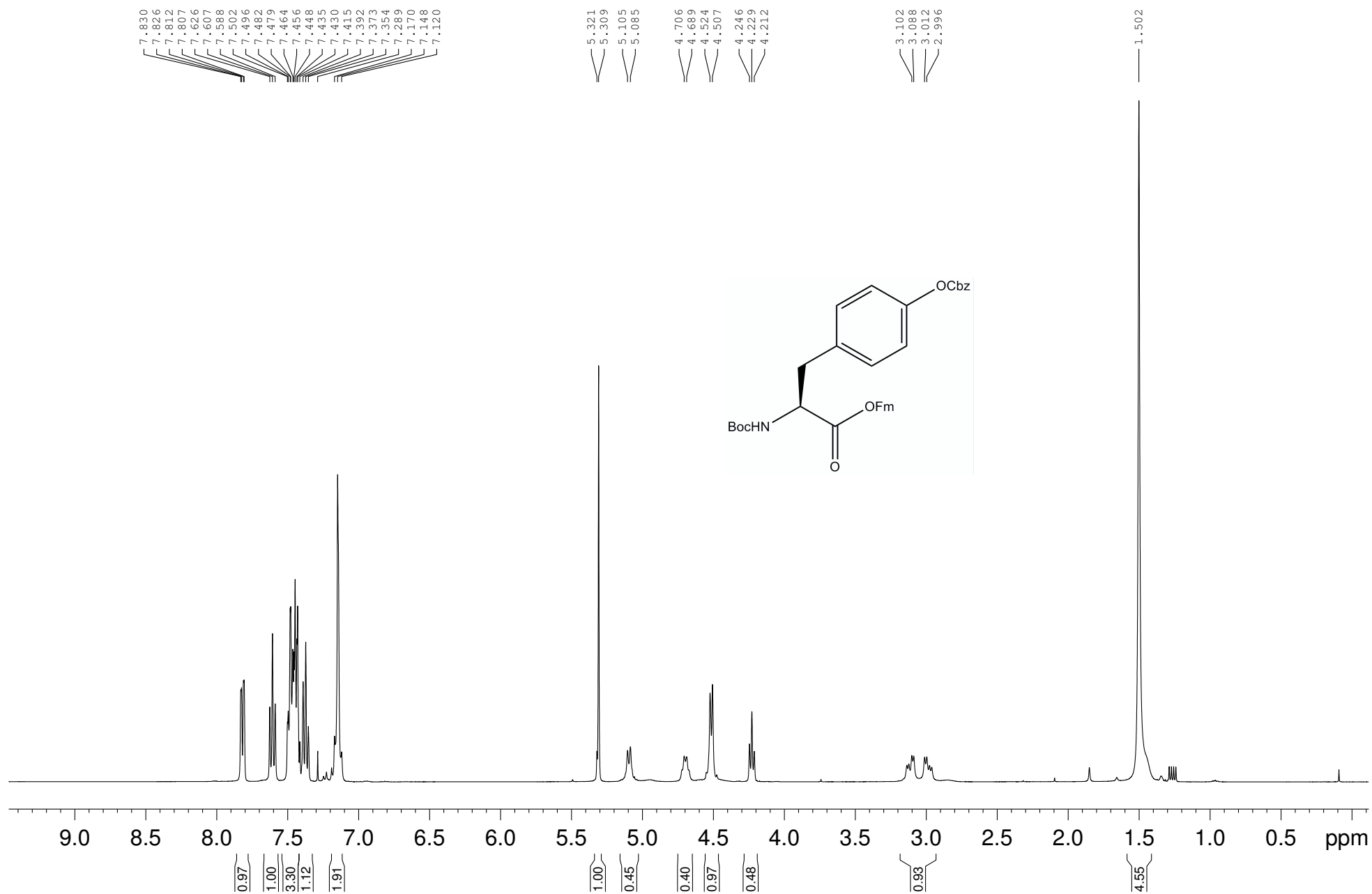
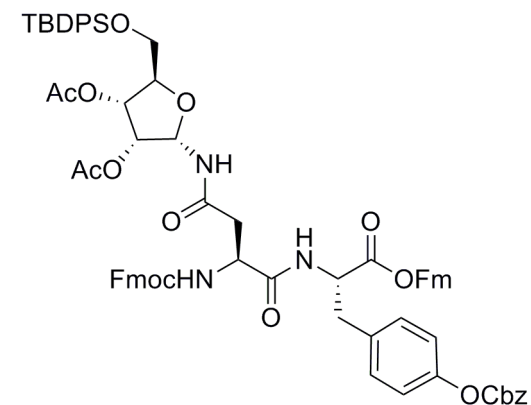
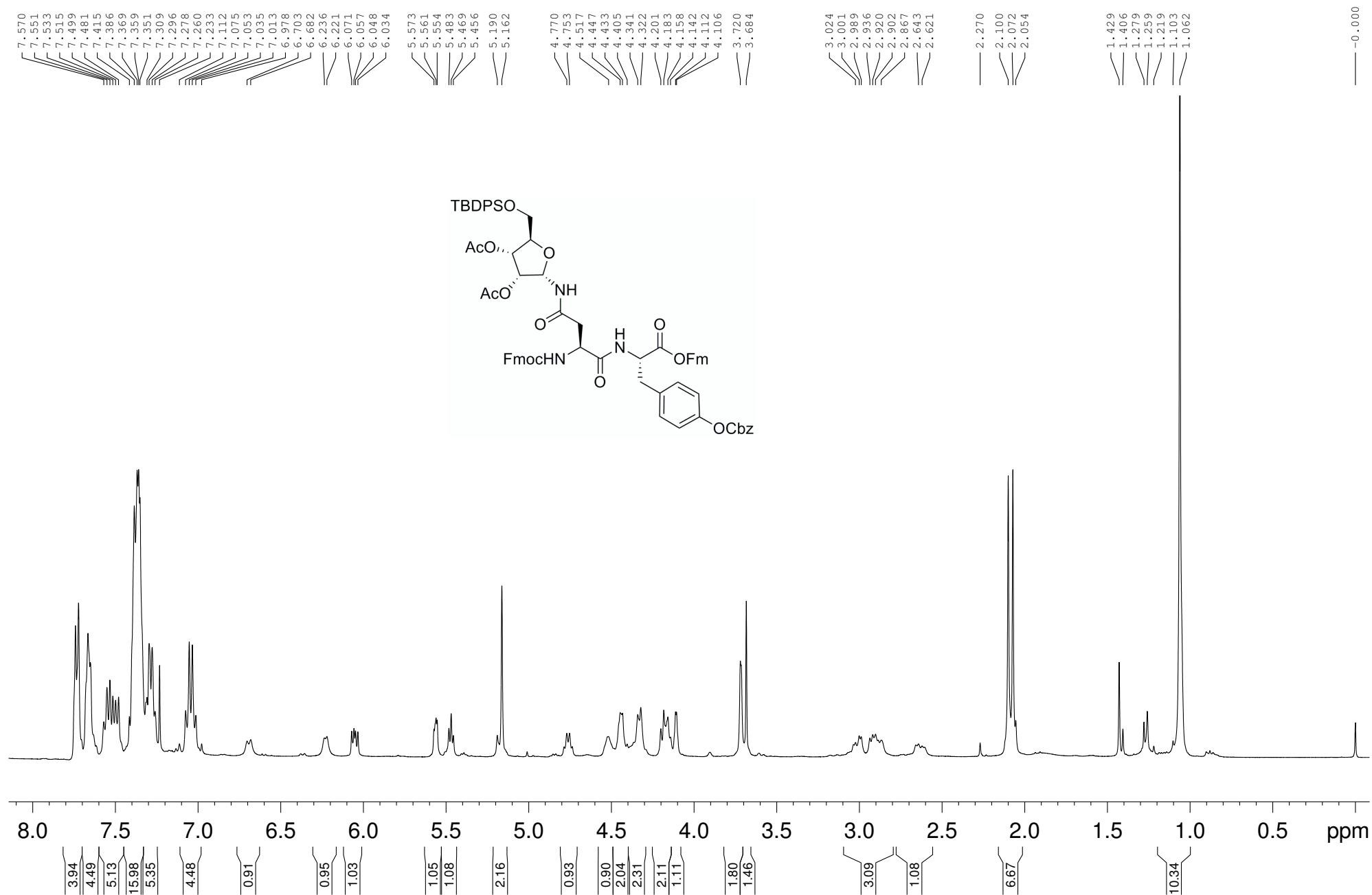


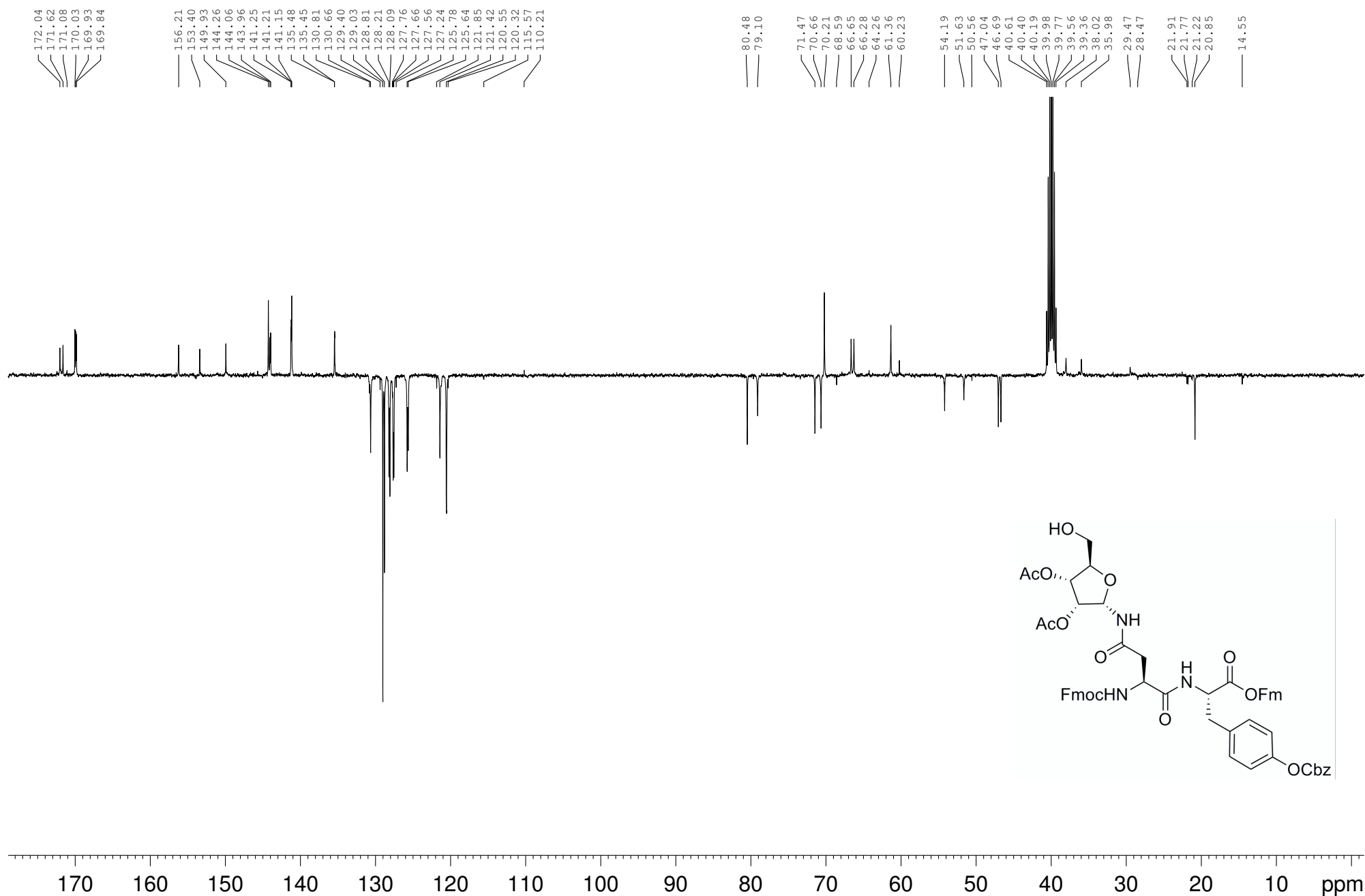
Figure 1: Evolution of the average number of nodes per cluster, N_c , as a function of the number of nodes, N , for different values of the parameter α . The curves show that N_c increases sharply with N , and the values of N_c are higher for larger α .



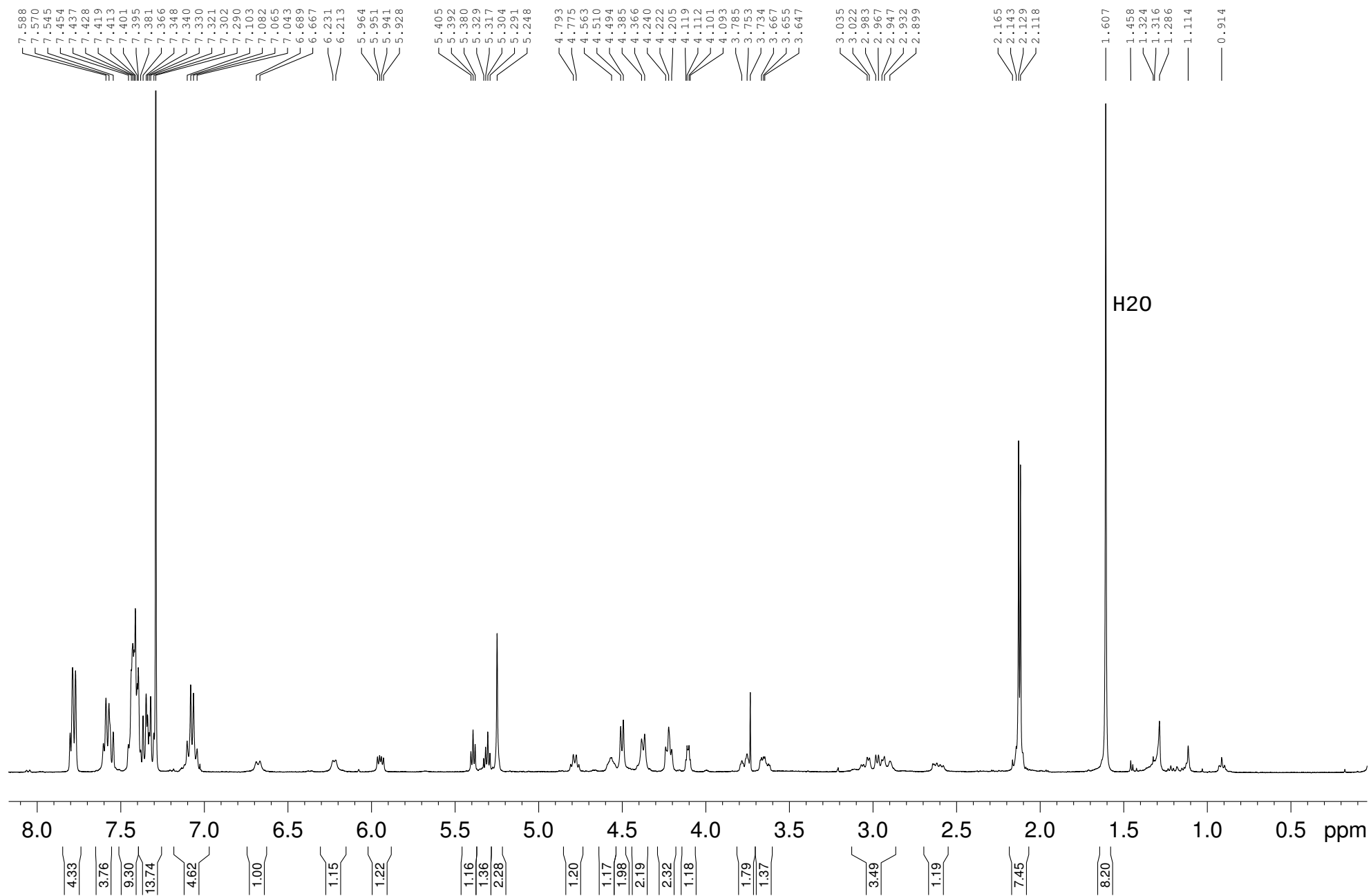
¹H-NMR, 400 MHz, MeOD-d₄/CDCl₃, compnd 11



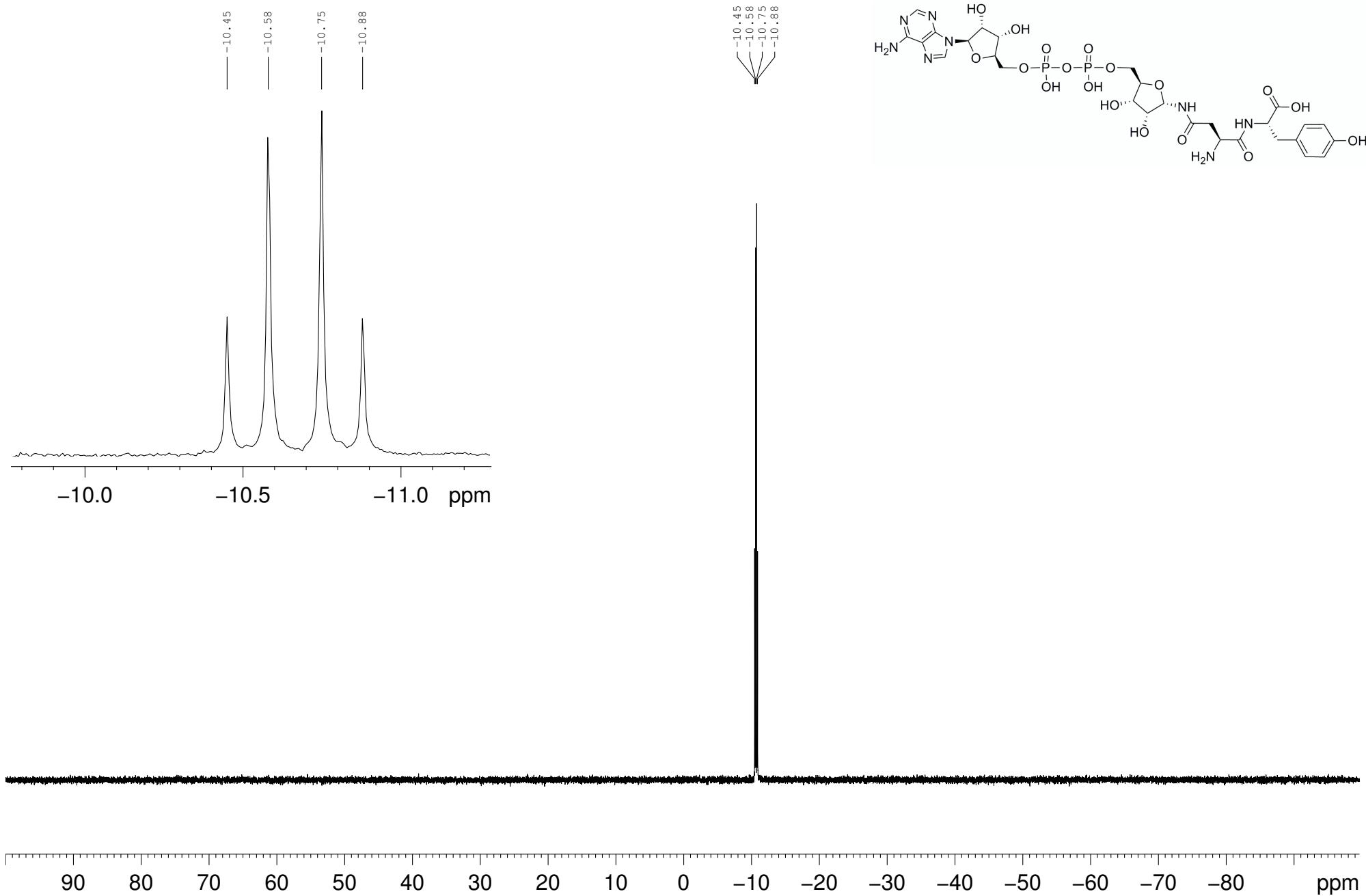
^{13}C -NMR, 100 MHz, DMSO- d_6 , cpmnd 12



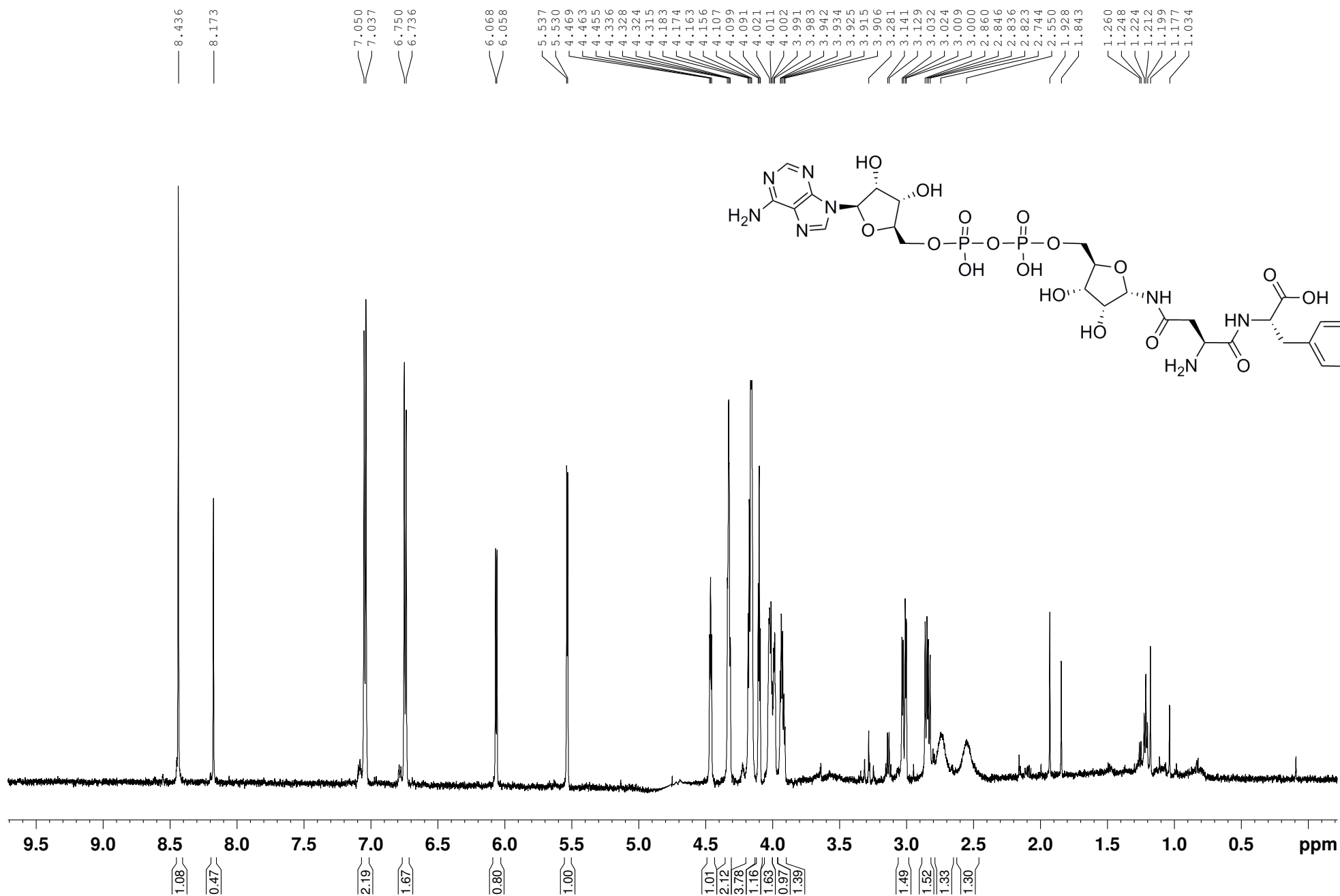
¹H-NMR, 400 MHz, CDCl₃, compnd 12



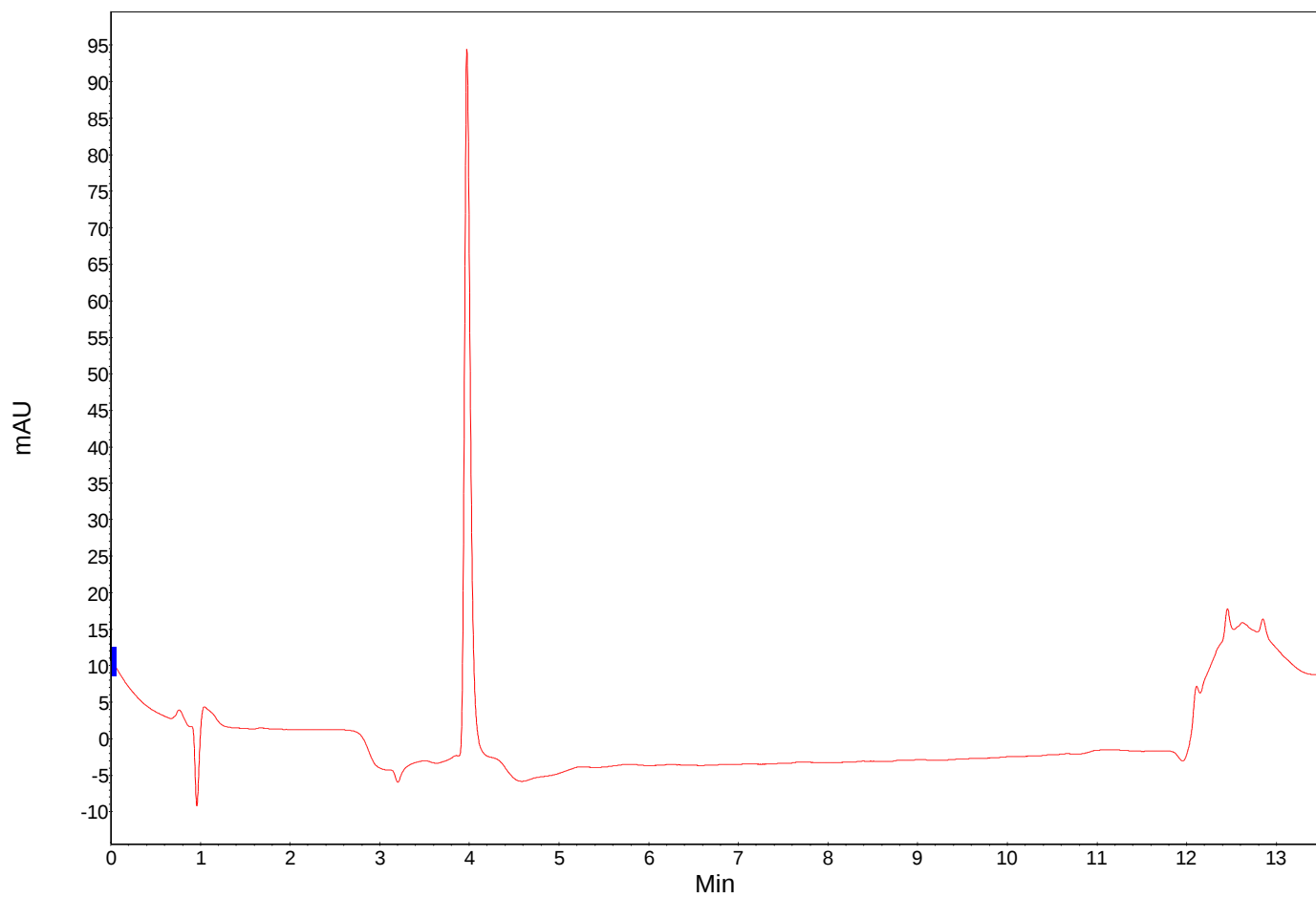
31P-NMR, D2O, 162 MHz, cpmnd 18



H-NMR, D₂O, 600 MHz, compnd. 18



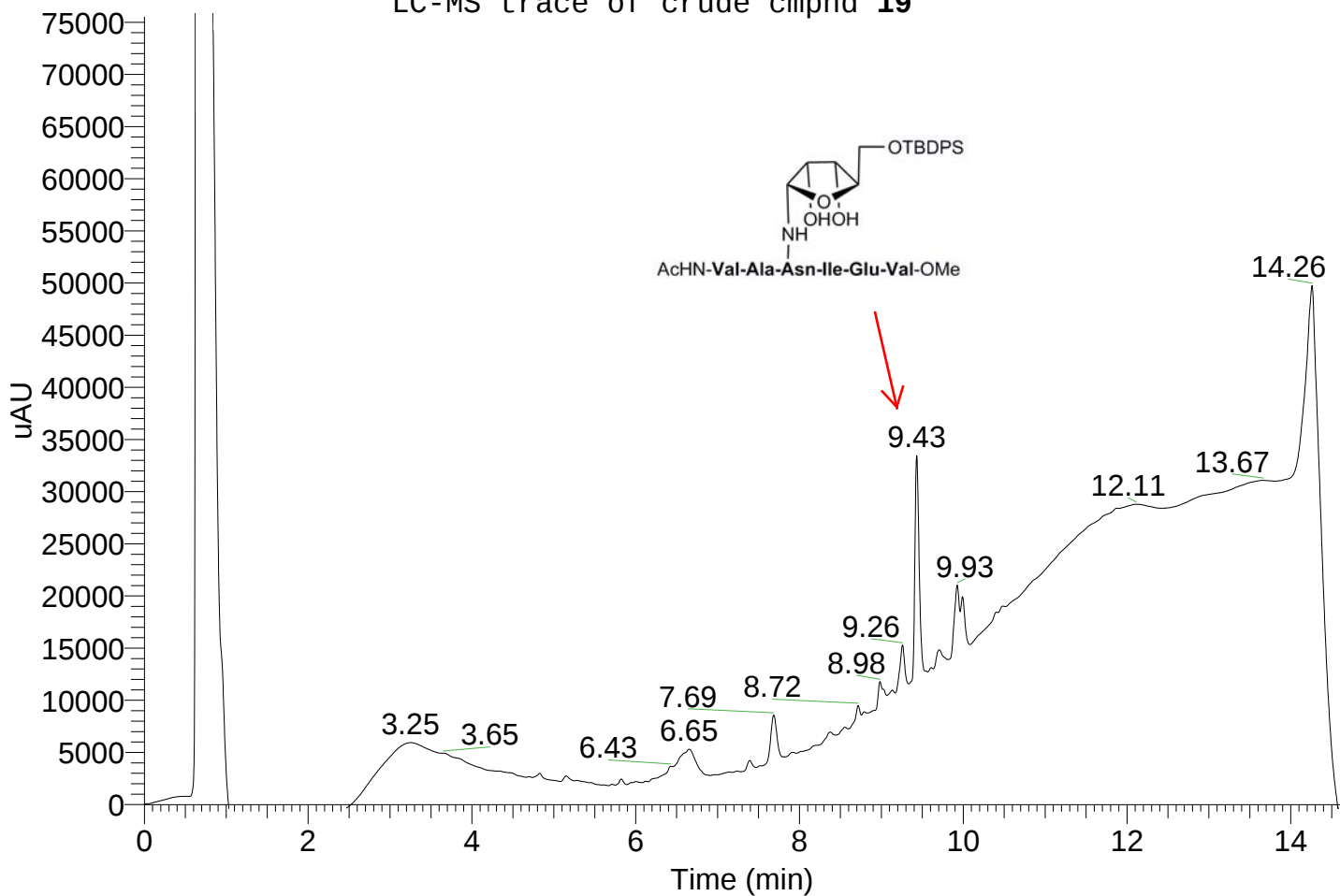
cmpnd 18, LC-MS (0 -50% B in 13.5 min), UV detection at 214 nm



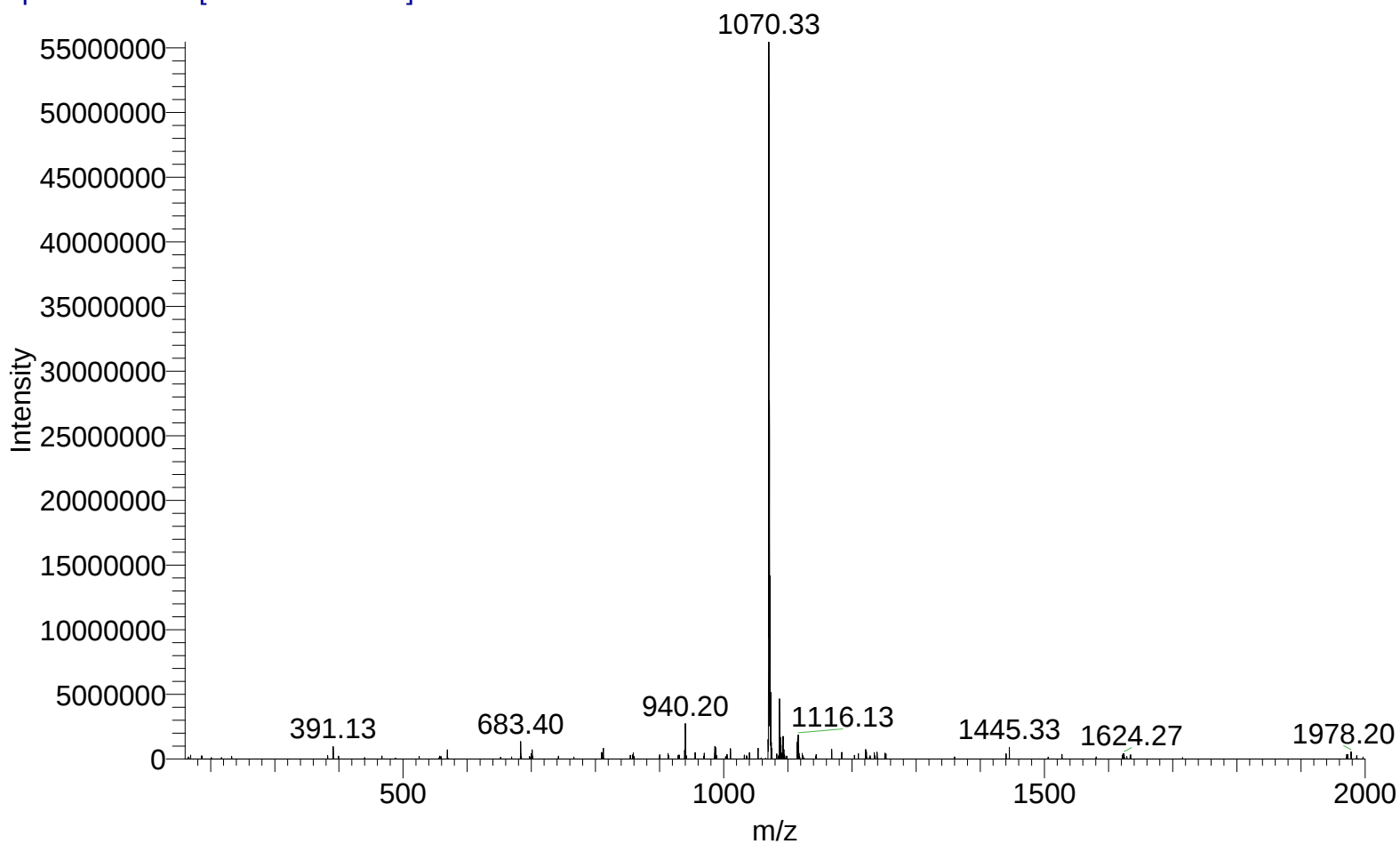
RT: 0.00 - 14.60

LC-MS trace of crude cmpnd **19**

NL:
3.02E5
Total Scan
PDA
0.00 - 14.60



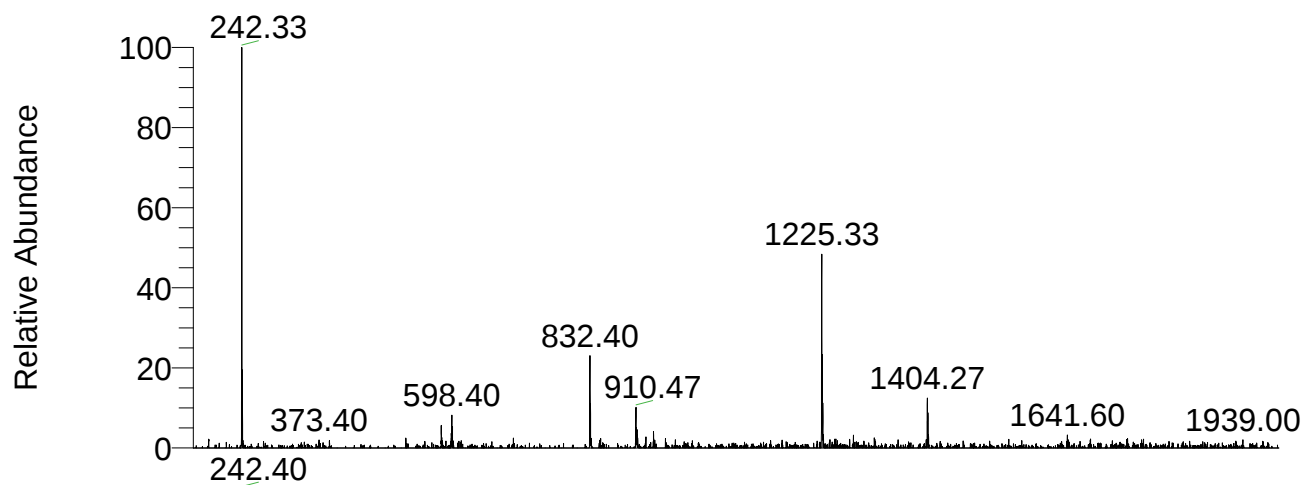
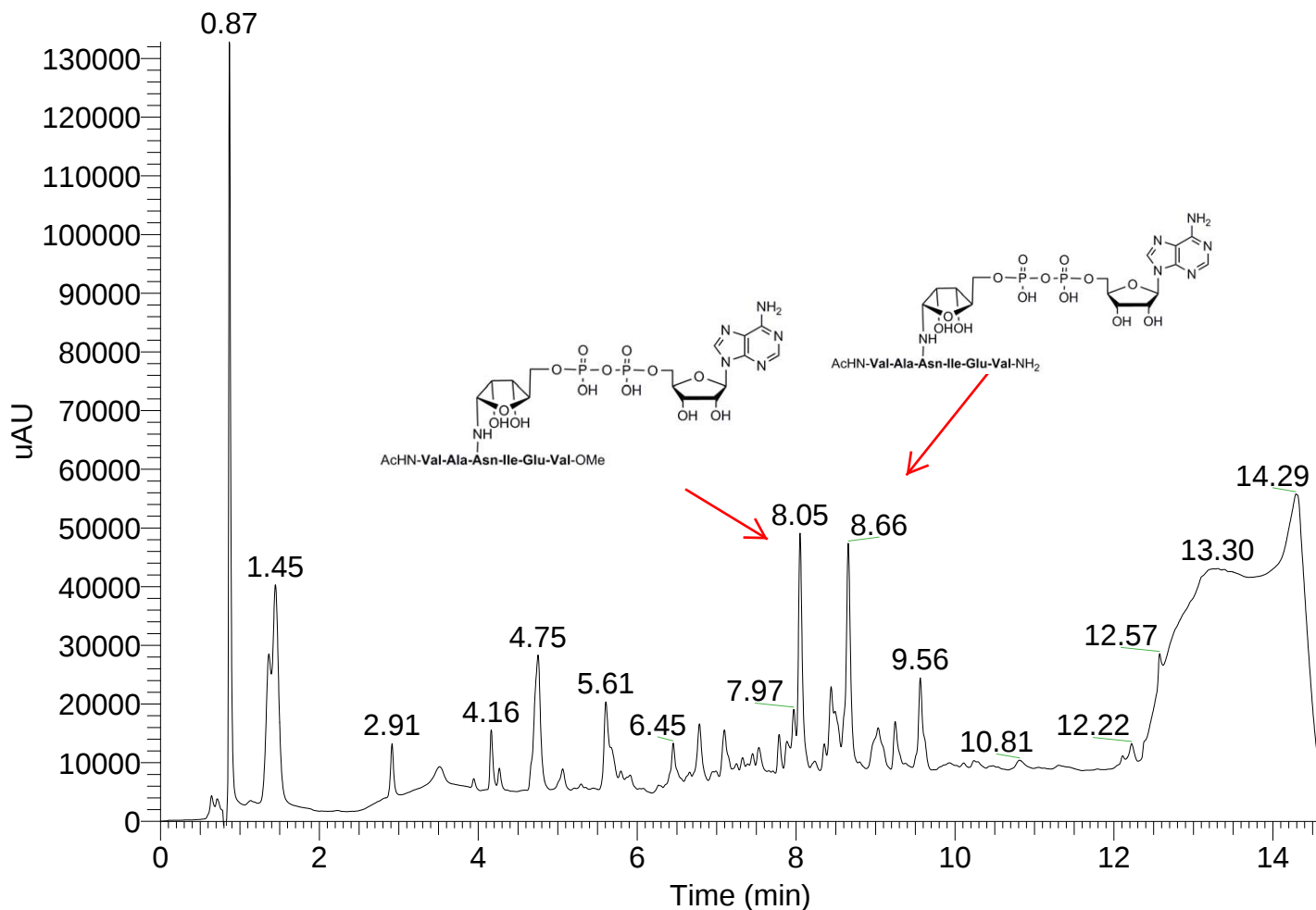
RT: 9.44 AV: 1 NL: 5.55E7
T: + p ESI Full ms [160.00-2000.00]



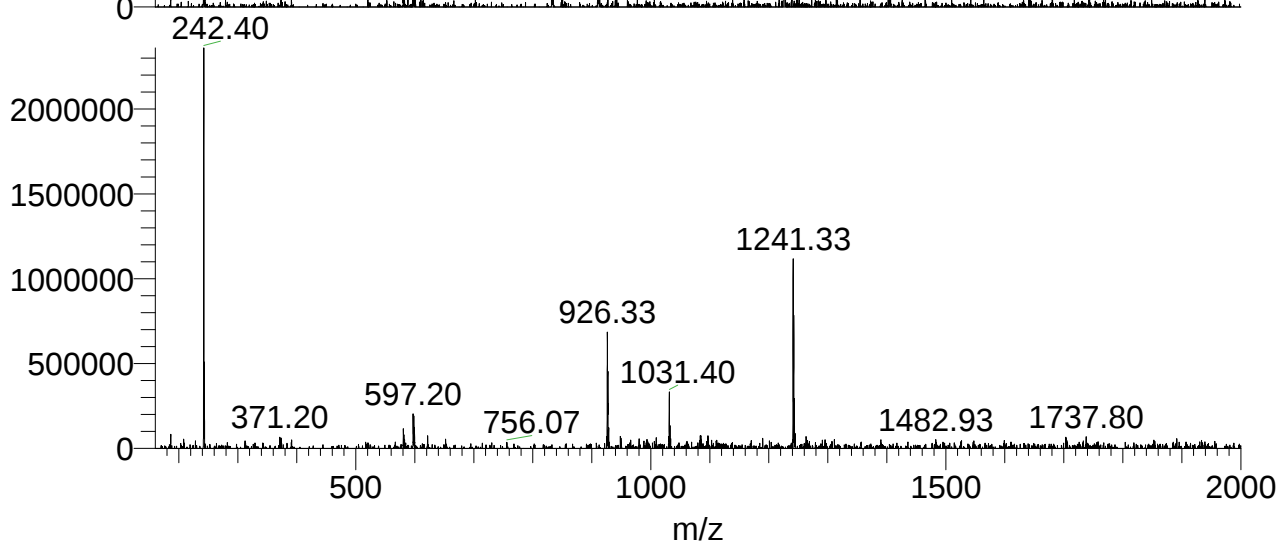
LC-MS trace of crude cmpnd. **21**

RT: 0.00 - 14.60

NL:
1.33E5
Total Scan
PDA

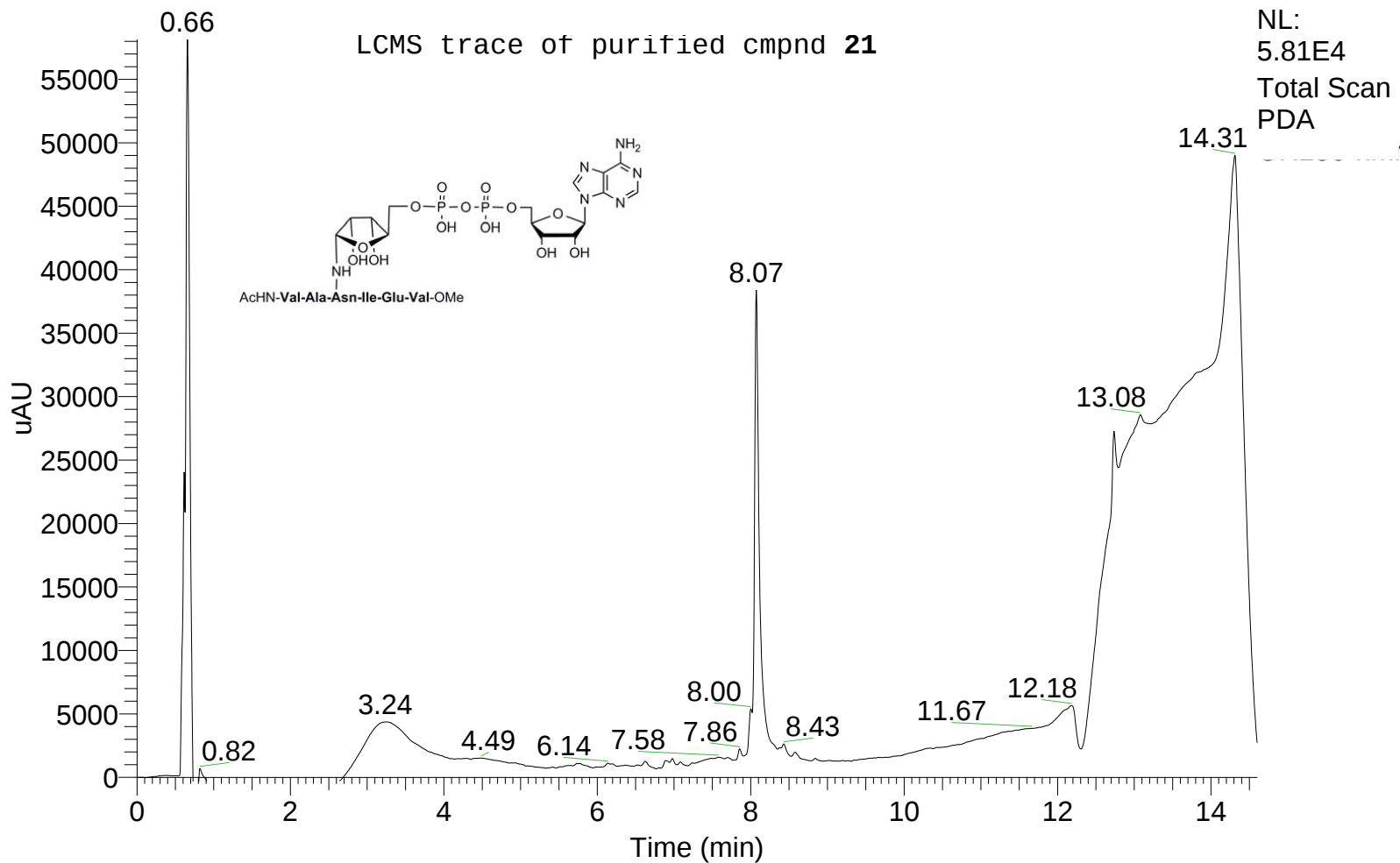


NL:
2.41E6
crude#450 RT:
8.66 AV: 1 T: + p
ESI Full ms
[160.00-2000.00]

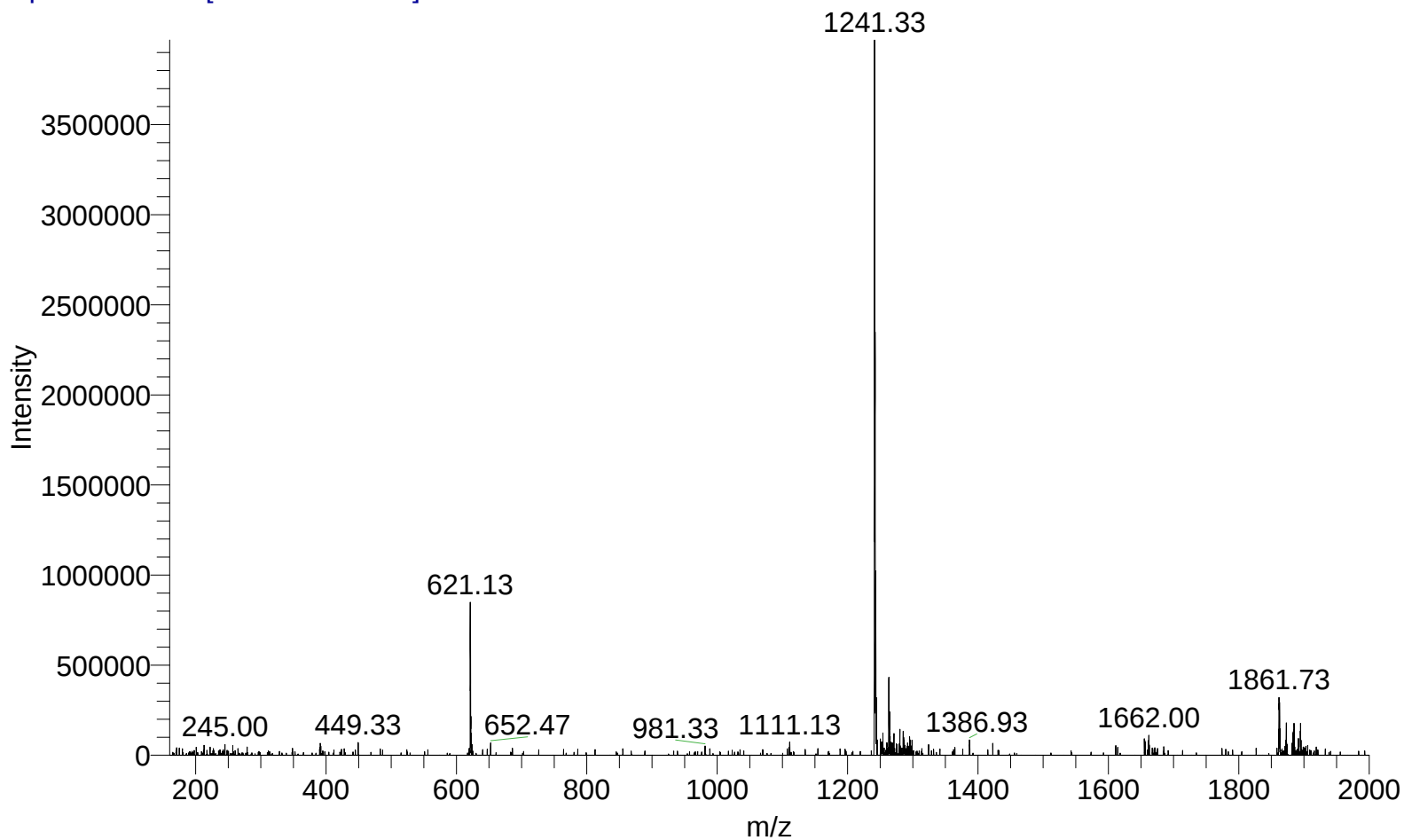


NL:
2.36E6
crude#418 RT:
8.04 AV: 1 T: + p
ESI Full ms
[160.00-2000.00]

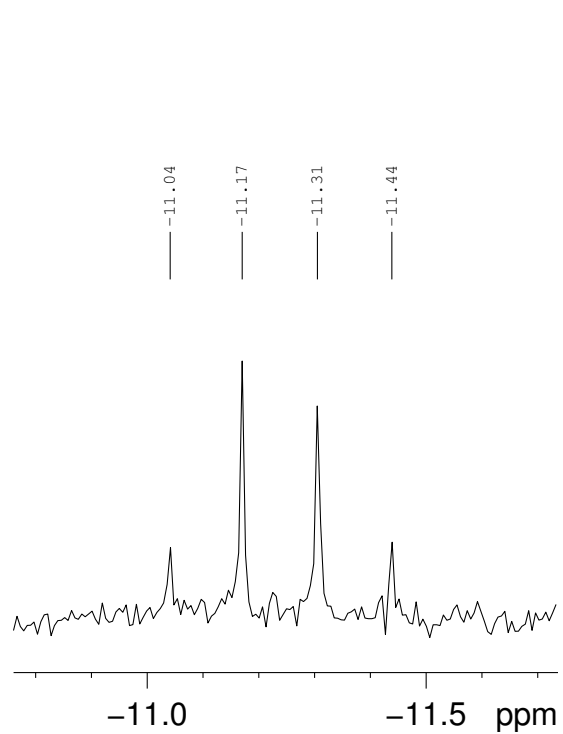
RT: 0.00 - 14.60



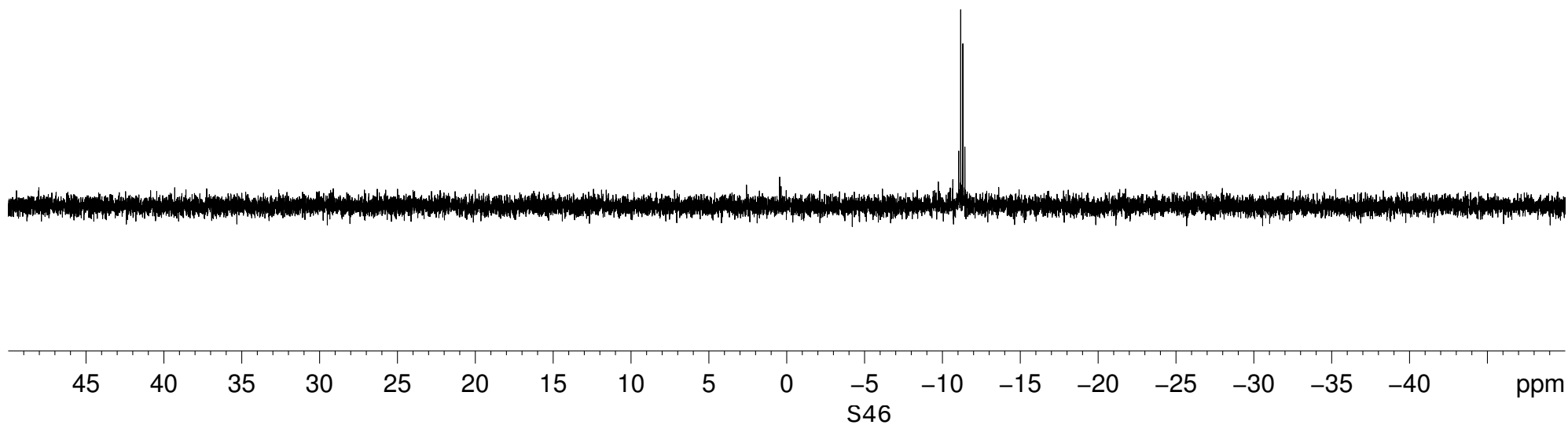
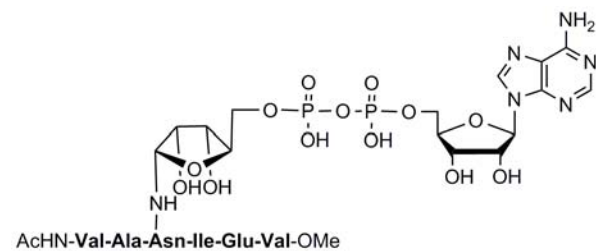
RT: 8.09 AV: 1 NL: 3.97E6
T: + p ESI Full ms [160.00-2000.00]



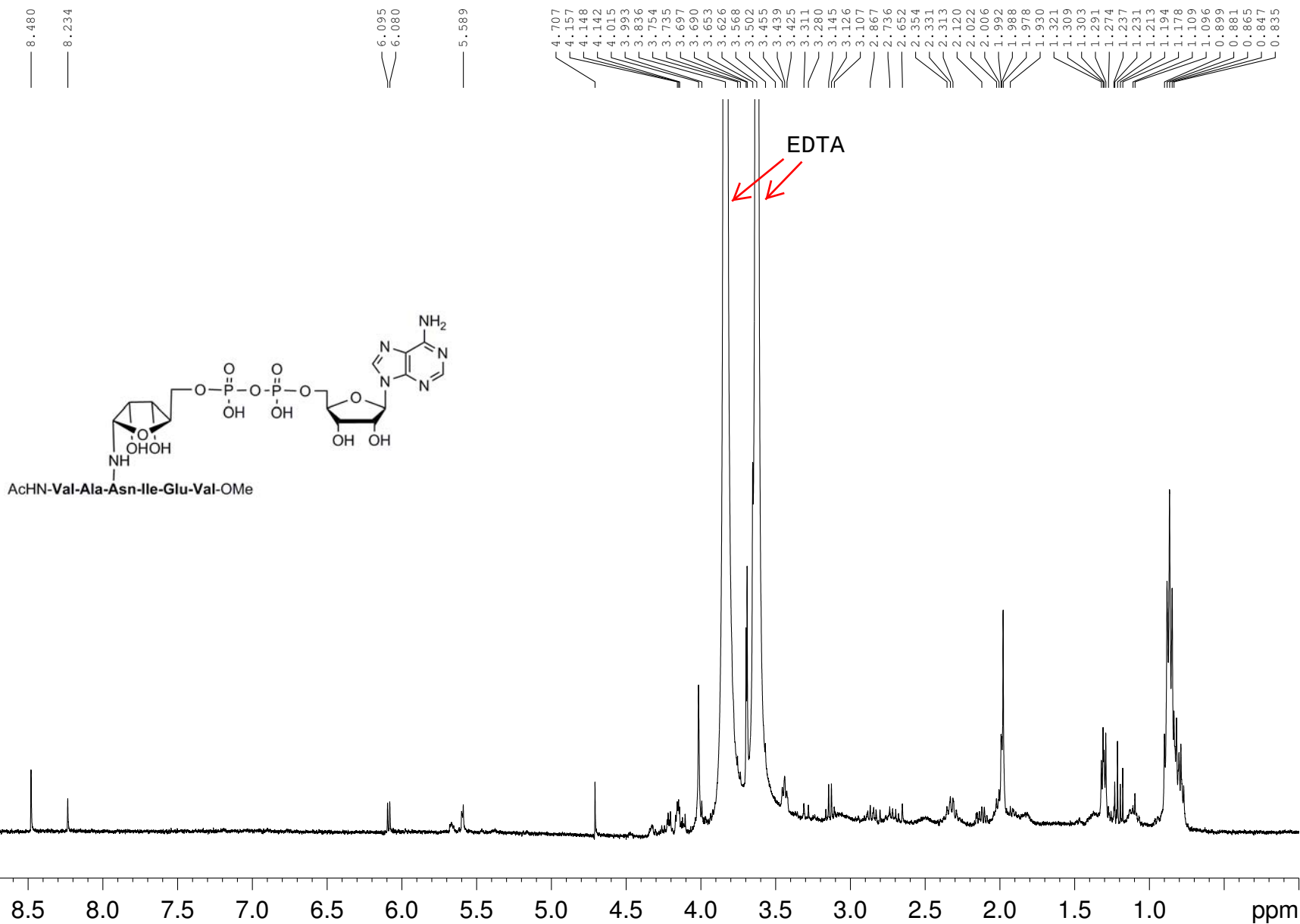
31p-nmr. 162 MHz, D2O, cmpnd 21



-11.04
-11.17
-11.31
-11.44

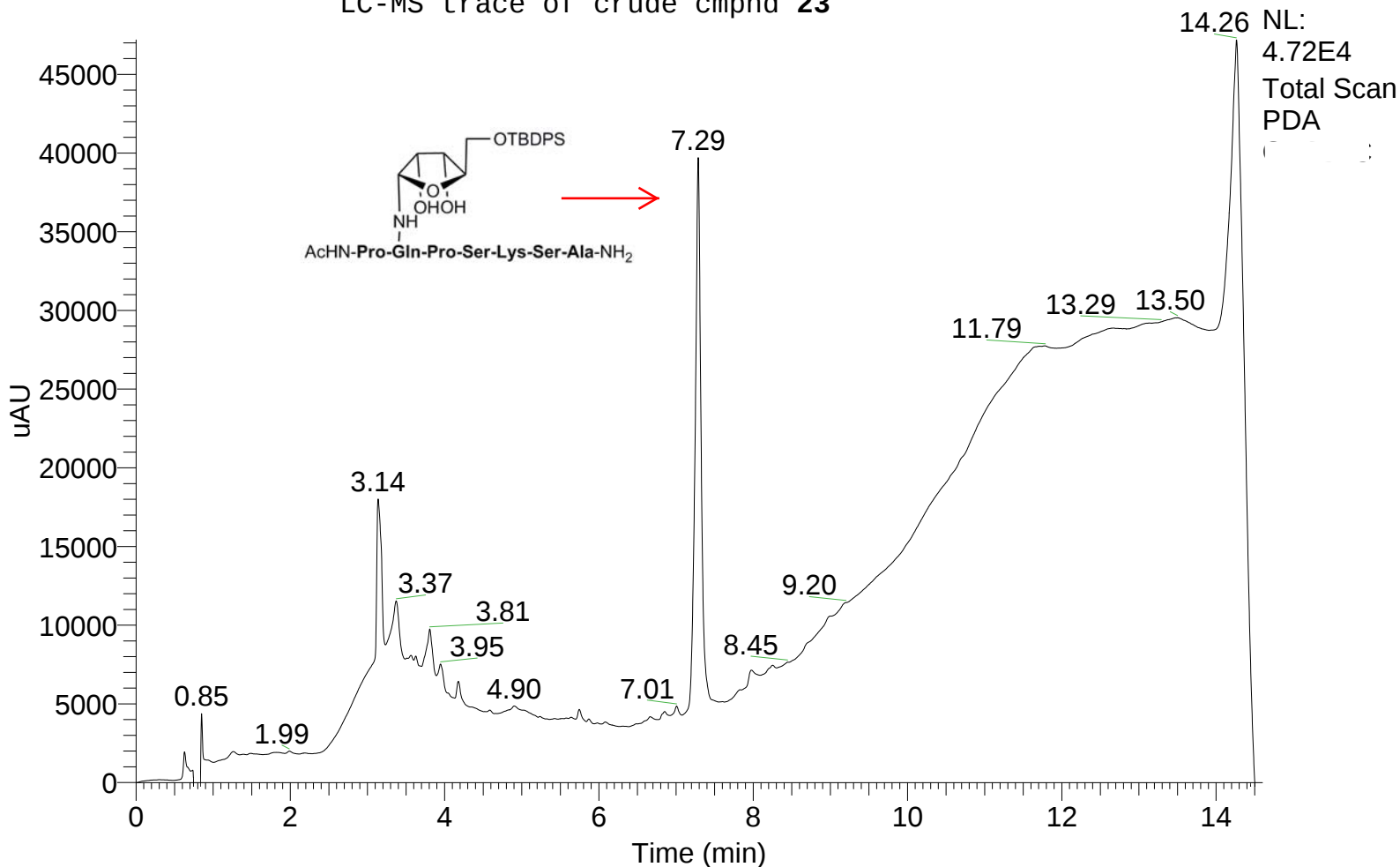


¹H-NMR, 400 MHz, D₂O, cmpnd 21H

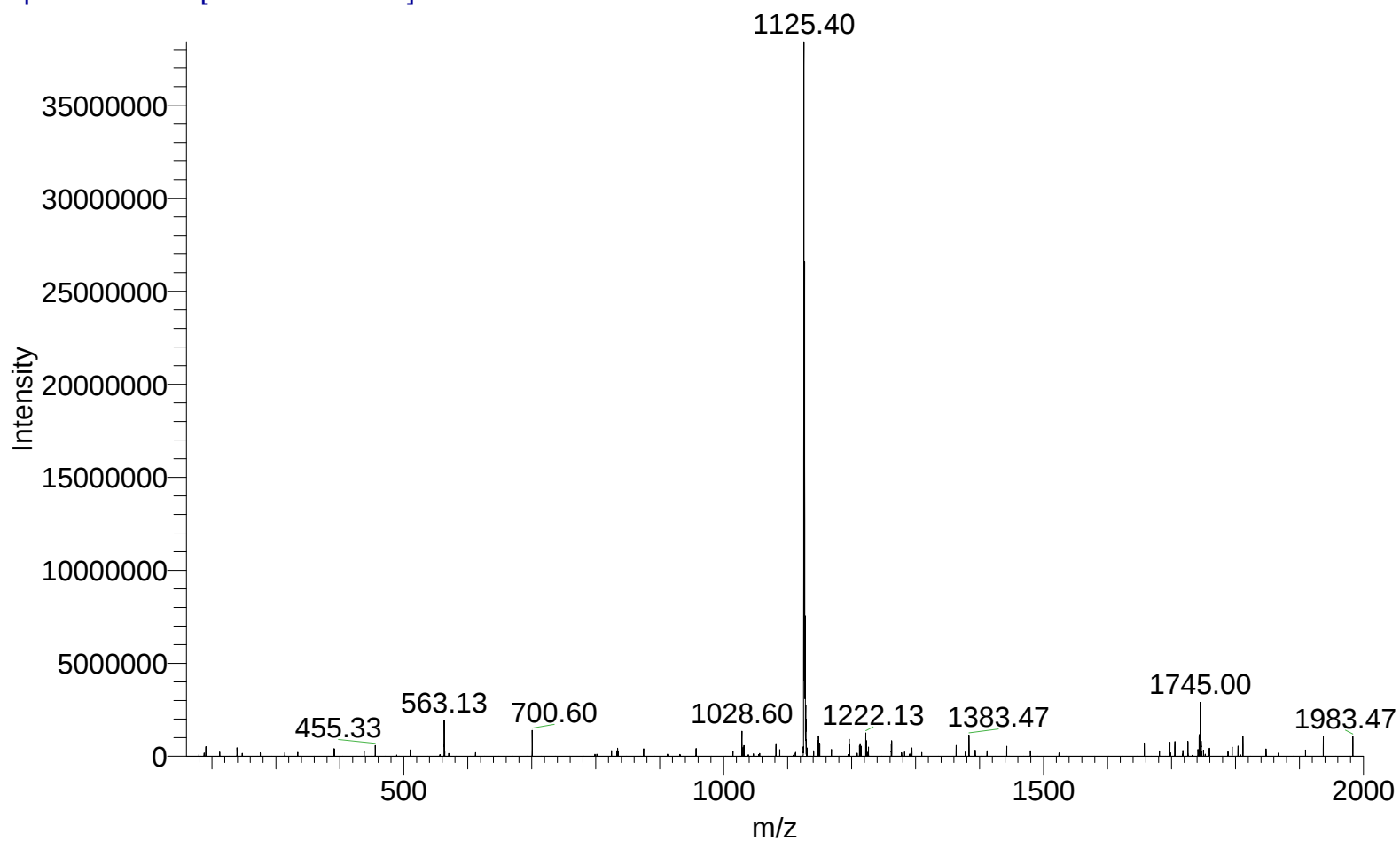


RT: 0.00 - 14.60

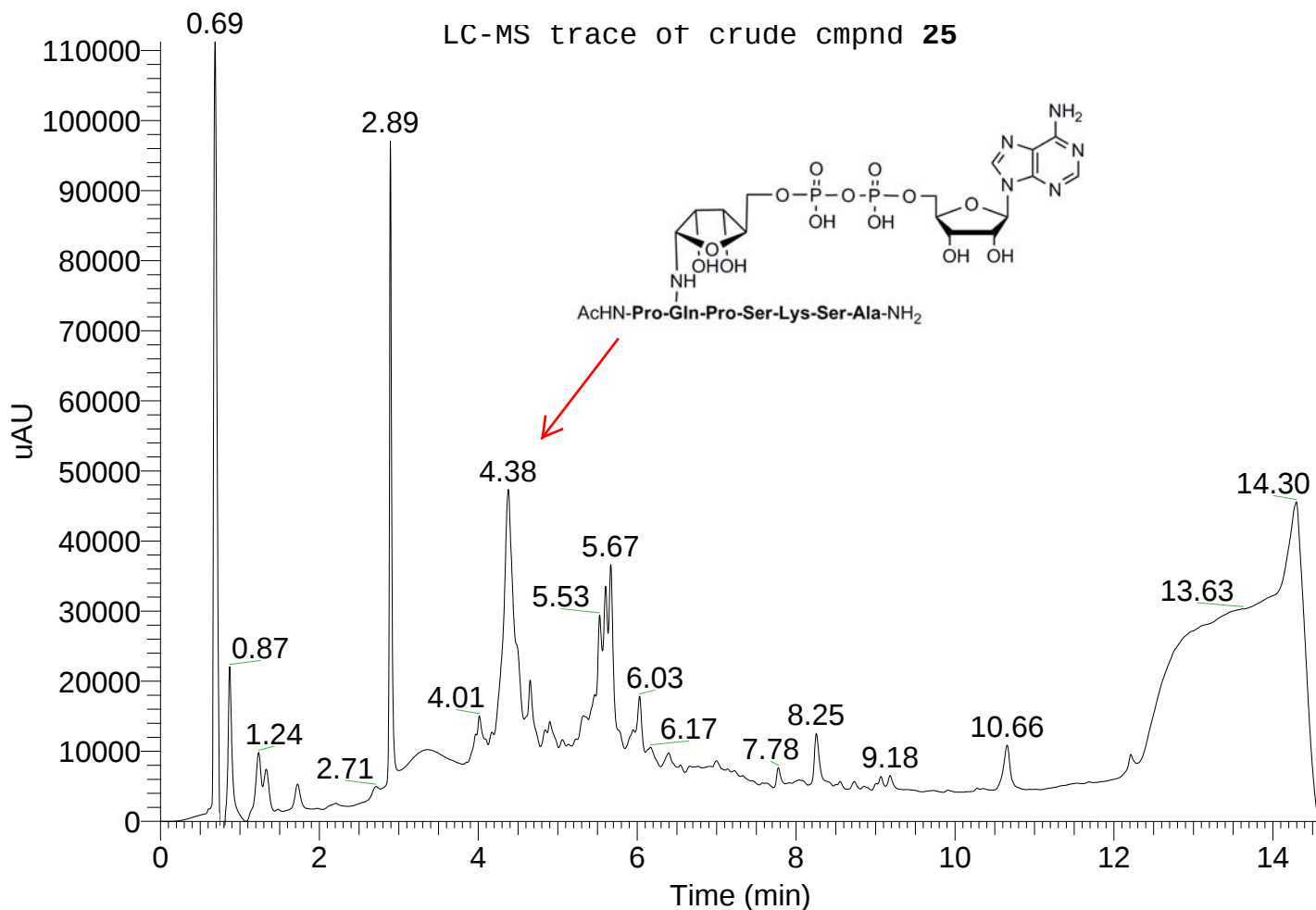
LC-MS trace of crude cmpnd 23



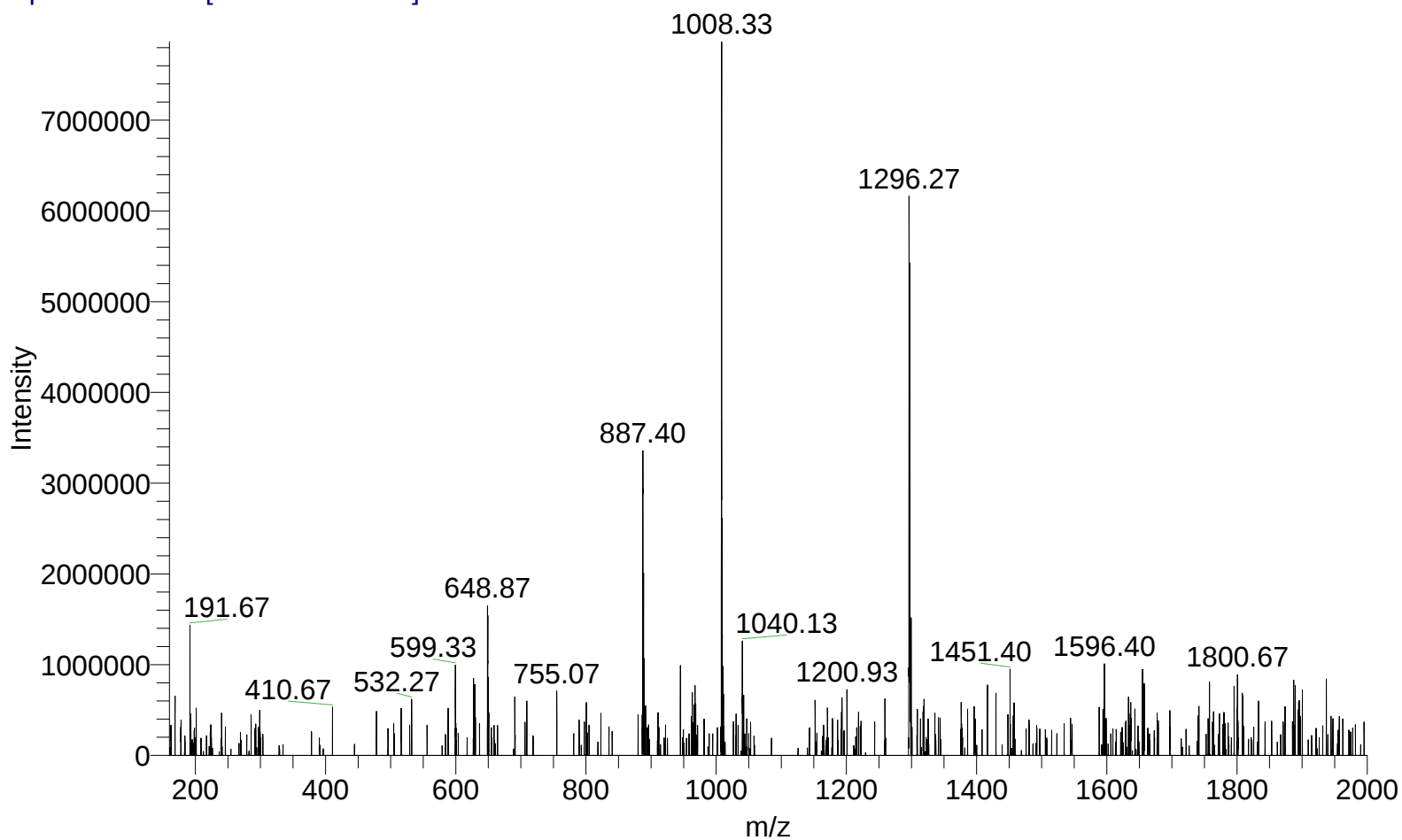
RT: 7.27 AV: 1 NL: 3.84E7
T: + p ESI Full ms [160.00-2000.00]



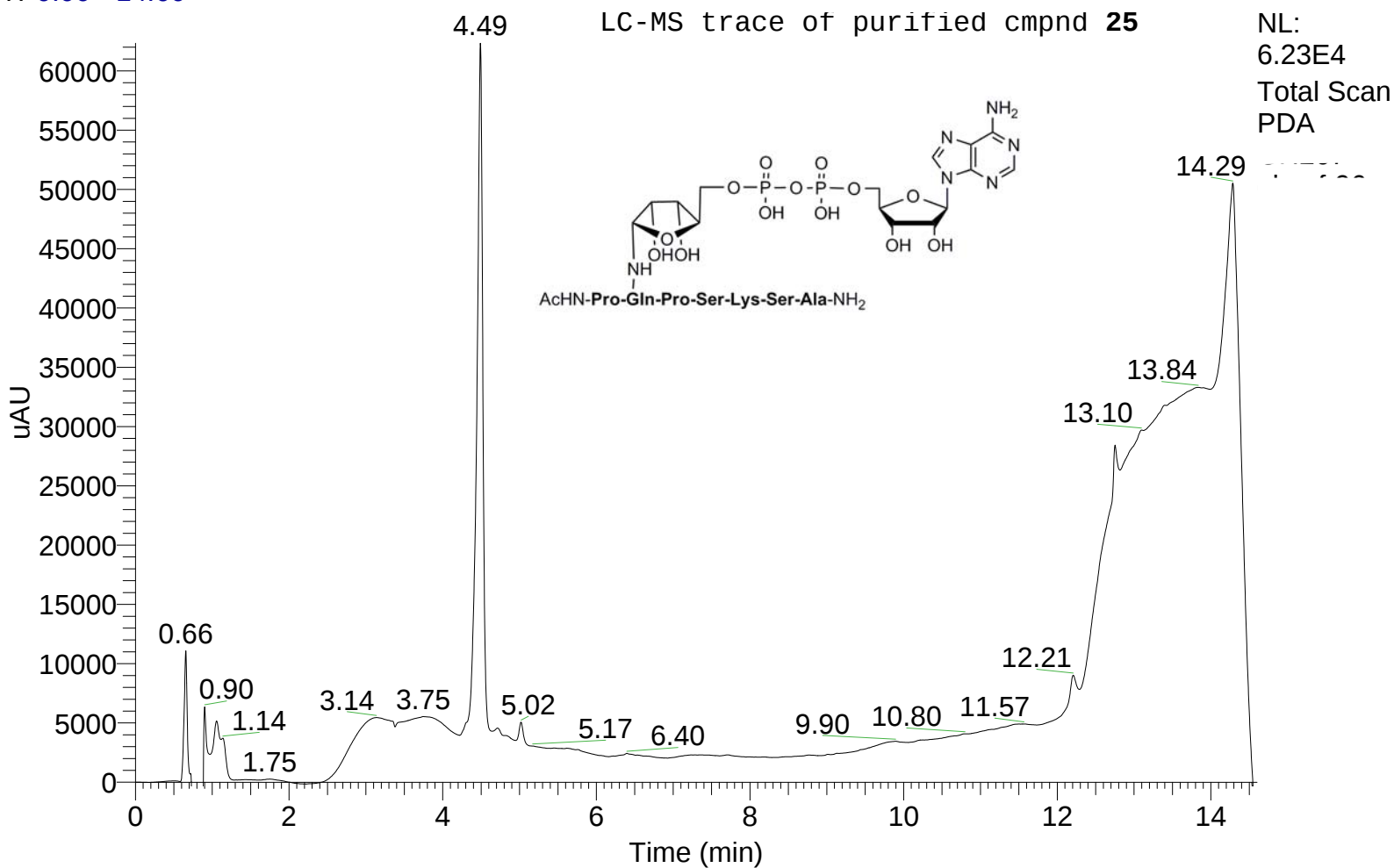
RT: 0.00 - 14.60



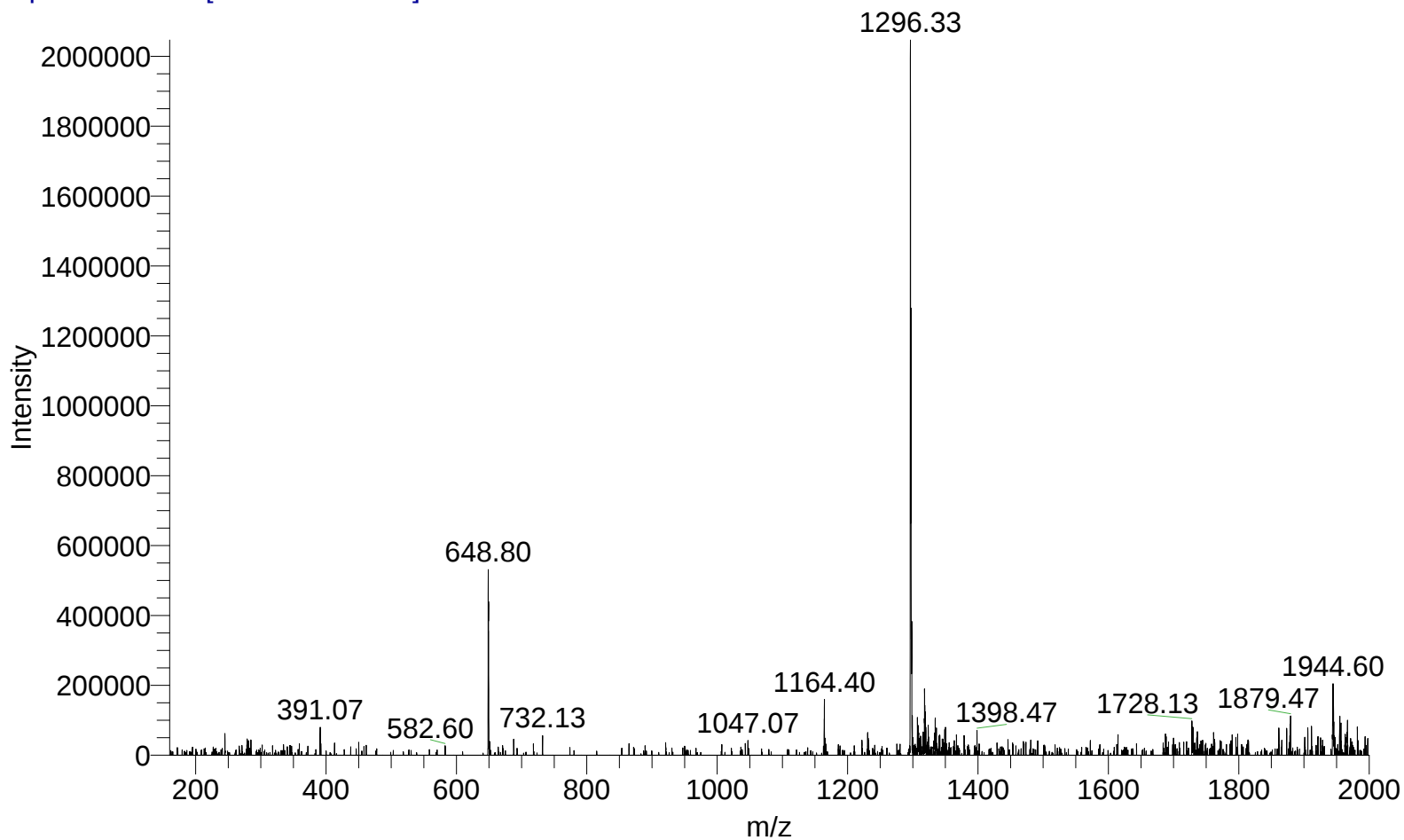
RT: 4.38 AV: 1 NL: 7.86E6
T: + p ESI Full ms [160.00-2000.00]



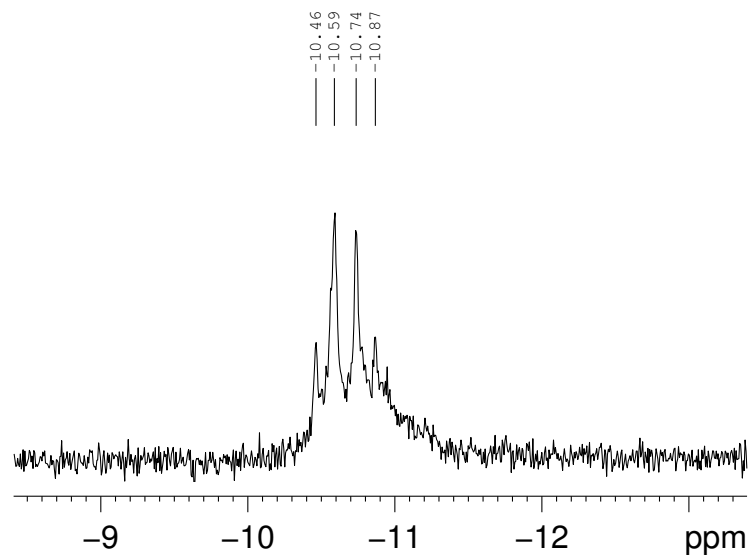
RT: 0.00 - 14.60



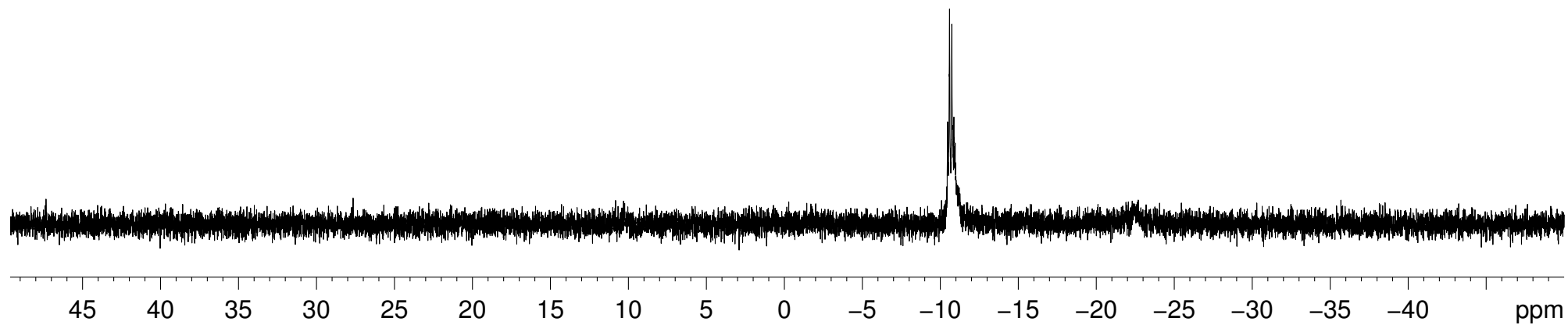
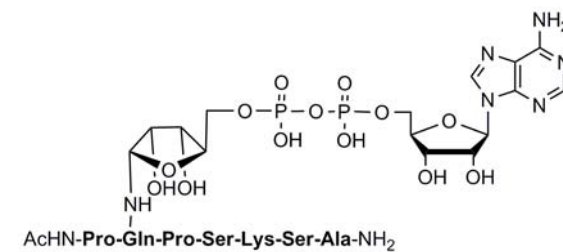
RT: 4.48 AV: 1 NL: 2.05E6
T: + p ESI Full ms [160.00-2000.00]



³¹P-NMR, 162 MHz, D₂O, compnd 25



-10.46
-10.59
-10.74
-10.74
-10.87



¹H-NMR, 400 MHz, D₂O, compnd 25

