

Supporting Information

Shared biosynthesis of the saliniketals and rifamycins in *Salinispora arenicola* is controlled by the *sare1259*-encoded cytochrome P450

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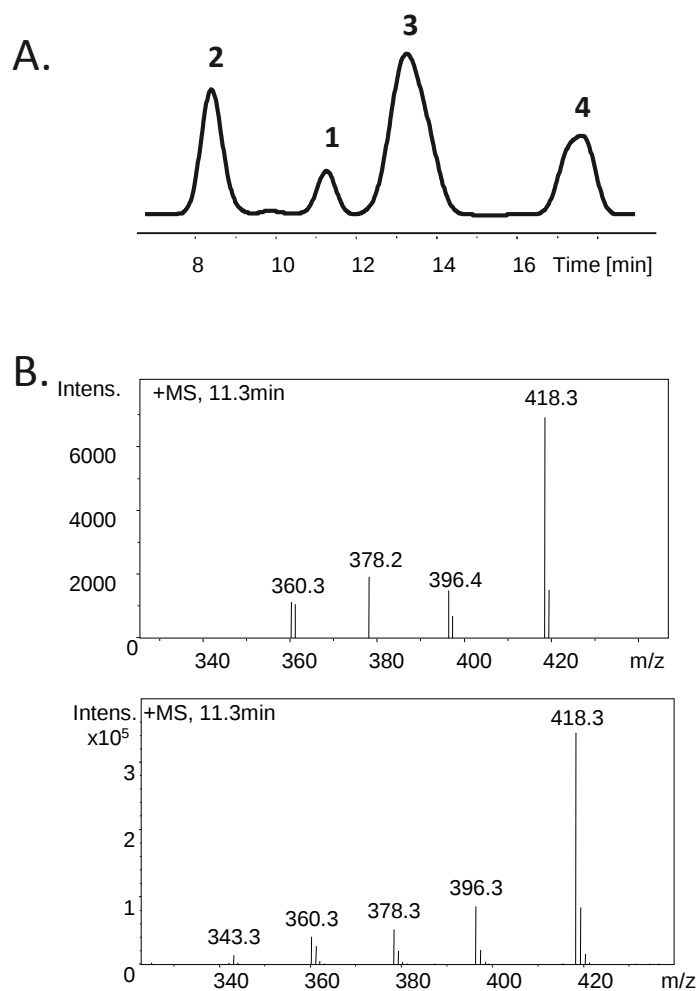


Figure S1. HPLC-MS analysis of *A. mediterranei* S699. (A) Extracted ion chromatogram for all masses corresponding to the m/z of the sodium adducts of the saliniketals (**1–2**) and rifamycins (**3–8**) produced by *A. mediterranei* S699. (B) MS fragmentation pattern for **1** from *A. mediterranei* S699 (top panel) and authentic standard (bottom panel).

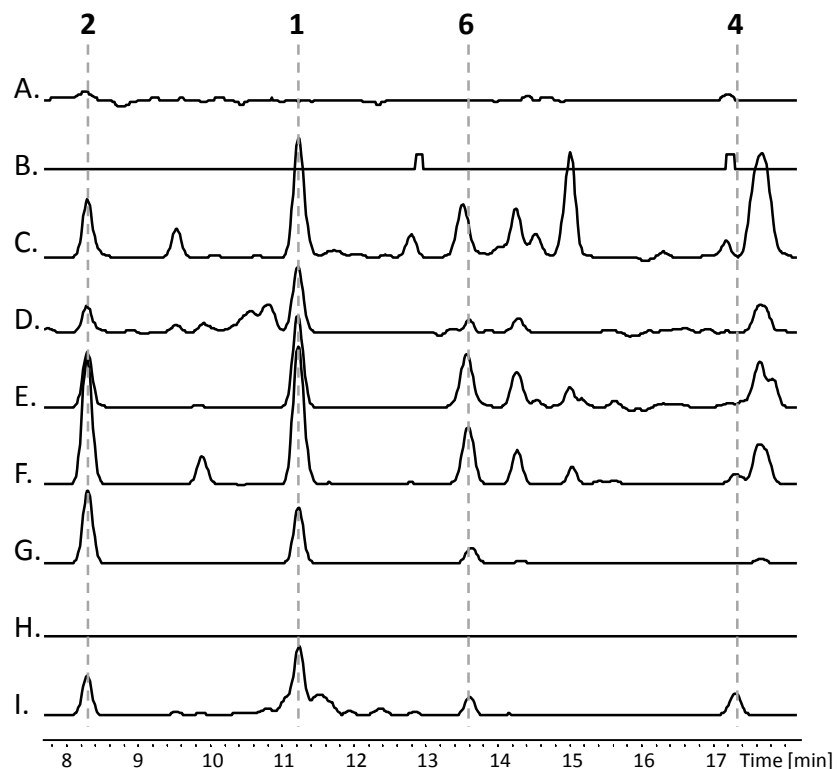


Figure S2. HPLC-MS analysis of SA-*rif* mutants generated in this study. Extracted ion chromatogram for all masses corresponding to the m/z of the sodium adducts of the saliniketals (1–2) and rifamycins (3–8) produced by (A) *sare1242::Apr^R*, (B) *sare1243::Apr^R*, (C) *sare1263::Apr^R*, (D) *sare1264::Apr^R*, (E) *sare1265::Apr^R*, (F) *sare1266::Apr^R*, (G) *sare1267::Apr^R*, (H) *sare1272/1273::Apr^R*, and (I) *sare1274::Apr^R*. Unlabeled peaks correspond to non-*rif* related compounds and/or unconfirmed *rif* shunt products.

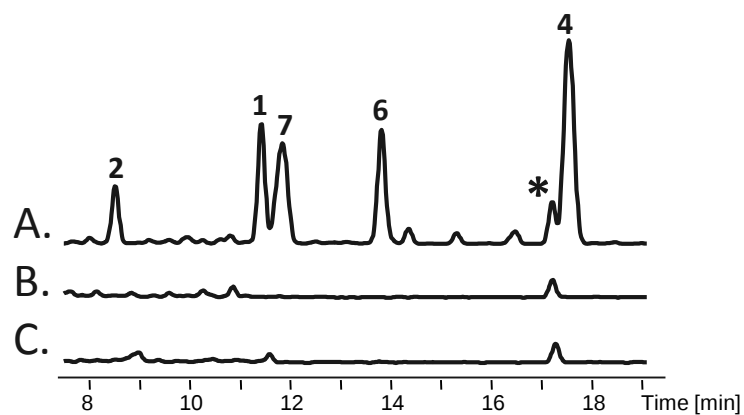


Figure S3. HPLC-MS analysis of the chemical complementation of the *S. arenicola rif-orf0* mutant strain *sare1245::apr^R*. EIC for all masses corresponding to the *m/z* of the sodium adducts of the saliniketals (**1–2**) and rifamycins (**3–8**) for (A) *sare1245::apr^R* chemically complemented with **7**, (B) *sare1245::apr^R* chemically complemented with AHBA, and (C) *sare1245::apr^R* negative control. (* indicates unrelated compounds that share an *m/z* ratio with one of the compounds **1–8**)

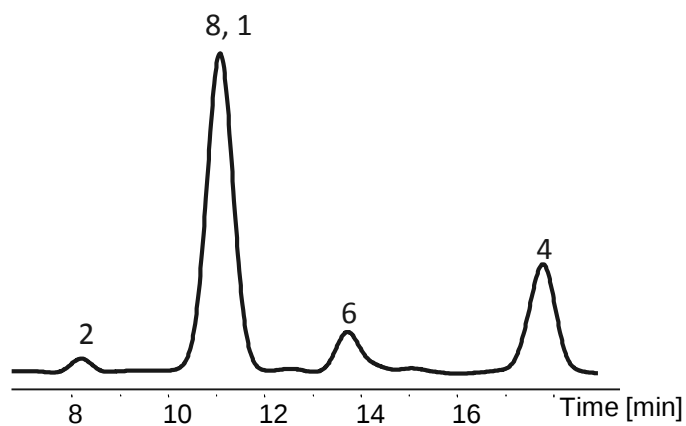


Figure S4. HPLC-MS analysis of the *A. mediterranei rif-orf5* mutant strain MT45025H. EIC for all masses corresponding to the m/z of the sodium adducts of the saliniketals (**1–2**) and rifamycins (**3–8**) produced by MT45025H cultured for 10 days in A1 media at 28°C and 225 rpm.

Figure S5. ^{13}C NMR spectrum (126 MHz, $\text{MeOH-}d_4$) of saliniketal A (**1**) from $[\text{U-}^{13}\text{C}]$ propionate feeding study.

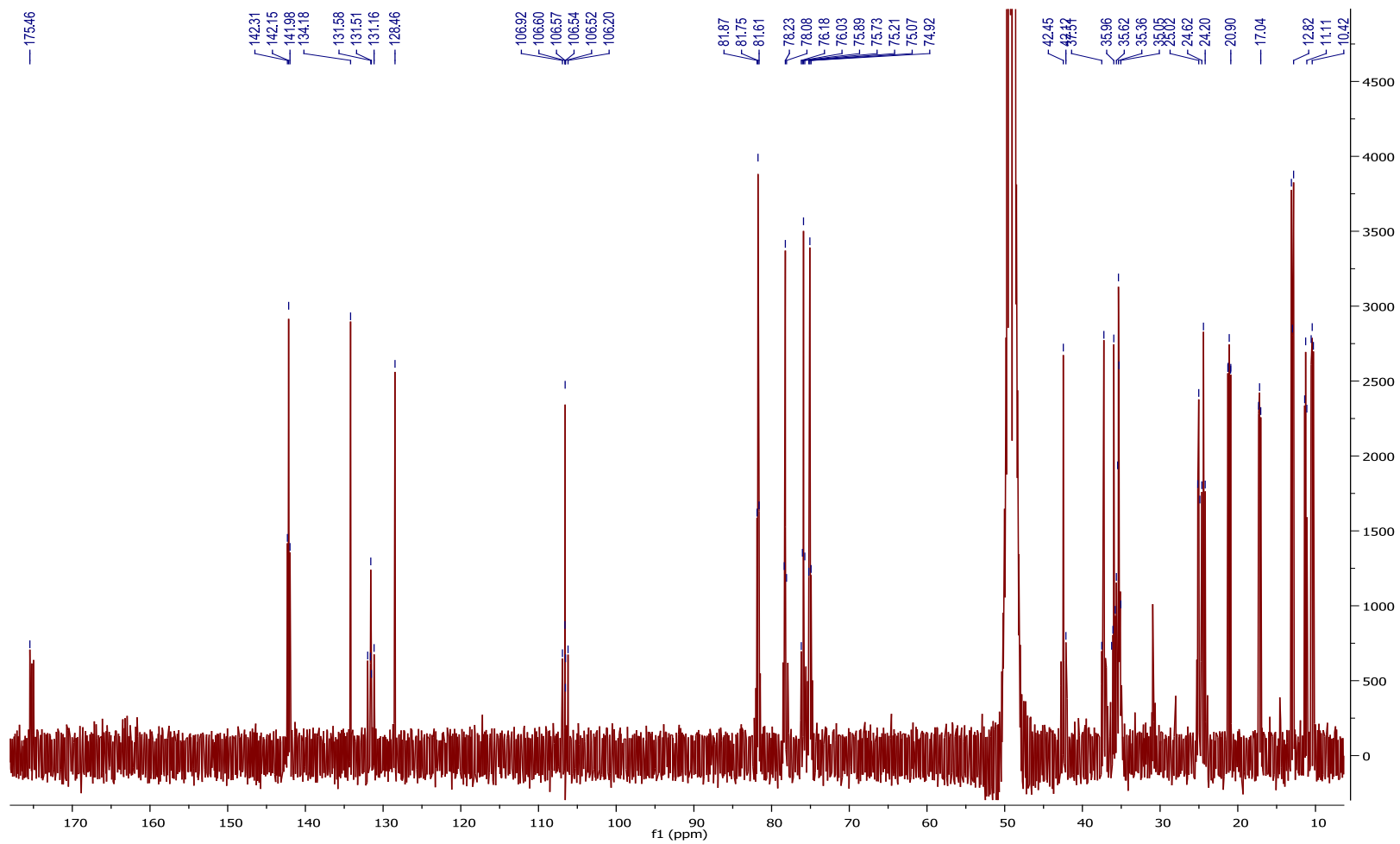


Figure S6. Partial ^{13}C NMR spectrum (126 MHz, $\text{MeOH-}d_4$) of saliniketal A (**1**) from $[\text{U-}^{13}\text{C}]$ propionate feeding study (127-177 ppm)

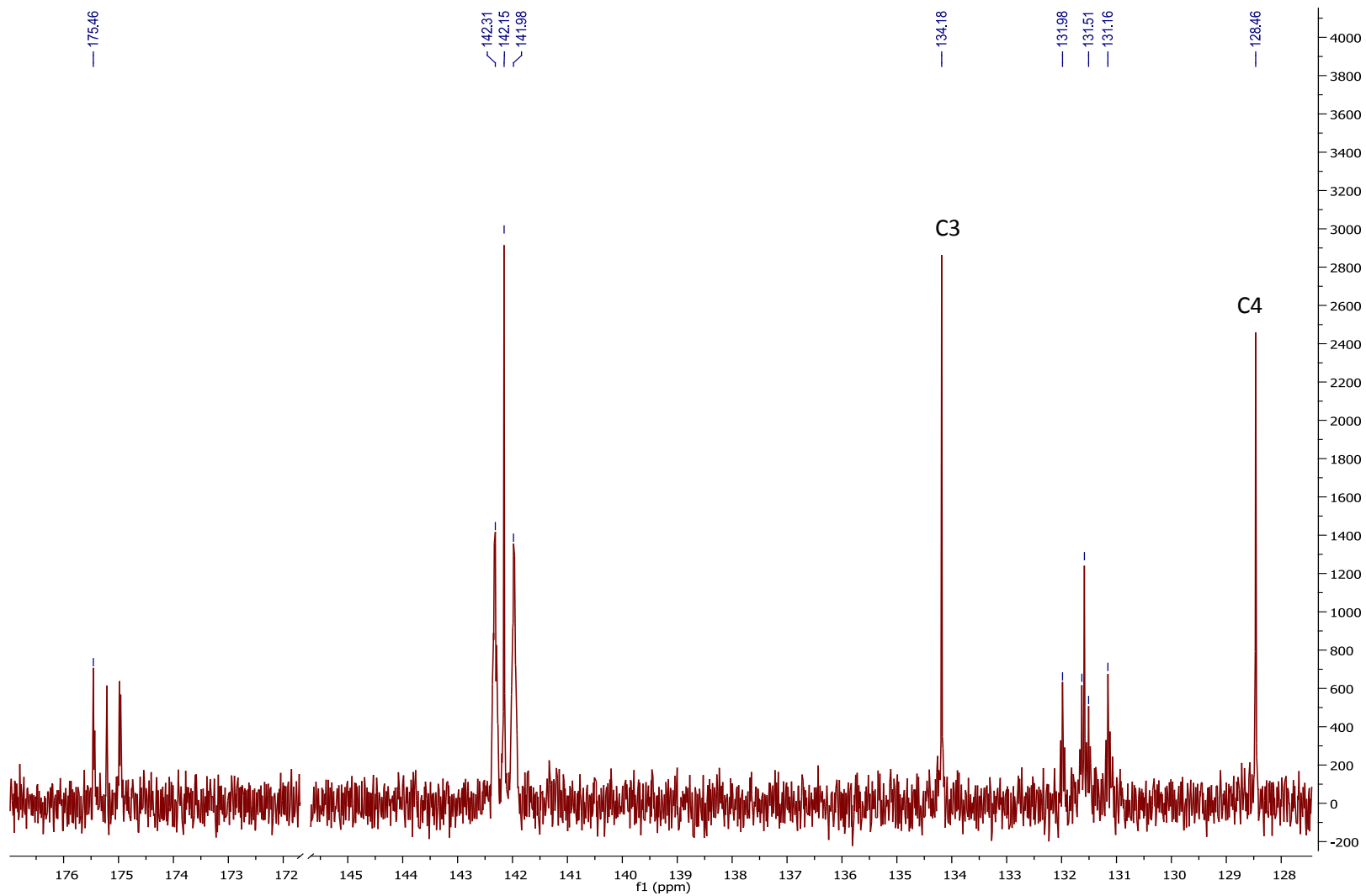


Figure S7. ^1H NMR spectrum (600 MHz, $\text{DMSO-}d_6$) of 34a-deoxy-rifamycin W (**7**) from *sare1259::apr^R* mutant

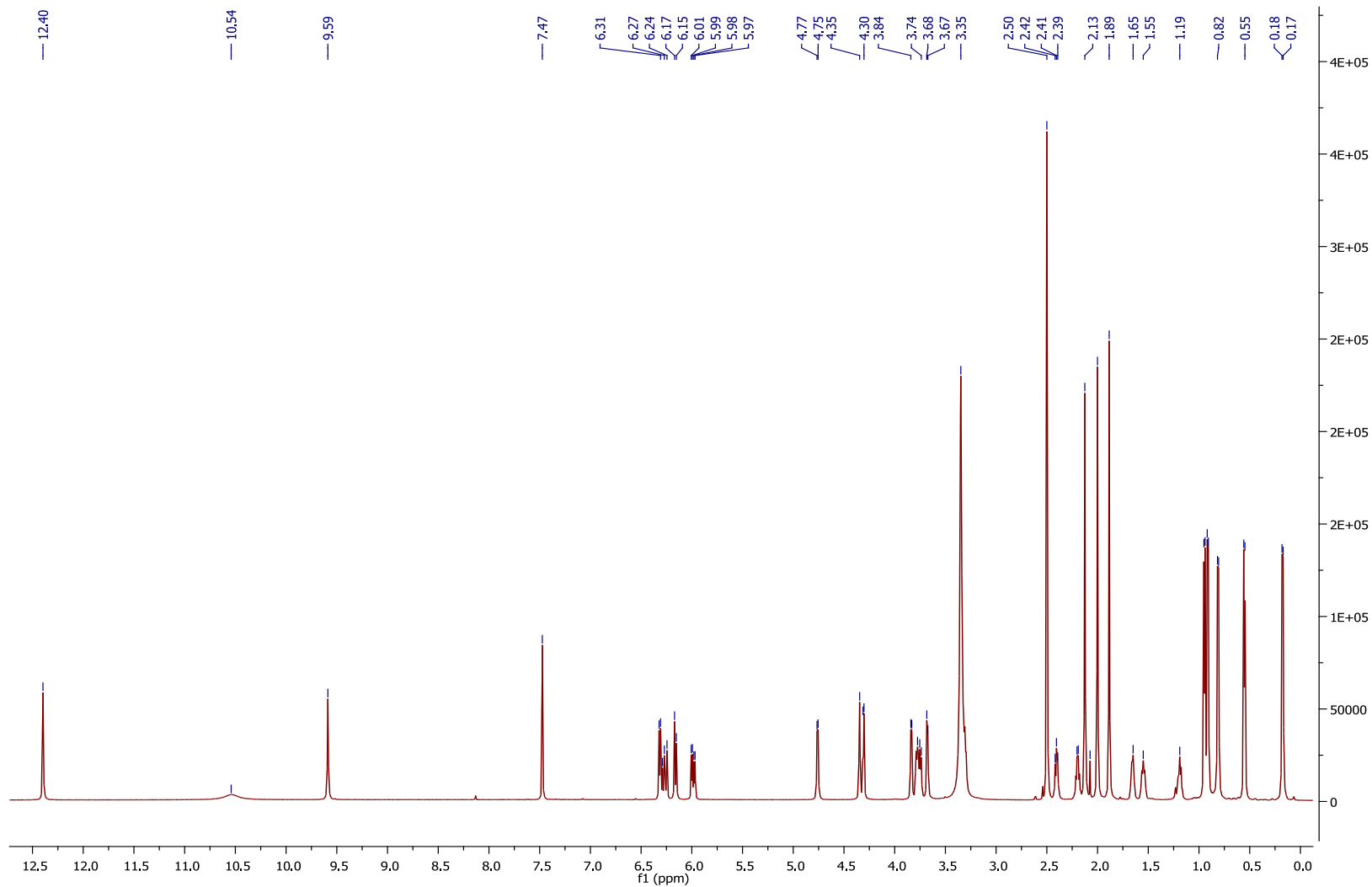


Figure S8. gHSQC spectrum (600 MHz, DMSO- d_6) of 34a-deoxy-rifamycin W (7) from *sare1259::apr^R* mutant

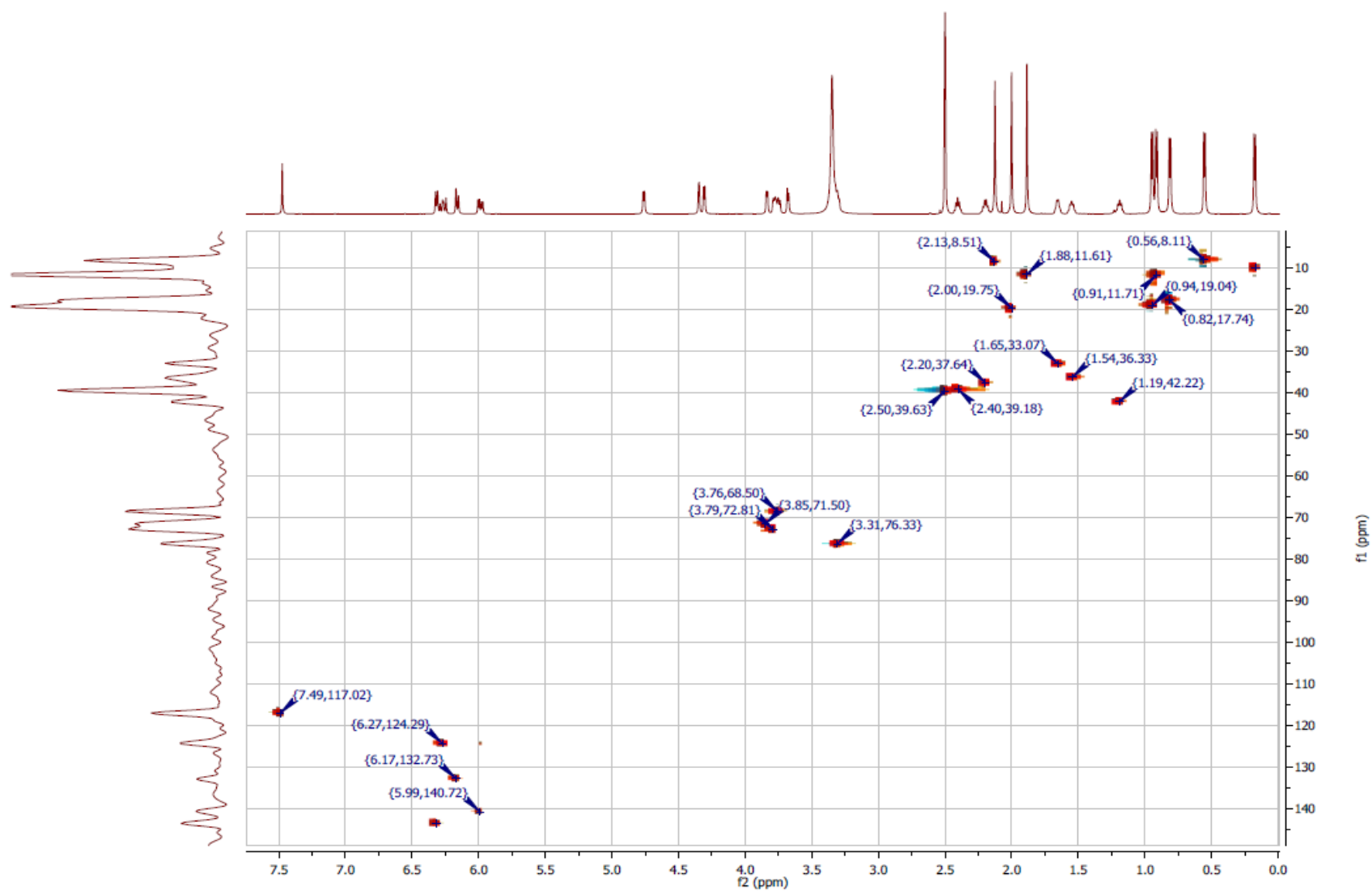


Figure S9. gHMBC spectrum (600 MHz, DMSO-*d*₆) of 34a-deoxy-rifamycin W (**7**) from *sare1259::apr^R* mutant

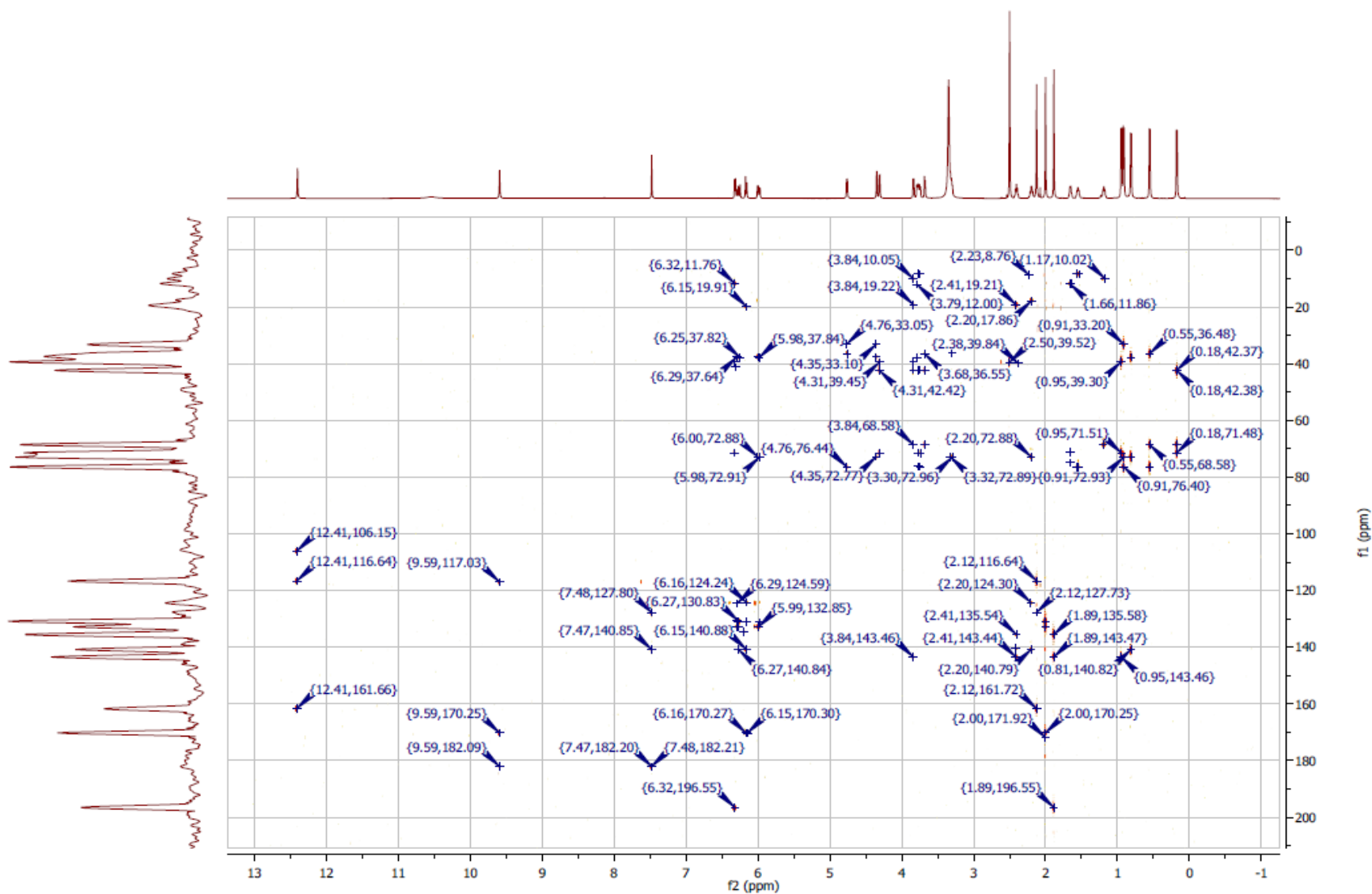


Table S1. Primers synthesized for gene inactivation experiments and verification of double crossover mutants in the SA-*rif* gene cluster.

Primers	Oligonucleotides
sare1242KOF	5'-cgtgggccccgcaggtggaggaagggctggaaagcaatgattccggggatccgtcgacc-3'
sare1242KOR	5'-tcatgcgggcgactcctttcacgggcacttccacggcacgtgtaggctggagctgcttc-3'
sare1243KOF	5'-atgagagttgccgtcctgtccccggcgccacgaacgcgccattccggggatccgtcgacc-3'
sare1243KOR	5'-ccgttcgttcctcgcgccggccgggggcccggcgccgtcatgtaggctggagctgcttc-3'
sare1245KOF	5'-ctcatccgcacaccatcgccgagtgagaaggatgccatgattccggggatccgtcgacc-3'
sare1245KOR	5'-gcactcaggaagaacgctcaccagcagacgggcagggtttgtaggctggagctgcttc-3'
sare1255KOF	5'-cgttcacgcatttctcaccggagaattcggagagacatgattccggggatccgtcgacc-3'
sare1255KOR	5'-gccccgacgaccgccaccggacgctcatgcgctggccactgtaggctggagctgcttc-3'
sare1258KOF	5'-cacgggcggtatcgaccgatccgtgctggcaggcggtcatgattccggggatccgtcgacc-3'
sare1258KOR	5'-gcgggccagtgcgaccggggatcagtgcgacaggggatcatgtaggctggagctgcttc-3'
sare1259KOF	5'-ggatcgccggcttttcgacacgagagggaggggctcatgattccggggatccgtcgacc-3'
sare1259KOR	5'-aggcacctcgggtcagggccagccgcccggcggtgtgctcatgtaggctggagctgcttc-3'
sare1260KOF	5'-gtgcatattgccgcacgtagtagagggaggcctcgcgcttgattccggggatccgtcgacc-3'
sare1260KOR	5'-cgatgcacacttcccggttcgcgcgccacctcacggcggttgtaggctggagctgcttc-3'
sare1262KOF	5'-tccaccggacttcgcacgctgattggagaccgattcgtgattccggggatccgtcgacc-3'
sare1262KOR	5'-gcgcacgcccgtgacgcccggcggtacgccgacggtcatgtaggctggagctgcttc-3'
sare1263KOF	5'-gcctcgagagaacctcacgaccgctgggagacgacttgattccggggatccgtcgacc-3'
sare1263KOR	5'-acccgcctcgccgcccggggcccggtgggacggacgcctcatgtaggctggagctgcttc-3'
sare1264KOF	5'-cagcgacagcccacactcgaccgtaggggatacgacatgattccggggatccgtcgacc-3'
sare1264KOR	5'-ctggtcacgatgcccgagtggaaccgaacccacccttcatgtaggctggagctgcttc-3'
sare1265KOF	5'-cggcgacgtgggtgaggtgttcgggggatcccggccgtgattccggggatccgtcgacc-3'
sare1265KOR	5'-gccggcccactctggcgggcgccctccggcgattgggtcatgtaggctggagctgcttc-3'
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sare1267KOF	5'-accggtacgcccaggtacgacgtcaggggggctcgtgtgattccggggatccgtcgacc-3'
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orf15KOF	5'-acaccaggcgaacaccgcactgtccaggcagtgcaatgattccggggatccgtcgacc-3'
orf15KOR	5'-tagccccgggtcgggtcgggagcacgggcttcgacgggtcgtgtaggctggagctgcttc-3'
sare1274KOF	5'-caccggggggcgaacgcgcgcggggtacggcacccgtgattccggggatccgtcgacc-3'
sare1274KOR	5'-cccgcgccggtcgacgctacggccgagcggcggtatgtcatgtaggctggagctgcttc-3'
sare1242CKF	5'-ccggcacccggtgaactacct-3'
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sare1259CKR	5'-cgttgaaccgccaggaccg-3'
sare1260CKF	5'-gagcgagggatgtgtggccg-3'
sare1260CKR	5'-ggagcaccacgggtcccgtcatc-3'
sare1262CKF	5'-acgtcccaccgaccgactgg-3'
sare1262CKR	5'-cagtgtctcgtgaccggggct-3'
sare1263CKF	5'-tgggtccggctgtttcgtccc-3'
sare1263CKR	5'-gctggtgacgtcggcaagct-3'
sare1264CKF	5'-agcgagttgcggtggctacg-3'
sare1264CKR	5'-gtgctgggcaggctcaggtgc-3'
sare1265CKF	5'-gcccagcacatcaccggagg-3'
sare1265CKR	5'-acagctcagcgggtgtgcggg-3'
sare1266CKF	5'-gtcctcgaagtggatcggcc-3'
sare1266CKR	5'-ggcatgtcggccctactctc-3'
sare1267CKF	5'-ggaacccggaacacgagacc-3'
sare1267CKR	5'-tacgagctccggcgccggtg-3'
orf15CKF	5'-tgatcaaccacgaccgctg-3'
orf15CKR	5'-ctcggccgtagcgtcgacc-3'
sare1274CKF	5'-agcaccggcttcgaccggtc-3'
sare1274CKR	5'-tcaggccccgaccagcgaac-3'

Complete reference 30.

Trefzer, A.; Jungmann, V.; Molnar, I.; Botejue, A.; Buckel, D.; Frey, G.; Hill, D. S.; Jorg, M.; Ligon, J. M.; Mason, D.; Moore, D.; Pachlatko, J. P.; Richardson, T. H.; Spangenberg, P.; Wall, M. A.; Zirkle, R.; Stege, J. T. *Appl. Environ. Microbiol.* **2007**, *73*, 4317.