

Supporting Information

Catalytic Formation of Silyl Enol Ethers and Its Applications for Aldol-Type Condensation and Aminomethylation Reactions

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General Information. All operations were carried out in an inert-atmosphere glove box or by using standard high vacuum and Schlenk techniques unless otherwise noted. Toluene, hexanes and Et₂O were distilled from purple solutions of sodium and benzophenone immediately prior to use. The NMR solvents were dried from activated molecular sieves (4 Å). All organic substrates were received from commercial sources and used without further purification. The ¹H, ²H, ¹³C and ³¹P NMR spectra were recorded on a 300 or 400 MHz Varian FT-NMR spectrometer. GC and GC-MS spectra were recorded from a Hewlett-Packard HP 6890 and Agilent 6850 spectrometers, respectively. Elemental analysis was performed at the Midwest Microlab, Indianapolis, IN.

Representative Procedure of the Catalytic Reaction: Silyl Enol Ether Formation. In a glove box, a ketone (2.0 mmol), CH₂=CHSiMe₃ (4.0 mmol) and complex **1** (7 mg, 0.5 mol %) were dissolved in toluene (3 mL) in a 25 mL Schlenk tube equipped with a magnetic stirring bar. The tube was brought out of the glove box, and was stirred in an oil bath set at 120 °C for 8-15 h. The tube was cooled to room temperature, and the crude product mixture was analyzed by GC-MS. For the detection of ethylene, the oven temperature of GC-MS was set at 25 °C (retention time = 1-2 min).

Aldol Condensation Reaction. The experiment was performed by following a reported procedure.¹ After evaporation of the solvent from the silyl enol ether solution, the crude product residue of **2** was dissolved in CH₂Cl₂ (2-3 mL). In a separate 100 mL Schlenk flask, TiCl₄ (3.0 mmol) was added to a cooled CH₂Cl₂ solution (5 mL) of 4-nitrobenzaldehyde (3.0 mmol) at 0 °C. After stirring for 15 min, the solution was cooled to -78 °C, and the CH₂Cl₂ solution of **2** was added dropwise via a syringe to the reaction flask. After stirring at -40 °C for 1 h, water (3 mmol) was added to the reaction flask, and the resulting mixture was stirred at 0 °C for 8 h. The reaction mixture was quenched by adding saturated Na₂CO₃ solution (10 mL), and the organic layer was extracted with Et₂O (3 × 20 mL). The ether solution was dried with anhydrous MgSO₄, and the Aldol product **3** was isolated by a column chromatograph on silica gel (*n*-hexanes/CH₂Cl₂).

Acylation Reaction. The experiment was performed by following a reported procedure.² The crude product residue of **2** (2.0 mmol) was dissolved in CH₂Cl₂ (2-3 mL). In a separate 100 mL Schlenk flask, TiCl₄ (3.0 mmol) was added to a cooled CH₂Cl₂ (5 mL) solution of acyl chloride (2.5 mmol) at 0 °C. After stirring for about 15 min, the crude product solution of **2** was added dropwise at -78 °C, and the mixture was further stirred for 1 h at -40 °C. The reaction mixture was quenched by adding aqueous NH₄Cl solution (10 mL), and the organic layer was extracted with Et₂O (3 × 20 mL). The combined ether solution was dried by anhydrous MgSO₄, and the product **4** was isolated by a column chromatograph on silica gel (EtOAc/*n*-hexanes).

Fluorination Reaction. The experiment was performed by following a reported procedure.³ In a 50 mL Schlenk flask, the crude product residue of **2** (2.0 mmol) was dissolved in CH₃CN (2 mL), and the solution was cooled to 0 °C in an ice bath. Selectfluor® (2.0 mmol) was added in several portions to the solution under N₂ purge. The reaction mixture was stirred while it was allowed to gradually warm to room temperature over 8 h. The solvent was evaporated, water (10 mL) was added to the residue, and the organic layer was extracted with Et₂O (3 × 20 mL). The combined organic layers were washed with brine solution, and dried over anhydrous Na₂SO₄. The analytically pure product **5** was isolated by a column chromatography on silica gel (EtOAc/*n*-hexanes).

Aminomethylation Reaction. The experiment was performed by following a reported procedure.⁴ In a 100 mL Schlenk flask, *t*-BuOOH (0.10 mL, 5-6 M in decane) was added dropwise to a mixture of *N,N*-dimethylaniline (1.5 mmol), CuBr (0.025 μmol) and the crude product **2** (0.50 mmol) dissolved in CH₃CN (5 mL) at room temperature. The resulting mixture was stirred at 50 °C for 12 h. The mixture was filtered through a pad of celite, and the solvent was removed under a reduced pressure. The residue was purified by a column chromatography on silica gel to afford the desired product **6**.

Deuterium Labeling Study. In a glove box, C₆D₅COCD₃ (26 mg) with CH₂=CHSiMe₃ (40 mg, 2.0 equiv) and **1** (0.5 mol %) were dissolved in toluene-*d*₈ (0.5 mL) in a J-Young NMR tube with a Teflon screw cap. The tube was brought out of the glove box, and was stirred in an

oil bath set at 120 °C for 12 h. The tube was cooled to room temperature, and the crude product mixture was analyzed by ^1H and ^2H NMR (Figure S1).

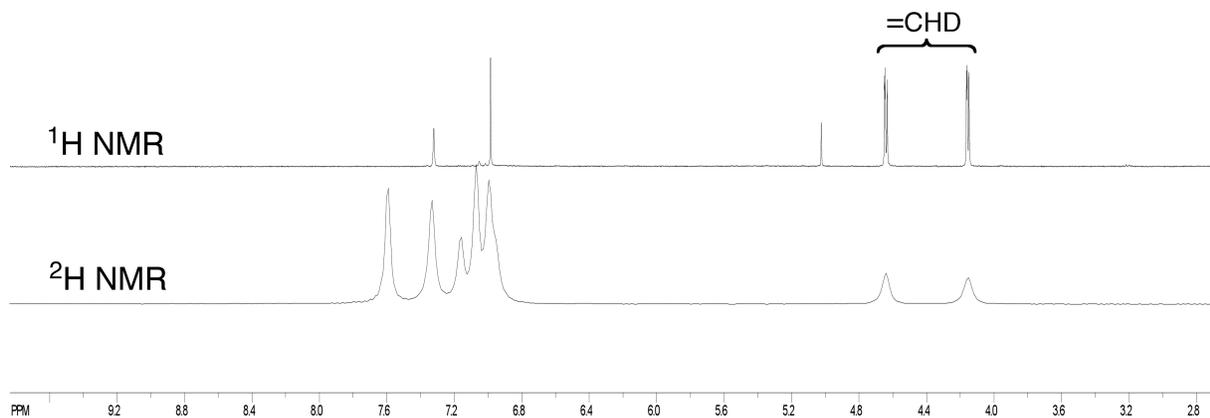


Figure S1. The ^1H and ^2H NMR Spectra of **3a-d**.

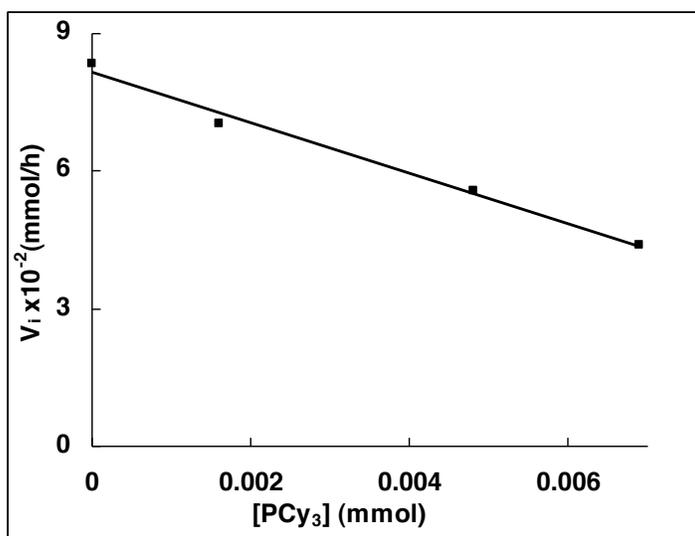


Figure S2. Plot of the Initial Rate (v_i) vs $[\text{PCy}_3]$ for the Coupling Reaction of Acetophenone and $\text{CH}_2=\text{CHSiMe}_3$.

Phosphine Inhibition Study. In a glove box, acetophenone (0.20 mmol), $\text{CH}_2=\text{CHSiMe}_3$ (0.40 mmol), **1** (1 mg, 0.5 mol %) and C_6Me_6 (2 mg, internal standard) were dissolved in

toluene- d_8 (0.5 mL) solution in a J-Young NMR tube with a Teflon screw cap. A predissolved PCy_3 in toluene- d_8 solution (5 μL , 1.0 M) was added to the tube via syringe. The tube was brought out of the glove box and was heated in an oil bath set at 120 $^\circ\text{C}$. The reaction was monitored by ^1H NMR in 30 min intervals. The rate was measured by the ^1H integration of the product peak, and was normalized against the internal standard peak. The k_{obs} was estimated from the first order plot of $\ln[\text{product}]$ vs reaction time.

Hammett Study. In the glove box, **1** (0.5 mol %), *para*-X- $\text{C}_6\text{H}_4\text{COMe}$ (0.20 mmol), $\text{CH}_2=\text{CHSiMe}_3$ (0.40 mmol) and C_6Me_6 (26 mg, internal standard) were dissolved in toluene- d_8 (0.5 mL) in a J-Young NMR tube with a Teflon screw cap. The tube was brought out of the box and was immersed in an oil bath set at 120 $^\circ\text{C}$. The reaction progress was monitored by ^1H NMR in 30 min intervals by measuring the ^1H integration of the product peaks, which were normalized against the internal standard peak. The k_{obs} was estimated from a first-order plot of $\ln[\text{product}]$ vs reaction time (Figure S3).

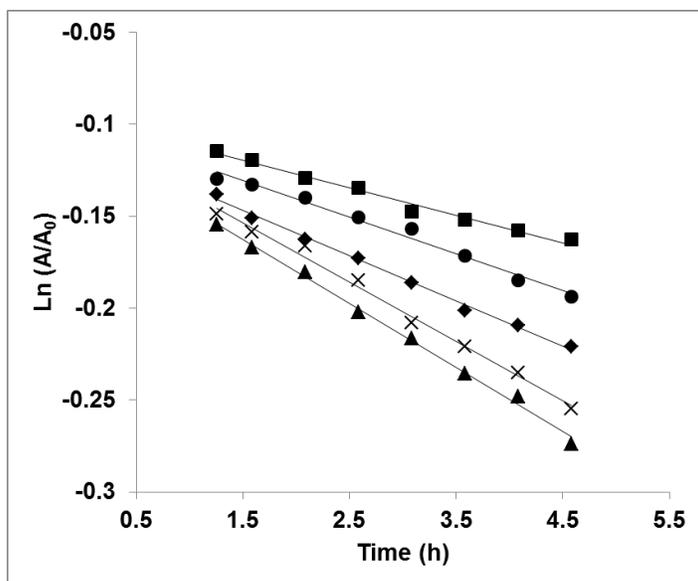


Figure S3. First-Order Plots of $\ln[\text{product}]$ vs Reaction Time for the Coupling Reaction of *para*-Substituted $p\text{-X-C}_6\text{H}_4\text{COCH}_3$ (X = OMe(■), CH_3 (●), H(◆), Cl(✕), Br(▲)) with $\text{CH}_2=\text{CHSiMe}_3$.

Characterization Data of Organic Products

For **3a**: ^1H NMR (400 MHz, CDCl_3) δ 8.16 (d, $J = 8.9$ Hz, 2H), 7.92 (d, $J = 7.8$ Hz, 2H), 7.50 (d, $J = 8.9$ Hz, 2H), 7.5-7.4 (m, 3H), 5.44 (s, 1H), 3.99 (s, 1H), 3.35 (m, 2H) ppm. $^{13}\text{C}\{^1\text{H}\}$ NMR (75 MHz, CDCl_3) δ 199.6, 150.5, 147.4, 136.3, 134.2, 129.0, 126.7, 123.9, 69.3, 47.2 ppm. GC-MS $m/z = 271$ (M^+). The ^1H and ^{13}C NMR spectral data are in good agreement with the literature data.⁵

For **3b**: ^1H NMR (300 MHz, CDCl_3) δ 8.16 (d, $J = 8.9$ Hz, 2H), 7.81 (d, $J = 8.1$ Hz, 2H), 7.57 (d, $J = 8.9$ Hz, 2H), 7.23 (d, $J = 8.1$ Hz, 2H), 5.44 (m, 1H), 4.04 (s, 1H), 3.33 (m, 2H), 2.39 (s, 3H) ppm. $^{13}\text{C}\{^1\text{H}\}$ NMR (100 MHz, CDCl_3) δ 199.1, 150.5, 147.2, 145.0, 133.7, 129.5, 128.3, 126.6, 123.7, 69.3, 46.8, 21.7 ppm. GC-MS $m/z = 285$ (M^+). The ^1H and ^{13}C NMR spectral data are in good agreement with the literature data.⁶

For **3c**: ^1H NMR (300 MHz, CDCl_3) δ 8.16 (d, $J = 8.9$ Hz, 2H), 7.88 (d, $J = 8.4$ Hz, 2H), 7.57 (d, $J = 8.9$ Hz, 2H), 6.90 (d, $J = 8.4$ Hz, 2H), 5.44 (m, $J = 4.0$ Hz, 1H), 4.10 (s, 1H), 3.85 (s, 3H), 3.31 (m, 2H) ppm. $^{13}\text{C}\{^1\text{H}\}$ NMR (75 MHz, CDCl_3) δ 198.2, 164.4, 150.7, 147.4, 130.8, 129.5, 126.8, 124.0, 114.2, 69.6, 55.8, 46.8 ppm. GC-MS $m/z = 301$ (M^+). The ^1H and ^{13}C NMR spectral data are in good agreement with the literature data.⁶

For **3d**: ^1H NMR (300 MHz, CDCl_3) δ 8.16 (d, $J = 8.9$ Hz, 2H), 7.88 (d, $J = 8.4$ Hz, 2H), 7.57 (d, $J = 8.9$ Hz, 2H), 6.90 (d, $J = 8.4$ Hz, 2H), 5.44 (m, 1H), 4.10 (s, 1H), 3.65 (m, 1H), 3.31 (m, 2H) ppm. $^{13}\text{C}\{^1\text{H}\}$ NMR (75 MHz, CDCl_3) δ 198.2, 164.4, 150.7, 147.4, 130.8, 129.5, 126.8, 124.0, 114.2, 69.6, 47.3 ppm. GC-MS $m/z = 350$ (M^+). Anal. Calcd for $\text{C}_{15}\text{H}_{12}\text{NO}_4\text{Br}$: C, 51.45; H, 3.45. Found C, 51.26; H, 3.54.

For **3e**: ^1H NMR (300 MHz, CDCl_3) δ 8.21 (d, $J = 8.8$ Hz, 2H), 7.88 (d, $J = 8.6$ Hz, 2H), 7.60 (d, $J = 8.8$ Hz, 2H), 7.44 (d, $J = 8.8$ Hz, 2H), 5.45 (m, 1H), 3.77 (s, 1H), 3.33 (m, 2H) ppm.

$^{13}\text{C}\{^1\text{H}\}$ NMR (75 MHz, CDCl_3) δ 198.4, 150.3, 147.6, 140.8, 134.7, 129.8, 129.4, 126.8, 124.1, 69.3, 47.3 ppm. GC-MS $m/z = 305$ (M^+). The ^1H and ^{13}C NMR spectral data are in good agreement with the literature data.⁶

For **3f**: ^1H NMR (300 MHz, CDCl_3) δ 8.44 (s, 1H), 8.22 (d, $J = 8.8$ Hz, 2H), 7.93 (m, 4H), 7.64 (d, $J = 8.8$ Hz, 2H), 7.61 (m, 3H), 5.51 (m, 1H), 3.97 (s, 1H), 3.51 (m, 2H) ppm. $^{13}\text{C}\{^1\text{H}\}$ NMR (75 MHz, CDCl_3) δ 199.6, 150.5, 147.5, 136.2, 133.7, 132.6, 130.5, 129.9, 129.3, 128.1, 127.3, 126.8, 124.1, 123.6, 69.7, 47.3 ppm. GC-MS $m/z = 321$ (M^+). Anal. Calcd for $\text{C}_{19}\text{H}_{15}\text{NO}_4$: C, 71.02; H, 4.71. Found C, 70.91; H, 4.63.

For **3g**: ^1H NMR (300 MHz, CDCl_3) δ 8.23 (d, $J = 8.8$ Hz, 2H), 7.61 (d, $J = 8.8$ Hz, 2H), 7.26 (m, 3H), 6.95 (m, 1H), 5.43 (m, 1H), 3.95 (s, 1H), 3.35 (m, 2H), 3.00 (s, 6H) ppm. $^{13}\text{C}\{^1\text{H}\}$ NMR (75 MHz, CDCl_3) δ 200.7, 150.9, 150.6, 147.5, 137.1, 129.7, 126.8, 124.0, 118.0, 116.6, 111.0, 69.6, 47.2, 40.7 ppm. GC-MS $m/z = 314$ (M^+). Anal. Calcd for $\text{C}_{17}\text{H}_{18}\text{N}_2\text{O}_4$: C, 64.96; H, 5.77. Found C, 65.14; H, 5.47.

For **3h** (*syn*): ^1H NMR (400 MHz, CDCl_3) δ 8.16 (d, $J = 8.8$ Hz, 2H), 7.93 (d, $J = 8.9$ Hz, 2H), 7.62-7.44 (m, 5H), 5.34 (d, $J = 2.3$ Hz, 1H), 4.08 (s, 1H), 3.79 (m, 1H), 1.15 (d, $J = 7.2$ Hz, 3H) ppm. $^{13}\text{C}\{^1\text{H}\}$ NMR (75 MHz, CDCl_3) δ 205.3, 149.3, 147.1, 135.1, 134.0, 129.1, 128.7, 127.1, 123.7, 72.5, 46.8, 11.2 ppm. GC-MS $m/z = 285$ (M^+). The ^1H and ^{13}C NMR spectral data are in good agreement with the literature data.⁷

For **3h** (*anti*): ^1H NMR (400 MHz, CDCl_3) δ 8.16 (d, $J = 8.8$ Hz, 2H), 7.93 (d, $J = 8.9$ Hz, 2H), 7.62-7.44 (m, 5H), 5.08 (d, $J = 7.2$ Hz, 1H), 4.08 (s, 1H), 3.82 (m, 1H), 1.15 (d, $J = 7.2$ Hz, 3H) ppm. $^{13}\text{C}\{^1\text{H}\}$ NMR (75 MHz, CDCl_3) δ 204.4, 149.8, 147.6, 136.1, 133.8, 129.0, 128.6, 127.8, 123.8, 75.9, 47.8, 15.9 ppm. GC-MS $m/z = 285$ (M^+). The ^1H and ^{13}C NMR spectral data are in good agreement with the literature data.⁷

For **3i** (*syn*): ^1H NMR (300 MHz, CDCl_3) δ 8.3-6.8 (m, 14H), 5.70 (d, $J = 3.4$ Hz 1H), 4.78 (d, $J = 3.4$ Hz, 1H), 3.90 (s, 1H) ppm. $^{13}\text{C}\{^1\text{H}\}$ NMR (75 MHz, CDCl_3) δ 202.1, 148.8, 147.6, 146.0, 138.1, 129.4, 133.4, 132.1, 130.4, 128.7, 128.3, 127.2, 124.1, 74.5, 60.0 ppm. GC-MS $m/z = 347$ (M^+). The ^1H and ^{13}C NMR spectral data are in good agreement with the literature data.⁸

For **3i** (*anti*): ^1H NMR (300 MHz, CDCl_3) δ 8.10-6.85 (m, 14H), 5.55 (d, $J = 8.9$ Hz, 1H), 4.68 (d, $J = 3.4$ Hz, 1H), 3.50 (s, 1H) ppm. $^{13}\text{C}\{^1\text{H}\}$ NMR (75 MHz, CDCl_3) δ 198.1, 148.0, 147.0, 145.3, 137.9, 129.3, 133.5, 132.1, 130.4, 128.7, 128.4, 127.5, 124.3, 76.5, 62.3 ppm. GC-MS $m/z = 347$ (M^+). The ^1H and ^{13}C NMR spectral data are in good agreement with the literature data.⁸

For **3j**: ^1H NMR (400 MHz, CDCl_3) δ 8.2-7.4 (m, 8H), 5.06 (s, 1H), 4.98 (d, $J = 8.9$ Hz, 1H), 3.00 (m, 1H), 2.73 (m, 2H) ppm. $^{13}\text{C}\{^1\text{H}\}$ NMR (100 MHz, CDCl_3) δ 209.0, 153.6, 148.7, 148.0, 136.2, 136.1, 128.2, 128.0, 126.8, 124.5, 124.0, 75.0, 53.1, 29.7 ppm. GC-MS $m/z = 283$ (M^+). Anal. Calcd for $\text{C}_{16}\text{H}_{13}\text{NO}_4$: C, 67.84; H, 4.63. Found C, 67.97; H, 4.51.

For **3k**: ^1H NMR (300 MHz, CDCl_3) δ 8.21 (d, $J = 8.8$ Hz, 2H), 8.05 (d, $J = 8.0$ Hz, 2H), 7.58 (d, $J = 8.8$ Hz, 2H), 7.54-7.23 (m, 3H), 5.11 (d, $J = 8.0$ Hz, 1H), 5.02 (s, 1H), 2.89 (m, 2H), 2.75 (m, 1H), 1.68 (m, 2H) ppm. $^{13}\text{C}\{^1\text{H}\}$ NMR (75 MHz, CDCl_3) δ 201.6, 148.7, 147.8, 144.4, 134.6, 132.2, 129.0, 128.3, 127.8, 127.2, 123.8, 74.8, 53.9, 28.9, 26.2 ppm. GC-MS $m/z = 297$ (M^+). Anal. Calcd for $\text{C}_{17}\text{H}_{15}\text{NO}_4$: C, 68.68; H, 5.09. Found C, 68.97; H, 4.93.

For **3l**: ^1H NMR (400 MHz, CDCl_3) δ 7.6-7.4 (m, 12H), 3.09 (s, 2H) ppm. $^{13}\text{C}\{^1\text{H}\}$ NMR (100 MHz, CDCl_3) δ 196.5, 137.5, 135.9, 134.0, 130.9, 128.9, 129.6, 26.7 ppm. GC-MS $m/z = 260$ (M^+). The ^1H and ^{13}C NMR spectral data are in good agreement with the literature data.⁹

For **3m**: ^1H NMR (400 MHz, CDCl_3) δ 7.82 (s, 2H), 7.52-7.32 (m, 10H), 2.93 (t, $J = 6.1$ Hz, 2H), 1.78 (t, $J = 6.1$ Hz, 2H) ppm. $^{13}\text{C}\{^1\text{H}\}$ NMR (75 MHz, CDCl_3) δ 190.3, 137.0, 136.2, 136.0,

130.4, 128.4, 128.6, 28.5, 23.0 ppm. GC-MS $m/z = 274$ (M^+). The ^1H and ^{13}C NMR spectral data are in good agreement with the literature data.⁹

For **3n**: ^1H NMR (300 MHz, CDCl_3) δ 7.82 (s, 2H), 7.52-7.32 (m, 10H), 3.15 (d, $J = 15.6$ Hz, 2H), 2.45 (t, $J = 13.8$ Hz, 2H), 1.48 (m, 1H), 0.96 (s, 9H) ppm. $^{13}\text{C}\{^1\text{H}\}$ NMR (75 MHz, CDCl_3) δ 190.9, 137.0, 136.4, 136.2, 130.6, 128.8, 128.7, 44.6, 32.8, 29.8, 27.5 ppm. GC-MS $m/z = 330$ (M^+). The ^1H and ^{13}C NMR spectral data are in good agreement with the literature data.¹⁰

For **3o**: ^1H NMR (300 MHz, CDCl_3) δ 8.16 (d, $J = 8.9$ Hz, 2H), 7.50 (d, $J = 8.9$ Hz, 2H), 5.26 (s, 1H), 3.73 (s, 1H), 2.44 (m, $J = 13.8$ Hz, 1H), 2.60-1.20 (m, 10H) ppm. $^{13}\text{C}\{^1\text{H}\}$ NMR (75 MHz, CDCl_3) δ 217.7, 149.8, 147.2, 126.9, 123.6, 72.6, 57.4, 44.0, 29.3, 29.2, 24.0, 23.7 ppm. GC-MS $m/z = 263$ (M^+). The ^1H and ^{13}C NMR spectral data are in good agreement with the literature data.¹¹

For **3p** (*syn*): ^1H NMR (300 MHz, CDCl_3) δ 8.10 (d, $J = 8.9$ Hz, 2H), 7.32 (d, $J = 8.9$ Hz, 2H), 5.26 (s, 1H), 4.00 (s, 1H), 3.26-1.68 (m, 8H), 1.37 (s, 3H) ppm. $^{13}\text{C}\{^1\text{H}\}$ NMR (75 MHz, CDCl_3) δ 214.4, 148.1, 146.8, 130.1, 123.9, 77.7, 47.9, 42.9, 35.7, 34.7, 25.3, 22.5 ppm. GC-MS $m/z = 263$ (M^+). The ^1H and ^{13}C NMR spectral data are in good agreement with the literature data.¹²

For **3p** (*anti*): ^1H NMR (300 MHz, CDCl_3) δ 8.13 (d, $J = 8.9$ Hz, 2H), 7.47 (d, $J = 8.9$ Hz, 2H), 5.03 (s, 1H), 4.25 (s, 1H), 3.26-1.68 (m, 8H), 1.14 (s, 3H) ppm. $^{13}\text{C}\{^1\text{H}\}$ NMR (75 MHz, CDCl_3) δ 219.0, 147.6, 146.8, 129.3, 123.0, 76.9, 52.7, 39.2, 37.1, 27.5, 20.7, 16.1 ppm. GC-MS $m/z = 263$ (M^+). The ^1H and ^{13}C NMR spectral data are in good agreement with the literature data.¹²

For **3q** (*syn*): ^1H NMR (300 MHz, CDCl_3) δ 8.04 (d, $J = 8.9$ Hz, 2H), 7.41 (d, $J = 8.9$ Hz, 2H), 5.17 (s, 1H), 3.97 (s, 1H), 2.81 (m, 1H), 2.02 (s, 3H), 1.44 (m, 2H), 1.09 (m, 4H), 0.69 (t, J

= 6.6 Hz, 3H) ppm. $^{13}\text{C}\{^1\text{H}\}$ NMR (75 MHz, CDCl_3) δ 211.1, 150.6, 147.2, 126.5, 123.6, 69.0, 50.6, 43.6, 31.2, 23.1, 22.4, 13.9 ppm. GC-MS m/z = 265 (M^+). The ^1H and ^{13}C NMR spectral data are in good agreement with the literature data.¹³

For **3q** (*anti*): ^1H NMR (300 MHz, CDCl_3) δ 8.04 (d, J = 8.9 Hz, 2H), 7.41 (d, J = 8.9 Hz, 2H), 4.94 (s, 1H), 3.74 (s, 1H), 2.81 (m, 1H), 2.02 (s, 3H), 1.44 (m, 2H), 1.09 (m, 4H), 0.69 (t, J = 6.6 Hz, 3H) ppm. $^{13}\text{C}\{^1\text{H}\}$ NMR (75 MHz, CDCl_3) δ 213.1, 149.8, 147.1, 127.1, 123.4, 72.9, 58.8, 31.5, 27.3, 23.4, 13.8 ppm. GC-MS m/z = 265 (M^+). The ^1H and ^{13}C NMR spectral data are in good agreement with the literature data.¹³

For **4a**: ^1H NMR (400 MHz, CDCl_3) δ 7.9-7.4 (m, 5H), 6.18 (s, 1H), 4.05 (s, 1H), 2.18 (s, 3H) ppm. $^{13}\text{C}\{^1\text{H}\}$ NMR (75 MHz, CDCl_3) δ 193.8, 183.3, 134.8, 132.3, 128.6, 127.0, 96.7, 25.9 ppm. GC-MS m/z = 162 (M^+). The ^1H and ^{13}C NMR spectral data are in good agreement with the literature data.²

For **5a**: ^1H NMR (300 MHz, CDCl_3) δ 7.8-7.2 (m, 5H), 5.55 (d, J_{HF} = 46.9 Hz, 2H); $^{13}\text{C}\{^1\text{H}\}$ NMR (75 MHz, CDCl_3) δ 193.1 (d, J_{CF} = 15.0 Hz), 145.4, 132.4, 130.4 and 127.6, 84.0 (d, J_{CF} = 188.9 Hz) ppm. GC-MS m/z = 138 (M^+). The ^1H and ^{13}C NMR spectral data are in good agreement with the literature data.³

For **5j**: ^1H NMR (300 MHz, CDCl_3) δ 7.70-7.30 (m, 4H), 5.14 (m, J_{HF} = 51.0 Hz, 1H), 3.35 (m, 1H), 3.08 (m, 1H) ppm. $^{13}\text{C}\{^1\text{H}\}$ NMR (75 MHz, CDCl_3) δ 199.9 (d, J_{CF} = 14.9 Hz), 149.7, 136.4, 133.7, 128.3, 126.9, 124.4, 91.7 (d, J_{CF} = 189.9 Hz), 33.2 (d, J_{CF} = 21.4 Hz) ppm. GC-MS m/z = 150 (M^+). The ^1H and ^{13}C NMR spectral data are in good agreement with the literature data.³

For **5k**: ^1H NMR (400 MHz, CDCl_3) δ 8.0-7.25 (m, 4H), 5.14 (ddd, J_{HF} = 46.9 Hz, J_{HH} = 12.8, 5.2 Hz, 1H), 3.12 (m, 2H), 2.56 (m, 1H), 2.34 (m, 1H) ppm. $^{13}\text{C}\{^1\text{H}\}$ NMR (100 MHz, CDCl_3) δ 193.6 (d, J_{CF} = 14.8 Hz), 143.2, 134.4, 131.4, 128.9, 128.0, 127.4, 91.2 (d, J_{CF} = 188.4

Hz), 30.4 (d, $J_{CF} = 19.4$ Hz), 27.1 (d, $J_{CF} = 11.6$ Hz) ppm. GC-MS $m/z = 164$ (M^+). The 1H and ^{13}C NMR spectral data are in good agreement with the literature data.³

For **6a**: 1H NMR (300 MHz, $CDCl_3$) δ 7.95-6.72 (m, 10H), 3.85 (t, $J = 7.0$ Hz, 2H), 3.24 (t, $J = 7.0$ Hz, 2H), 2.98 (s, 3H) ppm. $^{13}C\{^1H\}$ NMR (75 MHz, $CDCl_3$) δ 199.7, 148.8, 137.1, 133.4, 129.5, 128.8, 128.2, 116.7, 112.6, 48.1, 38.7, 35.3 ppm. GC-MS $m/z = 239$ (M^+). The 1H and ^{13}C NMR spectral data are in good agreement with the literature data.⁴

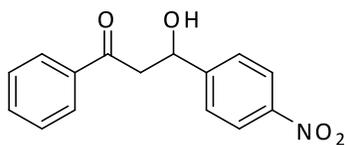
For **6j**: 1H NMR (300 MHz, $CDCl_3$) δ 7.26-6.75 (m, 5H), 3.89 (dd, $J = 15.1, 4.5$ Hz, 1H), 3.28 (dd, $J = 15.1, 8.0$ Hz, 1H), 2.96 (s, 3H), 2.53-2.45 (m, 1H), 2.37-1.63 (m, 6H) ppm. $^{13}C\{^1H\}$ NMR (75 MHz, $CDCl_3$) δ 220.2, 148.8, 129.3, 116.5, 112.3, 52.5, 48.4, 39.2, 38.1, 29.2, 20.8 ppm. GC-MS $m/z = 203$ (M^+). The 1H and ^{13}C NMR spectral data are in good agreement with the literature data.⁴

For **6k**: 1H NMR (300 MHz, $CDCl_3$) δ 7.30-6.70 (m, 5H), 3.89 (dd, $J = 15.1, 5.6$ Hz, 1H), 3.26 (dd, $J = 15.3, 7.2$ Hz, 1H), 3.02 (s, 3H), 2.78 (m, 1H), 2.50-1.42 (m, 8H) ppm. $^{13}C\{^1H\}$ NMR (75 MHz, $CDCl_3$) δ 212.5, 149.0, 129.2, 115.9, 111.7, 52.2, 49.2, 42.3, 39.6, 32.5, 27.9, 25.0 ppm. GC-MS $m/z = 217$ (M^+). The 1H and ^{13}C NMR spectral data are in good agreement with the literature data.⁴

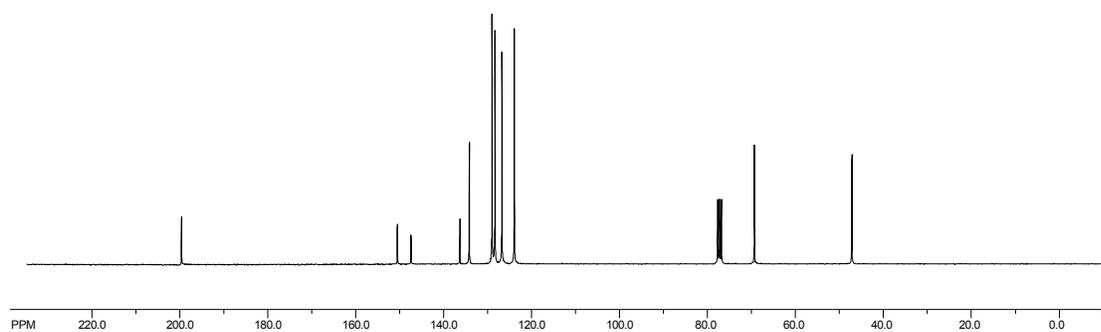
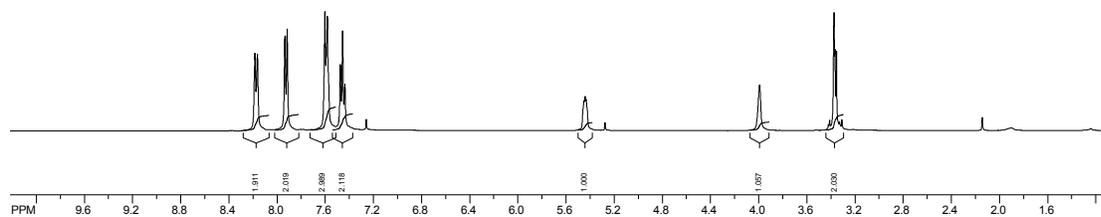
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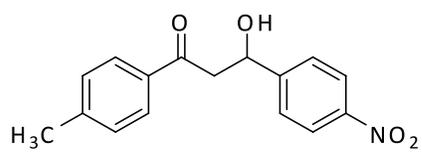
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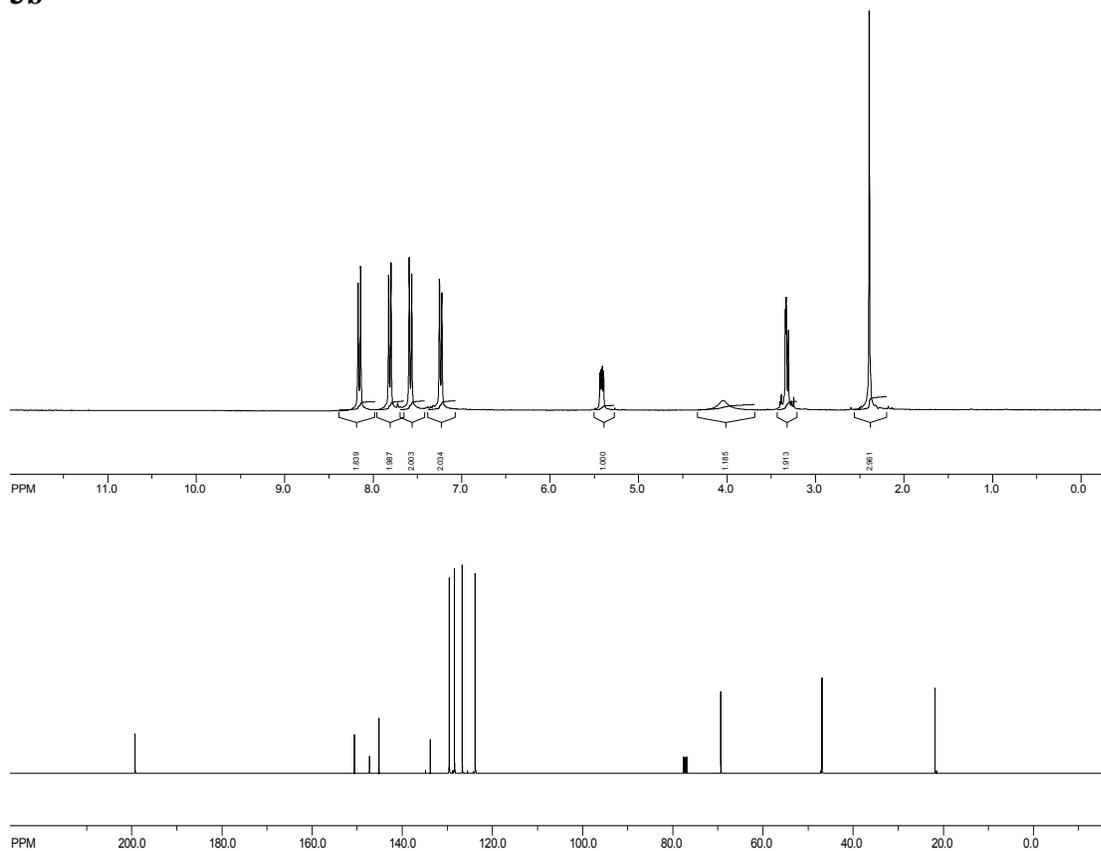


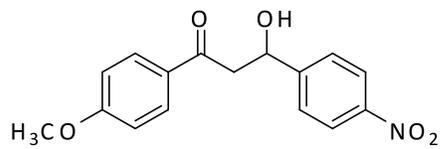
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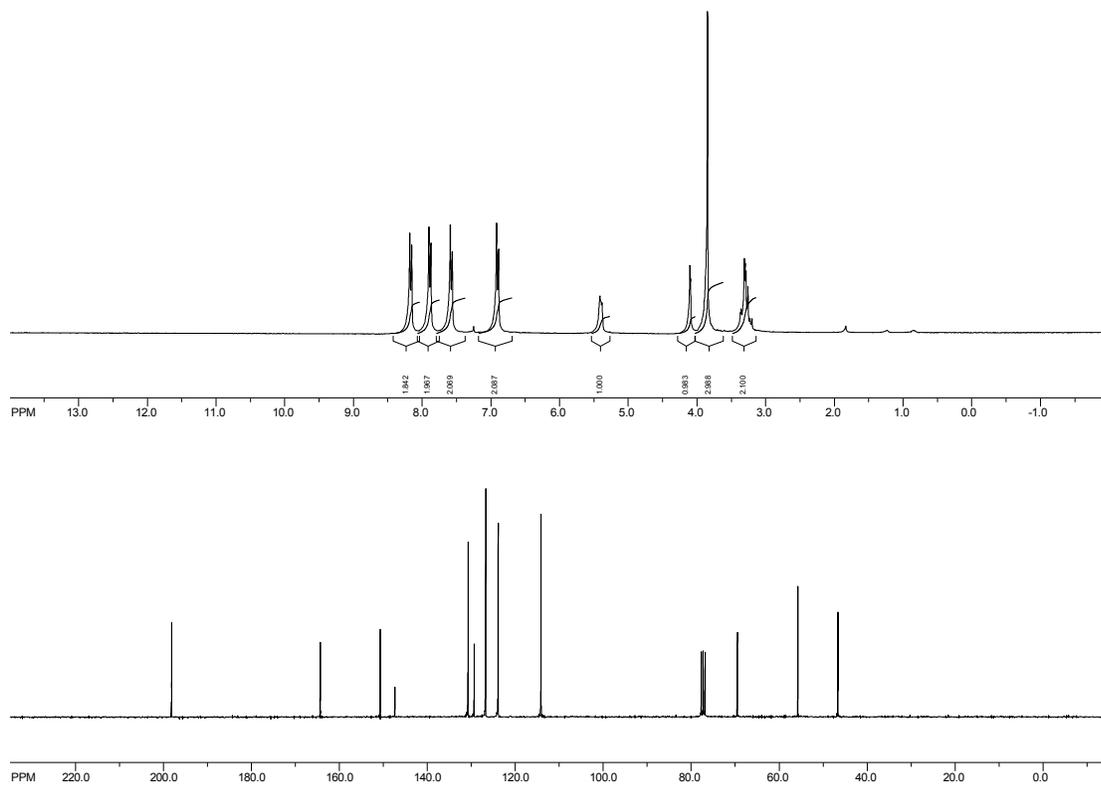


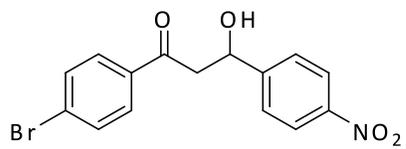
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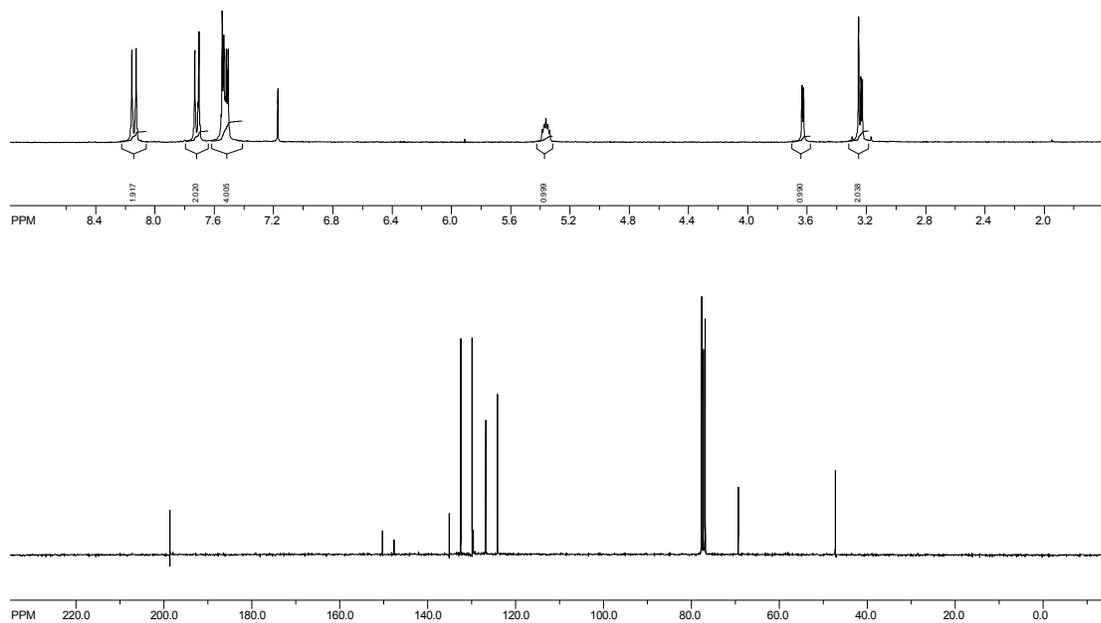


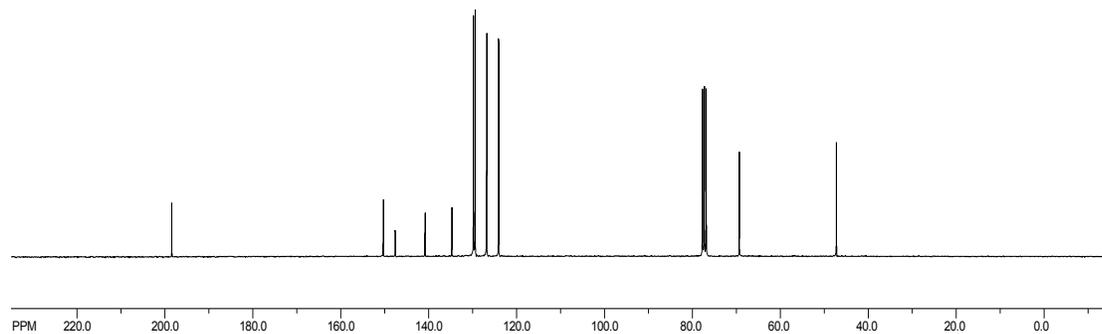
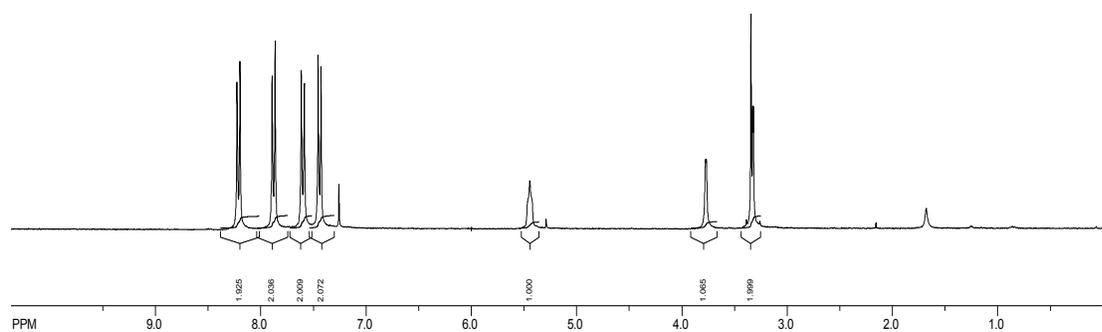
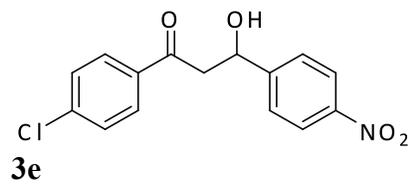
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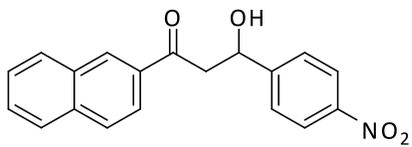




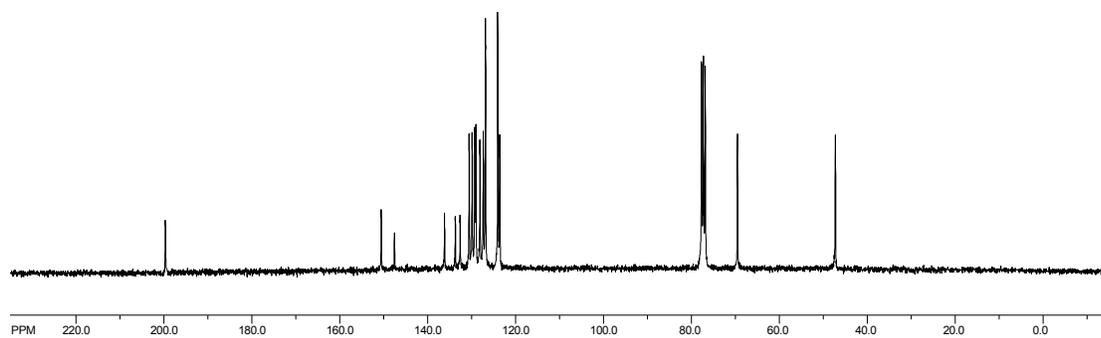
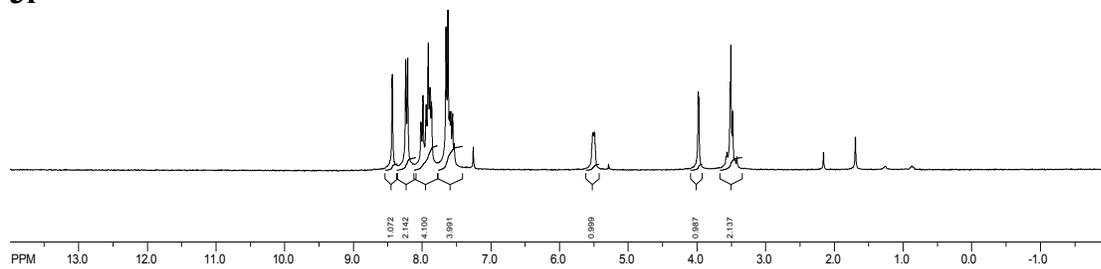
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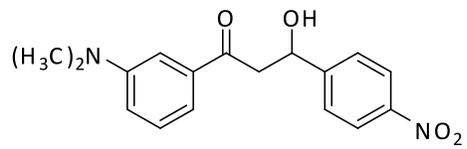




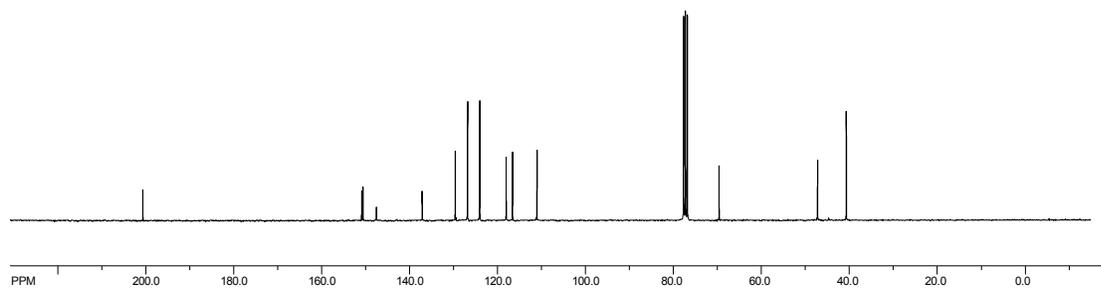
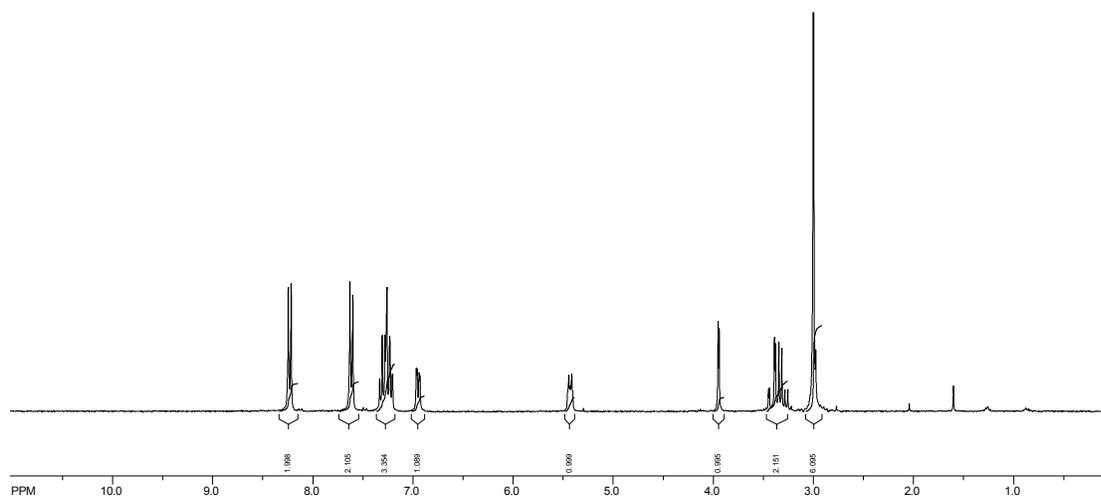


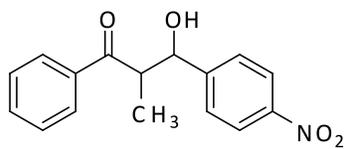
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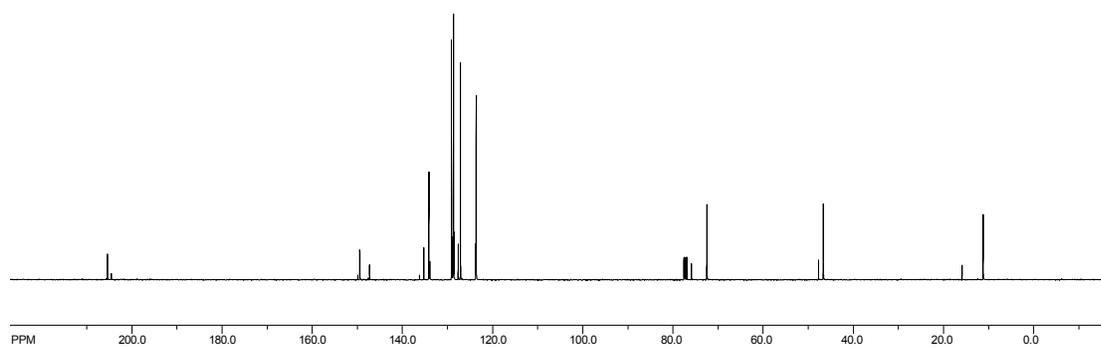
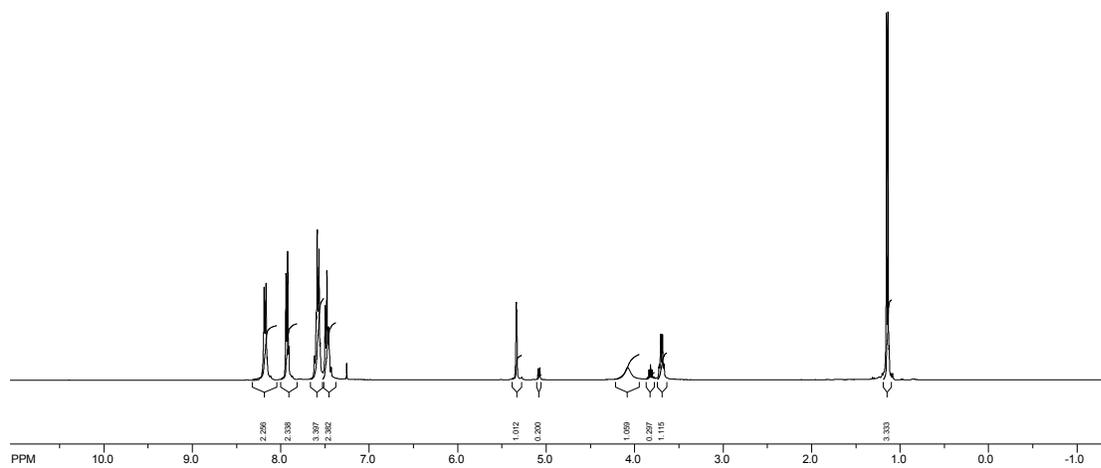


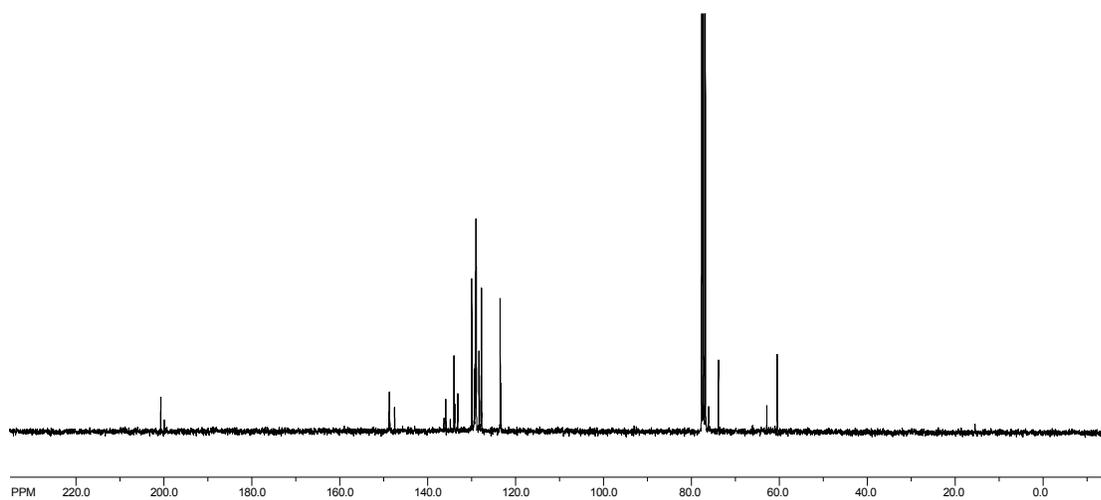
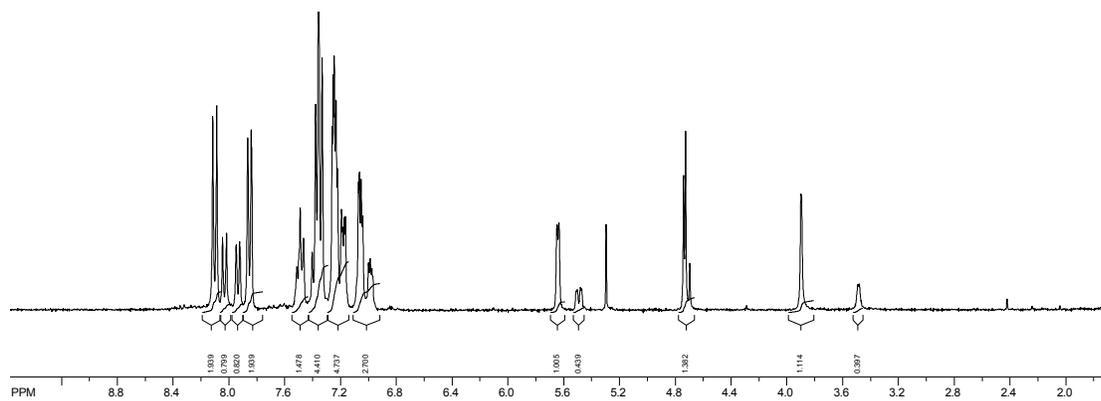
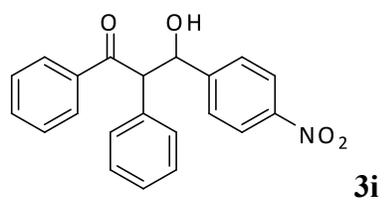
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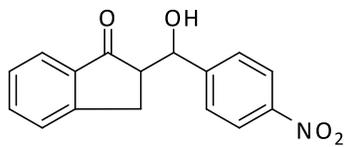




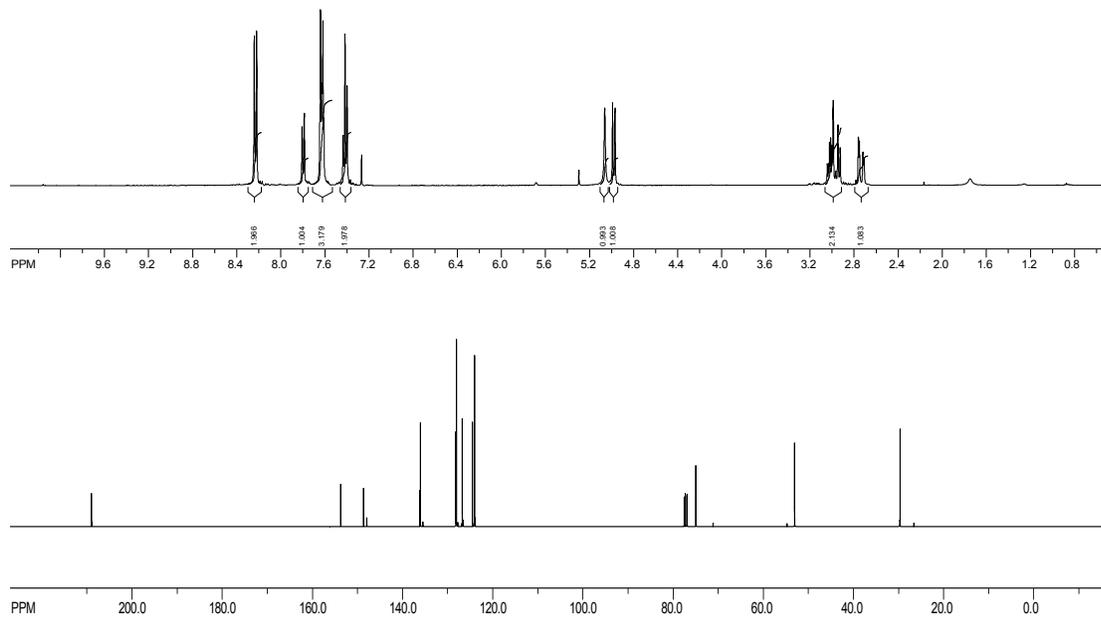
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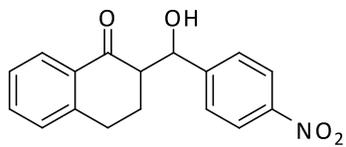




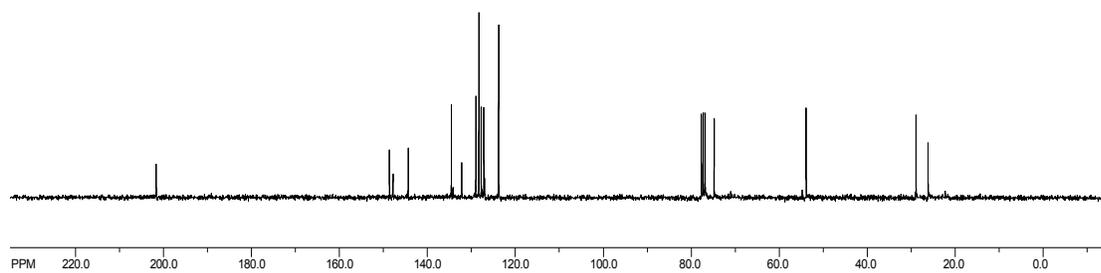
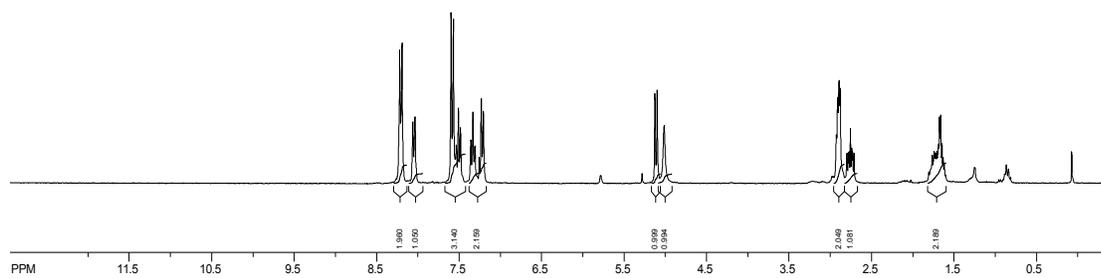


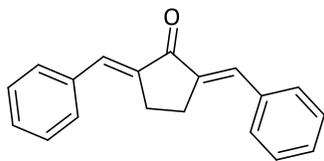
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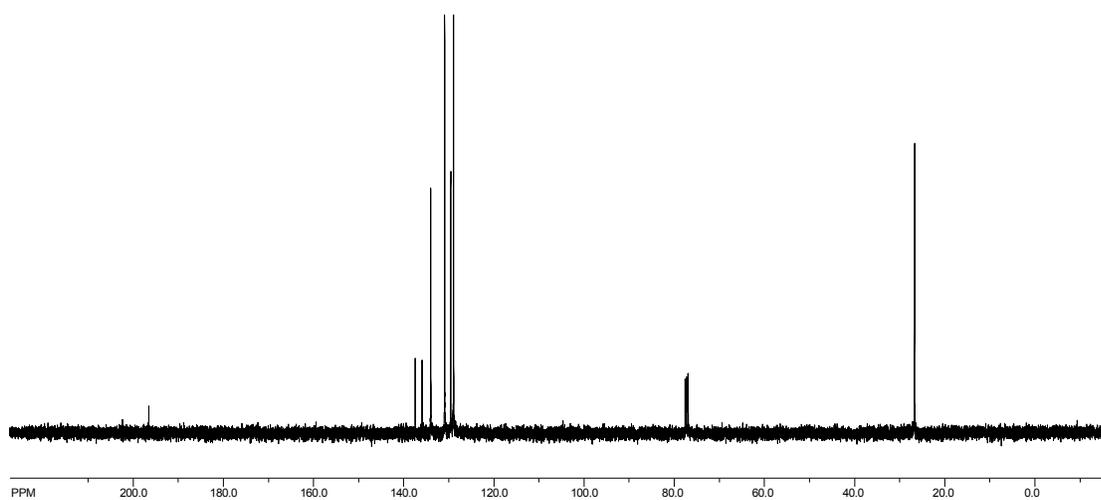
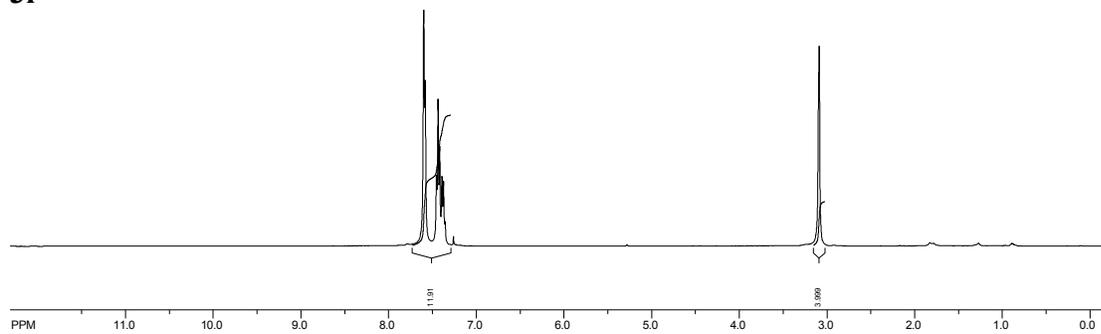


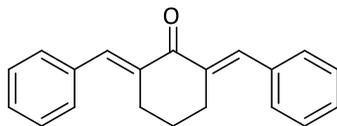
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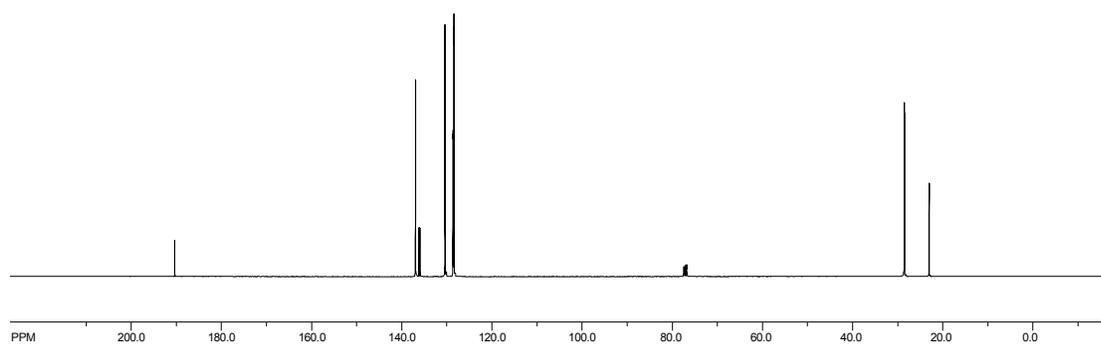
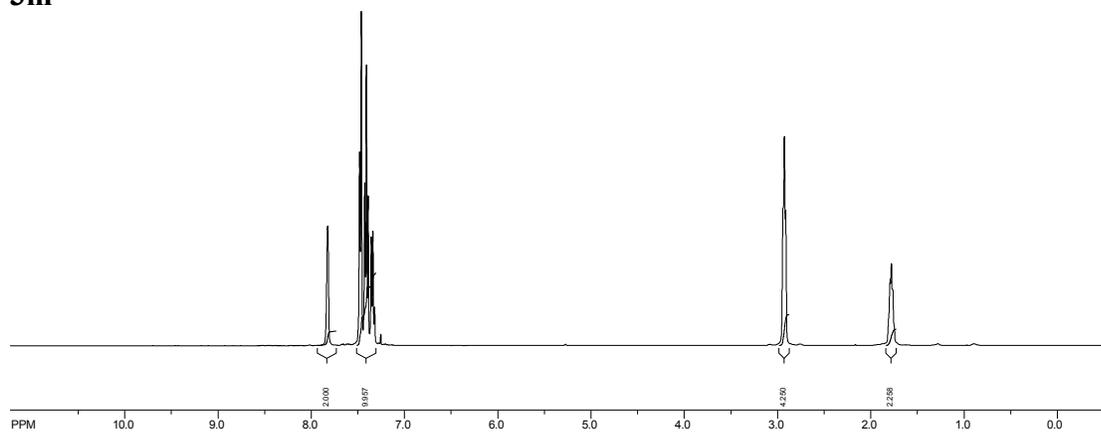


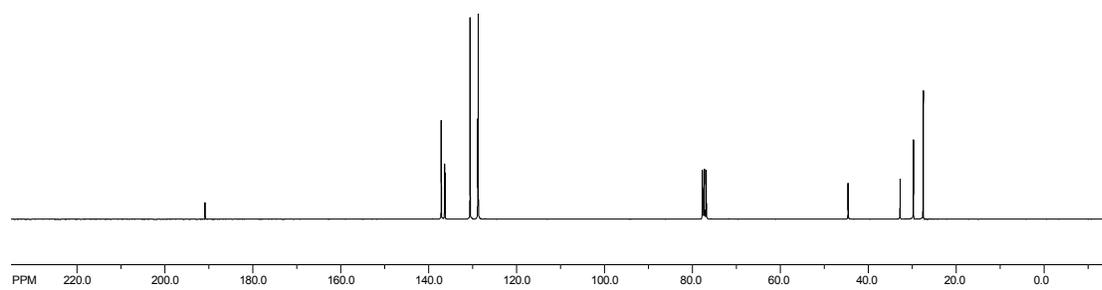
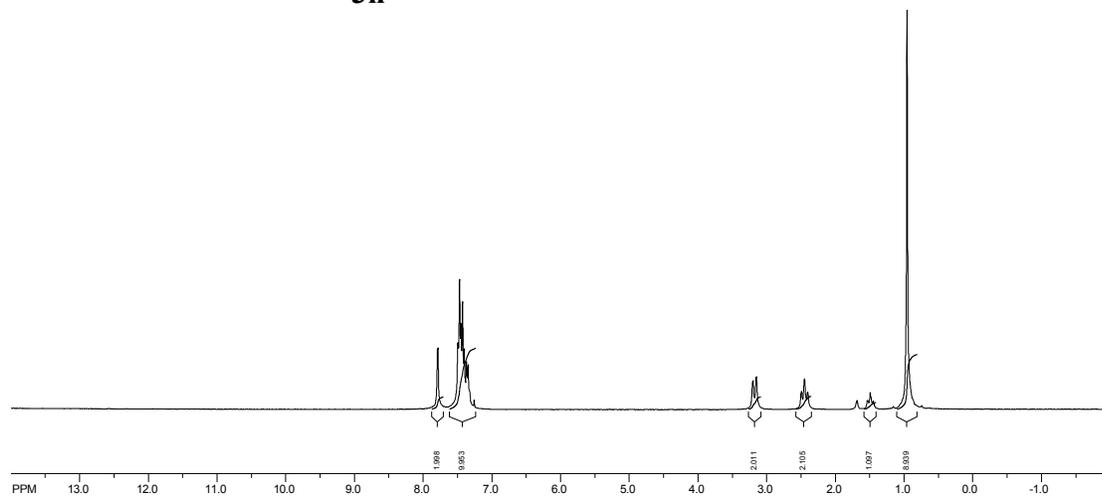
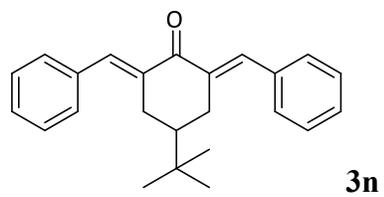
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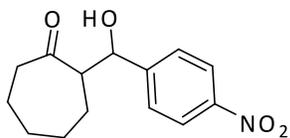




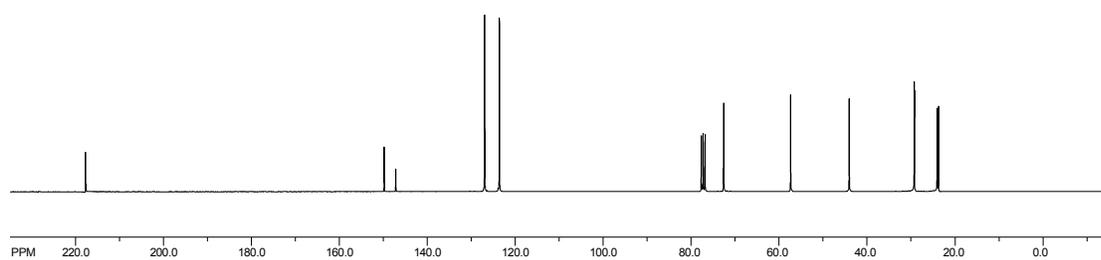
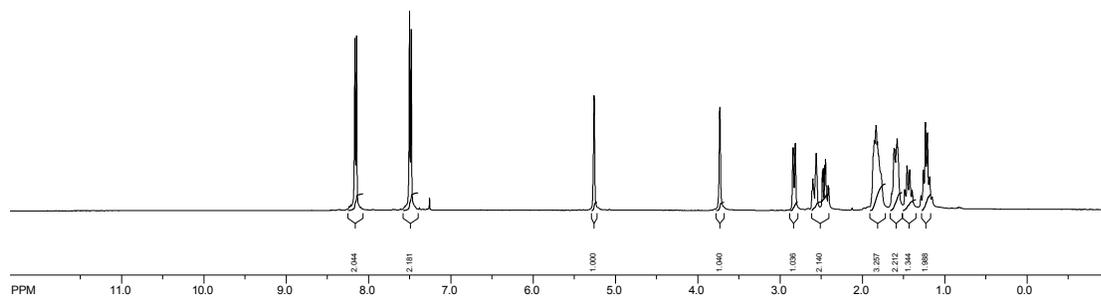
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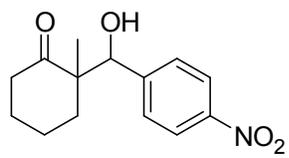




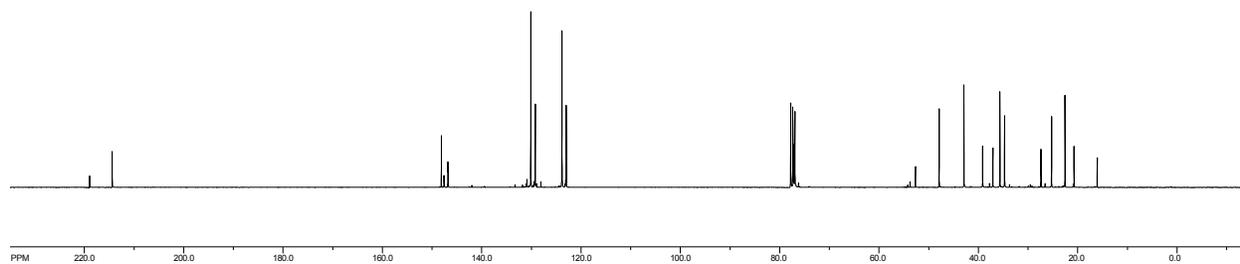
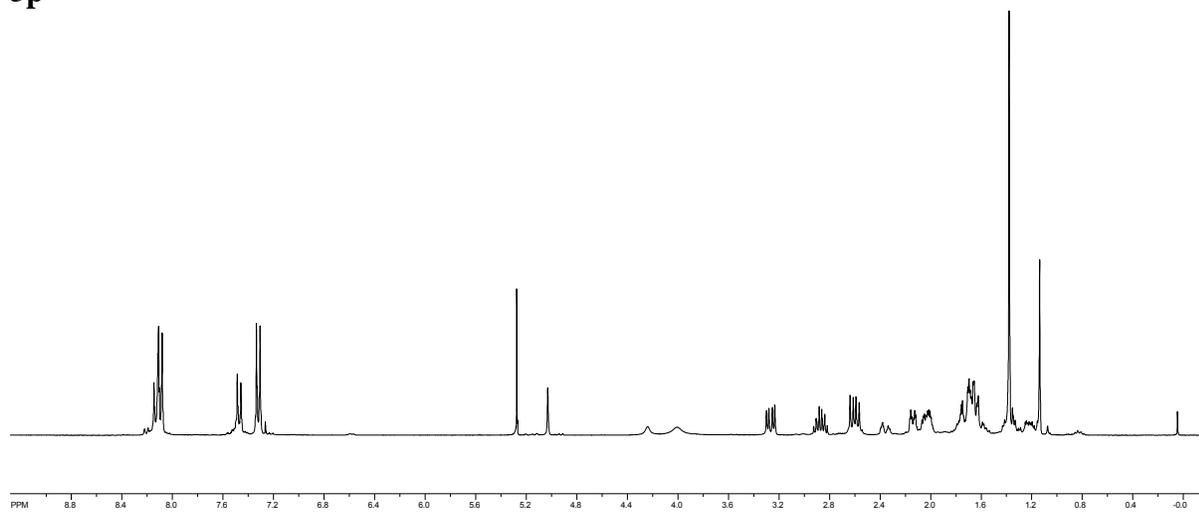


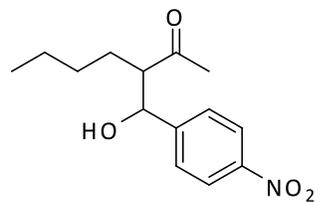
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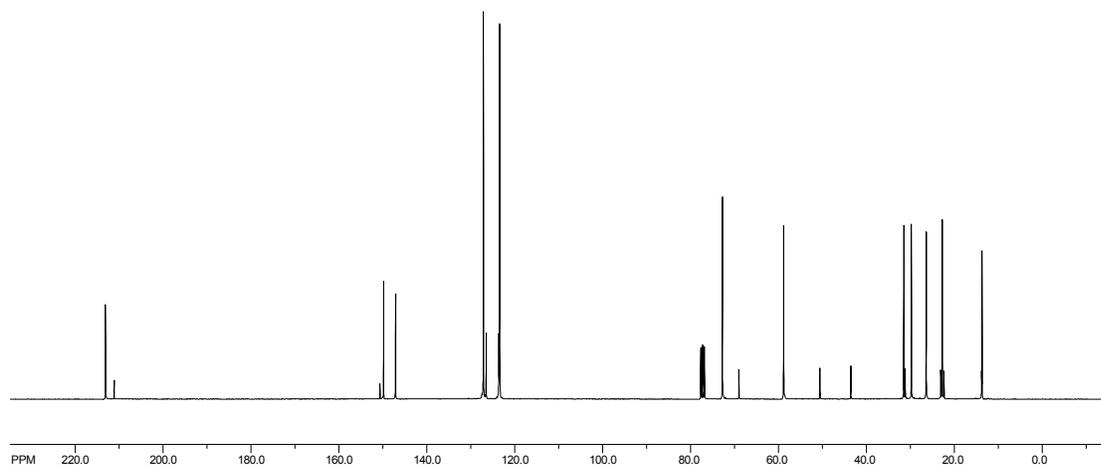
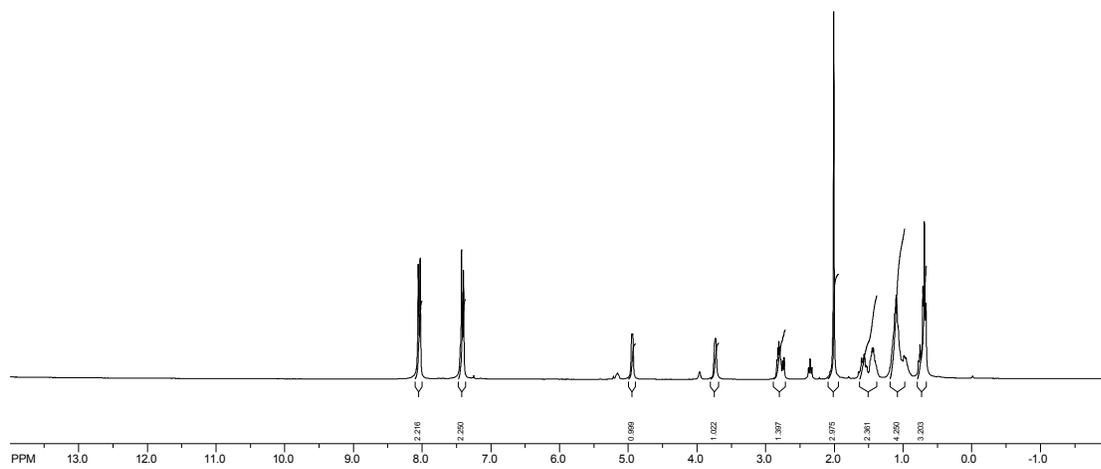


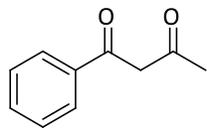
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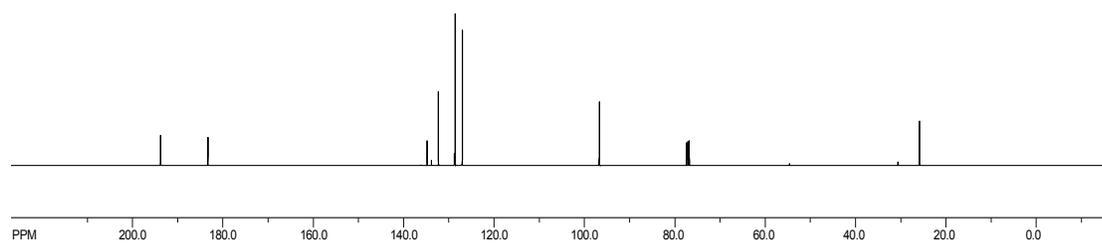
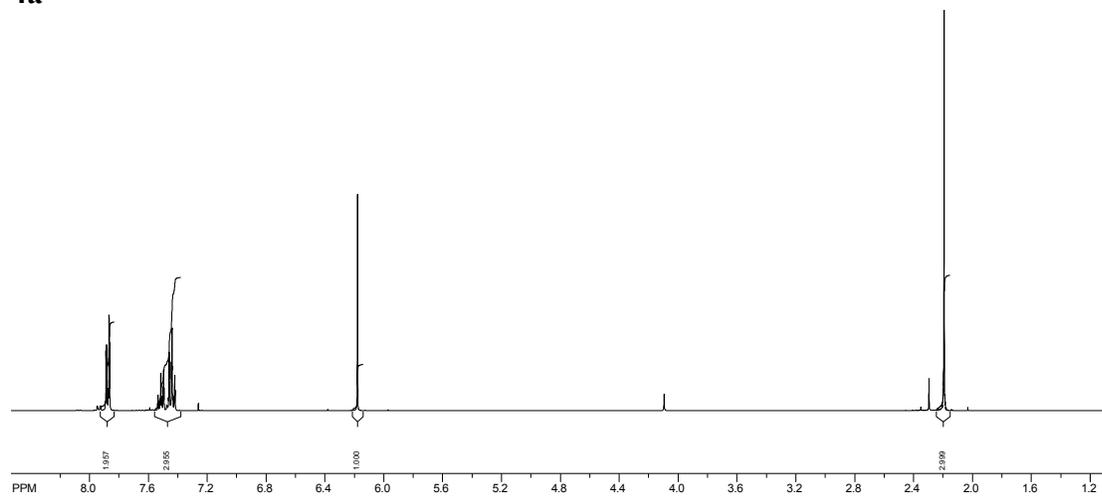


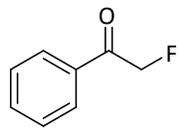
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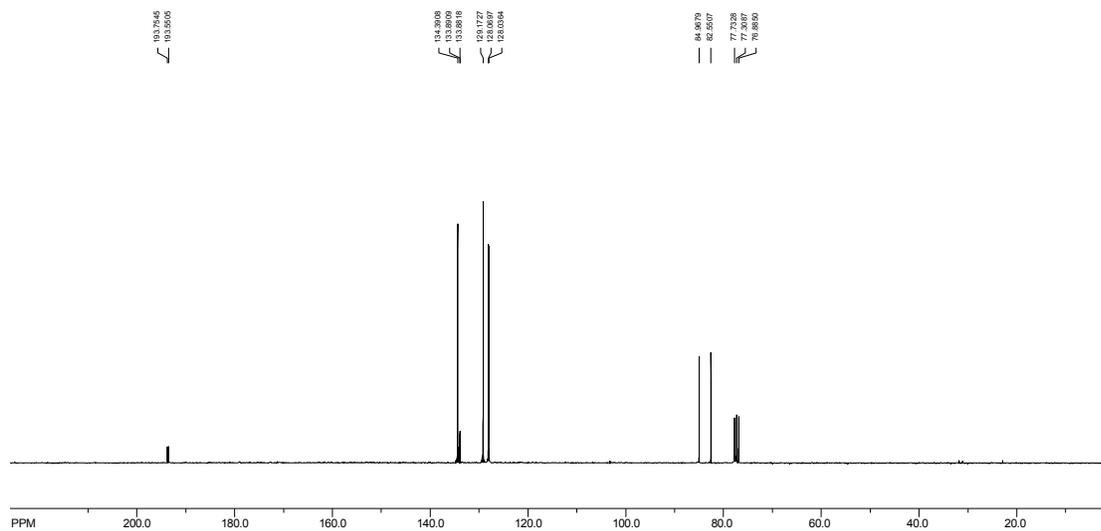
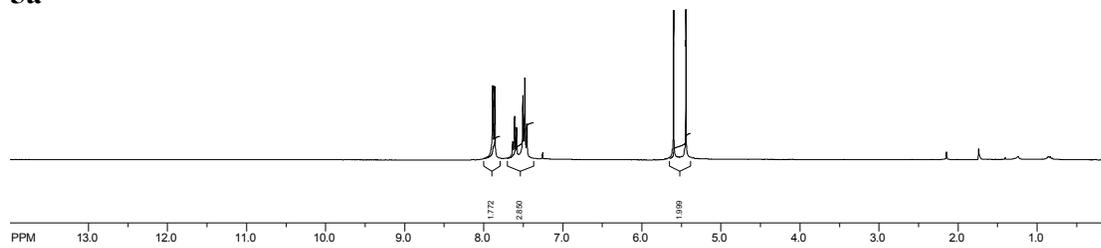


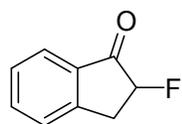
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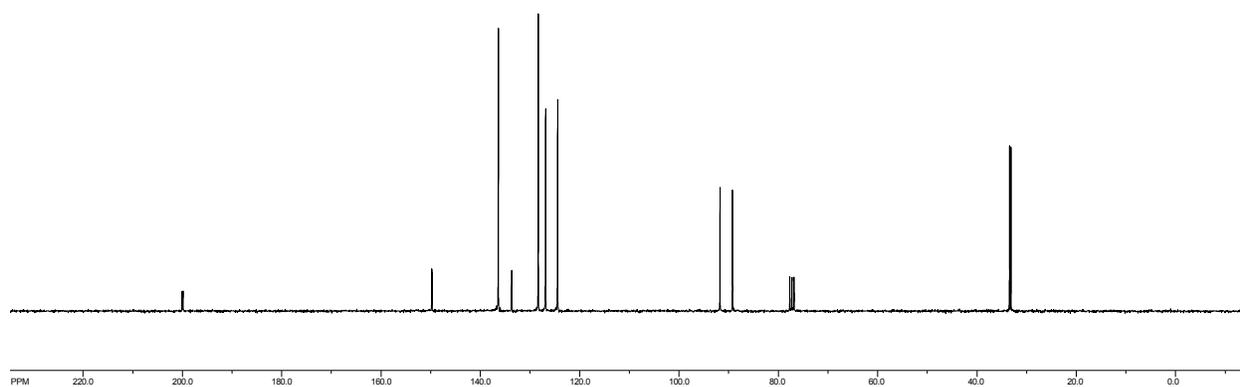
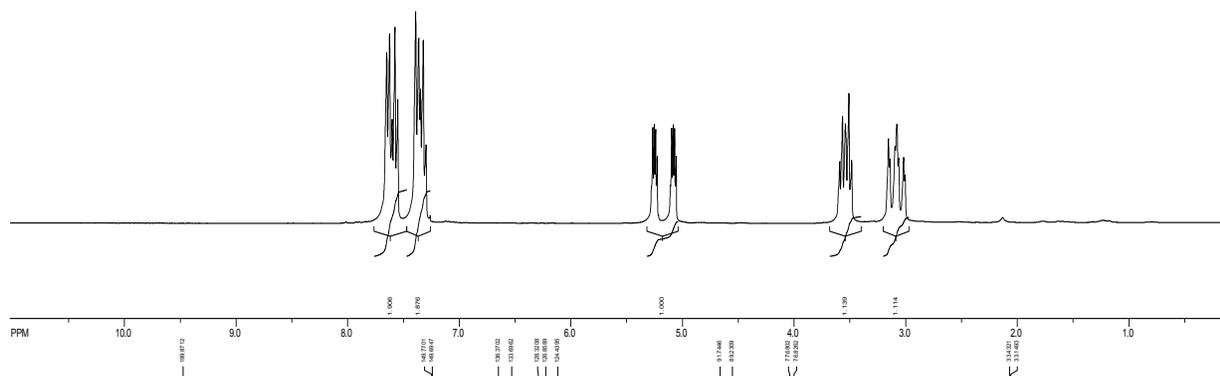


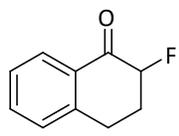
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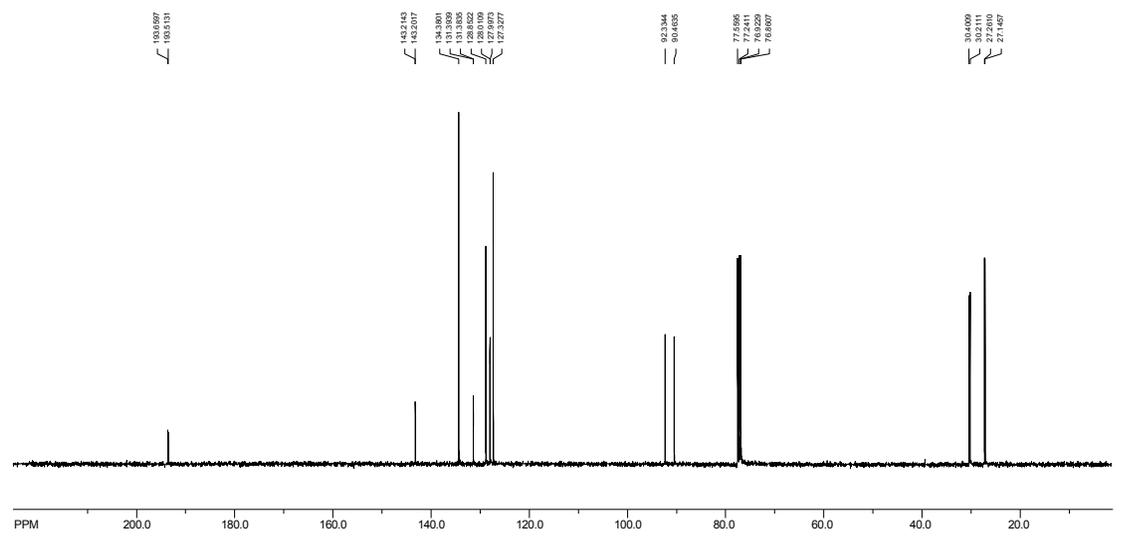
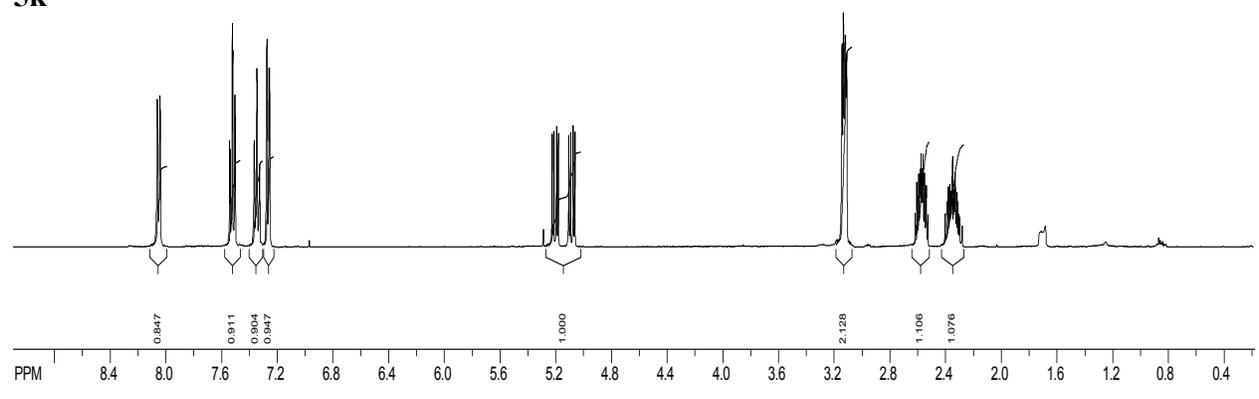


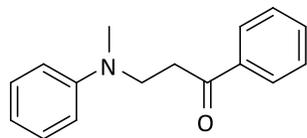
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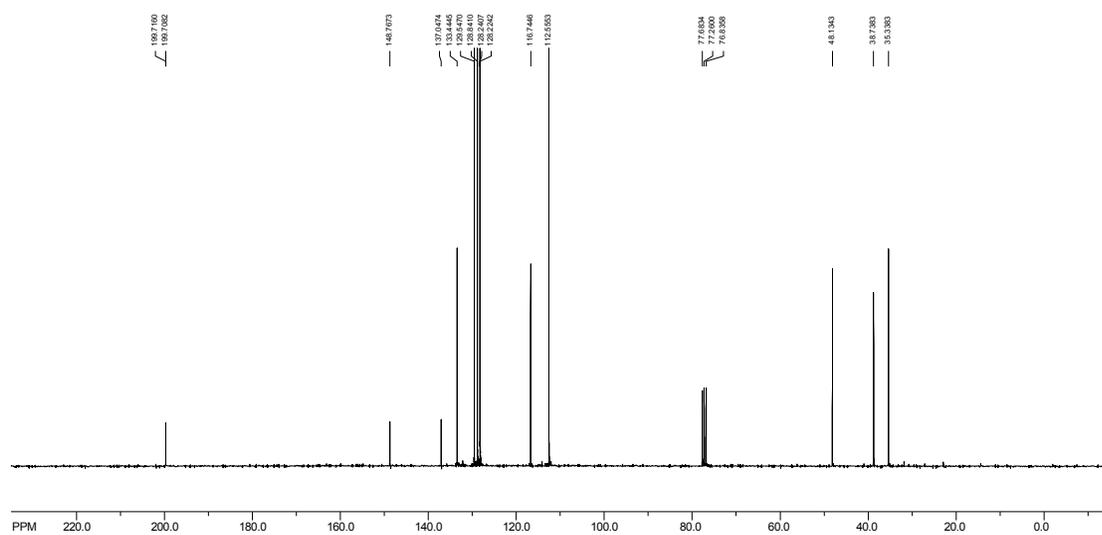
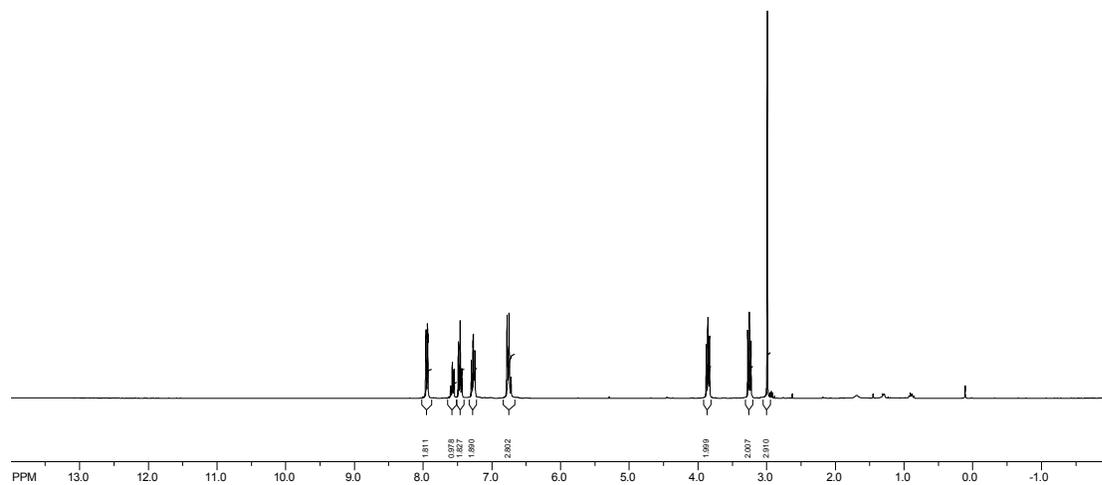


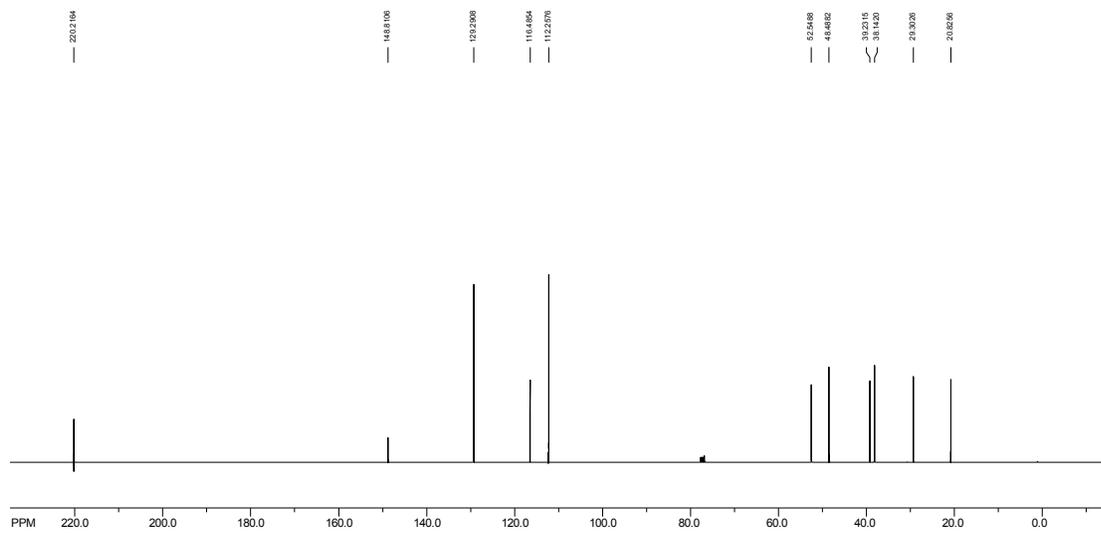
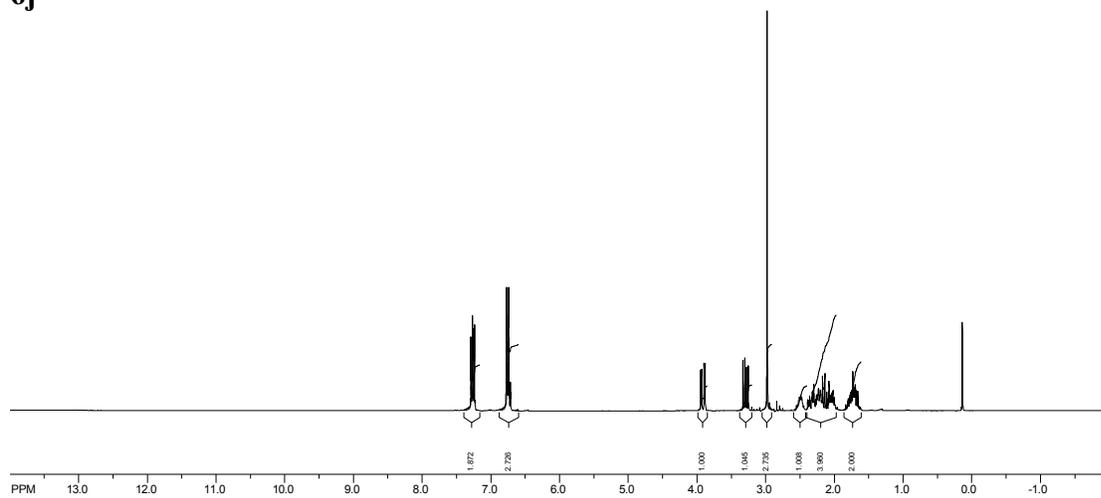
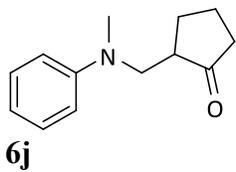
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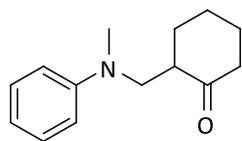




6a







6k

