Supplementary data

Lipid analysis

The presence of lipids in FHP was assessed by extracting all lipid content according to the method of Bligh and Dyer (Bligh, E. G., and Dyer, W. J. (1959) *Can J Biochem Physiol* **37**, 911-917). Lipid extraction was performed using 0.2 ml and 0.3 ml of pure FHP. The chloroform phase containing the lipid extract of the protein was analyzed then by one-dimensional Thin Layer Chromatography (TLC) on silica G plates with n-butanol/water/acetic acid (4:2:1, v/v) as a solvent for migration. Commercial phospholipid standards (phosphatidylethanolamine (PE) and phosphatidylglycerol (PG)) were included to permit identification of the phospholipids based on mobility. The reference standards were diluted with chloroform (PE) or methanol (PG) so as to contain known concentration of the particular component. The lipid spots are then detected by exposure to iodine vapour. Species that co-migrated with the solvent front also accumulated either in the lipid extract or in the phosphatidylethanolamine standard; these were probably degraded lipids or fatty acids.

Supplementary figure

FIGURE S1: **Lipid analysis.** Thin layer chromatogram of lipid extracts from purified FHP and of standard lipids and spotted on the pre-coated plates. 0.2 ml (70 nmole of heme protein; lane 1) and 0.3 ml (85 nmole of heme protein; lane 2) of purified protein was used for extraction. 50µg of phosphatidylethanolamine (PE) and phosphatidylglycerol (PG) were applied to the chromatogram. Phosphatidylglycerol but not phosphatidylethanolamine could be clearly identified.