

SUPPORTING INFORMATION

1. NMR analysis of the synthesized PLL₂₀-g_{3.5}-PEG_{2/3.4}-TG

The synthesized PLL₂₀-g_{3.5}-PEG_{2/3.4}-TG copolymer was analyzed by ¹H NMR (D₂O) and resulted in a copolymer with the following structure: PLL₂₀-g_{4.4}-PEG_{2/3.4}-TG_{25%}.

¹H NMR (D₂O) d:

- 0.83 (TG-peptide, -CH₃ valine side chains, d),
- 1.18-1.48 and 1.48-1.81 (PLL, -C_αHC_βH₂C_γH₂C_δH₂C_εH₂NH₂, m),
- 1.81-2.48 (TG-peptide, side chains),
- 2.90 (PLL, -C_αHC_βH₂C_γH₂C_δH₂C_εH₂NH₂, t),
- 3.05-3.14 (PLL and TG-peptide, -C_αHC_βH₂C_γH₂C_δH₂C_εH₂NHCO and side chains, m),
- 3.28 (mPEG, -OCH₃, s),
- 3.60 (mPEG and PEG, -OCH₂CH₂-, m),
- 4.05 (TG-peptide, -NHC_αHCO-, d),
- 4.21 (PLL and TG-peptide, -NHC_αHCO-, m),
- 4.32 (TG-peptide, -NHC_αHCO-, t),
- 4.45 (TG-peptide, -NHC_αHCO-, t)

The grafting ratios were calculated from the NMR spectrum. The ratio between mPEG₂₀₀₀ and PEG₃₄₀₀, to which the TG-peptide has been coupled, was determined from the integral value of the methoxy group of mPEG₂₀₀₀ peak at 3.28 ppm and from the integral value of the ethyleneoxide group of both mPEG₂₀₀₀ and PEG₃₄₀₀ peaks between 3.42-3.73 ppm. Based on the signal from the methoxy group of mPEG₂₀₀₀, the value for one proton of mPEG₂₀₀₀ is 0.36. This results in a total integral value for mPEG₂₀₀₀ with 45 ethylene oxide units of 64.80 and leaves a signal of 36.03 for the 77 ethylene oxide unit of PEG₃₄₀₀. This represents a value for one proton of PEG₃₄₀₀ of 0.12. Therefore the proportion of mPEG₂₀₀₀ to PEG₃₄₀₀ is 0.36 to 0.12 which is a ratio of 3 to 1.

Based on the signal at 0.83 ppm from the four methyl groups of the two valine residues in the TG-peptide and on the signals at 4.05, 4.32 and 4.45 ppm representing six protons from the C_αH groups in the TG-peptide, one proton from the TG-peptide has an integral value of 0.12. This value is the same as that of the PEG₃₄₀₀. Therefore, each PEG₃₄₀₀ is functionalized with one TG-peptide.

To calculate the ratio between the PEGs and PLL, the integral values from the C_βH₂, C_γH₂ and C_δH₂ groups of PLL representing 6 protons are used. One proton from PLL is represented by a value of 2.13 and one proton of the PEGs is represented by a value of 0.36 and 0.12. This results in a PLL-PEG grafting ratio of 4.4 to 1.

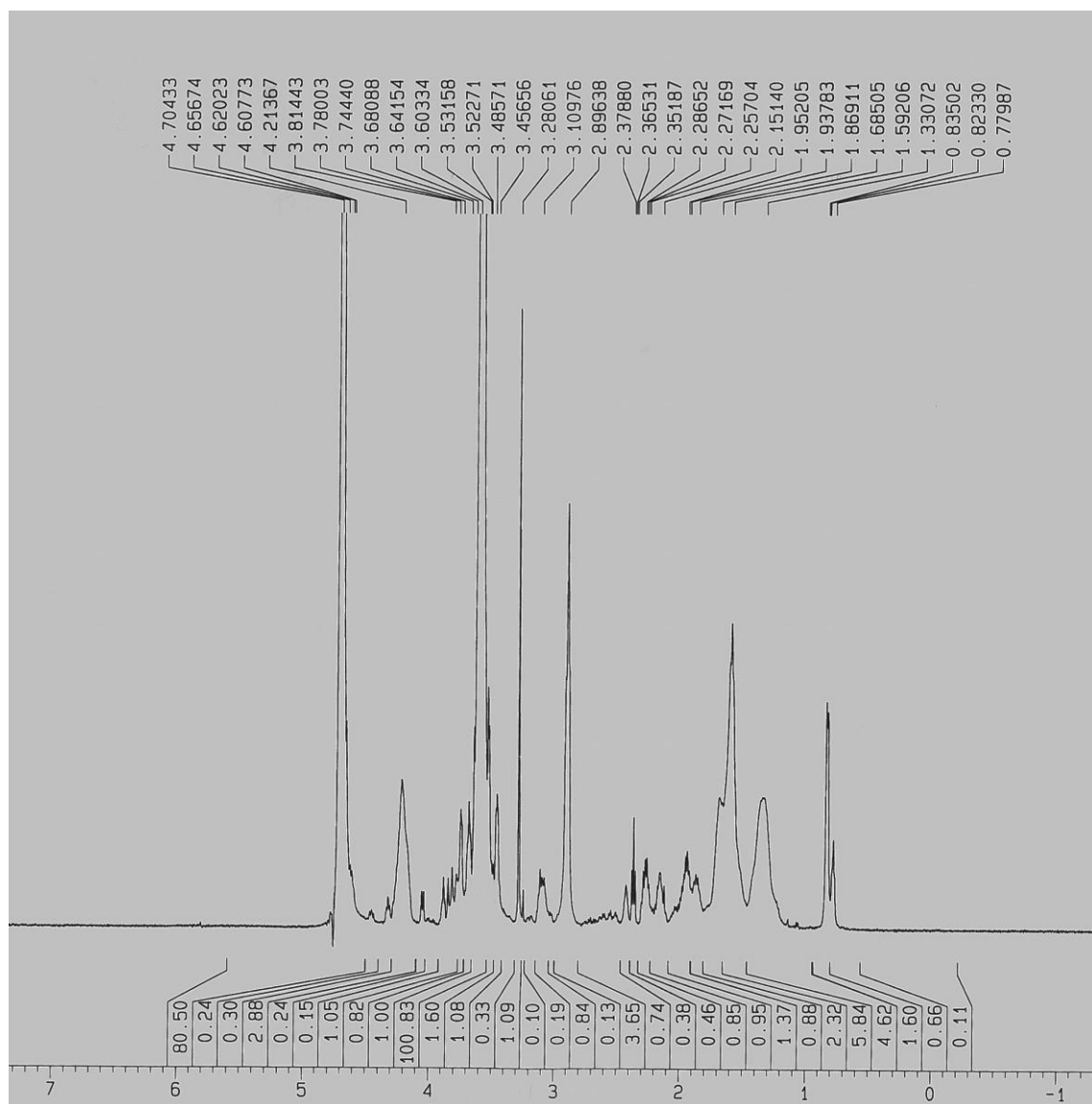


Figure S1. NMR of PLL-g-PEG-TG.

2. **In situ OWLS monitoring and quantification of PLL-g-PEG-TG adsorption on a titanium-coated waveguide and degree of resistance of the polymeric adlayer to serum adsorption.**

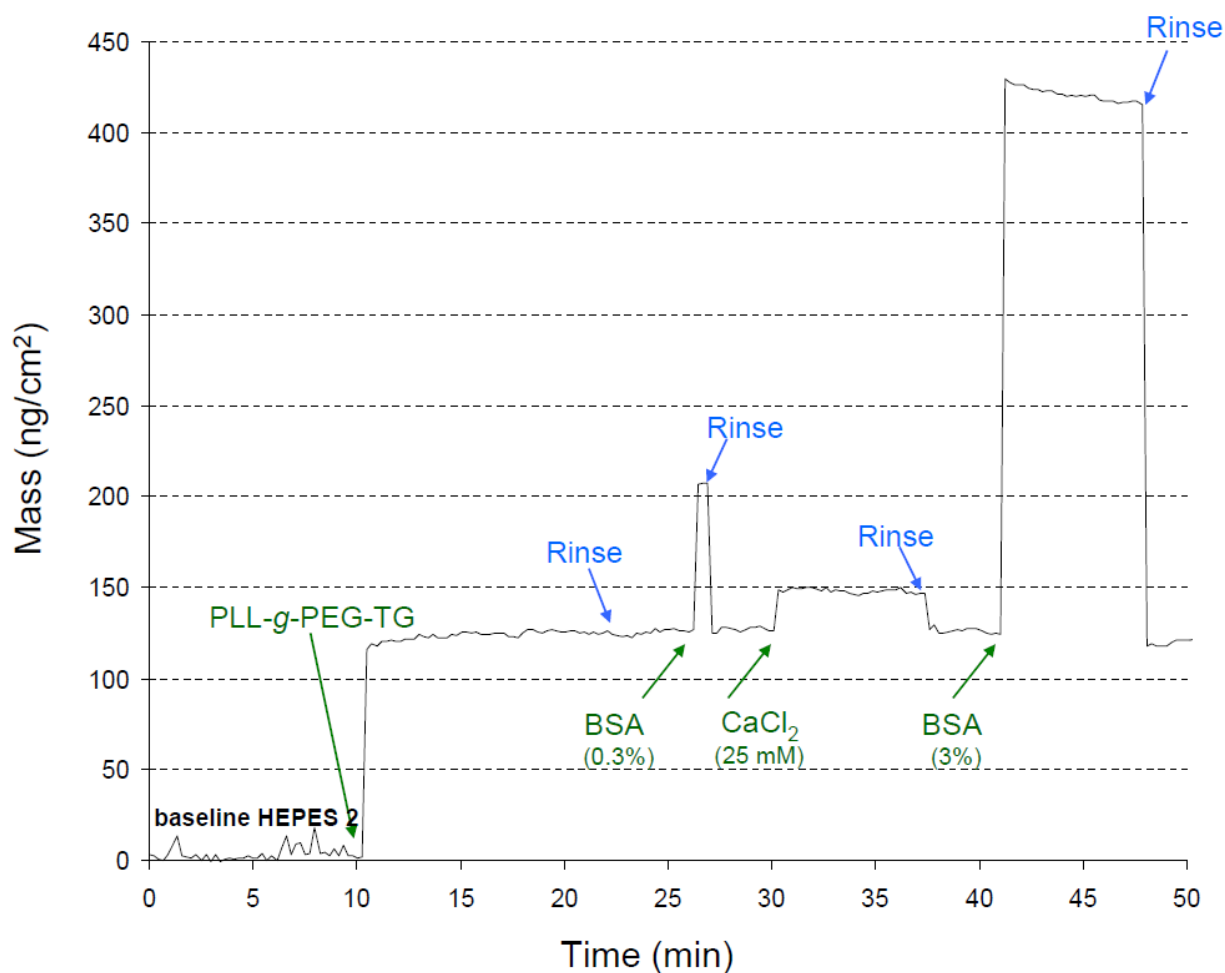


Figure S2. In situ OWLS PLL-g-PEG-TG adsorption and polymeric adlayer resistance to serum proteins ($dn/dc = 0.139$).

3. Summary of PLL-g-PEG-TG characterization by ¹H-NMR and OWLS and theoretical peptide densities distance between them on the modified surfaces. Immobilization calculations on the surface.

PLL-g-PEG-TG characterization by H-NMR and OWLS. Theoretical peptide densities based on these experimental values

PLL-g-PEG-TG characterization

Concept	Remarks	Units	Values
Molecular weight PLL	(mol wt PLL)	g/mol	20,000
N° Lys units			96
Molecular weight Lys UNIT	(mol wt Lys)	g/mol	208.3
Molecular weight PLL-g-PEG-TG	calculated (0)	g/mol	78,513.7
Molecular weight non-functionalized PEG side chains		g/mol	2,000
Molecular weight functionalized PEG side chains		g/mol	3,400.0
Molecular weight TG		g/mol	1,358.5
Grafting ration	(g)		4.4
Total number of PEG side chains			22
Fraction of functionalized PEG	(P _{PEG-TG})		25%
Fraction of non-functionalized PEG	(P _{PEG})		75%

In situ OWLS measurements

Concept	Remarks	Units	Values
Mass of PLL-g-PEG-TG adsorbed on a TiO ₂ waveguide	(m _a)	ng/cm ²	132 ± 5
BSA adsorption (non-fouling properties of the PLL-g-PEG-TG adlayer)		ng/cm ²	1.8 ± 1.2
Mass of PLL-g-PEG adsorbed on a Nb:O _x waveguide	(m _a)	ng/cm ²	170

Theoretical TG peptide density and distance between the moieties on a surface

Concept	Remarks	Units	Values
TG peptide density on a TiO ₂ surface	(p)	pmol/cm ²	9.1
Distance between the TG moieties on a TiO ₂ surface	calculated (2)	nm	4.6
TG peptide density on a Nb:O _x surface	(p)	pmol/cm ²	11.7
Distance between the TG moieties on a Nb:O _x surface	calculated (2)	nm	4.0

Maximum theoretical peptide densities on a surface and Immobilization amounts.

Concept	Remarks	Units	Values
Molecular weight 8-arm Lys-PEG		g/mol	53,744.0
Molecular weight Lys-FITC		g/mol	966.2
Molecular weight Lys-RGD		g/mol	1,018.3
Mass of 8-arm Lys-PEG immobilized on a TiO ₂ surface (one-step).	calculated (3)	ng/cm ²	487.3
Mass of Lys-FITC immobilized on a TiO ₂ surface (one-step).	calculated (3)	ng/cm ²	8.8
Mass of Lys-RGD immobilized on a TiO ₂ surface	calculated (3)	ng/cm ²	9.2
Mass of Lys-RGD immobilized on a Nb:O _x surface	calculated (3)	ng/cm ²	11.9
Mass of 8-arm Lys-PEG immobilized on a TiO ₂ surface (one-step).	calculated (3')	ng/cm ²	39.0
Molecular weight TG-VEGF			32,000.0
Molecular weight 8-arm TG-PEG			50,868.0
Mass of TG-VEGF immobilized on a TiO ₂ surface (two-steps).	calculated (4); 1mol Lys-PEG to 1mol TG-VEGF	ng/cm ²	15.7 ± 2
Mass of TG-VEGF immobilized on a TiO ₂ surface (two-steps).	calculated (4'); 1mol Lys-PEG to 4mol TG-VEGF	ng/cm ²	62.9 ± 7.9
Mass of TG-PEG immobilized on a TiO ₂ surface (two-steps).	calculated (5); 1mol Lys-PEG to 1mol TG-PEG	ng/cm ²	25 ± 3.1
Mass of TG-PEG immobilized on a TiO ₂ surface (two-steps).	calculated (5'); 1mol Lys-PEG to 4mol TG-PEG	ng/cm ²	100 ± 12.5
Mass of LYS-PEG immobilized on a TiO ₂ surface (three-steps).	calculated (6); 1mol TG-PEG to 1mol Lys-PEG	ng/cm ²	25.6 ± 3.7
Mass of LYS-PEG immobilized on a TiO ₂ surface (three-steps).	calculated (6'); 1mol TG-PEG to 4mol Lys-PEG	ng/cm ²	102.3 ± 14.8

- (0) [mol wt (PLL) + [mol wt (PLL) / mol wt (Lys) * (1 / g)] * [(P_{PEG} / 100) * mol wt (PEG)] + [(P_{PEG-TG} / 100) * mol wt (PEG-TG)]]
- (1) [[mol wt (PLL) / (mol wt (Lys) * g)] * [(m_a / mol wt (Pol)) * P_{PEG-TG}]]
- (2) [[(2 / 3) * (1 / (p * Avogadro's number))]]¹⁰
- (3) [p / mol wt (immobilized peptide via FXIIla)]; 100 % efficiency of the enzyme, all TG domains on the surface react with a Lys substrate
- (3') [p / mol wt (immobilized peptide via FXIIla) * 0.8]; 80 % efficiency of the enzyme, 1/10 TG domains on the surface react with a Lys substrate
- (4) [experimental 8-arm Lys-PEG immobilized (26.4 ± 3.3) / mol wt (8-arm Lys-PEG) * mol wt (TG-VEGF)]
- (4') [experimental 8-arm Lys-PEG immobilized (26.4 ± 3.3) / mol wt (8-arm Lys-PEG) * mol wt (TG-VEGF)]
- (5) [experimental 8-arm Lys-PEG immobilized (26.4 ± 3.3) / mol wt (8-arm Lys-PEG) * mol wt (8-arm TG-PEG)]
- (5') [experimental 8-arm Lys-PEG immobilized (26.4 ± 3.3) / mol wt (8-arm Lys-PEG) * mol wt (8-arm TG-PEG)]
- (6) [experimental 8-arm TG-PEG immobilized (24.2 ± 3.5) / mol wt (8-arm TG-PEG) * mol wt (8-arm Lys-PEG)]
- (6') [experimental 8-arm TG-PEG immobilized (24.2 ± 3.5) / mol wt (8-arm TG-PEG) * mol wt (8-arm Lys-PEG)]

Figure S3. Summary of PLL-g-PEG-TG characterization by ¹H-NMR and OWLS. Maximum theoretical peptide densities and distance between the TG moieties on the modified surfaces. Immobilization amounts (theoretical based on experimental values).

4. Representative OWLS experiment of enzymatic immobilization.

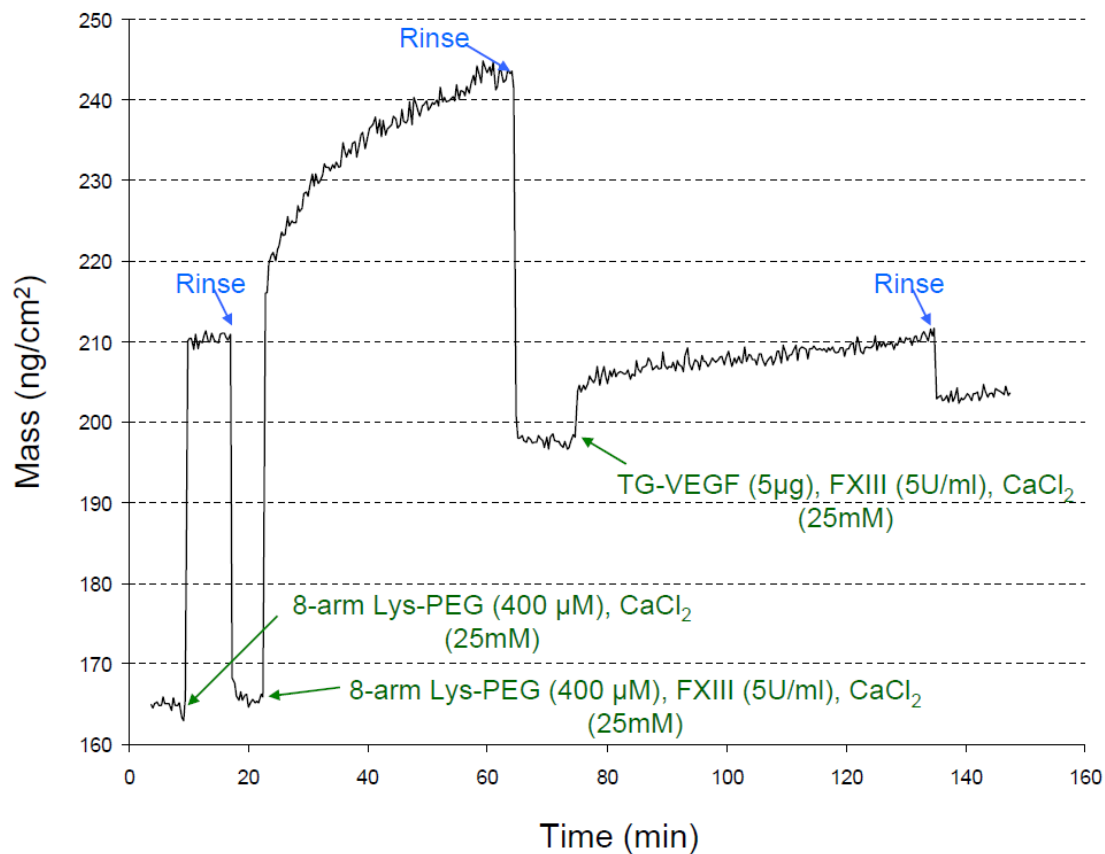


Figure S4. *In situ* OWLS immobilization of 8-arm Lys-PEG followed by TG-VEGF₁₂₁. First a negative control is shown (8-arm Lys-PEG in absence of FXIII). No absorption is observed. Next, immobilization of 8-arm Lys-PEG is monitored and followed by immobilization of TG-VEGF₁₂₁. The dn/dc for this graph is 0.139 (for PEG-based molecules. A dn/dc = 0.182 was considered for calculations involving TG-VEGF as mention in the Materials and Methods section). (Not real starting time).